



## Analyst

### Paper

# Analysis of soluble or titanium dioxide derived titanium levels in human whole blood: consensus from an inter-laboratory comparison

D. Koller <sup>a</sup>, P. Bramhall <sup>b</sup>, J. Devoy <sup>c</sup>, H. Goenaga-Infante <sup>d</sup>, C.F. Harrington <sup>e</sup>, S. Nuñez <sup>d</sup>, J. Morton <sup>f</sup>, E. Leese <sup>f</sup>, B. Sampson <sup>g</sup>, J. Rodgers and J. J. Powell <sup>a</sup>

### Supplementary Information for:

Received 00th January 20xx,  
Accepted 00th January 20xx

DOI: 10.1039/x0xx00000x

[www.rsc.org/](http://www.rsc.org/)

<sup>a</sup> Biomineral Research Group, Department of Veterinary Medicine, University of Cambridge, Madingley Road, Cambridge, CB3 0ES, UK and MRC Elsie Widdowson Laboratory (MRC-EWL), 120 Fulbourn Road, Cambridge, CB1 9NL, UK.  
Email: [jjp37@cam.ac.uk](mailto:jjp37@cam.ac.uk)

<sup>b</sup> University Hospital of Wales TRACE ELEMENT LABORATORY, Department of Medical Biochemistry and Immunology, Heath Park, Cardiff, CF14 4XW, UK.

<sup>c</sup> INRS, Unité de Génération d'atmosphères et de Chimie Analytique Toxicologique, Rue du Morvan CS60027, 54519 Vandoeuvre-lès-Nancy, France.

<sup>d</sup> LGC Limited, Inorganic Analysis, Queens Road Teddington, Middlesex, TW11 0LY, UK.

<sup>e</sup> SAS Trace Element Centre, Surrey Research Park, 15 Frederick Sanger Road, Guildford, Surrey, GU2 7YD, UK.

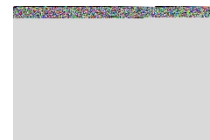
<sup>f</sup> Health and Safety Executive, Biological Monitoring, Harpur Hill, Buxton, SK17 9JN, UK.

<sup>g</sup> Charing Cross Hospital, SAS Trace Element Laboratory, Ground Floor Medical Oncology Block, Fulham Palace Road, London, W6 8RF, UK

† Footnotes relating to the title and/or authors should appear here.

Electronic Supplementary Information (ESI) available: [details of any supplementary information available should be included here]. See DOI: 10.1039/x0xx00000x

## Analyst



## Paper

Table S 1 Whole blood sample collection; number of vials and estimated collection volume

Date taken	Fasted. Number of vials	Non-fasted. Number of vials	Fasted. Volume (mL)	Non-fasted. Volume. (mL)
13/11/2015	7	-	63	-
16/11/2015	5	-	45	-
18/11/2015	5	5	45	45
27/11/2015	4	-	36	-
30/11/2015	10	-	90	-
01/12/2015	5	-	45	-
02/12/2015	-	5	-	45
	Total pooled	volume (mL)	Fasted 324	Non-fasted 90

Table S 2 Sample preparation and quality control for L1

## L1 – ICP-MS/MS

<b>Reagents</b>	Ammonia solution 0.2% (NH <sub>3</sub> ) supplied by Romil Ltd (Cambridge, UK), Triton® X-100 0.01% supplied by Fisher Scientific (Loughborough, UK), EDTA 0.01%. Sc, Ga, Y, Ge at 20ppb supplied by Inorganic Ventures; used as a universal diluent for samples and standards (1 in 15 dilution).
<b>Sample handling</b>	Whole blood samples were stored frozen at -20°C until analysis, thawed and equilibrated to laboratory temperature before sampling. Sample tubes were shaken gently to mix it thoroughly prior to opening.
<b>QC</b>	Custom Whole Blood control manufactured by UTAK Labs USA. Lot No. A5541 3 Levels 3, 15 & 30 µg/L
<b>Calibration type</b>	Custom Aqueous Calibration VWR Lot No. D13872
<b>Standards</b>	Commercial bespoke Calibrators from VWR
<b>Dilution</b>	Standards, samples and QCs materials were all diluted 1:15 fold with the diluent.
<b>Diluent, IS add.</b>	0.01%EDTA, 0.01%Tritonx100, 0.20% NH <sub>3</sub> with internal standards of Sc, Ga, Ge and Y at 20ppb (typically stick with Ga )
<b>Rinse/Wash solution</b>	1% NH <sub>3</sub> sol.
<b>Calibration verification</b>	N/A

Table S 3 Sample preparation and quality control for L2

L2 – ICP-MS/MS	
<b>Reagents</b>	Triton® X-100 supplied by Fisher Scientific (Loughborough, UK). Single standard TraceSELECT 1000 mg/L titanium and germanium were supplied by Fluka Analytical (Buchs, CH). Blank human blood was collected in house under Method development ethics. For nanoparticle spike a TiO <sub>2</sub> dispersion from Sigma (7000347) was used.
<b>Sample handling</b>	Whole blood samples were stored frozen at -20°C until analysis. Samples were allowed to reach room temperature while mixed on a roller mixer before analysis. Seronorm TM Trace Elements in Whole Blood level 1, Lot 1103128 (add. approx. value: Ti=14µg/L) and Level 2, Lot 1406264 (add. approx. value: Ti=10.3µg/L) from Sero AS (Billingstad, Norway). Additional QC were spiked Seronorm standards and 3 spiked samples, which were selected at random. All spiked QCs were spiked at 10µg/L and analysed once per batch of 20 samples.
<b>Calibration type</b>	External matrix matched calibration
<b>Standards</b>	Using the 1000 mg/L single standard of titanium a 200 µg/L single standard stock solutions was prepared. Working standard levels: 1, 5, 10, 20, 30, 50
<b>Dilution</b>	Standards, samples and QCs materials were all diluted 20 fold with the diluent. All standards were matrix matched with blank human blood.
<b>Diluent, IS add.</b>	Diluent containing 5 µg/L Ge internal standard: 0.005% v/v Triton® X-100
<b>Rinse/Wash solution</b>	0.005% v/v Triton® X-100
<b>Calibration verification</b>	A 10 µg/L calibration standard is analysed after each set of 10 samples.

Table S 4 Sample preparation and quality control for L3

L3 – SQ-ICP-MS	
<b>Reagents</b>	Ammonia solution (NH <sub>3</sub> ) supplied by Romil Ltd (Cambridge, UK), Triton® X-100 supplied by Fisher Scientific (Loughborough, UK). Single standard BDH ICP-MS grade 1000 mg/L titanium and germanium were supplied from VWR (Lutterworth, UK). Blank horse blood supplied from TCS Biosciences (Buckingham, UK).
<b>Sample handling</b>	Whole blood samples were frozen at -20°C upon receipt until analysis. Samples were allowed to reach room temperature, then mixed on a roller mixer before analysis.
<b>QC</b>	Quality control material was Seronorm TM Trace Elements in Whole Blood level 1, Lot 1103128 from Sero AS (Billingstad, Norway). No certified range, mean concentration of 14 µg/L.
<b>Calibration type</b>	External matrix matched calibration
<b>Standards</b>	Using the 1000 mg/L single standard of titanium a 10 mg/L and 1 mg/L single standard stock solutions were prepared. Working standard levels: 0.5, 2.5, 5, 25, 50, 125, 250, 500 and 1250
<b>Dilution</b>	Standards, samples and QCs materials were all diluted 50 fold with an alkaline diluent. All standards were matrix matched with blank horse blood.
<b>Diluent, IS add.</b>	Diluent containing 800 ng/L Ge internal standard: 1% v/v ammonia and 0.1% v/v Triton® X-100
<b>Rinse/Wash solution</b>	0.025% v/v ammonia solution and 0.05% v/v Triton® X-100.
<b>Calibration verification</b>	A 50 µg/L calibration standard is analysed at the beginning and end of the analytical run in addition to every 10 samples

Table S 5 Sample preparation and quality control for L4

L4 – HR ICP-MS	
<b>Reagents</b>	TMAH (electronic grade from Alfa Aesar, Ga and Ge internal standard 1000ppm from Alfa, TritonX-100 from Romil. MilliQ system used for water. Equine horse serum for calibration from Selbourne biologicals with concentration assigned by repeated standard addition calibration.
<b>Sample handling</b>	Samples stored at -20°C until analysed. Samples mixed by repeated inversion and by vortex mixing before analysis.
<b>QC</b>	Seronorm L1 batch 140263 (10.3 µg/L) L2 batch 1103129 (15 µg/L) and L3 batch 1112691 (12.8 µg/L) and ClinCheck (Recipe, Germany) batch 445 with user assigned concentrations of 0.78 (L1), 0.809 (L2) and 1.01 (L3) µg/L.
<b>Calibration type</b>	Matrix matched calibration
<b>Standards</b>	Standards prepared in matrix matched serum or whole blood with base concentrations assigned by standard addition analysis
<b>Dilution</b>	Samples, standards and Qc diluted 1:20 in diluent as below
<b>Diluent, IS add.</b>	Diluent contains 0.5% TMAH and 0.005% Triton with Ge and Ga added to 5 µg/L.

<b>Rinse/Wash solution</b>	As sample diluent without IS added.
<b>Calibration verification</b>	A 50 µg/L calibration standard is analysed at the beginning and end of the analytical run in addition to every 10 samples

Table S 6 Sample preparation and quality control for L5

L5 – ICP-OES	
<b>Reagents</b>	Single element stock solutions (1000 mg/L) of Ti and Y traceable to NIST (Certipur, Merck) were used to prepare calibration standards. All solutions were prepared in dilute HNO <sub>3</sub> (0.5 % v/v) prepared using trace element free concentrated acid (Primarplus, Trace Element Grade, Fisher) and reverse osmosis deionised water >18.2 MΩ cm resistivity (Millipore, Synergy System). Prior to the analysis of patient samples, a solution of Zn (2 mg/L) traceable to NIST (Certipur, Merck) was used to check the system performance, as recommended by the instrument manufacturer.
<b>Sample handling</b>	Whole blood samples were frozen at -20 °C upon receipt at the SAS Trace Element Laboratory, Guildford until analysis. Samples were allowed to reach room temperature, then mixed on a roller mixer before analysis.
<b>QC</b>	The IQC materials used were prepared in house and values assigned using high-resolution ICP-MS in HR mode (thanks to Dominique Debellis, Thermo Scientific). The IQC materials were prepared from equine whole blood in EDTA (TCS Biosciences Ltd.) and bovine calf serum (Selbourne Biological Services). Commercially available serum and blood reference materials (Seronom L2, Sero) and a candidate certified reference material (LGC8276, LGC) were also used.
<b>Calibration type</b>	External calibration. The instrument software generated a linear regression fit for the data using an unweighted linear equation and provided values for the slope, intercept and correlation coefficient.
<b>Standards</b>	Intermediate Ti standards with concentrations of 0, 0.5, 1.0, 2.0, 4.0, 8.0 and 16.0 µg/L (microgram per litre) were prepared in reverse osmosis water by dilution from an intermediate stock standard (10 mg/L). The calibration standards were made from these by diluting 1:5 with the diluent used for the samples, so that the internal standard concentration was the same in both standards and samples. The blood and serum IQC specimens were also diluted 1:5 with the same diluent.
<b>Dilution</b>	1: 5 in 0.5% v/v HNO <sub>3</sub> with Y as internal standard at 8 µg/L (diluent); the samples were centrifuged to remove precipitate
<b>Diluent, IS add.</b>	0.5% v/v HNO <sub>3</sub> with Y as internal standard at 8 µg/L
<b>Rinse/Wash solution</b>	2% v/v nitric acid and 0.5% v/v Triton® X-100
<b>Calibration verification</b>	The 5th calibration standard (8µg/L, dil.1:5) was analysed at the beginning of the analytical run in addition to every 5 samples

Table S 7 Sample preparation and quality control for L6

L6 – SQ ICP-MS	
<b>Reagents</b>	Certified water was used (Trace Metals 3, RTC, QC1448). All chemicals used in the study were of analytical grade or higher. Nitric acid was used to prepare 2.0 % HNO <sub>3</sub> (v/v) with ultrapure water. All single element stock solutions (1000 mg/L) was delivered by SCP Science and certified for purity and concentration.
<b>Sample handling</b>	The sample was digested in a 50 mL reactor. When digestion finished the sample was transferred in a 50 mL tube. The reactor was washed 3 times with about 10 mL of ultra-pure water. Then, the volume was reduced to 500 µL on a hotplate at 90°C, in order to reduce HF concentration. Finally, the sample was recuperated in a 50 mL tube with 25 mL of ultrapure water.
<b>Digestion</b>	2g od sample were digested with 8 mL HNO <sub>3</sub> and 2 mL HF. Ambient temperature to 170°C in 10 min, then 25 min at 170°C. Finally, 45 min from 170 °C to ambient temperature.
<b>Digestion system</b>	Microwave Assisted Reaction System (MARS) Express
<b>QC</b>	A certified water was used (Trace Metals 3) and diluted at 0.124 µg/L and 1.24 µg/L. About 20 % of deviation were accepted. All the Blank reactors were analysed in order to verify the absence of contamination. Seronom TM Trace Elements in Whole Blood level 1, Lot 1103128 (add.approx. value: Ti = 14 µg/L) was also used.
<b>Calibration type</b>	external calibration
<b>Standards</b>	Titanium standard solutions for ICP–MS calibration were prepared (at concentration levels of 50 to 10000 ng/L) by diluting a 10 g/L titanium standard stock solution with 2% (v/v) HNO <sub>3</sub> .
<b>Dilution</b>	2g in 25mL
<b>Diluent, IS add.</b>	An internal standard solution containing 100 µg/L of Ge was prepared by diluting a 1000 mg/L internal standard stock solution with 2% (v/v) HNO <sub>3</sub> . The internal standard was added to all samples and standard solutions.
<b>Rinse/Wash solution</b>	A 2 % (v/v) HNO <sub>3</sub> was used between two samples, to rinse injection system. All the reactors were soaked in a 20 % (v/v) HNO <sub>3</sub> . Then a blank procedure was run in the same conditions than the samples.
<b>Calibration verification</b>	A certified water was used (Trace Metals 3) and diluted at 0.124 µg/L and 1.24 µg/L. About 20 % of deviation were accepted.

Table S 8 Sample preparation and quality control for L7

L7 – ICP-MS/MS	
<b>Reagents</b>	Single standard 984 mg/kg titanium and scandium were supplied by VHG (Manchester, UK). For nanoparticle spike a $\text{TiO}_2$ dispersion from Sigma (7000347) was used.
<b>Sample handling</b>	Whole blood samples were stored frozen at $-80^\circ\text{C}$ until analysis, thawed and equilibrated to laboratory temperature before sampling. Sample tubes were shaken gently to mix it thoroughly prior to opening.
<b>Digestion</b>	Approximately 0.3 g of sample was accurately weighed into PTFE microwave digestion vessels, followed by the addition of 0.6 g $\text{HNO}_3$ , 0.12 g $\text{H}_2\text{O}_2$ and 0.015 g HF. After digestion, the digest solutions were made up to 3 g with ultrapure water ( $>18.2 \text{ M}\Omega\text{cm}^{-1}$ , Elga Maxima, UK) and analysed without further dilution
<b>Digestion system</b>	Ethos EZ microwave system with micro-sampling inserts (Milestone, Sorisole, Italy)
<b>QC</b>	Seronorm TM Trace Elements in Whole Blood level 2, Lot 1406264 and 1103129 (add. approx. value: $\text{Ti}=10.3 \mu\text{g/L}$ and $\text{Ti}=15 \mu\text{g/L}$ ) from Sero AS (Billingstad, Norway). In addition Seronorm QC were spiked with Ti ionic and nanoparticles to check for recovery.
<b>Calibration type</b>	External calibration
<b>Standards</b>	Using the 984 mg/kg single standard of titanium a 94 mg/kg single standard stock solutions was prepared in water. From this a second dilution was prepared at $954 \mu\text{g/kg}$ . From this a third dilution was prepared at a concentration of $9 \mu\text{g/kg}$ in 20% $\text{HNO}_3$ Ti stock, which then was used to prepare standards at 0.1, 0.5, 1, 2, and $5 \mu\text{g/kg}$ in 20% $\text{HNO}_3$ .
<b>Dilution</b>	The digests were analysed directly by ICP-MS/MS with external calibration using $1 \mu\text{g/kg}$ Sc as internal standard. Dilution: 0.3g in 3g.
<b>Diluent, IS add.</b>	After the digestion, samples were diluted up to 3 g with ultrapure water ( $> 18.2 \text{ M}\Omega\text{cm}^{-1}$ , Elga, Maxima, UK) and 0.1 g of Sc was added as internal standard to have a final concentration of $1 \mu\text{g/kg}$ Sc
<b>Rinse/Wash solution</b>	5% v/v $\text{HNO}_3$
<b>Calibration verification</b>	A $1 \mu\text{g/kg}$ calibration standard is analysed after each set of 5 samples and the calibration curve was measured at the beginning and at the end of the run

Table S 9 Instrument settings for Labs 1-7