

## **Electronic Supplementary Information (ESI†)**

**Real time detection of the nerve agent simulant diethylchlorophosphate by non fluorophoric small molecules generating cyclization induced fluorogenic response**

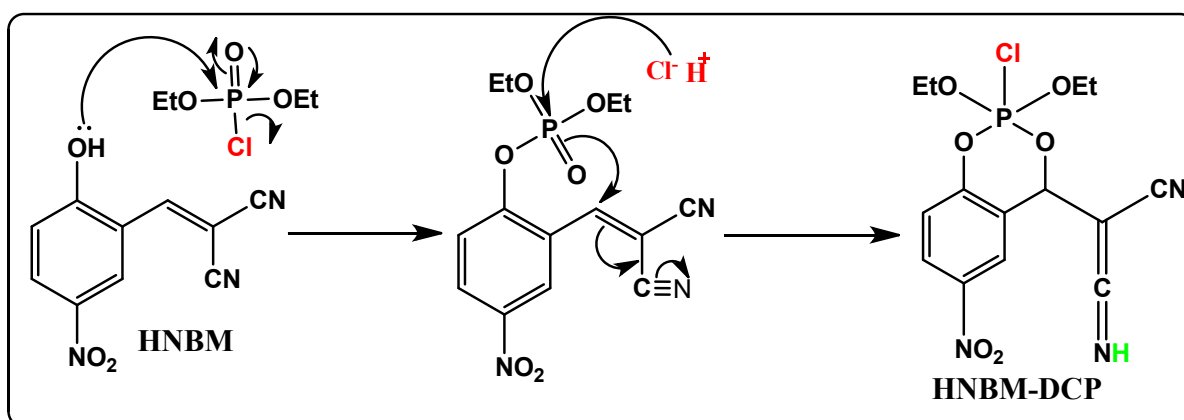
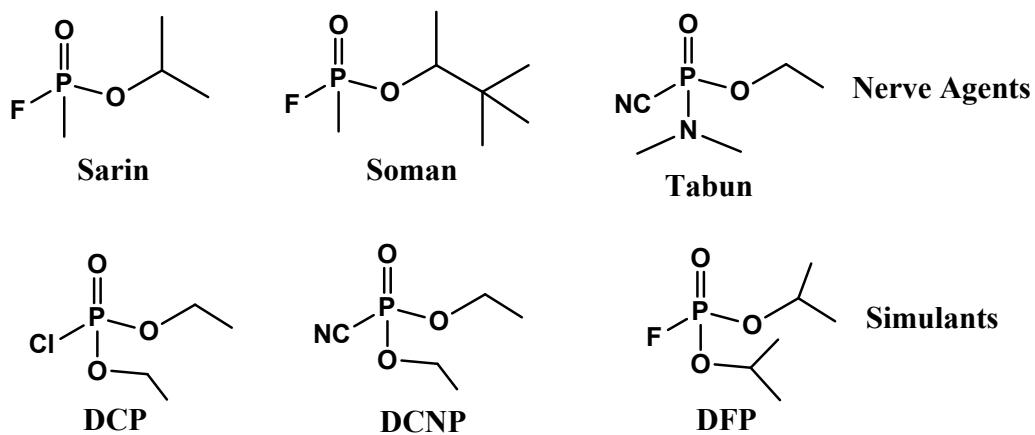
**Syed Samim Ali,<sup>a</sup> Ankita Gangopadhyay,<sup>a</sup> Ajoy Kumar Pramanik,<sup>a</sup> Sandip Kumar Samanta,<sup>a</sup>  
Uday Narayan Guria,<sup>a</sup> Srimanta Manna<sup>a</sup> and Ajit Kumar Mahapatra<sup>\*a</sup>**

<sup>a</sup>Department of Chemistry, Indian Institute of Engineering Science and Technology, Shibpur,  
Howrah – 711103, India.

\*Corresponding author: Tel.: +91 33 2668 4561; fax: +91 33 26684564;

E-mail: mahapatra574@gmail.com

**Chemical structure of nerve agents and Simulants:**



**Fig. S1** Plausible Mechanism of DCP binding

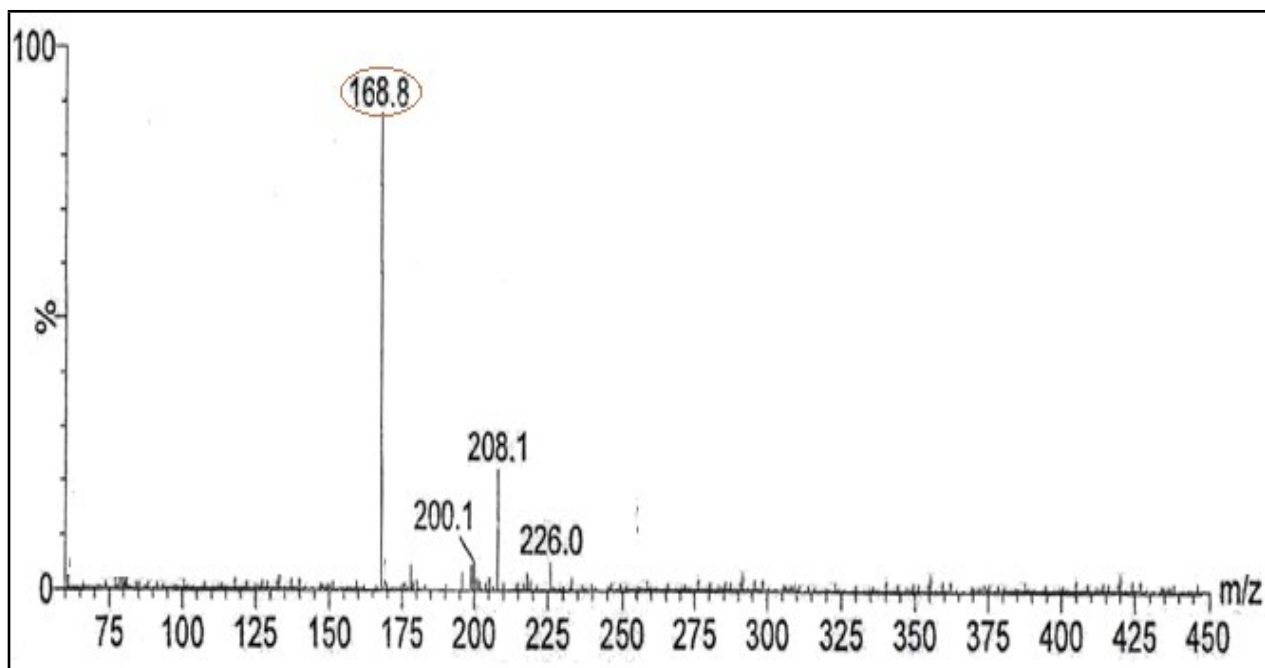


Fig. S2 LCMS of compound 2a

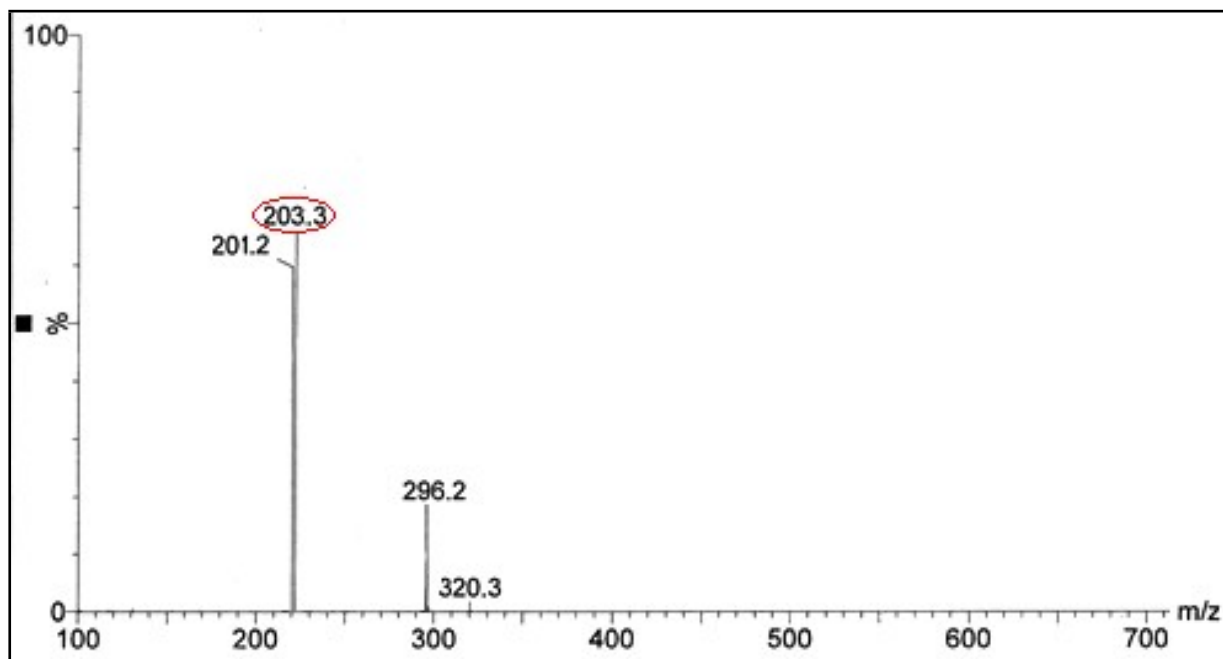


Fig. S3 LCMS of compound 2b

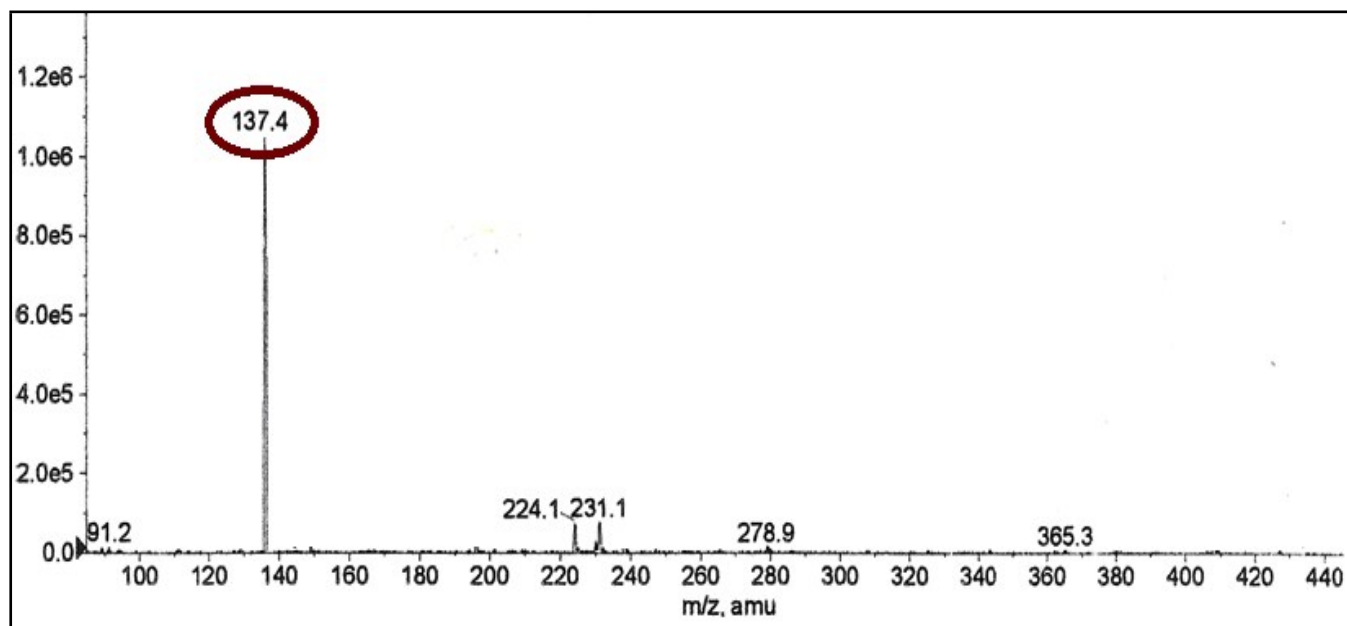


Fig. S4 LCMS of compound 2c

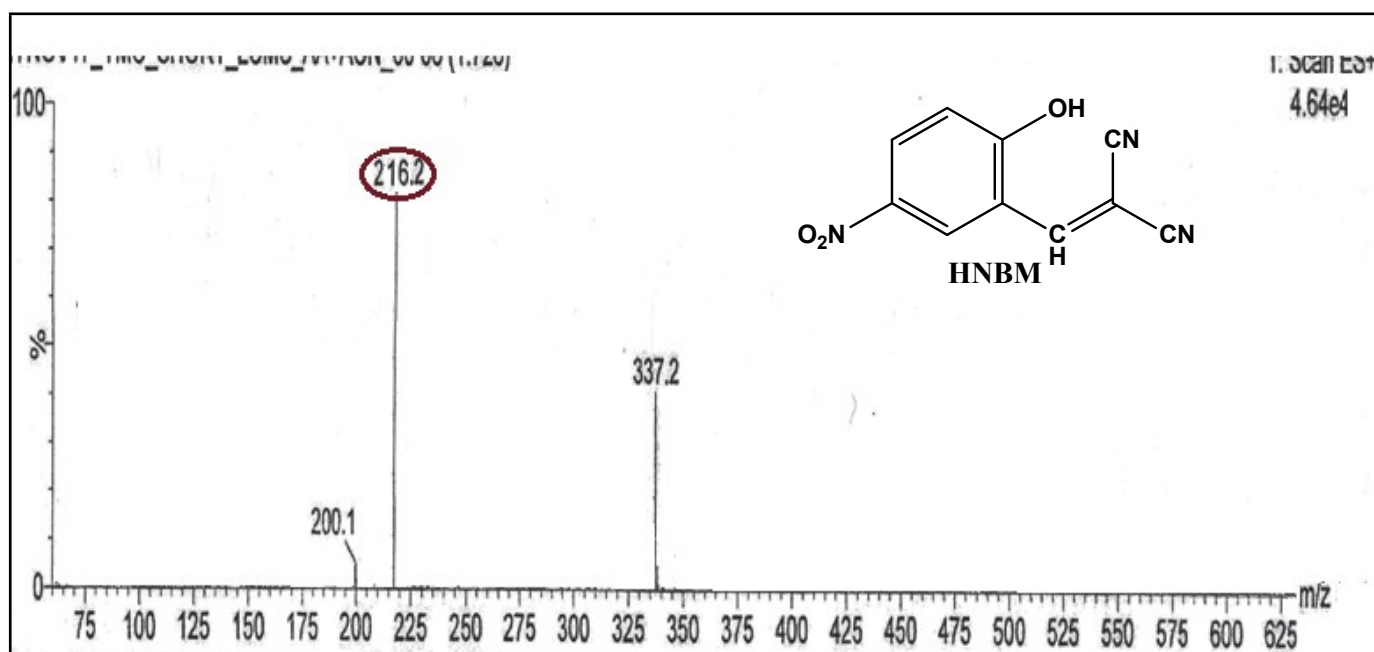


Fig. S5 LCMS of compound HNBM

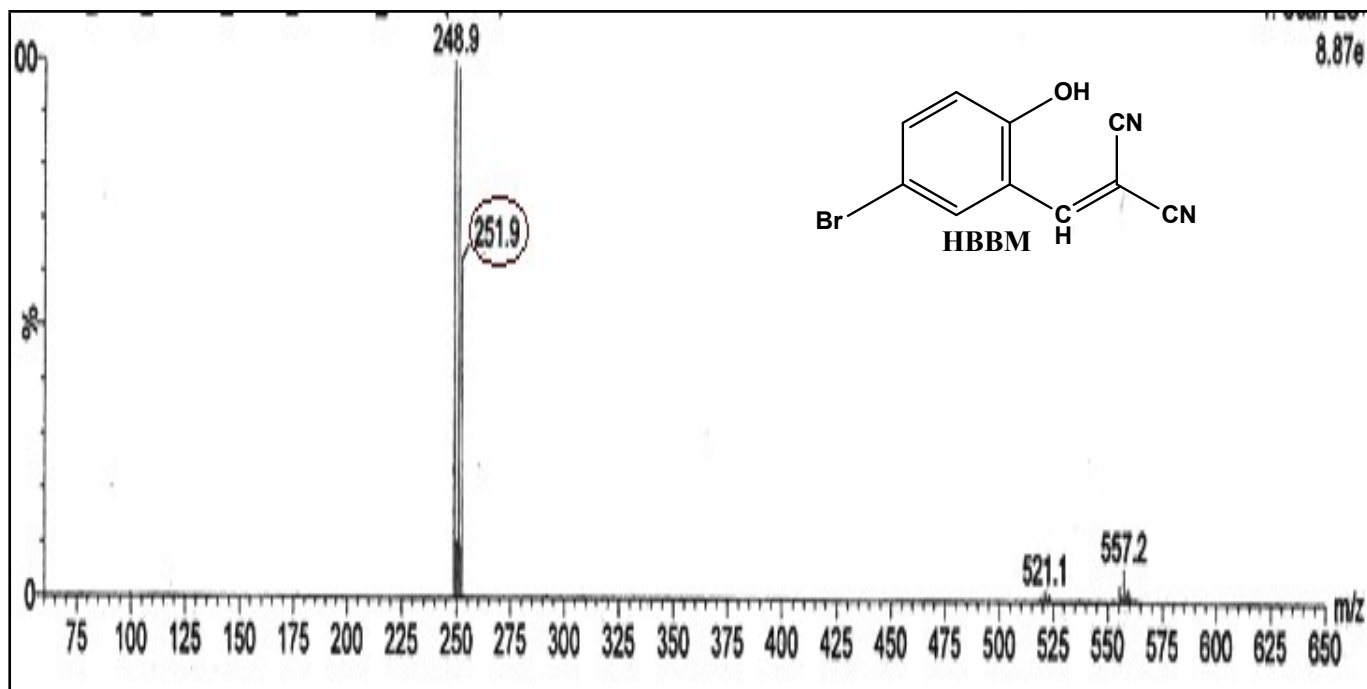


Fig. S6 LCMS of compound HBBM

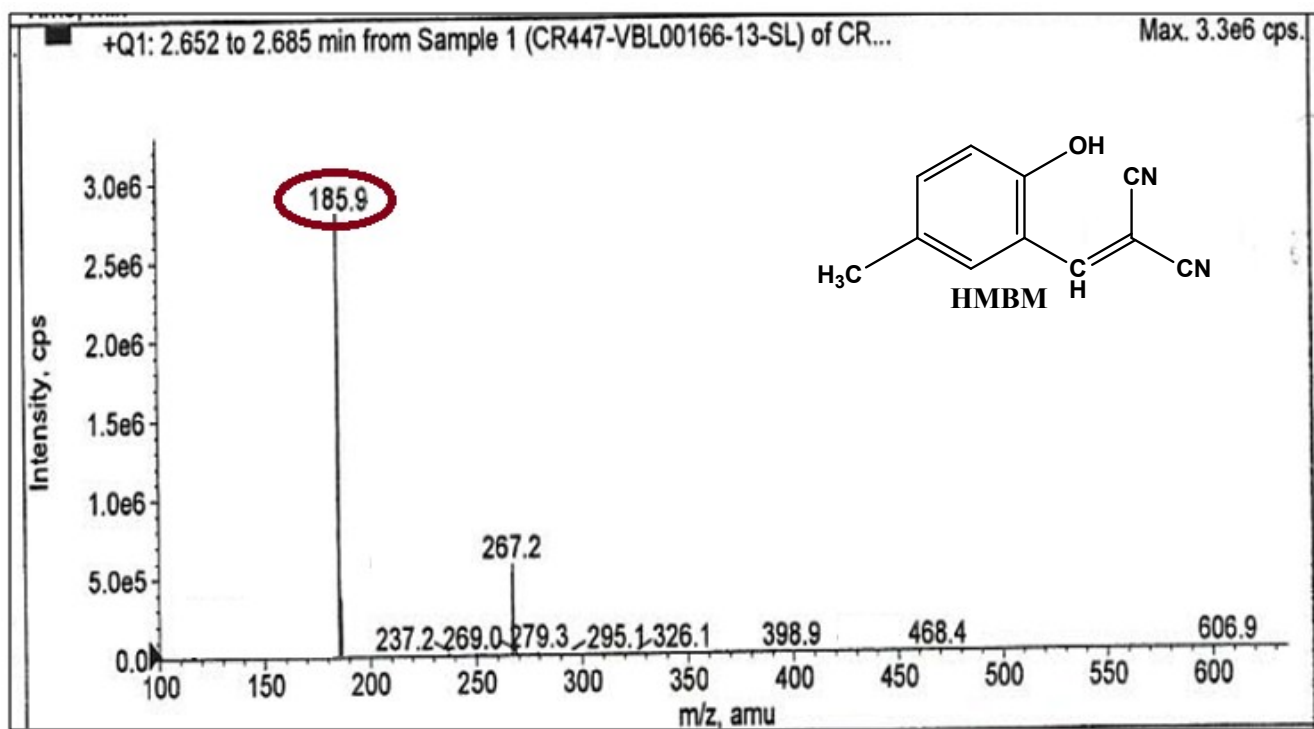


Fig. S7 LCMS of compound HMBM

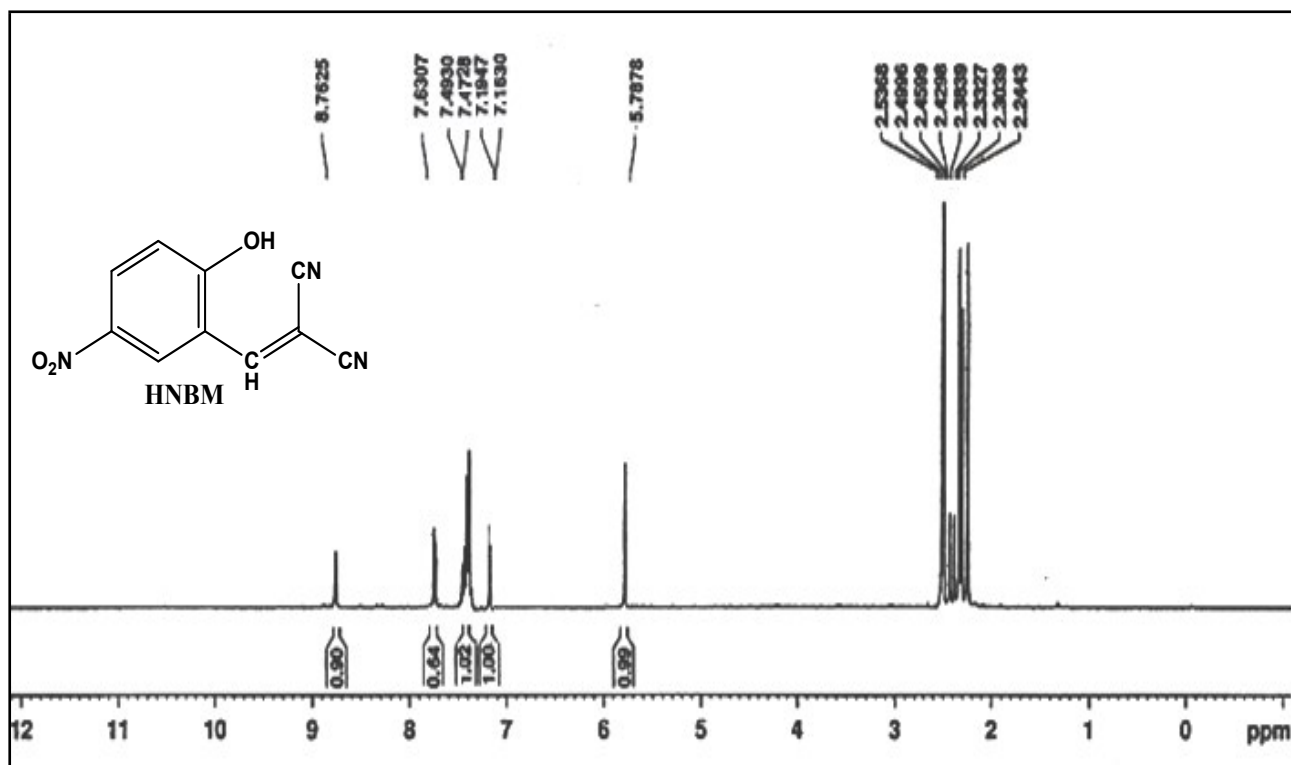


Fig. S8 <sup>1</sup>H NMR of HNBM in (d<sub>6</sub>-DMSO).

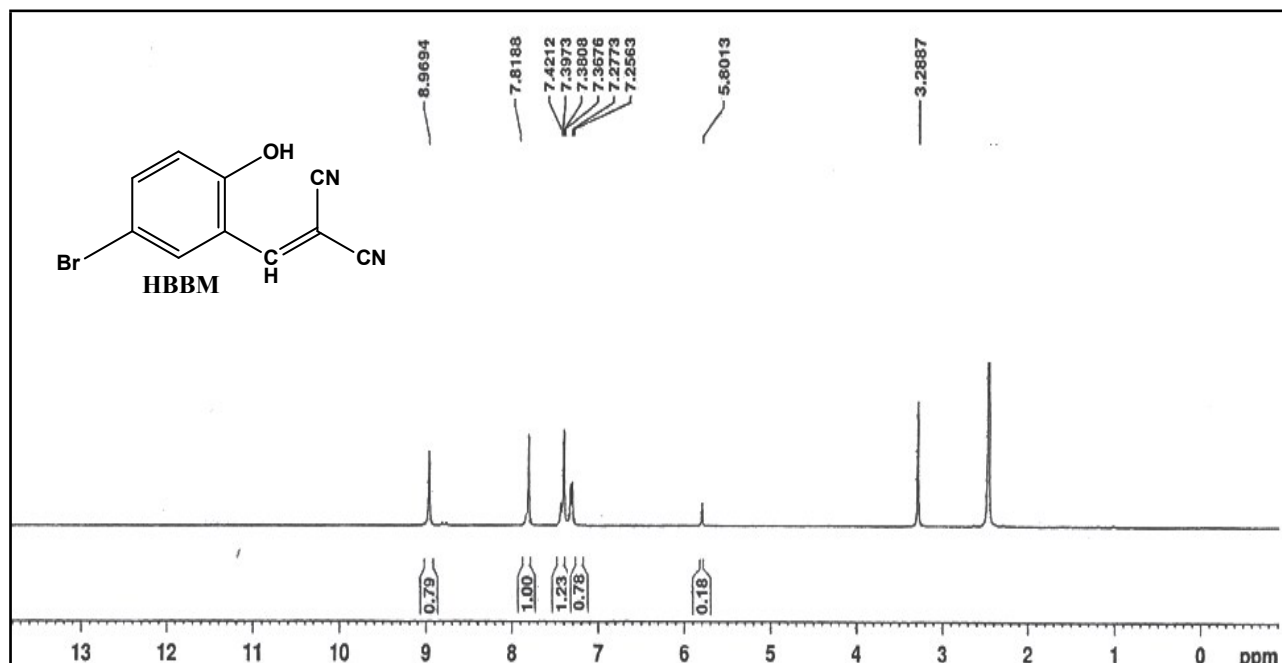


Fig. S9 <sup>1</sup>H NMR of HBBM in (d<sub>6</sub>-DMSO).

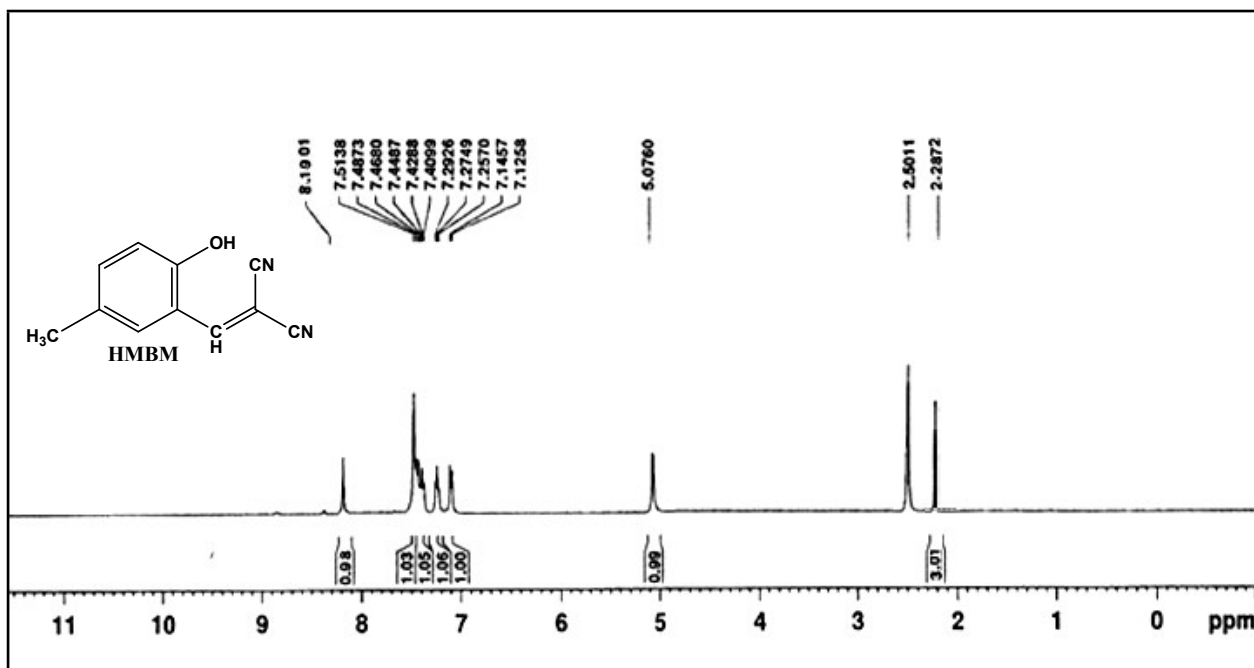


Fig. S10  $^1\text{H}$  NMR of HMBM in (d<sub>6</sub>-DMSO).

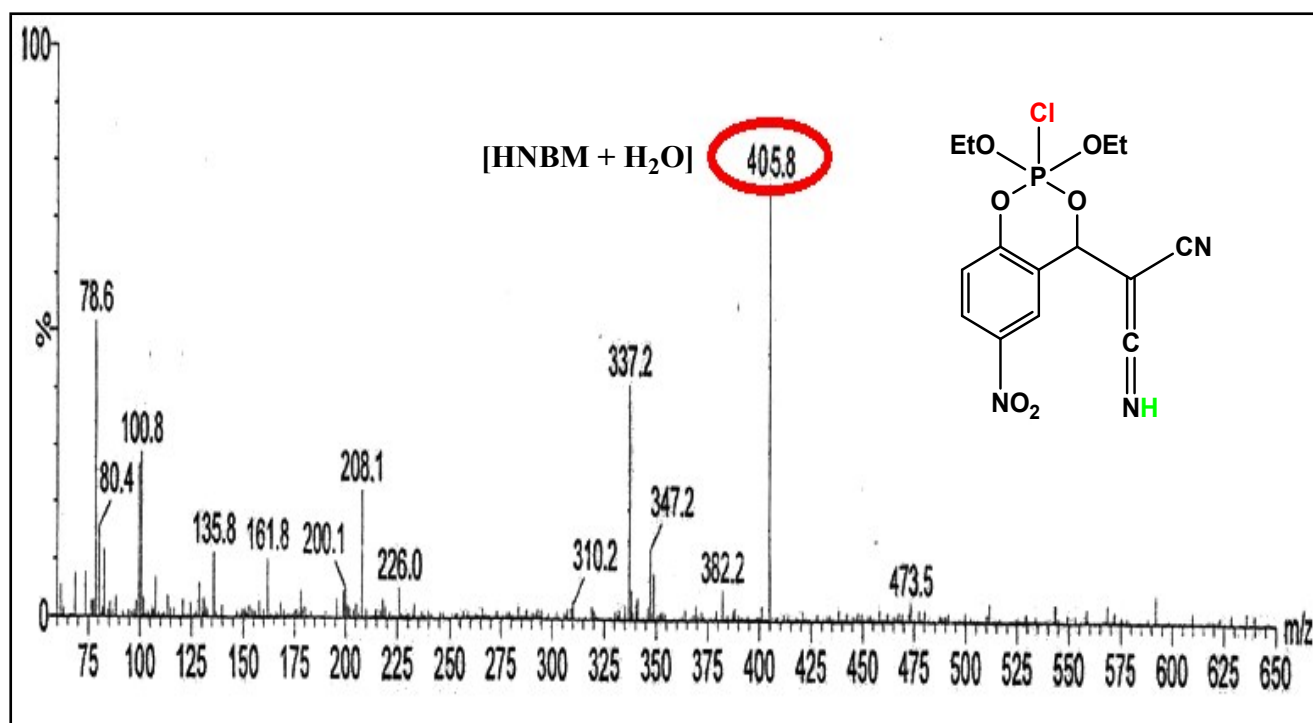


Fig. S11 LCMS spectra of HNBM-DCP Complex.

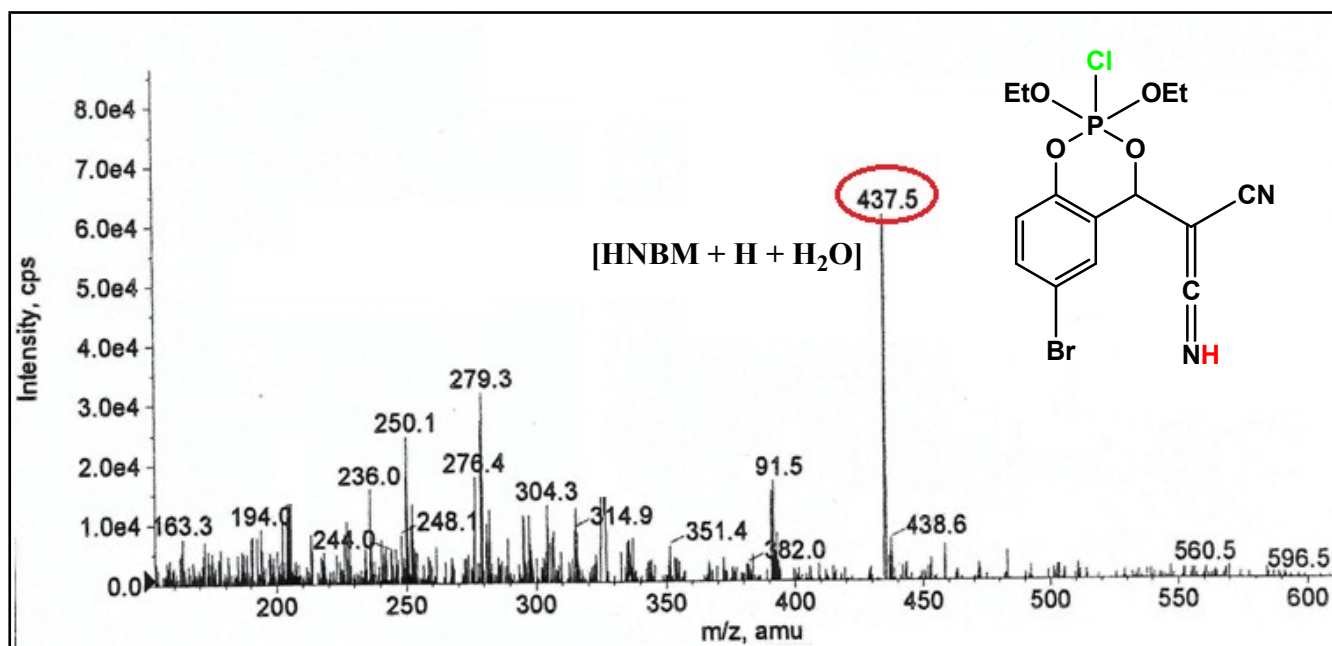


Fig. S12 LCMS spectra of HBBM-DCP Complex .

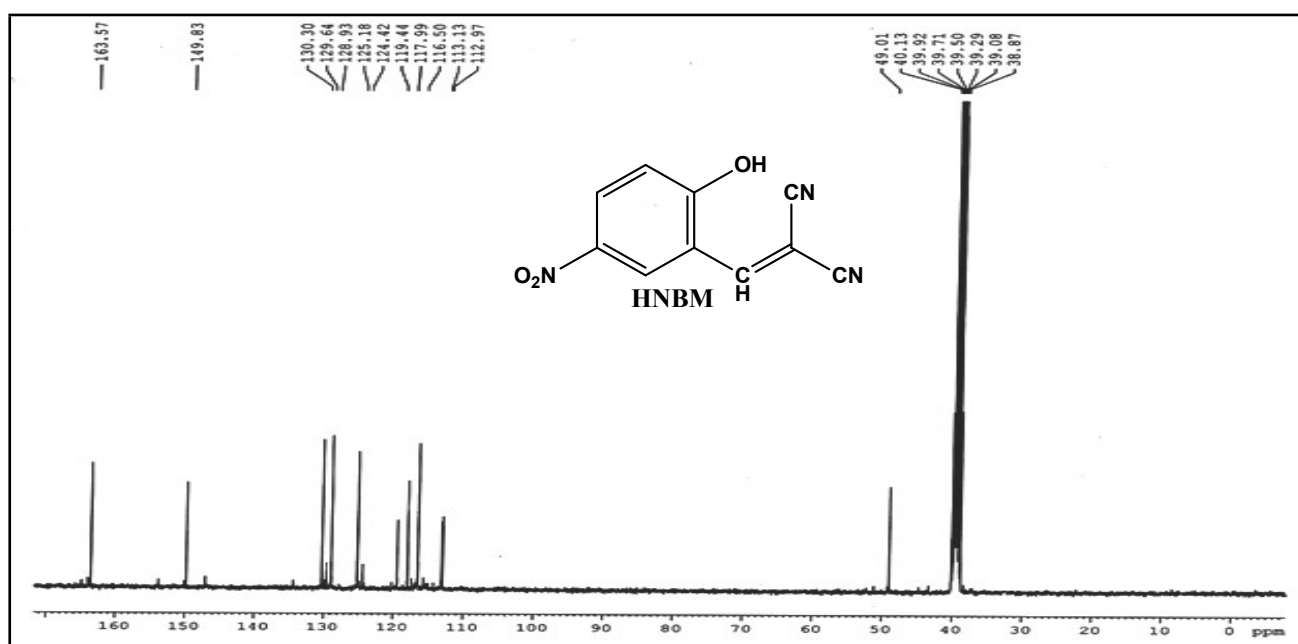


Fig. S13 <sup>13</sup>C NMR spectra of HNBM.



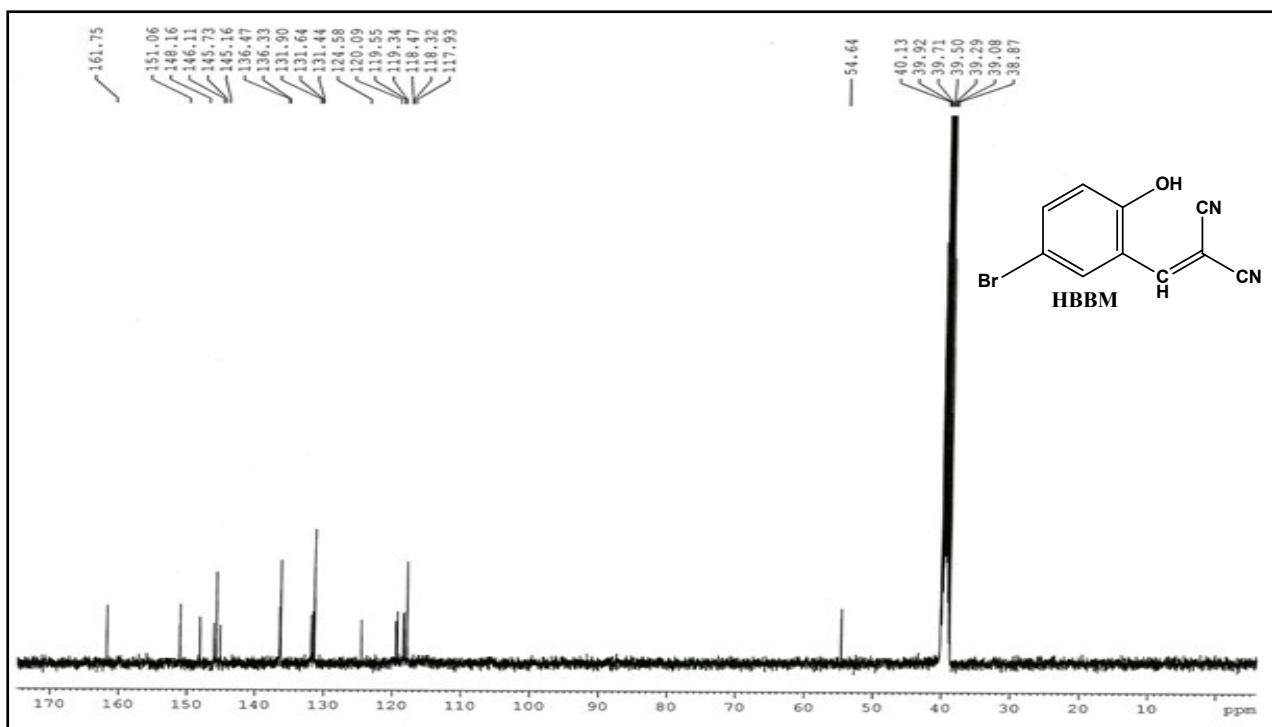


Fig. S14  $\text{C}^{13}$  NMR spectra of HBBM.

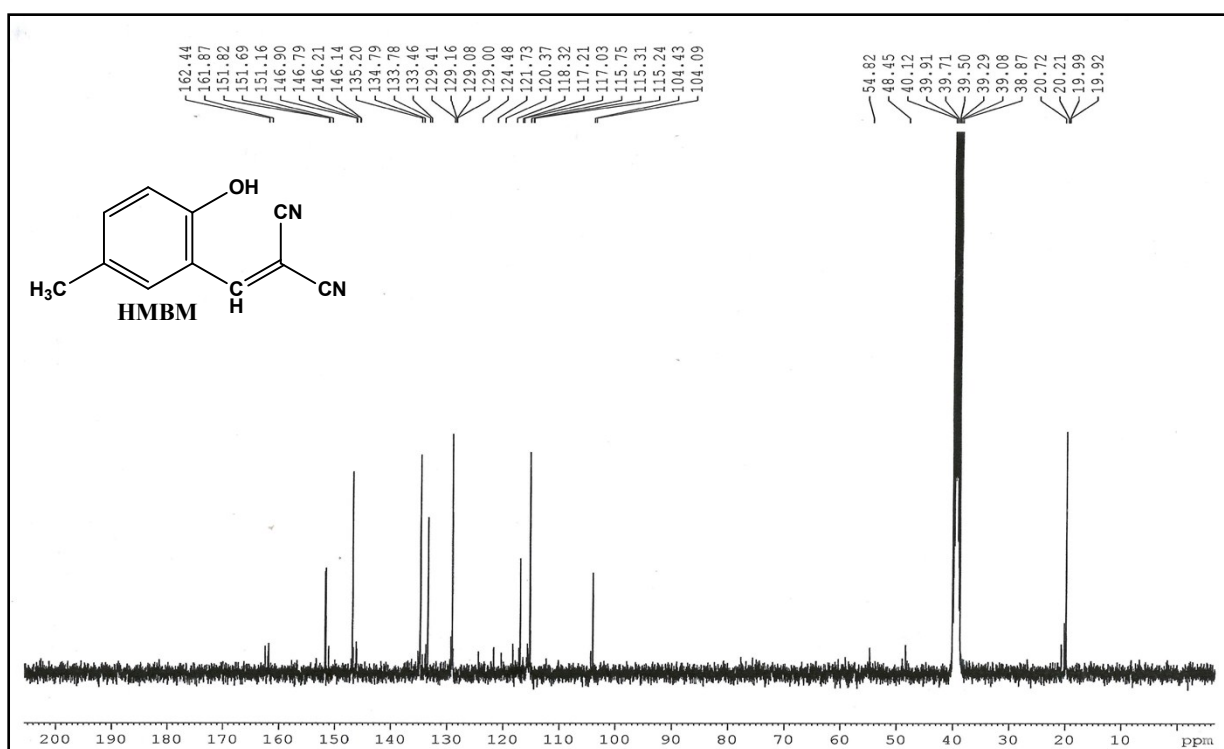
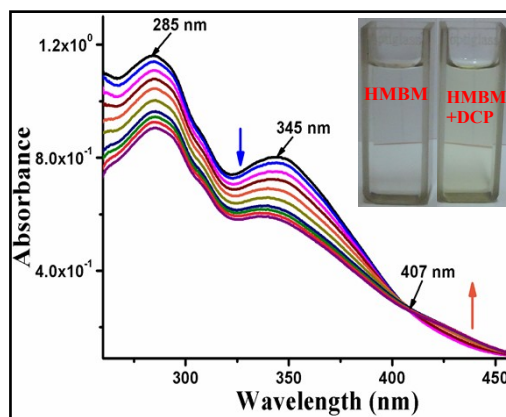
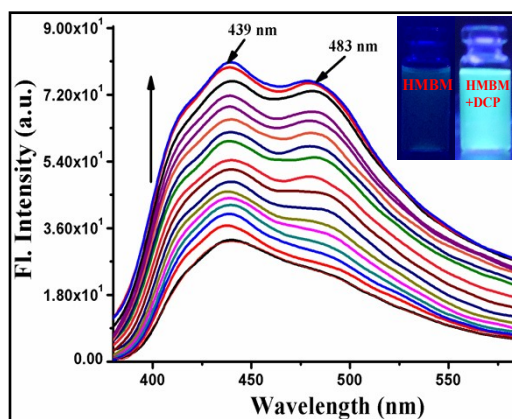


Fig. S15  $\text{C}^{13}$  NMR spectra of HMBM.



**Fig. S16** Absorption spectra of **HMBM** (2  $\mu\text{M}$ ) in acetonitrile–water (7:3) upon addition of increasing amount of DCP (40  $\mu\text{M}$ ).



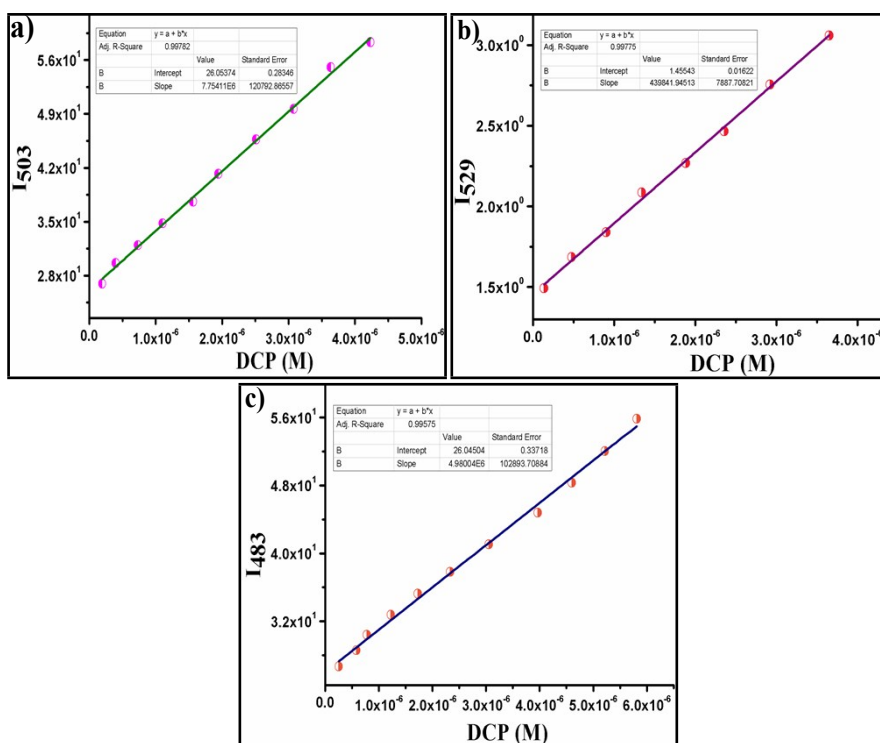
**Fig. S17** Fluorescence spectra of **HMBM** (2  $\mu\text{M}$ ) in acetonitrile–water (7:3) upon addition of increasing amount of DCP (40  $\mu\text{M}$ ),  $\lambda_{\text{exc}} = 345 \text{ nm}$ .

### Calculation of Detection limit:

The detection limit (DL) of **NTBT** for DCP were determined from the following equation:

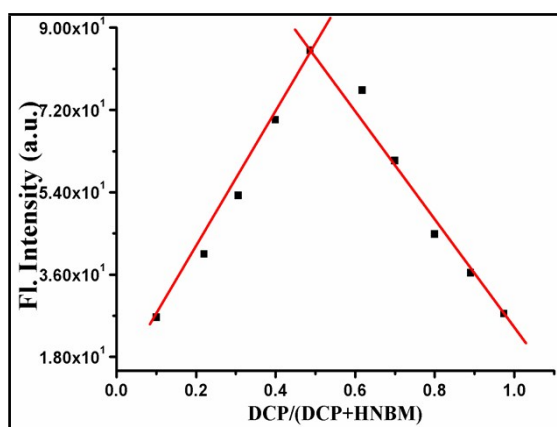
$$\text{DL} = K * \text{Sb1}/\text{S}$$

Where  $K = 2$  or  $3$  (we take  $3$  in this case);  $\text{Sb1}$  is the standard deviation of the blank solution;  $\text{S}$  is the slope of the calibration curve.

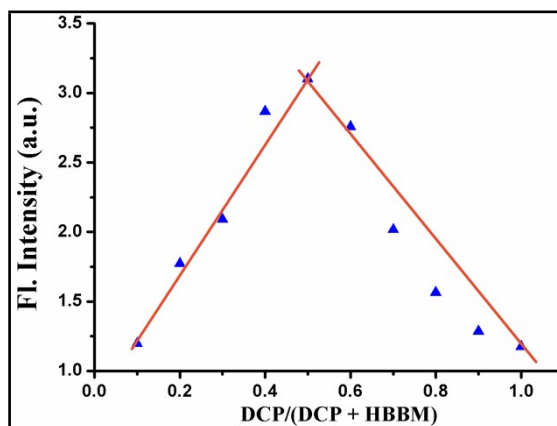


**Fig. S18** (a) LOD calculated from fluorescence data for **HNB** with DCP in  $\text{CH}_3\text{CN}$  at  $\lambda_{\text{ex}} = 373 \text{ nm}$ ,  $\lambda_{\text{em}} = 503 \text{ nm}$ . LOD is  $0.10 \mu\text{M}$  (b) LOD calculated from fluorescence data for **HBB** with DCP in  $\text{CH}_3\text{CN}$  at  $\lambda_{\text{ex}} = 398 \text{ nm}$ ,  $\lambda_{\text{em}} = 529 \text{ nm}$ . LOD is  $0.11 \mu\text{M}$  (c) LOD calculated from fluorescence data for **HMB** with DCP in  $\text{CH}_3\text{CN}$  at  $\lambda_{\text{ex}} = 345 \text{ nm}$ ,  $\lambda_{\text{em}} = 483 \text{ nm}$ . LOD is  $0.20 \mu\text{M}$ .

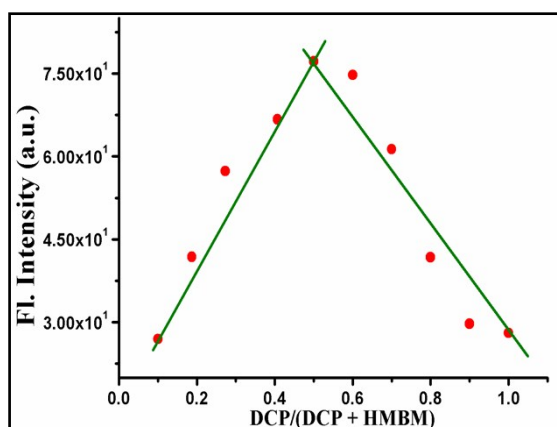
**Job's Plot:**



**Fig. S19** Fluorescence Job's plot of **HNB** with DCP.



**Fig. S20** Fluorescence Job's plot of **HBBM** with **DCP**.



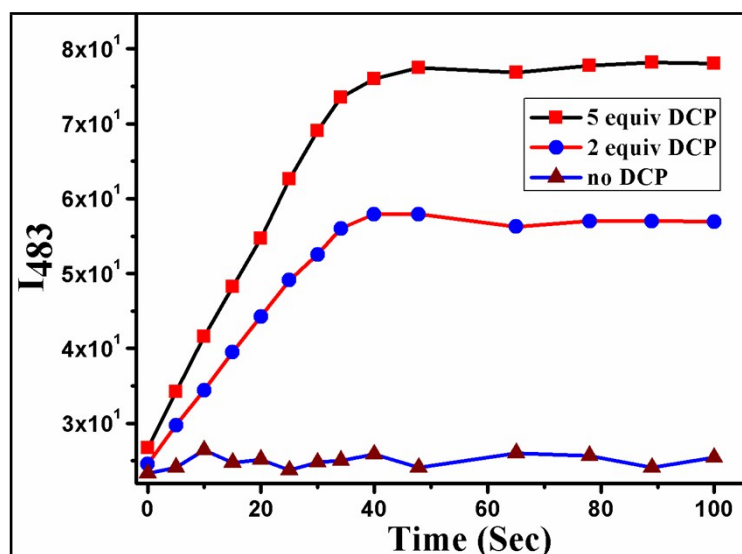
**Fig. S21** Fluorescence Job's plot of **HMBM** with **DCP**.

**Kinetic Study:**

The rate of the cyclization was determined by fitting the fluorescence intensities of the samples to the Pseudo-First Order Equation (1):

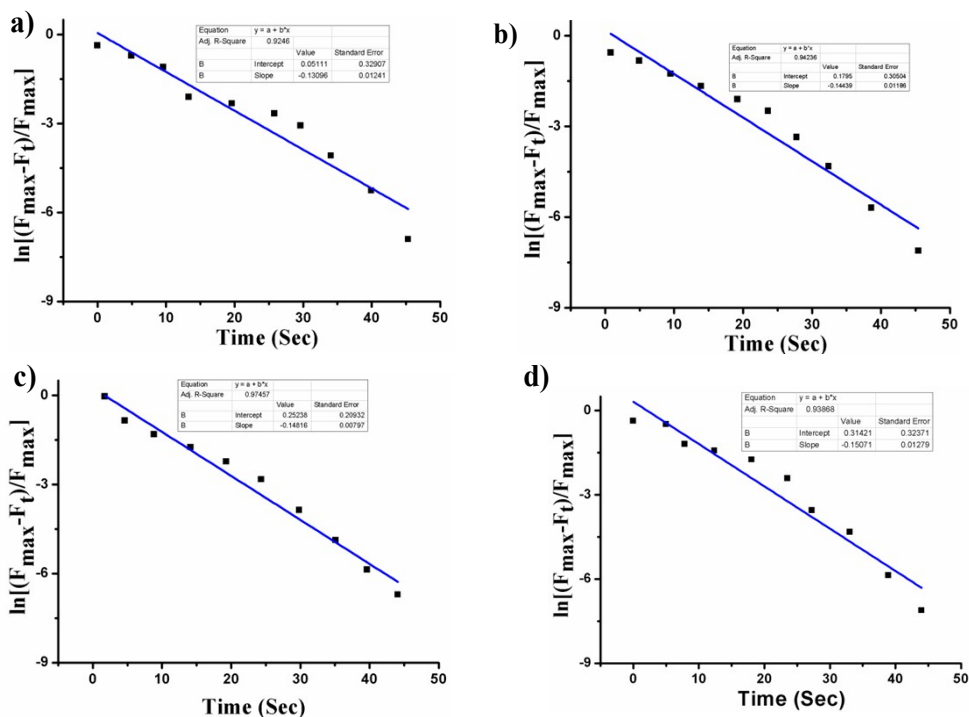
$$\ln(F_{\max} - F_t) / F_{\max} = -kt \dots \dots \dots (1)$$

Where  $F(t)$  and  $F(\max)$  are the fluorescence intensities at the monitoring wavelengths at times  $t$  and the maxima values which are the last fluorescence intensities when the cyclization of **NTBT** reached the conversion of 100%. The  $k$  is the apparent rate constant.

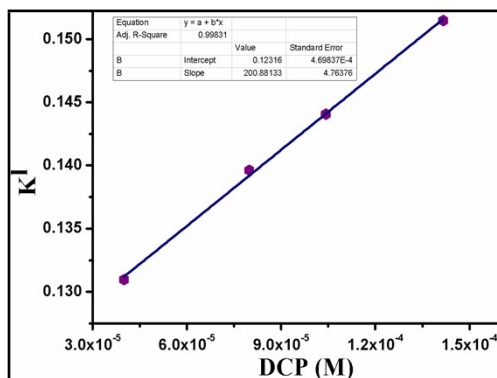


**Fig. S22** Time-dependent fluorescence intensity of **HMBM** ( $2 \mu\text{M}$ ) at 483 nm with incremental (0- 5 equiv) addition of DCP ( $40 \mu\text{M}$ ) excitation at 345 nm.

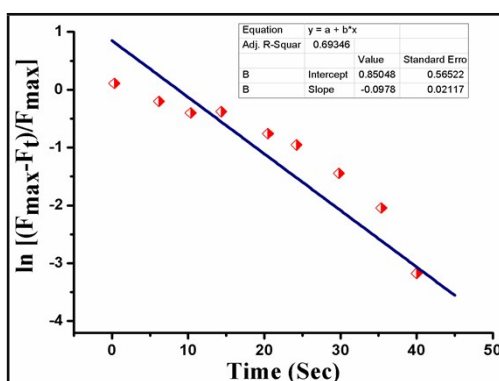
**Rate Constant:**



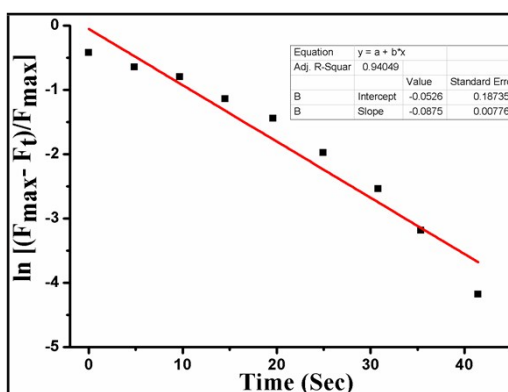
**Fig. S23** (a) Pseudo first-order kinetic plot of reaction of **HNBM** ( $0.2 \mu\text{M}$ ) with various concentration of DCP a)  $40 \mu\text{M}$  b)  $80 \mu\text{M}$  c)  $100 \mu\text{M}$  d)  $150 \mu\text{M}$  in acetonitrile. Thus the Pseudo first-order rate constant of the reaction at  $25^\circ\text{C}$ ,  $K = -0.15071 \text{ Sec}^{-1}$ .



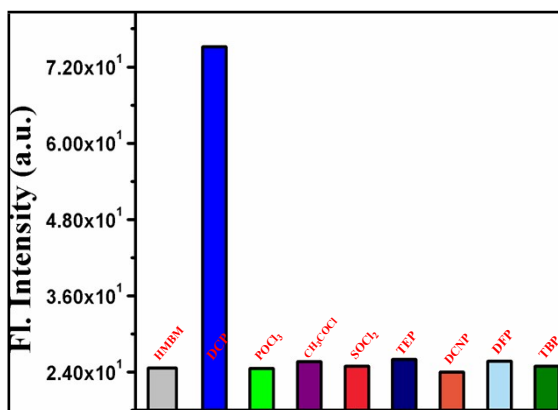
**Fig. S24** Plot of the observed  $k'$  versus the concentration of DCP for the pseudo first-order reaction of **HNBM** ( $2 \mu\text{M}$ ) with varying concentration of DCP ( $0\text{-}150 \mu\text{M}$ ). **Slope =  $200.88 \text{ M}^{-1}\text{Sec}^{-1}$ .**



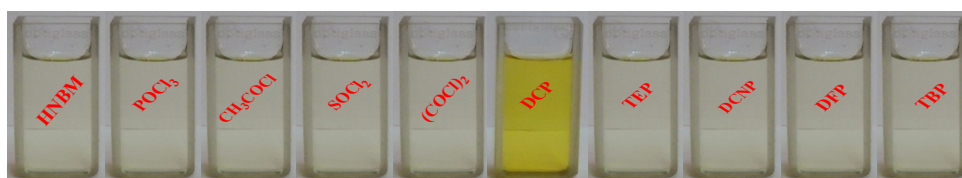
**Fig. S25** Pseudo first-order kinetic plot of reaction of **HBBM** ( $2 \mu\text{M}$ ) with DCP ( $40 \mu\text{M}$ ). Rate constant of the reaction at  $25^\circ\text{C}$ ,  $K = -0.097 \text{ Sec}^{-1}$



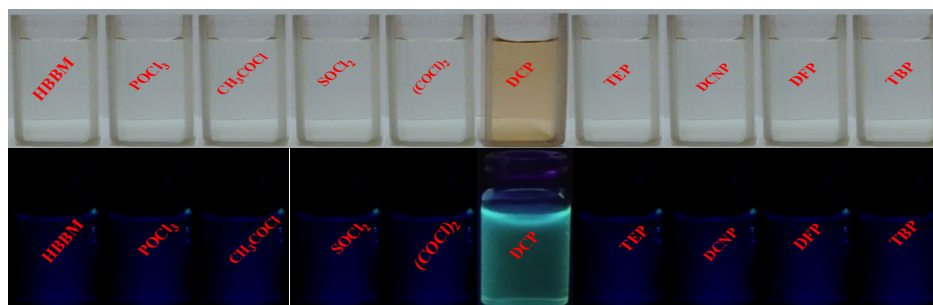
**Fig. S26** Pseudo first-order kinetic plot of reaction of **HMBM** ( $2 \mu\text{M}$ ) with DCP ( $40 \mu\text{M}$ ). Rate constant of the reaction at  $25^\circ\text{C}$ ,  $K = -0.087 \text{ Sec}^{-1}$ .



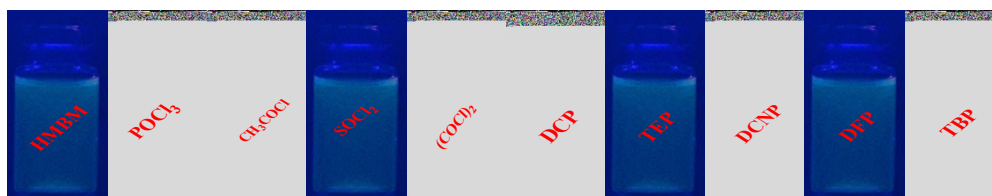
**Fig. S27** Fluorescence intensity response of **HNBMM** (2  $\mu\text{M}$ ) to DCP (40  $\mu\text{M}$ ) and other interferents (100  $\mu\text{M}$ ). The emission intensity was measured at  $\lambda_{\text{em}} = 483 \text{ nm}$ .



**Fig. S28** The visible color changes of **HNBMM** in aq.  $\text{CH}_3\text{CN}$  ( $\text{CH}_3\text{CN}:\text{H}_2\text{O} = 7:3 \text{ v/v}$ , 10 mM HEPES buffer, pH = 7.4) upon addition of various interferents.



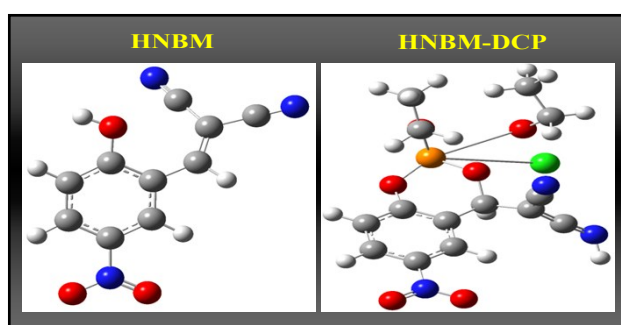
**Fig. S29** The visible color (top) and fluorescence (bottom) changes of **HNBMM** in aq.  $\text{CH}_3\text{CN}$  ( $\text{CH}_3\text{CN}:\text{H}_2\text{O} = 7:3 \text{ v/v}$ , 10 mM HEPES buffer, pH = 7.4) upon addition of various interferents.



**Fig. S30** Fluorescence changes of **HNBMM** in aq.  $\text{CH}_3\text{CN}$  ( $\text{CH}_3\text{CN}:\text{H}_2\text{O} = 7:3 \text{ v/v}$ , 10 mM HEPES buffer, pH = 7.4) upon addition of various interferents.

## Computational details:

Geometries have been optimized using the B3LYP/6-31G (d, p) level of theory. The geometries are verified as proper minima by frequency calculations. Time-dependent density functional theory calculation has also been performed at the same level of theory. All calculations have been carried out using Gaussian 09 program. The numbers in parentheses are the excitation energy in wavelength. [b] Oscillator strength. [c] H stands for HOMO and L stands for LUMO.



**Fig. S31** Energy Optimized structures of **HNBM** and **HNBM-DCP**

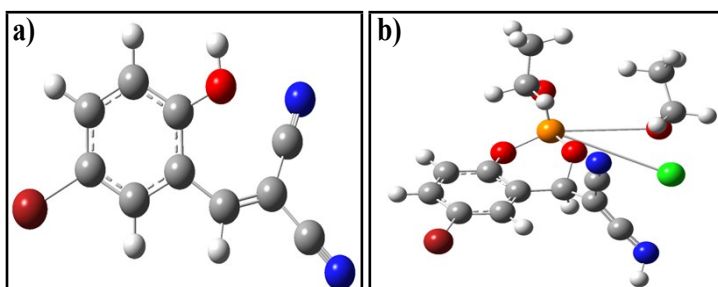
**Table S1:** Selected electronic excitation energies (eV), oscillator strengths (f) and main configurations of **HNBM** and **HNBM-DCP**. The data were calculated by TDDFT//B3LYP/6-311+G (*d,p*) based on the optimized ground state geometries.[a] Only selected excited states were considered. The numbers in parentheses are the excitation energy in wavelength. [b] Oscillator strength. [c] H stands for HOMO and L stands for LUMO.

Molecules	Electronic Transition	Excitation Energy <sup>a</sup>	f <sup>b</sup>	Composition <sup>c</sup>	(composition) %
<b>HNBM</b>	S <sub>0</sub> → S <sub>1</sub>	3.1886 eV 346.09 nm	0.1275	H → L+1	74.8
	S <sub>0</sub> → S <sub>6</sub>	4.2434 eV 311.16 nm	0.2891	H -1 → L	68.6
<b>HNBM-DCP</b>	S <sub>0</sub> → S <sub>1</sub>	3.9418 eV 396.98 nm	0.2140	H → L	64.3
	S <sub>0</sub> → S <sub>2</sub>	4.2187 eV 369.86 nm	0.2504	H-1 → L	66.2

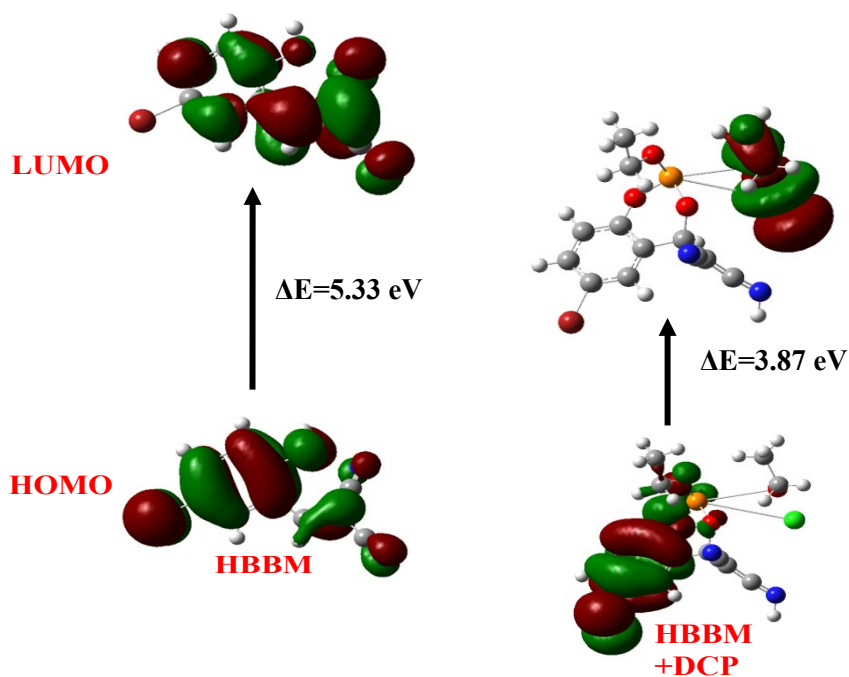


**Table S2:** Energies of the highest occupied molecular orbital (HOMO) and lowest unoccupied molecular orbital (LUMO) of **HNBM** and **HNBM - DCP**.

Species	$E_{\text{HOMO}}$ (a.u)	$E_{\text{LUMO}}$ (a.u)	$\Delta E$ (a.u)	$\Delta E$ (eV)	$\Delta E$ (kcal/mol)
<b>HNBM</b>	-0.27131	-0.1147	0.1565	4.2593	98.2037
<b>HNBM-DCP</b>	-0.29688	-0.18415	0.11273	3.06749	70.7380



**Fig. S32** Energy Optimized structures of **HBBM** and **HBBM-DCP**



**Fig. S33** Energy difference in the respective HOMO and LUMO of **HBBM** and **HBBM-DCP**.

**Table S3:** Selected electronic excitation energies (eV), oscillator strengths (f) and main configurations of **HBBM** and **HBBM-DCP**. The data were calculated by TDDFT//B3LYP/6-311+G (d,p) based on the optimized ground state geometries. [a] Only selected excited states were considered. The numbers in parentheses are the excitation energy in wavelength. [b] Oscillator strength. [c] H stands for HOMO and L stands for LUMO.

Molecules	Electronic Transition	Excitation Energy <sup>a</sup>	f <sup>b</sup>	Composition <sup>c</sup>	(composition) %
<b>HBBM</b>	S <sub>0</sub> → S <sub>1</sub>	3.1702 eV 349.09 nm	0.2075	H → L	93.8
	S <sub>0</sub> → S <sub>9</sub>	4.0234 eV 307.16 nm	0.5191	H -1 → L	93.6
<b>HBBM-DCP</b>	S <sub>0</sub> → S <sub>1</sub>	3.9018 eV 389.98 nm	0.2140	H → L	56.3
	S <sub>0</sub> → S <sub>2</sub>	4.8187 eV 369.86 nm	0.2504	H-1 → L	51.2

**Table S4:** Energies of the highest occupied molecular orbital (HOMO) and lowest unoccupied molecular orbital (LUMO) of **HBBM** and **HBBM - DCP**.

Species	E <sub>HOMO</sub> (a.u)	E <sub>LUMO</sub> (a.u)	ΔE(a.u)	ΔE(eV)	ΔE(kcal/mol)
<b>HBBM</b>	-0.23596	-0.03986	0.1961	5.3360	122.05275
<b>HBBM-DCP</b>	-0.24668	-0.10415	0.1425	3.8783	89.36631

### Quantum yield calculation:

Here, the quantum yield  $\phi$  was measured by using the following equation:

$$\phi_x = \phi_s (F_x / F_s)(A_s / A_x)(n_x^2 / n_s^2)$$

Where,

X & S indicate the unknown and standard solution respectively,  $\phi$  = quantum yield,

F = area under the emission curve, A = absorbance at the excitation wave length,

n = index of refraction of the solvent. Here  $\phi$  measurements were performed using Fluorescein in 0.1 M NaOH as standard ( $\phi = 0.79$ ) and anthracene in ethanol as standard (0.27) were used. For standard (s) Fluorescein in 0.1 M NaOH and anthracene in ethanol the following values were determined:

$$n_s = 1.3330 \text{ (for 0.1 M NaOH); } n_x = 1.344 \text{ (for CH}_3\text{CN); } \phi = 0.79.$$

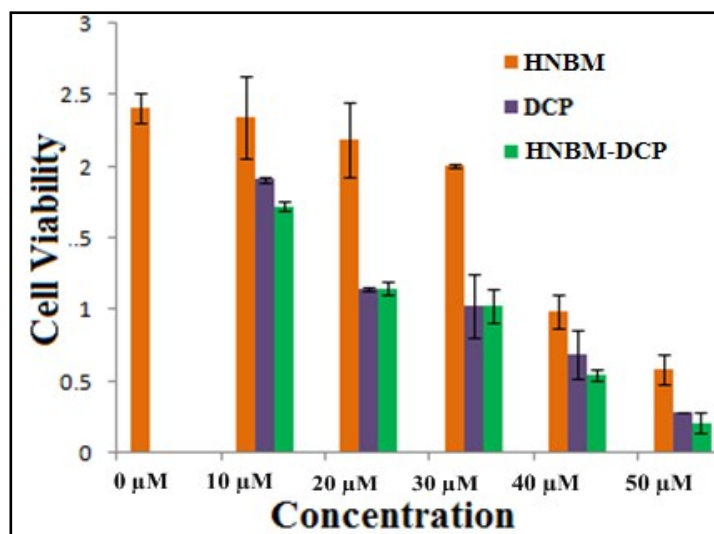
$$n_s = 1.5948 \text{ (ethanol); } n_x = 1.344 \text{ (for CH}_3\text{CN); } \phi = 0.27.$$

Using the above equation, we calculated quantum yield of probes.

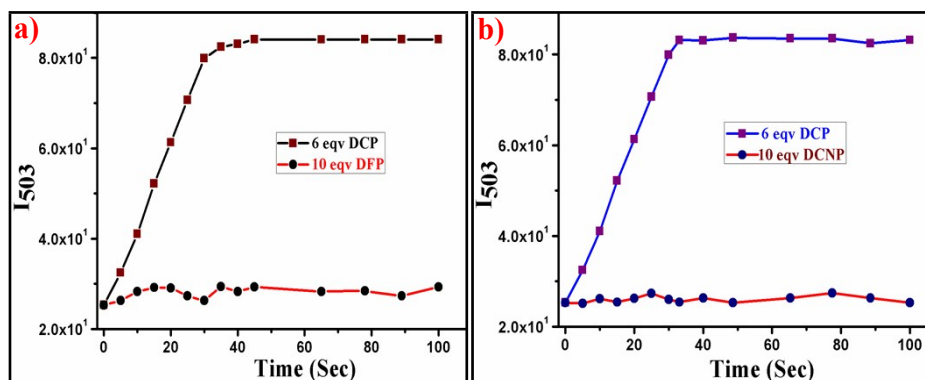
Probes	Quantum yield ( $\phi$ )
HNBM	0.03
HNBM-DCP at 503 nm	0.17
HBBM	0.008
HBBM-DCP at 529 nm	0.02
HMBM	0.025
HNBM-DCP at 483 nm	0.135

**Table S5:** Quantum yield data

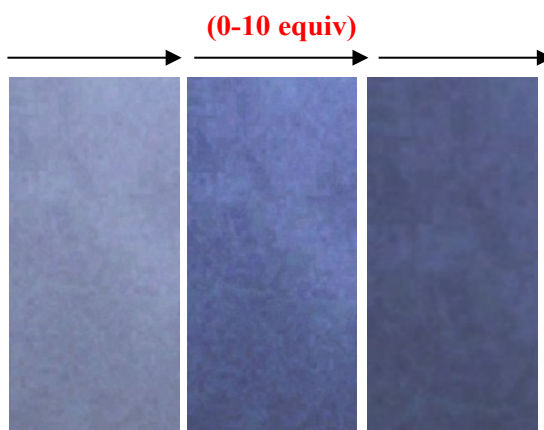
**MTT assay:**



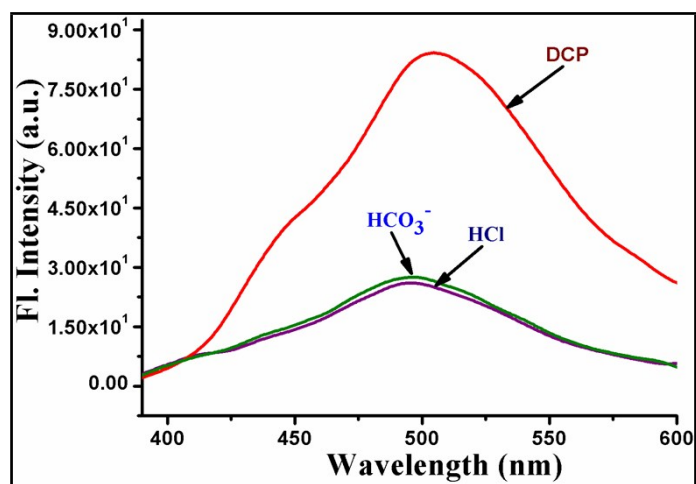
**Fig.S34** Cell viability assay of **Hep-2** cells to observe the cytotoxic effect of **HNBM** and **DCP**.



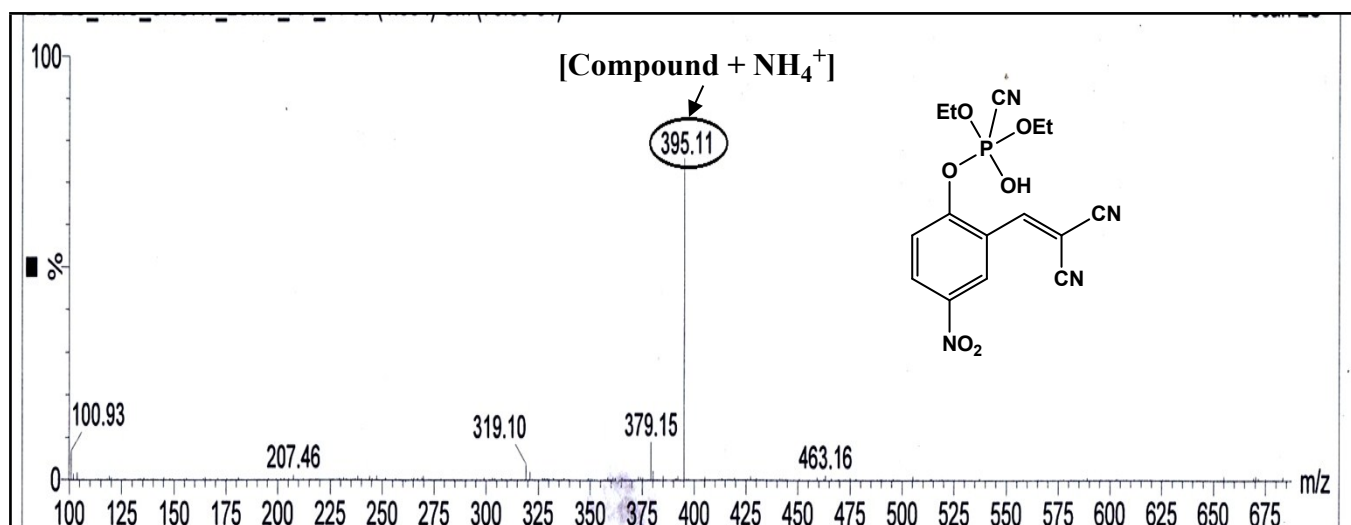
**Fig. S35** a) Time-dependent fluorescence intensity of **HNBM** ( $2 \mu\text{M}$ ) at 503 nm with 6 equiv addition of DCP ( $40 \mu\text{M}$ ) and 10 equiv addition of DFP, excitation at 373 nm. b) Time-dependent fluorescence intensity of **HNBM** ( $2 \mu\text{M}$ ) at 503 nm with 6 equiv addition of DCP ( $40 \mu\text{M}$ ) and 10 equiv addition of DCNP, excitation at 373 nm.



**Fig. S36** Fluorescence photos of **HNBM** soaked cellulose test papers have been exposed to various concentration (0- 10 equiv) of HCl for 1 min.



**Fig. S37** Comparative fluorescence spectra of **HNBM** (2  $\mu\text{M}$ ) in acetonitrile-water (10 mM HEPES buffer, 7:3 V/V, pH 7.4, at 25°C) upon addition of DCP (40  $\mu\text{M}$ ),  $\text{HCO}_3^-$  (100  $\mu\text{M}$ ) and HCl (100  $\mu\text{M}$ ),  $\lambda_{\text{exc}} = 373 \text{ nm}$ .



**Fig. S38** LCMS of **HNBM-DCNP**.

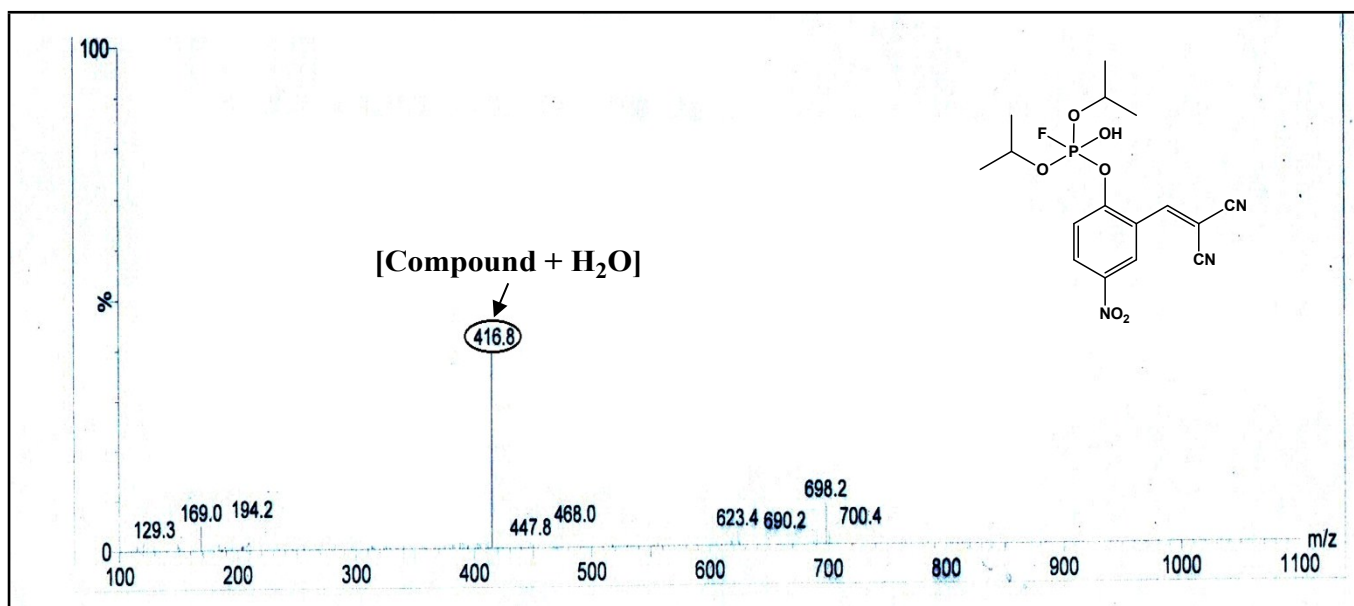


Fig. S39 LCMS of HNBM-DFP.