# Supplementary Information 

Ratiometric Red-emission Fluorescent Detection of $\mathbf{A l}^{3+}$ in Pure Aqueous
Solution and Live Cells by Fluorescent Peptidyl Probe using AggregationInduced emission

Lok Nath Neupane ${ }^{\text {a }}$, Pramod Kumar Mehta ${ }^{\text {a }}$, Semin Oh ${ }^{\text {a }}$, See-Hyoung Park ${ }^{\text {b }}$, Keun -Hyeung Lee ${ }^{\text {a,* }}$
${ }^{\text {a }}$ Center for Design and Applications of Molecular Catalysts, Department of Chemistry and Chemical Engineering, Inha University, Incheon, 402-751, South Korea. ${ }^{\mathrm{b}}$ Department of Bio and Chemical Engineering, Hongik University, Sejong 30016, South Korea.

Email address: leekh@inha.ac.kr
Fax number: +82-32-8675604
Phone number: +82-32-8607674

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## DLS measurement and CD measurement

The size distribution in an aqueous buffered solution ( 1 mM Tris, pH 7.0 ) was measured using a laser diffraction particle size analyzer (ELSZ, Otsuka Electronics, Osaka, Japan). The measurements were carried out by $90^{\circ}$ dynamic light scattering at 25 ${ }^{\circ} \mathrm{C}$.

Circular dichroism experiments were performed using JASCO 815 CD spectropolarimeter (Jasco, Tokyo, Japan). All the data were collected from 600 to 350 nm at a scan rate of $100 \mathrm{~nm} / \mathrm{min}$ at 0.5 nm data intervals and are presented as an average of three successive scans unless specified.

## Determination of detection limit and dissociation constant

The detection limit of $\mathbf{1}$ to $\mathrm{Al}^{3+}$ was calculated based on a fluorescence titration. To determine the $\mathrm{S} / \mathrm{N}$ ratio, the fluorescence emission intensity ratio $\left(\mathrm{I}_{600} / \mathrm{I}_{535}\right)$ of $5 \mu \mathrm{M}$ of $\mathbf{1}$ in aqueous solutions was measured 10 times, and the standard deviation of the blank measurements was determined. Three separate measurements of the emission intensity ratio were measured in the presence of increasing $\mathrm{Al}^{3+}$ concentrations, and the mean intensity ratio was plotted as a function of the $\mathrm{Al}^{3+}$ concentration to determine the slope. The detection limit was calculated using the following equation:

Detection limit $=3 \sigma / \mathrm{m}$
where $\sigma$ is the standard deviation of the intensity ratio of $\mathbf{1}$ in the absence of $\mathrm{Al}^{3+}$, and m is the slope of the emission intensity ratio $\left(\mathrm{I}_{600} / \mathrm{I}_{535}\right)$ of $5 \mu \mathrm{M}$ of $\mathbf{1}$ as a function of the $\mathrm{Al}^{3+}$ concentration. ${ }^{1}$

The dissociation constant was calculated based on the titration curve of the probe with metal ion. The fluorescence signal, F , is related to the equilibrium concentration of the complex (HL) between Host (H) and metal ion (L) by the following expression:
$\mathrm{F}=\mathrm{F}_{\mathrm{o}}+\Delta \mathrm{F} \times[\mathrm{HL}]$
$[\mathrm{HL}]=0.5 \times\left[\mathrm{K}_{\mathrm{D}}+\mathrm{L}_{\mathrm{T}}+\mathrm{H}_{\mathrm{T}}-\left\{\left(-\mathrm{K}_{\mathrm{D}}-\mathrm{L}_{\mathrm{T}}-\mathrm{H}_{\mathrm{T}}\right)^{2}-4 \mathrm{~L}_{\mathrm{T}} \mathrm{H}_{\mathrm{T}}\right\}^{1 / 2}\right]$
where $F_{o}$ is the fluorescence of the probe only and $\Delta F$ is the change in fluorescence due to the formation of HL. The dissociation constant was determined by a nonlinear least square fit of the data with the equation. ${ }^{2}$ The dissociation constant was calculated based on fluorescence titration curve of the probe with the metal ion using the modified BenesiHildebrand equation. The dissociation constant was calculated using the following equation. ${ }^{3,4}$
$\log \left(\mathrm{I}_{\mathrm{x}}-\mathrm{I}_{0} / \mathrm{I}_{\text {max }}-\mathrm{I}_{0}\right)=\mathrm{n} \times \log [\mathrm{M}]-\log \mathrm{K}_{\mathrm{d}}$
where $I_{0}$ is the fluorescence of the probe only, $I$ is the change in fluorescence due to the formation of complex, $I_{\max }$ is the final fluorescence emission intensity, $M$ is the concentration of $\mathrm{Al}^{3+}$ ions, and n is the slope.

## Measurement of Quantum yields

Fluorescence quantum yields of $\mathbf{1}$ in the presence and absence of $\mathrm{Al}^{3+}$ were obtained by using fluorescein as a standard. Each of the sample solution were prepared in distilled water and the absorbance were recorded in 10 mM tris buffer solution in different 10 mm quartz cell. The fluorescence spectrums of the solutions were recorded with the excitation wavelength of 470 nm and the relative fluorescence was determined by the area of the fluorescence emission. Where fluorescein used as a standard and its known quantum yield value is $0.91 .{ }^{5}$ Finally, quantum yield of $\mathbf{1}$ in the absence and presence of $\mathrm{Al}^{3+}$ were calculated. ${ }^{6}$

## Transmission Electron Microscopy (TEM) measurements

Transmission electron microscopy (TEM) was performed using a Philips CM 200 operated at an acceleration voltage of 120 kV . The sample was prepared by dropping 5 $\mu \mathrm{L}$ of the complex of $\mathbf{1}$ and $\mathrm{Al}^{3+}$ on a $300-$ mesh copper grid coated with carbon followed by staining with phosphotungstic acid ( $1 \mathrm{wt} \%$ ). TEM grids were completely dried in vacuum desiccator before TEM measurements.


Scheme S1. Synthetic scheme of 4


Scheme S2. Synthetic scheme of $\mathbf{1}$


Fig. S1. HPLC chromatogram of 4
+MS, 0.1-0.4min \#3-22

| Intens. $\times 10^{5}-$3 <br> 2 <br> 2 <br> 2 <br>  | $\left[4+\mathrm{Na}^{+}\right]^{+}$ <br> Calculated Mass $=430.1196$ <br> Observed Mass $=430.1199$ |  |  |  |  |  |  |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| 100 | 200 | 300 | 400 | 500 | 600 | 70 |  | 800 |  |  | m/z |
| Meas. m/z | \# | Ion Formula | $\mathrm{m} / \mathrm{z}$ | err [ppm] | mSigma | \# Sigma | Score | rdb | $e^{-}$Conf | N-Rule |  |
| 430.1199 | 1 | C20H16N9OS | 430.1193 | -1.5 | 5.7 | 1 | 100.00 | 17.5 | even | ok |  |
|  | 2 | C19H20N5O5S | 430.1180 | 4.6 | 7.8 | 2 | 41.47 | 12.5 | even | ok |  |
|  | 3 | C24H20N3O3S | 430.1220 | 4.7 | 18.1 | 3 | 32.44 | 16.5 | even | ok |  |
|  | 4 | C18H24NO9S | 430.1166 | -7.7 | 19.2 | 4 | 10.00 | 7.5 | even | ok |  |
|  | 5 | C17H20N9OS2 | 430.1227 | -6.3 | 21.2 | 5 | 21.32 | 12.5 | even | ok |  |
|  | 6 | C16H24N5O5S2 | 430.1213 | -3.2 | 27.3 | 6 | 52.40 | 7.5 | even | ok |  |
|  | 7 | C19H28NO4S3 | 430.1175 | 5.7 | 44.3 | 7 | 15.26 | 6.5 | even | ok |  |
|  | 8 | C16H32NO4S4 | 430.1209 | -2.1 | 64.0 | 8 | 23.81 | 1.5 | even | ok |  |
|  | 1 | C18H17N9NaOS | 430.1169 | -7.1 | 5.8 | 1 | 15.30 | 14.5 | even | ok |  |
|  | 2 | C22H21N3NaO3S | 430.1196 | 0.8 | 7.4 | 2 | 100.00 | 13.5 | even | ok |  |
|  | 3 | C19H25N3NaO3S2 | 430.1230 | 7.0 | 23.0 | 3 | 14.19 | 8.5 | even | ok |  |
|  | 4 | C27H21NNaOS | 430.1236 | -8.5 | 30.0 | 4 | 4.89 | 17.5 | even | ok |  |

Fig. S2. HRMS (ESI-TOF) spectrum of 4


Fig. S3. ${ }^{1} \mathrm{H}$ NMR of 4


Fig. S4. ${ }^{13} \mathrm{C}$ NMR of 4


Fig. S5. HPLC chromatogram of $\mathbf{1}$
+MS, 0.3-0.5min \#19-28


Fig. S6. HRMS (ESI-TOF) spectrum of 1


Fig. S7. ${ }^{1} \mathrm{H}$ NMR of $\mathbf{1}$


Fig. S8. ${ }^{13} \mathrm{C}$ NMR of $\mathbf{1}$


Fig. S9. Intensity ratio change $\left(\mathrm{I}_{600} / \mathrm{I}_{535}\right)$ of $\mathbf{1}(5 \mu \mathrm{M})$ as a function of $\mathrm{Al}^{3+}$ in aqueous buffered solution ( 10 mM Tris, $\mathrm{pH} 7.0)\left(\lambda_{\mathrm{ex}}=470 \mathrm{~nm}\right)$.


Fig. S10. Fluorescence emission spectra of $1(5 \mu \mathrm{M})$ with increasing concentration of $\mathrm{Al}^{3+}$ in the presence of (a) $\mathrm{Cu}^{2+}$ (12 equiv), (b) $\mathrm{Cr}^{3+}$ (12 equiv) and (c) $\mathrm{Fe}^{3+}$ (12 equiv) in aqueous buffered solution ( 10 mM Tris, pH 7.0 ).


Fig. S11. Non-linear least-squares fitting of the emission intensity of $\mathbf{1}(5 \mu \mathrm{M})$ with increasing concentration of $\mathrm{Al}^{3+}$ in aqueous buffered solution ( 10 mM Tris, pH 7.0 ).


Figure S12. Benesi-Hildebrand plot for determination of the binding stoichiometry and binding constant of $\mathbf{1}(5 \mu \mathrm{M})$ for $\mathrm{Al}^{3+}$ in aqueous buffered solution ( 10 mM Tris, pH 7.0 ). Emission intensity at 535 nm was used in the plot.


Fig. S13. Fluorescence emission spectra of $\mathbf{1}(5 \mu \mathrm{M})$ with $\mathrm{Al}^{3+}$ (12 equiv) in the presence of increasing concentration of EDTA in aqueous buffered solution ( 10 mM Tris, pH 7.0 ) $\left(\lambda_{\mathrm{ex}}=470 \mathrm{~nm}\right.$, slit $\left.=12 / 10 \mathrm{~nm}\right)$.


Fig. S14. Partial ${ }^{1} \mathrm{H}$ NMR spectra $(400 \mathrm{MHz})$ of $\mathbf{1}(5 \mathrm{mM})$ with increasing concentration of $\mathrm{Al}^{3+}$ in DMSO- $d_{6} / \mathrm{D}_{2} \mathrm{O}$ ( $\mathrm{v} / \mathrm{v}=4: 1$ ) containing 10 mM ammonium formate.


Fig. S15. UV-vis absorption spectra of $\mathbf{1}(5 \mu \mathrm{M})$ upon the gradual addition of $\mathrm{Al}^{3+}(0-80 \mu \mathrm{M})$ in aqueous buffered solution ( 10 mM Tris, pH 7.0 ).


Fig. S16. Fluorescence spectra of $\mathbf{1}(5 \mu \mathrm{M})$ in the absence and presence of $\mathrm{Al}^{3+}$ (12 equiv) with a different excitation wavelength


Fig. S17. IR spectra of $\mathbf{1}$ in the absence and presence of $\mathrm{Al}^{3+}$.


Fig. S18. Fluorescence spectra of $\mathbf{1}(5 \mu \mathrm{M})$ with increasing concentration of $\mathrm{Al}^{3+}$ in aqueous buffered solution (10 mM Hexamine buffer, pH 6.0).


Fig. S19. Linear relationship between the emission intensity ratio $\left(\mathrm{I}_{600} / \mathrm{I}_{535}\right)$ of $\mathbf{1}(5 \mu \mathrm{M})$ and the concentration of $\mathrm{Al}^{3+}$ $(0-5000 \mathrm{nM})$ in aqueous buffered solution ( 10 mM Tris, pH 7.0 ).


Fig. S20. Fluorescence emission spectra of $\mathbf{1}(5 \mu \mathrm{M})$ with increasing concentration of $\mathrm{Al}^{3+}(0-65 \mu \mathrm{M})$ in aqueous buffered solution ( 10 mM Tris, pH 7.0 ) containing (a) $10 \%(\mathrm{v} / \mathrm{v})$ tap water and (b) $10 \%(\mathrm{v} / \mathrm{v})$ ground water $\left(\lambda_{\mathrm{ex}}=\right.$ 470 nm ).


Fig. S21. Linear relationship between the emission intensity ratio $\left(\mathrm{I}_{600} / \mathrm{I}_{535}\right)$ of $\mathbf{1}(5 \mu \mathrm{M})$ and the concentration of $\mathrm{Al}^{3+}$ $(0-5000 \mathrm{nM})$ in aqueous buffered solution ( 10 mM Tris, pH 7.0 ) containing $10 \%(\mathrm{v} / \mathrm{v})$ tap water and $10 \%(\mathrm{v} / \mathrm{v})$ groundwater, respectively.


Fig. S22. Fluorescence emission spectra of $\mathbf{1}(5 \mu \mathrm{M})$ with $\mathrm{Al}^{3+}$ ( 12 equiv) in the presence of amino acids $(100 \mu \mathrm{M})$ and biothiols $(100 \mu \mathrm{M})$.


Fig. S23. Fluorescence emission spectra of $\mathbf{1}(5 \mu \mathrm{M})$ in the presence of $\mathrm{Al}^{3+}$ (12 equiv) with increasing concentration of (a) Cys, (b) Hcy, and (c) GSH.


Fig. S24. Emission intensity ratio changes of $\mathbf{1}(5 \mu \mathrm{M})$ by $\mathrm{Al}^{3+}$ (12 equiv) in the presence of (a) citric acid with $\mathrm{Fe}^{3+}$ and (b) citric acid with $\mathrm{Zn}^{2+}$.


Fig. S25. Emission intensity ratio changes of $\mathbf{1}(5 \mu \mathrm{M})$ by $\mathrm{Al}^{3+}$ (12 equiv) in the presence of olie acid and $\mathrm{Fe}^{3+}$.


Fig. S26. Emission intensity ratio changes of $\mathbf{1}(5 \mu \mathrm{M})$ by $\mathrm{Al}^{3+}$ (12 equiv) in the presence of ATP and $\mathrm{Fe}^{3+}$.


Fig. S27. MTT assay for the viability of MDA-MB-231 cells in DMEM $10 \%$ FBS treated with $\mathbf{1}, \mathbf{1}+\mathrm{Al}\left(\mathrm{NO}_{3}\right)_{3}$ and $\mathbf{1}+\mathrm{Al}\left(\mathrm{NO}_{3}\right)_{3}+$ EDTA for 24 h . The results are based on three separate MTT assays. The concentration of $\mathbf{1}$, $\mathrm{Al}\left(\mathrm{NO}_{3}\right)_{3}$ and EDTA is $10 \mu \mathrm{M}, 50 \mu \mathrm{M}$ and $200 \mu \mathrm{M}$, respectively.

## References

1 G. L. Long and J. D. Winefordner, Anal. Chem., 1983, 55, 712A-724A.
2 R. Reddi, T. R. Guzman, R. M. Breece, D. L. Tierney and B. R. Gibney, J. Am. Chem. Soc., 2007, 129, 12815-12827.
3 H. Y. Lin, P. Y. Cheng, C. F. Wan and A. T. Wu, Analyst, 2012, 137, 4415-4417.
4 A. Roy, S. Dey and P. Roy, Sens. Actuators B, 2016, 237, 628-642.
5 L. Porres, A. Holland, L.-O. Palsson, A. P. Monkman, C. Kemp and A. Beeby, J. Fluores., 2006, 16, 267-272.
6 G. A. Vehar, A. V. Reddy and J. H. Freisheim, Biochemistry, 1976, 15, 2512-2518.

