

Supplementary information

Rapid screening of colistin-resistant *Escherichia coli*, *Acinetobacter baumannii* and *Pseudomonas aeruginosa* by use of Raman spectroscopy and hierarchical cluster analysis

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Table S1 The antibiogram of the clinical isolates of *E. coli* (µg/ml)

isolates	specimen	ampicillin /sulbactam	ampicillin	amikacin	gentamicin	tobramycin	aztreonam	cefazolin	ciprofloxacin	levofloxacin	ceftriaxone	ceftazidime	cefepime	cefotetan	trimethoprim /sulfamethoxazole
E1	Sputum	4	8	2	1	1	1	4	4	8	1	1	1	4	20
E2	Wound secretion	32	32	2	1	1	64	64	4	8	64	16	64	4	320
E3	Urine	32	32	2	1	1	64	64	4	8	64	64	64	4	320
E4	Blood	32	32	4	16	8	16	64	4	8	64	64	4	4	320
E5	Urine	2	2	2	1	1	1	4	0.25	0.25	1	1	1	4	20
E6	Ascitic fluid	16	32	2	16	16	64	64	4	8	64	16	64	4	320
E7	Urine	32	32	2	16	8	64	64	4	8	64	64	64	64	320
E8	Urine	32	32	2	1	1	64	64	4	8	64	64	64	4	320
E9	Urine	32	32	2	1	1	64	64	4	8	64	32	32	4	320
E10	Urine	8	32	4	1	1	1	64	0.25	1	64	1	1	1	20
E11	Urine	4	8	2	1	1	1	4	0.25	0.25	1	1	1	4	20
E12	Urine	32	32	2	16	8	1	4	0.5	1	1	1	1	4	320
E13	Blood	16	32	2	16	8	1	4	4	8	1	1	1	4	320
E14	Sputum	32	32	2	1	1	16	64	4	8	16	64	1	32	320
E15	Sputum	32	32	2	1	1	4	64	0.25	1	64	1	2	4	20
E16	Urine	16	32	2	1	1	1	4	0.25	0.25	1	1	1	4	20
E17	Ascitic fluid	2	2	2	1	1	1	4	0.25	1	1	1	1	4	20
E18	Bile	2	4	2	1	1	1	4	0.25	0.25	1	1	1	4	20
E19	Blood	4	32	2	16	4	1	64	0.25	1	16	16	1	4	20
E20	Urine	16	32	2	16	8	1	4	0.25	1	1	1	1	4	320
E21	Bile	32	32	2	16	8	64	64	4	8	64	16	64	4	320
E22	Bile	16	16	2	1	1	1	8	4	4	1	1	1	4	20

E23	Sputum	16	32	2	1	1	1	64	1	1	1	4	1	4	20
E24	Sputum	16	32	2	1	1	16	64	0.25	1	64	4	2	4	320
E25	Ascitic fluid	32	32	2	1	1	2	64	4	8	32	1	2	4	20
E26	Sputum	32	32	2	16	8	64	64	4	8	64	64	64	4	20
E27	Urine	32	32	2	16	8	1	4	4	8	1	1	1	4	20
E28	Blood	4	32	2	1	1	4	64	4	8	64	4	1	4	320
E29	Ascitic fluid	16	32	4	16	16	4	64	4	8	64	4	2	4	320
E30	Urine	32	32	2	1	1	1	64	4	8	64	1	1	4	320

30 clinical isolates of *E. coli* were collected from October, 2017 to December, 2017 in Southwest Hospital (Chongqing, China). Antibiotic susceptibility tests were performed by the Vitek 2 system. All of the bacteria exhibit different antibiograms and none of the same isolate was obtained repeatedly.

Table S2 The antibiogram of the clinical isolates of *P. aeruginosa* (μg/ml)

isolates	specimen	imipenem	meropenem	piperacillin /tazobactam	amikacin	gentamicin	tobramycin	piperacillin	aztreonam	ciprofloxacin	levofloxacin	ceftriaxone	ceftazidime	cefepime	cefotetan	trimethoprim/sulfamethoxazole
P1	Sputum	2	1	4	2	1	1	4	8	0.25	1	64	4	2	64	160
P2	Sputum	16	16	16	2	1	1	8	32	0.5	1	64	8	8	64	320
P3	Sputum	16	0.25	8	2	1	1	4	32	1	1	64	8	8	64	320
P4	Sputum	2	0.25	4	2	1	1	4	32	0.25	1	32	4	2	64	80
P5	Sputum	1	1	64	2	1	1	32	32	1	2	64	4	8	64	320
P6	Sputum	2	0.25	4	2	1	1	4	32	0.25	0.5	64	4	2	64	80
P7	Sputum	4	1	8	8	4	1	8	32	0.5	1	64	4	2	64	160
P8	Sputum	1	0.25	4	16	1	1	4	32	0.5	0.25	64	1	2	64	80
P9	Sputum	4	1	4	2	1	1	4	32	0.25	0.5	64	2	1	64	80
P10	Fester	1	0.25	4	2	1	1	4	32	0.5	0.5	64	4	1	64	80
P11	Bile	2	0.25	16	2	1	1	32	32	0.25	0.5	64	16	4	64	80

P12	Ascitic fluid	2	0.25	4	2	1	1	4	32	0.5	0.5	64	4	2	64	80
P13	Sputum	1	0.25	4	16	8	1	4	32	0.25	1	64	4	2	64	160
P14	Sputum	4	0.25	4	2	1	1	4	4	0.25	0.5	64	4	1	64	80
P15	Fester	2	0.25	64	32	16	16	16	32	4	8	64	4	8	64	320
P16	Sputum	2	0.5	4	2	1	1	4	4	0.25	1	16	2	2	64	80
P17	Sputum	16	8	4	2	1	1	4	8	0.25	1	64	4	2	64	160
P18	Sputum	2	0.25	32	2	1	1	4	64	1	1	64	16	16	64	320
P19	Sputum	2	0.25	4	8	4	1	4	64	0.5	1	64	4	8	64	80
P20	Fester	2	0.5	4	16	8	2	4	2	0.25	1	16	1	2	64	80
P21	Sputum	16	4	4	2	2	1	8	8	0.25	0.5	16	4	2	64	80
P22	Sputum	2	2	120	8	2	1	128	16	0.25	1	64	64	16	64	80
P23	Sputum	16	16	120	32	16	16	128	16	1	4	64	4	8	64	320
P24	Sputum	1	0.25	4	2	1	1	4	16	0.25	0.5	64	2	1	64	320
P25	Sputum	1	0.25	64	16	8	2	4	64	0.5	1	64	8	32	64	320
P26	Sputum	8	8	8	8	4	1	8	2	0.25	1	64	4	4	64	160
P27	Sputum	8	8	8	8	2	1	8	4	0.25	1	64	4	2	64	80
P28	Sputum	4	1	4	2	1	1	4	8	0.5	1	64	4	2	64	160
P29	Throat swab	16	4	8	8	1	1	4	4	0.25	0.5	64	4	2	64	80
P30	Ascitic fluid	16	4	4	2	1	1	4	8	0.25	1	64	4	2	64	80

30 clinical isolates of *P. aeruginosa* were collected from October, 2017 to December, 2017 in Southwest Hospital (Chongqing, China). Antibiotic susceptibility tests were performed by the Vitek 2 system. All of the bacteria exhibit different antibiograms and none of the same isolate was obtained repeatedly.

Table S3 The antibiogram of the clinical isolates of *A. baumannii* (µg/ml)

isolates	specimen	macroclant	imipenem	ampicillin /sulbactam	ampicillin	gentamicin	tobramycin	aztreonam	ciprofloxacin	levofloxacin	ceftriaxone	ceftazidime	cefepime	trimethoprim/sulfamethoxazole
A1	Sputum	512	2	32	32	8	1	64	4	8	64	64	32	40
A2	Skin swab	512	1	2	8	1	1	16	0.25	0.25	8	4	2	20
A3	Throat swab	512	1	2	16	1	1	16	0.25	0.25	16	4	2	20
A4	Sputum	512	1	2	16	1	1	64	0.25	0.25	8	4	2	20
A5	Sputum	512	1	2	16	1	1	16	0.25	0.25	8	4	2	20
A6	Sputum	512	1	2	16	1	1	32	0.25	0.25	16	4	2	20
A7	Sputum	256	1	2	16	1	1	32	0.5	0.25	16	4	2	20
A8	Ascitic fluid	512	16	32	32	8	2	32	4R	4	64	64	64	320
A9	Sputum	512	16	32	32	1	1	32	4R	8	64	64	64	320
A10	Sputum	512	1	2	32	1	1	16	0.25	0.25	8	4	2	20
A11	Sputum	128	1	2	16	1	1	16	0.25	0.25	8	4	2	20
A12	Sputum	512	16	32	32	1	1	64	4	4	64	64	64	320
A13	Sputum	512	16	32	32	4	1	32	4	8	64	64	64	20
A14	Sputum	512	32	32	32	2	1	64	4	4	64	64	64	320
A15	Wound secretion	512	16	32	32	8	1	64	4	8	64	16	64	320
A16	Sputum	512	1	0.25	16	1	1	16	0.25	0.25	8	4	1	20
A17	Sputum	256	1	2	16	2	1	16	0.25	0.25	16	4	2	20
A18	Sputum	256	1	2	16	1	1	32	0.25	0.25	16	4	2	20

A19	Sputum	512	1	2	32	1	1	16	0.25	0.25	16	16	8	20
A20	Sputum	512	1	2	32	1	1	32	0.5	0.25	32	8	4	20
A21	Bile	512	1	2	32	1	1	64	0.25	0.25	16	8	4	20
A22	Sputum	512	16	32	32	16	16	64	4	4	64	64	64	320
A23	Ascitic fluid	512	16	32	32	16	16	64	4	8	64	64	64	320
A24	Sputum	512	16	32	32	16	16	64	4	8	64	64	64	20
A25	Ascitic fluid	512	8	32	32	1	1	64	4	4	64	64	64	320
A26	Sputum	512	16	32	32	16	16	64	4	2	64	64	64	320
A27	Sputum	512	16	32	32	16	16	64	4	4	64	64	64	20
A28	Sputum	256	16	32	32	16	16	64	4	2	64	64	64	320
A29	Sputum	256	1	2	16	1	1	16	0.25	0.25	16	4	2	20
A30	Sputum	512	1	2	16	1	1	32	0.25	0.25	16	8	4	20

30 clinical isolates of *A. baumannii* were collected from October, 2017 to December, 2017 in Southwest Hospital (Chongqing, China). Antibiotic susceptibility tests were performed by the Vitek 2 system. All of the bacteria exhibit different antibiograms and none of the same isolate was obtained repeatedly.

Table S4 Detection of colistin resistance in *E. coli* by BMD method and the new method

Isolates	BMD method (µg/ml)	the new method (µg/ml)	Isolates	BMD method (µg/ml)	the new method (µg/ml)
E1	2	Susceptible	E22	≤0.25	Susceptible
E2	0.5	Susceptible	E23	1	Susceptible
E3	≤0.25	Susceptible	E24	0.5	Susceptible
E4	≤0.25	Susceptible	E25	0.5	Susceptible
E5	≤0.25	Susceptible	E26	0.5	Susceptible
E6	0.5	Susceptible	E27	0.5	Susceptible
E7	≤0.25	Susceptible	E28	0.5	Resistant (false positive)
E8	≤0.25	Susceptible	E29	≤0.25	Susceptible
E9	≤0.25	Susceptible	E30	≤0.25	Susceptible
E10	≤0.25	Susceptible	E2R	8	Resistant
E11	1	Susceptible	E3R	4	Resistant
E12	1	Resistant (false positive)	E6R	≥32	Susceptible (false negative)
E13	≤0.25	Susceptible	E8R	4	Resistant
E14	≤0.25	Susceptible	E9R	16	Resistant
E15	≤0.25	Susceptible	E10R	4	Resistant
E16	0.5	Susceptible	E18R	16	Resistant
E17	0≤.25	Susceptible	E20R	≥32	Resistant
E18	≤0.25	Susceptible	E22R	8	Resistant
E19	≤0.25	Susceptible	E24R	8	Resistant
E20	≤0.25	Susceptible	E25R	8	Resistant
E21	≤0.25	Susceptible	E28R	8	Resistant

E2R is a resistant isolate induced from E2 with a serial passages exposed to colistin, so are the other resistant isolates. Of all the 30 susceptible (negative) tested by the new method, results of 28 susceptible corresponded to the standard BMD method, except 2 susceptible (E12, E28) were defined as resistant (false positive) ;Results of 11 resistant (positive) corresponded to the standard BMD method, only 1 resistant(E6R) was defined as susceptible (false negative). The detection specificity is 93.3% and the detection sensitivity is 91.7%.

Table S5 Detection of colistin resistance in *P. aeruginosa* by BMD method and the new method

Isolates	BMD method (µg/ml)	the new method (µg/ml)	Isolates	BMD method (µg/ml)	the new method (µg/ml)
P1	2	Susceptible	P22	1	Susceptible
P2	1	Susceptible	P23	2	Susceptible
P3	1	Susceptible	P24	2	Susceptible
P4	2	Susceptible	P25	2	Susceptible
P5	2	Susceptible	P26	2	Susceptible
P6	1	Susceptible	P27	2	Susceptible
P7	1	Susceptible	P28	1	Susceptible
P8	1	Susceptible	P29	0.5	Susceptible
P9	0.5	Susceptible	P30	≤0.25	Susceptible
P10	≤0.25	Resistant (false positive)	P1R	≥32	Susceptible (false negative)
P11	1	Susceptible	P2R	4	Resistant
P12	2	Susceptible	P7R	4	Resistant
P13	2	Susceptible	P8R	4	Resistant
P14	1	Susceptible	P11R	8	Resistant
P15	1	Susceptible	P15R	8	Resistant
P16	1	Susceptible	P17R	8	Resistant
P17	1	Susceptible	P18R	8	Resistant
P18	1	Susceptible	P21R	16	Resistant
P19	0.5	Susceptible	P25R	≥32	Resistant
P20	0.5	Resistant (false positive)			
P21	2	Susceptible			

P1R is a resistant isolate induced from P1 with a serial passages exposed to colistin, so are the other resistant isolates. Of all the 30 susceptible (negative) tested by the new method, results of 28 susceptible corresponded to the standard BMD method, except 2 susceptible (P10, E20) were defined as resistant (false positive) ;Results of 9 resistant (positive) corresponded to the standard BMD method, only 1 resistant (P1R) was defined as susceptible (false negative). The detection specificity is 93.3% and the detection sensitivity is 90.0%.

Table S6 Detection of colistin resistance in *A. baumannii* by BMD method and the new method

Isolates	BMD method (μg/ml)	the new method (μg/ml)	Isolates	BMD method (μg/ml)	the new method (μg/ml)
A1	1	Susceptible	A22	0.5	Susceptible
A2	1	Susceptible	A23	0.5	Susceptible
A3	1	Resistant (false positive)	A24	1	Resistant (false positive)
A4	0.5	Susceptible	A25	≤0.25	Susceptible
A5	1	Susceptible	A26	≤0.25	Susceptible
A6	0.5	Susceptible	A27	1	Susceptible
A7	0.5	Resistant (false positive)	A28	1	Susceptible
A8	0.5	Susceptible	A29	0.5	Susceptible
A9	0.5	Susceptible	A30	0.5	Susceptible
A10	≤0.25	Susceptible	A1R	8	Susceptible(false negative)
A11	0.25	Susceptible	A5R	4	Resistant
A12	≤0.25	Susceptible	A6R	4	Resistant
A13	0.5	Susceptible	A7R	16	Resistant
A14	0.5	Susceptible	A9R	≥32	Resistant
A15	0.5	Susceptible	A15R	4	Resistant
A16	2	Susceptible	A17R	≥32	Resistant
A17	1	Susceptible	A18R	16	Resistant
A18	1	Susceptible	A20R	8	Resistant
A19	0.5	Resistant (false positive)	A25R	8	Resistant
A20	0.5	Susceptible	A28R	8	Resistant
A21	0.5	Susceptible			

A1R is a resistant isolate induced from A1 with a serial passages exposed to colistin, so are the other resistant isolates. Of all the 30 susceptible (negative) tested by the new method, results of 26 susceptible corresponded to the standard BMD method, except 4 susceptible (A3, A7, A19 and A24) were defined as resistant (false positive) ; Results of 10 resistant (positive) corresponded to the standard BMD method, only 1 resistant (A1R) was defined as susceptible (false negative). The detection specificity is 86.7% and the detection sensitivity is 90.9%.

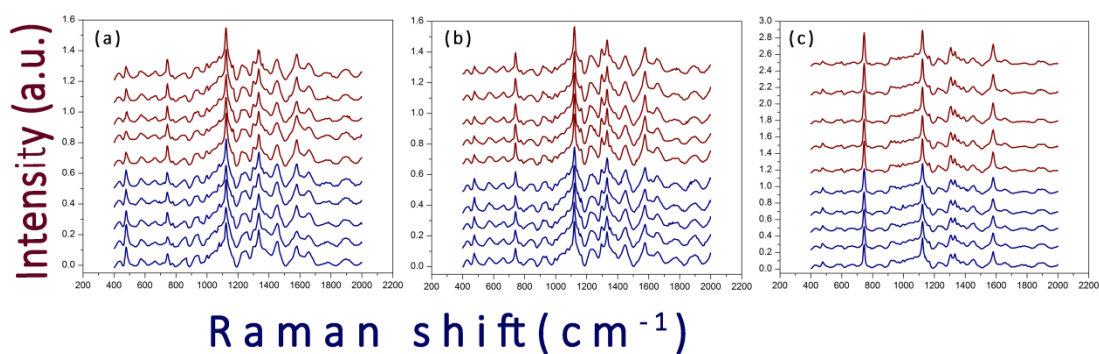


Figure S1. The 5 spectra of bacteria treated with colistin (blue) and without colistin (brown), (a) *E. coli*, (b) *A. baumannii*, (c) *P. aeruginosa*.

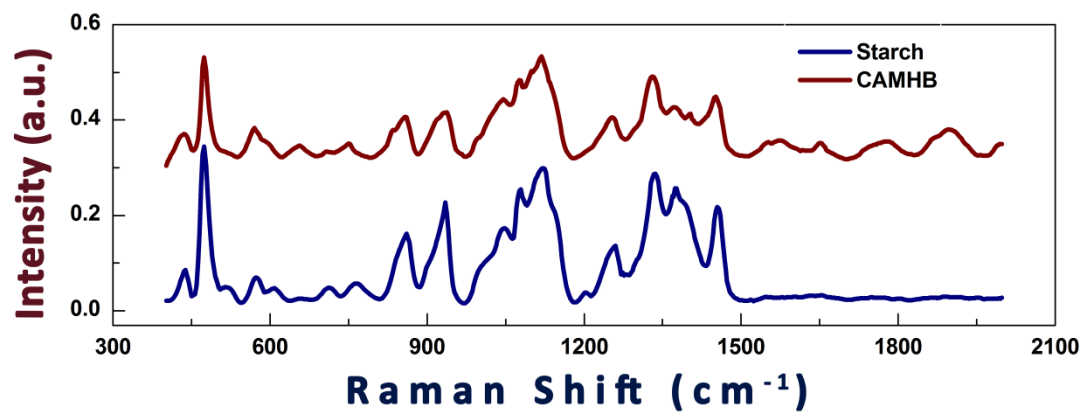


Figure S2. Raman spectra of the pure starch and the deposit of CAMHB. One spectrum is an average of five measurements.

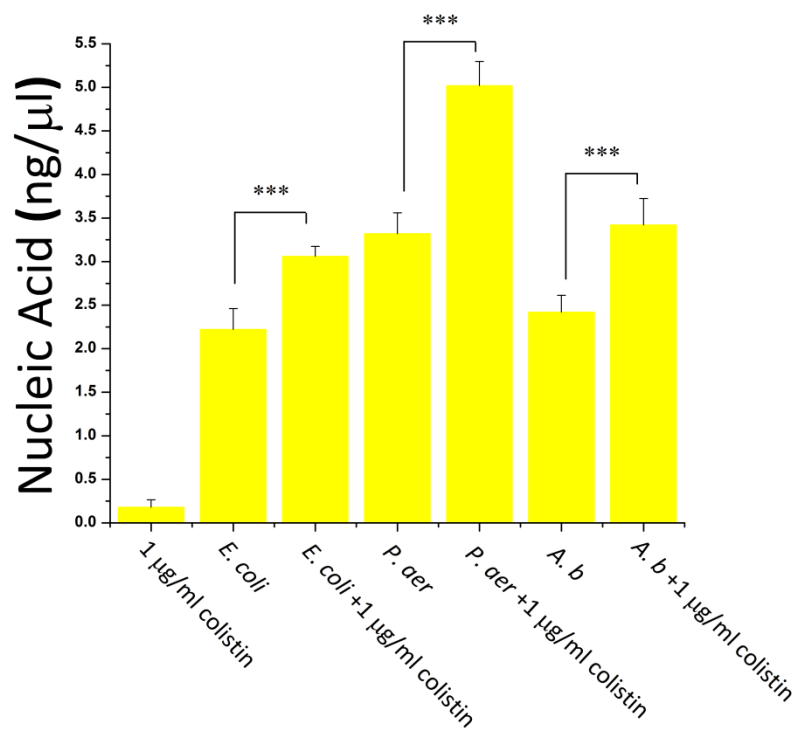


Figure S3. Nucleic acid concentrations of the supernatant of bacterial suspension after incubation with and without 1 μg/ml of colistin sulfate. *** $P < 0.05$ is the result of student's test analysis and error bar is the standard deviation of mean for three samples.

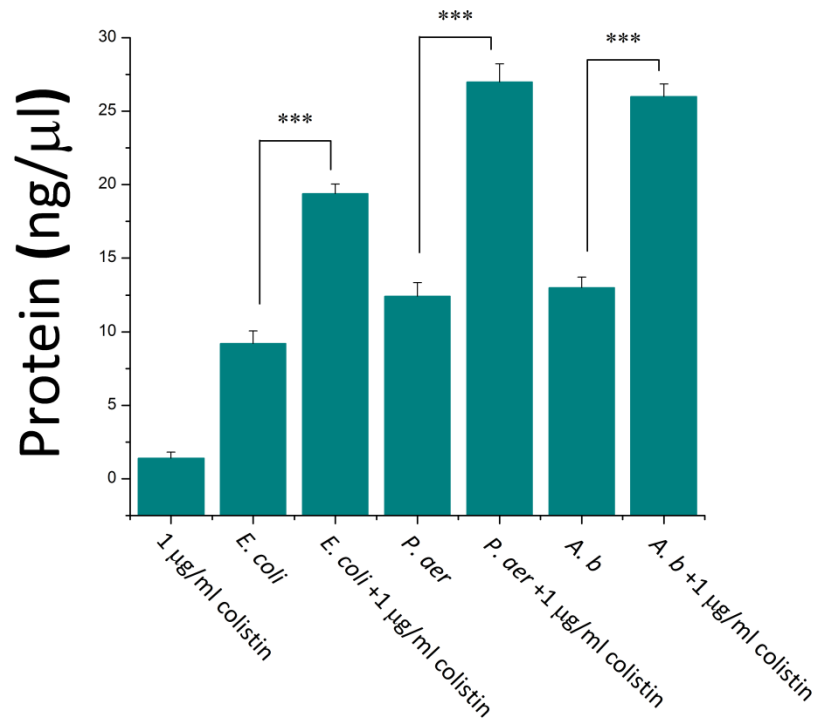
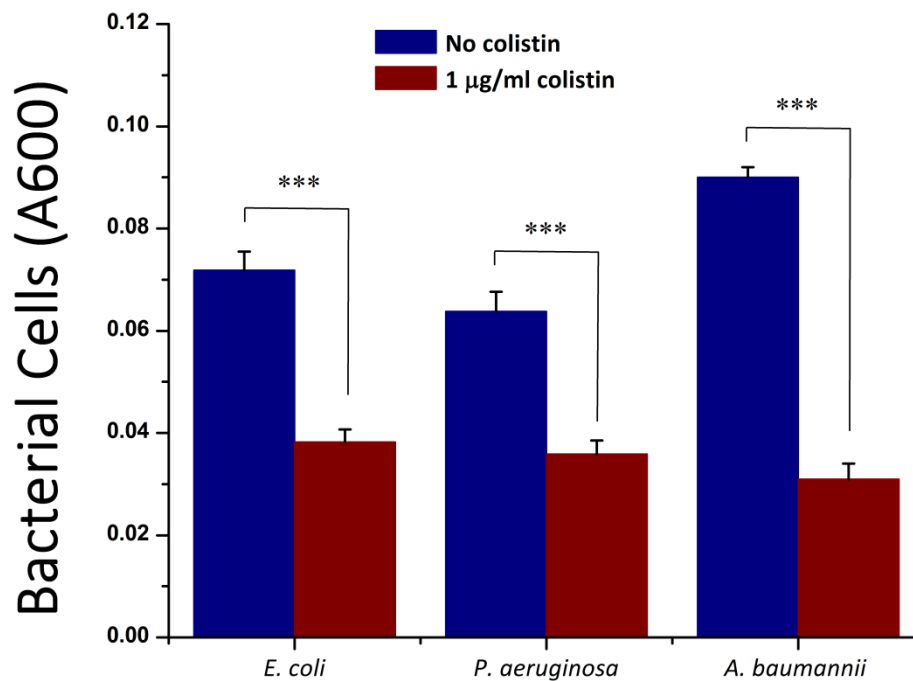


Figure S4. Protein concentrations of the supernatant of bacterial suspension after incubation with and without 1 $\mu\text{g/ml}$ of colistin sulfate. *** $P < 0.05$ is the result of student's test analysis and error bar is the standard deviation of mean for three samples.



Figures S5. The number of bacterial cells in the suspension of *E. coli* ATCC 25922, *P. aeruginosa* ATCC 27853 and *A. baumannii* ATCC 19606 treated with and without 1 $\mu\text{g/ml}$ of colistin sulfate. *** $P < 0.05$ is the result of student's test analysis and error bar is the standard deviation of mean for three samples.

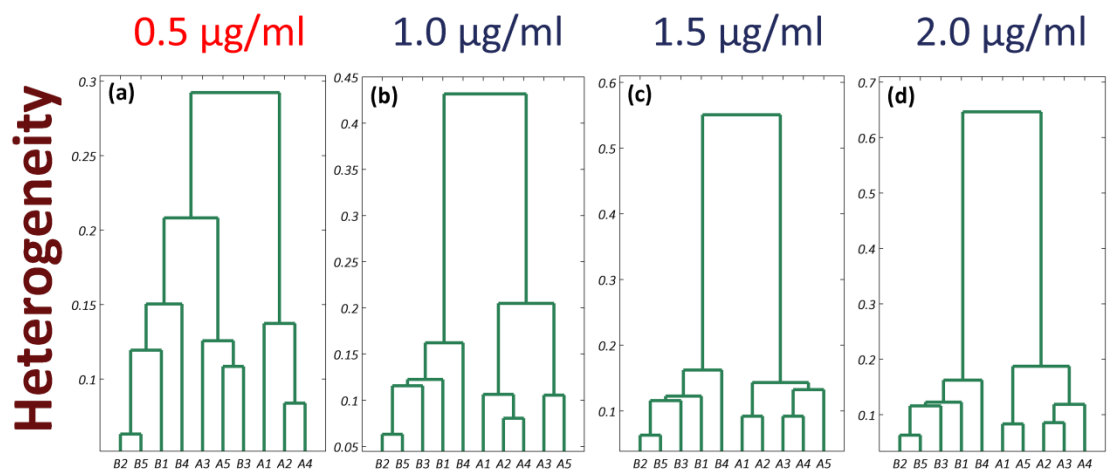


Figure S6. HCA dendrogram of the Raman spectra of *A. baumannii* ATCC 19606 treated with and without colistin, A1 to A5 are spectra of the bacteria exposed to the drug, B1 to B5 are spectra of unexposed bacteria, the concentration of colistin sulfate used in the test is 0.5 µg/ml (a), 1.0 µg/ml (b), 1.5 µg/ml (c) and 2.0 µg/ml (d) respectively.