## **Electronic Supplementary Information**

## Amplified colorimetric detection of tetracycline based on enzyme-linked aptamer

## assay with multivalent HRP-mimicking DNAzyme

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Name	Sequence (5' to 3')	Modification
Bio-aptamer	CGTACGGAATTCGCTAGCCCCCGGCAGGCCACGGC otamer TTGGGTTGGTCCCACTGCGCGTGGATCCGAGCTCCA CGTG	
H1	AGGGCGGGTGGGTGTTTAAGTTGGAGAATTGTACTT AAACACCTTCTTCTTGGGT	3-biotin
H2	H2 TGGGTCAATTCTCCAACTTAAACTAGAAGAAGGTGT TTAAGTTGGGTAGGGCGGG	
Trigger	AGAAGAAGGTGTTTAAGTA	/

 Table S1. Sequences of oligonucleotides uesd in this work



**Fig. S1** UV-vis absorption spectra of multivalent HRP-mimicking DNAzyme synthesized with 300 nM G-quadruplex-rich nanowires solution, 300 ng/mL SA, 300 nM Bio-aptamer, and 20 µM hemin. a: 300 nM G-quadruplex-rich nanowires solution; b: multivalent HRP-mimicking DNAzyme; c: 300 ng/mL SA; d : 300 nM Bio-aptamer; e: 20 µM hemin



Fig. S2 (A) UV-vis absorption spectra of TC-BSA solution before (a) and after (b) immobilization, the concentration of TC-BSA was 10  $\mu$ g/mL; (B) UV-vis absorption spectra of TC-BSA solution before (a) and after (b) immobilization, the concentration of TC-BSA was 50  $\mu$ g/mL

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**Fig. S3** Effect of coating buffer on the detection system, 10 mM Bicarbonate buffer (CB, 3 mM Na<sub>2</sub>CO<sub>3</sub>, 7 mM NaHCO<sub>3</sub>, pH 9.6), 10 mM phosphate buffer saline (PBS, 100  $\mu$ M KH<sub>2</sub>PO<sub>4</sub>, 500  $\mu$ M Na<sub>2</sub>HPO<sub>4</sub>(12H<sub>2</sub>O), 10 mM NaCl, 200 mM KCl, pH 7.4), and 10 mM Tris–HCl buffer with different pH.



**Fig. S4** Effect of blocking agents on the detection system, including 0.05% BSA, 1% BSA, and 0.05% defat dried milk



**Fig. S5** Effect of binding buffers on the detection system, buffer A: 20 mM Tris-HCl (pH 8.0), buffer B: 20 mM Tris-HCl (pH 8.0), 100 mM NaCl; buffer C: 20 mM Tris-HCl (pH 8.0), 100 mM NaCl, 2 mM MgCl<sub>2</sub>; buffer D: 20 mM Tris-HCl (pH 8.0), 100 mM NaCl, 2 mM MgCl<sub>2</sub>, 1 mM CaCl<sub>2</sub>



Fig. S6 Indirect competitive assay for TC detection using different concentrations of aptamer modified HRP-mimicking DNAzyme system (5, 10, 20 nM), the concentration of TC-BSA was  $4 \mu g/mL$ 



Fig. S7 Indirect competitive assay for TC detection in buffer using different concentrations of TC-BSA conjugate (2, 4, 8  $\mu$ g/mL), the concentration of aptamer modified HRP-mimicking DNAzyme was 10 nM



Fig. S8 Effect of competitive reaction time on the detection system, the concentration of TC-BSA and aptamer modified HRP-mimicking DNAzyme was 4  $\mu$ g/mL and 10 nM, respectively

Method	Keywords	Liner range	LOD	Ref.
ELISA	Biotin-avodin, HRP	3.16×10 <sup>-10</sup> -3.16×10 <sup>-7</sup> M	0.048 µg/ml	1
ELISA	Aptamer, HRP	0.01-100 ng/ml	9.6×10 <sup>-3</sup> ng/ml	2
HPLC	Liquid-liquid micro- extraction	5-15 μg/mL	0.95-3.6 μg/mL	3
HPLC	Solid phase extraction, Validation	50-500 ng/mL	21 ng/mL	4
Colorimeter	Colorimeter	0.5 -10 μg/mL	1.5 μg/mL	5
ELISA	Class-Specific monoclonal antibody	0.26 -2.0 μg/L	15 µg/L	6
Electrochemistry	Electrochemica aptasensor, M-shape structure	≤ 3000 nM	$\approx$ 329 ng/mL	7
Electrochemistry	PtNPs/C/GCE.	9.99-44.0 μM	$\approx$ 1.9 mg/ml	8
Electrochemistry	Aptamer biosensor	0.1-100 ng/mL	1 ng/mL	9
ELISA	Multivalent HRP- mimicking DNAzyme systems, Aptamer	10-2-10 <sup>4</sup> ng/mL	8.1×10 <sup>-2</sup> ng/ml	This work

 Table S2. Comparision of different methods for TC detection

Sample	Spiked concentration (ng/mL)	Detected concentration (ng/mL, mean±SD)	Recovery (%)
1	0.1	$0.086 \pm 0.0032$	86
2	1	$1.014 \pm 0.0012$	101.4
3	10	9.33±0.0021	93.3

Table S3. Recovery study

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