

Cascade toehold-mediated strand displacement strategy for label-free and sensitivity non-enzymatic recycling amplification detection of the *HIV-1* gene

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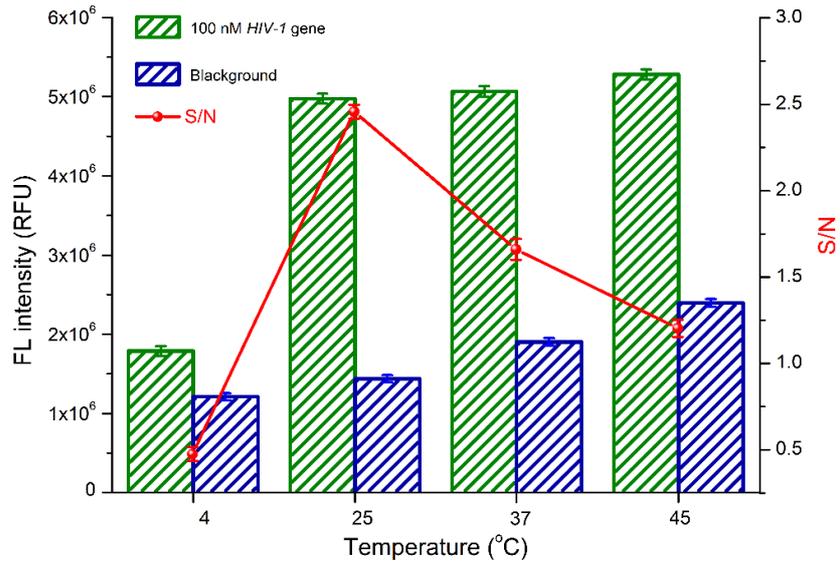


Fig. S1 Effect of the reaction temperature on the response of the sensing system. The histograms represent the fluorescence intensity of the solution in the presence of 100 nM *HIV-1* gene (green) and in the absence of the *HIV-1* gene (blue), respectively. The red line represents the S/N ratio. The corresponding error bars represent the standard deviation of three independent measurements obtained at each reaction temperature.

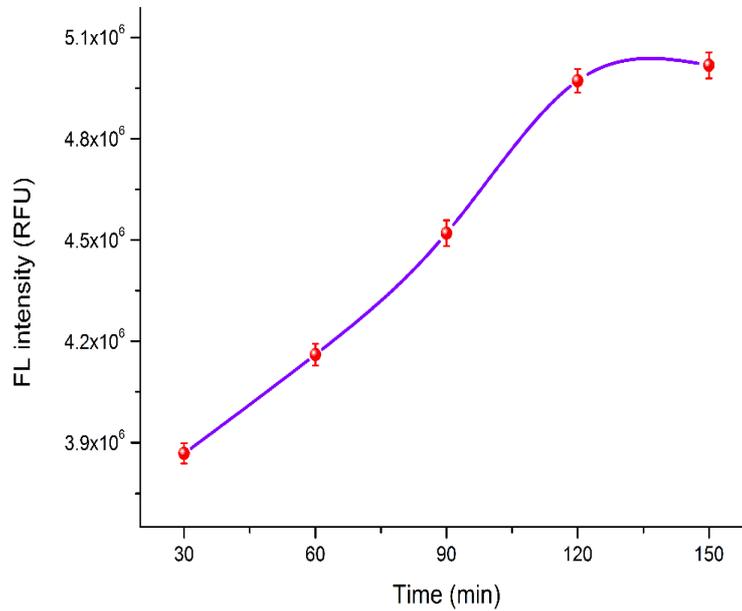


Fig. S2 Effect of the reaction time of the signal amplification on the fluorescence intensity of the proposed method for the detection of the *HIV-1* gene (100 nM). Reactions were performed at room temperature.

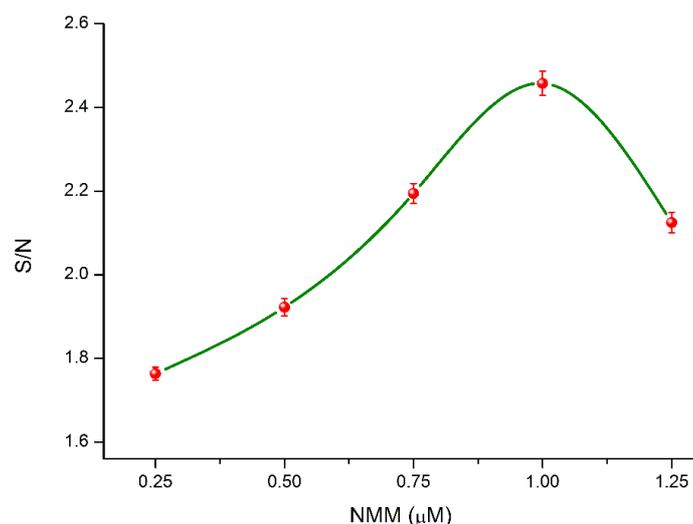


Fig. S3 Effect of the NMM concentration on the performance of the sensing system. The *HIV-1* gene concentration is 100 nM. Incubation temperature and reaction time were 25 °C and 120 min, respectively.

Table. S1 Comparison of the analytical methods capable of sensing the *HIV-1* gene

Method	Linear range	Detection Limit	References
Electrochemical biosensor based on amperometry	10 pM–50 nM	18 pM	1
Electrochemical biosensor based on a nafion-graphene composite	40 nM–2.56 μM	5 nM	2
Electrochemical biosensor based on a Closed Bipolar	100 pM–300 nM	30 pM	3
Electrochemical biosensor based on a triplex-oligonucleotide probe	0.1-10 nM	54 pM	4
Colorimetry biosensor base on a multi-amplification Nanoplatfrom	10 pM-120 nM	4.8 pM	5
Fluorescence biosensor base on grapheme oxide	7–80 nM	3 nM	6
Fluorescence biosensor base on fluorescence polarization	50 pM-8 nM	9.12 pM	7
Fluorescence biosensor base on TMSDRs	10 pM-1 μM	1.9 pM	This work

Reference

1. X. Gao, X. Wang, Y. Li, J. He and H. Yu, *Anal. Chem.*, 2018, **90**, pp 8147-8153.
2. B. Li , Z. Li, B. Situ, Z. Dai, Q. Liu, Q. Wang, D. Gu and L. Zheng, *Biosens. Bioelectron.*, 2014, **52**, 330-336.
3. R. Poorghasem, R. S. Saberi, M. Shayan, M. A. Mehrgardi and A. Kiani, *Electrochim. Acta.*, 2016, **222**, 1483-1490.
4. X. Wang, A. Jiang, T. Hou and F. Li, *Anal. Chim. Acta.*, 2015, **890**, 91-97.
5. Y. Long, C. Zhou, C. Wang, H. Cai, C. Yin, Q. Yang and D. Xiao, *Sci. Rep.*, 2016, **6**, 23949.
6. L. Chen, L. Song, Y. Zhang, P. Wang, Z. Xiao, Y. Guo and F. Cao, *ACS Appl. Mater. Inter.*, 2016, **8**, 11255-11261.
7. L. Wang, J. Tian, Y. Huang, X. Lin, W. Yang, Y. Zhao and S. Zhao, *Microchim. Acta.*, 2016, **183**, 2147-2153.