Supplementary Information for

Methodologies for the analysis of pesticides and pharmaceuticals in sediments and plant tissue

Pedro N. Carvalho^{1*†}, Yang Zhang^{1,2‡}, Tao Lyu^{1§}, Carlos A. Arias¹, Kai Bester³, Hans Brix¹

¹Department of Bioscience, Aarhus University, Ole Worms Allé 1, Building 1135, 8000 Aarhus C., Denmark

² College of Life Science, South China Normal University, Guangzhou 510631, PR China ³ Department of Environmental Science, Aarhus University, Frederiksborgvej 399, 4000 Roskilde, Denmark

* Corresponding author: Tel.: Tel.: +45 87158462 E-mail address: <u>pedro.carvalho@envs.au.dk</u> ORCID: 0000-0002-7131-9102

Table of Contents:

Table S1. Compounds studied description and chemical properties

M&M1. Solid-phase extraction procedures

Table S2. Instrument (HPLC-DAD) analytical figures of merit (LOD, LOQ, linearity range and R²)

Figure S1. Typical chromatograms for sediment extracts

Figure S2. Typical chromatograms for plant extracts

 ${\bf Table S3.} Comparison of the present method characteristics for sediment analysis with relevant multi-residue methods published$

Table S4. Comparison of the present method characteristics for plant analysis with other multi-residue methods published

Figure S3. Imazalil concentration in sediment samples

Figure S4. Ibuprofen concentration in plant tissue (leaves and roots)

[†]Current address: Department of Environmental Science, Aarhus University, Frederiksborgvej 399, 4000 Roskilde, Denmark

^{*} Current address: School of environmental Science and Engineering, Southern University of Science and Technology, Shenzhen 518055, PR China

[§] Current address: School of Animal, Rural and Environmental Sciences, Nottingham Trent University, Nottinghamshire NG25 oQF, UK

Systematic (common) name	Common	CASno.	Structural formula	MW	Log K _{ow}
Family / Application	addreviation			(g mol-1)	
Pesticides					
1,2-Benzisothiazol-3(2H)-one (Benzisothiazolinone)	BIT	2634-33-5	\sim	151	0.64 ^a
Isothiazolinones / Microbiocide and fungicide			NH		
Methyl-benzimidazol-2-yl carbamate (Carbendazim)	CD	10605-21-7		191	1.55 ^a
Carbamates / Fungicide			NH		
3-(3,4-dichloro-phenyl)-1,1-dimethylurea (Diuron)	DR	330-54-1		233	2.67 ^a
Phenylureas / herbicide			CI 0		
(<i>RS</i>)-1-[2-(Allyloxy)-2-(2,4- dichlorophenyl)ethyl]-1 <i>H</i> -imidazole (Imazalil)	IMZ	35554-44-0		297	4.10 ^a
Imidazoles / Fungicide and veterinary medicine					
2-(4-Chloro-2-methylphenoxy)propionic acid (Mecoprop)	МСРР	7085-19-0		214	2.9 4 ^a
Isothiazolinones / Herbicide			сі о он		
1-(4-Chlorophenyl)-4,4-dimethyl-3-(1 <i>H</i> - 1,2,4-triazol-1-ylmethyl)-3-pentanol (Tebuconazole)	TBU	107534-96-3	N HO	307	3.89 ª
Triazoles / Fungicide			CI		

Table S1. Compounds studied description and chemical properties

N ² -tert-butyl-N ⁴ -ethyl-6-(methylthio)- 1,3,5-triazine-2,4-diamine (Terbutryn) Triazines / Algaecide and herbicide	ТВ	886-50-0		241	3. 77 ^a
Antibacterial (analysed together with th	e pesticides)				
5-Chloro-2-(2,4-dichlorophenoxy)phenol (Triclosan)	TCS	3380-34-5	CI OH	289	4.66 ^a
Phenoxy phenols / Antimicrobial and fungicide			CI		
Pharmaceuticals					
5 <i>H</i> -Dibenz[<i>b</i> , <i>f</i>]azepine-5-carboxamide (Carbamazepine)	CBZ	298-46-4		236	2.25 ^a
- / Anticonvulsant			O NH ₂		
2-[(2,6- Dichlorophenyl)amino]benzeneacetic acid sodium salt (Diclofenac)	DCF	15307-79-6		296	4.02 ^a
- / Nonsteroidal anti-inflammatory drug (NSAID)					
(<i>RS</i>)-2-(4-(2- methylpropyl)phenyl)propanoic acid (Ibuprofen)	IBP	15687-27-1	ОН	206	3.79 ^a
- / Nonsteroidal anti-inflammatory drug (NSAID)					
1- <i>N</i> ,3- <i>N</i> -bis(2,3-dihydroxypropyl)-5-[<i>N</i> - (2,3-dihydroxypropyl)acetamido]-2,4,6- triiodobenzene-1,3-dicarboxamide (Iohexol)	IHE	66108-95-0	ОН	821	-4.16 ^b
- / Radiocontrast agents					

1- <i>N</i> ,3- <i>N-bis</i> (2,3-dihydroxypropyl)-5-(2- hydroxy- <i>N</i> -methylacetamido)-2,4,6- triiodobenzene-1,3-dicarboxamide (Iomeprol)	IME	78649-41-9		777	-3.08 ^b
- / Radiocontrast agents					
1- <i>N</i> ,3- <i>N-bis</i> (1,3-dihydroxypropan-2-yl)-5- [(2 <i>S</i>)-2-hydroxypropanamido]-2,4,6- triiodobenzene-1,3-dicarboxamide (Iopamidol)	IPA	60166-93-0		777	-2.54 ^b
- / Radiocontrast agents			ОН ОН		
1- <i>N</i> ,3- <i>N-bis</i> (2,3-dihydroxypropyl)-2,4,6- triiodo-5-(2-methoxyacetamido)-1- <i>N</i> - methylbenzene-1,3-dicarboxamide (Iopromide)	IPR	73334-07-3		791	-2.95 ^b
- / Radiocontrast agents					
(2S)-2-(6-methoxynaphthalen-2- yl)propanoic acid (Naproxen)	NPX	22204-53-1	ОН	230	3.10 ^a
- / Nonsteroidal anti-inflammatory drug (NSAID)					
(±)-1-Isopropylamino-3-(1-naphthyloxy)- 2-propanol hydrochloride (Propanolol)	PPL	318-98-9	OH H O N	259	2.60 ^a
- / Beta blocker					

^a Values from EPISuites ^b Values from ACD/Labs

M&M1. Solid-phase extraction (SPE) procedures

Strata-X cartridges were conditioned with 5 mL of MeOH followed by 5 mL of MilliQ water and a 100 mL sample (water:methanol 95:5, v/v) was loaded. After washing with 5 mL of water:methanol (95:5, v/v), the cartridges were dried for 30 min prior to being eluted with 5 mL of methanol:formic acid (90:10). Florisil cartridges were conditioned with 5 mL of *n*-hexane followed by loading of 5 mL methanolic samples. After washing with 5 mL of *n*-hexane:acetone (99:1, v/v), cartridges were dried for 5 min prior to being eluted with 5 mL of *n*-hexane:acetone (90:10, v/v).

Destinides	λ	Retention time	Linear range	R ²	ILOD	ILOQ
resticides	(nm)	(min)	(mg L-1)	(n=8)	(mg L-1)	(mg L-1)
Carbendazim	240	9.6	0.2 -10	0.9968	0.05	0.2
Benzoisothiazolinone	240	10.7	0.1 -10	0.9964	0.01	0.05
Imazalil	220	11.5	0.5 -10	0.9974	0.2	0.5
Terbutryn	240	12.2	0.1 -10	0.9957	0.01	0.05
Diuron	240	12.5	0.1 -10	0.9951	0.01	0.05
Mecoprop	220	13.0	0.3 -10	0.9965	0.1	0.3
Tebuconazole	220	14.3	0.4 -10	0.9959	0.1	0.4
Triclosan	240	15.3	0.2 -10	0.9967	0.08	0.2
Pharmaceuticals	λ	Retention time	Linear range	R ²	LOD	LOQ
	(nm)	(min)	(mg L-1)	(n=8)	(mg L-1)	(mg L-1)
Iopamidol	240	8.4	0.2 -10	0.9951	0.05	0.2
Iohexol	240	8.9	0.3 -10	0.9952	0.1	0.3
Iomeprol	240	9.0	0.3 -10	0.9947	0.1	0.3
Iopromide	240	9.4	0.2 -10	0.9943	0.05	0.2
Propanolol	240	11.2	0.2 -10	0.9947	0.03	0.2
Carbamazepine	240	12.4	0.1 -10	0.9945	0.05	0.1
Naproxen	240	13.5	0.1 -10	0.9942	0.05	0.1
Ibuprofen	220	13.8	0.3 -10	0.9951	0.1	0.3
Diclofenac	240	14.7	0.2 -10	0.9964	0.05	0.2

Table S2. Instrument (HPLC-DAD) analytical figures of merit (LOD, LOQ,
linearity range and R^2)*

* determined before any sample extraction and clean-up step using analytical standards and methanolic solutions



Figure S1. Typical chromatogram (DAD λ = 220 nm; background blank corrected) obtained for standard solution (**■**) and hexane (**■**); methanol:formic acid 96:4 v:v (**■**); methanol:acetone 95:5 v:v (**■**) extracts of spiked sediment (0.5 µg micropollutant g_{dry sediment}⁻¹ spiking level, theoretically 10 ng of micropollutant injected) after 30 min of ultrasonication



Figure S2. Typical chromatogram (DAD $\lambda = 220$ nm; background blank corrected) obtained for standard solution (**■**) and methanol: acetone 95:5 v:v extracts of spiked plant material (5 µg micropollutant gdry sediment⁻¹ spiking level; theoretically 10 ng of micropollutant injected) (**■**) and non-spiked plant material (**■**) after 30 min of ultrasonication, no clean-up

Pesticides					
Reference	Methodology	LOD (ng g-1)	LOQ (ng g-1)	RSD (%)	Recovery (%)
This work	USE-HPLC-DAD	5 - 100	25 - 250	4 - 30	53 – 101
[13]	PLE-LL-LC-HRMS/MS	0.01 - 4	0.03 - 14	1 - 30	57 - 139
[15]	USE-SPE-LC-MS/MS	0.7 - 1.2	-	9 - 12	68 - 102
[29]	*LC-MS/MS	3	10	26 - 45	40 - 60
[34]	QuEChERS-LC-MS/MS	0.1 - 2	1 - 6	8	46 - 102
[37]	MAE-SPME-GC-MS	0.01 - 0.1	0.02 - 0.45	7 - 17	86 - 112
[39]	SLE-LC-MS/MS	-	0.1 - 49	3 - 28	40 - 125
[40]	PLE-SPE-LC-MS/MS	0.02 - 17		2 - 17	67 - 118
Pl	narmaceuticals				
Reference	Methodology	LOD (ng g-1)	LOQ (ng g-1)	RSD (%)	Recovery (%)
This work	USE-HPLC-DAD	15 -50	50 - 150	6 - 12	50 - 98
[11]	MAE-GC-MS	30 - 80	-	< 11	25 - 81
[11]	MAE-HPLC-DAD	< 167	-	< 11	> 70
[13]	PLE-LL-LC-HRMS/MS	0.01 - 4	0.03 - 14	1 - 30	57 - 139
[15]	USE-SPE-LC-MS/MS	0.9 - 9.4	-	6 - 14	41 - 92
[21]	USE-SPE-LC-MS/MS	0.08 - 4.2	-	-	41 - 151
[30]	MAE-SPE-GC-MS	0.3 - 5.7	0.9 – 17.1	0.7 - 14.7	> 50
[35]	MAE- LC-MS/MS	0.3	1.0	1 - 8	98 - 103
[36]	USE-SPE-LC-MS/MS	-	0.6 – 5.6	-	< 10 - 343
[38]	USE-SPE-HPLC-DAD/FL	-	1 - 187	-	< 15 - 103

Table S3. Comparison of the present method characteristics for sedimentanalysis with relevant multi-residue methods published

USE – Ultrasonic solvent extraction, HPLC – High performance liquid chromatography, DAD diode array detector, PLE - pressurized liquid extraction, LL – liquid-liquid partitioning clean-up, LCliquid chromatography, HRMS/MS high resolution Orbitrap mass spectrometry, SPE – solidphase extraction, MS/MS – tandem mass spectrometry, *extraction not stated, QuEChERS - Quick Easy Cheap Effective Rugged Safe method, DMAE - dynamic microwave-assisted extraction, CFME - continuous-flow microextraction, GC – gas chromatography, MS – mass spectrometry, MAE – microwave assisted extraction, SPME – solid-phase microextraction, SLE – solid-liquid extraction, FL - fluorescence detectors,

Pesticides					
Reference	Methodology	LOD (ng g ⁻¹)	LOQ (ng g ⁻¹)	RSD (%)	Recovery (%)
This work	USE-HPLC-DAD	50 - 1000	250 - 2500	4 - 30	53 – 101
[29]	*LC-MS/MS	3	10	21 - 43	45 - 50
[41]	DMAE-CFME-GC-MS	0.6 – 1.6	2 - 5	9	81 - 107
[42]	dispersive-SPE-LC-MS/MS		10	3 - 16	72 - 104
[43]	QuEChERS-LC-MS/MS		10 - 50	< 20	80 - 136
[44]	SLE-SO-SPE-GC-MS/MS		10	< 20	70 - 120
-1	· 1				
Pharmaceut	ticals				
Pharmaceut Reference	Methodology	LOD (ng g ⁻¹)	LOQ (ng g-1)	RSD (%)	Recovery (%)
Pharmaceut Reference This work	Methodology USE-HPLC-DAD	LOD (ng g ⁻¹) 150 - 500	LOQ (ng g ⁻¹) 500 - 1500	RSD (%) 6 - 12	Recovery (%) 50 - 98
Pharmaceut Reference This work [14]	Methodology USE-HPLC-DAD USE-SPE-LC-MS/MS	LOD (ng g ⁻¹) 150 - 500 0.5 - 1.5	LOQ (ng g ⁻¹) 500 - 1500 2 - 4	RSD (%) 6 - 12 -	Recovery (%) 50 - 98 73 - 92
Pharmaceut Reference This work [14] [14]	Methodology USE-HPLC-DAD USE-SPE-LC-MS/MS BE-SPE-GC-MS	LOD (ng g ⁻¹) 150 - 500 0.5 - 1.5 10 - 20	LOQ (ng g ⁻¹) 500 - 1500 2 - 4 20 - 75	RSD (%) 6 - 12 - -	Recovery (%) 50 - 98 73 - 92 15 - 98
Pharmaceut Reference This work [14] [14] [14]	Methodology USE-HPLC-DAD USE-SPE-LC-MS/MS BE-SPE-GC-MS PLE- SPE-LC-MS	LOD (ng g ⁻¹) 150 - 500 0.5 - 1.5 10 - 20 2 - 23	LOQ (ng g ⁻¹) 500 - 1500 2 - 4 20 - 75 -	RSD (%) 6 - 12 - -	Recovery (%) 50 - 98 73 - 92 15 - 98 46 - 176
Pharmaceut Reference This work [14] [14] [14] [14]	Methodology USE-HPLC-DAD USE-SPE-LC-MS/MS BE-SPE-GC-MS PLE- SPE-LC-MS BE-SPE-LC-MS/MS	LOD (ng g ⁻¹) 150 - 500 0.5 - 1.5 10 - 20 2 - 23 25	LOQ (ng g ⁻¹) 500 - 1500 2 - 4 20 - 75 - -	RSD (%) 6 - 12 - - - -	Recovery (%) 50 - 98 73 - 92 15 - 98 46 - 176 40 - 120
Pharmaceut Reference This work [14] [14] [14] [14] [14]	Methodology USE-HPLC-DAD USE-SPE-LC-MS/MS BE-SPE-GC-MS PLE- SPE-LC-MS BE-SPE-LC-MS/MS PLE- SPE-GC-MS	LOD (ng g ⁻¹) 150 - 500 0.5 - 1.5 10 - 20 2 - 23 25 7 - 58	LOQ (ng g ⁻¹) 500 - 1500 2 - 4 20 - 75 - - -	RSD (%) 6 - 12 - - - - - -	Recovery (%) 50 - 98 73 - 92 15 - 98 46 - 176 40 - 120 46 - 94
Pharmaceut Reference This work [14] [14] [14] [14] [14] [45]	Methodology USE-HPLC-DAD USE-SPE-LC-MS/MS BE-SPE-GC-MS PLE- SPE-LC-MS BE-SPE-LC-MS/MS PLE- SPE-GC-MS PLE-LC-MS/MS	LOD (ng g ⁻¹) 150 - 500 0.5 - 1.5 10 - 20 2 - 23 25 7 - 58 2 - 12	LOQ (ng g ⁻¹) 500 - 1500 2 - 4 20 - 75 - - - - -	RSD (%) 6 - 12 - - - - - - - - < 20	Recovery (%) 50 - 98 73 - 92 15 - 98 46 - 176 40 - 120 46 - 94 70 - 134

Table S4. Comparison of the present method characteristics for plant analysis with other multi-residue methods published

USE – Ultrasonic solvent extraction, HPLC – High performance liquid chromatography, DAD diode array detector, dispersive-SPE - dispersive solid phase extraction, LC- liquid chromatography, MS/MS – tandem mass spectrometry, *extraction not stated, SLE – solid-liquid extraction, SO – salting out, SPE – solid-phase extraction, GC – gas chromatography, QuEChERS - Quick Easy Cheap Effective Rugged Safe method, BE – buffer extraction, MS – mass spectrometry, PLE pressurized liquid extraction



Figure S3. Imazalil concentration in substrate samples (sediment) from 6 different constructed wetland bed mesocosms (unplanted and planted with different plants) continuously run over 9 months under various hydraulic loading rates and imazalil concentration of both 10 and 100 μ g L⁻¹ in the influent



Figure S4. Ibuprofen concentration in plant tissue (leaves and roots) in different plant species after exposure for 24 days in spiked hydroponic media (water concentration of 10 mg L⁻¹)

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