

Supporting Information for

Label-free and sensitive detection of Uracil-DNA glycosylase using exponential real-time rolling circle amplification

Yuan Xu, Yun-Xi Cui, Qiu-Ge Zhao, An-Na Tang*, and De-Ming Kong*

*State Key Laboratory of Medicinal Chemical Biology, Tianjin Key Laboratory of
Biosensing and Molecular Recognition, Collaborative Innovation Center of Chemical
Science and Engineering, College of Chemistry, Nankai University, Tianjin, 300071,
PR China*

Submitted to **Analytical Methods**, April. 02, 2018

* Correspondence: Dr. An-Na Tang, College of Chemistry, Nankai University, 94 Weijin Road,

Tianjin, 300071, P. R. China

E-mail: tanganna@nankai.edu.cn

Fax: +86-22-23500938

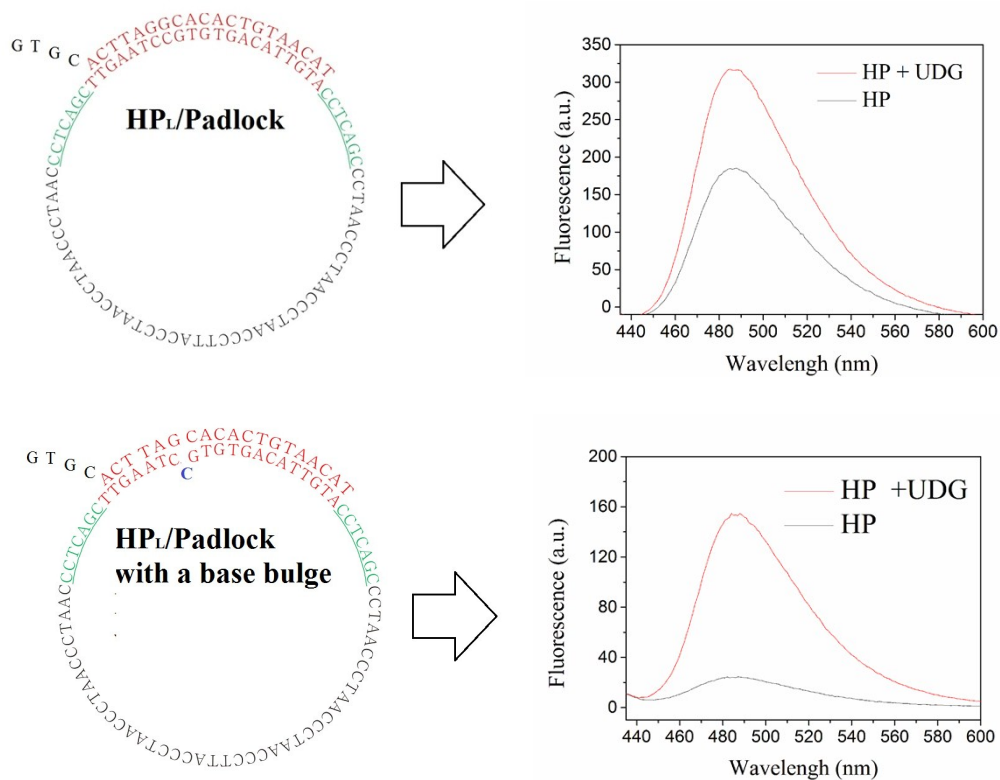
Dr. De-Ming Kong, College of Chemistry, Nankai University, 94 Weijin Road, Tianjin, 300071, P. R.

China

E-mail: kongdem@nankai.edu.cn

Fax: +86-22-23502458

1. HP_L/Padlock complex with a base bulge



Scheme S1. Comparison of HP_L/Padlock complexes without or with a base bulge. (Left), Schematic representation of HP_L/Padlock complexes; (Right) Fluorescence signals given by corresponding UDG-sensing systems.

2. Optimization of exponential RCA-based UDG-sensing platform

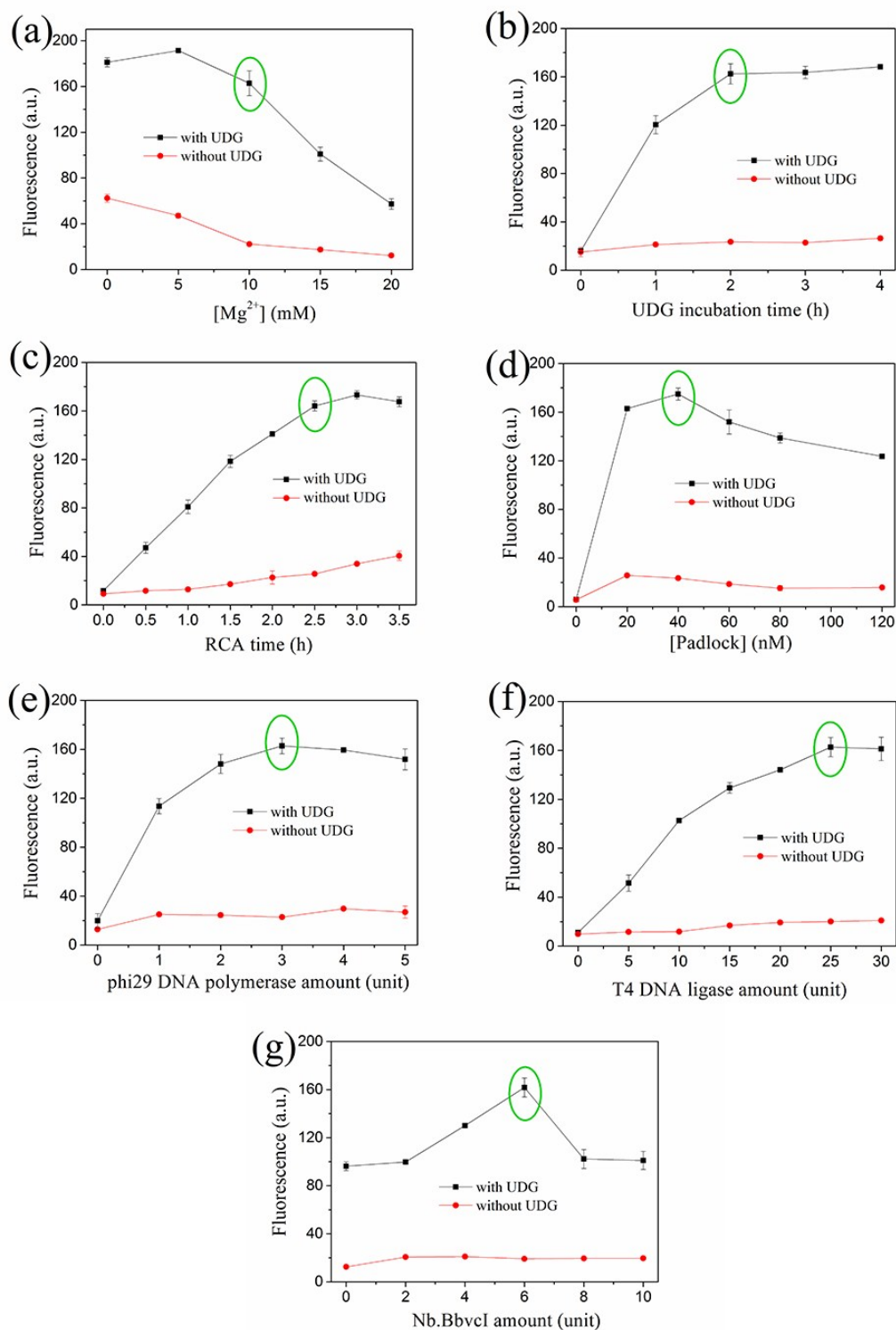


Fig. S1. Optimization of Mg^{2+} concentration (a), UDG incubation time (b), RCA reaction time (c), **Padlock** concentration (d) amounts of phi29 DNA polymerase (e), T4 DNA ligase (f) and Nb.BbvCI (g) in exponential RCA-based UDG-sensing system. The data were expressed as mean \pm σ (n=3).

3. Linear RCA-based UDG-sensing platform

3.1 Optimization of experimental conditions

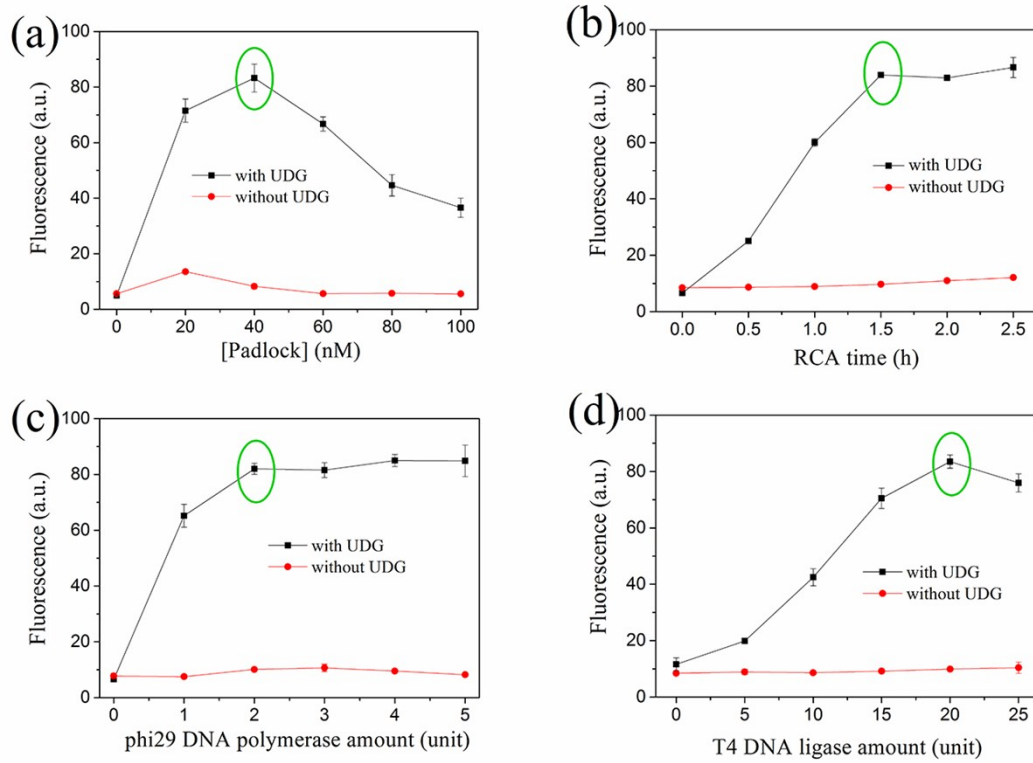


Fig. S2. Optimization of **Padlock** concentration (a), RCA reaction time (b), amounts of phi29 DNA polymerase (c) and T4 DNA ligase (d) in linearly amplified RCA-based UDG-sensing system. The data were expressed as mean \pm σ (n=3).

3.2 UDG activity detection using linear RCA-based sensing platform

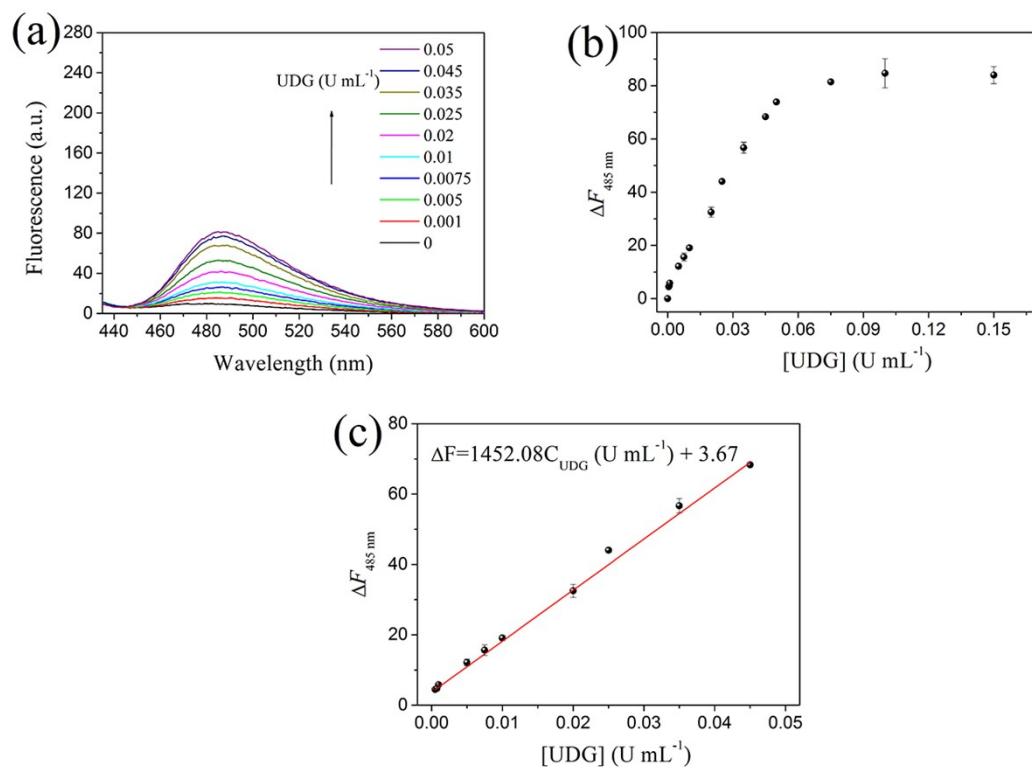


Fig. S3. UDG activity detection by linear RCA-based sensing platform. (a) Fluorescence spectra of the sensing systems containing different concentrations of UDG. (b) UDG activity-dependent fluorescence intensity change (ΔF) at 485 nm. (c) The linear relationship between ΔF and UDG concentration in the range of $1 \times 10^{-3} \sim 0.045$ U mL⁻¹. The data were expressed as mean \pm σ (n=3).

4. Comparison of our method with other reported ones

Table S1. Detection limits and real sample applications of some UDG detection methods

Method	System	Detection limit (U mL ⁻¹)	Real sample application	Reference
Colorimetric	Formation of a G-quadruplex–cofactor complex	0.008	Not given	[12]
Colorimetric	Nicking enzyme assisted chain replace amplification	0.02	Not given	[13]
Luminescent	UDG assisted pyridinium luminescent switch-on molecular probe	0.005	Diluted serum samples with spiked UDG	[14]
Fluorescence	Enzyme-assisted bicyclic cascade signal	0.0001	Cell extract	[16]
Fluorescence	Intrinsically fluorescent nucleotides	0.0025	Cell extract	[17]
Fluorescence	Inserted fluorescence probe	0.0008	Cell extract	[18]
Fluorescence	Synthesis a G-quadruplex probe	0.02	Not given	[19]
Fluorescence	Real time monitoring uracil excision via labeled molecular beacons	0.005	Cell extract with spiked UDG	[21]
Fluorescence	Real-time strand displacement amplification	6×10 ⁻⁵	Cell extract	[25]
Fluorescence	Real-time exponential RCA	5.5×10 ⁻⁵	Cell extract	This work