

Electronic Supplementary Information (ESI) for:

**Sensitive electrochemical aptasensor for detection of Aflatoxin B2  
based on polyacrylamide/phytic acid/polydopamine hydrogel  
modified screen printed carbon electrode**

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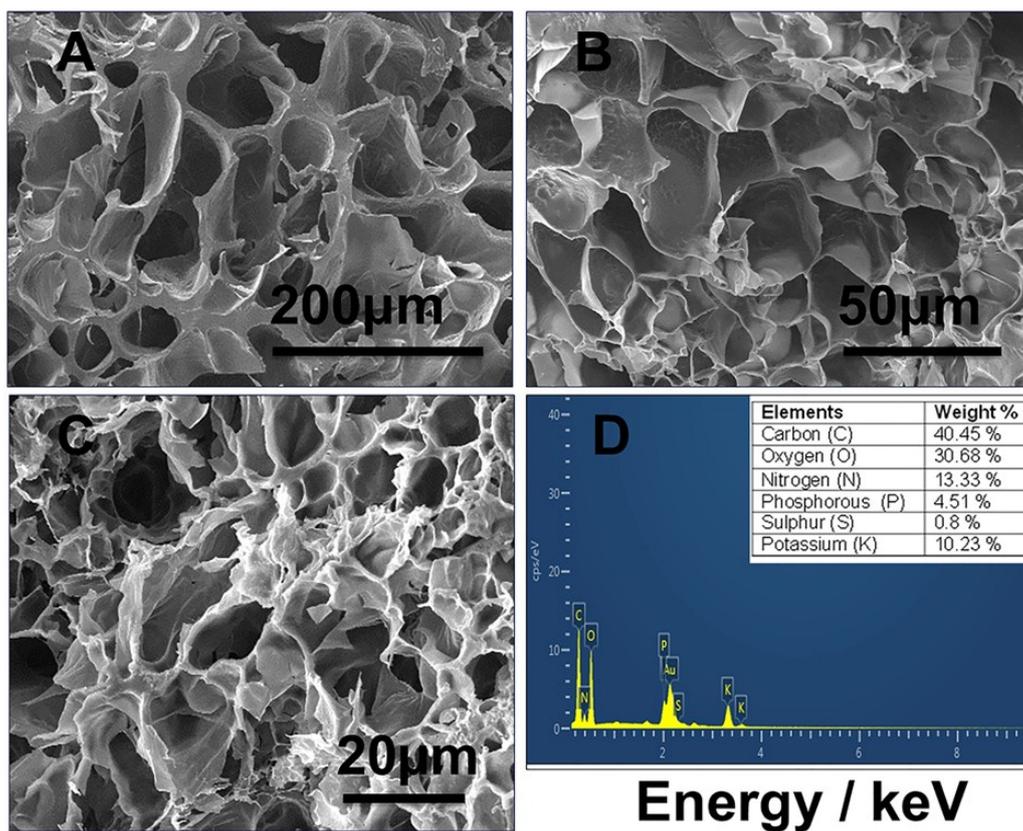
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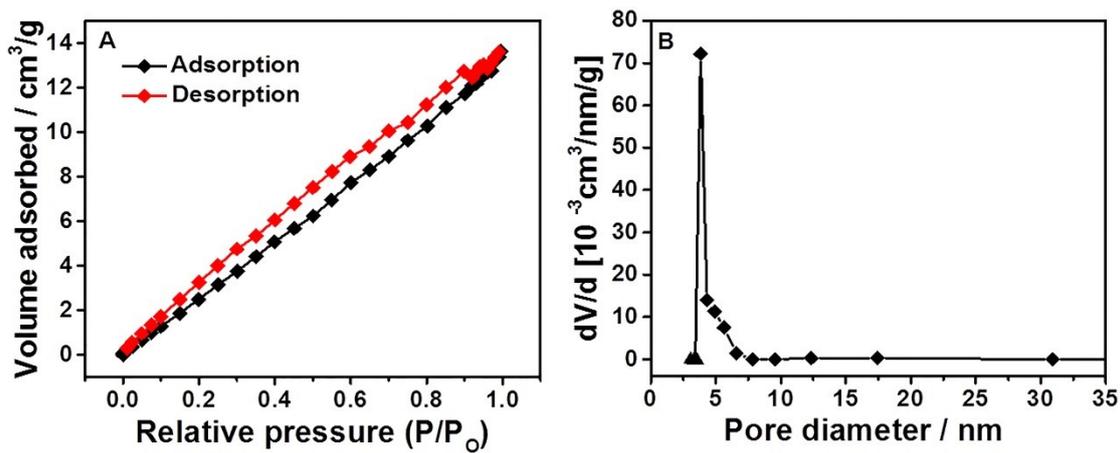
## 2. Additional Figures



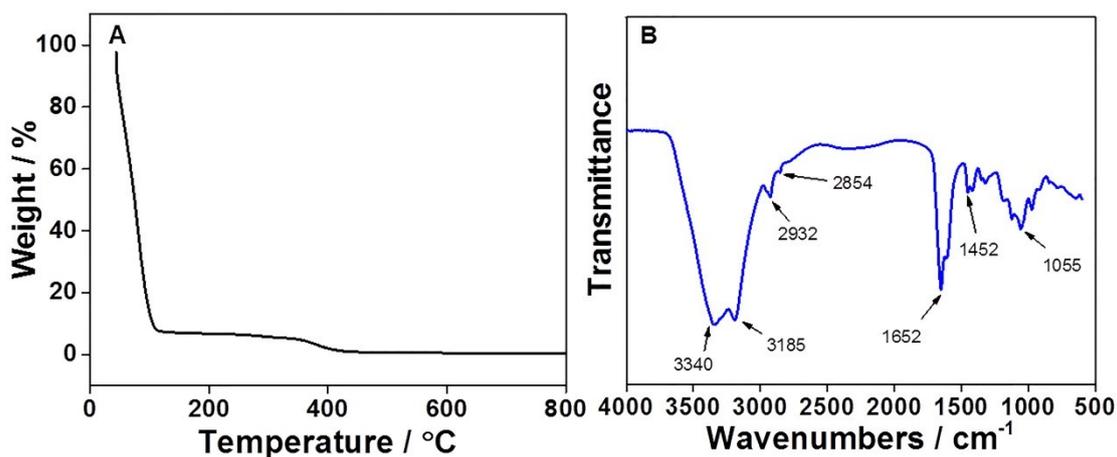
**Fig.S1** A photograph of the PAM/PA/PDA hydrogel inside a glass vial.



**Fig.S2** (A, B and C) SEM micrographs of PAM/PA/PDA hydrogel of different magnification and (D) EDX of PAM/PA/PDA hydrogel



**Fig.S3** (A) N<sub>2</sub> adsorption-desorption isotherms, (B) BJH pore-size distribution of dehydrated PAM/PA/PDA hydrogel.



**Fig.S4** (A) Thermal gravimetric analysis of PAM/PA/PDA hydrogel and (B) FTIR spectrum of PAM/PA/PDA hydrogel.

The H<sub>2</sub>O content of the PAM/PA/PDA hydrogel was around 90 % (wt/wt) (as shown in Fig.S4A). The high H<sub>2</sub>O content arises from hydrophilic functional groups attached to the polymeric backbone, which provides a favourable microenvironment for immobilizing biomolecules.<sup>S1,S2</sup> The FTIR spectrum of PAM/PA/PDA hydrogel is shown in Fig.S4B. The characteristic peak at 3340 cm<sup>-1</sup> and 3185 cm<sup>-1</sup> are assigned to asymmetric and symmetric stretching vibrations of NH<sub>2</sub>.<sup>S3</sup> Whereas the broad peak around 3300 cm<sup>-1</sup> is attributed to stretching vibrations of OH that indicate an abundance of hydroxyl groups.<sup>S4</sup> In addition, the strong absorption peaks at 1652 cm<sup>-1</sup> and 1609 cm<sup>-1</sup> are arised from the vibrations of C=O and C=C, respectively.<sup>S5</sup> Beside the band at 1452 cm<sup>-1</sup> is associated with the vibration of C-N (C-N stretching for primary amide), and the characteristic bands of PO<sub>4</sub><sup>3-</sup> are appeared at 1055 cm<sup>-1</sup>. This result confirmed presence of abundant functional groups on the surface of PAM/PA/PDA hydrogel which is considered advantageous for immobilizing aptamer onto an electrode surface.

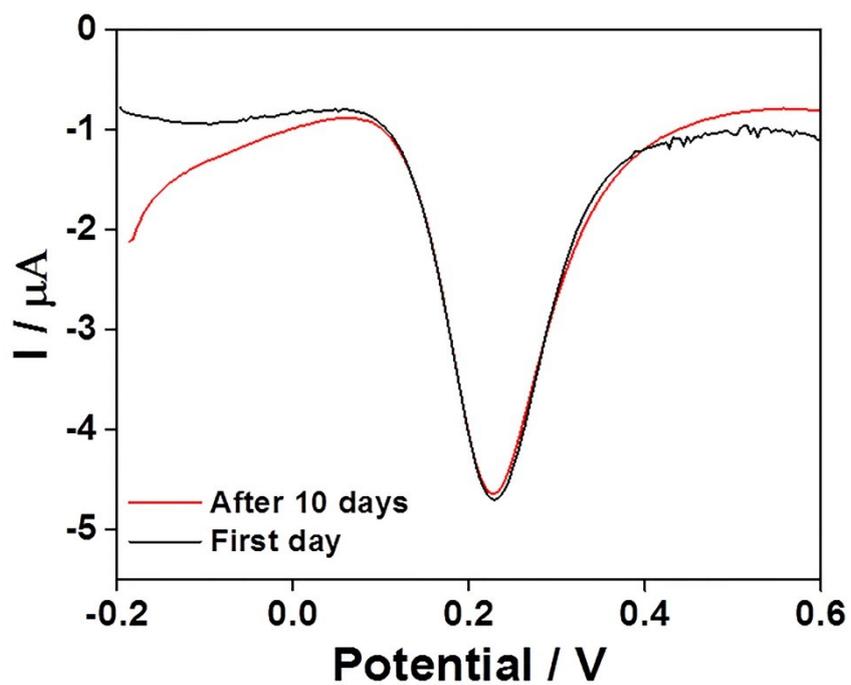
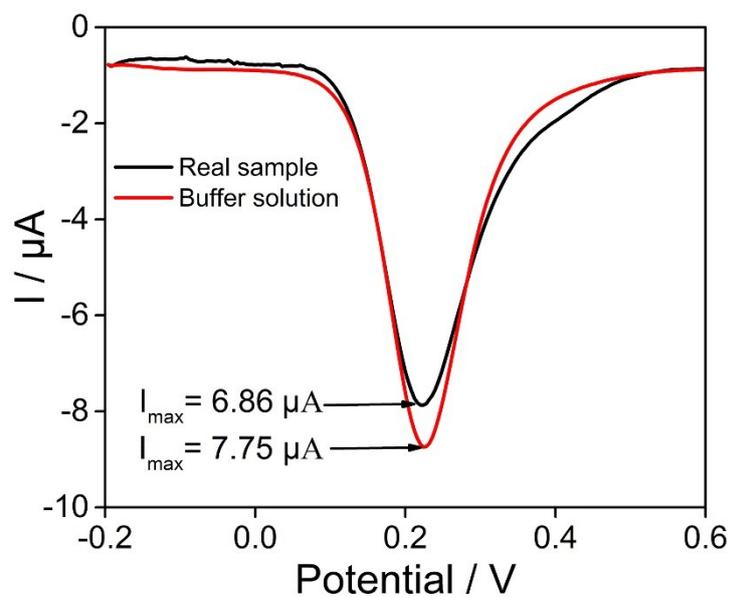


Fig.S5 Stability of the proposed SPCE/PAM/PA/PDA/Apt.



**Fig.S6** DPV responses of the SPCE/PAM/PA/PDA/Apt in buffer and real samples containing  $[\text{Fe}(\text{CN})_6]^{3-/4-}$  redox couple ( $5 \text{ mmol L}^{-1}$ ) without AFB2.

### 3. Additional Table

**Table S1.** Optimization of precursor concentrations and reaction conditions for preparation of PAM/PA/PDA hydrogel

PAM/PA/PDA hydrogels	DA/AM (wt%)	AM (g)	KPS/AM (wt%)	TMEDA ( $\mu$ L)	MBA/AM (wt%)	Water ( $\mu$ L)	PA/AM (wt%)	Time (min)
A	0.5	0.28	8	20	0.27	250	0.31	1
B	0.5	0.28	8	5	0.27	250	0.31	3
C	0.5	0.28	8	1	0.27	250	0.31	8
D	0.8	0.28	8	2	0.27	250	0.31	15
E	0.8	0.28	8	0.5	0.27	250	0.31	120
F	4.65	0.28	8	0.5	0.27	250	0.31	-

**Chemicals used:** dopamine (DA), acrylamide (AM), potassium peroxydisulfate (KPS), N, N'-Methylene bisacrylamide (MBA) and tetramethylethylenediamine (TMEDA).

The polymerization parameters such as reaction time, monomer concentration, and initiator concentration were optimized to obtain PAM/PA/PDA hydrogels of desired properties. As shown in Table S1, the PAM/PA/PDA hydrogel cannot form when the weight ratios of DA/AM were higher than 0.8wt%, because the reductive DA molecules affected the activity of the initiator (KPS) and thus retard the polymerization of AM monomers.<sup>S6</sup> In this regard, row D was selected for preparation of PAM/PA/PDA hydrogel by taking into consideration the modification time of SPCEs.

#### 4. Additional References

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