Supplementary Data

Electronic supplementary information

Movie S1 Video showing trapping of hydrogels in microfluidic chip.

Fig. S1. (A) Photograph of the toaster-sized reader used for flow automation and fluorescence detection (B) Fluidic mechanism showing how liquids are dispensed into the channel and flown through suction pumping. (C) Device for hydrogel fabrication off chip.



Fig. S2. (A) Image of a hydrogel sensor captured at the end of the assay. (B) A uniform signal is seen across the length of the gel as measured by the fluorescent intensity from end-to-end.







Fig. S3. Bland altman analysis of (A) β -HCG (B) FSH (C) LH (D) PRL hormone tests.

	Hydrogel size (µm)						
β-HCG	Before	After drying	Recovery				
	drying	and hydration	(%)				
Length	525	505	96.2				
Width	356	354	99.4				
Height	140	129	92.1				
FSH							
Length	510	497	97.5				
Width	343	336	98.0				
Height	135	125	92.6				
LH							
Length	487	490	100.6				
Width	345	337	97.7				
Height	139	136	97.8				
PRL							
Length	510	476	93.3				
Width	363	339	93.4				
Height	143	139	97.2				
Negative							
Length	510	470	92.2				
Width	355	334	94.1				
Height	135	131	97.0				

Table S1. Size recovery of hydrogels post drying and hydration.

Note: Hydrogel size recovery was tested after 1 week of storage of dried hydrogels at 4° C in vacuum. N=3, Standard deviation was $\leq 10\%$.

Table S2. Intra-batch precision: assay precision tested within same batch of hydrogels for three different hormone concentrations in triplicate.

FSH	x	σ	CV			
(mIU/mL)	А	0	C.v.			
5	4.9	0.2	4.6			
10	8.3	0.8	1.6			
50	43.4	9.7	3.6			
LH (mIU/mL)						
5	5.35	0.4	8.1			
25	22.4	1.2	5.5			
100	99.5	9.5	9.6			
PRL (ng/mL)						
10	9.77	0.4	3.7			
25	22.9	0.2	0.9			
100	104.73	10	9.5			
β-hCG (mIU/mL)						
25	30.45	0.63	2.08			
500	495.33	39.2	7.91			
1000	998.71	11.26	1.12			

Hormone	Concentration	Ν	Х	σ	C.V.
FSH	50	11	99.9	7.77	7.78
LH	100	7	101	9.96	9.86
PRL	100	5	98	8.86	9.04
β-HCG	100	7	97.71	7.38	7.56

Table S3. Inter-batch precision: Assay precision tested within different hydrogelbatches for one hormone concentration in triplicate.

		Hormone used				
		FSH	LH	TSH	β-HCG	PRL
	FSH	100	0.7	1.25	0.62	ND
Capture	LH	0	100	1.25	0.28	ND
antibody	PRL	0	0	0	0	100
	β-hCG	0	0	0	100	ND

Table S4. Cross-reactivity (%) of capture antibody was tested using highestconcentration of hormones used in the study.

N=3, Standard deviation was $\leq 10\%$.

Hormone	Х	σ	C.V.
β-hCG	452	5.65	1.25
FSH	45.42	1.34	2.94
LH	83.3	0.91	1.09
PRL	161.73	8.91	5.51

Table S5. Effect of hemoglobin interference on assay.

Calibrator concentration used for interference study was β -hCG- 500 mIU/mL, FSH- 50 mIU/mL, LH- 100 mIU/mL and PRL- 150 ng/mL.

Table S6. Stability of β -HCG, FSH, LH, and PRL capture antibody in hydrogels	
stored and dried in the presence of stabilizers.	

	Stabilizer and hydrogels containing capture antibody						
Day	Liquid plate sealer	0.45 M Trehalose + 1% PEG 3.4 K					
	HCG	FSH	LH	PRL			
0	100	100	100	100			
7	105	97	98	108			
10	ND	100	101	80			
15	97	92	85	83			
21	ND	88	94	76			
30	99	ND	ND	ND			
45	95	ND	ND	ND			

N=3, Standard deviation was \leq 10%. ND: not determined.

		FSH		LH		H PRL			
Days	Х	σ	C.V.	Х	σ	C.V.	Х	σ	C.V.
80	95.5	4.9	10.7	94.6	6.2	6.4	86.7	11.7	13.5
120	99.3	0.8	1.8	105.1	3.3	3.9	103.6	13.1	12.7
180	75.9	1.8	4.6	83.5	4.2	4.5	113.1	2.6	2.3

Table S7. Stability of FSH, LH and PRL detection antibody on conjugate pad stored in vacuum desiccator at 4°C.

Table S8. Cost of different cartridge elements for a batch of 20,000 car	tridges
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Item	Cost (USD)
Plastic disposables (including cartridge and other consumables)	0.53
Antibodies and other reagents (for 3 tests FSH, LH, Prolactin)	0.59
Packaging	0.09
Total	1.21