

## Supporting information for:

# Simple and rational design of polymer nano-platform for high performance of HCV related miR-122 reduction in liver

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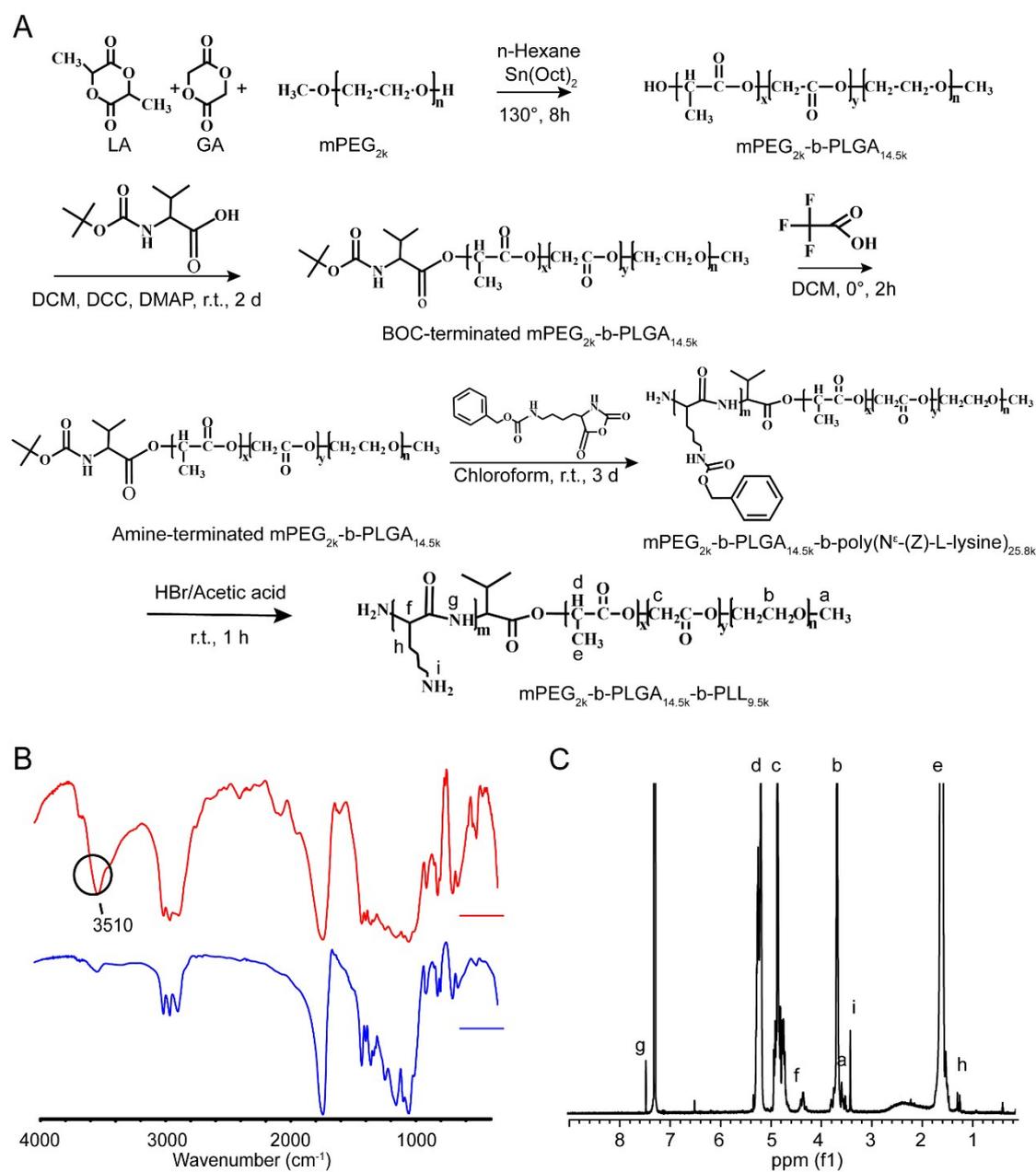
## Supplementary Figures:

**Table S1.** AN-loaded mPEG-b-PLGA-b-PLL nanoparticle formulations and properties

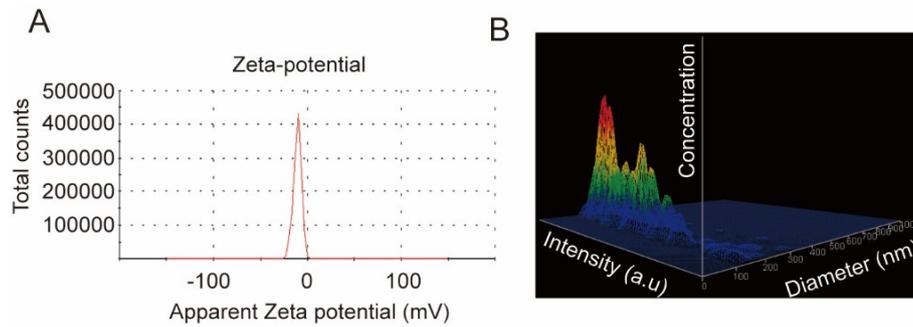
Formulation	antimiR in feed (nmol)	EE (%)	DL (%)	Size(nm)	PDI	Zeta potential(mV)
NP <sub>2.5</sub>	2.5	> 90	0.213	118.5	0.204	+22
NP <sub>5.0</sub>	5	> 85	0.426	136.2	0.241	-5.4
NP <sub>10.0</sub>	10	> 80	0.851	155.7	0.268	-12

**Table S2.** IN-loaded mPEG-b-PLGA-b-PLL nanoparticle formulations and properties

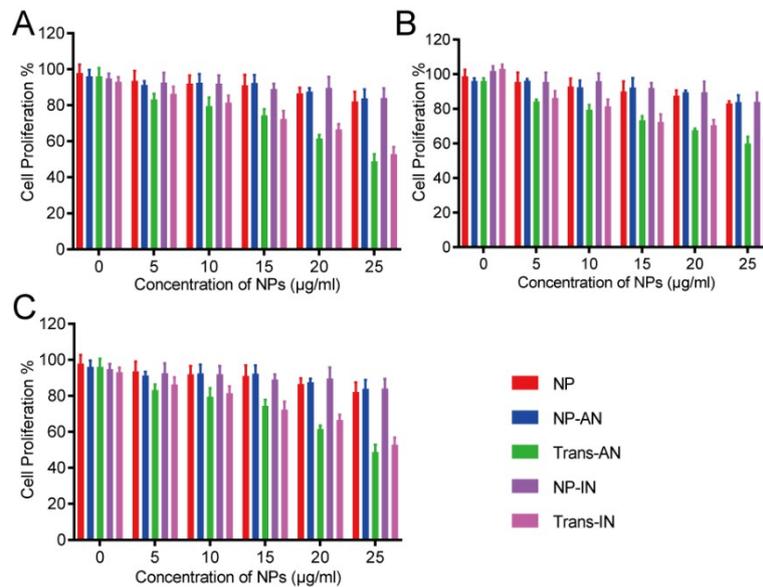
Formulation	antimiR in feed (nmol)	EE (%)	DL (%)	Size(nm)	PDI	Zeta potential(mV)
NP <sub>2.5</sub>	2.5	> 90	0.204	120.9	0.192	+24
NP <sub>5.0</sub>	5	> 80	0.417	140.6	0.217	-4.2
NP <sub>10.0</sub>	10	> 80	0.824	153.1	0.253	-10.5



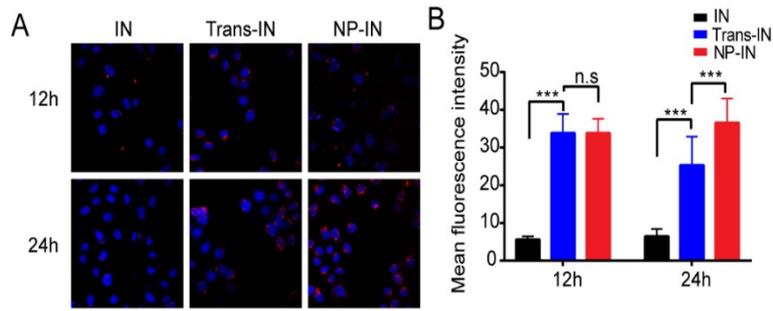
**Fig. S1** Synthesis of mPEG-PLGA-b-PLL block copolymer and characteristics of mPEG-PLGA-b-PLL block copolymer (A). FTIR spectrum of mPEG-PLGA (Red), mPEG-PLGA-b-PLL (Blue) (B). The absorption peak at  $3510\text{ cm}^{-1}$  was attributed to the  $\text{-NH-}$  stretch vibration, indicating the mPEG-PLGA-b-PLL was synthesized.  $^1\text{H}$  NMR spectrum of mPEG-PLGA-b-PLL block copolymer (C). The peaks at 4.36 ppm (f,  $\text{-CH-}$ ), 3.38 ppm (i,  $\text{-CH}_2\text{-}$ ) and 1.35 ppm (h,  $\text{-CH}_2\text{-}$ ) were attributed to protons of the lysine segment. The peaks at 7.87 (g,  $\text{-NH-}$ ) ppm were assigned to protons of the amino group of the lysine group (C).



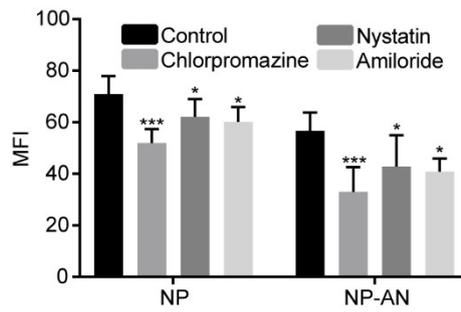
**Fig. S2** Characteristics of NPs. (A) The Zeta-potential of NP-AN was -12 mV. (B) The size distribution results of Nanosight showed similar sizes in the distribution results.



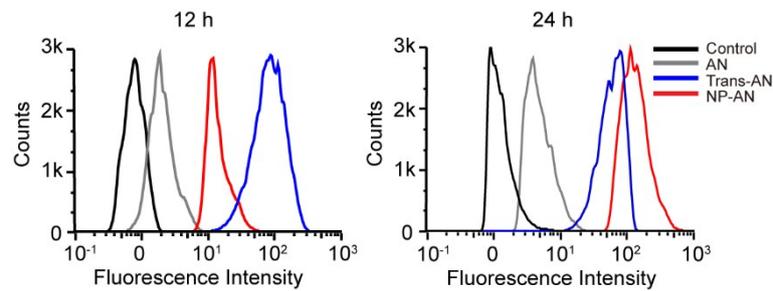
**Fig. S3** (A), (B), (C) MTT results of NPs for 24, 48 and 72 h. The viability of cells incubated with anti-miRs-loaded NPs were all above 80%, which showed nearly no toxicity to LO2 cells. Data are presented as the means  $\pm$  SD. (n=3). NP, NP-AN, Trans-AN, NP-IN and Trans-IN means injected with empty nanoparticles, antagomir-loaded nanoparticles, transfection agent with antagomir-122, inhibitor-loaded nanoparticles and transfection agent with inhibitor-122.



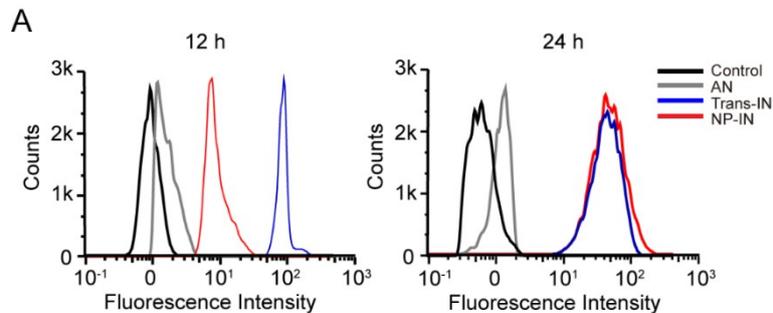
**Fig. S4** Cellular uptake of NPs and lysosome escape of NPs. (A) LO2 cells treated with different IN, transfection agent with IN and NP-IN for 12 and 24h. (B) Quantitative results of cellular uptake of A, which further qualified the results from A.



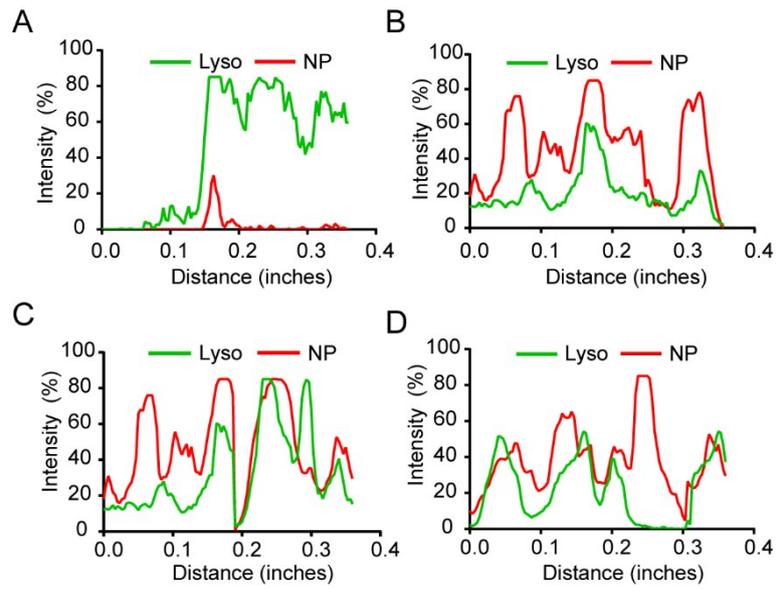
**Fig. S5** Investigation of internalization pathways of NP and NP-AN.



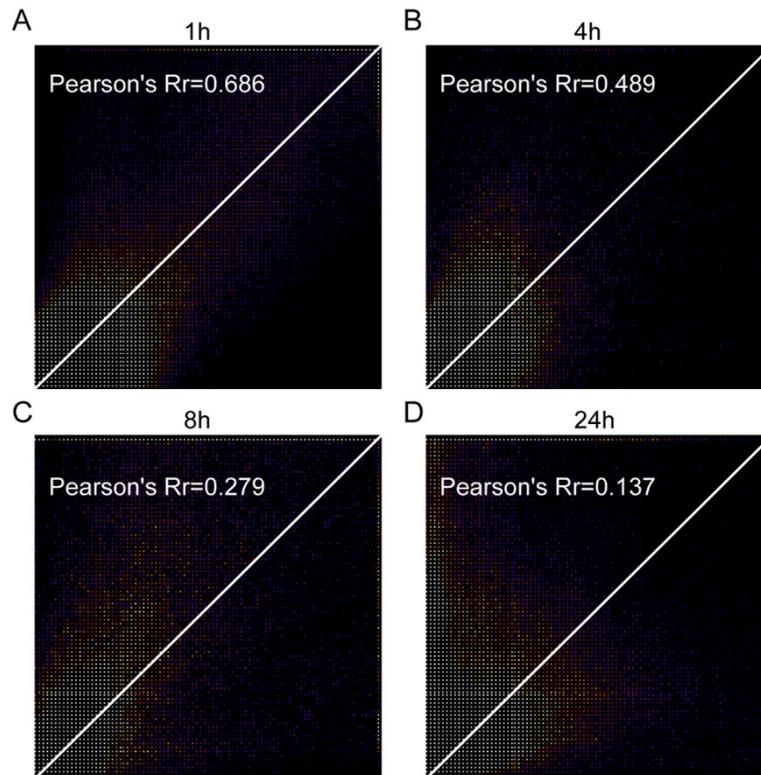
**Fig. S6** Cellular uptake results measured by flowcytometry of NP-AN. The result was consistent with the result at (A). (D) showed that NPs were trapped at lysosome at 4 h, as the red and green fluorescence were overlaid together.



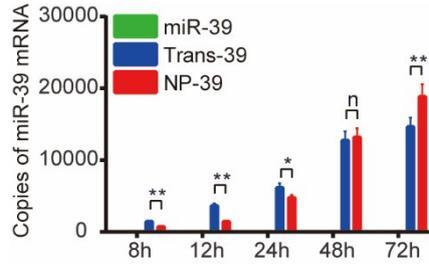
**Fig. S7** Cellular uptake results measured by flowcytometry. (A) LO2 cells treated with IN with/without NPs and transfection agent for 12h. (B) LO2 cells treated with inhibitor with/without NPs and transfection agent for 24h.



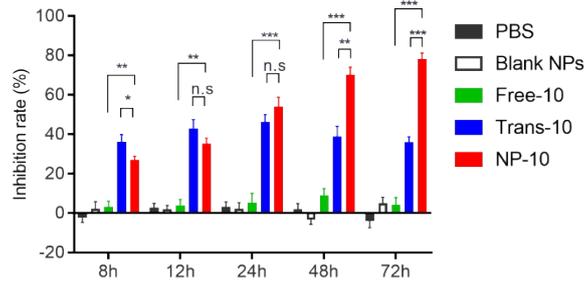
**Fig. S8** Lysosome escape was performed at 8 and 12 h, while the green and red fluorescence began to depart after 1 h (A), 8 h (B), 12 h (C) and 24 h (D).



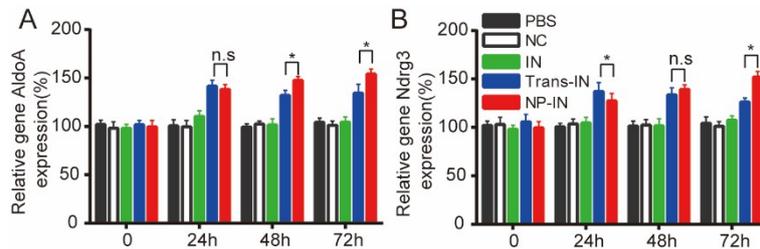
**Fig. S9** Scatter plot of Figure 2D. (A) Scatter plot calculated from Figure 2D merged picture of 1h; (B) Scatter plot calculated from Figure 2D merged picture of 4h; (C) Scatter plot calculated from Figure 2D merged picture of 8h; (D) Scatter plot calculated from Figure 2D merged picture of 24h.



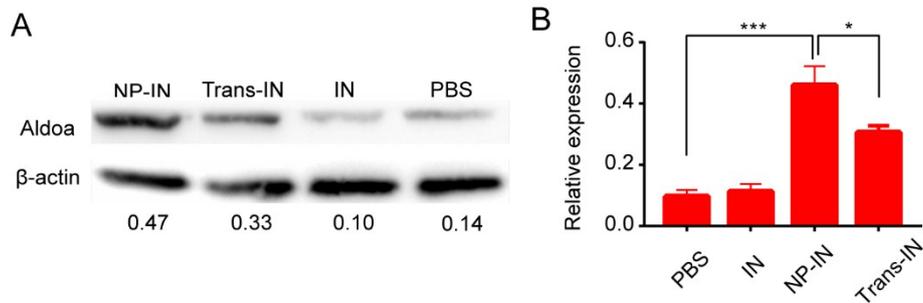
**Fig. S10** Exogenous microRNA transfection in vitro. LO2 cells treated with miR-39 mimic (miR-39), transfection agent with miR-39 mimic (Trans-39) and miR-39 mimic loaded NPs (NP-39).



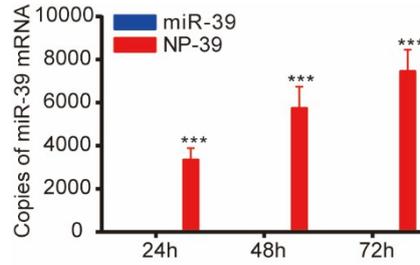
**Fig. S11** LO2 cells treated with inhibitor (IN), transfection agent with IN and IN loaded NPs (NP-IN), miR-122 inhibition were measured.



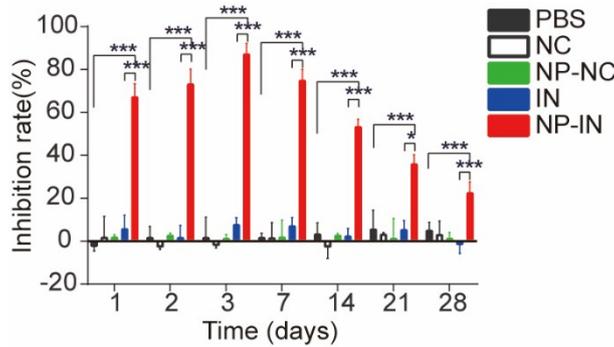
**Fig. S12** LO2 cells treated with inhibitor (IN), transfection agent with IN and IN loaded NPs (NP-IN), Aldoa and Ndr3 expression were measured. (A) Relative Aldoa expression after incubation. (B) Relative Ndr3 expression after incubation.



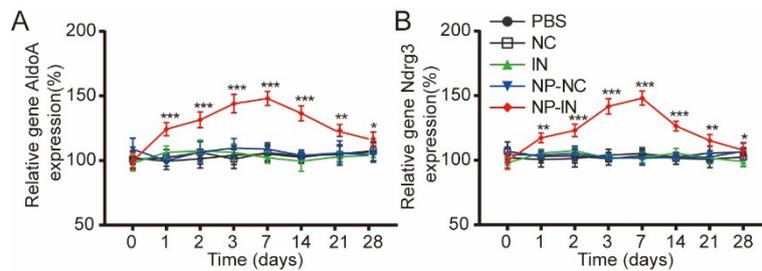
**Fig. S13** (A) The Aldoa protein expression after different treatments. Aldoa protein level increased dramatically within 3 weeks after administrated with NP-IN. (B) Quantitative results of A



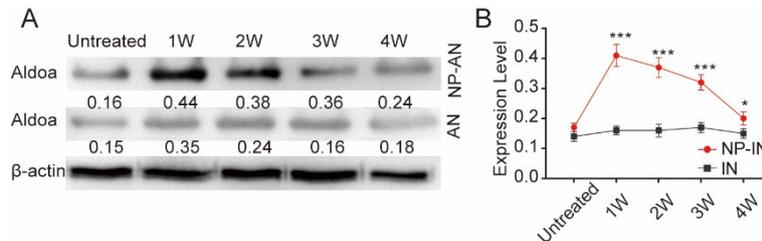
**Fig. S14** Exogenous microRNA transfection *in vivo*. Balb/c mice injected with miR-39 mimic (miR-39), transfection agent with miR-39 mimic (Trans-39) and miR-39 mimic loaded NPs (NP-39).



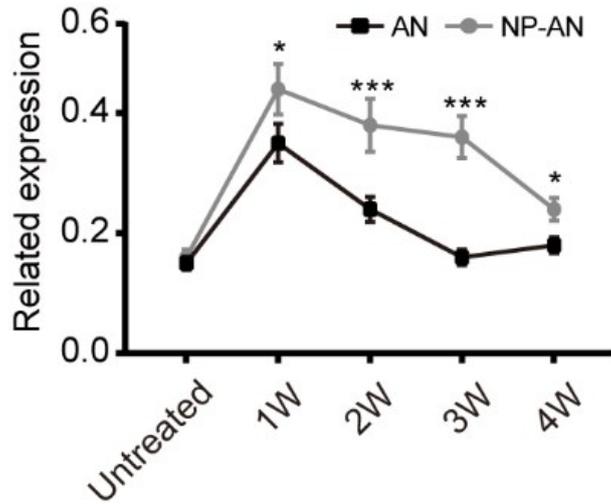
**Fig. S15** Mice treated with different naked IN, transfection agent and NP-IN, miR-122 inhibition were measured.



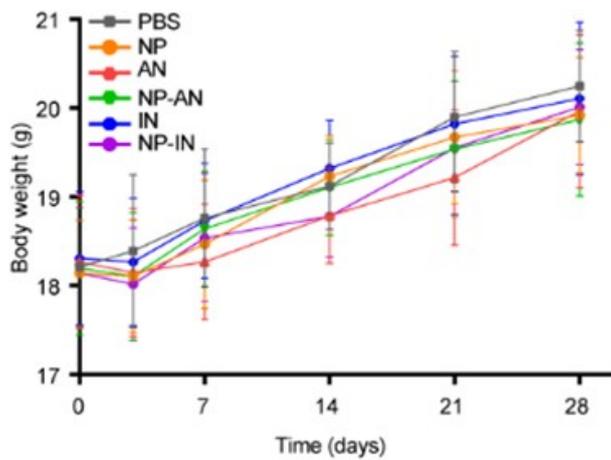
**Fig. S16** *In vivo* miR-122 knockdown efficiency and relative gene expression after injected with NP-IN. (A) The Aldoa gene expression after injected with NP-IN. (B) The Aldoa gene expression after injected with NP-IN.



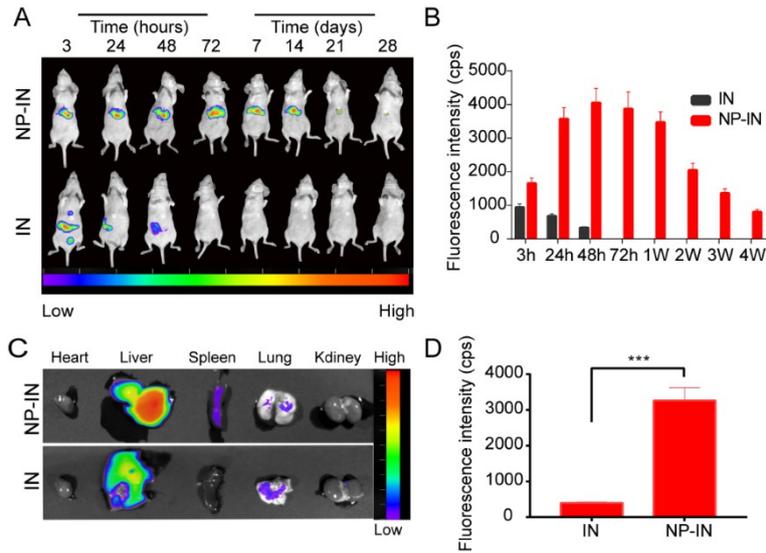
**Fig. S17** (A) The Aldoa protein expression after different treatments. Aldoa protein level increased dramatically within 3 weeks after administrated with NP-IN. (B) Quantitative results of A. Naked IN group, there was no significant change compared to the control; The NP-IN also have strong influence on Aldoa protein.



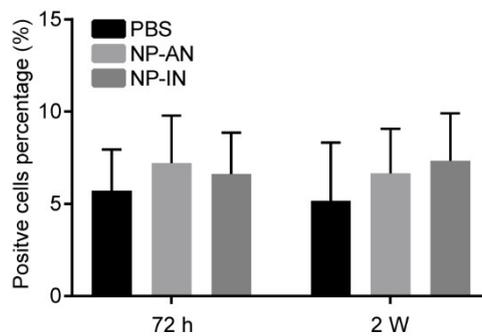
**Fig. S18** Quantitative results of NP-AN, which showed that it have strong influence on Aldoa protein than free AN.



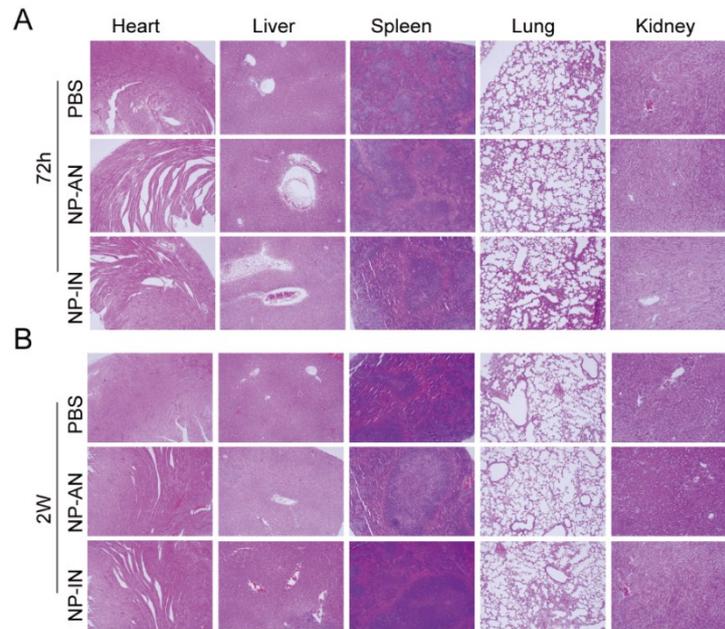
**Fig. S19** Body weight changes after administrated with PBS, blank NP, AN, IN NP-AN and NP-IN. All groups showed no significant impact on the body weight of mice. PBS, AN, NP-AN, IN and NP-IN means injected with PBS, antagomir-122, antagomir-122 loading nanoparticles, inhibitor-122 and inhibitor-122 loading nanoparticles.



**Fig. S20** *In vivo* targeting and organ distribution of NP-IN. (A) Fluorescence images of mice after i.v. administered NP. NP-IN could retained at liver for over 28 days, as the fluorescence results showed, while the IN retained only for 1 days. (B) Quantitative results of mice after different treatments. Further confirmed the results of A. Data are presented as the means  $\pm$  SD. (n=6). (C) Ex vivo fluorescence images of organs harvested from treated mice at 72 h. NP-IN targeted at liver. Fluorescence of IN was much lower than NP-IN and even detected at lung. (D) Quantitative results of organs harvested from mice after 72 h of treatments. Further confirmed the results showed at B. Data are presented as the means  $\pm$  SD. (n=6). \*P<0.05, \*\*P<0.01, \*\*\*P<0.005. IN and NP-IN means injected with inhibitor-122 and inhibitor-122 loading nanoparticles.



**Fig. S21** Quantification of TUNEL assay. No obvious apoptosis was observed in NP-AN and NP-IN comparing to PBS group. \*P<0.05, \*\*P<0.01, \*\*\*P<0.005.



**Fig. S22** HE staining of organs obtained from treated mice at 72 hours (A) and 2 weeks (B). No apparent tissue damage could be observed in both 72 hours and 2 week HE slices.