Electronic Supplementary Information for

Combining pyclen framework with conjugated antenna for the design of europium and samarium luminescent bioprobes

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Ligand and complex synthesis

- Materials and methods

Reagents were purchased from ACROS Organics and from Aldrich Chemical Co and used without further purification. Dialysis membranes (cut-off 100-500 Da) were purchased from spectrumlabs. All solvents were dried and distilled prior to use according to standard methods. Methyl 3,6,9,15-tetraazabicyclo[9.3.1]pentadecane-1(15),11,13-triene-3-acetate 1¹ and methyl 4-((4-(2-(2-(2-methoxyethoxy)ethoxy)phenyl)ethynyl)-6-

(((methylsulfonyl)oxy)methyl)picolinate 2^2 were synthesized as previously described. Analytic HPLC was performed on a Prominence Shimadzu HPLC/LCMS-2020 equipped with a UV SPD-20 A detector. The chromatographic system employs HPLC (Vision HT C18 HL 5 μ 250× 4.6 mm) with H₂O and MeCN as eluents [isocratic 100% H₂O (5 min), linear gradient from 0 to 90% MeCN (10 min), isocratic 90% MeCN (5 min)] at a flow rate of 1 mL/min and UV detection at 254 and 350 nm. NMR spectra were recorded at the "Services communs" of the University of Brest. ¹H and ¹³C NMR spectra were recorded using Bruker Avance 500 (500 MHz), Bruker Avance 400 (400 MHz), or BrukerAMX-3 300 (300 MHz) spectrometers. HRMS analyses were realized on a HRMS Q-Tof MaXis, sources ESI, APCI, APPI, nano-ESI (at the Institute of Organic and Analytic Chemistry – ICOA in Orléans).

Synthesis of (3):

Dimethyl 9-[(methoxycarbonyl)methyl]-3,6,9,15-tetraazabicyclo[9.3.1]pentadecane-1(15),11,13-triene-3,6-di[methylene-4-((4-(2-(2-(2-methoxyethoxy)ethoxy)phenyl) ethynyl)picolinate]



A solution of compound 1 (106 mg, 0.38 mmol) and K_2CO_3 (211 mg, 1.52 mmol, 4 eq) in CH₃CN (9.5 mL) was stirred at room temperature for 30 min. To this solution was added dropwise a solution of compound 2 (396 mg, 0.78 mmol, 2.05 eq) in CH₃CN (14 mL). The reaction mixture was stirred at room temperature for 15h and solvents were evaporated to dryness. The residue was taken up in CH₂Cl₂ and the residual salts were filtered on celite.

Solvents were evaporated to dryness and the crude was purified by chromatography on neutral alumina (eluent: CH₂Cl₂/MeOH 100/0 to 100/1.5) to give **3** (166 mg, 0.15 mmol, 39%) as a brown oil. Rf (CH₂Cl₂/MeOH 100/3) = 0.29. ¹H NMR spectrum could not be described because of its complexity. ¹³C NMR (75 MHz, CDCl₃) δ 171.6, 165.2, 164.4, 160.1, 159.9, 159.8, 158.7, 158.4, 157.8, 147.4, 147.3, 137.9, 134.4, 134.3, 133.5, 133.4, 127.8, 127.6, 125.1, 124.9, 121.3, 120.7, 114.7, 114.6, 113.3, 113.2, 97.0, 96.6, 84.7, 84.6, 71.7, 70.6, 70.4, 70.3, 69.3, 67.4, 62.5, 61.9, 61.6, 61.3, 59.3, 58.8, 57.1, 55.6, 54.8, 52.9, 52.7, 51.5. ESI-HR-MS (positive, MeOH) *m/z* calcd. for [C₆₀H₇₃N₆O₁₄]⁺: 1101.5179, found: 1101.5183, [M+H]⁺; calcd. for [C₆₀H₇₄N₆O₁₄]²⁺: 551.2626, found: 551.2631, [M+2H]²⁺; calcd. for [C₆₀H₇₂N₆NaO₁₄]⁺: 1123.4999, found: 1123.5003, [M+Na]⁺.

Synthesis of (L^{5b}):

9-[acetate]-3,6,9,15-tetraazabicyclo[9.3.1]pentadecane-1(15),11,13-triene-3,6di[methylene-4-((4-(2-(2-(2-methoxyethoxy)ethoxy)ethoxy)phenyl)ethynyl)picolinate]



To a solution of compound **3** (130 mg, 0.12 mmol) in THF/MeOH (8/2 v/v, 7.9 mL) was added 1M KOH (1.57 mL). The reaction mixture was stirred at reflux for 22h. Solvents were evaporated and the residue was submitted to a dialysis for 15h (cut-off 100-500 Da) to give L^{5b} as a brown oil. ¹H NMR (500 MHz, D₂O) δ 7.74 (d, J = 1.0 Hz, 1H), 7.61 (d, J = 0.7 Hz, 1H), 7.47 (t, J = 7.7 Hz, 1H), 7.18 (s, 1H), 7.13 (d, J = 8.6 Hz, 2H), 7.08 (d, J = 8.5 Hz, 2H), 6.93 (d, J = 7.7 Hz, 1H), 6.87 (s, 1H), 6.76 (dd, J = 8.8, 3.0 Hz, 4H), 6.73 (d, J = 7.6 Hz, 1H), 4.08-3.51 (m, 36H), 3.31 (s, 3H), 3.31 (s, 3H), some signals of the pyclen skeleton could not be observed.¹³C NMR (126 MHz, D₂O) δ 181.2, 174.2, 174.0, 161.9, 161.8, 161.2, 160.2, 156.9, 156.4, 154.0, 140.9, 136.7, 136.4, 136.1, 135.7, 131.2, 129.7, 127.9, 126.7, 123.7, 123.3, 117.5, 116.6, 116.5, 98.8, 97.8, 88.7, 88.1, 73.8, 72.7, 72.4, 72.3, 71.7, 70.0, 69.9, 65.9, 61.1, 60.9, 60.0, 59.7, 59.2, 58.2, 57.3, 54.5, 50.5. ESI-HR-MS (positive, H₂O, M corresponds to the three carboxylic acids form of formula C₅₇H₆₆N₆O₁₄) *m/z* calcd. for [C₅₇H₆₄FeN₆O₁₄]⁺: 1112.3824,

found: 1112.3837, $[M-2H+Fe]^+$; calcd. for $[C_{57}H_{67}N_6O_{14}]^+$: 1059.4709, found: 1059.4720, $[M+H]^+$; calcd. for $[C_{57}H_{65}FeN_6O_{14}]^{2+}$: 556.6949, found: 556.6965, $[M-H+Fe]^{2+}$; calcd. for $[C_{57}H_{68}N_6O_{14}]^{2+}$: 530.2391, found: 530.2397, $[M+2H]^{2+}$

Synthesis of Eu complex [EuL^{5b}]

Compound L^{5b} (25 mg, 21.3 µmol) was dissolved in H₂O (10 mL). The pH was adjusted to 6.4 with HCl 0.2M before addition of EuCl₃.6H₂O (15 mg, 1.5 eq). The pH was then adjusted to 5.8 with KOH 1M. The reaction mixture was stirred at room temperature for 1h and the pH checked again. The reaction mixture was stirred at reflux for 20h and solvents were evaporated to dryness. Purification of the residue by flash chromatography (cartridge C18 from Reveleris, Gradient: H₂O/MeOH 100/0 to 2/8 over 35 min) gave the desired Eu-complex [EuL^{5b}] (8 mg, 6.6 µmol, 31%) as a yellow solid. ESI-HR-MS (positive, MeOH) *m/z* calcd. for [C₅₇H₆₄EuN₆O₁₄]⁺: 1209.3687, found: 1209.3708, [M+H]⁺; calcd. for [C₅₇H₆₅EuN₆O₁₄]²⁺: 605.1880, found: 605.1891, [M+2H]²⁺.

Synthesis of Sm complex [SmL^{5b}]

Compound L^{5b} (22 mg, 18.7 µmol) was dissolved in H₂O (10 mL). The pH was controlled (8.3) before addition of SmCl₃.6H₂O (10 mg, 1.5 eq). The pH was then adjusted to 5.6 with KOH 1M. The reaction mixture was stirred at reflux for 1h and the pH checked again. The reaction mixture was stirred at reflux for 17h, resulting in the formation of a solid on the flask's walls. The reaction mixture was filtered on cotton and rinsed with H₂O. The solid stuck on the cotton was dissolved with MeOH and combined with the solid on the flask's walls to give the Sm complex [SmL^{5b}] (13 mg, 10.8 µmol, 58%) as a brown oil. ESI-HR-MS (positive, MeOH) *m/z* calcd. for [C₅₇H₆₄N₆O₁₄Sm]⁺: 1208.3672, found: 1208.3688, [M+H]⁺; calcd. for [C₅₇H₆₅N₆O₁₄Sm]²⁺: 604.6872, found: 604.6876, [M+2H]²⁺.



Figure S1: ¹H NMR of compound 3 (500 MHz, CDCl₃, 25°C).



Figure S2: ¹³C NMR of compound 3 (75 MHz, CDCl₃, 25°C).



Figure S3: HRMS of compound 3.



Figure S4: ¹H NMR of compound L^{5b} (500 MHz, D₂O, 25°C).



Figure S5: ¹³C NMR of compound L^{5b} (125 MHz, D₂O, 25°C).



Meas. m/z	Ζ	#	Ion Formula	m/z	err [ppm]	mSigma	rdb	e ⁻ Conf
2+ 5			C75H58CaN6O	549.214352	1.3	84.5	51.0	even
556.696527	2+	1	C61H69FeO16	556.696201	-0.4	8.9	28.5	odd
	2+	2	C57H65FeN6O14	556.694858	-2.9	11.6	29.5	odd
	2+	3	C62H65FeN4O12	556.696869	0.8	15.5	33.5	odd
	2+	4	C63H61FeN8O8	556.697538	2.0	23.9	38.5	odd
	2+	5	C74H61FeN2O5	556.695944	-0.8	66.2	46.5	odd
	2+	6	C75H57FeN6O	556.696613	0.4	75.7	51.5	odd
	2+	7	C79H61FeO3	556.697956	2.8	87.6	50.5	odd
1059.472006	1+	1	C57H67N6O14	1059.470977	-1.0	32.9	28.0	even
	1+	2	C61H71O16	1059.473663	1.6	33.7	27.0	even
	1+	3	C70H59N8O3	1059.470464	-1.5	70.2	46.0	even
	1+	4	C74H63N2O5	1059.473150	1.1	84.7	45.0	even
1112.383700	1+	1	C61H68FeO16	1112.385125	1.4	8.3	28.5	odd
	1+	2	C57H64FeN6O14	1112.382440	-1.0	15.3	29.5	odd
	1+	3	C70H56FeN8O3	1112.381927	-1.4	50.1	47.5	odd
	1+	4	C74H60FeN2O5	1112.384612	1.0	61.5	46.5	odd
	1+	5	C75H56FeN6O	1112.385949	-889.9	663.5	51.5	odd

Figure S6: HRMS of compound L^{5b} . Peak at 338 is due to the presence of erucamide in Eppendorf's.



Figure S7: Analytical HPLC of the [EuL^{5b}] complex. Column: Vision HT C18 HL 5 μ 250× 4.6 mm. Gradient: 100% H₂O 0-5 min, 0-90% ACN 5-15 min, 90% ACN 15-20 min, 100% H₂O 20-25 min. Flow : 1mL/min. retention time = 15.186 min.



Figure S8: HRMS of the [EuL^{5b}] complex. Peak at 338 is due to the presence of erucamide in Eppendorf's.



Figure S9: Analytical HPLC of the [SmL^{5b}] complex. Column: Vision HT C18 HL 5 μ 250× 4.6 mm. Gradient: 100% H₂O 0-5 min, 0-90% ACN 5-15 min, 90% ACN 15-20 min, 100% H₂O 20-25 min. Flow : 1mL/min. retention time = 15.278 min.



Figure S10: HRMS of the [SmL^{5b}] complex. Peak at 338 is due to the presence of erucamide in Eppendorf's.

Computational details

DFT geometry optimizations were carried out with the Gaussian 09 (revision D.01) package³ tightening self-consistent field convergence thresholds (10-10 a.u.) and geometry optimization (10-5 a.u.) convergence thresholds. Calculations were realized on the $[YL^{5b'}]$ model. The hybrid PBE0 functional has been used.⁴ The "Stuttgart/Dresden" basis sets and effective core potentials were used to describe the yttrium atom,⁵ whereas all other atoms were described with the SVP basis sets.⁶

Table S1. Continuous shape measurements of the O_3N_6 coordination polyhedron in [YL^{5b'}] performed with SHAPE 2.⁷

EP-9	OPY-9	HBPY-9	JTC-9	JCCU-9	CCU-9	JCSAPR-9
(Enneagon)	(Octagonal pyramid)	(Heptagonal bipyramid)	(Triangular cupola)	(Capped	(Capped	(Capped sq. antiprism)
				cube)	cube)	
D_{9h}	$\mathrm{C}_{8\mathrm{v}}$	D_{7h}	C_{3v}	C_{4v}	C_{4v}	C_{4v}
34.033	22.836	16.619	10.540	9.239	8.281	2.614
CSAPR-9	JTCTPR-9	TCTPR-9	JTDIC-9	HH-9	MFF-9	
(Capped sq.	(Tricapped trigonal	(Tricapped	(Tridiminished icosahedron)	(Hula-hoop)	(Muffin)	
antiprism)	prism)	trigonal		C_{2v}	C_s	
C_{4v}	D_{2h}	prism)	C_{3v}			
	231	D_{3h}				
1.896	2.68843	2.05745	10.238	10.617	2.197	

Photophysical measurements in solution

Absorption spectra were recorded on a JASCO V-650 spectrophotometer as solutions in spectrophotometric-grade methanol or water (ca. 10^{-5} or 10^{-6} mol.L⁻¹). Emission spectra were measured by using a Horiba–Jobin–Yvon Fluorolog-3 fluorimeter. Spectra were corrected for both excitation-source light-intensity variation and emission spectral responses. Luminescence lifetimes were obtained by pulsed excitation with a FL-1040 UP xenon lamp. Luminescence quantum yields, Φ , were measured with dilute solutions in water or organic solvents with an absorbance of less than 0.1 by using equation (1):

$$\frac{\Phi_x}{\Phi_r} = \frac{A_r(\lambda)}{A_x(\lambda)} \frac{n_x^2}{n_r^2} \frac{D_x}{D_r}$$
(1)

in which A is the absorbance at the excitation wavelength (λ), n is the refractive index, and D is the integrated luminescence intensity; r and x stand for reference and sample, respectively The reference is quinine bisulfate in a 1N aqueous solution of sulfuric acid ($\Phi_r = 0.546$). Excitation of reference and sample compounds was performed at the same wavelength.

Practically, for each sample, series of measurements are performed with different absorbance ranging from 0.1 to 0.01. The plot of the integrated luminescence intensity vs. absorbance gives straight line with excellent correlation coefficients (figure S11) and the slope S can be determined. Equation (1) becomes (2):

$$\frac{\Phi_x}{\Phi_r} = \frac{S_x(\lambda)}{S_r(\lambda)} \frac{n_x^2}{n_r^2}$$
(2)

The hydration number q is determined for europium complexes by measuring the lifetime in H_2O and D_2O according to the equation (3):⁸

$$q = 1.11 (k(H_2O) - k(D_2O) - 0.31)$$



Figure S11. Determination of the quantum yield of $[EuL^{5b}]$ in MeOH (red), H₂O (blue), CH₂Cl₂ (magenta) and the reference quinine sulfate in a 1N aqueous solution of sulfuric acid (black). The straight lines correspond to the linear fit.



Figure S12. Determination of the quantum yield of $[SmL^{5b}]$ in MeOH (red), H₂O (blue), and the reference quinine sulfate in a 1N aqueous solution of sulfuric acid (black). The straight lines correspond to the linear fit.



Figure S13. Normalized absorption (blue), excitation (dotted) and emission spectra (red) of **3** in MeOH ($\lambda_{exc} = 325$ nm) at room temperature. The harmonic scattering at 650 nm is indicated by the *.

Table S2. Relative intensity of the $[EuL^i]$ complexes measured in MeOH at room temperature. r^i represents the ratio of area of the j=2 over j=4 band for the $[EuL^i]$ complex and %j=2 and %j=4 represent the percentages of the area of the j band over the entire spectrum.

			Ai					
complex	j =0	j = 1	j = 2	j = 3	j = 4	r ⁱ	% j=2	% j=4
[EuL ^{5b}]	0.0	1.0	7.2	0.4	2.4	3.0	65.4	21.6
[EuL ^{3b}]+	0.1	1.0	5.2	0.9	2.1	2.5	56.5	22.5
[EuL ^{2b}]	0.1	1.0	9.1	0.4	0.8	10.9	79.2	7.3
$[EuL^{1a}]^+$	0.1	1.0	2.7	0.2	2.0	1.4	45.0	33.3

Cell culturing and treatment

We used the T24 human epithelial bladder cancer cell line (ATCC no. HBT-4). In our experiments, T24 cells were cultured in 25 cm² tissue-culture flasks (T25) at 378C in a humidified atmosphere with 5% CO₂. They were incubated in RPMI 1640 supplemented with 100 UmL⁻¹ penicillin, 100 µg.mL⁻¹ streptomycin, and 10% fetal calf serum (complete medium). Cells were grown to near confluence in the culture flasks and then suspended with a solution of 0.05% trypsin–ethylenediaminetetraacetic acid (EDTA; Sigma). Cells were placed on a LabTek I chambered cover glass (Nunc) at low cell density in complete culture medium 24 h before experiments. After being washed with phosphate-buffered saline (PBS), cells were fixed with PFA (3% in PBS) for 10 min, permeabilized with PBS containing 0.5% Triton X100 for 10 min, and then washed with PBS.

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