

Supporting Information

Dehydroxylation of alcohols for nucleophilic substitution

Jia Chen, Jin-Hong Lin, and Ji-Chang Xiao*

* Key Laboratory of Organofluorine Chemistry, Shanghai Institute of Organic Chemistry, Chinese Academy of Sciences, 345 Lingling Road, Shanghai 200032, China
Tel: +86-21-54925340 Fax: +86-21-64166128 Email: jchxiao@sioc.ac.cn

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1. General information

Solvents and reagents were purchased from commercial sources and used as received unless otherwise noted. ^1H , ^{13}C , ^{19}F and ^{31}P NMR spectra were detected on a 500 MHz, 400MHz or 300 MHz NMR spectrometer. Data for ^1H NMR, ^{13}C NMR, ^{19}F NMR and ^{31}P NMR were recorded as follows: chemical shift (δ , ppm), multiplicity (s = singlet, d = doublet, t = triplet, m = multiplet, q = quartet, coupling constant (J) in Hz). Mass spectra were obtained on a GC-MS or LC-MS. High resolution mass data were recorded on a high resolution mass spectrometer in the EI, ESI or DART mode.

2. Evidence for the proposed mechanism

Usually, Ph_3P would readily undergo alkylation with alkyl iodides to give alkylphosphonium salts. Surprisingly, Ph_3P was almost completely transformed into $\text{Ph}_3\text{P}=\text{O}$, and neither monophosphonium salt ($\text{Ph}_3\text{P}^+\text{CH}_2\text{CH}_2\text{I I}^-$) nor diphosphonium salt ($\text{Ph}_3\text{P}^+\text{CH}_2\text{CH}_2\text{P}^+\text{Ph}_3 \text{ 2I}^-$) was observed in the dehydroxy-functionalization reaction. Furthermore, diphosphonium salt ($\text{Ph}_3\text{P}^+\text{CH}_2\text{CH}_2\text{P}^+\text{Ph}_3 \text{ 2I}^-$) could not promote dehydroxy-functionlization, suggesting that the alkylation process did not occur. NMR spectroscopy was then used to monitor the reaction of Ph_3P with $\text{ICH}_2\text{CH}_2\text{I}$. Mixing Ph_3P (0.2 mmol) and $\text{ICH}_2\text{CH}_2\text{I}$ (0.2 mmol) in DMF (2 mL) at room temperature would immediately led to the rapid evolution of gas and the complete conversion of both $\text{ICH}_2\text{CH}_2\text{I}$ and Ph_3P . ^1H NMR analysis of the reaction mixture revealed that $\text{ICH}_2\text{CH}_2\text{I}$ was converted into ethylene ($\text{CH}_2=\text{CH}_2$) (Figure S1). Ph_3P was transformed into two new species (**A** and **B**), the signals of which detected by ^{31}P NMR spectroscopy appeared at 25.5 ppm and 11.9 ppm, respectively. The signal at 25.5 ppm was assigned to $\text{Ph}_3\text{P}=\text{O}$, as the intensity of this signal in ^{31}P NMR spectrum was increased by external addition of $\text{Ph}_3\text{P}=\text{O}$ into the system. Intermediate **B** is so fragile that the attempts at its isolation or further characterization were unsuccessful. Its structure is proposed to be a pentacoordinate species (Figure S1). $\text{Ph}_3\text{P}=\text{O}$ should be formed from this species with the simultaneous formation of Vilsmeier–Haack type intermediate **C**. After intermediates **A** and **B** were generated, the subsequent addition of phenylbutanol and sodium phenolate gave the desired product in 28% yield, indicating that both species **B** and **C** are key intermediates of the reaction. This low yield is because these two species are unstable and would readily undergo side reactions in the absence of substrates. If substrates are added before intermediates **B** and **C** are formed, high yields would be obtained, as shown in Tables 1 and 2.

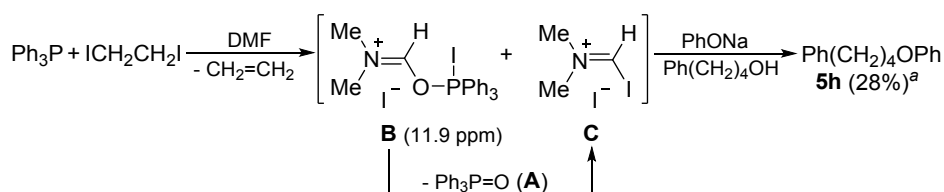


Figure S1. The reaction of Ph_3P with $\text{ICH}_2\text{CH}_2\text{I}$ in DMF. ^aThe yield was determined by ^1H NMR spectroscopy.

The reaction of Ph_3P with $\text{ICH}_2\text{CH}_2\text{I}$ to furnish intermediate **B** occurs so fast that no direct evidence could be collected to elucidate this process. Fortunately, we found that the conversion in chloroform proceeded slowly. Therefore, the reaction in CDCl_3 in a NMR tube was monitored by ^{31}P NMR spectroscopy. New phosphorus species **D** and **E** were detected immediately after mixing Ph_3P and $\text{ICH}_2\text{CH}_2\text{I}$ (Figure S2-A). They were too unstable to be isolated for characterization, and thus their structures were determined based on their ^{31}P NMR signals. The formation of intermediates **D** and **E** should be because of a strong P-I halogen bond.¹ In intermediate **D**, the phosphorus atom carries slightly positive charge, therefore a downfield shift was observed in ^{31}P NMR spectroscopy. Intermediate **E** is close to a pentacoordinate phosphorus species, thus its signal would move upfield.² This intermediate could be either a linear or a cyclic structure, depending on the interaction of terminal Ph_3P and CH_2I moiety. Intermediates **D** and **E** were slowly transformed into two new species **F** and **G**, with ^{31}P NMR signals appearing at +45.8 ppm and -18.7 ppm, respectively (Figure S2-B). **F** and **G** were determined to be Ph_3PI_2 and $[\text{Ph}_3\text{P}^+\text{I I}^-]$, respectively, based on the reported phosphorus signals of Ph_3PI_2 ³ and $[\text{Ph}_3\text{P}^+\text{I I}^-]$ ³⁻⁴.

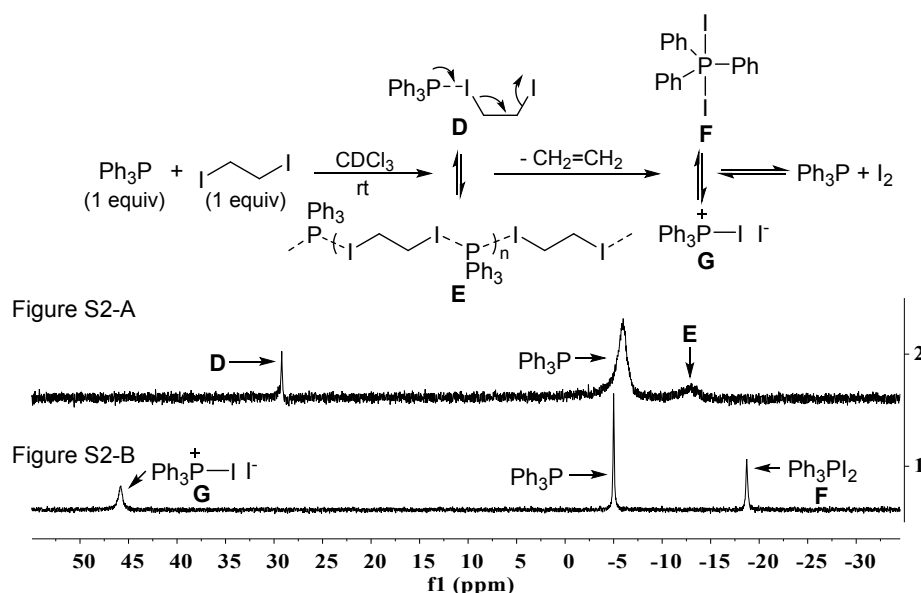
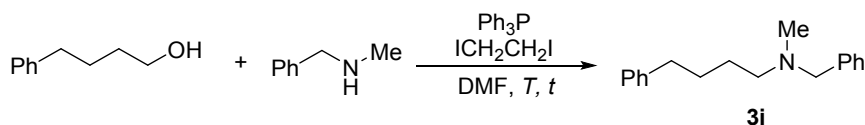


Figure S2. The reaction of Ph_3P with $\text{ICH}_2\text{CH}_2\text{I}$ in CDCl_3 .

3. Screening reaction conditions of dehydroxy-amination of alcohols



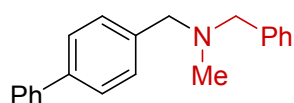
Entry	Ratio ^a	T (°C)	t (hour)	Yield (%) ^b
1	1:3:1.2:1.2	r.t.	10 h	66
2	1:3:1.2:1.2	50	10 h	63
3	1:3:1.2:1.2	80	10 h	62
4	1:3.5:1.2:1.2	r.t.	10 h	68
5	1:4:1.2:1.2	r.t.	10 h	75
6	1:4:1.1:1.1	r.t.	10 h	55
7	1:4:1.3:1.3	r.t.	10 h	70
8	1:4:1.5:1.5	r.t.	10 h	72
9	1:4:1.2:1.2	r.t.	2	44
10	1:4:1.2:1.2	r.t.	4	45
11	1:4:1.2:1.2	r.t.	6	56

Reaction conditions: **1** (0.5 mmol), **2**, Ph₃P and ICH₂CH₂I in DMF (5 mL). ^aMolar ratio of 1:2:Ph₃P:ICH₂CH₂I. ^bDetermined by ¹H NMR with the use of anisole as an internal standard.

4. General procedure for dehydroxy-functionalization of alcohols

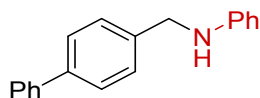
4.1. General procedure for dehydroxylation of alcohols to form C-N and C-S bonds

Alcohol **1a** (1.0 equiv, 0.5 mmol, 92.1 mg), triphenylphosphine (1.2 equiv, 0.6 mmol, 157.4 mg) and anhydrous DMF (5.0 mL) were added into a 10 mL sealed tube under a N₂ atmosphere. 1,2-Diiodoethane (1.2 equiv, 0.6 mmol, 169.1 mg) was then added and the resulting mixture was stirred for around 1 min until the 1,2-diiodoethane was completely dissolved. Amine **2a** (3.0 equiv, 1.5 mmol, 181.8 mg) was added subsequently and the mixture was stirred at room temperature for 10-12 h. Dichloromethane (20 mL) was added and the resulting solution was washed with water (20 mL x 3). The organic layer was dried over Na₂SO₄. After filtration, the solvent was removed by concentration under reduced pressure. The residue was subjected to flash column chromatography to afford the pure product **3a**.

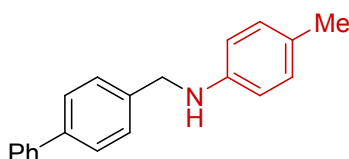


1-([1,1'-biphenyl]-4-yl)-N-benzyl-N-methylmethanamine (**3a**):⁵ 96%; white solid. ¹H NMR (400 MHz, chloroform-*d*) δ 7.64 – 7.55 (m, 4H), 7.49 – 7.38 (m, 6H), 7.38 –

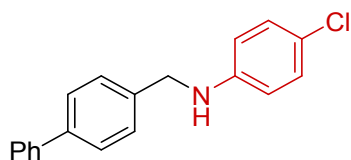
7.31 (m, 3H), 7.30 – 7.23 (m, 1H), 3.58 (s, 2H), 3.57 (s, 2H), 2.24 (s, 3H). LC-MS (ESI) Calcd. for C₂₁H₂₂N [M+H]⁺ 288.2; found 288.2.



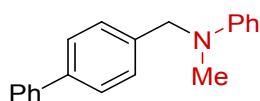
N-([1,1'-biphenyl]-4-ylmethyl)aniline (**3b**).⁵ 4 equiv of N-nucleophile was used as the nucleophile; the reaction temperature was 50 °C. 90%; white solid. ¹H NMR (400 MHz, chloroform-*d*) δ 7.57 (t, *J* = 7.0 Hz, 4H), 7.46 – 7.39 (m, 4H), 7.33 (t, *J* = 7.3 Hz, 1H), 7.18 (t, *J* = 7.8 Hz, 2H), 6.72 (t, *J* = 7.3 Hz, 1H), 6.65 (d, *J* = 7.9 Hz, 2H), 4.36 (s, 2H), 4.05 (s, 1H).



N-([1,1'-biphenyl]-4-ylmethyl)-4-methylaniline (**3c**). 4 equiv of N-nucleophile was used as the nucleophile; the reaction temperature was 50 °C. 94%; white solid. m.p. 122 – 123 °C. ¹H NMR (400 MHz, chloroform-*d*) δ 7.62 – 7.52 (m, 4H), 7.49 – 7.39 (m, 4H), 7.39 – 7.29 (tt, *J* = 7.4, 1.6 Hz, 1H), 6.99 (d, *J* = 8.0 Hz, 2H), 6.59 (d, *J* = 8.4 Hz, 2H), 4.35 (s, 2H), 3.94 (s, 1H), 2.24 (s, 3H). ¹³C NMR (101 MHz, chloroform-*d*) δ 145.9, 140.9, 140.1, 138.8, 129.8, 128.8, 127.9, 127.4, 127.2, 127.1, 126.8, 113.0, 48.3, 20.4. IR (neat) ν = 3392, 2924, 2852, 1612, 1520, 1406, 1928, 1156, 1080, 826, 815, 771, 727, 697, 512 cm⁻¹. HRMS (EI) Calcd. for C₂₀H₁₉N [M]⁺: 273.1517; found 273.1517.

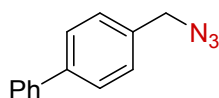


N-([1,1'-biphenyl]-4-ylmethyl)-4-chloroaniline (**3d**). 4 equiv of N-nucleophile was used as the nucleophile; the reaction temperature was 50 °C. 82%; white solid. m.p. 124 – 125 °C. ¹H NMR (400 MHz, chloroform-*d*) δ 7.68 – 7.51 (m, 4H), 7.50 – 7.38 (m, 4H), 7.35 (t, *J* = 7.3 Hz, 1H), 7.12 (d, *J* = 8.7 Hz, 2H), 6.57 (d, *J* = 8.7 Hz, 2H), 4.34 (s, 2H), 4.09 (s, 1H). ¹³C NMR (101 MHz, chloroform-*d*) δ 146.6, 140.8, 140.4, 138.0, 129.1, 128.8, 127.8, 127.45, 127.35, 127.1, 122.2, 114.0, 48.1. IR (neat) ν = 3383, 3026, 2843, 1594, 1494, 1400, 1339, 1157, 1077, 1006, 815, 762, 689, 669, 505 cm⁻¹. HRMS (EI) Calcd. for C₁₉H₁₆NCl [M]⁺: 293.0971; found 293.0963.

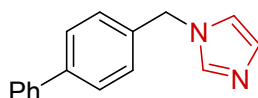


N-([1,1'-biphenyl]-4-ylmethyl)-*N*-methylaniline (**3e**).⁶ 4 equiv of N-nucleophile was used as the nucleophile; the reaction temperature was 50 °C. quantitative; white solid.

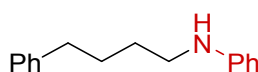
^1H NMR (400 MHz, chloroform-*d*) δ 7.63 – 7.52 (m, 4H), 7.44 (t, J = 7.6 Hz, 2H), 7.39 – 7.28 (m, 3H), 7.29 – 7.17 (m, 2H), 6.80 (d, J = 8.0 Hz, 2H), 6.74 (t, J = 7.3 Hz, 1H), 4.59 (s, 2H), 3.06 (s, 3H).



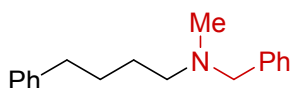
4-(azidomethyl)-1,1'-biphenyl (**3f**).⁷ NaN_3 was used as the nucleophile. quantitative; white solid. ^1H NMR (400 MHz, chloroform-*d*) δ 7.65 – 7.52 (m, 4H), 7.49 – 7.42 (m, 2H), 7.41 – 7.31 (m, 3H), 4.38 (s, 2H).



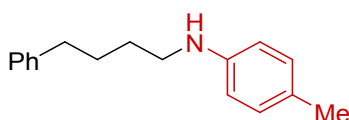
1-([1,1'-biphenyl]-4-ylmethyl)-1H-imidazole (**3g**).⁸ 4 equiv of N-nucleophile was used as the nucleophile; the reaction temperature was 50 °C. 78%; white solid. ^1H NMR (400 MHz, chloroform-*d*) δ 7.61 – 7.50 (m, 5H), 7.43 (t, J = 7.4 Hz, 2H), 7.34 (t, J = 7.3 Hz, 1H), 7.21 (d, J = 7.8 Hz, 2H), 7.10 (s, 1H), 6.92 (s, 1H), 5.14 (s, 2H).



N-(4-phenylbutyl)aniline (**3h**).⁹ 4 equiv of N-nucleophile was used as the nucleophile; the reaction temperature was 50 °C. 71%; yellow liquid. ^1H NMR (400 MHz, chloroform-*d*) δ 7.32 – 7.25 (m, 2H), 7.25 – 7.12 (m, 5H), 6.68 (t, J = 7.1 Hz, 1H), 6.58 (d, J = 8.1 Hz, 2H), 3.57 (s, 1H), 3.12 (t, J = 6.9 Hz, 2H), 2.66 (t, J = 7.4 Hz, 2H), 1.80 – 1.59 (m, 4H).

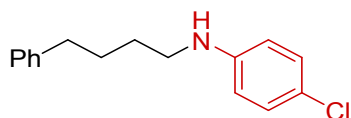


N-benzyl-*N*-methyl-4-phenylbutan-1-amine (**3i**).¹⁰ 4 equiv of N-nucleophile was used as the nucleophile. 59%; colorless liquid. ^1H NMR (400 MHz, chloroform-*d*) δ 7.34 – 7.28 (m, 4H), 7.28 – 7.21 (m, 3H), 7.20 – 7.13 (m, 3H), 3.47 (s, 2H), 2.61 (t, J = 7.5 Hz, 2H), 2.38 (t, J = 7.2 Hz, 2H), 2.17 (s, 3H), 1.73 – 1.48 (m, 4H).

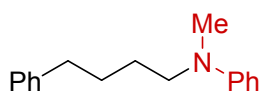


4-methyl-*N*-(4-phenylbutyl)aniline (**3j**). 4 equiv of N-nucleophile was used as the nucleophile; the reaction temperature was 50 °C. 74%; white solid. m.p. 32 – 33 °C. ^1H NMR (400 MHz, chloroform-*d*) δ 7.32 – 7.25 (m, 2H), 7.22 – 7.11 (m, 3H), 6.98 (d, J = 8.0 Hz, 2H), 6.52 (d, J = 8.3 Hz, 2H), 3.44 (s, 1H), 3.10 (t, J = 6.9 Hz, 2H), 2.65 (t, J = 7.4 Hz, 2H), 2.23 (s, 3H), 1.79 – 1.58 (m, 4H). ^{13}C NMR (101 MHz, chloroform-*d*) δ 146.2, 142.2, 129.7, 128.4, 128.3, 126.4, 125.8, 112.9, 44.2, 35.7,

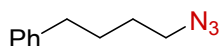
29.2, 29.0, 20.4. IR (neat) ν = 3406, 3061, 3024, 2932, 2857, 1619, 1495, 1478, 1453, 1373, 1317, 1250, 1181, 1125, 806, 747, 698, 508 cm^{-1} . HRMS (EI) Calcd. for $\text{C}_{17}\text{H}_{21}\text{N}$ $[\text{M}]^+$: 239.1674; found 239.1681.



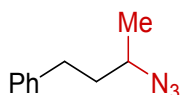
4-chloro-*N*-(4-phenylbutyl)aniline (**3k**). 4 equiv of N-nucleophile was used as the nucleophile; the reaction temperature was 50 °C. 53%; yellow liquid. ^1H NMR (400 MHz, chloroform-*d*) δ 7.29 (t, J = 7.4 Hz, 2H), 7.23 – 7.15 (m, 3H), 7.10 (d, J = 8.5 Hz, 2H), 6.49 (d, J = 8.4 Hz, 2H), 3.57 (s, 1H), 3.09 (t, J = 7.0 Hz, 2H), 2.66 (t, J = 7.5 Hz, 2H), 1.81 – 1.57 (m, 4H). ^{13}C NMR (101 MHz, chloroform-*d*) δ 147.0, 142.1, 129.0, 128.4, 128.4, 125.9, 121.6, 113.7, 44.0, 35.6, 29.0, 28.9. IR (neat) ν = 3415, 3261, 3061, 3025, 2935, 2857, 1600, 1502, 1475, 1401, 1320, 1248, 1177, 1090, 814, 748, 699, 669, 504 cm^{-1} . HRMS (EI) Calcd. for $\text{C}_{16}\text{H}_{18}\text{NCl}$ $[\text{M}]^+$: 259.1128; found 259.1123.



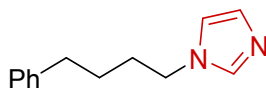
N-methyl-*N*-(4-phenylbutyl)aniline (**3l**).¹¹ 4 equiv of N-nucleophile was used as the nucleophile; the reaction temperature was 50 °C. 80%; yellow liquid. ^1H NMR (400 MHz, chloroform-*d*) δ 7.32 – 7.26 (m, 2H), 7.26 – 7.13 (m, 5H), 6.72 – 6.64 (m, 3H), 3.33 (t, J = 6.9 Hz, 2H), 2.91 (s, 3H), 2.64 (t, J = 7.3 Hz, 2H), 1.73 – 1.56 (m, 4H).



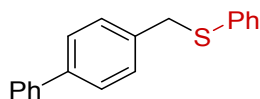
(4-azidobutyl)benzene (**3m**).¹² NaN_3 was used as the nucleophile. Irrespective of whether a base (2 equiv of 2,6-lutidine) was used or not, a quantitative yield was obtained. If the base is used, it should be added after NaN_3 . colorless liquid. ^1H NMR (400 MHz, chloroform-*d*) δ 7.28 (t, J = 7.4 Hz, 2H), 7.22 – 7.14 (m, 3H), 3.27 (t, J = 6.6 Hz, 2H), 2.64 (t, J = 7.4 Hz, 2H), 1.77 – 1.57 (m, 4H).



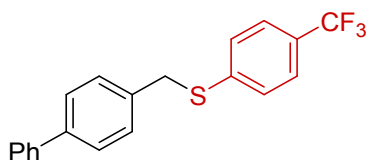
(3-azidobutyl)benzene (**3n**).¹³ NaN_3 was used as the nucleophile. 98%; colorless liquid. ^1H NMR (400 MHz, chloroform-*d*) δ 7.28 (t, J = 7.4 Hz, 2H), 7.22 – 7.15 (m, 3H), 3.43 (h, J = 6.6 Hz, 1H), 2.80 – 2.59 (m, 2H), 1.88 – 1.69 (m, 2H), 1.28 (d, J = 6.5 Hz, 3H).



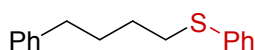
1-(4-phenylbutyl)-1H-imidazole (**3o**).¹⁴ 4 equiv of N-nucleophile was used as the nucleophile; the reaction temperature was 50 °C. 82%; yellow liquid. ¹H NMR (400 MHz, chloroform-*d*) δ 7.92 (s, 1H), 7.34 – 7.25 (m, 2H), 7.20 (tt, *J* = 7.3, 1.5 Hz, 1H), 7.17 – 7.10 (m, 3H), 6.92 (s, 1H), 3.96 (t, *J* = 7.2 Hz, 2H), 2.64 (t, *J* = 7.2 Hz, 2H), 1.82 (p, *J* = 7.3 Hz, 2H), 1.63 (p, *J* = 7.5 Hz, 2H).



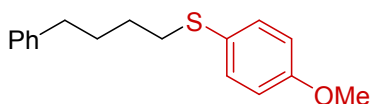
([1,1'-biphenyl]-4-ylmethyl)(phenyl)sulfane (**3p**).⁵ PhS⁻Na⁺ was used as the nucleophile. 93%; white solid. ¹H NMR (400 MHz, chloroform-*d*) δ 7.56 (d, *J* = 7.5 Hz, 2H), 7.51 (d, *J* = 8.2 Hz, 2H), 7.42 (t, *J* = 7.7 Hz, 2H), 7.38 – 7.29 (m, 5H), 7.29 – 7.22 (m, 3H), 7.21 – 7.14 (m, 1H), 4.15 (s, 2H). GC-MS (EI) Calcd. for C₁₉H₁₆S [M]⁺ 276.1; found 276.1.



([1,1'-biphenyl]-4-ylmethyl)(4-(trifluoromethyl)phenyl)sulfane (**3q**): 85%; white solid. m.p. 161 – 162 °C. ¹H NMR (400 MHz, chloroform-*d*) δ 7.63 – 7.52 (m, 4H), 7.49 (d, *J* = 8.2 Hz, 2H), 7.47 – 7.31 (m, 7H), 4.22 (s, 2H). ¹⁹F NMR (376 MHz, chloroform-*d*) δ -62.45 (s, 3F). ¹³C NMR (101 MHz, chloroform-*d*) δ 142.0, 142.0, 140.55, 140.48, 135.4, 129.2, 128.8, 128.0, 127.8 (q, *J* = 33.3 Hz), 127.4, 127.0, 125.7 (q, *J* = 3.8 Hz), 124.1 (q, *J* = 272.7 Hz), 37.4. IR (neat) ν = 1654, 1403, 1322, 1176, 1157, 1118, 845, 823, 773, 745, 689, 590, 495 cm⁻¹. HRMS (EI) Calcd. for C₂₀H₁₅F₃S [M]⁺: 344.0847; found 344.0835.

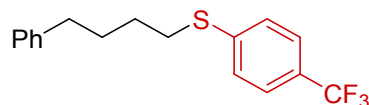


phenyl(4-phenylbutyl)sulfane (**3r**).¹⁵ PhS⁻Na⁺ was used as the nucleophile. 95%; colorless liquid. ¹H NMR (400 MHz, chloroform-*d*) δ 7.34 – 7.23 (m, 6H), 7.21 – 7.13 (m, 4H), 2.93 (t, *J* = 7.1 Hz, 2H), 2.62 (t, *J* = 7.4 Hz, 2H), 1.85 – 1.62 (m, 4H). GC-MS (EI) Calcd. for C₁₆H₁₈S [M]⁺ 242.1; found 242.1.

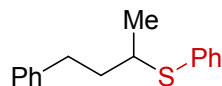


(4-methoxyphenyl)(4-phenylbutyl)sulfane (**3s**): 96%; colorless liquid. ¹H NMR (400 MHz, chloroform-*d*) δ 7.36 – 7.22 (m, 4H), 7.21 – 7.09 (m, 3H), 6.83 (d, *J* = 8.7 Hz, 2H), 3.79 (s, 3H), 2.83 (t, *J* = 7.2 Hz, 2H), 2.60 (t, *J* = 7.6 Hz, 2H), 1.73 (p, *J* = 7.0 Hz, 2H), 1.62 (p, *J* = 7.2 Hz, 2H). ¹³C NMR (101 MHz, chloroform-*d*) δ 158.8, 142.2, 133.2, 128.4, 128.3, 126.7, 125.8, 114.5, 55.3, 35.8, 35.4, 30.4, 28.8. IR (neat) ν = 3061, 3025, 3001, 2934, 2855, 2835, 1592, 1570, 1493, 1461, 1440, 1284, 1245, 1179,

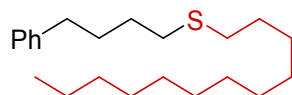
1103, 1032, 826, 747, 699, 638, 625, 526 cm^{-1} . HRMS (EI) Calcd. for $\text{C}_{17}\text{H}_{20}\text{OS}$ $[\text{M}]^+$: 272.1235; found 272.1229.



(4-phenylbutyl)(4-(trifluoromethyl)phenyl)sulfane (**3t**): quantitative; colorless liquid. ^1H NMR (400 MHz, chloroform-*d*) δ 7.49 (d, $J = 8.2$ Hz, 2H), 7.36 – 7.22 (m, 4H), 7.23 – 7.10 (m, 3H), 2.97 (t, $J = 7.0$ Hz, 2H), 2.64 (t, $J = 7.3$ Hz, 2H), 1.90 – 1.62 (m, 4H). ^{19}F NMR (376 MHz, chloroform-*d*) δ -62.41 (s, 3F). ^{13}C NMR (101 MHz, chloroform-*d*) δ 142.5, 142.5, 141.8, 128.4, 127.3, 127.2 (q, $J = 33.3$ Hz), 125.9, 125.6 (q, $J = 3.8$ Hz), 124.2 (q, $J = 272.7$ Hz), 35.3, 32.4, 30.4, 28.3. IR (neat) $\nu = 3086, 3063, 3027, 2934, 2858, 1607, 1497, 1454, 1402, 1328, 1166, 1126, 1095, 1030, 1013, 824, 745, 699, 590, 494$ cm^{-1} . HRMS (EI) Calcd. for $\text{C}_{17}\text{H}_{20}\text{OS}$ $[\text{M}]^+$: 310.1003; found 310.1004.



phenyl(4-phenylbutan-2-yl)sulfane (**3u**).¹⁶ PhS^-Na^+ was used as the nucleophile. 40%; colorless liquid. ^1H NMR (400 MHz, chloroform-*d*) δ 7.39 – 7.33 (m, 2H), 7.31 – 7.24 (m, 4H), 7.24 – 7.14 (m, 4H), 3.20 (h, $J = 6.7$ Hz, 1H), 2.87 – 2.70 (m, 2H), 1.99 – 1.87 (m, 1H), 1.87 – 1.76 (m, 1H), 1.32 (d, $J = 6.7$ Hz, 3H).

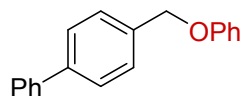


dodecyl(4-phenylbutyl)sulfane (**3v**): 80%; colorless liquid. ^1H NMR (400 MHz, chloroform-*d*) δ 7.30 – 7.23 (m, 2H), 7.20 – 7.12 (m, 3H), 2.62 (t, $J = 7.5$ Hz, 2H), 2.51 (t, $J = 7.3$ Hz, 2H), 2.47 (t, $J = 7.4$ Hz, 2H), 1.77 – 1.66 (m, 2H), 1.66 – 1.58 (m, 2H), 1.58 – 1.50 (m, 2H), 1.40 – 1.19 (m, 18H), 0.87 (t, $J = 6.8$ Hz, 3H). ^{13}C NMR (101 MHz, Chloroform-*d*) δ 142.3, 128.4, 128.3, 125.7, 35.5, 32.2, 32.0, 31.9, 30.6, 29.7, 29.68, 29.65, 29.63, 29.56, 29.4, 29.28, 29.26, 29.0, 22.7, 14.1. IR (neat) $\nu = 3085, 3062, 3026, 2925, 2853, 1604, 1496, 1454, 1377, 1261, 1030, 743, 698$ cm^{-1} . HRMS (EI) Calcd. for $\text{C}_{22}\text{H}_{38}\text{S}$ $[\text{M}]^+$: 334.2694; found 334.2699.

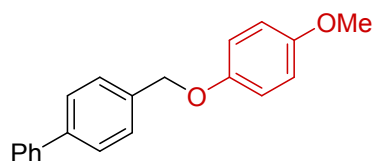
4.2. General procedure for dehydroxylation of alcohols to form C-O and C-X (X = Cl, Br, I) bonds

Alcohol **1a** (1.0 equiv, 0.5 mmol, 92.1 mg), triphenylphosphine (1.2 equiv, 0.6 mmol, 157.4 mg) and anhydrous DMF (5.0 mL) were added into a 10 mL sealed tube under a N_2 atmosphere. 1,2-Diiodoethane (1.2 equiv, 0.6 mmol, 169.1 mg) was then added and the resulting mixture was stirred for around 1 min until the 1,2-diiodoethane was completely dissolved. Sodium phenolate **4a** (3.0 equiv, 1.5 mmol, 174.1 mg) was added subsequently and the mixture was stirred at room temperature for 10-12 h.

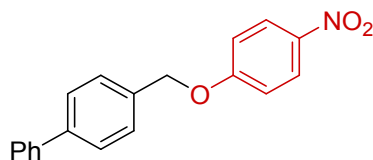
Dichloromethane (20 mL) was added and the resulting solution was washed with water (20 mL x 3). The organic layer was dried over Na₂SO₄. After filtration, the solvent was removed by concentration under reduced pressure. The residue was subjected to flash column chromatography to afford the pure product **5a**.



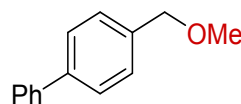
4-(phenoxy)methyl-1,1'-biphenyl (**5a**):¹⁷ 98%; white solid. ¹H NMR (400 MHz, chloroform-*d*) δ 7.60 (t, *J* = 7.4 Hz, 4H), 7.51 (d, *J* = 8.1 Hz, 2H), 7.44 (t, *J* = 7.5 Hz, 2H), 7.38 – 7.26 (m, 3H), 7.04 – 6.93 (m, 3H), 5.10 (s, 2H). GC-MS (EI) Calcd. for C₁₉H₁₆O [M]⁺ 260.1; found 260.1.



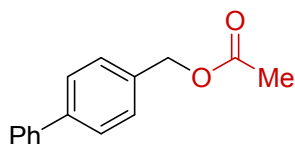
4-((4-methoxyphenoxy)methyl)-1,1'-biphenyl (**5b**): 83%; white solid. m.p. 162 – 163 °C. ¹H NMR (400 MHz, chloroform-*d*) δ 7.64 – 7.57 (m, 4H), 7.50 (d, *J* = 8.0 Hz, 2H), 7.44 (t, *J* = 7.5 Hz, 2H), 7.35 (t, *J* = 7.3 Hz, 1H), 6.94 (d, *J* = 9.0 Hz, 2H), 6.85 (d, *J* = 9.1 Hz, 2H), 5.05 (s, 2H), 3.77 (s, 3H). ¹³C NMR (101 MHz, chloroform-*d*) δ 154.0, 153.0, 140.9, 140.8, 136.3, 128.8, 128.0, 127.37, 127.35, 127.2, 115.9, 114.7, 70.5, 55.7. IR (neat) ν = 2995, 2958, 2927, 2833, 1506, 1488, 1438, 1407, 1388, 1290, 1232, 1220, 1178, 1127, 1112, 1039, 1016, 877, 824, 767, 745, 717, 688, 524 cm⁻¹. HRMS (EI) Calcd. for C₂₀H₁₈O₂ [M]⁺: 290.1307; found 290.1310.



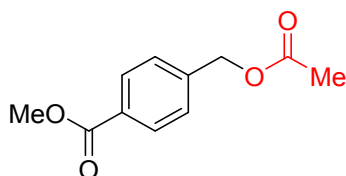
4-((4-nitrophenoxy)methyl)-1,1'-biphenyl (**5c**):¹⁸ 76%; yellow solid. ¹H NMR (400 MHz, chloroform-*d*) δ 8.21 (d, *J* = 9.1 Hz, 2H), 7.63 (d, *J* = 8.1 Hz, 2H), 7.59 (d, *J* = 7.5 Hz, 2H), 7.52 – 7.41 (m, 4H), 7.36 (t, *J* = 7.3 Hz, 1H), 7.04 (d, *J* = 9.1 Hz, 2H), 5.19 (s, 2H).



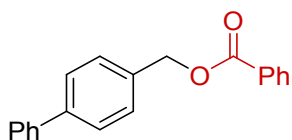
4-(methoxymethyl)-1,1'-biphenyl (**5d**):¹⁹ 92%; colorless liquid. ¹H NMR (400 MHz, chloroform-*d*) δ 7.62 – 7.54 (m, 4H), 7.47 – 7.38 (m, 4H), 7.34 (tt, *J* = 7.3, 1.5 Hz, 1H), 4.50 (s, 2H), 3.42 (s, 3H).



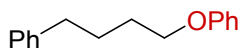
[1,1'-biphenyl]-4-ylmethyl acetate (**5e**).²⁰ $\text{CH}_3\text{CO}_2\text{K}^+$ was used as the nucleophile. 99%; colorless liquid. ^1H NMR (400 MHz, chloroform-*d*) δ 7.62 – 7.55 (m, 4H), 7.48 – 7.40 (m, 4H), 7.38 – 7.32 (m, 1H), 5.15 (s, 2H), 2.12 (s, 3H).



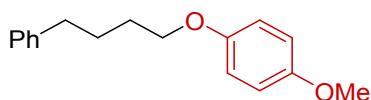
methyl 4-(acetoxymethyl)benzoate (**5f**).²¹ $\text{CH}_3\text{CO}_2\text{K}^+$ was used as the nucleophile. 96%; colorless liquid. ^1H NMR (400 MHz, Chloroform-*d*) δ 8.01 (d, J = 8.2 Hz, 2H), 7.39 (d, J = 8.2 Hz, 2H), 5.13 (s, 2H), 3.90 (s, 3H), 2.11 (s, 3H).



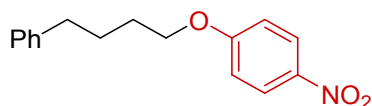
[1,1'-biphenyl]-4-ylmethyl benzoate (**5g**).²² quantitative; white solid. ^1H NMR (400 MHz, chloroform-*d*) δ 8.14 – 8.06 (m, 2H), 7.64 – 7.57 (m, 4H), 7.57 – 7.50 (m, 3H), 7.48 – 7.41 (m, 4H), 7.35 (tt, J = 7.3, 1.6 Hz, 1H), 5.41 (s, 2H).



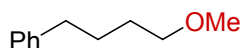
(4-phenoxybutyl)benzene (**5h**).²³ quantitative; white solid. ^1H NMR (400 MHz, chloroform-*d*) δ 7.32 – 7.23 (m, 4H), 7.23 – 7.15 (m, 3H), 6.96 – 6.86 (m, 3H), 3.96 (t, J = 5.9 Hz, 2H), 2.69 (t, J = 7.0 Hz, 2H), 1.96 – 1.67 (m, 4H).



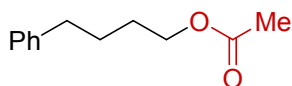
1-methoxy-4-(4-phenylbutoxy)benzene (**5i**): quantitative; colorless liquid. ^1H NMR (400 MHz, chloroform-*d*) δ 7.28 (t, J = 7.5 Hz, 2H), 7.22 – 7.15 (m, 3H), 6.84 – 6.80 (m, 4H), 3.92 (t, J = 4.9 Hz, 2H), 3.76 (s, 3H), 2.68 (t, J = 6.5 Hz, 2H), 1.85 – 1.76 (m, 4H). ^{13}C NMR (101 MHz, chloroform-*d*) δ 153.7, 153.2, 142.3, 128.4, 128.3, 125.8, 115.4, 114.6, 68.4, 55.8, 35.6, 29.0, 27.9. IR (neat) ν = 3025, 2999, 2939, 2861, 2832, 1508, 1465, 1389, 1288, 1232, 1180, 1107, 1040, 825, 745, 699, 524 cm^{-1} . HRMS (EI) Calcd. for $\text{C}_{17}\text{H}_{20}\text{O}_2$ $[\text{M}]^+$: 256.1463; found 256.1470.



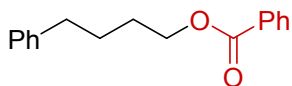
1-nitro-4-(4-phenylbutoxy)benzene (**5j**):²⁴ 46%; yellow liquid. ¹H NMR (400 MHz, chloroform-*d*) δ 8.17 (d, J = 9.3 Hz, 2H), 7.32 – 7.25 (m, 2H), 7.22 – 7.16 (m, 3H), 6.91 (d, J = 9.2 Hz, 2H), 4.04 (t, J = 5.8 Hz, 2H), 2.69 (t, J = 7.0 Hz, 2H), 1.92 – 1.74 (m, 4H).



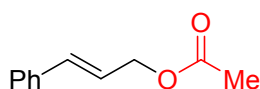
(4-methoxybutyl)benzene (**5k**):²⁵ 40%; colorless liquid. ¹H NMR (400 MHz, chloroform-*d*) δ 7.30 – 7.23 (m, 2H), 7.19 – 7.13 (m, 3H), 3.37 (t, J = 6.3 Hz, 2H), 3.31 (s, 3H), 2.62 (t, J = 7.5 Hz, 2H), 1.73 – 1.57 (m, 4H).



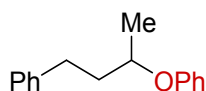
4-phenylbutyl acetate (**5l**):²⁶ CH₃CO₂⁻K⁺ was used as the nucleophile. quantitative; colorless liquid. ¹H NMR (400 MHz, chloroform-*d*) δ 7.30 – 7.25 (m, 2H), 7.21 – 7.14 (m, 3H), 4.07 (t, J = 6.2 Hz, 2H), 2.63 (t, J = 7.1 Hz, 2H), 2.03 (s, 3H), 1.73 – 1.62 (m, 4H).



4-phenylbutyl benzoate (**5m**):²⁷ 60%; colorless liquid. ¹H NMR (400 MHz, chloroform-*d*) δ 8.06 – 8.00 (m, 2H), 7.54 (t, J = 7.4 Hz, 1H), 7.43 (t, J = 7.8 Hz, 2H), 7.31 – 7.25 (m, 2H), 7.22 – 7.15 (m, 3H), 4.33 (t, J = 6.0 Hz, 2H), 2.68 (t, J = 7.0 Hz, 2H), 1.88 – 1.73 (m, 4H).

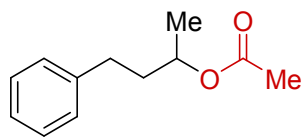


cinnamyl acetate (**5n**):²¹ CH₃CO₂⁻K⁺ was used as the nucleophile. 99%; colorless liquid. ¹H NMR (400 MHz, Chloroform-*d*) δ 7.42 – 7.35 (m, 2H), 7.35 – 7.28 (m, 2H), 7.28 – 7.20 (m, 1H), 6.64 (d, J = 15.9 Hz, 1H), 6.27 (dt, J = 15.9, 6.5 Hz, 1H), 4.71 (dd, J = 6.5, 1.1 Hz, 2H), 2.09 (s, 3H).

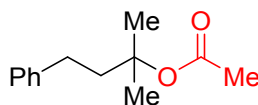


(3-phenoxybutyl)benzene (**5o**):²⁸ 32%; colorless liquid. ¹H NMR (400 MHz, chloroform-*d*) δ 7.30 – 7.25 (m, 4H), 7.21 – 7.14 (m, 3H), 6.92 (t, J = 7.3 Hz, 1H), 6.86 (d, J = 7.9 Hz, 2H), 4.36 (h, J = 6.0 Hz, 1H), 2.87 – 2.66 (m, 2H), 2.14 – 2.01 (m,

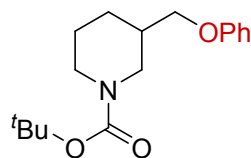
1H), 1.96 – 1.82 (m, 1H), 1.32 (d, $J = 6.1$ Hz, 3H).



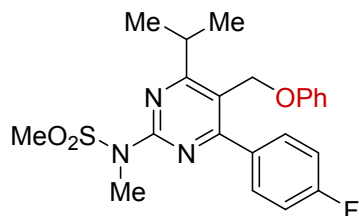
4-phenylbutan-2-yl acetate (**5p**).¹⁶ $\text{CH}_3\text{CO}_2\text{K}^+$ was used as the nucleophile. 40%; colorless liquid. ^1H NMR (400 MHz, chloroform- d) δ 7.30 – 7.23 (m, 2H), 7.20 – 7.13 (m, 3H), 4.92 (h, $J = 6.3$ Hz, 1H), 2.73 – 2.53 (m, 2H), 2.01 (s, 3H), 1.98 – 1.86 (m, 1H), 1.86 – 1.73 (m, 1H), 1.23 (d, $J = 6.3$ Hz, 3H).



2-methyl-4-phenylbutan-2-yl acetate (**5q**).¹⁶ $\text{CH}_3\text{CO}_2\text{K}^+$ was used as the nucleophile; 60 °C of reaction temperature and 5 h of reaction time were used. 44%; colorless liquid. ^1H NMR (400 MHz, Chloroform- d) δ 7.30 – 7.23 (m, 2H), 7.21 – 7.13 (m, 3H), 2.67 – 2.59 (m, 2H), 2.10 – 2.01 (m, 2H), 1.96 (s, 3H), 1.49 (s, 6H).

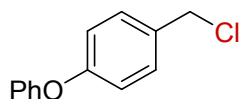


tert-butyl 3-(phenoxy)methylpiperidine-1-carboxylate (**5r**): 55%; white solid. m.p. 57 – 58 °C. ^1H NMR (400 MHz, DMSO- d_6) δ 7.29 (t, $J = 7.8$ Hz, 2H), 6.98 – 6.88 (m, 3H), 4.21 – 3.53 (m, 4H), 2.89 (br, 2H), 1.95 – 1.73 (m, 2H), 1.72 – 1.56 (m, 1H), 1.56 – 1.05 (m, 11H). ^{13}C NMR (101 MHz, chloroform- d) δ 158.9, 155.0, 129.4, 120.7, 114.5, 79.4, 69.9, 47.2, 44.5, 35.9, 28.4, 27.4, 24.5. IR (neat) $\nu = 3447, 2975, 2930, 2857, 1692, 1599, 1587, 1497, 1469, 1421, 1365, 1266, 1242, 1175, 1148, 1041, 753, 691, 668$ cm^{-1} . HRMS (EI) Calcd. for $\text{C}_{17}\text{H}_{25}\text{NO}_3$ $[\text{M}]^+$: 291.1834; found 291.1842.

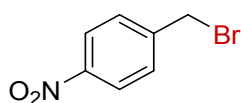


N-(4-(4-fluorophenyl)-6-isopropyl-5-(phenoxy)methylpyrimidin-2-yl)-*N*-methylmethanesulfonamide (**5s**): 77%; white solid. m.p. 129 – 130 °C. ^1H NMR (400 MHz, chloroform- d) δ 7.77 – 7.69 (m, 2H), 7.37 – 7.29 (m, 2H), 7.08 (t, $J = 8.7$ Hz, 2H), 7.03 (t, $J = 7.4$ Hz, 1H), 6.94 (d, $J = 8.6$ Hz, 2H), 4.89 (s, 2H), 3.59 (s, 3H), 3.52 (s, 3H), 3.40 – 3.26 (m, 1H), 1.32 (d, $J = 6.7$ Hz, 6H). ^{19}F NMR (376 MHz, chloroform- d) δ -110.85 – -110.98 (m, 1F). ^{13}C NMR (101 MHz, chloroform- d) δ

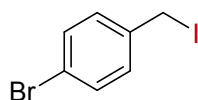
178.4, 166.6, 163.8 (d, $J = 250.3$ Hz), 158.4, 158.0, 133.8 (d, $J = 3.1$ Hz), 131.4 (d, $J = 8.5$ Hz), 129.7, 121.6, 117.5, 115.6 (d, $J = 21.7$ Hz), 114.7, 63.3, 42.5, 33.1, 31.8, 22.1. IR (neat) $\nu = 2974, 2931, 1872, 1605, 1594, 1553, 1480, 1339, 1231, 1155, 1069, 1029, 957, 896, 848, 820, 772, 756, 691, 600, 575, 521, 495$ cm⁻¹. HRMS (EI) Calcd. for C₂₂H₂₄N₃O₃FS [M]⁺: 429.1522; found 429.1528.



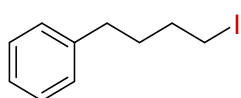
1-(chloromethyl)-4-phenoxybenzene (**5t**).²⁹ ⁿBu₄N⁺Cl⁻ was used as the nucleophile. 55%; colorless liquid. ¹H NMR (400 MHz, chloroform-*d*) δ 7.40 – 7.30 (m, 4H), 7.12 (t, $J = 7.4$ Hz, 1H), 7.02 (d, $J = 7.7$ Hz, 2H), 6.97 (d, $J = 8.6$ Hz, 2H), 4.58 (s, 2H).



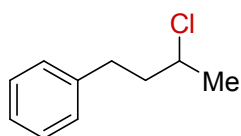
1-(bromomethyl)-4-nitrobenzene (**5u**).³⁰ ⁿBu₄N⁺Br⁻ was used as the nucleophile. quantitative; white solid. ¹H NMR (400 MHz, chloroform-*d*) δ 8.14 (d, $J = 8.7$ Hz, 2H), 7.51 (d, $J = 8.7$ Hz, 2H), 4.46 (s, 2H).



1-bromo-4-(iodomethyl)benzene (**5v**).³¹ NaI was not required as the iodide anion was generated from ICH₂CH₂I; 60 °C of reaction temperature and 2 h of reaction time in CH₃CN were used. 92%; white solid. ¹H NMR (500 MHz, chloroform-*d*) δ 7.44 – 7.40 (m, 2H), 7.26 – 7.23 (m, 2H), 4.40 (s, 2H).

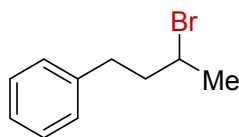


(4-iodobutyl)benzene (**5w**).¹² NaI was not required as the iodide anion was generated from ICH₂CH₂I; 60 °C of reaction temperature and 2 h of reaction time in CH₃CN were used. 85%; yellow liquid. ¹H NMR (400 MHz, chloroform-*d*) δ 7.28 (t, $J = 7.4$ Hz, 2H), 7.23 – 7.15 (m, 3H), 3.20 (t, $J = 6.9$ Hz, 2H), 2.63 (t, $J = 7.5$ Hz, 2H), 1.92 – 1.80 (m, 2H), 1.80 – 1.68 (m, 2H).

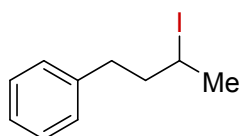


(3-chlorobutyl)benzene (**5x**).³² ⁿBu₄N⁺Cl⁻ was used as the nucleophile. 79%; colorless liquid. ¹H NMR (400 MHz, chloroform-*d*) δ 7.33 – 7.25 (m, 2H), 7.23 – 7.16 (m, 3H), 3.99 (h, $J = 6.5$ Hz, 1H), 2.90 – 2.80 (m, 1H), 2.79 – 2.69 (m, 1H), 2.05 – 1.97 (m,

2H), 1.53 (d, $J = 6.5$ Hz, 3H).



(3-bromobutyl)benzene (**5y**).³³ $n\text{Bu}_4\text{N}^+\text{Br}^-$ was used as the nucleophile. 62%; colorless liquid. ^1H NMR (400 MHz, chloroform- d) δ 7.32 – 7.26 (m, 2H), 7.23 – 7.16 (m, 3H), 4.13 – 4.02 (m, 1H), 2.91 – 2.81 (m, 1H), 2.79 – 2.69 (m, 1H), 2.19 – 1.98 (m, 2H), 1.72 (d, $J = 6.7$ Hz, 3H).



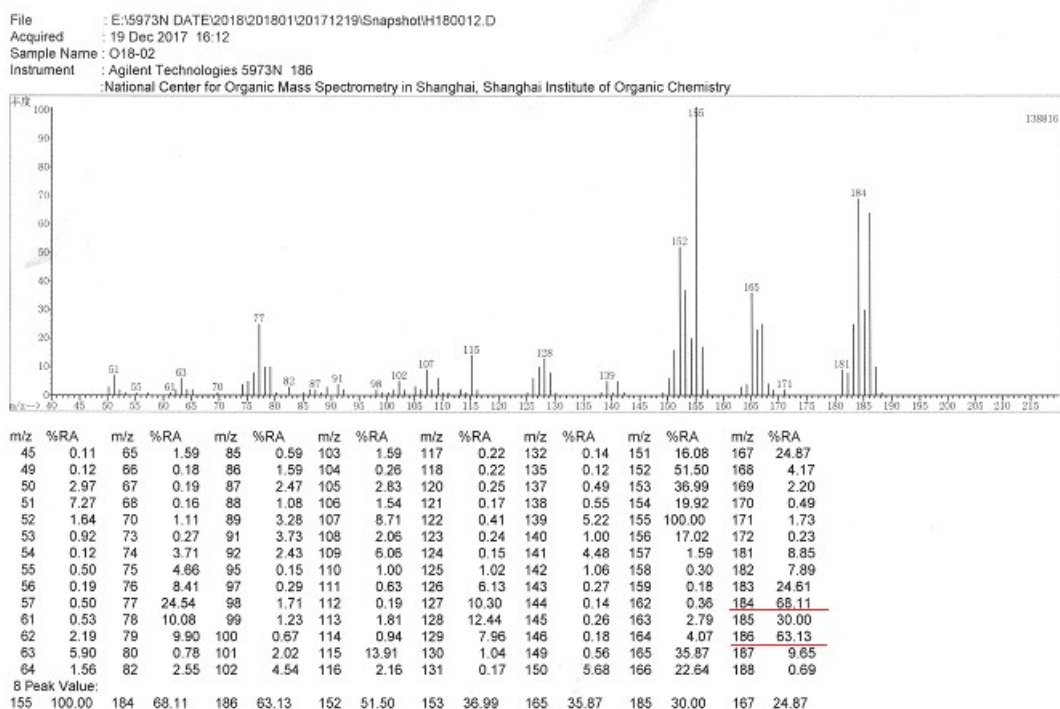
(3-iodobutyl)benzene (**5z**).³⁴ NaI was not required as the iodide anion was generated from $\text{ICH}_2\text{CH}_2\text{I}$; 60 °C of reaction temperature and 2 h of reaction time in CH_3CN were used. 35%; colorless liquid. ^1H NMR (400 MHz, chloroform- d) δ 7.32 – 7.26 (m, 2H), 7.24 – 7.16 (m, 3H), 4.19 – 4.05 (m, 1H), 2.90 – 2.80 (m, 1H), 2.75 – 2.64 (m, 1H), 2.22 – 2.10 (m, 1H), 1.95 (d, $J = 6.8$ Hz, 3H), 1.93 – 1.82 (m, 1H).

5 The synthesis of ^{18}O -1a

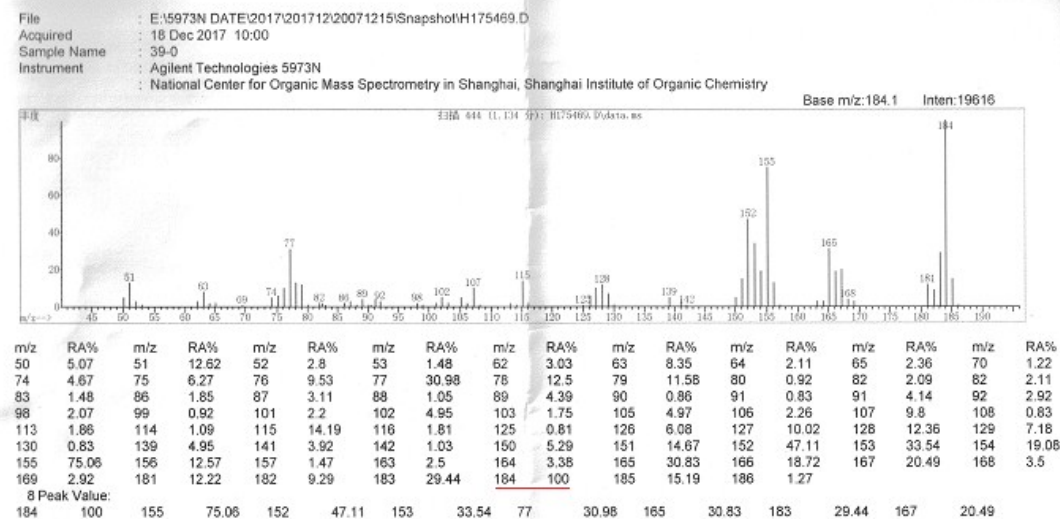
NaH (0.6 mmol, 14.4 mg) was added into a solution of H_2^{18}O (^{18}O content: 95%, 0.6 mmol, 12.0 mg) in anhydrous DMF (1.0 mL) under N_2 atmosphere. The mixture was stirred for around 3 min to afford the suspension of Na^{18}OH in anhydrous DMF.

Alcohol **1a** (1.0 equiv, 0.2 mmol, 36.8 mg), triphenylphosphine (1.2 equiv, 0.24 mmol, 62.9 mg) and anhydrous DMF (1.0 mL) were added into a 10 mL sealed tube under a N_2 atmosphere. 1,2-Diiodoethane (1.2 equiv, 0.24 mmol, 67.6 mg) was then added and the resulting mixture was stirred for around 1 min until the 1,2-diiodoethane was completely dissolved. The above Na^{18}OH suspension was added subsequently and the mixture was stirred at 50 °C for 5 h. Dichloromethane (20 mL) was added and the resulting solution was washed with water (20 mL x 3). The organic layer was dried over Na_2SO_4 . After filtration, the solvent was removed by concentration under reduced pressure. The residue was subjected to flash column chromatography to afford product **^{18}O -1a** (25.0 mg, 67%). MS-EI analysis of the product showed that $^{18}\text{O}:^{16}\text{O} = 50:50$.

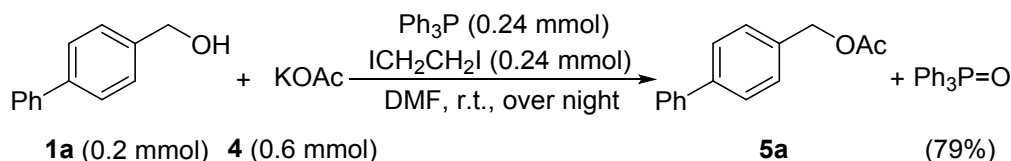
MS-EI spectrum of the ^{18}O -product:



MS-EI spectrum of the substrate **1a**:

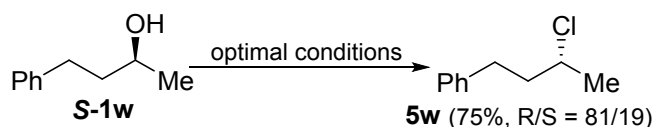


6. Isolation of Ph₃P=O



Esterification of alcohol **1a** (0.2 mmol) with KOAc occurred well under the optimal conditions. After the reaction was completed, 53.0 mg of Ph₃P=O was isolated in 79% yield (based on Ph₃P). ¹H NMR (400 MHz, chloroform-*d*) δ 7.72 – 7.60 (m, 6H), 7.58 – 7.48 (m, 3H), 7.49 – 7.37 (m, 6H). ³¹P NMR (162 MHz, chloroform-*d*) δ 29.86 (s, 1P).

7. Inversion of configuration



S-1w (0.5 mmol) was converted into **R-5w** under the optimized conditions. Enantiomeric excess was determined by chiral GC with a CP-chirasil-DEX CB column (length 25 m, diameter 0.25 mm, 0.25 μm layer thickness) and nitrogen as carrier gas. Injector temperature was 100 °C. Major enantiomer *rt* = 36.417 min, minor enantiomer *rt* = 37.465 min. GC analysis from the isolated product showed an *er* of 81:19.

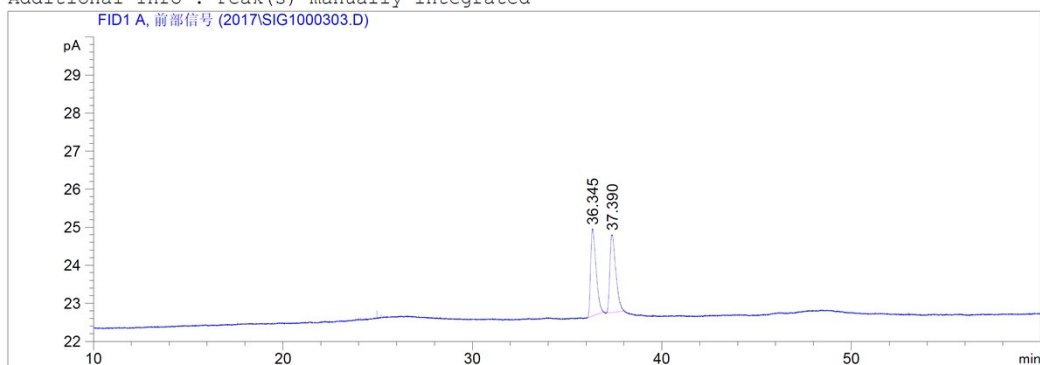
[α]_D²⁵ = -37.4 (*c* = 1.15 g/100 mL, CHCl₃, *er* 81:19).

Lit. [α]_D²⁵ = -36.4 (*c* = 1.00 g/100 mL, CHCl₃, *R*-ent, no enantiopurity given).³²

Gas chromatogram of a racemic sample:

Data File C:\CHEM32\1\DATA\2017\SIG1000303.D
Sample Name: 08-90 R/S

```
=====
Acq. Operator   : YZR
Acq. Instrument : 仪器 1                      Location : Vial 1
Injection Date  : 9/25/2017 12:32:29 PM      Inj Volume : Manually
Acq. Method     : C:\CHEM32\1\METHODS\TEST2-1.M
Last changed    : 9/25/2017 1:41:10 PM by YZR
                  (modified after loading)
Analysis Method : C:\CHEM32\1\METHODS\TEST2-1.M
Last changed    : 4/20/2018 3:20:34 PM by lx1
                  (modified after loading)
Additional Info  : Peak(s) manually integrated
FID1 A, 前部信号 (2017\SIG1000303.D)
```



Area Percent Report

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=====
Sorted By      : Signal
Multiplier:    : 1.0000
Dilution:      : 1.0000
Use Multiplier & Dilution Factor with ISTDs
```

Signal 1: FID1 A, 前部信号

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	36.345	BB	0.2435	45.28983	2.28254	50.45764
2	37.390	BB	0.2907	44.46829	2.03692	49.54236

Totals : 89.75812 4.31946

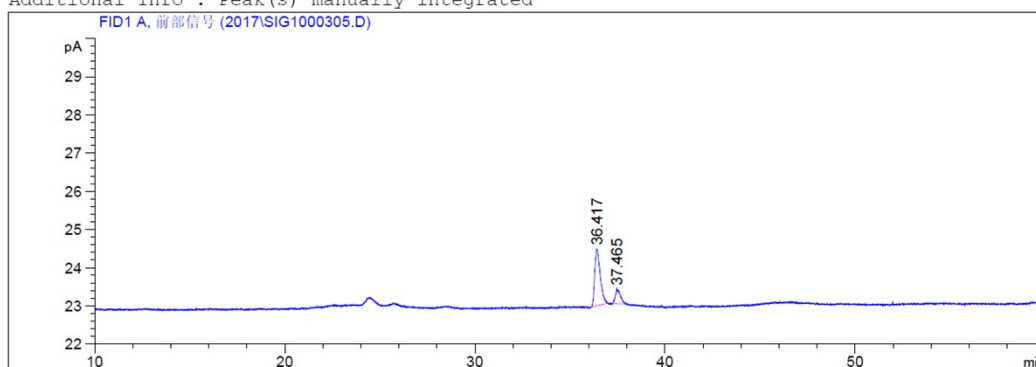
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*** End of Report ***
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Gas chromatogram of the enantioenriched sample:

Data File C:\CHEM32\1\DATA\2017\SIG1000305.D
Sample Name: 0899

```
=====
Acq. Operator   : YZR
Acq. Instrument : 仪器 1                      Location : Vial 1
Injection Date  : 9/25/2017 2:50:23 PM
                                           Inj Volume : Manually

Acq. Method     : C:\CHEM32\1\METHODS\TEST2-1.M
Last changed    : 9/25/2017 2:49:20 PM by YZR
                  (modified after loading)
Analysis Method : C:\CHEM32\1\METHODS\TEST2-1.M
Last changed    : 4/20/2018 3:20:34 PM by lxl
                  (modified after loading)
Additional Info  : Peak(s) manually integrated
```



Area Percent Report

```
=====
Sorted By      : Signal
Multiplier:    : 1.0000
Dilution:      : 1.0000
Use Multiplier & Dilution Factor with ISTDs
```

Signal 1: FID1 A, 前部信号

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	36.417	BB	0.2578	29.82327	1.47542	81.04265
2	37.465	BB	0.2300	6.97621	3.60126e-1	18.95735

Totals : 36.79947 1.83555

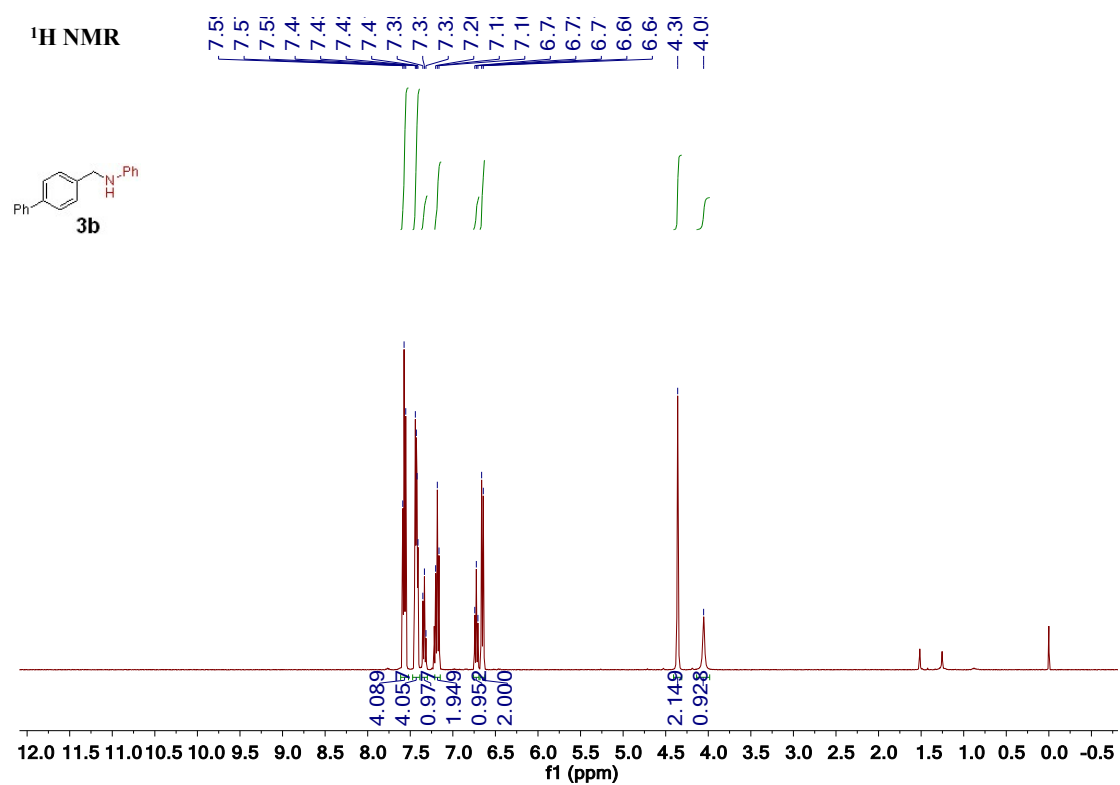
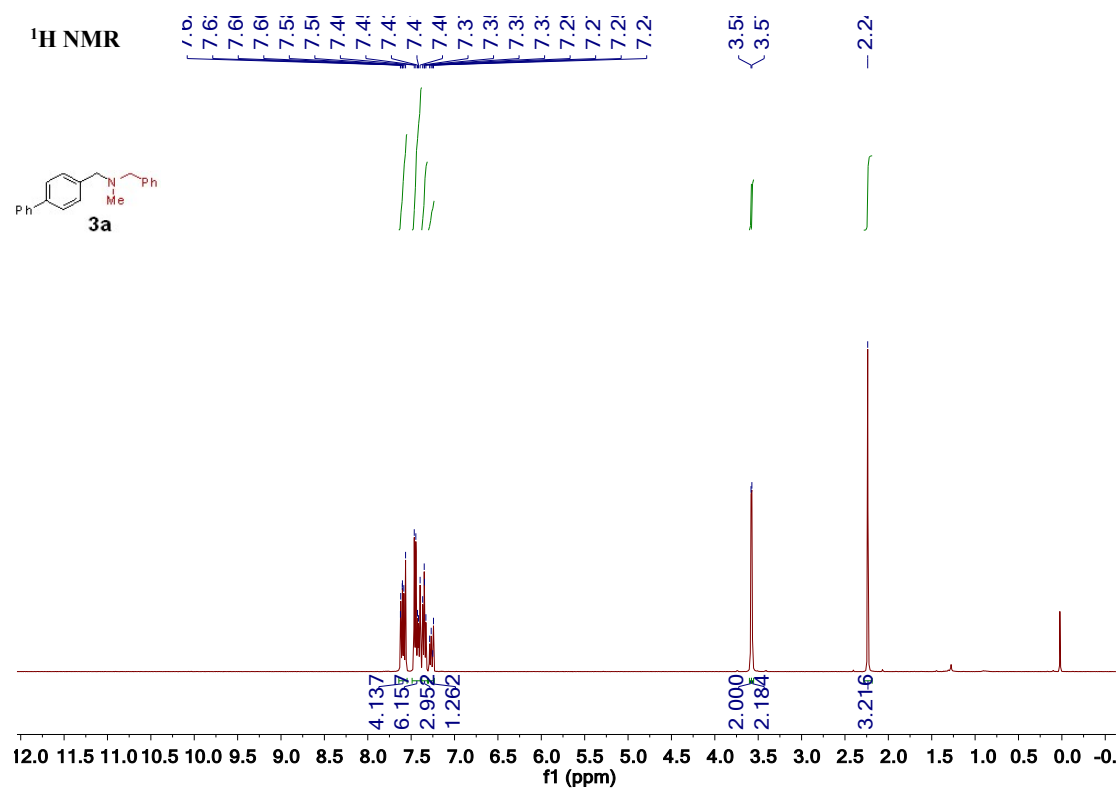
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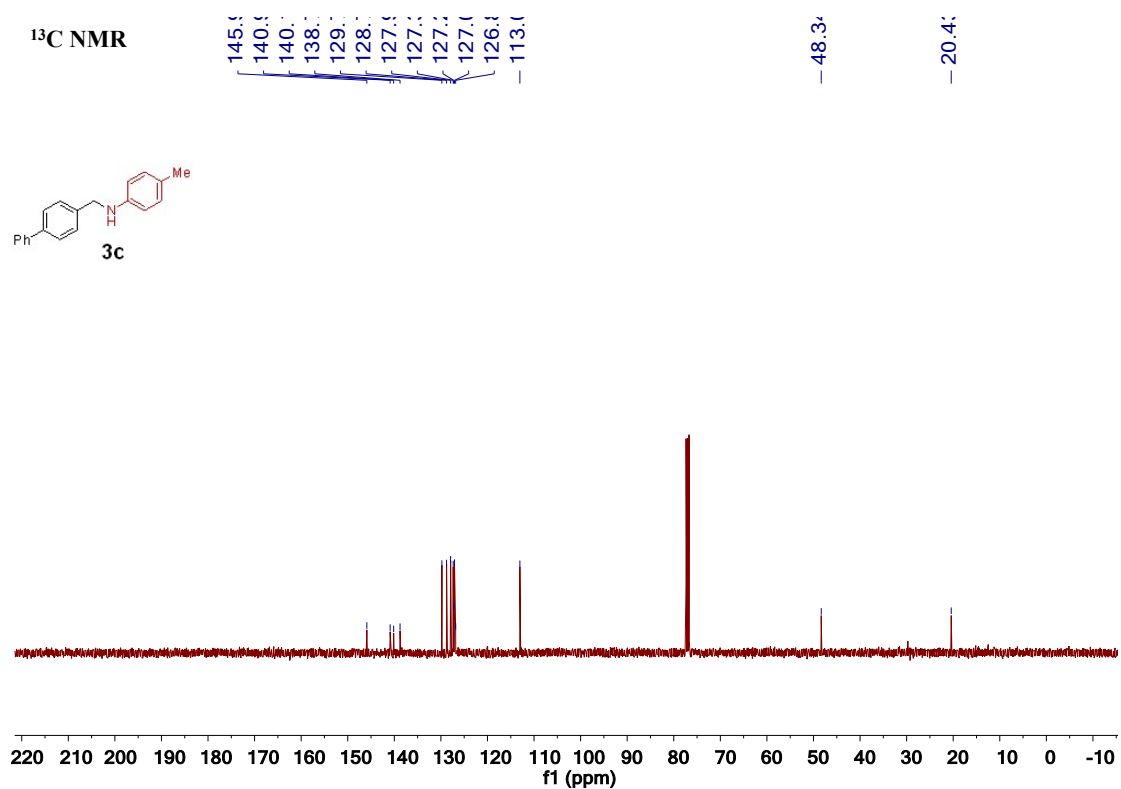
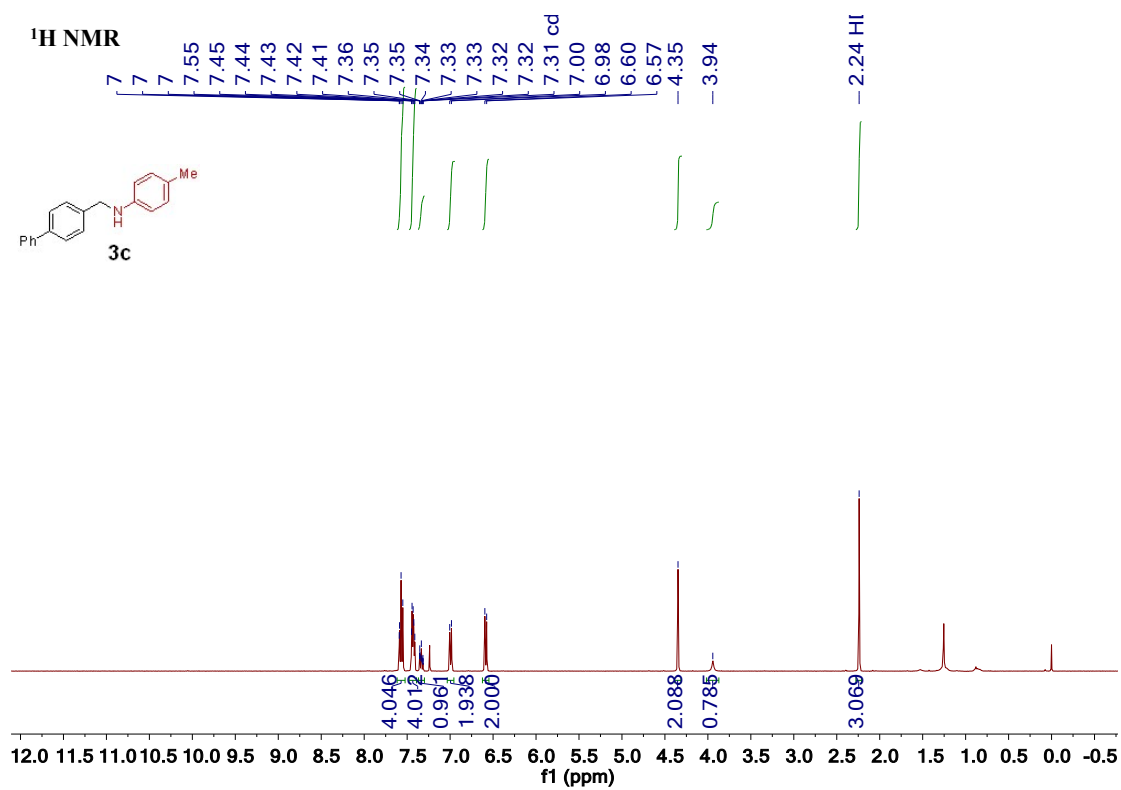
8. References and notes

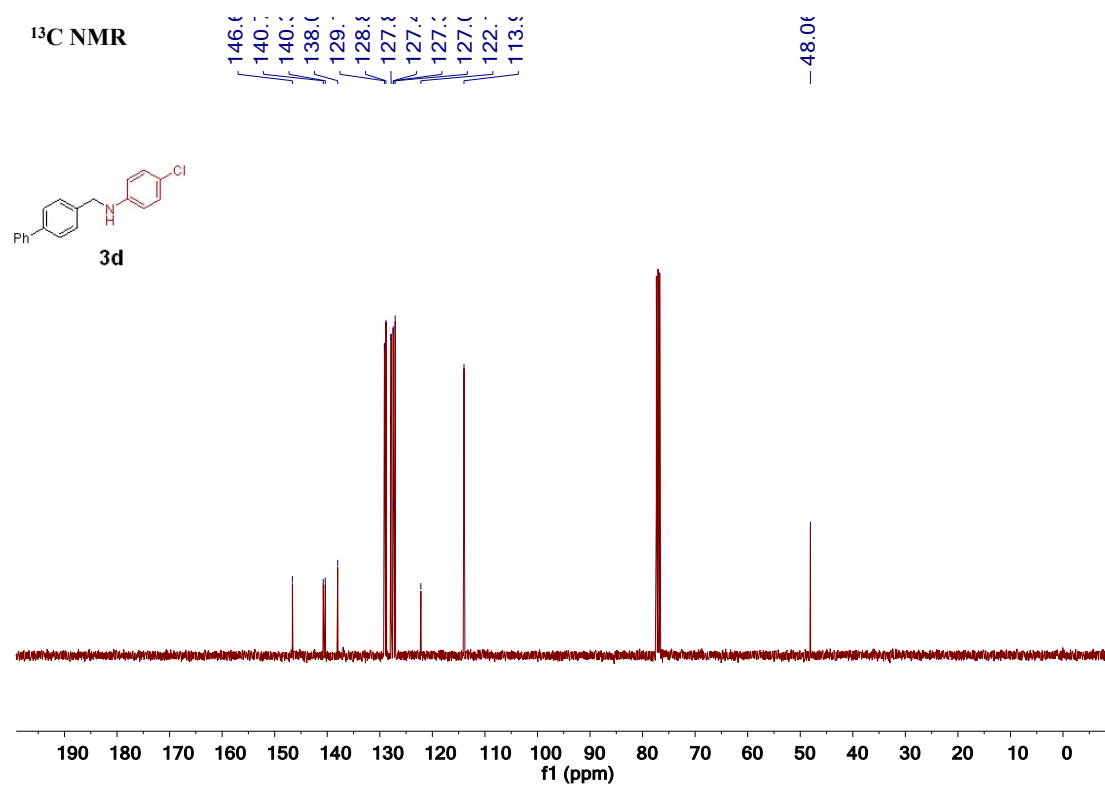
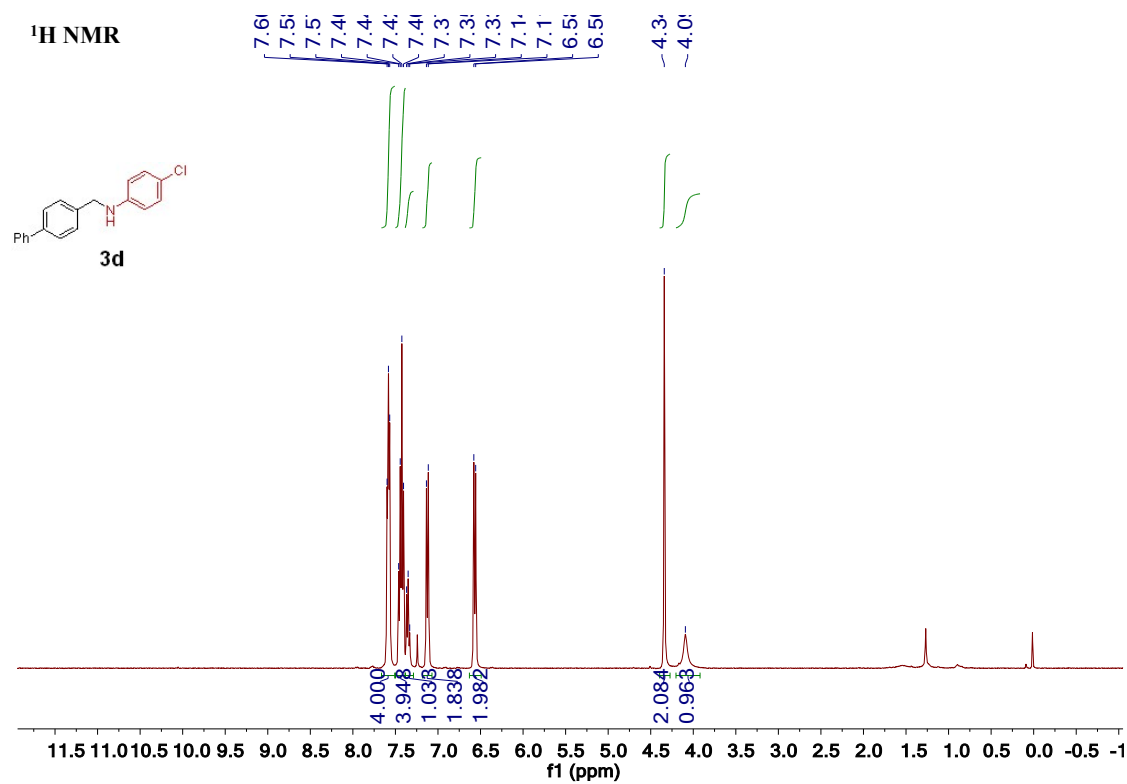
1. L. C. Gilday, S. W. Robinson, T. A. Barendt, M. J. Langton, B. R. Mullaney and P. D. Beer, *Chem. Rev.*, 2015, **115**, 7118-7195.
2. (a) R. Appel and H. Schoeler, *Chem. Ber.*, 1977, **110**, 2382-2384; (b) M. A. H. A. Al-Juboori, P. N. Gates and A. S. Muir, *J. Chem. Soc., Chem. Commun.*, 1991, 1270-1271; (c) Q. Yin, Y. Ye, G. Tang and Y. F. Zhao, *Spectrochim. Acta A.*, 2006, **63**, 192-195.
3. P. J. Garegg, T. Regberg, J. Stawinski and R. Stromberg, *J. Chem. Soc., Perkin Trans. 2*, 1987, **0**, 271-274.
4. S. P. Morcillo, L. Alvarez de Cienfuegos, A. J. Mota, J. Justicia and R. Robles, *J. Org. Chem.*, 2011, **76**, 2277-2281.
5. P. A. Champagne, J. Pomarole, M.-È. Thérien, Y. Benhassine, S. Beaulieu, C. Y. Legault and J.-F. Paquin, *Org. Lett.*, 2013, **15**, 2210-2213.
6. Y. Y. Gui, L. L. Liao, L. Sun, Z. Zhang, J. H. Ye, G. Shen, Z. P. Lu, W. J. Zhou and D. G. Yu, *Chem. Commun.*, 2017, **53**, 1192-1195.
7. S. Pramanik and P. Ghorai, *Org. Lett.*, 2014, **16**, 2104-2107.
8. X. Hua, J. Masson-Makdissi, R. J. Sullivan and S. G. Newman, *Org. Lett.*, 2016, **18**, 5312-5315.
9. Z. Chen, H. Zeng, S. A. Girard, F. Wang, N. Chen and C.-J. Li, *Angew. Chem. Int. Ed.*, 2015, **54**, 14487-14491.
10. S. Zhu, N. Niljianskul and S. L. Buchwald, *J. Am. Chem. Soc.*, 2013, **135**, 15746-15749.
11. S. M. Bronner and R. H. Grubbs, *Chem. Sci.*, 2014, **5**, 101-106.
12. L. Candish, E. A. Standley, A. Gomez-Suarez, S. Mukherjee and F. Glorius, *Chem. Eur. J.*, 2016, **22**, 9971-9974.
13. C. Liu, X. Wang, Z. Li, L. Cui and C. Li, *J. Am. Chem. Soc.*, 2015, **137**, 9820-9823.
14. C. P. Owen, I. Shahid, M. S. Olusanjo, C. H. Patel, S. Dhanani and S. Ahmed, *J. Steroid Biochem. Mol. Biol.*, 2008, **111**, 117-127.
15. T. Tamai, K. Fujiwara, S. Higashimae, A. Nomoto and A. Ogawa, *Org. Lett.*, 2016, **18**, 2114-2117.
16. Y. Nishimoto, A. Okita, M. Yasuda and A. Baba, *Org. Lett.*, 2012, **14**, 1846-1849.
17. M. Thangaraj, S. S. Bhojgude, M. V. Mane and A. T. Biju, *Chem. Commun.*, 2016, **52**, 1665-1668.
18. M. de Candia, F. Liantonio, A. Carotti, R. De Cristofaro and C. Altomare, *J. Med. Chem.*, 2009, **52**, 1018-1028.
19. B.-T. Guan, S.-K. Xiang, B.-Q. Wang, Z.-P. Sun, Y. Wang, K.-Q. Zhao and Z.-J. Shi, *J. Am. Chem. Soc.*, 2008, **130**, 3268-3269.
20. T.-C. Chang and S. J. Yu, *Synthetic Commun.*, 2015, **45**, 651-662.
21. T. Mino, T. Hasegawa, Y. Shirae, M. Sakamoto and T. Fujita, *J. Org. Chem.*, 2007, **692**, 4389-4396.
22. S. Burugupalli, S. Shah, P. L. van der Peet, S. Arora, J. M. White and S. J.

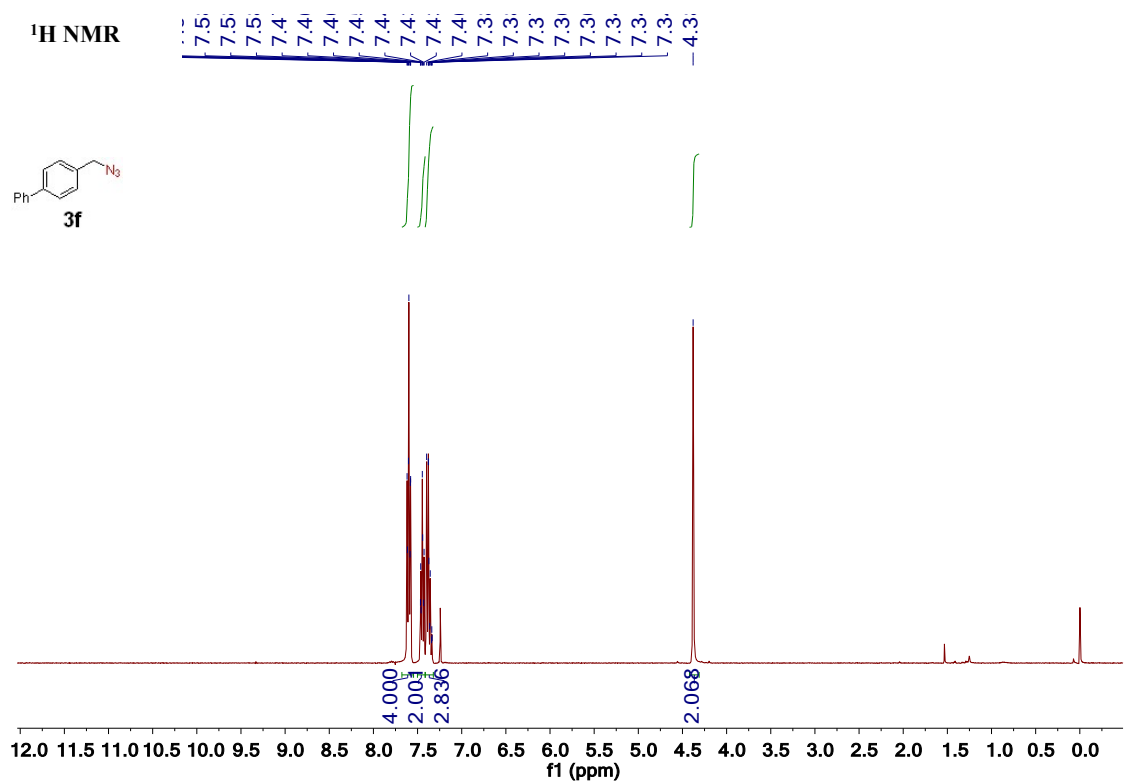
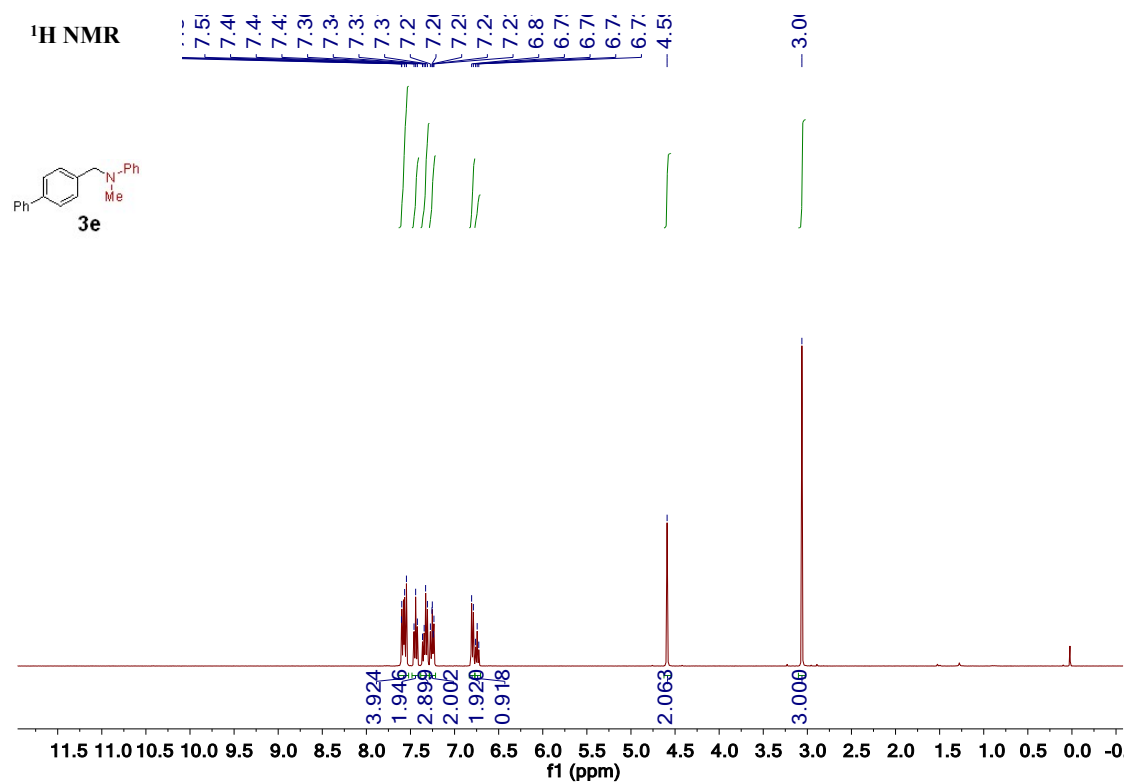
- Williams, *Org. Biomol. Chem.*, 2016, **14**, 97-104.
23. E. M. v. Duzee and H. Adkins, *J. Am. Chem. Soc.*, 1935, **57**, 147-151.
 24. A. Catalano, A. Carocci, I. Defrenza, M. Muraglia, A. Carrieri, F. Van Bambeke, A. Rosato, F. Corbo and C. Franchini, *Eur. J. Med. Chem.*, 2013, **64**, 357-364.
 25. M. Yato, K. Homma and A. Ishida, *Tetrahedron*, 2001, **57**, 5353-5359.
 26. P. Lu, T. Hou, X. Gu and P. Li, *Org. Lett.*, 2015, **17**, 1954-1957.
 27. J. D. St. Denis, C. C. G. Scully, C. F. Lee and A. K. Yudin, *Org. Lett.*, 2014, **16**, 1338-1341.
 28. C.-G. Yang and C. He, *J. Am. Chem. Soc.*, 2005, **127**, 6966-6967.
 29. M. A. Santos, S. M. Marques, T. Tuccinardi, P. Carelli, L. Panelli and A. Rossello, *Bioorg. Med. Chem.*, 2006, **14**, 7539-7550.
 30. T. V. Nguyen and A. Bekensir, *Org. Lett.*, 2014, **16**, 1720-1723.
 31. D. Das, J. M. H Anal and L. Rokhum, *J. Chem. Sci.*, 2016, **128**, 1695-1701.
 32. P. H. Huy, S. Motsch and S. M. Kappler, *Angew. Chem. Int. Ed.*, 2016, **55**, 10145-10149.
 33. X. Ma, M. Diane, G. Ralph, C. Chen and M. R. Biscoe, *Angew. Chem. Int. Ed.*, 2017, **56**, 12663-12667.
 34. A. R. Ellwood and M. J. Porter, *J. Org. Chem.*, 2009, **74**, 7982-7985.

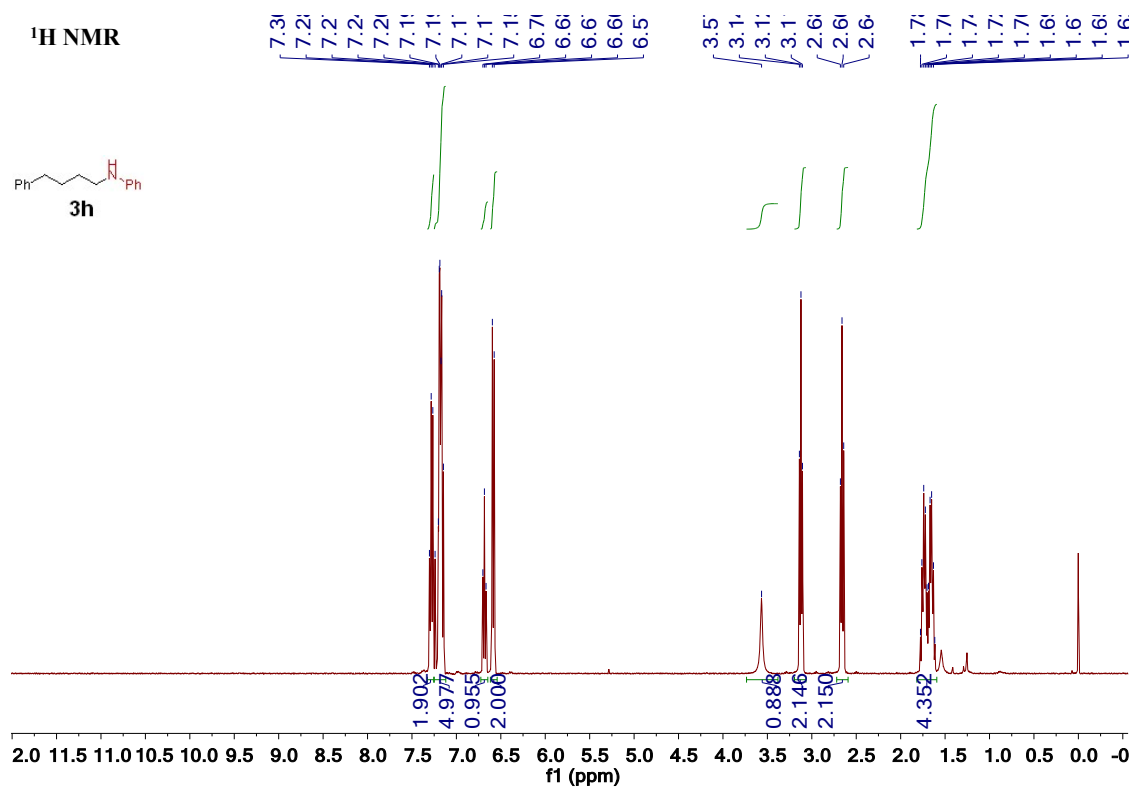
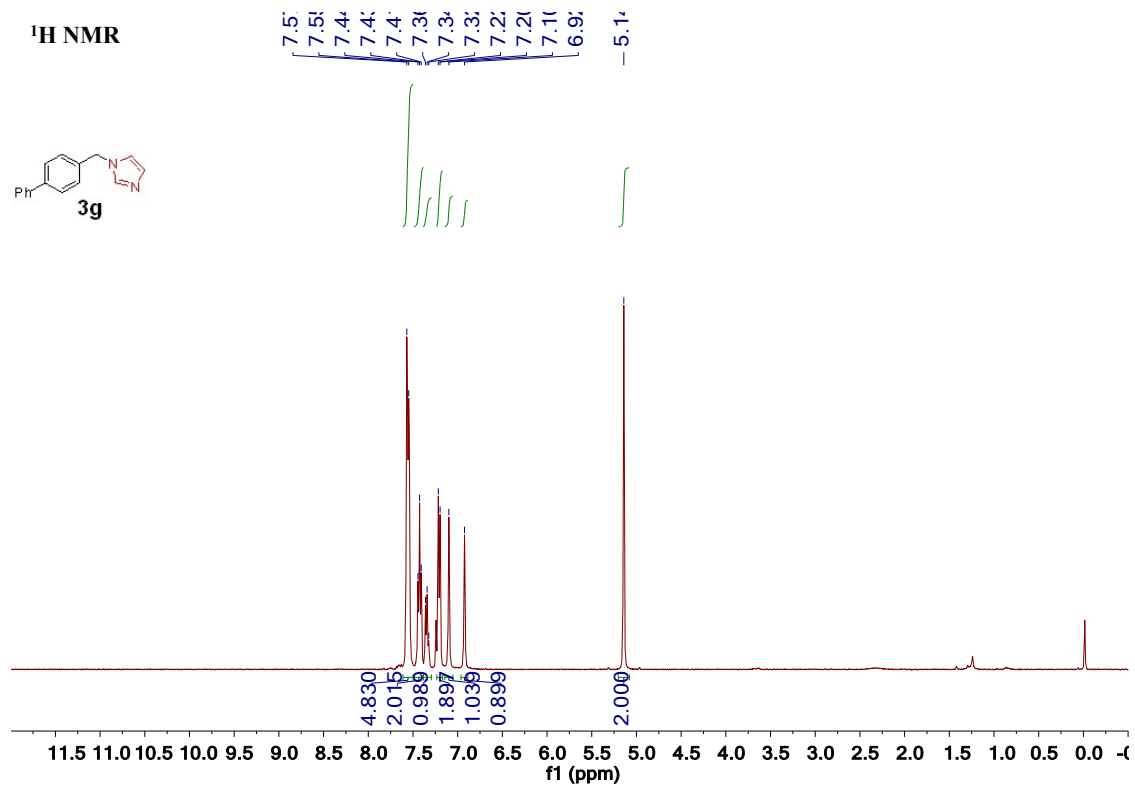
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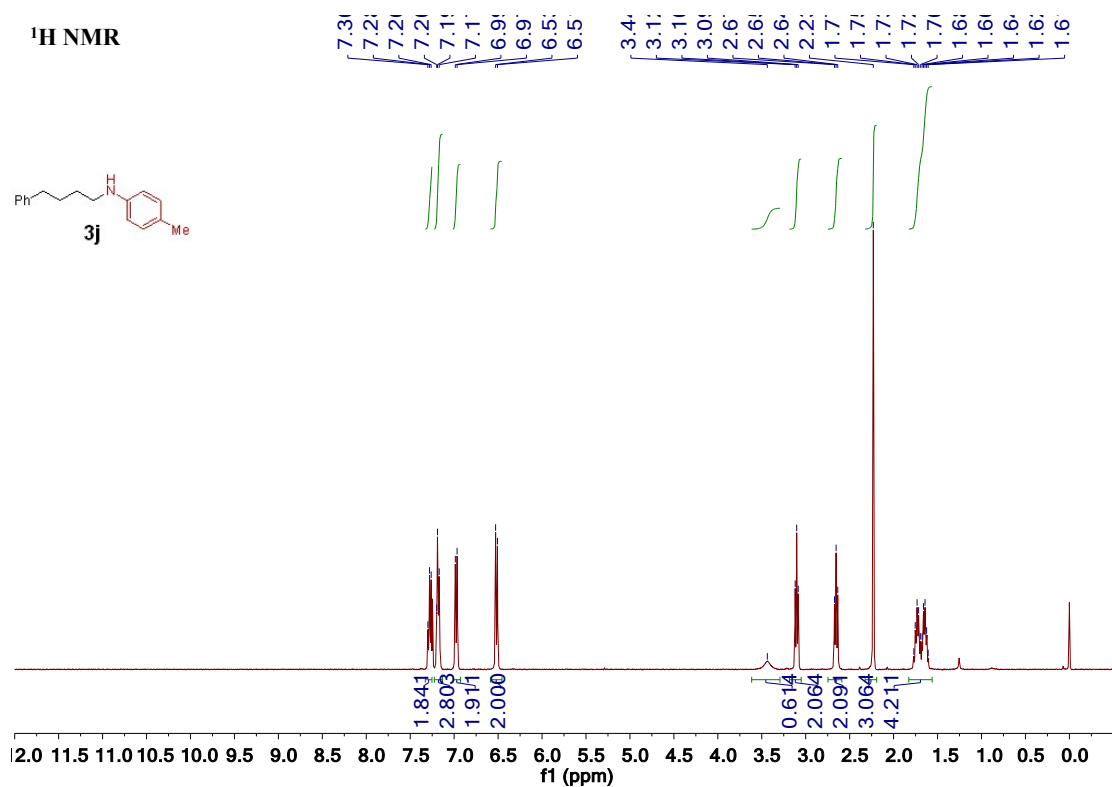
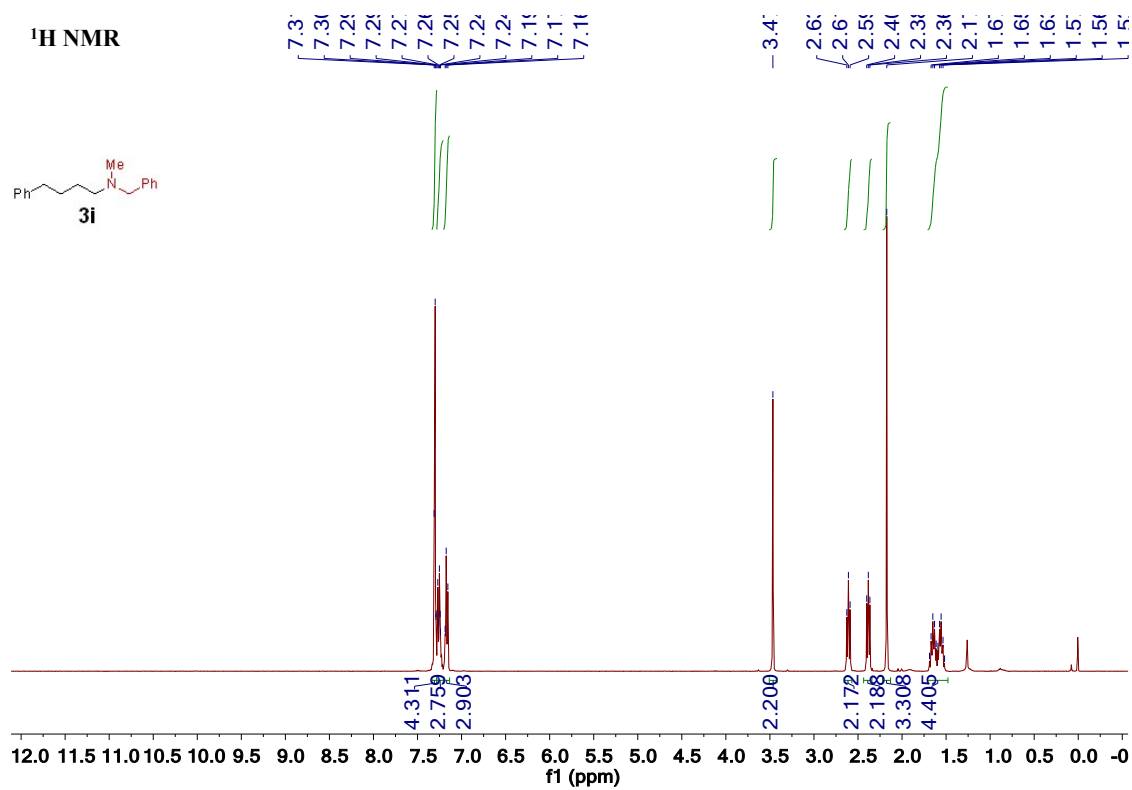


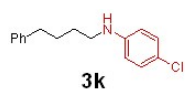
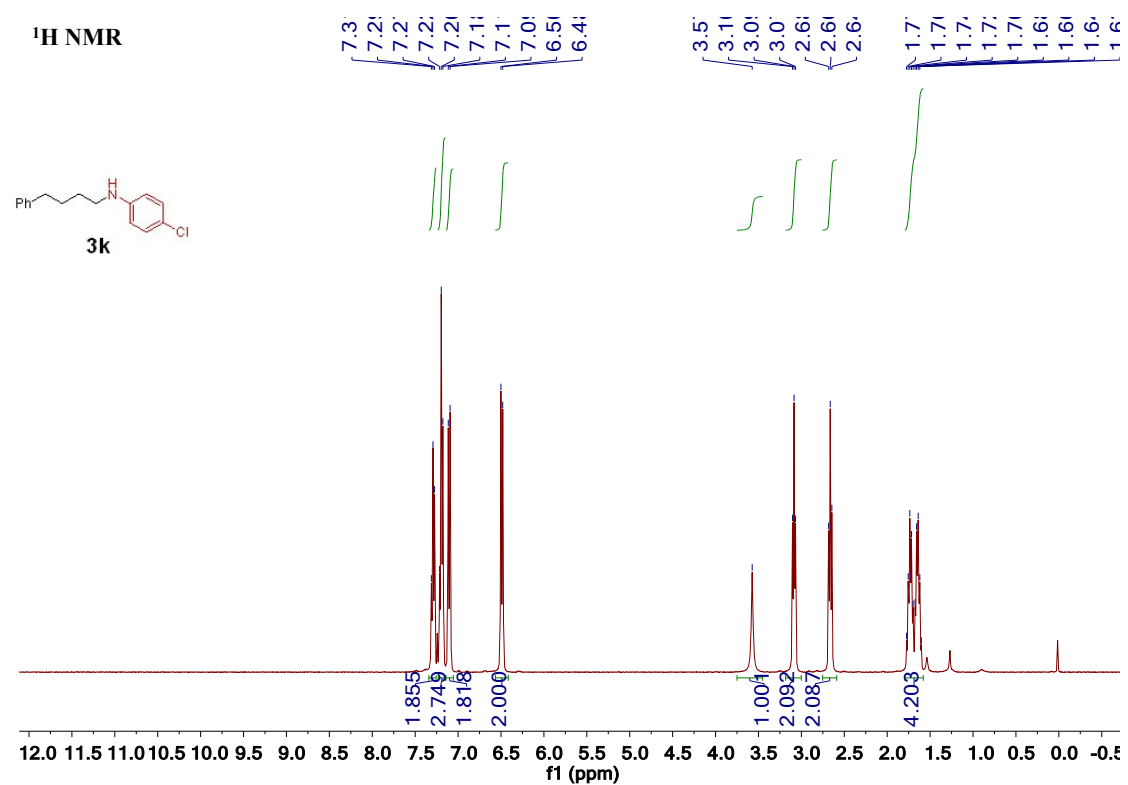
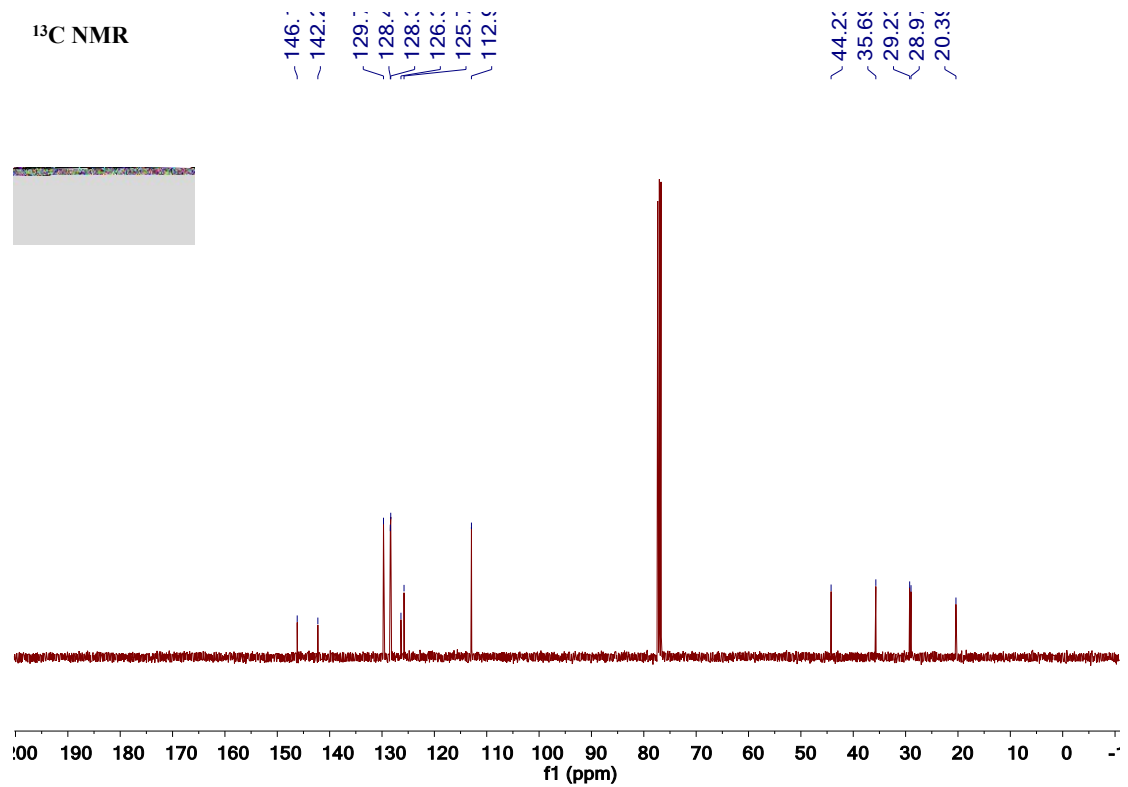


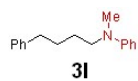
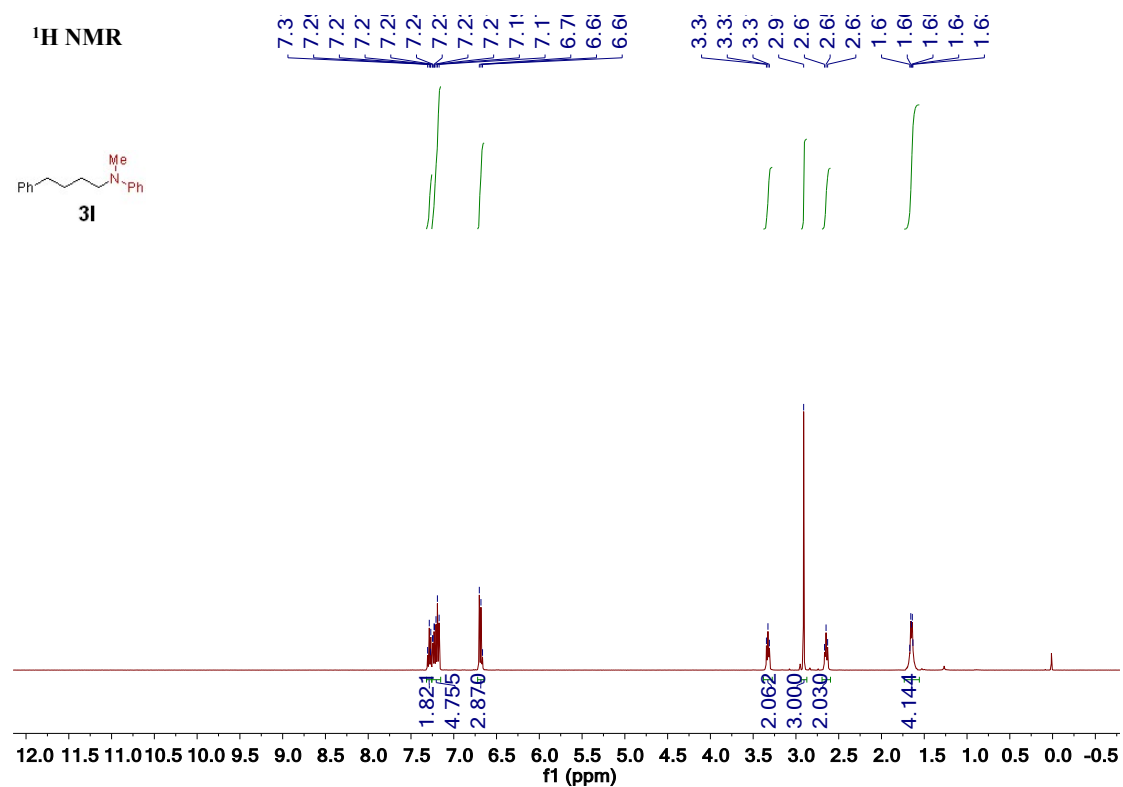
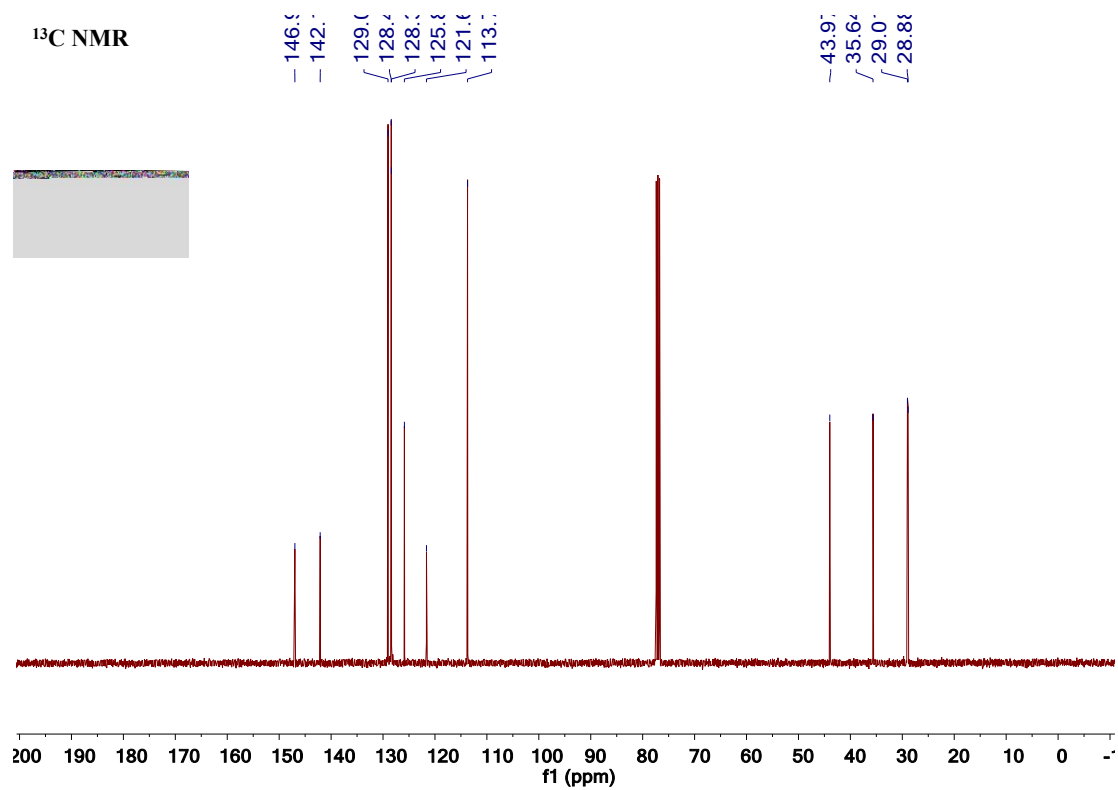


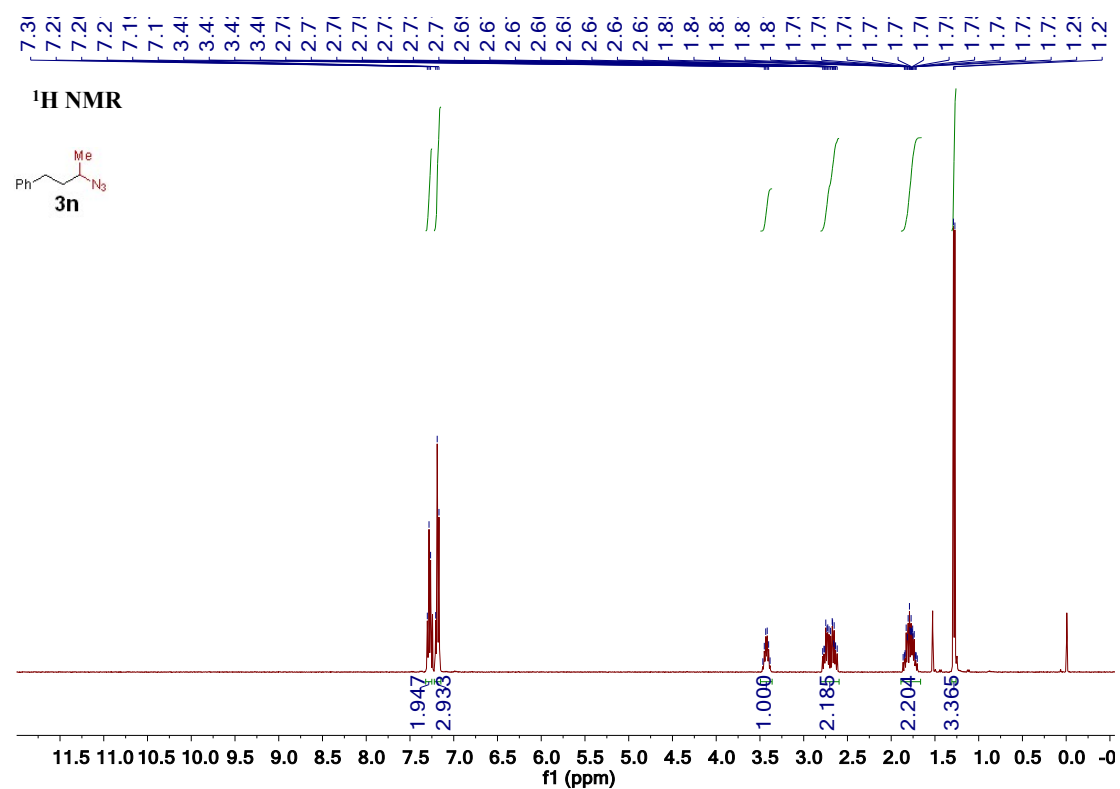
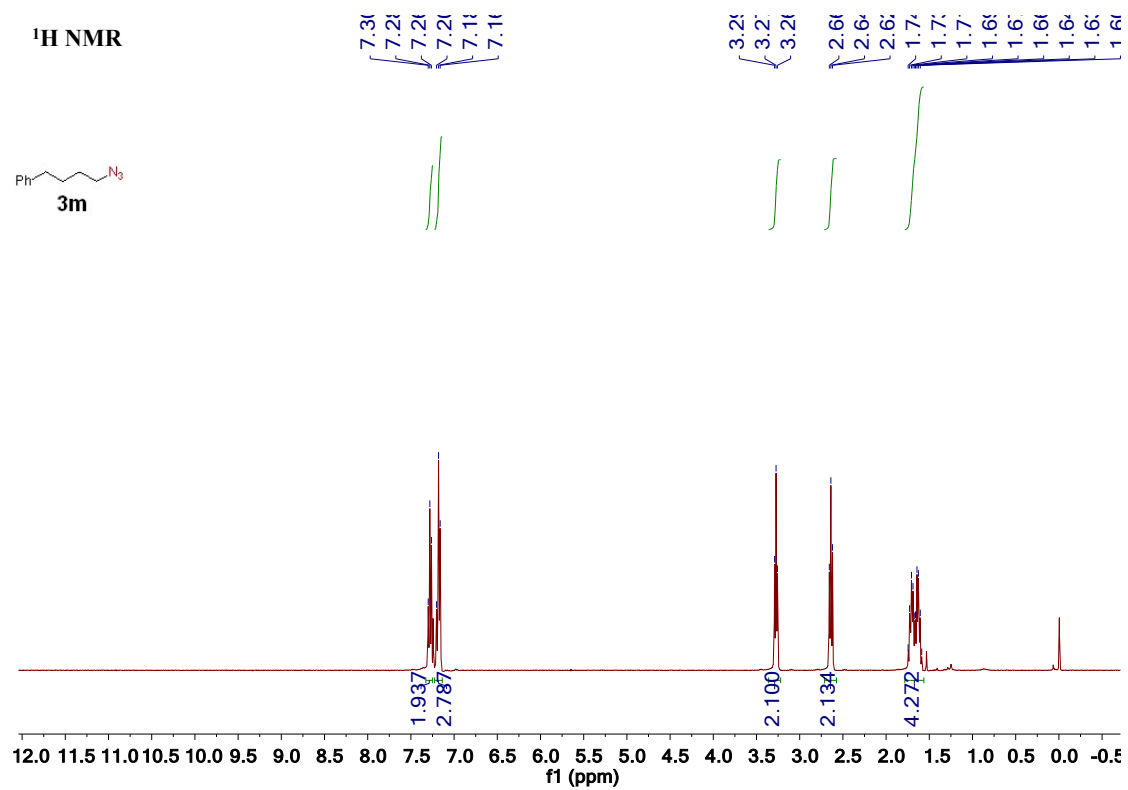


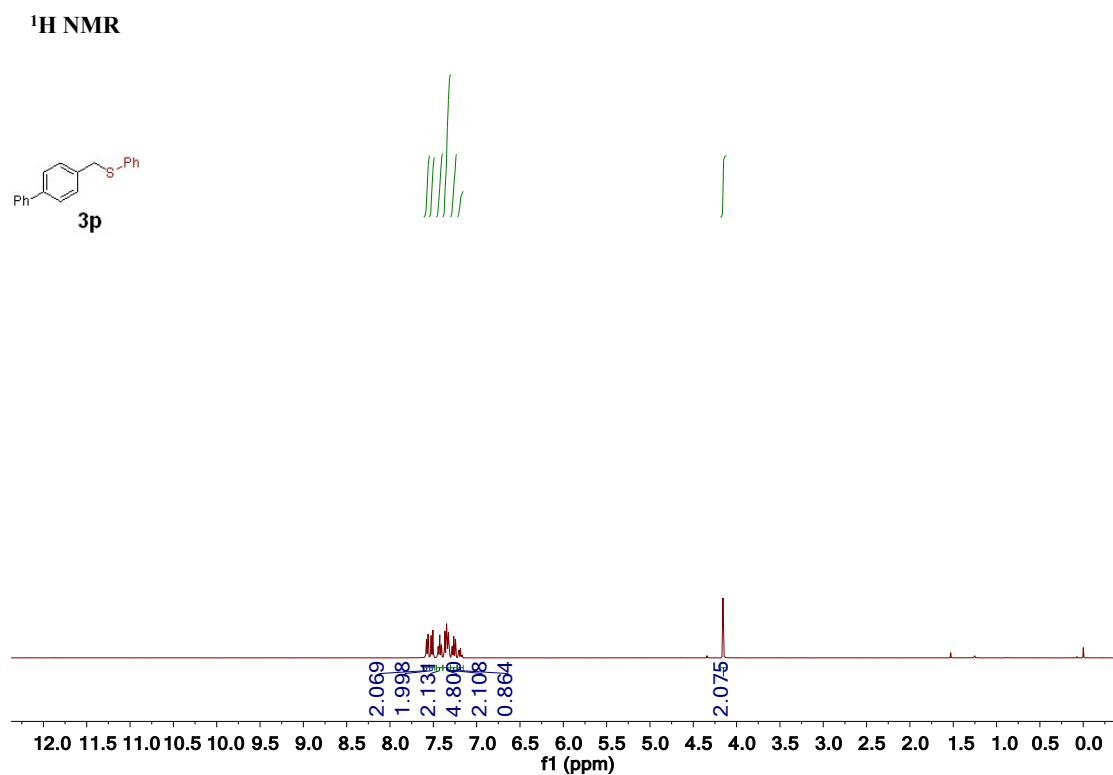
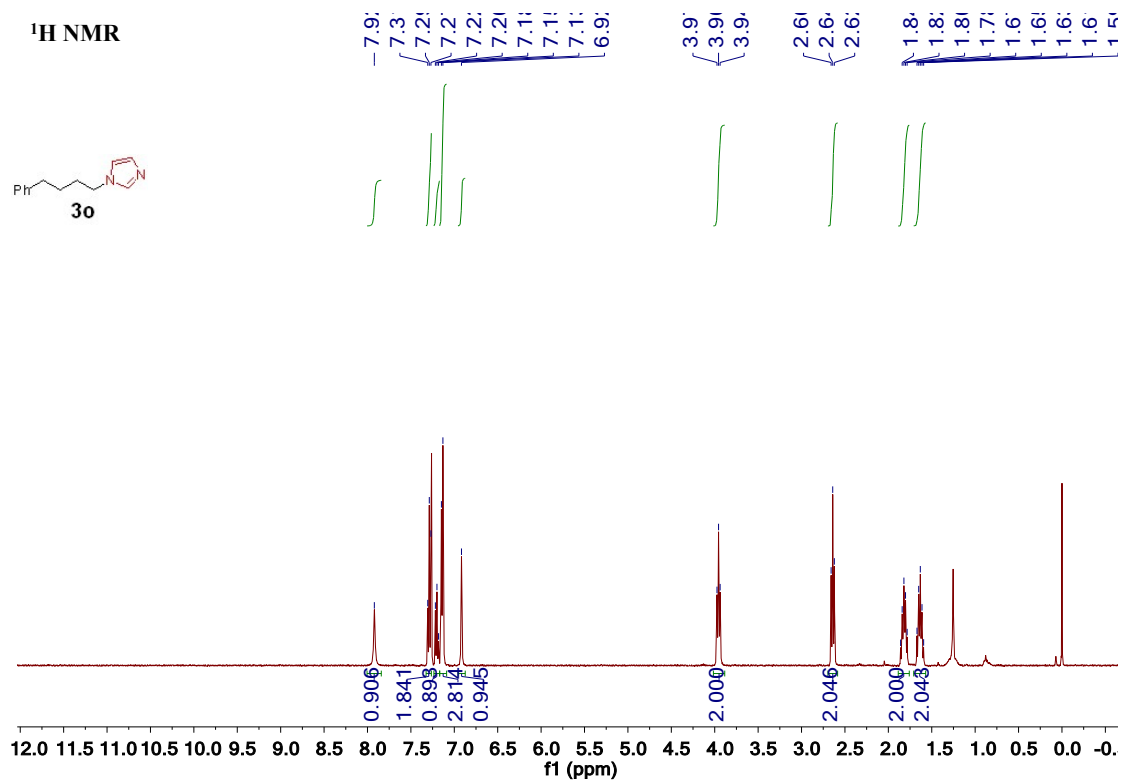


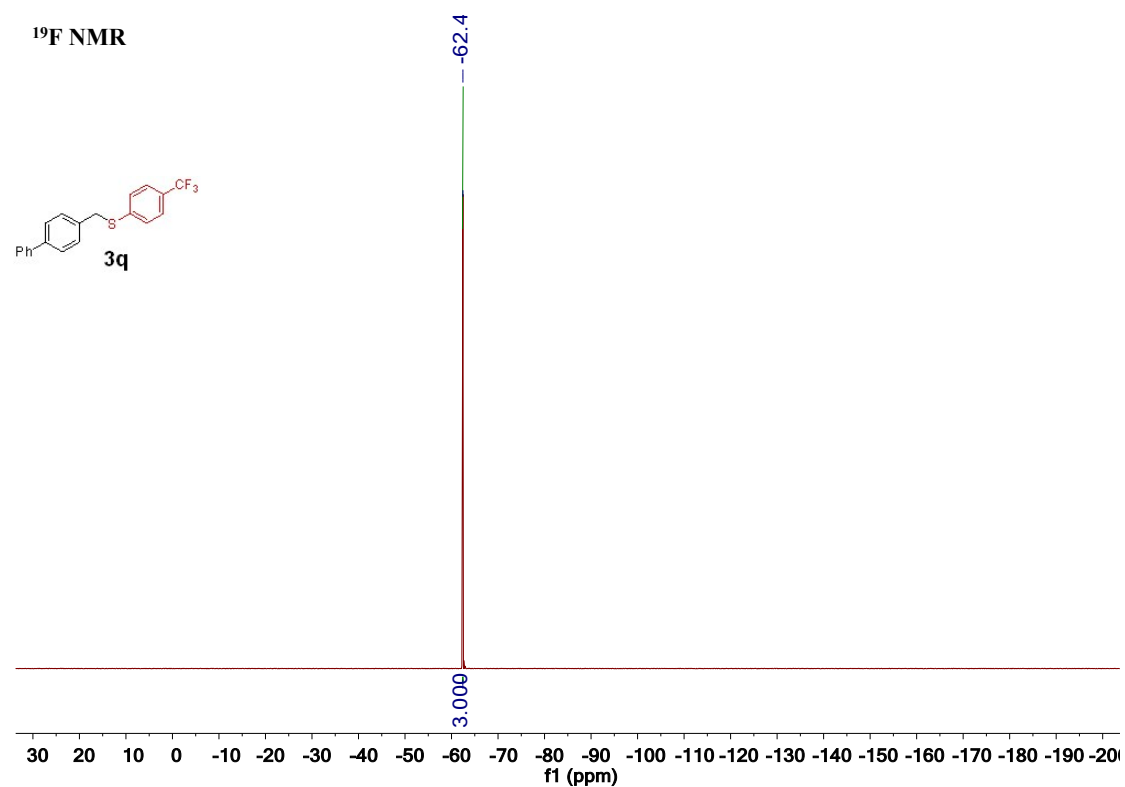
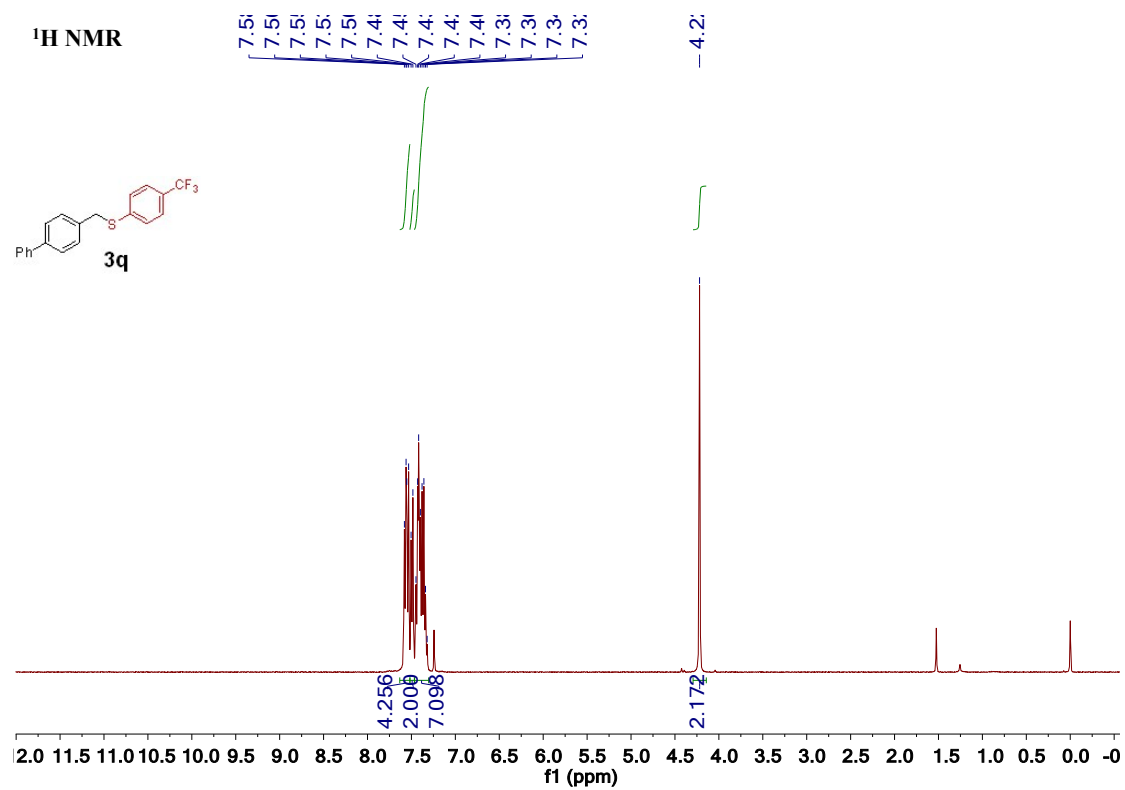


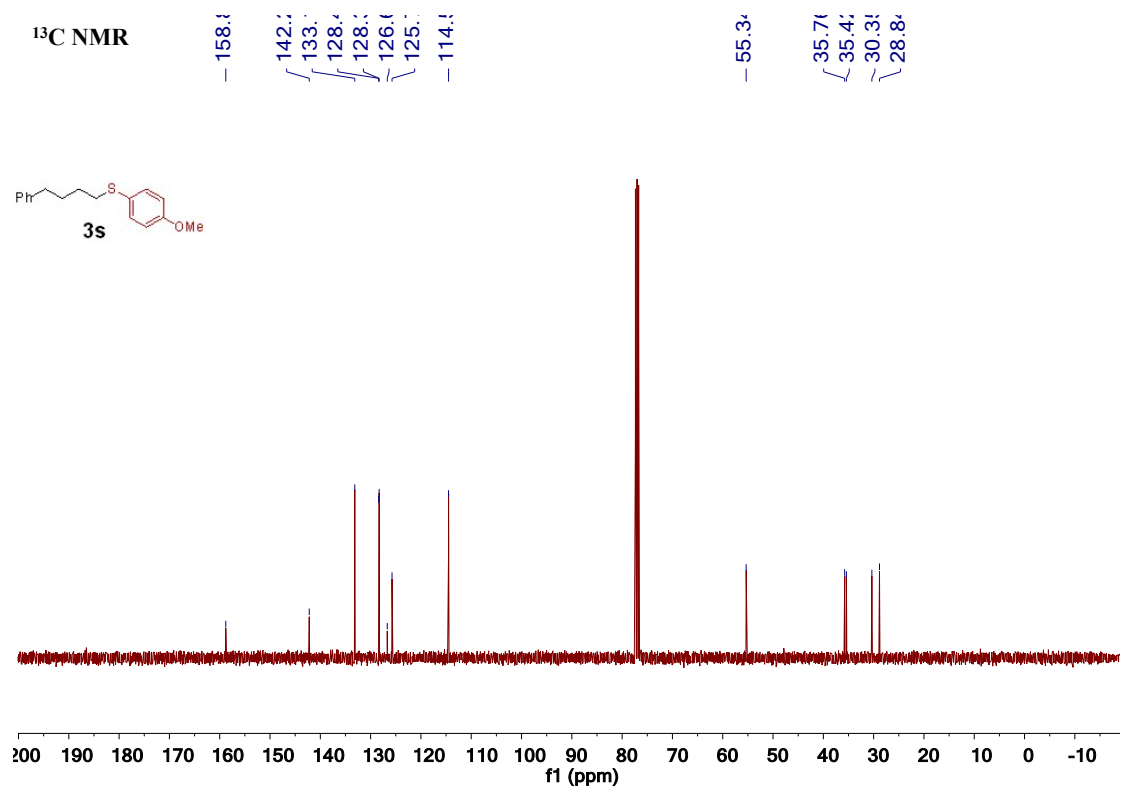
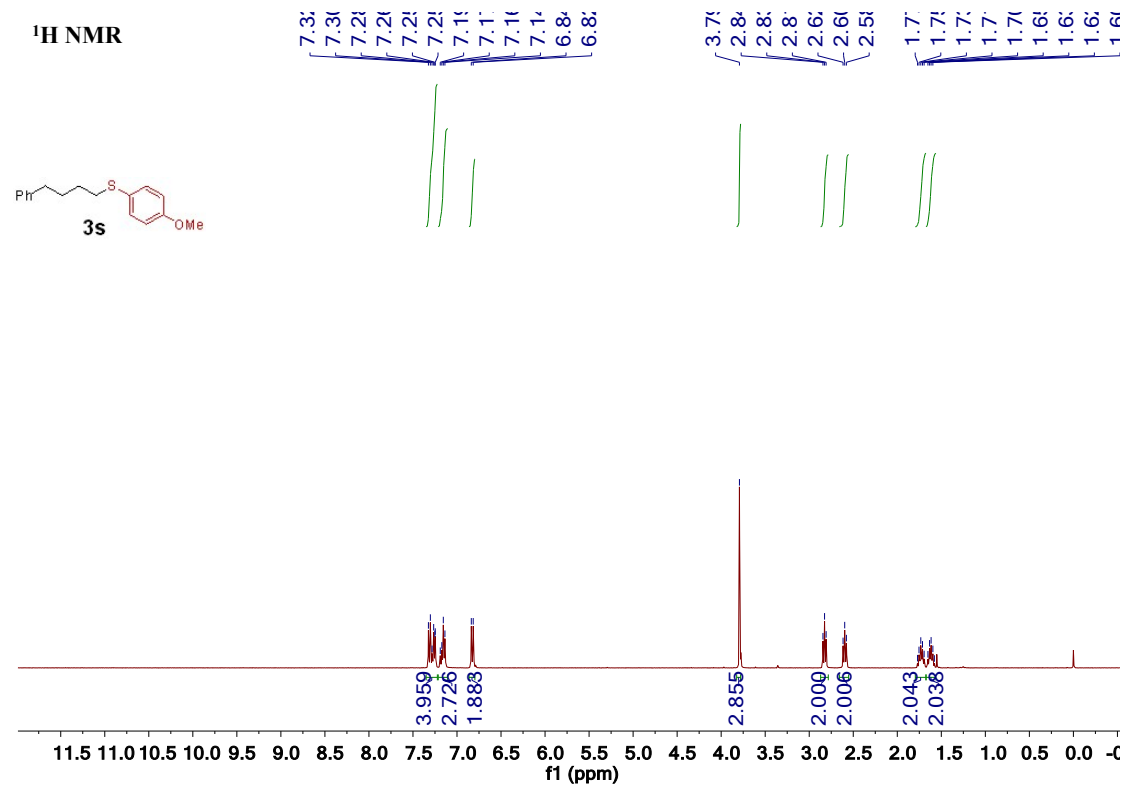


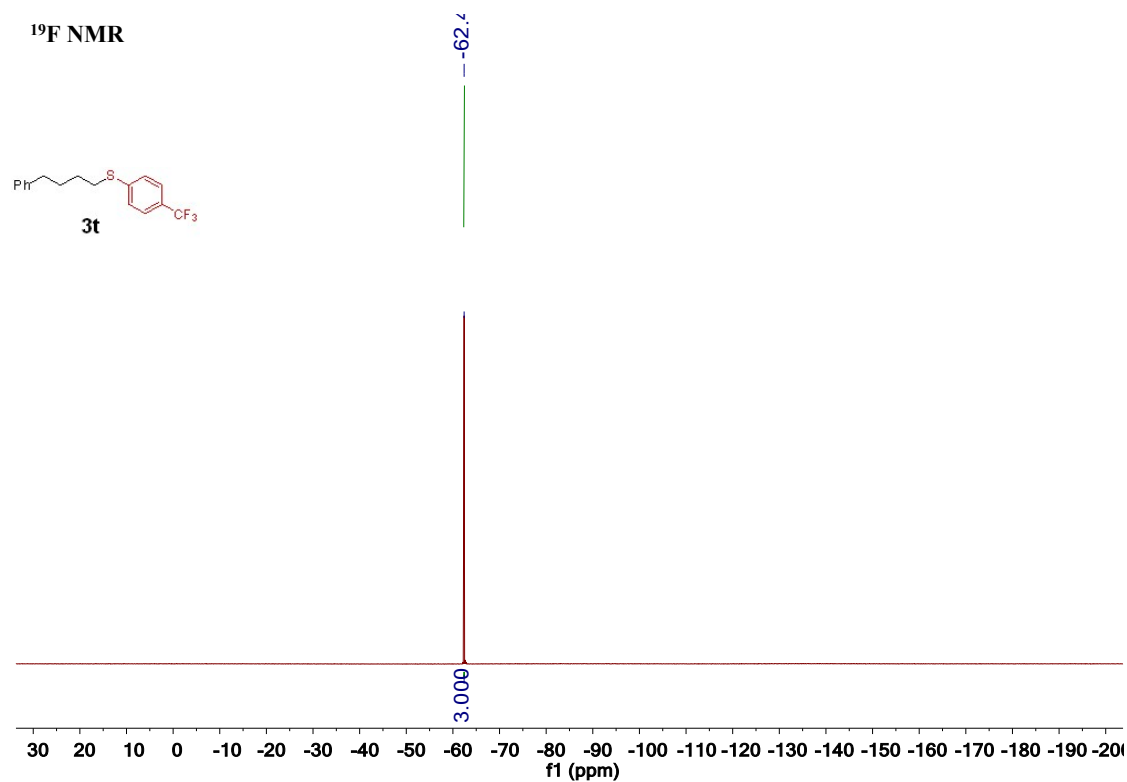
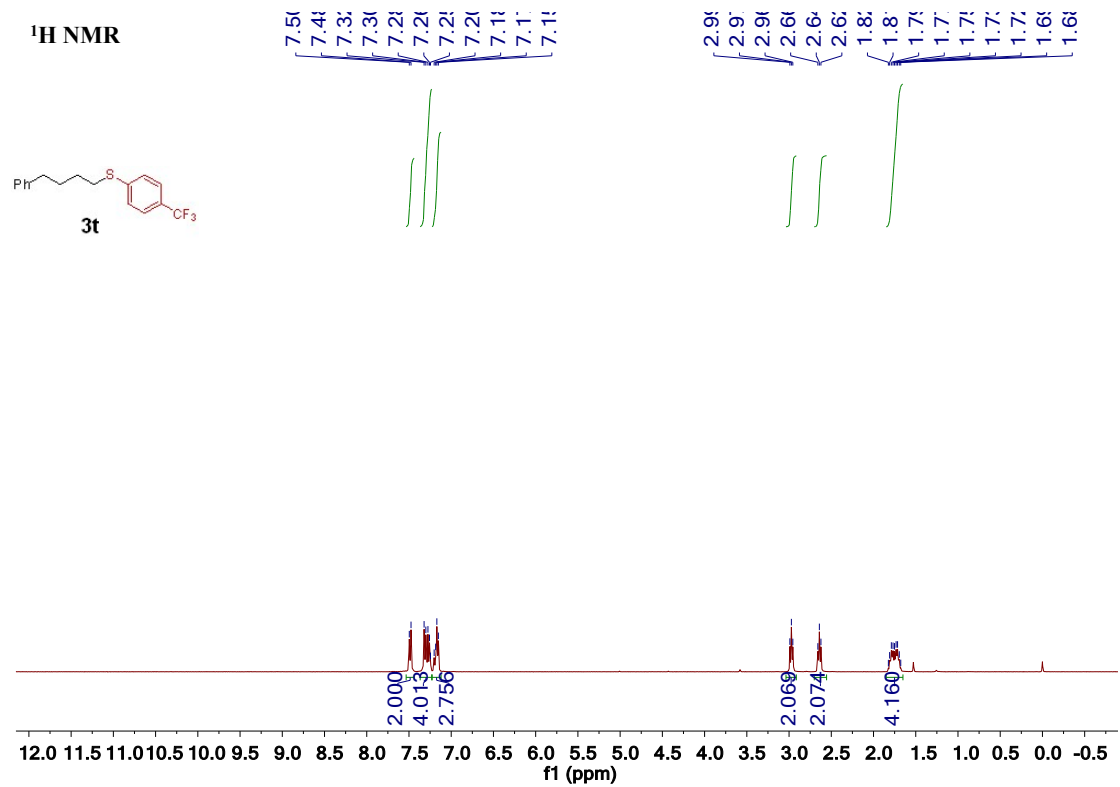


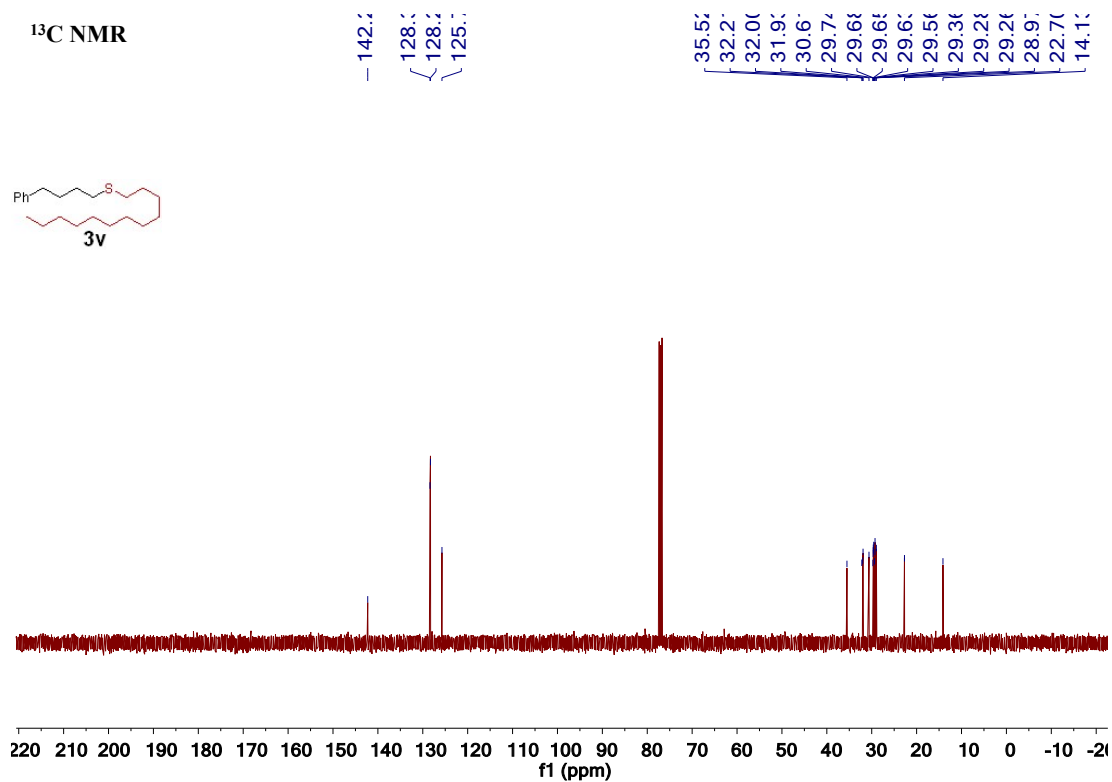
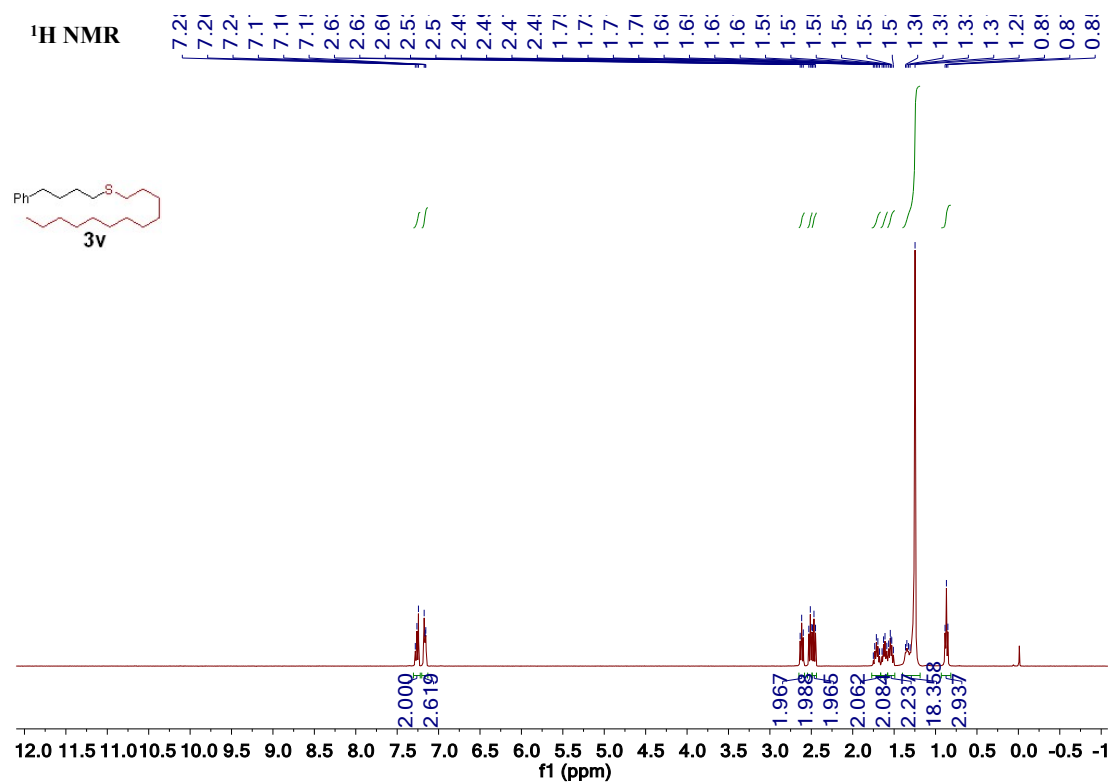


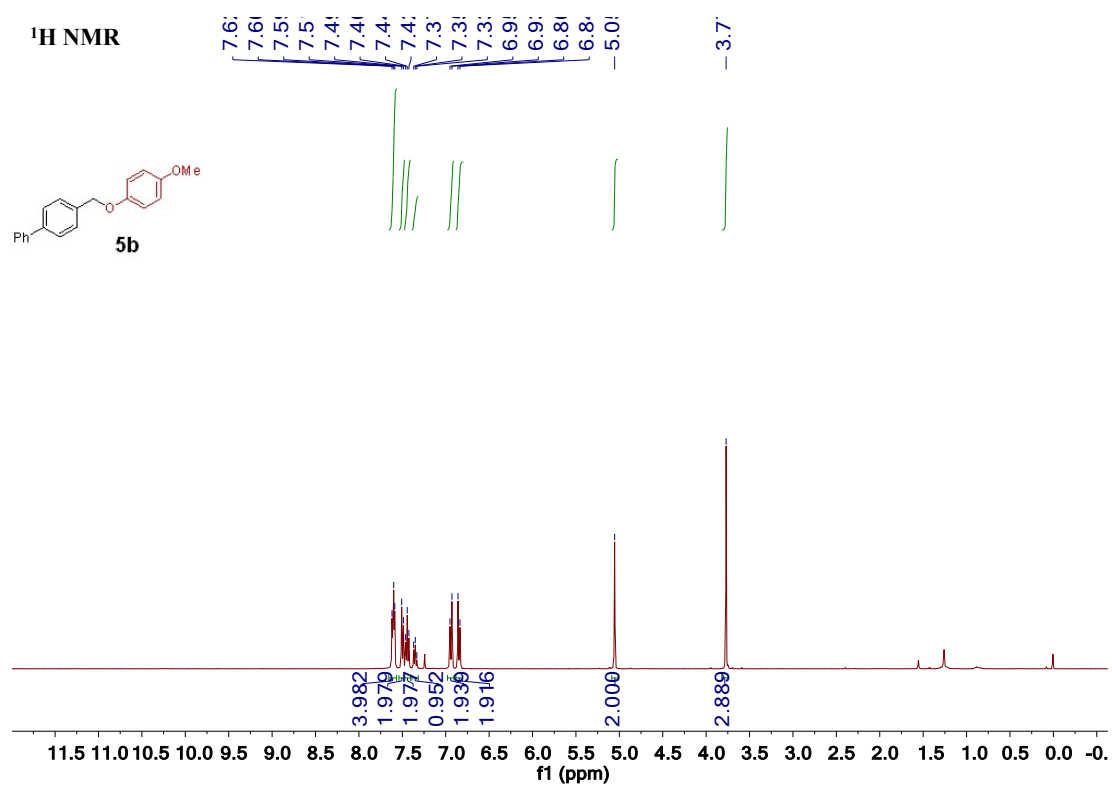
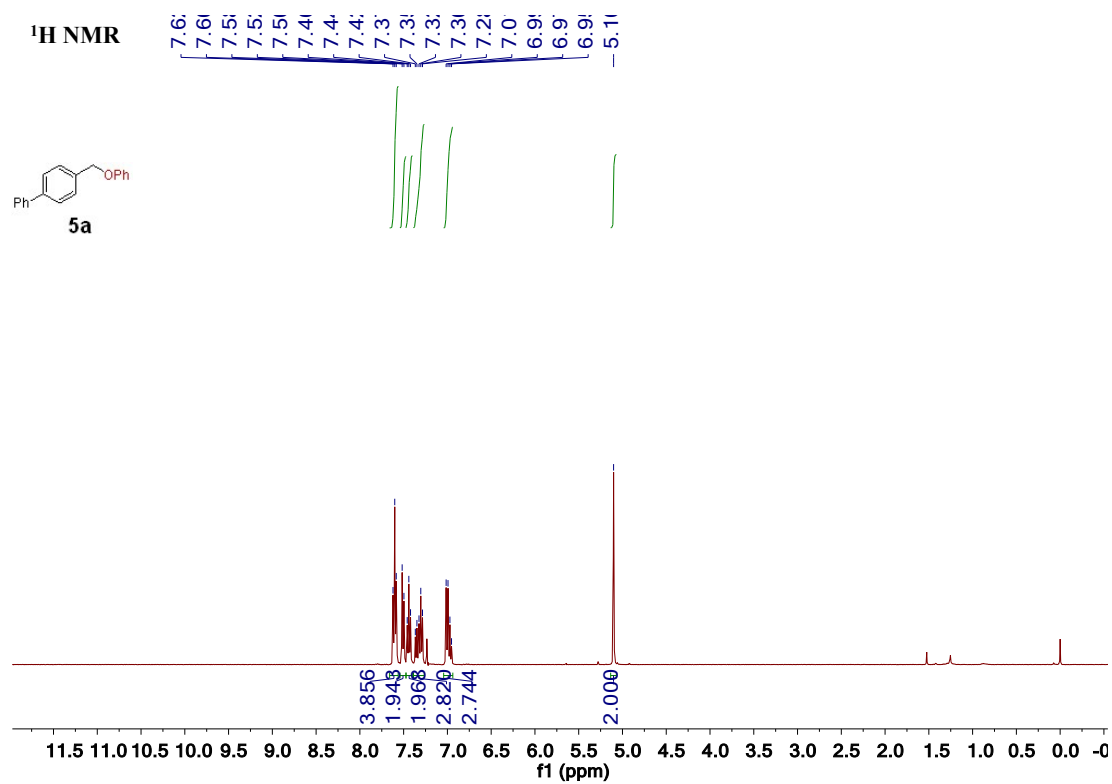


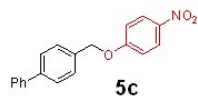
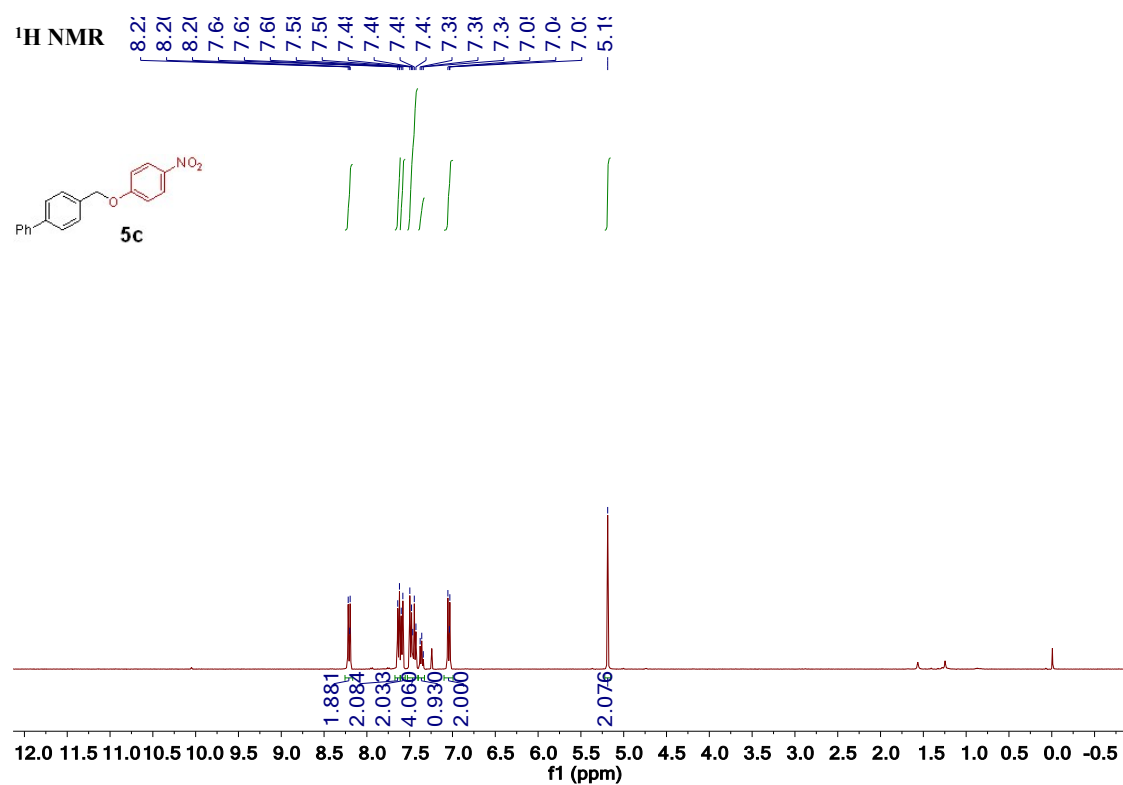
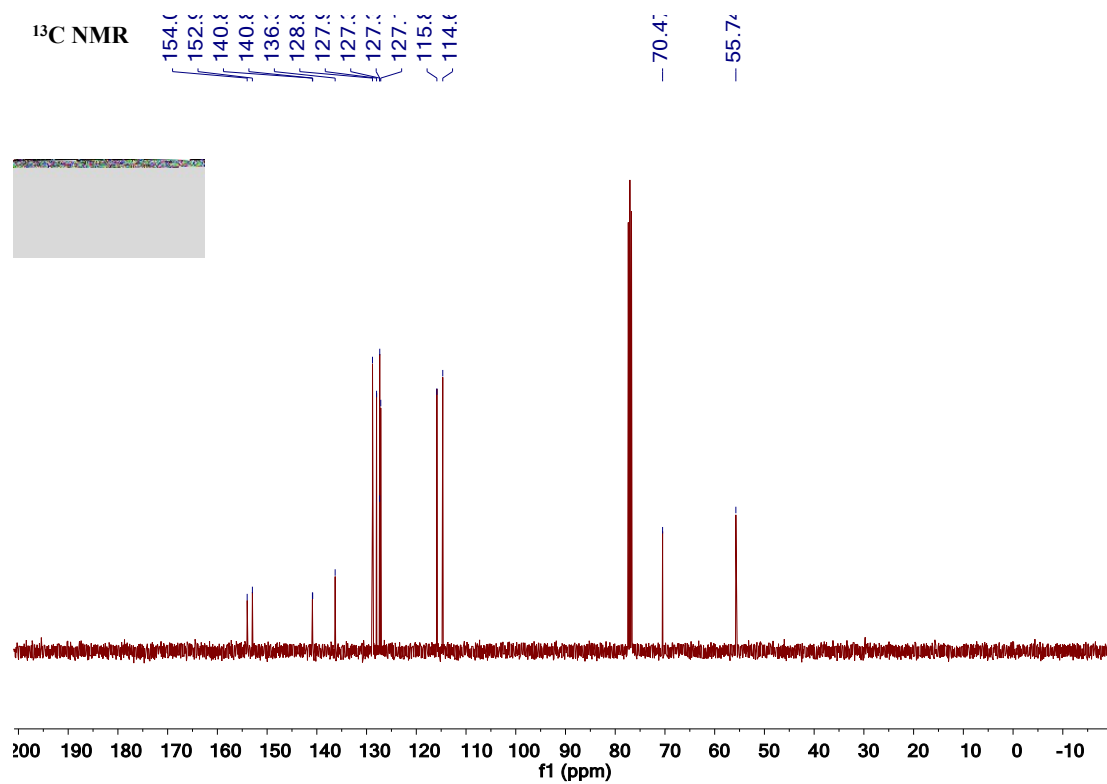


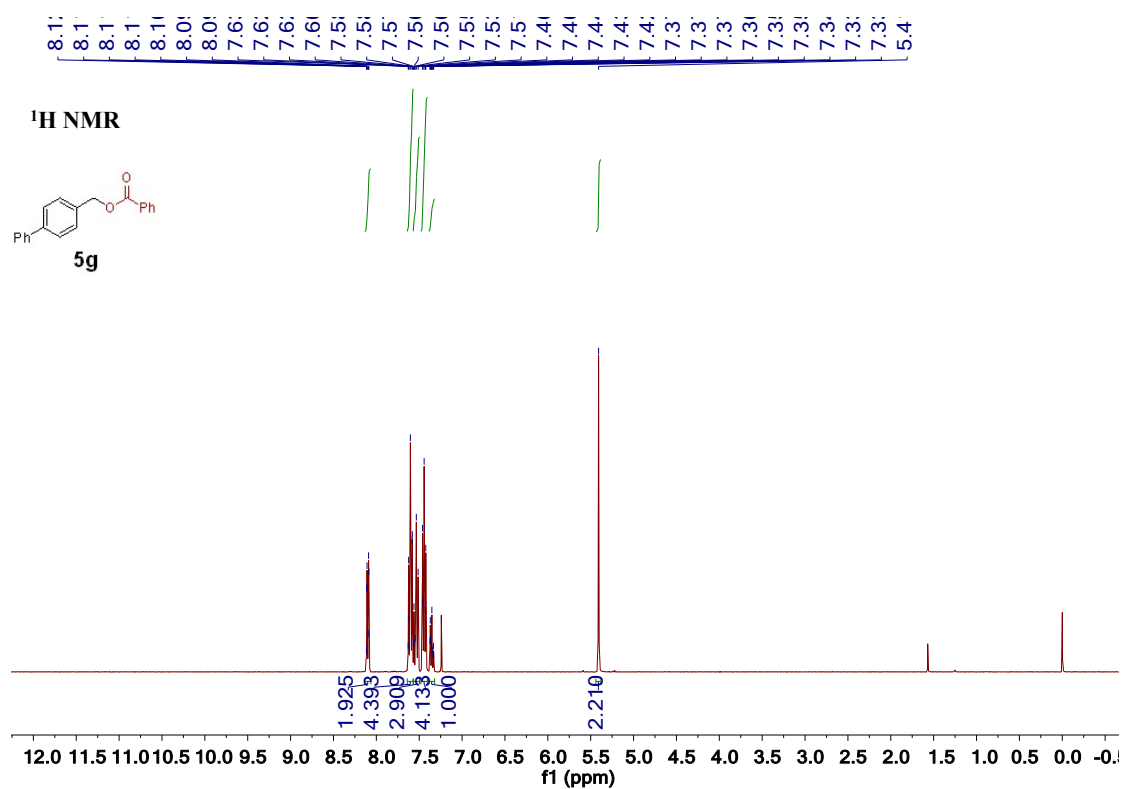
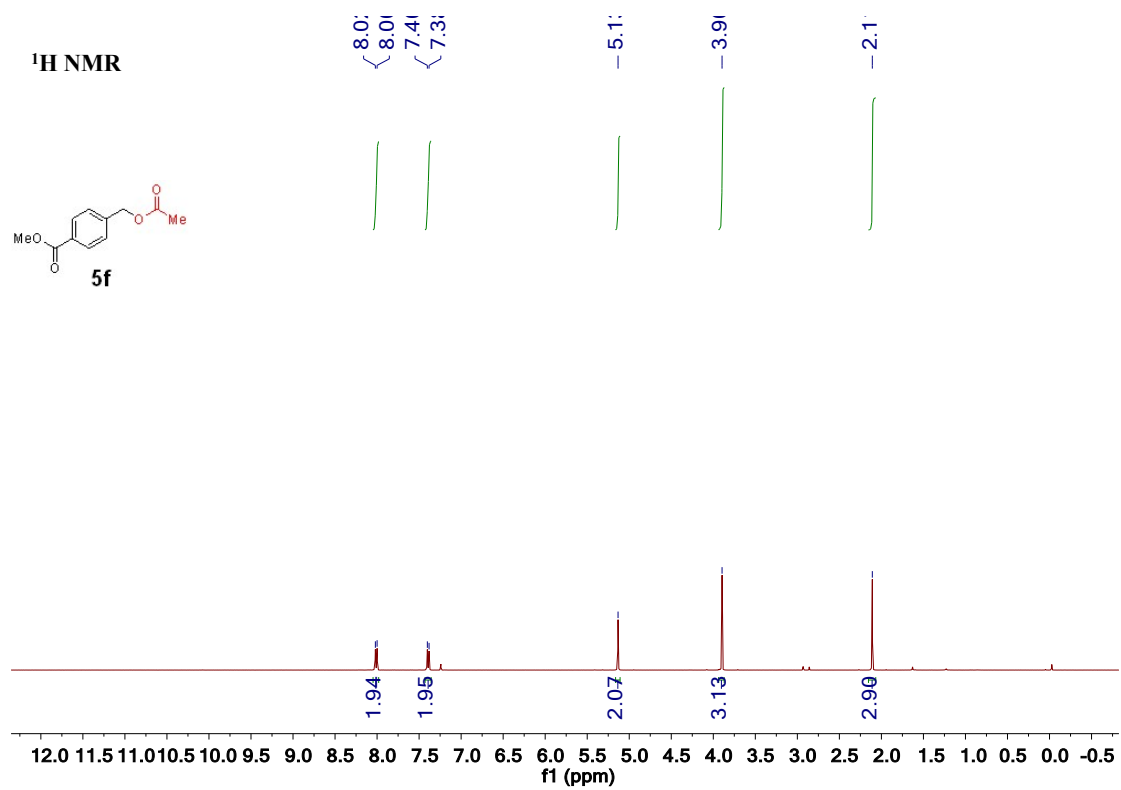


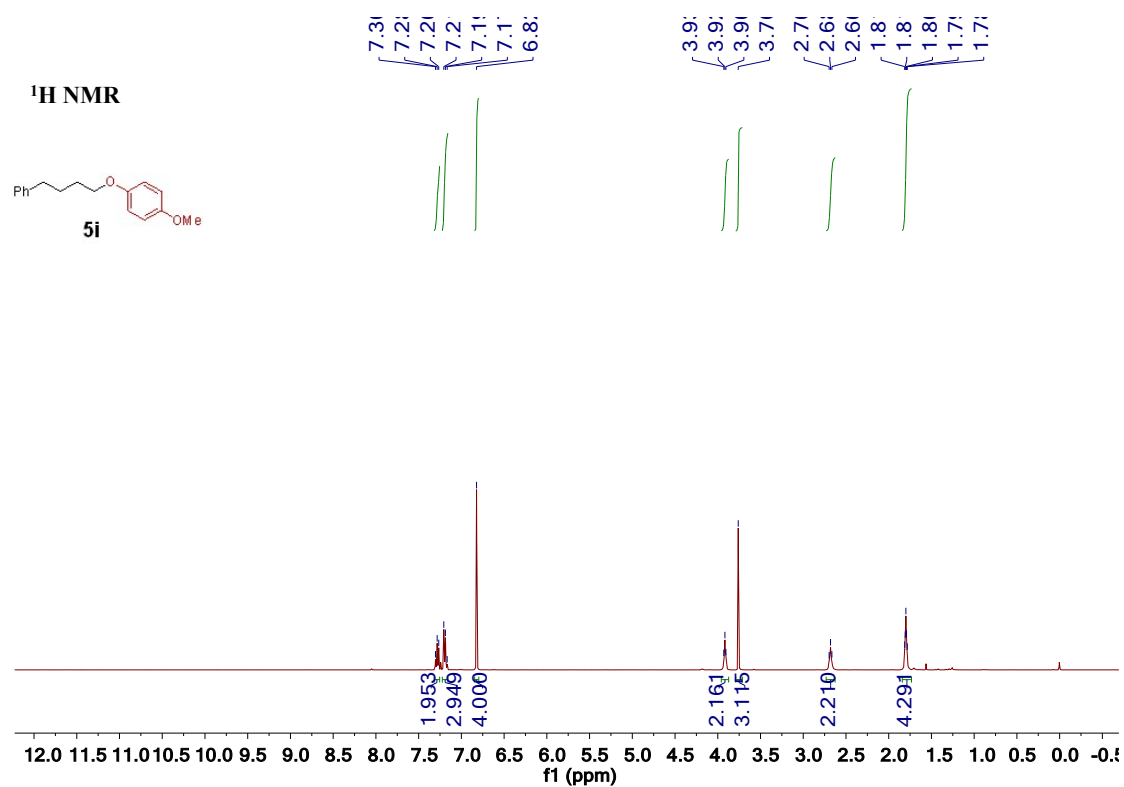
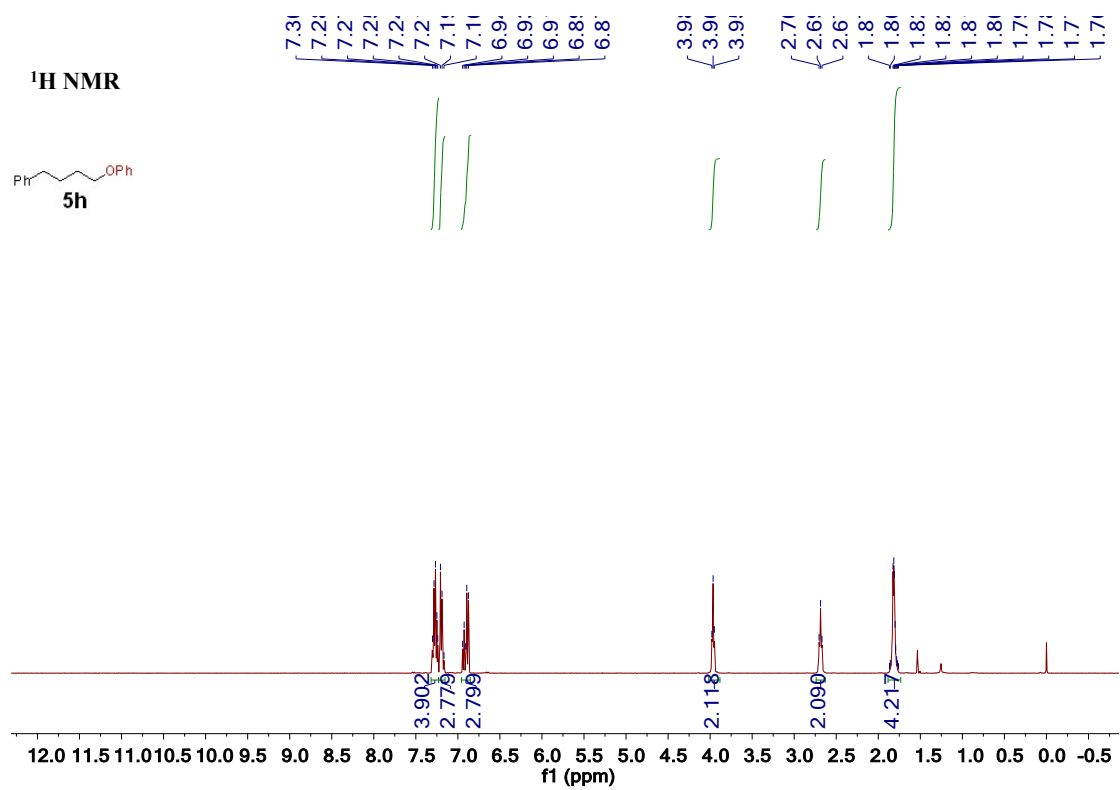


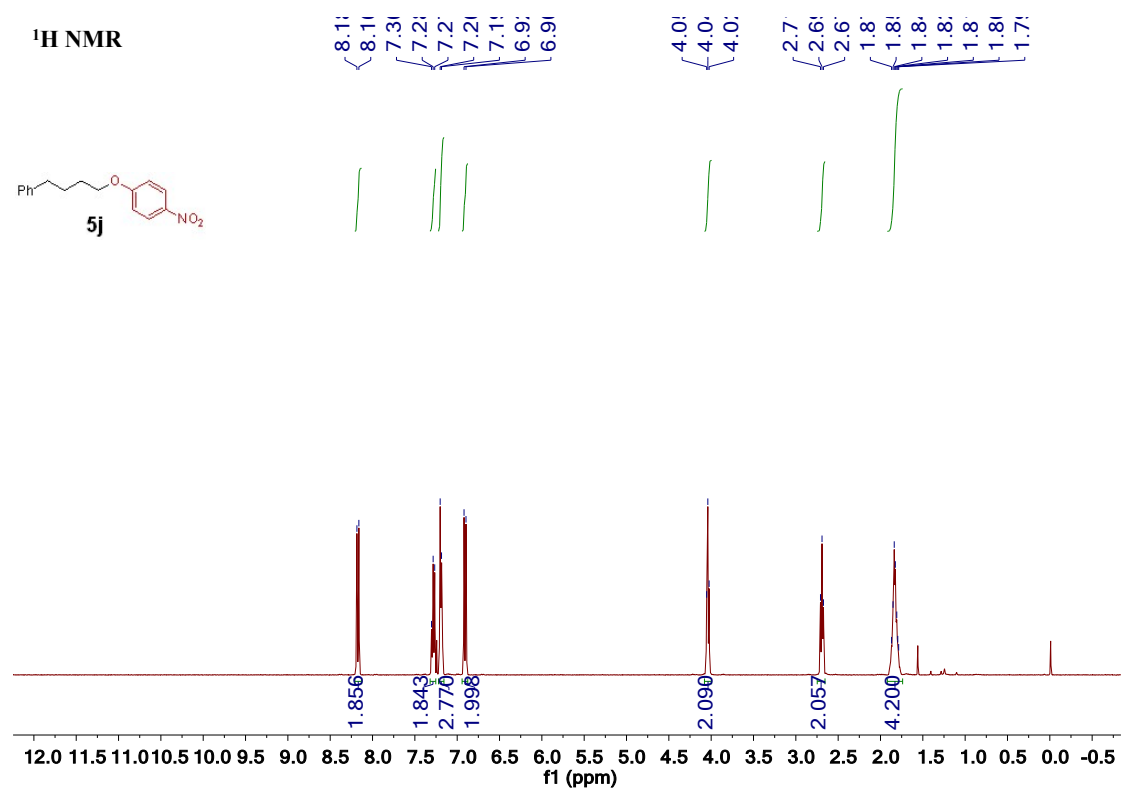
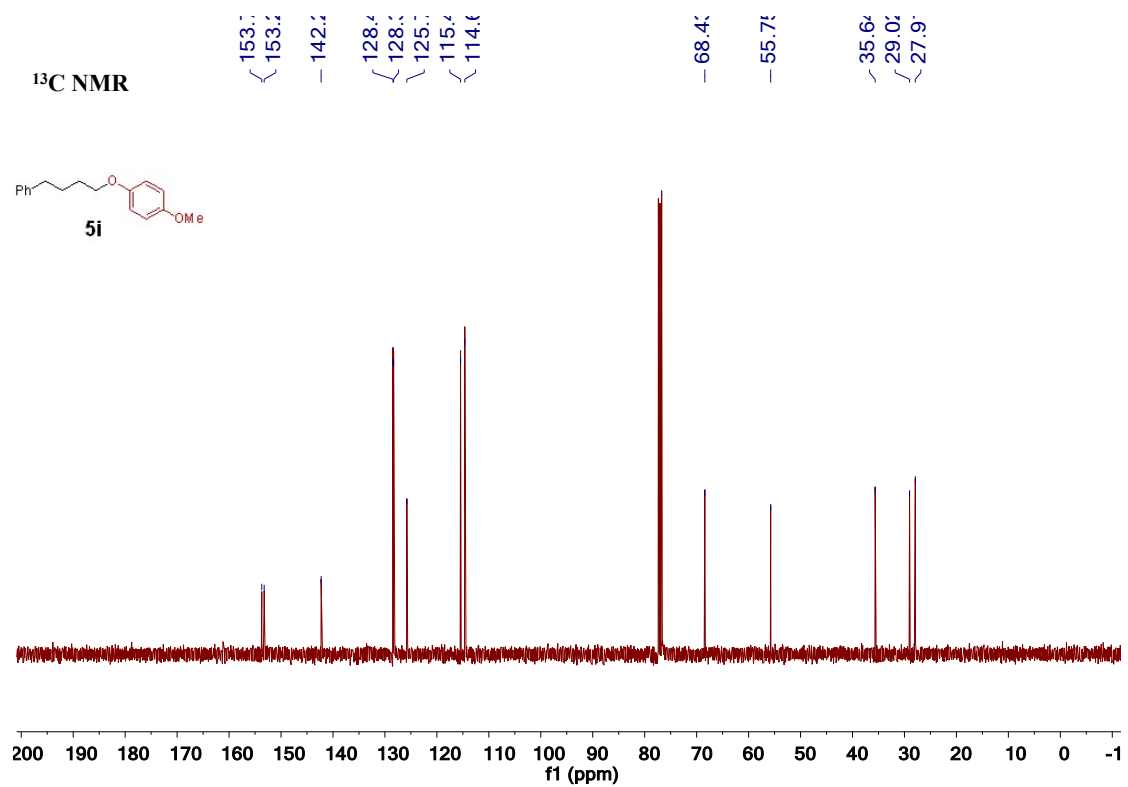


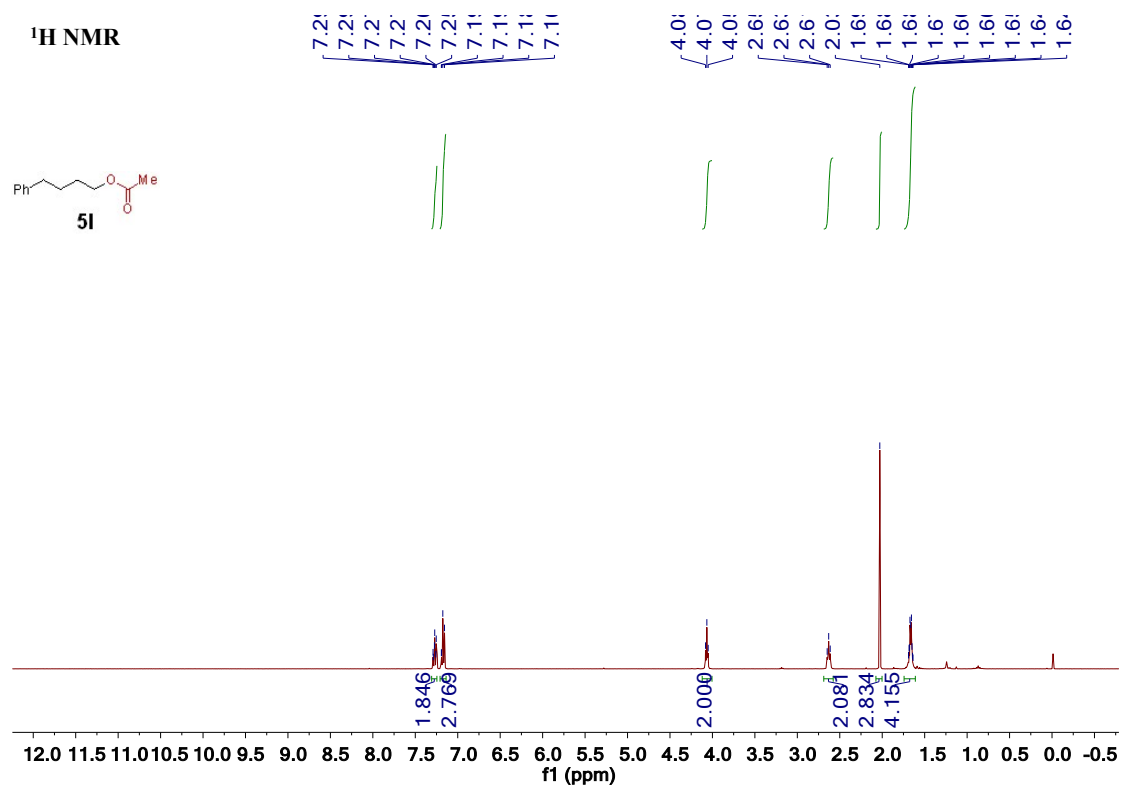
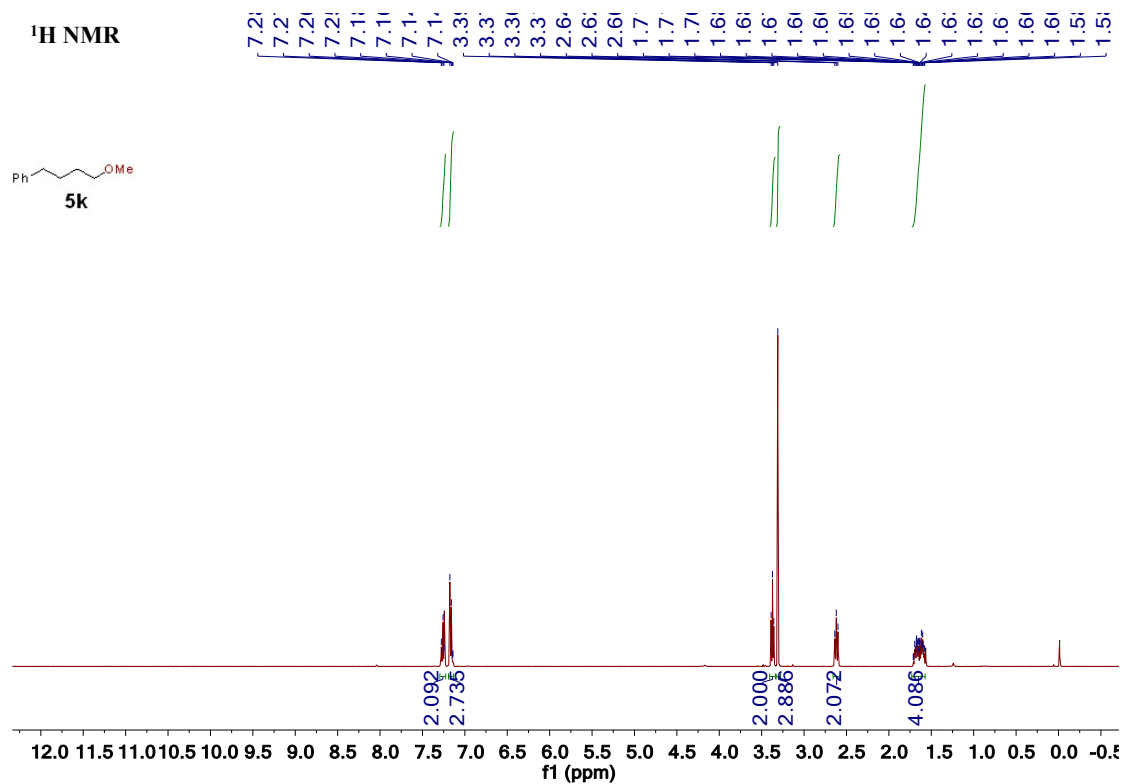


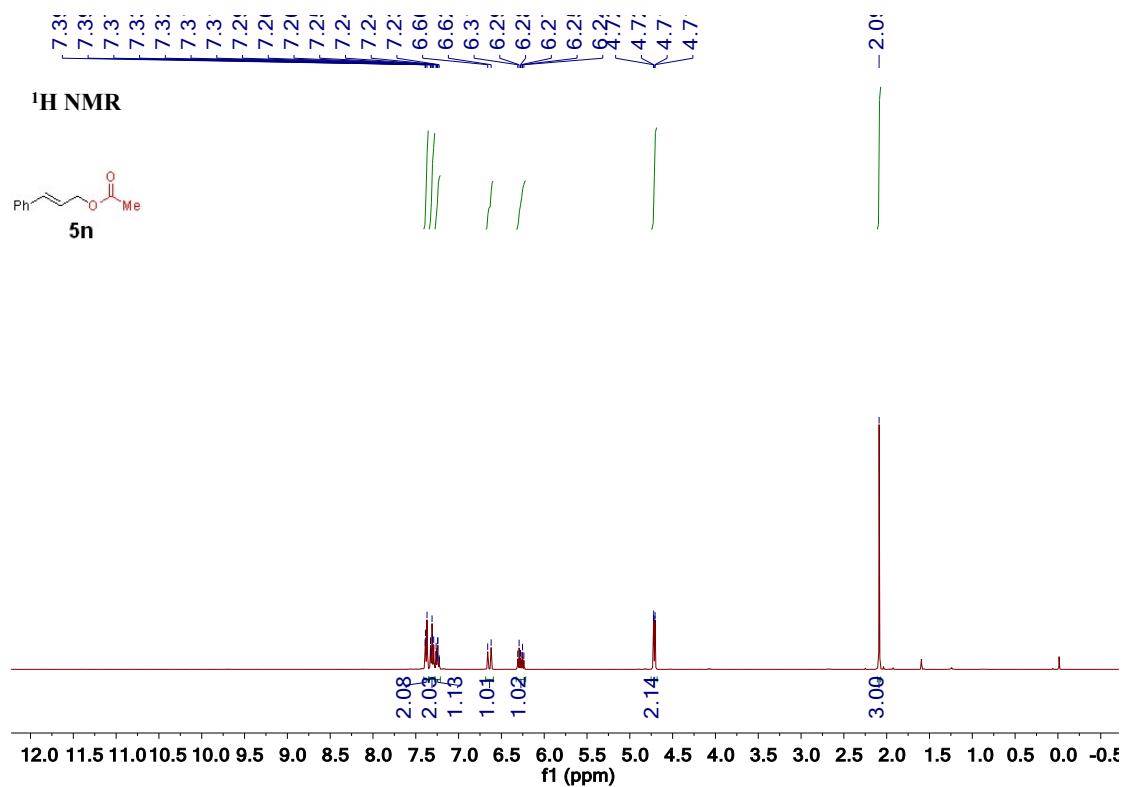
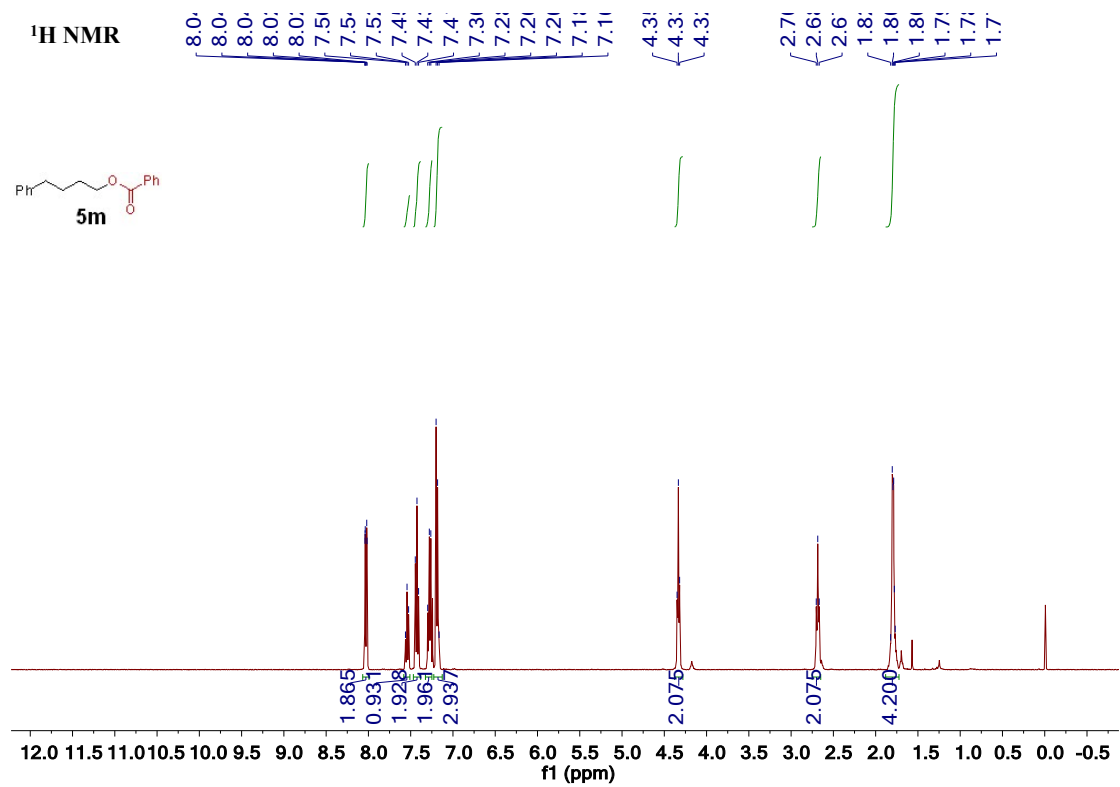


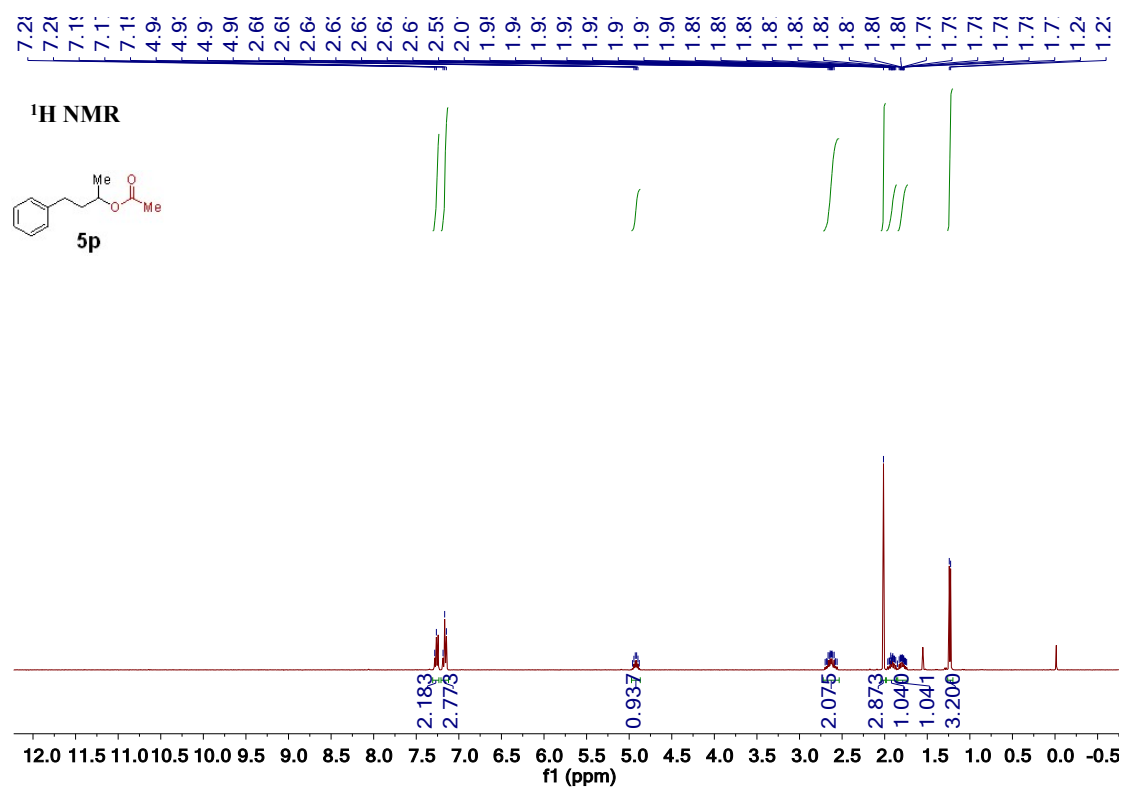
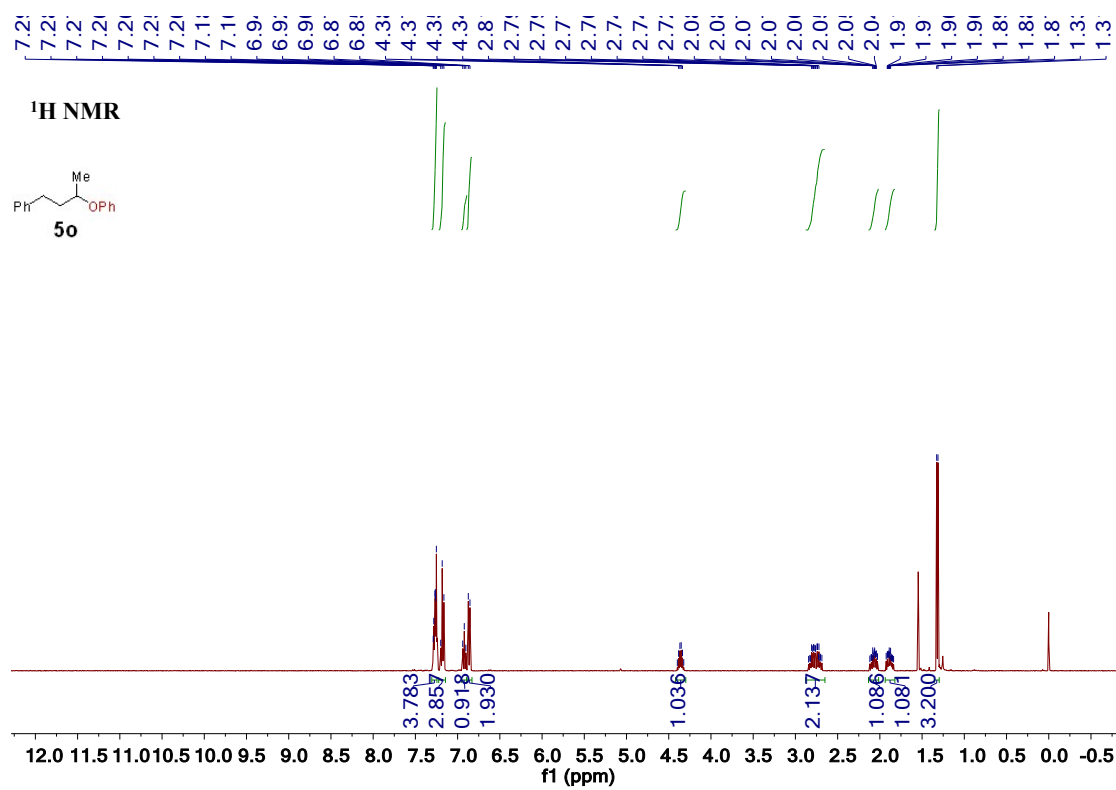


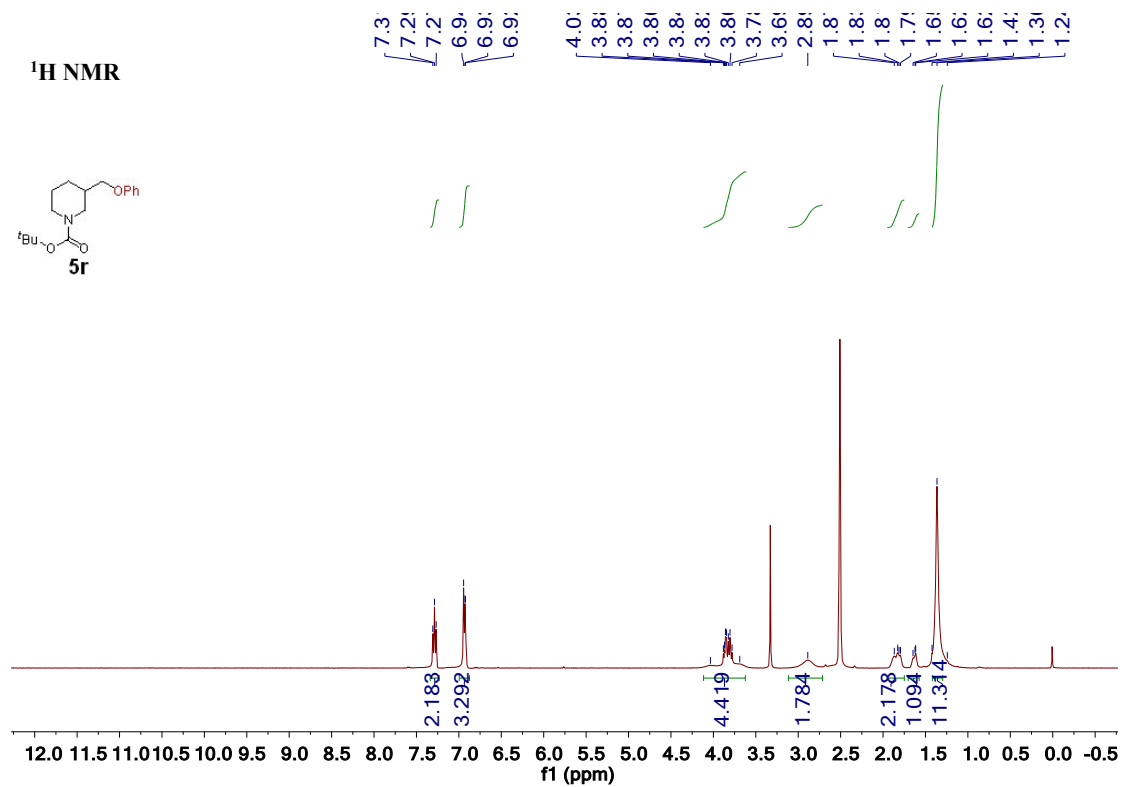
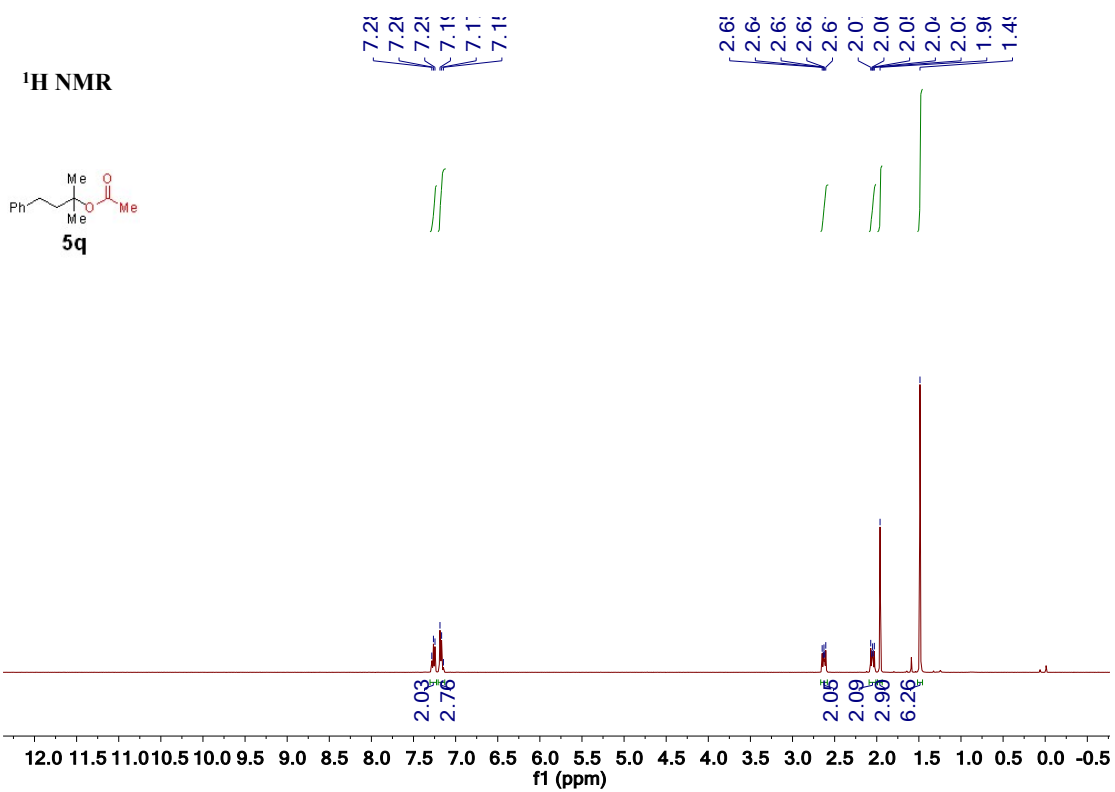




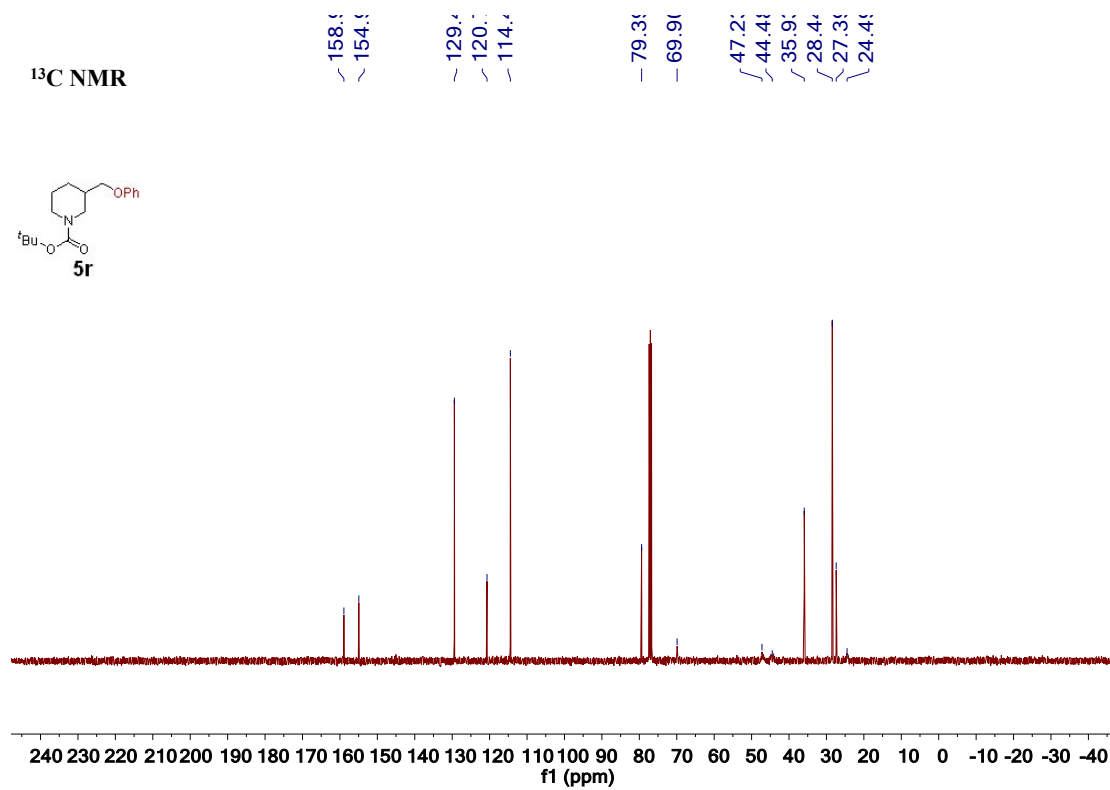
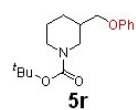




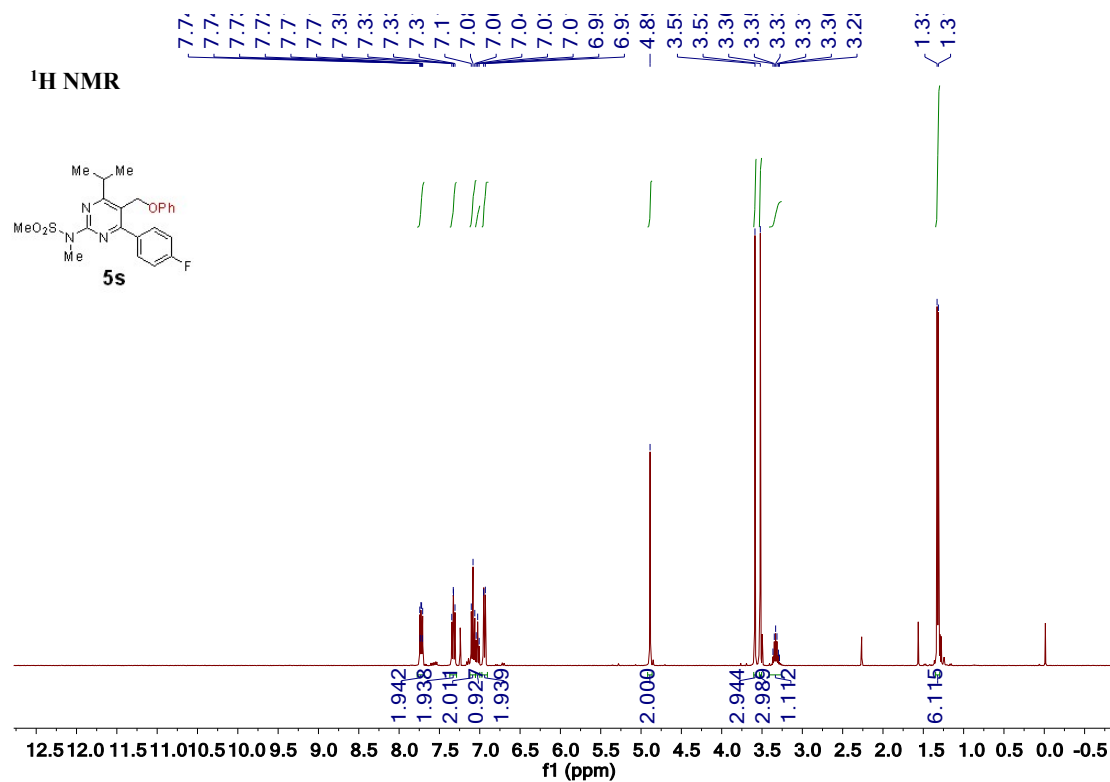
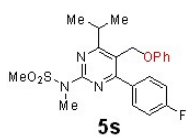




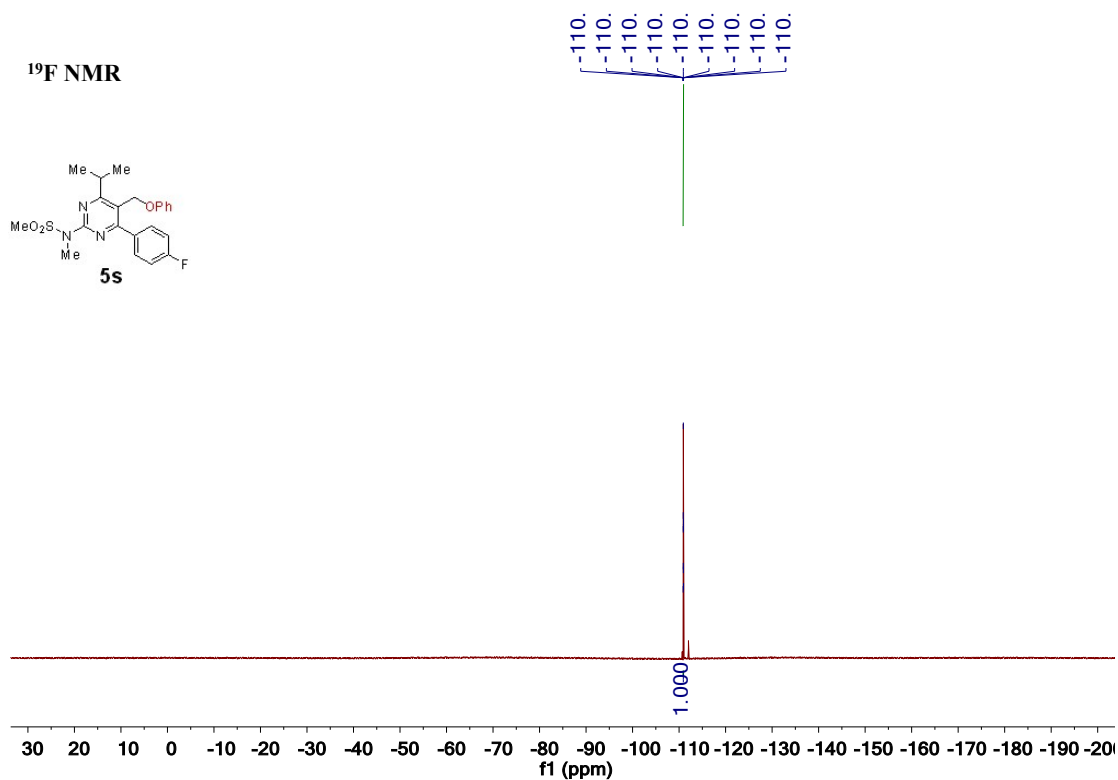
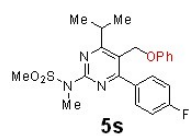
¹³C NMR



¹H NMR



^{19}F NMR



^{13}C NMR

