Electronic Supplementary Information

for

Dual Roles of Ethyl Bromodifluoroacetate in the Formation

of Fluorine-containing Heteroaromatic Compounds a

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1. General information

All chemicals were purchased from Adamas Reagent, energy chemical company (BrCF₂COOEt, BrCF₂COOH, ClCF₂COONa), J&K Scientific Ltd, Bide Pharmatech Ltd and Tansoole, Shuya company (BrCF₂PO(OEt)₂). Unless otherwise stated, all experiments were conducted in a sealed tube under N₂ atmosphere. Reactions were monitored by TLC or GC-MS analysis. Flash column chromatography was performed over silica gel (200-300 mesh).

¹H-NMR and ¹³C-NMR spectra were recorded in CDCl₃ on a Bruker Avance 500 spectrometer (500 MHz ¹H, 125 MHz ¹³C, 470 MHz ⁹F) at room temperature. Chemical shifts were reported in ppm on the scale relative to CDCl₃ (δ = 7.26 for ¹H-NMR, δ = 77.00 for ¹³C-NMR) as an internal reference. Coupling constants (*J*) were reported in Hertz (Hz).

2. Optimization of experiment conditions for 3a

Table S1. The reaction of without the additive compared with additive



Table S2. Optimization of the bases

Ar ² NH ₂ 1a	+ Frid COOEt Cu(OTf) ₂ (10 mol%) 1,10-phen (12 mol%) base (3 equiv) 120 °C, 20 h, CH ₃ CN Frid 3a	-COOEt
entry	base yie	eld (%)
1	K ₂ CO ₃	78%
2	Na ₂ CO ₃	86% (^b 82%)
3	NaHCO ₃	73%
4	K ₃ PO ₄	67%
5	Na ₃ PO ₄	80%
6	Cs ₂ CO ₃	57%
7	NaOAc	63%
8	Li ₂ CO ₃	49%
Poaction condition:	1a (0.2 mmol.) 2 (3 equiv) $Cu(OTf)$ (10 mel%) 1.10 phon (12 mel%)	base (3 equiv)

Reaction condition:, **1a** (0.2 mmol), **2** (3 equiv), Cu(OTf)₂ (10 mol%), 1,10-phen (12 mol%), base (3 equiv), GC yield. ^{*b*} isolated yield

Table S3. Optimization of the Cu salt for reaction

Ar ¹ NH ₂ 1a	Br [Cu] (10 mol%) 1,10-phen (12 mol%) 1,10-phen (12 mol%) Na ₂ CO ₃ (3 equiv) 120 °C, 20 h, CH ₃ CN 2 120 °C, 20 h, CH ₃ CN	COOEt 3a
entry	[Cu]	yield (%)
1	CuSO ₄	67%
2	Cu(OTf) ₂	86% (^b 82%)
3	Cu(OAc) ₂	68%
4	CuCl ₂	60%
5	Cu(NO ₃) ₂	73%
6	Cu(acac) ₂	70%
7	CuCN	47%
8	CuCl	58%
9	CuBr	54%
10	Cul	66%

Reaction condition:, **1a** (0.2 mmol), **2** (3 equiv), Cu salt (10 mol%), 1,10-phen (12 mol%), Na₂CO₃ (3 equiv), GC yield. ^{*b*} isolated yield

Table S4. Optimization of the solvent for reaction

Ar ² NH ₂ 1a	+ Free COOEt Cu(OTf) ₂ (10 1,10-phen (1 Na ₂ CO ₃ (3 120 °C, 20 h	equiv)
entry	solvent	yield (%)
1	dioxane	77%
2	CH ₃ CN	86% (^b 82%)
3	toluene	trace
4	THF	79%
5	acetone	68%
6	DCE	70%
7	DMF	33%
8	DMSO	trace
9	CH₃OH	61%
10	DMOE	42%

Reaction condition:, **1a** (0.2 mmol), **2** (3 equiv), Cu salt (10 mol%), 1,10-phen (12 mol%), Na₂CO₃ (3 equiv), GC yield. ^{*b*} isolated yield, DMOE=1,2-Dimethoxyethane

Table S5. Optimization of the ligand for reaction



entry	ligand	yield (%)
1	L1	69%
2	L2	61%
3	L3	66%
4	L4	68%
5	L5	77%
6	L6	74%
7	L7	71%
8	L8	78%
9	L9	86% (^b 82%)

Reaction condition:, **1a** (0.2 mmol), **2** (3 equiv), Cu(OTf)₂ (10 mol%), ligand (12 mol%), Na₂CO₃ (3 equiv), GC yield. ^{*b*} isolated yield,



Table S6. Screening of the different diflouromethyl compounds

3. General procedure for starting materials

(1). General experimental procedures for substrates 1



GP-I: In a dry 50 mL round bottom flask, phenylboronic acid (1.3 eq.), K_2CO_3 (4.0 eq.) and Pd(PPh₃)₄ (0.1 eq.) were dissolved in a mixture of toluene / water / ethanol (3:2:1, 0.1 M). 2-Bromoaniline (1.0 eq.) was added and the resulting mixture was heated to 95 °C for 20 hours. After cooling to room temperature, the biphasic solution was diluted with saturated aqueous NH₄Cl and CH₂Cl₂ and the phases were separated. The aqueous phase was extracted twice with CH₂Cl₂ (50 mL) and the combined organic phases were washed with water (1 x 50 mL) and saturated aqueous NaHCO₃ (1 x 50 mL). The organic phases were dried over Na₂SO₄ and filtered. The filtrate was concentrated in vacuo to afford the crude product. Purification by column chromatography on silica gel (petroleum ether / ethyl acetate) afforded the corresponding products.¹



GP-II 2-Iodoaniline (10 mmol, 1.0 equiv), p-tolylboronic acid (12 mmol, 1.2 equiv) were added to a dry Schlenk flask. The flask was evacuated and backfilled with pure N_2 for 3 times. DME (10 mL) and aqueous solution of K_2CO_3 (2 M, 20 mL) were added with syringe and the mixture was stirred for 30 min at room temperature under N_2 atmosphere. To the stirred mixture, $PdCl_2(PPh_3)_2$ (0.2 mmol, 140 mg, 0.02 equiv) in DME (10 mL) was added with syringe at room temperature and the mixture was stirred at 80 °C for 12 h under N_2 atmosphere (monitored by TLC). After the reaction was complete, the mixture was then cooled to room temperature and diluted with EtOAc (20 mL). The aqueous layer was extracted with EtOAc for 3 times (20 mL × 3). Then the organic phase was combinated and dried over anhydrous MgSO₄. The solvent was removed under reduced pressure and the residue was purified by column chromatography on silica gel by using a 30:1 mixture of petroleum ether/EtOAc as an eluent to provide amine .²

(2). General experimental procedures for substrates 5



To the stirred solution of KOH (6 g) in 24 mL of water, benzothioazole (3 mmol) was added and refluxed for 17 h. After cooling to room temperature, MeI (3 mmol) was added drop wise and stirring was continued for an additional 1 h. The resultant reaction mixture extracted with diethyl ether (3 x 25 mL) combined organic layers dried over Na_2SO_4 , filtered and concentrated in vacuum. Purification of the crude product was achieved by flash column chromatography using petrol ether/ethyl acetate (15:1) as eluent.

4. General process for the synthesis of B



In a dried Schlenk tube were placed **1** (0.2 mol, 1 equiv), Na₂CO₃ (0.6 mol, 3 equiv), Cu(OTf)₂ (0.02 mmol 0.1 equiv), 1,10-phen (0.024 mmol 0.12 equiv). **2** (0.6 mmol, 3 equiv) and solvent is added the mixture under N₂ atmosphere. The resulting mixture was stirred at 120 °C for 20 h. Upon completion of the reaction, the solvent was evaporated under reduced pressure and the residue was purified by flash column chromatograph (silica gel, petroleum ether:EtOAc = 100:1, v/v) to give the desired product **3**.

5. Crystal data of 3a

Crystallographic data for compound **3a** (CCDC-1820711) has been deposited with the Cambridge Crystallographic Data Centre, Copies of the data can be obtained, free of charge, on application to CCDC (Email:deposit@ccdc.cam.ac.uk).



Bond precision:	C-C = 0.0031 A	Wavelength=0.71073		
Cell: Temperature:	alpha=90	b=18.0049(15) beta=90		
lemperature:	296 K			
	Calculated	Reporte	d	
Volume	2841.2(4)	2841.2(4)	
Space group	Pbca	Pbca		
Hall group		-P 2ac	2ab	
Moiety formula	C17 H13 F2 N O2	C17 H13	F2 N 02	
Sum formula	C17 H13 F2 N O2	C17 H13	F2 N 02	
Mr	301.28	301.29		
Dx,g cm-3	1.409	1.409		
Z	8	8		
Mu (mm-1)	0.110	0.110		
F000	1248.0	1248.8		
F000'	1248.76			
h,k,lmax	9,21,22	9,21,22		
Nref	2494	2487		
Tmin, Tmax		0.419,1	.000	
Tmin'				
Correction method= # Reported T Limits: Tmin=0.419 Tmax=1.000 AbsCorr = MULTI-SCAN				
Data completeness= 0.997 Theta(max) = 24.990			990	
R(reflections) = 0.0445(1797) wR2(reflections) = 0.1237(2487				
S = 1.061 Npar= 199				

SMe NH ₂ 4a	+ F ^w F	Br COOEt 2	Cu(OTf) ₂ (10 mc 1,10-phen (12 m Na ₂ CO ₃ (4 eq B ₂ pin ₂ (30 mol CH ₃ CN, 80 °	uiv) %)	N F F	COOEt
entry	[Cu]	ligand	additive	base	solvent	yield (%)
1	Cu(OTf) ₂	1,10-phen		Na ₂ CO ₃	CH ₃ CN	74
2	Cu(OTf) ₂	1,10-phen	$B_2 pin_2$	Na ₂ CO ₃	CH ₃ CN	83 (78) ^b
3	Cu(OTf) ₂	1,10-phen	$B_2 pin_2$	Na ₂ CO ₃	THF	67
4	CuSO ₄	1,10-phen	$B_2 pin_2$	Na ₂ CO ₃	CH ₃ CN	47
5	Cu(OTf) ₂	Xantphos	$B_2 pin_2$	Na ₂ CO ₃	CH ₃ CN	60
6	Cu(OTf) ₂	1,10-phen	$B_2 pin_2$	K ₂ CO ₃	CH ₃ CN	69
7	Cu(OTf) ₂	1,10-phen	$B_2 pin_2$	DBU	CH₃CN	23
8 ^c	Cu(OTf) ₂	1,10-phen	$B_2 pin_2$	Na ₂ CO ₃	CH ₃ CN	54

6. Optimization of experiment conditions for 5

^a Reaction conditions: **4a** (0.2 mmol), ethyl bromodifluoroacetate (**2**) (3 equiv), [Cu] (10 mol%), ligand (12 mol%), B₂pin₂ (30 mol%), base (4 equiv), CH₃CN (2 mL) under N₂ atmosphere at 80 °C for 24 h. GC yields. ^b Isolated yields. ^c 12 h

7. General process for the synthesis of 5



In a dried Schlenk tube were placed **4** (0.2 mol, 1 equiv), Na₂CO₃ (0.8 mol, 4 equiv), Cu(OTf)₂ (0.02 mmol 0.1 equiv), 1,10-phen (0.024 mmol 0.12 equiv). **2** (0.6 mmol, 3 equiv), B₂pin₂ (0.06 mmol 0.3 equiv) and solvent CH₃CN is added the mixture under N₂ atmosphere. The resulting mixture was stirred at 80 °C for 24 h. Upon completion of the reaction, the solvent was evaporated under reduced pressure and the residue was purified by flash column chromatograph (silica gel, petroleum ether:EtOAc = 100:1, v/v) to give the desired product **5**.

8. Control experiments to figure out the key intermediate for this transformation



9. Control experiments to figure out the formation of isocyanides



from amines



10. Control experiments and radical trapping experiments.

11. Computation

Computational Methods

All calculations were carried out with the Gaussian 09 program. Geometries were optimized using the B3LYP density functional with the LANL2DZ basis set for Br and the 6-31G basis set for other atoms. Harmonic frequency analysis was conducted at the same level of theory to verify the stationary points to be real minima or saddle points and to obtain the thermal corrections at 298.15 K. Intrinsic reaction coordinate (IRC) calculations were carried out to ensure that the transition states connect the correct reactants and products. Single-point energies were calculated at optimized gas-phase geometries at the M06-2X/6-311G(d,p) level and solvent effects were introduced with the SMD approach. In the single-point calculations, thermal corrections were added to obtain Gibbs free energies (kcal/mol).

Gaussian 09, Revision D.01, Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Scalmani, G.; Barone, V.; Mennucci, B.; Petersson, G. A.; Nakatsuji, H.; Caricato, M.; Li, X.; Hratchian, H. P.; Izmaylov, A. F.; Bloino, J.; Zheng, G.; Sonnenberg, J. L.; Hada, M.; Ehara, M.; Toyota, K.; Fukuda, R.; Hasegawa, J.; Ishida, M.; Nakajima, T.; Honda, Y.; Kitao, O.; Nakai, H.; Vreven, T.; Montgomery, J. A., Jr.; Peralta, J. E.; Ogliaro, F.; Bearpark, M.; Heyd, J. J.; Brothers, E.; Kudin, K. N.; Staroverov, V. N.; Kobayashi, R.; Normand, J.; Raghavachari, K.; Rendell, A.; Burant, J. C.; Iyengar, S. S.; Tomasi, J.; Cossi, M.; Rega, N.; Millam, N. J.; Klene, M.; Knox, J. E.; Cross, J. B.; Bakken, V.; Adamo, C.; Jaramillo, J.; Gomperts, R.; Stratmann, R. E.; Yazyev, O.; Austin, A. J.; Cammi, R.; Pomelli, C.; Ochterski, J. W.; Martin, R. L.; Morokuma, K.; Zakrzewski, V. G.; Voth,

G. A.; Salvador, P.; Dannenberg, J. J.; Dapprich, S.; Daniels, A. D.; Farkas, Ö.; Foresman, J. B.; Ortiz, J. V.; Cioslowski, J.; Fox, D. J. Gaussian, Inc., Wallingford CT, 2010.

Computational Results

Aniline was used as model molecule of amine substrate for computational study. Free energies are given in kcal/mol.



12. Characterization data for products

ethyl 2,2-difluoro-2-(phenanthridin-6-yl)acetate (3a)³



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a white solid (49.3 mg 82%). ¹H NMR (500 MHz, CDCl₃) δ 8.67 (d, *J* = 8.4 Hz,

1H), 8.61 – 8.53 (m, 2H), 8.12 (dd, J = 5.9, 3.6 Hz, 1H), 7.94 – 7.87 (m, 1H), 7.78 – 7.71 (m, 3H), 4.57 (q, J = 7.1 Hz, 2H), 1.48 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 163.7 (t, J = 31.3 Hz), 150.15 (t, J = 28.8 Hz), 141.7, 133.8, 131.2, 130.8, 129.0, 128.9, 127.8, 126.23 (t, J = 4.8 Hz), 124.8, 122.5, 122.3 (t, J = 2.5 Hz), 122.0, 115.81 (t, J = 252.5 Hz), 63.0, 14.1. ¹⁹F NMR (470 MHz, CDCl₃) δ -98.8.

ethyl 2,2-difluoro-2-(8-methylphenanthridin-6-yl)acetate (3b)



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a white solid (82%). ¹H NMR (500 MHz, CDCl₃) δ 8.59 – 8.50 (m, 2H), 8.32 (d, *J* = 0.7

Hz, 1H), 8.15 – 8.05 (m, 1H), 7.77 – 7.68 (m, 3H), 4.57 (q, J = 7.1 Hz, 2H), 2.62 (s, 3H), 1.48 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) 163.7 (t, J = 30 Hz), 149.9 (t, J = 30 Hz), 141.5, 138.0, 133.1, 131.8, 130.8, 128.8, 128.5, 125.5 (t, J = 3.8 Hz), 125.0, 125.0 (t, J = 2.5 Hz), 122.4 121.9, 116.0 (t, J = 252.5 Hz), 63.0, 21.9, 14.1. ¹⁹F NMR (470 MHz, CDCl₃) δ -98.9.

HRMS (ESI, m/z) calcd for C₁₈H₁₆F₂NO₂[M+H]⁺: 316.1149; found: 316.1145.

ethyl 2-(8-ethylphenanthridin-6-yl)-2,2-difluoroacetate (3c)



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a white solid (83%). ¹H NMR (500 MHz, CDCl₃) δ 8.62 (d, *J* = 8.5 Hz, 1H), 8.60 – 8.50

(m, 1H), 8.36 (d, J = 1.1 Hz, 1H), 8.16 – 8.09 (m, 1H), 7.83 – 7.71 (m, 3H), 4.59 (q, J = 7.1 Hz, 2H), 2.95 (q, J = 7.6 Hz, 2H), 1.50 (t, J = 7.1 Hz, 3H), 1.41 (t, J = 7.6 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 163.8 (t, J = 31.3 Hz), 150.0 (t, J = 28.8 Hz), 144.2, 141.5, 132.0, 132.0, 130.8, 128.8, 128.5, 125.0, 124.40 (t, J = 5.0 Hz), 122.6 (t, J = 2.5 Hz), 122.5, 121.9, 115.9 (t, J = 251.3 Hz), 63.0, 29.2, 15.5, 14.1. ¹⁹F NMR (470 MHz, CDCl₃) δ -99.0.

HRMS (ESI, m/z) calcd for C₁₉H₁₈F₂NO₂[M+H]⁺: 330.1306; found: 330.1302.

ethyl 2,2-difluoro-2-(8-isopropylphenanthridin-6-yl)acetate (3d)



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a white solid (83%).

 $\begin{array}{|c|c|c|c|c|} \hline N & CF_2COOEt \\ \hline \ ^1H \ NMR \ (500 \ MHz, \ CDCl_3) \ \delta \ 8.61 \ (d, \ J = 8.6 \ Hz, \ 1H), \\ \hline 8.58 - 8.52 \ (m, \ 1H), \ 8.37 \ (d, \ J = 1.8 \ Hz, \ 1H), \ 8.16 - 8.07 \ (m, \ 1H), \ 7.81 \ (dd, \ J = 8.6, \ 1.7 \ Hz, \ 1H), \ 7.76 - 7.63 \ (m, \ 2H), \ 4.56 \ (q, \ J = 7.1 \ Hz, \ 2H), \ 3.19 \ (dt, \ J = 13.8, \ 6.9 \ Hz, \ 1H), \ 1.47 \ (t, \ J = 7.1 \ Hz, \ 3H), \ 1.40 \ (d, \ J = 6.9 \ Hz, \ 6H). \ ^{13}C \ NMR \ (125 \ MHz, \ CDCl_3) \ \delta \ 163.8 \ (t, \ J = 31.3 \ Hz), \ 150.0 \ (t, \ J = 28.8 \ Hz), \ 148.8, \ 141.5, \ 132.2, \ 130.8, \ 130.5, \ 128.8, \ 128.5, \ 125.0, \ 123.10 \ (t, \ J = 5.0 \ Hz), \ 122.6 \ (t, \ J = 5 \ Hz), \ 122.6, \ 121.9, \ 115.9 \ (t, \ J = 252.5 \ Hz), \ 63.0, \ 34.4, \ 23.9, \ 14.1. \ ^{19}F \ NMR \ (470 \ MHz, \ CDCl_3) \ \delta \ -99.1. \ HRMS \ (ESI, \ m/z) \ calcd \ for \ C_{20}H_{20}F_2NO_2[M+H]^+; \ 344.1462; \ found: \ 344.1460. \ \end{array}$

ethyl 2,2-difluoro-2-(8-propylphenanthridin-6-yl)acetate (3e)



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a yellow oil (77%). ¹H NMR (500 MHz, CDCl₃) δ 8.58 (d, *J* = 8.5 Hz, 1H), 8.56 - 8.51

(m, 1H), 8.32 (d, J = 1.5 Hz, 1H), 8.22 – 8.01 (m, 1H), 7.79 – 7.65 (m, 3H), 4.57 (q, J = 7.1 Hz, 2H), 2.93 – 2.80 (m, 2H), 1.84 – 1.73 (m, 2H), 1.48 (t, J = 7.1 Hz, 3H), 1.01 (t, J = 7.3 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 163.8 (t, J = 30 Hz), 149.94 (t, J = 28.8 Hz), 142.7, 141.5, 132.4, 132.0, 130.8, 128.8, 128.5, 125.1 (t, J = 6.3 Hz), 125.0, 122.5 (t, J = 2.5 Hz), 122.4, 121.9, 115.9 (t, J = 252.5 Hz), 63.0, 38.2, 24.5, 14.1, 13.8. ¹⁹F NMR (470 MHz, CDCl₃) δ -99.1.

HRMS (ESI, m/z) calcd for C₂₀H₂₀F₂NO₂[M+H]⁺: 344.1462; found: 344.1458.

ethyl 2-(8-(tert-butyl)phenanthridin-6-yl)-2,2-difluoroacetate (3f)



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a yellow oil (76%). ¹H NMR (500 MHz, CDCl₃) δ 8.60 (d, *J* = 8.8 Hz, 1H), 8.54 (dd, *J* = 5.7, 3.7 Hz, 2H), 8.16 - 8.07 (m, 1H), 7.98 (dd, *J* = 8.8,

1.9 Hz, 1H), 7.76 – 7.68 (m, 2H), 4.56 (q, J = 7.1 Hz, 2H), 1.52 – 1.43 (m, 12H). ¹³C NMR (125 MHz, CDCl₃) δ 163.6 (t, J = 30 Hz), 151.0, 150.2 (t, J = 28.8 Hz), 141.6, 131.8, 130.8, 129.7, 128.8, 128.6, 124.9, 122.4 (t, J = 1.3 Hz), 122.3, 121.9, 121.8 (t, J = 5.0 Hz), 115.84 (t, J = 252.5 Hz), 63.0, 35.3, 31.2, 14.1. ¹⁹F NMR (470 MHz, CDCl₃) δ -99.2.

HRMS (ESI, m/z) calcd for C₂₁H₂₂F₂NO₂[M+H]⁺: 358.1619; found: 358.1615.

Ethyl 2,2-difluoro-2-(8-(methylthio)phenanthridin-6-yl)acetate (3g)



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a white solid (78%). ¹H NMR (500 MHz, CDCl₃) δ 8.65 – 8.39 (m, 2H), 8.25 (d, *J* = 1.9 Hz, 1H), 8.16 – 8.01 (m, 1H), 7.72 (ddt, *J* = 14.3, 6.8, 3.4

Hz, 3H), 4.56 (q, J = 7.1 Hz, 2H), 2.64 (s, 3H), 1.47 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 163.60 (t, J = 31.3 Hz), 149.08 (t, J = 28.8 Hz), 141.4, 139.6, 131.1, 130.9, 130.0, 129.1, 128.7, 124.8, 122.9 (t, J = 17.5 Hz), 122.7, 121.7, 121.1 (t, J = 5.0 Hz), 115.8, 63.1, 15.4, 14.1. ¹⁹F NMR (470 MHz, CDCl₃) δ -99.33 (s). HRMS (ESI, m/z) calcd for C₁₈H₁₆F₂NO₂S[M+H]⁺: 348.0870; found: 348.0876.

ethyl 2,2-difluoro-2-(8-methoxyphenanthridin-6-yl)acetate (3h)⁴



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a white solid (58.3 mg, 88%). ¹H NMR (500 MHz, CDCl₃) δ 8.55 (d, *J* = 9.2 Hz, 1H), 8.49 – 8.44 (m, 1H), 8.08 (dd, *J* = 7.9, 1.5 Hz, 1H), 7.89 –

7.83 (m, 1H), 7.74 – 7.64 (m, 2H), 7.51 (dd, J = 9.1, 2.6 Hz, 1H), 4.57 (q, J = 7.1 Hz, 2H), 3.99 (s, 3H), 1.48 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 163.7 (t, J = 31.3 Hz), 158.8, 148.1 (t, J = 28.8 Hz), 140.9, 130.7, 128.9, 128.3, 127.9, 125.0, 124.0, 123.6 (t, J = 2.5 Hz), 122.3, 121.5, 115.9 (t, J = 251.3 Hz), 105.7 (t, J = 5 Hz), 63.0, 55.5, 14.1. ¹⁹F NMR (470 MHz, CDCl₃) δ -99.9.

ethyl 2,2-difluoro-2-(8-fluorophenanthridin-6-yl)acetate (3i)



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a white solid (47 mg, 74%). ¹H NMR (500 MHz, CDCl₃) δ 8.63 (dd, *J* = 9.2, 5.3 Hz, 1H), 8.52 - 8.42 (m, 1H), 8.17 (ddd, *J* = 9.7, 4.3,

2.0 Hz, 1H), 8.12 – 7.96 (m, 1H), 7.79 – 7.69 (m, 2H), 7.63 (ddd, J = 9.1, 8.0, 2.6 Hz, 1H), 4.58 (q, J = 7.1 Hz, 2H), 1.49 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 163.37 (t, J = 31.3 Hz), 162.3, 160.4, 149.4 (t, J = 28.8 Hz), 149.3 (t, J = 30.0 Hz), 141.4, 130.9, 130.6, 129.4, 128.9, 125.1 (d, J = 7.5 Hz), 124.4, 123.4 (t, J = 1.25 Hz), 123.3 (t, J = 1.25 Hz), 121.8, 120.8 (d, J = 23.8 Hz), 115.6 (t, J = 252.5 Hz), 111.2 (t, J = 5 Hz), 111.0(t, J = 5 Hz) 63.1, 14.1. ¹⁹F NMR (470 MHz, CDCl₃) δ -99.3, -110.3. HRMS (ESI, m/z) calcd for C₁₇H₁₃F₃NO₂[M+H]⁺: 320.0898; found: 320.0894.

ethyl 2-(8-chlorophenanthridin-6-yl)-2,2-difluoroacetate (3j)⁴



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a white solid (77%). ¹H NMR (500 MHz, CDCl₃) δ 8.59 (d, *J* = 8.9 Hz, 1H), 8.55 – 8.43 (m, 2H), 8.09 (dd, *J* = 5.9, 3.6 Hz, 1H), 7.83 (dd, *J*

= 8.9, 2.1 Hz, 1H), 7.80 – 7.71 (m, 2H), 4.58 (q, J = 7.1 Hz, 2H), 1.49 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 163.33 (t, J = 30.0 Hz), 149.2 (t, J = 28.8 Hz), 141.6, 134.1, 132.2, 131.9, 131.0, 129.4, 129.4, 125.6 (t, J = 5 Hz), 124.3, 124.2, 123.1 (t, J =1.25 Hz), 121.9, 115.6 (t, J = 252.5 Hz), 63.1, 14.1. ¹⁹F NMR (470 MHz, CDCl₃) δ -98.8.

ethyl 2-(8-bromophenanthridin-6-yl)-2,2-difluoroacetate (3k)



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a white solid (72%). ¹H NMR (500 MHz, CDCl₃) δ 8.65 (dd, *J* = 3.7, 1.8 Hz, 1H), 8.55 - 8.40 (m, 2H), 8.24 - 8.01 (m, 1H), 7.93 (dd, *J* =

8.9, 2.0 Hz, 1H), 7.74 (dt, J = 5.5, 3.2 Hz, 2H), 4.58 (q, J = 7.1 Hz, 2H), 1.49 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 163.32 (t, J = 30.0 Hz), 149.00 (t, J = 31.3 Hz), 141.6, 134.5, 132.5, 130.9, 129.4, 129.4, 128.6 (t, J = 5 Hz), 124.2, 123.34 (t, J = 2.5 Hz), 122.2, 121.9, 115.6(t, J = 252.5 Hz), 63.2, 14.1. ¹⁹F NMR (470 MHz, CDCl₃) δ -98.6.

HRMS (ESI, m/z) calcd for C₁₇H₁₃BrF₂NO₂[M+H]⁺: 380.0098; found: 380.0093.

ethyl 2,2-difluoro-2-(8-phenylphenanthridin-6-yl)acetate (31)⁴



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a yellow oil (73%). ¹H NMR (500 MHz, CDCl₃) δ 8.77 (dd, *J* = 7.7, 5.4 Hz, 2H), 8.67 – 8.59 (m, 1H), 8.23 – 8.11 (m, 2H), 7.84 – 7.76 (m, 4H),

7.61 – 7.54 (m, 2H), 7.51 – 7.44 (m, 1H), 4.61 (q, *J* = 7.1 Hz, 2H), 1.51 (t, *J* = 7.1 Hz,

3H). 13C NMR (125 MHz, CDCl₃) δ 163.7 (t, J = 31.3 Hz), 150.24 (t, J = 28.8 Hz),

141.7, 140.6, 139.9, 132.9, 130.9, 130.5, 129.2, 129.0, 129.0, 128.1, 127.5, 124.7, 124.09 (t, *J* = 4.8 Hz), 123.1, 122.7 (t, *J* = 1.7 Hz), 122.1, 115.9 (t, *J* = 253.8 Hz), 63.1,

14.2. 19F NMR (470 MHz, CDCl₃) δ -98.8.

6-(2-ethoxy-1,1-difluoro-2-oxoethyl)phenanthridine-8-

carboxylate (3m)



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a white solid (72%). ¹H NMR (500 MHz, CDCl₃) δ 9.26 (d, *J* = 1.6 Hz, 1H),

8.71 (d, J = 8.7 Hz, 1H), 8.60 (dd, J = 7.0, 2.5 Hz, 1H), 8.51 (dd, J = 8.7, 1.5 Hz, 1H), 8.17 – 8.07 (m, 1H), 7.81 (pd, J = 7.1, 3.5 Hz, 2H), 4.61 (q, J = 7.1 Hz, 2H), 4.52 (q, J = 7.1 Hz, 2H), 1.51 (td, J = 7.1, 3.8 Hz, 6H). ¹³C NMR (125 MHz, CDCl₃) δ 165.7, 163.41 (t, J = 30.0 Hz), 150.6 (t, J = 30 Hz), 142.4, 136.6, 131.1, 130.9, 130.1, 129.7, 129.3, 128.4 (t, J = 5.0 Hz), 124.2, 122.8, 122.6, 121.8 (t, J = 1.25 Hz), 115.6 (t, J = 252.5 Hz), 63.1, 61.7, 14.4, 14.1. ¹⁹F NMR (470 MHz, CDCl₃) δ -98.3.

HRMS (ESI, m/z) calcd for C₂₀H₁₈F₂NO₄[M+H]⁺: 374.1204; found: 374.1199.

Ethyl2-(8-acetylphenanthridin-6-yl)-2,2-difluoroacetate(3n)



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a white solid (76%). ¹H NMR (500 MHz, CDCl₃) δ 9.05 (d, *J* = 1.6 Hz, 1H), 8.63 (d, *J* = 8.7 Hz, 1H), 8.52 (dd, *J* = 8.1, 1.4 Hz, 1H), 8.38

(dd, J = 8.7, 1.7 Hz, 1H), 8.13 - 8.00 (m, 1H), 7.76 (pd, J = 7.1, 1.6 Hz, 2H), 4.58 (q, J = 7.1 Hz, 2H), 2.76 (s, 3H), 1.49 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 197.0,163.3 (t, J = 30 Hz), 150.4 (t, J = 28.8 Hz), 142.4, 136.6, 135.7, 130.9, 130.2, 129.4, 129.4, 127.37 (t, J = 5.0 Hz), 124.0, 123.1, 122.6, 121.7 (t, J = 1.3 Hz), 115.59 (t, J = 252.5 Hz), 63.2, 26.6, 14.1. ¹⁹F NMR (470 MHz, CDCl₃) δ -98.1. HRMS (ESI, m/z) calcd for C₁₉H₁₆F₂NO₃[M+H]⁺: 344.1094; found: 344.1098.

ethyl 2,2-difluoro-2-(8-(trifluoromethyl)phenanthridin-6-yl)acetate

(30)



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a white solid (71%). ¹H NMR (500 MHz, CDCl₃) δ 8.83 (s, 1H), 8.79 (d, *J* = 8.8

Hz, 1H), 8.59 (dd, J = 6.7, 2.8 Hz, 1H), 8.18 – 8.11 (m, 1H), 8.09 (dd, J = 8.7, 1.5 Hz, 1H), 7.86 – 7.77 (m, 2H), 4.58 (q, J = 7.1 Hz, 2H), 1.49 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 163.2 (t, J = 30 Hz), 150.1 (t, J = 28.8 Hz), 142.3, 136.0, 131.1, 130.3, 129.9, 129.6, 127.14 (dd, J = 2.5 Hz), 124.8 (t, J = 1.3 Hz), 123.9 (q, J = 3.8

Ethyl

Hz), 123.6, 122.7, 122.4, 121.6 (t, J = 2.5 Hz), 115.5 (t, J = 252.5 Hz), 63.2, 14.1. ¹⁹F NMR (470 MHz, CDCl₃) δ -62.4, -98.3. HRMS (ESI, m/z) calcd for C₁₈H₁₃F₅NO₂[M+H]⁺: 370.0866; found: 370.0862.

ethyl 2-(7,9-dimethylphenanthridin-6-yl)-2,2-difluoroacetate (3p)



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a white solid (78%). ¹H NMR (500 MHz, CDCl₃) δ 8.51 (dd, *J* = 4.9, 3.8 Hz, 1H), 8.31 (s, 1H), 8.07 - 7.93 (m, 1H), 7.77 - 7.62 (m, 2H),

7.40 (s, 1H), 4.54 (q, J = 7.1 Hz, 2H), 2.95 (t, J = 2.9 Hz, 3H), 2.56 (s, 3H), 1.46 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 164.7 (t, J = 32.5 Hz), 149.16 (t, J = 32.5 Hz), 140.9, 140.5, 136.2, 135.8, 134.3, 130.0, 128.7, 128.7 125.1, 122.3, 121.0, 120.4, 117.14 (t, J = 256.3 Hz), 62.6, 23.9 (t, J = 12.5 Hz), 21.8, 14.1. ¹⁹F NMR (470 MHz, CDCl₃) δ -96.1.

HRMS (ESI, m/z) calcd for C₁₉H₁₈F₂NO₂[M+H]⁺: 330.1306; found: 330.1300.

ethyl 2-(benzo[i]phenanthridin-5-yl)-2,2-difluoroacetate (3q)⁴



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a white solid (63%). ¹H NMR (500 MHz, CDCl₃) δ 9.11 (d, *J* = 8.5 Hz, 1H), 8.67 – 8.54

(m, 2H), 8.20 (d, *J* = 8.9 Hz, 1H), 8.17 – 8.10 (m, 1H), 8.00 (dd, *J* = 7.9, 1.3 Hz, 1H), 7.83 – 7.68 (m, 4H), 4.46 (q, *J* = 7.1 Hz, 2H), 1.35 (t, *J* = 7.1 Hz, 3H). 13C NMR (125

MHz, CDCl₃) δ 164.29 (t, J = 32.5 Hz), 148.20 (t, J = 30.0 Hz), 142.0, 135.2, 133.1, 132.9, 129.9, 129.4, 128.8, 128.4 (t, J = 16.3 Hz), 128.4, 128.2, 127.4, 127.3, 124.7, 122.7, 120.8, 119.8, 117.1 (t, J = 257.5 Hz), 62.7, 14.0. ¹⁹F NMR (470 MHz, CDCl₃) δ -98.4.

ethyl 2-(9-chlorophenanthridin-6-yl)-2,2-difluoroacetate (3r)



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a white solid (74%).¹H NMR (500 MHz, CDCl₃) δ 8.63 (d, *J* = 2.0 Hz, 1H), 8.54 – 8.44 (m, 2H), 8.19 – 8.02 (m, 1H), 7.86 – 7.75 (m, 2H),

7.71 (dd, J = 8.9, 2.1 Hz, 1H), 4.57 (q, J = 7.1 Hz, 2H), 1.48 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 163.4 (t, J = 30 Hz),149.8 (t, H = 10.0 Hz), 142.1, 138.0,

135.3, 131.0, 129.7, 129.2, 128.6, 127.9 (t, J = 6.3 Hz), 123.8, 122.3, 122.1, 120.6 (t, J = 1.3 Hz), 115.6 (t, J = 252.5 Hz), 63.1, 14.1. ¹⁹F NMR (470 MHz, CDCl₃) δ -98.6. HRMS (ESI, m/z) calcd for C₁₇H₁₃ClF₂NO₂[M+H]⁺: 336.0603; found: 336.0595.

Ethyl 2-([1,3]dioxolo[4,5-j]phenanthridin-6-yl)-2,2-difluoroacetate (3s)



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a white solid (84%). ¹H NMR (500 MHz, CDCl₃) δ 8.35 (dd, *J* = 6.2, 3.5 Hz, 1H), 8.06 (dd, *J* = 6.2, 3.4 Hz, 1H), 7.94 (s, 1H), 7.85 (s, 1H),

7.68 (dd, J = 6.3, 3.3 Hz, 2H), 6.19 (s, 2H), 4.56 (q, J = 7.1 Hz, 2H), 1.48 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 163.69 (t, J = 31.3 Hz), 151.5, 148.6 (t, J = 12.5 Hz), 141.5, 132.2, 130.7, 128.5, 128.4, 125.0, 121.9, 118.9 (t, J = 1.3 Hz), 116.0 (t, J = 252.5 Hz), 103.5 (t, J = 5.0 Hz), 102.3, 100.3, 63.0, 14.1. ¹⁹F NMR (470 MHz, CDCl₃) δ -99.1.

HRMS (ESI, m/z) calcd for C₁₈H₁₄F₂NO₄[M+H]⁺: 346.0891; found: 346.0886.

ethyl 2,2-difluoro-2-(9-methoxyphenanthridin-6-yl)acetate (3t)



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a white solid (85%). ¹H NMR (500 MHz, CDCl₃) δ 8.51 – 8.41 (m, 2H), 8.08 (d, *J* = 7.7 Hz, 1H), 7.93 (d, *J* = 2.0 Hz, 1H), 7.77 – 7.65 (m, 2H), 7.32 (dd, *J* = 9.2, 2.5 Hz, 1H), 4.56 (q, *J* = 7.1 Hz,

2H), 4.04 (s, 3H), 1.47 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 163.75 (t, J = 31.3 Hz), 161.6, 149.5 (t, J = 28.8 Hz), 142.1, 136.3, 130.8, 129.1, 128.3, 128.1 (t, J = 5 Hz), 124.6, 122.1, 118.2, 117.12 (t, J = 1.3 Hz), 115.8 (t, J = 252.5 Hz), 103.1, 63.0, 55.6, 14.1. ¹⁹F NMR (470 MHz, CDCl₃) δ -98.6.

HRMS (ESI, m/z) calcd for C₁₈H₁₆F₂NO₃[M+H]⁺: 332.1098; found: 332.1098.

ethyl 2-(2-cyanophenanthridin-6-yl)-2,2-difluoroacetate (3u)



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a white solid (71%). ¹H NMR (500 MHz, CDCl₃) δ 8.90 (d, *J* = 1.5 Hz,

1H), 8.61 (dd, J = 19.9, 8.3 Hz, 2H), 8.18 (d, J = 8.4 Hz, 1H), 8.05 – 7.97 (m, 1H), 7.92 (dd, J = 8.4, 1.7 Hz, 1H), 7.89 – 7.81 (m, 1H), 4.57 (q, J = 7.1 Hz, 2H), 1.47 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 163.1 (t, J = 31.3 Hz), 153.1 (t, J = 28.8 Hz), 143.2, 132.9, 132.5, 132.0, 130.6, 129.3, 127.8, 126.7 (t, J = 5.0 Hz), 125.0,

122.6 (t, J = 2.5 Hz), 122.5, 118.5, 115.2 (t, J = 253.8 Hz), 112.4, 63.3, 14.1. ¹⁹F NMR (470 MHz, CDCl₃) δ -98.9. HRMS (ESI, m/z) calcd for C₁₈H₁₃F₂N₂O₂[M+H]⁺: 327.0945; found: 327.0942.

ethyl 2,2-difluoro-2-(3-fluorophenanthridin-6-yl)acetate (3v)



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a white solid (86%). ¹H NMR (500 MHz, CDCl₃) δ 8.50 (t, *J* = 7.2 Hz, 2H),

8.45 (dd, J = 9.1, 5.7 Hz, 1H), 7.84 (ddd, J = 8.3, 7.1, 1.1 Hz, 1H), 7.77 – 7.61 (m, 2H), 7.45 (ddd, J = 9.0, 8.1, 2.7 Hz, 1H), 4.58 (q, J = 7.1 Hz, 2H), 1.48 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 163.6, 163.5 (t, J = 15.4 Hz), 161.6, 151.4 (t, J = 28.8 Hz), 142.9 (d, J = 11.3 Hz), 133.6, 131.6, 127.7, 126.3 (t, J = 5.0 Hz), 124.1 (d, J = 8.8 Hz) 122.2, 121.8, 121.5 (d, J = 2.5 Hz), 118.1(d, J = 23.8 Hz), 115.7 (t, J = 253.8 Hz), 115.1 (d, J = 21.3 Hz), 63.1, 14.1. ¹⁹F NMR (470 MHz, CDCl₃) δ -98.7, -111.2.

HRMS (ESI, m/z) calcd for C₁₇H₁₃F₃NO₂[M+H]⁺: 320.0898; found: 320.0896.

ethyl 2,2-difluoro-2-(3-methoxyphenanthridin-6-yl)acetate (3w)⁵



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a white solid (78%). ¹H NMR (500 MHz, CDCl₃) δ 8.56 (d, *J* =

8.4 Hz, 1H), 8.51 (d, J = 8.2 Hz, 1H), 8.45 (d, J = 9.1 Hz, 1H), 7.85 (ddd, J = 8.3, 7.1, 1.1 Hz, 1H), 7.67 (ddd, J = 8.2, 7.0, 1.1 Hz, 1H), 7.49 (d, J = 2.7 Hz, 1H), 7.37 (dd, J = 9.0, 2.7 Hz, 1H), 4.56 (q, J = 7.1 Hz, 2H), 3.97 (s, 3H), 1.46 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 163.8 (t, J = 31.3 Hz), 160.3, 150.5 (t, J = 27.5 Hz), 143.5, 134.1, 131.2, 126.8, 126.2 (t, J = 5.0 Hz), 123.2, 122.0, 121.4 (t, J = 1.3 Hz), 120.1, 119.0, 115.7 (t, J = 252.5 Hz), 110.4, 63.0, 55.7, 14.1. ¹⁹F NMR (470 MHz, CDCl₃) δ -98.8.

HRMS (ESI, m/z) calcd for C₁₀H₁₄N₃O₂[M+H]⁺: 208.1081; found: 208.1080.

ethyl 2,2-difluoro-2-(2-(trifluoromethyl)phenanthridin-6-yl)acetate

(3x)



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a white solid (79%). ¹H NMR (500 MHz, CDCl₃) δ 8.85 (s, 1H),

8.70 (d, J = 8.4 Hz, 1H), 8.60 (d, J = 8.3 Hz, 1H), 8.23 (d, J = 8.5 Hz, 1H), 8.04 – 7.89 (m, 2H), 7.83 (ddd, J = 8.2, 7.1, 1.0 Hz, 1H), 4.57 (q, J = 7.1 Hz, 2H), 1.48 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 163.3 (t, J = 30 Hz),152.4 (t, J = 28.8 Hz), 143.1, 133.6, 132.0, 131.8, 130.6 (dd, J = 32.5 Hz), 128.8, 126.61 (t, J = 5.0 Hz), 125.1 (m, J = 2.5 Hz), 124.6, 122.9, 122.6, 120.8 (t, J = 3.8 Hz), 119.9 (q, J = 5 Hz), 115.4 (t, J = 253.8 Hz), 63.2, 14.1. ¹⁹F NMR (470 MHz, CDCl₃) δ -62.1, -98.9. HRMS (ESI, m/z) calcd for C₁₈H₁₃F₅NO₂[M+H]⁺: 370.0866; found: 370.0859.

ethyl 2,2-difluoro-2-(4-fluorophenanthridin-6-yl)acetate (3y)



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a white solid (86%). ¹H NMR (500 MHz, CDCl₃) δ 8.63 – 8.54 (m, 2H), 8.29 (d, *J* = 8.4 Hz, 1H), 7.90 (ddd, *J* = 8.3, 7.1, 1.2 Hz, 1H), 7.77

(ddd, J = 8.3, 7.1, 1.1 Hz, 1H), 7.66 (td, J = 8.1, 5.2 Hz, 1H), 7.40 (ddd, J = 9.3, 7.9, 1.0 Hz, 1H), 4.60 (q, J = 7.2 Hz, 2H), 1.50 (t, J = 7.2 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 163.5 (t, J = 30.0 Hz), 160.0, 157.9, 150.3 (t, J = 31.3 Hz), 133.3, 131.7, 131.5 (d, J = 10.0 Hz), 129.1 (d, J = 8.8 Hz) 128.5, 126.7, 126.4 (t, J = 5.0 Hz), 122.9, 122.4 (t, J = 2.5 Hz), 117.5 (d, J = 3.8 Hz), 115.8 (t, J = 252.5 Hz), 114.1 (d, J = 18.8 Hz), 63.3, 14.0. ¹⁹F NMR (470 MHz, CDCl₃) δ -98.7, -122.3.

HRMS (ESI, m/z) calcd for C₁₇H₁₃F₃NO₂[M+H]⁺: 320.0898; found: 320.0891.

ethyl 2,2-difluoro-2-(2-(trifluoromethoxy)phenanthridin-6-yl)acetate

(3z)



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a white solid (76%). ¹H NMR (500 MHz, CDCl₃) δ

8.68 – 8.55 (m, 2H), 8.39 (d, J = 1.8 Hz, 1H), 8.18 (d, J = 8.9 Hz, 1H), 8.05 – 7.91 (m, 1H), 7.89 – 7.79 (m, 1H), 7.63 (dd, J = 8.9, 1.5 Hz, 1H), 4.59 (q, J = 7.1 Hz, 2H), 1.50 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 163.4 (t, J = 30.0 Hz), 150.7 (t, J = 28.8 Hz), 149.1, 140.0, 133.3, 132.9, 131.6, 128.8, 126.49 (t, J = 5.0 Hz), 126.0, 122.6, 122.4 (t, J = 1.3 Hz), 122.3, 121.6, 117.6 (t, J = 252.5 Hz), 113.6, 63.1, 14.1. ¹⁹F NMR (470 MHz, CDCl₃) δ -57.6, -98.9.

HRMS (ESI, m/z) calcd for C₁₈H₁₃F₅NO₃[M+H]⁺: 386.0816; found: 386.0812.

ethyl 2-(benzo[c][1,5]naphthyridin-6-yl)-2,2-difluoroacetate (3aa)



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a white solid (77%). ¹H NMR (500 MHz, CDCl₃) δ 9.36 – 9.27 (m, 1H), 9.07 (dd,

J = 4.3, 1.7 Hz, 1H), 8.58 (d, J = 8.3 Hz, 1H), 8.41 (dd, J = 8.3, 1.7 Hz, 1H), 8.01 (ddd, J = 8.2, 7.1, 1.0 Hz, 1H), 7.88 (ddd, J = 8.4, 7.1, 1.3 Hz, 1H), 7.70 (dd, J = 8.3, 4.3 Hz, 1H), 4.58 (q, J = 7.1 Hz, 2H), 1.48 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 163.4 (t, J = 30.0 Hz), 151.2, 151.1 (t, J = 28.8 Hz), 141.5, 137.8, 136.6, 134.8, 131.8, 129.6, 125.6 (t, J = 5.0 Hz), 124.35 (t, J = 1.3 Hz), 124.2, 124.1, 115.3 (t, J = 252.5 Hz), 63.2, 14.1. ¹⁹F NMR (470 MHz, CDCl₃) δ -99.0.

HRMS (ESI, m/z) calcd for $C_{16}H_{13}F_2N_2O_2[M+H]^+$: 303.0945; found: 303.0942.

2,2'-(pyrrolo[1,2-a]quinoxaline-1,4-diyl)bis(2,2-

difluoroacetate) (3ab)

Diethyl



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a white solid (73%). ¹H NMR (500 MHz, CDCl₃) δ 8.19 (d, *J* = 8.6 Hz, 1H), 8.03 (dd, *J* = 8.0, 1.5 Hz, 1H), 7.64 (ddd, *J* = 8.7,

7.3, 1.6 Hz, 1H), 7.58 – 7.51 (m, 1H), 7.31 – 7.22 (m, 2H), 4.48 (q, J = 7.1 Hz, 2H), 4.37 (q, J = 7.1 Hz, 2H), 1.40 (t, J = 7.1 Hz, 3H), 1.26 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 162.9 (t, J = 33.8 Hz), 162.6 (t, J = 31.3 Hz), 135.1, 131.5, 130.1, 128.0, 126.4, 125.7, 122.4 (t, J = 31.3 Hz), 118.7 (t, J = 6.3 Hz), 116.8 (t, J = 7.5 Hz), 115.1, 113.1, 111.1 (t, J = 246.3 Hz), 107.34 (t, J = 3.8 Hz), 64.0, 63.4, 14.0, 13.8. ¹⁹F NMR (470 MHz, CDCl₃) δ -92.7, -105.1.

HRMS (ESI, m/z) calcd for C₁₉H₁₇F₄N₂O₄[M+H]⁺: 413.1124; found: 413.1117.

ethyl 2-(benzo[*d*]thiazol-2-yl)-2,2-difluoroacetate (5a)



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a white solid (78%). ¹H NMR

(500 MHz, CDCl₃) δ 8.16 (d, *J* = 7.8 Hz, 1H), 8.02 – 7.96 (m, 1H), 7.58 (ddd, *J* = 8.3, 7.3, 1.3 Hz, 1H), 7.55 – 7.50 (m, 1H), 4.44 (q, *J* = 7.1 Hz, 2H), 1.37 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 161.7 (t, *J* = 32.5 Hz), 160.3 (t, *J* = 32.5 Hz), 152.5, 135.1, 127.0, 127.0, 124.8, 122.0, 110.3 (t, *J* = 251.3 Hz), 64.0, 13.9. ¹⁹F NMR (470 MHz, CDCl₃) δ -98.4.

HRMS (ESI, m/z) calcd for $C_{11}H_{10}F_2NO_2S[M+H]^+$: 258.0400; found: 258.0396.

ethyl 2,2-difluoro-2-(5-methoxybenzo[d]thiazol-2-yl)acetate (5b)



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a white solid

(77%). ¹H NMR (500 MHz, CDCl₃) δ 8.01 (d, *J* = 9.0 Hz, 1H), 7.37 (d, *J* = 2.5 Hz, 1H), 7.16 (dd, *J* = 9.1, 2.5 Hz, 1H), 4.44 (q, *J* = 7.1 Hz, 2H), 3.90 (s, 3H), 1.37 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 161.9 (t, *J* = 31.3 Hz), 159.0, 157.3 (t, *J* = 32.5 Hz), 147.0, 136.8, 125.3, 117.0, 110.3 (t, *J* = 25.3 Hz), 103.6, 63.9, 55.9, 13.9. ¹⁹F NMR (470 MHz, CDCl₃) δ -98.0.

HRMS (ESI, m/z) calcd for $C_{12}H_{12}F_2NO_3S[M+H]^+$: 288.0506; found: 288.0502.

ethyl 2-(5-ethoxybenzo[d]thiazol-2-yl)-2,2-difluoroacetate (5c)



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a white solid

(82%). ¹H NMR (500 MHz, CDCl₃) δ 8.00 (d, *J* = 9.0 Hz, 1H), 7.36 (d, *J* = 2.5 Hz, 1H), 7.15 (dd, *J* = 9.0, 2.5 Hz, 1H), 4.44 (q, *J* = 7.1 Hz, 2H), 4.11 (q, *J* = 7.0 Hz, 2H), 1.47 (t, *J* = 7.0 Hz, 3H), 1.37 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 161.9 (t, *J* = 32.5 Hz), 158.4, 157.1 (t, *J* = 32.5Hz), 146.9, 136.8, 125.3, 117.4, 108.3 (t, *J* = 251.3 Hz), 64.2, 63.9, 14.7, 13.9. ¹⁹F NMR (470 MHz, CDCl₃) δ -98.0. HRMS (ESI, m/z) calcd for C₁₃H₁₄F₂NO₃S[M+H]⁺: 302.0662; found: 302.0658.

ethyl 2,2-difluoro-2-(5-(trifluoromethoxy)benzo[d]thiazol-2-yl)acetate

(5d)



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a white solid

(85%). ¹H NMR (500 MHz, CDCl₃) δ 8.16 (d, *J* = 9.0 Hz, 1H), 7.85 (d, *J* = 1.2 Hz, 1H), 7.45 (dd, *J* = 9.0, 1.6 Hz, 1H), 4.44 (q, *J* = 7.1 Hz, 2H), 1.38 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 161.5 (t, *J* = 32.5 Hz), 161.4 (t, *J* = 31.3 Hz), 150.8, 147.8, 136.1, 125.8, 121.1, 114.3, 109.9 (t, *J* = 252.5 Hz), 64.1, 13.9. ¹⁹F NMR (470 MHz, CDCl₃) δ -58.0, -98.5.

HRMS (ESI, m/z) calcd for $C_{12}H_9F_5NO_3S[M+H]^+$: 342.0223; found: 342.0220.

ethyl 2,2-difluoro-2-(5-methylbenzo[d]thiazol-2-yl)acetate (5e)



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a white solid (79%).

¹H NMR (500 MHz, CDCl₃) δ 8.02 (d, J = 8.4 Hz, 1H), 7.75 (s, 1H), 7.37 (dd, J = 8.4, 1.5 Hz, 1H), 4.44 (q, J = 7.1 Hz, 2H), 2.52 (s, 3H), 1.37 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 161.8 (t, J = 31.3 Hz), 159.1 (t, J = 32.5 Hz), 150.6, 137.5, 135.4, 128.7, 124.2, 121.5, 110.3 (t, J = 251.3 Hz), 63.9, 21.7, 13.9. ¹⁹F NMR (470 MHz, CDCl₃) δ -98.3.

HRMS (ESI, m/z) calcd for $C_{12}H_{12}F_2NO_2S[M+H]^+$: 272.0557; found: 272.0556.

ethyl 2-(5-bromobenzo[d]thiazol-2-yl)-2,2-difluoroacetate (5f)



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a white solid (67%).

¹H NMR (500 MHz, CDCl₃) δ 8.13 (d, J = 1.8 Hz, 1H), 8.00 (d, J = 8.8 Hz, 1H), 7.68 (dd, J = 8.8, 1.9 Hz, 1H), 4.44 (q, J = 7.1 Hz, 2H), 1.37 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 161.5 (t, J = 32.5 Hz), 160.8 (t, J = 32.5 Hz), 151.3, 136.7, 130.7, 125.8, 124.7, 121.0, 110.0 (t, J = 252.5 Hz), 64.1, 13.9. ¹⁹F NMR (470 MHz, CDCl₃) δ -98.43 (s).

HRMS (ESI, m/z) calcd for C₁₁H₁₀BrF₂NO₂S[M+H]⁺: 335.9505; found: 335.9507.

ethyl 2-(6-chlorobenzo[d]thiazol-2-yl)-2,2-difluoroacetate (5g)



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a yiellow oil (75%).

¹H NMR (500 MHz, CDCl₃) δ 7.92 (dd, J = 8.9, 5.0 Hz, 1H), 7.83 (dd, J = 9.1, 2.5 Hz, 1H), 7.31 (td, J = 8.8, 2.5 Hz, 1H), 4.44 (q, J = 7.1 Hz, 2H), 1.38 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 163.0, 162.7 (t, J = 32.5 Hz), 161.5 (t, J = 31.3 Hz), 161.0, 153.4 (d, J = 12.2 Hz), 130.6, 122.9 (d, J = 9.9 Hz), 116.3 (d, J = 25 Hz), 110.82 (d, J = 23.8 Hz), 110.00 (t, J = 251.3 Hz), 64.1, 13.9. ¹⁹F NMR (470 MHz, CDCl₃) δ -98.6, -114.0.

HRMS (ESI, m/z) calcd for C₁₁H₉F₃NO₂S[M+H]⁺: 276.0301; found: 276.0303.

ethyl 2-(4,6-difluorobenzo[d]thiazol-2-yl)-2,2-difluoroacetate (5h)



The reaction was performed following the general procedure. The residue was purified by flash column chromatograph (silica gel, petroleum ether:AcOEt = 100:1, v/v) to give the product as a yiellow oil (75%). ¹H NMR (500 MHz, CDCl₃) δ 7.48 (ddd, J = 7.5, 2.3,

1.3 Hz, 1H), 7.09 (ddd, J = 9.9, 9.2, 2.3 Hz, 1H), 4.45 (q, J = 7.1 Hz, 2H), 1.38 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 160.63 – 160.16 (m), 138.1,109.8 (t, J = 250 Hz), 104.14 (d, J = 5.0 Hz), 104.03 (dd, J = 26.8, 5.0 Hz), 104.28 – 103.46 (m), 64.2, 13.9. ¹⁹F NMR (470 MHz, CDCl₃) δ -97.9, -108.45 (d, J = 7.0 Hz), -114.71 (d, J = 7.1 Hz).

HRMS (ESI, m/z) calcd for $C_{11}H_8F_4NO_2S[M+H]^+$: 294.0212; found: 294.0208.

(2'-isocyano-[1,1'-biphenyl]-4-yl)(methyl)sulfane (8)

¹H NMR (500 MHz, CDCl₃) δ 7.51 – 7.32 (m, 8H), 2.54 (s, 3H). ¹³C NMR (125MHz, CDCl₃) δ 166.5, 139.3, 138.2, 133.5, 130.4 129.6, 129.3, 128.0, 126.2, 15.5.



(2'-isocyano-[1,1'-biphenyl]-4-yl)(methyl)sulfane

(11)



¹H NMR (500 MHz, CDCl₃) δ 7.49 – 7.31 (m, 4H), 7.01 – 6.95 (m, 2H), 6.92 (t, *J* = 5.0 Hz, 1H), 6.03 (s, 2H). ¹³C NMR (125 MHz, CDCl₃) δ 166.5, 147.5, 138.5, 130.8, 130.5, 129.5, 127.9, 122.9, 109.5, 108.5, 101.4.

1-(difluoromethyl)-1H-benzo[d]imidazole (10)

¹H NMR (500 MHz, CDCl₃) δ 8.12 (s, 1H), 7.93 – 7.76 (m, 1H), 7.61 (dd, J = 5.4, 3.6 Hz, 1H), 7.37 (ddd, J = 85.8, 55.8, 43.2 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 143.9, 139.1, 130.6, 124.8, 124.2, 121.0, 111.1, 109.0 (t, J = 248.8 Hz) ¹⁹F NMR (470 MHz, CDCl₃) δ -93.7.

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14. NMR spectroscopic data



ethyl 2,2-difluoro-2-(phenanthridin-6-yl)acetate (3a)





ethyl 2,2-difluoro-2-(8-methylphenanthridin-6-yl)acetate (3b)







ethyl 2-(8-ethylphenanthridin-6-yl)-2,2-difluoroacetate (3c)





ethyl 2,2-difluoro-2-(8-isopropylphenanthridin-6-yl)acetate (3d)








ethyl 2,2-difluoro-2-(8-propylphenanthridin-6-yl)acetate (3e)





ethyl 2-(8-(tert-butyl)phenanthridin-6-yl)-2,2-difluoroacetate (3f)







ethyl 2,2-difluoro-2-(8-(methylthio)phenanthridin-6-yl)acetate (3g)





ethyl 2,2-difluoro-2-(8-methoxyphenanthridin-6-yl)acetate (3h)









ethyl 2,2-difluoro-2-(8-fluorophenanthridin-6-yl)acetate (3i)





ethyl 2-(8-chlorophenanthridin-6-yl)-2,2-difluoroacetate (3j)









ethyl 2-(8-bromophenanthridin-6-yl)-2,2-difluoroacetate (3k)





ethyl 2,2-difluoro-2-(8-phenylphenanthridin-6-yl)acetate (3l)







Ethyl 6-(2-ethoxy-1,1-difluoro-2-oxoethyl)phenanthridine-8-

carboxylate (3m)





ethyl 2-(8-acetylphenanthridin-6-yl)-2,2-difluoroacetate(3n)







ethyl 2,2-difluoro-2-(8-(trifluoromethyl)phenanthridin-6-yl)acetate

(30)





ethyl 2-(7,9-dimethylphenanthridin-6-yl)-2,2-difluoroacetate (3p)







ethyl 2-(benzo[i]phenanthridin-5-yl)-2,2-difluoroacetate (3q)





ethyl 2-(9-chlorophenanthridin-6-yl)-2,2-difluoroacetate (3r)







Ethyl 2-([1,3]dioxolo[4,5-j]phenanthridin-6-yl)-2,2-difluoroacetate (3s)



ethyl 2,2-difluoro-2-(9-methoxyphenanthridin-6-yl)acetate (3t)









ethyl 2-(2-cyanophenanthridin-6-yl)-2,2-difluoroacetate (3u)





ethyl 2,2-difluoro-2-(3-fluorophenanthridin-6-yl)acetate (3v)









ethyl 2,2-difluoro-2-(3-methoxyphenanthridin-6-yl)acetate (3w)





ethyl 2,2-difluoro-2-(2-(trifluoromethyl)phenanthridin-6-yl)acetate

(3x)









ethyl 2,2-difluoro-2-(4-fluorophenanthridin-6-yl)acetate (3y)





ethyl 2,2-difluoro-2-(2-(trifluoromethoxy)phenanthridin-6-yl)acetate











ethyl 2-(benzo[c][1,5]naphthyridin-6-yl)-2,2-difluoroacetate (3aa)







(3ab)






ethyl 2-(benzo[d]thiazol-2-yl)-2,2-difluoroacetate (5a)



ethyl 2,2-difluoro-2-(5-methoxybenzo[d]thiazol-2-yl)acetate (5b)







ethyl 2-(5-ethoxybenzo[d]thiazol-2-yl)-2,2-difluoroacetate (5c)





(5d)









ethyl 2,2-difluoro-2-(5-methylbenzo[d]thiazol-2-yl)acetate (5e)



ethyl 2-(5-bromobenzo[d]thiazol-2-yl)-2,2-difluoroacetate (5f)









ethyl 2-(6-chlorobenzo[d]thiazol-2-yl)-2,2-difluoroacetate (5g)





ethyl 2-(4,6-difluorobenzo[d]thiazol-2-yl)-2,2-difluoroacetate (5h)







(2'-isocyano-[1,1'-biphenyl]-4-yl)(methyl)sulfane (8)



(2'-isocyano-[1,1'-biphenyl]-4-yl)(methyl)sulfane (11)



1-(difluoromethyl)-1*H*-benzo[*d*]imidazole (10)

