

Supporting Information

Squaryl group modified phosphoglycolipid analogs as potential modulators of GPR55

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1. Biological experiments

1-1. Materials and methods

Animals

All experimental protocols using animals were approved by the Wako Animal Experiments Committee at RIKEN. Fertilized Boris Brown chicken eggs were obtained from a local supplier (Inoue Poultry Farm, Sagamihara, Japan) and stored in a 38 °C rocking incubator until the embryos had developed to Hamburger and Hamilton Stage 36 (*Hamburger and Hamilton, 1951*).

Cell culture

Dorsal root ganglia (DRGs) were dissected from Stage 36 chick embryos and DRG sensory neurons were dissociated into single cells by trypsinization and manual trituration. Dissociated neurons were then suspended in Leibowitz's L15 medium (Life Technologies, CA) with N2 Supplement (Life Technologies), 20 ng/ml nerve growth factor (NGF; Promega, WI) and 750 µg/ml bovine serum albumin (Life Technologies), and seeded onto 3 cm glass-based dishes that had been coated with 9 µg/cm² mouse laminin (Life Technologies). Seeded dishes were placed in a 37 °C incubator with 5% CO₂ for 1 – 4 hours before use in the turning assay. As described previously (*Guy et al., 2015*), these neuronal cultures allowed us to selectively analyse NGF-responsive nociceptive axons. Test compounds were dissolved in a vehicle of 0.001% (v/v) DMSO/PBS and used at an in-pipette concentration of 10 or 20 µM. Where used, specific GPR55 inhibitor ML193 (*Kotsikorou et al, 2013*; Tocris Bioscience, Bristol, England) was dissolved in DMSO and added to the cell culture medium to give a final bath concentration of 10 µM, at least 20 minutes before start of the turning assay. For control experiments, an equivalent volume of DMSO (0.001% (v/v)) was added to the cell culture medium 20 minutes before the assay.

In vitro axon turning assay

Axon turning assays (*Lohof et al., 1992*) were performed as described previously (*Guy et al., 2015*) with minor modifications. On an Olympus IX51 inverted microscope (Olympus Corporation, Tokyo, Japan), a growth cone, the tip of single extending axons, was introduced to a microscopic concentration gradient of DFQ test compound and imaged for 45 minutes using a QICAM Fast 1394 (QImaging, Burnaby, Canada) CCD digital camera controlled by Metavue software (version 7.8.2.0, Molecular Devices, CA). The microscopic concentration gradient was produced by repeated pulsatile ejection of DFQ solution through a glass micropipette (borosilicate glass with filament,

catalog # BF100-50-10, Sutter Instrument, CA) the tip of which was placed 100 μm away from the growth cone. Micropipettes were fashioned using a P-97 Flaming/Brown micropipette puller (Sutter Instrument), designed to have a steep shoulder and an approximate tip diameter of 1 – 2 μm .

Positive nitrogen gas pressure of 1 – 4 psi was applied to the test solution within the pipette by connection to a PV820 Picopump (World Precision Instruments, FL) electrically gated pressure control system. The frequency (2 Hz) and duration (20 ms) of positive pressure was controlled by a pulse generator (model AWG-50, ELMOS, Osaka, Japan). This pulsatile ejection of test solution from the pipette tip induces and maintains a microscopic concentration gradient of test compound in the culture medium for the duration of the assay (Lohof *et al*, 1992, Zheng *et al*, 1994). At a distance of 100 μm from the pipette tip, test compound concentration is approximately 1000 times lower than the in-pipette concentration ((Lohof *et al*, 1992, Ming *et al*, 1997). At the end of the assay, the turning angle (the angle between the growth cone's start position and its final position, measured relative to its axon) and axon extension (in microns) were calculated using Metavue software (**Figure S-1**). Growth cones that bifurcated, collapsed, retracted or failed to extend more than 10 μm during the 45 minutes of the assay were discounted from the study. All turning assays in the initial exploratory experiment (**Figure 2b**) were carried out with the experimenter blind to the identity of the compound being tested.

Statistics

Statistical analyses were carried out using GraphPad Prism (version 6.0c, GraphPad Software Inc, CA). Experimental datasets were tested for normality by D'Agostino and Pearson omnibus test, and for homoskedasticity using Bartlett's test. Datasets that failed normality or equal variance tests (**Figure 2c**, Compound **1A** turning angle) were analyzed using Kruskal-Wallis test, with Dunn's multiple comparisons post-test. All three datasets shown in **Figure S-2** were found to have normal distribution and equal variances, and were analyzed by 1-way ANOVA with Tukey's multiple comparisons post-test. Detailed statistical test data can be provided on request.

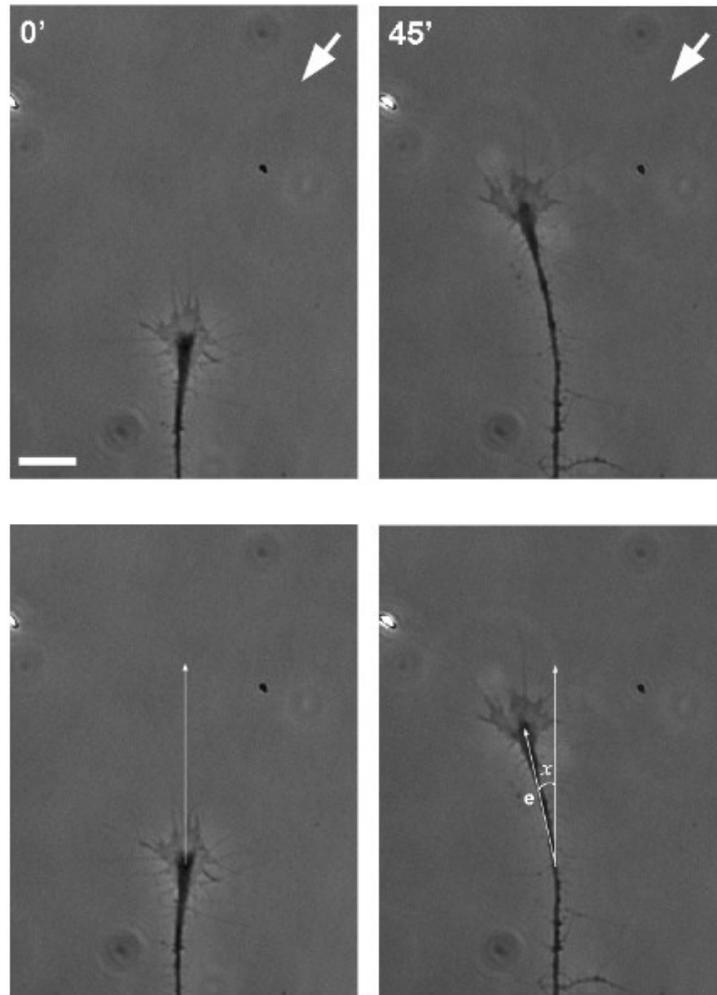


Figure S-1. Method of calculation of turning angle and axon extension. A: Representative images of start (left panel) and end (right panel) of chemorepulsive axon turning observed during turning assay. Numbers in each panel show the minutes after onset of concentration gradient, arrows indicate the source of the gradient. Scale bar, 10 μm . B: the same axonal growth cone as shown in A, with a white line with arrowhead superimposed on the image to show the initial direction of growth (left panel). A second straight line was traced from the center of the growth cone at the beginning of the assay to the center of the growth cone at the end of the assay (right panel). Turning angle (x) between this line and the original direction of growth was calculated using Metavue software. Axon extension (e) was measured as the length of the second straight line in microns, using the same program.

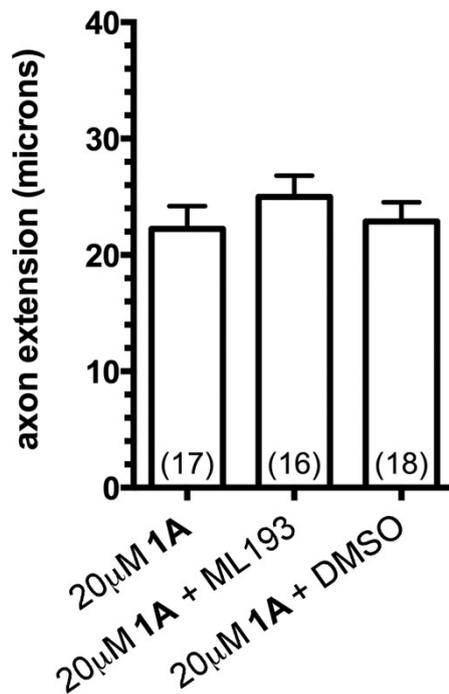


Figure S-2. Axon growth is unaffected by compound 1A or treatment with ML193. Bars show mean axon extension in microns \pm SEM, numbers in parentheses indicate the number of axons tested. No significant difference in axon extension was observed.

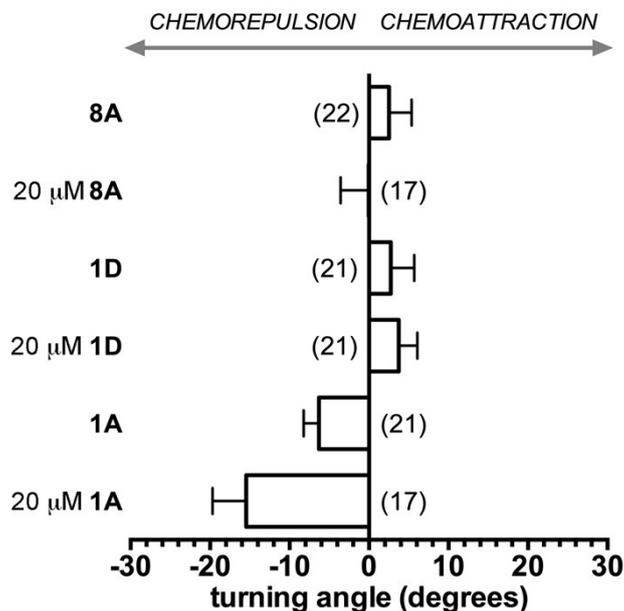


Figure S-3. Compounds 1D and 8A were not active in axon turning assays. Compared to compound 1A which displayed a dose-dependent increase in chemorepulsive activity, 1D and 8A did not show significant chemotropic activity even when tested at the higher in-pipette concentration of 20 µM. Bars show mean axon turning angle \pm SEM, numbers in parentheses indicate the number of axons tested. Data for 8A, 1D, 1A and 20 µM 1A bars are the same experimental data as shown in Figure 2b and 2d.

References

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Zheng, J. Q.; Felder, M.; Connor, J. A.; Poo, M. M. Turning of Nerve Growth Cones Induced by Neurotransmitters. *Nature* **1994**, *368* (6467), 140–144.

Ming, G. L.; Song, H. J.; Berninger, B.; Holt, C. E.; Tessier-Lavigne, M.; Poo, M. M. cAMP-Dependent Growth Cone Guidance by Netrin-1. *Neuron* **1997**, *19* (6), 1225–1235.

2. Quantum mechanics calculations

Quantum mechanics simulations were performed utilizing the Gaussian 09 program on the RIKEN internal HOKUSAI cluster. All geometries were optimized at the MP2/6-31+G(d) level followed by single point energy calculations. The dihedral angle of the anomeric linkage is referred to as φ , measured between O(ring)-C(ring)-O(ester)-P and O(ring)-C(ring)-N-C(double bond) in compound **2** and **3** respectively. The inner dihedral angle of the phosphate on the carbohydrate side in **2** is referred to as ψ composed of C(ring)-O(ester)-P-O(ester), while the second inner phosphate dihedral is referred to as ω , measured between O(ester)-P-O(ester)-C(methyl). Potential energy scans of **3** were conducted with all three defined dihedrals φ , ψ and ω constrained. At each set of φ and ψ three different positions of ω , namely gauche⁺, gauche⁻ and anti, were evaluated and the lowest energy conformation was selected to generate the plot in figure 1a and the corresponding supporting figure S4. In case of compound **3**, ψ is defined as C(ring)-N-C(double bond)-C(carbonyl). During potential energy scans of **3** only φ and ψ were restrained and utilized to generate figure 1b. All potential energy plots were prepared utilizing the GNUplot 4.6 software package.

The lowest energy conformation of **2** at each of the three identified potential energy minima was screened against all conformations of **3**. To rank the degree of similarity, first the heavy atoms of the tetrahydropyran ring of each structure were superimposed. Next the shortest distance between either of the phosphate carbonyl oxygen of **2** with either of the carbonyl oxygen of **3** was determined, as well as the distance between the methyl group carbon of the **2** and **3**. The top 8 structures of **3** with the smallest distance difference compared to the respective conformation of **2** were plotted on the energy landscape of **3** to identify the corresponding conformational space. The structures were visualized with the VMD [Humphrey, A. Dalke, K. Schulten; J. Molec. Graphics, 1996, 14, 33-38] software package and initially ranked utilizing an in-house script.

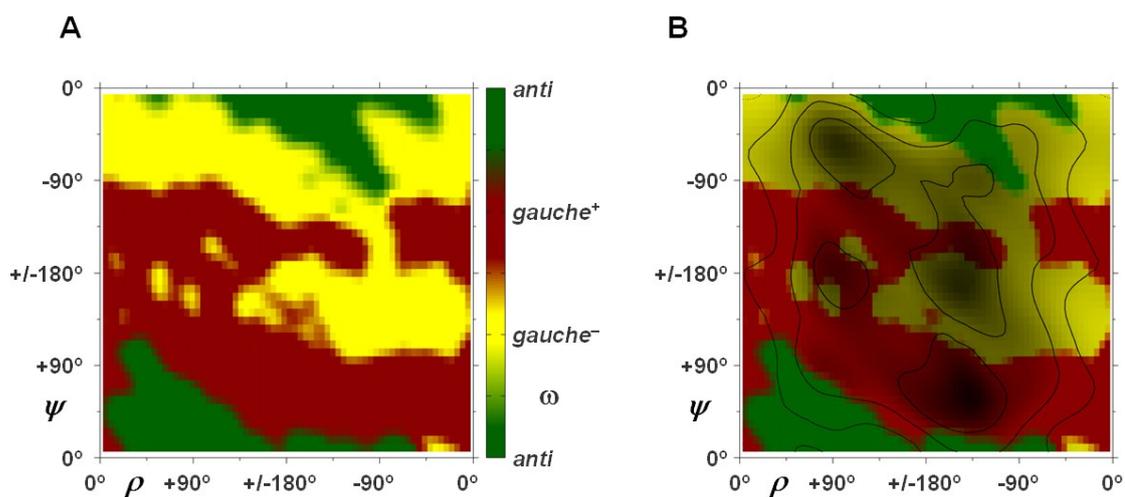


Figure S4 (A) Heatmap of second inner dihedral phosphate angle of compound **2**; green, anti; yellow, gauche⁻, red, gauche⁺; (B) overlay with conformational energy, dark areas indicate regions of low energy.

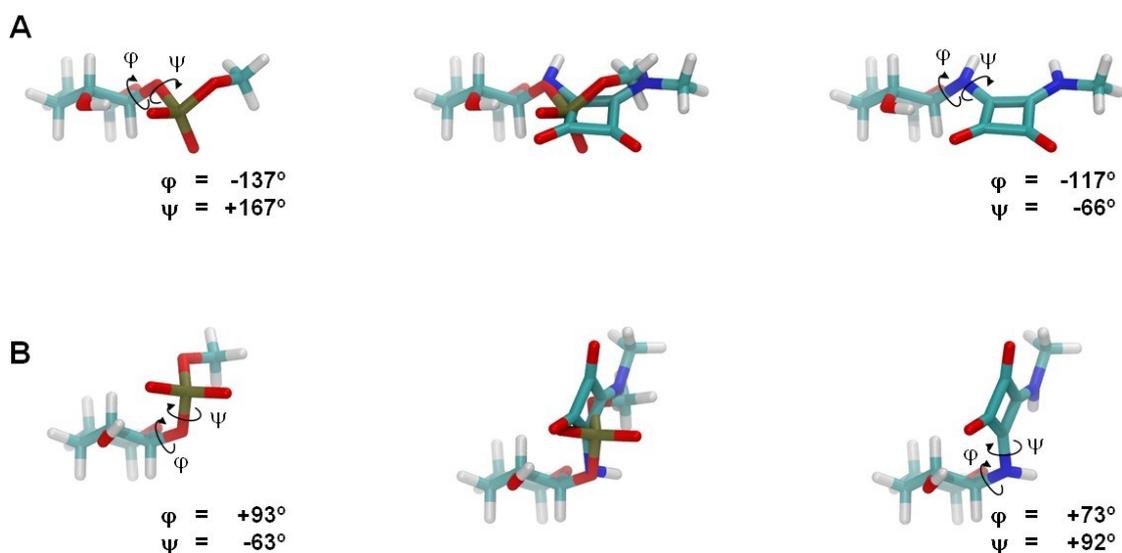


Figure S5: Lowest energy conformation of compound **2** in potential energy minima II (A) and III (B) is superimposed with close matching conformation of compound **3**.

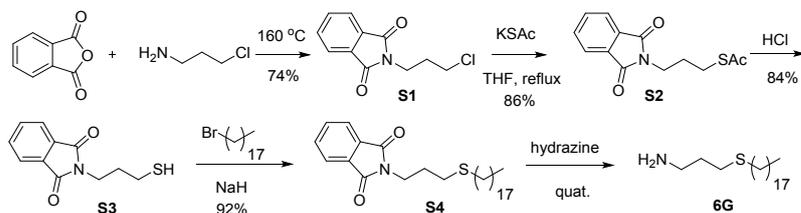
3. Chemical experiments and compound characterization

3-1. General procedures

All reactions sensitive to air and/or moisture were carried out under nitrogen or argon atmosphere with anhydrous solvents. Column chromatography was performed on silica gel 60 (EM Science, 40–100 mesh). Reactions were monitored by thin-layer chromatography (TLC) on Kieselgel 60 F254 (EM Science), and compounds were detected by examination under UV light and by charring with 10% sulfuric acid in MeOH. Solvents were removed under reduced pressure at <40 °C. All reactions were carried out under an argon atmosphere. ¹H NMR and ¹³C NMR spectra were recorded on a Bruker Advance 500 spectrometer with a cryoprobe and on a JEOL ECX 400 spectrometer. ¹H NMR spectra were recorded in CDCl₃ referenced to TMS at 0.00 ppm or to CHCl₃ at 7.24 ppm and to H₂O at 4.79 ppm, and ¹³C NMR spectra were referenced to CDCl₃ at 77.0 ppm or native scale. Assignments were made by standard 2D experiments. MALDI-TOF mass spectra were recorded on a Bruker Autoflex Speed spectrometer and a SHIMADZU Kompact MALDI AXIMA-CFR spectrometer with 2-hydroxy-5-methoxybenzoic acid as the matrix. HRMS determinations were made the use of a JEOL AccuTOF JMS-T700LCK mass spectrometer with CF₃CO₂Na as the internal standard. All other reagents were purchased from Kanto Chemicals Co. Inc., Tokyo Chemical Industries Co., Ltd., Wako Pure Chemical Industries Ltd., and Aldrich Chemical Co.

3-2. Synthesis and compound characterization

Scheme S1. Synthesis of 3-(octadecylthio)propan-1-amine (**6G**):



2-(3-(3-chloropropyl)-1*H*-isindole-1,3(2*H*)-dione (**S1**):¹

Phthalic anhydride (2.96 g, 20 mmol) and 3-chloropropylamine hydrochloric salt (2.86 g, 22 mmol) were heated in an oil bath at 160 °C. The reaction mixture was maintained at the same temperature for 15 min. Subsequently, it was cooled to room temperature, and 30 ml H₂O were added before solidification, to form slurry. The product was filtered, washed twice with H₂O, and purified by flash chromatography on silica gel (eluent, *n*-hexane: EtOAc = 10:1 to 6:1) afforded the title compound **S1** (3.30 g, 74% yield) as a white solid. ¹H NMR (CDCl₃, 400 MHz): δ 7.79–7.65 (m, 4H), 3.78 (t, *J* = 6.88 Hz, 2H), 3.52 (t, *J* = 6.44 Hz, 2H), 2.15–2.08 (m, 2H); ¹³C NMR (CDCl₃, 100 MHz): δ 168.29, 134.13, 132.05, 123.35, 42.14, 35.68, 31.51; HRMS (ESI) *m/z* [M+Na]⁺ calculated for C₁₁H₁₀ClNO₂Na 246.0298, found 246.0294.

S-(3-(1,3-Dioxoisindolin-2-yl)propyl)ethanethioate (**S2**):²

To a stirred solution of **S1** (2.23 g, 11 mmol) in THF (120 mL) was added potassium thioacetate (33 mmol), and the mixture was refluxed for 4 h at 85 °C. And the solvent was evaporated to dryness in vacuo, and then added water (100 mL), and the organic materials were extracted with EtOAc (3 × 30 mL). The combined extracts were washed with water and brine, dried over anhydrous MgSO₄, and concentrated in vacuo. The crude product was purified by flash chromatography on silica gel (eluent, *n*-hexane: EtOAc = 10:1 to 5:1) to give compound **S2** as white powder (2.49 g, 86% yield). ¹H NMR (CDCl₃, 400 MHz): δ 7.78–7.64 (m, 4H), 3.68 (t, *J* = 6.88 Hz, 2H), 2.82 (t, *J* = 7.32 Hz, 2H), 2.25 (s, 3H), 1.93–1.86 (m, 2H); ¹³C NMR (CDCl₃, 100 MHz): δ 168.29, 134.13, 132.05, 123.35, 42.14, 35.68, 31.51; HRMS (ESI) *m/z* [M+Na]⁺ calculated for C₁₃H₁₃NO₃SNa 286.0514, found 286.0523.

2-(3-(3-Mercaptopropyl)isindoline-1,3-dione (**S3**):

A solution of **S2** (2 g, 7.6 mmol) in anhydrous methanol (23 mL) was first degassed and refilled with N₂. Concentrated HCl (3 mL) was added to the solution, and the resulting mixture was refluxed for 5 h under N₂ atmosphere until no more starting material could be detected by TLC. The reaction mixture was quenched with water (20 mL) and extracted with EtOAc (3 × 30 mL). The organic layer was dried over anhydrous Na₂SO₄ and concentrated in vacuo to give crude product, which was purified by flash chromatography on silica gel (eluent, *n*-hexane: EtOAc = 10:1 to 5:1) to give compound **S3** as white powder (1.41 g, 84% yield).

¹H NMR (CDCl₃, 400 MHz): δ 7.81–7.66 (m, 4H), 3.76 (t, *J* = 6.88 Hz, 2H), 2.50 (q, *J* = 7.32 Hz, 2H), 1.98–1.91 (m, 2H); ¹³C NMR (CDCl₃, 100 MHz): δ 168.45, 134.09, 132.06, 123.34, 36.46, 32.78, 21.96; HRMS (ESI) *m/z* [M+Na]⁺ calculated for C₁₁H₁₁NO₂SNa 244.0408, found 244.0402.

2-(3-(octadecylthio)propyl)isindoline-1,3-dione (**S4**):

1-Bromooctadecane (2.12 g, 6.36 mmol) was added to a mixture containing sodium hydride (424

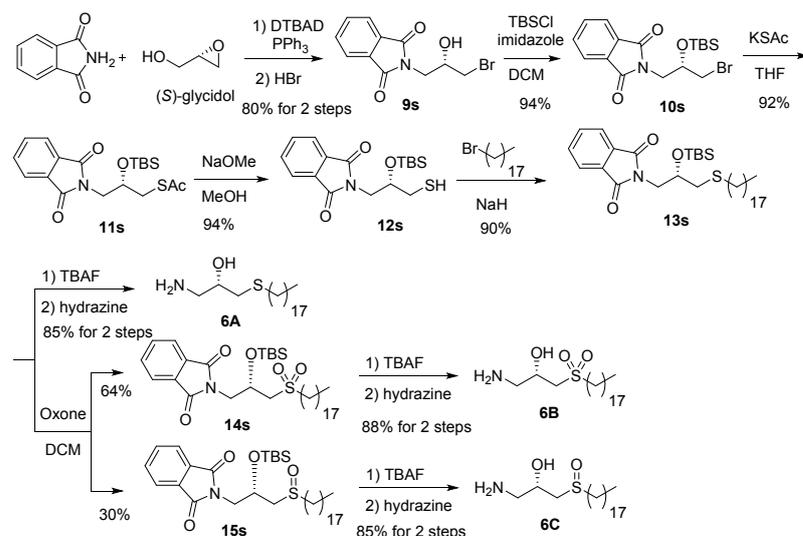
mg, 10.6 mmol) and compound **S3** (1.17 g, 5.3 mmol) in dry THF (15 mL). The mixture was stirred at room temperature for 2 h. The reaction mixture was cooled to 0 °C and water (20 mL) was added. The solution was extracted with EtOAc (3 × 30 mL). The organic layer was separated, dried with anhydrous Na₂SO₄, and evaporated under vacuum. The crude product was purified using flash chromatography on silica gel (eluent, *n*-hexane: EtOAc = 10:1 to 5:1) to give compound **S3** as white powder (2.3 g, 92% yield).

¹H NMR (CDCl₃, 400 MHz): δ 7.85–7.70 (m, 4H), 3.79 (t, *J* = 7.32 Hz, 2H), 2.56–2.48 (m, 2H), 1.99–1.92 (m, 2H), 1.58–1.51 (m, 2H), 1.34–1.24 (m, 32H), 0.87 (t, *J* = 6.40 Hz, 3H); ¹³C NMR (CDCl₃, 100 MHz): δ 168.46, 134.04, 132.19, 123.34, 37.31, 32.16, 32.06, 29.79, 29.71, 29.66, 29.63, 29.46, 29.34, 29.00, 28.55, 22.79, 14.23; HRMS (ESI) *m/z* [M+Na]⁺ calculated for C₂₉H₄₇NO₂SNa 496.3225, found 496.3234.

3-(octadecylthio)propan-1-amine (**S5**):

N-Cinnamylphthalimide **S4** (473 mg, 1 mmol) was mixed with hydrazine hydrate monohydrate (250 mg, 5 mmol) and MeOH (10 mL), and the mixture was stirred at 70 °C for 4 h. After the solvent was removed by a rotary evaporator, the solid residue was treated with 1 N NaOH (15 mL), and the resulting mixture was extracted with EtOAc (3 × 30 mL) and dried over anhydrous Na₂SO₄. Evaporation of the solvent under reduced pressure afforded the crude product **S5**, which were used for next step without purification.

Scheme S2. Synthesis of compounds **6A**, **6B** and **6C**:



(2*S*)-3-Bromo-2-hydroxypropan-1-ammonium bromide (**9s**).³

Triphenyl phosphine (6.88 g, 26.25 mmol) was added to a stirred solution of phthalimide (3.86 g, 47 mmol) in THF (40 mL) at 0 °C, followed by (*S*)-(2)-glycidol (2.24 g, 30.3 mmol). Di-tert-butyl azodicarboxylate (6.04 g, 26.25 mmol) was added dropwise and the solution stirred for 5 h at room temperature. The solvent was removed under reduced pressure, dissolved in methanol (20 mL) and cooled to 0 °C. Hydrobromic acid (48% aq., 8.35 mL) was added slowly to the rapidly stirred solution until no further effervescence was observed and a white precipitate formed. The solution was diluted with water (200 mL) and filtered. The aqueous layer extracted with CH₂Cl₂ (3 × 30 mL) and the solvent was removed under reduced pressure. The residue was purified using flash chromatography on silica gel (eluent, *n*-hexane: EtOAc = 10:1 to 5:1) to give compound **9s** as white powder (5.96 g, 80% yield for two steps).

¹H NMR (CDCl₃, 400 MHz): δ 7.85–7.70 (m, 4H), 3.79 (t, *J* = 7.32 Hz, 2H), 2.56–2.48 (m, 2H), 1.99–1.92 (m, 2H), 1.58–1.51 (m, 2H), 1.34–1.24 (m, 32H), 0.87 (t, *J* = 6.40 Hz, 3H); ¹³C NMR (CDCl₃, 100 MHz): δ 168.46, 134.04, 132.19, 123.34, 37.31, 32.16, 32.06, 29.79, 29.71, 29.66, 29.63, 29.46, 29.34, 29.00, 28.55, 22.79, 14.23; HRMS (ESI) *m/z* [M+Na]⁺ calculated for C₁₁H₁₀BrNO₃Na 305.9742, found 305.9733.

(*S*)-2-(3-bromo-2-((tert-butyldimethylsilyl)oxy)propyl)isoindoline-1,3-dione (**10s**).

To a solution of compound **9s** (3.90 g, 13.8 mmol) in CH₂Cl₂ (20 mL) containing imidazole (2.34 g, 34.50 mmol) was added TBSCl (3.10 g, 20.60 mmol) dropwise under N₂ at room temperature. The solution was stirred at room temperature for 8 h, concentrated in vacuo and purified using flash chromatography on silica gel (eluent, *n*-hexane: EtOAc = 20:1 to 10:1) to give compound **10s** as white powder (5.14 g, 94% yield).

¹H NMR (CDCl₃, 400 MHz): δ 7.87–7.70 (m, 4H), 4.25–4.19 (m, 1H), 3.855 (dd, *J* = 2.28, 6.88 Hz, 2H), 3.37 (d, *J* = 5.04 Hz, 3H), 0.82 (s, 9H), 0.08 (s, 3H), -0.08 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz): δ 169.29, 134.22, 132.06, 123.41, 69.38, 42.70, 35.41, 25.66, 17.91, -4.66, -4.86; HRMS (ESI) *m/z* [M+Na]⁺ calculated for C₁₇H₂₄BrNO₃SiNa 420.0607, found 420.0602.

(*S*)-*S*-2-((tert-butyldimethylsilyl)oxy)-3-(1,3-dioxoisoindolin-2-yl)propyl ethanethioate (**11s**):

To a stirred solution of **10s** (4.08 g, 10.25 mmol) in THF (60 mL) was added potassium thioacetate (3.51 g, 30.75 mmol), and the mixture was refluxed for 8 h at 85 °C. And the solvent was evaporated to dryness in vacuo, and then added water (100 mL), and the organic materials were extracted with EtOAc (3 × 30 mL). The combined extracts were washed with water and brine, dried over anhydrous Na₂SO₄, and concentrated in vacuo. The crude product was purified by flash chromatography on silica gel (eluent, *n*-hexane: EtOAc = 10:1 to 5:1) to give compound **11s** as white powder (3.71 g, 92% yield).

¹H NMR (CDCl₃, 400 MHz): δ 7.84–7.68 (m, 4H), 4.16–4.10 (m, 1H), 3.78–3.67 (m, 2H), 3.08–2.96 (m, 2H), 2.32 (s, 3H), 0.80 (s, 9H), 0.05 (s, 3H), -0.11 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz): δ 195.28, 168.32, 134.12, 132.10, 123.34, 68.79, 42.81, 34.19, 30.62, 25.67, 17.86, -4.72, -4.89; HRMS (ESI) *m/z* [M+Na]⁺ calculated for C₁₉H₂₇NO₄SSiNa 416.1328, found 416.1322.

(*S*)-2-(2-((tert-butyldimethylsilyl)oxy)-3-mercaptopropyl)isoindoline-1,3-dione (**12s**):

A solution of **11s** (3.7 g, 9.41 mmol) in anhydrous methanol (35 mL) was first degassed and refilled with N₂. NaOMe (2.5 g, 47 mmol) was added to the solution, and the resulting mixture was stirred at room temperature for 0.5 h under N₂ atmosphere until no more starting material could be detected by TLC. The reaction mixture was quenched with Dowerx 50WX2 hydrogen form and filtered through celite. The organic layer was dried over anhydrous Na₂SO₄ and concentrated in vacuo to give crude product, which was purified by flash chromatography on silica gel (eluent, *n*-hexane: EtOAc = 10:1 to 5:1) to give compound **12s** as white powder (3.10 g, 94% yield).

¹H NMR (CDCl₃, 400 MHz): δ 7.86–7.69 (m, 4H), 4.14–4.09 (m, 1H), 3.91–3.79 (m, 2H), 2.70–2.55 (m, 2H), 0.83 (s, 9H), 0.06 (s, 3H), -0.05 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz): δ 168.40, 134.15, 132.10, 123.37, 70.40, 42.21, 29.88, 25.76, 17.99, -4.63, -4.75; HRMS (ESI) *m/z* [M+Na]⁺ calculated for C₁₇H₂₅NO₃SSiNa 374.1222, found 374.1216.

(*S*)-2-(2-((tert-butyldimethylsilyl)oxy)-3-(octadecylthio)propyl)isoindoline-1,3-dione (**13s**):

1-Bromooctadecane (366 mg, 0.91 mmol) was added to a mixture containing sodium hydride (73 mg, 10.6 mmol) and compound **12s** (320 mg, 1.1 mmol) in dry THF (3 mL). The mixture was stirred at room temperature for 2 h. The reaction mixture was cooled to 0 °C and water (20 mL) was added. The solution was extracted with EtOAc (3 × 30 mL). The organic layer was separated, dried with

anhydrous Na₂SO₄, and evaporated under vacuum. The crude product was purified using flash chromatography on silica gel (eluent, *n*-hexane: EtOAc = 20:1 to 10:1) to give compound **13s** as white powder (493 mg, 90% yield).

¹H NMR (CDCl₃, 400 MHz): δ 7.86–7.68 (m, 4H), 4.19–4.13 (m, 1H), 3.93–3.75 (m, 2H), 2.68–2.53 (m, 4H), 1.59–1.52 (m, 2H), 1.36–1.18 (m, 32H), 0.86 (t, *J* = 6.88 Hz, 3H), 0.79 (s, 9H), 0.04 (s, 3H), -0.13 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz): δ 168.40, 134.03, 132.20, 123.26, 69.30, 43.24, 37.57, 33.12, 32.02, 29.79, 29.69, 29.62, 29.46, 29.34, 28.96, 25.72, 22.78, 17.88, 14.22, -4.56, -4.87; HRMS (ESI) *m/z* [M+Na]⁺ calculated for C₃₅H₆₁NO₃SSiNa 626.4039, found 626.4041.

(*S*)-2-(2-hydroxy-3-(octadecylthio)propyl)isoindoline-1,3-dione (**13s_1**):

To a solution of compound **13s** (90 mg, 0.15 mmol) in THF (4 mL) was added a 1 M solution of tetrabutylammonium fluoride in THF (225 μL, 0.225 mmol) and AcOH (17 μL) at room temperature. The solution was stirred for overnight and diluted with 10 mL of EtOAc solution. The organic layer was separated and washed with saturated NaHCO₃ (3 × 10 mL). The water extract was washed with EtOAc solution (2 × 10 mL), and the organic layers were combined and dried over Na₂SO₄. The solvent was evaporated *in vacuo*, and the residue was chromatographed over silica gel (eluent, *n*-hexane: EtOAc = 10:1 to 5:1) to give **13s_1** as white powder (493 mg, 85% yield).

¹H NMR (CDCl₃, 400 MHz): δ 7.87–7.72 (m, 4H), 4.06–3.99 (m, 1H), 3.89–3.82 (m, 2H), 2.85–2.34 (m, 4H), 1.96 (br, 1H), 1.67–1.53 (m, 2H), 1.43–1.11 (m, 32H), 0.89 (t, *J* = 6.88 Hz, 3H); ¹³C NMR (CDCl₃, 100 MHz): δ 168.72, 134.23, 132.04, 123.56, 68.34, 43.05, 37.31, 32.03, 29.78, 29.62, 29.47, 29.31, 28.90, 22.79, 14.23; HRMS (ESI) *m/z* [M+Na]⁺ calculated for C₂₉H₄₇NO₃SNa 512.3174, found 512.3170.

(*S*)-1-amino-3-(octadecylthio)propan-2-ol (**6A**):

N-Cinnamylphthalimide **13s_1** (473 mg, 1 mmol) was mixed with hydrazine hydrate monohydrate (250 mg, 5 mmol) and MeOH (10 mL), and the mixture was stirred at 70 °C for 4 h. After the solvent was removed by a rotary evaporator, the solid residue was treated with 1 N NaOH (15 mL), and the resulting mixture was extracted with EtOAc (3 × 30 mL) and dried over anhydrous Na₂SO₄. Evaporation of the solvent under reduced pressure afforded the crude product **6A**, which were used for next step without purification.

(*S*)-2-2-((tert-butyldimethylsilyloxy)-3-(octadecylsulfonyl)propyl)isoindoline-1,3-dione (**14s**) and 2-((2*S*)-2-((tert-butyldimethylsilyloxy)-3-(octadecylsulfinyl)propyl)isoindoline-1,3-dione (**15s**):

To **13s** (416 mg, 0.69 mmol) in DCM (20 mL) at room temperature was added OXONE (1.27 g, 1.04 mmol). The resulting cloudy white reaction mixture was stirred vigorously at room temperature for 8 h and then poured onto brine and the layers separated. The aqueous phase was extracted with EtOAc and the combined organic extracts dried over Na₂SO₄. The solvent was evaporated *in vacuo*, and the residue was chromatographed over silica gel (eluent, *n*-hexane: EtOAc = 10:1 to 5:1) to give **14s** (280 mg, 64% yield) as white powder and **15s** (128 mg, 30% yield) as white powder. Data for sulfone **14s**: ¹H NMR (CDCl₃, 400 MHz): δ 7.85–7.70 (m, 4H), 4.57–4.51 (m, 1H), 3.81 (d, *J* = 6.84 Hz, 2H), 3.24–2.99 (m, 4H), 1.84–1.76 (m, 2H), 1.38–1.23 (m, 30H), 0.88 (s, 9H), 0.86 (t, *J* = 6.88 Hz, 3H), 0.18 (s, 3H), 0.16 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz): δ 168.20, 134.40, 131.88, 123.58, 65.67, 58.08, 54.97, 43.09, 32.01, 29.79, 29.67, 29.59, 29.45, 29.35, 29.19, 28.53, 25.80, 22.78, 21.68, 17.89, 14.22, -4.59, -4.70; HRMS (ESI) *m/z* [M+Na]⁺ calculated for C₃₅H₆₁NO₅SSiNa 658.3937, found 658.3932.

Data for s sulfoxide **15s**: ¹H NMR (CDCl₃, 400 MHz): δ 7.86–7.68 (m, 4H), 4.48–4.42 (m, 1H), 3.94–3.78 (m, 2H), 2.95–2.65 (m, 4H), 1.76–1.71 (m, 2H), 1.41–1.23 (m, 32H), 0.90–0.83 (m, 12H), 0.20 (s, 3H), 0.19 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz): δ 168.23, 134.33, 131.97, 123.52, 65.16, 59.53, 53.31, 43.41, 32.01, 29.79, 29.61, 29.45, 29.25, 28.88, 25.82, 25.72, 22.86, 22.78, 18.10, 14.22, -4.42, -4.86; HRMS (ESI) m/z [M+Na]⁺ calculated for C₃₅H₆₁NO₄SSiNa 642.3988, found 642.3981.

(*S*)-2-(2-hydroxy-3-(octadecylsulfonyl)propyl)isoindoline-1,3-dione (**14s_1**):

To a solution of compound **14s** (230 mg, 0.362 mmol) in THF (7 mL) was added a 1 M solution of tetrabutylammonium fluoride in THF (1.1 mL, 0.225 mmol) and AcOH (84 μL) at room temperature. The solution was stirred for overnight and diluted with 10 mL of EtOAc solution. The organic layer was separated and washed with saturated NaHCO₃ (3 × 10 mL). The water extract was washed with EtOAc solution (2 × 10 mL), and the organic layers were combined and dried over Na₂SO₄. The solvent was evaporated *in vacuo*, and the residue was chromatographed over silica gel (eluent, *n*-hexane: EtOAc = 5:1 to 2:1) to give **14s_1** as white powder (166 mg, 88% yield).

¹H NMR (CDCl₃, 400 MHz): δ 7.86–7.75 (m, 4H), 4.60–4.57 (m, 1H), 3.88 (t, *J* = 5.48 Hz, 2H), 3.26–3.05 (m, 4H), 1.82–1.72 (m, 2H), 1.41–1.24 (m, 30H), 0.86 (t, *J* = 6.88 Hz, 3H); ¹³C NMR (CDCl₃, 100 MHz): δ 168.68, 134.52, 131.80, 123.76, 65.61, 57.01, 54.87, 43.20, 32.02, 29.79, 29.69, 29.60, 29.46, 29.37, 29.14, 28.51, 22.78, 21.81, 14.23; HRMS (ESI) m/z [M+Na]⁺ calculated for C₂₉H₄₇NO₅SNa 544.3073, found 544.3067.

(*S*)-1-amino-3-(octadecylsulfonyl)propan-2-ol (**6B**):

N-Cinnamylphthalimide **14s_1** (125 mg, 1 mmol) was mixed with hydrazine hydrate monohydrate (72 μL) and EtOH (5 mL), and the mixture was stirred at 80 °C for 4 h. After the solvent was removed by a rotary evaporator, the solid residue was treated with 1 N NaOH (15 mL), and the resulting mixture was extracted with EtOAc (3 × 30 mL) and dried over anhydrous Na₂SO₄. Evaporation of the solvent under reduced pressure afforded the crude product **6B**, which were used for next step without purification.

2-((2*S*)-2-hydroxy-3-(octadecylsulfinyl)propyl)isoindoline-1,3-dione (**15s_1**):

To a solution of compound **15s** (130 mg, 0.21 mmol) in THF (4 mL) was added a 1 M solution of tetrabutylammonium fluoride in THF (630 μL, 0.63 mmol) and AcOH (97 μL) at room temperature. The solution was stirred for overnight and diluted with 10 mL of EtOAc solution. The organic layer was separated and washed with saturated NaHCO₃ (3 × 10 mL). The water extract was washed with EtOAc solution (2 × 10 mL), and the organic layers were combined and dried over Na₂SO₄. The solvent was evaporated *in vacuo*, and the residue was chromatographed over silica gel (eluent, *n*-hexane: EtOAc = 5:1 to 2:1) to give **15s_1** as white powder (90 mg, 85% yield).

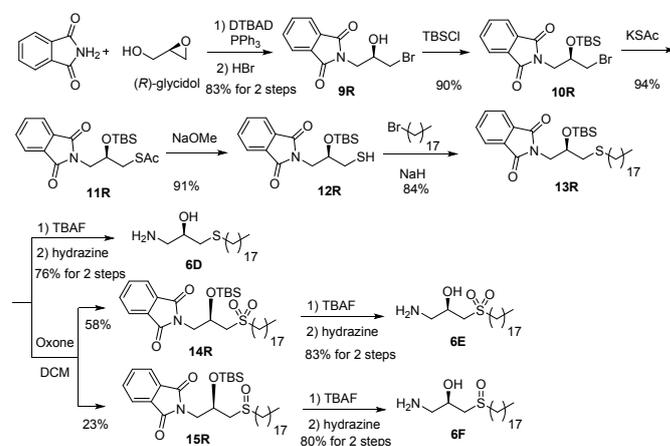
¹H NMR (CDCl₃, 400 MHz): δ 7.86–7.71 (m, 4H), 4.63–4.57 (m, 1H), 3.28 (br, 2H), 2.89–2.68 (m, 4H), 1.82–1.72 (m, 2H), 1.47–1.24 (m, 30H), 0.86 (t, *J* = 6.88 Hz, 3H); ¹³C NMR (CDCl₃, 100 MHz): δ 168.66, 134.34, 131.96, 123.63, 67.07, 65.27, 54.19, 53.67, 43.56, 43.30, 32.01, 29.79, 29.62, 29.44, 29.25, 28.85, 22.77, 22.43, 14.21; HRMS (ESI) m/z [M+Na]⁺ calculated for C₂₉H₄₇NO₄SNa 528.3123, found 528.3129.

(2*S*)-1-amino-3-(octadecylsulfinyl)propan-2-ol (**6C**):

N-Cinnamylphthalimide **15s_1** (75 mg, 0.15 mmol) was mixed with hydrazine hydrate monohydrate (90 μL) and EtOH (10 mL), and the mixture was stirred at 80 °C for overnight. After the solvent was

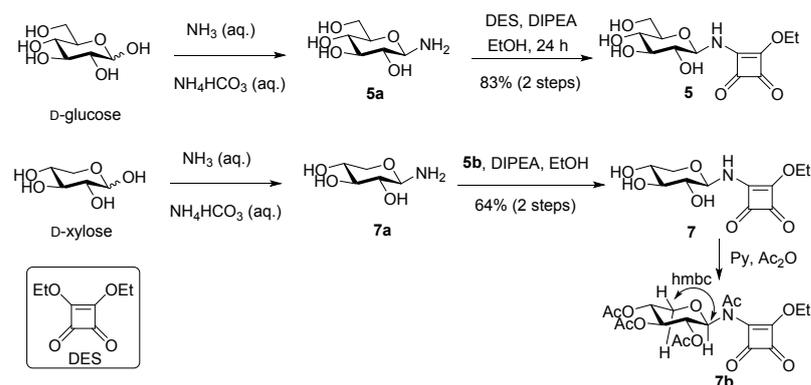
removed by a rotary evaporator, the solid residue was treated with 1 N NaOH (15 mL), and the resulting mixture was extracted with EtOAc (3 × 30 mL) and dried over anhydrous Na₂SO₄. Evaporation of the solvent under reduced pressure afforded the crude product **6C**, which were used for next step without purification.

Scheme S3. Synthesis of compounds **6D**, **6E** and **6F**:



In a similar manner, compounds **6D**, **6E** and **6F** were prepared from (*R*)-glycidol as starting material.

Scheme S4. Synthesis of sugar squaramides:⁴



3-ethoxy-4-(glucosylamino)cyclobut-3-ene-1,2-dione **5**.

To an aqueous solution of D-glucose (2.5 g, 13.9 mmol) containing NH₄HCO₃ (1.1 g), commercial aq. NH₃ (68 mL) was added. This solution was heated during 36 h at 42 °C, concentrated to half volume, and then freeze-dried, which were used for next step without purification.

DIPEA (1.23 mL, 6.95 mmol) was added dropwise to a vigorously stirred solution of diethyl squarate (DES; 3.07 mL, 20.85 mmol) and crude glycosylamine **5a** in EtOH (30 mL). Then the reaction mixture was stirred at room temperature for 12 hours. Evaporation of the solvent under reduced pressure afforded the crude product, which was purified by flash chromatography on silica gel (eluent, CHCl₃: MeOH = 10:1 to 5:1) to give compound **5** as yellow powder (3.5 g, 83% yield for two steps).

¹H NMR (CD₃OD, 400 MHz): δ 4.75–4.70 (m, 2H), 3.84–3.62 (m, 2H), 3.40–3.33 (m, 5H), 1.44 (t, *J* = 7.36 Hz, 3H); ¹³C NMR (CD₃OD, 100 MHz): δ 193.5, 188.95, 178.03, 174.04, 78.64, 77.28, 73.09, 69.83, 69.75, 61.23, 58.57, 14.78; HRMS (ESI) *m/z* [M+Na]⁺ calculated for C₁₂H₁₇NO₈Na 326.0852, found 326.0848.

3-ethoxy-4-(xylosylamino)cyclobut-3-ene-1,2-dione **7**.

In a similar manner, compound **7** was prepared from D-xylose as starting material (63% yield for two steps).

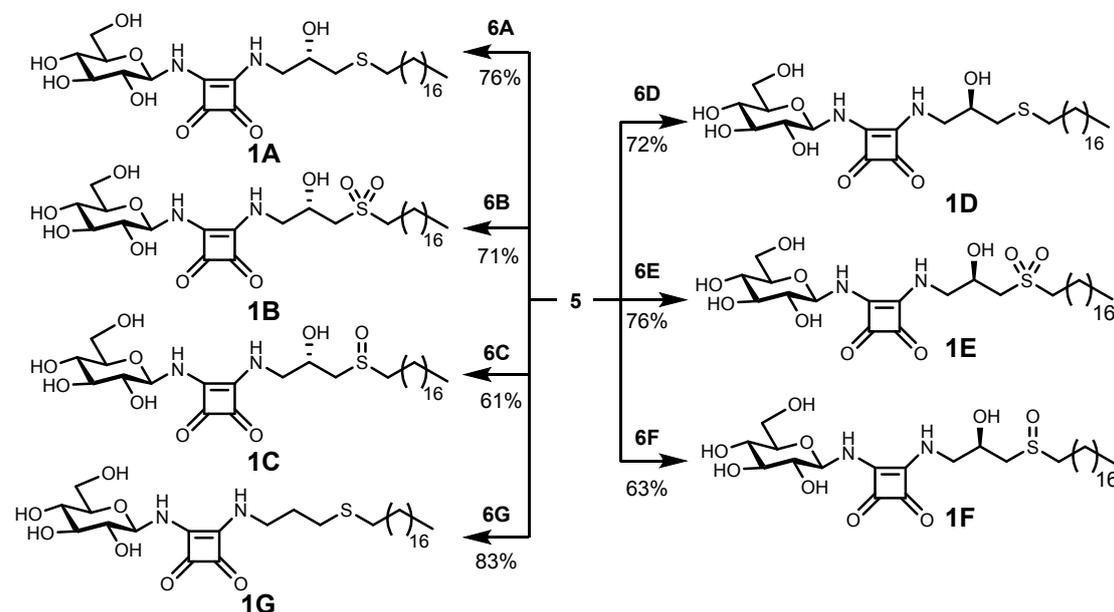
^1H NMR (CD_3OD , 400 MHz): δ 4.78–4.70 (m, 2H), 3.85 (dd, $J = 11.44, 5.48$ Hz, 1H), 3.53–3.47 (m, 1H), 3.36–3.26 (m, 4H), 1.44 (t, $J = 6.88$ Hz, 3H); ^{13}C NMR (CD_3OD , 100 MHz): δ 188.14, 184.69, 178.07, 173.97, 85.21, 84.49, 77.30, 72.97, 69.79, 69.53, 67.38, 14.78; HRMS (ESI) m/z $[\text{M}+\text{Na}]^+$ calculated for $\text{C}_{11}\text{H}_{15}\text{NO}_7\text{Na}$ 296.0746, found 296.0747.

(2R,3R,4S,5R)-2-(N-(2-ethoxy-3,4-dioxocyclobut-1-en-1-yl)acetamido)tetrahydro-2H-pyran-3,4,5-triyl triacetate **7b**

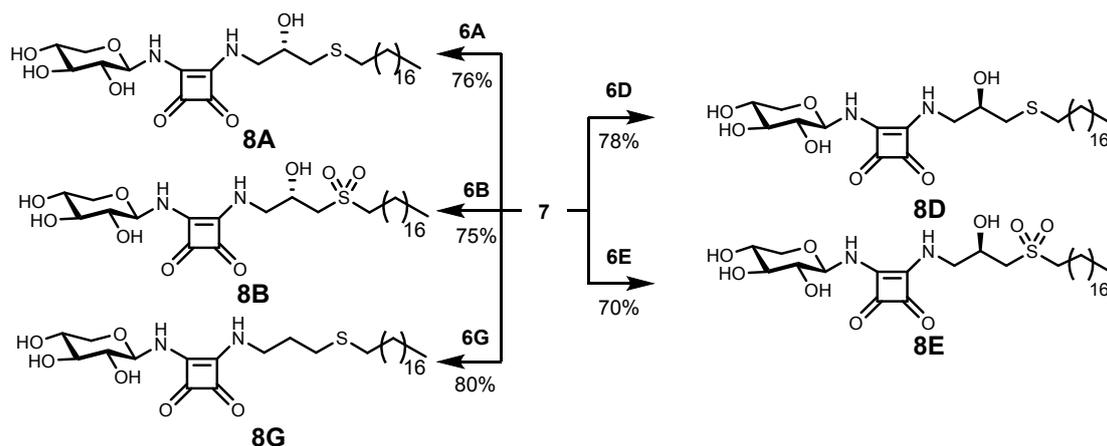
To further confirm the conformation of compound **7**, fully protected acetyl group of compound **7b** was prepared: Ac_2O (0.5 mL) was added dropwise to a vigorously stirred solution of compound **7** (100 mg, 0.36 mmol) in pyridine (1 mL). Then the reaction mixture was stirred at room temperature for 12 hours. Evaporation of the solvent under reduced pressure afforded the crude product, which was purified by flash chromatography on silica gel (eluent, *n*-hexane: EtOAc = 5:1 to 2:1) to give compound **7b** as yellow powder (118 mg, 73% yield).

^1H NMR (CDCl_3 , 400 MHz): δ 5.78 (d, $J = 9.64$ Hz, 1H, H-1), 5.45 (t, $J = 9.64$ Hz, 1H, H-2), 5.31 (t, $J = 9.60$ Hz, 1H, H-3), 5.08–4.97 (m, 3H, H-4 and CH_2), 4.14 (dd, $J = 11.48, 5.52$ Hz, 1H, H-5a), 3.44 (t, $J = 11.00$ Hz, 1H, H-5e), 2.36 (s, 3H, NAc), 2.04 (s, 3H, OAc), 2.03 (s, 3H, OAc), 1.96 (s, 3H, OAc), 1.57 (t, $J = 7.32$ Hz, 3H); ^{13}C NMR (CDCl_3 , 100 MHz): δ 189.52, 187.10, 186.40, 170.30, 170.01, 169.91, 169.13, 82.37, 72.47, 72.00, 70.11, 68.22, 65.29, 24.06, 20.72, 20.40, 15.88; HRMS (ESI) m/z $[\text{M}+\text{Na}]^+$ calculated for $\text{C}_{19}\text{H}_{23}\text{NO}_{11}\text{Na}$ 464.1169, found 464.11687.

Scheme S5. Synthesis of squaramide analogues:



Scheme S6. Synthesis of squaramide analogues:



General synthetic procedure for the synthesis of squaramide analogues **1A-1G**.

DIPEA (0.5 equiv.) was added dropwise to a vigorously stirred solution of glycosylamine (1 equiv.) and the appropriate amine (1.2 equiv.) in EtOH (20 mL/mmol). Then the reaction mixture was stirred at 40 °C for 12-36 hours. Evaporation of the solvent under reduced pressure afforded the crude product, which was purified by flash chromatography on silica gel (eluent, CHCl₃: MeOH = 10:1 to 5:1) to give title compounds **1A-1G** as white powder.

3-(((S)-2-hydroxy-3-(octadecylthio)propyl)amino)-4-(glucosylamino)cyclobut-3-ene-1,2-dione 1A: Following the general procedure, starting with **3** (15.0 mg, 0.05 mmol), **6A** (22.0 mg, 0.06 mmol) and DIPEA (5 μL, 0.025 mmol) in 1 mL of dry ethanol (reaction time: 24 h), compound **1A** was obtained as white powder (23 mg, 76% yield).

¹H NMR (pyridine-d₅, 400 MHz): δ 4.42–3.88 (m, 7H), 3.62–3.55 (m, 1H), 2.86 (d, *J* = 6.44 Hz, 2H), 2.55 (t, *J* = 7.36 Hz, 2H), 1.51–1.44 (m, 2H), 1.29–1.06 (m, 32H), 0.76 (t, *J* = 6.40 Hz, 3H); ¹³C NMR (pyridine-d₅, 100 MHz): δ 185.82, 183.95, 170.08, 169.35, 85.67, 80.08, 78.52, 75.07, 70.93, 70.59, 61.64, 49.37, 37.23, 32.79, 32.92, 29.97, 29.87, 29.71, 29.68, 29.51, 29.40, 29.37, 29.29, 22.84, 14.19; HRMS (ESI) *m/z* [M+Na]⁺ calculated for C₃₁H₅₆N₂O₈SNa 639.3655, found 639.3663.

3-(((S)-2-hydroxy-3-(octadecylsulfonyl)propyl)amino)-4-(glucosylamino)cyclobut-3-ene-1,2-dione 1B:

Following the general procedure, starting with **3** (10.0 mg, 0.033 mmol), **6B** (16.0 mg, 0.04 mmol) and DIPEA (3 μL, 0.017 mmol) in 1 mL of dry ethanol (reaction time: 36 h), compound **1B** was obtained as white powder (15 mg, 71% yield).

¹H NMR (pyridine-d₅, 400 MHz): δ 4.40–4.01 (m, 7H), 3.77–3.1 (m, 1H), 3.62–3.41 (m, 4H), 1.93–1.85 (m, 2H), 1.38–1.06 (m, 32H), 0.76 (t, *J* = 6.40 Hz, 3H); ¹³C NMR (pyridine-d₅, 100 MHz): δ 185.44, 183.80, 169.69, 168.82, 85.69, 80.03, 78.16, 75.11, 74.11, 70.49, 66.76, 57.42, 54.90, 49.44, 31.90, 29.78, 29.70, 29.61, 29.39, 29.20, 28.54, 22.73, 21.96, 14.10; HRMS (ESI) *m/z* [M+Na]⁺ calculated for C₃₁H₅₆N₂O₁₀SNa 671.3553, found 671.3546.

3-(((2S)-2-hydroxy-3-(octadecylsulfinyl)propyl)amino)-4-(glucosylamino)cyclobut-3-ene-1,2-dione 1C:

Following the general procedure, starting with **3** (10.0 mg, 0.033 mmol), **6C** (15.0 mg, 0.04 mmol) and DIPEA (3 μL, 0.017 mmol) in 1 mL of dry ethanol (reaction time: 36 h), compound **1C** was obtained as white powder (12 mg, 61% yield).

¹H NMR (pyridine-d₅, 400 MHz): δ 4.71–4.78 (m, 1H), 4.42–4.06 (m, 7H), 3.31–3.18 (m, 2H), 2.94–2.77 (m, 2H), 1.62–1.74 (m, 2H), 1.25–1.09 (m, 32H), 0.76 (t, *J* = 6.88 Hz, 3H); ¹³C NMR

(pyridine- d_5 , 100 MHz): δ 185.49, 183.88, 169.68, 168.79, 85.55, 79.90, 78.43, 74.97, 70.67, 66.46, 52.36, 49.52, 31.91, 29.79, 29.71, 29.52, 29.39, 28.85, 22.99, 22.73, 14.10; HRMS (ESI) m/z $[M+Na]^+$ calculated for $C_{31}H_{56}N_2O_9SNa$ 655.3604, found 655.3605.

3-(((R)-2-hydroxy-3-(octadecylthio)propyl)amino)-4-(glucosylamino)cyclobut-3-ene-1,2-dione **1D**:

Following the general procedure, starting with **3** (15.0 mg, 0.05 mmol), **6D** (22.0 mg, 0.06 mmol) and DIPEA (5 μ L, 0.025 mmol) in 1 mL of dry ethanol (reaction time: 24 h), compound **1D** was obtained as white powder (22 mg, 72% yield).

1H NMR (pyridine- d_5 , 400 MHz): δ 4.38–3.88 (m, 7H), 3.70–3.37 (m, 1H), 2.95–2.48 (m, 4H), 1.53–1.41 (m, 2H), 1.29–1.06 (m, 32H), 0.79 (t, $J = 6.88$ Hz, 3H); ^{13}C NMR (pyridine- d_5 , 100 MHz): δ 185.91, 184.24, 170.17, 168.97, 85.97, 80.29, 78.66, 75.37, 71.04, 70.61, 49.46, 37.36, 32.89, 31.95, 29.83, 29.76, 29.72, 29.44, 29.40, 29.00, 22.77, 14.12; HRMS (ESI) m/z $[M+Na]^+$ calculated for $C_{31}H_{56}N_2O_8SNa$ 639.3655, found 639.3658.

3-(((R)-2-hydroxy-3-(octadecylsulfonyl)propyl)amino)-4-(glucosylamino)cyclobut-3-ene-1,2-dione **1E**:

Following the general procedure, starting with **3** (10.0 mg, 0.033 mmol), **6E** (16.0 mg, 0.04 mmol) and DIPEA (3 μ L, 0.017 mmol) in 1 mL of dry ethanol (reaction time: 36 h), compound **1E** was obtained as white powder (16 mg, 76% yield).

1H NMR (pyridine- d_5 , 400 MHz): δ 4.46–4.04 (m, 7H), 3.75–3.69 (m, 1H), 3.60–3.42 (m, 4H), 1.92–1.88 (m, 2H), 1.29–1.09 (m, 32H), 0.80 (t, $J = 7.32$ Hz, 3H); ^{13}C NMR (pyridine- d_5 , 100 MHz): δ 186.03, 184.05, 169.97, 169.09, 86.03, 80.30, 78.57, 75.38, 70.64, 66.91, 61.36, 57.64, 54.96, 49.64, 31.94, 29.82, 29.74, 29.65, 29.43, 29.23, 22.76, 22.06, 14.10; HRMS (ESI) m/z $[M+Na]^+$ calculated for $C_{31}H_{56}N_2O_{10}SNa$ 671.3553, found 671.3568.

3-(((2R)-2-hydroxy-3-(octadecylsulfinyl)propyl)amino)-4-(glucosylamino)cyclobut-3-ene-1,2-dione **1F**:

Following the general procedure, starting with **3** (10.0 mg, 0.033 mmol), **6F** (15.0 mg, 0.04 mmol) and DIPEA (3 μ L, 0.017 mmol) in 1 mL of dry ethanol (reaction time: 36 h), compound **1F** was obtained as white powder (13 mg, 63% yield).

1H NMR (pyridine- d_5 , 400 MHz): δ 4.80–4.74 (m, 1H), 4.40–4.04 (m, 7H), 3.32–3.09 (m, 2H), 2.96–2.68 (m, 2H), 1.68–1.56 (m, 2H), 1.38–1.05 (m, 32H), 0.76 (t, $J = 6.88$ Hz, 3H); ^{13}C NMR (pyridine- d_5 , 100 MHz): δ 185.74, 183.80, 169.89, 168.57, 85.51, 79.87, 78.33, 74.97, 70.67, 66.57, 66.51, 57.93, 52.92, 52.41, 49.90, 49.16, 31.90, 29.79, 29.70, 29.51, 29.39, 28.86, 23.06, 22.97, 22.72, 14.10; HRMS (ESI) m/z $[M+Na]^+$ calculated for $C_{31}H_{56}N_2O_9SNa$ 655.3604, found 655.3596.

3-(((3-(octadecylthio)propyl)amino)-4-(glucosylamino)cyclobut-3-ene-1,2-dione **1G**:

Following the general procedure, starting with **3** (61.0 mg, 0.2 mmol), **6G** (82.0 mg, 0.24 mmol) and DIPEA (18 μ L, 0.1 mmol) in 4 mL of dry ethanol (reaction time: 18 h), compound **1G** was obtained as white powder (99 mg, 83% yield).

1H NMR (pyridine- d_5 , 400 MHz): δ 4.38–3.79 (m, 7H), 2.90–2.88 (m, 1H), 2.58–2.33 (m, 4H), 1.95–1.80 (m, 2H), 1.53–1.42 (m, 2H), 1.29–1.06 (m, 32H), 0.76 (t, $J = 6.88$ Hz, 3H); ^{13}C NMR (pyridine- d_5 , 100 MHz): δ 185.33, 183.84, 169.75, 168.53, 85.65, 79.90, 78.32, 75.05, 70.59, 61.05, 43.20, 40.57, 32.12, 31.90, 31.63, 29.78, 29.70, 29.39, 29.36, 29.00, 28.96, 22.73, 14.10; HRMS (ESI) m/z $[M+Na]^+$ calculated for $C_{31}H_{56}N_2O_7SNa$ 623.3706, found 623.3698.

3-(((S)-2-hydroxy-3-(octadecylthio)propyl)amino)-4-(xylosylamino)cyclobut-3-ene-1,2-dione **8A**:

Following the general procedure, starting with **7** (13.7 mg, 0.05 mmol), **6A** (22.0 mg, 0.06 mmol) and DIPEA (5 μ L, 0.025 mmol) in 1 mL of dry ethanol (reaction time: 24 h), compound **8A** was obtained as white powder (22 mg, 76% yield).

^1H NMR (pyridine- d_5 , 400 MHz): δ 4.27–3.79 (m, 6H), 2.90–2.83 (m, 2H), 2.58–2.46 (m, 3H), 1.50–1.44 (m, 2H), 1.29–1.06 (m, 32H), 0.76 (t, $J = 6.88$ Hz, 3H); ^{13}C NMR (pyridine- d_5 , 100 MHz): δ 185.26, 183.59, 169.73, 169.01, 85.89, 78.54, 74.53, 70.62, 70.56, 68.51, 49.25, 40.50, 36.86, 32.80, 31.90, 29.79, 29.71, 29.65, 29.39, 29.36, 28.96, 22.73, 14.10; HRMS (ESI) m/z $[\text{M}+\text{Na}]^+$ calculated for $\text{C}_{30}\text{H}_{54}\text{N}_2\text{O}_7\text{SNa}$ 609.3549, found 609.3544.

3-(((S)-2-hydroxy-3-(octadecylsulfonyl)propyl)amino)-4-(xylosylamino)cyclobut-3-ene-1,2-dione **8B**:

Following the general procedure, starting with **7** (9.0 mg, 0.033 mmol), **6B** (16.0 mg, 0.04 mmol) and DIPEA (3 μ L, 0.017 mmol) in 1 mL of dry ethanol (reaction time: 36 h), compound **8B** was obtained as white powder (15 mg, 75% yield).

^1H NMR (pyridine- d_5 , 400 MHz): δ 4.73–4.84 (m, 1H), 4.23–4.31 (m, 1H), 4.18–3.85 (m, 5H), 3.81–3.67 (m, 2H), 3.56–3.51 (m, 1H), 3.44–3.39 (m, 2H), 1.96–1.88 (m, 2H), 1.35–1.06 (m, 32H), 0.78 (t, $J = 7.32$ Hz, 3H); ^{13}C NMR (pyridine- d_5 , 100 MHz): δ 184.96, 183.82, 169.74, 169.52, 85.99, 78.60, 74.00, 70.67, 68.55, 66.47, 57.21, 54.94, 49.68, 40.25, 31.92, 29.80, 29.72, 29.63, 29.41, 29.21, 28.57, 22.74, 22.01, 14.09; HRMS (ESI) m/z $[\text{M}+\text{Na}]^+$ calculated for $\text{C}_{30}\text{H}_{54}\text{N}_2\text{O}_9\text{SNa}$ 641.3448, found 641.3435.

3-((3-(octadecylthio)propyl)amino)-4-(xylosylamino)cyclobut-3-ene-1,2-dione **8G**:

Following the general procedure, starting with **7** (55.0 mg, 0.2 mmol), **6G** (82.0 mg, 0.24 mmol) and DIPEA (18 μ L, 0.1 mmol) in 4 mL of dry ethanol (reaction time: 24 h), compound **8G** was obtained as white powder (91 mg, 80% yield).

^1H NMR (pyridine- d_5 , 400 MHz): δ 4.28–3.71 (m, 6H), 2.51 (t, $J = 7.32$ Hz, 2H), 2.38 (t, $J = 7.76$ Hz, 2H), 1.88–1.81 (m, 2H), 1.48–1.41 (m, 2H), 1.27–1.08 (m, 32H), 0.76 (t, $J = 6.88$ Hz, 3H); ^{13}C NMR (pyridine- d_5 , 100 MHz): δ 185.43, 183.98, 169.56, 169.08, 86.28, 78.78, 74.72, 70.78, 68.61, 43.25, 31.95, 31.90, 31.46, 29.83, 29.76, 29.71, 29.44, 29.38, 28.99, 22.80, 22.77, 14.11; HRMS (ESI) m/z $[\text{M}+\text{Na}]^+$ calculated for $\text{C}_{30}\text{H}_{54}\text{N}_2\text{O}_6\text{SNa}$ 593.3600, found 593.3589.

3-(((R)-2-hydroxy-3-(octadecylthio)propyl)amino)-4-(xylosylamino)cyclobut-3-ene-1,2-dione **8D**:

Following the general procedure, starting with **7** (13.7 mg, 0.05 mmol), **6D** (22.0 mg, 0.06 mmol) and DIPEA (5 μ L, 0.025 mmol) in 1 mL of dry ethanol (reaction time: 24 h), compound **8D** was obtained as white powder (23 mg, 78% yield).

^1H NMR (pyridine- d_5 , 400 MHz): δ 4.29–4.04 (m, 6H), 3.84–3.82 (m, 2H), 2.86–2.78 (m, 2H), 2.55–2.51 (m, 2H), 1.51–1.44 (m, 2H), 1.29–1.06 (m, 32H), 0.76 (t, $J = 6.88$ Hz, 3H); ^{13}C NMR (pyridine- d_5 , 100 MHz): δ 185.26, 183.61, 169.56, 168.96, 85.87, 78.56, 74.63, 70.60, 70.48, 68.53, 49.18, 36.88, 32.79, 31.90, 29.79, 29.71, 29.66, 29.39, 29.35, 28.96, 22.73, 14.10; HRMS (ESI) m/z $[\text{M}+\text{Na}]^+$ calculated for $\text{C}_{30}\text{H}_{54}\text{N}_2\text{O}_7\text{SNa}$ 609.3549, found 609.3549.

3-(((R)-2-hydroxy-3-(octadecylsulfonyl)propyl)amino)-4-(xylosylamino)cyclobut-3-ene-1,2-dione **8E**:

Following the general procedure, starting with **7** (9.0 mg, 0.033 mmol), **6E** (16.0 mg, 0.04 mmol) and DIPEA (3 μ L, 0.017 mmol) in 1 mL of dry ethanol (reaction time: 36 h), compound **8E** was obtained as white powder (14 mg, 70% yield).

^1H NMR (pyridine- d_5 , 400 MHz): δ 4.78–4.84 (m, 1H), 4.27–3.85 (m, 5H), 3.81–3.72 (m, 2H), 3.69–3.56 (m, 1H), 3.44–3.39 (m, 2H), 1.93–1.87 (m, 2H), 1.35–1.06 (m, 32H), 0.78 (t, $J = 7.32$

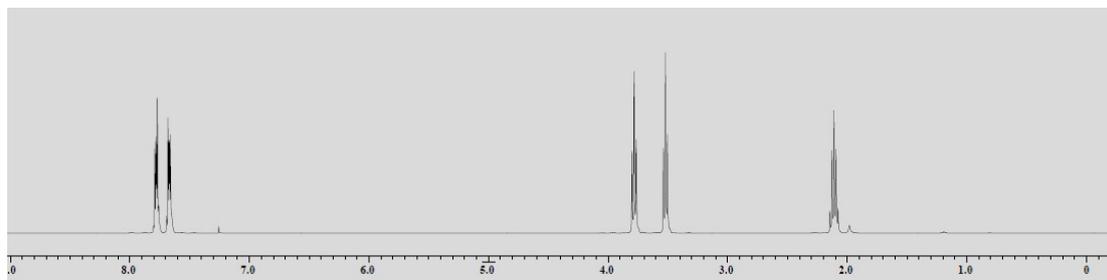
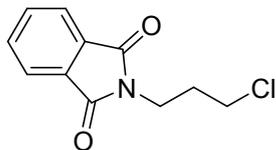
Hz, 3H); ¹³C NMR (pyridine-d₅, 100 MHz): δ 185.43, 183.05, 169.46, 169.08, 85.88, 78.50, 74.54, 70.60, 68.48, 66.50, 57.24, 54.94, 49.64, 31.90, 29.77, 29.69, 29.61, 29.38, 29.19, 28.55, 22.72, 21.93, 14.08; HRMS (ESI) m/z [M+Na]⁺ calculated for C₃₀H₅₄N₂O₉SNa 641.3448, found 641.3459.

References:

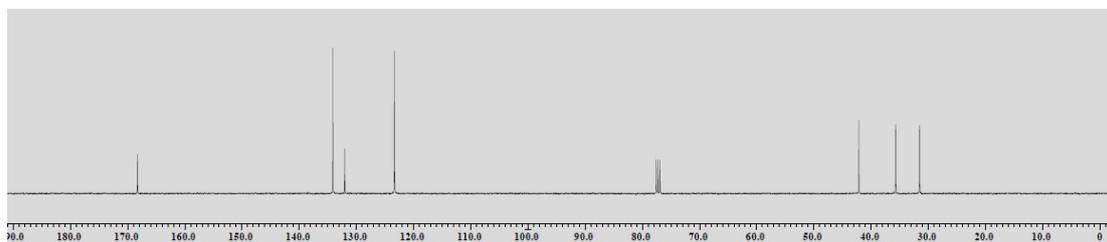
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2. K. Hu, Y. Qi, J. Zhao, H. Jiang, X. Chen, J. Ren, *Eur. J. Med. Chem.* **2013**, *64*, 529-539.
3. a) Ball, D. H. Williams, J. M. Long, L., *Jr. J. Org. Chem.* **1963**, *28*, 1589. b) C. I. Harding, D. J. Dixon, S. V. Ley *Tetrahedron* **2004**, *60*, 7679–7692.
4. A. Lubineau, J. Aug, B. Drouillat, *Carbohydrate Research* **1995**, *266*, 211-219.

NMR spectra of all new compounds

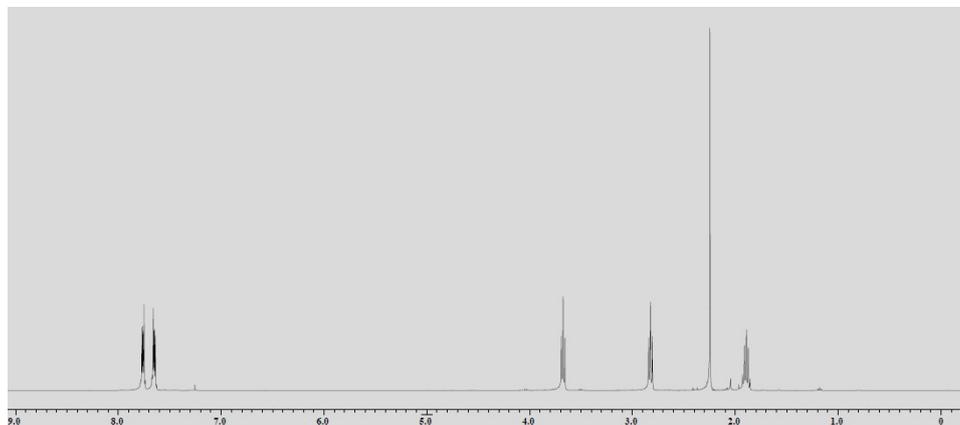
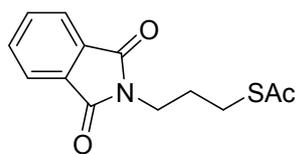
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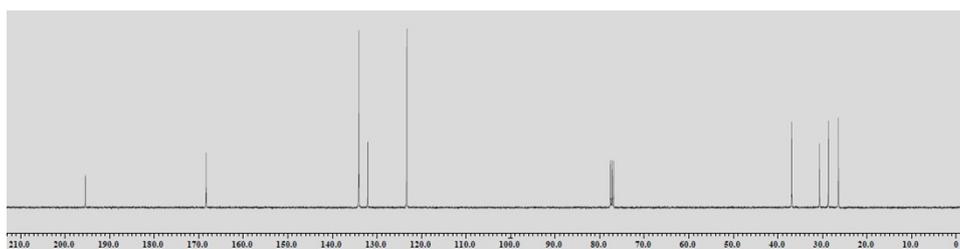
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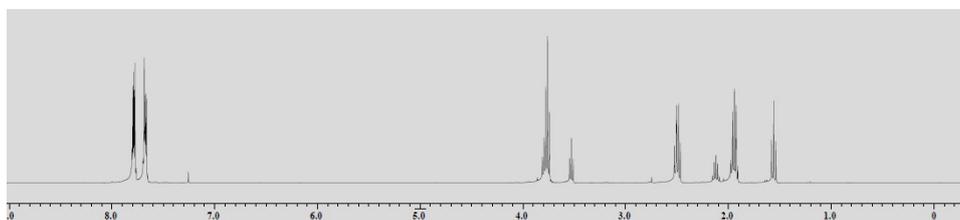
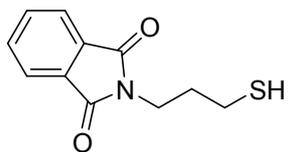
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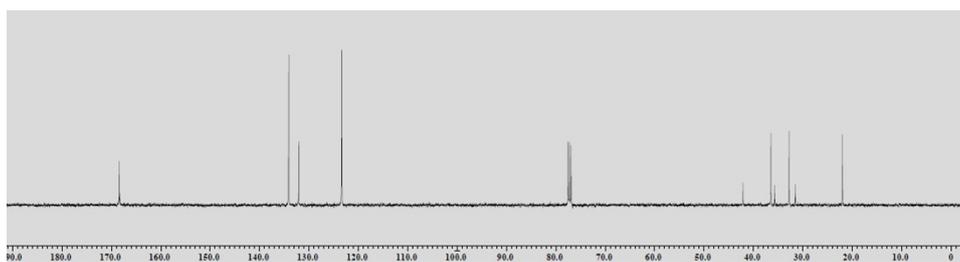
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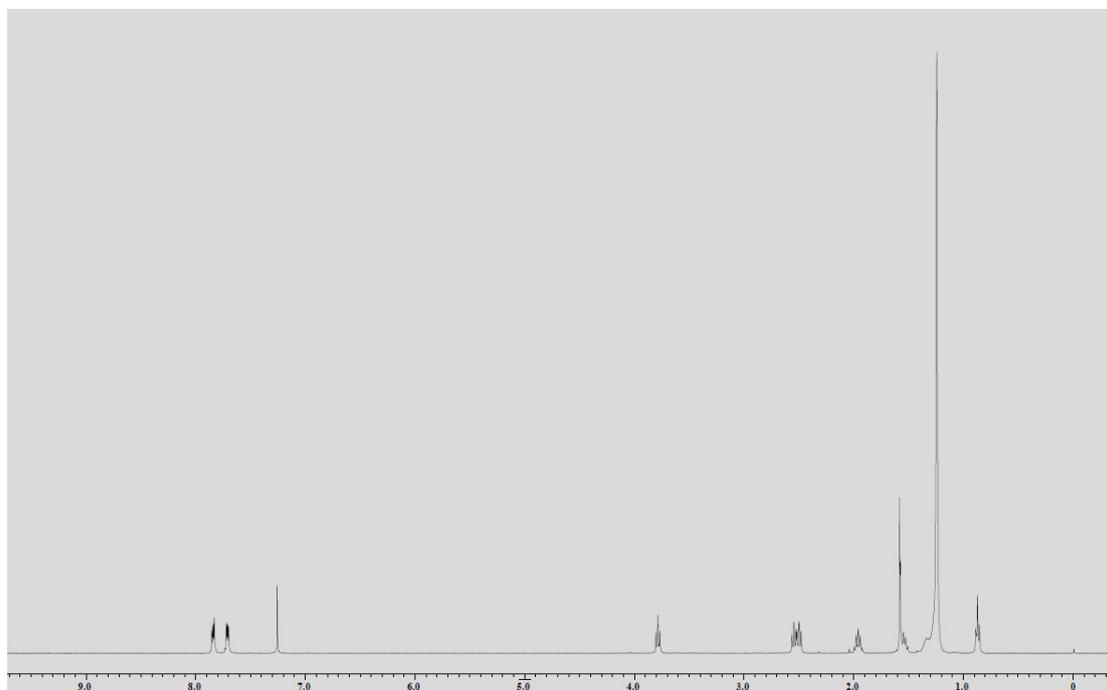
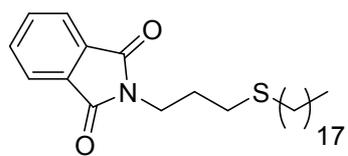
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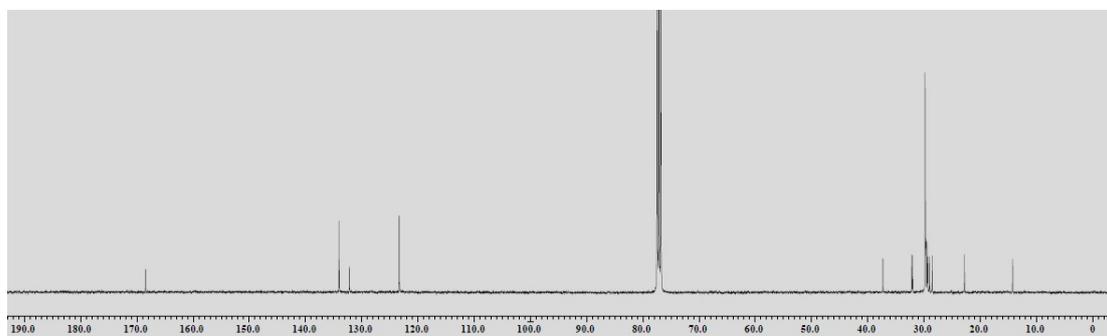
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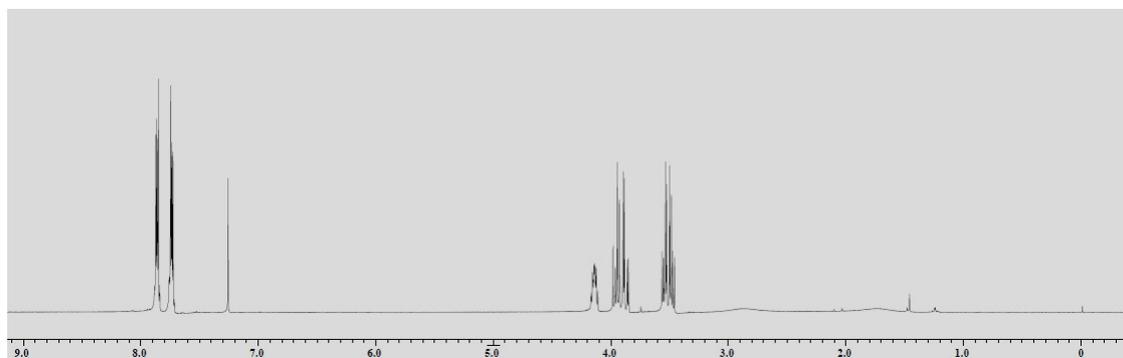
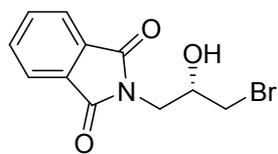
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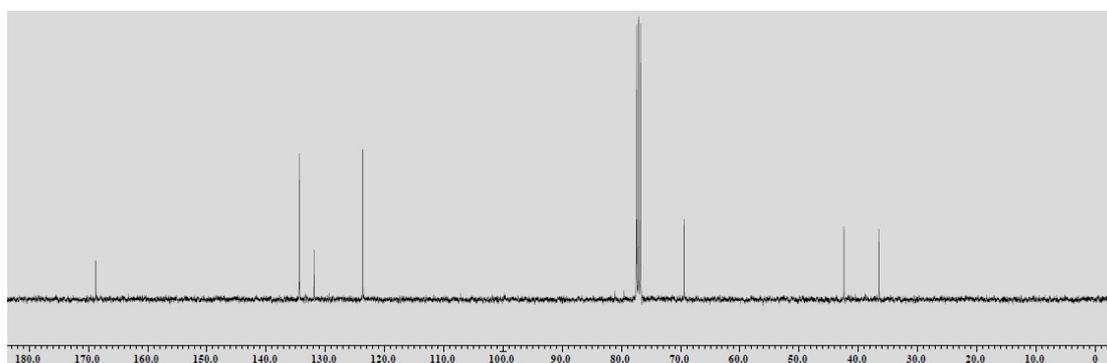
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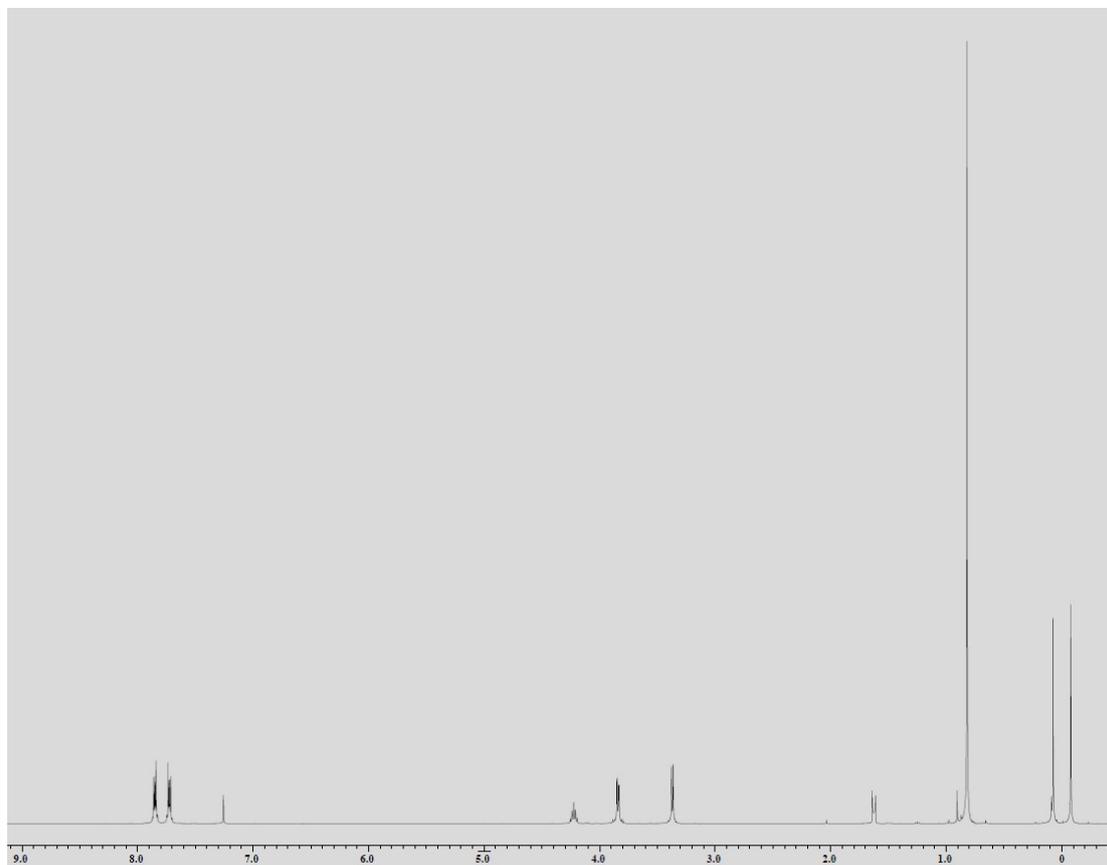
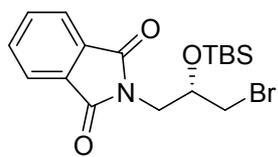
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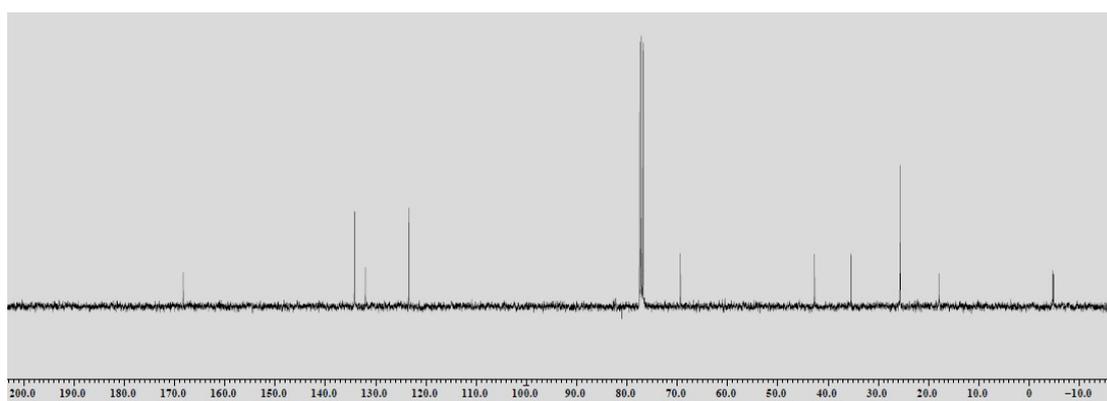
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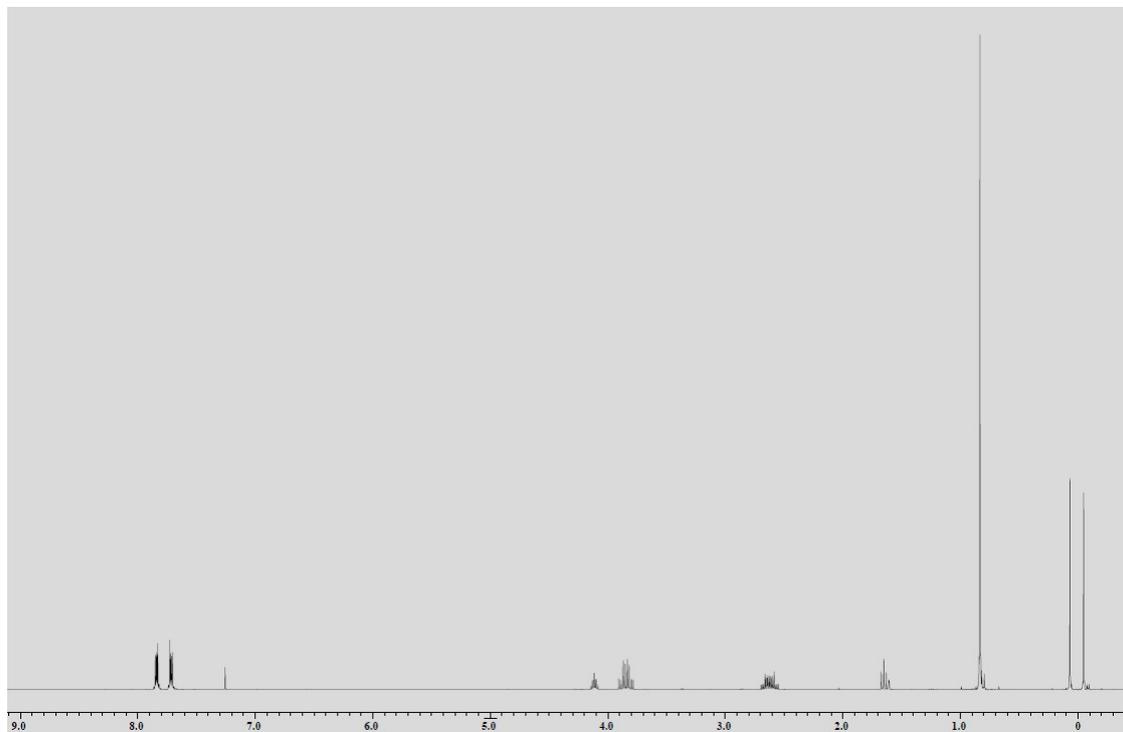
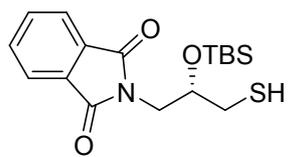
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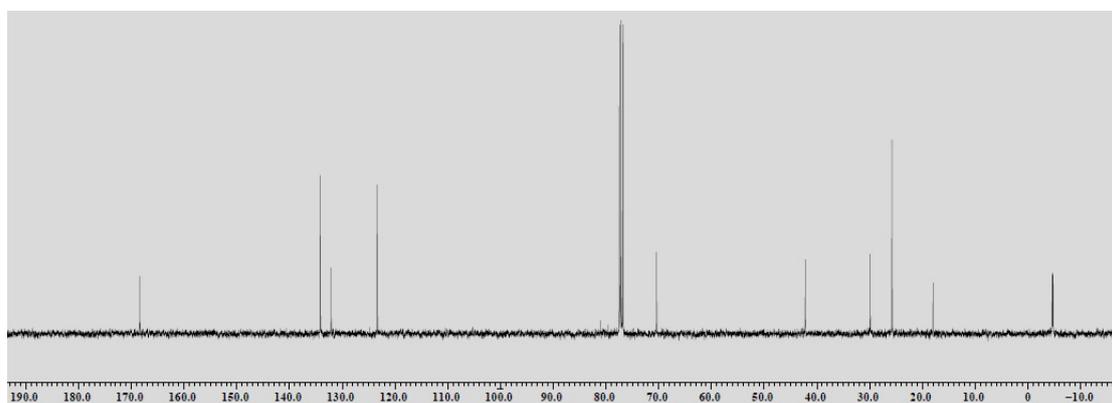
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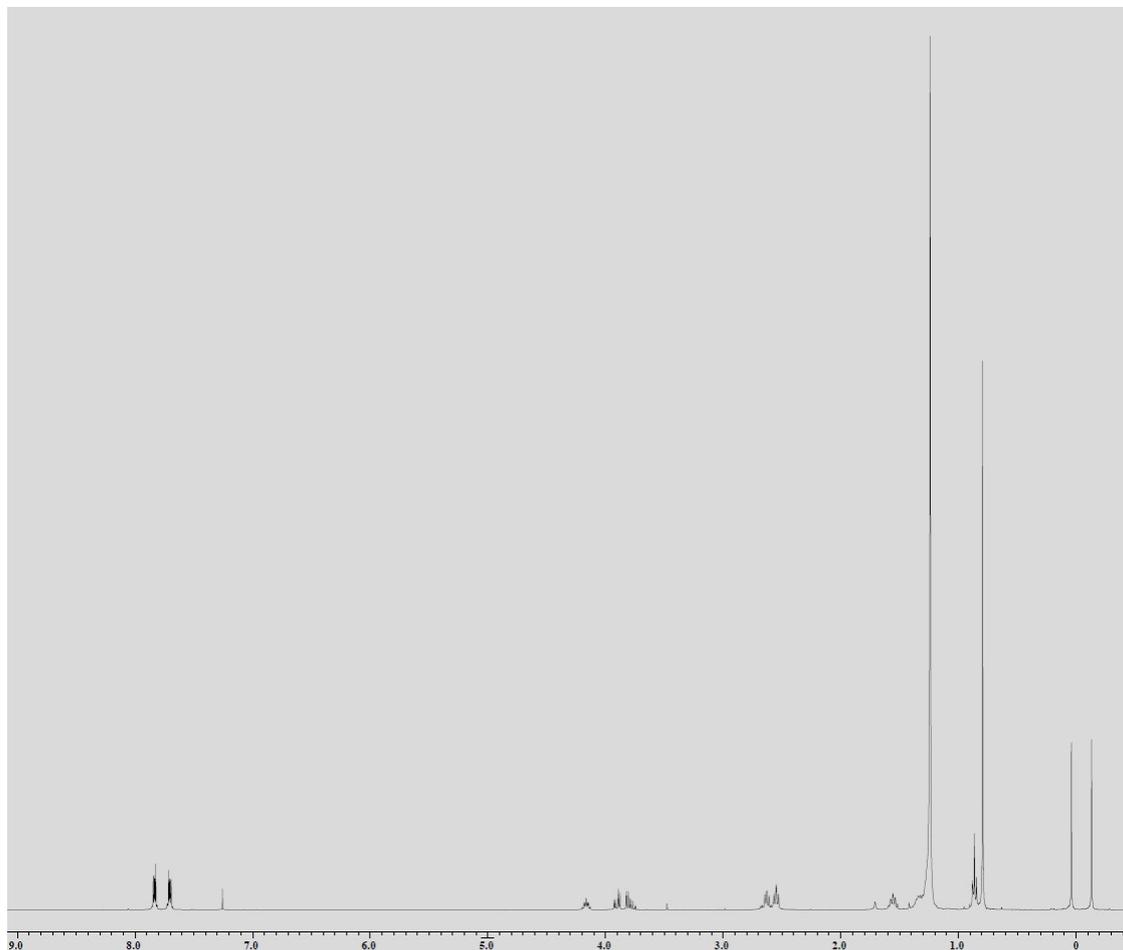
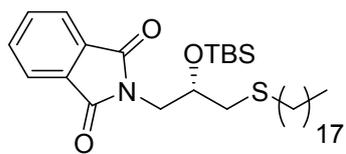
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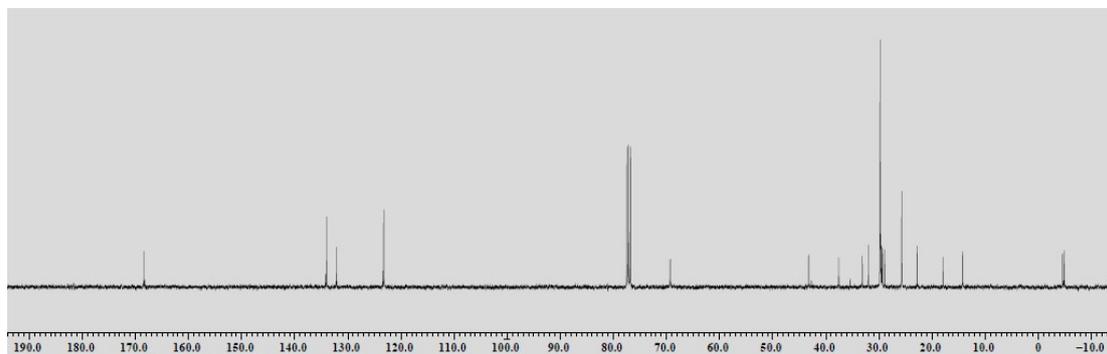
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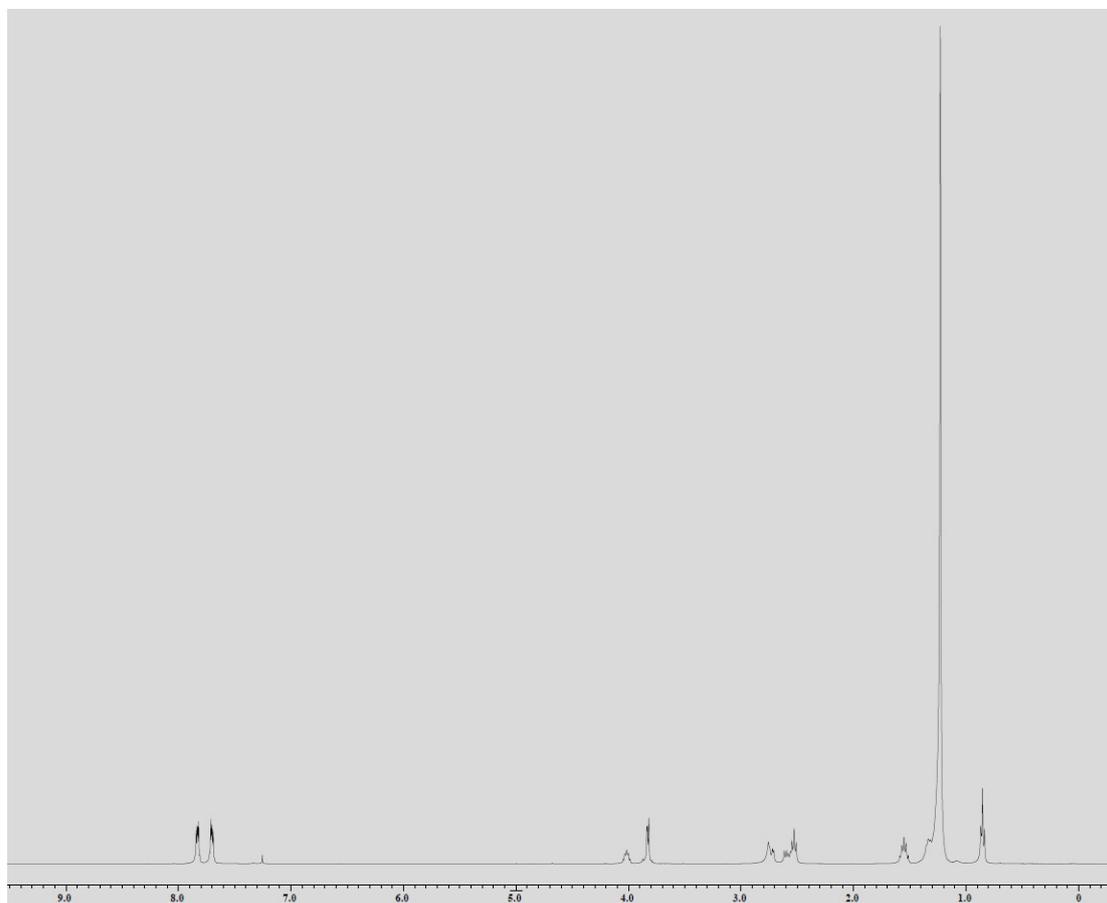
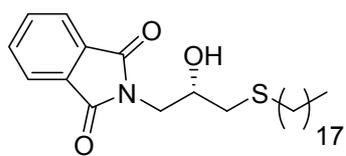
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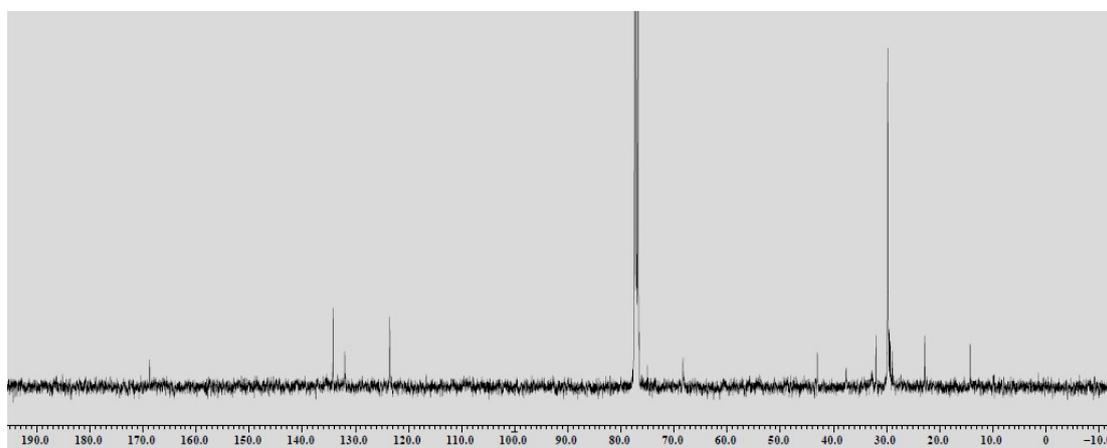
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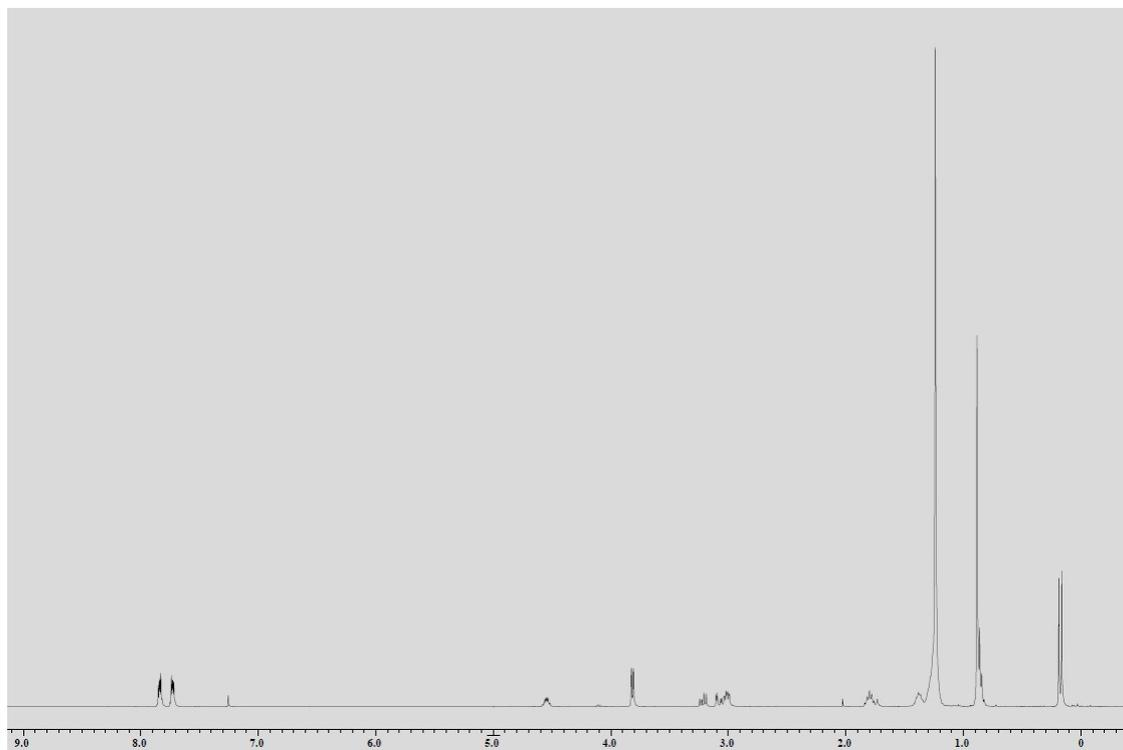
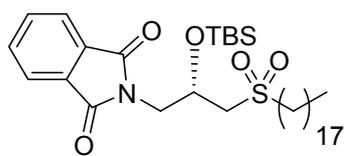
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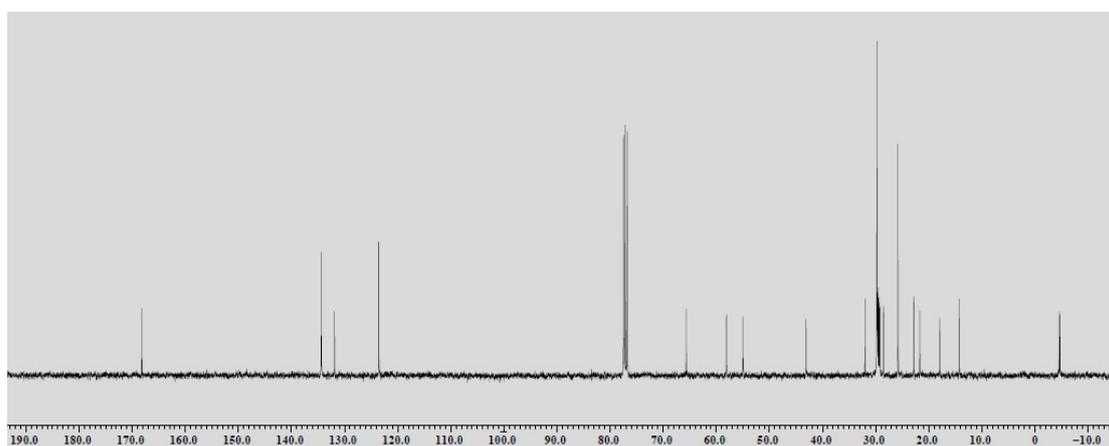
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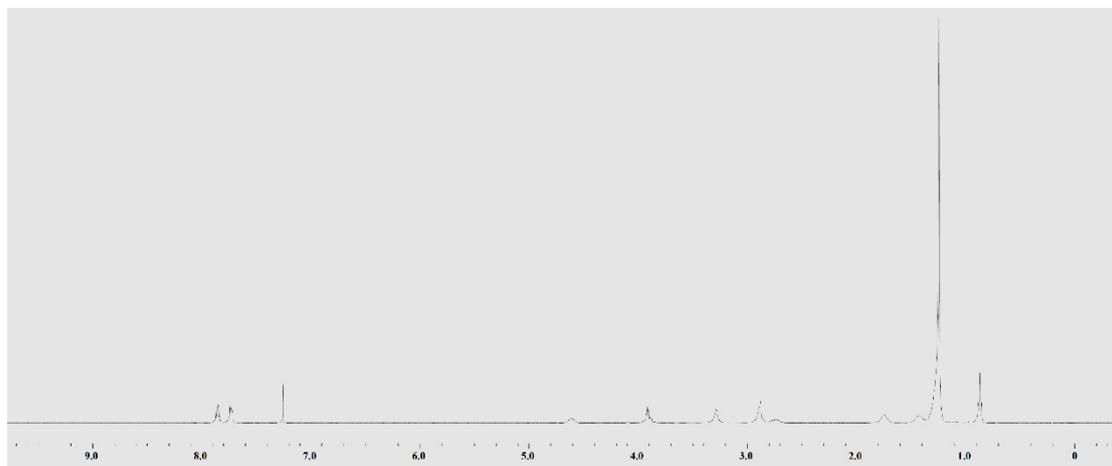
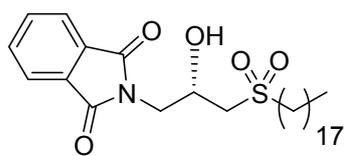
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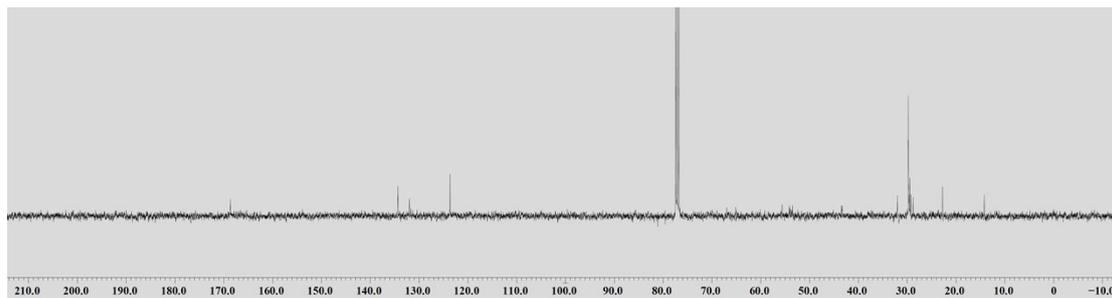
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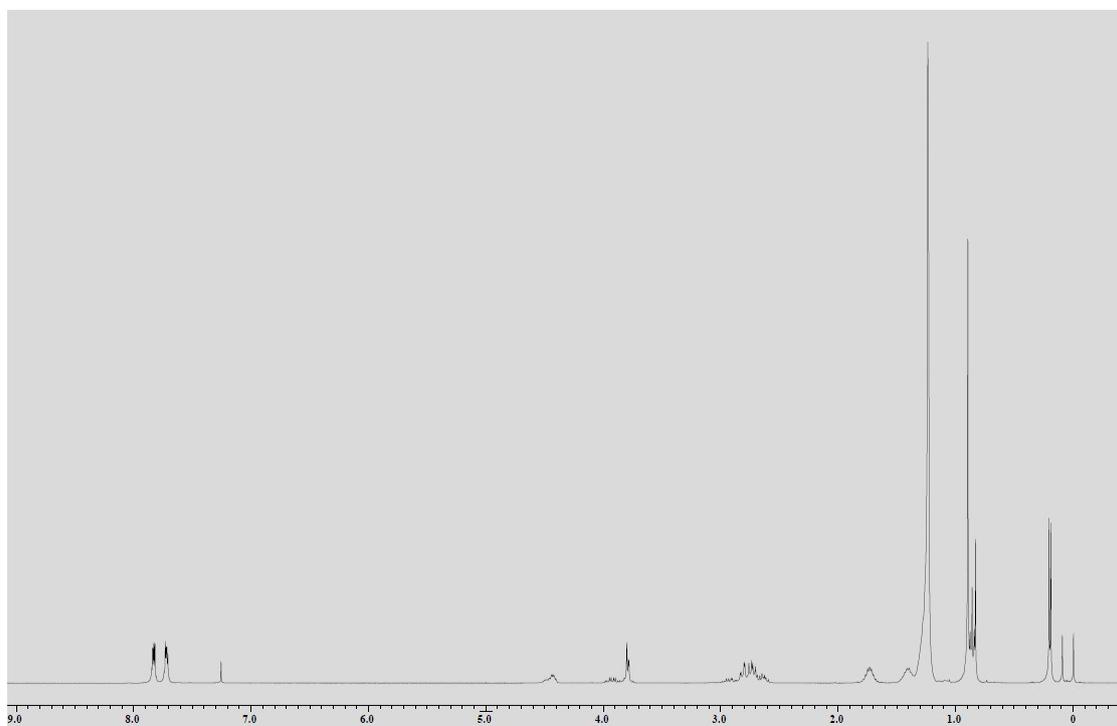
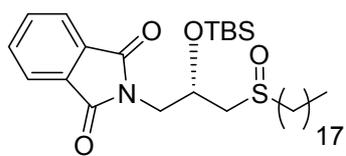
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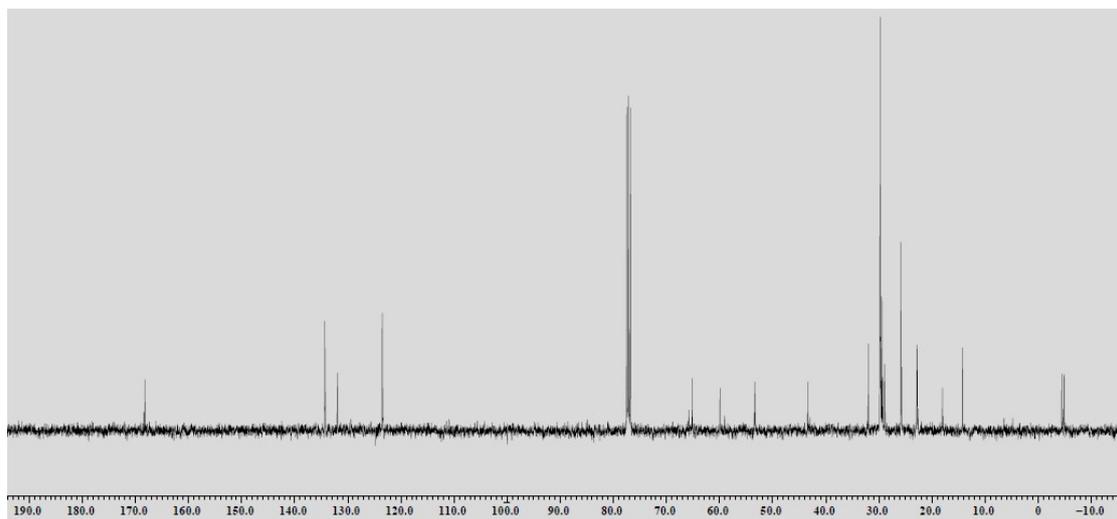
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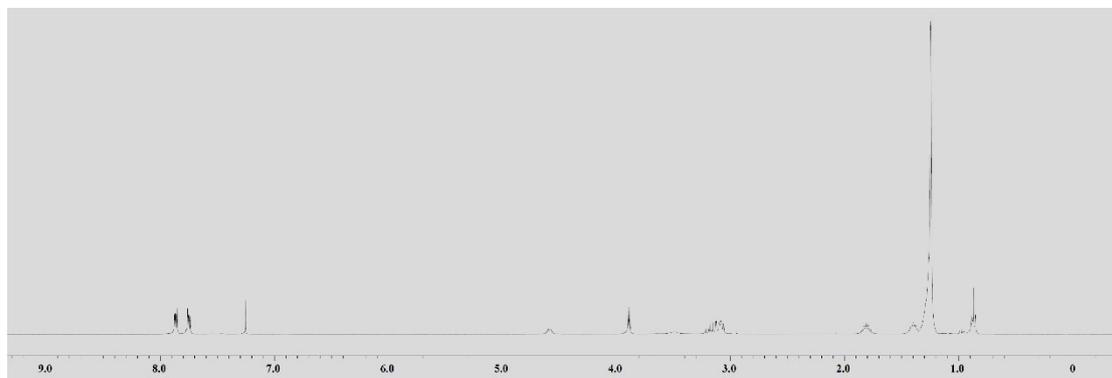
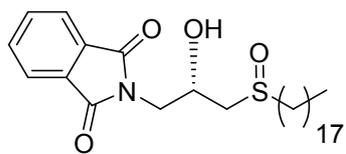
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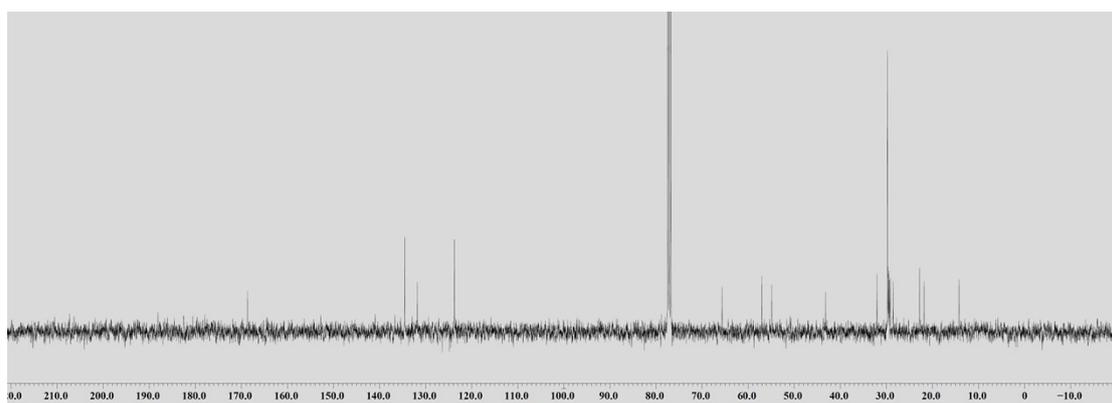
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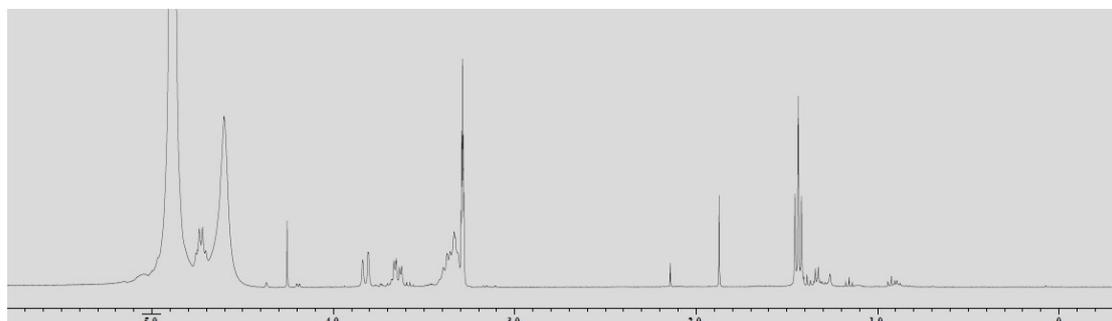
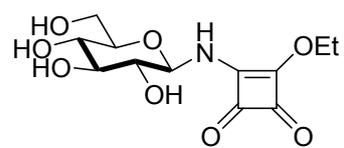
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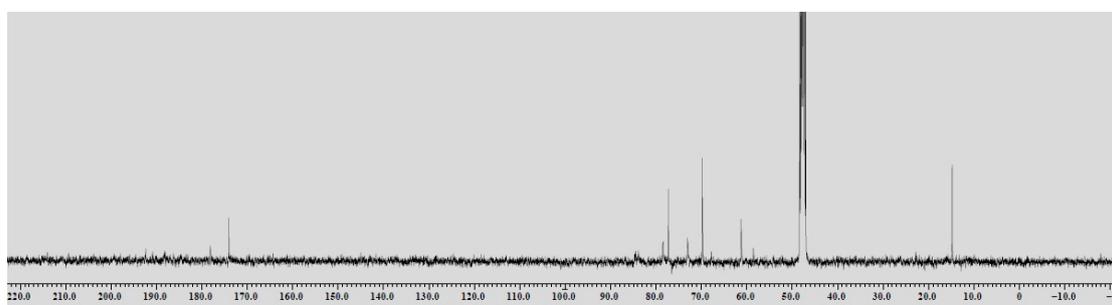
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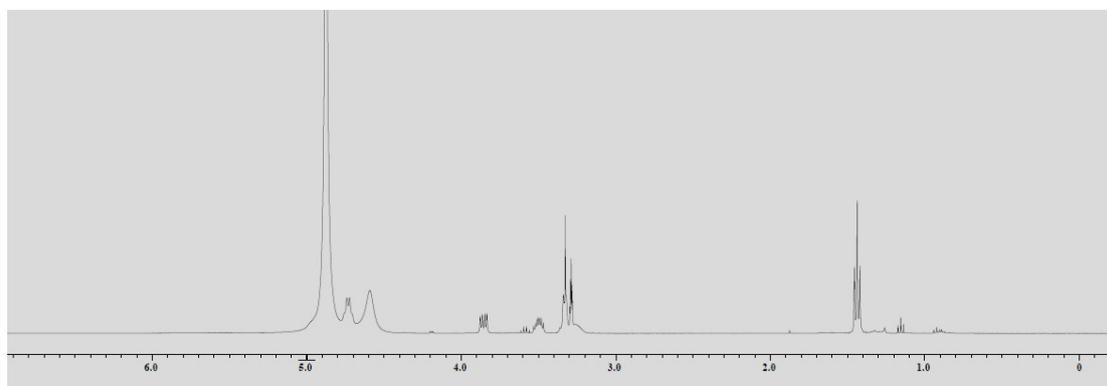
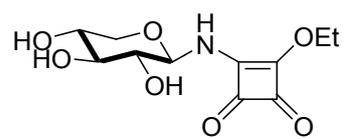
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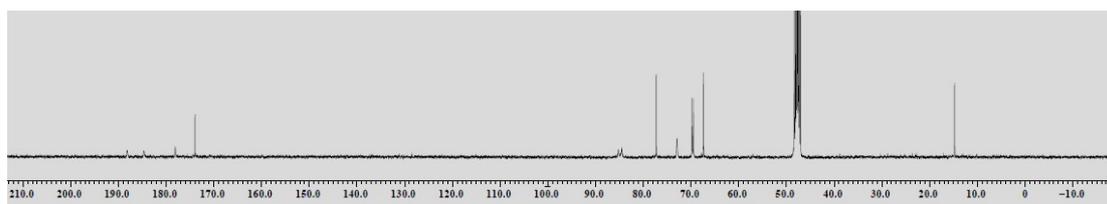
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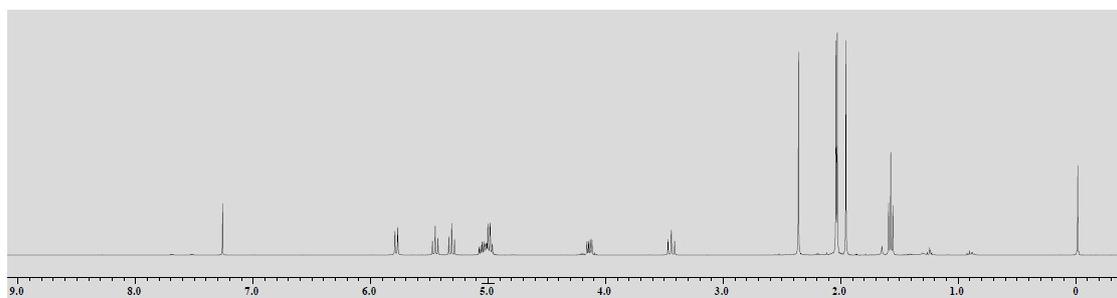
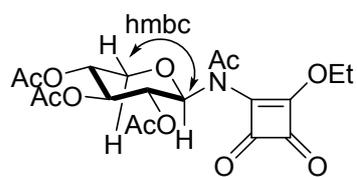
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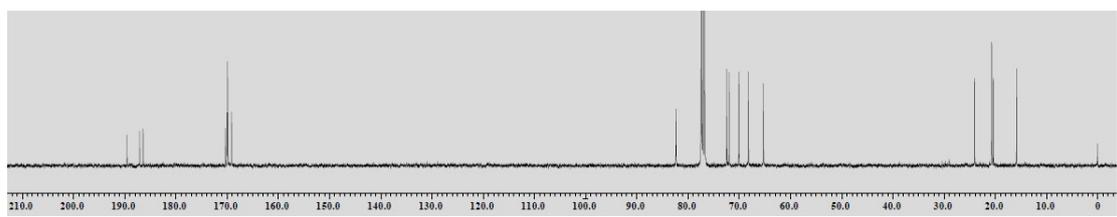
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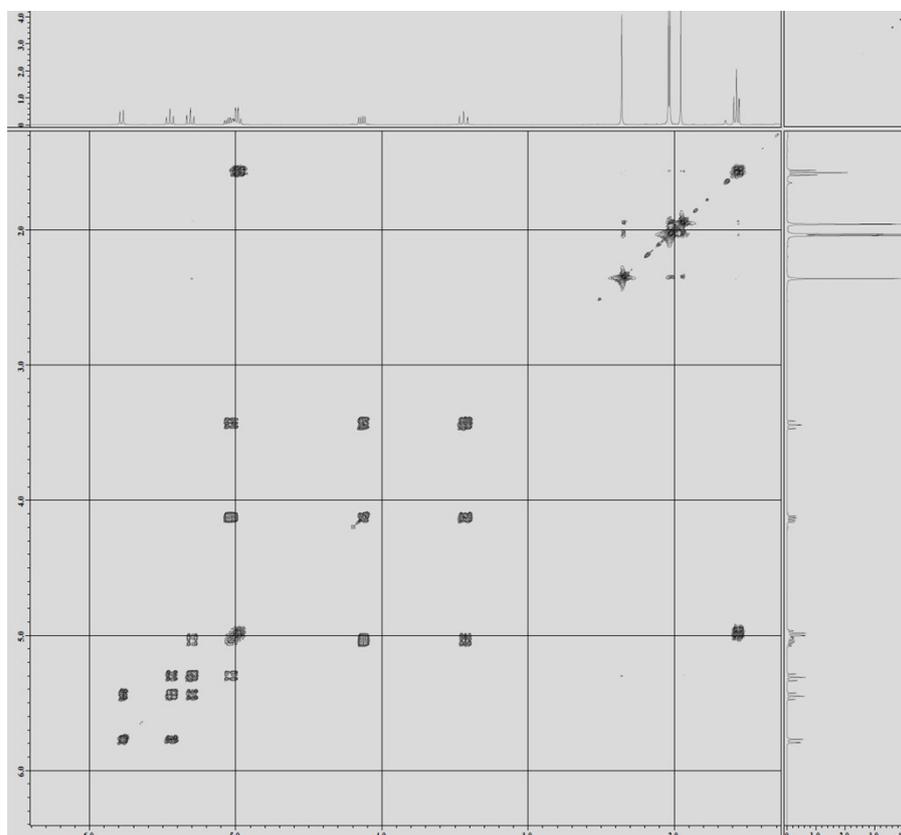
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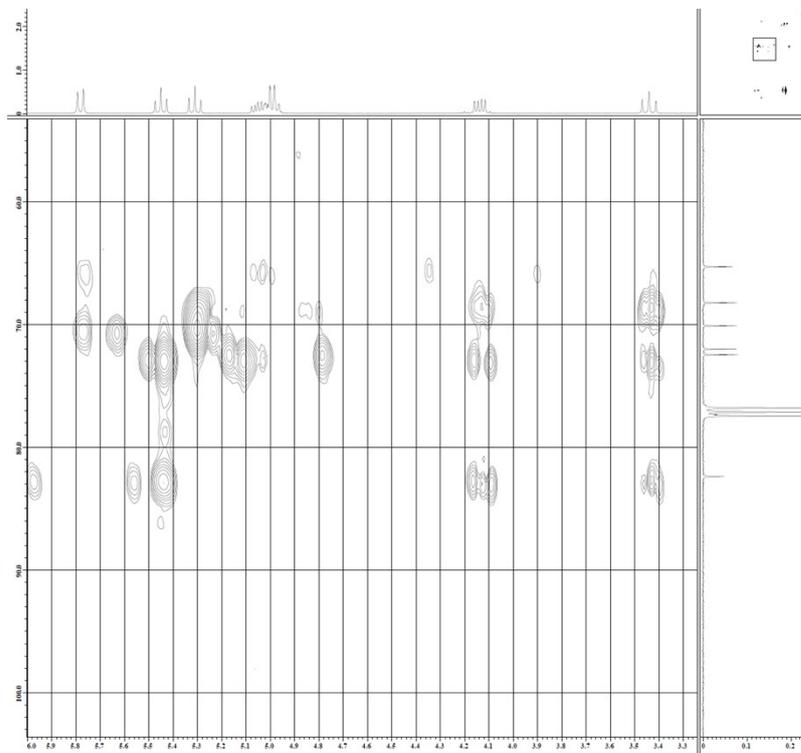
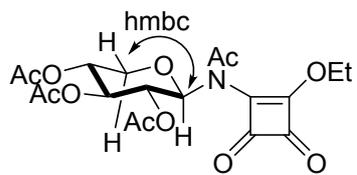
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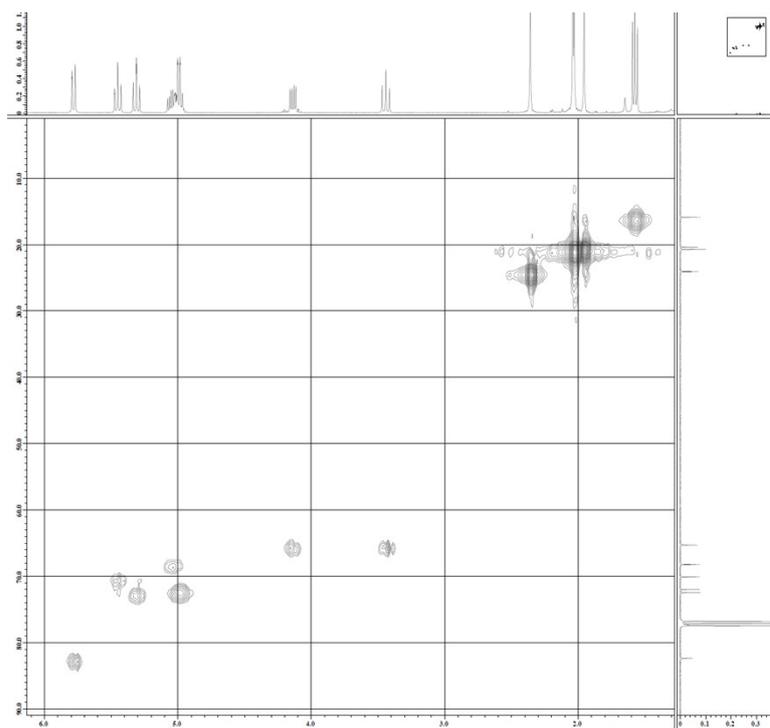
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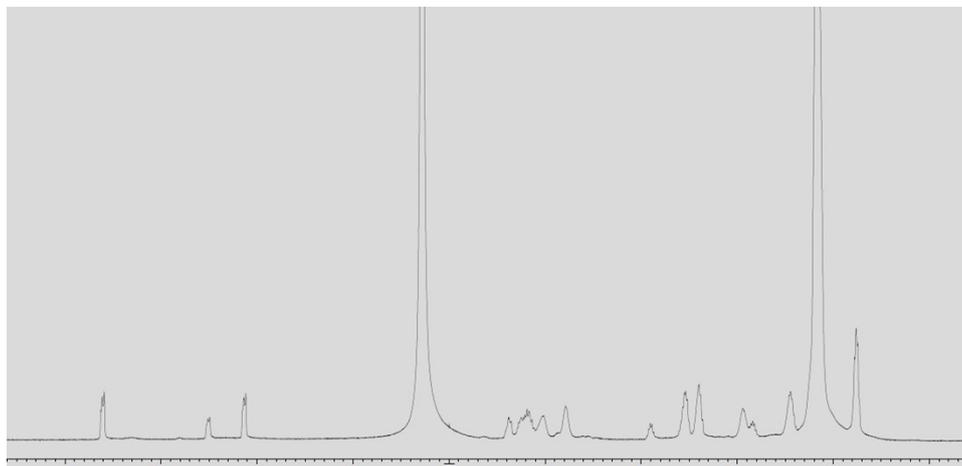
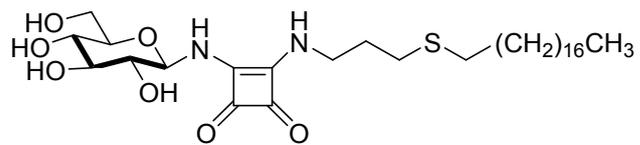
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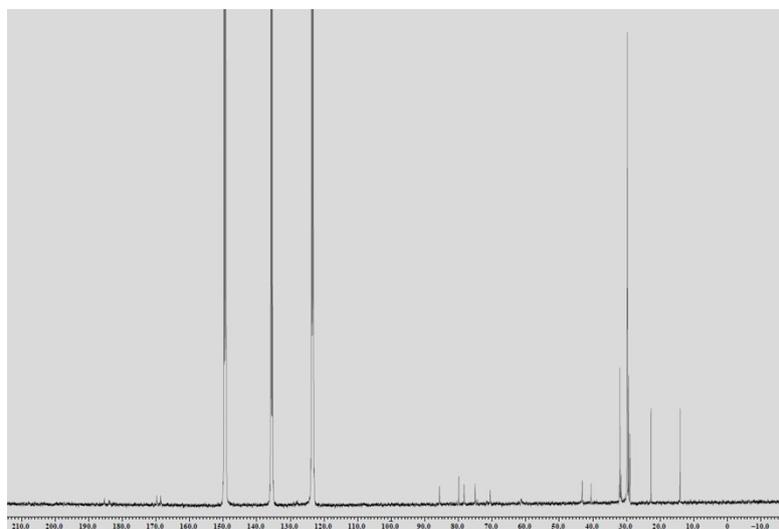
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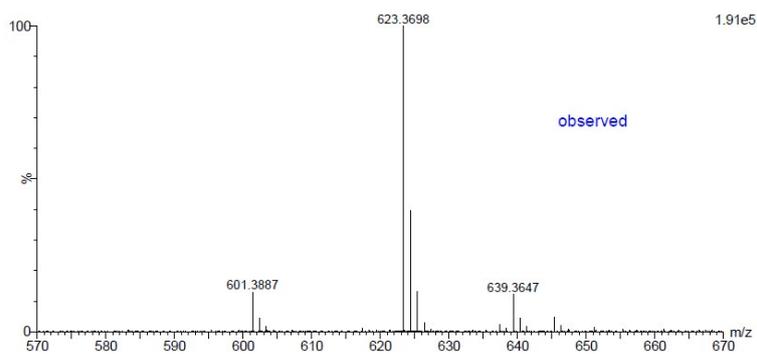
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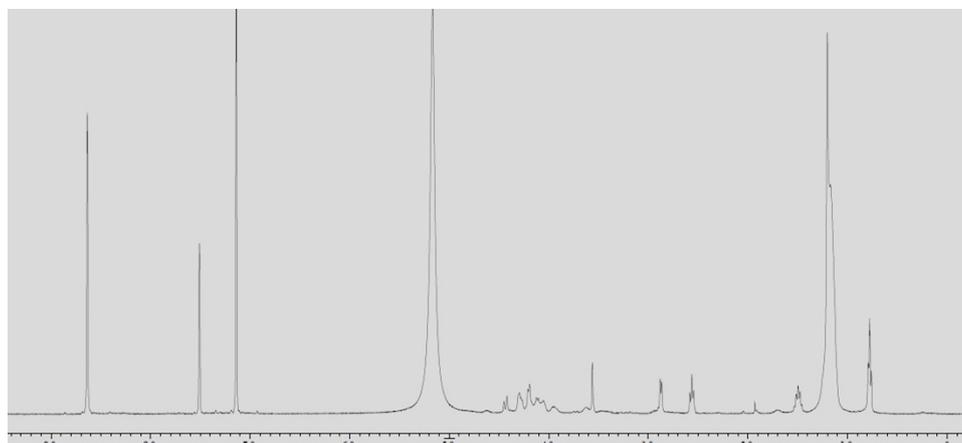
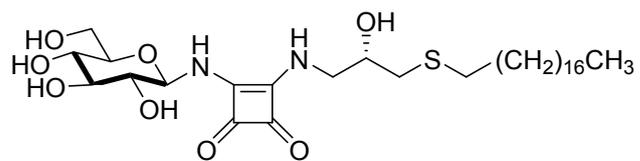
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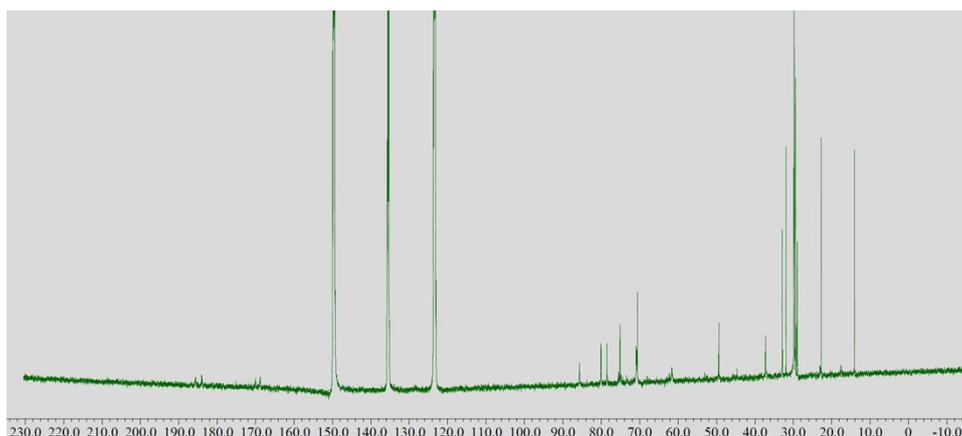
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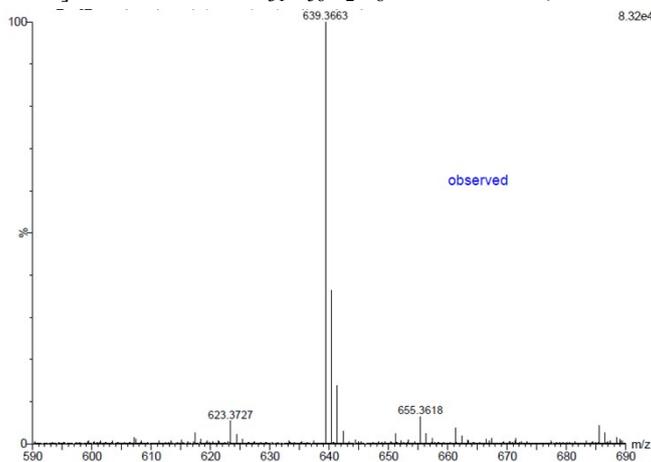
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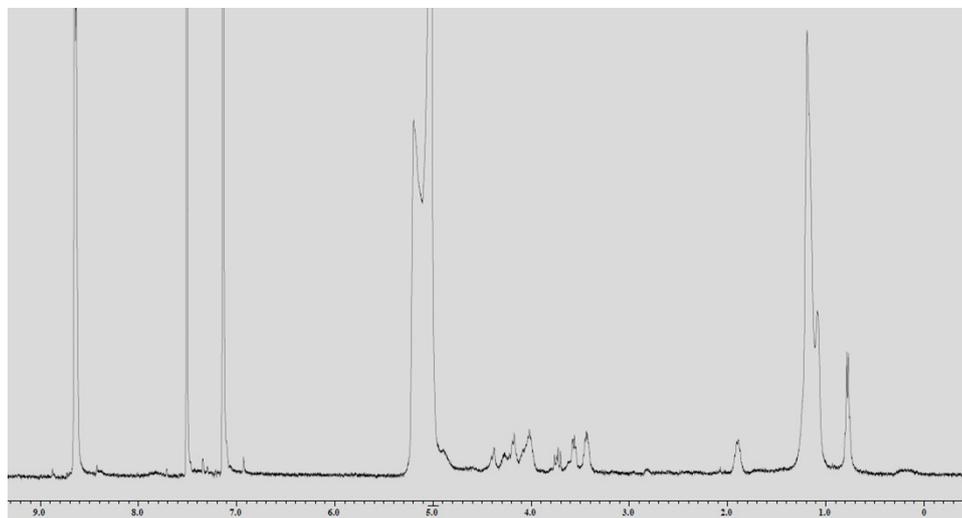
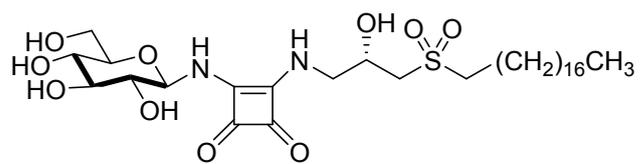
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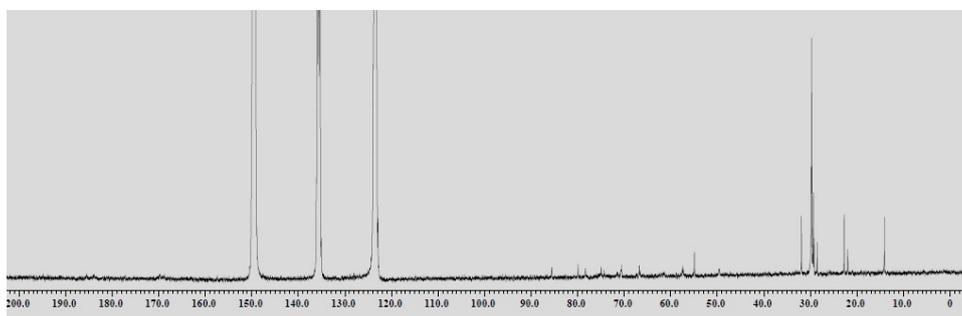
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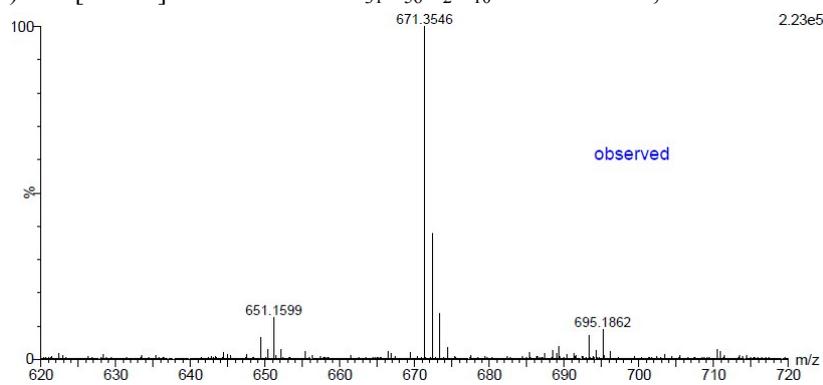
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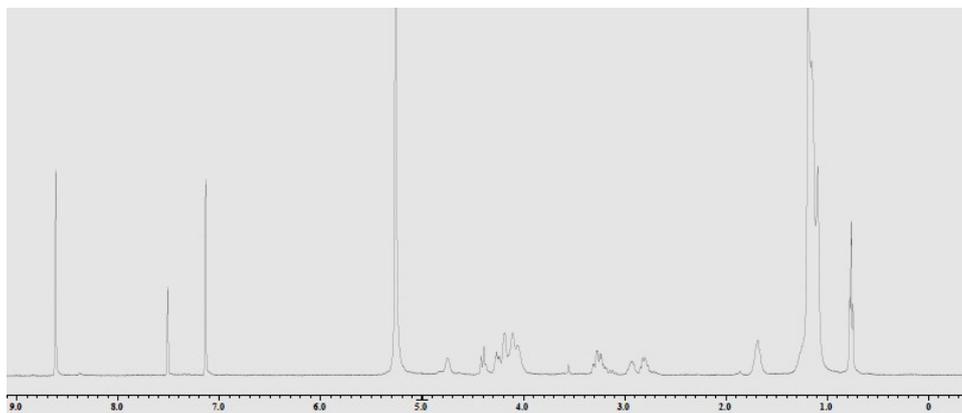
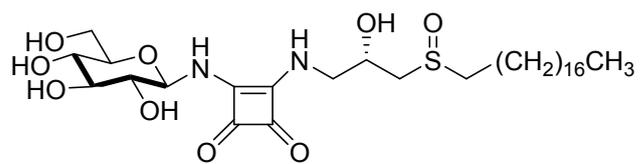
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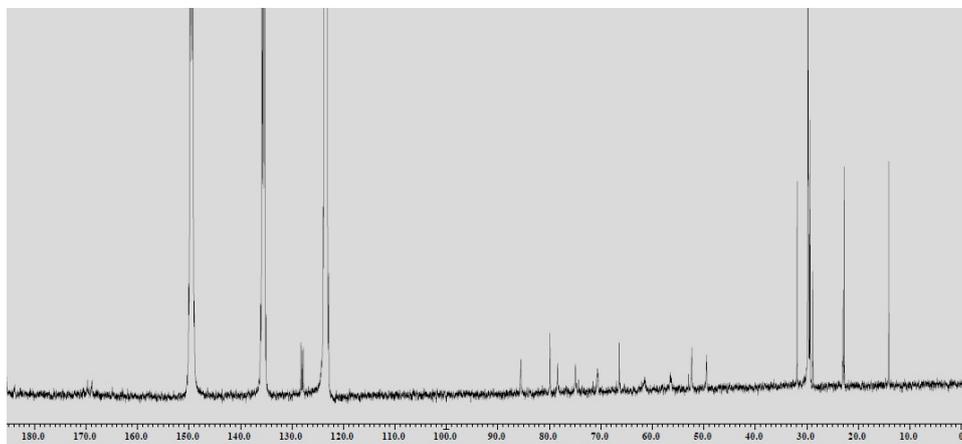
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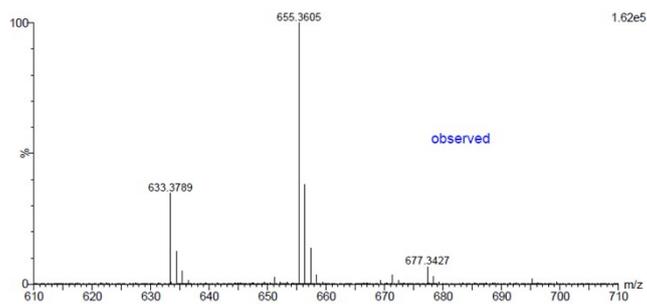
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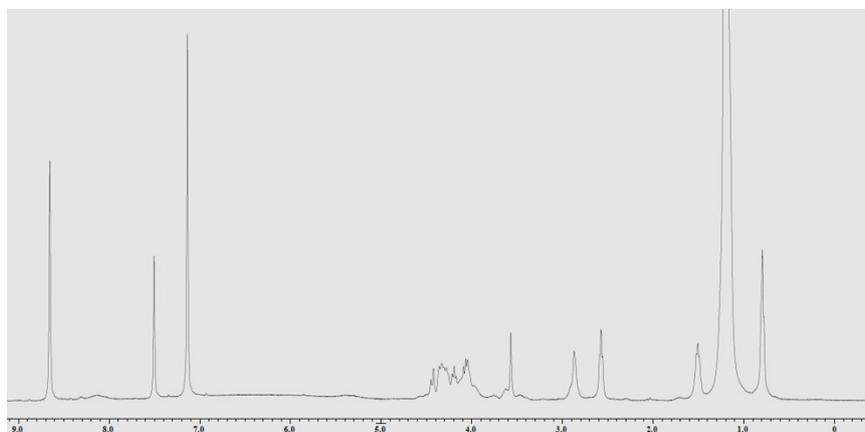
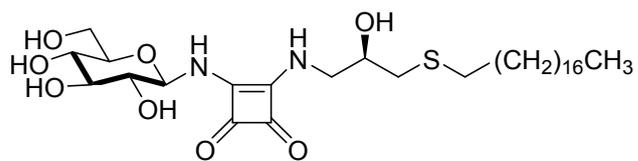
^{13}C NMR, 100 MHz, pyridine- d_5



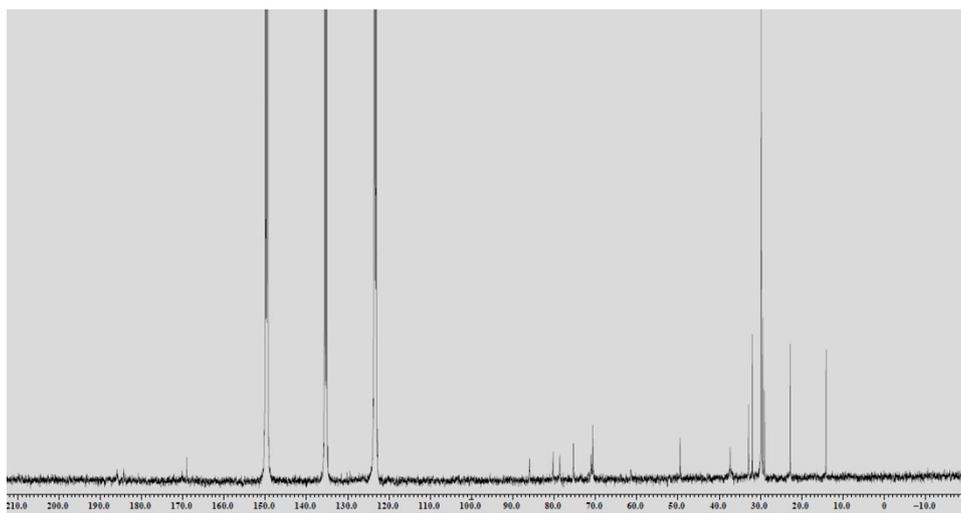
HRMS (ESI) m/z $[\text{M}+\text{Na}]^+$ calculated for $\text{C}_{31}\text{H}_{56}\text{N}_2\text{O}_9\text{SNa}$ 655.3604, found 655.3605.



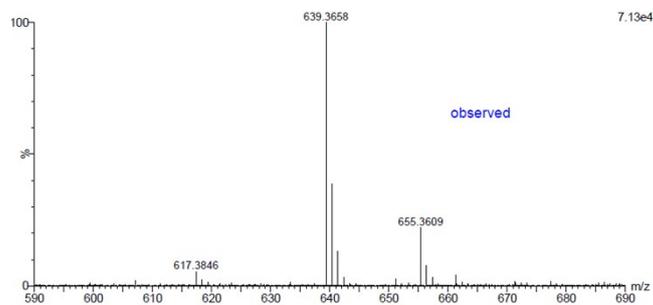
^1H NMR, 400 MHz, pyridine- d_5



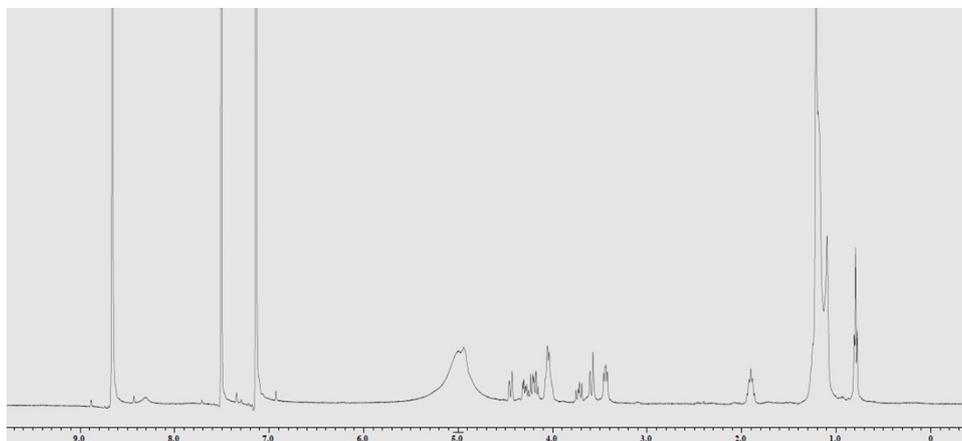
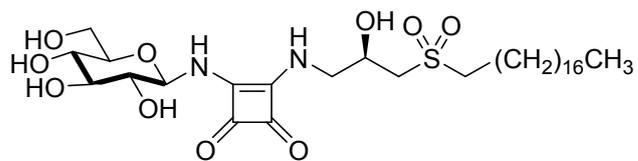
^{13}C NMR, 100 MHz, pyridine- d_5



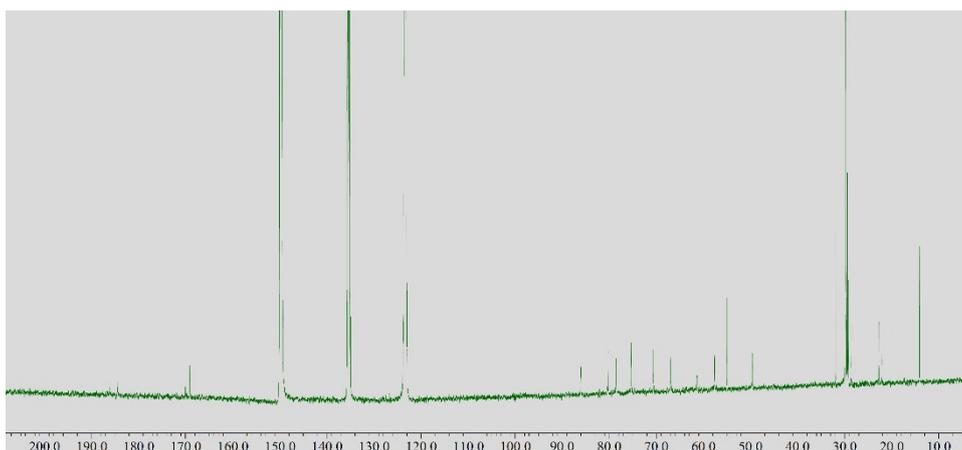
HRMS (ESI) m/z $[\text{M}+\text{Na}]^+$ calculated for $\text{C}_{31}\text{H}_{56}\text{N}_2\text{O}_8\text{SNa}$ 639.3655, found 639.3658.



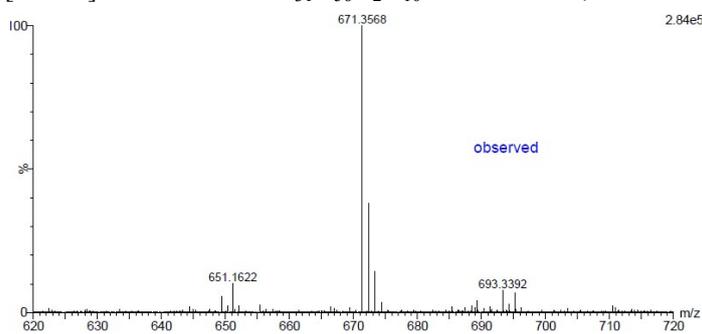
^1H NMR, 400 MHz, pyridine- d_5



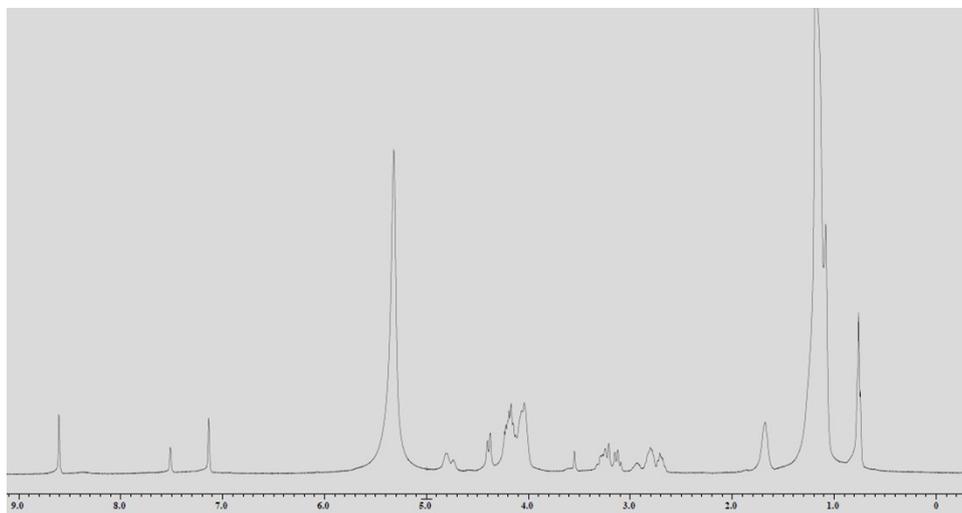
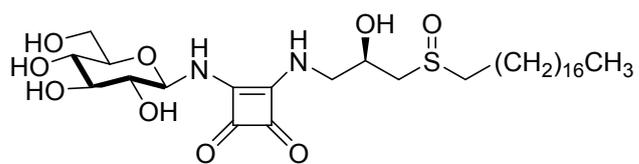
^{13}C NMR, 100 MHz, pyridine- d_5



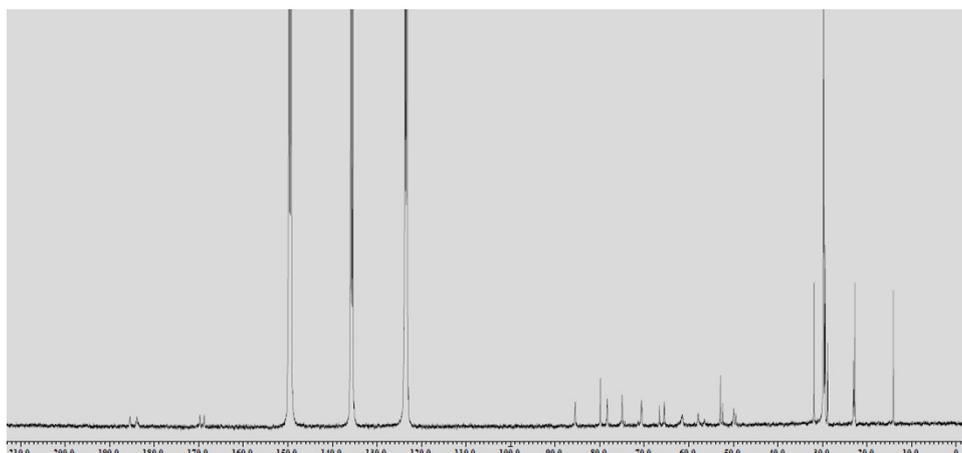
HRMS (ESI) m/z $[\text{M}+\text{Na}]^+$ calculated for $\text{C}_{31}\text{H}_{56}\text{N}_2\text{O}_{10}\text{SNa}$ 671.3553, found 671.3568.



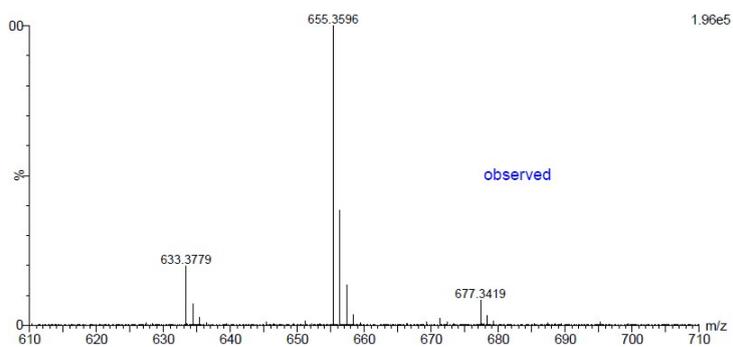
^1H NMR, 400 MHz, pyridine- d_5



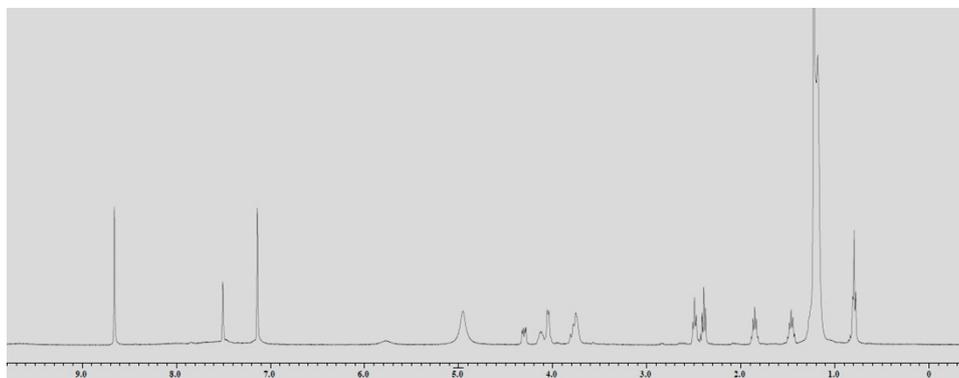
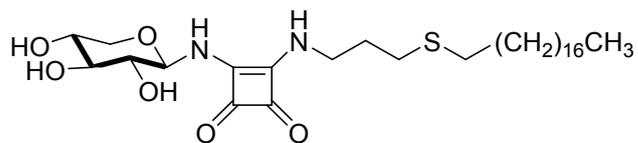
^{13}C NMR, 100 MHz, pyridine- d_5



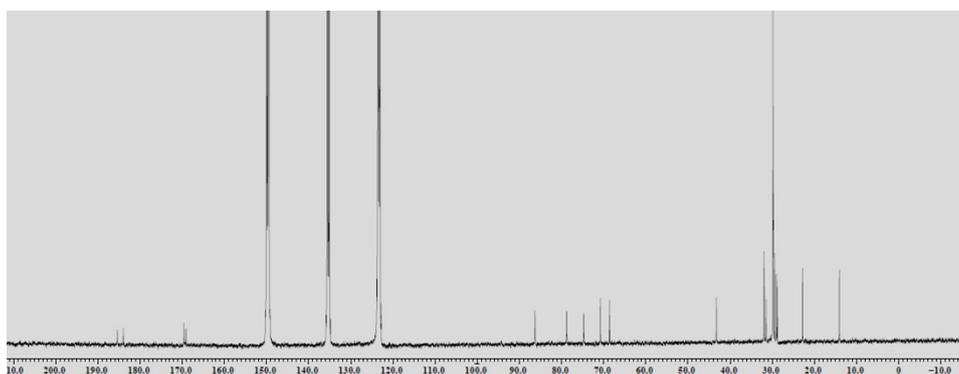
HRMS (ESI) m/z $[\text{M}+\text{Na}]^+$ calculated for $\text{C}_{31}\text{H}_{56}\text{N}_2\text{O}_9\text{SNa}$ 655.3604, found 655.3596.



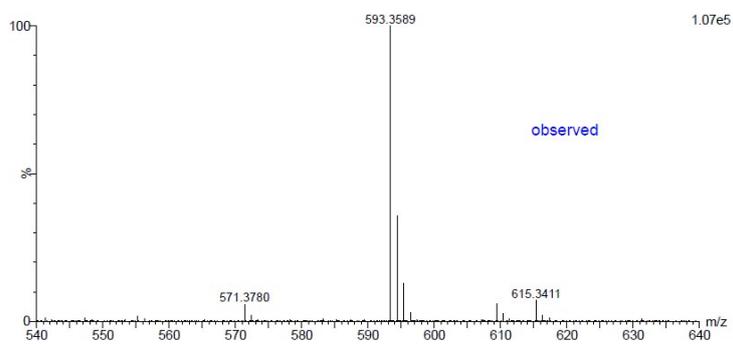
^1H NMR, 400 MHz, pyridine- d_5



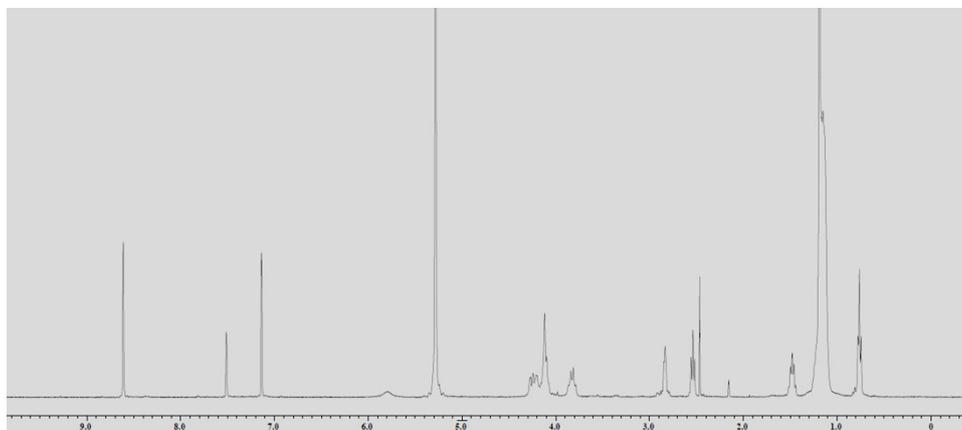
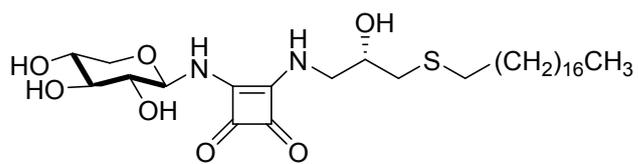
^{13}C NMR, 100 MHz, pyridine- d_5



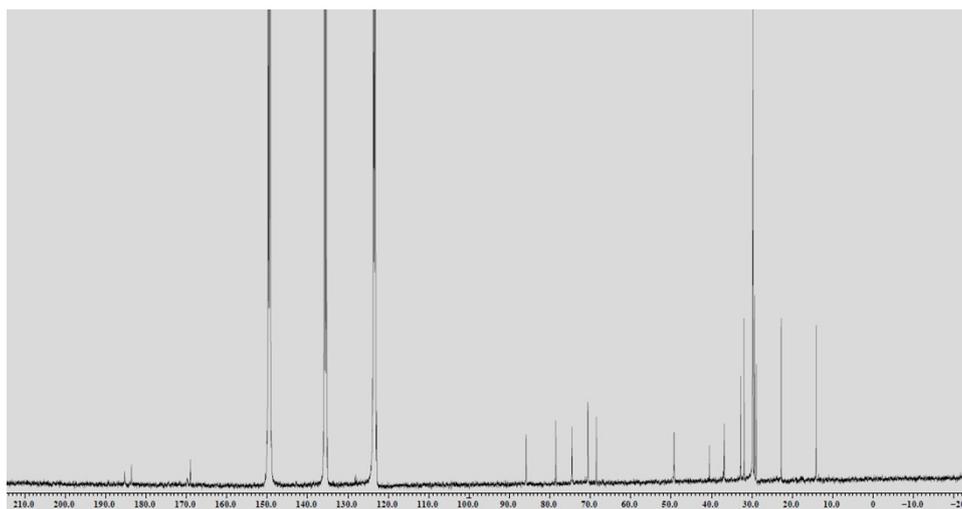
HRMS (ESI) m/z $[\text{M}+\text{Na}]^+$ calculated for $\text{C}_{30}\text{H}_{54}\text{N}_2\text{O}_6\text{SNa}$ 593.3600, found 593.3589.



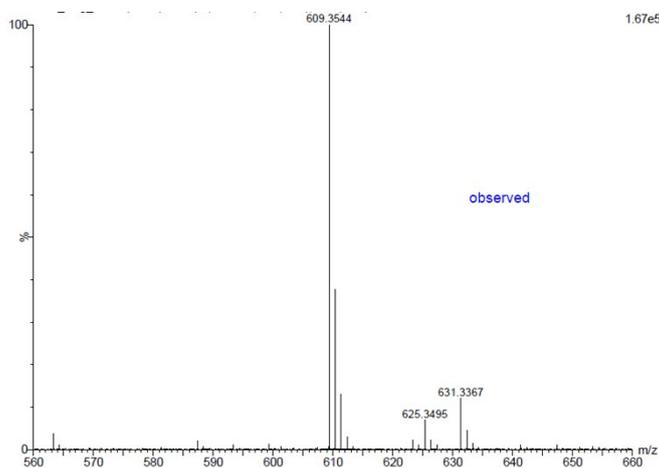
^1H NMR, 400 MHz, pyridine- d_5



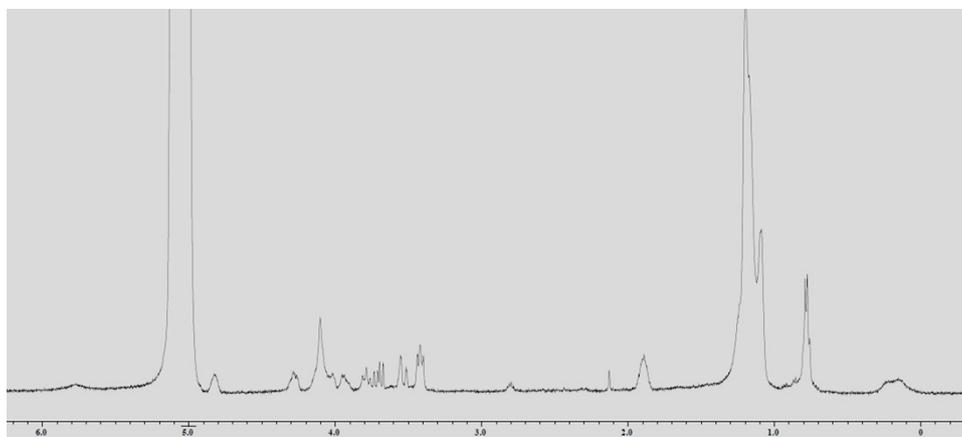
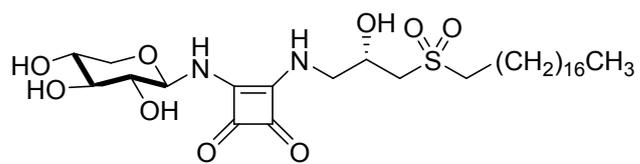
^{13}C NMR, 100 MHz, pyridine- d_5



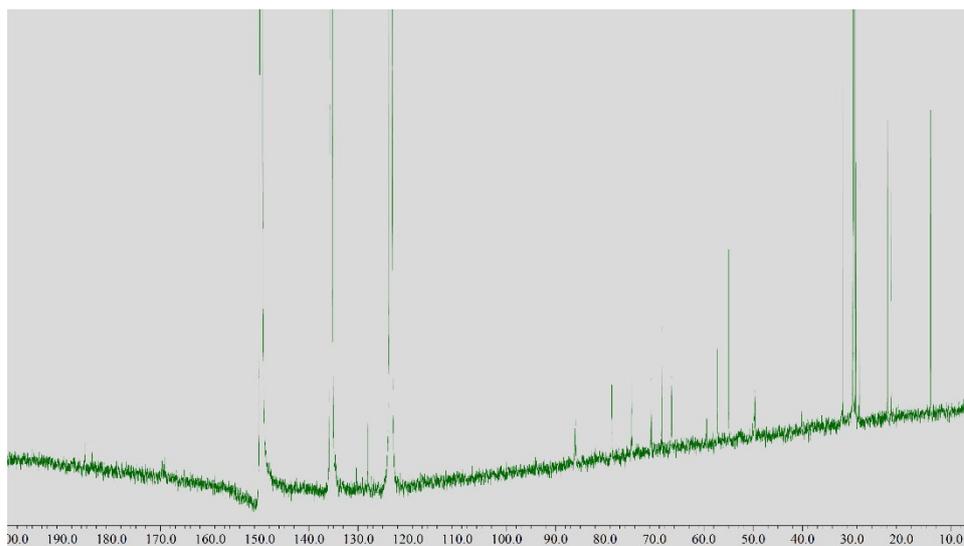
HRMS (ESI) m/z $[\text{M}+\text{Na}]^+$ calculated for $\text{C}_{30}\text{H}_{54}\text{N}_2\text{O}_7\text{SNa}$ 609.3549, found 609.3544.



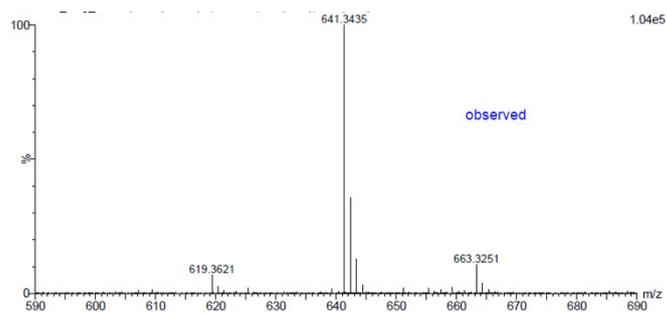
^1H NMR, 400 MHz, pyridine- d_5



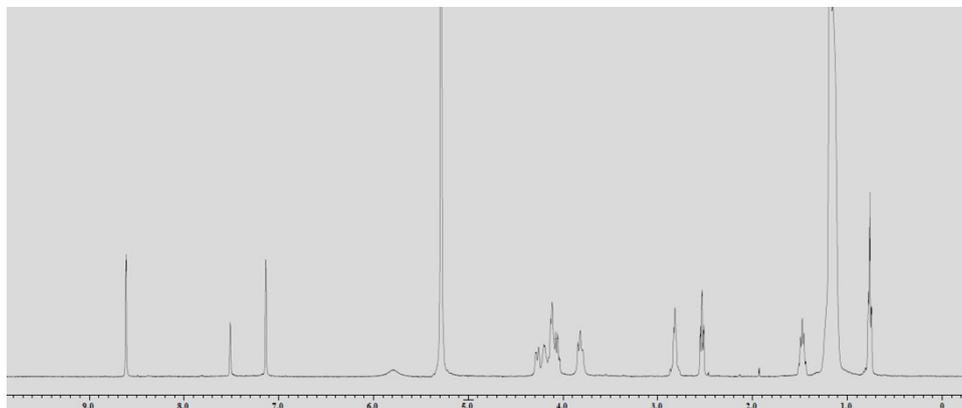
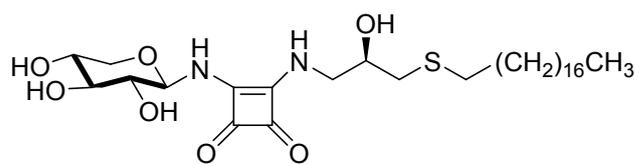
^{13}C NMR, 100 MHz, pyridine- d_5



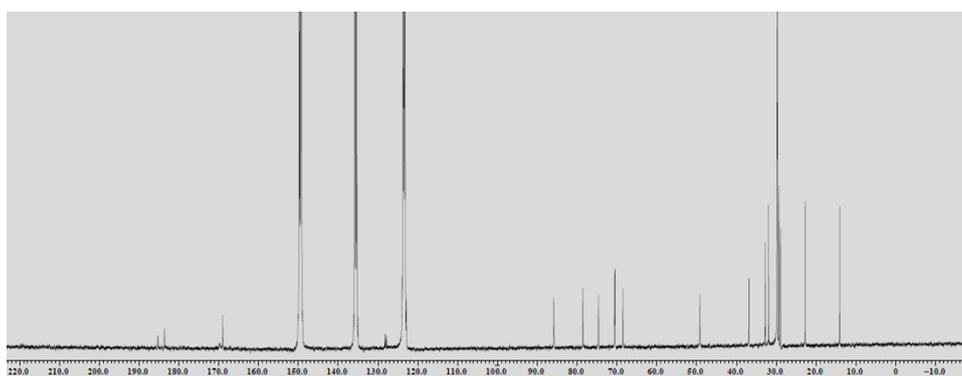
HRMS (ESI) m/z $[\text{M}+\text{Na}]^+$ calculated for $\text{C}_{30}\text{H}_{54}\text{N}_2\text{O}_9\text{SNa}$ 641.3448, found 641.3435.



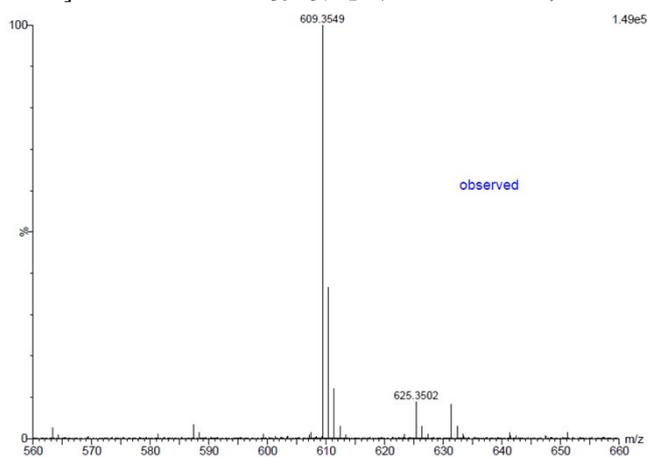
^1H NMR, 400 MHz, pyridine- d_5



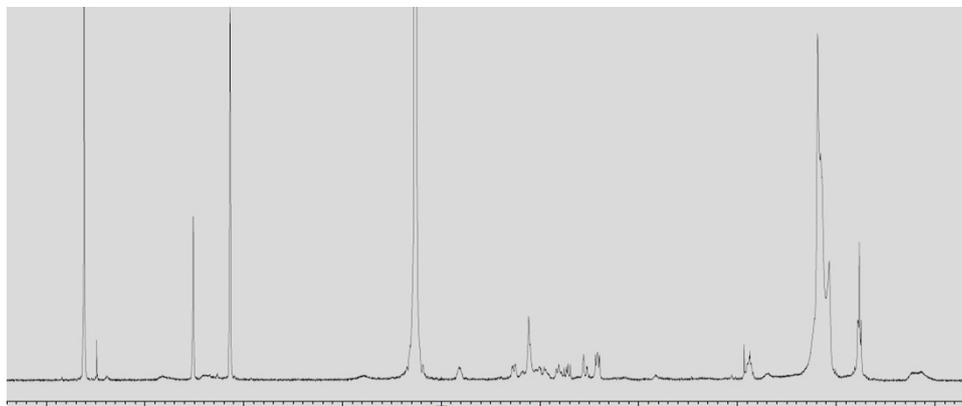
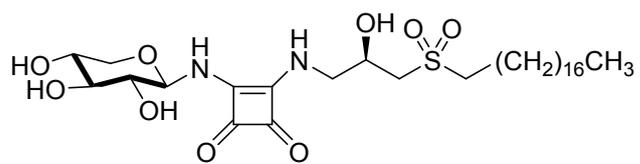
^{13}C NMR, 100 MHz, pyridine- d_5



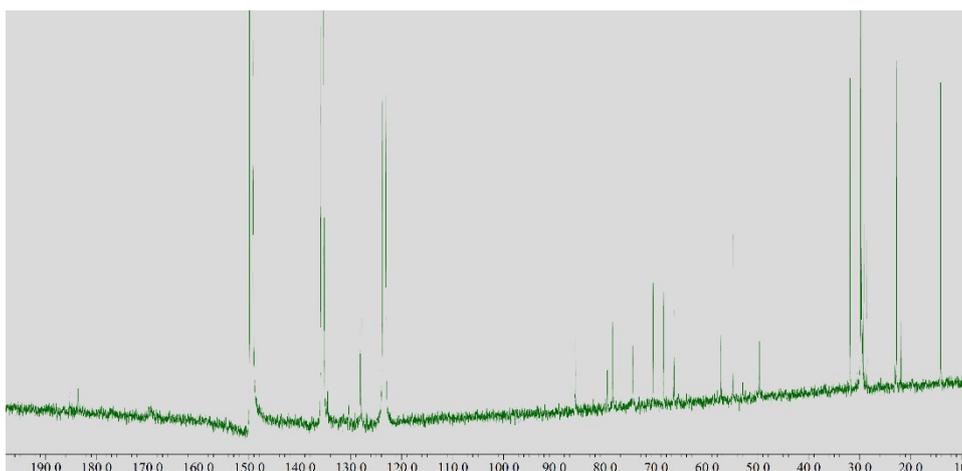
HRMS (ESI) m/z $[\text{M}+\text{Na}]^+$ calculated for $\text{C}_{30}\text{H}_{54}\text{N}_2\text{O}_7\text{SNa}$ 609.3549, found 609.3549



^1H NMR, 400 MHz, pyridine- d_5



^{13}C NMR, 100 MHz, pyridine- d_5



HRMS (ESI) m/z $[\text{M}+\text{Na}]^+$ calculated for $\text{C}_{30}\text{H}_{54}\text{N}_2\text{O}_9\text{SNa}$ 641.3448, found 641.3459.

