

Supporting Information

**The Open d-Shell Enforces the Active Space in 3d Metal Catalysis:
Highly Enantioselective Chromium(II) Pincer Catalysed
Hydrosilylation of Ketones**

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Contents

1. General Remarks	3
2. Synthesis of the Precatalysts $R(\text{PdmBox})\text{Cr}(\text{CH}_2\text{SiMe}_3)$	5
2.1 Synthesis of the $R(\text{PdmBox})\text{CrCl}$ Complexes	5
2.1.1 $[(R)\text{-Ph}(\text{PdmBox})\text{CrCl}]$	5
2.1.2 $[(S)\text{-iPr}(\text{PdmBox})\text{CrCl}]$	6
2.2 Syntheses of the $R(\text{PdmBox})\text{Cr}(\text{CH}_2\text{SiMe}_3)$ Complexes	6
2.2.1 $[(R)\text{-Ph}(\text{PdmBox})\text{Cr}(\text{CH}_2\text{SiMe}_3)]$	6
2.2.2 $[(S)\text{-iPr}(\text{PdmBox})\text{Cr}(\text{CH}_2\text{SiMe}_3)]$	7
3. Catalysis	8
3.1 Catalytic Enantioselective Reduction	8
3.2 Enantioselective Hydrosilylation of Ketones	10
3.3 Characterization of Alcohols	10
4. Mechanistic Studies	20
4.1 Radical Trap Experiments	20
4.2 Labelling Experiments	20
4.3 Kinetic Isotope Effect	21
4.4 Hammett-Correlation	22
5. Chromatographic Data	24
6. Crystal Structure Data	32
7. Literature	33

1. General Remarks

Reactions

Unless stated otherwise, all reactions were carried out under inert conditions in heat-gun dried glassware and under an atmosphere of argon using standard Schlenk techniques or inside of a Glovebox (M. Braun Unilab 2000). All chemicals were bought from commercial suppliers (Acros, Sigma-Aldrich, Alfa Aesar or ABCR) and were used without any further purification, unless mentioned otherwise. Dry solvents were taped from a solvent purification system (M. Braun SPS-800) and used immediately. Manual degassing of solvents, if needed, was done by performing three consecutive freeze-pump-thaw cycles. Dry DMF was purchased from Sigma-Aldrich. Deuterated solvents were purchased from Deutero GmbH or Sigma-Aldrich and dried over sodium (C_6D_6), distilled, degassed and stored under an atmosphere of argon. All chromium salts were purchased with a trace metal purity of 99.9 %. The PdmBox-ligands,¹ (tmeda)CrCl₂² and deuterated Silane³ were synthesized according to reported procedures.

Analytics

Nuclear magnetic resonance (NMR) spectra were recorded on Bruker Avance II 400 and Bruker Avance III 600 at room temperature. Chemical shifts are reported in ppm, coupling constants in Hz. Chemical shifts were referenced to residual solvent protons and the ¹³C isotope of deuterated solvents.⁴ Multiplicities are indicated as: s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet. Atom numbering is illustrated in the respective figure shown above each procedure.

Mass spectrometry (MS) and high-resolution MS were obtained at the mass spectroscopy department of the University of Heidelberg. Electron spray ionization (ESI) was carried out on a Bruker ApexQe hybrid 9.4 T FT-IVR machine, liquid injection field desorption ionization (LIFDI) was performed on a JEOL JMS- 700 instrument.

Elemental analysis (EA) for C, H, and N was performed at a facility of the Chemistry Department at the University of Heidelberg on a vario MICRO Cube or vario EL Cube.

High Performance Liquid Chromatography (HPLC) measurements were carried out on Agilent Technologies 1260 Infinity HPLC equipped with solvent pump, auto-sampler, membrane solvent degasser and DAD detector.

Silica gel (SiO₂, pore size 60 Å) for flash column chromatography (FCC) was purchased from Sigma-Aldrich. Thin Layer chromatography (TLC) was performed with Polygram® SIL G/UV₂₅₄ purchased from Macherey-Nagel. Components were visualized by fluorescence quenching during irradiation with UV light (254 nm) or were revealed with Hanessian's stain.

X-ray Crystal Structure Determinations

Crystal data and details of the structure determinations are compiled in Table S2. Full shells of intensity data were collected at low temperature with an Agilent Technologies Supernova-E CCD diffractometer (Mo-K α radiation, microfocus X-ray tube, multilayer mirror optics). Detector frames (typically ω -, occasionally ϕ -scans, scan width 0.4°) were integrated by profile fitting.^{5,6} Data were corrected for air and detector absorption, Lorentz and polarization effects⁶ and scaled essentially by application of appropriate spherical harmonic functions.^{6,7,8} Absorption by the crystal was treated with a semiempirical multiscan method (as part of the scaling process), and augmented by a spherical correction.^{7,8} The structures were solved by the charge flip procedure⁹ and refined by full-matrix least squares methods based on F^2 against all unique reflections.¹⁰ All non-hydrogen atoms were given anisotropic displacement parameters. Hydrogen atoms were input at calculated positions and refined with a riding model.¹¹

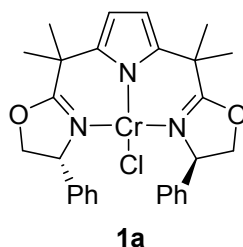
CCDC 1850233-1850234 contains the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via https://www.ccdc.cam.ac.uk/data_request/cif.

2. Synthesis of the Precatalysts $^R(\text{PdmBox})\text{Cr}(\text{CH}_2\text{SiMe}_3)$

2.1 Synthesis of the $^R(\text{PdmBox})\text{CrCl}$ Complexes

The protioligand $^R(\text{PdmBox})\text{H}$ (2.09 mmol, 1.0 eq.) was dissolved in THF (20 mL) and LiHMDS (2.30 mmol, 1.1 eq.) dissolved in THF (5 mL) was slowly added and the mixture was stirred for 30 min at rt. Subsequently, a suspension of $(\text{tmeda})\text{CrCl}_2$ (2.09 mmol, 1.0 eq.) in THF (10 mL) was added dropwise to the reaction mixture and stirring was continued for 12 h at rt. Afterwards, the reaction was filtered, the solvent was removed, and the residue was redissolved in a mixture of toluene/pentane (1:3) and filtered over celite. The filtrate was evaporated to afford $^R(\text{PdmBox})\text{CrCl}$ complex as dark-violet solid. Single crystals suitable for X-ray diffraction were obtained from a saturated solution of **2b** in *n*-pentane at $-40\text{ }^\circ\text{C}$.

2.1.1 [R]-Ph(**PdmBox**)CrCl]



Yield: dark-violet solid (582.3 mg, 62 %).

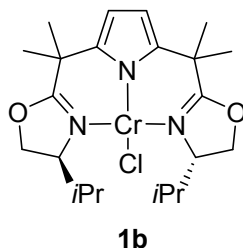
^1H NMR (C_6D_6 , 600.13 MHz, 295 K, paramagnetic): δ [ppm] = 23.34, 18.53, 5.25, 2.41, -37.81 .

Magnetic Susceptibility (Evans, C_6D_6 , 295 K)¹²: $\mu_{\text{eff}} = 4.75\ \mu_{\text{B}}$.

EA: calcd. C: 63.69 %, H: 5.72 %, N: 7.96 %; found: C: 63.09 %, H: 5.93 %, N: 7.71 %.

HR-MS (ESI⁺): m/z calcd. for $[\text{C}_{28}\text{H}_{30}\text{ClCrN}_3\text{O}_2]$, $[\text{M}]^+$: 527.1422; found: 527.1421.

2.1.2 [(*S*)-*i*Pr(PdmBox)CrCl]



Yield: dark-violet solid (497.3. mg, 64 %).

¹H NMR (C₆D₆, 600.13 MHz, 295 K, paramagnetic): δ [ppm] = 32.03, 26.09, 19.23, 3.62, −36.95.

Magnetic Susceptibility (Evans, C₆D₆, 295 K)¹²: μ_{eff} = 4.70 μ_B.

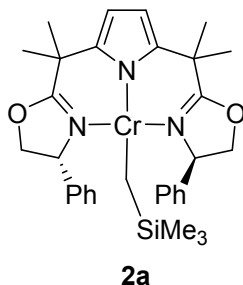
EA: calcd. C: 57.45 %, H: 7.45 %, N: 9.14 %; found: C: 57.29 %, H: 7.31 %, N: 8.91 %.

HR-MS (ESI⁺): m/z calcd. for [C₂₂H₃₂ClCrN₃O₂]⁺, [M]⁺: 459.1817; found: 459.1818.

2.2 Syntheses of the ^R(PdmBox)Cr(CH₂SiMe₃) Complexes

^R(PdmBox)CrCl complex (1.09 mmol, 1.00 eq.) was dissolved in toluene (50 mL), a solution of Mg(CH₂SiMe₃)₂(THF)₂ (1.10 mmol, 1.01 eq.) in toluene (20 mL) was added and the reaction was stirred for 12 h at rt. The reaction mixture was filtered, the solvent was removed, and the residue was redissolved in a mixture of toluene/pentane (1:10) and filtered over Celite. The filtrate was evaporated to afford ^R(PdmBox)Cr(CH₂SiMe₃) complex as dark-red solid. Single crystals suitable for X-ray diffraction were obtained from a saturated solution of **3b** in *n*-hexane at −40 °C.

2.2.1 [(*R*)-Ph(PdmBox)Cr(CH₂SiMe₃)]



Yield: dark-red solid (442.7 mg, 70 %).

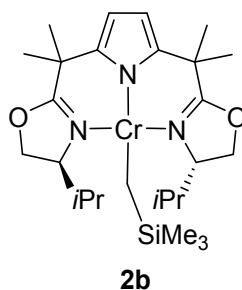
¹H NMR (C₆D₆, 600.13 MHz, 295 K, paramagnetic): δ [ppm] = 21.97, 20.21, 9.08 4.58, −24.46.

Magnetic Susceptibility (Evans, C₆D₆, 295 K)¹²: μ_{eff} = 4.73 μ_B.

EA: calcd. C: 66.29, H: 7.13, N: 7.25; found: C: 64.94, H: 6.90, N: 7.49.¹

MS (LIFDI⁺): m/z calcd. for [C₂₉H₃₁CrN₃O₂]⁺, [M-HSiMe₃]⁺: 505.6; found: 505.6.

2.2.2 [(*S*)-*i*Pr(PdmBox)Cr(CH₂SiMe₃)]



Yield: dark-red solid (409.7 mg, 73 %).

¹H NMR (C₆D₆, 600.13 MHz, 295 K, paramagnetic): δ [ppm] = 31.93, 21.99, 14.72, 6.78, 4.39, –25.59.

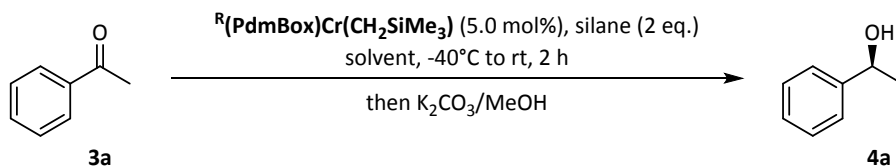
EA: calcd. C: 61.02 %, H: 8.86 %, N: 8.21 %; found: C: 59.28 %, H: 8.23 %, N: 7.93 %.¹

MS (LIFDI⁺): m/z calcd. for [C₂₃H₃₅CrN₃O₂]⁺, [M-HSiMe₃]⁺: 437.2; found: 437.2.

¹ Systematical and analytical bias due to the high air and moist sensitivity of the alkyl complexes.

3. Catalysis

3.1 Catalytic Enantioselective Reduction



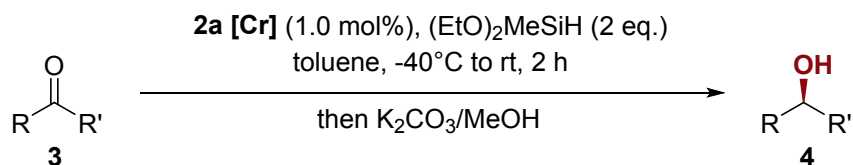
A solution of the precatalyst $R(PdmBox)Cr(CH_2SiMe_3)$ **2a** or **2b** (6.20 μ mol, 5.0 mol%; down to 0.12 μ mol, 0.1 mol%) and acetophenone **3a** (15.0 mg, 0.12 mmol, 1.0 eq.) in 1 mL solvent was cooled to $-40^\circ C$. Neat silane (0.25 mmol, 2.0 eq.) was added in one portion and the mixture was warmed to rt over a period of 2 h. The resulting silyl ether was hydrolyzed by adding a saturated solution of K_2CO_3 in MeOH (2 mL). The mixture was stirred for 1 h and filtered through a pad of silica. The residue was eluted with DCM and analyzed by chiral HPLC.

Table S1 Optimizing the reaction conditions for the hydrosilylation of ketones.

#	R	Solvent	Silane	Cat. load. [mol%]	Conv. [%] ^a	ee [%] ^a
1	(<i>R</i>)-Ph	toluene	(EtO) ₂ MeSiH	5.0	> 99	95 (<i>S</i>)
2	(<i>S</i>)- <i>i</i> Pr	toluene	(EtO) ₂ MeSiH	5.0	> 99	75 (<i>R</i>)
3	(<i>R</i>)-Ph	toluene	(EtO) ₃ SiH	5.0	> 99	83 (<i>S</i>)
4	(<i>R</i>)-Ph	toluene	<i>n</i> BuSiH ₃	5.0	> 99	84 (<i>S</i>)
5	(<i>R</i>)-Ph	toluene	PMHS ^b	5.0	58	84 (<i>S</i>)
6 ^c	(<i>R</i>)-Ph	toluene	PhSiH ₃	5.0	> 99	70 (<i>S</i>)
7 ^c	(<i>R</i>)-Ph	toluene	Ph ₂ SiH ₂	5.0	0	n.d.
8 ^c	(<i>R</i>)-Ph	toluene	Me ₂ PhSiH	5.0	0	n.d.
9	(<i>R</i>)-Ph	<i>n</i> -pentane	(EtO) ₂ MeSiH	5.0	> 99	90 (<i>S</i>)
10	(<i>R</i>)-Ph	<i>n</i> -hexane	(EtO) ₂ MeSiH	5.0	> 99	90 (<i>S</i>)
11	(<i>R</i>)-Ph	Et ₂ O	(EtO) ₂ MeSiH	5.0	> 99	90 (<i>S</i>)
12	(<i>R</i>)-Ph	THF	(EtO) ₂ MeSiH	5.0	> 99	86 (<i>S</i>)
13	(<i>R</i>)-Ph	tmeda	(EtO) ₂ MeSiH	5.0	> 99	80 (<i>S</i>)
14	(<i>R</i>)-Ph	DCM	(EtO) ₂ MeSiH	5.0	0	n.d.
15	(<i>R</i>)-Ph	MeCN	(EtO) ₂ MeSiH	5.0	0	n.d.
16	(<i>R</i>)-Ph	toluene	(EtO) ₂ MeSiH	2.5	> 99	95 (<i>S</i>)
17	(<i>R</i>)-Ph	toluene	(EtO) ₂ MeSiH	1.0	> 99	95 (<i>S</i>)
18	(<i>R</i>)-Ph	toluene	(EtO) ₂ MeSiH	0.5	> 99	95 (<i>S</i>)
19	(<i>R</i>)-Ph	toluene	(EtO) ₂ MeSiH	0.25	53	95 (<i>S</i>)
20	(<i>R</i>)-Ph	toluene	(EtO) ₂ MeSiH	0.1	0	n.d.

^aDetermined by chiral HPLC. ^bPMHS = Poly(methylhydrosiloxane). ^cWork-up with NaOH in *i*PrOH.

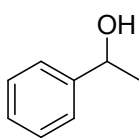
3.2 Enantioselective Hydrosilylation of Ketones



A solution of the precatalyst **2a** (0.72mg, 5.2μmol, 1.0 mol%) and the substrate **3** (0.52mmol, 1.0 eq.) in 1 mL toluene was cooled to -40 °C in a cold bath. Neat (EtO)₂MeSiH (168 μl, 1.04 mmol, 2.0 eq.) was added in one portion and the mixture was allowed to warm to rt in that bath over a period of 2 h. The resulting silyl ether was hydrolyzed by adding a saturated solution of K₂CO₃ in MeOH (2 mL). The mixture was stirred for 1 h and filtered through a pad of silica. The residue was eluted with DCM. The organic layer was brined (5 mL) and the aqueous phase was washed with ethyl acetate (2 × 3 mL). The resulting alcohol **5** was purified by FCC (SiO₂, PE/EA 10:1) and analyzed by NMR spectroscopy and chiral HPLC or chiral GC. The absolute configuration was determined by comparison with reported data and commercially available samples.¹³

3.3 Characterization of Alcohols

4a 1-Phenylethanol



Yield: colorless liquid (56.2 mg, 89 %).

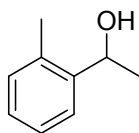
¹H NMR (CDCl₃, 600.24 MHz, 295 K): δ [ppm] = 7.36-7.32 (m, 4H), 7.26-7.23 (m, 1H), 4.87 (q, ³J_{H,H} = 6.5 Hz, 1H), 1.91 (s, 1H), 1.48 (d, ³J_{H,H} = 6.5 Hz, 1H).

¹³C NMR (CDCl₃, 150.93 MHz, 295 K): δ [ppm] = 145.8, 128.5, 127.5, 125.4, 70.4, 25.1.

Chromatography: Chiralcel OD-H (Hexane/*i*PrOH 98:2, 1.0 mL/min, 20 °C, λ = 210 nm); t_R = 15.2 min (*R*), t_R = 19.9 min (*S*); 95 %ee (*S*).

4b

1-(2-Tolyl)ethanol



Yield: colorless liquid (61.8 mg, 87 %).

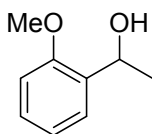
¹H NMR (CDCl₃, 600.24 MHz, 295K): δ [ppm] = 7.50 (d, ³J_{H,H} = 7.5 Hz, 1H), 7.25-7.22 (m, 1H), 7.19-7.13 (m, 2H), 5.15 (q, ³J_{H,H} = 6.5 Hz, 1H), 2.35 (s, 3H), 1.66 (s, 1H), 1.47 (d, ³J_{H,H} = 6.5 Hz, 1H).

¹³C NMR (CDCl₃, 150.93 MHz, 295K): δ [ppm] = 143.8, 134.3, 130.4, 127.2, 126.4, 124.5, 66.8, 23.9, 18.9.

Chromatography: Chiralpak AD-H (Hexane/*i*PrOH 98:2, 1.0 mL/min, 20 °C, λ = 210 nm); t_R = 14.2 min (*R*), t_R = 16.4 min (*S*); 98 %ee (*S*).

4c

1-(2-Methoxyphenyl)ethanol



Yield: colorless solid (69.9 mg, 89 %).

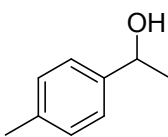
¹H NMR (CDCl₃, 600.24 MHz, 295K): δ [ppm] = 7.33-7.32 (m, 1H), 7.26-7.23 (m, 1H), 6.96-6.95 (m, 1H), 6.88 (d, ³J_{H,H} = 8.2 Hz, 1H), 5.09 (q, ³J_{H,H} = 6.5 Hz, 1H), 3.87 (s, 3H), 2.55 (s, 1H), 1.50 (d, ³J_{H,H} = 6.5 Hz, 1H).

¹³C NMR (CDCl₃, 150.93 MHz, 295K): δ [ppm] = 166.9, 150.9, 129.8, 129.3, 125.3, 70.0, 52.1, 25.3.

Chromatography: Chiralcel OD-H (Hexane/*i*PrOH 99:1, 1.0 mL/min, 25 °C, λ = 210 nm); t_R = 25.6 min (*S*), t_R = 27.9 min (*R*); 94 %ee (*S*).

4d

1-(4-Tolyl)ethanol



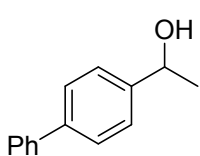
Yield: colorless liquid (63.7 mg, 91 %).

¹H NMR (CDCl₃, 600.24 MHz, 295K): δ [ppm] = 7.24 (d, ³J_{H,H} = 8.0 Hz, 2H), 7.15 (d, ³J_{H,H} = 8.0 Hz, 2H), 4.85 (q, ³J_{H,H} = 6.5 Hz, 1H), 2.32 (s, 3H), 1.75 (s, 1H), 1.46 (d, ³J_{H,H} = 6.5 Hz, 1H).

¹³C NMR (CDCl₃, 150.93 MHz, 295K): δ [ppm] = 142.8, 137.2, 129.2, 125.4, 70.3, 25.1, 21.1.

Chromatography: Chiralcel OD-H (Hexane/*i*PrOH 98:2, 1.0mL/min, 20 °C, λ = 210 nm); t_R = 15.1 min (*S*), t_R = 17.8 min (*R*); 98 %ee (*S*).

4e 1-([1,1'-Biphenyl]-4-yl)ethanol



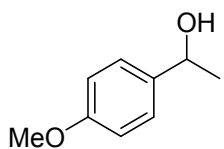
Yield: colorless solid (94.7 mg, 92 %).

¹H NMR (CDCl₃, 600.24 MHz, 295K): δ [ppm] = 7.60-7.58 (m, 4H), 7.46-7.42 (m, 4H), 7.36-7.33 (m, 1H), 4.97 (q, ³J_{H,H} = 6.5 Hz, 1H), 1.75 (s, 1H), 1.55 (d, ³J_{H,H} = 6.5 Hz, 1H).

¹³C NMR (CDCl₃, 150.93 MHz, 295K): δ [ppm] = 144.8, 140.9, 140.5, 128.8, 127.3, 127.1, 125.8, 70.2, 25.2.

Chromatography: Chiralpak AD-H (Hexane/*i*PrOH 95:5, 0.8 mL/min, 20 °C, λ = 230 nm); t_R = 16.1 min (*S*), t_R = 17.6 min (*R*); 90 %ee (*S*).

4f 1-(4-Methoxyphenyl)ethanol



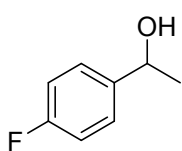
Yield: yellow liquid (73.1 mg, 91 %).

¹H NMR (CDCl₃, 600.24 MHz, 295K): δ [ppm] = 7.27 (d, ³J_{H,H} = 8.2 Hz, 2H), 6.89 (d, ³J_{H,H} = 8.0 Hz, 2H), 4.87 (q, ³J_{H,H} = 6.5 Hz, 1H), 3.81 (s, 3H), 1.72 (s, 1H), 1.50 (d, ³J_{H,H} = 6.5 Hz, 1H).

¹³C NMR (CDCl₃, 150.93 MHz, 295K): δ [ppm] = 158.5, 137.7, 126.4, 113.6, 69.8, 55.3, 25.1.

Chromatography: Chiralcel OD-H (Hexane/*i*PrOH 98:2, 1.0 mL/min, 10 °C, λ = 210 nm); t_R = 25.1 min (*R*), t_R = 26.8 min (*S*); 90 %ee (*S*).

4g 1-(4-Fluorophenyl)ethanol



Yield: colorless liquid (64.3 mg, 87 %).

^1H NMR (CDCl_3 , 600.24 MHz, 295K): δ [ppm] = 7.36-7.32 (m, 2H), 7.05-7.00 (m, 2H), 4.90 (q, $^3J_{\text{H,H}} = 6.4$ Hz, 1H), 1.82 (s, 1H), 1.48 (d, $^3J_{\text{H,H}} = 6.5$ Hz, 1H).

^{13}C NMR (CDCl_3 , 150.93 MHz, 295K): δ [ppm] = 162.1 (d, $J = 262.0$ Hz), 141.4 (d, $J = 3.4$ Hz), 127.9 (d, $J = 9.7$ Hz), 115.3 (d, $J = 23.7$ Hz), 69.8, 25.3.

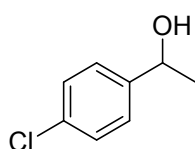
^{19}F NMR (CDCl_3 , 150.93 MHz, 295K): δ [ppm] = -115.3.

Chromatography: Chiraldex BP-M (mode: isothermal, 170 kPa, 120 °C);

$t_R = 16.4$ min (*S*), $t_R = 16.9$ min (*R*); 90 %ee (*S*).

4h

1-(4-Chlorophenyl)ethanol



Yield: slightly yellow liquid (71.4 mg, 88 %).

^1H NMR (CDCl_3 , 600.24 MHz, 295K): δ [ppm] = 7.24-7.20 (m, 4H), 4.79 (q, $^3J_{\text{H,H}} = 6.5$ Hz, 1H), 2.07 (s, 1H), 1.40 (d, $^3J_{\text{H,H}} = 6.5$ Hz, 1H).

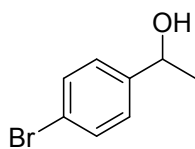
^{13}C NMR (CDCl_3 , 150.93 MHz, 295K): δ [ppm] = 144.3, 133.0, 128.6, 126.8, 69.7, 25.3.

Chromatography: Chiralcel OD-H (Hexane/*i*PrOH 95:5, 0.7 mL/min, 20 °C,

$\lambda = 210$ nm); $t_R = 11.3$ min (*S*), $t_R = 12.1$ min (*R*); 89 %ee (*S*).

4i

1-(4-Bromophenyl)ethanol



Yield: yellow liquid (97.2 mg, 92 %).

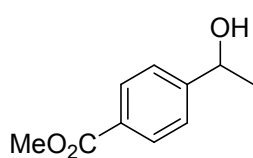
^1H NMR (CDCl_3 , 600.24 MHz, 295K): δ [ppm] = 7.47 (d, $^3J_{\text{H,H}} = 8.0$ Hz, 2H), 7.26 (d, $^3J_{\text{H,H}} = 8.0$ Hz, 2H), 4.87 (q, $^3J_{\text{H,H}} = 6.5$ Hz, 1H), 1.76 (s, 1H), 1.47 (d, $^3J_{\text{H,H}} = 6.5$ Hz, 1H).

^{13}C NMR (CDCl_3 , 150.93 MHz, 295K): δ [ppm] = 144.7, 131.6, 127.2, 121.2, 69.8, 25.3.

Chromatography: Chiralcel OD-H (Hexane/*i*PrOH 95:5, 1.0 mL/min, 20 °C,

$\lambda = 210$ nm); $t_R = 8.6$ min (*S*), $t_R = 9.2$ min (*R*); 86 %ee (*S*).

4j

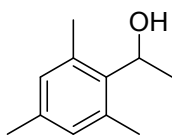
Methyl 4-(1-Hydroxyethyl)benzoate**Yield:** colorless liquid (29.1 mg, 31 %).

¹H NMR (CDCl₃, 600.24 MHz, 295K): δ [ppm] = 8.02 (d, ³J_{H,H} = 8.2 Hz, 2H), 7.45 (d, ³J_{H,H} = 8.0 Hz, 2H), 4.96 (q, ³J_{H,H} = 6.5 Hz, 1H), 3.91 (s, 3H), 1.82 (s, 1H), 1.50 (d, ³J_{H,H} = 6.5 Hz, 1H).

¹³C NMR (CDCl₃, 150.93 MHz, 295K): δ [ppm] = 166.9, 150.9, 129.8, 129.3, 125.3, 70.0, 52.1, 25.3.

Chromatography: Chiralpak AD-H (Hexane/*i*PrOH 95:5, 1.0 mL/min, 20 °C, λ = 215 nm); t_R = 17.2 min (*R*), t_R = 18.4 min (*S*); 50 %ee (*S*).

4k

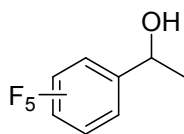
1-(2,4,6-Trimethylphenyl)ethanol**Yield:** colorless solid (72.6 mg, 86 %).

¹H NMR (CDCl₃, 600.24 MHz, 295K): δ [ppm] = 6.82 (s, 2H), 5.36 (q, ³J_{H,H} = 6.5 Hz, 1H), 2.41 (s, 6H), 2.25 (s, 3H), 1.62 (s, 1H), 1.52 (d, ³J_{H,H} = 6.5 Hz, 1H).

¹³C NMR (CDCl₃, 150.93 MHz, 295K): δ [ppm] = 137.6, 136.5, 135.7, 130.2, 67.5, 52.1, 21.6, 20.7, 20.5.

Chromatography: Chiralcel OD-H (Hexane/*i*PrOH 99:1, 0.8 mL/min, 20 °C, λ = 210 nm); t_R = 21.9 min (*S*), t_R = 25.5 min (*R*); 85 %ee (*S*).

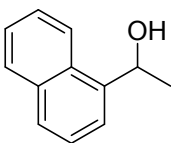
4l

1-(2,3,4,5,6-Pentafluorophenyl)ethanol**Yield:** no conv.

Chromatography: Chiralpak AD-H (Hexane/*i*PrOH 99:1, 1.0 mL/min, 20 °C, λ = 210 nm); t_R = 16.7 min (*S*), t_R = 20.1 min (*R*).

4m

1-(Naphthalen-1-yl)ethanol



Yield: colorless solid (80.1 mg, 90 %).

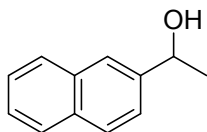
¹H NMR (CDCl₃, 600.24 MHz, 295K): δ [ppm] = 8.10 (d, ³J_{H,H} = 8.2 Hz, 1H), 7.88 (d, ³J_{H,H} = 8.2 Hz, 1H), 7.78 (d, ³J_{H,H} = 8.2 Hz, 1H), 7.67 (d, ³J_{H,H} = 8.2 Hz, 1H), 7.54-7.46 (m, 3H), 5.64 (q, ³J_{H,H} = 6.5 Hz, 1H), 2.21 (s, 1H), 1.66 (d, ³J_{H,H} = 6.5 Hz, 3H).

¹³C NMR (CDCl₃, 150.93 MHz, 295K): δ [ppm] = 141.4, 133.8, 130.3, 128.9, 127.9, 127.9, 126.0, 125.6, 123.2, 122.0, 67.1, 24.3.

Chromatography: Chiralcel OD-H (Hexane/*i*PrOH 90:10, 0.8 mL/min, 20 °C, λ = 210 nm); t_R = 12.3 min (*R*), t_R = 19.5 min (*S*); 96 %ee (*S*).

4n

1-(Naphthalen-2-yl)ethanol



Yield: colorless solid (79.3 mg, 88 %).

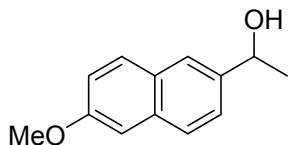
¹H NMR (CDCl₃, 600.24 MHz, 295K): δ [ppm] = 7.85-7.81 (m, 4H), 7.52-7.45 (m, 3H), 5.08 (q, ³J_{H,H} = 6.5 Hz, 1H), 1.88 (s, 1H), 1.59 (d, ³J_{H,H} = 6.5 Hz, 3H).

¹³C NMR (CDCl₃, 150.93 MHz, 295K): δ [ppm] = 143.1, 133.2, 132.9, 128.3, 127.9, 127.7, 126.1, 125.8, 123.8, 70.6, 52.1, 25.2.

Chromatography: Chiralcel OD-H (Hexane/*i*PrOH 95:5, 1.0 mL/min, 20 °C, λ = 210 nm); t_R = 16.5 min (*R*), t_R = 17.8 min (*S*); 90 %ee (*S*).

4o

9-Methoxy-1-(Naphthalen-2-yl)ethanol



Yield: colorless solid (95.8 mg, 90 %).

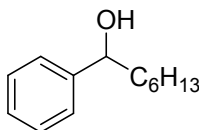
¹H NMR (CDCl₃, 600.24 MHz, 295K): δ [ppm] = 7.74-7.71 (m, 3H), 7.48-7.46 (m, 1H), 7.16-7.12 (m, 2H), 5.04 (q, ³J_{H,H} = 6.5 Hz, 1H), 3.91 (s, 3H), 1.89 (s, 1H), 1.57 (d, ³J_{H,H} = 6.5 Hz, 3H).

¹³C NMR (CDCl₃, 150.93 MHz, 295K): δ [ppm] = 157.6, 140.9, 134.0, 129.4, 128.7, 127.2, 124.4, 123.8, 118.9, 70.5, 55.2, 20.7, 25.1.

Chromatography: Chiralcel OD-H (Hexane/*i*PrOH 95:5, 0.8 mL/min, 20 °C, $\lambda = 210$ nm); $t_R = 19.5$ min (*S*), $t_R = 28.0$ min (*R*); 92 %ee (*S*).

4p

1-Phenylheptanol



Yield: colorless liquid (89.3 mg, 90 %).

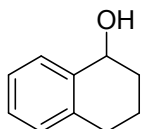
^1H NMR (CDCl_3 , 600.24 MHz, 295 K): δ [ppm] = 7.35-7.26 (m, 5H), 4.66 (dd, $^3J_{\text{H,H}} = 7.0$ Hz, 1H), 1.83-1.77 (m, 1H), 1.73-1.66 (m, 1H), 1.58 (s, 1H), 1.43-1.37 (m, 1H), 1.33-1.23 (m, 7H), 0.87 (t, $^3J_{\text{H,H}} = 6.5$ Hz, 3H).

^{13}C NMR (CDCl_3 , 150.93 MHz, 295 K): δ [ppm] = 144.9, 128.4, 127.5, 125.9, 74.7, 39.1, 31.7, 29.2, 25.8, 22.6.

Chromatography: Chiralcel OD-H (Hexane/*i*PrOH 99:1, 0.8 mL/min, 20 °C, $\lambda = 210$ nm); $t_R = 18.1$ min (*R*), $t_R = 22.8$ min (*S*); 86 %ee (*S*).

4q

1,2,3,4-Tetrahydronaphthalen-1-ol



Yield: yellow liquid (72.8 mg, 93 %).

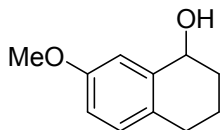
^1H NMR (CDCl_3 , 600.24 MHz, 295 K): δ [ppm] = 7.46-7.44 (m, 1H), 7.25-7.22 (m, 2H), 7.15-7.12 (m, 1H), 4.82 (t, $^3J_{\text{H,H}} = 4.5$ Hz, 1H), 2.89-2.73 (m, 2H), 2.06-1.92 (m, 3H), 1.84-1.79 (m, 1H), 1.76 (s, 1H).

^{13}C NMR (CDCl_3 , 150.93 MHz, 295 K): δ [ppm] = 138.7, 137.1, 129.0, 128.7, 127.6, 126.2, 68.1, 32.3, 29.3, 18.8.

Chromatography: Chiralcel OD-H (Hexane/*i*PrOH 99:1, 1.0 mL/min, 20 °C, $\lambda = 215$ nm); $t_R = 23.9$ min (*S*), $t_R = 27.5$ min (*R*); 93 %ee (*S*).

4r

7-Methoxy-1,2,3,4-tetrahydronaphthalen-1-ol



Yield: yellowish solid (77.2 mg, 91 %).

^1H NMR (CDCl_3 , 600.24 MHz, 295 K): δ [ppm] = 7.02 (d, $^3J_{\text{H,H}} = 8.3$ Hz, 1H), 6.96 (d, $^3J_{\text{H,H}} = 2.4$ Hz, 1H), 6.78 (dd, $^3J_{\text{H,H}} = 8.4$ Hz, $^3J_{\text{H,H}} = 2.4$ Hz, 1H), 4.74

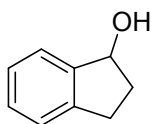
(t, $^3J_{\text{H,H}} = 5.4$ Hz, 1H), 3.79 (s, 3H), 2.77-2.63 (m, 2H), 2.04-1.91 (m, 2H), 1.88-1.84 (m, 1H), 1.78-1.74 (m, 1H), 1.67 (s, 1H).

^{13}C NMR (CDCl_3 , 150.93 MHz, 295 K): δ [ppm] = 157.6, 139.5, 129.7, 128.9, 114.2, 112.4, 68.5, 55.4, 32.5, 28.6, 19.3.

Chromatography: Chiralcel OD-H (Hexane/*i*PrOH 95:5, 0.7 mL/min, 20 °C, $\lambda = 215$ nm); $t_R = 42.8$ min (*S*), $t_R = 50.2$ min (*R*); 87 %ee (*S*).

4s

2,3-Dihydro-1H-inden-1-ol



Yield: slightly yellow liquid (69.4 mg, 89 %).

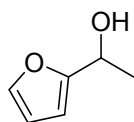
^1H NMR (CDCl_3 , 600.24 MHz, 295 K): δ [ppm] = 7.42 (d, $^3J_{\text{H,H}} = 6.5$ Hz, 1H), 7.28-7.23 (m, 3H), 5.26 (t, $^3J_{\text{H,H}} = 6.2$ Hz, 1H), 3.09-3.01 (m, 1H), 2.85-2.80 (m, 1H), 2.52-2.48 (m, 1H), 1.99-1.92 (m, 1H), 1.64 (s, 1H).

^{13}C NMR (CDCl_3 , 150.93 MHz, 295 K): δ [ppm] = 144.9, 143.4, 128.4, 126.7, 124.9, 124.2, 76.5, 36.0, 29.8.

Chromatography: Chiralcel OD-H (Hexane/*i*PrOH 98:2, 1.0 mL/min, 20 °C, $\lambda = 210$ nm); $t_R = 17.6$ min (*S*), $t_R = 20.1$ min (*R*); 94 %ee (*S*).

4t

1-(Furan-2-yl)ethanol



Yield: red liquid (40.7 mg, 67 %).

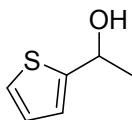
^1H NMR (CDCl_3 , 600.24 MHz, 295 K): δ [ppm] = 7.37 (m, 1H), 6.32 (dd, $^3J_{\text{H,H}} = 6.5$ Hz, $^3J_{\text{H,H}} = 3.7$ Hz, 1H), 6.22 (d, $^3J_{\text{H,H}} = 6.5$ Hz, 1H), 4.88 (q, $^3J_{\text{H,H}} = 6.5$ Hz, 1H), 2.56 (s, 1H), 1.54 (d, $^3J_{\text{H,H}} = 6.4$ Hz, 3H).

^{13}C NMR (CDCl_3 , 150.93 MHz, 295 K): δ [ppm] = 149.3, 126.2, 124.4, 124.0, 123.5, 69.3, 65.3, 25.2.

Chromatography: Chiralcel OJ-H (Hexane/*i*PrOH 99:1, 1.0 mL/min, 20 °C, $\lambda = 215$ nm); $t_R = 30.3$ min (*S*), $t_R = 33.3$ min (*R*); 90 %ee (*S*).

4u

1-(Thiophen-2-yl)ethanol



Yield: yellow liquid (59.3 mg, 90 %).

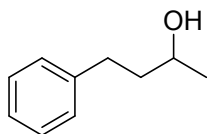
¹H NMR (CDCl₃, 600.24 MHz, 295 K): δ [ppm] = 7.24 (dd, $^3J_{\text{H,H}} = 6.5$ Hz, $^3J_{\text{H,H}} = 3.7$ Hz, 1H), 6.99-6.93 (m, 2H), 5.14 (q, $^3J_{\text{H,H}} = 6.4$ Hz, 1H), 1.99 (s, 1H), 1.61 (d, $^3J_{\text{H,H}} = 6.4$ Hz, 3H).

¹³C NMR (CDCl₃, 150.93 MHz, 295 K): δ [ppm] = 149.6, 126.7, 124.5, 124.0, 123.6, 70.3, 66.3, 25.3.

Chromatography: Chiralcel OJ-H (Hexane/*i*PrOH 99:1, 1.0 mL/min, 20 °C, $\lambda = 215$ nm); $t_R = 38.7$ min (*S*), $t_R = 46.7$ min (*R*); 90 %ee (*S*).

4v

4-Phenylbutan-2-ol



Yield: colorless liquid (63.2 mg, 84 %).

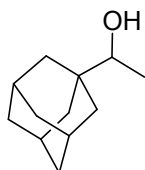
¹H NMR (CDCl₃, 600.24 MHz, 295 K): δ [ppm] = 7.32-7.30 (m, 2H), 7.23-7.20 (m, 3H), 3.86 (sext., $^3J_{\text{H,H}} = 6.2$ Hz, 1H), 2.81-2.76 (m, 1H), 2.72-2.67 (m, 1H), 1.85-1.75 (m, 2H), 1.59 (s, 1H), 1.26 (d, $^3J_{\text{H,H}} = 6.2$ Hz, 3H).

¹³C NMR (CDCl₃, 150.93 MHz, 295 K): δ [ppm] = 142.1, 128.4, 125.8, 67.5, 40.8, 32.2, 23.6.

Chromatography: Chiralpak AD-H (Hexane/*i*PrOH 96:4, 0.7 mL/min, 10 °C, $\lambda = 215$ nm); $t_R = 13.8$ min (*R*), $t_R = 14.6$ min (*S*); 30 %ee (*S*).

4w

1-(Adamantan-1-yl)ethanol



Yield: colorless liquid (76.3 mg, 81 %).

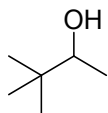
¹H NMR (CDCl₃, 600.24 MHz, 295 K): δ [ppm] = 3.28 (q, $^3J_{\text{H,H}} = 6.5$ Hz, 1H), 1.99 (m, 3H), 1.72-1.63 (m, 6H), 1.60-1.57 (m, 3H), 1.49-1.46 (m, 3H), 1.42 (s, 1H), 1.09 (d, $^3J_{\text{H,H}} = 6.5$ Hz, 3H).

¹³C NMR (CDCl₃, 150.93 MHz, 295 K): δ [ppm] = 75.8, 37.7, 37.2, 36.6, 28.3, 16.5.

Chromatography: Analysis of the benzoic ester, Chiralpak AD-H (Hexane/*i*PrOH 98:2, 1.0 mL/min, 25 °C, λ = 230 nm); t_R = 5.0 min (*R*), t_R = 5.7 min (*S*); 95 %ee (*S*).

4x

3,3-Dimethylbutan-2-ol



Yield: colorless liquid (32.4 mg, 62 %).

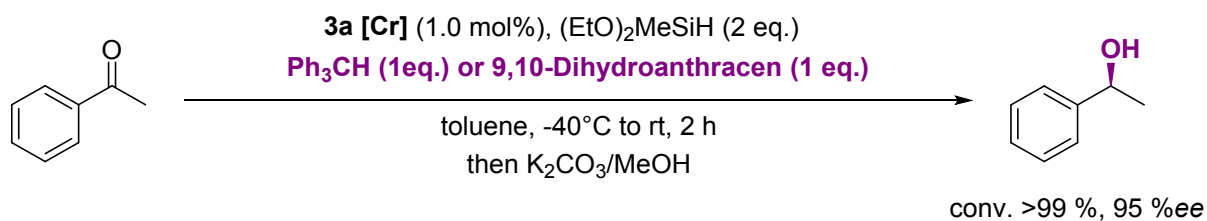
^1H NMR (CDCl_3 , 600.24 MHz, 295 K): δ [ppm] = 3.46 (q, $^3J_{\text{H,H}}$ = 6.5 Hz, 1H), 1.62 (s, 1H), 1.17 (d, $^3J_{\text{H,H}}$ = 6.5 Hz, 3H), 0.89 (m, 9H).

^{13}C NMR (CDCl_3 , 150.93 MHz, 295 K): δ [ppm] = 75.6, 34.9, 25.5, 17.9.

Chromatography: Analysis of the benzoic ester Chiralcel OD-H (Hexane/*i*PrOH 100:0, 0.5 mL/min, 25 °C, λ = 230 nm); t_R = 10.0 min (*R*), t_R = 10.7 min (*S*); 90 %ee (*S*).

4. Mechanistic Studies

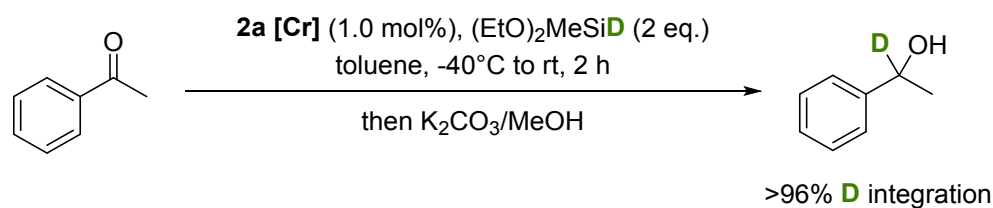
4.1 Radical Trap Experiments



The reduction of acetophenone **3a** (15.0 mg, 124 μ mol, 1.0 eq.) was accomplished by the general procedure with a radical trap reagent (triphenylmethane (124 μ mol, 1.0 eq.) or 9,10-dihydroanthracene (124 μ mol, 1.0 eq.)). The obtained product was analyzed by chiral HPLC.

Chromatography: Chiralcel OD-H (Hexane/*i*PrOH 98:2, 1.0 mL/min, 20 °C, λ = 210 nm); t_R = 15.2 min (*R*), t_R = 19.9 min (*S*); 95 % ee (*S*).

4.2 Labelling Experiments



The reduction of acetophenone **3a** (15.0 mg, 124 μ mol, 1.0 eq.) was carried out analogous to the general procedure with deuterated diethoxymethylsilane (42.0 μ l, 240 μ mol, 2.0 eq., 2% residual H). The obtained product **4a-d₁** was characterized by NMR spectroscopy.

¹H NMR (CDCl₃, 600.24 MHz, 295 K): δ [ppm] = 7.36-7.23 (m, 5H), 4.88 (q, ³*J*_{H,H} = 6.5 Hz, 0.037 H), 1.91 (s, 1H), 1.48 (d, ³*J*_{H,H} = 6.5 Hz, 1H).

^2H NMR (CDCl_3 , 92.12 MHz, 295 K): δ [ppm] = 4.87.

^{13}C NMR (CDCl_3 , 150.93 MHz, 295 K): δ [ppm] = 145.8, 128.5, 127.5, 125.4, 68.9 (t, $^2J_{\text{D,C}} = 22$ Hz), 25.1.

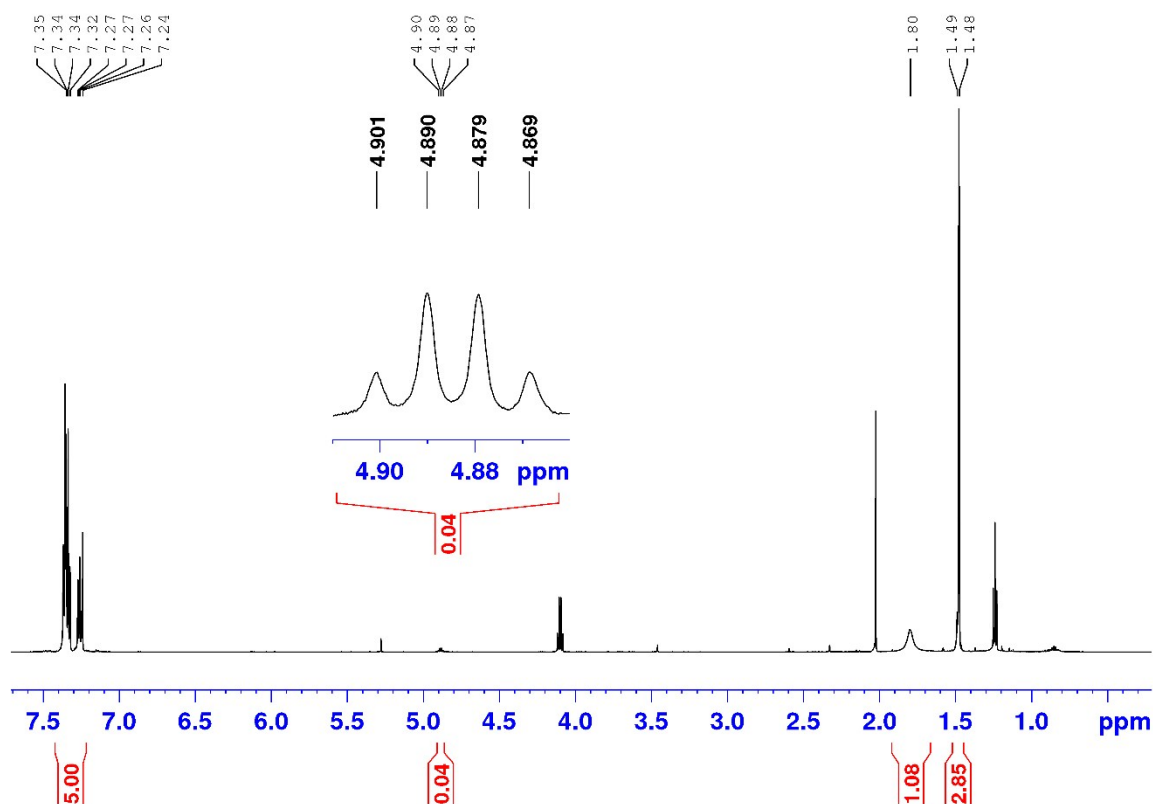
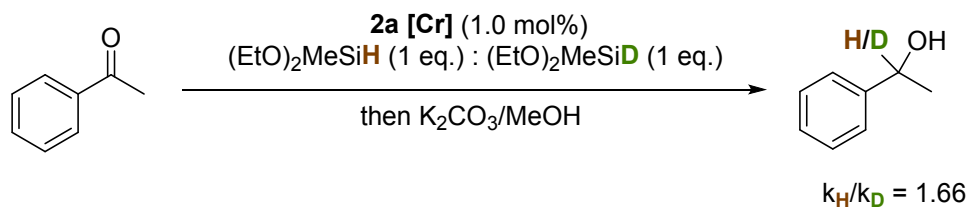


Figure S1: ^1H NMR spectrum of the deuterated acetophenone **4a-d₁** with zoom at the quartet region at 4.88 ppm.

4.3 Kinetic Isotope Effect



The reduction of acetophenone **3a** (15.0 mg, 124 μmol , 1.0 eq.) was carried out analogous to the general procedure with a mixture of equal amounts of diethoxymethylsilane (20.0 μL , 124 μmol , 1.0 eq.) and deuterated diethoxymethylsilane (20.0 μL , 124 μmol , 1.0 eq.). The ratio of deuterated to normal product was determined by ^1H NMR spectroscopy. The KIE was calculated using the following equation.

$$KIE = \frac{k_H}{k_D} = \frac{\ln(1 - F_H)}{\ln(1 - F_D)}$$

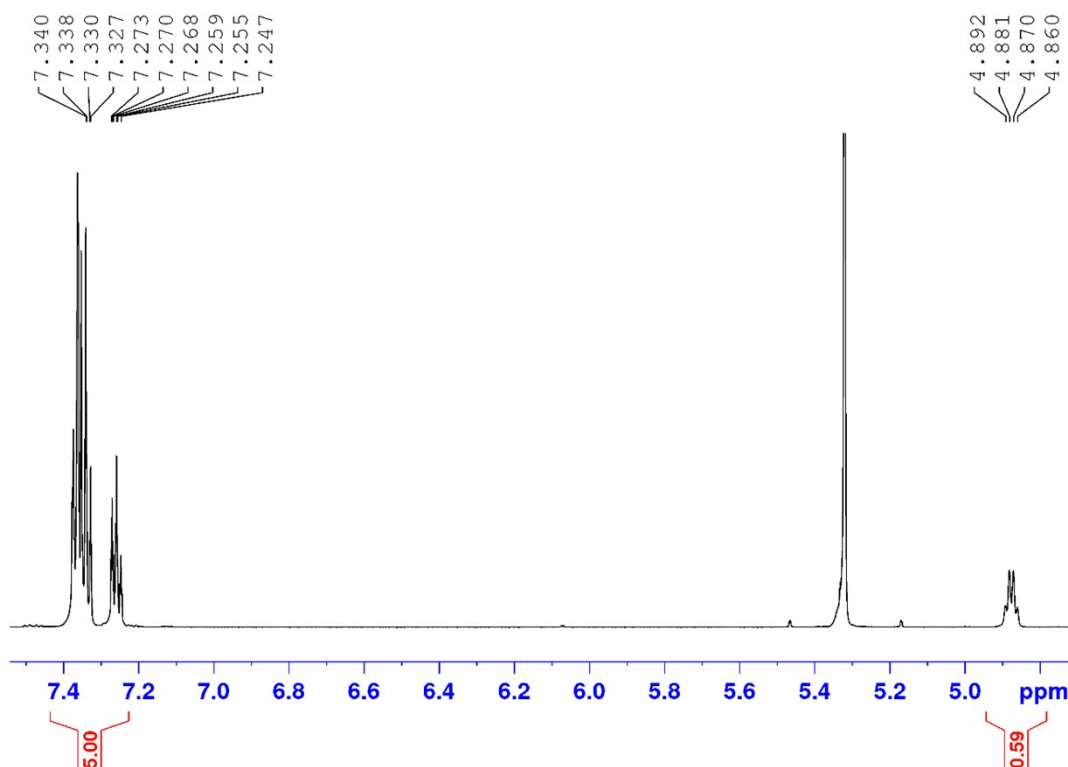


Figure S2: Extract of the ^1H NMR spectrum of the partially deuterated 1-phenylethanol (CD_2Cl_2 , 600.24 MHz, 295 K).

KIEs of *p*-Br and *p*-OMe derivatives were calculated in analogy to the protocol outlined above.

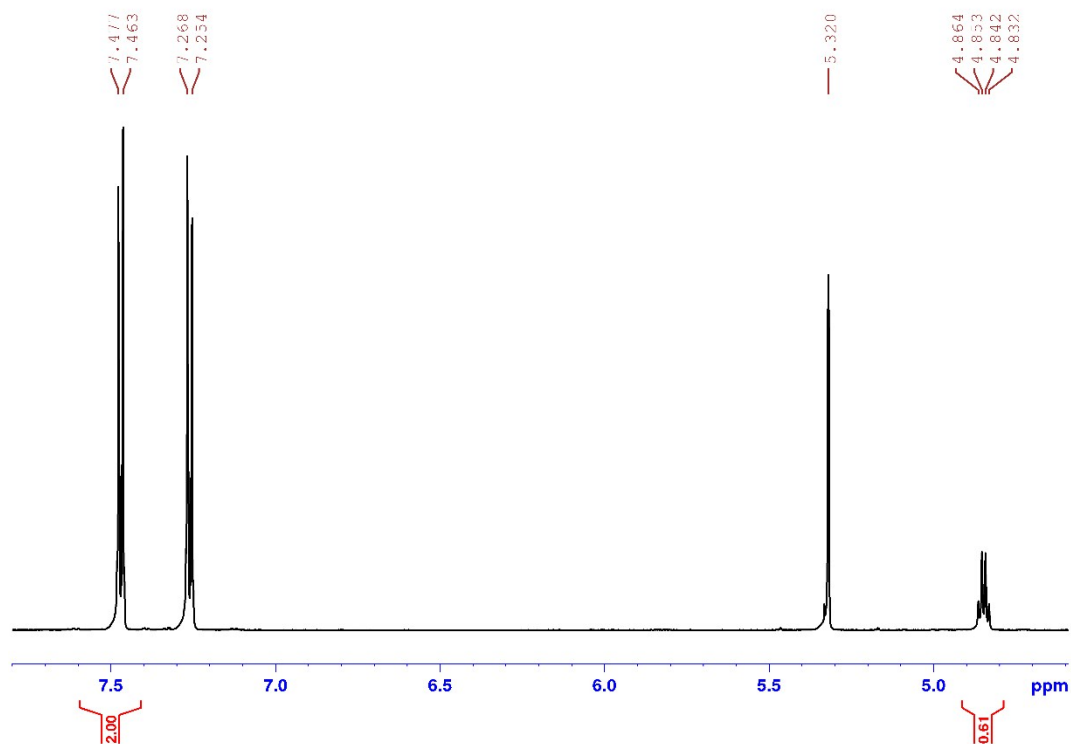


Figure S3: Extract of the ^1H NMR spectrum of the partially deuterated 1-(4-bromophenyl)ethanol (CD_2Cl_2 , 600.24 MHz, 295 K).

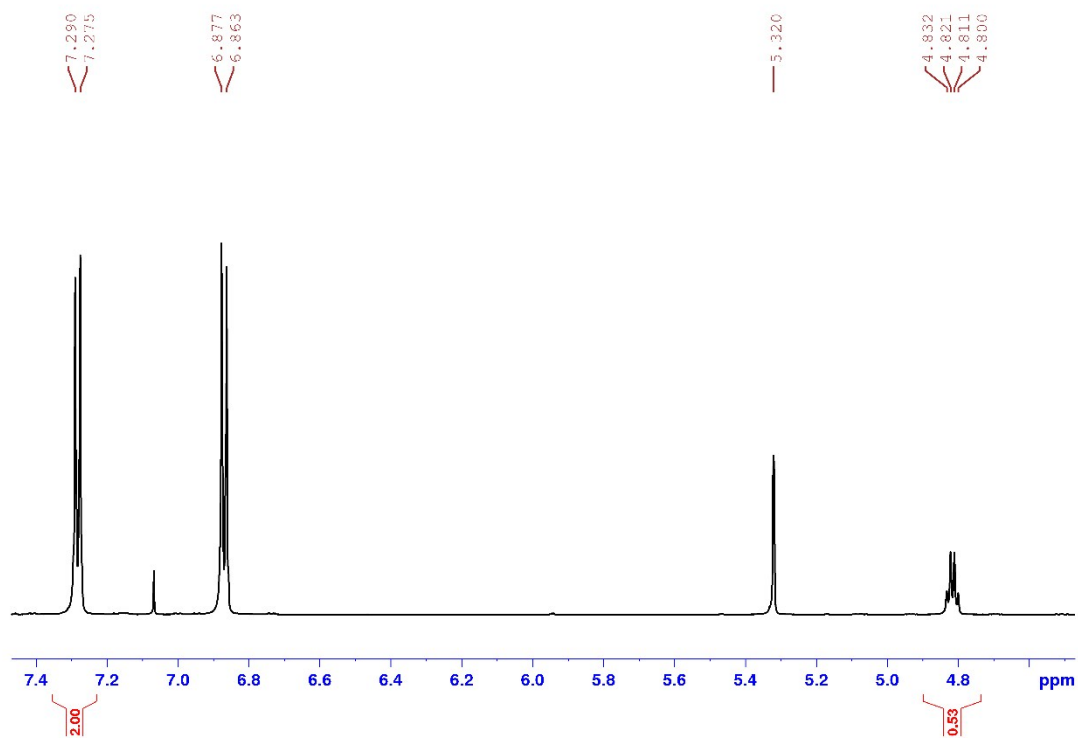


Figure S4: Extract of the ^1H NMR spectrum of the partially deuterated 1-(4-methoxyphenyl)ethanol (CD_2Cl_2 , 600.24 MHz, 295 K).

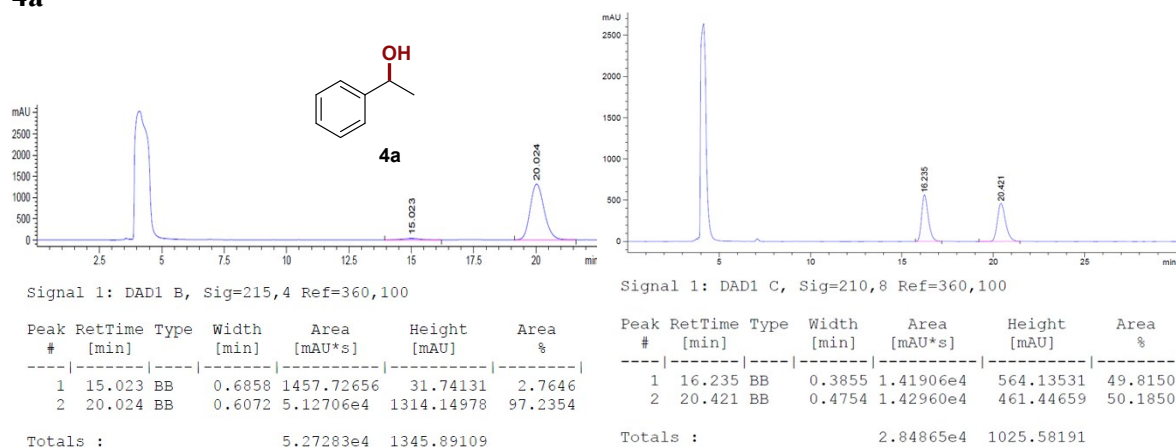
4.4 Hammett-Correlation

The precatalyst (0.72 mg, 2.6 μmol , 1.0 mol%), acetophenone 12 (15.0 mg, 124 μmol , 1.0 eq.) and one of each para substituted ketone (20mg – 35 mg each, 140 μmol – 180 μmol each, 1.0 eq. each) were dissolved in benzene- d_6 (0.5 mL). Diethoxymethylsilane (20.0 μl , 124 μmol) was added and the reaction was stirred for 2 h at rt. Afterwards, three drops of a tetrabutylammonium fluoride solution (1.0 M in THF) were added to each sample. Subsequently, all samples were analyzed by ^{13}C NMR spectroscopy (D_1 -time = 5 s). The relative rate constants were determined from the NMR integral ratio of the product peaks using the following equation. The σ -values were taken from literature.¹⁴

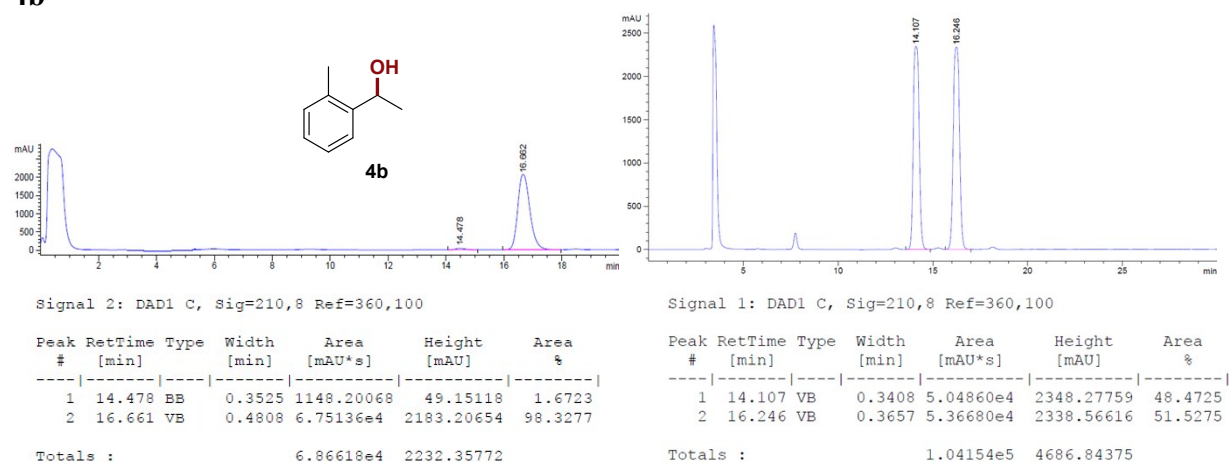
$$\frac{k_R}{k_H} = \frac{\ln \left(\frac{c_{R,t}}{c_{R,t=0}} \right)}{\ln \left(\frac{c_{H,t}}{c_{H,t=0}} \right)}$$

5. Chromatographic Data

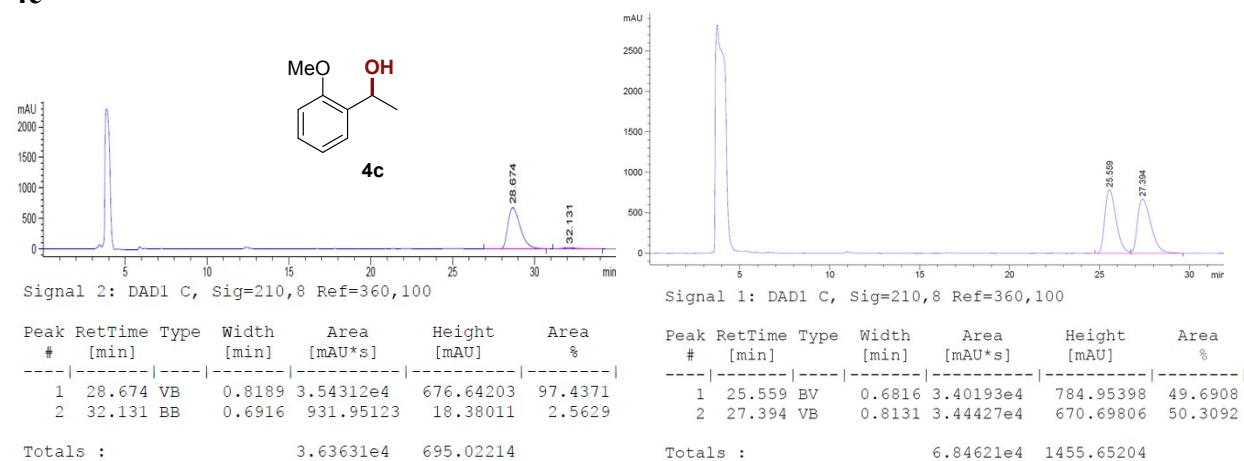
4a



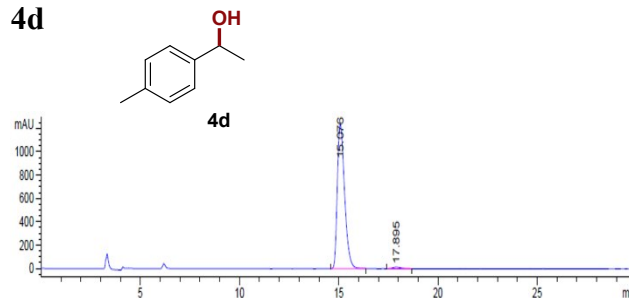
4b



4c



4d

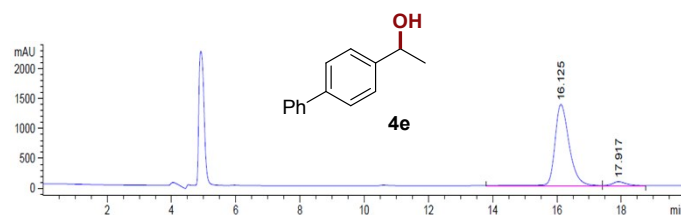


Signal 2: DAD1 C, Sig=210,8 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	15.076	BB	0.3968	3.08068e4	1211.06824	98.3216
2	17.884	BV	0.4845	525.88599	15.71191	1.6784

Totals : 3.13327e4 1226.78015

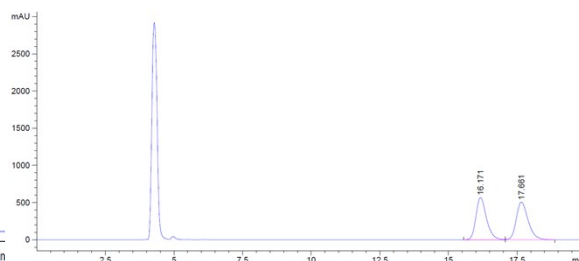
4e



Signal 1: DAD1 B, Sig=230,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	16.125	BB	0.4665	4.15811e4	1360.79980	95.7412
2	17.917	BB	0.4309	1849.61389	65.21682	4.2588

Totals : 4.34307e4 1426.01662

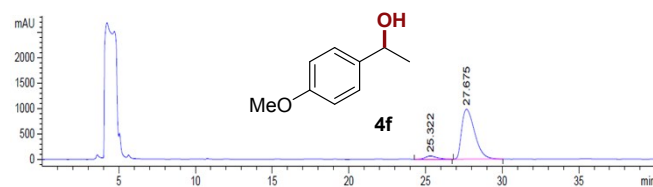


Signal 1: DAD1 B, Sig=230,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	16.171	BV	0.4061	1.51945e4	568.00879	49.9848
2	17.661	VB	0.4602	1.52037e4	506.53748	50.0152

Totals : 3.03982e4 1074.54626

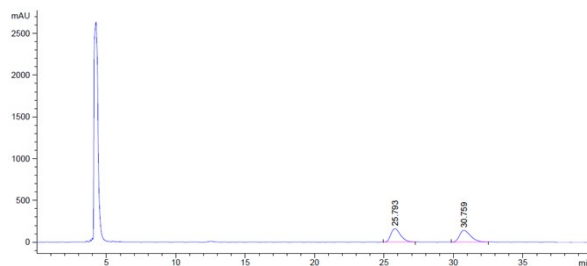
4f



Signal 2: DAD1 C, Sig=210,8 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	25.322	VB	0.7574	3083.91235	61.67646	5.0870
2	27.675	BB	0.9444	5.75396e4	980.84735	94.9130

Totals : 6.06235e4 1042.52382

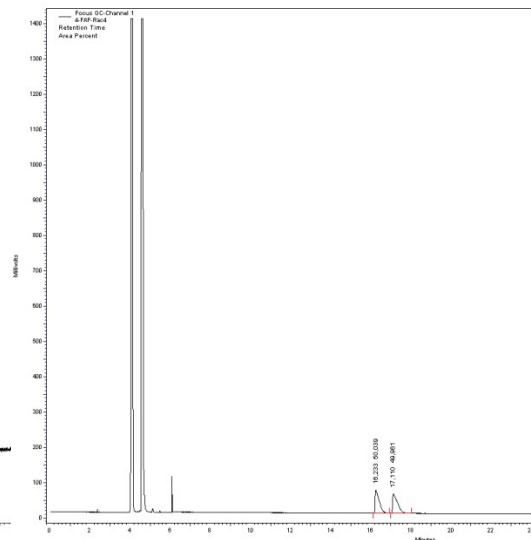
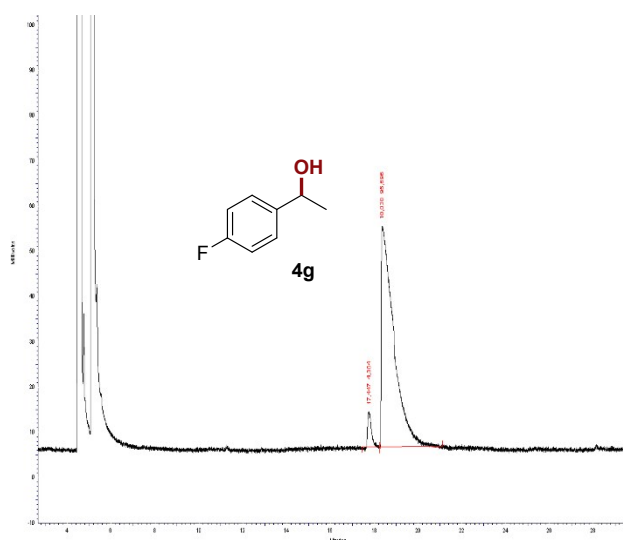


Signal 1: DAD1 C, Sig=210,8 Ref=360,100

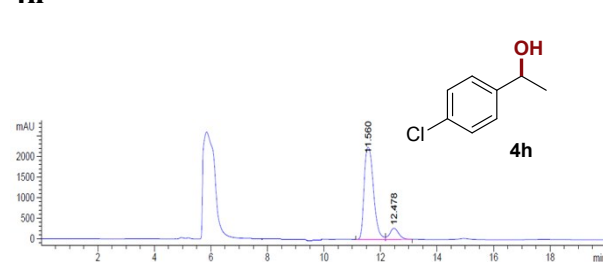
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	25.793	BB	0.7417	7765.30859	161.27690	49.8027
2	30.759	BB	0.8659	7826.82031	142.26196	50.1973

Totals : 1.55921e4 303.53886

4g



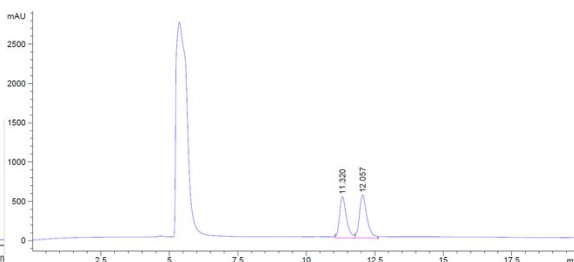
4h



Signal 3: DAD1 C, Sig=210,8 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.342	MM	0.2816	2.19660e4	1300.26270	96.9659
2	12.072	MM	0.2745	687.32568	41.72610	3.0341

Totals : 2.26533e4 1341.98880

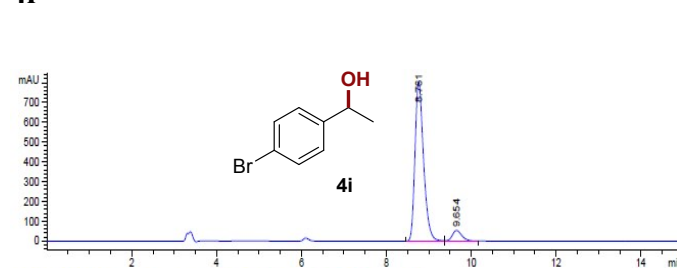


Signal 3: DAD1 C, Sig=210,8 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.320	MF	0.2907	9168.56348	525.70673	46.8626
2	12.057	FM	0.3172	1.03962e4	546.27844	53.1374

Totals : 1.95648e4 1071.98517

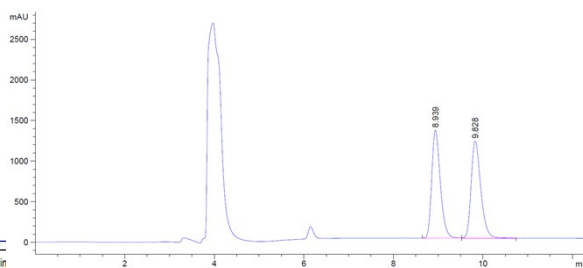
4i



Signal 1: DAD1 C, Sig=210,8 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	8.761	BV	0.2154	9576.69043	683.63574	93.0471
2	9.654	VB	0.2391	715.61804	45.60443	6.9529

Totals : 1.02923e4 729.24017

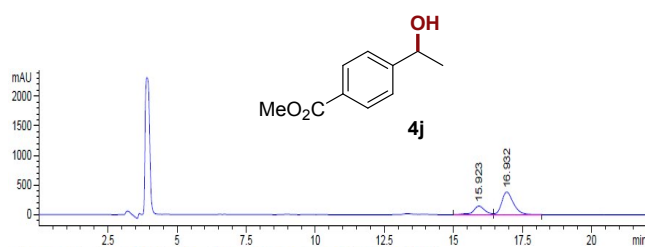


Signal 1: DAD1 C, Sig=210,8 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	8.939	BV	0.2032	1.77246e4	1332.23499	49.8299
2	9.828	VB	0.2279	1.78456e4	1197.48718	50.1701

Totals : 3.55702e4 2529.72217

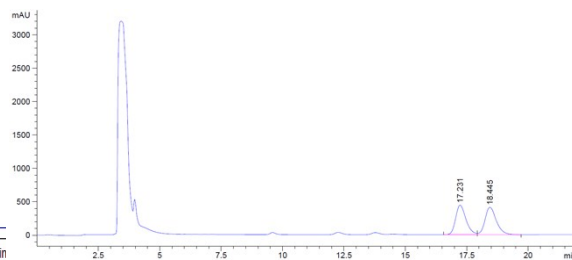
4j



Signal 3: DAD1 D, Sig=215,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	15.923	MM	0.4323	2840.11377	109.48695	24.5643
2	16.932	MM	0.4675	8721.85059	310.91879	75.4357

Totals : 1.15620e4 420.40575

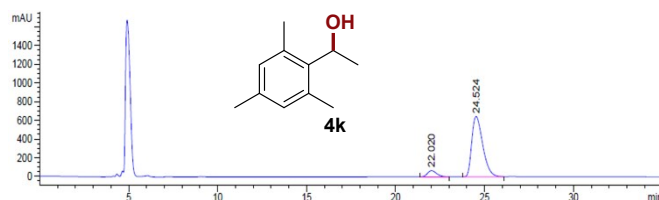


Signal 3: DAD1 D, Sig=215,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	17.231	BV	0.4472	1.29377e4	442.44095	49.6298
2	18.445	VB	0.4872	1.31307e4	410.49481	50.3702

Totals : 2.60684e4 852.93576

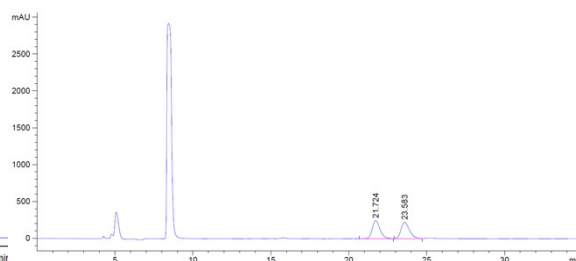
4k



Signal 1: DAD1 C, Sig=210,8 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	22.020	BB	0.5500	2358.32593	66.13913	7.8811
2	24.524	BB	0.6703	2.75654e4	645.44666	92.1189

Totals : 2.99237e4 711.58578

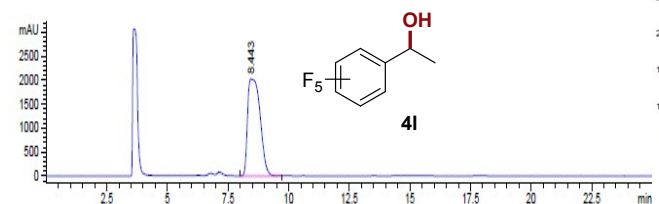


Signal 1: DAD1 C, Sig=210,8 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	21.724	BB	0.5333	8574.23926	246.83929	50.4017
2	23.583	BB	0.5754	8437.55762	224.03615	49.5983

Totals : 1.70118e4 470.87544

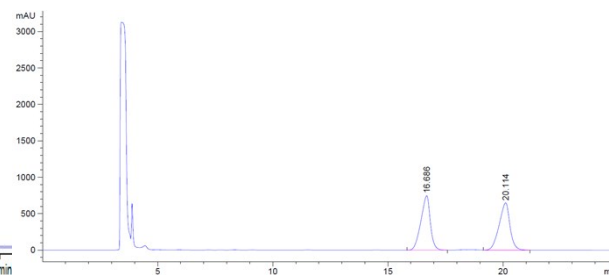
4l



Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	8.443	BB	0.4769	7.27329e4	2025.50549	100.0000

Totals : 7.27329e4 2025.50549

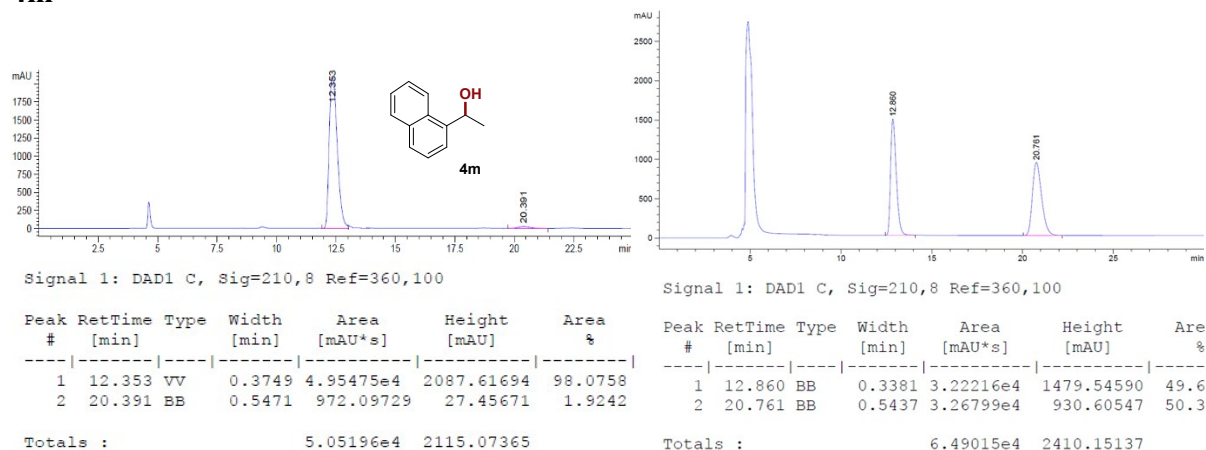


Signal 1: DAD1 A, Sig=254,4 Ref=360,100

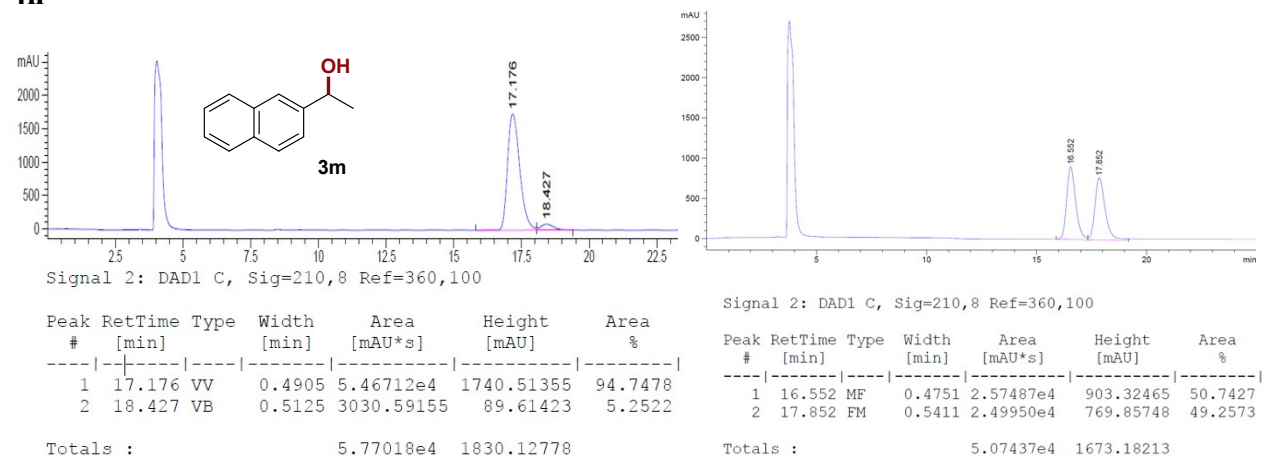
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	16.686	BB	0.4208	2.11732e4	746.75647	50.0781
2	20.114	VB	0.4893	2.11071e4	649.14795	49.9219

Totals : 4.22803e4 1395.90442

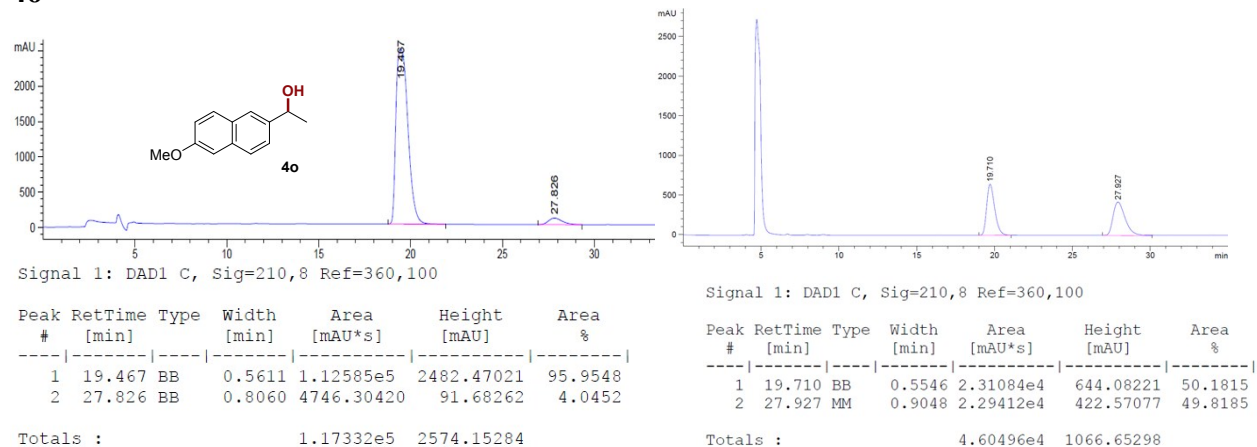
4m



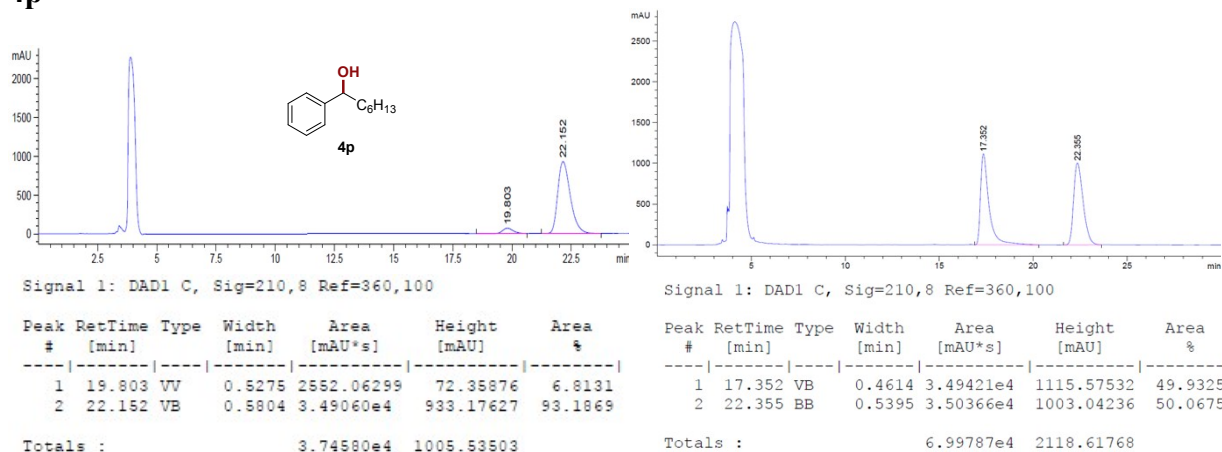
4n



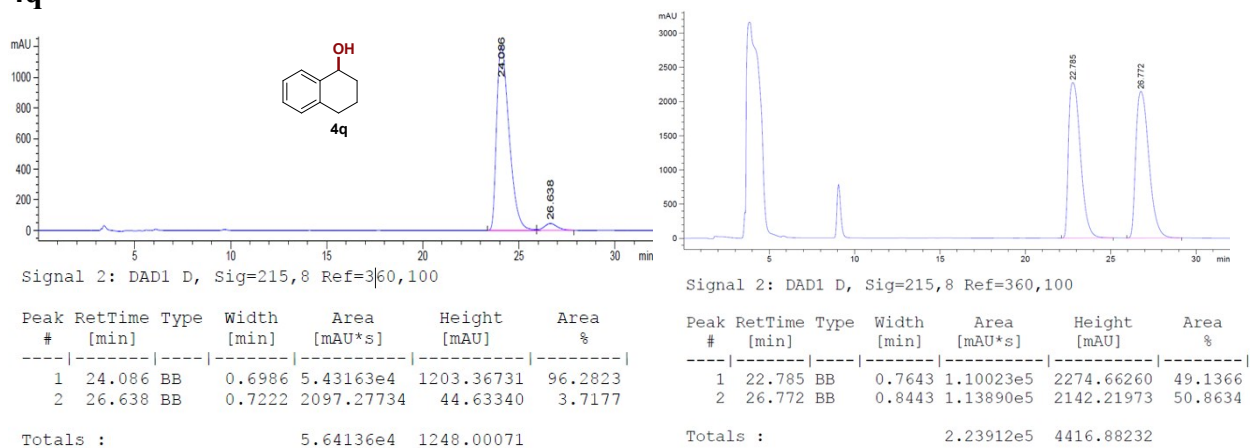
4o



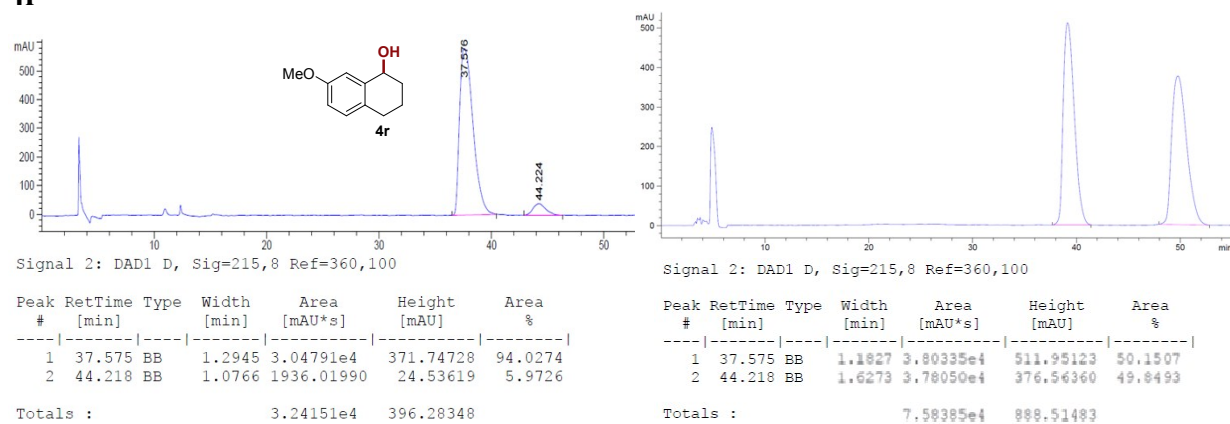
4p



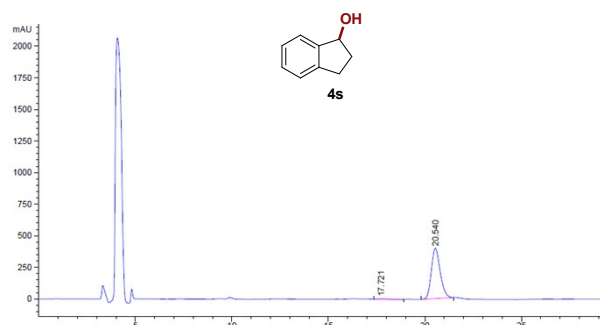
4q



4r

Data agree with reported racemate.^{13a}

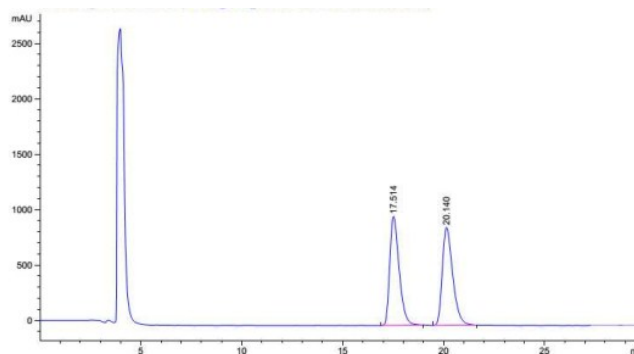
4s



Signal 1: DAD1 C, Sig=210,8 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	17.721	VB	0.4191	156.59662	5.20371	1.2124
2	20.540	BB	0.4924	1.27602e4	399.72852	98.7876

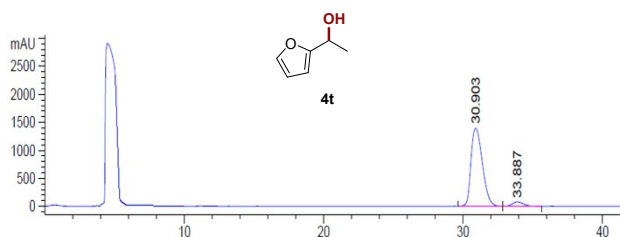
Totals : 1.29168e4 404.93223



Signal 1: DAD1 C, Sig=210,8 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	17.514	BB	0.4903	3.15029e4	981.91467	49.9212
2	20.140	BB	0.5502	3.16023e4	881.56042	50.0788

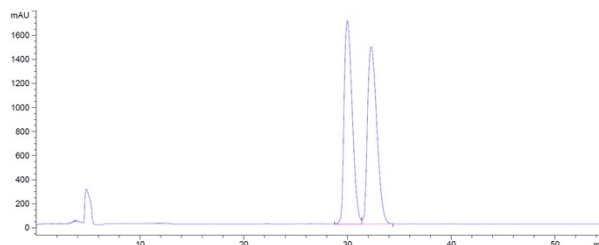
4t



Signal 1: DAD1 B, Sig=215,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	30.903	BB	0.9097	7.79231e4	1369.66309	94.6282
2	33.885	BB	0.8887	4423.52637	77.41204	5.3718

Totals : 8.23466e4 1447.07513

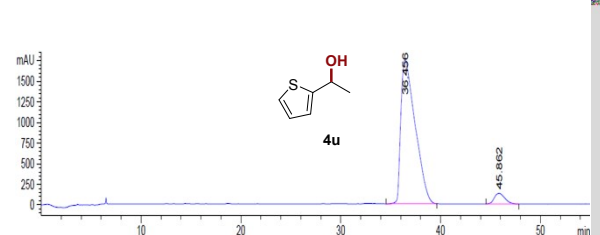


Signal 1: DAD1 B, Sig=215,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	29.977	BV	0.8781	9.20907e4	1689.02271	49.8811
2	32.274	VB	1.0035	9.25296e4	1471.79517	50.1189

Totals : 1.84620e5 3160.81787

4u



Signal 1: DAD1 B, Sig=215,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	36.456	VB	1.4150	1.76906e5	1768.56421	94.8289
2	45.864	BB	1.1162	9646.81250	132.05803	5.1711

Totals : 1.86553e5 1900.62224

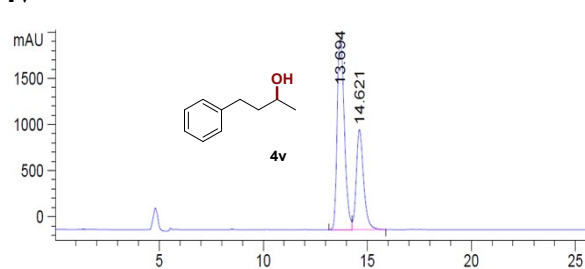


Signal 1: DAD1 B, Sig=215,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	39.151	BB	1.1827	3.80335e4	511.95123	50.1507
2	49.775	BB	1.6273	3.78050e4	376.56360	49.8493

Totals : 7.58385e4 888.51483

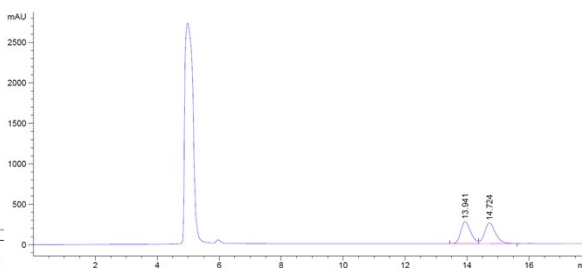
4v



Signal 3: DAD1 D, Sig=215,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	13.694	BV	0.3813	5.01619e4	2065.83276	65.3926
2	14.621	VB	0.3710	2.65469e4	1086.53491	34.6074

Totals : 7.67088e4 3152.36768

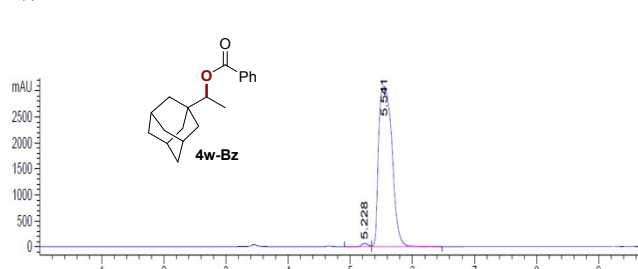


Signal 3: DAD1 D, Sig=215,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	13.941	BV	0.3382	6033.42822	272.59311	49.2202
2	14.724	VB	0.3663	6224.60791	257.23038	50.7798

Totals : 1.22580e4 529.82349

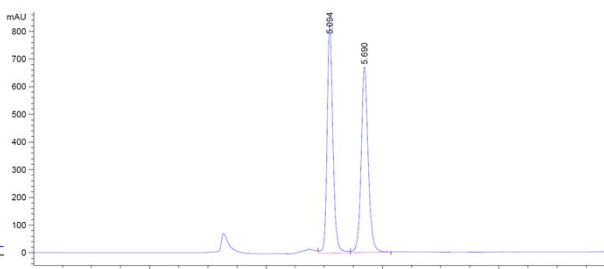
4w



Signal 1: DAD1 B, Sig=230,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	5.382	VB	0.1296	2562.64771	306.47476	97.5699
2	5.929	BB	0.1381	63.82467	7.02082	2.4301

Totals : 2626.47237 313.49558

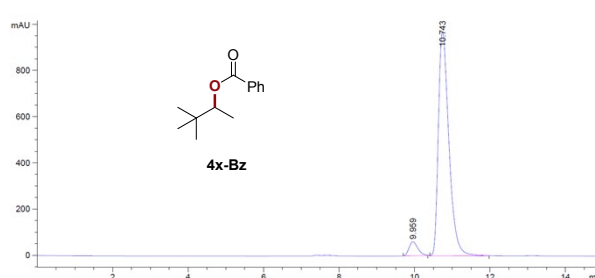


Signal 1: DAD1 B, Sig=230,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	5.094	VV	0.1018	5676.69824	829.47650	50.3031
2	5.690	VB	0.1256	5608.28369	670.55634	49.6969

Totals : 1.12850e4 1500.03284

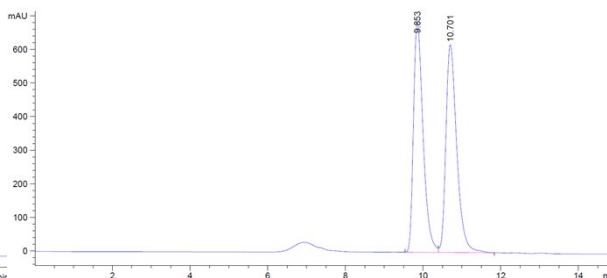
4x



Signal 1: DAD1 B, Sig=230,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	9.959	MM	0.2712	980.92780	60.29387	4.9686
2	10.743	VB	0.2956	1.87614e4	970.42322	95.0314

Totals : 1.97424e4 1030.71708



Signal 1: DAD1 B, Sig=230,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	9.853	BV	0.2648	1.17669e4	683.75092	49.4732
2	10.701	VB	0.2966	1.20175e4	618.83972	50.5268

Totals : 2.37844e4 1302.59064

6. Crystal Structure Data

Table S2. Details of the crystal structure determinations of **1b** and **2b**.

	1b	2b
formula	C ₂₂ H ₃₄ ClCrN ₃ O ₂	C ₂₆ H ₄₅ CrN ₃ O ₂ Si
crystal system	orthorhombic	orthorhombic
space group	<i>P</i> 2 ₁ 2 ₁ 2 ₁	<i>P</i> 2 ₁ 2 ₁ 2 ₁
<i>a</i> /Å	9.01882(13)	11.7996(7)
<i>b</i> /Å	13.64803(16)	14.2516(10)
<i>c</i> /Å	19.2371(2)	17.9837(12)
<i>V</i> /Å ³	2367.88(5)	3024.2(3)
<i>Z</i>	4	4
<i>M_r</i>	459.97	511.74
<i>F</i> ₀₀₀	976	1104
δ_c /Mg·m ⁻³	1.290	1.124
<i>m</i> /mm ⁻¹	0.618	0.442
max., min. transmission factors	1.116, 0.911	1.000, 0.411
X-radiation, λ /Å	Mo-K α , 0.71073	
data collect. temperatur. /K	120(1)	220(1)
θ range /°	2.1 to 32.4	2.2 to 25.2 °
index ranges <i>h,k,l</i>	-13 ... 13, -20 ... 20, -28 ... 28	-14 ... 14, -17 ... 17, -21 ... 21
reflections measured	193770	38247
unique [<i>R</i> _{int}]	8348 [0.0691]	5422 [0.1452]
observed [<i>I</i> ≥ 2σ(<i>I</i>)]	7583	3677
data / restraints / parameters	8348 / 0 / 270	5422 / 0 / 309
GooF on <i>F</i> ²	1.041	1.056
R indices [<i>F</i> > 4σ(<i>F</i>)] <i>R</i> (<i>F</i>), <i>wR</i> (<i>F</i> ²)	0.0378, 0.0912	0.0645, 0.1553
R indices (all data) <i>R</i> (<i>F</i>), <i>wR</i> (<i>F</i> ²)	0.0449, 0.0946	0.1000, 0.1794
absolute structure parameter	-0.019(5)	0.02(3)
largest residual peaks /e·Å ⁻³	0.502, -0.232	0.577, -0.452
CCDC deposition number	1850233	1850234

7. Literature

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