Electronic Supporting Information

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General remarks

All reagents were obtained from commercial sources and used as received. Pluronic F-127 (Mw = 12,600, EO106PO70EO106) was supplied by Sigma-Aldrich Co. LLC. Technical grade petroleum ether (40-60°C bp.) and ethyl acetate were used for chromatography column.

The images of surface morphologies and the energy-dispersive spectrometry (EDS) spectra were acquired by scanning electron microscope (SEM) with FEI Quanta 400 FEG. The structure and crystallinity of the as-synthesized catalysts was tested by powder X-ray diffraction (XRD) analysis on a D8 Advance diffractometer (Bruker, Germany) . X-ray photoelectron spectra (XPS) was analyzed on the Thermo ESCALAB 250XI instrument.

^{1}H NMR spectra were recorded in $CDCl_3$ at ambient temperature on Bruker AVANCE I 300 or 400 spectrometers at 300.1 or 400.1 MHz, using the solvent as internal standard (7.26 ppm). ^{13}C NMR spectra were obtained at 75 MHz and referenced to the internal solvent signals (central peak is 77.2 ppm). Chemical shift (δ) and coupling constants (*J*) are given in ppm and in Hz, respectively. The peak patterns are indicated as follows: s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet, and br. for broad.

GC analyses were performed with GC-14C (Shimadzu) equipped with a 30-m capillary column (Supelco, SPB-5, fused silica capillary column, 30 M*0.25 mm*0.25 mm film thickness), was used with N_2/air as vector gas. GCMS were measured by GCMS-7890A-5975C (Agilent) with GC-7890A equipped with a 30-m capillary column (HP-5ms, fused silica capillary column, 30 M*0.25 mm*0.25 mm film thickness), was used with helium as vector gas.

The following GC conditions were used: initial temperature 80 °C, for 2 minutes, then rate 20 °C/min. until 260 °C and 260°C for 20 minutes.

Preparation of the Ru-OMC catalyst

During the fabrication process of catalysts, 9.45 g Pluronic F127 was firstly dissolved in ethanol solution under stirred conditions to form a transparent solution A. Then, 16.5 g resorcinol was completely dissolved in water and obtained solution was slowly added in the solution A and continuously stirred to obtain solution B. Subsequently, 13.5 g formaldehyde (37 wt%) was added to solution B slowly. After stirring for 30 min, 2.3 ml HCl was added slowly as a catalyst to solution B, and the mixtures was stirred continuously for 6 h. The obtained precursor solution presents two separated phases, the upper transparent phase was discarded, while the lower yellow phase was considered as polymer-rich resorcinol formaldehyde resin precursor. The $RuCl_3$ was then added to this precursor with 5 wt% Ru contents. Finally, the precursors with different Ru contents were dried in an oven at 90 ℃ for 12 h. The resulted polymers were then carbonized at 1000 ℃ for 2 h with a heating rate 2 ℃/min under a N_2 atmosphere with the flow rate of 80 mL/min. Finally, the Ru-OMC catalysts were obtained after cooling down the materials to room temperature. The obtained catalysts were denoted as Ru-OMC.

Characterization of Ru-OMC catalysts

The images of surface morphologies and the energy-dispersive spectrometry (EDS) spectra were acquired by scanning electron microscope (SEM) with FEI Quanta 400 FEG. The structure and crystallinity of the as-synthesized catalysts was tested by powder X-ray diffraction (XRD) analysis on a D8 Advance diffractometer (Bruker, Germany) in the 2θ range of 10º~80º. Small-angle XRD was also performed at 2θ from 0.5º to 5º. X-ray photoelectron spectra (XPS) was analyzed on the Thermo ESCALAB 250XI instrument using Al Kα (*hv* =1486.6 eV) from an X-ray source operating at 15 kV and 10 mA. The C(1s) line at 284.6 eV was used as the binding energy reference.

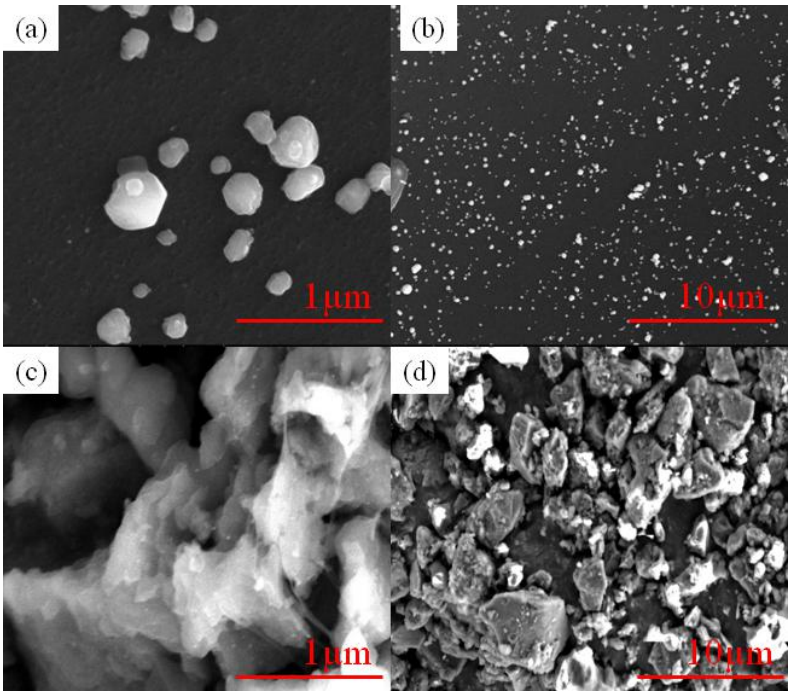


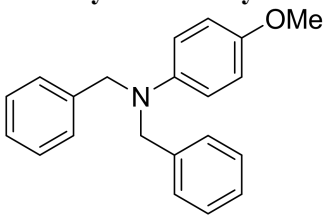
Fig. S1. The SEM micrographs of (a), (b) as-synthesized Ru-OMC catalyst and (c) , (d) reused for 14 times.

General procedure for Ru-OMC catalyzed selective reductive aminations of aldehydes with imines

Ru-OMC (5 wt%, 1 mol%, 20 mg), imine (0.5 mmol), aldehyde (0.75 mmol), $PhSiH_3$ (0.75 mmol, 90 μL) and MeOH (2 mL) were introduced in a tube under air, equipped with magnetic stirring bar and was stirred at 40 ºC. After 16 h, the conversion of the reaction was analyzed by gas chromatography. The solvent was then evaporated under vacuum and the desired product was purified by using a silica gel chromatography column and a mixture of petrol ether/ethyl acetate as eluent.

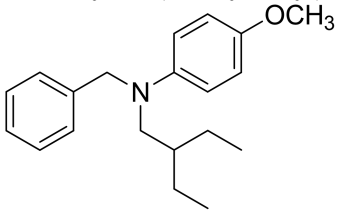
Characterization data of substrates

***N*-benzyl-4-methoxy-*N*-benzylaniline[1] (3a)**



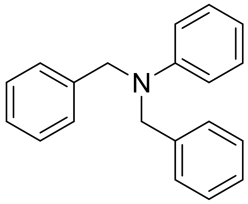
Light green solid, yield = 82%, 124 mg, ^{1}H NMR (300 MHz, $CDCl_3$): δ = 7.41-7.28 (m, 10H), 6.85-6.76 (m, 4H), 4.64 (s, 4H), 3.79 (s, 3H). $^{13}C\{^1H\}$ NMR (75 MHz, $CDCl_3$): δ = 151.8, 143.9, 139.1, 128.7, 127.1, 127.0, 114.8, 114.6, 55.8, 55.3. MS (EI): m/z: 303 (70, M^+), 212 (70), 91 (100), 65 (20).

***N*-benzyl-*N*-(2-ethylbutyl)-4-methoxyaniline (3b)**



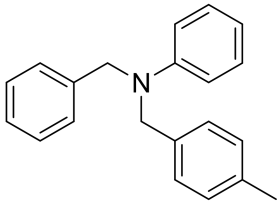
Colorless solid, yield = 85%, 126 mg, ^{1}H NMR (300 MHz, $CDCl_3$): δ = 7.35-7.22 (m, 5H), 6.81 (d, 2H, *J* = 9.3 Hz), 6.70 (d, 2H, *J* = 9.0 Hz), 4.53 (s, 2H), 3.77 (s, 3H), 3.24 (d, 2H, *J* = 7.2 Hz), 1.75-1.73 (m, 1H), 1.49-1.30 (m, 4H), 0.91 (t, 6H, *J* = 7.5 Hz). $^{13}C\{^1H\}$ NMR (75 MHz, $CDCl_3$): δ = 151.4, 143.9, 139.5, 128.6, 127.1, 126.7, 114.8, 114.7, 56.6, 56.3, 55.9, 39.4, 23.7, 10.9.

***N,N*-dibenzylaniline[2] (3c)**



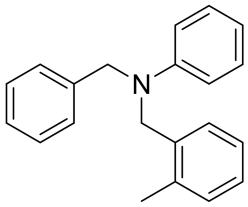
Colorless solid, yield = 86%, 117 mg, ^{1}H NMR (300 MHz, $CDCl_3$): δ = 7.42-7.21 (m, 12H), 6.82-6.75 (m, 3H), 4.72 (s, 4H). $^{13}C\{^1H\}$ NMR (75 MHz, $CDCl_3$): δ = 149.3, 138.7, 129.4, 128.8, 127.0, 126.8, 116.9, 112.6, 54.3. MS (EI): m/z: 273 (30, M^+), 196 (25), 182 (25), 91 (100), 65 (10).

***N*-benzyl-*N*-(4-methylbenzyl)aniline[1] (3d)**



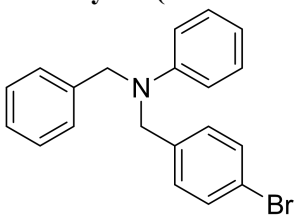
Orange solid, yield = 58%, 83 mg, ^{1}H NMR (300 MHz, $CDCl_3$): δ = 7.39-7.19 (m, 11H), 6.81-6.75(m, 3H), 4.69 (s, 4H), 2.412 (s, 3H). $^{13}C\{^1H\}$ NMR (75 MHz, $CDCl_3$): δ = 149.3, 138.7, 136.5, 135.5, 129.4, 129.3, 128.7, 126.9, 126.7, 124.5, 116.8, 112.6, 54.2, 54.0, 21.1. MS (EI): m/z: 287 (40, M^+), 196 (10), 105 (100), 91 (55), 77 (35).

***N*-benzyl-*N*-(2-methylbenzyl)aniline[1] (3e)**



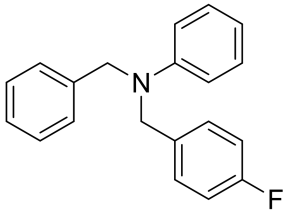
White solid, yield = 45%, 65 mg, ^{1}H NMR (300 MHz, $CDCl_3$): δ = 7.37-7.22 (m, 11H), 6.74-6.72(m, 3H), 4.72 (s, 2H), 4.62 (s, 2H), 2.32 (s, 3H). $^{13}C\{^{1}H\}$ NMR (75 MHz, $CDCl_3$): δ = 149.2, 138.7, 135.7, 130.5, 129.4, 128.8, 127.0, 126.9, 126.8, 126.3, 126.1, 116.8, 116.6, 112.4, 54.3, 52.5, 19.1. MS (EI): m/z: 287 (45, M^+), 183 (90), 105 (100), 91 (80), 77 (50).

***N*-benzyl-*N*-(4-bromobenzyl)aniline[1] (3f)**



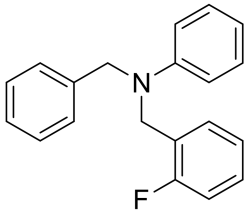
Green solid, yield = 62%, 109 mg, ^{1}H NMR (300 MHz, $CDCl_3$): δ = 7.51-7.17 (m, 11H), 6.80-6.77 (m, 3H), 4.69 (s, 2H), 4.64 (s, 2H). $^{13}C\{^{1}H\}$ NMR (75 MHz, $CDCl_3$): δ = 149.0, 138.4, 137.8, 131.9, 129.4, 128.8, 128.6, 127.2, 126.9, 120.8, 117.3, 112.8, 54.5, 53.9. MS (EI): m/z: 351 (30, M^+), 207 (10), 169 (35), 91 (100), 77 (55).

***N*-benzyl-*N*-(4-fluorobenzyl)aniline[1] (3g)**



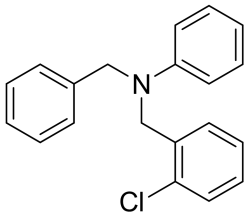
Light yellow solid, yield = 83%, 121 mg, ^{1}H NMR (300 MHz, $CDCl_3$): δ = 7.38-7.21 (m, 9H), 7.09-7.03 (m, 2H), 6.81-6.78 (m, 3H), 4.69 (s, 2H), 4.66 (s, 2H). $^{13}C\{^{1}H\}$ NMR (75 MHz, $CDCl_3$): δ = 163.7 (d, J_{CF} = 243.1Hz), 149.2, 138.6, 134.3, 129.4, 128.8, 128.5 (d, J_{CF} = 7.9Hz), 127.1, 126.9, 117.2, 115.8 (d, J_{CF} = 21.3Hz), 112.8, 54.4, 53.8. MS (EI): m/z: 291 (50, M^+), 200 (10), 109 (100), 91 (80), 77 (25).

***N*-benzyl-*N*-(2-fluorobenzyl)aniline[1] (3h)**



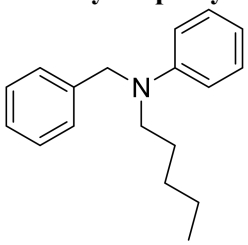
White solid, yield = 78%, 113 mg, ^{1}H NMR (300 MHz, $CDCl_3$): δ = 7.41-7.10 (m, 11H), 6.79-6.76 (m, 3H), 4.77 (s, 2H), 4.73 (s, 2H). $^{13}C\{^{1}H\}$ NMR (75 MHz, $CDCl_3$): δ = 162.59 (d, J_{CF} = 243.75Hz), 148.7, 138.4, 139.3, 128.7, 128.5, 128.4, 127.0, 126.7, 125.4 (d, J_{CF} = 13.5Hz), 124.2 (d, J_{CF} = 3.75Hz), 117.0, 115.5 (d, J_{CF} = 21.0Hz), 112.5, 54.5, 48.5. MS (EI): m/z: 291 (45, M^+), 200 (35),109 (55), 91 (100), 77 (35).

***N*-benzyl-*N*-(2-chlorobenzyl)aniline[1] (3i)**



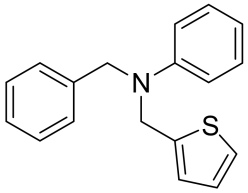
White solid, yield = 51%, 78 mg, ^{1}H NMR (300 MHz, $CDCl_3$): δ = 7.45-7.19 (m, 11H), 6.79-6.70 (m, 3H), 4.75 (s, 2H), 4.74 (s, 2H). $^{13}C\{^1H\}$ NMR (75 MHz, $CDCl_3$): δ = 148.7, 138.4, 135.5, 133.2, 129.8, 129.4, 128.9, 128.3, 127.9, 127.2, 127.1, 126.8, 117.2, 112.5, 54.7, 52.8. MS (EI): m/z: 307 (50, M^+), 180 (15), 125 (55), 91 (100), 77 (50).

N-benzyl-N-pentylaniline[3] (3j)



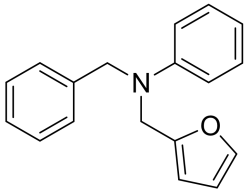
Light yellow oil, yield = 72%, 91 mg, ^{1}H NMR (300 MHz, $CDCl_3$): δ = 7.40-7.22 (m, 7H), 6.76-6.70 (m, 3H), 4.61 (s, 2H), 3.45 (t, 2H, *J* = 7.5 Hz), 1.79-1.69 (m, 2H), 1.47-1.34 (m, 4H), 0.98 (t, 3H, *J* = 6.6 Hz). $^{13}C\{^1H\}$ NMR (75 MHz, $CDCl_3$): δ = 148.7, 139.3, 129.3, 128.7, 126.9, 126.7, 116.0, 112.2, 54.6, 51.5, 29.5, 27.0, 22.8, 14.3. MS (EI): m/z: 253 (20, M^+), 196 (65), 91 (100), 77 (15).

***N*-benzyl-*N*-(thiophen-2-ylmethyl)aniline[1] (3k)**



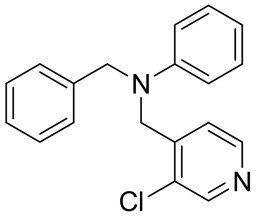
Light yellow oil, yield = 68%, 95 mg, ^{1}H NMR (300 MHz, $CDCl_3$): δ = 7.43-7.25 (m, 8H), 7.04-6.80 (m, 5H), 4.82 (s, 2H), 4.69 (s, 2H). $^{13}C\{^1H\}$ NMR (75 MHz, $CDCl_3$): δ = 148.8, 142.6, 138.6, 129.4, 128.8, 127.1, 127.0, 126.9, 125.2, 124.5, 117.6, 113.5, 54.2, 50.0. MS (EI): m/z: 279 (50, M^+), 97 (100), 77 (20), 51 (10).

***N*-benzyl-*N*-(furan-2-ylmethyl)aniline (3l)**



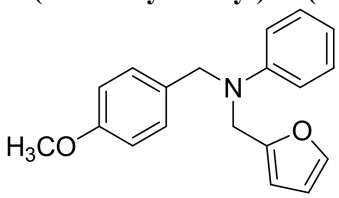
Light yellow oil, yield = 71%, 93 mg, ^{1}H NMR (300 MHz, $CDCl_3$): δ = 7.42-7.27 (m, 8H), 6.92-6.80 (m, 3H), 6.42-6.31 (m, 1H), 6.28-6.19 (m, 1H), 4.68 (s, 2H), 4.60 (s, 2H). $^{13}C\{^1H\}$ NMR (75 MHz, $CDCl_3$): δ = 152.4, 149.0, 142.1, 138.8, 129.3, 128.8, 127.1, 126.9, 117.3, 113.1, 110.4, 107.6, 54.4, 47.8. MS (EI): m/z: 263 (75, M^+), 220 (15), 91 (50), 81 (100).

N-benzyl-N-((3-chloropyridin-4-yl)methyl)aniline (3m)



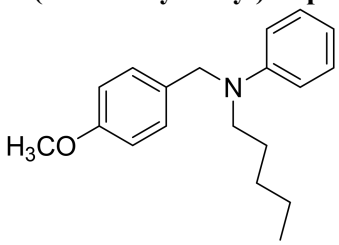
Light yellow solid, yield = 55%, 85 mg, ^{1}H NMR (300 MHz, $CDCl_3$): δ = 8.34-8.35 (m, 1H), 7.58 (d, 1H, *J* = 7.6 Hz), 7.20-7.41 (m, 8H), 6.68-6.81 (m, 3H), 4.72 (s, 2H), 4.70 (s, 2H). ^{13}C{^{1}H} NMR (75 MHz, $CDCl_3$): δ = 150.3, 148.3, 148.1, 138.0, 136.8, 132.4, 129.6, 129.0, 127.3, 126.7, 122.8, 117.6, 112.4, 54.8, 52.2. MS (EI): m/z: 308 (60, M^+), 231 (20), 126 (25), 91 (100), 65 (10).

N-(furan-2-ylmethyl)-N-(4-methoxybenzyl)aniline (3n)



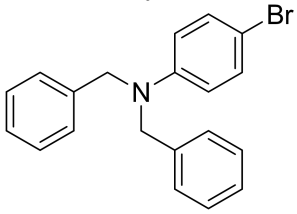
Light yellow oil, yield = 69%, 101 mg, ^{1}H NMR (300 MHz, $CDCl_3$): δ = 7.42-7.41 (m, 1H), 7.29-7.22 (m, 4H), 6.94-6.89 (m, 4H), 6.79 (t, 1H, *J* = 7.2 Hz), 6.37-6.35 (m, 1H), 6.22-6.21 (m, 1H), 4.60 (s, 2H), 4.56 (s, 2H), 3.85 (s, 3H). ^{13}C{^{1}H} NMR (75 MHz, $CDCl_3$): δ = 158.8, 152.5, 149.0, 142.0, 130.6, 129.3, 128.1, 117.3, 114.1, 113.2, 110.4, 107.6, 55.4, 53.8, 47.6. MS (EI): m/z: 293 (35, M^+), 121 (100), 81 (10).

N-(4-methoxybenzyl)-N-pentylaniline (3o)



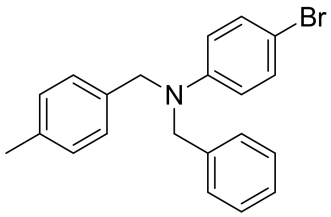
Light yellow oil, yield = 77%, 109 mg, ^{1}H NMR (300 MHz, $CDCl_3$): δ = 7.28-7.20 (m, 4H), 6.94-6.90 (m, 2H), 6.77-6.71 (m, 3H), 4.55 (s, 2H), 3.85 (s, 3H), 1.78-1.68 (m, 2H), 1.47-1.36 (m, 4H), 0.99 (t, 3H, *J* = 6.6 Hz). ^{13}C{^{1}H} NMR (75 MHz, $CDCl_3$): δ = 158.6, 148.8, 131.1, 129.3, 127.8, 116.0, 114.1, 112.3, 55.4, 54.0, 51.3, 29.5, 26.9, 22.7, 14.3. MS (EI): m/z: 283 (20, M^+), 226 (10), 121 (100), 77 (15).

***N,N*-dibenzyl-4-bromoaniline[4] (3p)**



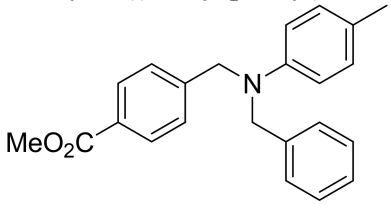
Brown solid, yield = 33%, 58 mg, ^{1}H NMR (300 MHz, $CDCl_3$): δ = 7.44-7.29 (m, 12H), 6.69 (d, 2H, *J* = 8.7 Hz), 4.71 (s, 4H). ^{13}C{^{1}H} NMR (75 MHz, $CDCl_3$): δ = 148.2, 138.1, 132.0, 128.9, 127.2, 126.7, 114.4, 108.8, 54.6. MS (EI): m/z: 351 (20, M^+), 260 (5), 91 (100), 65 (20).

***N*-benzyl-4-bromo-*N*-(4-methylbenzyl)aniline[1] (3q)**



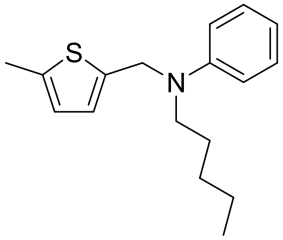
Light yellow solid, yield = 50%, 92 mg, ^{1}H NMR (300 MHz, $CDCl_3$): δ = 7.46-7.20 (m, 11H), 6.70 (d, 2H, *J* = 8.7 Hz), 4.72 (s, 2H), 4.70 (s, 2H), 2.45 (s, 3H). $^{13}C\{^{1}H\}$ NMR (75 MHz, $CDCl_3$): δ = 148.2, 138.2, 136.8, 134.9, 131.9, 129.6, 128.9, 127.2, 126.7, 114.3, 108.7, 54.5, 54.3, 21.2. MS (EI): m/z: 365 (20, M^+), 207 (5), 105 (100), 91, (45), 77 (20).

Methyl 4-((benzyl(*p*-tolyl)amino)methyl)benzoate[1] (3r)



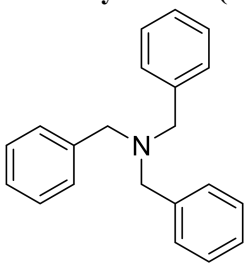
White solid, yield = 66%, 114 mg, ^{1}H NMR (300 MHz, $CDCl_3$): δ = 8.01 (d, 2H, *J* = 7.8 Hz), 7.36-7.29 (m, 7H), 7.01 (d, 2H, *J* = 8.1 Hz), 6.67 (d, 2H, *J* = 8.7 Hz), 4.67 (s, 2H), 4.65 (s, 2H), 3.93 (s, 3H), 2.26 (s, 3H). $^{13}C\{^{1}H\}$ NMR (75 MHz, $CDCl_3$): δ = 167.1, 146.9, 144.7, 138.7, 130.1, 129.9, 129.0, 128.8, 127.1, 126.9, 126.8, 126.5, 113.0, 54.9, 54.6, 52.2, 20.4. MS (EI): m/z: 345 (45, M^+), 253 (10), 195 (25), 91 (100), 65 (25).

N-((5-methylthiophen-2-yl)methyl)-N-pentylaniline (3s)



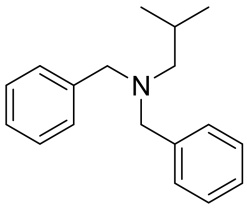
Light yellow oil, yield = 74%, 106 mg, ^{1}H NMR (300 MHz, $CDCl_3$): δ = 7.29-7.25 (m, 2H), 6.83-6.62 (m, 5H), 4.63 (s, 2H), 3.39 (t, 2H, *J* = 7.6 Hz), 2.47 (s, 3H), 1.70-1.68 (m, 2H), 1.43-1.39 (m, 4H), 0.98 (t, 3H, *J* = 7.2 Hz). $^{13}C\{^{1}H\}$ NMR (75 MHz, $CDCl_3$): δ = 148.4, 140.6, 138.7, 129.3, 124.8, 124.5, 116.6, 112.8, 51.0, 50.3, 29.5, 27.1, 22.8, 15.5, 14.3. MS (EI): m/z: 273 (25, M^+), 216 (10), 111 (100), 77 (15).

Tribenzylamine[5] (3t)



Light yellow solid, yield = 64%, 92 mg, ^{1}H NMR (300 MHz, $CDCl_3$): δ = 7.51-7.29 (m, 15H), 3.65 (s, 6H). $^{13}C\{^{1}H\}$ NMR (75 MHz, $CDCl_3$): δ = 139.8, 128.9, 128.4, 127.0, 58.1. MS (EI): m/z: 287 (30, M^+), 210 (35), 91 (100), 65 (15).

***N*,*N*-dibenzyl-2-methylpropan-1-amine[6] (3u)**



Light yellow solid, yield = 80%, 101 mg, ^{1}H NMR (300 MHz, $CDCl_3$): δ = 7.53-7.32 (m, 10H), 3.65 (s, 4H), 2.29 (d, 2H, *J* = 7.5 Hz), 2.05-1.96 (m, 1H), 1.02-0.99 (m, 6H). $^{13}C\{^{1}H\}$ NMR (75 MHz, $CDCl_3$): δ = 140.2, 129.0, 128.3, 126.9, 62.4, 59.0, 26.3, 21.0. MS (EI): m/z: 253 (5, M^+), 210 (60), 91 (100), 65 (15).

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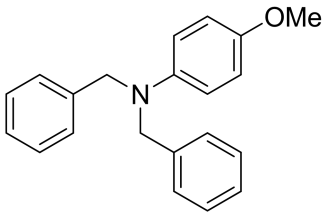
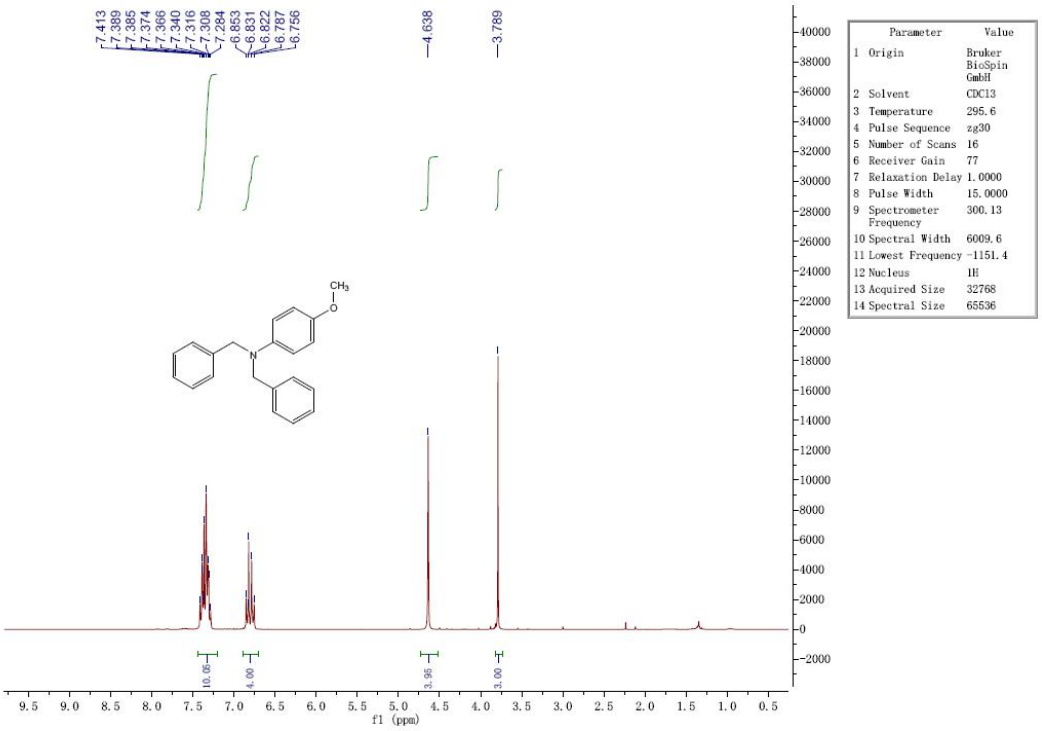
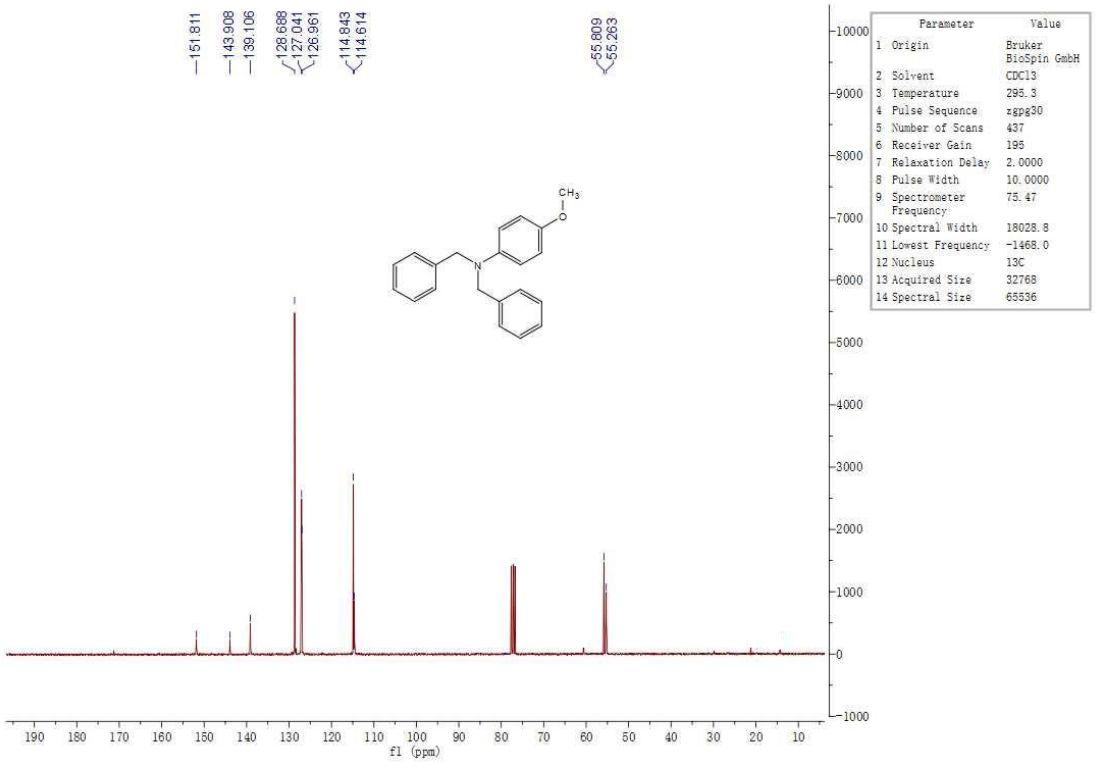
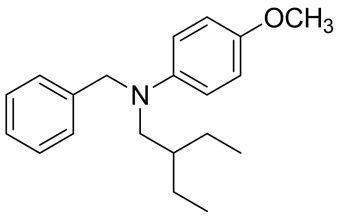
***N*-benzyl-4-methoxy-*N*-benzylaniline (3a)**

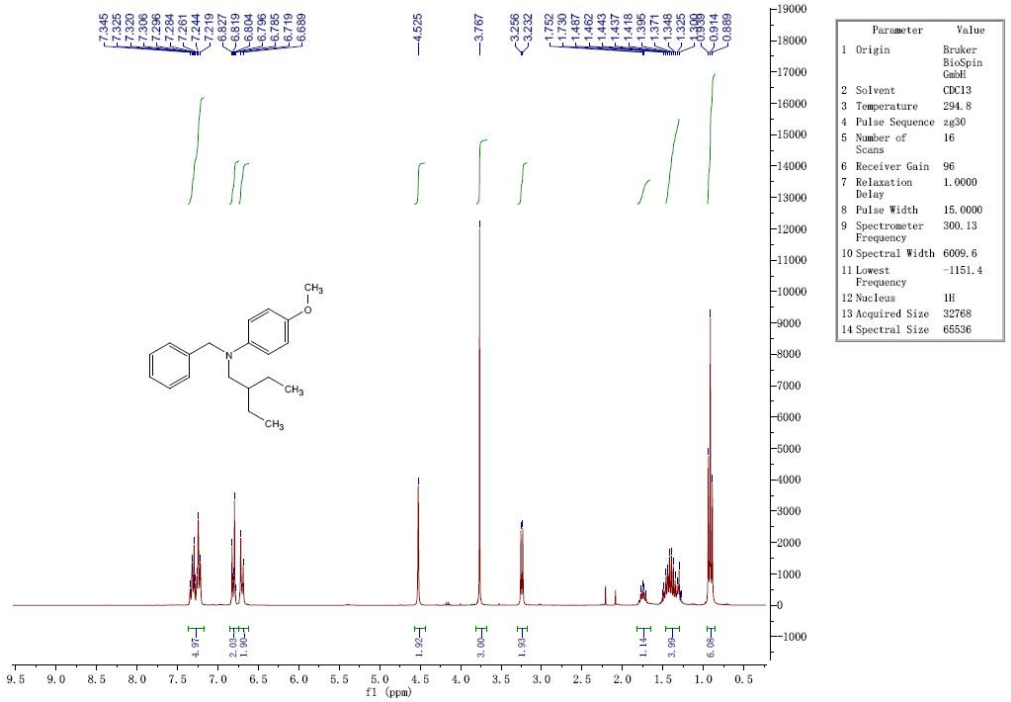






N-benzyl-*N*-(2-ethylbutyl)-4-methoxyaniline (3b)





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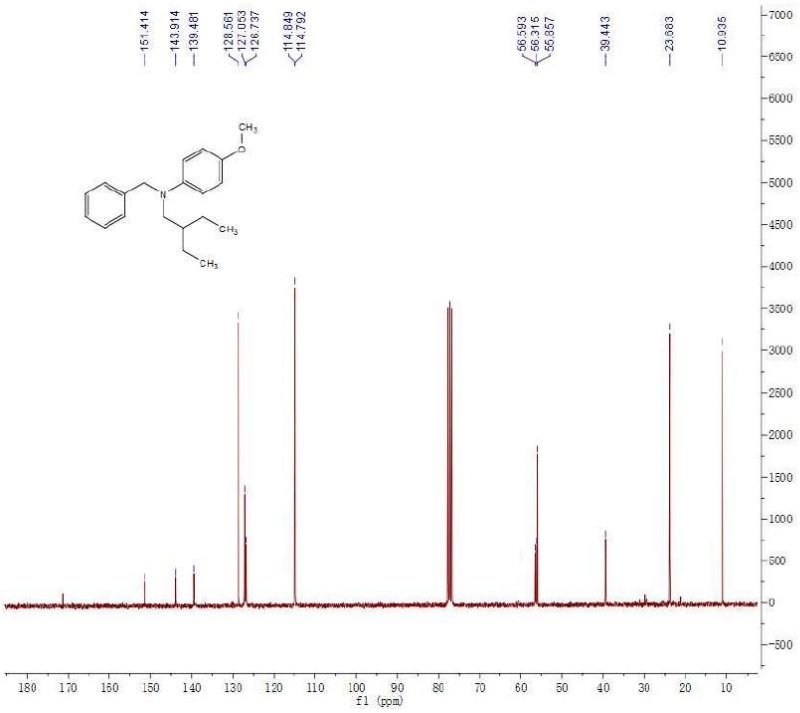
5	Number of Scans	16
6	Receiver Gain	96
7	Relaxation Delay	1.0000
8	Pulse Width	15.0000
9	Spectrometer Frequency	300.13
10	Spectral Width	6009.6
11	Lowest Frequency	-1151.4
12	Nucleus	1H
13	Acquired Size	32768
14	Spectral Size	65536



	Parameter	Value
1	Origin	Bruker BioSpin GmbH
2	Solvent	CDCl3
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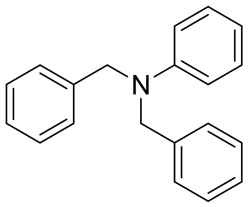
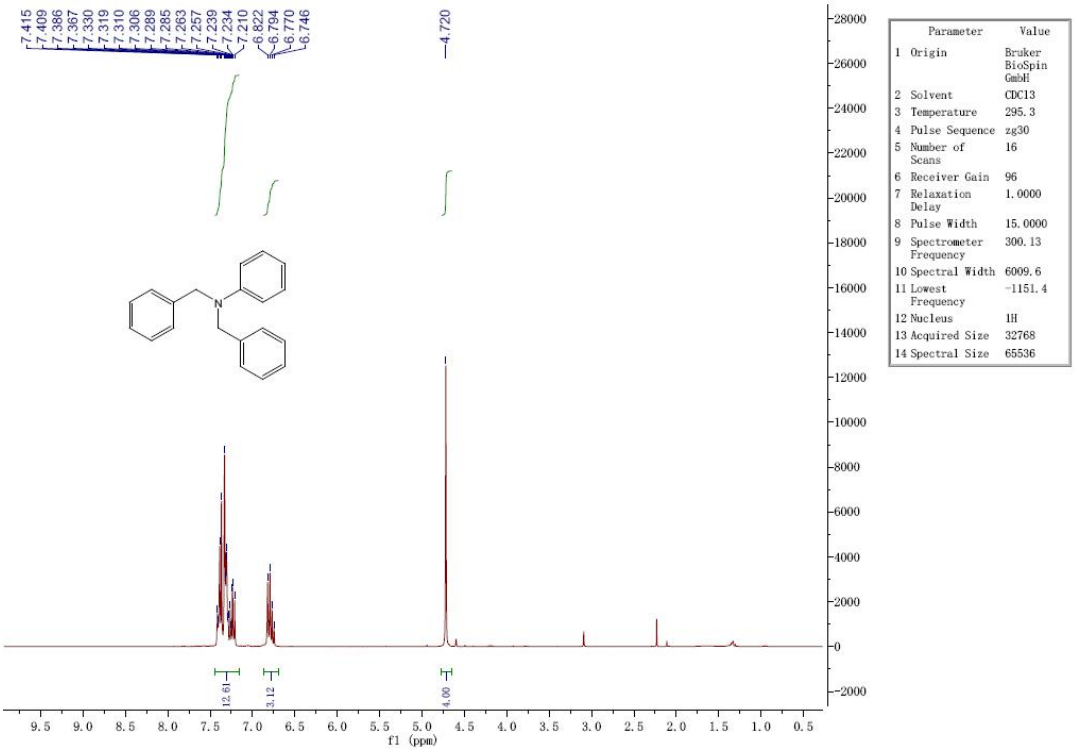
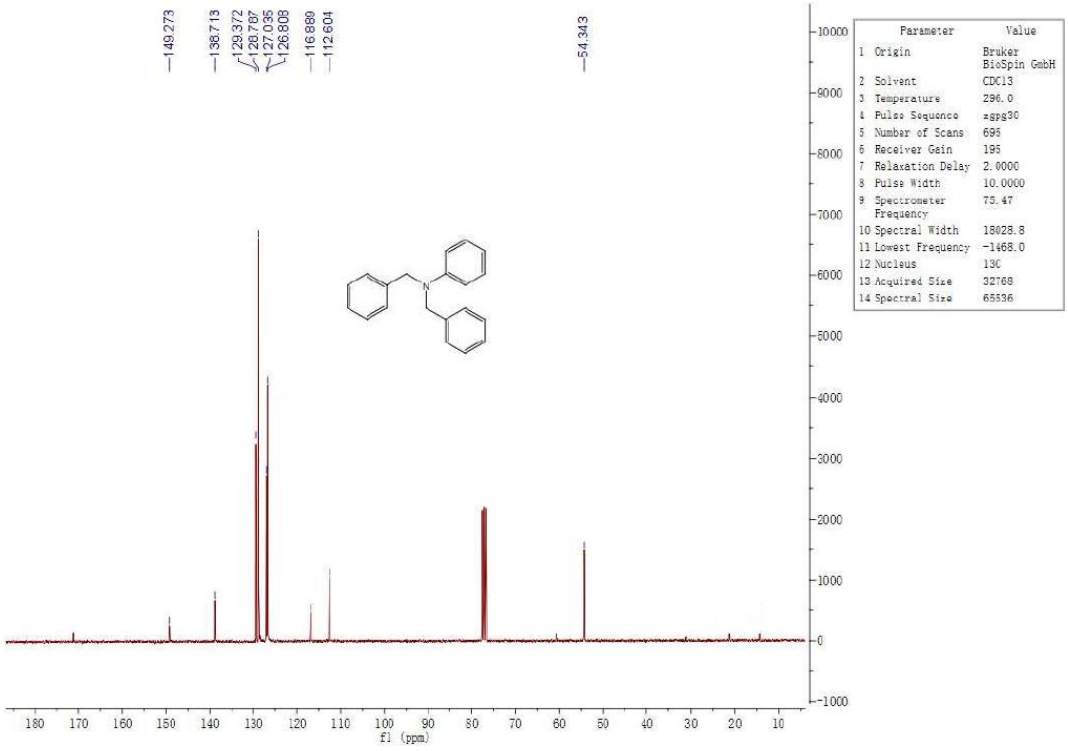
4	Pulse Sequence	zgpg30
5	Number of Scans	1024
6	Receiver Gain	195
7	Relaxation Delay	2.0000
8	Pulse Width	10.0000
9	Spectrometer Frequency	75.47
10	Spectral Width	18028.8
11	Lowest Frequency	-1468.0
12	Nucleus	13C
13	Acquired Size	32768
14	Spectral Size	65536

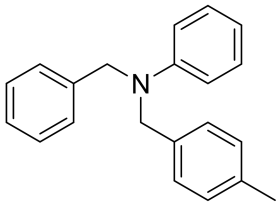
***N,N*-dibenzylaniline (3c)**

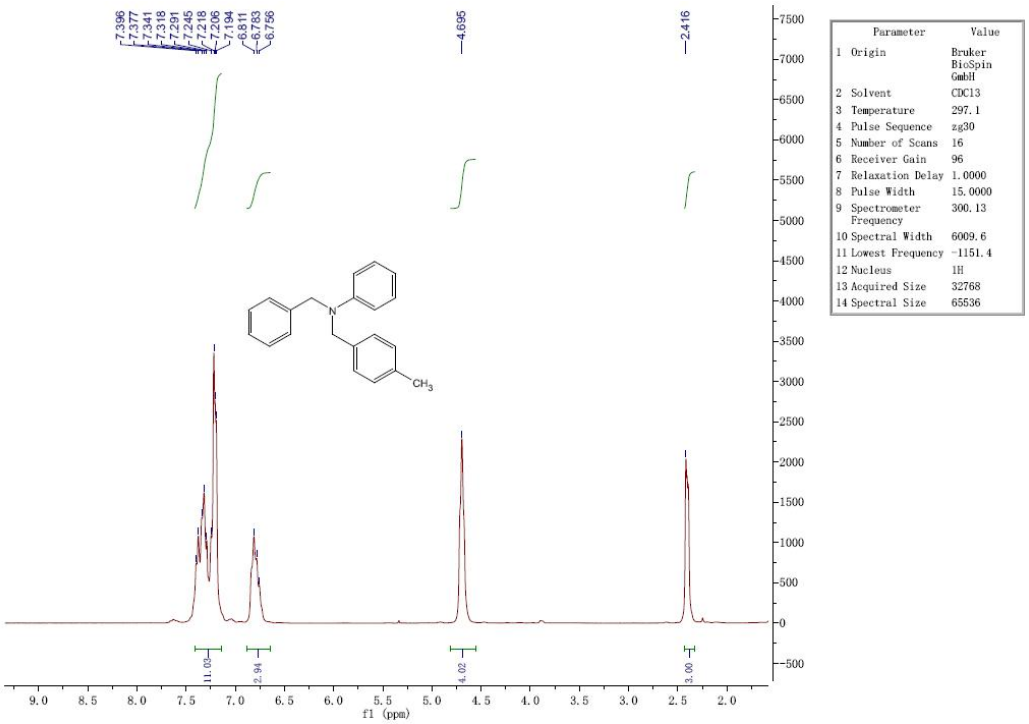






N-benzyl-*N*-(4-methylbenzyl)aniline (3d)





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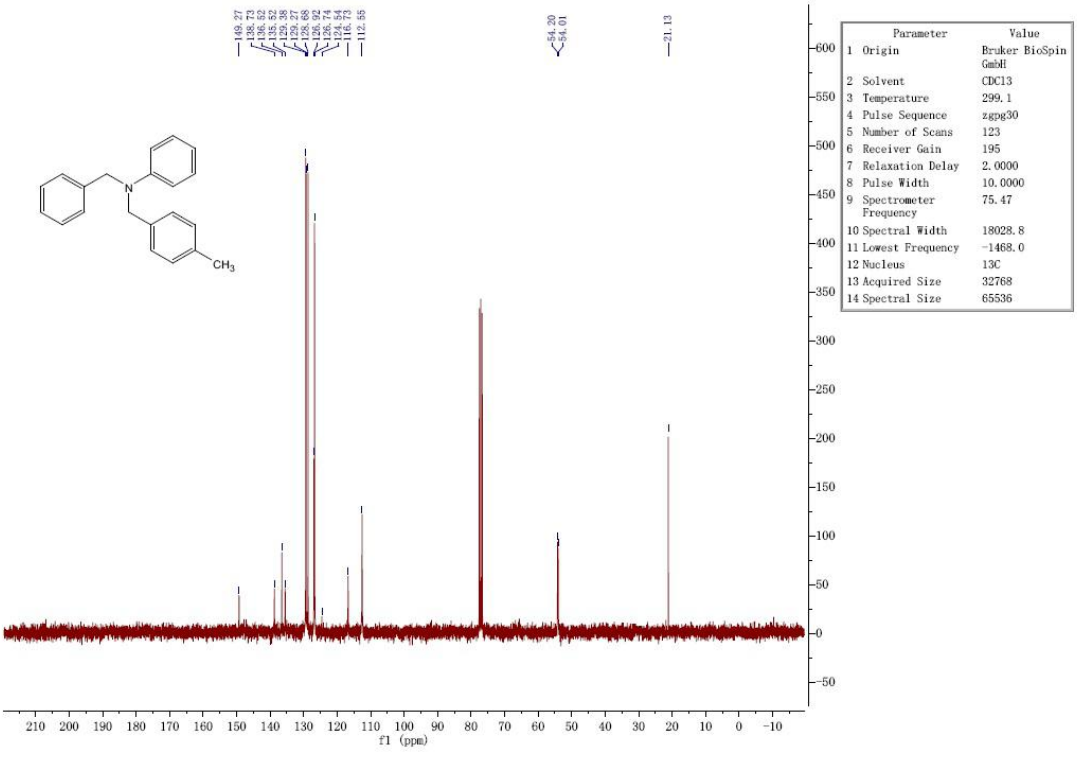
1	Origin	Bruker BioSpin GmbH
2	Solvent	CDCl3
3	Temperature	297.1
4	Pulse Sequence	zg30
5	Number of Scans	16
6	Receiver Gain	96
7	Relaxation Delay	1.0000
8	Pulse Width	15.0000
9	Spectrometer Frequency	300.13
10	Spectral Width	6009.6
11	Lowest Frequency	-1151.4
12	Nucleus	1H
13	Acquired Size	32768
14	Spectral Size	65536



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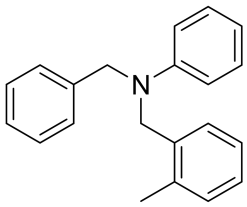
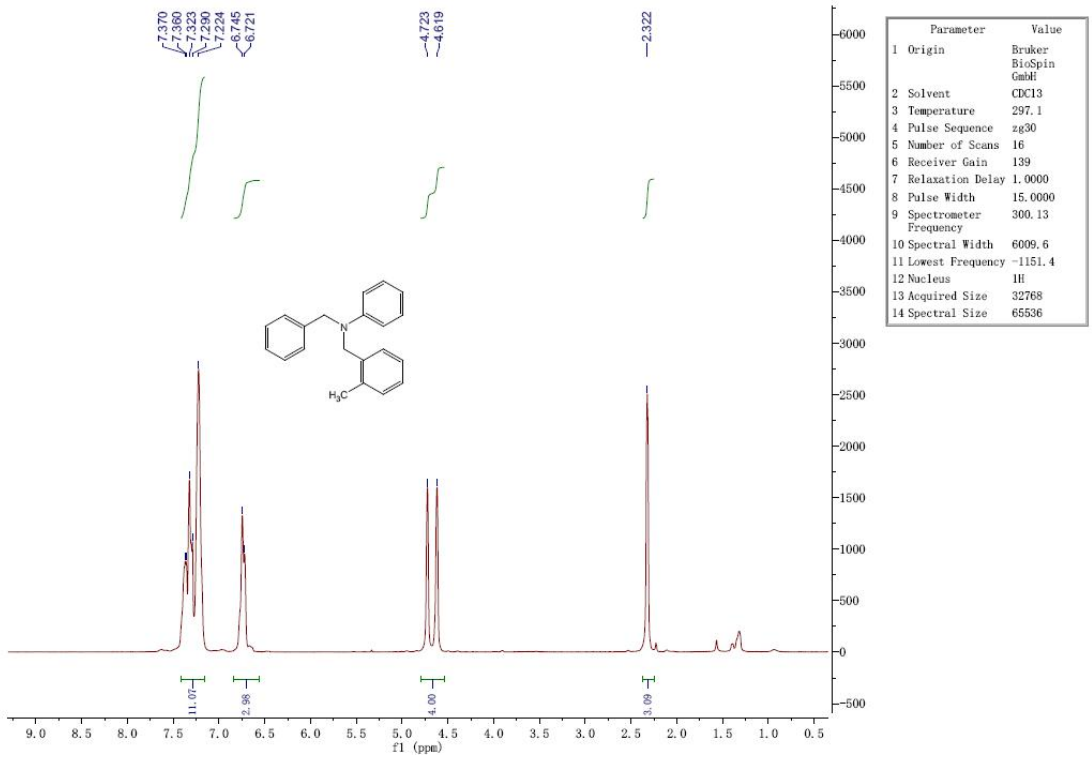
7	Relaxation Delay	2.0000
8	Pulse Width	10.0000
9	Spectrometer Frequency	75.47
10	Spectral Width	18028.8
11	Lowest Frequency	-1468.0
12	Nucleus	13C
13	Acquired Size	32768
14	Spectral Size	65536

N-benzyl-*N*-(2-methylbenzyl)aniline (3e)





	Parameter	Value
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2	Solvent	CDCl3
3	Temperature	297.1
4	Pulse Sequence	zg30
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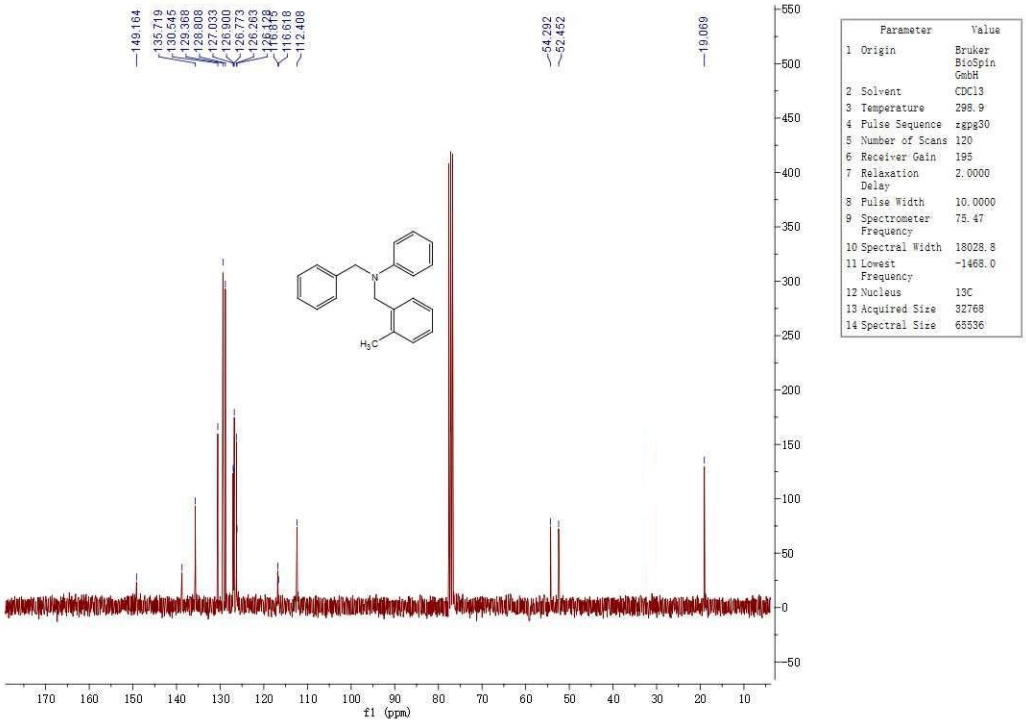
8	Pulse Width	15.0000
9	Spectrometer Frequency	300.13
10	Spectral Width	6009.6
11	Lowest Frequency	-1151.4
12	Nucleus	1H
13	Acquired Size	32768
14	Spectral Size	65536



	Parameter	Value
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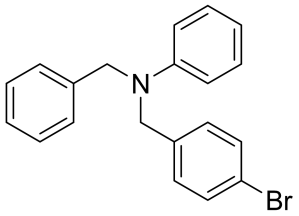
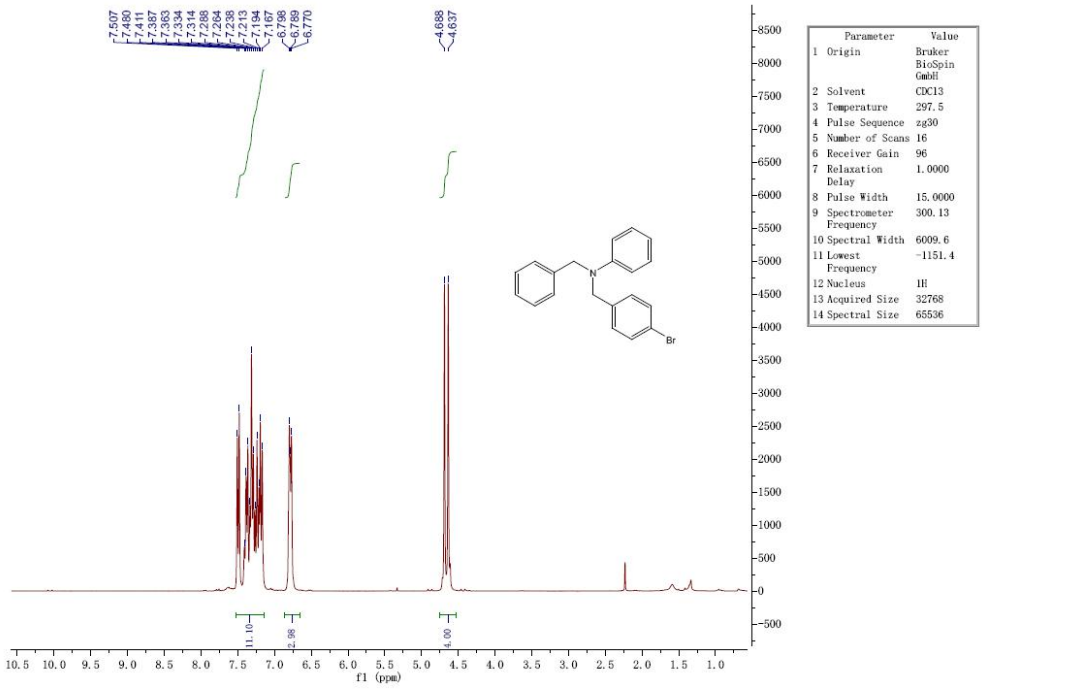
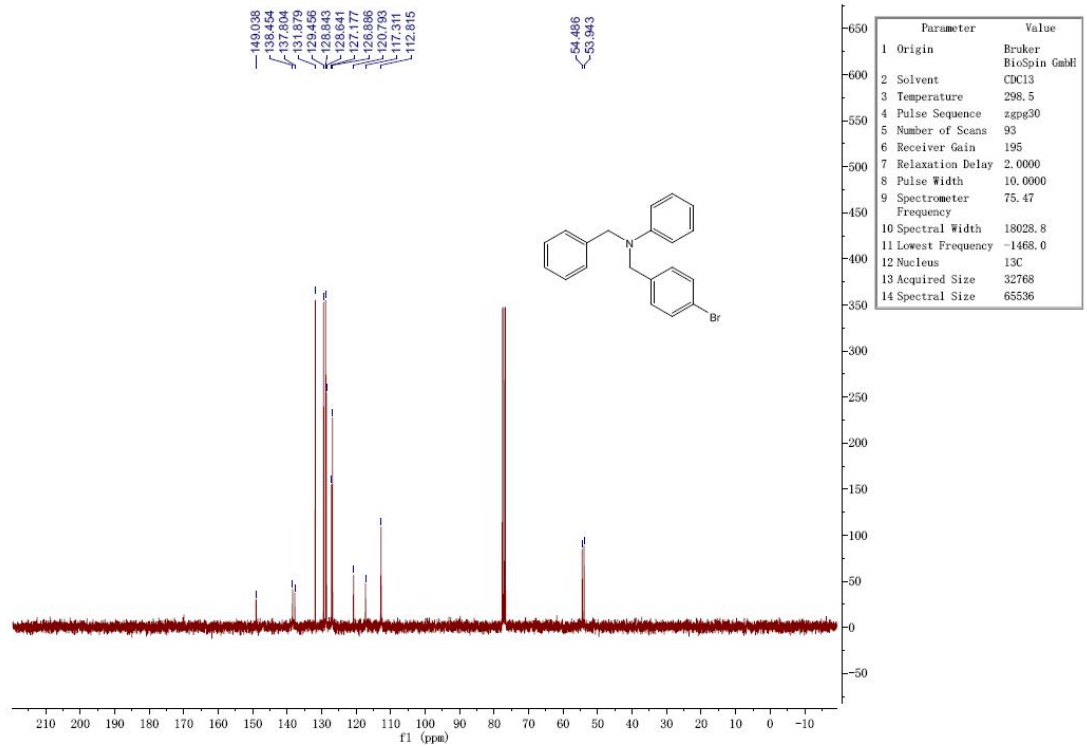
8	Pulse Width	10.0000
9	Spectrometer Frequency	75.47
10	Spectral Width	18028.8
11	Lowest Frequency	-1468.0
12	Nucleus	13C
13	Acquired Size	32768
14	Spectral Size	65536

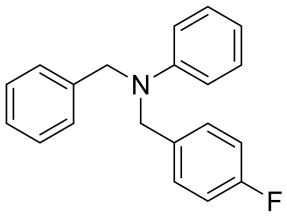
N-benzyl-*N*-(4-bromobenzyl)aniline (3f)

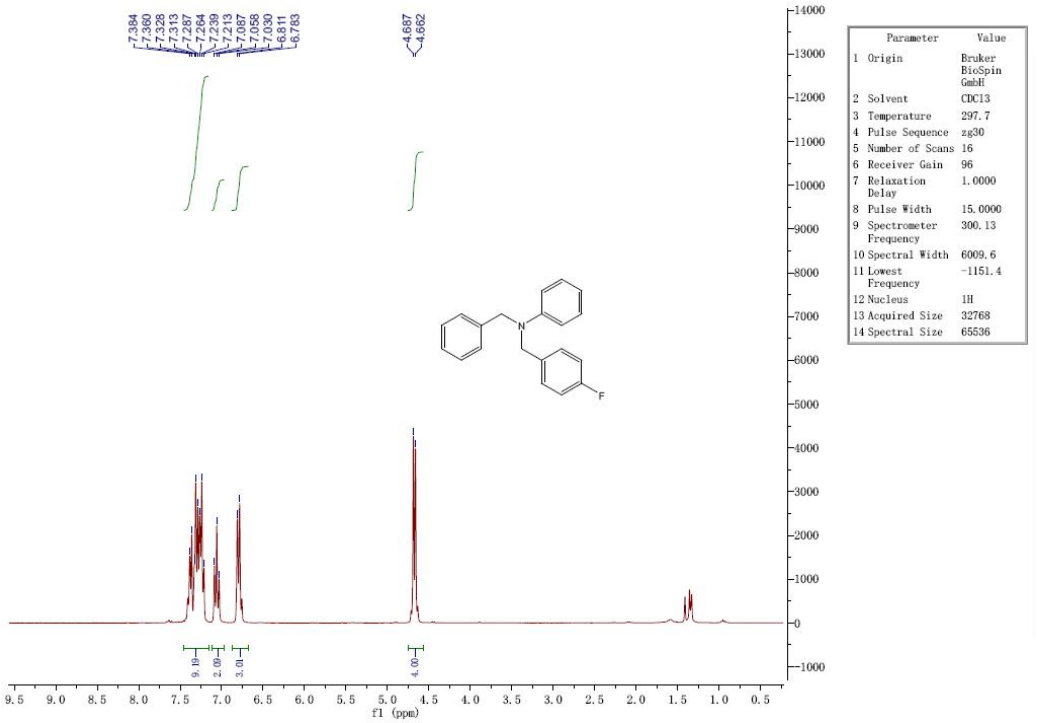






N-benzyl-*N*-(4-fluorobenzyl)aniline (3g)





	Parameter	Value
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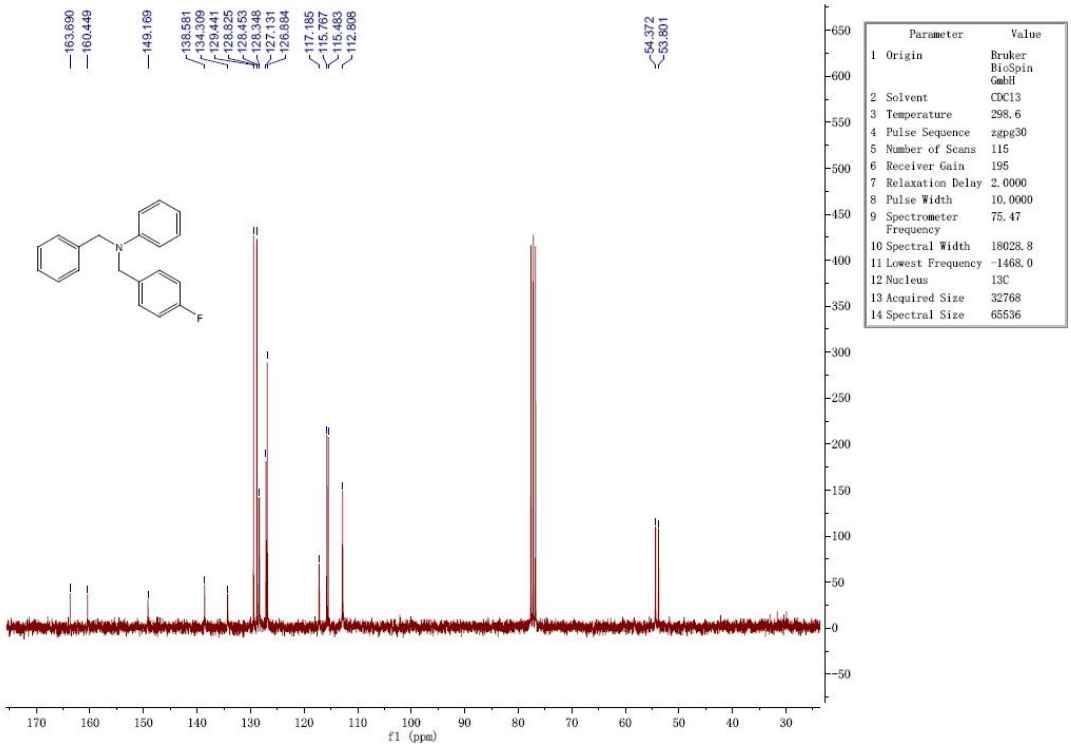
8	Pulse Width	15.0000
9	Spectrometer Frequency	300.13
10	Spectral Width	6009.6
11	Lowest Frequency	-1151.4
12	Nucleus	1H
13	Acquired Size	32768
14	Spectral Size	65536



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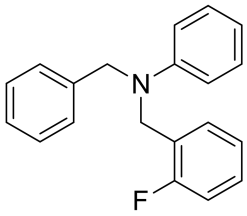
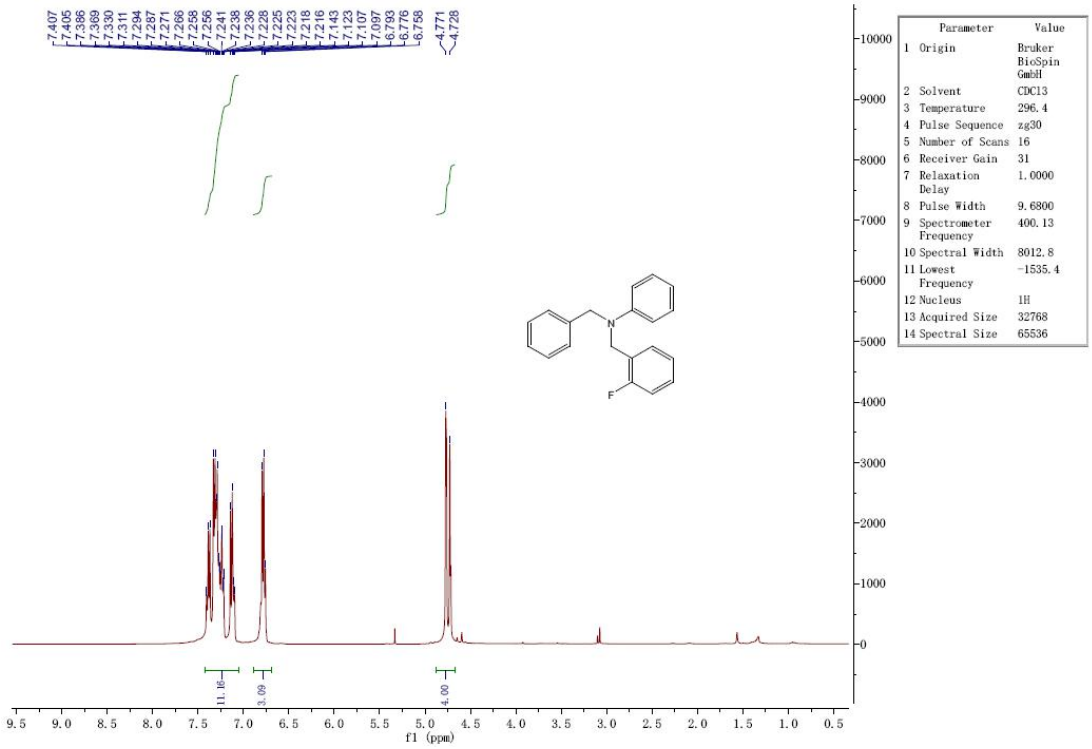
8	Pulse Width	10.0000
9	Spectrometer Frequency	75.47
10	Spectral Width	18028.8
11	Lowest Frequency	-1468.0
12	Nucleus	13C
13	Acquired Size	32768
14	Spectral Size	65536

N-benzyl-*N*-(2-fluorobenzyl)aniline (3h)





	Parameter	Value
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4	Pulse Sequence	zg30
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6	Receiver Gain	31
7	Relaxation Delay	1.0000
8	Pulse Width	9.6800
9	Spectrometer Frequency	400.13
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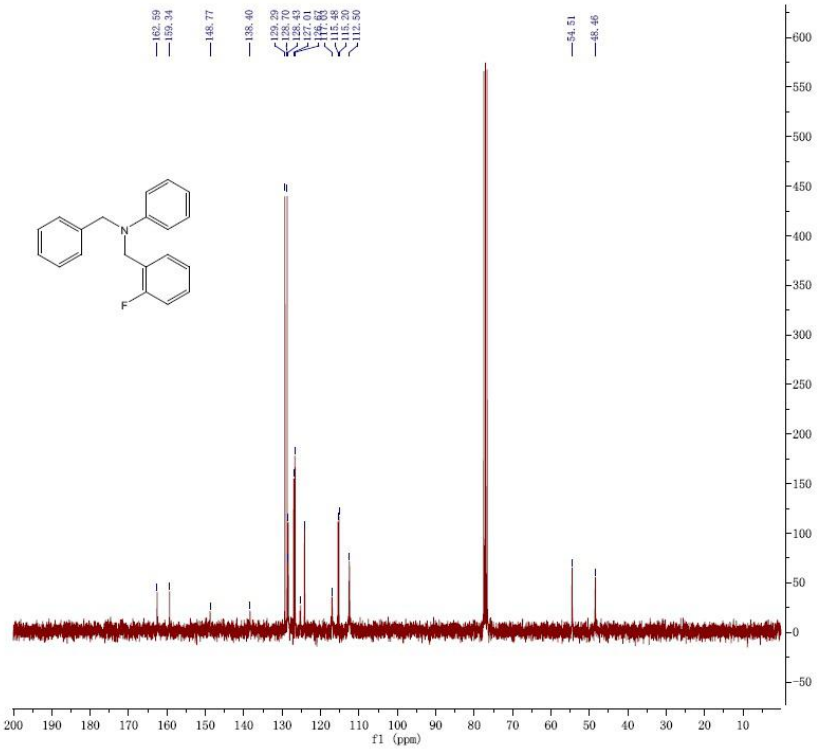
11	Lowest Frequency	-1535.4
12	Nucleus	1H
13	Acquired Size	32768
14	Spectral Size	65536



	Parameter	Value
1	Origin	Bruker BioSpin GmbH
2	Solvent	CDCl3
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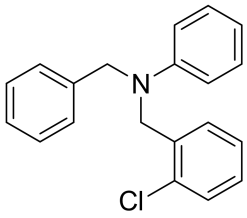
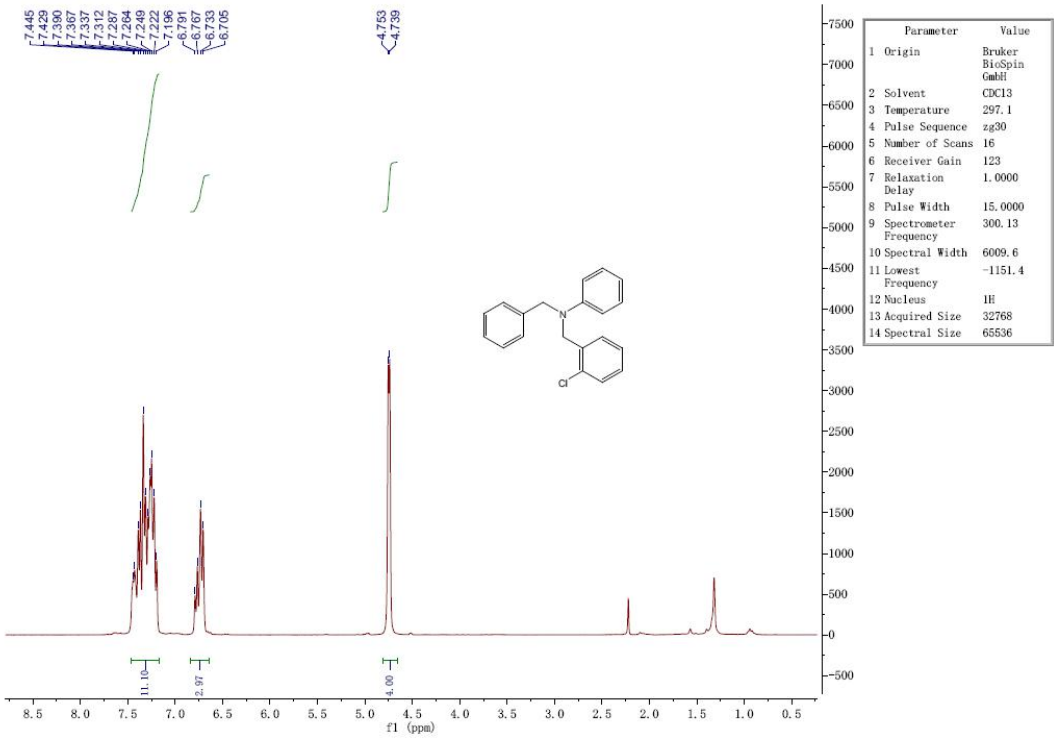
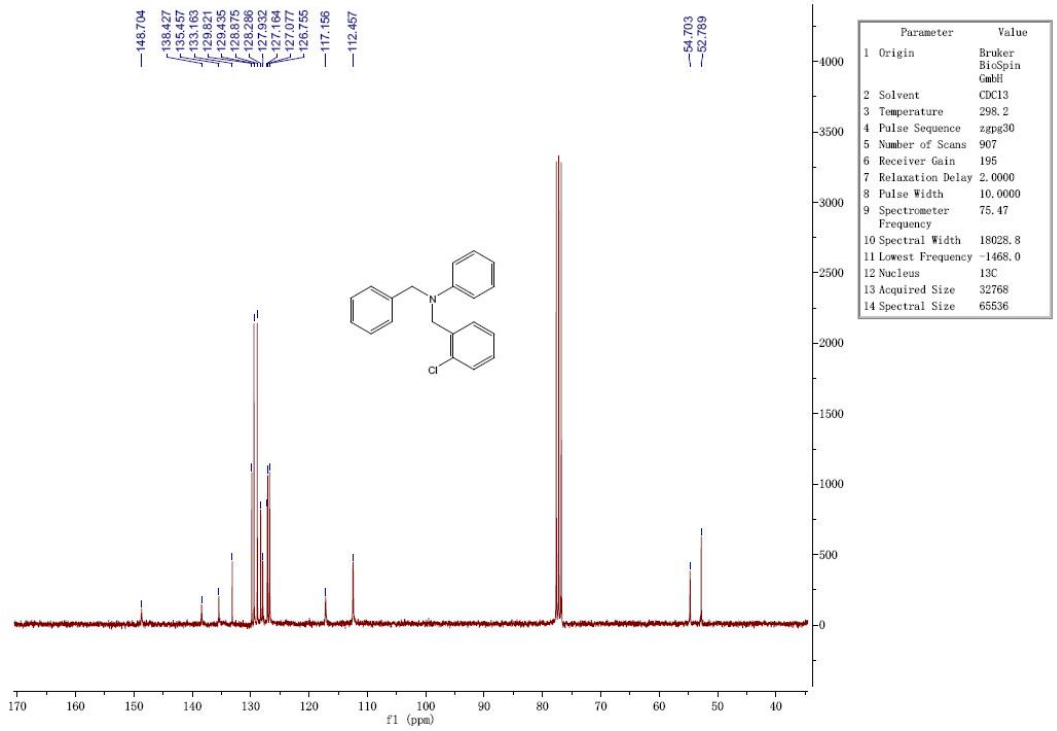
6	Receiver Gain	195
7	Relaxation Delay	2.0000
8	Pulse Width	10.0000
9	Spectrometer Frequency	75.47
10	Spectral Width	18028.8
11	Lowest Frequency	-1468.0
12	Nucleus	13C
13	Acquired Size	32768
14	Spectral Size	65536

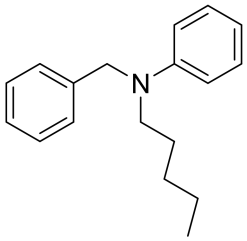
N-benzyl-*N*-(2-chlorobenzyl)aniline (3i)

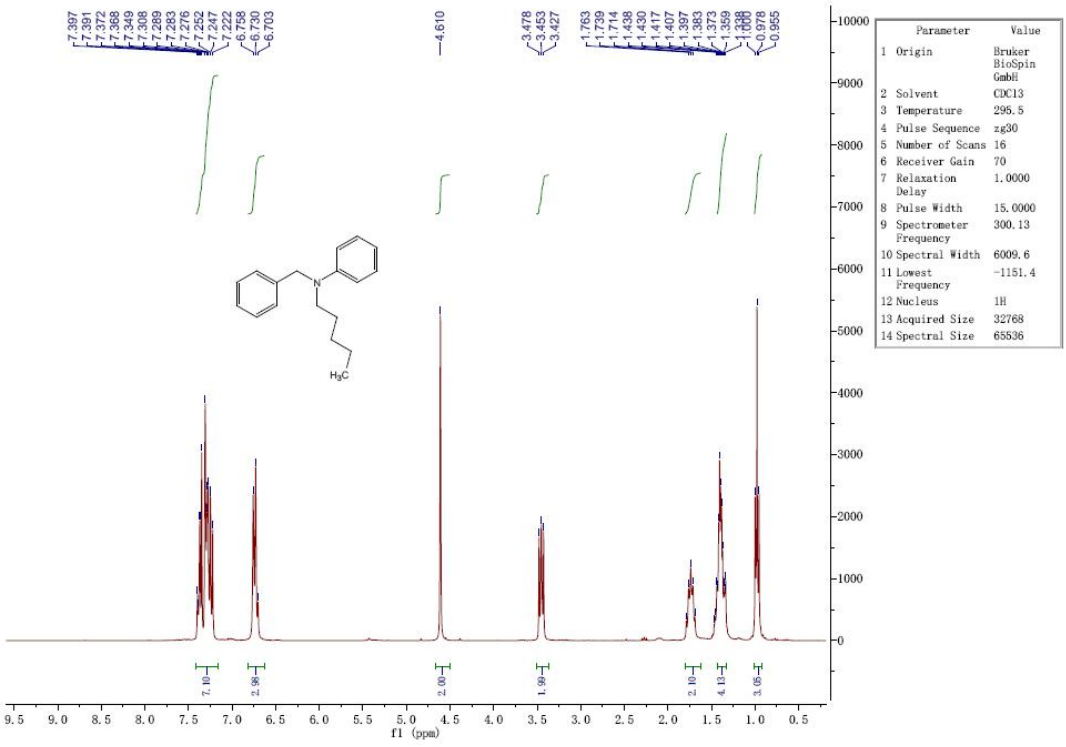


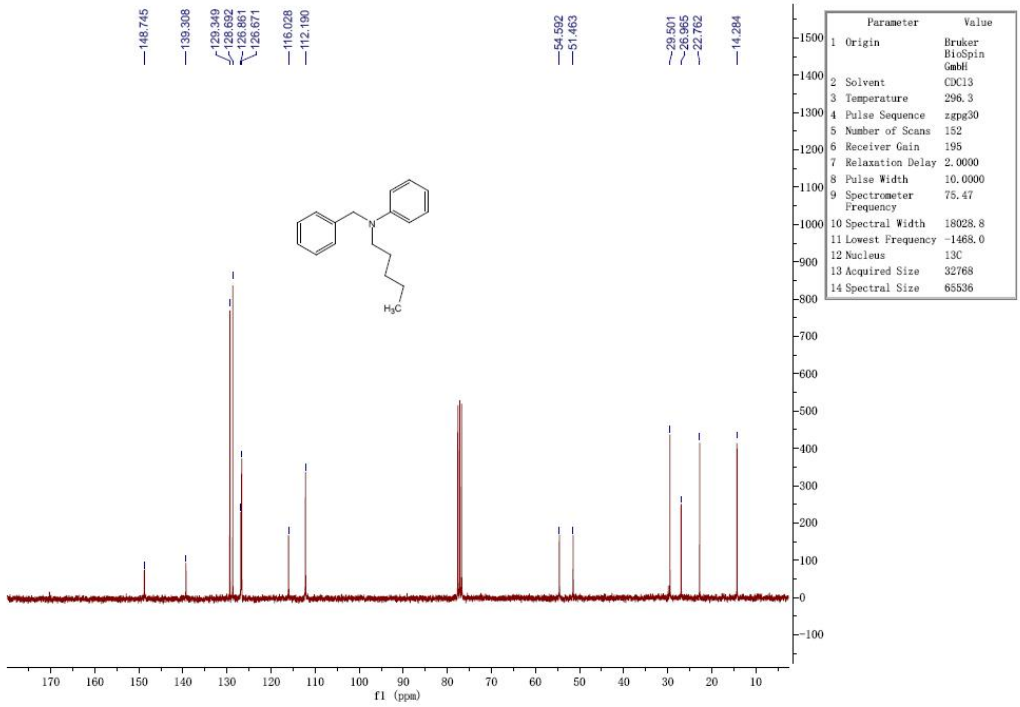




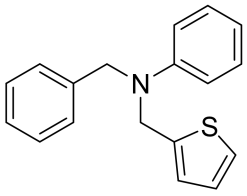
N-benzyl-N-pentylaniline (3j)

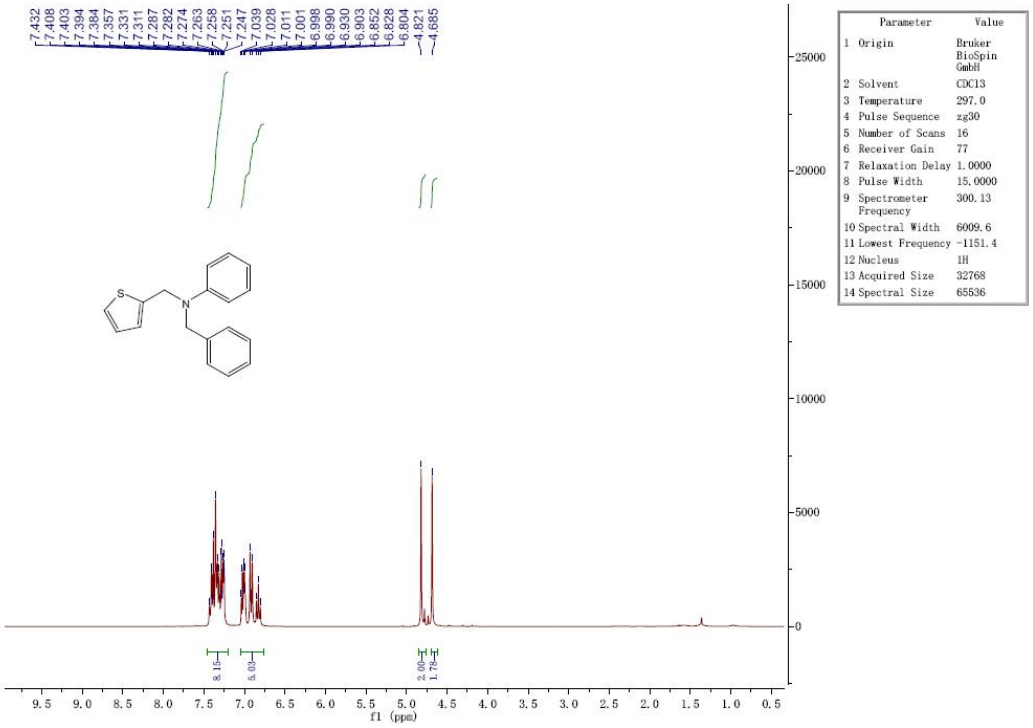






***N*-benzyl-*N*-(thiophen-2-ylmethyl)aniline (3k)**





	Parameter	Value
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6	Receiver Gain	77
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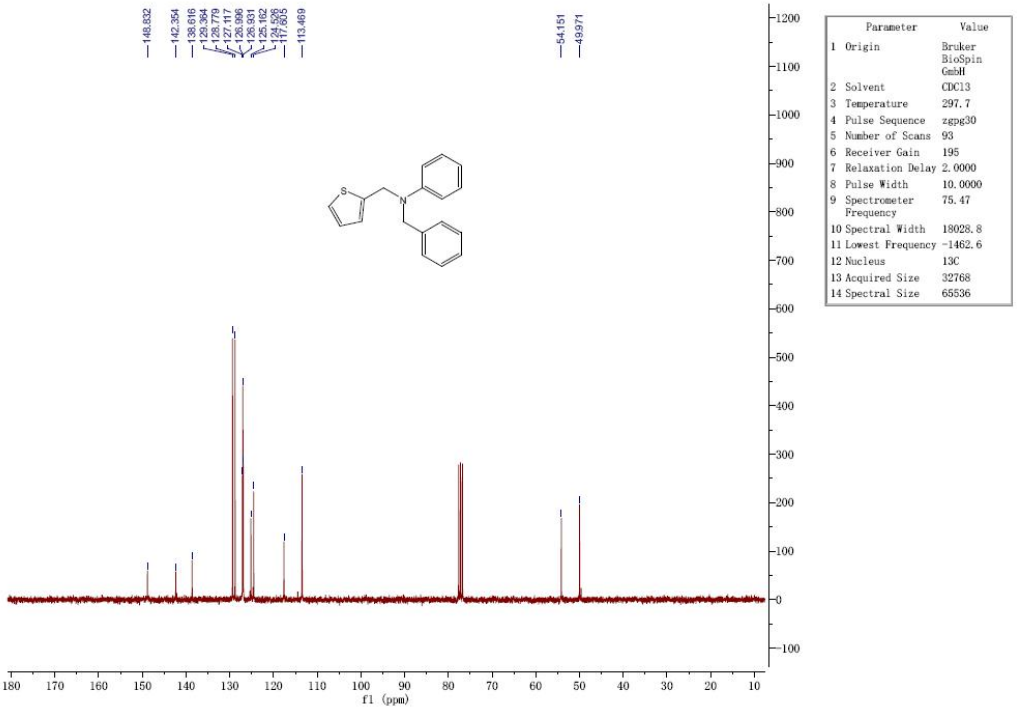
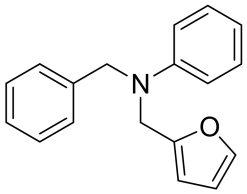
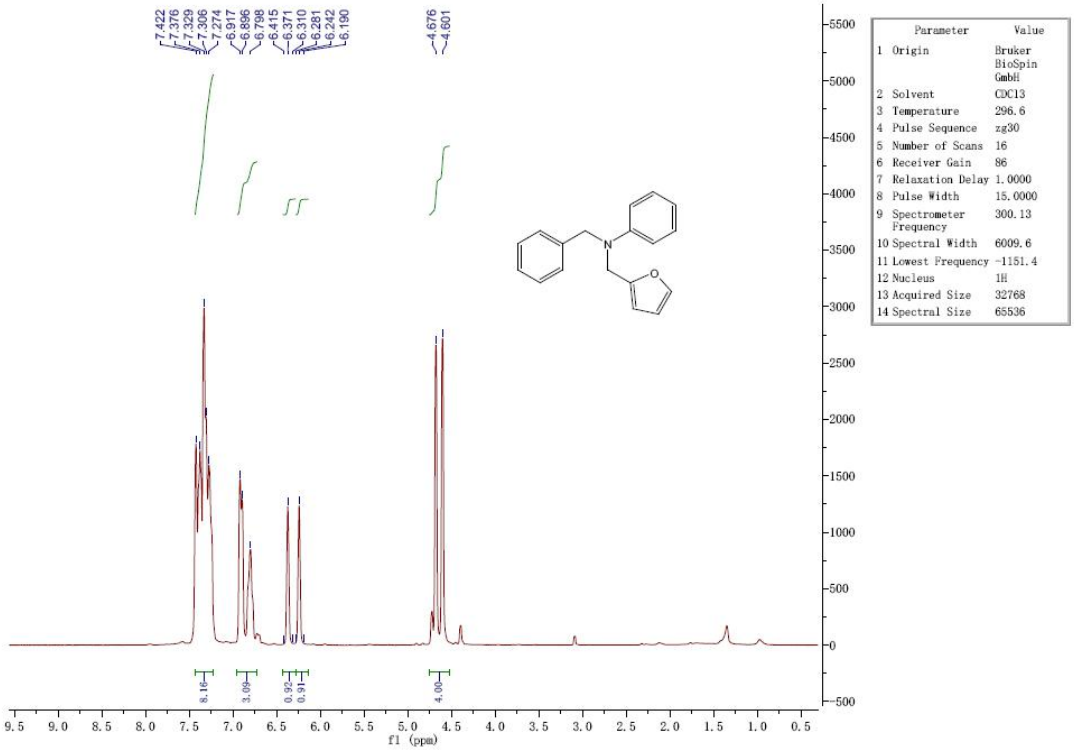
8	Pulse Width	15.0000
9	Spectrometer Frequency	300.13
10	Spectral Width	6009.6
11	Lowest Frequency	-1151.4
12	Nucleus	1H
13	Acquired Size	32768
14	Spectral Size	65536



	Parameter	Value
1	Origin	Bruker BioSpin GmbH
2	Solvent	CDCl3
3	Temperature	297.7
4	Pulse Sequence	zgpg30
5	Number of Scans	93
6	Receiver Gain	195
7	Relaxation Delay	2.0000
8	Pulse Width	10.0000
9	Spectrometer Frequency	75.47
10	Spectral Width	18028.8
11	Lowest Frequency	-1462.6
12	Nucleus	13C
13	Acquired Size	32768
14	Spectral Size	65536

***N*-benzyl-*N*-(furan-2-ylmethyl)aniline (3l)**





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2	Solvent	CDCl3
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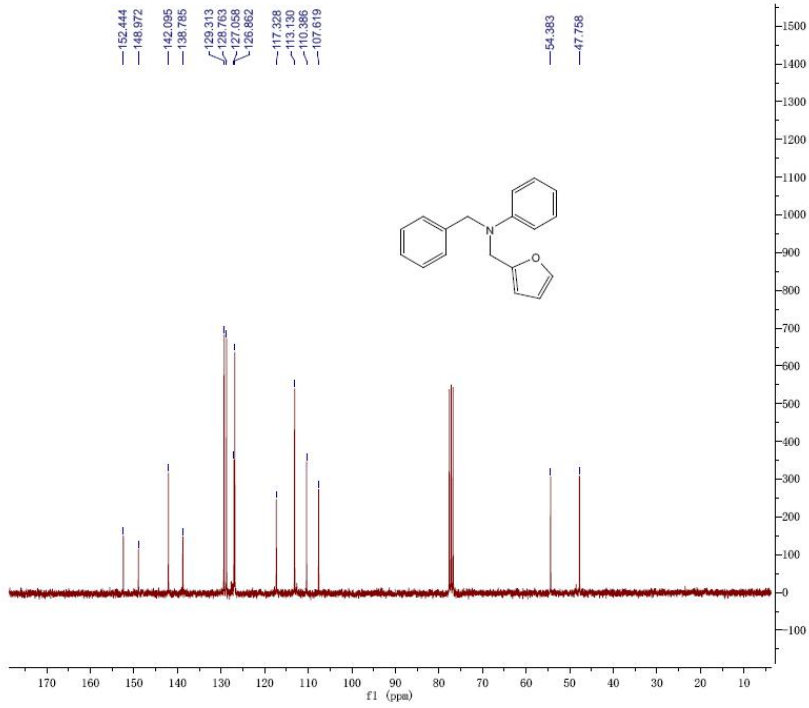
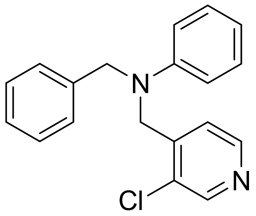
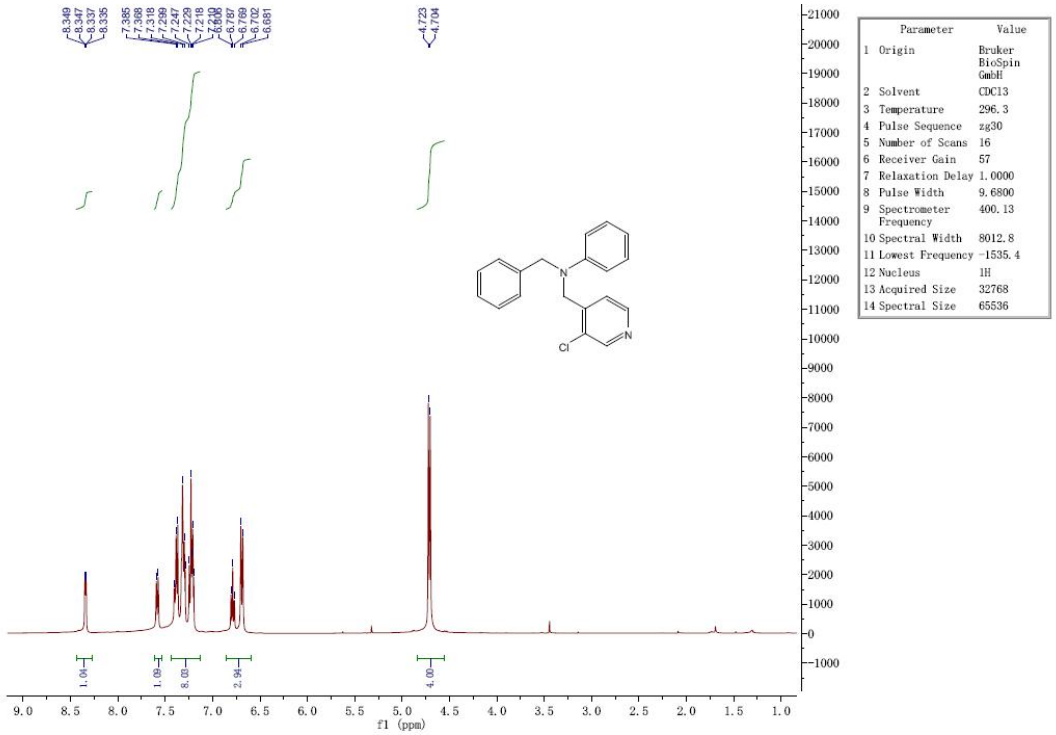
4	Pulse Sequence	zg30
5	Number of Scans	16
6	Receiver Gain	86
7	Relaxation Delay	1.0000
8	Pulse Width	15.0000
9	Spectrometer Frequency	300.13
10	Spectral Width	6009.6
11	Lowest Frequency	-1151.4
12	Nucleus	1H
13	Acquired Size	32768
14	Spectral Size	65536



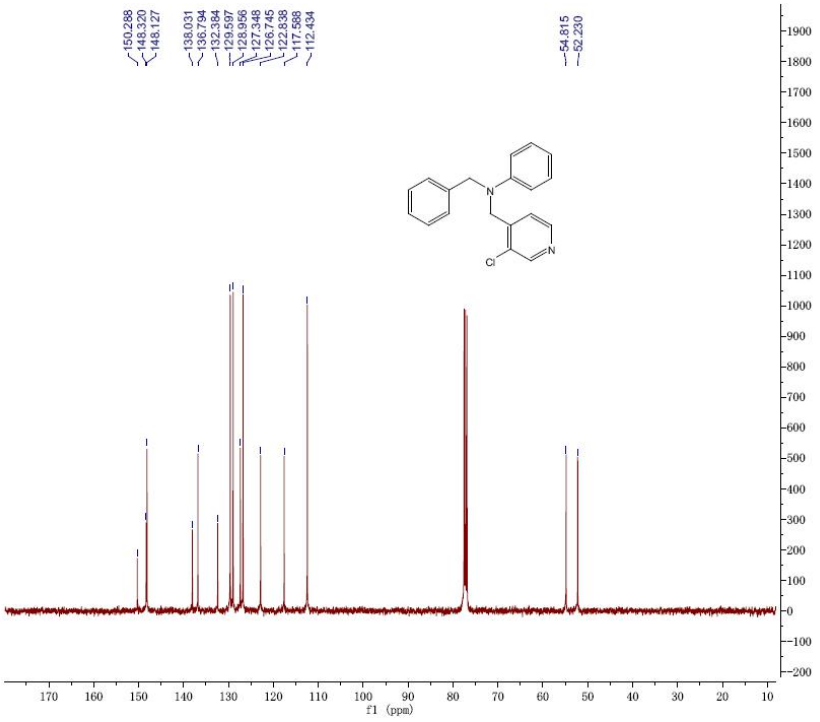
	Parameter	Value
1	Origin	Bruker BioSpin GmbH
2	Solvent	CDCl3
3	Temperature	296.5
4	Pulse Sequence	zgpg30
5	Number of Scans	197
6	Receiver Gain	195
7	Relaxation Delay	2.0000
8	Pulse Width	10.0000
9	Spectrometer Frequency	75.47
10	Spectral Width	18028.8
11	Lowest Frequency	-1468.0
12	Nucleus	13C
13	Acquired Size	32768
14	Spectral Size	65536

N-benzyl-N-((3-chloropyridin-4-yl)methyl)aniline (3m)



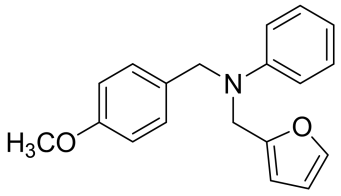


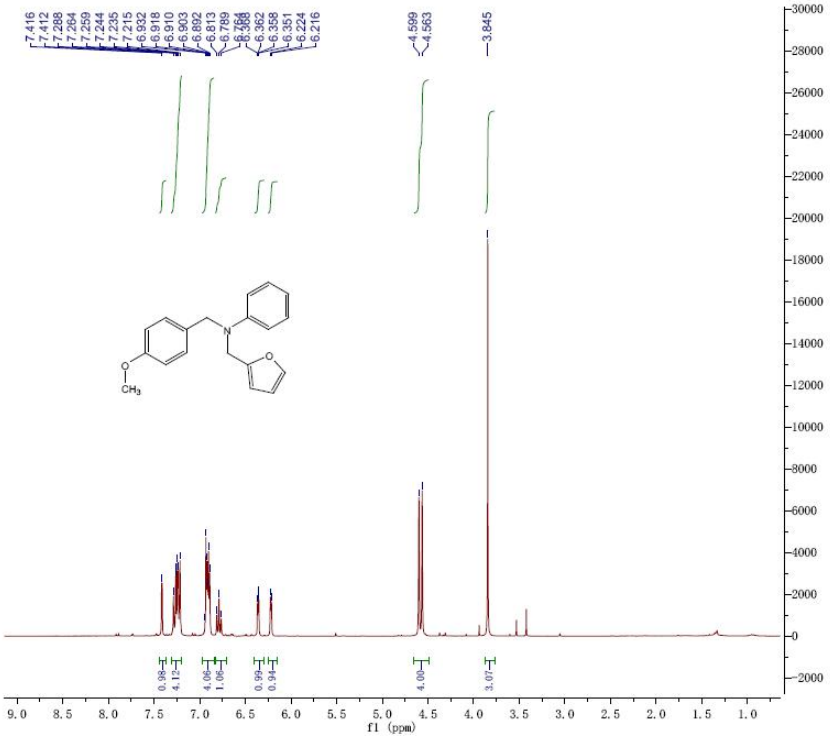
	Parameter	Value
1	Origin	Bruker BioSpin GmbH
2	Solvent	CDCl3
3	Temperature	296.3
4	Pulse Sequence	zg30
5	Number of Scans	16
6	Receiver Gain	57
7	Relaxation Delay	1.0000
8	Pulse Width	9.6800
9	Spectrometer Frequency	400.13
10	Spectral Width	8012.8
11	Lowest Frequency	-1535.4
12	Nucleus	1H
13	Acquired Size	32768
14	Spectral Size	65536



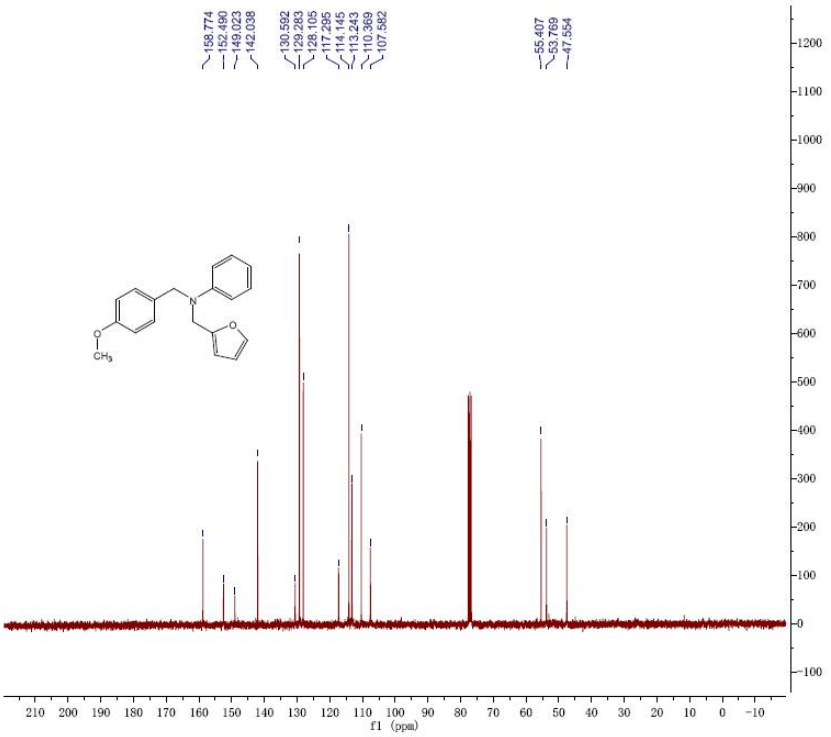
	Parameter	Value
1	Origin	Bruker BioSpin GmbH
2	Solvent	CDCl3
3	Temperature	296.9
4	Pulse Sequence	zgpg30
5	Number of Scans	300
6	Receiver Gain	208
7	Relaxation Delay	2.0000
8	Pulse Width	9.9500
9	Spectrometer Frequency	100.62
10	Spectral Width	24038.5
11	Lowest Frequency	-1958.0
12	Nucleus	13C
13	Acquired Size	32768
14	Spectral Size	65536

N-(furan-2-ylmethyl)-N-(4-methoxybenzyl)aniline (3n)



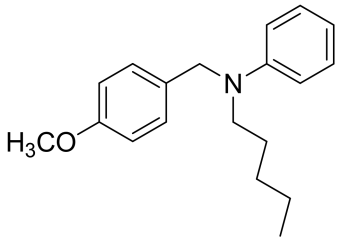


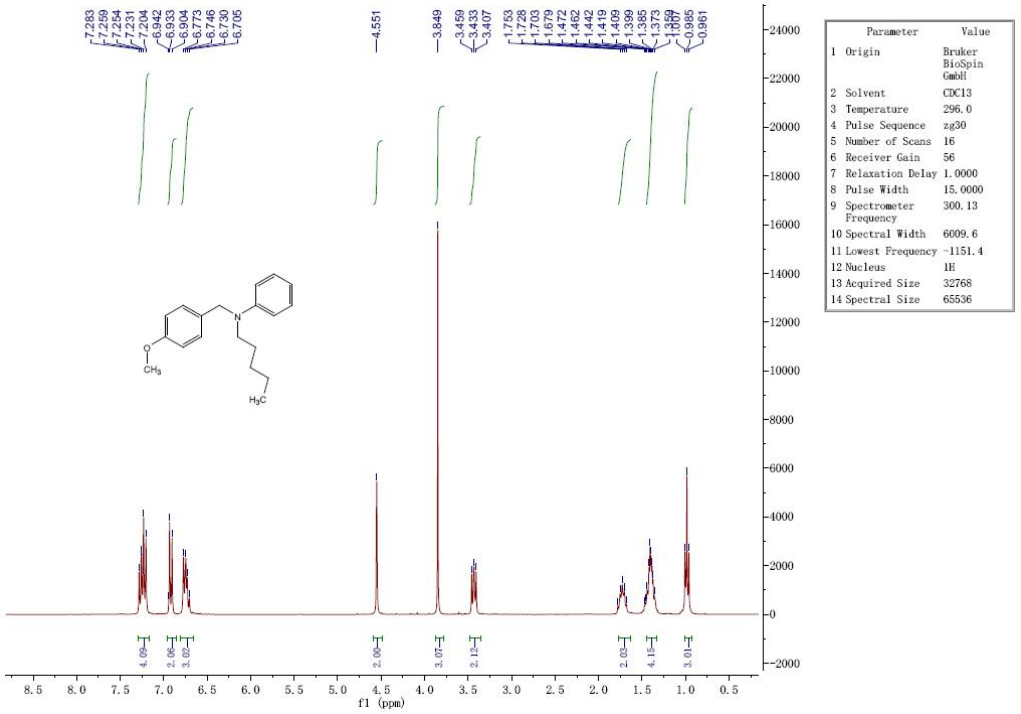
	Parameter	Value
1	Origin	Bruker BioSpin GmbH
2	Solvent	CDCl3
3	Temperature	295.5
4	Pulse Sequence	zg30
5	Number of Scans	16
6	Receiver Gain	77
7	Relaxation Delay	1.0000
8	Pulse Width	15.0000
9	Spectrometer Frequency	300.13
10	Spectral Width	6009.6
11	Lowest Frequency	-1151.4
12	Nucleus	1H
13	Acquired Size	32768
14	Spectral Size	65536

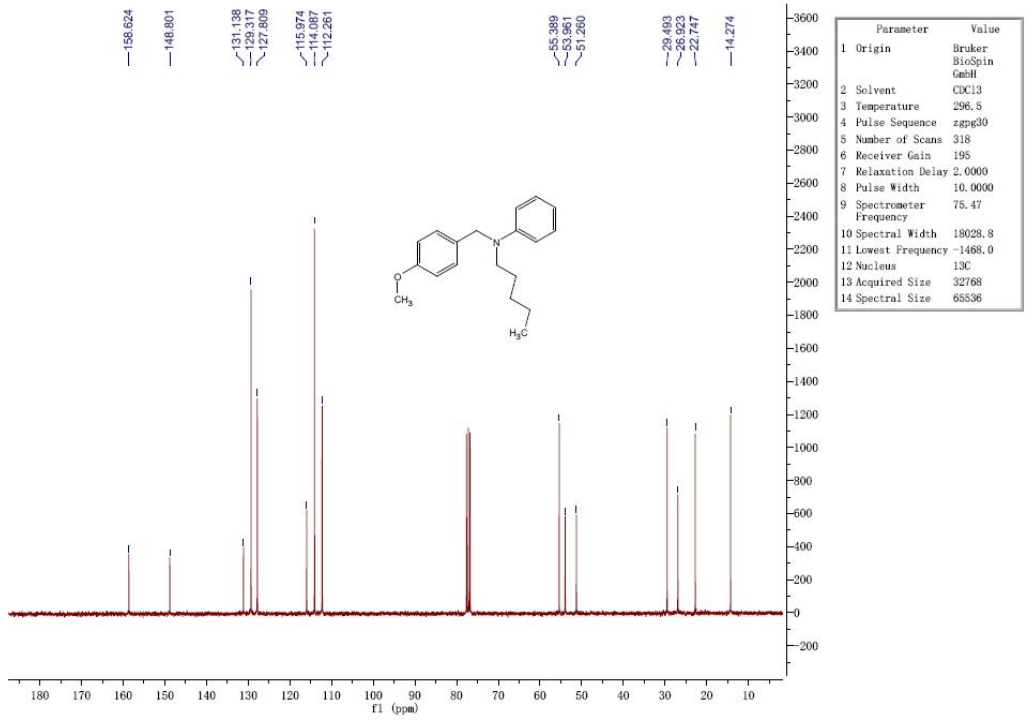


	Parameter	Value
1	Origin	Bruker BioSpin GmbH
2	Solvent	CDCl3
3	Temperature	296.3
4	Pulse Sequence	zgpg30
5	Number of Scans	139
6	Receiver Gain	195
7	Relaxation Delay	2.0000
8	Pulse Width	10.0000
9	Spectrometer Frequency	75.47
10	Spectral Width	18028.8
11	Lowest Frequency	-1468.0
12	Nucleus	13C
13	Acquired Size	32768
14	Spectral Size	65536

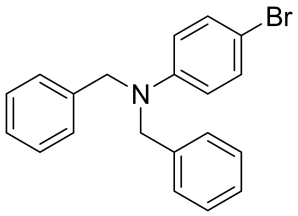
N-(4-methoxybenzyl)-N-pentylaniline (3o)

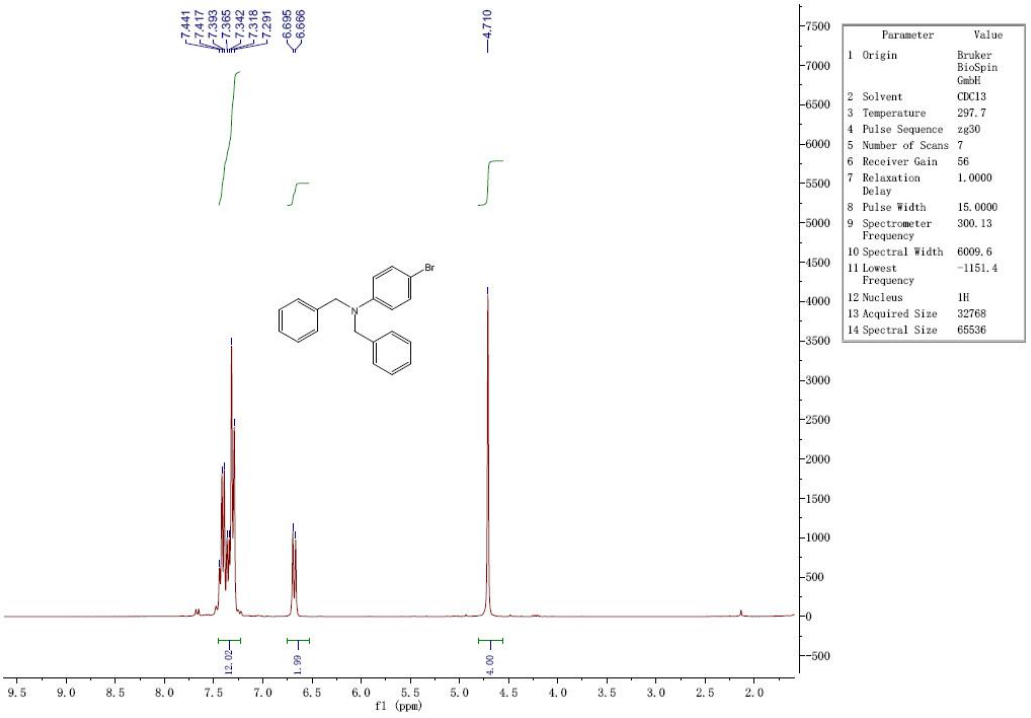




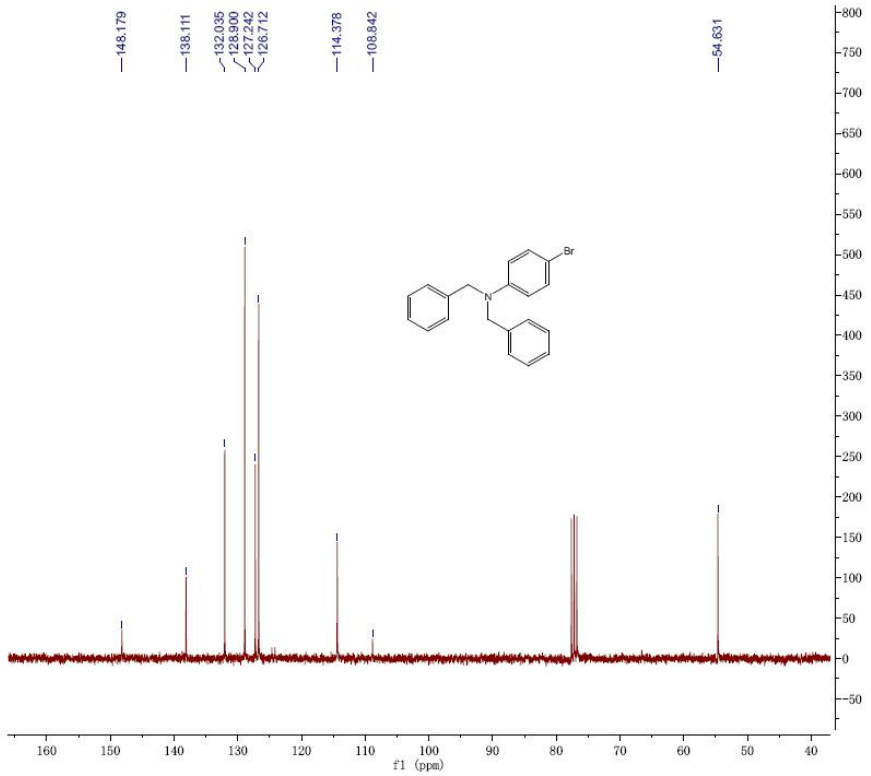


***N,N*-dibenzyl-4-bromoaniline (3p)**



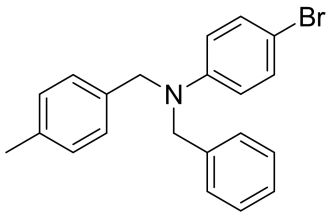


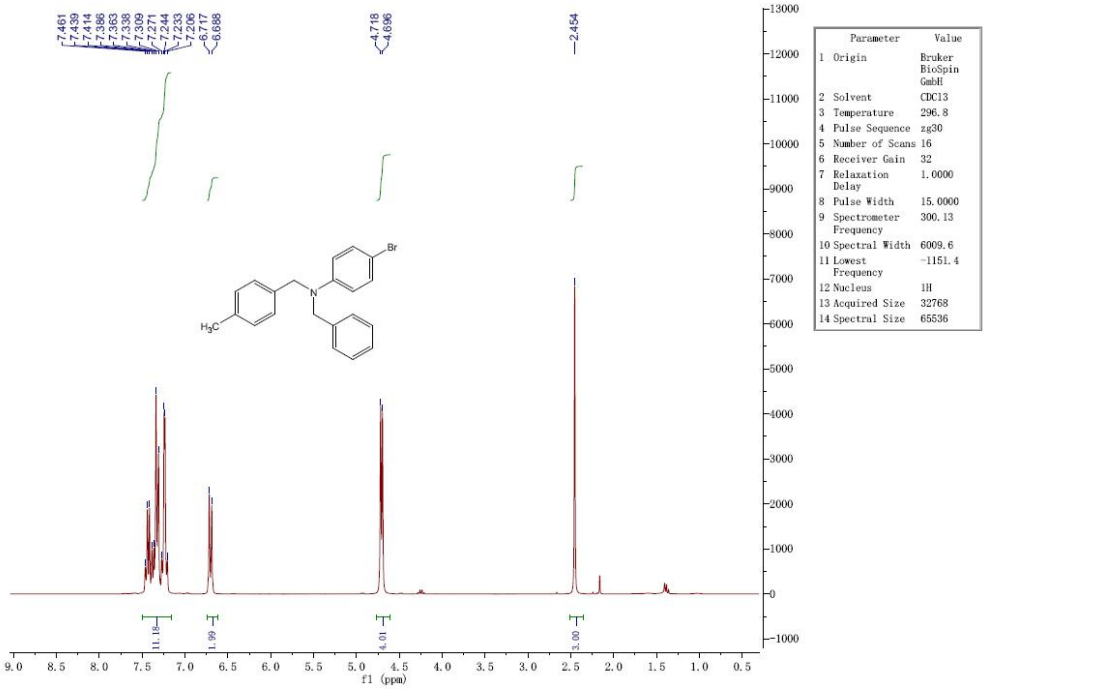
	Parameter	Value
1	Origin	Bruker BioSpin GmbH
2	Solvent	CDCl3
3	Temperature	297.7
4	Pulse Sequence	zg30
5	Number of Scans	7
6	Receiver Gain	56
7	Relaxation Delay	1.0000
8	Pulse Width	15.0000
9	Spectrometer Frequency	300.13
10	Spectral Width	6009.6
11	Lowest Frequency	-1151.4
12	Nucleus	1H
13	Acquired Size	32768
14	Spectral Size	65536



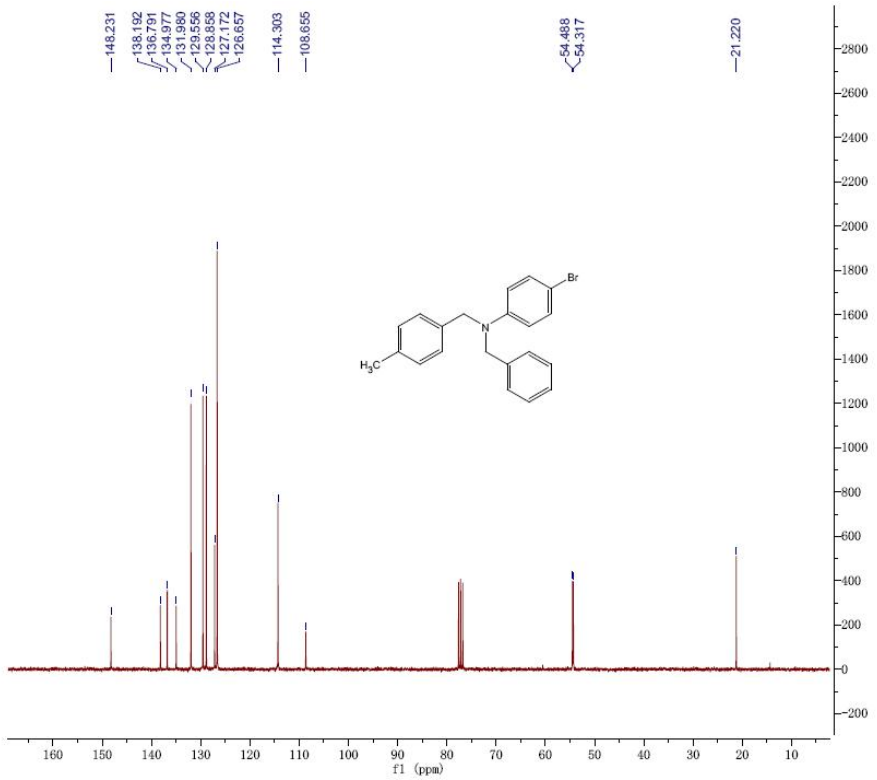
	Parameter	Value
1	Origin	Bruker BioSpin GmbH
2	Solvent	CDCl3
3	Temperature	298.7
4	Pulse Sequence	zgpg30
5	Number of Scans	63
6	Receiver Gain	195
7	Relaxation Delay	2.0000
8	Pulse Width	10.0000
9	Spectrometer Frequency	75.48
10	Spectral Width	18028.8
11	Lowest Frequency	-1468.0
12	Nucleus	13C
13	Acquired Size	32768
14	Spectral Size	65536

N-benzyl-4-bromo-*N*-(4-methylbenzyl)aniline (3q)



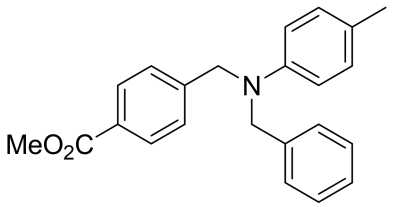


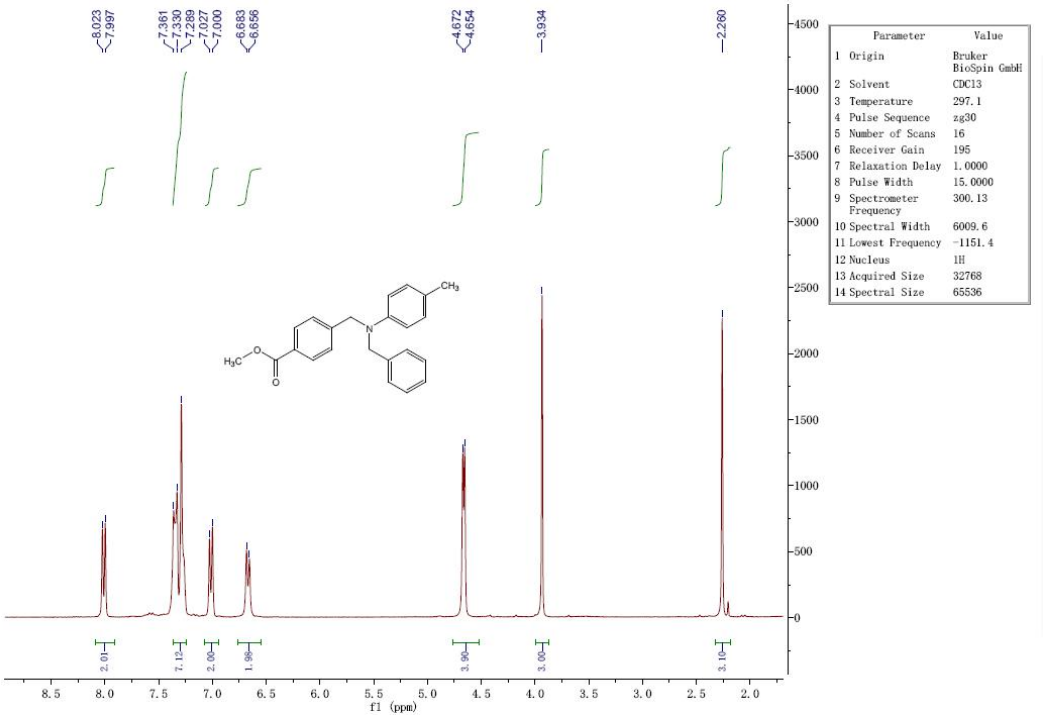
	Parameter	Value
1	Origin	Bruker BioSpin GmbH
2	Solvent	CDCl3
3	Temperature	296.8
4	Pulse Sequence	zg30
5	Number of Scans	16
6	Receiver Gain	32
7	Relaxation Delay	1.0000
8	Pulse Width	15.0000
9	Spectrometer Frequency	300.13
10	Spectral Width	6009.6
11	Lowest Frequency	-1151.4
12	Nucleus	1H
13	Acquired Size	32768
14	Spectral Size	65536

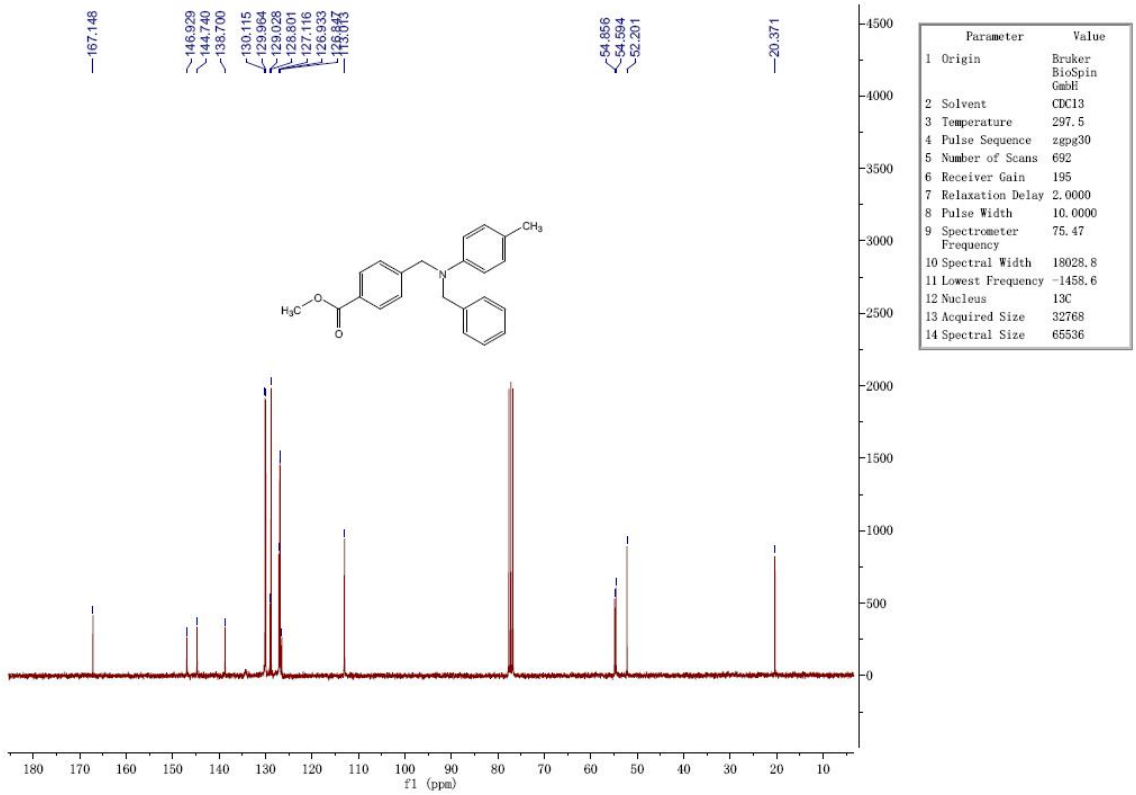


	Parameter	Value
1	Origin	Bruker BioSpin GmbH
2	Solvent	CDCl3
3	Temperature	297.6
4	Pulse Sequence	zgpg30
5	Number of Scans	132
6	Receiver Gain	195
7	Relaxation Delay	2.0000
8	Pulse Width	10.0000
9	Spectrometer Frequency	75.47
10	Spectral Width	18028.8
11	Lowest Frequency	-1468.0
12	Nucleus	13C
13	Acquired Size	32768
14	Spectral Size	65536

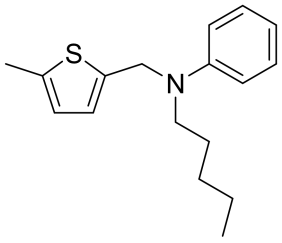
Methyl 4-((benzyl(*p*-tolyl)amino)methyl)benzoate (3r)

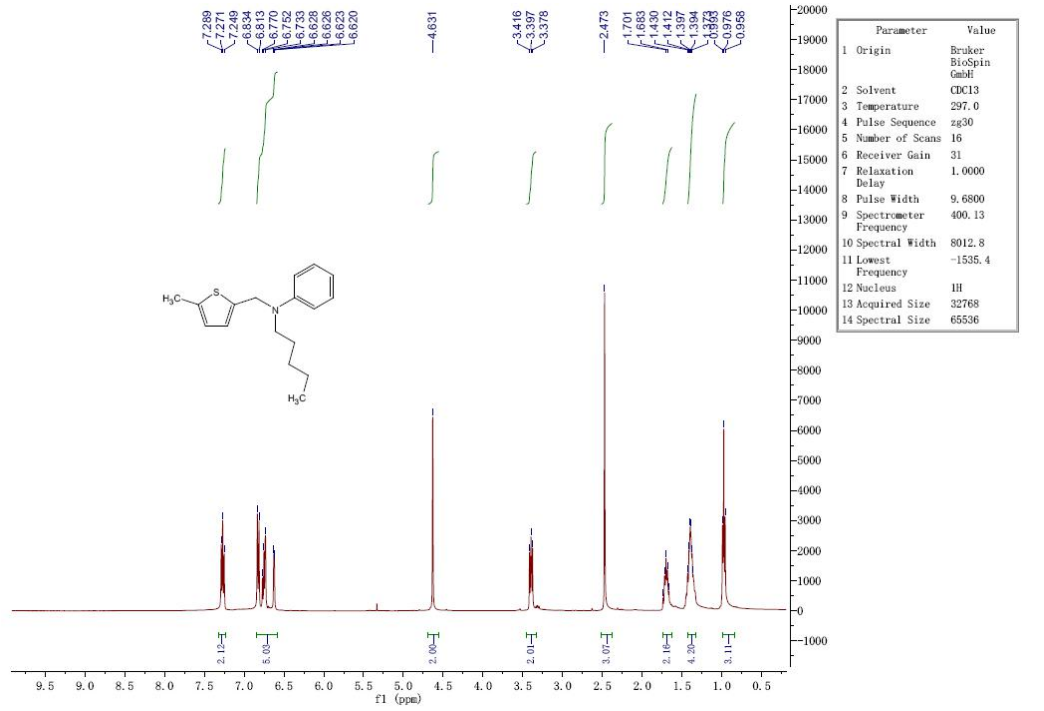




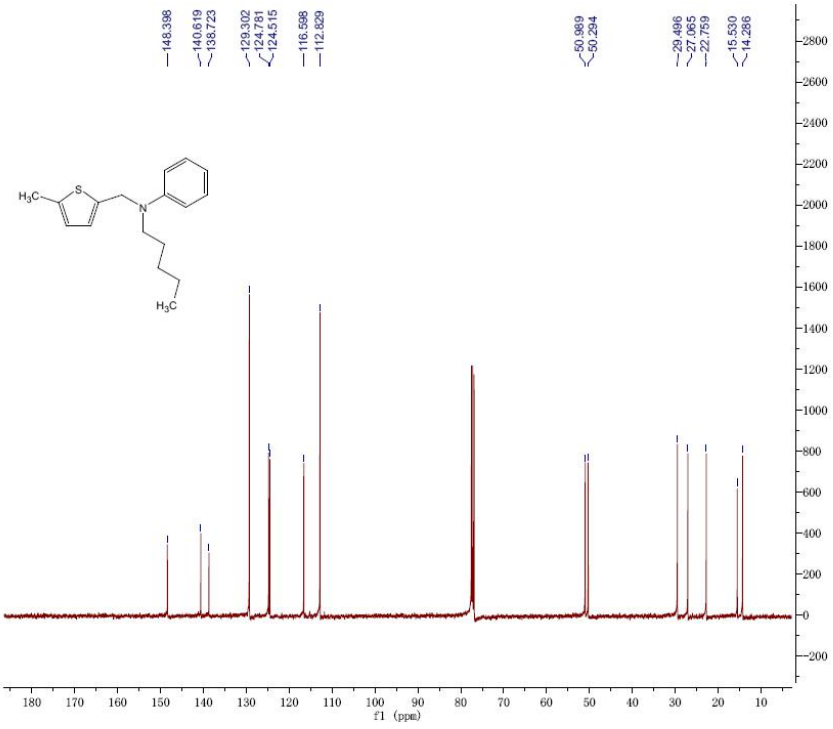


N-((5-methylthiophen-2-yl)methyl)-N-pentylaniline (3s)



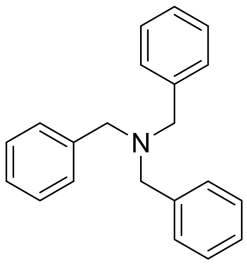


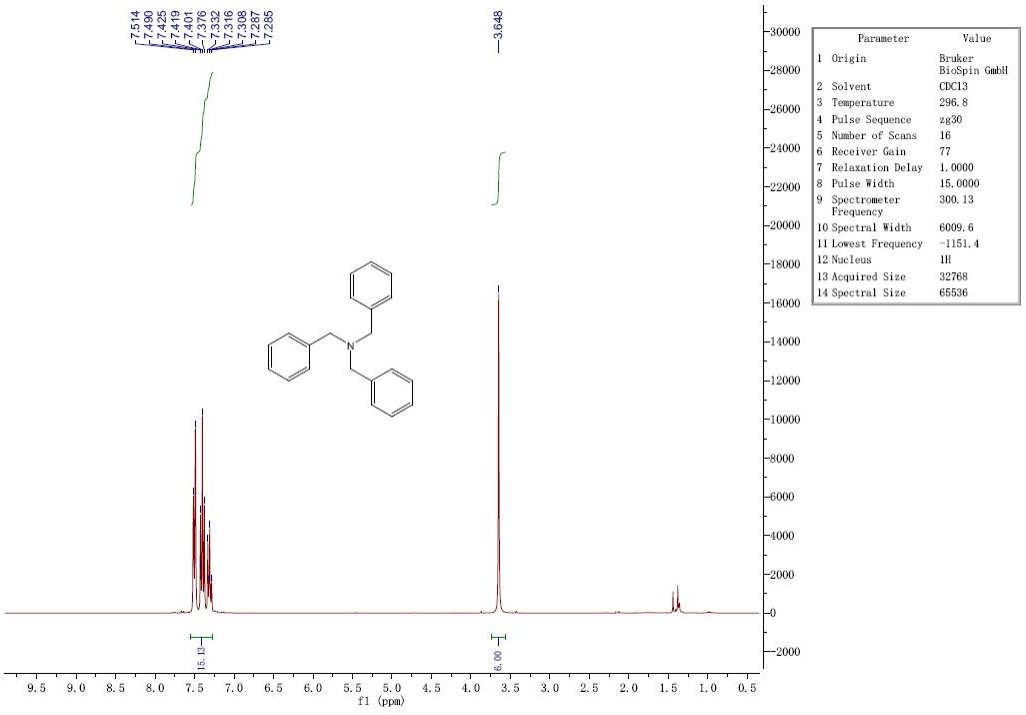
	Parameter	Value
1	Origin	Bruker BioSpin GmbH
2	Solvent	CDCl3
3	Temperature	297.0
4	Pulse Sequence	zg30
5	Number of Scans	16
6	Receiver Gain	31
7	Relaxation Delay	1.0000
8	Pulse Width	9.6800
9	Spectrometer Frequency	400.13
10	Spectral Width	8012.8
11	Lowest Frequency	-1535.4
12	Nucleus	1H
13	Acquired Size	32768
14	Spectral Size	65536



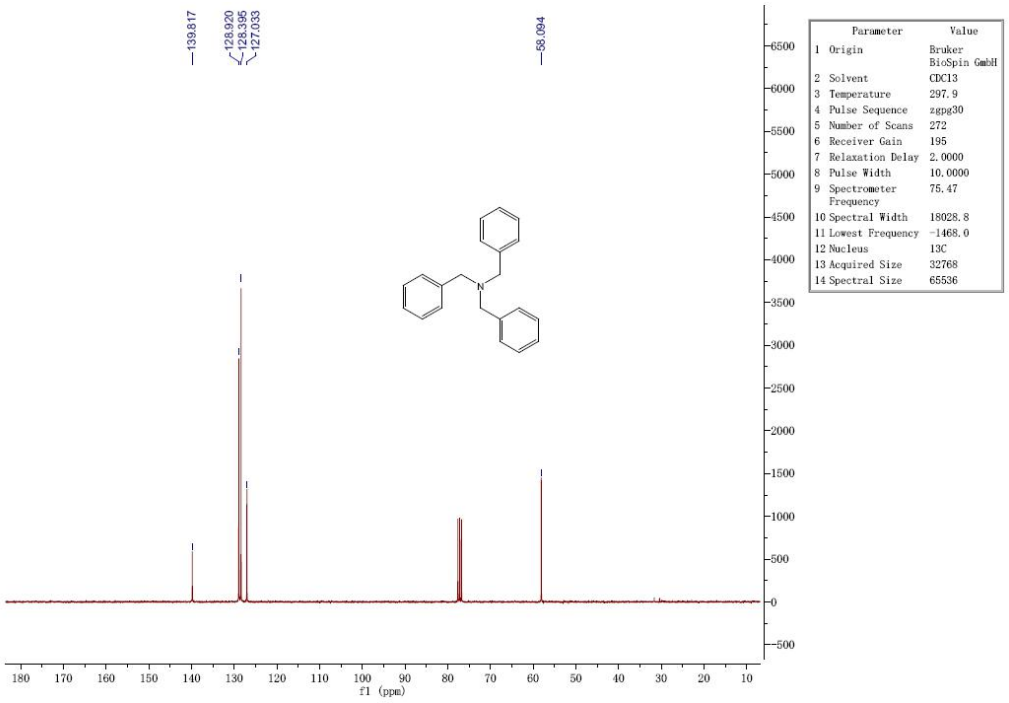
	Parameter	Value
1	Origin	Bruker BioSpin GmbH
2	Solvent	CDCl3
3	Temperature	297.6
4	Pulse Sequence	zgpg30
5	Number of Scans	300
6	Receiver Gain	208
7	Relaxation Delay	2.0000
8	Pulse Width	9.9500
9	Spectrometer Frequency	100.62
10	Spectral Width	24038.5
11	Lowest Frequency	-1946.4
12	Nucleus	13C
13	Acquired Size	32768
14	Spectral Size	65536

Tribenzylamine (3t)



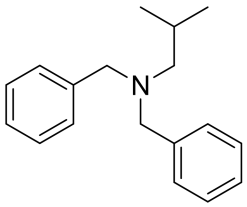


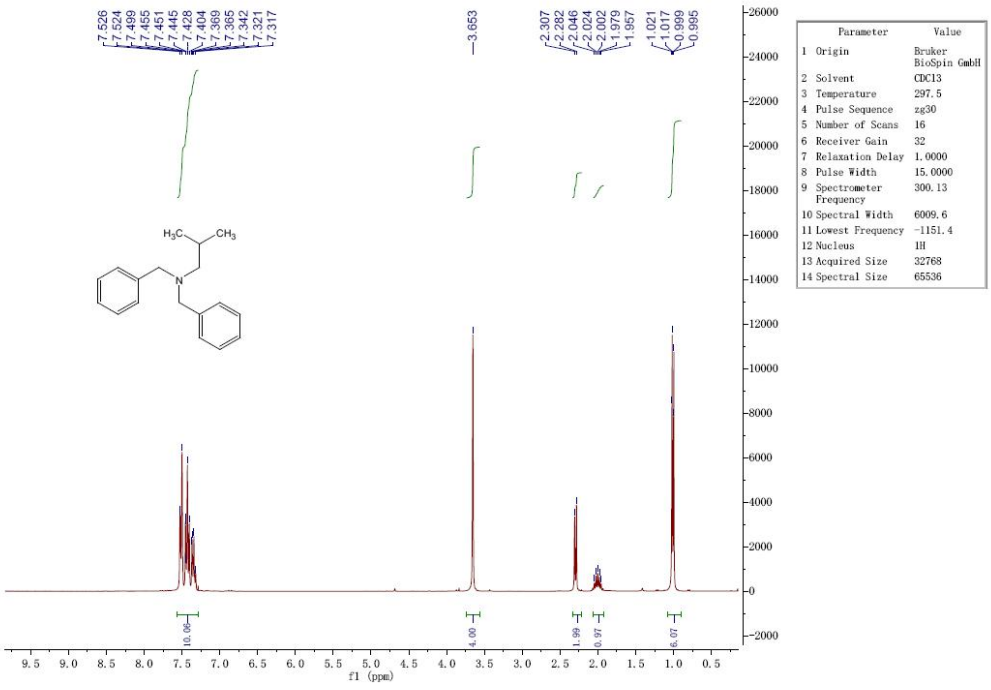
	Parameter	Value
1	Origin	Bruker BioSpin GmbH
2	Solvent	CDCl3
3	Temperature	296.8
4	Pulse Sequence	zg30
5	Number of Scans	16
6	Receiver Gain	77
7	Relaxation Delay	1.0000
8	Pulse Width	15.0000
9	Spectrometer Frequency	300.13
10	Spectral Width	6009.6
11	Lowest Frequency	-1151.4
12	Nucleus	1H
13	Acquired Size	32768
14	Spectral Size	65536



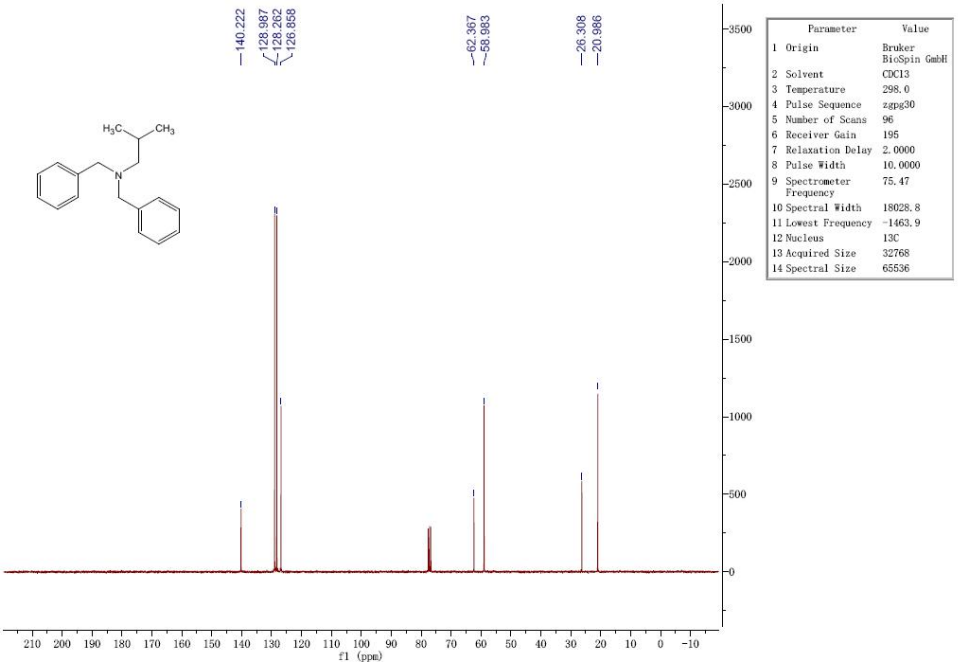
	Parameter	Value
1	Origin	Bruker BioSpin GmbH
2	Solvent	CDCl3
3	Temperature	297.9
4	Pulse Sequence	zgpg30
5	Number of Scans	272
6	Receiver Gain	195
7	Relaxation Delay	2.0000
8	Pulse Width	10.0000
9	Spectrometer Frequency	75.47
10	Spectral Width	18028.8
11	Lowest Frequency	-1468.0
12	Nucleus	13C
13	Acquired Size	32768
14	Spectral Size	65536

***N*,*N*-dibenzyl-2-methylpropan-1-amine (3u)**





	Parameter	Value
1	Origin	Bruker BioSpin GmbH
2	Solvent	CDCl3
3	Temperature	297.5
4	Pulse Sequence	zg30
5	Number of Scans	16
6	Receiver Gain	32
7	Relaxation Delay	1.0000
8	Pulse Width	15.0000
9	Spectrometer Frequency	300.13
10	Spectral Width	6009.6
11	Lowest Frequency	-1151.4
12	Nucleus	1H
13	Acquired Size	32768
14	Spectral Size	65536



	Parameter	Value
1	Origin	Bruker BioSpin GmbH
2	Solvent	CDCl3
3	Temperature	298.0
4	Pulse Sequence	zgpg30
5	Number of Scans	96
6	Receiver Gain	195
7	Relaxation Delay	2.0000
8	Pulse Width	10.0000
9	Spectrometer Frequency	75.47
10	Spectral Width	18028.8
11	Lowest Frequency	-1463.9
12	Nucleus	13C
13	Acquired Size	32768
14	Spectral Size	65536