

Relay Tricyclic Pd(II)/Ag(I) Catalysis: Design of a Four-Component Reaction Driven by Nitrene-Transfer on Isocyanide Yields Inhibitors of EGFR

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Table of Contents:

	Page no
Experimental Procedures	
S1 General Considerations	S2
S2 Detailed Results of screening	S2
S3 General procedures	S6
S4 Analytical data of compounds	S6
S5 Control Experiments	S15
S5.1 Radical trap	S15
S5.2 Evolution of species	S16
S5.3 Chemical kinetics of 4-CR	S16
S5.4 Experiments on H/D Exchange	S20
S5.5 Experiment with labelled substrates	S21
S5.6 Proposed Reaction Mechanism	S21
S6 Experimental Biology	S22
S6.1 Cell Culture and reagents	S22
S6.2 Evaluation of anticancer activity of the synthesized compounds by MTT cytotoxicity Assay	S22
S6.3 Human Peripheral Blood Mononuclear Cells (HPBMCs) culturing and treatment by investigational molecules to assess cytotoxicity of investigational molecules	S23
S6.4 Pyrazolo[1,5-c]quinazoline as EGFR inhibitors	S24
S6.5 Measurement of intracellular reactive oxygen species (ROS)	S25
S6.6 Determination of mitochondrial membrane potential (μM)	S26
S6.7 Cell apoptosis assay using Propidium Iodide	S26
S7 Molecular docking studies	S27
S8 Crystallographic Analysis	S28
Result and Discussion	
S9 Pharmacological activity and literature search for synthesis of pyrazolo[1,5-c]quinazolines	S34
S10 Construction of diversity-rich library of pyrazolo[1,5-c]quinazolines	S36
S11 References	S37
S12 Copies of ^1H , ^{13}C NMR and HRMS spectra	S38

Experimental Procedures

S1. General Considerations

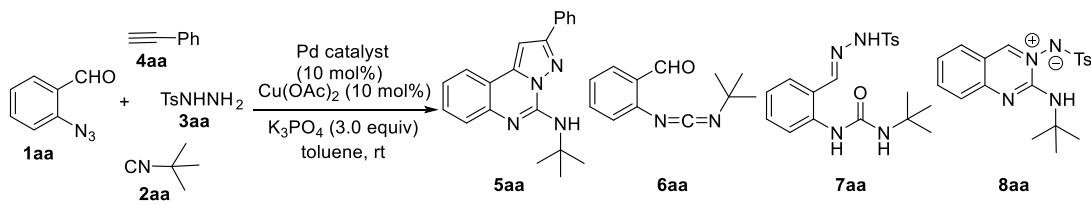
All other reagents were purchased from Aldrich or Spectrochem used as such without purification. Analytical TLC was performed using 2.5*5cm plated coated with a 0.25mm thickness of silica gel (60F-254 Merck) and visualization was accomplished with UV light or I_2 / KMnO_4 staining. ^1H and ^{13}C NMR spectra were obtained from Bruker's Ascend 500 MHz spectrophotometer operating at 500.3 MHz for ^1H and 125.8 MHz for ^{13}C experiments. The chemical shifts are reported in ppm scale relative to residual CDCl_3 (δ 7.269 ppm) for ^1H and residual TMS. ^{13}C NMR spectra were reported relative to CDCl_3 (δ 77.00 ppm). Melting Points were recorded on BuchiM-655 Melting Point Apparatus and are uncorrected. High-resolution mass spectra (HRMS) were taken in the ESI positive ion mode. The abbreviations for multiplicities are used as: s=singlet, d=doublet, t=triplet, q=quartet, dd=doublet of doublets, m=multiplet, br s=broad singlet, dt=doublet of triplets, tt=triplet of triplets, ddd=doublet of doublets. All starting materials were prepared as reported in the literature.¹

S2. Detailed Results of Screening

S2.1 Table S1: Initial Screening

We started our investigation using 2-azido benzaldehyde **1aa** (1 equiv), *tert.* butyl isocyanide **2aa** (1.2 equiv), tosyl hydrazide **3aa** (1 equiv) and phenylacetylene **4aa** (1.5 equiv) as a benchmark reaction using PdCl_2 (10 mol%),

AgOTf (10 mol%), as catalytic system, in toluene with K_3PO_4 as a base, which did not lead to the formation of desired product **5aa** (Table S1, Entry 1).

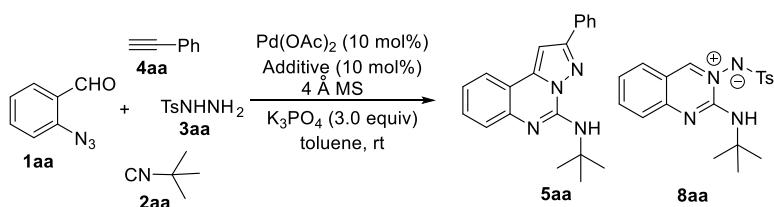


S.No.	Catalyst	Yield ^a				
		5aa	6aa	7aa	8aa	1aa
1.	PdCl ₂	-	-	-	-	95
2.	Pd(PPh ₃) ₂ Cl ₂	5	45	-	-	45
3.	Pd(PPh ₃) ₄	7	25	40	-	24
4.	Pd ₂ (dba) ₃	20	20	30	-	10
5.	Pd(OAc) ₂	33	-	50	5	-
6.	Pd(OAc) ₂	75 ^b	-	-	13	-

^aIsolated yield, ^baddition of 4 Å MS

On extensive screening of various Pd sources, we found that $Pd_2(dba)_3$ and $Pd(OAc)_2$ could afford the desired product **5aa** in 20% and 33% isolated yields. Of these, $Pd(OAc)_2$ proved to be optimal because of complete consumption of starting material (Entry 5). We observed the formation of **7aa** as a major side product (Characterized by single-crystal X-ray diffraction analysis, refer Section 6), formed mainly due to condensation of **6aa** and **3aa** followed by nucleophilic attack of water. The origin of **7aa** could be due to the nucleophilic attack of water on carbodiimide **6aa**. Gratifyingly, a substantial improvement in the yield was observed on employing the 4 Å MS (Entry 6).

S2.2 Table S2: Additive effect

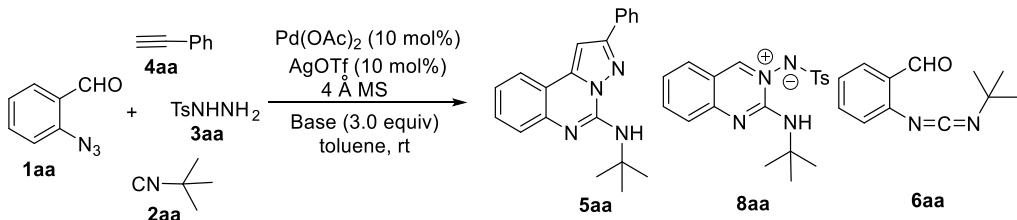


S.No.	Additive	Yield ^a	
		5aa	8aa
1.	$Cu(OAc)_2$	75	13
2.	CuI	63	21
3.	CuBr	55	28
4.	CuCl	59	23
5.	AgOTf	96	-
6.	AgOAc	78	15
7.	Ag_2CO_3	72	21
8.	$AgSbF_6$	15	68

^aIsolated yield

To enhance the reactivity, we surveyed a variety of additives which revealed that both silver and copper salts facilitate this reaction. Of these, AgOTf was the most efficient additive for this strategy by overcomes the formation of a side product.

S2.3 Table S3: Base effect

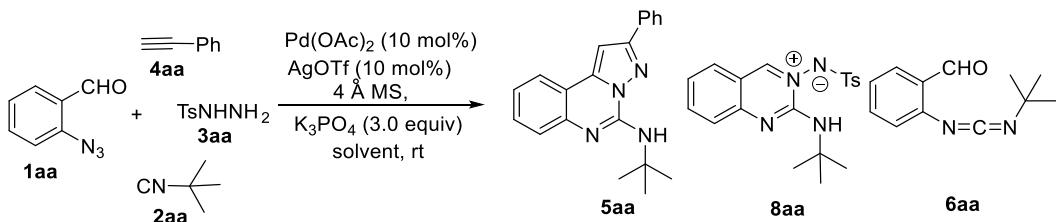


S.No.	Base	Yield ^a		
		5aa	8aa	6aa
1.	DABCO	82	-	12
2.	NaOAc	38	31	25
3.	Cs ₂ CO ₃	35 ^b	-	30
4.	KO'Bu	48	-	40
5.	DBU	64	-	26
6.	K ₃ PO ₄	96	-	-

^aIsolated yield, ^b1aa recovered.

The choice of base significantly affected the final outcome of this reaction. K₃PO₄ as a base produced best results among various bases screened in Table S3.

S2.4 Table S4: Solvent Effect

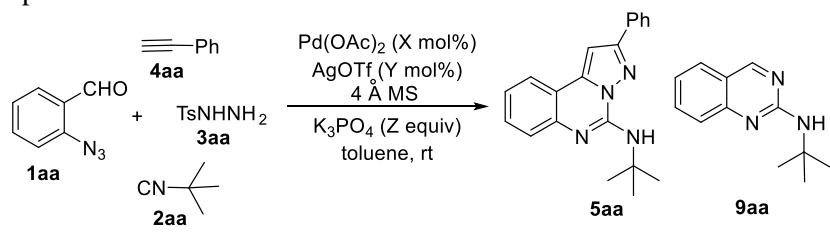


S.No.	Solvent	Yield ^a		
		5aa	8aa	6aa
1.	THF	62	20	-
2.	DCE	30	40	25
3.	DMF	- ^b	-	-
4.	DMSO	-	47 ^b	-
5.	dioxane	49	15	12
6.	toluene	96	-	-

^aIsolated yield, ^b1aa recovered

Next, we screened various solvents for 4-CR. The highest yield of **5aa** was obtained in toluene.

S2.5 Table S5: Final optimization

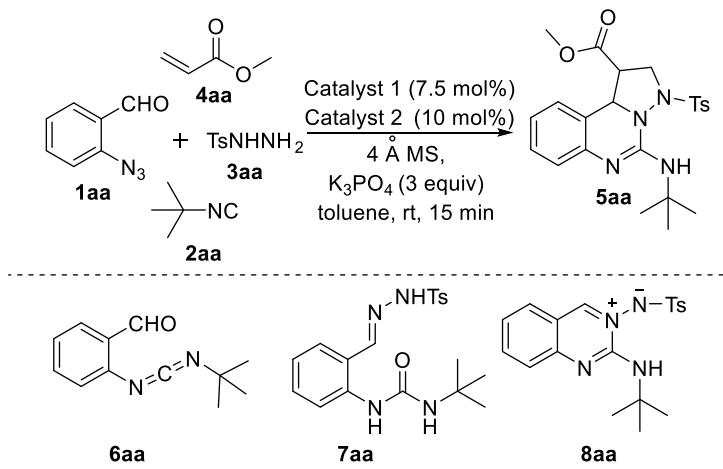


S. No	X	Y	Z	Yield ^a (%)
1.	10	10	3	96
2.	7.5	10	3	95
3.	5	10	3	61
4.	7.5	7.5	3	74
5.	7.5	5	3	43
6.	7.5	10	2	61
7.	7.5	10	1	55
8.	7.5	10	3	25 ^b

^aIsolated yield, ^bMajor side product isolated at 60 °C

Next, we scrutinized the stoichiometric screening of the catalyst, additive and base and observed that decreased in the catalytic loading of Pd(OAc)₂ from 10 mol % to 7.5 mol % resulted the desired product **5aa** in comparable yields, but there is substantial reduction in the yield, when 5 mol % of either of the catalysts was employed (entry 3). Furthermore, decrease in the loading of additive and base lead to diminished yields. Evidently, there is a detrimental effect of increasing the temperature from rt to 60 °C, which afforded the lower yield of **5aa** with major side product **9aa** that was characterized by single-crystal X-ray diffraction analysis (refer section S8).

S2.6 Table S6: Optimization of reaction condition for alkene.



Entry	Catalyst 1	Catalyst 2	Additives	isolated yield ^a (%)				
				5aa	6aa	7aa	8aa	1aa
1.	PdCl ₂	Cu(OAc) ₂	-	-	-	-	-	95
2.	Pd(PPh ₃) ₂ Cl ₂	Cu(OAc) ₂	-	10	40	-	-	44
3.	Pd(PPh ₃) ₄	Cu(OAc) ₂	-	5	20	50	-	21
4.	Pd ₂ (dba) ₃	Cu(OAc) ₂	-	15	20	25	-	10
5.	Pd(OAc) ₂	Cu(OAc) ₂	-	30	-	45	5	-

6.	Pd(OAc) ₂	Cu(OAc) ₂	4 Å MS	80	-	-	10	-
7.	Pd(OAc) ₂	CuI	4 Å MS	71	-	-	20	-
8.	Pd(OAc) ₂	CuBr	4 Å MS	65	-	-	30	-
9.	Pd(OAc) ₂	CuCl	4 Å MS	55	-	-	27	-
10.	Pd(OAc) ₂	AgOTf	4 Å MS	95	-	-	-	-
11.	Pd(OAc) ₂	AgOAc	4 Å MS	70	-	-	25	-
12.	Pd(OAc) ₂	Ag ₂ CO ₃	4 Å MS	65	-	-	30	-
13.	Pd(OAc) ₂	AgSbF ₆	4 Å MS	42	-	-	25	-
14.	-	AgOTf	4 Å MS	-	-	-	-	91
15.	Pd(OAc) ₂	-	4 Å MS	38	-	-	55	-
16.	Pd(OAc) ₂	AgOTf,	4 Å MS	93 ^b	-	-	-	-

^aReaction Condition: **1aa** (1 equiv), **2aa** (1.2 equiv), **3aa** (1 equiv), **26aa** (1 equiv). ^b 7.5 mol % of Pd(OAc)₂

Thorough optimisation studies for 4CR of alkene were carried out. An analogous reaction condition was required for this reaction compared to reactions involving alkynes. We observed that both palladium and co-catalyst (either Cu or Ag) were essential for this transformation.

S3. General Procedure for the synthesis of compounds **5** and **26**

A mixture of 2-Azidobenzaldehyde (1.0 equiv), isocyanide (1.2 equiv), Pd(OAc)₂(7.5 mol%), 4 Å MS, TsNHNH₂ (1.0 equiv), alkyne/alkenes (1.5 equiv), AgOTf (10 mol%) and K₃PO₄ (3.0 equiv) in toluene was added to a 20 mL Schlenk tube. The formed mixture was stirred at rt for 15 min. The reaction mixture was diluted with ethyl acetate (15 mL) and washed with water (10 mL). The organic layer was separated, dried on Na₂SO₄ and evaporated under vacuum. The crude product, so obtained, was purified by column chromatography to afford the desired product.

S4. Analytical Data for the compounds **5aa-5hb** and **26aa-26ha**

5aa: *N-(tert-butyl)-2-phenylpyrazolo[1,5-*c*]quinazolin-5-amine*

Pale yellow solid, Yield: 0.102 g (95%), m.p.: 83-85 °C. ¹H NMR (500 MHz, CDCl₃): δ 8.04 (d, 2H, aromatic C-H, *J* = 7.0 Hz), 7.91 (dd, 1H, aromatic C-H, *J* = 7.85, 1.1 Hz), 7.67 (d, 1H, aromatic C-H, *J* = 8.2 Hz), 7.54-7.48 (m, 3H, aromatic C-H), 7.44-7.42 (m, 1H, aromatic C-H), 7.29 (td, 1H, aromatic C-H, *J* = 8.0, 1.1 Hz), 7.19 (s, 1H, aromatic C-H), 6.50 (br s, 1H, N-H), 1.69 (s, 9H, sp³ C-H). ¹³C NMR (125 MHz, CDCl₃): δ 153.4, 142.2, 141.9, 140.9, 132.6, 129.7, 128.9, 128.8, 126.6, 126.1, 123.1, 122.8, 115.9, 95.5, 51.9, 29.1. HRMS (ESI) calcd for C₂₀H₂₁N₄ [M+H]⁺ 317.1761, found 317.1755.

5ab: *N-(tert-butyl)-2-(4-methoxyphenyl)pyrazolo[1,5-*c*]quinazolin-5-amine*

White solid, Yield: 0.113 g (96%), m.p.: 138-140 °C. ¹H NMR (500 MHz, CDCl₃): δ 7.96 (d, 2H, aromatic C-H, *J* = 8.8 Hz), 7.90 (dd, 1H, aromatic C-H, *J* = 7.8, 1.2 Hz), 7.66 (d, 1H, aromatic C-H, *J* = 7.9 Hz), 7.51 (td, 1H, aromatic C-H, *J* = 7.2, 1.4 Hz), 7.27 (td, 1H, aromatic C-H, *J* = 8.0, 1.0 Hz), 7.11 (s, 1H, aromatic C-H), 7.02 (d, 2H, aromatic C-H, *J* = 8.7 Hz), 6.48 (br s, 1H, N-H), 3.89 (s, 3H, OMe), 1.68 (s, 9H, sp³ C-H). ¹³C NMR (125 MHz, CDCl₃): δ 160.3, 153.2, 142.2, 141.9, 140.9, 129.6, 127.9, 126.1, 125.3, 123.1, 122.7, 115.9, 114.1, 94.9, 55.4, 51.9, 29.1. HRMS (ESI) calcd for C₂₁H₂₃N₄O [M+H]⁺ 347.1867, found 347.1865.

5ac: *2-(4-cyanophenyl)-N-(tert-butyl)pyrazolo[1,5-*c*]quinazolin-5-amine*

White solid, Yield: 0.093 g (80%), m.p.: 214-216 °C. ¹H NMR (500 MHz, CDCl₃): δ 8.12 (d, 2H, aromatic C-H, *J* = 8.4 Hz), 7.90 (dd, 1H, aromatic C-H, *J* = 7.8, 1.1 Hz), 7.76 (d, 2H, aromatic C-H, *J* = 8.3 Hz), 7.68 (d, 1H, *J* = 8.0 Hz), 7.54 (td, 1H, aromatic C-H, *J* = 7.5, 1.5 Hz), 7.30 (td, 1H, aromatic C-H, *J* = 7.9, 0.9 Hz), 7.21 (s, 1H, aromatic C-H), 6.44 (br s, 1H, N-H), 1.69 (s, 9H, sp³ C-H). ¹³C NMR (125 MHz, CDCl₃): δ 151.2, 141.9, 141.8, 141.4, 137.0, 132.6, 130.0, 127.0, 126.3, 123.1, 123.0, 118.8, 115.7, 112.2, 95.9, 52.1, 29.1.

5ad: *N-(tert-butyl)-2-(4-nitrophenyl)pyrazolo[1,5-*c*]quinazolin-5-amine*

Yellow solid, Yield: 0.120 g (97%), m.p.: 209-211 °C. ¹H NMR (500 MHz, CDCl₃): δ 8.35 (d, 2H, aromatic C-H, *J* = 8.3 Hz), 8.19 (d, 2H, aromatic C-H, *J* = 8.1 Hz), 7.92 (d, 1H, aromatic C-H, *J* = 7.8 Hz), 7.69 (d, 1H, aromatic C-H, *J* = 8.2 Hz), 7.55 (t, 1H, aromatic C-H, *J* = 7.2 Hz), 7.31 (t, 1H, aromatic C-H, *J* = 7.4 Hz), 7.27 (s, 1H, aromatic C-H), 6.46 (br s, 1H, N-H), 1.69 (s, 9H, sp³ C-H). ¹³C NMR (125 MHz, CDCl₃): δ 150.8, 147.9, 141.8,

141.5, 138.9, 130.1, 127.2, 126.3, 124.2, 123.12, 123.13, 115.7, 96.2, 52.2, 29.1. HRMS (ESI) calcd for C₂₀H₂₀N₅O₂ [M+H]⁺ 362.1612, found 362.1600.

5ae: 2-(4-bromophenyl)-N-(tert-butyl)pyrazolo[1,5-c]quinazolin-5-amine

Yellow solid, Yield: 0.102 g (76%), m.p.: 146-148 °C. ¹H NMR (500 MHz, CDCl₃): δ 7.90-7.88 (m, 3H, aromatic C-H), 7.68 (d, 1H, aromatic C-H, J = 8.2 Hz), 7.61 (d, 2H, aromatic C-H, J = 8.4 Hz), 7.53 (td, 1H, aromatic C-H, J = 8.3, 1.3 Hz), 7.28 (td, 1H, aromatic C-H, J = 7.9, 0.8 Hz), 7.14 (s, 1H, aromatic C-H), 6.46 (br s, 1H, N-H), 1.69 (s, 9H, sp³ C-H). ¹³C NMR (125 MHz, CDCl₃): δ 152.2, 142.0, 141.9, 141.1, 131.9, 131.6, 129.8, 128.1, 126.2, 123.1, 122.9, 122.8, 115.8, 95.4, 52.0, 29.1. HRMS (ESI) calcd for C₂₀H₂₀BrN₄ [M+H]⁺ 395.0866, found 395.0860.

5ba: 2-phenyl-N-(2,4,4-trimethylpentan-2-yl)pyrazolo[1,5-c]quinazolin-5-amine

White solid, Yield: 0.115 g (91%), m.p.: 82-83 °C. ¹H NMR (500 MHz, CDCl₃): δ 8.03 (d, 2H, aromatic C-H, J = 7.1 Hz), 7.92 (dd, 1H, aromatic C-H, J = 7.8, 1.2 Hz), 7.67 (d, 1H, aromatic C-H, J = 8.15 Hz), 7.54-7.49 (m, 3H, aromatic C-H), 7.44-7.41 (m, 1H, aromatic C-H), 7.28 (td, 1H, aromatic C-H, J = 8.0, 1.0 Hz), 7.19 (s, 1H, aromatic C-H), 6.67 (br s, 1H, N-H), 2.09 (s, 2H, sp³ C-H), 1.76 (s, 6H, sp³ C-H), 1.11 (s, 9H, sp³ C-H). ¹³C NMR (125 MHz, CDCl₃): δ 153.3, 142.1, 141.9, 140.8, 132.7, 129.6, 128.9, 128.8, 126.6, 126.2, 123.1, 122.6, 115.9, 95.4, 55.8, 52.4, 31.9, 31.7, 29.3. HRMS (ESI) calcd for C₂₄H₂₉N₄ [M+H]⁺ 373.2387, found 373.2366.

5bb: 1-(4-((2,4,4-trimethylpentan-2-yl)amino)pyrazolo[1,5-c]quinazolin-2-yl)phenyl)ethan-1-one

White solid, Yield: 0.131 g (93%), m.p.: 155-157 °C. ¹H NMR (500 MHz, CDCl₃): δ 8.09 (d, 4H, aromatic C-H, J = 8.2 Hz), 7.91 (d, 1H, aromatic C-H, J = 7.1 Hz), 7.67 (d, 1H, aromatic C-H, J = 7.9 Hz), 7.53 (t, 1H, aromatic C-H, J = 7.2 Hz), 7.29-7.25 (m, 2H, aromatic C-H), 6.64 (br s, 1H, N-H), 2.67 (s, 3H, sp³ C-H), 2.08 (s, 2H, sp³ C-H), 1.75 (s, 6H, sp³ C-H), 1.10 (s, 9H, sp³ C-H). ¹³C NMR (125 MHz, CDCl₃): δ 197.6, 151.9, 141.9, 141.8, 141.1, 137.2, 137.1, 129.8, 128.9, 126.6, 126.2, 123.1, 122.8, 115.8, 95.9, 55.8, 52.5, 31.9, 31.6, 29.3, 26.7. HRMS (ESI) calcd for C₂₆H₃₁N₄O [M+H]⁺ 415.2493, found 415.2490.

5bc: 2-(3-nitrophenyl)-N-(2,4,4-trimethylpentan-2-yl)pyrazolo[1,5-c]quinazolin-5-amine

Pale yellow solid, Yield: 0.122 g (86%), m.p.: 152-154 °C. ¹H NMR (500 MHz, CDCl₃): δ 8.86 (t, 1H, aromatic C-H, J = 1.8 Hz), 8.33 (d, 1H, aromatic C-H, J = 7.7 Hz), 8.26 (dd, 1H, aromatic C-H, J = 8.1, 1.3 Hz), 7.92 (dd, 1H, aromatic C-H, J = 7.8, 1.0 Hz), 7.69-7.65 (m, 2H, aromatic C-H), 7.55 (td, 1H, aromatic C-H, J = 8.3, 1.3 Hz), 7.31 (td, 1H, aromatic C-H, J = 0.9, 7.95 Hz), 7.26 (s, 1H, aromatic C-H), 6.62 (br s, 1H, N-H), 2.09 (s, 2H, sp³ C-H), 1.76 (s, 6H, sp³ C-H), 1.10 (s, 9H, sp³ C-H). ¹³C NMR (125 MHz, CDCl₃): δ 150.8, 148.8, 141.9, 141.8, 141.4, 134.6, 132.3, 130.0, 129.7, 126.3, 123.3, 123.1, 122.9, 121.4, 115.7, 95.7, 55.9, 52.3, 31.9, 31.6, 29.4. HRMS (ESI) calcd for C₂₄H₂₈N₅O₂ [M+H]⁺ 418.2238, found 418.2226. Despite repeated drying on high vacuum, a significant solvent peak is observed in ¹H and ¹³C NMR spectra. The isolated yield has been modified to 86 %.

5bd: 2-(2-bromophenyl)-N-(2,4,4-trimethylpentan-2-yl)pyrazolo[1,5-c]quinazolin-5-amine

Yellow solid, Yield: 0.118 g (77%), m.p.: 95-96.5 °C. ¹H NMR (500 MHz, CDCl₃): δ 7.92 (dd, 1H, aromatic C-H, J = 7.7, 0.9 Hz), 7.82 (dd, 1H, aromatic C-H, J = 7.6, 1.5 Hz), 7.73 (dd, 1H, aromatic C-H, J = 8.0, 0.7 Hz), 7.67 (d, 1H, aromatic C-H, J = 8.1 Hz), 7.52 (td, 1H, aromatic C-H, J = 8.3, 1.3 Hz), 7.43 (td, 1H, aromatic C-H, J = 7.5, 0.9 Hz), 7.34 (s, 1H, aromatic C-H), 7.28 (t, 2H, aromatic C-H, J = 7.7 Hz), 6.59 (br s, 1H, N-H), 2.07 (s, 2H, sp³ C-H), 1.73 (s, 6H, sp³ C-H), 1.08 (s, 9H, sp³ C-H). ¹³C NMR (125 MHz, CDCl₃): δ 152.5, 141.89, 141.87, 139.9, 134.0, 133.8, 131.8, 129.9, 129.6, 127.4, 126.1, 123.2, 122.7, 122.5, 115.9, 99.4, 52.8, 52.1, 31.8, 31.6, 29.4. HRMS (ESI) calcd for C₂₄H₂₈BrN₄ [M+H]⁺ 451.1492, found 451.1489.

5be: 2-(2-aminophenyl)-N-(2,4,4-trimethylpentan-2-yl)pyrazolo[1,5-c]quinazolin-5-amine

Light brown solid, Yield: 0.112 g (85%), m.p.: 114-116 °C. ¹H NMR (500 MHz, CDCl₃): δ 7.93 (dd, 1H, aromatic C-H, J = 7.8, 1.0 Hz), 7.72 (dd, 1H, aromatic C-H, J = 7.7, 1.3 Hz), 7.68 (d, 1H, aromatic C-H, J = 8.1 Hz), 7.54 (td, 1H, aromatic C-H, J = 7.2, 1.3 Hz), 7.29 (td, 1H, aromatic C-H, J = 8.0, 1.0 Hz), 7.22 (td, 1H, aromatic C-H, J = 8.6, 1.4 Hz), 7.19 (s, 1H, aromatic C-H), 6.88-6.84 (m, 2H, aromatic C-H), 6.41 (br s, 1H, N-H), 5.38 (br s, 2H, N-H), 2.03 (s, 2H, sp³ C-H), 1.76 (s, 6H, sp³ C-H), 1.11 (s, 9H, sp³ C-H). ¹³C NMR (125 MHz, CDCl₃): δ 154.0, 145.2, 141.9, 141.8, 140.1, 129.8, 129.7, 129.6, 126.1, 123.3, 122.8, 117.8, 116.7, 115.8, 115.7, 96.6, 55.9, 53.1, 31.9, 31.7, 29.1. HRMS (ESI) calcd for C₂₄H₃₀N₅ [M+H]⁺ 388.2496, found 388.2494.

5ca: N-(tert-butyl)-2-butylpyrazolo[1,5-c]quinazolin-5-amine

Pale yellow oil, Yield: 0.076 g (76%). ¹H NMR (500 MHz, CDCl₃): δ 7.83 (dd, 1H, aromatic C-H, J = 7.8, 1.0 Hz), 7.65 (dd, 1H, aromatic C-H, J = 8.2, 0.5 Hz), 7.49 (td, 1H, aromatic C-H, J = 8.4, 1.4 Hz), 7.24 (td, 1H, aromatic C-H, J = 8.0, 1.1 Hz), 6.69 (s, 1H, aromatic C-H), 6.35 (br s, 1H, N-H), 2.84 (t, 2H, sp³ C-H, J = 7.7 Hz), 1.79 (quintet, 2H, sp³ C-H, J = 7.5 Hz), 1.67 (s, 9H, sp³ C-H), 1.50-1.44 (sextet, 2H, sp³ C-H, J = 7.4 Hz), 1.00 (t, 3H, sp³ C-H,

J = 7.4 Hz). ^{13}C NMR (125 MHz, CDCl_3): δ 156.4, 142.2, 141.9, 140.3, 129.3, 126.0, 123.0, 122.5, 116.0, 97.1, 51.9, 31.7, 29.1, 28.4, 22.6, 13.9. HRMS (ESI) calcd for $\text{C}_{18}\text{H}_{25}\text{N}_4$ [$\text{M}+\text{H}]^+$ 297.2074, found 297.2060.

5cb: *N*-(*tert*-butyl)-2-pentylpyrazolo[1,5-*c*]quinazolin-5-amine

Yellow oil, Yield: 0.077 g (73%). ^1H NMR (500 MHz, CDCl_3): δ 7.83 (dd, 1H, aromatic C-H, *J* = 7.8, 1.1 Hz), 7.65 (d, 1H, aromatic C-H, *J* = 8.1 Hz) 7.49 (td, 1H, aromatic C-H, *J* = 8.3, 1.3 Hz), 7.24 (td, 1H, aromatic C-H, *J* = 7.9, 1.0 Hz), 6.69 (s, 1H, aromatic C-H), 6.35 (br s, 1H, N-H), 2.83 (t, 2H, sp^3 C-H, *J* = 7.7 Hz), 1.81-1.77 (m, 2H, sp^3 C-H), 1.67 (s, 9H, sp^3 C-H), 1.45-1.39 (m, 4H, sp^3 C-H), 0.95 (t, 3H, sp^3 C-H, *J* = 7.0 Hz). ^{13}C NMR (125 MHz, CDCl_3): δ 156.5, 142.2, 141.9, 140.3, 129.3, 126.0, 123.0, 122.5, 115.9, 97.1, 51.9, 31.7, 29.3, 29.1, 28.6, 22.5, 14.1. HRMS (ESI) calcd for $\text{C}_{19}\text{H}_{27}\text{N}_4$ [$\text{M}+\text{H}]^+$ 311.2230, found 311.2217.

5cc: *N*-2-di-*tert*-butylpyrazolo[1,5-*c*]quinazolin-5-amine

White solid, Yield: 0.071 g (71%), m.p.: 99-101 °C. ^1H NMR (500 MHz, CDCl_3): δ 7.83 (dd, 1H, aromatic C-H, *J* = 7.8, 1.1 Hz), 7.62 (d, 1H, aromatic C-H, *J* = 8.1 Hz), 7.47 (td, 1H, aromatic C-H, *J* = 8.4, 1.4 Hz), 7.23 (td, 1H, aromatic C-H, *J* = 8.0, 1.0 Hz), 6.74 (s, 1H, aromatic C-H), 6.38 (br s, 1H, N-H), 1.66 (s, 9H, sp^3 C-H), 1.44 (s, 9H, sp^3 C-H). ^{13}C NMR (125 MHz, CDCl_3): δ 164.7, 142.3, 141.8, 139.9, 129.2, 125.9, 122.9, 122.4, 116.1, 94.9, 51.7, 32.6, 30.6, 29.1. HRMS (ESI) calcd for $\text{C}_{18}\text{H}_{25}\text{N}_4$ [$\text{M}+\text{H}]^+$ 297.2074, found 297.2055.

5cd: *N*-(*tert*-butyl)-2-(trimethylsilyl)pyrazolo[1,5-*c*]quinazolin-5-amine

Colorless oil, Yield: 0.049 g (46%). ^1H NMR (500 MHz, CDCl_3): δ 7.91 (d, 1H, aromatic C-H, *J* = 7.7 Hz), 7.69 (d, 1H, aromatic C-H, *J* = 8.1 Hz), 7.52 (t, 1H, aromatic C-H, *J* = 7.3 Hz), 7.29 (d, 1H, aromatic C-H, *J* = 7.8 Hz), 7.02 (s, 1H, aromatic C-H), 6.57 (br s, 1H, N-H), 1.70 (s, 9H, sp^3 C-H), 0.45 (s, 9H, sp^3 C-H). ^{13}C NMR (125 MHz, CDCl_3): δ 156.1, 142.4, 141.8, 139.7, 129.2, 125.9, 123.1, 122.6, 116.4, 104.4, 51.8, 29.1, -1.1. HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{25}\text{N}_4\text{Si}$ [$\text{M}+\text{H}]^+$ 313.1843, found 313.1836.

5cd': *N*-(*tert*-butyl)pyrazolo[1,5-*c*]quinazolin-5-amine

Transparent oil, Yield: 0.032 g (39%). ^1H NMR (500 MHz, CDCl_3): δ 7.91 (d, 1H, aromatic C-H, *J* = 2.0 Hz), 7.88 (dd, 1H, aromatic C-H, *J* = 7.8, 1.2 Hz), 7.66 (d, 1H, aromatic C-H, *J* = 8.2 Hz), 7.51 (td, 1H, aromatic C-H, *J* = 7.1, 1.4 Hz), 7.29-7.26 (m, 1H, aromatic C-H), 6.89 (d, 1H, aromatic C-H, *J* = 2.0 Hz), 6.40 (br s, 1H, N-H), 1.66 (s, 9H, sp^3 C-H). ^{13}C NMR (125 MHz, CDCl_3): δ 142.1, 141.7, 141.5, 139.8, 129.6, 126.1, 123.1, 122.8, 116.1, 98.3, 51.9, 29.1. HRMS (ESI) calcd for $\text{C}_{14}\text{H}_{17}\text{N}_4$ [$\text{M}+\text{H}]^+$ 241.1448, found 241.1435.

5ce: *N*-(*tert*-butyl)-2-(thiophen-2-yl)pyrazolo[1,5-*c*]quinazolin-5-amine

White solid, Yield: 0.079 g (72%), m.p.: 132-135 °C. ^1H NMR (500 MHz, CDCl_3): δ 7.88 (dd, 1H, aromatic C-H, *J* = 7.8, 1.3 Hz), 7.66 (d, 1H, aromatic C-H, *J* = 8.2 Hz), 7.59 (dd, 1H, aromatic C-H, *J* = 3.5, 0.8 Hz), 7.52 (td, 1H, aromatic C-H, *J* = 8.3, 1.2 Hz), 7.38 (dd, 1H, aromatic C-H, *J* = 5.0, 0.8 Hz), 7.29-7.26 (m, 1H, aromatic C-H), 7.15 (dd, 1H, aromatic C-H, *J* = 4.9, 3.6 Hz), 7.08 (s, 1H, aromatic C-H), 6.41 (br s, 1H, N-H), 1.68 (s, 9H, sp^3 C-H). ^{13}C NMR (125 MHz, CDCl_3): δ 148.6, 142.0, 141.0, 135.7, 129.8, 127.7, 126.16, 126.15, 125.8, 123.2, 122.8, 115.8, 95.5, 52.0, 29.1. HRMS (ESI) calcd for $\text{C}_{18}\text{H}_{19}\text{N}_4\text{S}$ [$\text{M}+\text{H}]^+$ 323.1325, found 323.1321.

5cf: *N*-(*tert*-butyl)-2-(pyridin-2-yl)pyrazolo[1,5-*c*]quinazolin-5-amine

Pale yellow solid, Yield: 0.092 g (85%), m.p.: 81-83 °C. ^1H NMR (500 MHz, CDCl_3): δ 8.74 (d, 1H, aromatic C-H, *J* = 4.3 Hz), 8.19 (d, 1H, aromatic C-H, *J* = 7.8 Hz), 7.93 (d, 1H, aromatic C-H, *J* = 7.8 Hz), 7.81 (t, 1H, aromatic, C-H, *J* = 7.7 Hz), 7.67 (d, 1H, aromatic C-H, *J* = 8.2 Hz), 7.55 (s, 1H, aromatic C-H), 7.52 (t, 1H, aromatic C-H, *J* = 8.0 Hz), 7.30 (q, 2H, aromatic C-H, *J* = 6.9 Hz), 6.49 (br s, 1H, N-H), 1.69 (s, 9H, sp^3 C-H). ^{13}C NMR (125 MHz, CDCl_3): δ 153.4, 151.6, 149.7, 142.1, 141.8, 141.1, 136.7, 129.7, 126.1, 123.4, 123.2, 122.9, 121.1, 116.1, 97.1, 52.1, 29.1. HRMS (ESI) calcd for $\text{C}_{19}\text{H}_{20}\text{N}_5$ [$\text{M}+\text{H}]^+$ 318.1713, found 318.1701.

5cg: ethyl 5-(*tert*-butylamino)pyrazolo[1,5-*c*]quinazoline-2-carboxylate

White solid, Yield: 0.051 g (48%), m.p.: 73.3-74.9 °C. ^1H NMR (500 MHz, CDCl_3): δ 7.89 (dd, 1H, aromatic C-H, *J* = 7.8, 1.1 Hz), 7.67 (d, 1H, aromatic C-H, *J* = 8.1 Hz), 7.55 (td, 1H, aromatic C-H, *J* = 8.4, 1.4 Hz), 7.39 (s, 1H, aromatic C-H), 7.30 (td, 1H, aromatic C-H, *J* = 8.0, 1.0 Hz), 6.48 (br s, 1H, N-H), 4.50 (q, 2H, sp^3 C-H, *J* = 7.1 Hz), 1.66 (s, 9H, sp^3 C-H), 1.48 (t, 3H, sp^3 C-H, *J* = 7.1 Hz). ^{13}C NMR (125 MHz, CDCl_3): δ 162.4, 145.0, 141.7, 141.6, 140.9, 130.2, 126.3, 123.3, 123.1, 115.8, 100.8, 61.6, 52.4, 28.9, 14.3. HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{21}\text{N}_4\text{O}_2$ [$\text{M}+\text{H}]^+$ 313.1659, found 313.1640.

5cg': ethyl 5-(*tert*-butylamino)pyrazolo[1,5-*c*]quinazoline-1-carboxylate

Transparent oil, Yield: 0.038g (36%). ^1H NMR (500 MHz, CDCl_3): δ 9.44 (dd, 1H, aromatic C-H, *J* = 8.2, 1.2 Hz), 8.37 (s, 1H, aromatic C-H), 7.69 (d, 1H, aromatic C-H, *J* = 7.4 Hz), 7.62 (td, 1H, aromatic C-H, *J* = 7.0, 1.3Hz), 7.35 (td, 1H, aromatic C-H, *J* = 8.2, 1.2 Hz), 6.51 (br s, 1H, N-H), 4.43 (q, 2H, sp^3 C-H, *J* = 7.1 Hz), 1.65 (s, 9H,

$\text{sp}^3 \text{C-H}$, 1.46 (t, 3H, $\text{sp}^3 \text{C-H}$, $J = 7.1$ Hz). ^{13}C NMR (125 MHz, CDCl_3): δ 163.3, 144.8, 143.5, 141.9, 140.7, 131.3, 127.3, 125.9, 123.1, 115.5, 108.9, 60.6, 52.2, 28.9, 14.4. HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{21}\text{N}_4\text{O}_2$ [$\text{M}+\text{H}]^+$ 313.1659, found 313.1644.

5ch: (5-(tert-butylamino)pyrazolo[1,5-c]quinazolin-2-yl)methyl acetate

Pale yellow solid, Yield: 0.093 g (88%), m.p.: 93.5-95 °C. ^1H NMR (500 MHz, CDCl_3): δ 7.84 (dd, 1H, aromatic C-H, $J = 7.8$, 1.1 Hz), 7.65 (d, 1H, aromatic C-H, $J = 8.1$ Hz), 7.51 (td, 1H, aromatic C-H, $J = 7.3$, 1.4 Hz), 7.27 (td, 1H, aromatic C-H, $J = 6.9$, 1.0 Hz), 6.90 (s, 1H, aromatic C-H), 6.34 (br s, 1H, N-H), 5.32 (s, 2H, $\text{sp}^3 \text{C-H}$), 2.16 (s, 3H, $\text{sp}^3 \text{C-H}$), 1.65 (s, 9H, $\text{sp}^3 \text{C-H}$). ^{13}C NMR (125 MHz, CDCl_3): δ 170.7, 150.1, 141.9, 141.8, 140.8, 129.8, 126.1, 123.1, 122.9, 115.8, 98.1, 60.1, 52.0, 29.0, 21.0. HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{21}\text{N}_4\text{O}_2$ [$\text{M}+\text{H}]^+$ 313.1659, found 313.1644.

5da: (5-((2,4,4-trimethylpentan-2-yl)amino)pyrazolo[1,5-c]quinazolin-2-yl)methanol

Pale yellow solid, Yield: 0.084 g (76%), m.p.: 116-117 °C. ^1H NMR (500 MHz, CDCl_3): δ 7.82 (dd, 1H, aromatic C-H, $J = 7.8$, 0.9 Hz), 7.66 (d, 1H, aromatic C-H, $J = 8.1$ Hz), 7.51 (td, 1H, aromatic C-H, $J = 8.3$, 1.3 Hz), 7.27-7.24 (m, 1H, aromatic C-H), 6.85 (s, 1H, aromatic C-H), 6.42 (br s, 1H, N-H), 4.91 (s, 2H, $\text{sp}^3 \text{C-H}$), 2.08 (s, 2H, $\text{sp}^3 \text{C-H}$), 1.71 (s, 6H, $\text{sp}^3 \text{C-H}$), 1.04 (s, 9H, $\text{sp}^3 \text{C-H}$). ^{13}C NMR (125 MHz, CDCl_3): δ 154.7, 141.8, 141.7, 140.7, 129.7, 126.1, 123.1, 122.7, 115.8, 96.4, 59.4, 55.8, 51.7, 31.7, 31.5, 29.5. HRMS (ESI) calcd for $\text{C}_{19}\text{H}_{27}\text{N}_4\text{O}$ [$\text{M}+\text{H}]^+$ 327.2180, found 327.2178. Despite repeated drying on high vacuum, a significant solvent peak is observed in ^1H and ^{13}C NMR spectra. The isolated yield has been modified to 76 %.

5db: N-((5-((2,4,4-trimethylpentan-2-yl)amino)pyrazolo[1,5-c]quinazolin-2-yl)methyl)benzamide

Pale yellow solid, Yield: 0.140 g (96%), m.p.: 110-112 °C. ^1H NMR (500 MHz, CDCl_3): δ 7.86-7.82 (m, 3H, aromatic C-H), 7.64 (d, 1H, aromatic C-H, $J = 8.0$ Hz), 7.55-7.45 (m, 4H, aromatic C-H), 7.27-7.24 (m, 1H, aromatic C-H), 6.87 (s, 1H, aromatic C-H), 6.77 (br s, 1H, N-H), 6.42 (br s, 1H, N-H), 4.88 (d, 1H, $\text{sp}^3 \text{C-H}$, $J = 5.3$ Hz), 2.04 (s, 2H, $\text{sp}^3 \text{C-H}$), 1.71 (s, 6H, $\text{sp}^3 \text{C-H}$), 1.06 (s, 9H, $\text{sp}^3 \text{C-H}$). ^{13}C NMR (125 MHz, CDCl_3): δ 167.4, 151.8, 141.8, 141.7, 140.9, 134.3, 131.7, 129.7, 128.6, 127.0, 126.1, 123.2, 122.8, 115.7, 97.1, 55.8, 52.3, 38.3, 31.8, 31.6, 29.3. HRMS (ESI) calcd for $\text{C}_{26}\text{H}_{32}\text{N}_5\text{O}$ [$\text{M}+\text{H}]^+$ 430.2602, found 430.2601.

5dc: 2-(naphthalen-1-yl)-N-(2,4,4-trimethylpentan-2-yl)pyrazolo[1,5-c]quinazolin-5-amine

Off-white solid, Yield: 0.113 g (93%), m.p.: 127-129.5 °C. ^1H NMR (500 MHz, CDCl_3): δ 8.62-8.61 (m, 1H, aromatic C-H), 7.97-7.95 (m, 3H, aromatic C-H), 7.87 (d, 1H, aromatic C-H, $J = 6.8$ Hz), 7.73 (d, 1H, aromatic C-H, $J = 8.1$ Hz), 7.61 (t, 1H, aromatic C-H, $J = 7.6$ Hz), 7.58-7.55 (m, 3H, aromatic C-H), 7.31 (t, 1H, aromatic C-H, $J = 7.4$ Hz), 7.19 (s, 1H, aromatic C-H), 6.71 (br s, 1H, N-H), 2.09 (s, 2H, $\text{sp}^3 \text{C-H}$), 1.78 (s, 6H, $\text{sp}^3 \text{C-H}$), 1.13 (s, 9H, $\text{sp}^3 \text{C-H}$). ^{13}C NMR (125 MHz, CDCl_3): δ 153.4, 142.1, 142.0, 140.3, 134.0, 131.5, 130.9, 129.7, 129.2, 128.4, 128.1, 126.6, 126.2, 126.1, 126.0, 125.4, 123.2, 122.7, 115.9, 99.5, 55.8, 52.5, 31.9, 31.6, 29.4. HRMS (ESI) calcd for $\text{C}_{28}\text{H}_{31}\text{N}_4$ [$\text{M}+\text{H}]^+$ 423.2543, found 423.2537.

5dd: 2-cyclopropyl-N-(2,4,4-trimethylpentan-2-yl)pyrazolo[1,5-c]quinazolin-5-amine

Yellow oil, Yield: 0.098 g (86%). ^1H NMR (500 MHz, CDCl_3): δ 7.79 (dd, 1H, aromatic C-H, $J = 7.8$, 1.2 Hz), 7.63 (d, 1H, aromatic C-H, $J = 8.2$ Hz), 7.48 (td, 1H, aromatic C-H, $J = 7.2$, 1.4 Hz), 7.23 (td, 1H, aromatic C-H, $J = 8.0$, 1.1Hz), 6.52 (s, 1H, aromatic C-H), 6.45 (br s, 1H, N-H), 2.14 (tt, 1H, $\text{sp}^3 \text{C-H}$, $J = 8.4$, 5.0 Hz), 2.07 (s, 2H, $\text{sp}^3 \text{C-H}$), 1.71 (s, 6H, $\text{sp}^3 \text{C-H}$), 1.09-1.08 (m, 2H, $\text{sp}^3 \text{C-H}$), 1.07 (s, 9H, $\text{sp}^3 \text{C-H}$) 0.94 (dt, 2H, $\text{sp}^3 \text{C-H}$, $J = 7.0$, 4.2 Hz). ^{13}C NMR (125 MHz, CDCl_3): δ 158.2, 141.95, 141.92, 140.3, 129.3, 126.0, 122.9, 122.3, 115.8, 94.6, 55.6, 52.1, 31.8, 31.6, 29.4, 9.7, 9.1. HRMS (ESI) calcd for $\text{C}_{21}\text{H}_{29}\text{N}_4$ [$\text{M}+\text{H}]^+$ 337.2387, found 337.2366.

5ea: 8-bromo-N-(tert-butyl)-2-phenylpyrazolo[1,5-c]quinazolin-5-amine

Pale yellow solid, Yield: 0.057 g (65%), m.p.: 168-170 °C. ^1H NMR (500 MHz, CDCl_3): δ 8.01 (d, 2H, aromatic C-H, $J = 7.0$ Hz), 7.84 (d, 1H, aromatic C-H, $J = 1.9$ Hz), 7.74 (d, 1H, aromatic C-H, $J = 8.4$ Hz), 7.50 (t, 2H, aromatic C-H, $J = 7.2$ Hz), 7.43 (t, 1H, aromatic C-H, $J = 6.2$ Hz), 7.37(dd,1H, aromatic C-H, $J = 8.3$, 1.9 Hz), 7.15 (s, 1H, aromatic C-H), 6.56 (br s, 1H, N-H), 1.68 (s, 9H, $\text{sp}^3 \text{C-H}$). ^{13}C NMR (125 MHz, CDCl_3): δ 153.7, 143.0, 142.6, 140.4, 132.4, 129.0, 128.8, 128.7, 126.6, 125.8, 124.3, 123.3, 114.7, 95.7, 52.2, 29.1. HRMS (ESI) calcd for $\text{C}_{20}\text{H}_{20}\text{BrN}_4$ [$\text{M}+\text{H}]^+$ 395.0866, found 395.0862.

5eb: N-(tert-butyl)-2-phenyl-8-(trifluoromethyl)pyrazolo[1,5-c]quinazolin-5-amine

White solid, Yield: 0.070 g (78%), m.p.: 54-56 °C. ^1H NMR (500 MHz, CDCl_3): δ 8.02 (d, 2H, aromatic C-H, $J = 7.2$ Hz), 7.99 (d, 1H, aromatic C-H, $J = 8.1$ Hz), 7.94 (s, 1H, aromatic C-H), 7.52-7.43 (m, 4H, aromatic C-H), 7.25 (s, 1H, aromatic C-H), 6.60 (br s, 1H, N-H), 1.69 (s, 9H, $\text{sp}^3 \text{C-H}$). ^{13}C NMR (125 MHz, CDCl_3): δ 153.8, 142.7, 141.7, 140.0, 132.2, 131.4 ($J_{\text{C-F}} = 18$ Hz), 129.1, 128.9, 126.7, 123.8, 123.4 (q, $J_{\text{C-F}} = 18$ Hz), 118.7 (q, $J_{\text{C-F}} = 3.7$ Hz), 118.2, 96.6, 52.3, 29.0. ^{19}F NMR (470 MHz, CDCl_3): δ -62.44. HRMS (ESI) calcd for $\text{C}_{21}\text{H}_{20}\text{F}_3\text{N}_4$

$[M+H]^+$ 385.1635, found 385.1630. Despite repeated drying on high vacuum, a significant solvent peak is observed in ^1H and ^{13}C NMR spectra. The isolated yield has been modified to 78 %.

5ec: *N*-(tert-butyl)-8-fluoro-2-phenylpyrazolo[1,5-*c*]quinazolin-5-amine

Off white solid, Yield: 0.068 g (67%), m.p.: 107-109 °C. ^1H NMR (500 MHz, CDCl_3): δ 8.01 (d, 2H, aromatic C-H, $J = 7.1$ Hz), 7.87 (dd, 1H, aromatic C-H, $J = 8.7, 6.1$ Hz), 7.49 (t, 2H, aromatic C-H, $J = 7.2$ Hz), 7.44-7.41 (m, 1H, aromatic C-H), 7.32 (dd, 1H, aromatic C-H, $J = 10.6, 2.5$ Hz), 7.12 (s, 1H, aromatic C-H), 7.02 (td, 1H, aromatic C-H, $J = 8.5, 2.5$ Hz), 6.56 (br s, 1H, N-H), 1.68 (s, 9H, sp^3 C-H). ^{13}C NMR (125 MHz, CDCl_3): δ 163.7 (d, $J_{\text{C}-\text{F}} = 245$ Hz), 153.6, 142.7, 140.6, 132.5, 128.9, 128.8, 126.3, 124.7 (d, $J_{\text{C}-\text{F}} = 10.0$ Hz), 112.6, 111.5, 111.3, 111.1, 95.2, 52.1, 29.1. ^{19}F NMR (470 MHz, CDCl_3): δ -110.9. HRMS (ESI) calcd for $\text{C}_{20}\text{H}_{20}\text{FN}_4$ $[M+H]^+$ 335.1667, found 335.1665.

5ed: *N*-(tert-butyl)-8-chloro-2-phenylpyrazolo[1,5-*c*]quinazolin-5-amine

White solid, Yield: 0.071 g (74%), m.p.: 134-136 °C. ^1H NMR (500 MHz, CDCl_3): δ 8.01 (d, 2H, aromatic C-H, $J = 7.2$ Hz), 7.79 (d, 1H, aromatic C-H, $J = 8.3$ Hz), 7.67 (d, 1H, aromatic C-H, $J = 1.9$ Hz), 7.50 (t, 2H, aromatic C-H, $J = 7.2$ Hz), 7.43 (t, 1H, aromatic C-H, $J = 7.2$ Hz), 7.23 (dd, 1H, aromatic C-H, $J = 8.3, 2.0$ Hz), 7.14 (s, 1H, aromatic C-H), 6.56 (br s, 1H, N-H), 1.68 (s, 9H, sp^3 C-H). ^{13}C NMR (125 MHz, CDCl_3): δ 153.7, 142.9, 142.7, 140.4, 135.1, 132.4, 129.0, 128.8, 126.6, 125.6, 124.2, 123.1, 114.4, 95.7, 52.2, 29.1. HRMS (ESI) calcd for $\text{C}_{20}\text{H}_{20}\text{ClN}_4$ $[M+H]^+$ 351.1371, found 351.1369.

5fa: dimethyl-5-(tert-butylamino)pyrazolo[1,5-*c*]quinazoline-1,2-dicarboxylate

White solid, Yield: 0.089 g (74%), m.p.: 159-161 °C. ^1H NMR (500 MHz, CDCl_3): δ 8.76 (dd, 1H, aromatic C-H, $J = 8.1, 0.9$ Hz), 7.69 (d, 1H, aromatic C-H, $J = 8.1$ Hz), 7.62 (td, 1H, aromatic C-H, $J = 8.3, 1.2$ Hz), 7.33 (td, 1H, aromatic C-H, $J = 8.1, 1.0$ Hz), 6.45 (br s, 1H, N-H), 4.03 (s, 3H, sp^3 C-H), 3.99 (s, 3H, sp^3 C-H), 1.64 (s, 9H, sp^3 C-H). ^{13}C NMR (125 MHz, CDCl_3): δ 163.7, 163.2, 145.8, 143.1, 141.3, 140.1, 131.5, 126.3, 125.5, 123.4, 114.9, 108.0, 52.9, 52.49, 52.48, 28.9.

5fb: dimethyl 5-(tert-butylamino)-8-chloropyrazolo[1,5-*c*]quinazoline-1,2-dicarboxylate

White solid, Yield: 0.087 g (81%), m.p.: 181-182 °C. ^1H NMR (500 MHz, CDCl_3): δ 8.82 (d, 1H, aromatic C-H, $J = 8.7$ Hz), 7.69 (d, 1H, aromatic C-H, $J = 2.1$ Hz), 7.28 (dd, 1H, aromatic C-H, $J = 8.8, 2.1$ Hz), 6.52 (br s, 1H, N-H), 4.03 (s, 3H, sp^3 C-H), 3.97 (s, 3H, sp^3 C-H), 1.63 (s, 9H, sp^3 C-H). ^{13}C NMR (125 MHz, CDCl_3): δ 163.4, 163.2, 146.4, 144.2, 141.8, 139.8, 137.3, 127.1, 125.7, 123.9, 113.4, 107.9, 53.0, 52.7, 52.5, 28.8. HRMS (ESI) calcd for $\text{C}_{18}\text{H}_{20}\text{ClN}_4\text{O}_4$ $[M+H]^+$ 391.1168, found 391.1165.

5ga: diethyl-5-(tert-butylamino)pyrazolo[1,5-*c*]quinazoline-1,2-dicarboxylate

Off-white solid, Yield: 0.110 g (84%), m.p.: 63-65 °C. ^1H NMR (500 MHz, CDCl_3): δ 8.83 (d, 1H, aromatic C-H, $J = 8.1$ Hz), 7.69 (d, 1H, aromatic C-H, $J = 8.1$ Hz), 7.61 (t, 1H, aromatic C-H, $J = 8.1$ Hz), 7.33 (t, 1H, aromatic C-H, $J = 8.0$ Hz), 6.46 (br s, 1H, N-H), 4.49 (q, 2H, sp^3 C-H, $J = 7.1$ Hz), 4.45 (q, 2H, sp^3 C-H, $J = 7.1$ Hz), 1.64 (s, 9H, sp^3 C-H), 1.45 (t, 3H, sp^3 C-H, $J = 7.1$ Hz), 1.42 (t, 3H, sp^3 C-H, $J = 7.1$ Hz). ^{13}C NMR (125 MHz, CDCl_3): δ 163.3, 162.9, 146.3, 143.1, 141.4, 140.0, 131.4, 126.3, 125.7, 123.4, 115.0, 108.1, 62.2, 61.5, 52.5, 28.9, 14.2, 14.1.

5gb: diethyl-8-bromo-5-(tert-butylamino)pyrazolo[1,5-*c*]quinazoline-1,2-dicarboxylate

Off-white solid, Yield: 0.075 g (74%), m.p.: 125-127 °C. ^1H NMR (500 MHz, CDCl_3): δ 8.82 (d, 1H, aromatic C-H, $J = 8.8$ Hz), 7.87 (d, 1H, aromatic C-H, $J = 2.0$ Hz), 7.42 (dd, 1H, aromatic C-H, $J = 8.7, 2.0$ Hz), 6.53 (br s, 1H, N-H), 4.49 (q, 2H, sp^3 C-H, $J = 7.1$ Hz), 4.43 (q, 2H, sp^3 C-H, $J = 7.1$ Hz), 1.63 (s, 9H, sp^3 C-H), 1.45 (t, 3H, sp^3 C-H, $J = 7.1$ Hz), 1.41 (t, 3H, sp^3 C-H, $J = 7.1$ Hz). ^{13}C NMR (125 MHz, CDCl_3): δ 162.925, 162.920, 146.9, 144.3, 141.9, 139.8, 128.8, 127.3, 126.5, 125.6, 113.8, 108.0, 62.3, 61.5, 52.7, 28.8, 14.2, 14.1.

5ha: dimethyl-5-((2,4,4-trimethylpentan-2-yl)amino)pyrazolo[1,5-*c*]quinazoline-1,2-dicarboxylate

White solid, Yield: 0.119 g (85%), m.p.: 103-105 °C. ^1H NMR (500 MHz, CDCl_3): δ 8.76 (dd, 1H, aromatic C-H, $J = 8.2, 1.0$ Hz), 7.70 (dd, 1H, aromatic C-H, $J = 8.2, 0.7$ Hz), 7.63-7.59 (m, 1H, aromatic C-H), 7.35-7.32 (m, 1H, aromatic C-H), 6.51 (br s, 1H, N-H), 4.03 (s, 3H, sp^3 C-H), 3.99 (s, 3H, sp^3 C-H), 2.08 (s, 2H, sp^3 C-H), 1.69 (s, 6H, sp^3 C-H), 1.02 (s, 9H, sp^3 C-H). ^{13}C NMR (125 MHz, CDCl_3): δ 163.8, 163.2., 145.8, 143.1, 141.2, 139.9, 131.4, 126.3, 125.5, 123.4, 114.8, 107.6, 56.2, 52.9, 52.5, 51.3, 31.8, 31.5, 29.4. HRMS (ESI) calcd for $\text{C}_{22}\text{H}_{29}\text{N}_4\text{O}_4$ $[M+H]^+$ 413.2184, found 413.2178. Despite repeated drying on high vacuum, a significant solvent peak is observed in ^1H and ^{13}C NMR spectra. The isolated yield has been modified to 85 %.

5hb: diethyl-5-((2,4,4-trimethylpentan-2-yl)amino)pyrazolo[1,5-*c*]quinazoline-1,2-dicarboxylate

White solid, Yield: 0.111 g (73%), m.p.: 176-178 °C. ^1H NMR (500 MHz, CDCl_3): δ 8.83 (dd, 1H, aromatic C-H, $J = 8.2, 1.0$ Hz), 7.69 (dd, 1H, aromatic C-H, $J = 8.0, 0.7$ Hz), 7.63-7.59 (m, 1H, aromatic C-H), 7.34-7.31 (m, 1H,

aromatic C-H), 6.53 (br s, 1H, N-H), 4.49 (q, 2H, sp^3 C-H, $J = 7.2$ Hz), 4.45 (q, 2H, sp^3 C-H, $J = 7.2$ Hz), 2.08 (s, 2H, sp^3 C-H), 1.69 (s, 6H, sp^3 C-H), 1.45 (t, 3H, sp^3 C-H, $J = 7.1$ Hz), 1.42 (t, 3H, sp^3 C-H, $J = 7.1$ Hz), 1.02 (s, 9H, sp^3 C-H). ^{13}C NMR (125 MHz, CDCl_3): δ 163.3, 162.9, 146.3, 143.1, 141.2, 139.9, 131.3, 126.3, 125.7, 123.3, 114.9, 108.1, 62.1, 61.5, 56.2, 51.4, 31.8, 31.5, 29.4, 14.2, 14.1. HRMS (ESI) calcd for $\text{C}_{24}\text{H}_{33}\text{N}_4\text{O}_4$ [M+H]⁺ 441.2497, found 441.2490.

26aa: Methyl 5-(*tert*-butylamino)-3-tosyl-1,2,3,10b-tetrahydropyrazolo[1,5-*c*]quinazoline-1-carboxylate

White solid, Yield: 0.144 g (93%), m.p.: 137-138 °C. ^1H NMR (500 MHz, CDCl_3): δ 7.85 (d, 2H, aromatic C-H, $J = 8.1$ Hz), 7.36 (d, 2H, aromatic C-H, $J = 8.0$ Hz), 7.12 (t, 1H, aromatic C-H, $J = 7.0$ Hz), 6.93 (d, 1H, aromatic C-H, $J = 7.8$ Hz), 6.80 (t, 1H, aromatic C-H, $J = 7.2$ Hz), 6.76 (d, 1H, aromatic C-H, $J = 6.8$ Hz), 5.21 (br s, 1H, N-H), 4.14 (d, 1H, sp^3 C-H, $J = 7.2$ Hz), 3.91 (dd, 1H, sp^3 C-H, $J = 12.1, 8.4$ Hz), 3.79 (dd, 1H, sp^3 C-H, $J = 12.1, 2.8$ Hz), 3.36 (td, 1H, sp^3 C-H, $J = 8.0, 2.8$ Hz), 3.21 (s, 3H, sp^3 OC-H), 2.47 (s, 3H, sp^3 C-H), 1.33 (s, 9H, sp^3 C-H). ^{13}C NMR (125 MHz, CDCl_3): δ 170.9, 148.9, 145.6, 142.4, 131.8, 130.1, 129.16, 129.14, 126.6, 123.7, 120.8, 117.5, 62.4, 52.6, 51.9, 51.2, 47.9, 28.9, 21.7. HRMS (ESI) calcd for $\text{C}_{23}\text{H}_{29}\text{N}_4\text{O}_4\text{S}$ [M+H]⁺ 457.1904, found 457.1894.

26ab: Methyl 8-bromo-5-(*tert*-butylamino)-3-tosyl-1,2,3,10b-tetrahydropyrazolo[1,5-*c*]quinazoline-1-carboxylate

Yellow solid, Yield: 0.101 g (85%), m.p.: 178-180 °C. ^1H NMR (500 MHz, CDCl_3): δ 7.83 (d, 2H, aromatic C-H, $J = 8.2$ Hz), 7.36 (d, 2H, aromatic C-H, $J = 8.0$ Hz), 7.10 (d, 1H, aromatic C-H, $J = 1.7$ Hz), 6.91 (dd, 1H, aromatic C-H, $J = 8.0, 1.8$ Hz), 6.62 (d, 1H, aromatic C-H, $J = 8.0$ Hz), 5.28 (br s, 1H, N-H), 4.10 (d, 1H, sp^3 C-H, $J = 7.2$ Hz), 3.90 (dd, 1H, sp^3 C-H, $J = 12.1, 8.3$ Hz), 3.77 (dd, 1H, sp^3 C-H, $J = 12.2, 2.8$ Hz), 3.35 (td, 1H, sp^3 C-H, $J = 8.1, 2.8$ Hz), 3.27 (s, 3H, sp^3 OC-H), 2.47 (s, 3H, sp^3 C-H), 1.31 (s, 9H, sp^3 C-H). ^{13}C NMR (125 MHz, CDCl_3): δ 170.8, 149.6, 145.8, 144.1, 131.7, 130.2, 129.1, 127.8, 126.4, 123.4, 122.6, 116.3, 61.9, 52.3, 52.1, 51.4, 47.9, 28.8, 21.7. HRMS (ESI) calcd for $\text{C}_{23}\text{H}_{28}\text{BrN}_4\text{O}_4\text{S}$ [M+H]⁺ 535.1009, found 535.1007.

26ac: Methyl 5-(*tert*-butylamino)-8-chloro-3-tosyl-1,2,3,10b-tetrahydropyrazolo[1,5-*c*]quinazoline-1-carboxylate

Yellow solid, Yield: 0.116 g (86%), m.p.: 173-175 °C, ^1H NMR (500 MHz, CDCl_3): δ 7.84 (d, 2H, aromatic C-H, $J = 8.2$ Hz), 7.36 (d, 2H, aromatic C-H, $J = 8.0$ Hz), 6.94 (d, 1H, aromatic C-H, $J = 2.0$ Hz), 6.76 (dd, 1H, aromatic C-H, $J = 8.1, 2.1$ Hz), 6.68 (d, 1H, aromatic C-H, $J = 8.1$ Hz), 5.28 (br s, 1H, N-H), 4.12 (d, 1H, sp^3 C-H, $J = 7.2$ Hz), 3.91 (dd, 1H, sp^3 C-H, $J = 12.2, 8.3$ Hz), 3.78 (dd, 1H, sp^3 C-H, $J = 12.2, 2.9$ Hz), 3.36 (td, 1H, sp^3 C-H, $J = 8.2, 2.9$ Hz), 3.27 (s, 3H, sp^3 OC-H), 2.47 (s, 3H, sp^3 C-H), 1.31 (s, 9H, sp^3 C-H). ^{13}C NMR (125 MHz, CDCl_3): δ 170.8, 149.6, 145.7, 143.9, 134.5, 131.7, 130.2, 129.1, 127.5, 123.4, 120.5, 115.9, 61.8, 52.4, 52.1, 51.4, 47.9, 28.8, 21.7. HRMS (ESI) calcd for $\text{C}_{23}\text{H}_{28}\text{ClN}_4\text{O}_4\text{S}$ [M+H]⁺ 491.1515, found 491.1510.

26ad: Methyl 5-(*tert*-butylamino)-8-fluoro-3-tosyl-1,2,3,10b-tetrahydropyrazolo[1,5-*c*]quinazoline-1-carboxylate

Colorless liquid, Yield: 0.130 g (91%), ^1H NMR (500 MHz, CDCl_3): δ 7.85 (d, 2H, aromatic C-H, $J = 8.1$ Hz), 7.37 (d, 2H, aromatic C-H, $J = 8.0$ Hz), 6.71 (dd, 1H, aromatic C-H, $J = 8.2, 6.4$ Hz), 6.63 (d, 1H, aromatic C-H, $J = 10.0$ Hz), 6.51 (td, 1H, aromatic C-H, $J = 8.4, 2.5$ Hz), 5.29 (br s, 1H, N-H), 4.14, (d, 1H, sp^3 C-H, $J = 7.2$ Hz), 3.91 (dd, 1H, sp^3 C-H, $J = 12.2, 8.3$ Hz), 3.78 (dd, 1H, sp^3 C-H, $J = 12.2, 2.8$ Hz), 3.34 (td, 1H, sp^3 C-H, $J = 8.0, 2.8$ Hz), 3.26 (s, 3H, sp^3 OC-H), 2.48 (s, 3H, sp^3 C-H), 1.32 (s, 9H, sp^3 C-H). ^{13}C NMR (125 MHz, CDCl_3): δ 170.9, 163.54 (d, $J_{\text{C-F}} = 243$ Hz), 149.5, 145.7, 131.8, 130.2, 129.2, 127.6 (d, $J_{\text{C-F}} = 10$ Hz), 113.2, 109.9 (d, $J_{\text{C-F}} = 15.2$ Hz), 107.6 (d, $J_{\text{C-F}} = 22.5$ Hz), 61.9, 52.4, 52.0, 51.4, 47.9, 28.8, 21.7. ^{19}F NMR (470 MHz, CDCl_3): δ -113.74. HRMS (ESI) calcd for $\text{C}_{23}\text{H}_{28}\text{FN}_4\text{O}_4\text{S}$ [M+H]⁺ 475.1810, found 475.1798.

26ae: Methyl 5-(*tert*-butylamino)-3-tosyl-8-(trifluoromethyl)-1,2,3,10b-tetrahydropyrazolo[1,5-*c*]quinazoline-1-carboxylate

Pale yellow Oil, Yield: 0.106 g (87%). ^1H NMR (500 MHz, CDCl_3): δ 7.85 (d, 2H, aromatic C-H, $J = 8.0$ Hz), 7.38 (d, 2H, aromatic C-H, $J = 7.8$ Hz), 7.18 (s, 1H, aromatic C-H), 7.03 (d, 1H, aromatic C-H, $J = 7.6$ Hz), 6.87 (d, 1H, aromatic C-H, $J = 7.6$ Hz), 5.29 (br s, 1H, N-H), 4.20 (d, 1H, sp^3 C-H, $J = 7.1$ Hz), 3.94 (dd, 1H, sp^3 C-H, $J = 12.1, 8.5$ Hz), 3.80 (dd, 1H, sp^3 C-H, $J = 12.1, 2.4$ Hz), 3.41 (t, 1H, sp^3 C-H, $J = 6.0$ Hz), 3.23 (s, 3H, sp^3 OC-H), 2.48 (s, 3H, sp^3 C-H), 1.32 (s, 9H, sp^3 C-H). ^{13}C NMR (125 MHz, CDCl_3): δ 170.7, 149.7, 145.8, 143.2, 131.7, 131.5 (q, $J_{\text{C-F}} = 31.5$ Hz), 130.2, 129.2, 127.0, 124.1 (q, $J_{\text{C-F}} = 270.7$ Hz), 120.9, 120.3 (d, $J_{\text{C-F}} = 4.6$ Hz), 116.9 (d, $J_{\text{C-F}} = 7.5$ Hz), 61.8, 52.4, 52.0, 51.5, 47.8, 28.8, 21.7. ^{19}F NMR (470 MHz, CDCl_3): δ -62.88. HRMS (ESI) calcd for $\text{C}_{24}\text{H}_{28}\text{F}_3\text{N}_4\text{O}_4\text{S}$ [M+H]⁺ 525.1778, found 525.1764.

26ba: methyl 5-(*tert*-butylamino)-3-(phenylsulfonyl)-1,2,3,10b-tetrahydropyrazolo[1,5-c]quinazoline-1-carboxylate

Off-white solid, Yield: 0.134 g (89%), m.p.: 130-132 °C. ¹H NMR (500 MHz, CDCl₃): δ 7.98 (dd, 2H, aromatic C-H, J = 8.5, 0.8 Hz), 7.71 (t, 1H, aromatic C-H, J = 7.5 Hz), 7.58 (t, 2H, aromatic C-H, J = 8.0 Hz), 7.13 (td, 1H, aromatic C-H, J = 8.7, 1.3 Hz), 6.94 (d, 1H, aromatic C-H, J = 7.8 Hz), 6.81, (td, 1H, aromatic C-H, J = 7.4, 1.1 Hz), 6.75 (dd, 1H, aromatic C-H, J = 7.3, 1.1 Hz), 5.19 (br s, 1H, N-H), 4.12 (d, 1H, sp³ C-H, J = 7.3 Hz), 3.94 (dd, 1H, sp³ C-H, J = 12.2, 8.3 Hz), 3.82 (dd, 1H, sp³ C-H, J = 12.2, 3.0 Hz), 3.36, (td, 1H, sp³ C-H, J = 8.2, 3.0 Hz), 3.22 (s, 3H, sp³ OC-H), 1.32 (s, 9H, sp³ C-H). ¹³C NMR (125 MHz, CDCl₃): δ 170.9, 148.8, 142.3, 134.9, 134.3, 129.5, 129.1, 128.1, 126.6, 123.7, 120.9, 117.4, 62.5, 52.5, 51.9, 51.2, 48.0, 28.9. HRMS (ESI) calcd for C₂₂H₂₇N₄O₄S [M+H]⁺ 443.1748, found 443.1735.

26bb: methyl 5-(*tert*-butylamino)-3-((4-iodophenyl)sulfonyl)-1,2,3,10b-tetrahydropyrazolo[1,5-c]quinazoline-1-carboxylate

White solid, Yield: 0.174 g (90%), m.p.: 147-149 °C. ¹H NMR (500 MHz, CDCl₃): δ 7.94 (d, 2H, aromatic C-H, J = 8.1 Hz), 7.68 (d, 2H, aromatic C-H, J = 8.1 Hz), 7.15 (t, 1H, aromatic C-H, J = 7.0 Hz), 7.95 (d, 1H, aromatic C-H, J = 7.8 Hz), 6.85-6.80 (m, 2H, aromatic C-H), 5.12 (brs, 1H, N-H), 4.21 (d, 1H, sp³ C-H, J = 7.1 Hz), 3.88 (dd, 1H, sp³ C-H, J = 12.0, 8.5 Hz), 3.82 (dd, 1H, sp³ C-H, J = 12.0, 2.2 Hz), 3.39, (td, 1H, sp³ C-H, J = 7.0, 5.5 Hz), 3.22 (s, 3H, sp³ OC-H), 1.33 (s, 9H, sp³ C-H). ¹³C NMR (125 MHz, CDCl₃): δ 170.8, 148.6, 142.2, 138.8, 134.5, 130.4, 129.3, 126.7, 123.8, 121.0, 117.3, 102.5, 62.6, 52.4, 52.0, 51.3, 47.9, 28.9. HRMS (ESI) calcd for C₂₂H₂₆IN₄O₄S [M+H]⁺ 569.0714, found 569.0700.

26bc: Methyl 3-((4-bromophenyl)sulfonyl)-5-(*tert*-butylamino)-1,2,3,10b-tetrahydropyrazolo[1,5-c]quinazoline-1-carboxylate

White solid, Yield: 0.160 g (91%), m.p.: 142-144 °C. ¹H NMR (500 MHz, CDCl₃): δ 7.84 (d, 2H, aromatic C-H, J = 8.5 Hz), 7.72 (d, 2H, aromatic C-H, J = 8.2 Hz), 7.15 (t, 1H, aromatic C-H, J = 6.6 Hz), 6.95 (d, 1H, aromatic C-H, J = 7.7 Hz), 6.86-6.80 (m, 2H, aromatic C-H), 5.13 (br s, 1H, N-H), 4.20 (d, 1H, sp³ C-H, J = 7.2 Hz), 3.89 (dd, 1H, sp³ C-H, J = 12.1, 8.1 Hz), 3.39 (td, 1H, sp³ C-H, J = 7.6, 2.4 Hz), 3.22 (s, 3H, sp³ OC-H), 1.33 (s, 9H, sp³ C-H). ¹³C NMR (125 MHz, CDCl₃): δ 170.8, 148.6, 142.2, 133.9, 132.8, 130.6, 129.9, 129.3, 126.6, 123.8, 121.0, 117.3, 62.6, 52.4, 52.0, 51.3, 48.0, 28.9. HRMS (ESI) calcd for C₂₂H₂₆BrN₄O₄S [M+H]⁺ 521.0853, found 521.0849.

26bd: Methyl 5-(*tert*-butylamino)-3-((4-nitrophenyl)sulfonyl)-1,2,3,10b-tetrahydropyrazolo[1,5-c]quinazoline-1-carboxylate

Yellow solid, Yield: 0.147 g (89%), m.p.: 131-133 °C. ¹H NMR (500 MHz, CDCl₃): δ 8.41 (d, 2H, aromatic C-H, J = 8.6 Hz), 8.19 (d, 2H, aromatic C-H, J = 8.6 Hz), 7.16 (t, 1H, aromatic C-H, J = 7.4 Hz), 6.96 (d, 1H, aromatic C-H, J = 7.9 Hz), 6.84 (t, 1H, aromatic C-H, J = 7.4 Hz), 6.78 (d, 1H, aromatic C-H, J = 7.0 Hz), 5.13 (br s, 1H, N-H), 4.13 (d, 1H, sp³ C-H, J = 7.1 Hz), 3.91-3.89 (m, 2H, sp³ C-H), 3.42-3.38 (m, 1H, sp³ C-H), 3.22 (s, 3H, sp³ OC-H), 1.33 (s, 9H, sp³ C-H). ¹³C NMR (125 MHz, CDCl₃): δ 170.5, 151.1, 148.2, 141.9, 140.6, 130.6, 129.4, 126.6, 124.5, 123.9, 121.3, 117.1, 62.9, 52.1, 52.0, 51.4, 48.2, 28.9. HRMS (ESI) calcd for C₂₂H₂₆N₅O₆S [M+H]⁺ 488.1599, found 488.1599.

26be: Methyl 5-(*tert*-butylamino)-3-(naphthalen-1-ylsulfonyl)-1,2,3,10b-tetrahydropyrazolo[1,5-c]quinazoline-1-carboxylate

Yellow solid, Yield: 0.149 g (89%), m.p.: 176-178 °C. ¹H NMR (500 MHz, CDCl₃): δ 8.89 (d, 1H, aromatic C-H, J = 8.1 Hz), 8.36 (d, 1H, aromatic C-H, J = 6.8 Hz), 8.18 (d, 1H, aromatic C-H, J = 7.6 Hz), 8.02 (d, 1H, aromatic C-H, J = 7.7 Hz), 7.77 (t, 1H, aromatic C-H, J = 6.6 Hz), 7.66 (t, 1H, aromatic C-H, J = 7.0 Hz), 7.58 (t, 1H, aromatic C-H, J = 7.1 Hz), 7.12 (t, 1H, aromatic C-H, J = 6.5 Hz), 6.96 (d, 1H, aromatic C-H, J = 6.5 Hz), 6.86 (d, 2H, aromatic C-H, J = 7.4 Hz), 5.05 (d, 1H, aromatic C-H, J = 7.2 Hz), 4.46 (t, 1H, sp³ C-H, J = 11.2 Hz), 4.32 (br s, 1H, N-H), 3.80 (d, 1H, sp³ C-H, J = 11.7 Hz), 3.62 (s, 1H, sp³ C-H), 3.19 (s, 3H, sp³ OC-H), 0.72 (s, 9H, sp³ C-H). ¹³C NMR (125 MHz, CDCl₃): δ 171.5, 148.2, 142.3, 135.9, 134.3, 133.6, 130.2, 129.6, 129.5, 129.0, 128.9, 127.4, 126.9, 124.6, 124.3, 123.5, 120.9, 117.9, 63.3, 52.2, 51.9, 50.5, 46.6, 28.2. HRMS (ESI) calcd for C₂₆H₂₉N₄O₄S [M+H]⁺ 493.1904, found 493.1897.

26bf: Methyl 5-(*tert*-butylamino)-3-((4-fluorophenyl)sulfonyl)-1,2,3,10b-tetrahydropyrazolo[1,5-c]quinazoline-1-carboxylate

Off-white solid, Yield: 0.131 g (84%), m.p.: 131-133 °C. ¹H NMR (500 MHz, CDCl₃): δ 8.03-7.99 (m, 2H, aromatic C-H), 7.27-7.23 (m, 2H, aromatic C-H), 7.14 (td, 1H, aromatic C-H, J = 8.8, 1.6 Hz), 6.96 (d, 1H, aromatic C-H, J = 7.8 Hz), 6.85-6.79 (m, 2H, aromatic C-H), 5.18 (br s, 1H, N-H), 4.17 (d, 1H, sp³ C-H, J = 7.2 Hz), 3.90 (dd, 1H, sp³ C-H, J = 12.1, 8.2 Hz), 3.83 (dd, 1H, sp³ C-H, J = 12.1, 3.0 Hz), 3.39 (td, 1H, sp³ C-H, J = 8.0, 3.0 Hz), 3.22

(s, 3H, sp^3 OC-H), 1.34 (s, 9H, sp^3 C-H). ^{13}C NMR (125 MHz, CDCl_3): δ 170.7, 167.3, 165.2, 148.7, 132.0 (d, $J_{\text{C}-\text{F}} = 9.6$ Hz), 130.9, 129.3, 126.6, 123.7, 121.0, 117.3, 116.8 (d, $J_{\text{C}-\text{F}} = 22.4$ Hz), 62.6, 52.4, 52.0, 51.3, 48.0, 28.9. HRMS (ESI) calcd for $\text{C}_{22}\text{H}_{26}\text{FN}_4\text{O}_4\text{S}$ [M+H]⁺ 461.1654, found 461.1644.

26ca: methyl 3-((4-methoxyphenyl)sulfonyl)-5-((2,4,4-trimethylpentan-2-yl)amino)-1,2,3,10b-tetrahydropyrazolo[1,5-c]quinazoline-1-carboxylate

Off-white solid, Yield: 0.159 g (89%), m.p.: 161-163 °C. ^1H NMR (500 MHz, CDCl_3): δ 7.87 (d, 2H, aromatic C-H, $J = 8.8$ Hz), 7.12 (t, 1H, aromatic C-H, $J = 7.1$ Hz), 7.00 (d, 2H, aromatic C-H, $J = 8.8$ Hz), 6.93 (d, 1H, aromatic C-H, $J = 7.8$ Hz), 6.80 (t, 1H, aromatic C-H, $J = 7.3$ Hz), 6.74 (d, 1H, aromatic C-H, $J = 7.0$ Hz), 5.39 (br s, 1H, N-H), 4.06 (d, 1H, sp^3 C-H, $J = 7.4$ Hz), 3.92-3.90 (m, 1H, sp^3 C-H), 3.88 (s, 3H, sp^3 OC-H), 3.77 (dd, 1H, sp^3 C-H, $J = 12.2, 3.1$ Hz), 3.35 (td, 1H, sp^3 C-H, $J = 8.0, 3.1$ Hz), 3.21 (s, 3H, sp^3 C-H), 2.04 (d, 1H, sp^3 C-H, $J = 14.7$ Hz), 1.48-1.45 (m, 4H, sp^3 C-H), 1.45 (s, 3H, sp^3 C-H), 1.02 (s, 9H, sp^3 C-H). ^{13}C NMR (125 MHz, CDCl_3): δ 171.0, 164.3, 148.6, 142.4, 131.3, 129.1, 126.6, 126.0, 123.7, 120.6, 117.5, 114.7, 62.4, 55.8, 55.1, 52.7, 52.1, 51.9, 48.3, 31.6, 31.5, 28.4. HRMS (ESI) calcd for $\text{C}_{27}\text{H}_{37}\text{N}_4\text{O}_5\text{S}$ [M+H]⁺ 529.2479, found 529.2465.

26cb: Methyl 3-((4-chlorophenyl)sulfonyl)-5-((2,4,4-trimethylpentan-2-yl)amino)-1,2,3,10b-tetrahydropyrazolo[1,5-c]quinazoline-1-carboxylate

Pale yellow solid, Yield: 0.161 g (89%), m.p.: 171-173 °C. ^1H NMR (500 MHz, CDCl_3): δ 7.90 (d, 2H, aromatic C-H, $J = 5.8$ Hz), 7.55 (d, 2H, aromatic C-H, $J = 5.7$ Hz), 7.14 (s, 1H, aromatic C-H), 6.95 (d, 1H, aromatic C-H, $J = 5.8$ Hz), 6.82-6.79 (m, 2H, aromatic C-H), 5.31 (br s, 1H, N-H), 4.09 (d, 1H, sp^3 C-H, $J = 4.7$ Hz), 3.90-3.81 (m, 2H, sp^3 C-H), 3.37 (s, 1H, sp^3 C-H), 3.22 (s, 3H, sp^3 OC-H), 2.02 (d, 1H, sp^3 C-H, $J = 14.3$ Hz), 1.49-1.40 (m, 7H, sp^3 C-H), 1.03 (s, 9H, sp^3 C-H). ^{13}C NMR (125 MHz, CDCl_3): δ 170.7, 148.2, 142.3, 141.3, 133.3, 130.5, 129.9, 129.2, 126.6, 123.8, 120.8, 117.3, 62.6, 55.2, 52.4, 52.2, 51.9, 48.4, 31.6, 31.5, 29.7, 28.3. HRMS (ESI) calcd for $\text{C}_{26}\text{H}_{34}\text{ClN}_4\text{O}_4\text{S}$ [M+H]⁺ 533.1984, found 533.1975.

26cc: Methyl 3-(benzylsulfonyl)-5-((2,4,4-trimethylpentan-2-yl)amino)-1,2,3,10b-tetrahydropyrazolo[1,5-c]quinazoline-1-carboxylate

White solid, Yield: 0.156 g (90%), m.p.: 143-145 °C. ^1H NMR (500 MHz, CDCl_3): δ 7.47-7.42 (m, 5H, aromatic C-H), 7.20 (td, 1H, aromatic C-H, $J = 7.8, 1.2$ Hz), 7.03 (d, 2H, aromatic C-H, $J = 7.8$ Hz), 7.00 (d, 2H, aromatic C-H, $J = 7.3$ Hz), 6.91 (t, 1H, aromatic C-H, $J = 7.32$ Hz), 5.53 (br s, 1H, N-H), 5.07 (d, 1H, sp^3 C-H, $J = 7.7$ Hz), 4.57 (d, 1H, sp^3 C-H, $J = 13.3$ Hz), 4.40 (d, 1H, sp^3 C-H, $J = 13.3$ Hz), 4.05 (dd, 1H, sp^3 C-H, $J = 12.2, 8.6$ Hz), 3.72 (dd, 1H, sp^3 C-H, $J = 12.3, 3.6$ Hz), 3.55 (td, 1H, sp^3 C-H, $J = 8.1, 3.7$ Hz), 3.25 (s, 3H, sp^3 OC-H), 2.11 (d, 1H, sp^3 C-H, $J = 14.3$ Hz), 1.59 (d, 1H, sp^3 C-H, $J = 14.8$ Hz), 1.53 (s, 6H, sp^3 C-H), 1.06 (s, 9H, sp^3 C-H). ^{13}C NMR (125 MHz, CDCl_3): δ 171.1, 148.2, 142.2, 131.0, 129.3, 129.2, 129.1, 126.9, 126.4, 123.8, 121.2, 117.8, 63.4, 55.6, 54.7, 52.6, 51.99, 51.96, 47.5, 31.6, 29.6, 28.3. HRMS (ESI) calcd for $\text{C}_{27}\text{H}_{37}\text{N}_4\text{O}_4\text{S}$ [M+H]⁺ 513.2530, found 513.2525.

26cd: Methyl 3-(octylsulfonyl)-5-((2,4,4-trimethylpentan-2-yl)amino)-1,2,3,10b-tetrahydropyrazolo[1,5-c]quinazoline-1-carboxylate

White solid, Yield: 0.167 g (92%), m.p.: 137-138 °C. ^1H NMR (500 MHz, CDCl_3): δ 7.17 (t, 1H, aromatic C-H, $J = 7.4$ Hz), 7.00 (d, 2H, aromatic C-H, $J = 8.9$ Hz), 6.89 (t, 1H, aromatic C-H, $J = 7.3$ Hz), 5.47 (br s, 1H, N-H), 5.03 (d, 1H, sp^3 C-H, $J = 7.8$ Hz), 4.07 (dd, 1H, sp^3 C-H, $J = 12.3, 8.5$ Hz), 3.64 (dd, 1H, sp^3 C-H, $J = 12.3, 3.7$ Hz), 3.58 (td, 1H, sp^3 C-H, $J = 8.1, 3.8$ Hz), 3.25 (s, 3H, sp^3 OC-H), 2.20 (d, 1H, sp^3 C-H, $J = 14.8$ Hz), 1.91 (quint, 2H, sp^3 C-H, $J = 7.8$ Hz), 1.59 (d, 1H, sp^3 C-H, $J = 14.8$ Hz), 1.52 (d, 6H, sp^3 C-H, $J = 3.6$ Hz), 1.47-1.42 (m, 2H, sp^3 C-H), 1.32-1.27 (m, 10H, sp^3 C-H), 1.07 (s, 9H, sp^3 C-H), 0.88 (t, 3H, sp^3 C-H, $J = 6.4$ Hz). ^{13}C NMR (125 MHz, CDCl_3): δ 171.3, 148.2, 142.1, 129.2, 127.0, 123.8, 121.2, 117.9, 63.1, 55.5, 52.4, 51.9, 51.8, 49.2, 46.5, 31.7, 31.6, 29.8, 28.98, 28.94, 28.53, 28.4, 22.7, 22.6, 14.0. HRMS (ESI) calcd for $\text{C}_{28}\text{H}_{47}\text{N}_4\text{O}_4\text{S}$ [M+H]⁺ 535.3313, found 535.3309.

26ce: Methyl 3-tosyl-5-((2,4,4-trimethylpentan-2-yl)amino)-1,2,3,10b-tetrahydropyrazolo[1,5-c]quinazoline-1-carboxylate

Yellow solid, Yield: 0.155 g (89%), m.p.: 132-134 °C. ^1H NMR (500 MHz, CDCl_3): δ 7.83 (d, 2H, aromatic C-H, $J = 8.2$ Hz), 7.35 (d, 2H, aromatic C-H, $J = 8.1$ Hz), 7.12 (td, 1H, aromatic C-H, $J = 8.3, 1.2$ Hz), 6.93 (d, 1H, aromatic C-H, $J = 7.8$ Hz), 6.79 (t, 1H, aromatic C-H, $J = 6.7$ Hz), 6.73 (d, 1H, aromatic C-H, $J = 6.7$ Hz), 5.36 (br s, 1H, N-H), 4.06 (d, 1H, sp^3 C-H, $J = 7.5$ Hz), 3.93 (dd, 1H, sp^3 C-H, $J = 12.2, 8.4$ Hz), 3.77 (dd, 1H, sp^3 C-H, $J = 12.2, 3.3$ Hz), 3.34 (td, 1H, sp^3 C-H, $J = 8.1, 3.3$ Hz), 3.21 (s, 3H, sp^3 OC-H), 2.46 (s, 3H, sp^3 C-H), 2.02 (d, 1H, sp^3 C-H, $J = 14.7$ Hz), 1.48 (d, 1H, sp^3 C-H, $J = 14.7$ Hz), 1.45 (s, 3H, sp^3 C-H), 1.39 (s, 3H, sp^3 C-H), 1.02 (s, 9H, sp^3 C-H). ^{13}C NMR (125 MHz, CDCl_3): δ 170.9, 148.5, 145.5, 142.4, 131.9, 130.1, 129.1, 126.6, 123.7,

120.6, 117.5, 62.4, 55.1, 52.6, 52.1, 51.9, 48.3, 31.6, 31.5, 29.7, 28.3, 21.7. HRMS (ESI) calcd for C₂₇H₃₇N₄O₄S [M+H]⁺ 513.2530, found 513.2515.

26da: Ethyl 5-(*tert*-butylamino)-3-tosyl-1,2,3,10b-tetrahydropyrazolo[1,5-*c*]quinazoline-1-carboxylate

White solid, Yield: 0.140 g (88%), m.p.: 153-155 °C. ¹H NMR (500 MHz, CDCl₃): δ 7.85 (d, 2H, aromatic C-H, J = 8.2 Hz), 7.35 (d, 2H, aromatic C-H, J = 8.1 Hz), 7.12 (td, 1H, aromatic C-H, J = 8.3, 2.3 Hz), 6.92 (d, 1H, aromatic C-H, J = 7.8 Hz), 6.81-6.77 (m, 2H, aromatic C-H), 5.20 (br s, 1H, N-H), 3.90 (dd, 1H, sp³ C-H, J = 8.2, 7.2 Hz), 3.81 (dd, 1H, sp³ C-H, J = 12.1, 3.0 Hz), 3.73-3.63 (m, 2H, sp³ C-H), 3.34 (td, 1H, sp³ C-H, J = 7.9, 3.0 Hz), 2.46 (s, 3H, sp³ C-H), 1.32 (s, 9H, sp³ C-H), 0.74 (t, 3H, sp³ C-H, J = 7.1 Hz). ¹³C NMR (125 MHz, CDCl₃): δ 170.7, 148.9, 145.5, 142.7, 131.9, 130.1, 129.2, 129.1, 126.7, 123.7, 120.7, 117.7, 62.4, 61.2, 52.2, 51.2, 48.0, 28.8, 21.7, 13.3. HRMS (ESI) calcd for C₂₄H₃₁N₄O₄S [M+H]⁺ 471.2061, found 471.2059. Despite repeated drying on high vacuum, a significant solvent peak is observed in ¹H and ¹³C NMR spectra. The isolated yield has been modified to 76 %.

26db: Butyl 5-(*tert*-butylamino)-3-tosyl-1,2,3,10b-tetrahydropyrazolo[1,5-*c*]quinazoline-1-carboxylate:

White solid, Yield: 0.158 g (93%), m.p.: 137-138 °C. ¹H NMR (500 MHz, CDCl₃): δ 7.84 (d, 2H, aromatic C-H, J = 8.2 Hz), 7.35 (d, 2H, aromatic C-H, J = 8.1 Hz), 7.13 (m, 1H, aromatic C-H, J = 8.5, 1.6 Hz), 6.93 (d, 1H, aromatic C-H, J = 7.8 Hz) 6.81-6.76 (m, 2H, aromatic C-H), 5.19 (br s, 1H, N-H), 4.13 (t, 1H, sp³ C-H, J = 7.1 Hz), 4.02 (t, 2H, sp³ C-H, J = 6.6 Hz), 3.91-3.83 (m, 3H, sp³ C-H), 3.80 (d, 1H, sp³ C-H, J = 12.1, 2.9 Hz), 3.35 (td, 1H, sp³ C-H, J = 7.9, 2.9 Hz), 2.46 (s, 3H, sp³ C-H), 2.19-2.16 (m, 1H, sp³ C-H), 1.98-1.94 (m, 1H, sp³ C-H), 1.57 (quintet, 1H, sp³ C-H, J = 6.8 Hz), 1.31 (s, 9H, sp³ C-H), 0.93 (t, 3H, sp³ C-H, J = 7.3 Hz). ¹³C NMR (125 MHz, CDCl₃): δ 170.5, 148.9, 145.6, 142.6, 131.9, 130.1, 129.2, 129.1, 126.8, 123.7, 120.7, 117.7, 62.4, 60.3, 52.0, 51.3, 47.9, 30.5, 28.8, 21.7, 19.0, 13.7.

26dc: *tert*-butyl 5-(*tert*-butylamino)-3-tosyl-1,2,3,10b-tetrahydropyrazolo[1,5-*c*]quinazoline-1-carboxylate

White solid, Yield: 0.155 g (92%), m.p.: 153-155 °C. ¹H NMR (500 MHz, CDCl₃): δ 7.84, (d, 2H, aromatic C-H, J = 8.1 Hz), 7.34 (d, 2H, aromatic C-H, J = 8.0 Hz), 7.13 (td, 1H, aromatic C-H, J = 8.3, 2.3 Hz), 6.92 (d, 1H, aromatic C-H, J = 7.8 Hz), 6.82-6.78 (m, 1H, aromatic C-H), 5.21 (br s, 1H, N-H), 4.10 (d, 1H, sp³ C-H, J = 7.2 Hz), 3.82-3.81 (m, 2H, sp³ C-H), 3.22 (td, 1H, sp³ C-H, J = 6.9, 4.3 Hz), 2.46 (s, 3H, sp³ C-H), 1.31 (s, 9H, sp³ C-H), 0.98 (s, 3H, sp³ C-H). ¹³C NMR (125 MHz, CDCl₃): δ 169.5, 149.0, 145.4, 142.9, 131.9, 130.0, 129.2, 128.9, 127.0, 123.7, 120.5, 118.2, 81.5, 62.3, 52.4, 51.2, 48.0, 28.8, 27.2, 21.7. HRMS (ESI) calcd for C₂₆H₃₅N₄O₄S [M+H]⁺ 499.2374, found 499.2367.

26dd: Phenyl 5-(*tert*-butylamino)-3-tosyl-1,2,3,10b-tetrahydropyrazolo[1,5-*c*]quinazoline-1 carboxylate

Pale yellow solid, Yield: 0.155 g (88%), m.p.: 177-179 °C. ¹H NMR (500 MHz, CDCl₃): δ 7.88 (d, 2H, aromatic C-H, J = 8.2 Hz), 7.38 (d, 2H, aromatic C-H, J = 8.0 Hz), 7.21-7.15 (m, 3H, aromatic C-H), 7.08 (t, 1H, aromatic C-H, J = 7.3 Hz), 7.02 (d, 1H, aromatic C-H, J = 7.4 Hz), 6.91 (d, 1H, aromatic C-H, J = 6.4 Hz), 6.83 (td, 1H, aromatic C-H, J = 7.3, 1.0 Hz), 6.43 (d, 2H, aromatic C-H, J = 7.6 Hz), 5.20 (br s, 1H, N-H), 4.35 (d, 1H, sp³ C-H, J = 7.3 Hz), 4.06 (dd, 1H, sp³ C-H, J = 12.3, 8.4 Hz), 3.92 (dd, 1H, sp³ C-H, J = 12.2, 3.0 Hz), 3.62 (td, 1H, sp³ C-H, J = 8.2, 3.0 Hz), 2.48 (s, 3H, sp³ C-H), 1.22 (s, 9H, sp³ C-H). ¹³C NMR (125 MHz, CDCl₃): δ 169.6, 150.0, 148.8, 145.7, 142.8, 131.9, 130.2, 129.4, 129.2, 129.1, 127.0, 125.8, 123.9, 121.2, 121.1, 117.3, 62.5, 52.3, 51.2, 48.0, 28.8, 21.7. HRMS (ESI) calcd for C₂₈H₃₁N₄O₄S [M+H]⁺ 519.2061, found 519.2059.

26de: Benzyl-5-(*tert*-butylamino)-3-tosyl-1,2,3,10b-tetrahydropyrazolo[1,5-*c*]quinazoline-1- carboxylate

Off-white solid, Yield: 0.159 g (88%), m.p.: 148-150 °C. ¹H NMR (500 MHz, CDCl₃): δ 7.77 (d, 2H, aromatic C-H, J = 7.9 Hz), 7.27 (d, 2H, aromatic C-H, J = 7.8 Hz), 7.18-7.17 (m, 3H, aromatic C-H), 7.05 (t, 1H, aromatic C-H, J = 6.1 Hz), 6.87-6.85 (m, 3H, aromatic C-H), 6.72-6.69 (m, 2H, aromatic C-H), 5.14 (br s, 1H, N-H), 4.60 (d, 1H, sp³ C-H, J = 12.1 Hz), 4.40 (d, 1H, sp³ C-H, J = 12.1 Hz), 4.07 (d, 1H, sp³ C-H, J = 7.1 Hz), 3.83 (dd, 1H, sp³ C-H, J = 11.9, 8.4 Hz), 3.75 (dd, 1H, sp³ C-H, J = 11.8, 2.3 Hz), 3.31 (t, 1H, sp³ C-H, J = 7.1 Hz), 2.38 (s, 3H, sp³ C-H), 1.24 (s, 9H, sp³ C-H). ¹³C NMR (125 MHz, CDCl₃): δ 170.7, 148.9, 145.6, 142.6, 134.9, 131.9, 130.1, 129.2, 129.1, 128.5, 128.3, 128.2, 126.7, 123.9, 120.8, 117.6, 67.1, 62.4, 52.2, 51.3, 48.2, 28.8, 21.7. HRMS (ESI) calcd for C₂₉H₃₃N₄O₄S [M+H]⁺ 533.2217, found 533.2211.

26ea: 1-(5-(*tert*-butylamino)-3-tosyl-1,2,3,10b-tetrahydropyrazolo[1,5-*c*]quinazolin-1-yl)ethan-1-one

Off-white solid, Yield: 0.134 g (79%), m.p.: 113-115 °C. ¹H NMR (500 MHz, CDCl₃): δ 7.84 (d, 2H, aromatic C-H, J = 8.2 Hz), 7.34 (d, 2H, aromatic C-H, J = 8.0 Hz), 7.16 (td, 1H, aromatic C-H, J = 8.9, 1.6 Hz), 6.95 (d, 1H, aromatic C-H, J = 7.8 Hz), 6.89 (dd, 1H, aromatic C-H, J = 7.4, 1.4 Hz), 6.84 (td, 1H, aromatic C-H, J = 7.3, 1.0 Hz), 5.22 (br s, 1H, N-H), 4.12 (t, 1H, sp³ C-H, J = 7.7 Hz), 3.86 (dd, 1H, sp³ C-H, J = 11.5, 1.9 Hz), 3.66 (dd, 1H, sp³ C-H, J = 11.5, 7.6 Hz), 3.49 (td, 1H, sp³ C-H, J = 7.4, 1.7 Hz), 2.46 (s, 3H, sp³ C-H), 1.56 (s, 3H, sp³ C-H),

1.31 (s, 9H, sp^3 C-H). ^{13}C NMR (125 MHz, CDCl_3): δ 205.7, 149.4, 145.4, 142.8, 131.8, 130.0, 129.6, 129.2, 126.2, 124.2, 120.9, 117.7, 62.6, 58.0, 51.3, 47.2, 30.5, 28.7, 21.7. HRMS (ESI) calcd for $\text{C}_{23}\text{H}_{29}\text{N}_4\text{O}_3\text{S}$ [$\text{M}+\text{H}]^+$ 441.1955 found 441.1951. Despite repeated drying on high vacuum, a significant solvent peak is observed in ^1H and ^{13}C NMR spectra. The isolated yield has been modified to 79 %.

26fa: Diethyl 5-(*tert*-butylamino)-3-tosyl-1,2,3,10b-tetrahydropyrazolo[1,5-c]quinazoline-1,2-dicarboxylate
White solid, Yield: 0.156 g (70%), m.p.: 133–135 °C. ^1H NMR (500 MHz, CDCl_3): δ 7.90 (d, 2H, aromatic C-H, J = 8.2 Hz), 7.37 (d, 2H, aromatic C-H, J = 8.0 Hz), 7.12 (td, 1H, aromatic C-H, J = 8.2, 1.2 Hz), 6.90 (d, 1H, aromatic C-H, J = 7.9 Hz), 6.85–6.79 (m, 2H, aromatic C-H), 5.42 (br s, 1H, N-H), 4.96 (d, 1H, sp^3 C-H, J = 8.8 Hz), 4.57 (d, 1H, sp^3 C-H, J = 6.8 Hz), 4.21 (q, 2H, sp^3 C-H, J = 7.1 Hz), 3.80 (dd, 1H, sp^3 C-H, J = 8.7, 7.0 Hz), 3.68–3.63 (m, 2H, sp^3 C-H), 2.47 (s, 3H, sp^3 C-H), 1.22 (s, 9H, sp^3 C-H), 0.87 (t, 3H, sp^3 C-H, J = 6.3 Hz), 0.67 (t, 3H, sp^3 C-H, J = 7.1 Hz). ^{13}C NMR (125 MHz, CDCl_3): δ 168.8, 167.8, 148.9, 145.6, 143.0, 132.4, 130.1, 129.4, 129.2, 126.7, 123.8, 120.6, 117.0, 63.8, 62.0, 61.2, 60.7, 55.6, 51.0, 28.6, 21.7, 14.1, 13.3. HRMS (ESI) calcd for $\text{C}_{27}\text{H}_{35}\text{N}_4\text{O}_6\text{S}$ [$\text{M}+\text{H}]^+$ 543.2272, found 543.2265. Despite repeated drying on high vacuum, a significant solvent peak is observed in ^1H and ^{13}C NMR spectra. The isolated yield has been modified to 70 %.

26ga: 5-(*tert*-butylamino)-3-((4-fluorophenyl)sulfonyl)-1,2,3,10b-tetrahydropyrazolo[1,5-c]quinazoline-1-carbonitrile

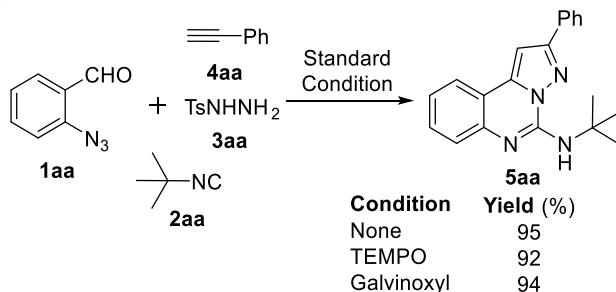
Yellow solid, Yield: 0.120 g (83%), m.p.: 179–181 °C. ^1H NMR (500 MHz, CDCl_3): δ 8.02–7.99 (m, 2H, aromatic C-H), 7.29–7.25 (m, 3H, aromatic C-H), 7.11 (d, 1H, aromatic C-H, J = 7.8 Hz), 6.91 (t, 1H, aromatic C-H, J = 7.3 Hz), 6.81 (d, 1H, aromatic C-H, J = 7.3 Hz), 5.10 (brs, 1H, N-H), 4.08 (d, 1H, sp^3 C-H, J = 5.0 Hz), 4.03 (dd, 1H, sp^3 C-H, J = 12.2, 8.9 Hz), 3.77 (dd, 1H, sp^3 C-H, J = 12.2, 1.7 Hz), 3.51 (t, 1H, sp^3 C-H, J = 6.0 Hz), 1.34 (s, 9H, sp^3 C-H). ^{13}C NMR (125 MHz, CDCl_3): δ 166.4 (d, $J_{\text{C}-\text{F}}$ = 257.5 Hz), 147.5, 142.3, 132.1 (d, $J_{\text{C}-\text{F}}$ = 11.25 Hz), 130.3, 125.9, 124.6, 121.8, 117.2, 116.9, 116.8, 116.2, 61.9, 51.6, 49.2, 38.9, 28.7. ^{19}F NMR (470 MHz, CDCl_3): δ -100.77. HRMS (ESI) calcd for $\text{C}_{21}\text{H}_{23}\text{FN}_5\text{O}_2\text{S}$ [$\text{M}+\text{H}]^+$ 428.1551, found 428.1540.

26ha: Methyl 3-benzoyl-5-(*tert*-butylamino)-1,2,3,10b-tetrahydropyrazolo[1,5-c]quinazoline-1-carboxylate
Colorless oil, Yield: 0.110 g (80%). ^1H NMR (500 MHz, CDCl_3): δ 7.81 (d, 2H, aromatic C-H, J = 7.5 Hz), 7.49–7.45 (m, 1H, aromatic C-H), 7.38 (t, 2H, aromatic C-H, J = 7.6 Hz), 7.16 (t, 1H, aromatic C-H, J = 7.4 Hz), 7.00 (d, 1H, aromatic C-H, J = 7.7 Hz), 6.94 (d, 1H, aromatic C-H, J = 7.2 Hz), 6.85 (t, 1H, aromatic C-H, J = 7.2 Hz), 5.09 (br s, 1H, N-H), 4.69 (s, 1H, sp^3 C-H), 4.50 (d, 1H, sp^3 C-H, J = 8.0 Hz), 3.66 (dd, 1H, sp^3 C-H, J = 11.5, 4.2 Hz), 3.53 (q, 1H, sp^3 C-H, J = 8.4 Hz), 3.23 (s, 3H, sp^3 OC-H), 1.47 (s, 9H, sp^3 C-H). ^{13}C NMR (125 MHz, CDCl_3): δ 173.7, 171.4, 148.8, 142.6, 132.6, 131.7, 130.5, 129.2, 128.6, 128.2, 126.7, 123.9, 121.1, 63.9, 51.8, 51.2, 46.2, 30.9, 29.2. HRMS (ESI) calcd for $\text{C}_{23}\text{H}_{27}\text{N}_4\text{O}_3$ [$\text{M}+\text{H}]^+$ 407.2078, found 407.2068.

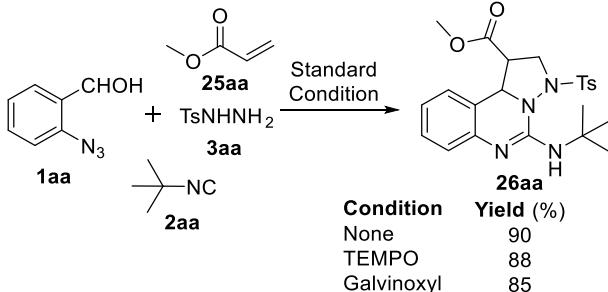
S5. Control Experiments for Elucidating Mechanism of 4-CR

S5.1. Radical Trap Experiment

A) For alkynes



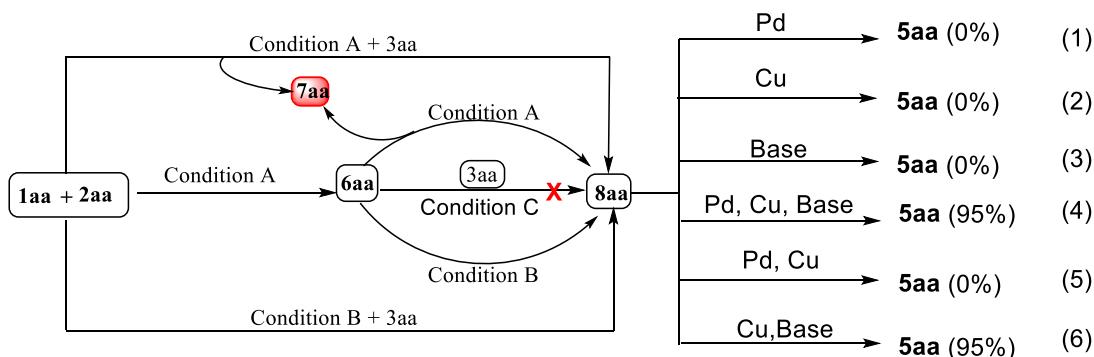
B) For activated alkenes



PROCEDURE: The reactions were carried out as per the protocol reported in section S3 for alkynes and acrylates in the presence of either TEMPO or Galvinoxyl (free radical scavenger). The products were purified by silica-gel column chromatography (EtOAc: Hexane) to produce **5aa** and **26aa** respectively. Radical scavengers did not alter the reaction pathway suggesting the involvement of a non-radical pathway (A & B).

S5.2 Evolution of species

Initially, a mixture of **1aa** and **2aa** was reacted under condition A leading to the formation of carbodiimide **6aa** that was characterized by ¹H NMR. The intermediate **6aa** was highly unstable and immediately converted to the corresponding urea upon prolonged exposure to air. Hence, it was generated *in situ*. Interestingly, **6aa**, generated *in situ*, or a mixture of **1aa** and **2aa** produced **8aa**, characterized by X-ray crystallographic studies (refer section S8), upon reacting with **3aa**, albeit in a low yield of 38%. On careful analysis of these two reaction mixtures, we observed a side product urea **7aa**, which was characterized by X-ray crystallographic studies (section S8). However, the yield was improved substantially after using 4 Å MS (condition B). Both these steps needed Palladium as a catalyst as **6aa** failed to produce **8aa** under the condition C. The conversion of **8aa** to **5aa** was dependent on both addition of second transition metal (Either Ag and Cu) and a suitable base.



Condition A: 7.5 mol% Pd(OAc)₂, toluene, rt; Condition B: 7.5 mol% Pd(OAc)₂, 4 Å MS, toluene, rt, Condition C: 4 Å MS, toluene, rt

6aa: 2-(((tert-butylimino)methylene)amino)benzaldehyde

¹H NMR (500 MHz, CDCl₃): δ 10.52 (s, 1H, aldehydic C-H), 7.84 (d, 1H, aromatic C-H, J = 7.7, 1.4 Hz), 7.51 (td, 1H, aromatic C-H, J = 8.0, 1.5 Hz), 7.25 (d, 1H, aromatic C-H, J = 8.0 Hz), 7.18 (t, 1H, aromatic C-H, J = 7.5 Hz), 1.45 (s, 9H, sp³ C-H).

7aa: (E)-N'-(2-(3-(tert-butyl)ureido)benzylidene)-4-methylbenzenesulfonohydrazide

¹H NMR (500 MHz, CDCl₃): δ 10.58 (s, 1H, aldehydic C-H), 8.32 (d, 1H, aromatic C-H, J = 7.7 Hz), 7.73-7.71 (m, 3H, aromatic C-H), 7.23-7.19 (m, 3H, aromatic C-H), 6.98 (d, 1H, aromatic C-H, J = 7.0 Hz), 6.81 (t, 1H, aromatic C-H, J = 5.8 Hz), 5.02 (br s, 1H, N-H), 2.32 (s, 3H, sp³ C-H), 1.45 (s, 9H, sp³ C-H).

8aa: 2-(tert-butylamino)quinazolin-3-i um-3-yl)(tosyl)amide

¹H NMR (500 MHz, CDCl₃): δ 9.40 (s, 1H, aromatic C-H), 7.84 (t, 1H, aromatic C-H, J = 7.3 Hz), 7.76 (d, 1H, aromatic C-H, J = 8.0 Hz), 7.60 (d, 1H, aromatic C-H, J = 8.5 Hz), 7.53 (d, 2H, aromatic C-H, J = 8.0 Hz), 7.38 (t, 1H, aromatic C-H, J = 7.4 Hz), 7.21 (s, 1H, aromatic C-H), 7.17 (d, 2H, aromatic C-H, J = 7.8 Hz), 2.36 (s, 3H, sp³ C-H), 1.18 (s, 9H, sp³ C-H). ¹³C NMR (125 MHz, CDCl₃): δ 154.8, 150.3, 148.2, 141.7, 141.1, 137.9, 129.6, 128.7, 126.1, 126.0, 124.9, 117.2, 52.2, 27.6, 21.3.

9aa: N-(tert-butyl)quinazolin-2-amine

Yellow solid, Yield: 0.037 g (55%), m.p.: 79-81 °C. ¹H NMR (500 MHz, CDCl₃): δ 8.93 (s, 1H, aromatic C-H), 7.67-7.62 (m, 2H, aromatic C-H), 7.57 (d, 1H, aromatic C-H, J = 8.3 Hz), 7.20 (td, 1H, aromatic C-H, J = 7.8, 0.9 Hz), 5.29 (br s, 1H, N-H), 1.53 (s, 9H, sp³ C-H). ¹³C NMR (125 MHz, CDCl₃): δ 161.4, 159.1, 151.9, 133.9, 127.4, 125.8, 122.3, 119.8, 51.1, 28.9.

S5.3 Chemical Kinetics of 4-CR

General Methods: Chemical kinetic study was performed by monitoring the reaction mixture using ¹H NMR (500 MHz) spectroscopy with dibenzyl ether as the internal standard. The spectra were processed and integrated with MestReNova (8.0.2) and the data was processed in Excel (2010). The graphs were plotted with OriginPro (8.6). The reported initial rates are the average of three independent experiments.

General Procedure: A reaction vial (10 mL) was charged with 2-azidobenzaldehyde (1.0 equiv), *tert*-butylisocyanide (1.2 equiv), Pd(OAc)₂ (7.5 mol %), 4 Å MS, TsNNH₂ (1 equiv), phenyl acetylene (1.5 equiv), AgOTf (10 mol%), K₃PO₄ (3.0 equiv) and toluene (0.5 mL). The reaction mixture was stirred at room temperature for the particular time interval. Then, the reaction mixture was filtered through a bed of Celite® and the solution so obtained was evaporated under reduced pressure. The mixture was then transferred into a standard 5 mm bore NMR tube and subjected to ¹H NMR (500 MHz) spectroscopy using dibenzyl ether (25 µL) as the internal standard.

Table S6. Characteristic ¹H NMR resonances used for monitoring chemical species.

Species	δ
1aa	10.36
5aa	7.19
8aa	9.40
6aa	10.52
10aa	8.01

S5.3.1 Four-component reaction (1aa + 2aa + 3aa + 4aa)

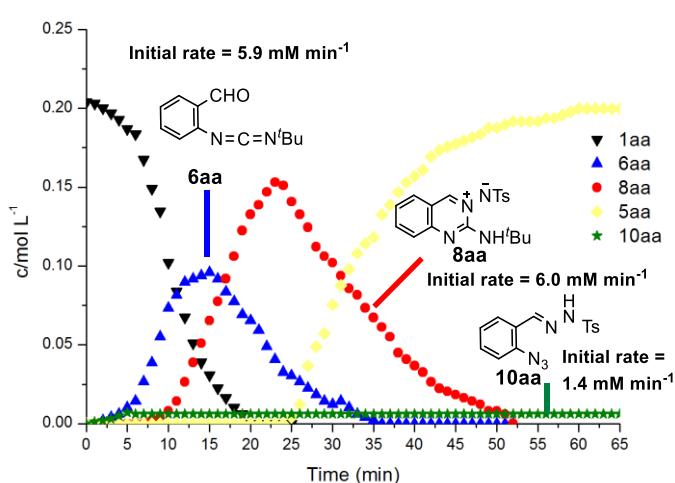
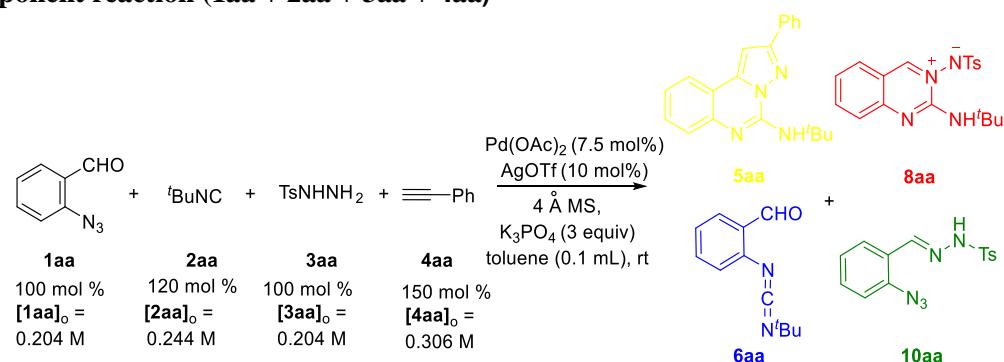
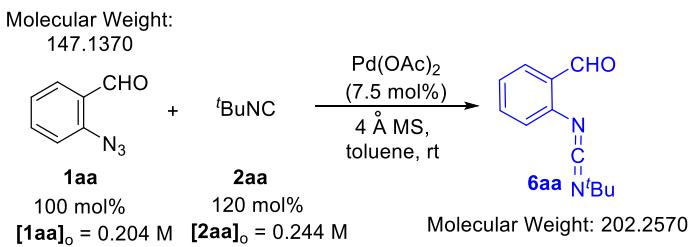


Figure S1. ¹H NMR spectroscopy profile of domino four CR

S5.3.2 Two-component reaction (1aa + 2aa)

Procedure: Same as section 5.3 with 2-azidobenzaldehyde (1.0 equiv), *tert*-butylisocyanide (1.2 equiv), Pd(OAc)₂ (7.5 mol%) and 4 Å MS in toluene.



Initial rate for formation of **6aa**: 9.8 mM min^{-1} (average of two experiments)

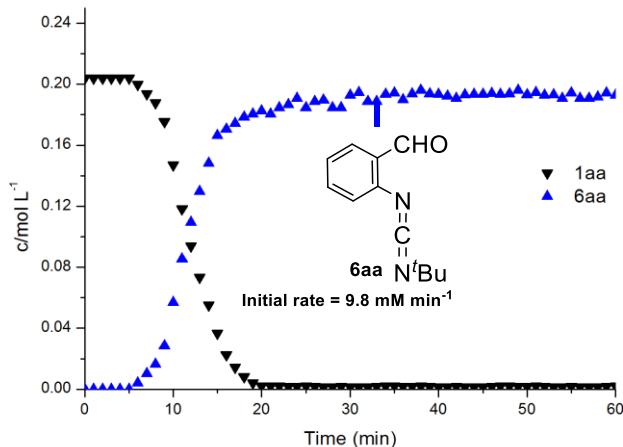
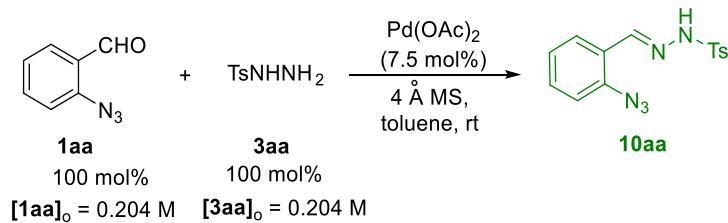


Figure S2. ^1H NMR spectroscopy profile of two CR (**1aa** + **2aa**)

S5.3.3 Two-component reaction (**1aa** + **3aa**)

Procedure: Same as section 5.3 with 2-azidobenzaldehyde (1.0 equiv), tosyl hydrazide (1.0 equiv), $\text{Pd}(\text{OAc})_2$ (7.5 mol%) and 4 Å MS in toluene.



Initial rate for formation of **10aa**: 5.9 mM min^{-1} (average of two experiments)

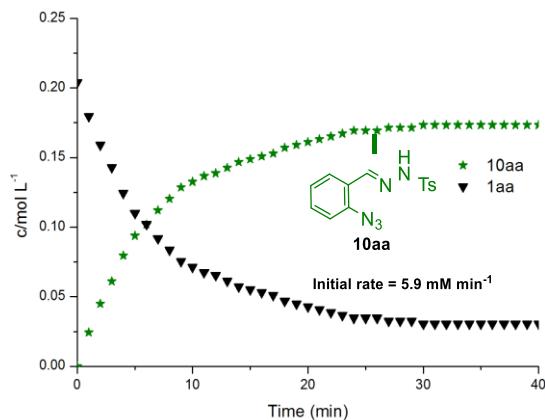
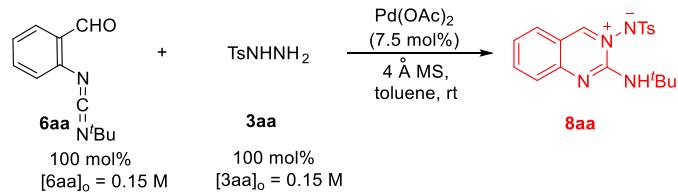


Figure S3. ^1H NMR spectroscopy profile of two CR (**1aa** + **3aa**)

S5.3.4 Two-component reaction (**6aa** + **3aa**)

Procedure: Same as section 5.3 with carbodiimide (1.0 equiv), tosyl hydrazide (1 equiv), $\text{Pd}(\text{OAc})_2$ (7.5 mol%) and 4 Å MS in toluene.



Initial rate for formation of **8aa**: 47.5 mM min^{-1} (average of two experiments)

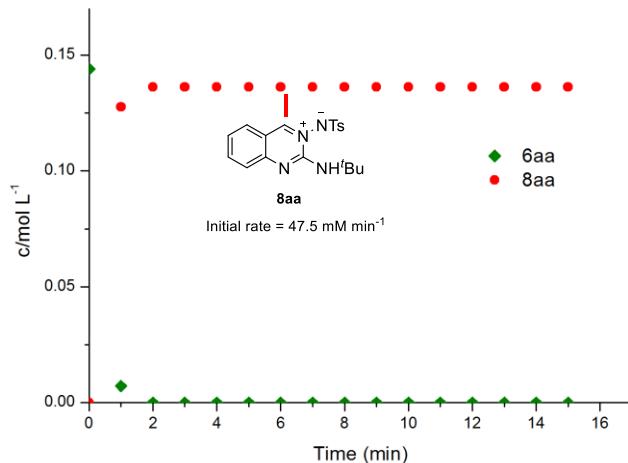


Figure S4. ^1H NMR spectroscopy profile of two CR (**6aa** + **3aa**)

S5.3.5. Two-component reaction (**10aa** + **2aa**)

Procedure: Same as section 5.3 with hydrazone (1.0 equiv), *tert*-butyl isocyanide (1 equiv), Pd(OAc)_2 (7.5 mol%) and 4 Å MS in toluene.

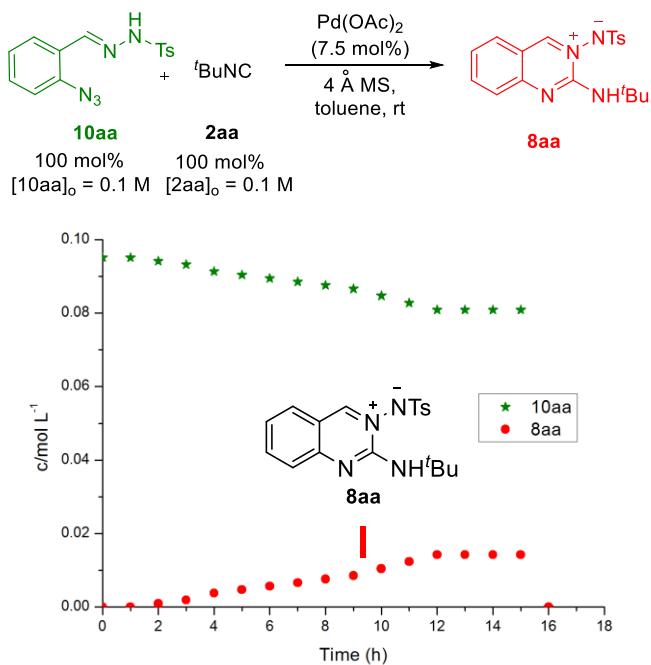


Figure S5. ^1H NMR spectroscopy profile of two CR (**10aa** + **2aa**)

S5.3.6. Result and discussion for kinetic studies

Two plausible routes can be drawn for the generation of the intermediate **8aa**:

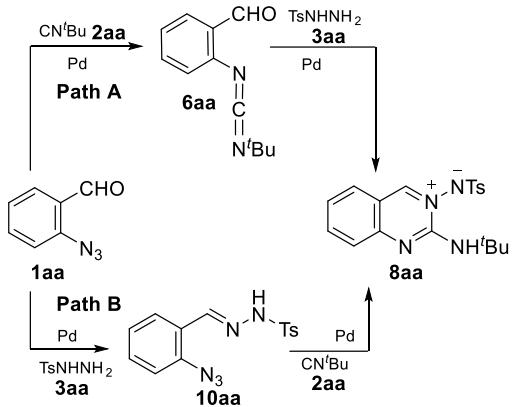
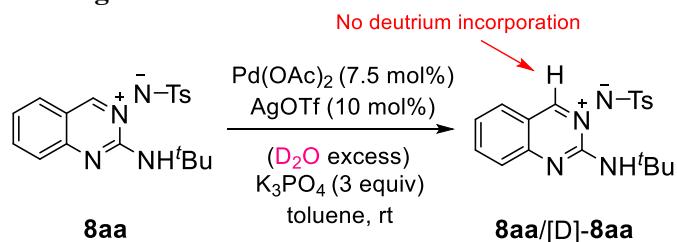


Figure S6: Proposed reaction pathways for the formation of **8aa**

To probe whether the formation of **8aa** could proceed through both path A and path B, or whether one of these pathways was dominating, kinetic studies of 4CR were carried out by monitoring the reaction progress with ^1H NMR spectroscopy, indicated that the formation of **6aa** (**1aa** + **2aa**) was four times faster than the formation of **10aa** (**1aa** + **3aa**). Each coupling step of the process then examined individually by two-component coupling reaction of **1aa** and **2aa** via Pd-catalyzed azide-isocyanide denitrogenative coupling reaction for the formation of **6aa** proceeds at comparable rates in both 4-CRs (5.9 mM min $^{-1}$, the blue line in Fig. S1) and individual 2-CR (9.8 mM min $^{-1}$, Fig. S2). In contrast, condensation of **1aa** and tosyl hydrazide **3aa** for the formation of **10aa** proceeded faster in the individual 2-CR (5.9 mM min $^{-1}$, Fig. S3) compared that of the 4-CR (1.4 mM min $^{-1}$, the green line in Fig. 3a). The reaction of carbodiimide **6aa** with **3aa** exhibited a very rapid conversion to **8aa** (Fig. S4). In addition, the azide-isocyanide denitrogenative coupling of **10aa** and **2aa** was extremely sluggish and unproductive (furnished **8aa** in 15% yields even after prolonging the reaction, Fig. S5). Thus kinetic profiling and isolation of reaction intermediates unequivocally ruled out path B for the formation of **8aa**. The sluggish rate of reaction of the competing concurrent catalytic processes, such as path B, ensured the time-resolution of the relay catalytic process.

S5.4 Experiments on H/D Exchange



8aa (30 mg, 1 equiv), Pd(OAc) $_2$ (2 mg, 7.5 mol%), AgOTf (7.6 mg, 10 mol%), K $_3$ PO $_4$ (51.5 mg, 3.0 equiv), D $_2$ O (5 equiv) and toluene were added to a 20 mL Schlenk tube. The mixture was stirred at rt for 1 h. The reaction mixture was diluted with ethyl acetate (15 mL) and evaporated under vacuum. The crude product was purified by column chromatography to afford the product. These results revealed that no deuterium atom was incorporated at the C-4 position of the intermediate **8aa**. The result indicates that the reaction is not going through C-H activation.

S5.4.1 Result and discussion

For the conversion of the azomethine imine **8aa** to the title compound **5aa**, a possibility of three reaction pathways emerged as depicted in Fig. S7: (1) dipolar cycloaddition; (2) C-H activation alkynylation; (3) direct alkynylation.

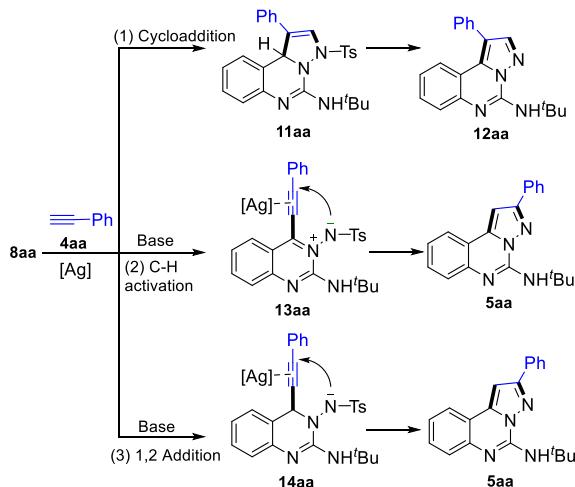
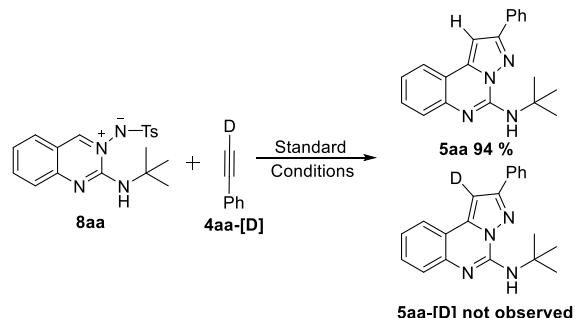


Figure S7: Three plausible pathways for the reaction of azomethine imine **8aa** and alkyne and their control experiments

The regiochemical outcome of 4-CR and H/D scrambling experiment (Section S5.4) ruled out dipolar cycloaddition and C-H activation/alkynylation routes, respectively. Hence control experiments supported a base-mediated direct alkynylation as a possible route for the conversion of the azomethine imine **8aa** to **5aa**.

S5.5 Experiment with labelled substrate

To the mixture of **8aa** (30 mg, 1 equiv), **4aa-[D]** (12.5 mg, 1.5 equiv), $\text{Pd}(\text{OAc})_2$ (2 mg, 7.5 mol%), AgOTf (7.6 mg, 10 mol%) and K_3PO_4 (51.5 mg, 3.0 equiv) in a 20 mL Schlenk tube was added toluene. The mixture was stirred at rt for 1 h. The reaction mixture was diluted with ethyl acetate (15 mL) and evaporated under vacuum. The crude product, so obtained, was purified by column chromatography to afford 24 mg (94%) of the product. No deuterium incorporation was observed, which implicate that AgOTf activated alkyne by converting it to acetylide, which undergoes 1,2 addition across azomethine imine of **8aa**.



No deuterium incorporation in **5aa** was observed when the deuterated substrate (ethynyl-d)-benzene was reacted with **8aa**. This observation supports the formation of acetylide during the reaction.

S5.6: Proposed Reaction Mechanism

Based on experimental studies and literature precedence,² a plausible mechanism for the four-component reaction mediated by three independent catalytic cycles of Pd- and Ag-metals can be proposed as depicted in Fig. S8. **1aa** and **2aa** enters the catalytic cycle A of palladium metal and furnishes a coordination complex **15**. Extrusion of dinitrogen from **15** generates nitrene **16**, which triggers concerted intramolecular isocyanide-transfer on nitrene to produce the carbodiimide **6aa**. The reactive carbodiimide enters catalytical cycle B of palladium metal, where its aldehydic functional group condenses with **3aa** to furnish the hydrazone **19**³ that further undergoes *6-exo-dig* cyclization to produce an air-stable azomethine imine **8aa**. The catalytic cycle C embarks with the formation of acetylide in the presence of base and Ag-salt. The acetylide undergoes 1,2-addition across azomethine imine **8aa** to produce **22**, which converts to **23** by an intramolecular attack of the amide on the alkyne to produce the five-membered ring. Subsequently, detosylative aromatization generates **24**. Protonolysis of the C-Ag bond upon aqueous workup generates the title compound **5aa**, explaining the lack or deuterium incorporation in the labelling studies.

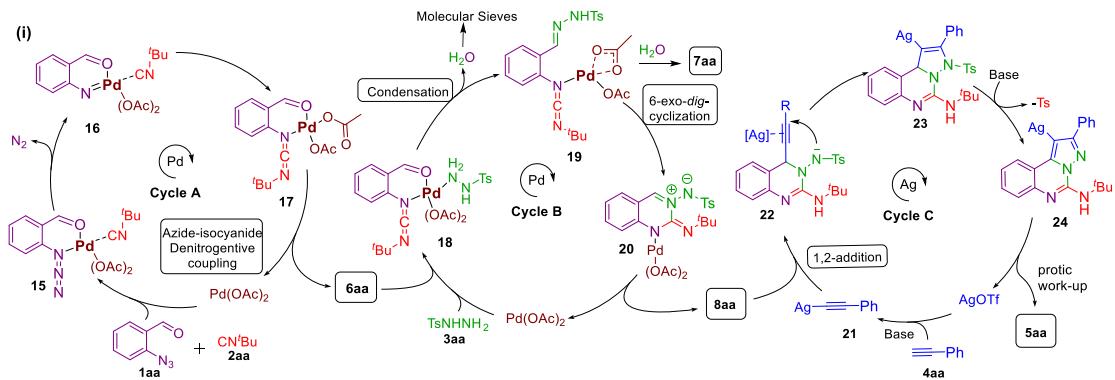


Figure S8: Plausible Reaction mechanism

S6. Experimental biology

6.1 Cell culture and reagents

Cancer cell lines A549 (Lung Cancer), HCT-116 (Colon Cancer), U-87 MG (Human Glioblastoma) were purchased from National Cell Repository at National Centre for Cell Sciences (NCCS, Pune, India). Cancer cells were maintained in Dulbecco's modified Eagle's medium (DMEM) containing 10% heat-inactivated fetal bovine serum (HI-FBS; Gibco, USA) containing penicillin and streptomycin at 100 units/ mL for each. RPMI 1640 and DMEM, Penicillin/ Streptomycin antibiotic solution, phosphate buffer saline, fetal bovine serum media and MTT dye for MTT assay were purchased from HiMedia; z-lyte kinase assay kit-tyr4 peptide for EGFR assay was procured from Invitrogen (catalogue no. PV3193); propidium iodide and RNase for cell cycle analysis were procured from Sigma Aldrich; DMSO, biological grade was purchased from SRL; JC-1 (5,5',6,6'-tetrachloro-1,1',3,3'-tetraethylbenzimidazol-carbocyanine iodide) from Life technologies (Thermo Fisher Scientific); 2,7-dichlorofluorescein diacetate (H2DCFDA) dye from Life technologies (Thermo Fisher Scientific). Automatic cell counter (Invitrogen), incubator (Galaxy, New Brunswick), centrifuge 5430 R (Eppendorf, Germany), laminar air flow (macro scientific works) for aseptic condition, inverted microscope (Magnus, Olympus) for visualization of the cancer cell, BD accuri C6 flow cytometer for cell cycle analysis and microplate reader (BioTek) for reading absorbance in various cell based studies were used.

S6.1.1 Culturing of the cancer cell lines

Cancer cell lines were treated in the DMEM medium. Trypsin was added to detach the cancer cell lines (trypsinization). Centrifugation was done at 1200 rpm at 37 °C for 5 minutes for harvesting the cells. Further, supernatant was disposed and resuspension of the cell pellet was done using 2 mL of the media. The cell number was counted using an automated cell counter. The cells were transferred to fresh media every day.

S6.1.2 Maintenance and sub-culturing of cancer cell lines

The maintenance of cultured cell lines was done in 25 cm² or 75 cm² flasks containing DMEM medium supplemented with 10% Fetal Bovine serum (FBS), 1X antibiotic solution and was incubated at 37 °C containing 5% CO₂, 2% O₂ and 95% humid atmosphere.

S6.2 Evaluation of anticancer activity of the synthesized compounds by MTT Cytotoxicity Assay

A549 (Lung Cancer), HCT-116 (Colon Cancer) and U-87 MG (Human Glioblastoma) cancer cell lines were utilized for the biological studies. Approximately, 10000 cells were seeded in each well of the 96 well plate. The plate was incubated at 37°C with 5% CO₂, 2% O₂ and 95% humid atmosphere for 24 h. At the end of the 24 h, treatments of investigational compounds were given to the cells in triplicate concentrations of 1 μM, 5 μM and 25 μM. The cells were further incubated for 48 h. The media was removed from each well. MTT (1 mg/ml in phosphate-buffered saline—PBS) stock solution was prepared and added as 10 μl/well. This was subsequently incubated in the dark for 4 h. The resulting intracellular precipitate (formazan) thus formed was dissolved in DMSO solution and was subjected to mild shaking so as to dissolve all the formazan crystals. The absorbance was measured at 570 nm using an ELISA plate reader (Bio-Rad, USA). Cytotoxicity was calculated in percentage by using the formula: Cytotoxicity (%) = (OD of control – OD of test/OD of control) x 100.

Among 29 compounds of pyrazolo[1,5-c]quinazolines **5** analyzed, compound **5ea**, **5bc**, **5aa**, **5hb**, **5bd**, **5be**, **5gb**, **5ec**, **5fb**, **5dd** and **5ba** showed activity of < 3 μM against A549 cells. Compounds **5aa**, **5hb**, **5be**, **5gb** and **5eb** exhibited activity of < 3 μM against HCT-116 cells whereas compound **5fb** and **5dd** were found to possess activity of < 3 μM against U-87 MG cells. In tetrahydropyrazolo[1,5-c]quinazoline **26**, 18 compounds were analyzed for

their antiproliferative potential. Compounds **26dd**, **26dc**, **26ab**, **26fa**, **26cc**, **26be** and **26ga** exhibited low micromolar activity of < 3 μ M against A549 cells. Compounds **26dd**, **26cc**, **26aa**, **26ba**, **26ca** and **26ga** were shown to be most active (< 3 μ M) toward HCT-116 cells. Compounds **26ea** and **26ga** exhibited activity of < 3 μ M against U-87 MG cells. The results are compiled in Table S7.

Table S7. Antiproliferative potential of the investigational compounds along with their minimal inhibitory concentrations at various cancer cell lines under study

entry	5	IC ₅₀ (μ M) ^a			entry	5/25	IC ₅₀ (μ M) ^a		
		A549	HCT-116	U-87MG			A549	HCT-116	U-87MG
1.	5aa	1.8 \pm 0.7	2.9 \pm 0.4	4.2 \pm 0.7	24.	5ec	1.4 \pm 0.7	4.8 \pm 0.5	4.3 \pm 0.2
2.	5ab	9.2 \pm 0.3	6.5 \pm 0.9	8.1 \pm 0.3	25.	5ed	14.3 \pm 0.1	17.2 \pm 0.6	13.2 \pm 0.4
3.	5ac	22.1 \pm 0.6	>25	20.7 \pm 0.3	26.	5fb	1.1 \pm 0.3	2.3 \pm 0.3	2.6 \pm 0.3
4.	5ad	13.9 \pm 0.5	7.7 \pm 0.5	7.3 \pm 0.6	27.	5gb	1.1 \pm 0.1	1.4 \pm 0.6	4.7 \pm 0.7
5.	5ae	11.6 \pm 0.9	4.9 \pm 0.6	8.8 \pm 0.5	28.	5hb	1.1 \pm 0.7	2.9 \pm 0.9	6.2 \pm 0.5
6.	5ba	9.6 \pm 0.7	5.3 \pm 0.6	9.3 \pm 0.7	29.	26aa	6.4 \pm 0.3	1.9 \pm 0.7	9.7 \pm 0.9
7.	5bb	22.1 \pm 0.6	7.3 \pm 0.2	5.4 \pm 0.9	30.	26ab	2.8 \pm 0.4	7.9 \pm 0.6	6.3 \pm 0.8
8.	5bc	2.3 \pm 0.9	4.7 \pm 0.5	13.2 \pm 0.8	31.	26ac	8.9 \pm 0.3	6.6 \pm 0.3	17.8 \pm 0.3
9.	5bd	1.4 \pm 0.3	5.3 \pm 0.7	3.6 \pm 0.7	32.	26ba	4.1 \pm 0.5	2.6 \pm 0.9	3.4 \pm 0.8
10.	5be	1.6 \pm 0.7	2.7 \pm 0.5	3.1 \pm 0.8	33.	26bc	1.4 \pm 0.9	3.4 \pm 0.8	7.6 \pm 0.8
11.	5cb	14.3 \pm 0.3	3.1 \pm 0.3	<25	34.	26be	1.5 \pm 0.9	3.7 \pm 0.8	3.9 \pm 0.4
12.	5cc	16.2 \pm 0.8	22.8 \pm 0.9	19.3 \pm 0.5	35.	26bf	17.2 \pm 0.7	9.2 \pm 0.7	<25
13.	5cd	2.2 \pm 0.4	7.4 \pm 0.5	5.3 \pm 0.9	36.	26ca	8.4 \pm 0.8	2.3 \pm 0.6	11.8 \pm 0.8
14.	5ce	9.3 \pm 0.7	6.5 \pm 0.4	9.3 \pm 0.8	37.	26cb	12.2 \pm 0.4	7.6 \pm 0.7	9.2 \pm 0.4
15.	5cf	21.6 \pm 0.8	16.3 \pm 0.5	11.4 \pm 0.4	38.	26cc	1.8 \pm 0.3	2.2 \pm 0.5	5.9 \pm 0.9
16.	5cg	18.6 \pm 0.9	11.2 \pm 0.4	9.8 \pm 0.3	39.	26cd	19.6 \pm 0.6	8.7 \pm 0.8	<25
17.	5ch	12.2 \pm 0.6	7.3 \pm 0.4	<25	40.	26ce	3.8 \pm 0.7	4.2 \pm 0.7	7.5 \pm 0.9
18.	5da	2.6 \pm 0.8	4.7 \pm 0.9	6.7 \pm 0.9	41.	26dc	1.8 \pm 0.7	7.6 \pm 0.7	8.4 \pm 0.3
19.	5db	23.2 \pm 0.9	13.4 \pm 0.3	<25	42.	26dd	1.3 \pm 0.5	1.8 \pm 0.7	3.9 \pm 0.9
20.	5dc	11.1 \pm 0.7	17.2 \pm 0.5	<25	43.	26de	17.1 \pm 0.5	9.7 \pm 0.8	16.7 \pm 0.7
21.	5dd	1.4 \pm 0.2	1.6 \pm 0.5	2.8 \pm 0.7	44.	26ea	1.3 \pm 0.5	1.7 \pm 0.5	2.3 \pm 0.9
22.	5ea	1.4 \pm 0.4	3.9 \pm 0.9	6.3 \pm 0.6	45.	26fa	1.6 \pm 0.3	6.4 \pm 0.3	3.9 \pm 0.8
23.	5eb	3.1 \pm 0.6	2.9 \pm 0.7	7.2 \pm 0.6	46.	26ga	1.1 \pm 0.5	1.4 \pm 0.8	2.2 \pm 0.3

^aValues are derived from averaging three independent experiments and each experiment was done in triplicate. Based upon the relevant MTT data four compounds **5fb**, **5dd**, **26ea** and **26ga** were selected for further biological studies.

S6.2.1 Structure-Activity Relationship of cytotoxic activity of pyrazolo[1,5-*c*]quinazolines

1. Substitutions on C-2 of pyrazoles ring were not well tolerated
2. Substrates with electron withdrawing substituents, such as ester and cyano, on C-1 of pyrazoles were the most potent, and substrates with bulky groups on the same position were inactive
3. Substituents on benzene ring are well tolerated

S6.3 Human Peripheral Blood Mononuclear Cells (HPBMCs) culturing and treatment by investigational molecules to assess the cytotoxicity of investigational molecules

In order to perform selectivity of investigational molecules toward cancer cells, we performed HPBMCs assay. The assay was strictly performed as per the protocol no. CUPB/cc/14/IEC/4483 approved by Institutional Ethics Committee of Central University of Punjab, Bathinda. The protocol used was standard operating procedure,

provided by Institutional Ethics Committee of Central University of Punjab according to guidelines issued by Indian Council of Medical Research (ICMR), Govt. of India.

Fresh Blood was drawn from healthy individual and HPBMCs were isolated from whole blood by lysing RBCs. The cells were then suspended in RPMI media supplemented with 10% fetal bovine serum (FBS), 1% antibiotic solution and were cultured and treated as per the MTT assay procedure discussed previously. Investigational compounds **5fb**, **5dd**, **26ea** and **26ga** do not cause any significant cytotoxicity towards normal HPBMCs even at the highest concentration of 25 μ M, implicating a pattern of selectivity to cancer cells (Figure S9).

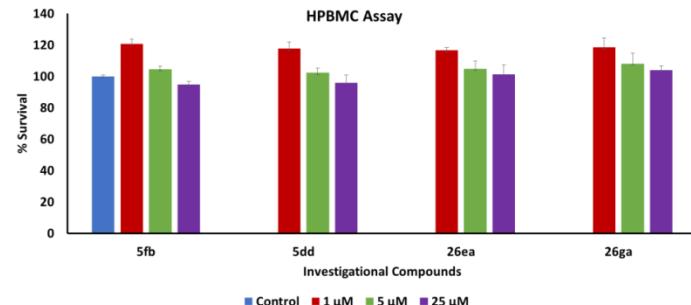


Figure S9: Graph depicting percentage survival of human peripheral blood mononuclear cells in response to treatment with selected investigational compounds for 48 h at concentrations of 1, 5 and 25 μ M. Data is expressed as mean values \pm S.D. of three independent experiments.

S6.4 Pyrazolo[1,5-*c*]quinazoline as EGFR inhibitors

S6.4.1. Structure Activity Relationship studies

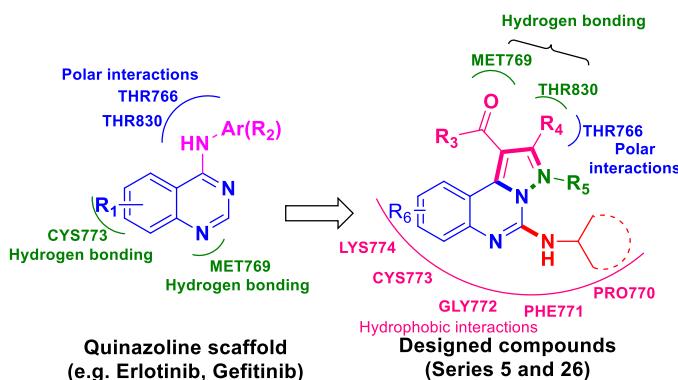


Figure S10: Design strategy for EGFR inhibitors (Series **5** and **26**, Scheme 2 and 3)

Owing to their structural resemblance to the quinazoline scaffold of gefitinib and erlotinib, the investigational compounds **5dd**, **5fb**, **26ea** and **26ga** were envisaged to posses EGFR inhibitory activity. EGFR is over-expressed in almost every cancer types and is a validated drug target. Small molecules tend to occupy the ATP binding kinase domain of EGFR and thus alter the cell signalling cascade. The quinazoline-based scaffolds are found to reside in the catalytic EGFR binding pocket, where the N1 atom of the quinazoline ring acts as a hydrogen bond acceptor in an interaction with Met-769, the N3 atom interacts with Thr-830 through a bridging water molecule, and the arylamino-substituents occupies the normally empty hydrophobic pocket within the active region of EGFR receptor. The pyrazole fusion was hypothesized to occupy the extra space in ATP binding cavity available in the active site of EGFR and may be exploited for extra affinity of quinazoline based EGFR inhibitors. In addition, a bulkier alkylamino substituent at C2 might reduce metabolic degradation of quinazoline nucleus and could promote fitting of the bulkier group in the hydrophobic pocket, thereby enhancing the overall stability of protein-ligand complex (figure S10).

S6.4.2 EGFR inhibitory assay

The Z-LYTE® kinase assay kit-tyr4 peptide (catalogue no. PV3193) was used for the assessment of EGFR Inhibitory potential of the investigational compounds **5fb**, **5dd**, **26ea** and **26ga**. The assay relies upon an enzymatic reaction through which auto phosphorylation and signalling activity of the EGFR is measured. The assay was done by our previous standardized protocol.⁴ Briefly, the inhibitory effect of compounds on EGFR was measured spectrophotometrically at 400, 445 and 520 nm, respectively. Erlotinib was used as a positive control. The reagents

were prepared according to catalogue no. PV3193, the assay mixture comprised of 133 µL kinase buffer, 0.5 µL kinase peptide mixtures, 0.5 µL phosphopeptide mixtures and 0.5 µL ATP test sample solution. The assay plates were prepared in triplicates. The investigational compounds **5fb**, **5dd**, **26ea** and **26ga** were serially diluted at a concentration of 250 nM, 750 nM, 1 µM, 5 µM and 25 µM in DMSO (~4%), were mixed and incubated at room temperature for 1 h. After incubation, it was treated with 5 µL of development solution and kept in the dark for another 1 h. The reaction was quenched with the addition of 5 µL stop solution. The emission ratio (Coumarin Emission (445 nm)/fluorescein emission (520 nm) signifying the extent of phosphorylation was done using the given equation.

$$\% \text{ phosphorylation} = 1 - \frac{(Emission\ ratio \times F100\%) - C100\%}{(C0\% - C100\%) + [Emission\ ratio (F100\% - F0\%)]}$$

Where C0% = Average coumarin emission signal of the 100% Phos. Control; C100% = Average coumarin emission signal of the 0% Phos. Control; F100% = Average Fluorescein emission signal of the 100% Phos. Control; F0% = Average Fluorescein emission signal of the 0% Phos. Control

Table S8. EGFR inhibitory activity of the investigational compounds

Investigational compounds	IC ₅₀ ^a Cancer Cell lines (µM)			EGFR Inhibitory Activity IC ₅₀ ^a (nM)
	A549	HCT-116	U-87MG	
5dd	1.4 ± 0.2	1.6 ± 0.5	2.8 ± 0.7	639.4 ± 0.34
5fb	1.1 ± 0.3	2.3 ± 0.3	2.6 ± 0.3	235 ± 0.92
26ea	1.3 ± 0.5	1.7 ± 0.5	2.3 ± 0.9	3060 ± 0.65
26ga	1.1 ± 0.5	1.4 ± 0.8	2.2 ± 0.3	625 ± 0.74
Erlotinib	-	-	-	423 ± 0.89

^aValues are derived from averaging three independent experiments and each experiment was done in triplicate

To our delight, **5fb** was found to be the most potent even with respect to the standard drug, erlotinib, while **5dd** and **26ga** showed comparable activity in the nanomolar range. To further validate these results, molecular docking studies were performed. Refer section S7 for details.

S6.5 Measurement of intracellular reactive oxygen species (ROS)

Apoptosis is linked to the increase in ROS levels and the loss of the mitochondrial membrane potential ($\Delta\Psi M$).⁵ The measurement of intracellular production of ROS was done using 2',7'-dichlorofluorescein diacetate (H2DCFDA) dye, a stable non-polar compound that readily diffuses into the cell and converted to 2',7'-dichlorofluorescin (DCF) upon cleavage of acetate groups by intracellular esterases and oxidation by peroxidase. Interestingly, the fluorescent intensity obtained is directly proportional to the amount of ROS produced by the cell. The lung cancer cell line A549 was treated with the investigational compounds **5fb**, **5dd**, **26ea** and **26ga** for 48 h. Then, cells were washed with PBS and incubated in H2DCFDA at 37°C for half an hour. Cells were lysed in alkaline solution and centrifuged for 10 min. The supernatant was transferred to a 96-well plate and fluorescence was measured at 5 and 25 µM concentration at Ex/Em: ~492–495/517–527 using microplate reader. Results were calculated as fluorescence intensity relative to control cells. A significant time- and dose-dependent increase in ROS level was observed with a positive control, erlotinib (Figure S11)

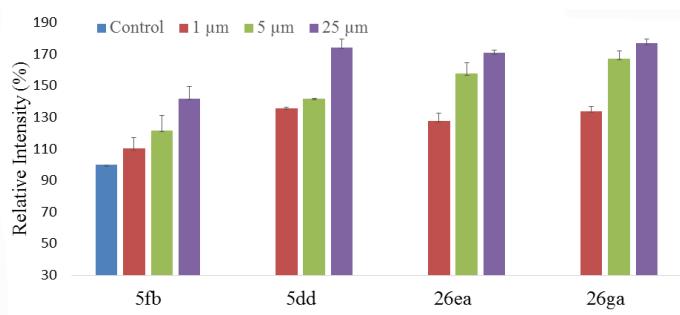


Figure S11: H2DCFDA based assay to measure intracellular ROS

S6.6 Determination of mitochondrial membrane potential (μ M)

A549 cells were seeded in 96 well plated and treated with 1, 5 and 25 mM concentrations of **5fb**, **5dd**, **26ea** and **26ga** for 48 h. Subsequently the cells were incubated with JC-1 dye (10 mL) (5,5',6,6'-tetrachloro-1,1',3,3'tetraethylbenzimi-dazolylcarbocyanine iodide) for 30 min. Cells were washed with PBS and quantitatively analyzed under fluorescent microplate reader (emission at 527 and 590). Cells pre-treated with **5fb** were analyzed under confocal microscopy (Figure S12). JC-1 dye selectively enters mitochondria and reversibly changes colour from red to green as μ M decreases (OD590/OD527 ratio). The results showed that all the compounds lead to imbalance OD590/OD527 ratios and thus altered the mitochondrial membrane (Figure S13).

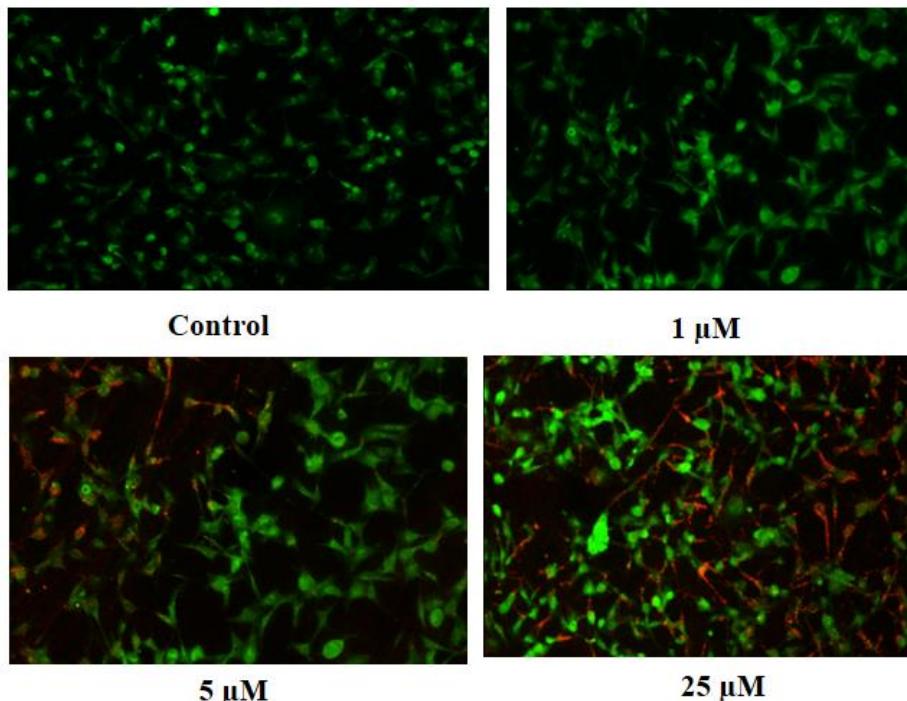


Figure S12: Confocal microscopy of cells treated with **5fb** and JC1 dye

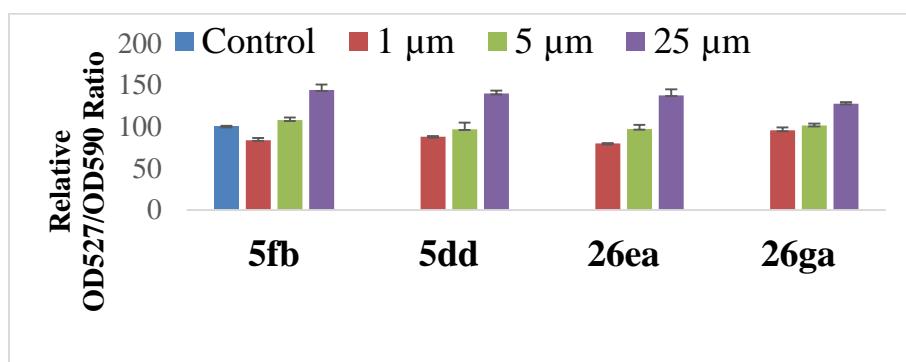


Figure S13: JC-1 dye-based assay to measure a change in mitochondrial membrane potential on A549 cancer cell line treated **5fb**, **5dd**, **26ea** and **26ga**.

S6.7 Cell apoptosis assay using Propidium Iodide

A549 cancer cells, pretreated with compound **5fb** and **26ga** at a concentration of 5 and 25 μ M for 24 h, were transferred in a manner to obtain 10^6 cells per tube. The cells were further centrifuged at 1200 rpm for 5 min and washed with 1X PBS. Thereafter cells were fixed using chilled ethanol and incubated for 3 h at -20 °C. Post 3 h, cells were subjected to centrifugation at 2500 rpm and washed once with 1 X PBS. Each tube was mixed with 50 μ L of propidium iodide (Sigma Aldrich) and 50 μ L Ribonuclease (10 mg/mL solution in water), and incubated for 30 minutes at room temperature in the dark. The DNA content was then measured using BD Accuri C6 flow cytometer. For the cells treated with **5fb**, a substantial rise in the G1 phase of cells, 32.5% and 43.77% at concentration of 5 and 25 μ M respectively, was observed compared to that of control (23.1%). Cells treated with

26ga exhibited similar results with profound G1 phase arrest at 36.32% and 40.65% at concentration of 5 and 25 μM respectively compared with the control (Figure S14-S15).

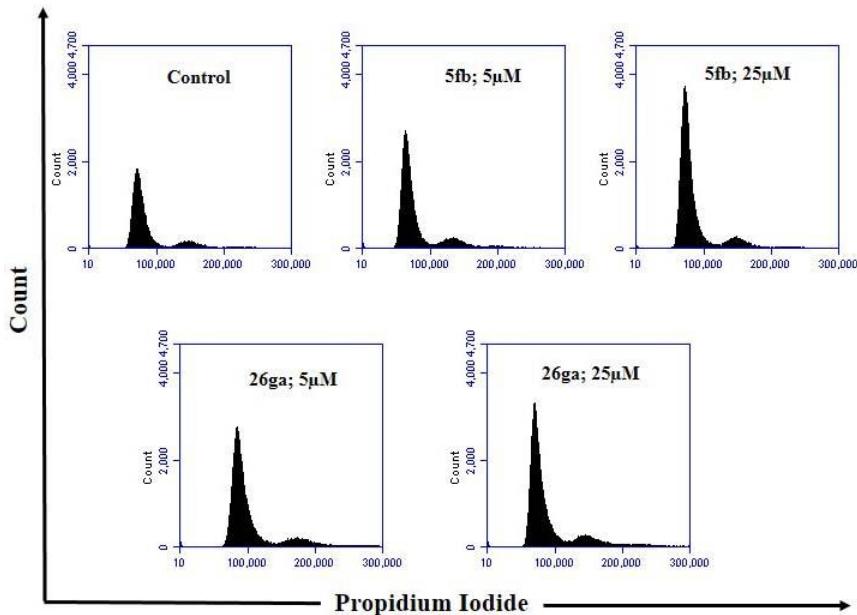


Figure S14: Cell cycle analysis using flow cytometry of A549 cancer cell line treated with **5fb** and **26ga**.

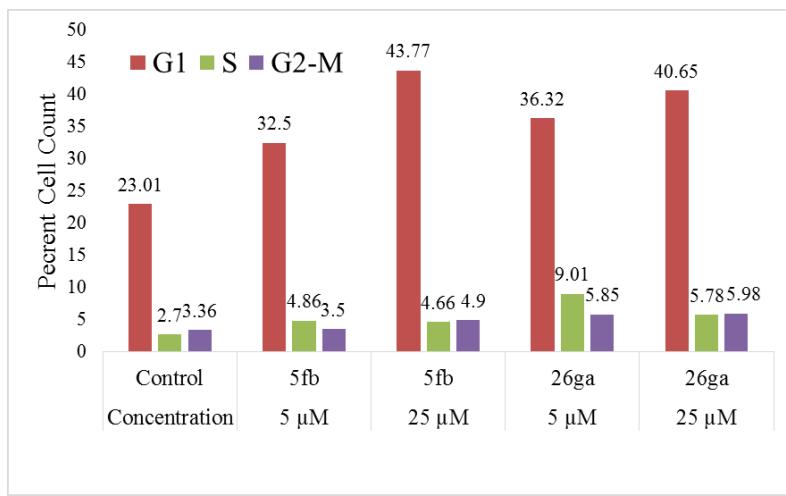


Figure S15: The bar graph represents the percent cell count (DNA) at various stages of cell cycle.

S7. Molecular Docking Studies

Molecular docking studies were performed using GLIDE module of Schrodinger Maestro Licensed software (11.1 version). The protein with PDB Id- 1M17 (2.6 \AA) was retrieved from the protein data bank (<https://www.rcsb.org/pdb/home/home.do>). Initial protein refinement involving bond orders and addition of hydrogen to the side chains was done using protein preparation wizard. Further refinement involved optimization and minimization using OPLS3 force field. Ligands were sketched using 2D sketcher implemented in Maestro 1 and Ligprep was used for ligand optimization and minimization. Further, receptor grid generation involved defining active site in the protein structure for which co-crystallized ligand co-ordinates were used. The bound erlotinib formed the centroid of the grid box. Docking studies were performed in Xtra Precision (XP) mode. Validation of docking protocol was performed by comparing the conformation of docked erlotinib against the conformation of co-crystallized erlotinib. The RMSD of the conformations was found to be 1.27 \AA and the residue interaction was found to be similar to the native ligand.

In order to investigate the underlying binding pattern, potential inhibitors **5fb**, **5dd**, **26ea** and **26ga** showing promising biological activity (section S6.4) were docked in the generated grid along with erlotinib. The resultant docked conformations resulted in binding affinity scores and residue interaction pattern (Table S9). **5fb** exhibited comparable binding to the wild type EGFR ATP binding site, and similar to erlotinib. It was observed that

compound **5fb** possessed higher affinity as compared to **5dd**, **26ea** and **26ga** despite being more active in the biological evaluation. The binding pattern of the highest active compound **5fb** figuring out the important interactions are represented in Figure S16.

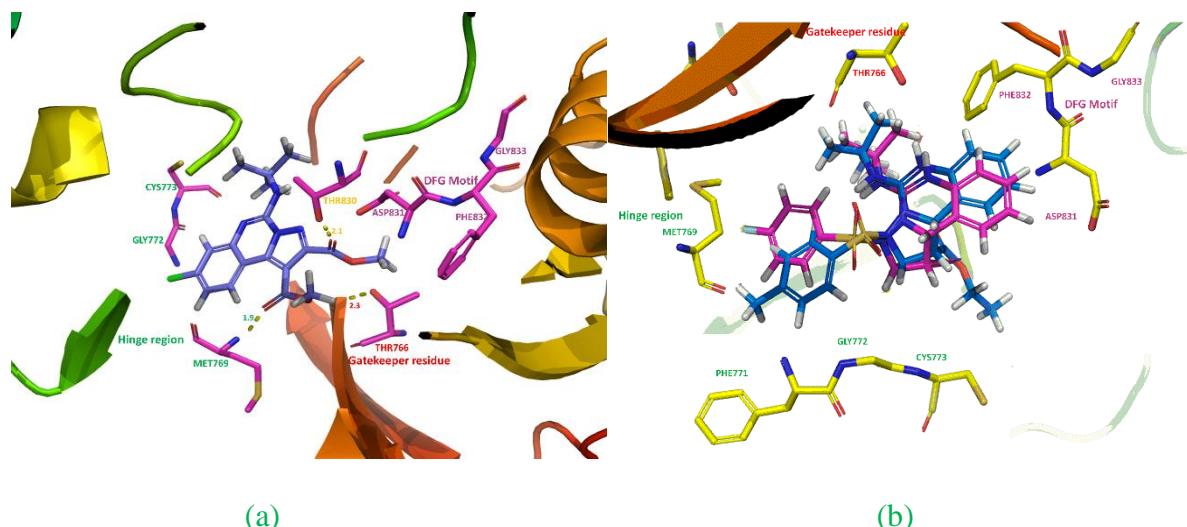


Figure S16: Binding pattern representation of (a) the most active compound, **5fb** (blue colour), and (b) **26ea** (blue colour) and **26ga** (pink colour) with the key amino acid interactions

Table S9. Representation of selected compounds docking scores and their important interaction within the catalytic domain of EGFR

Compound	Docking score	Important interacting with the residues				
		MET769	THR766	THR830	ASP831	CYS773
5fb	-7.793	+	+	+	-	-
5dd	-4.462	-	-	-	+	-
26ga	-3.806	-	-	-	+	+
26ea	-3.509	-	-	-	+	-

Though **5dd**, **26ea** and **26ga** binds to the ATP binding site of EGFR, no hydrogen bonding interactions with MET769 and CYS773, essential amino acid residues important for the EGFR inhibitory activity, are observed. This important interaction of hinge residue MET769 is observed in **5fb**. The carbonyl group of the acetate group attached to **5fb** forms hydrogen bond with the -NH of the hinge region residue MET769 of the active site. Additionally, hydrogen bonding is observed between the THR830 and another carbonyl of acetate group and hydrogen bonding of methyl with oxygen atom of THR766. These additional interactions might have enhanced the overall affinity of **5fb** with the ATP binding domain of EGFR, exhibiting a potent activity against the kinase.

S8. Crystallographic Information

S8.1 5ad

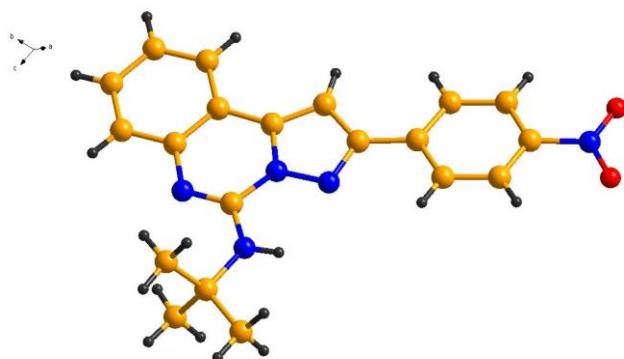


Figure S17. X-Ray crystal structure of **5ad**. (Carbon, Orange; Nitrogen, Blue; Oxygen, Red; Hydrogen, Black)

Table S10: Crystal data and structure refinement for **5ad**

Identification code	CCDC Number 1566282
Empirical formula	C ₄₁ H ₄₀ Cl ₂ N ₁₀ O ₄
Formula weight	807.73
Temperature/K	296.15
Crystal system	Monoclinic
Space group	C2/c
a/Å	27.5324(10)
b/Å	9.3673(3)
c/Å	19.5352(7)
α/°	90.00
β/°	128.8920(10)
γ/°	90.00
Volume/Å ³	3921.4(2)
Z	4
ρ _{calcg} /cm ³	1.368
μ/mm ⁻¹	0.222
F(000)	1688.0
Crystal size/mm ³	0.3 × 0.2 × 0.2
Radiation	MoKα ($\lambda = 0.71073$)
2Θ range for data collection/°	9.2 to 51
Index ranges	-33 ≤ h ≤ 33, -11 ≤ k ≤ 11, -23 ≤ l ≤ 23
Reflections collected	23824
Independent reflections	3307 [R _{int} = 0.0424, R _{sigma} = 0.0229]
Data/restraints/parameters	3307/0/261
Goodness-of-fit on F ²	1.325
Final R indexes [I>=2σ (I)]	R ₁ = 0.0551, wR ₂ = 0.1576
Final R indexes [all data]	R ₁ = 0.0660, wR ₂ = 0.1722
Largest diff. peak/hole / e Å ⁻³	0.56/-0.44

S8.2 8aa

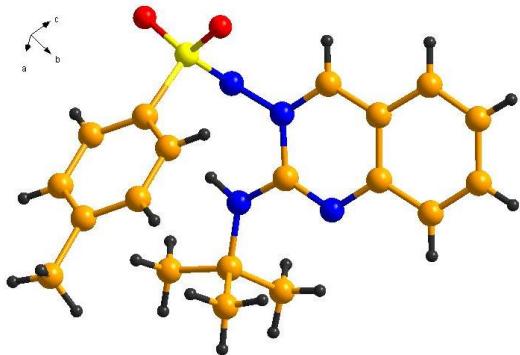


Figure S18. X-Ray crystal structure of compound **8aa**. (Carbon, Orange; Nitrogen, Blue; Oxygen, Red; Hydrogen, Black)

Table S11: Crystal data and structure refinement for **8aa**

Identification code	CCDC Number 1566284
Empirical formula	C ₁₉ H ₂₂ N ₄ O ₂ S
Formula weight	370.48
Temperature/K	296(2)
Crystal system	Triclinic
Space group	P-1
a/Å	7.8633(6)
b/Å	9.3267(7)
c/Å	13.5742(10)
α/°	84.166(4)
β/°	89.184(4)
γ/°	79.543(4)
Volume/Å ³	973.90(13)
Z	2
ρ _{calc} g/cm ³	1.2633
μ/mm ⁻¹	0.186
F(000)	392.4
Crystal size/mm ³	0.2 × 0.2 × 0.15
Radiation	Mo Kα ($\lambda = 0.71073$)
2Θ range for data collection/°	3.02 to 51
Index ranges	-10 ≤ h ≤ 11, -13 ≤ k ≤ 12, -19 ≤ l ≤ 19
Reflections collected	22709
Independent reflections	3626 [R _{int} = 0.12, R _{sigma} = 0.1059]
Data/restraints/parameters	3626/0/238
Goodness-of-fit on F ²	1.130

Final R indexes [I>=2σ (I)] R₁ = 0.0858, wR₂ = 0.2398

Final R indexes [all data] R₁ = 0.1076, wR₂ = 0.2778

Largest diff. peak/hole / e Å⁻³ 0.69/-1.46

S8.3. 7aa

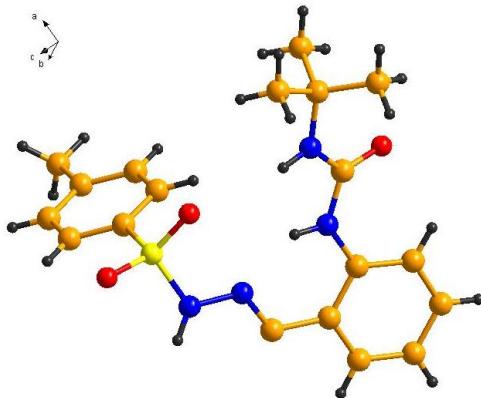


Figure S19. X-Ray crystal structure of 7aa. (Carbon, Orange; Nitrogen, Blue; Oxygen, Red; Hydrogen, Black)

Table S12. Crystal data and structure refinement for 7aa

Identification code	CCDC Number 1566285
Empirical formula	C ₁₉ H ₂₃ N ₄ O ₃ S
Formula weight	387.49
Temperature/K	100(2)
Crystal system	Monoclinic
Space group	C2/c
a/Å	34.877(2)
b/Å	9.0455(6)
c/Å	12.9419(9)
α/°	90
β/°	105.171(3)
γ/°	90
Volume/Å ³	3940.7(5)
Z	8
ρ _{calc} g/cm ³	1.3061
μ/mm ⁻¹	0.191
F(000)	1641.7
Crystal size/mm ³	0.2 × 0.2 × 0.2
Radiation	Mo Kα (λ = 0.71073)
2Θ range for data collection/°	2.42 to 50.98
Index ranges	-46 ≤ h ≤ 46, -10 ≤ k ≤ 12, -17 ≤ l ≤ 17

Reflections collected	29018
Independent reflections	3651 [$R_{\text{int}} = 0.0418$, $R_{\text{sigma}} = 0.0253$]
Data/restraints/parameters	3651/0/248
Goodness-of-fit on F^2	1.048
Final R indexes [$I \geq 2\sigma(I)$]	$R_1 = 0.0502$, $wR_2 = 0.1728$
Final R indexes [all data]	$R_1 = 0.0623$, $wR_2 = 0.2088$
Largest diff. peak/hole / e Å ⁻³	0.95/-0.71

S8.4. 9aa

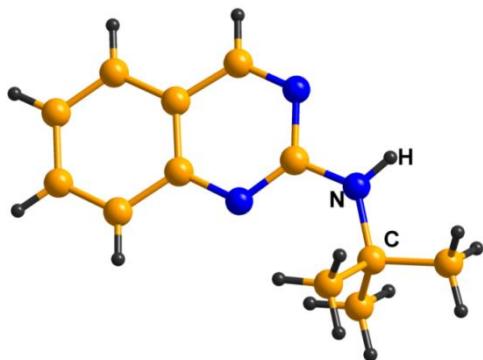


Figure S20 X-Ray crystal structure of compound **9aa**. (Carbon, Orange; Nitrogen, Blue; Oxygen, Red; Hydrogen, Black)

Table S13. Crystal data and structure refinement for **9aa**.

Identification code	CCDC number 1566113
Empirical formula	C ₁₂ H ₁₅ N ₃
Formula weight	201.27
Temperature/K	100(2)
Crystal system	Monoclinic
Space group	P2 ₁ /n
a/Å	8.3134(8)
b/Å	12.5189(13)
c/Å	11.0332(12)
α/°	90
β/°	107.377(3)
γ/°	90
Volume/Å ³	1095.9(2)
Z	4
ρ _{calc} g/cm ³	1.220
μ/mm ⁻¹	0.075
F(000)	432.0

Crystal size/mm ³	0.35 × 0.22 × 0.15
Radiation	MoKα ($\lambda = 0.71073$)
2Θ range for data collection/°	5.428 to 50.042
Index ranges	-9 ≤ h ≤ 9, -14 ≤ k ≤ 14, -13 ≤ l ≤ 12
Reflections collected	8589
Independent reflections	1924 [$R_{\text{int}} = 0.0494$, $R_{\text{sigma}} = 0.0447$]
Data/restraints/parameters	1924/0/143
Goodness-of-fit on F^2	1.032
Final R indexes [$I \geq 2\sigma(I)$]	$R_1 = 0.0446$, $wR_2 = 0.0950$
Final R indexes [all data]	$R_1 = 0.0705$, $wR_2 = 0.1060$
Largest diff. peak/hole / e Å ⁻³	0.15/-0.23

S8.5. 26be

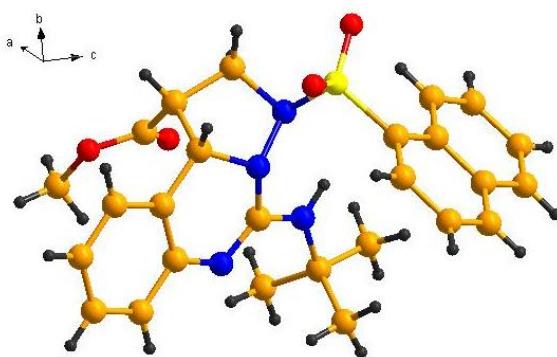


Figure S21. X-Ray crystal structure of **26be**. (Carbon, Orange; Nitrogen, Blue; Oxygen, Red; Hydrogen, Black)

Table S14. Crystal data and structure refinement for **26be**

Identification code	CCDC Number 1566283
Empirical formula	C ₂₆ H ₂₈ N ₄ O ₄ S
Formula weight	492.58
Temperature/K	296.15
Crystal system	Monoclinic
Space group	C2/c
a/Å	29.7473(8)
b/Å	10.1292(3)
c/Å	18.7181(5)
α/°	90.00
β/°	118.7280(10)
γ/°	90.00
Volume/Å ³	4945.8(2)

Z	8
ρ_{calc} g/cm ³	1.323
μ/mm^{-1}	0.171
F(000)	2080.0
Crystal size/mm ³	0.2 × 0.2 × 0.15
Radiation	MoKα ($\lambda = 0.71073$)
2Θ range for data collection/°	8.84 to 47.18
Index ranges	-33 ≤ h ≤ 33, -11 ≤ k ≤ 11, -21 ≤ l ≤ 21
Reflections collected	45616
Independent reflections	3519 [$R_{\text{int}} = 0.0474$, $R_{\text{sigma}} = 0.0203$]
Data/restraints/parameters	3519/0/320
Goodness-of-fit on F^2	1.282
Final R indexes [$I \geq 2\sigma(I)$]	$R_1 = 0.0515$, $wR_2 = 0.1518$
Final R indexes [all data]	$R_1 = 0.0605$, $wR_2 = 0.1621$
Largest diff. peak/hole / e Å ⁻³	1.04/-0.24

S9. Pharmacological activity and literature search for the synthesis of pyrazolo[1,5-c]quinazolines

S9.1: Pharmacological activity of pyrazolo[1,5-c]quinazolines reported in the literature

Asproni et al⁶ have reported the synthesis and SAR studies of the pyrazolo[1,5-c]quinazolines as novel potent phosphodiesterase 10A(PDE10A) inhibitors. In this series, the compound **1** showed the highest affinity for PDE10A enzyme with $IC_{50} = 16\text{nm}$. Pae et al⁷ employed structure-based virtual screening of pyrazolo quinazoline derivatives against Eg5 inhibitors. These derivatives exhibit growth inhibition in proliferation assays and induce monoastral spindles in cells with IC_{50} value of $6.3\text{ }\mu\text{m}$ (Fig.S22, 2). Kumar et al⁸ have rationally designed and synthesized some 5,6-dihydropyrazolo/ pyrazolo[1,5-c]quinazoline derivative and evaluated these compounds for *in vitro* xanthine oxidase inhibitory activity (Fig. S22, 3-5). Cecchi et al⁹ have reported the synthesis and binding activity of the 2-substituted pyrazolo[1,5-c]quinazolines as optimal site of benzodiazepine receptor activity (Fig. S22, 6-7). Varano et al^{10,11} described the binding affinity of pyrazolo[1,5-c]quinazolines towards AMPA (2-amino-3-(3-hydroxy-5-methylisoxazol-4-yl)-propionic acid), Gly/ NMDA (Glycine/N-methyl-D-aspartic acid) and KA (Kainic acid) receptor and also performed their docking studies (Fig.S22, 8)

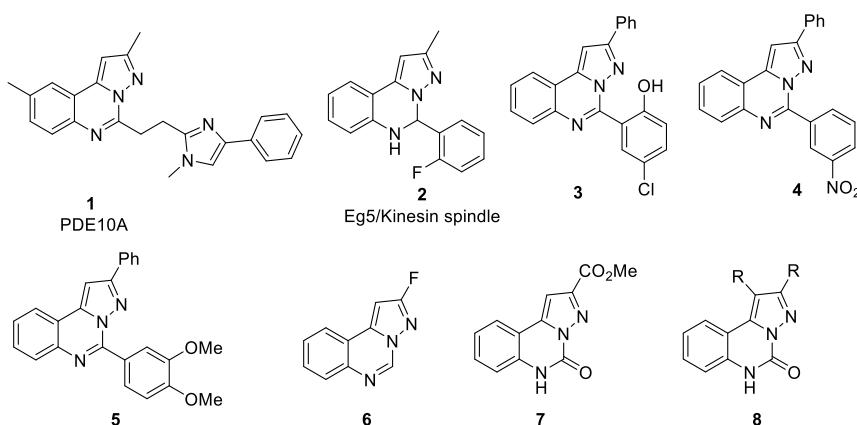


Figure S22: Pharmacological importance of pyrazolo[1,5-c]quinazoline

S9.2: Synthesis of pyrazolo[1,5-c]quinazolines reported in the literature

we envisaged a potential to develop a new higher-order multicomponent reaction by combining four versatile privileged synthons: 2-azidobenzaldehyde, isocyanide, sulfonyl hydrazides and alkynes. Herein, we report the realization of a four-component reaction through a Pd(II)/Ag(I) binary catalytic system for the synthesis of diverse pyrazolo[1,5-c]quinazolines (Fig. S23, Scheme 13) that are known to potential biological interest. Conventionally, pyrazolo[1,5-c]quinazolines are synthesized in a multi-step fashion by condensation of 2-pyrazoloanilines with triphosgene (Fig. S23, Scheme 1),^{9,12} glyoxylic acid (Fig. S23, Scheme 2),¹⁰ aldehydes (Fig. S23, Scheme 3),¹³ chloroacetylchloride (Fig. S23, Scheme 4)⁵ and cyanamide (Fig. S23, Scheme 5),¹⁴ Cu-mediated cyclization of 5-(2-bromoaryl)-1H-pyrazoles with carbonyl compounds in aqueous ammonia (Fig. S23, Scheme 6),¹⁵ reaction of 3-diazo-1,3-dihydro-2H-indol-2-one derivatives with either enaminones (Fig. S23, Scheme 7)¹⁶ or activated alkynes (Fig. S23, Scheme 8),¹⁷ two-component [3+2] cycloaddition reactions of *N*-aminoquinazoliumylides with olefins (Fig. S23, Scheme 9),¹⁸ 1,3-dipolar cycloaddition of *N*-unprotected ylideneoxindoles and dimethyl diazomethylphosphonate anion generated *in situ* from Bestmann-Ohira reagent (Fig. S23, Scheme 10),¹⁹ Au-catalysed intramolecular bicyclization of N-propargylic sulfonylhydrazones (Fig. S23, Scheme 11),²⁰ and Cu-catalyzed two-step reaction of 1-(2-bromophenyl)-3-alkylprop-2-yn-1-ones, hydrazines and amidines (Fig. S23, Scheme 12).²¹ To the best of our knowledge, there is no report of the multicomponent assembly of pyrazolo[1,5-c]quinazolines in one-pot to date.

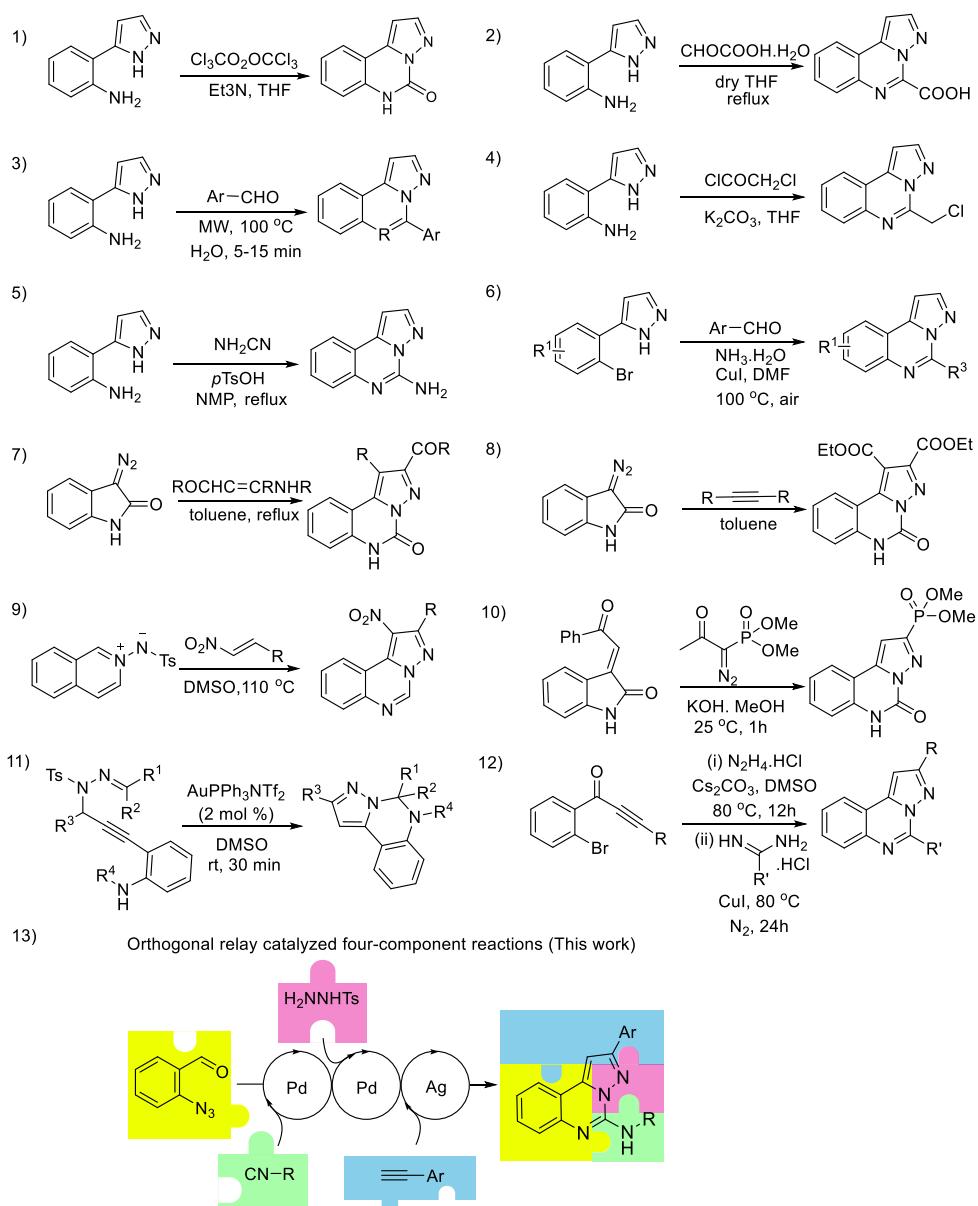
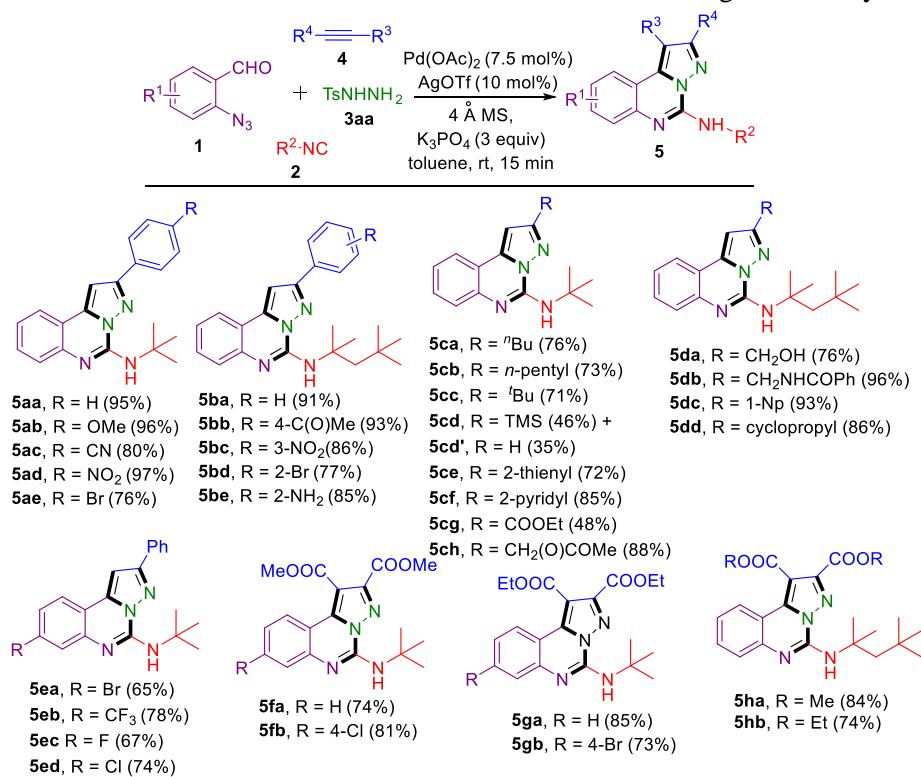


Figure S23: Various methods reported in the literature for the synthesis of pyrazolo[1,5-c]quinazolines.

Results and Discussion

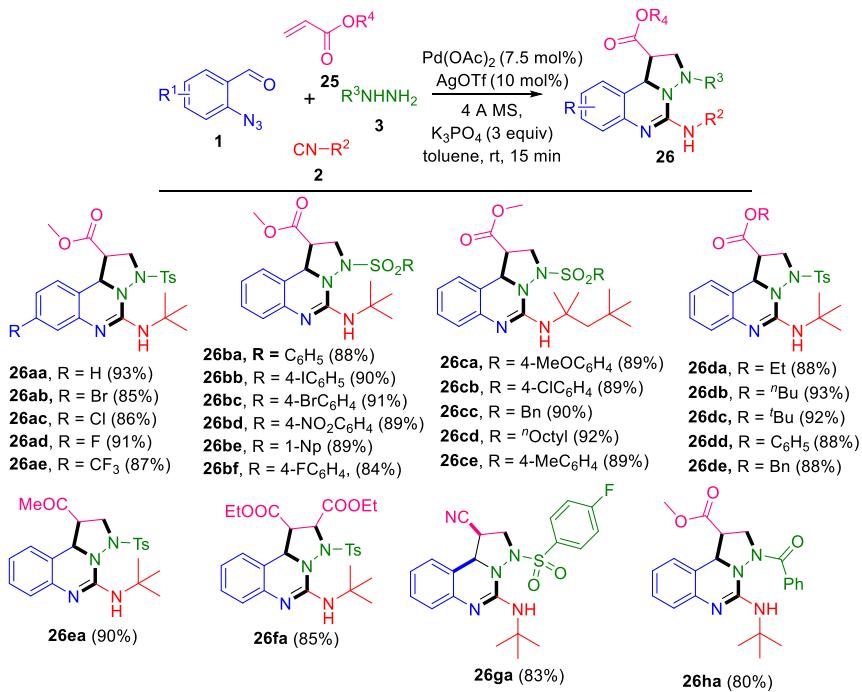
S10. Construction of the diversity-rich library of pyrazolo[1,5-c]quinazolines

The overall scope of this reaction was found to be wide (Scheme 2). Both aromatic and aliphatic terminal alkynes and electron deficient internal alkynes participated in 4CR. The chemical integrity of reactive functional groups such as cyano, ketone, ester and halides (Br and I) remained unperturbed in the standard reaction condition providing a chemical handle for further transformation. Fluorinated substitutions in **5eb**, **5ec** and **5fc** would contribute in enhancing their pharmacodynamics and pharmacokinetic properties. Interestingly, trimethylsilylacetylene participated the 4-CR to furnish **5cd** up to 46% isolated yield and a desilylated product **5cd'** was also observed with 35% isolated yield. Heteroarylalkynes such as 2-thienyl, 2-pyridyl were also subjected to the 4CR reaction to afford **5ce** and **5cf** respectively in high yields. Notably, electron deficient 2-azidobenzadehydes were compatible to deliver 4CR adduct. In contrast, electron rich 2-azidobenzadehydes did not afford desired product **5** due to low reactivity of corresponding carbodiimide intermediate **6**. The investigation underscores the high tolerance of the 4CR reaction to steric effect and excellent control over regioselectivity.



Scheme 2: Substrate Scope of Terminal and Internal Alkynes for 4CR

The 4CR could be elegantly extended to activated alkenes to furnish tetrahydropyrazolo[1,5-c]quinazolines **26** of four-point diversity exhibiting excellent atom economy (Scheme 3). Electron-deficient alkenes, such as acrylates and acrylonitrile participated efficiently in 4CR reaction to produce **26** in good to excellent yields with the formation of one diastereomer exclusively. A range of aryl and alkyl sulfonyl hydrazides were successfully incorporated. Both electron-donating and electron-withdrawing group on aryl sulfonyl hydrazides diversity were well tolerated. In all cases, the reaction proceeds with high regiochemical fidelity.



Scheme 3: Substrate Scope of alkenes for 4CR

S11. References

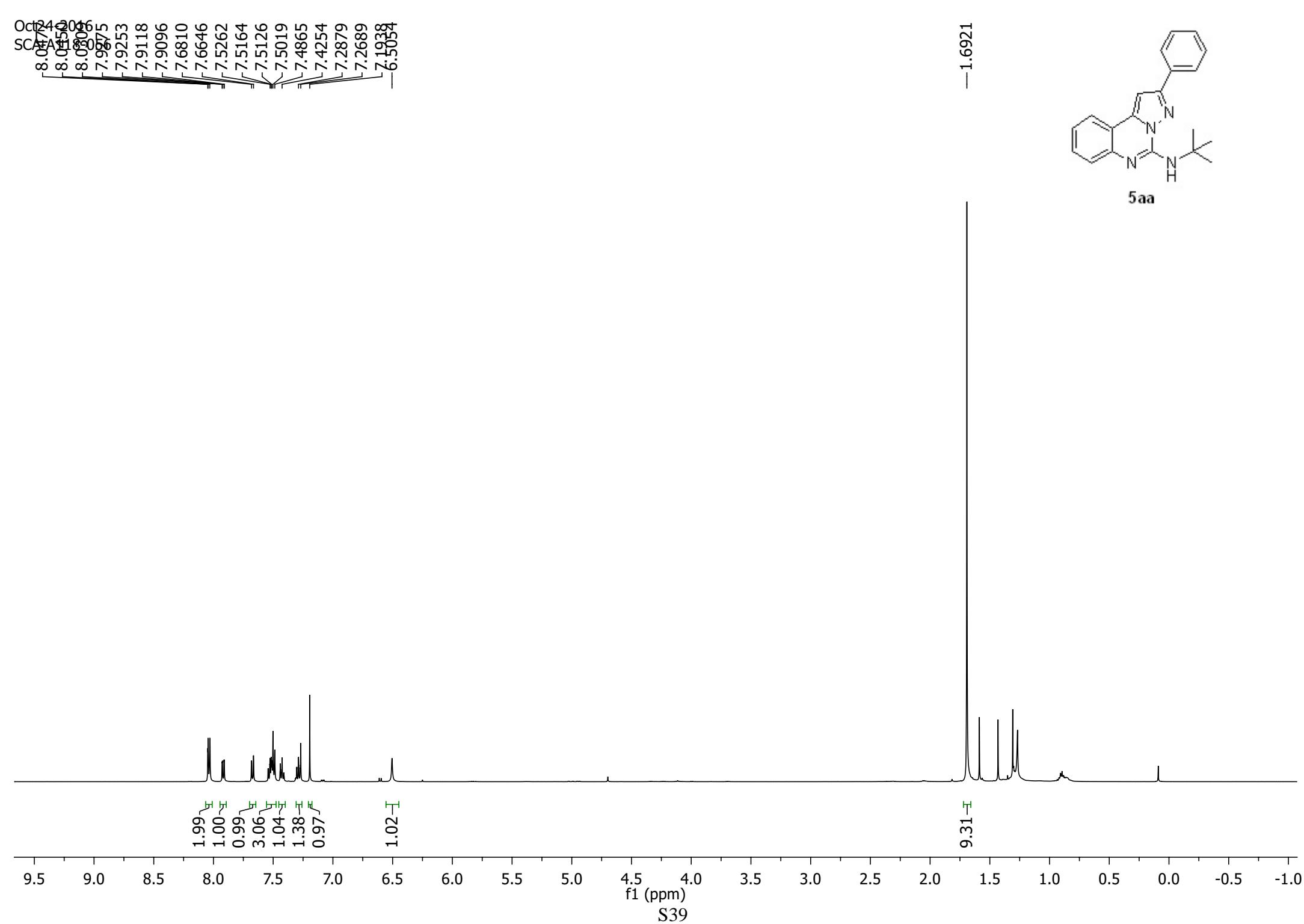
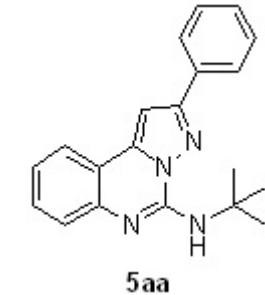
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- 3) The hydrazone **18** was found extremely prone towards hydration to produce a major side product urea **7aa**. The use of molecular sieves however could elegantly assist in avoiding this undesired event.
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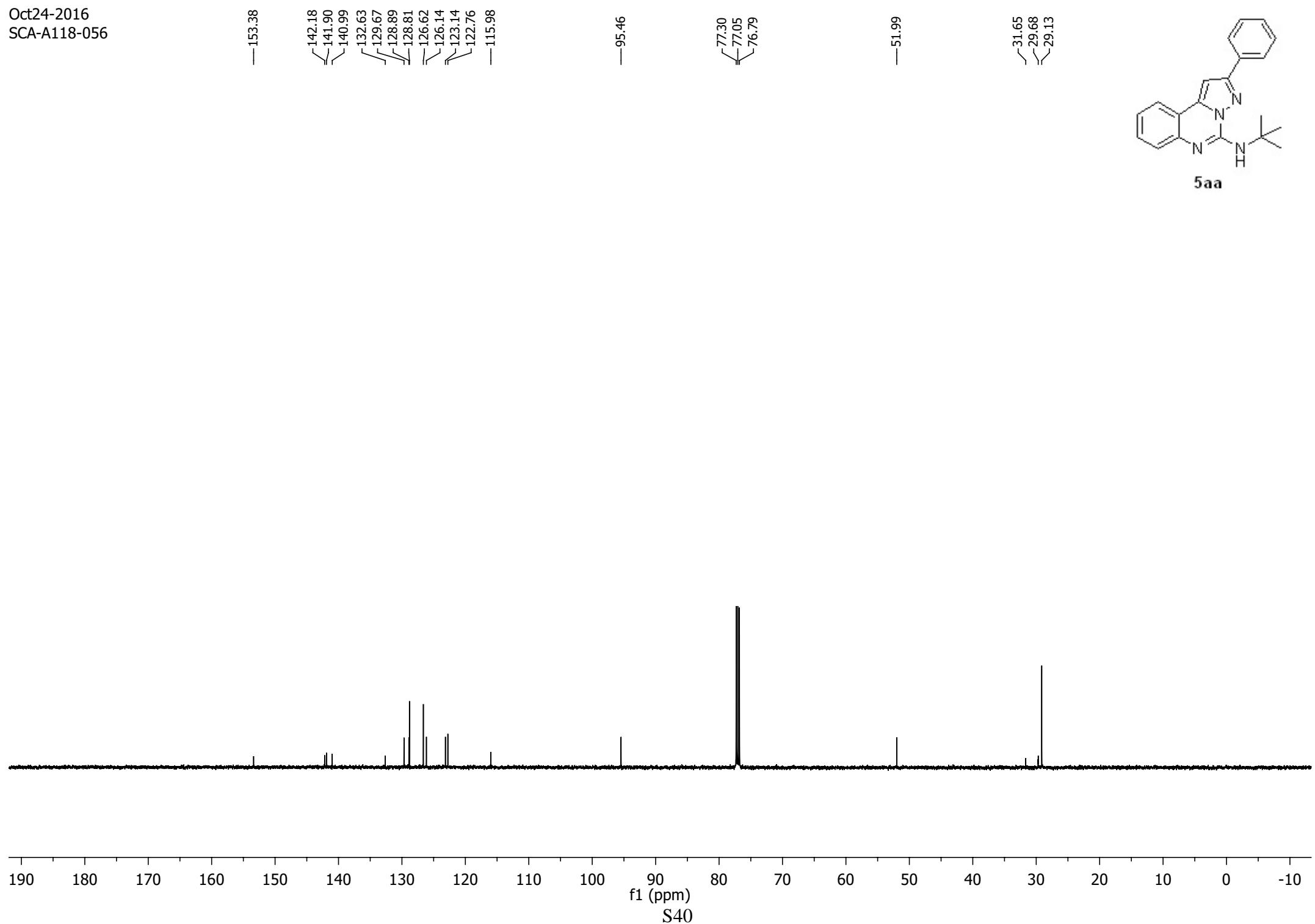
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S12. Copies of ^1H , ^{13}C NMR and HRMS spectra

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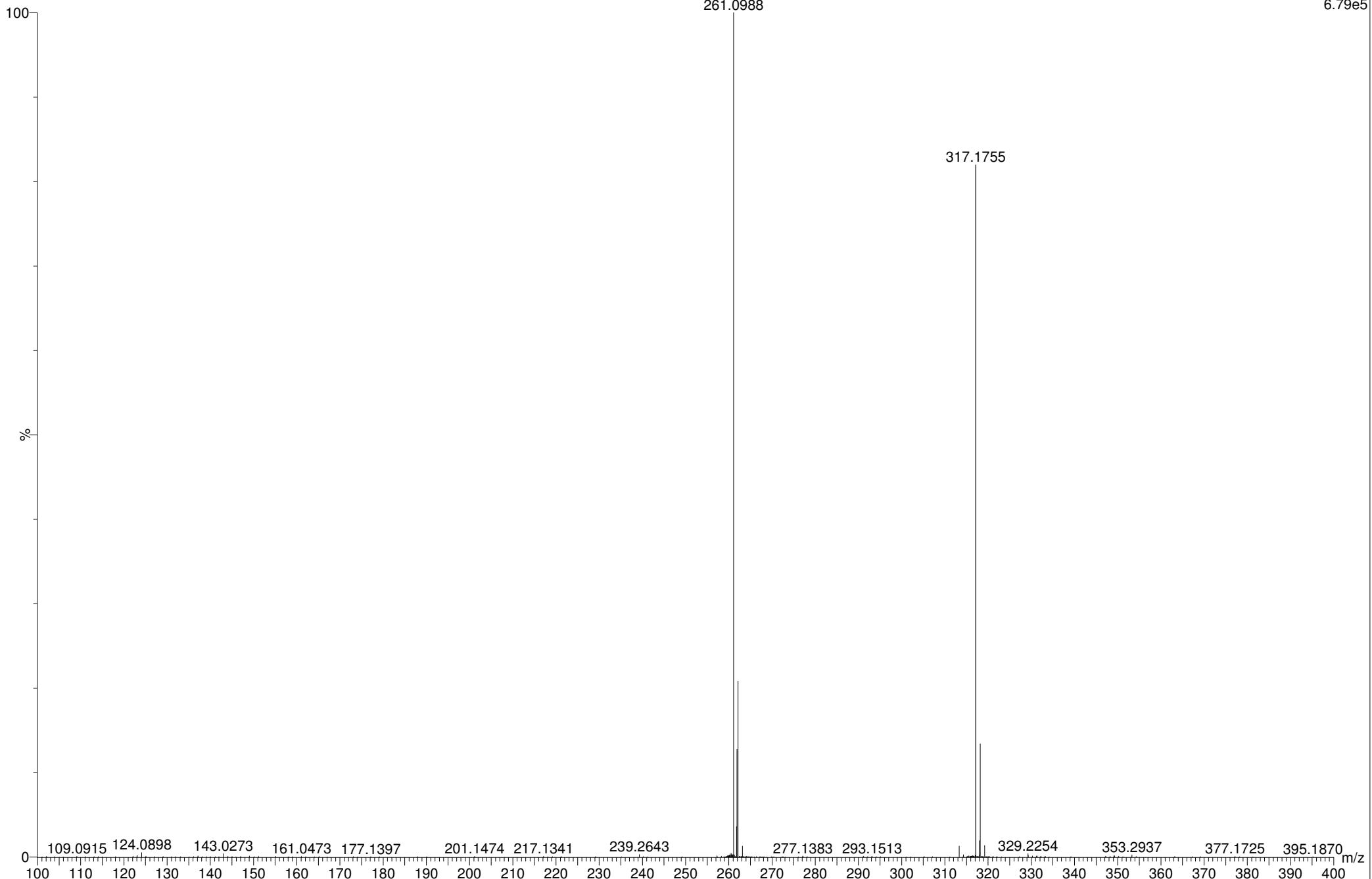
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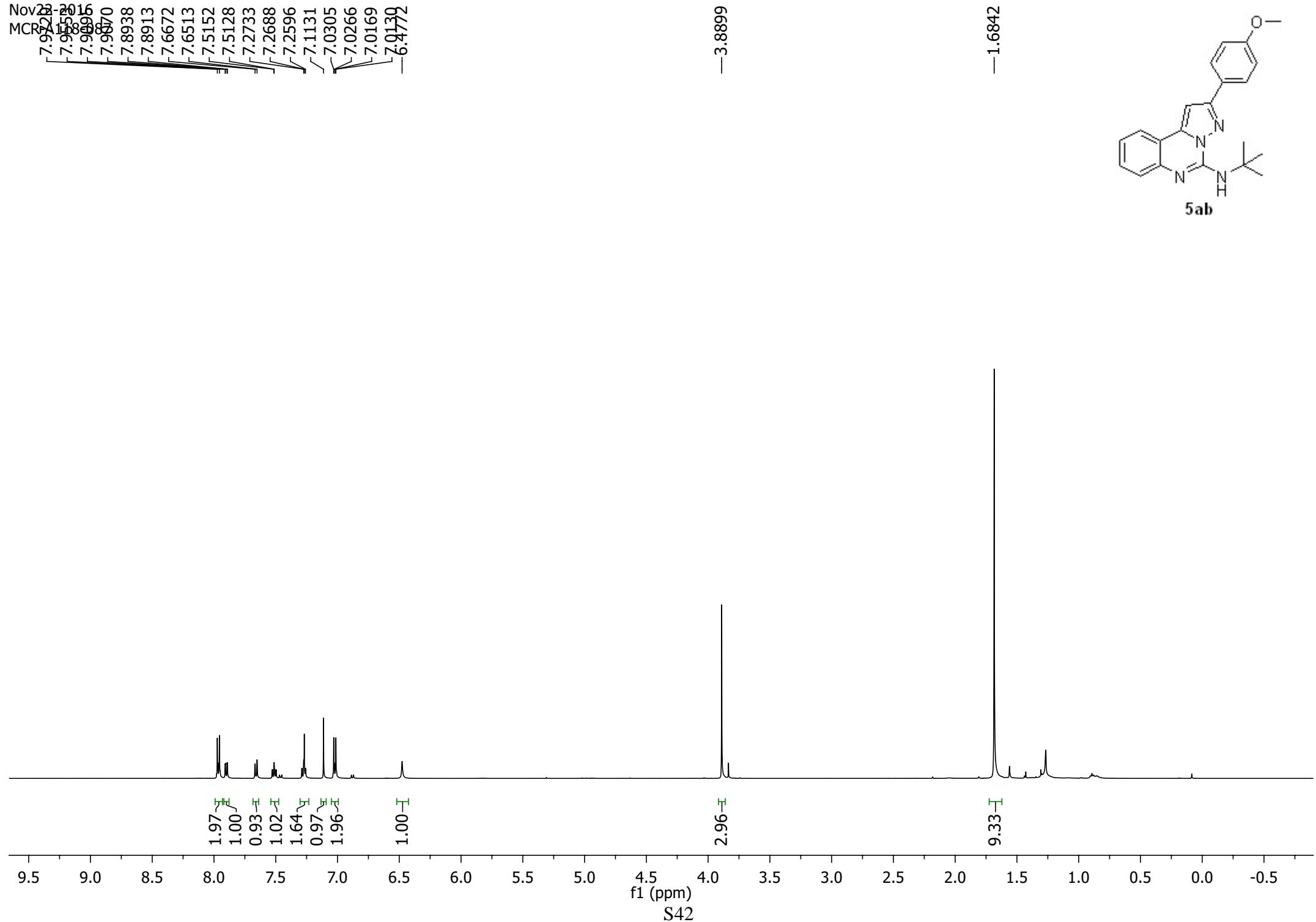
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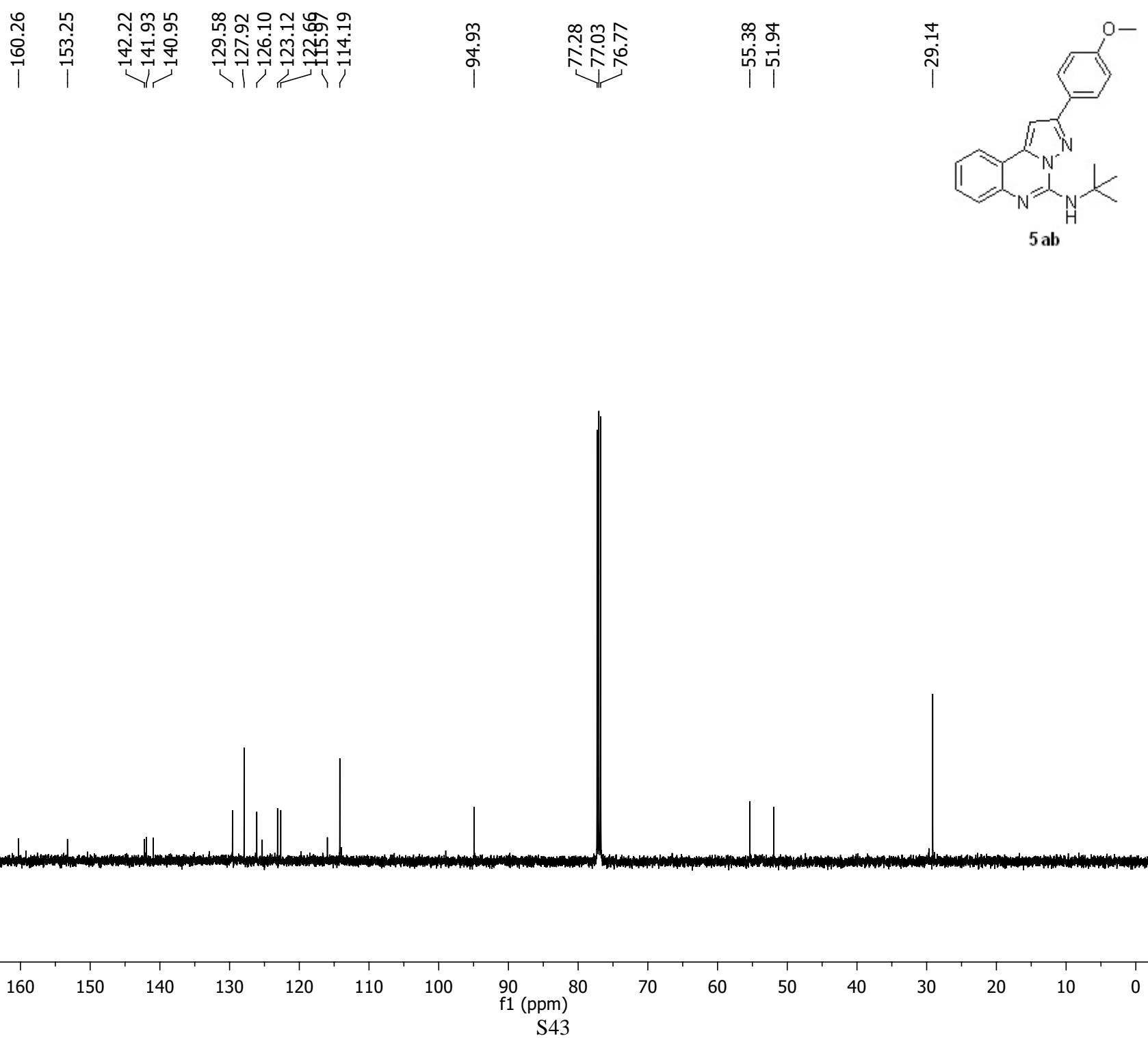
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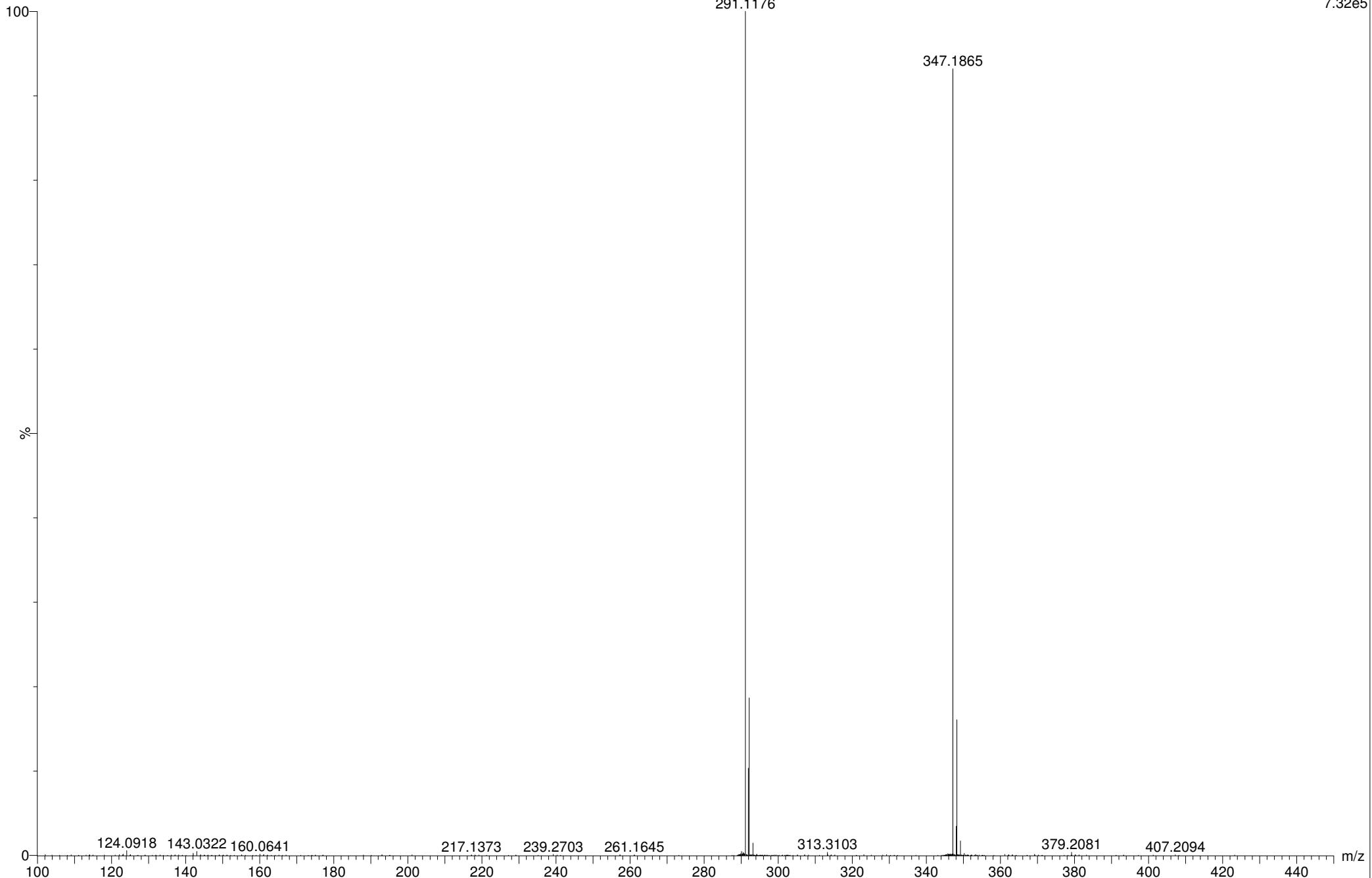
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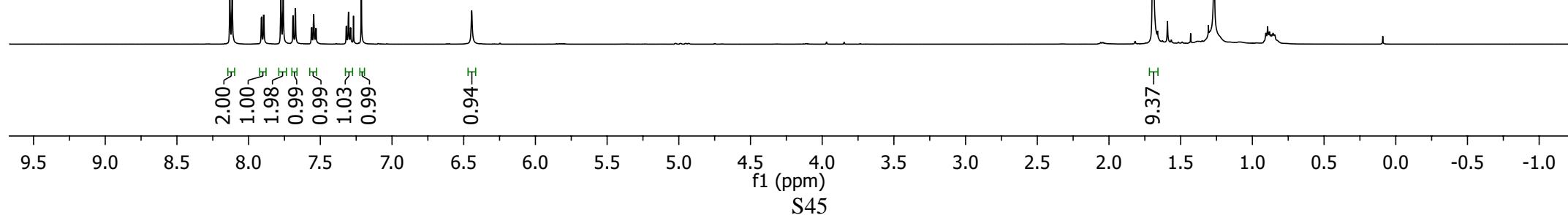
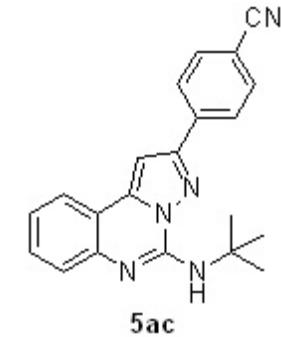
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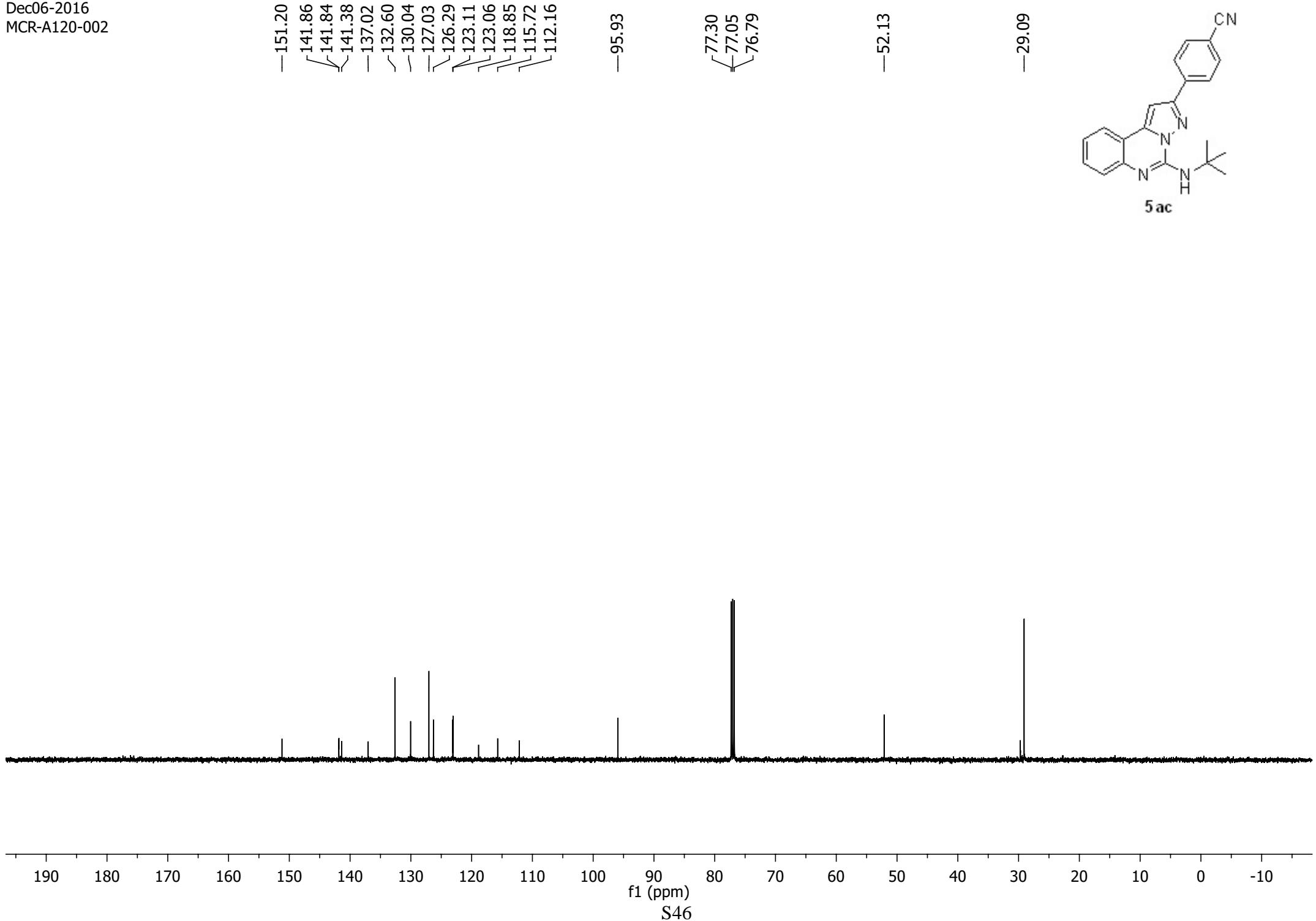
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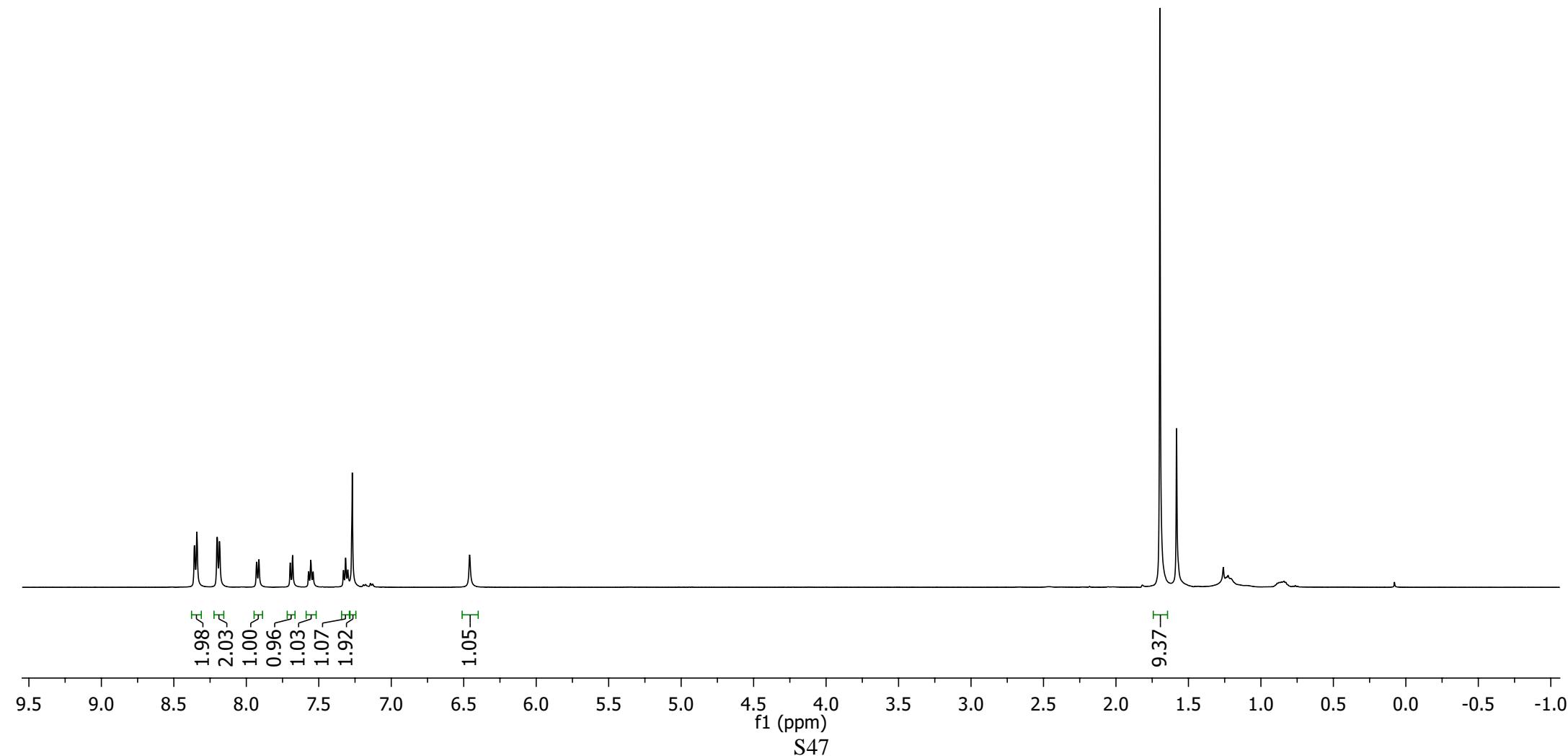
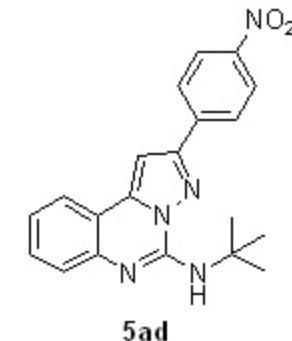


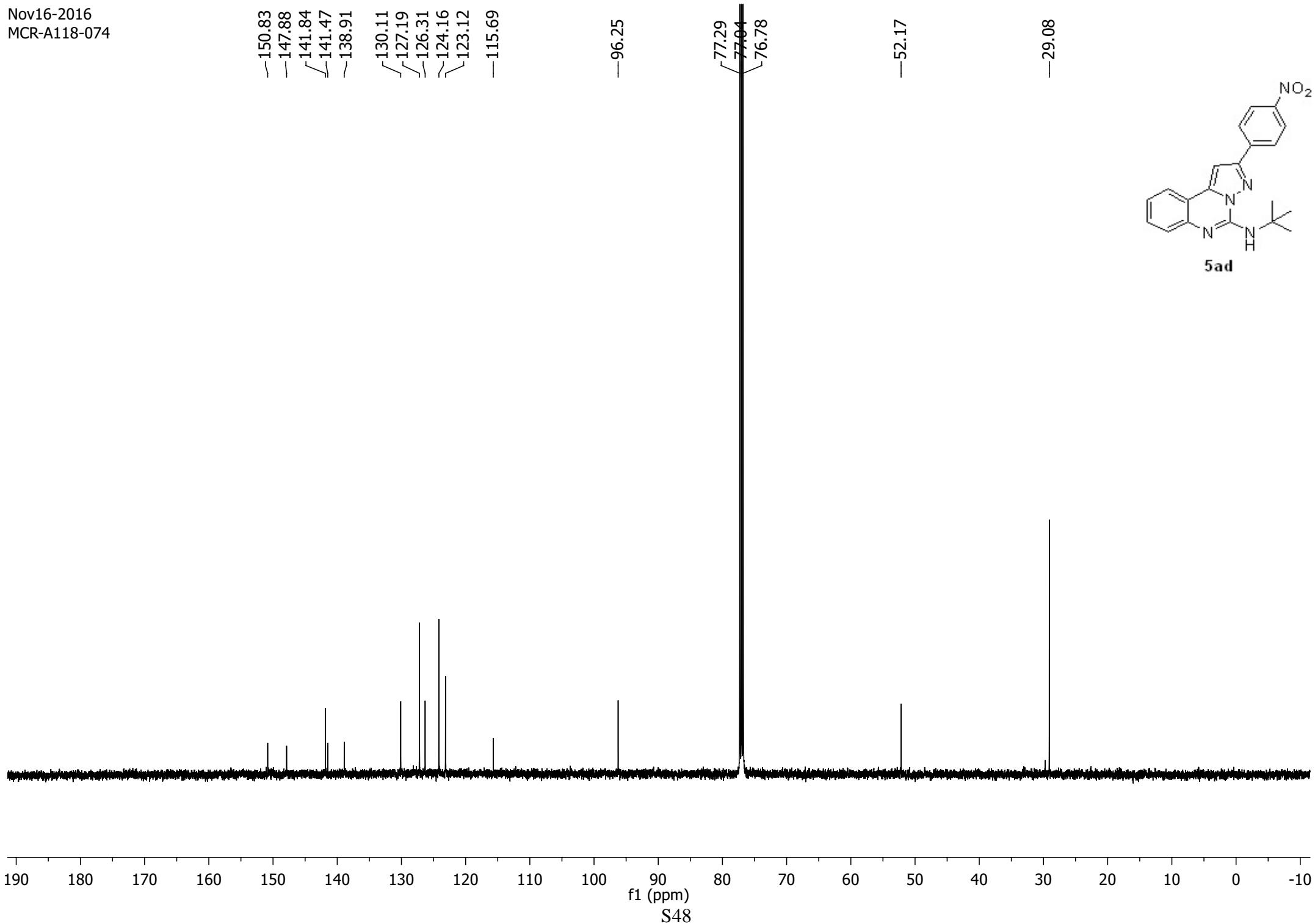


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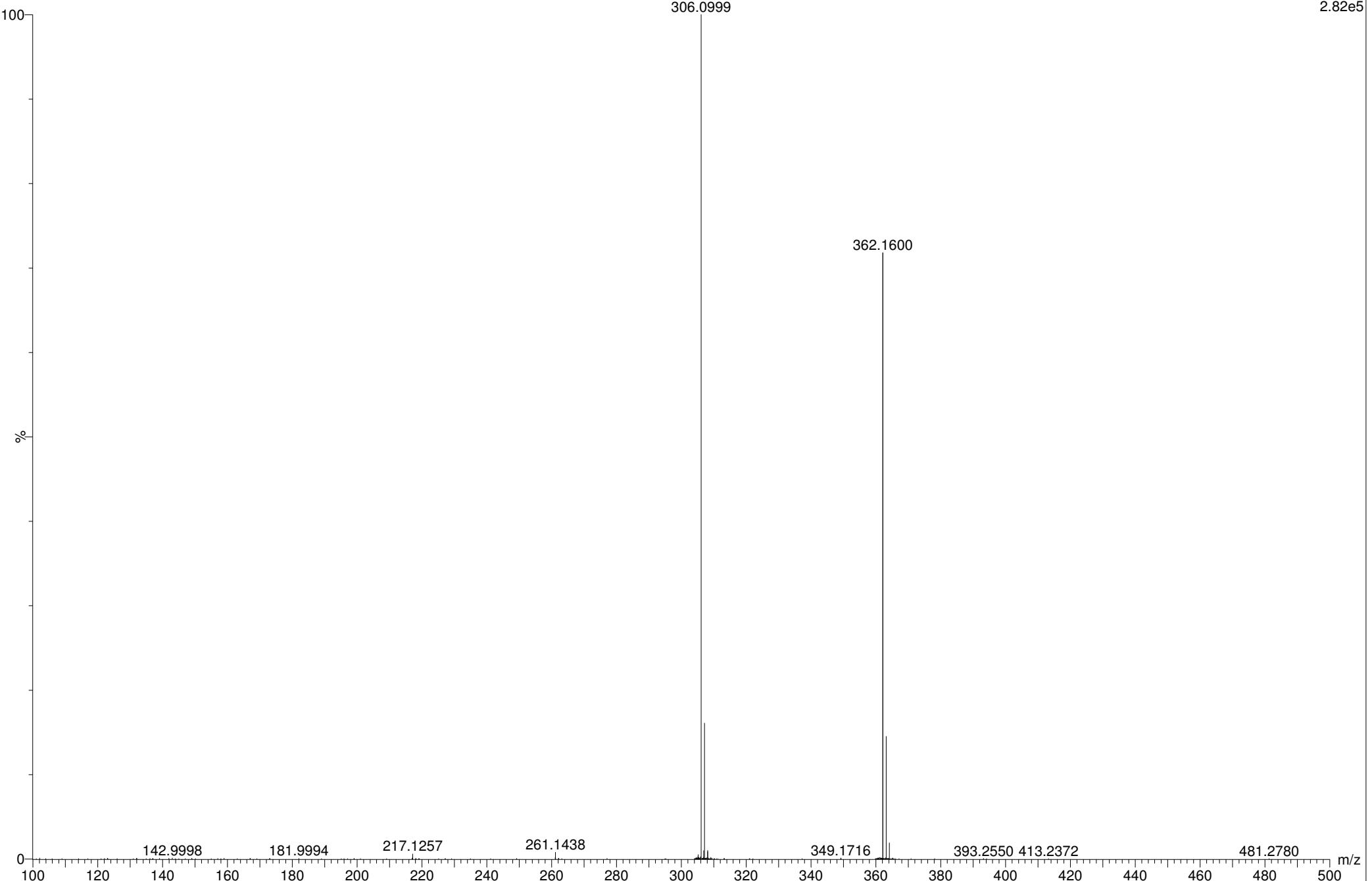
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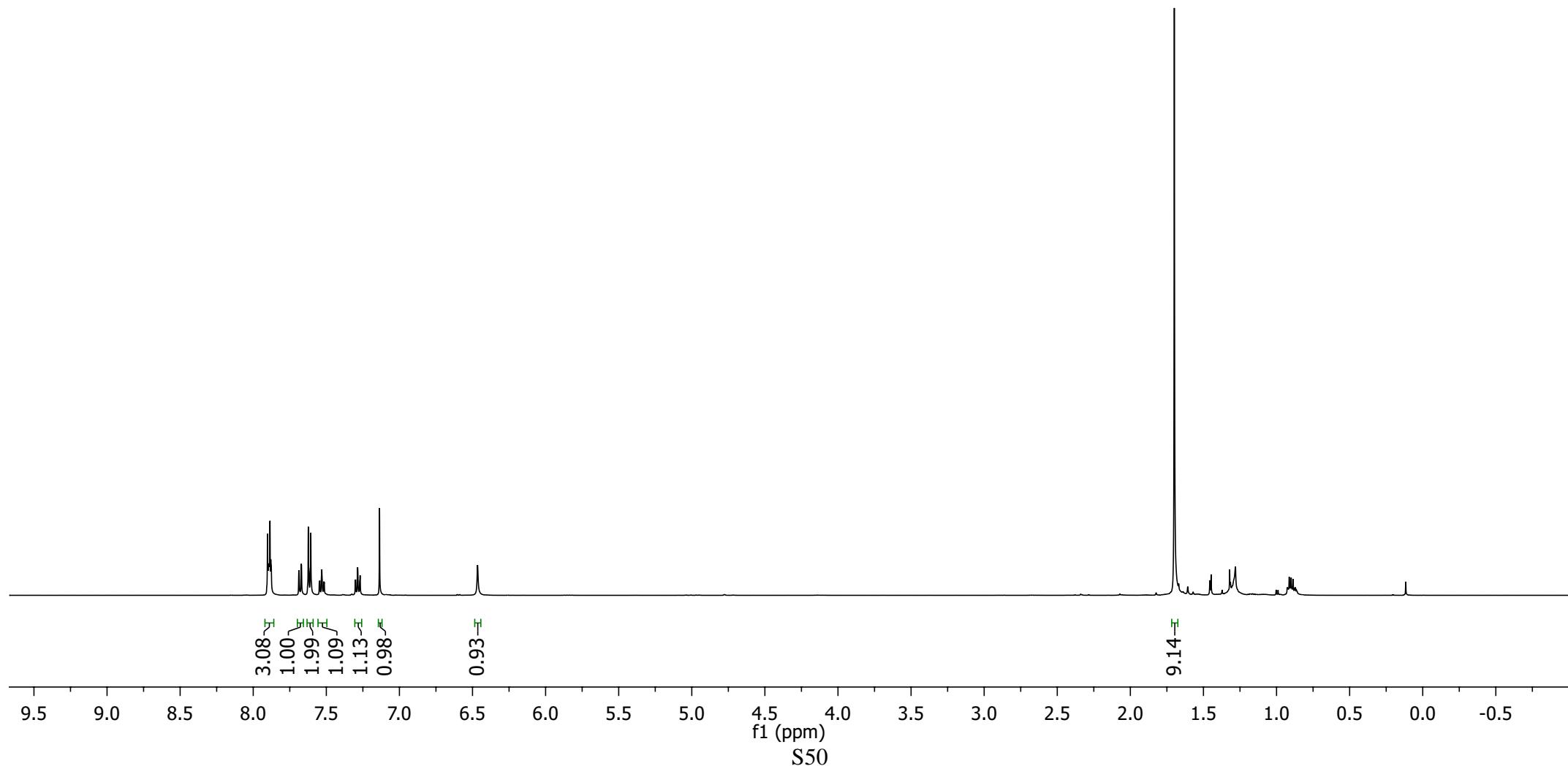
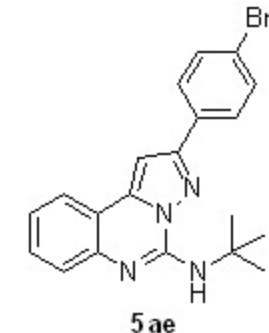
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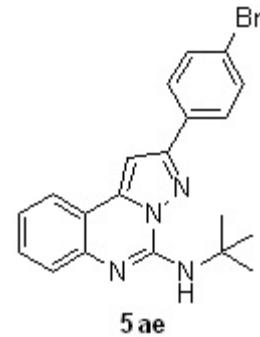
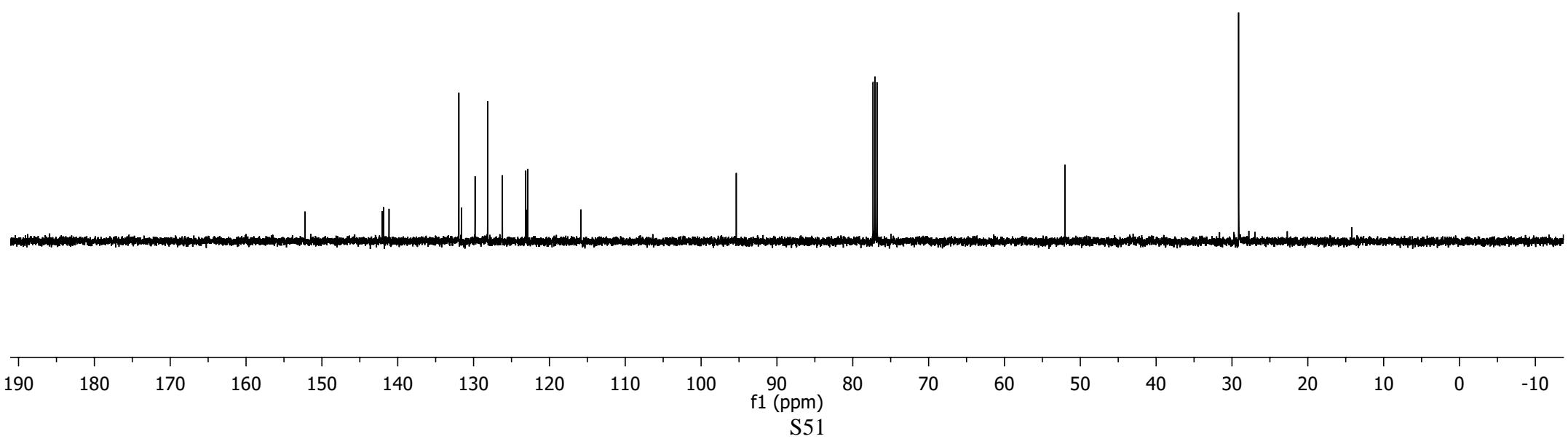


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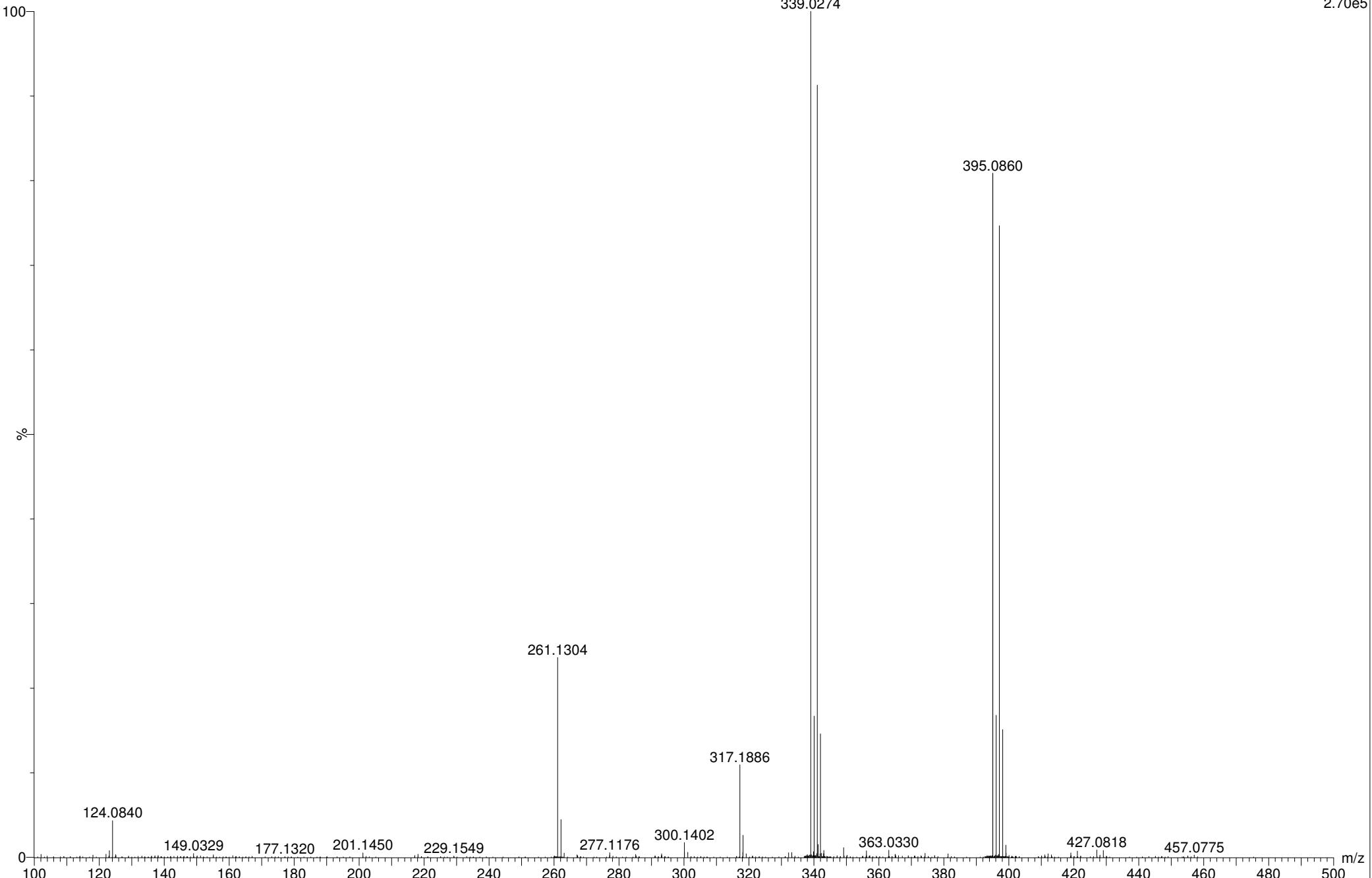
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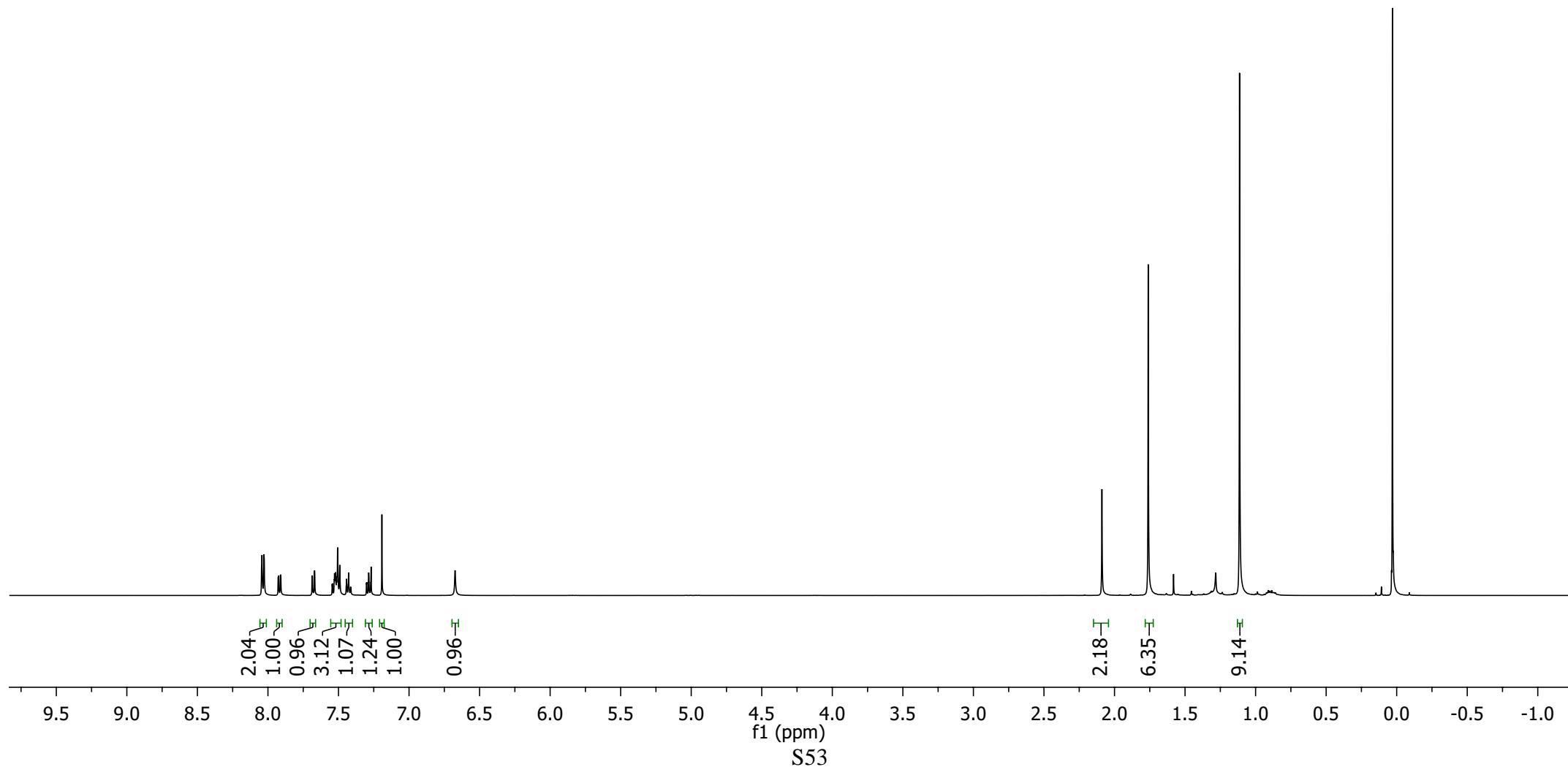
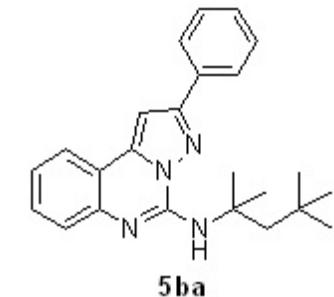
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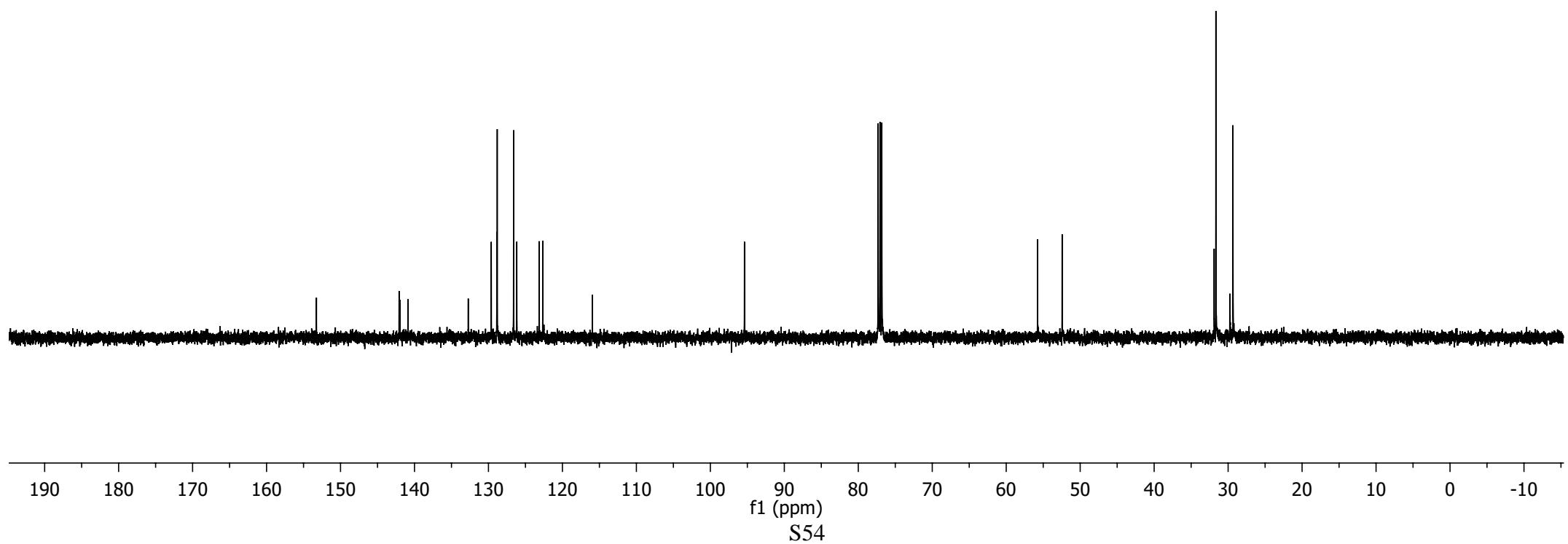
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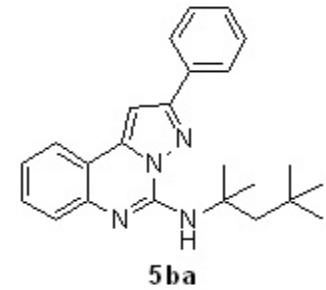
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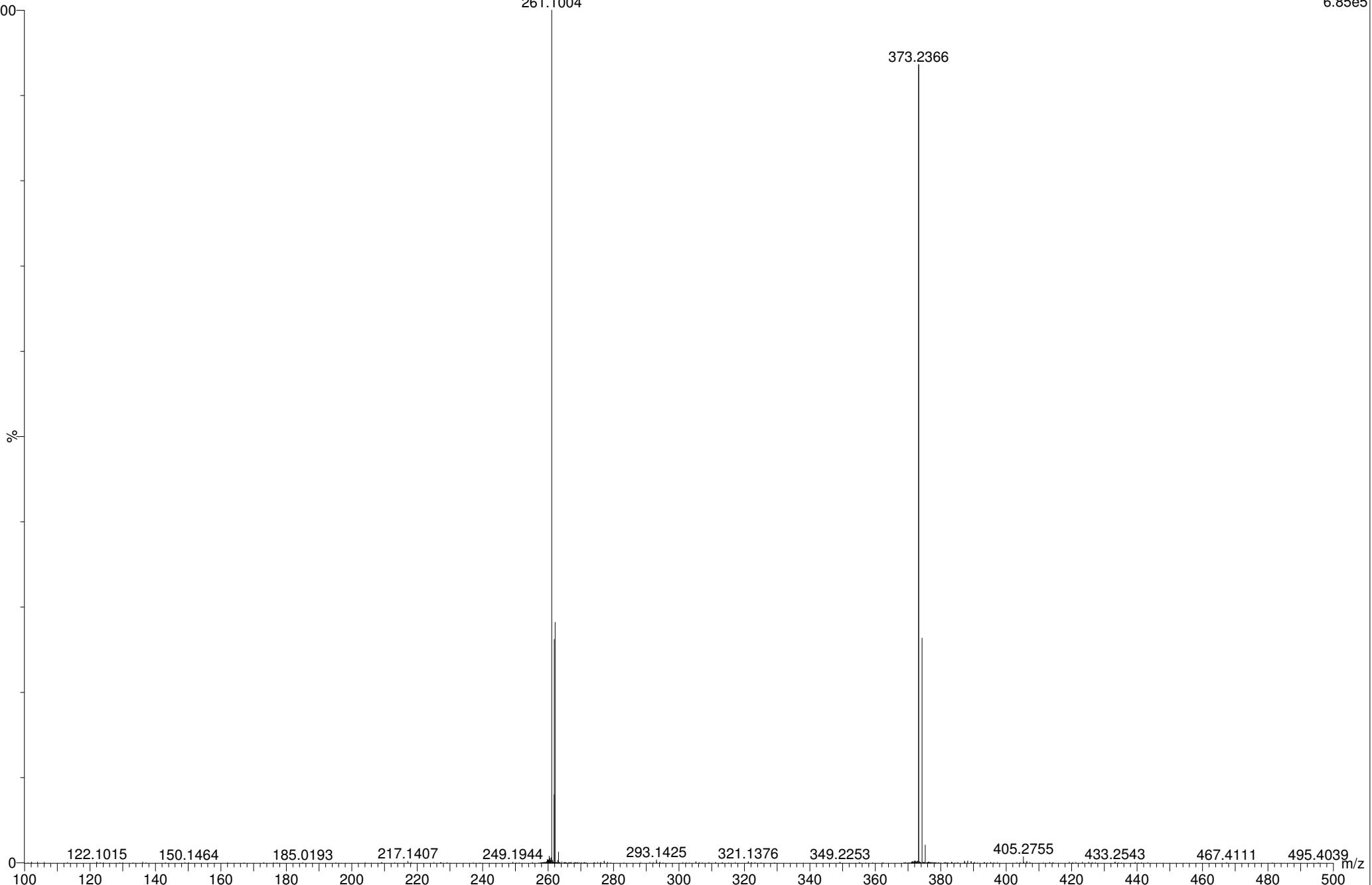
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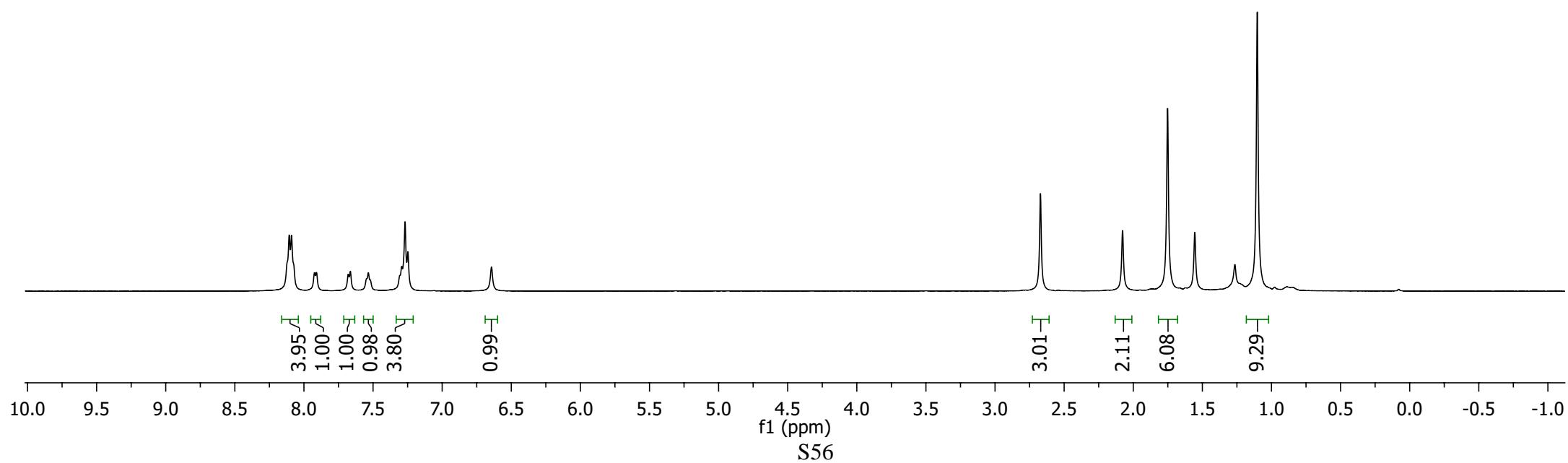
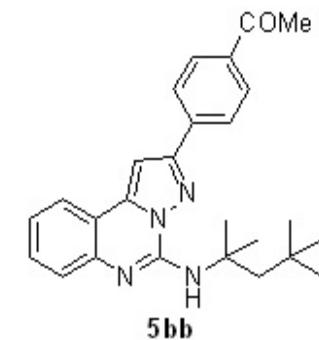
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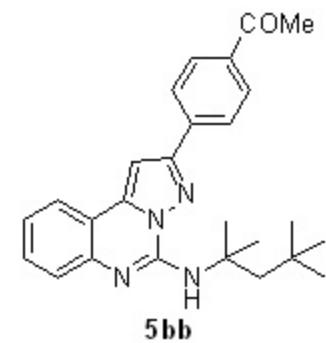
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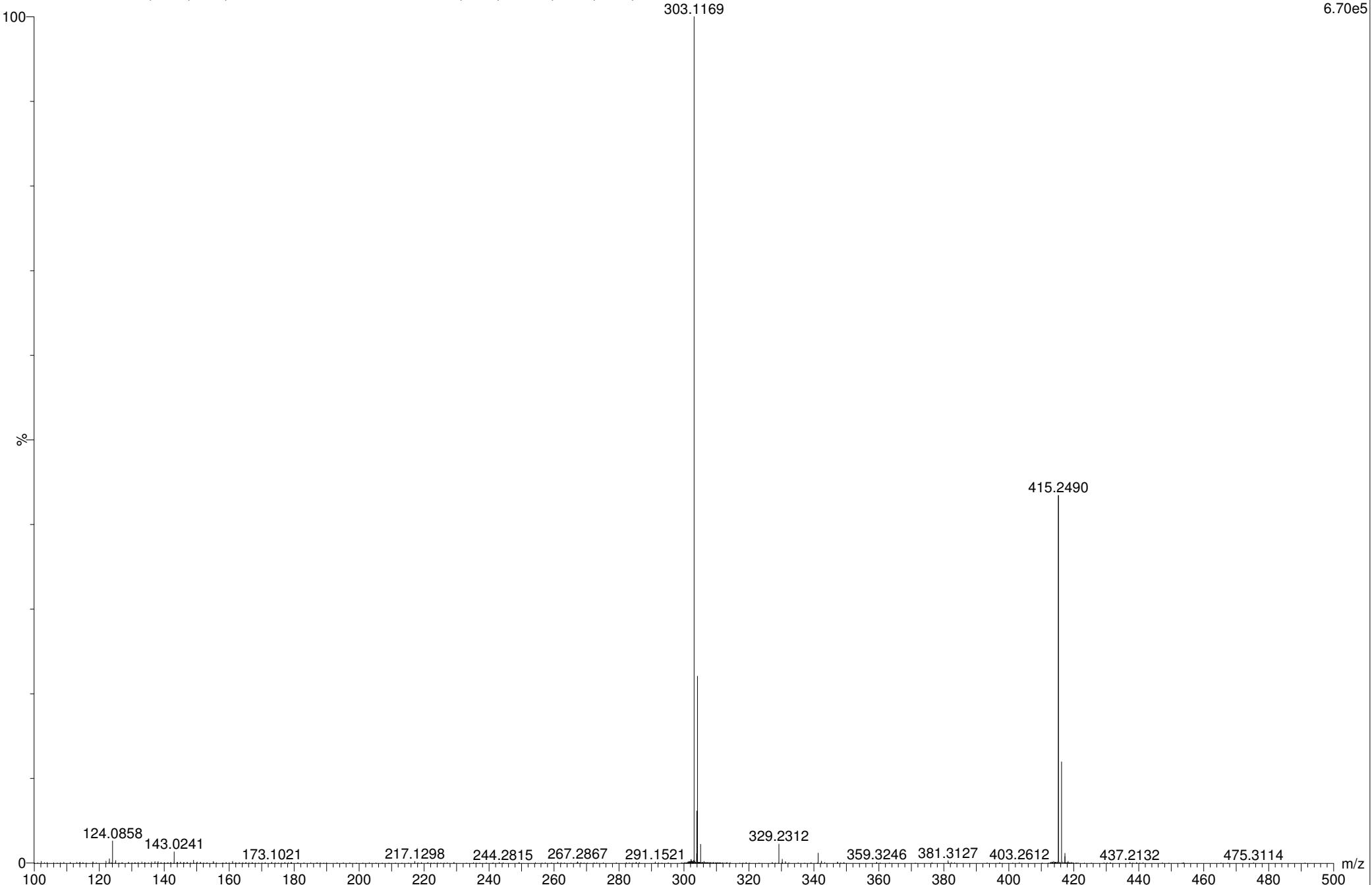
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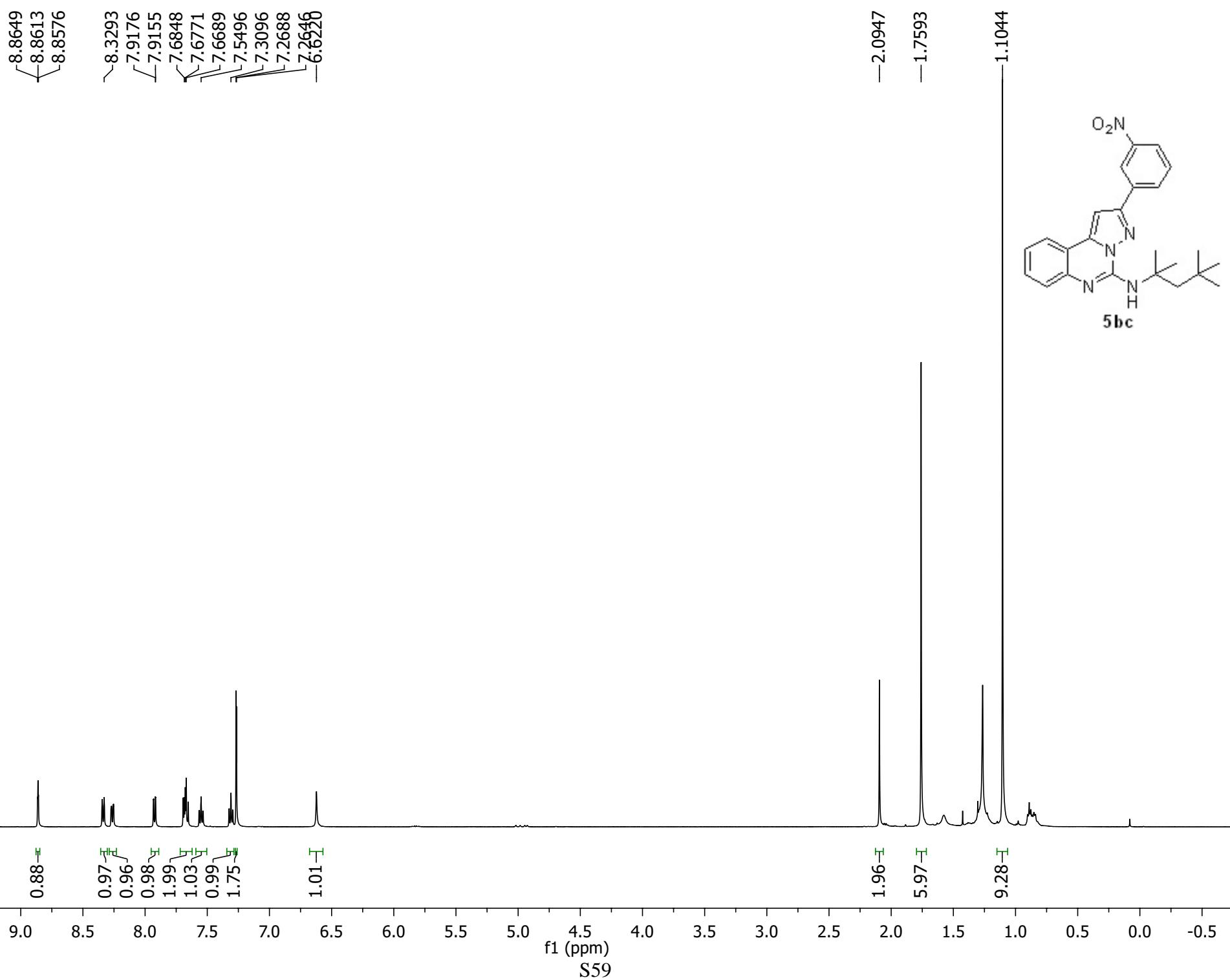
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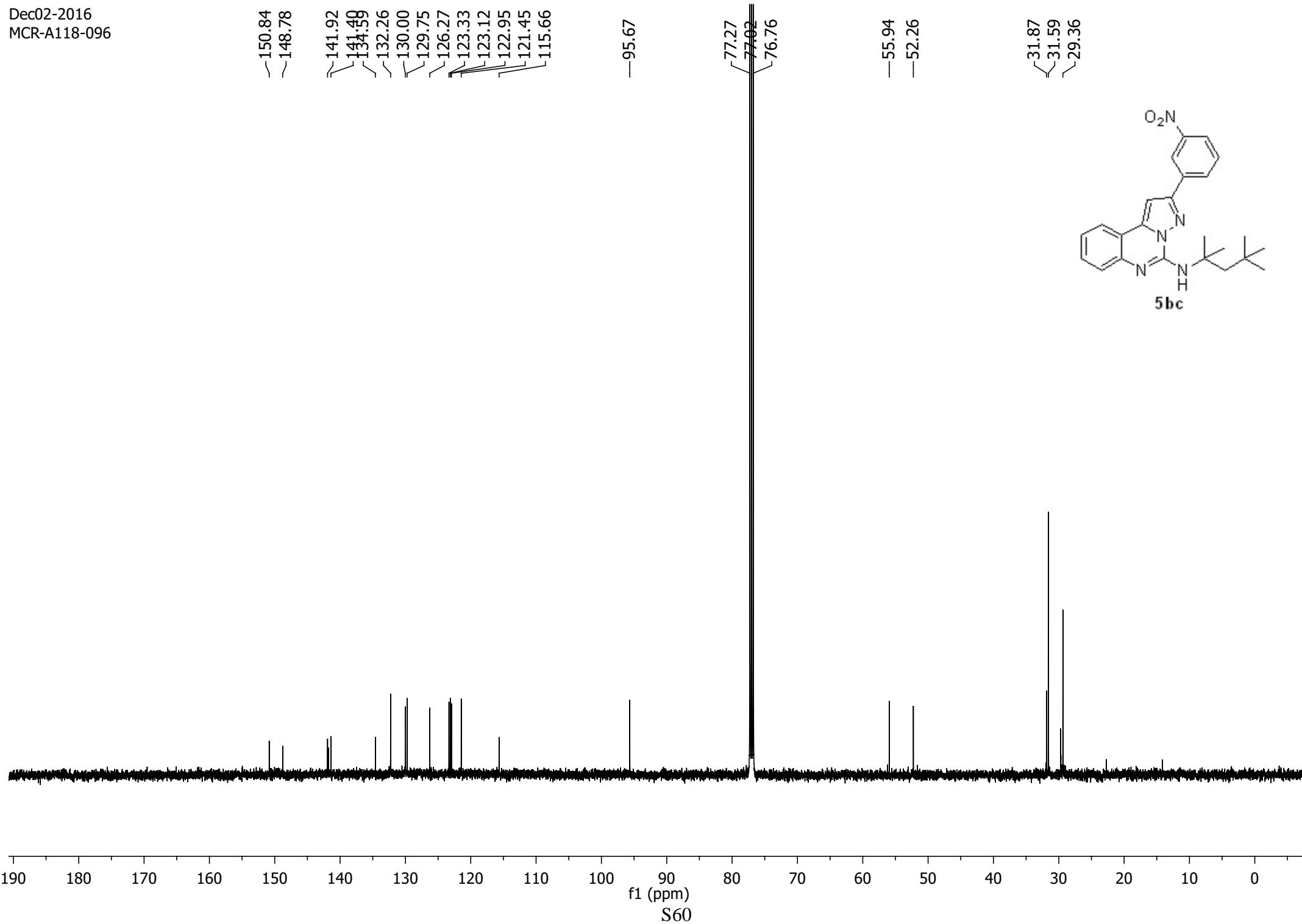
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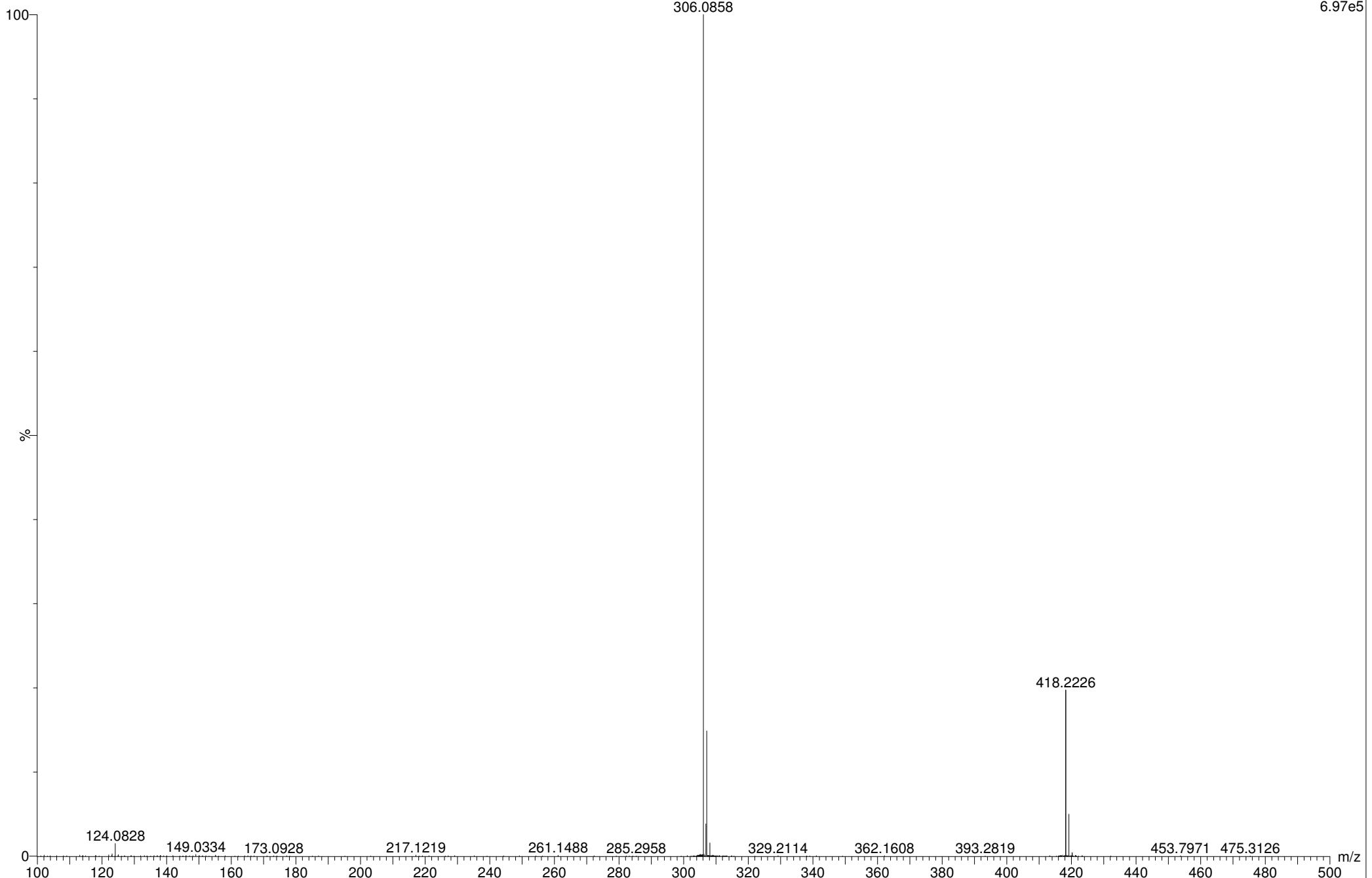
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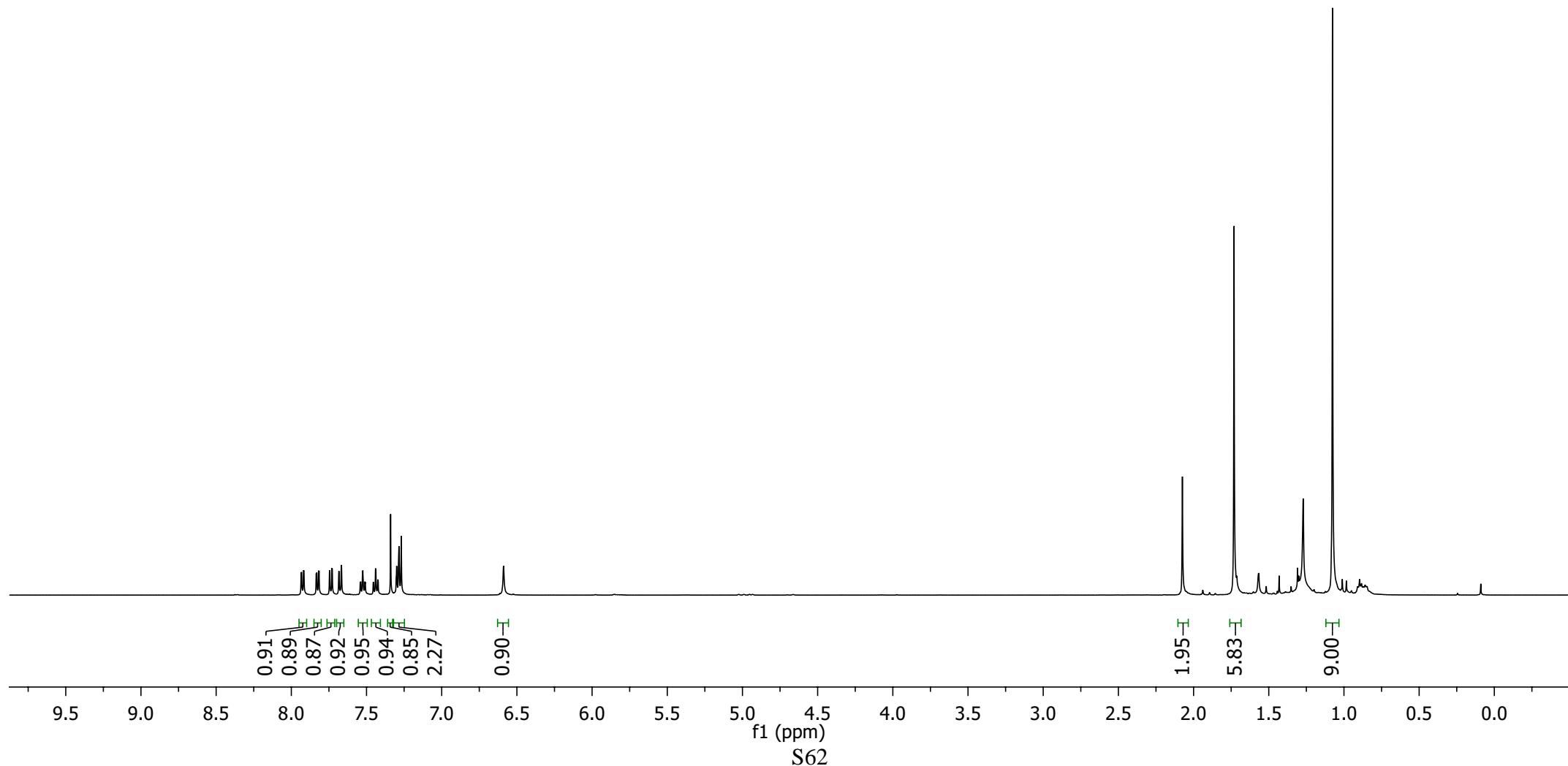
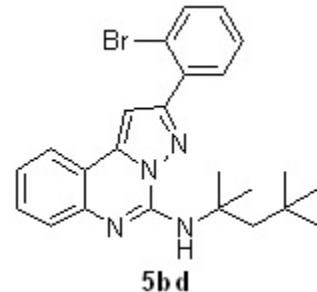


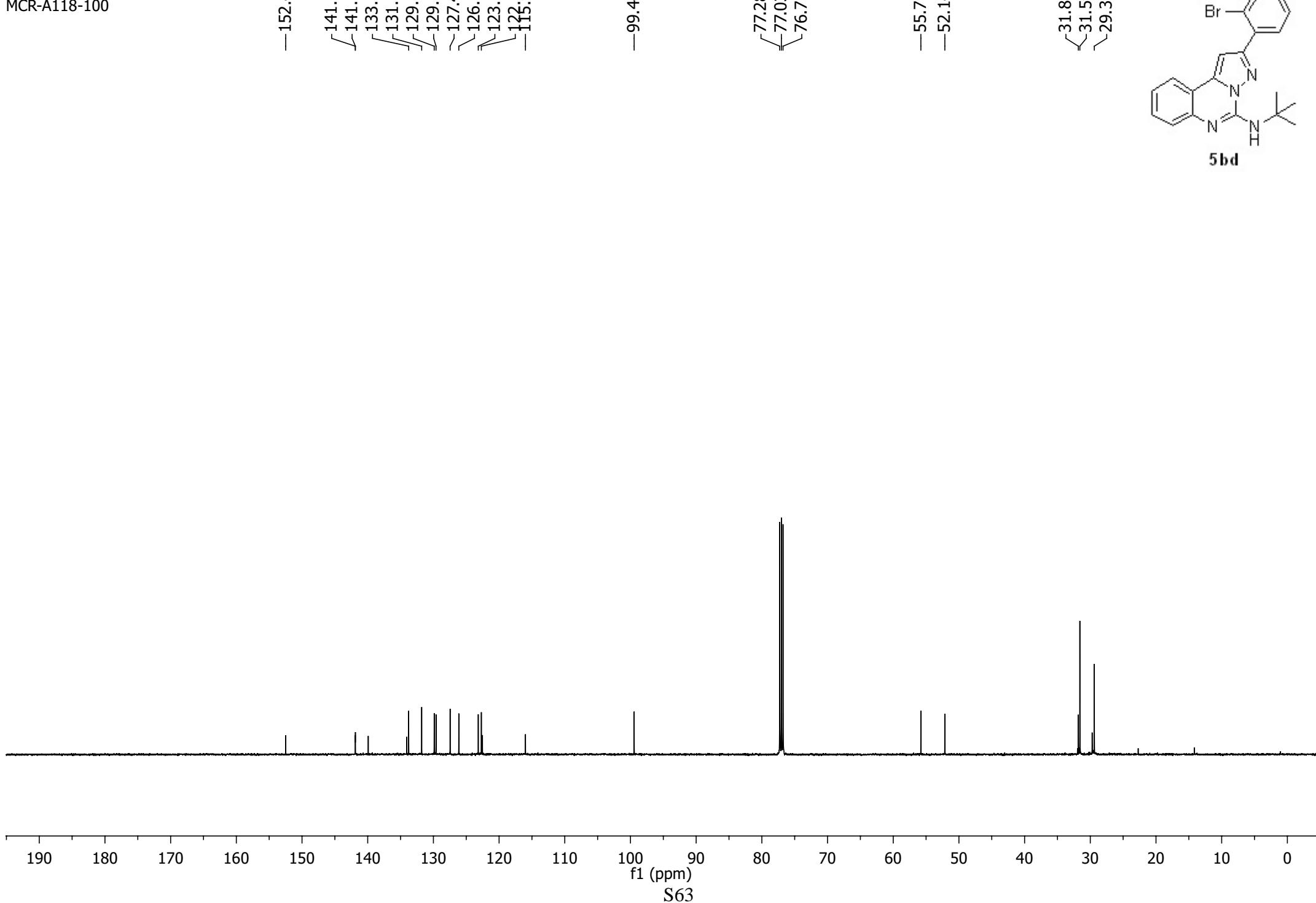
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7.9324
7.9186
7.9166
7.8912
7.8190
7.8158
7.7457
7.7442
7.7297
7.7282
7.6834
7.6671
7.5248
7.4399
7.4383
7.3397
7.2995
7.2841
7.2687
6.5888

-2.0744
-1.7310

-1.0763





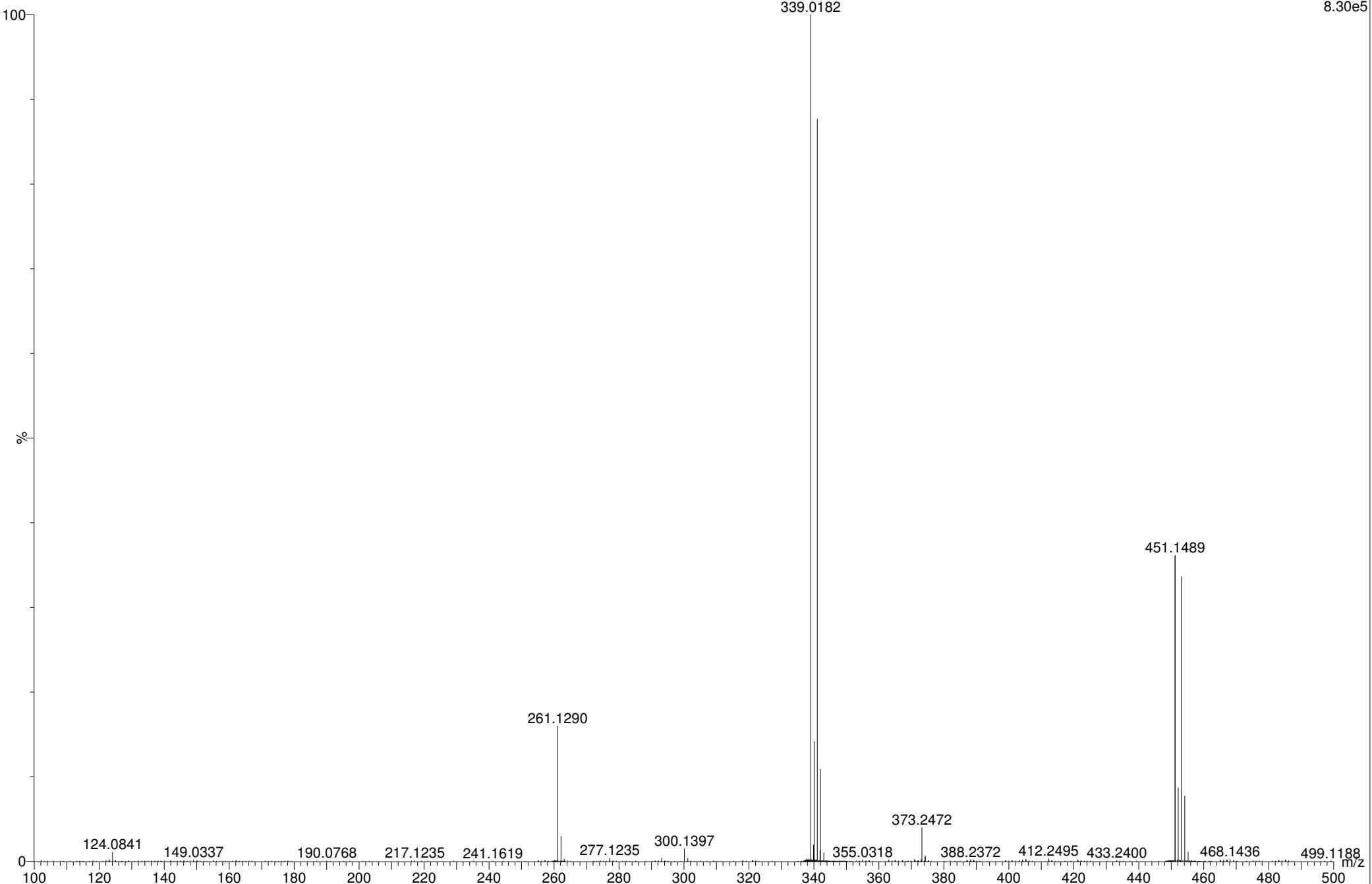
PROCESSED BY
PAWAN KUMAR

CSIR-IHBT
NPC&PD DIVISION

19-Jan-2017 14:28:38

MCR-A118-100_02 114 (2.113) AM (Cen,3, 80.00, Ar,5000.0,451.15,0.70); Sb (2,10.00); Cm (107:126)

1: TOF MS ES+
8.30e5



Dec 21 2016
MCR

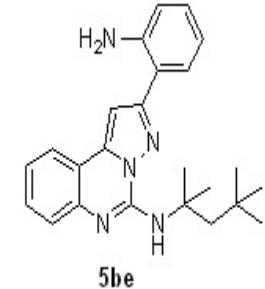
7.9416
7.9255
7.9238
7.7334
7.7308
7.7179
7.7153
7.6909
7.6747
7.5434
7.5411
7.2961
7.2239
7.2076
7.1991
6.8624
6.8498
6.8352

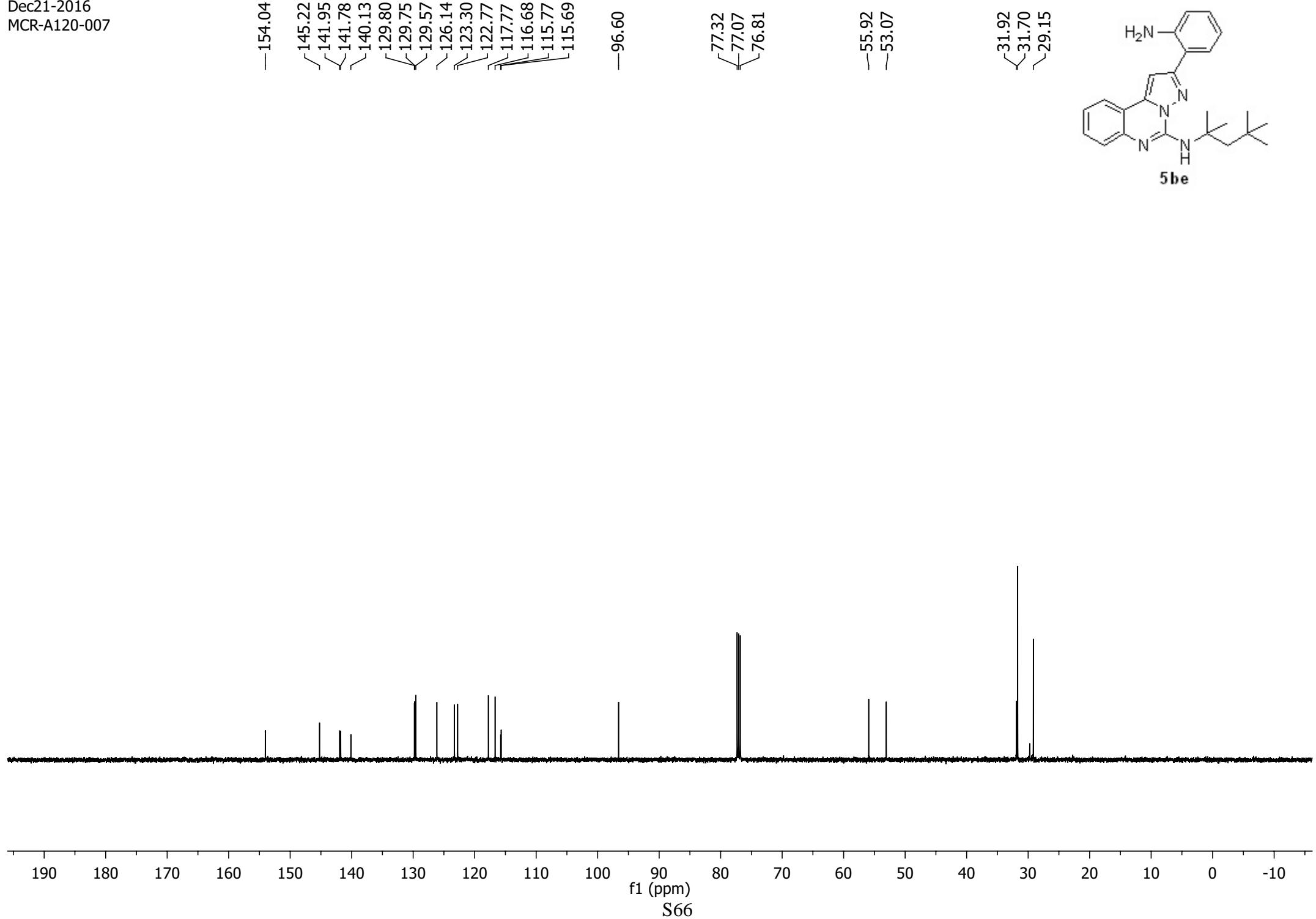
-5.3766

f1 (ppm)
S65

-2.0330
-1.7638

1.1153





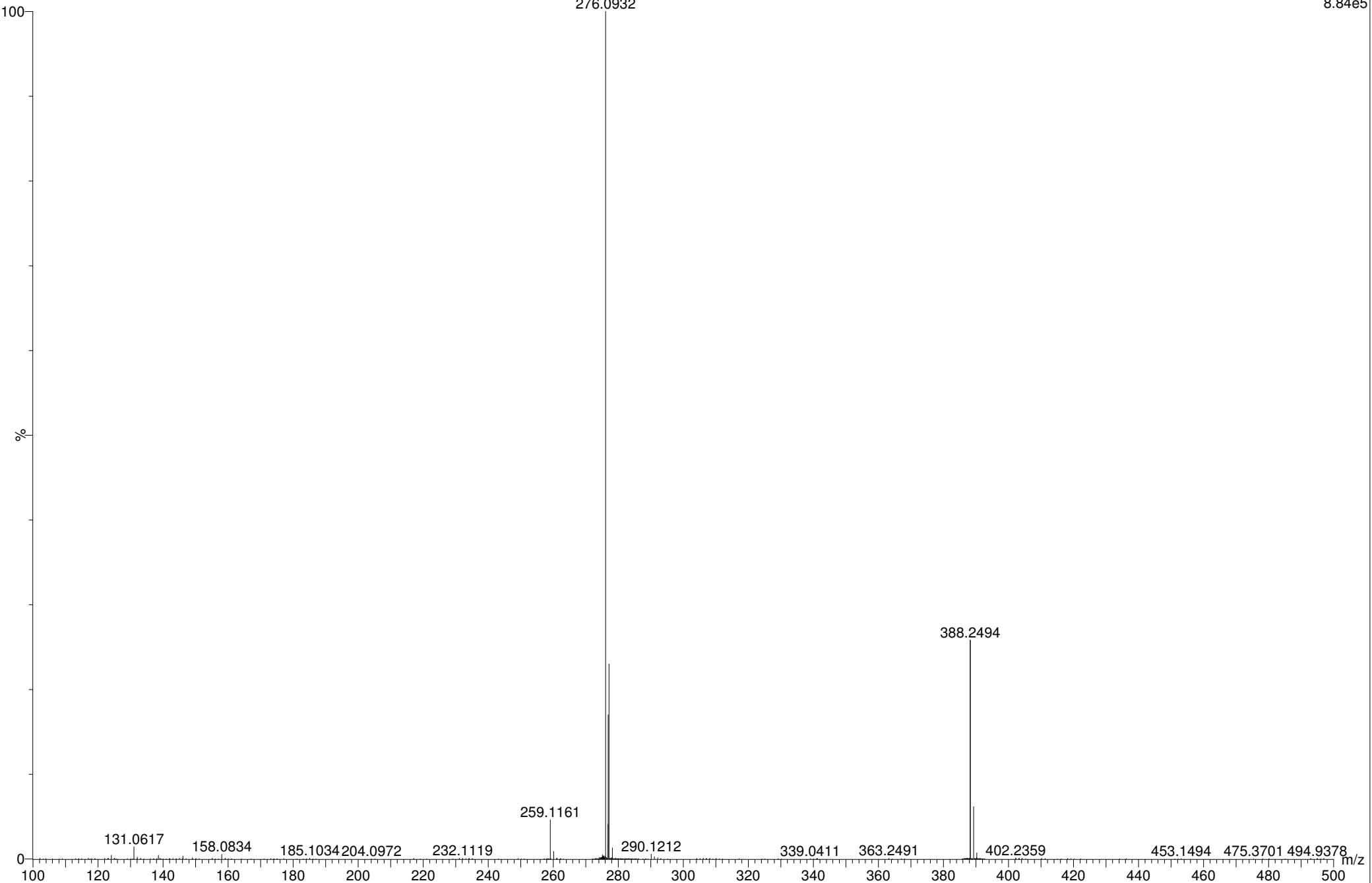
PROCESSED BY
PAWAN KUMAR

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NPC&PD DIVISION

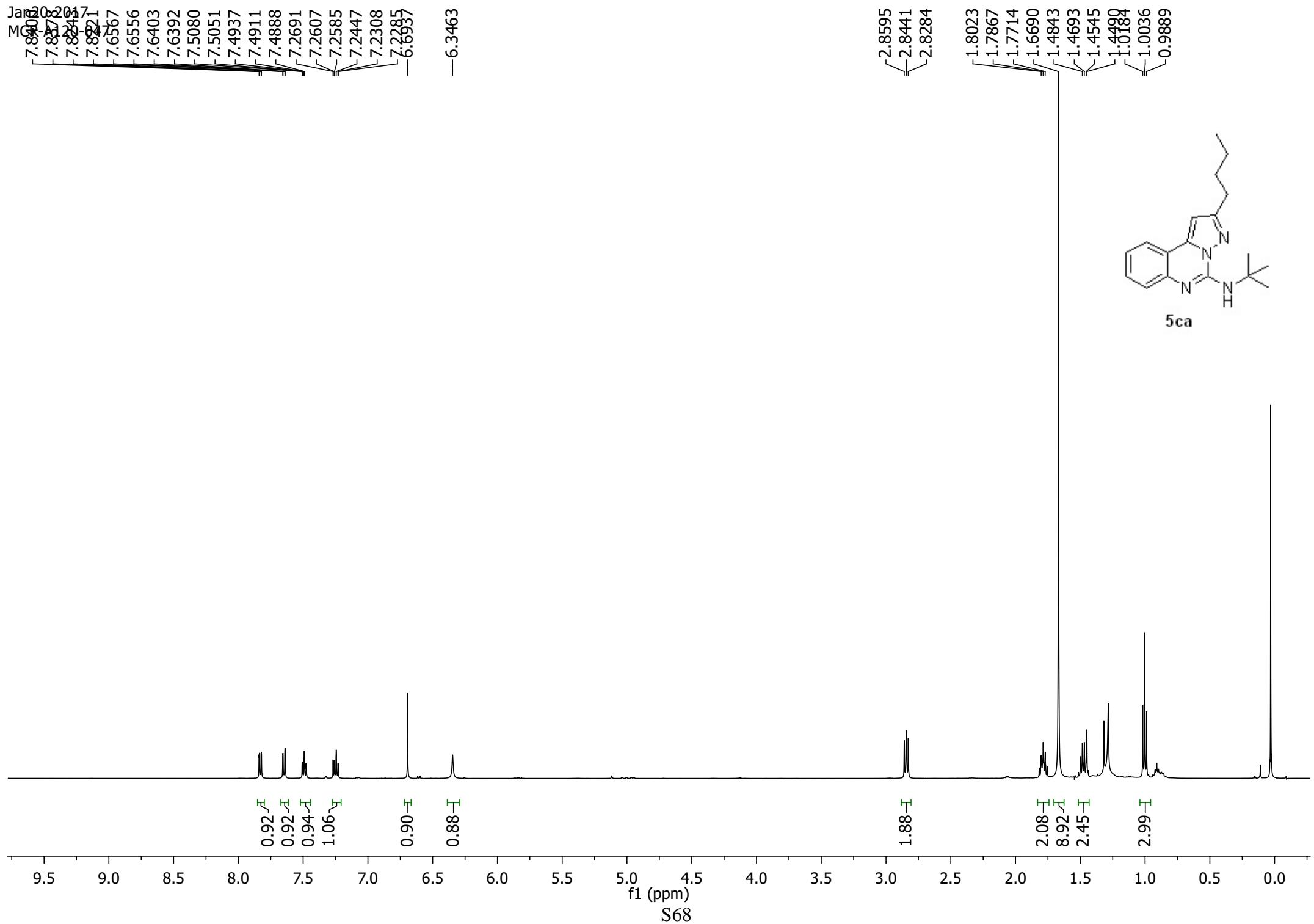
19-Jan-2017 14:32:39

MCR-A120-007_02 51 (0.945) AM (Cen,3, 80.00, Ar,5000.0,388.25,0.70); Sb (2,10.00); Cm (48:60)

1: TOF MS ES+
8.84e5



Jan 2001 7-1
M-G-A 7-24-22
7.890 [7.887] 7.892 7.6567
7.6556 7.6403 7.6392



-156.45

142.18
141.87
140.31

129.34
126.02
123.03
122.50
-116.00

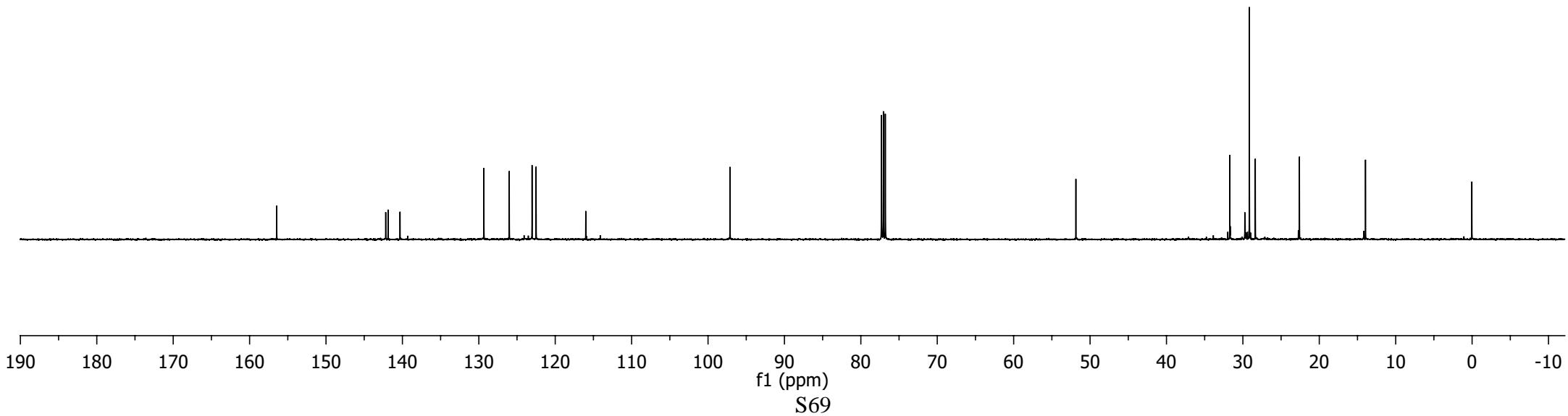
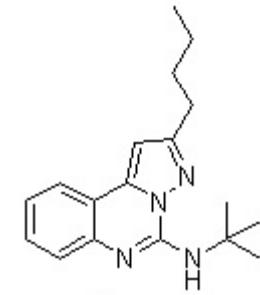
-97.12

77.30
77.05
76.79

-51.86

-31.72
29.15
28.37
-22.60

-13.95



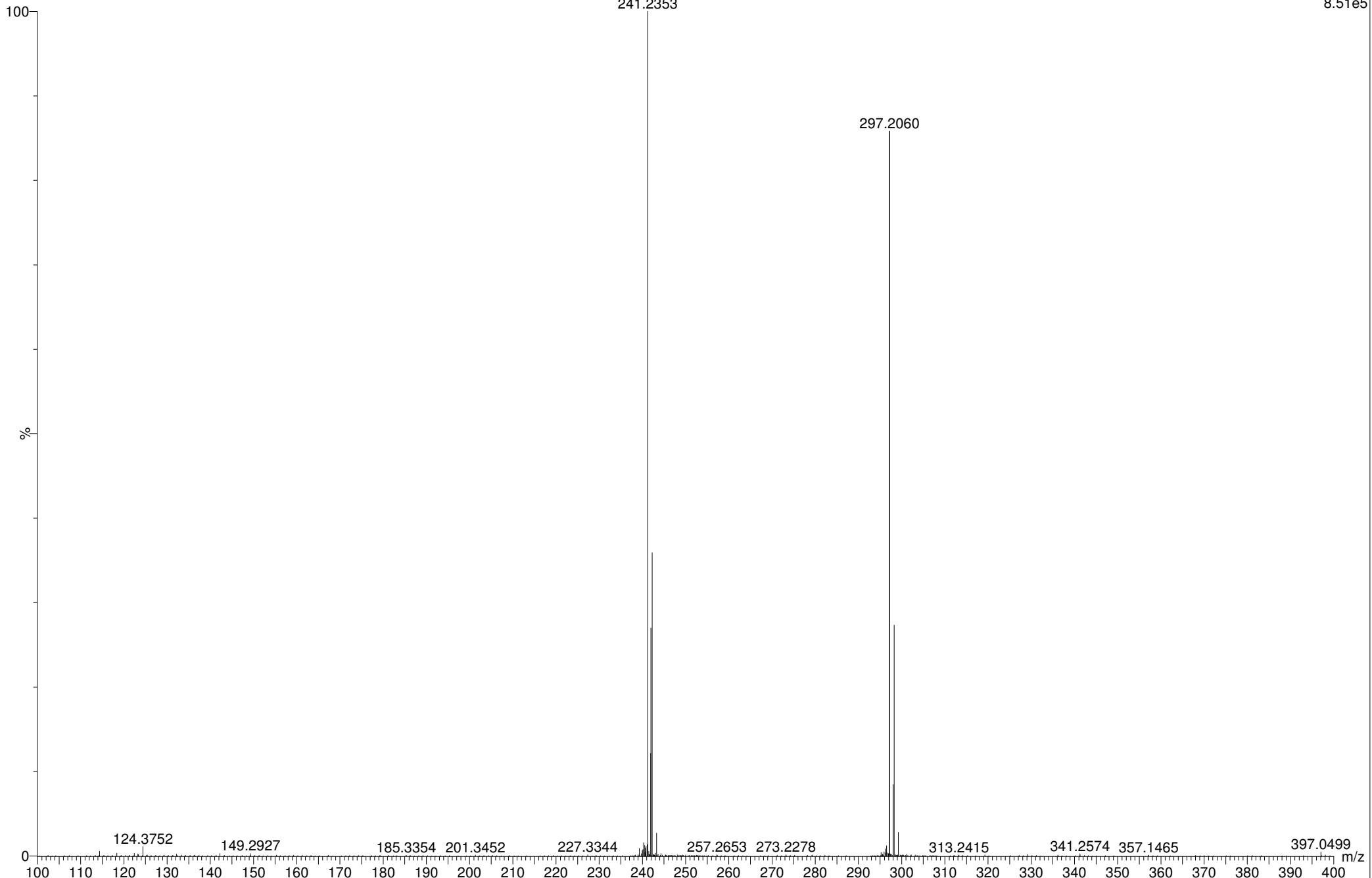
PROCESSED BY
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CSIR-IHBT
NPC&PD DIVISION

08-May-2017 18:32:14

MCR-A120-047_01A 117 (2.167) AM (Top,4, Ar,5000.0,297.21,0.70); Sb (2,10.00); Cm (105:137)

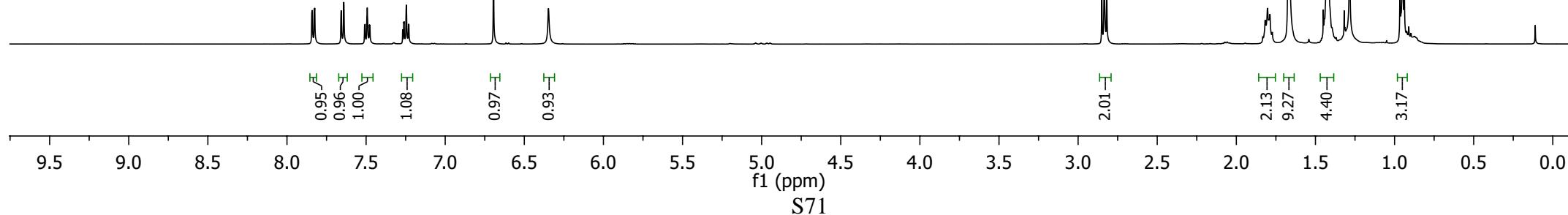
1: TOF MS ES+
8.51e5



Nov15-2016
MCR-A1

7.84206
7.84196
7.8269
7.8246
7.6573
7.6410
7.5093
7.5066
7.4926
7.4787
7.4759
7.2688
7.2617
7.2597
7.2458
7.2318
7.2298
6.6947

-6.3474



2.8501
2.8346
2.8189

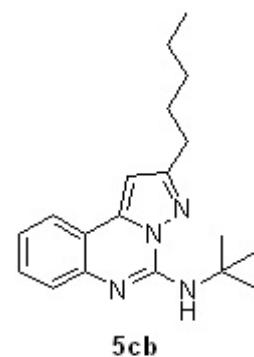
1.8030
1.7876

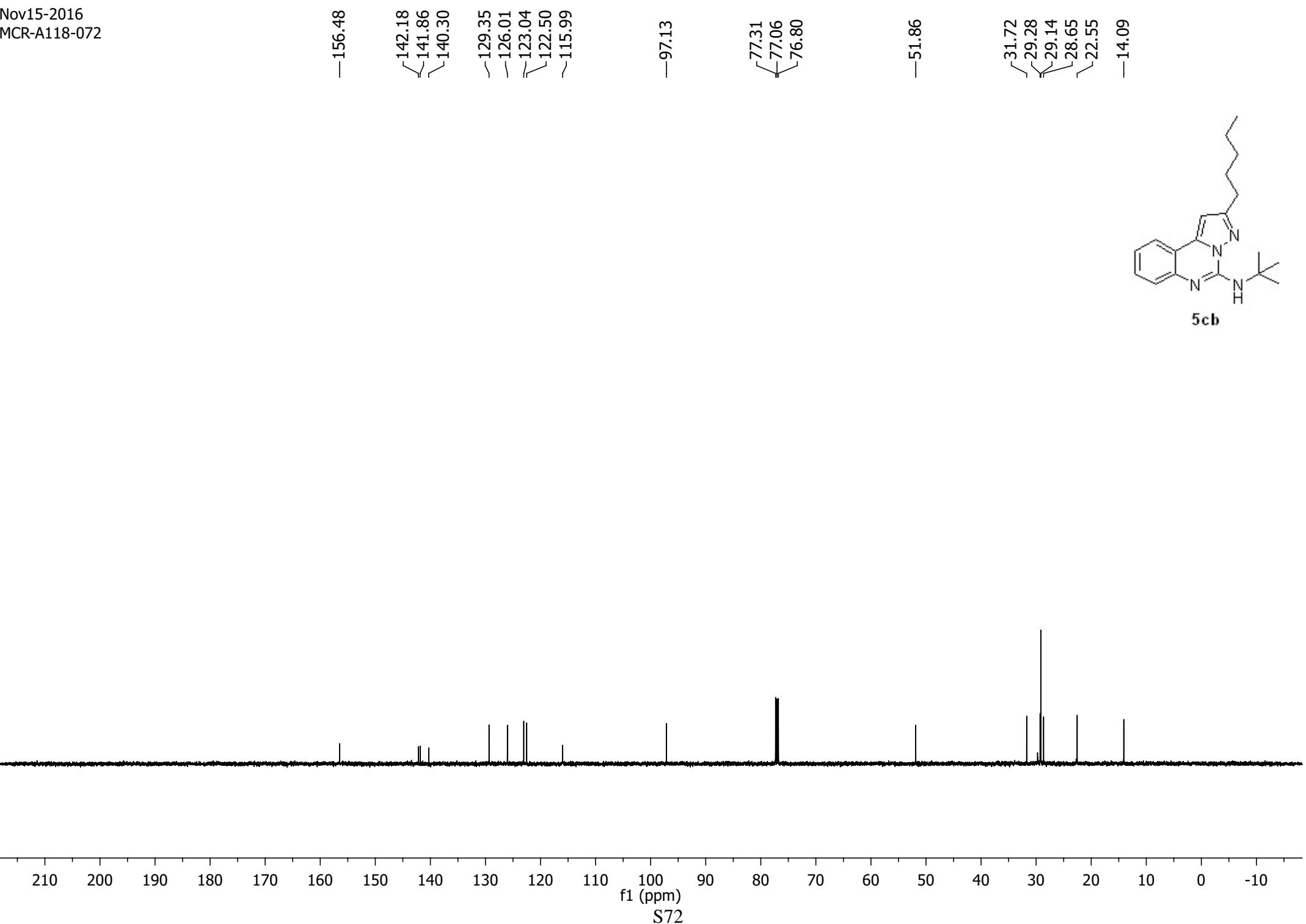
1.6699
1.4502

1.4438
1.4273

1.4202
0.9662

0.9522
0.9381





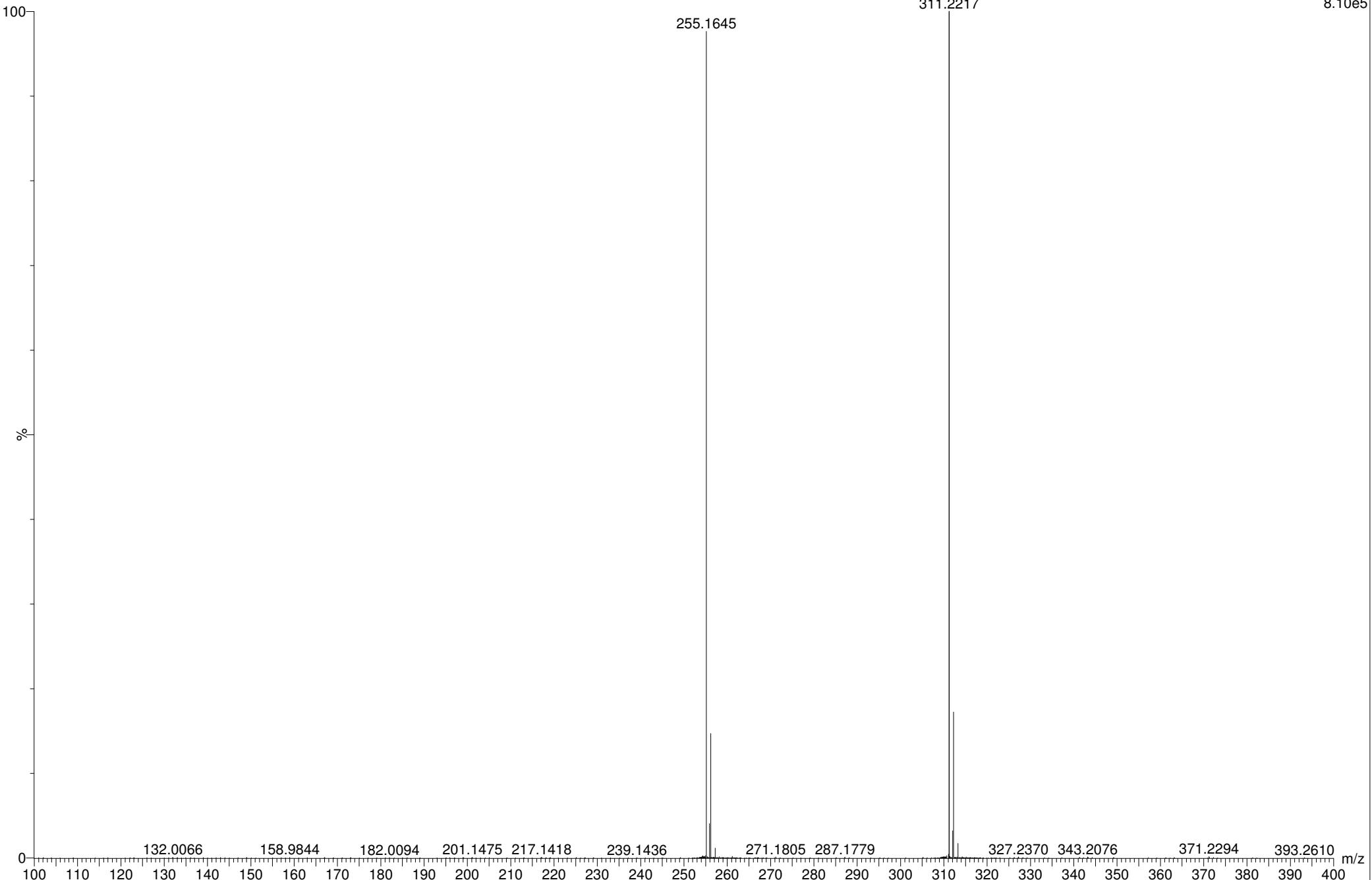
PROCESSED BY
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NPC&PD DIVISION

24-Nov-2016 17:07:38

MCR-A118-072 128 (2.375) AM (Cen,3, 80.00, Ar,5000.0,311.22,0.70); Sb (2,10.00); Cm (119:134)

1: TOF MS ES+
8.10e5



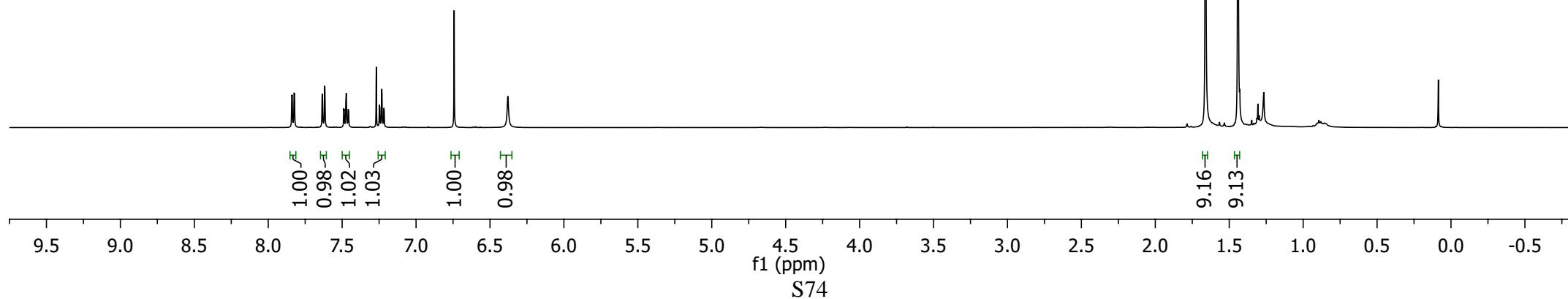
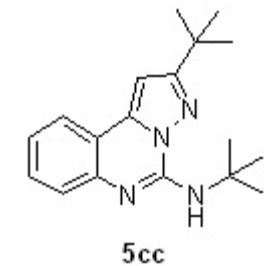
FID

MCR-A110

7.8199
7.8181
7.8243
7.8225
7.6343
7.6179
7.4889
7.4863
7.4724
7.4583
7.4557
7.2688
7.2475
7.2319
7.2176
6.7422

-6.3792

1.6603
1.4408
9.16
9.13



FID
MCR-A118-071

-164.67

142.30
141.78
139.94

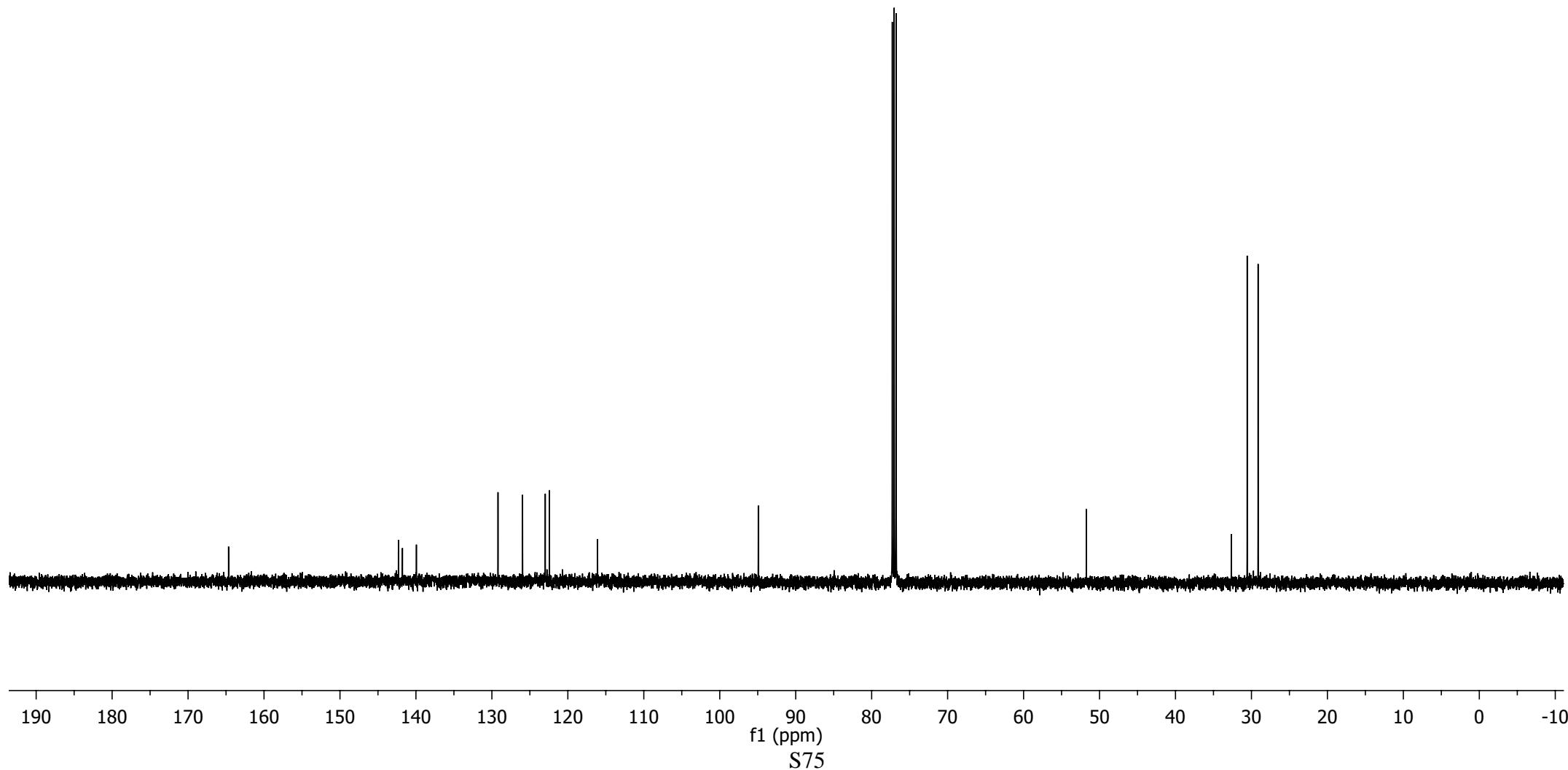
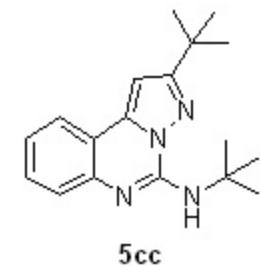
-129.19
-125.96
-122.97
-122.42
-116.09

-94.93

77.29
77.03
76.78

-51.74

32.65
30.56
29.13



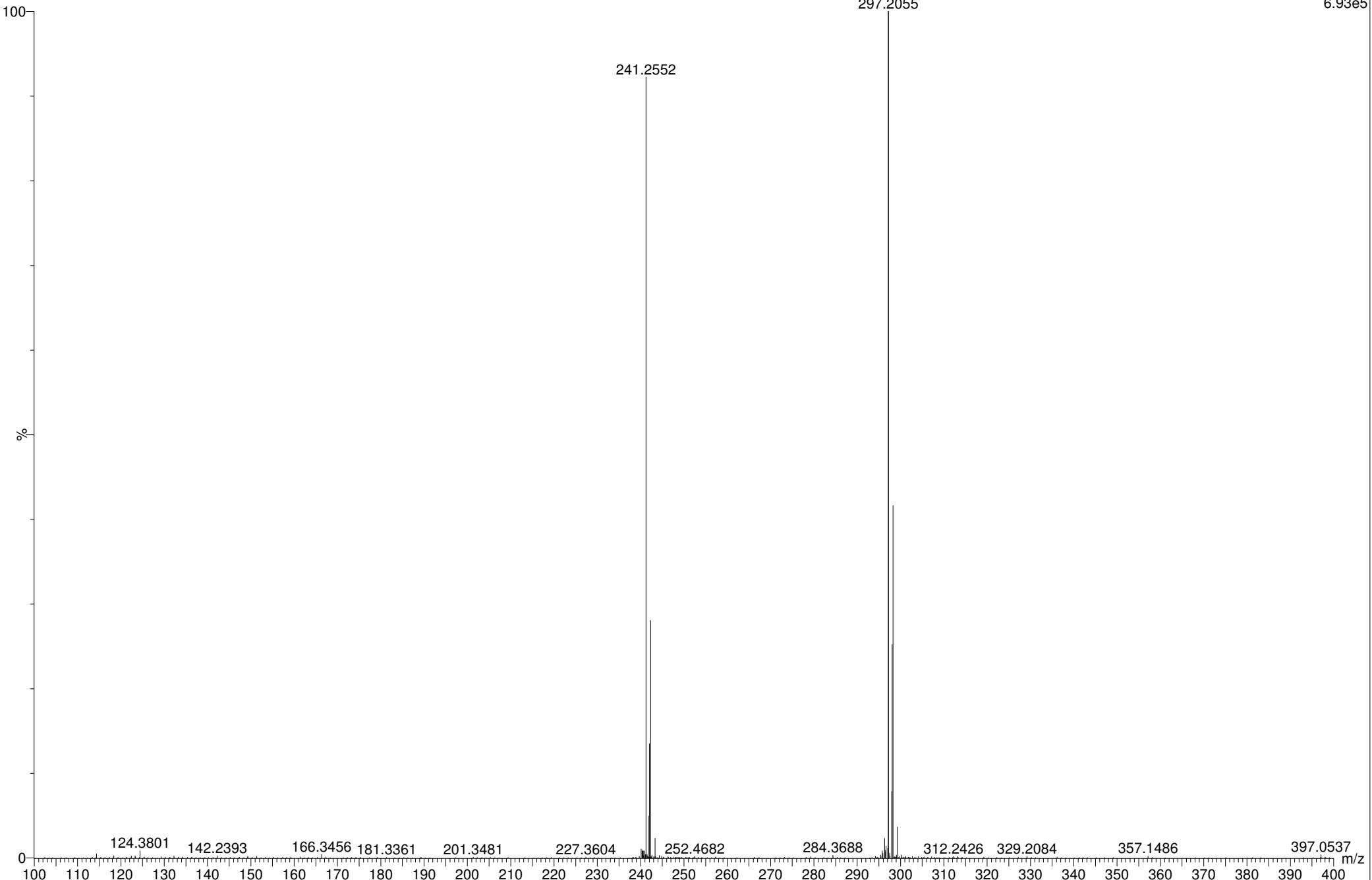
PROCESSED BY
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NPC&PD DIVISION

08-May-2017 17:55:40

MCR-A118-071_01A 114 (2.113) AM (Top,4, Ar,5000.0,297.21,0.70); Sb (2,10.00); Cm (104:128)

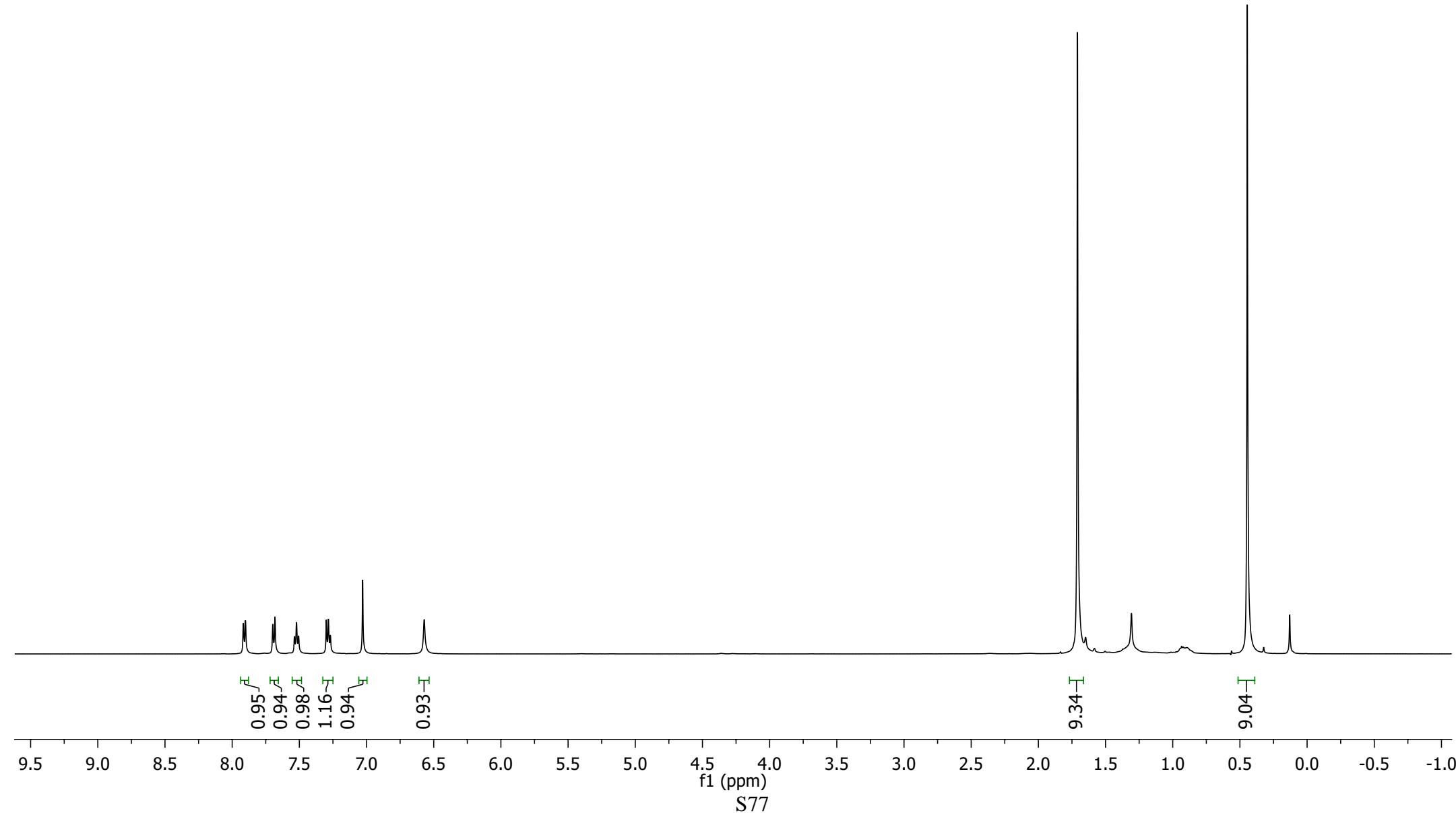
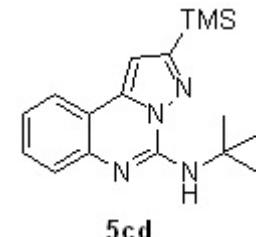
1: TOF MS ES+
6.93e5



7.9165
7.9011
7.6977
7.6815
7.5358
7.5211
7.5062
7.2994
7.2838
7.2691
7.0288
-6.5709

-1.7090

-0.4449



-156.13

142.37
141.83
139.69

-129.25
-125.92
-123.10
-122.56
-116.36

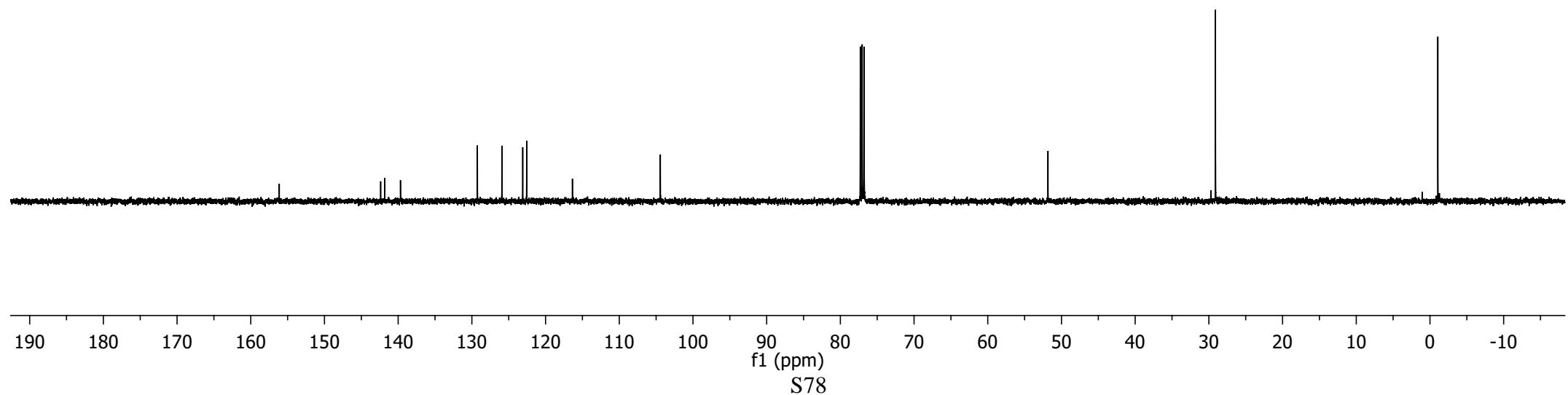
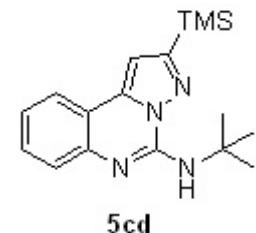
-104.43

77.29
77.04
76.79

-51.83

-29.10

-1.07



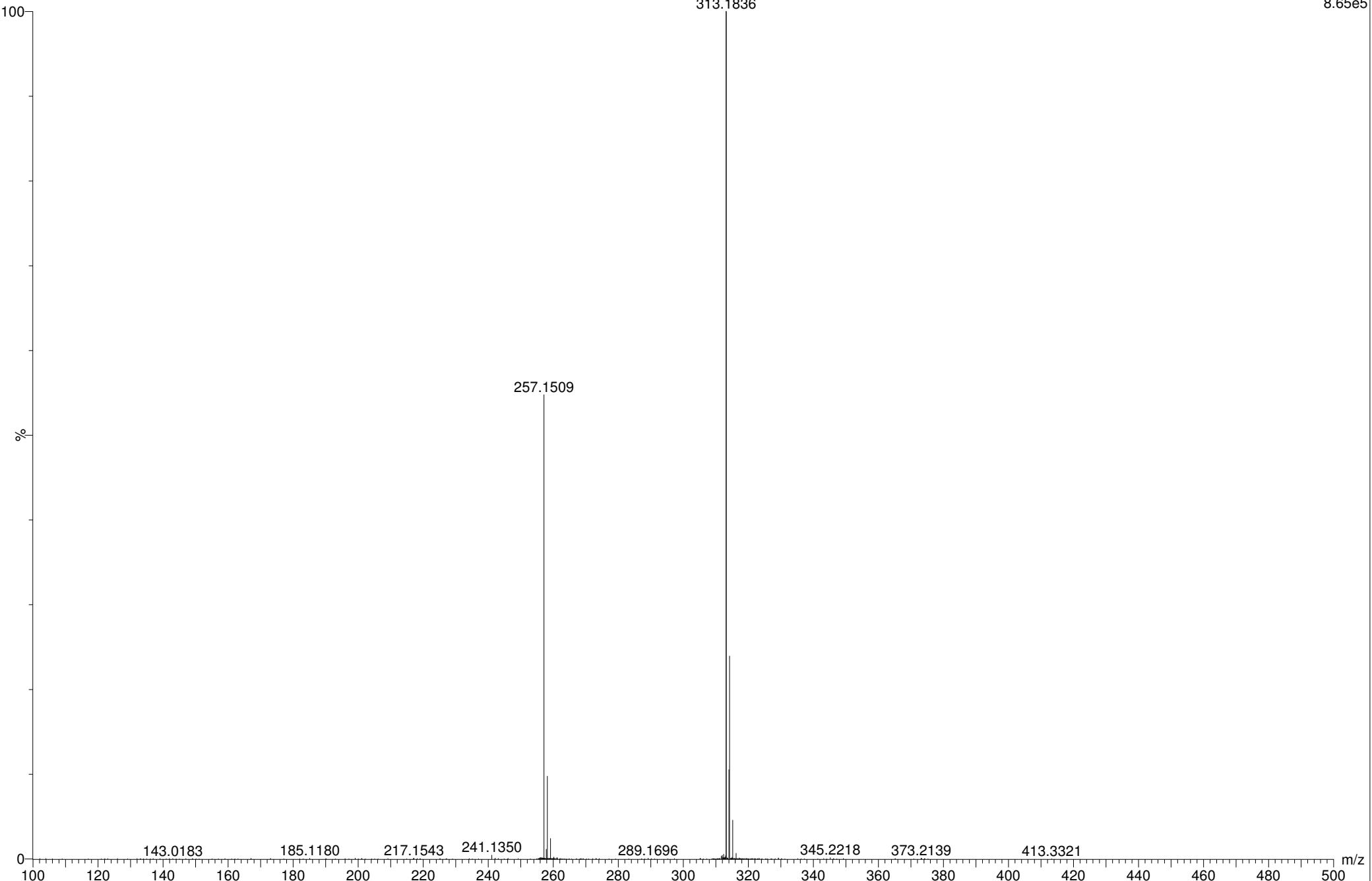
PROCESSED BY
PAWAN KUMAR

CSIR-IHBT
NPC&PD DIVISION

24-Nov-2016 16:35:00

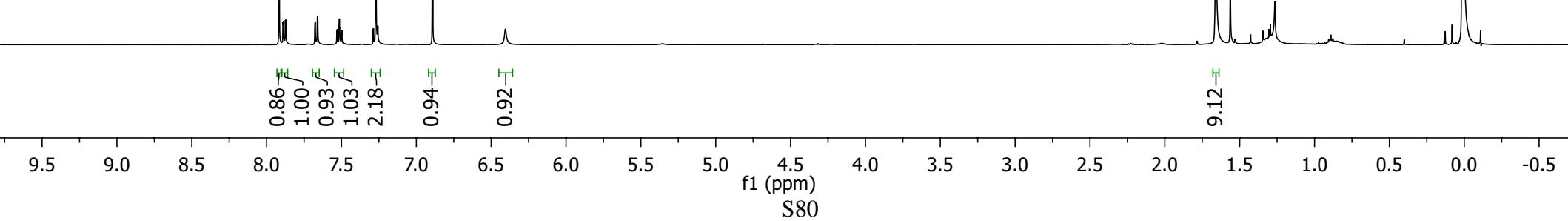
MCR-A118-70A 98 (1.818) AM (Cen,3, 80.00, Ar,5000.0,313.18,0.70); Sb (2,10.00); Cm (94:106)

1: TOF MS ES+
8.65e5

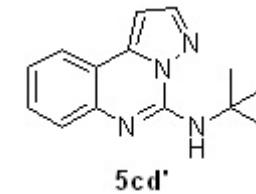


J_{AB} 1.16, 2.01, 1.7
M_{CH} 88
7.944
7.887
7.868
7.858
7.848
7.837
7.824
7.6759
7.6595
7.5281
7.5167
7.5141
7.5118
7.2888
7.2866
7.2689
7.2589
6.8946
6.8901

-6.4050



-1.6598



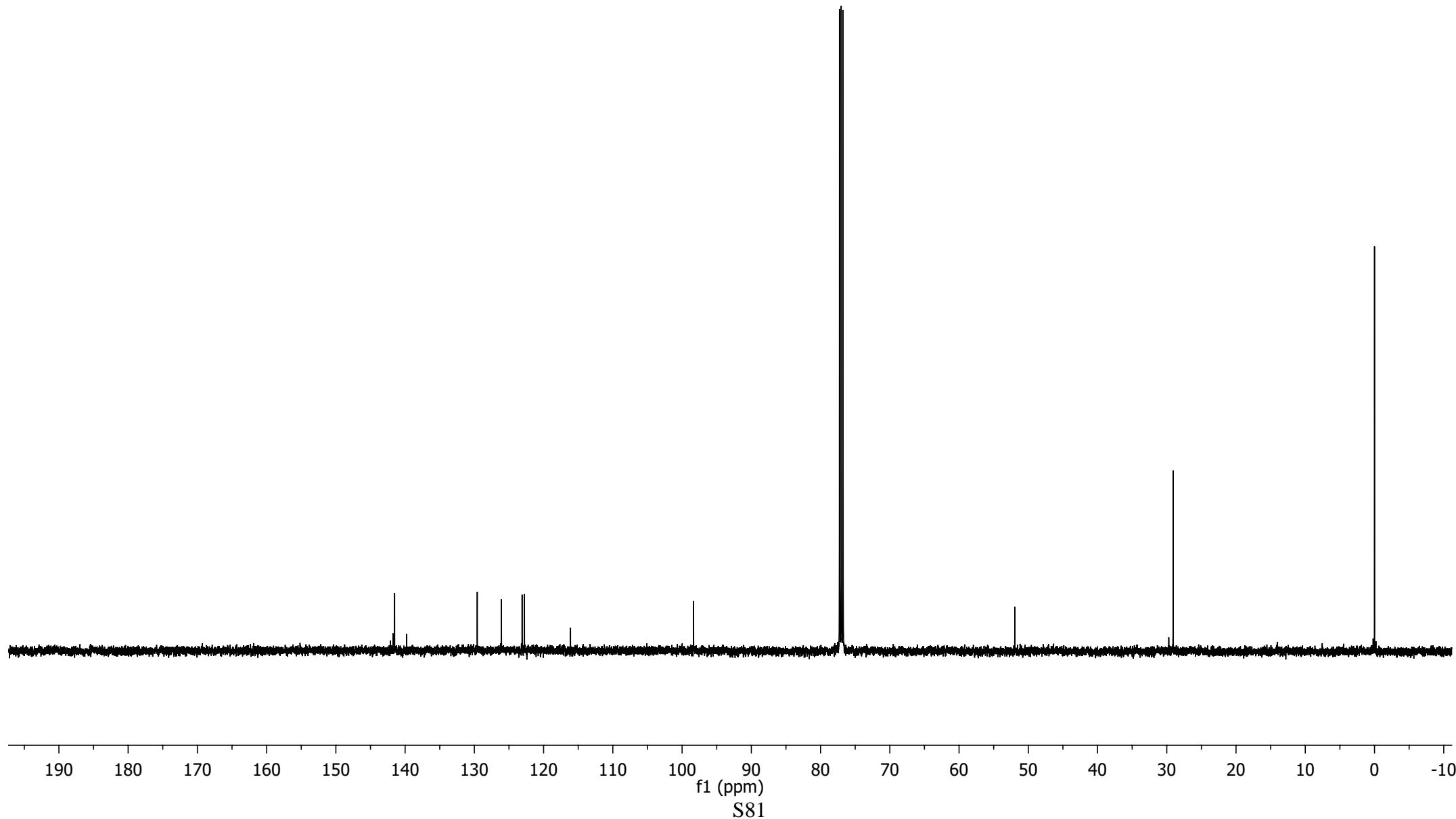
141.75
141.52
139.77
129.58
126.09
123.11
122.76
-116.11

-98.34

77.27
77.01
76.76

-51.95

-29.08



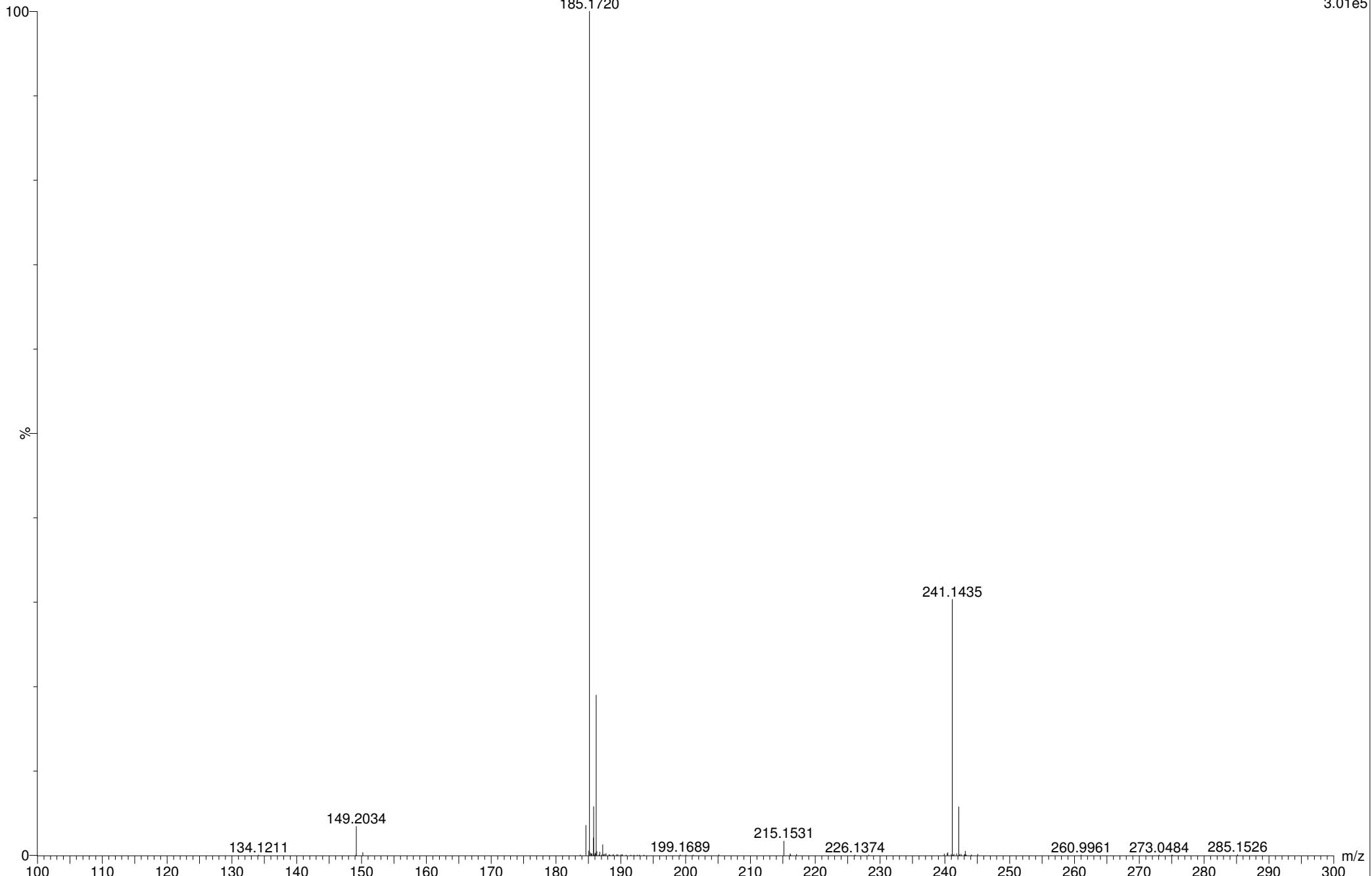
PROCESSED BY
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NPC&PD DIVISION

08-May-2017 18:36:16

MCR-A118-070B_01A 42 (0.778) AM (Top,4, Ar,5000.0,241.14,0.70); Sb (2,10.00); Cm (38:53-(64:69+25:33))

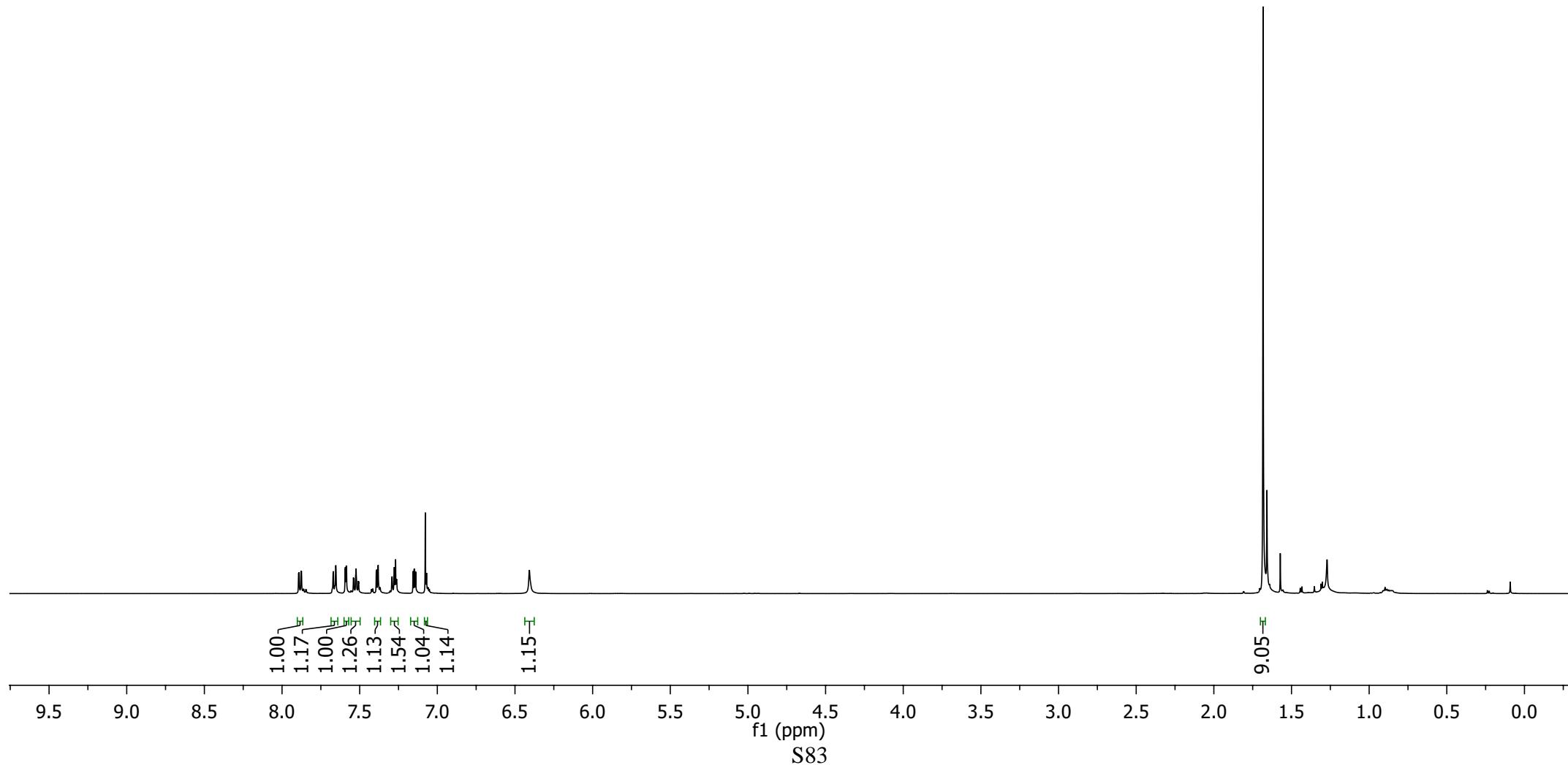
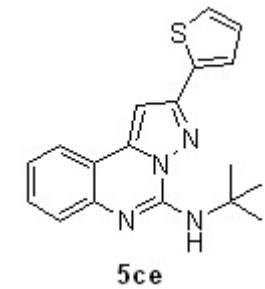
1: TOF MS ES+
3.01e5

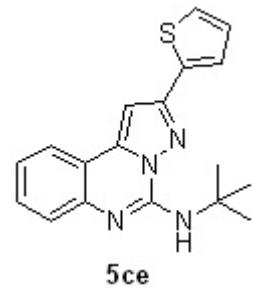
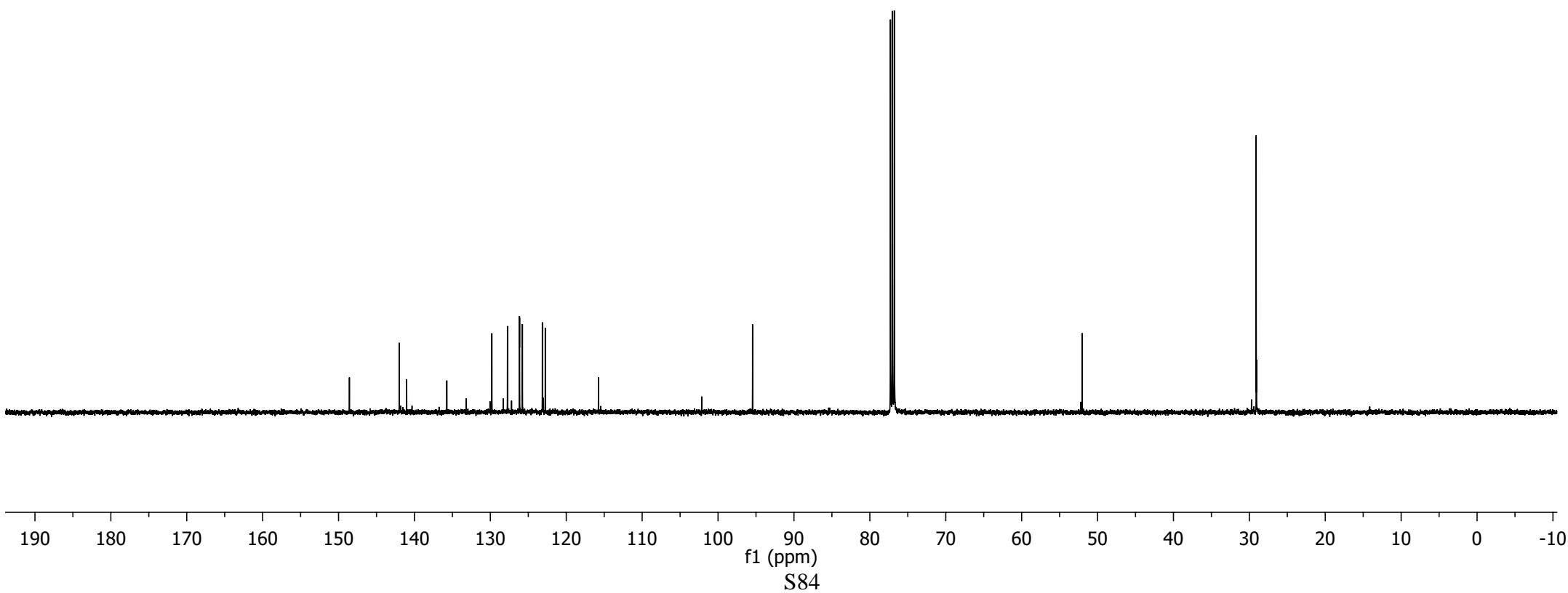


Dec03-2016
MCR-A174

7.8476
7.8474
7.8470
7.6536
7.5936
7.5920
7.5866
7.5849
7.5232
7.3927
7.3911
7.3826
7.3809
7.2772
7.2689
7.1565
7.1493
7.1466
7.1393
6.9074

-1.6831





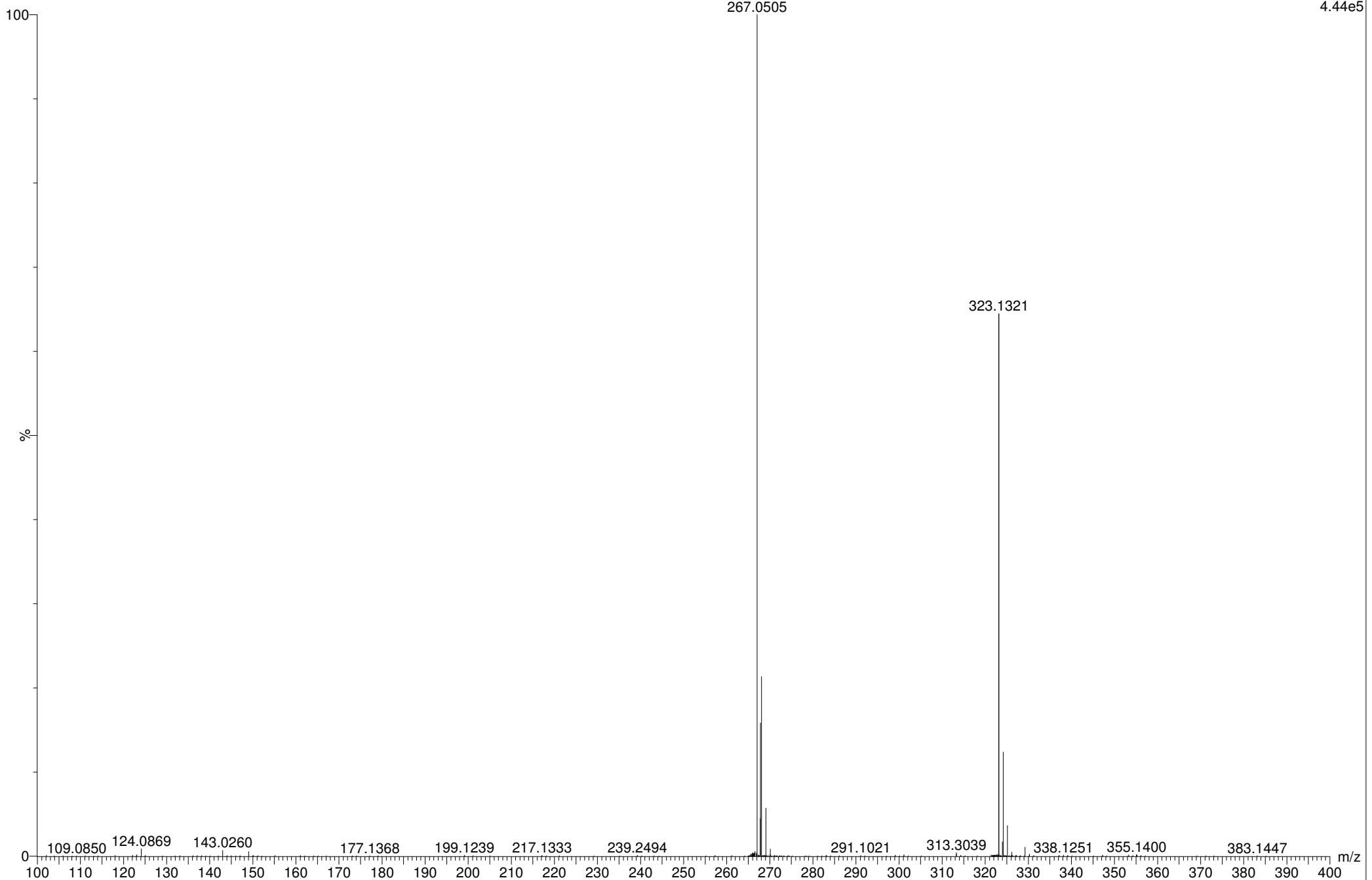
PROCESSED BY
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CSIR-IHBT
NPC&PD DIVISION

19-Jan-2017 14:01:11

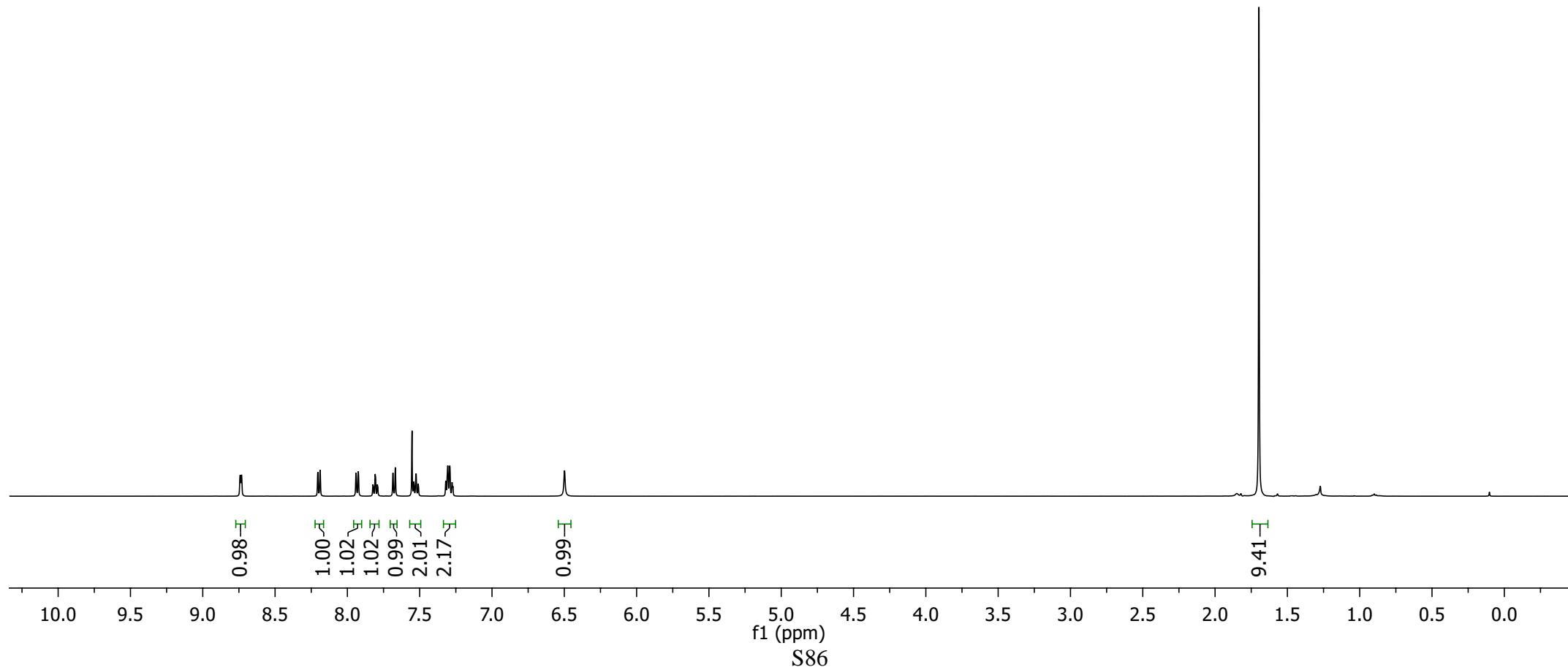
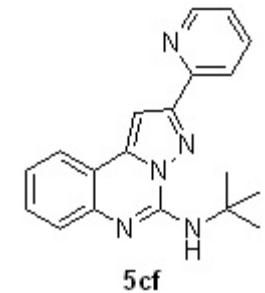
MCR-A118-103_02 41 (0.760) AM (Cen,3, 80.00, Ar,5000.0,323.13,0.70); Sb (2,10.00); Cm (34:42)

1: TOF MS ES+
4.44e5



8.7414
8.7319
8.2038
8.1880
7.9402
7.9245
7.8079
7.6849
7.6685
7.5528
7.5247
7.3059
6.4985

-1.6964



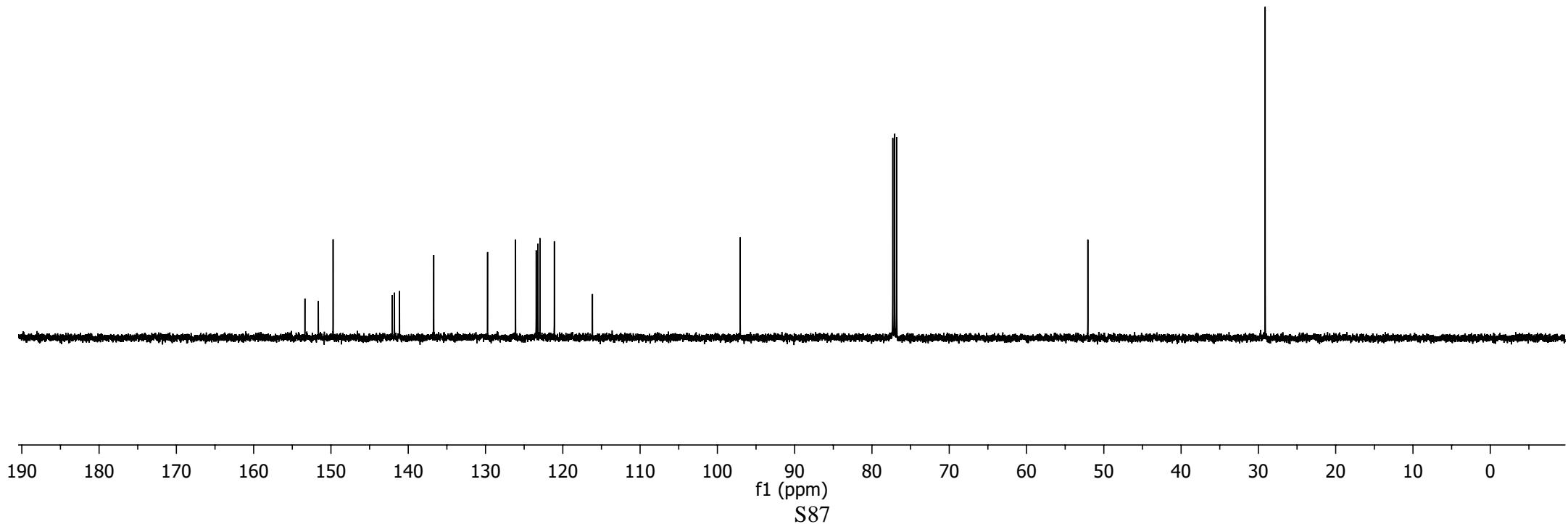
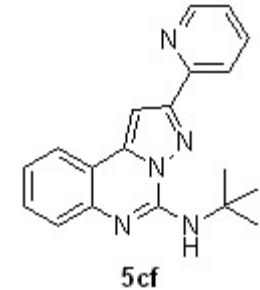
153.37
151.63
149.72
142.07
141.78
141.13
136.70
129.74
126.11
123.44
123.24
122.96
121.10
116.16

—97.05

77.30
77.05
76.80

—52.07

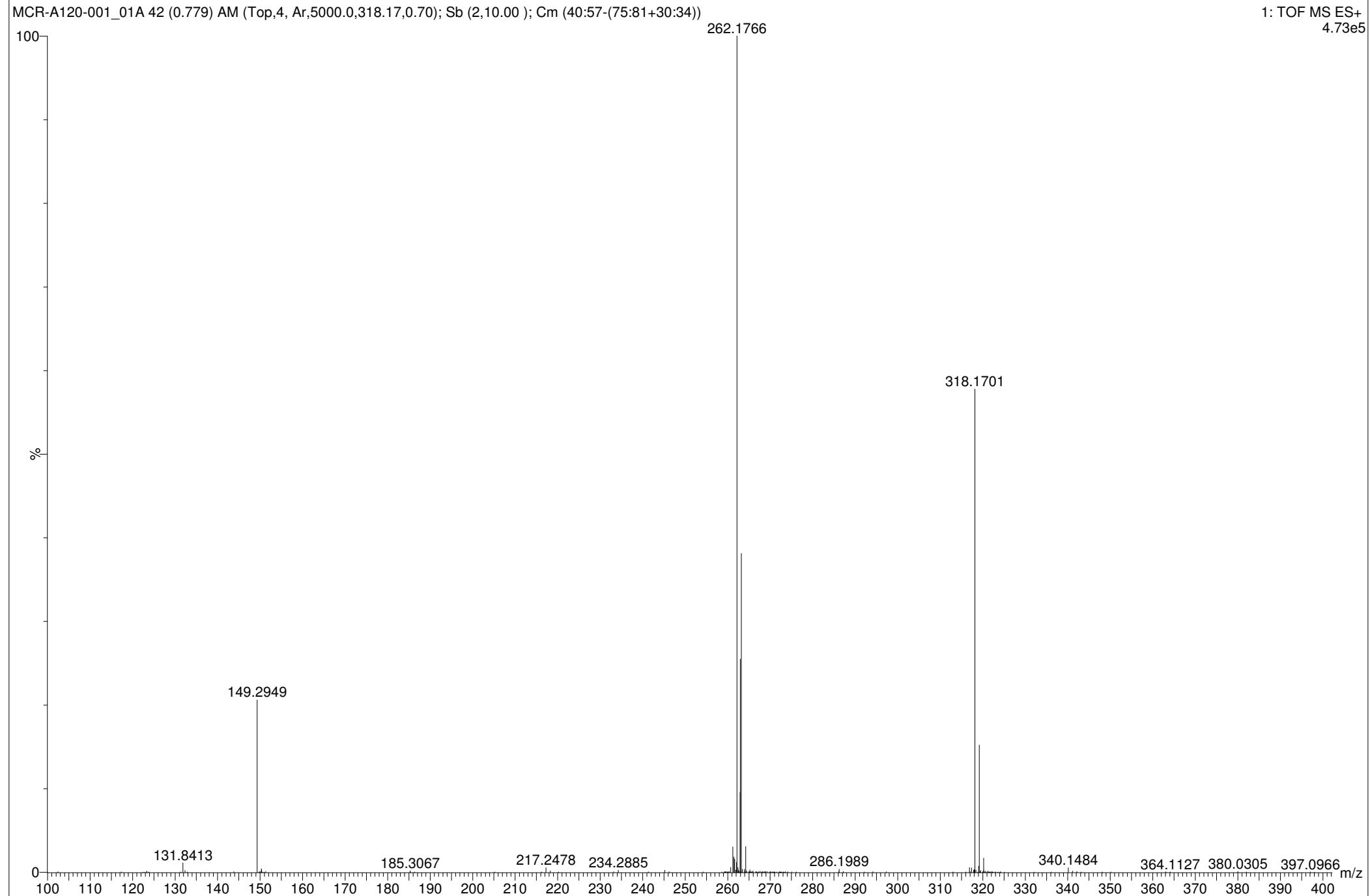
—29.12



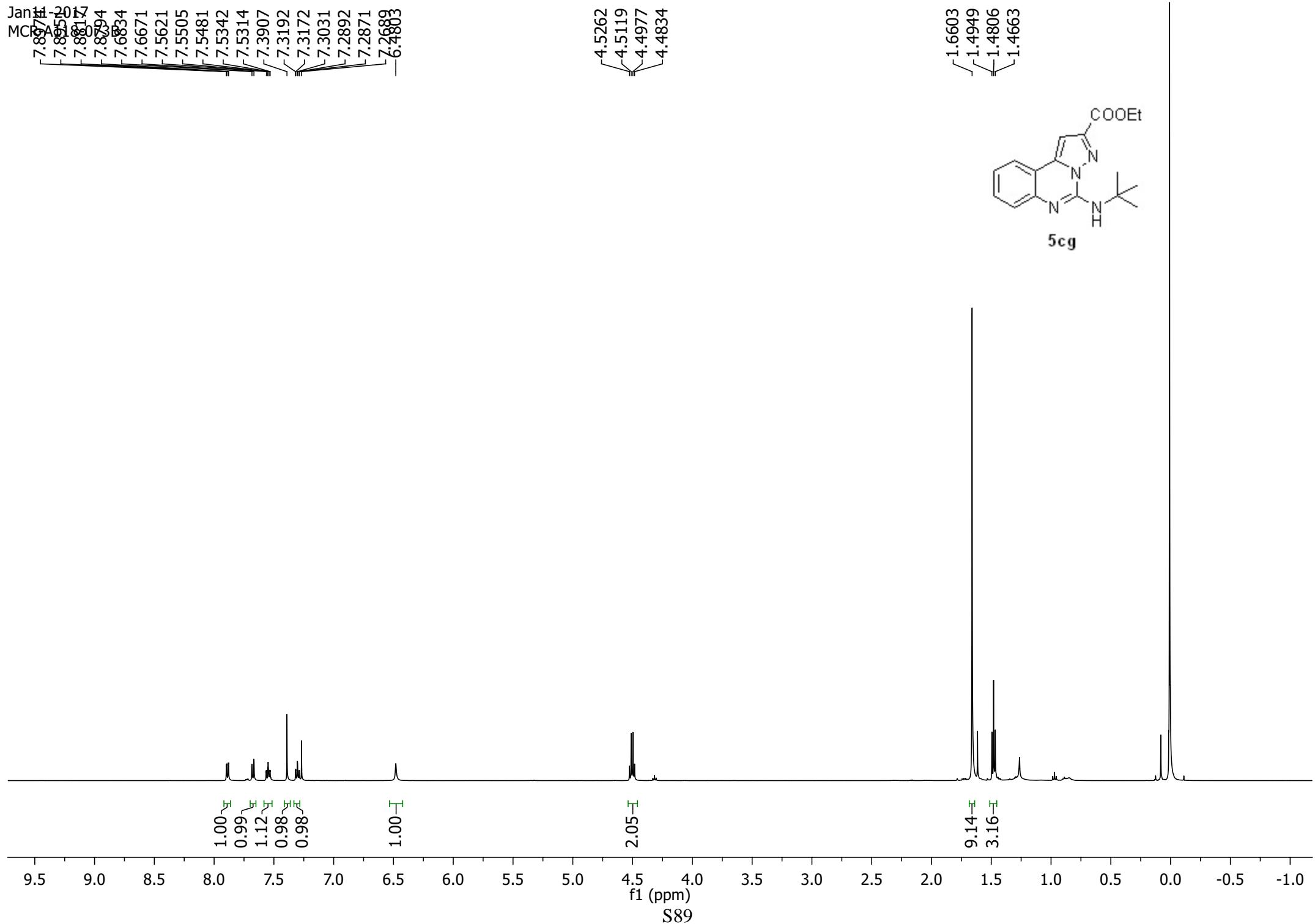
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08-May-2017 17:59:48



MCR-A-1 8073
88 88 88



-162.45

145.05
141.67
141.60
140.92
130.22
126.26
123.26
123.14
-115.77

-100.78

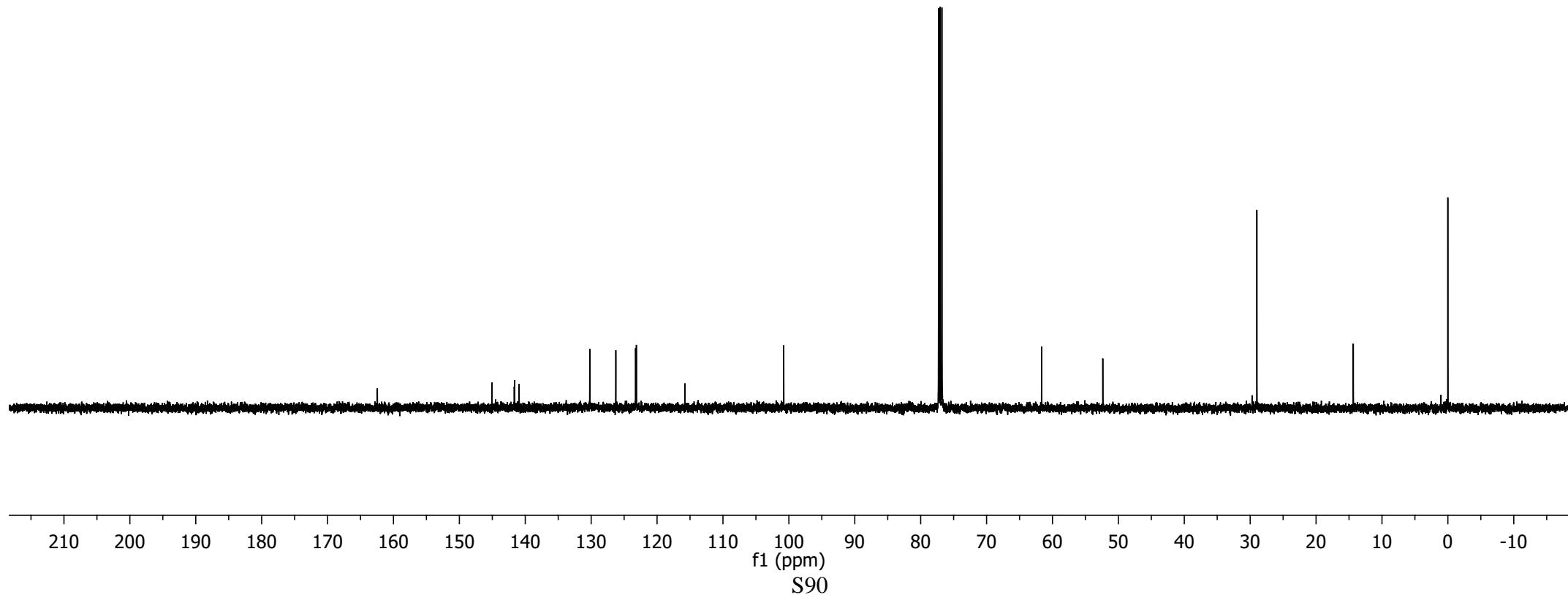
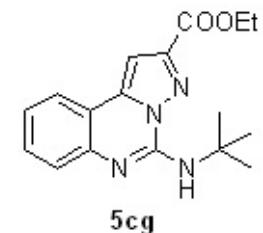
77.28
77.03
76.77

-61.65

-52.36

-28.98

-14.35



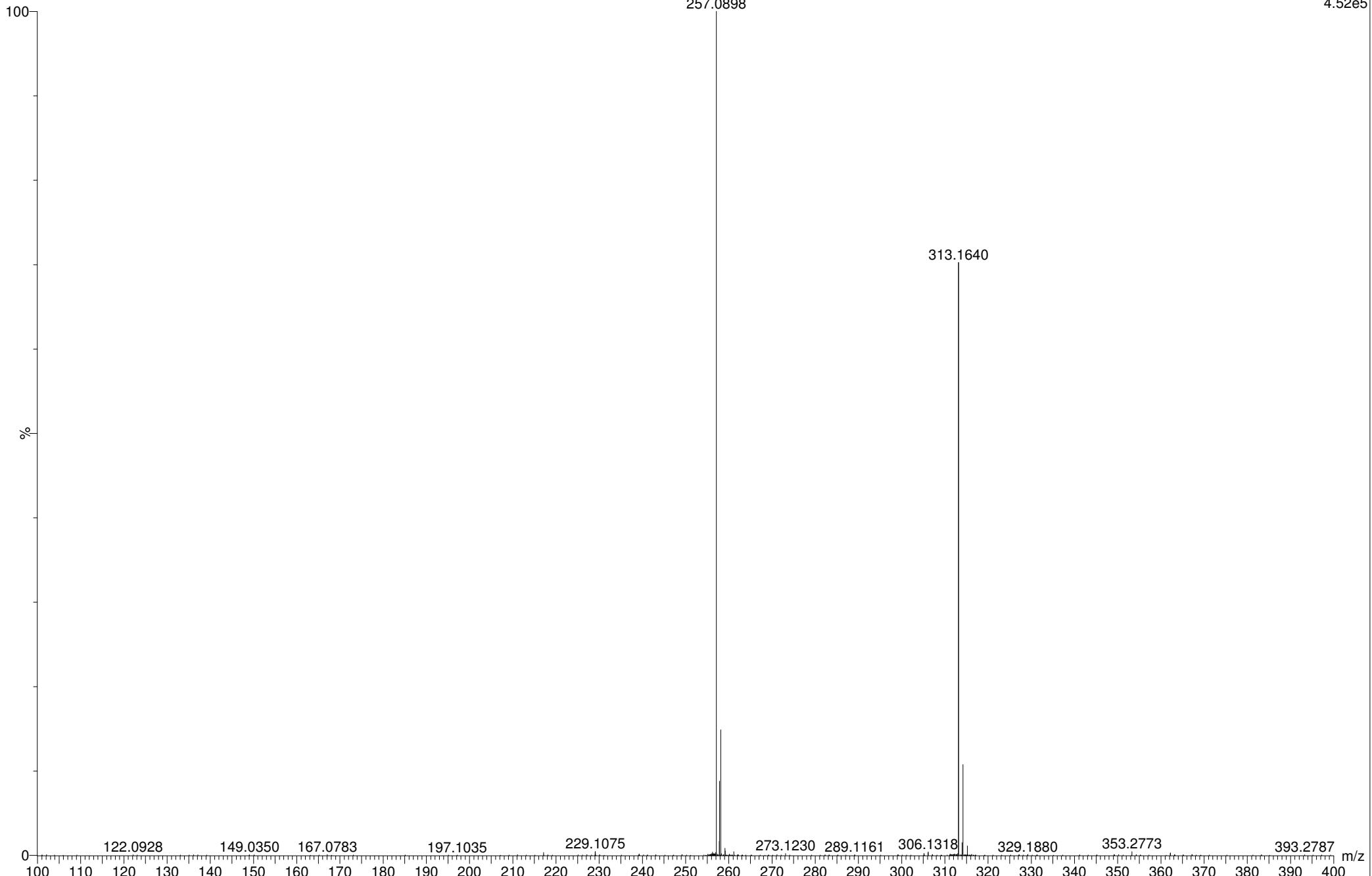
PROCESSED BY
PAWAN KUMAR

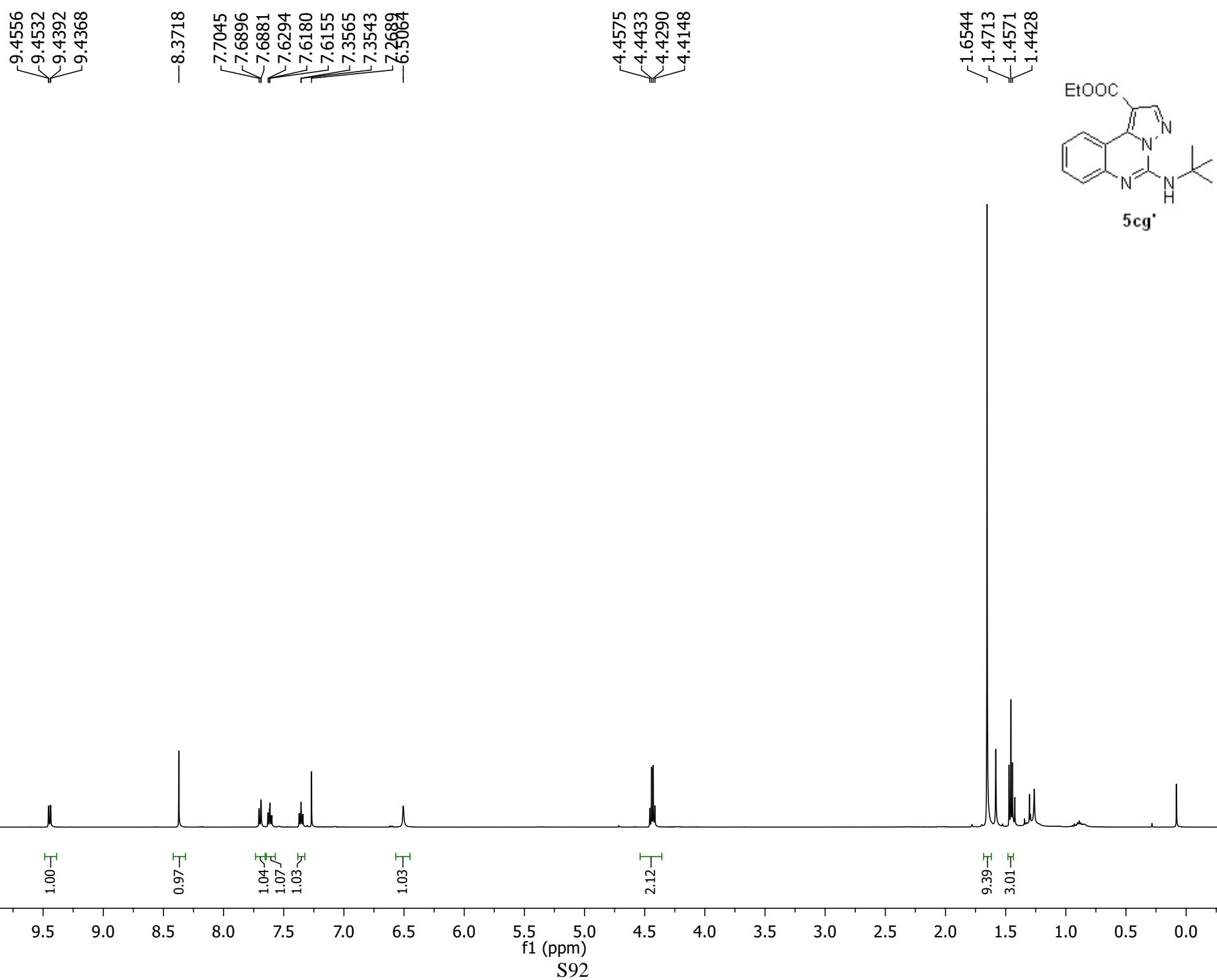
CSIR-IHBT
NPC&PD DIVISION

24-Nov-2016 17:11:42

MCR-A118-073A 37 (0.687) AM (Cen,3, 80.00, Ar,5000.0,313.16,0.70); Sb (2,10.00); Cm (35:41)

1: TOF MS ES+
4.52e5





—163.33

—144.76
—143.47
—141.97
—140.72

—131.27
—127.28
—125.98
—123.06

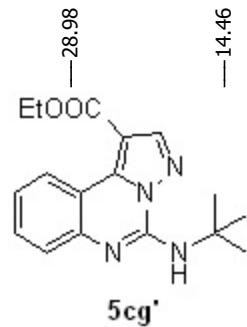
—115.53

—108.91

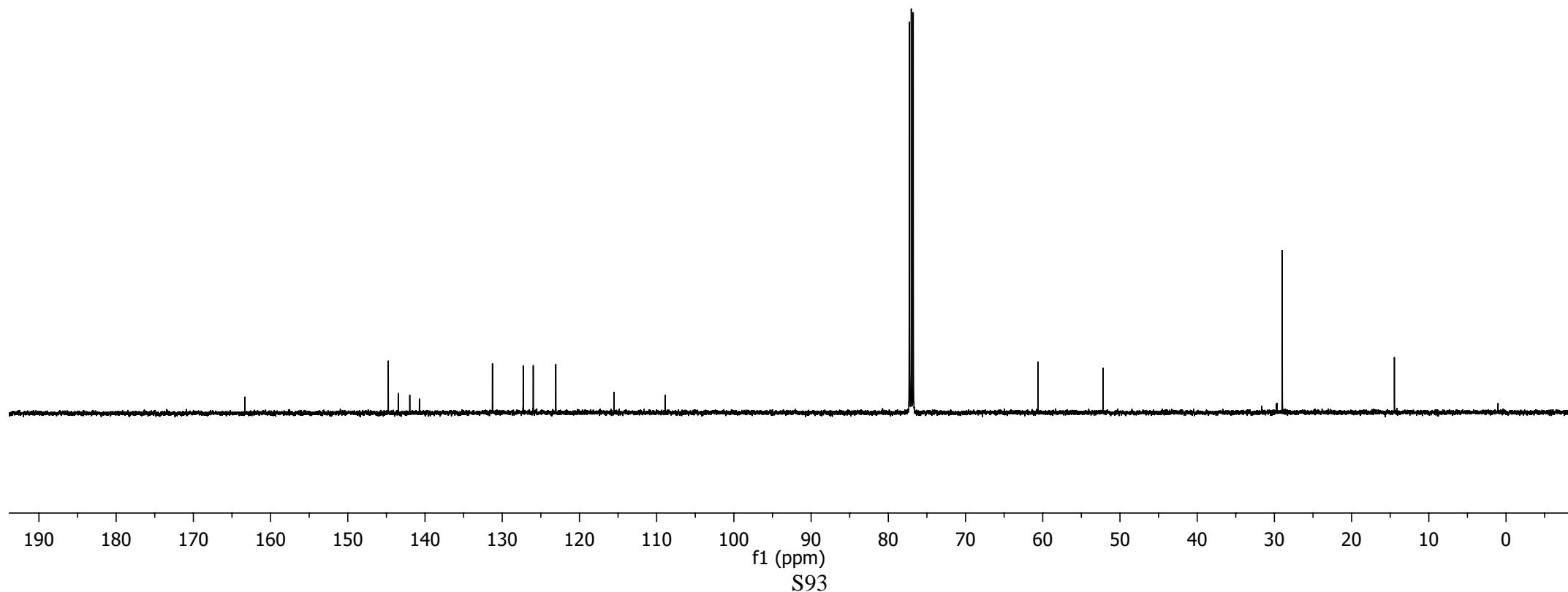
77.28
77.03
76.77

—60.62

—52.18



—14.46



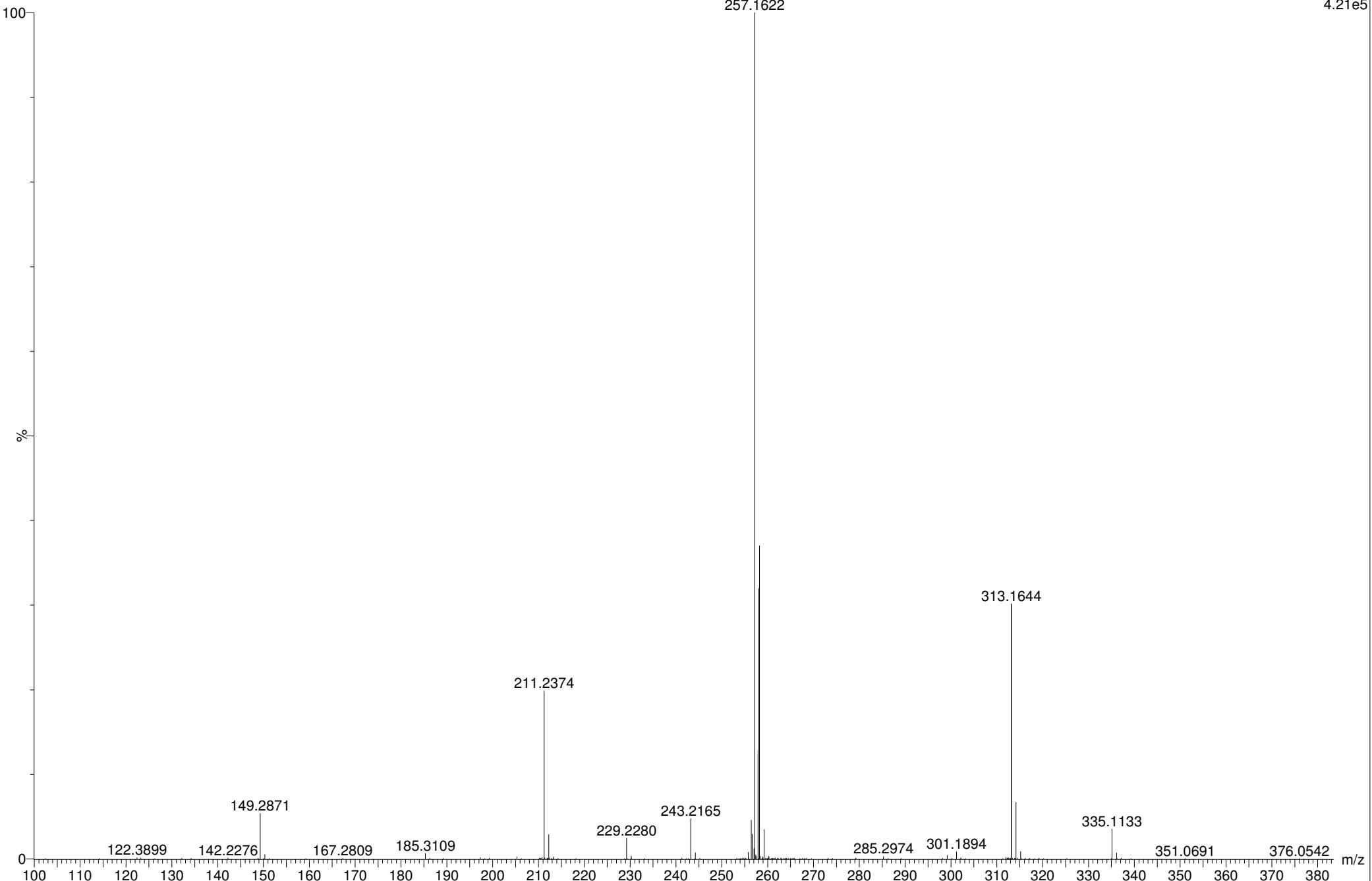
PROCESSED BY
PAWAN KUMAR

CSIR-IHBT
NPC&PD DIVISION

08-May-2017 18:24:13

MCR-A118-073B_01A 44 (0.816) AM (Top,4, Ar,5000.0,313.16,0.70); Sb (2,10.00); Cm (39:54-(71:75+24:28))

1: TOF MS ES+
4.21e5



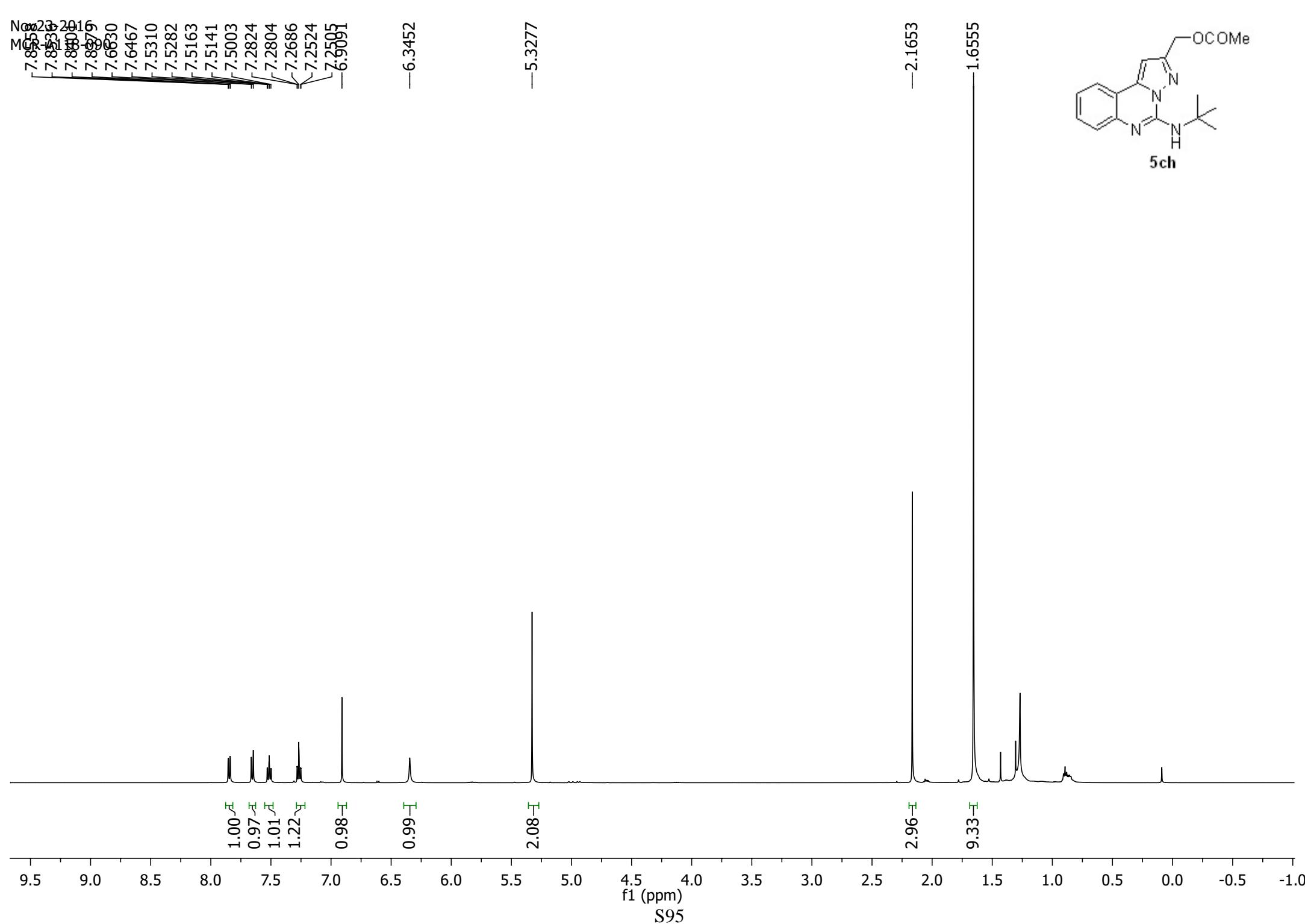
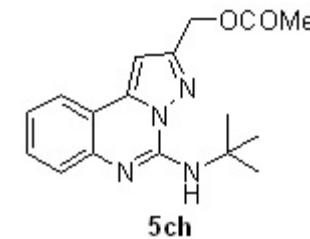
Noo236 2016
M95
7.8536
7.8507
7.8476
7.6630
7.6467
7.5310
7.5282
7.5163
7.5141
7.5003
7.2824
7.2804
7.2686
7.2524
7.2505

-6.3452

-5.3277

-2.1653

1.6555



-170.74

-150.06
141.87
141.77
140.78

-129.77
126.13
123.12
122.86

-115.83

-98.15

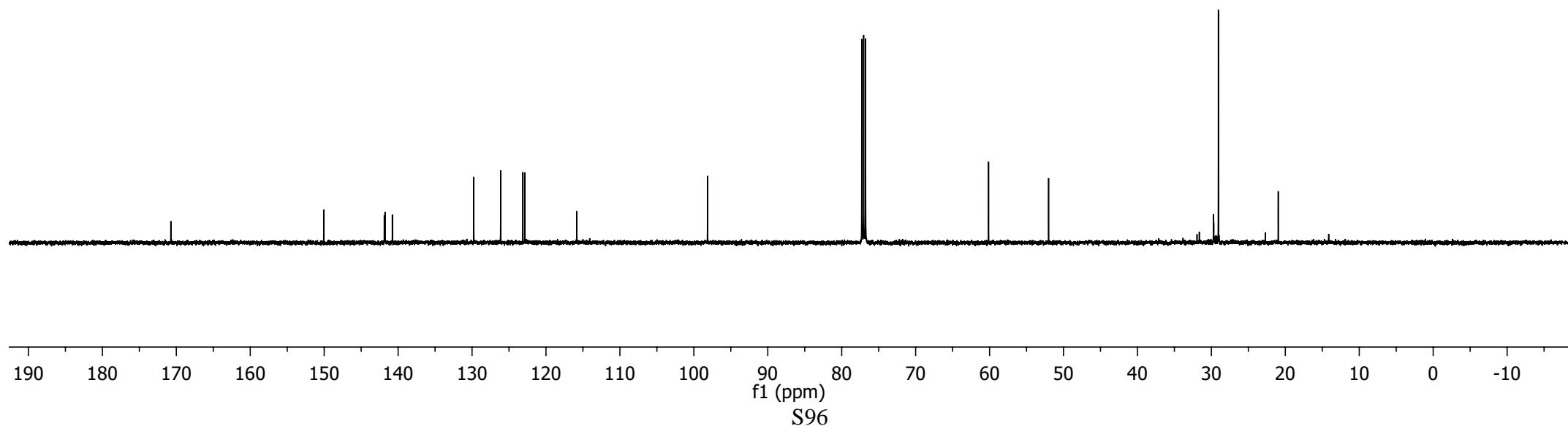
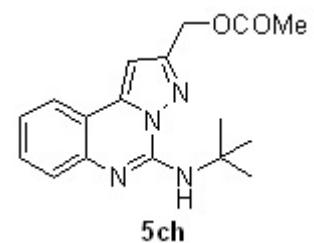
77.29
77.03
76.78

-60.13

-52.02

-29.05

-20.96



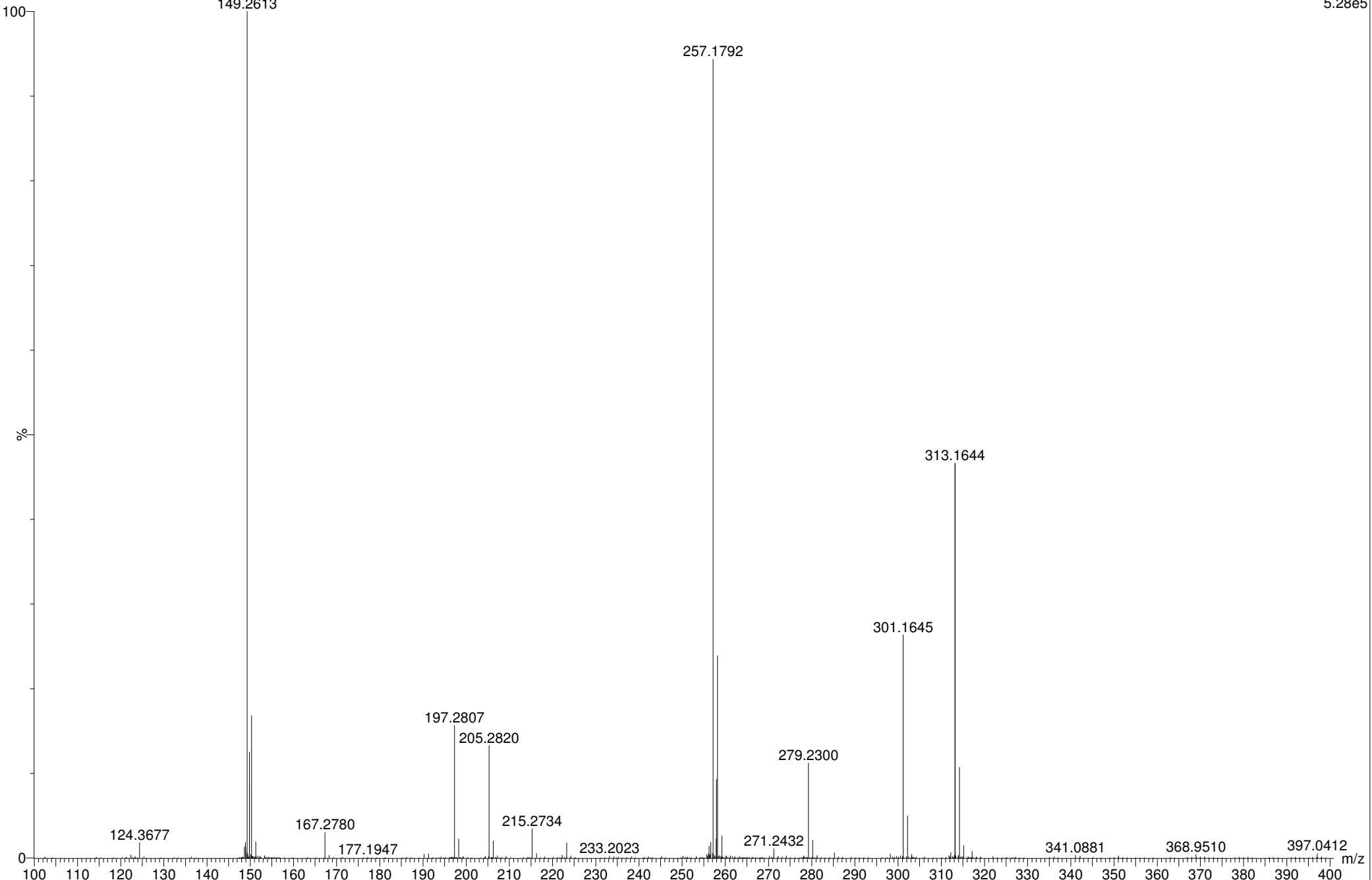
PROCESSED BY
PAWAN KUMAR

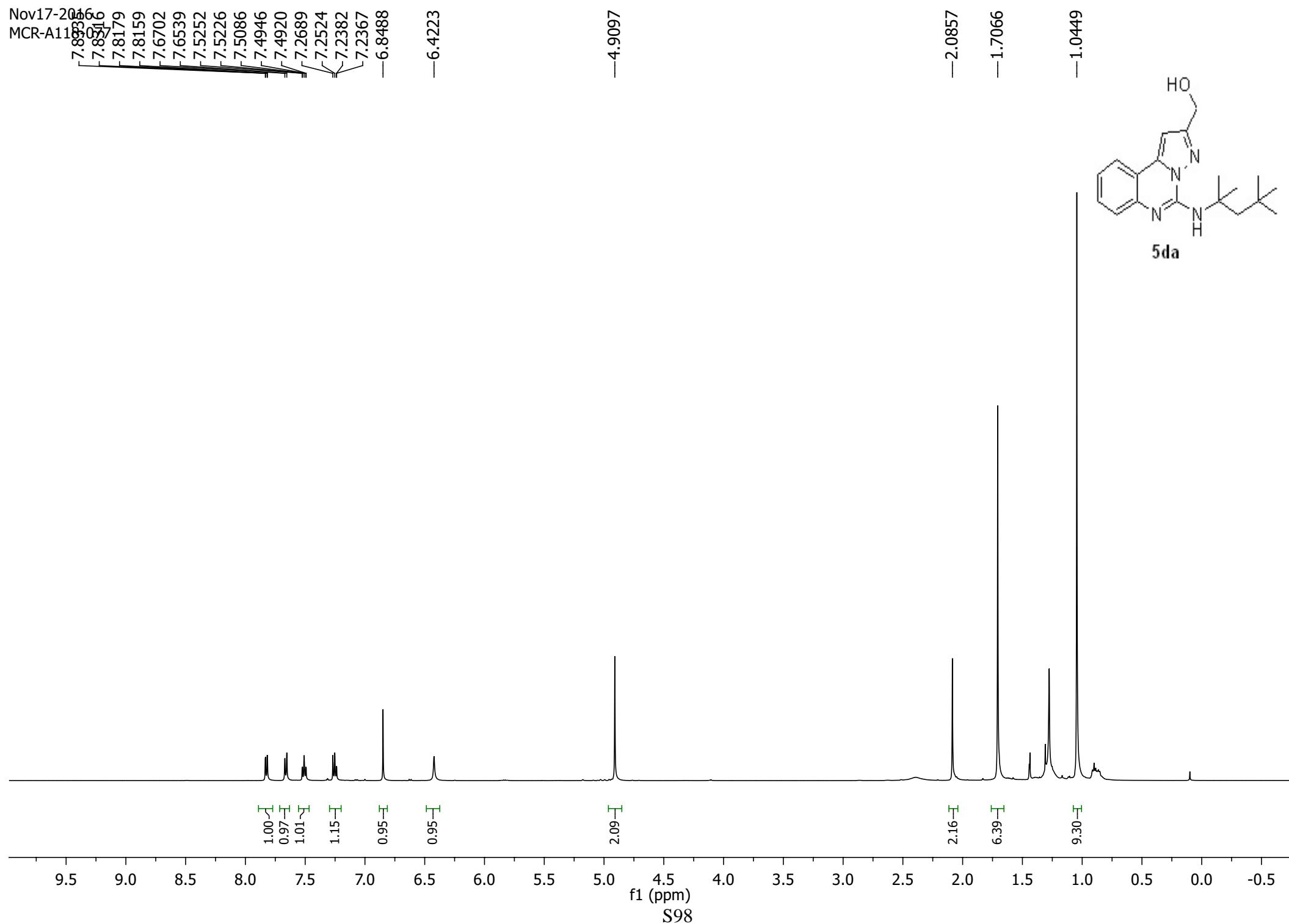
CSIR-IHBT
NPC&PD DIVISION

08-May-2017 17:47:37

MCR-A118-090_01A 38 (0.705) AM (Top,4, Ar,5000.0,313.16,0.70); Sb (2,10.00); Cm (31:61-(83:92+18:24))

1: TOF MS ES+
5.28e5





—154.67

141.85
141.78
140.73

129.68
126.07
123.10
122.71

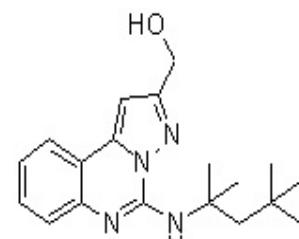
—115.83

—96.45

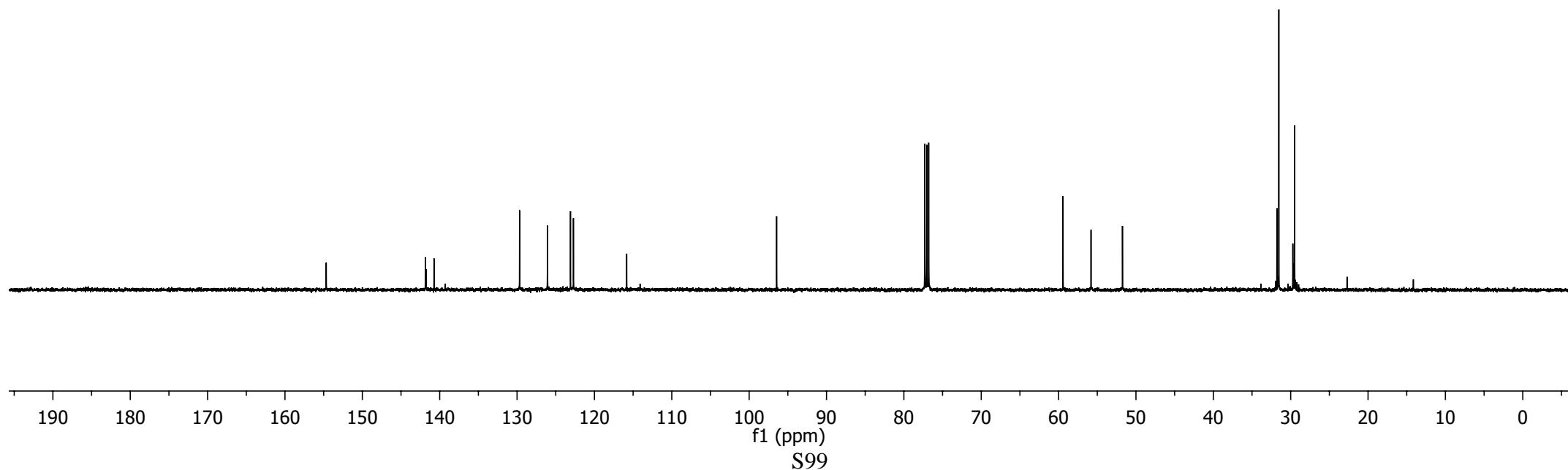
77.30
77.05
76.79

~59.43
~55.81
~51.75

31.78
31.53
29.49



5da



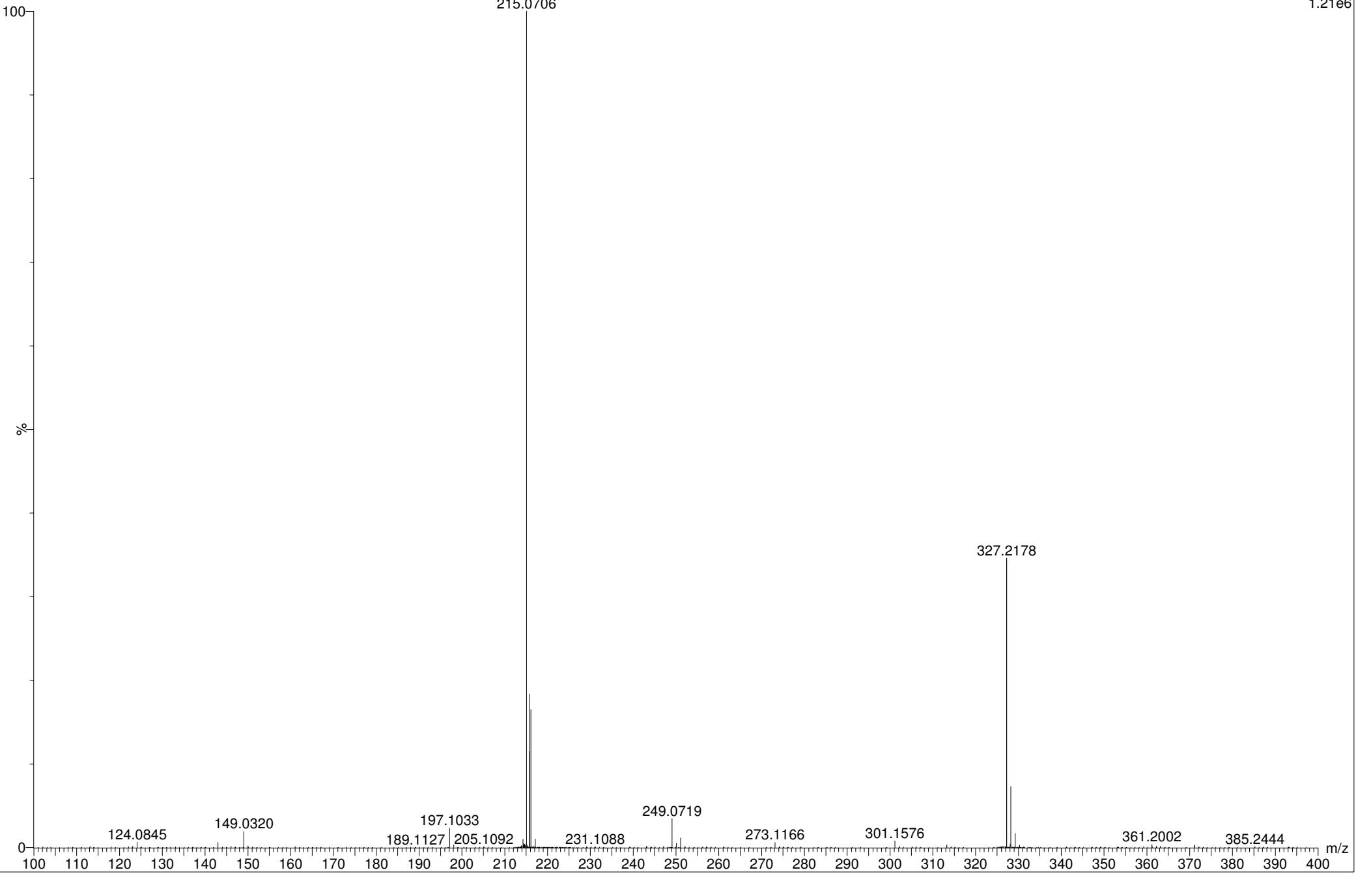
PROCESSED BY
PAWAN KUMAR

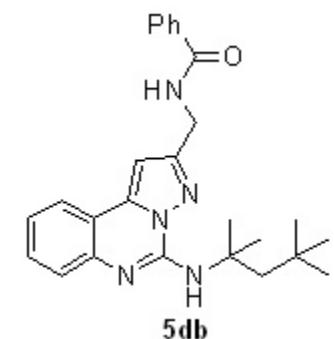
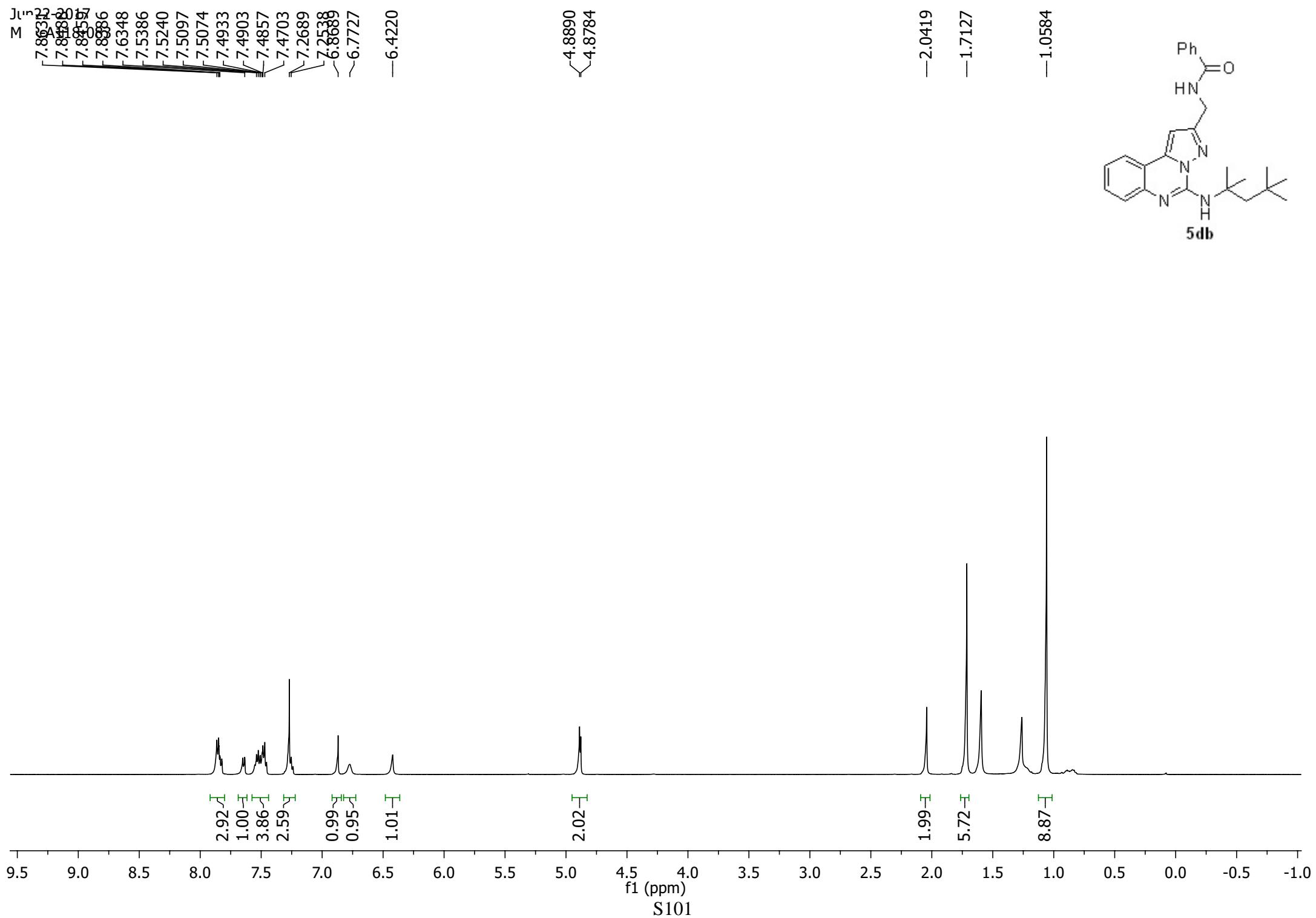
CSIR-IHBT
NPC&PD DIVISION

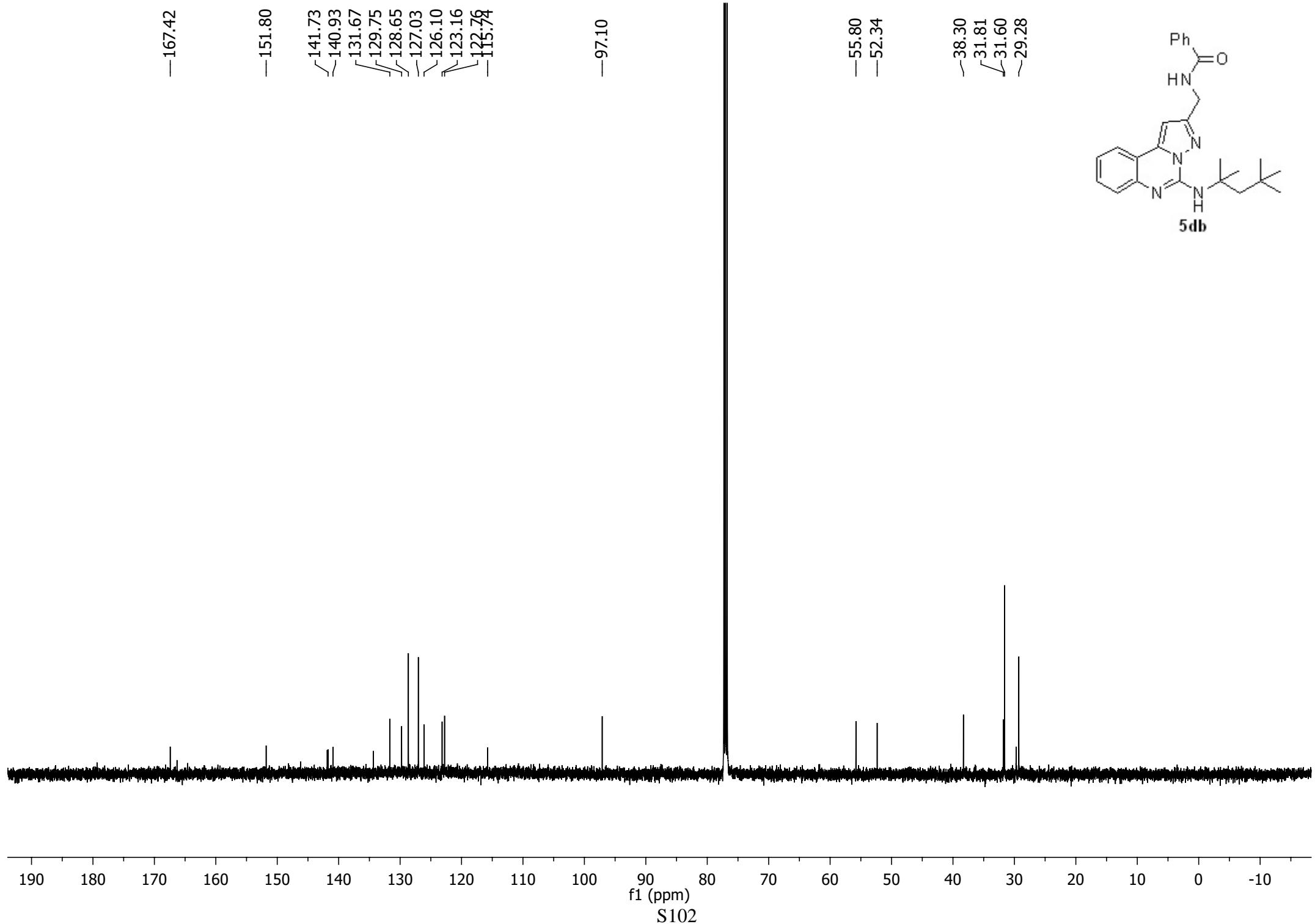
19-Jan-2017 14:49:04

MCR-A118-077_02 30 (0.556) AM (Cen,3, 80.00, Ar,5000.0,327.22,0.70); Sb (2,10.00); Cm (25:43)

1: TOF MS ES+
1.21e6







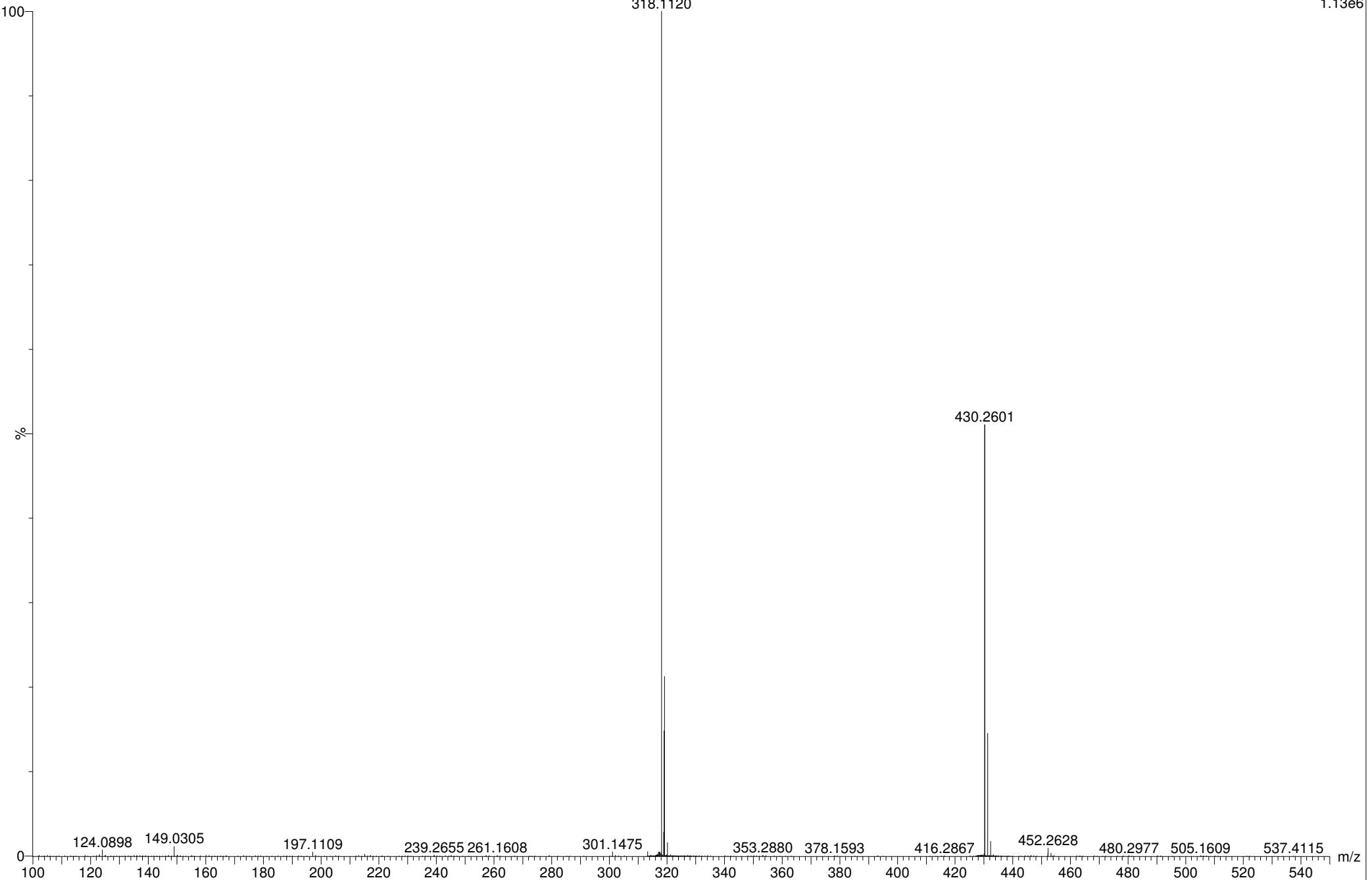
PROCESSED BY
PAWAN KUMAR

MCR-A118-083_02 33 (0.612) AM (Cen,3, 80.00, Ar,5000.0,430.26,0.70); Sb (2,10.00); Cm (28:45)

CSIR-IHBT
NPC&PD DIVISION

19-Jan-2017 14:53:04

1: TOF MS ES+
1.13e6



Nov22-2016

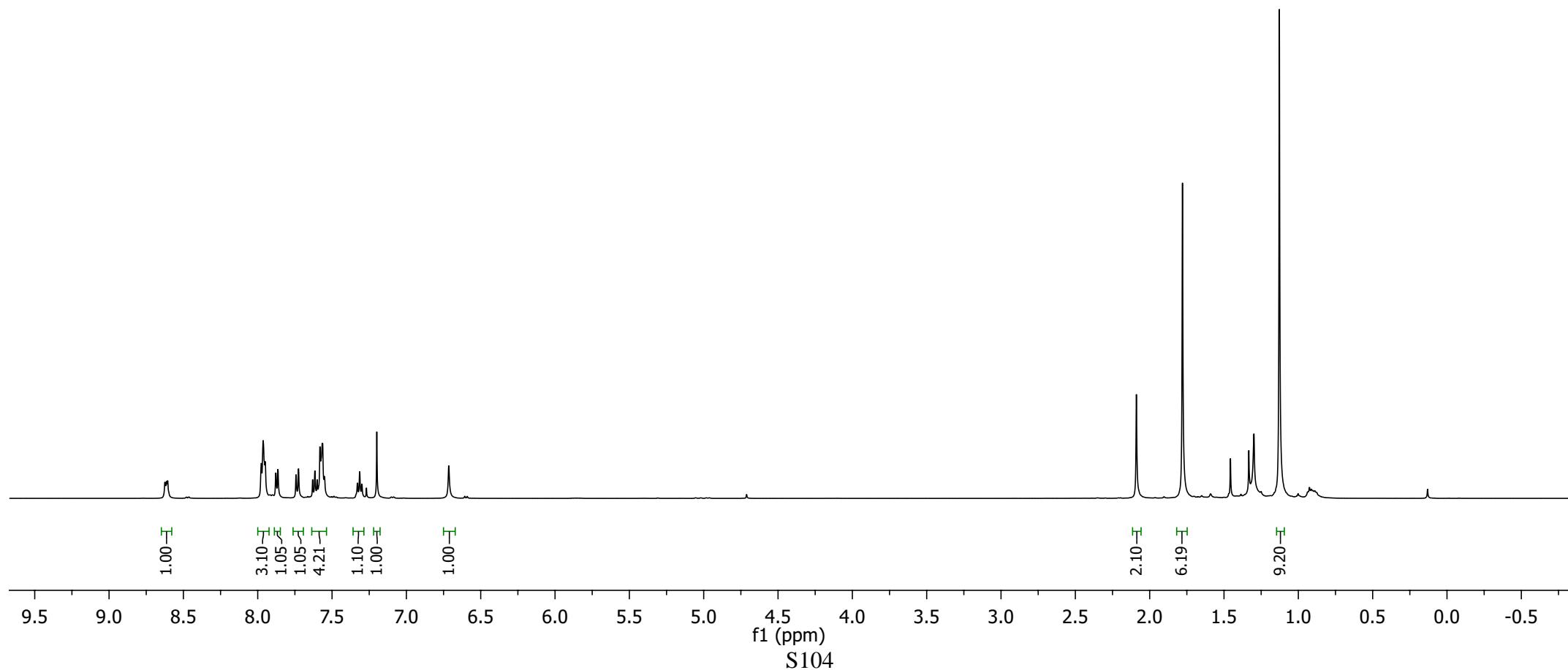
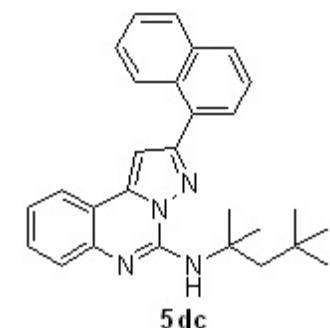
MCR-A118-07845

8.6245
8.6129
8.6056

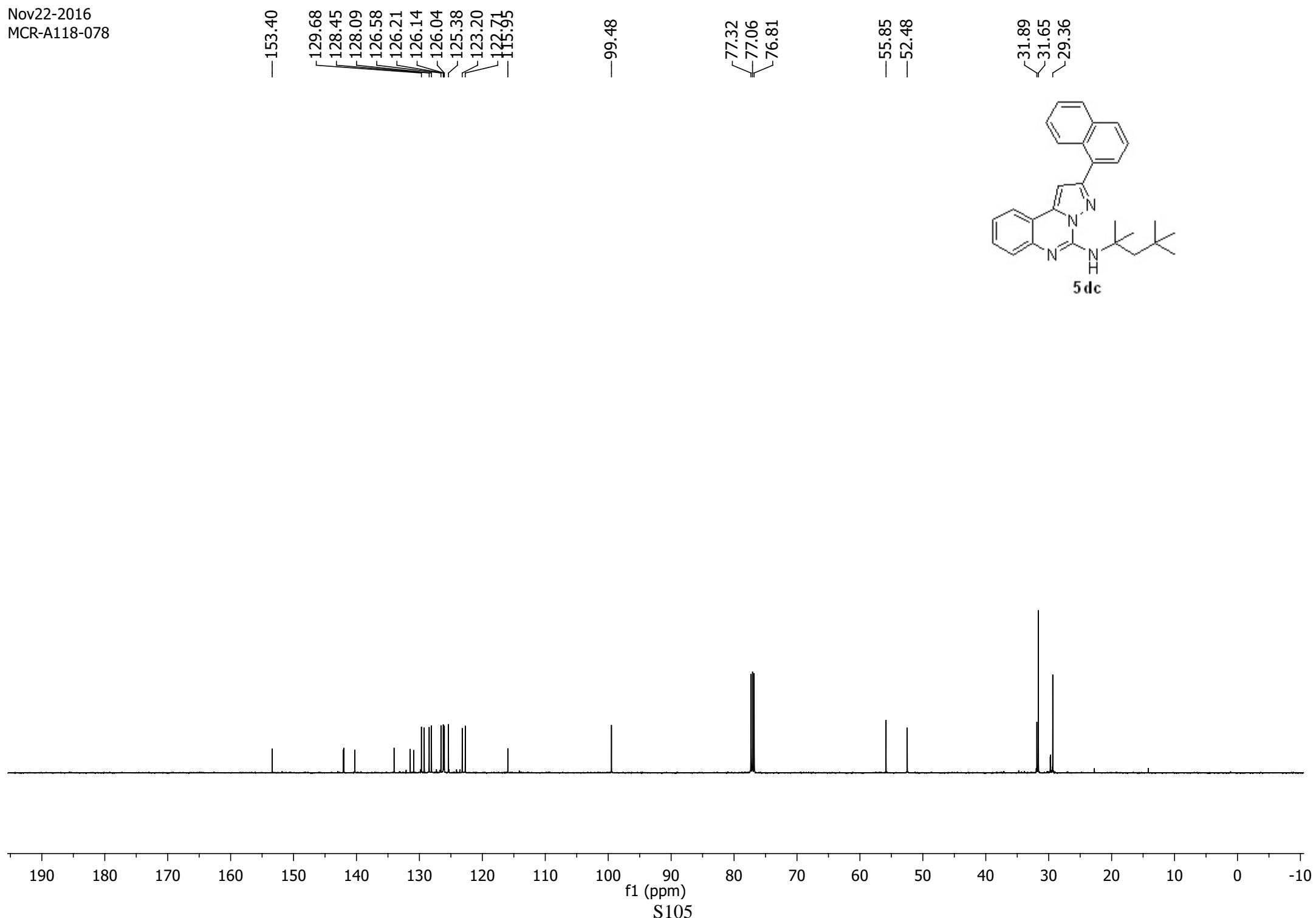
7.9757
7.9633
7.9504
7.8645
7.7257
7.5815
7.5660
7.5632
6.7198

-2.0894
-1.7794

-1.1278



Nov22-2016
MCR-A118-078



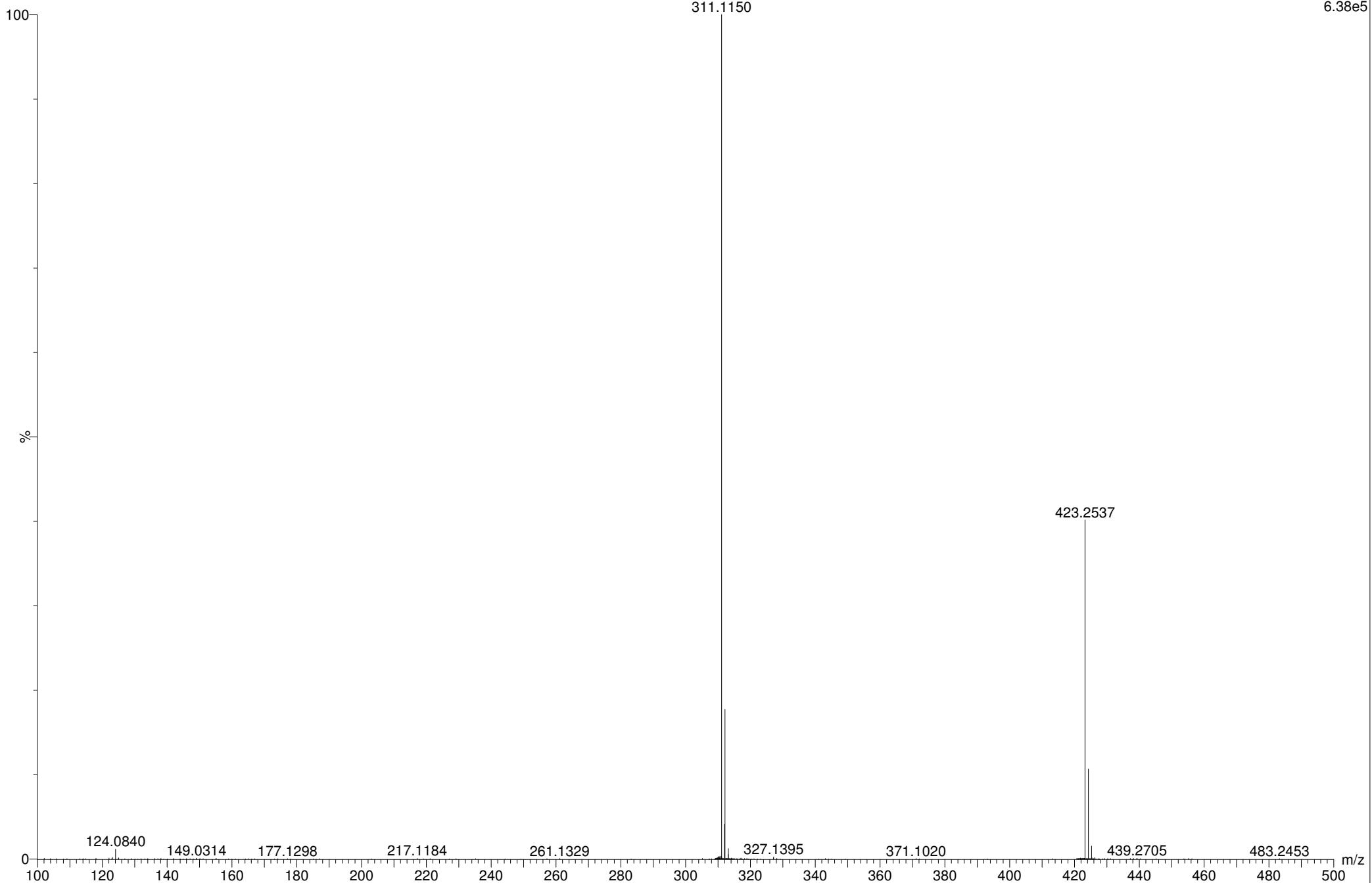
PROCESSED BY
PAWAN KUMAR

CSIR-IHBT
NPC&PD DIVISION

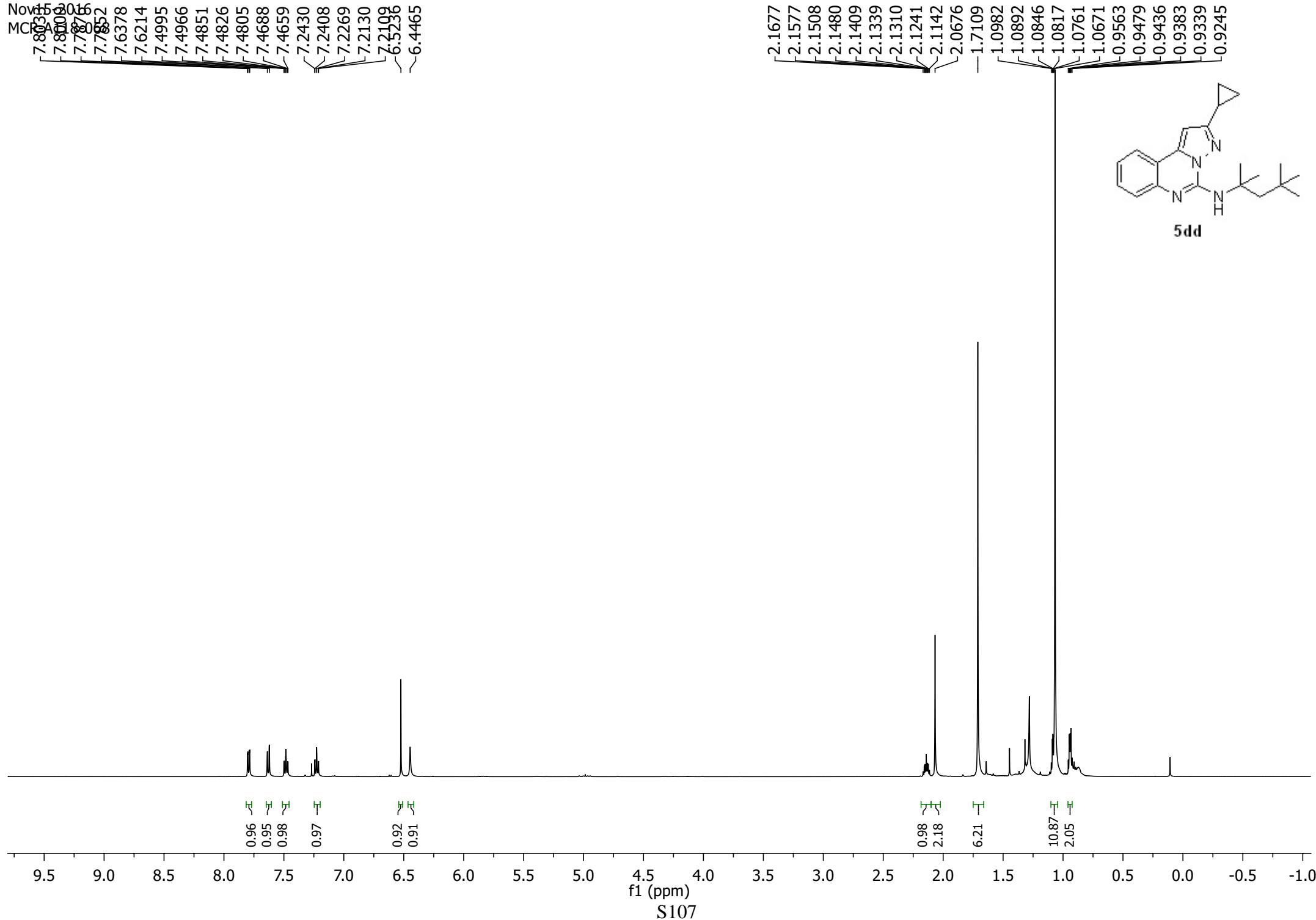
19-Jan-2017 13:18:04

MCR-A118-078_02 128 (2.373) AM (Cen,3, 80.00, Ar,5000.0,423.25,0.70); Sb (2,10.00); Cm (121:131)

1: TOF MS ES+
6.38e5



Nov 15 2016
MCR 303
87010
87052
6378
6214



-158.17

141.95
141.92
140.28

129.33
126.00
122.98
122.35

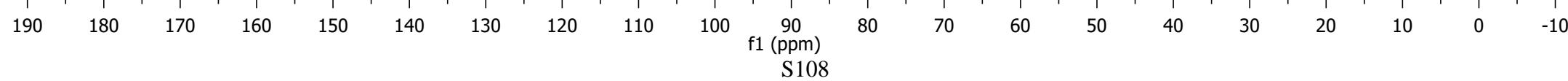
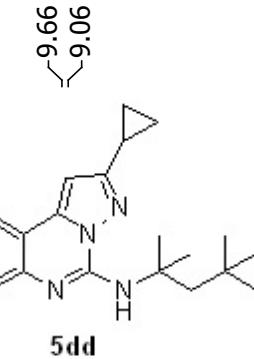
-115.80

-94.56

77.31
77.06
76.80

-55.60
-52.10

31.81
31.58
29.43



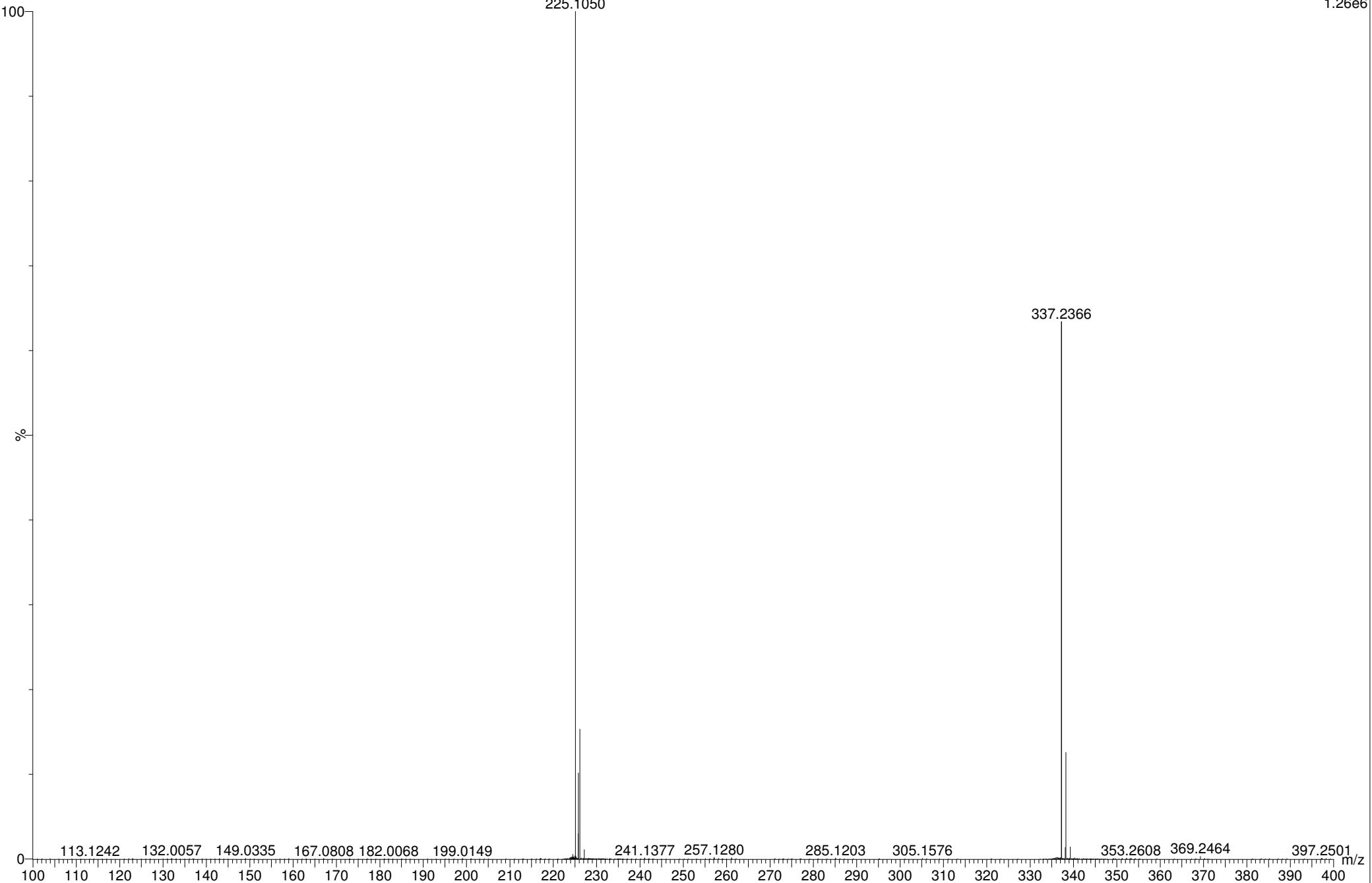
PROCESSED BY
PAWAN KUMAR

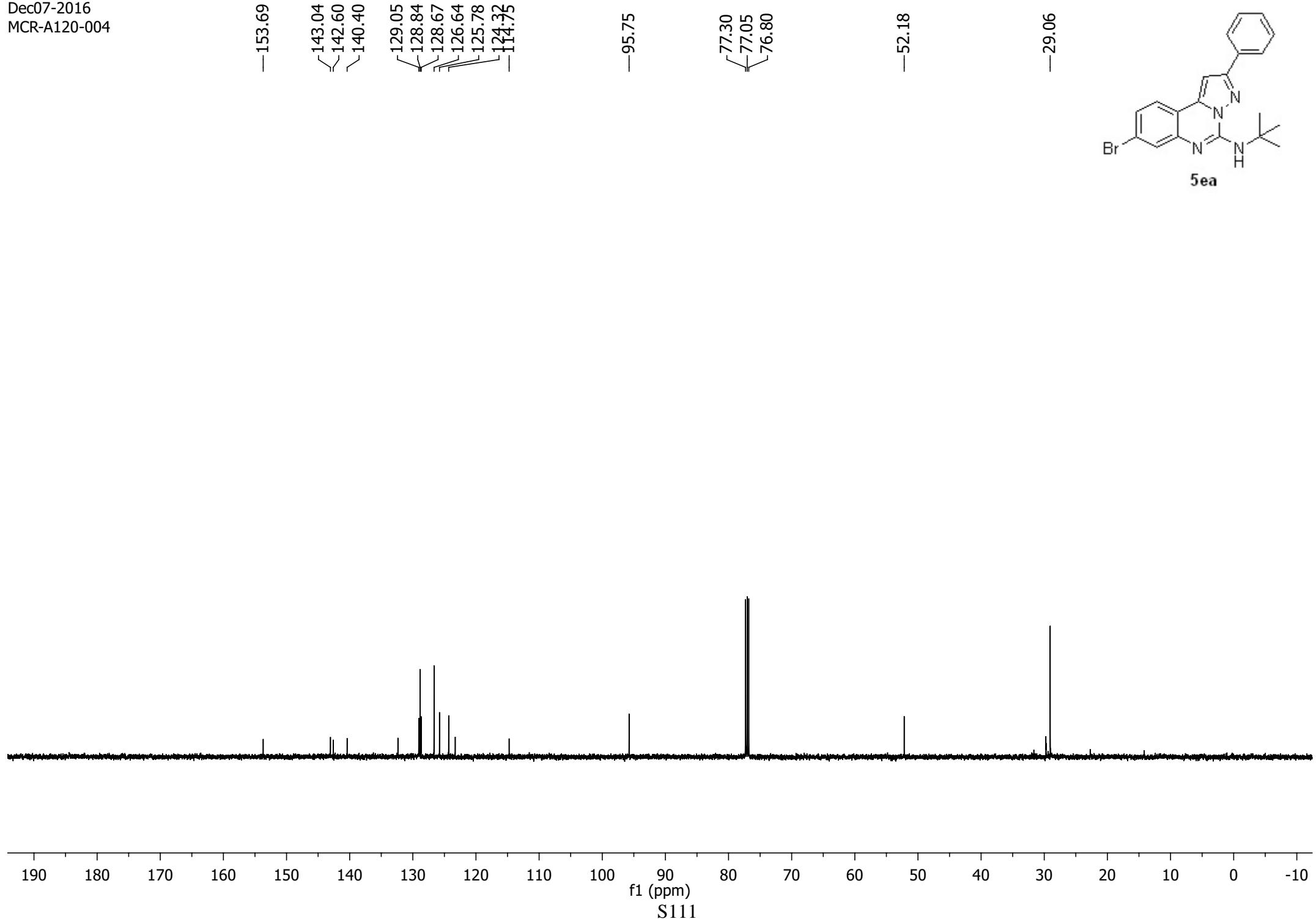
CSIR-IHBT
NPC&PD DIVISION

24-Nov-2016 16:22:44

MCR-A118-068 132 (2.449) AM (Cen,3, 80.00, Ar,5000.0,337.24,0.70); Sb (2,10.00); Cm (124:143)

1: TOF MS ES+
1.26e6





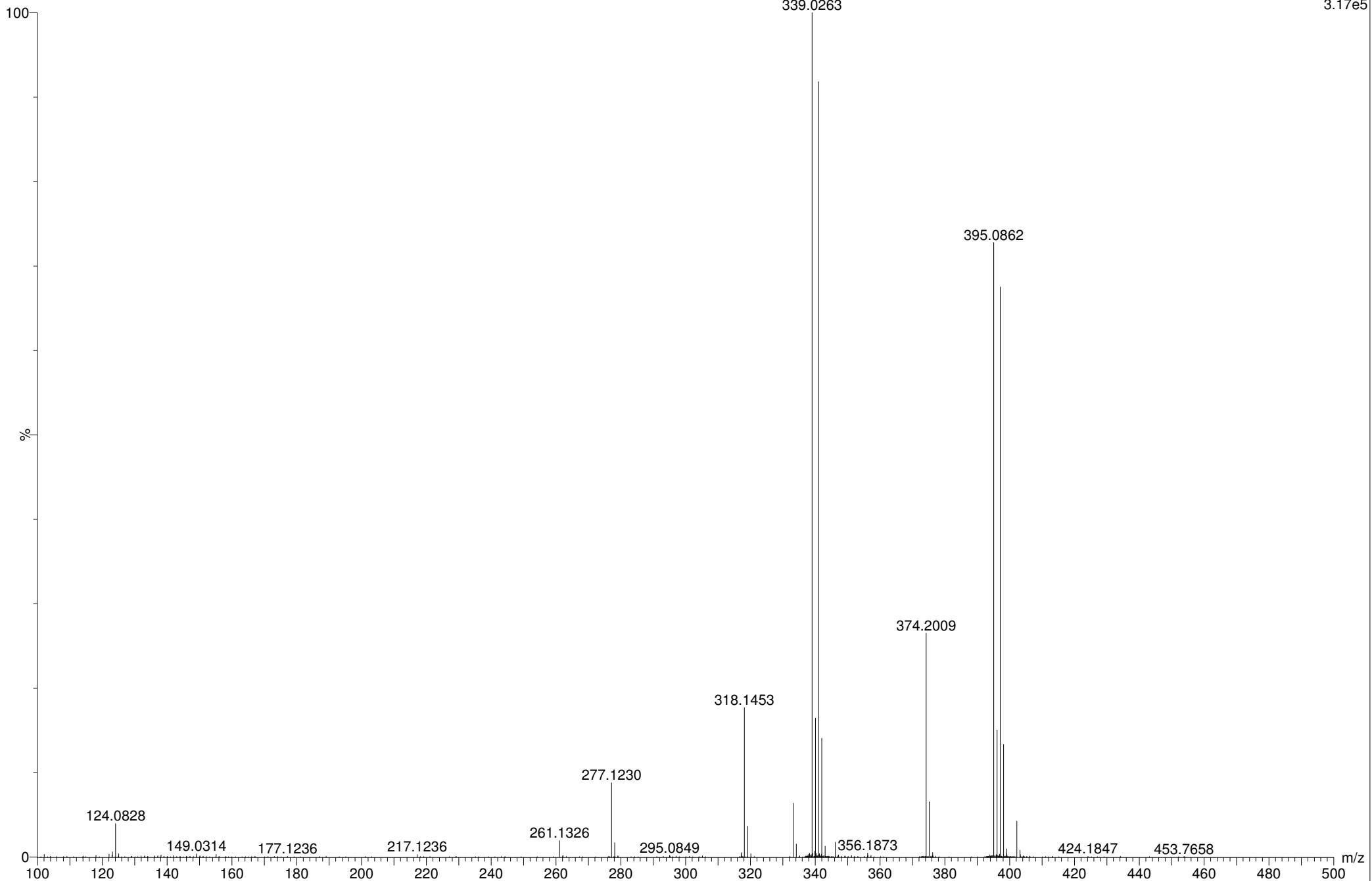
PROCESSED BY
PAWAN KUMAR

CSIR-IHBT
NPC&PD DIVISION

19-Jan-2017 13:10:14

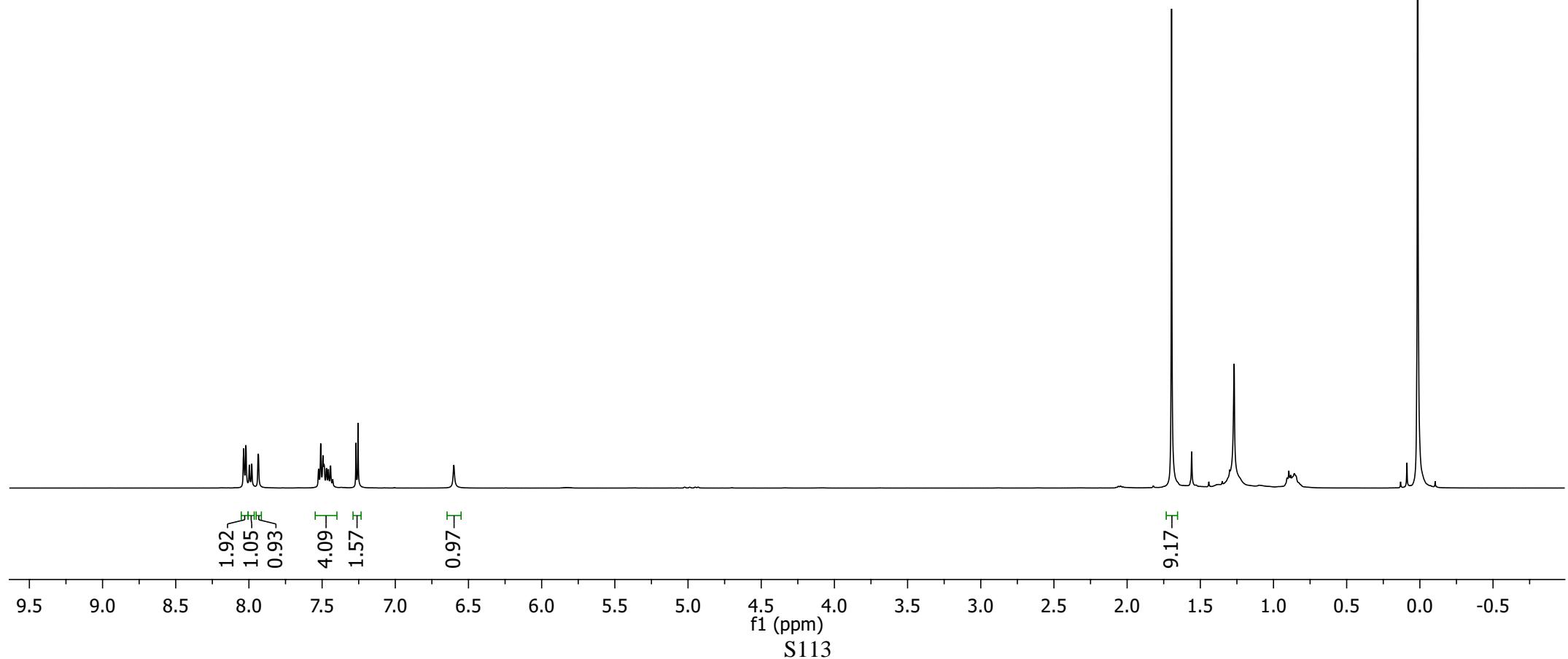
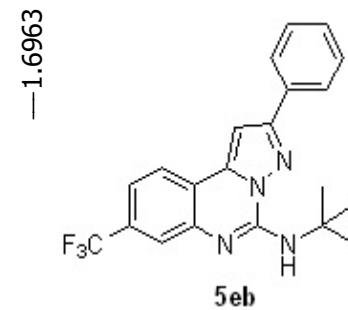
MCR-A120-004_02 73 (1.355) AM (Cen,3, 80.00, Ar,5000.0,395.09,0.70); Sb (2,10.00); Cm (67:83)

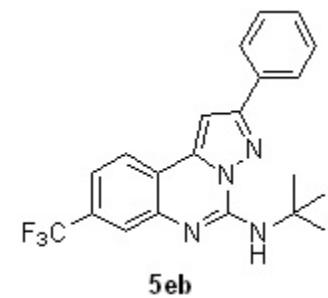
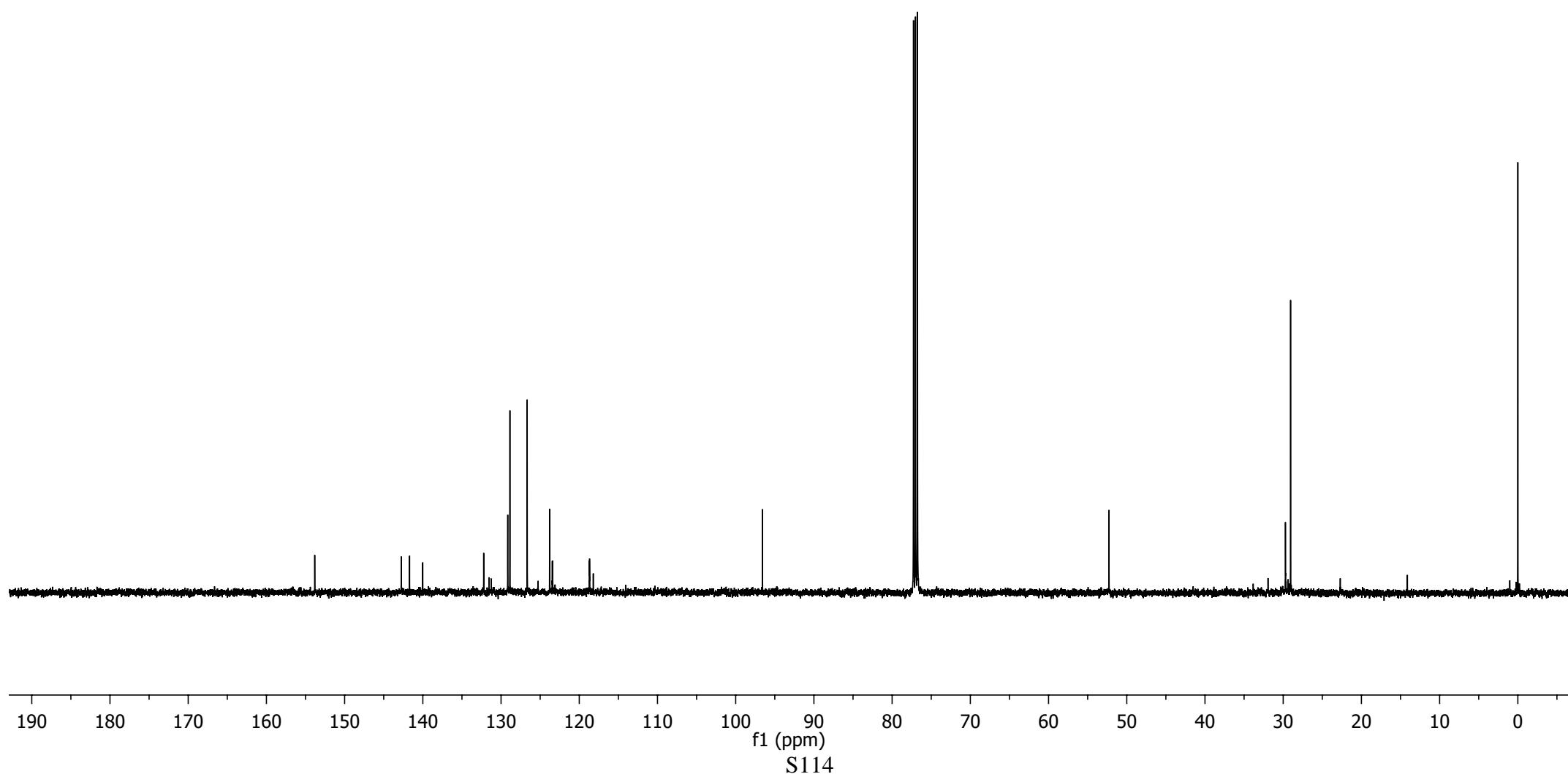
1: TOF MS ES+
3.17e5



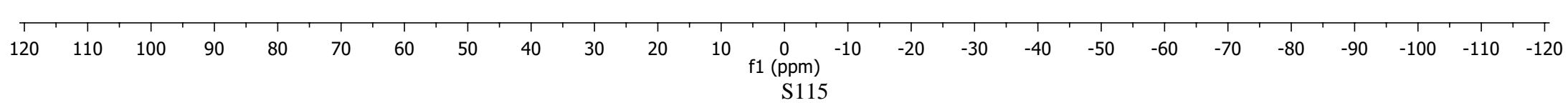
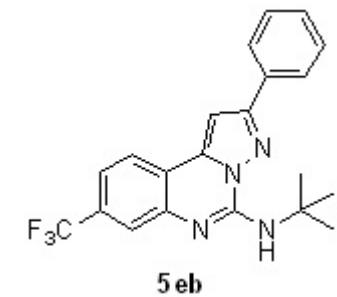
Jan03-2017
MCR-A120-026

8.0353
8.0209
7.9971
7.9808
7.9361
7.5097
7.4943
7.4880
7.4419
7.2688
6.6002





-62.44



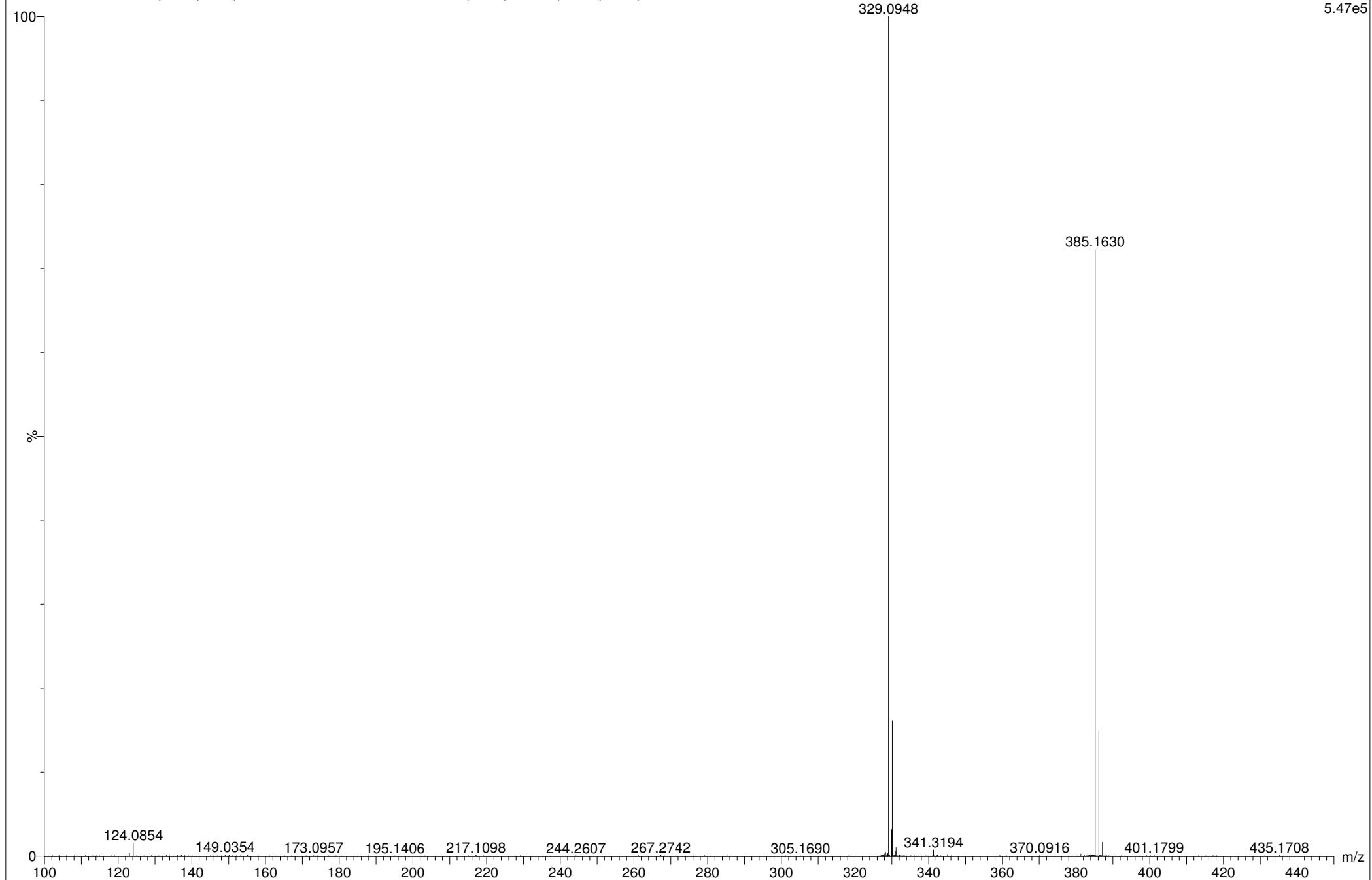
PROCESSED BY
PAWAN KUMAR

CSIR-IHBT
NPC&PD DIVISION

19-Jan-2017 15:01:03

MCR-A120-026_02 60 (1.112) AM (Cen,3, 80.00, Ar,5000.0,385.16,0.70); Sb (2,10.00); Cm (56:68)

1: TOF MS ES+
5.47e5



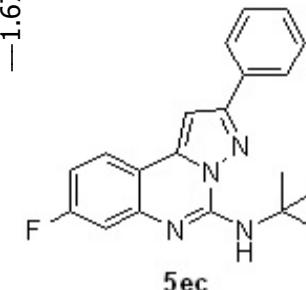
Dec30-2016

MCR-A120-016

8.0265
8.0238
8.0096
7.8831
7.8709
7.8657
7.8536

7.5116
7.4971
7.4818
7.4262
7.2690
6.1254

-1.6773

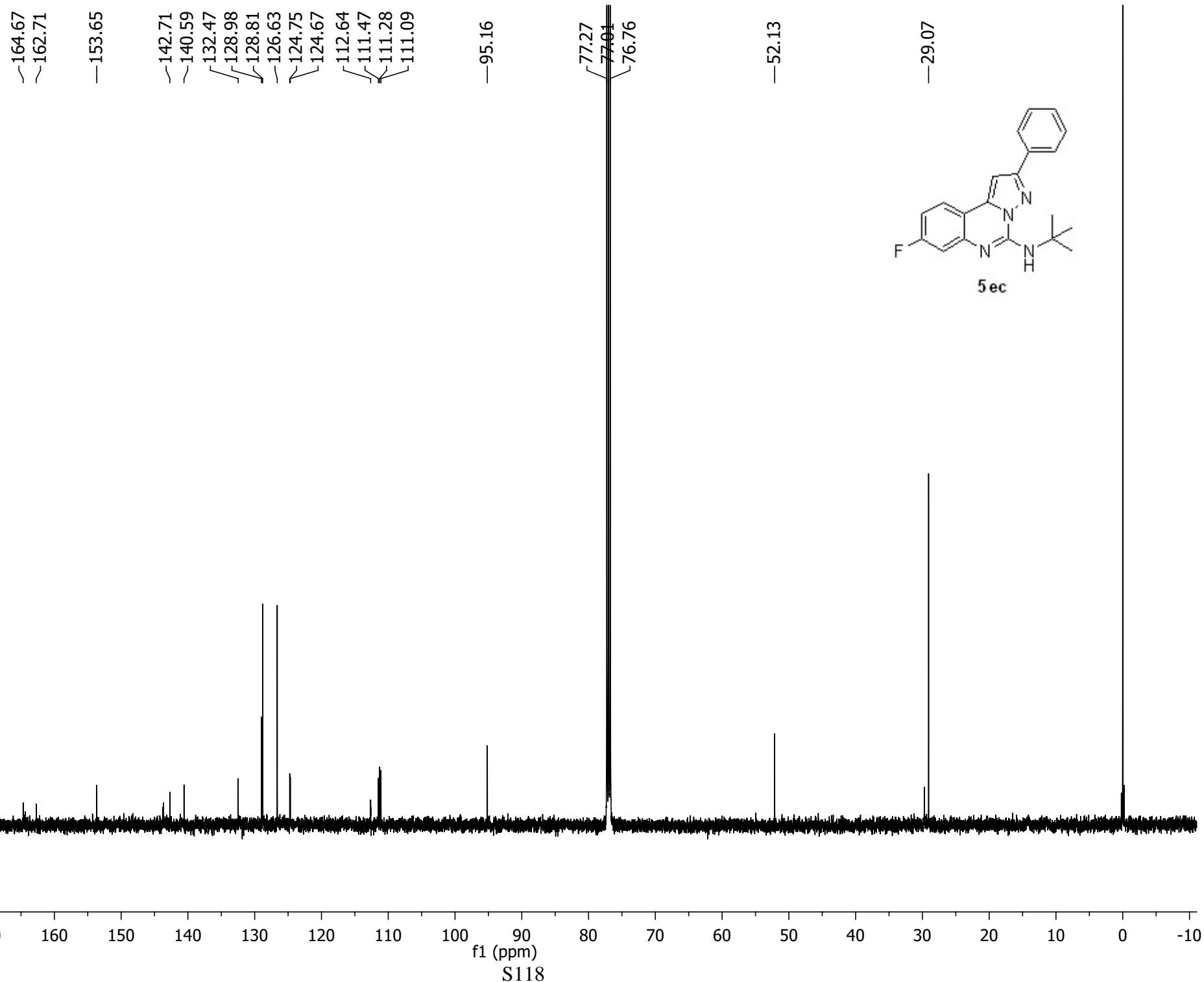


2.06
1.04
2.07
1.08
1.00
1.01
1.05
0.99

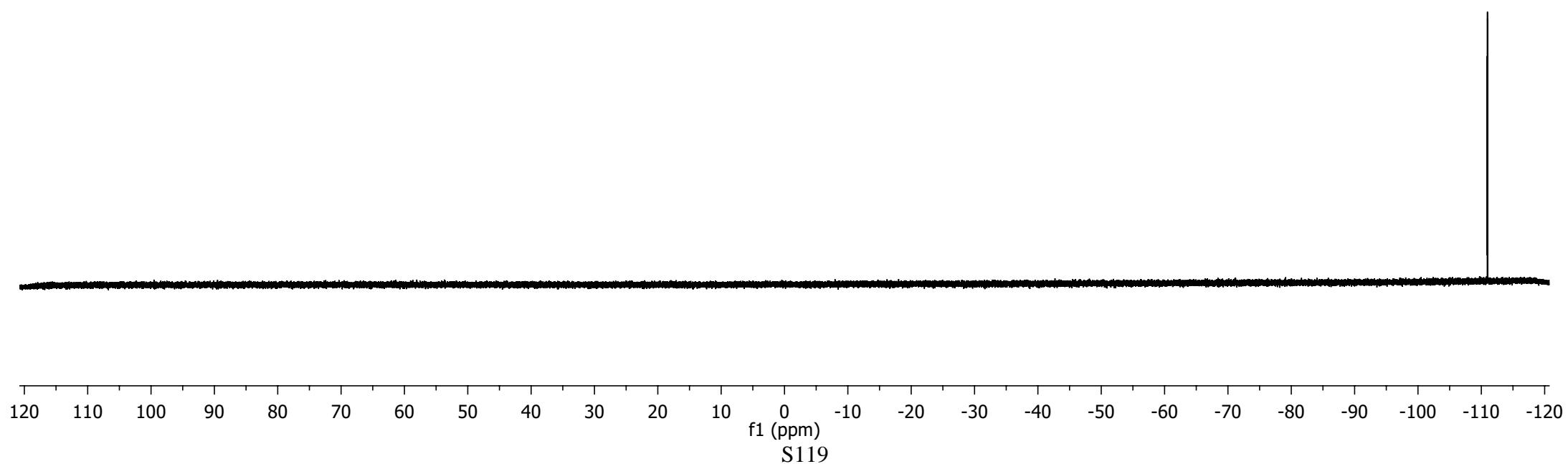
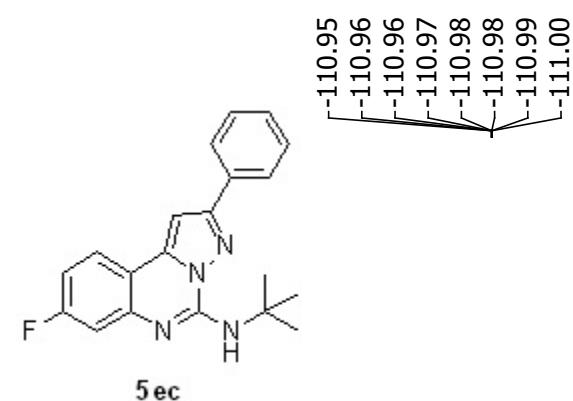
8.96

f1 (ppm)
S117

9.5 9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0 -0.5 -1.0



Dec31-2016
MCR-A120-016



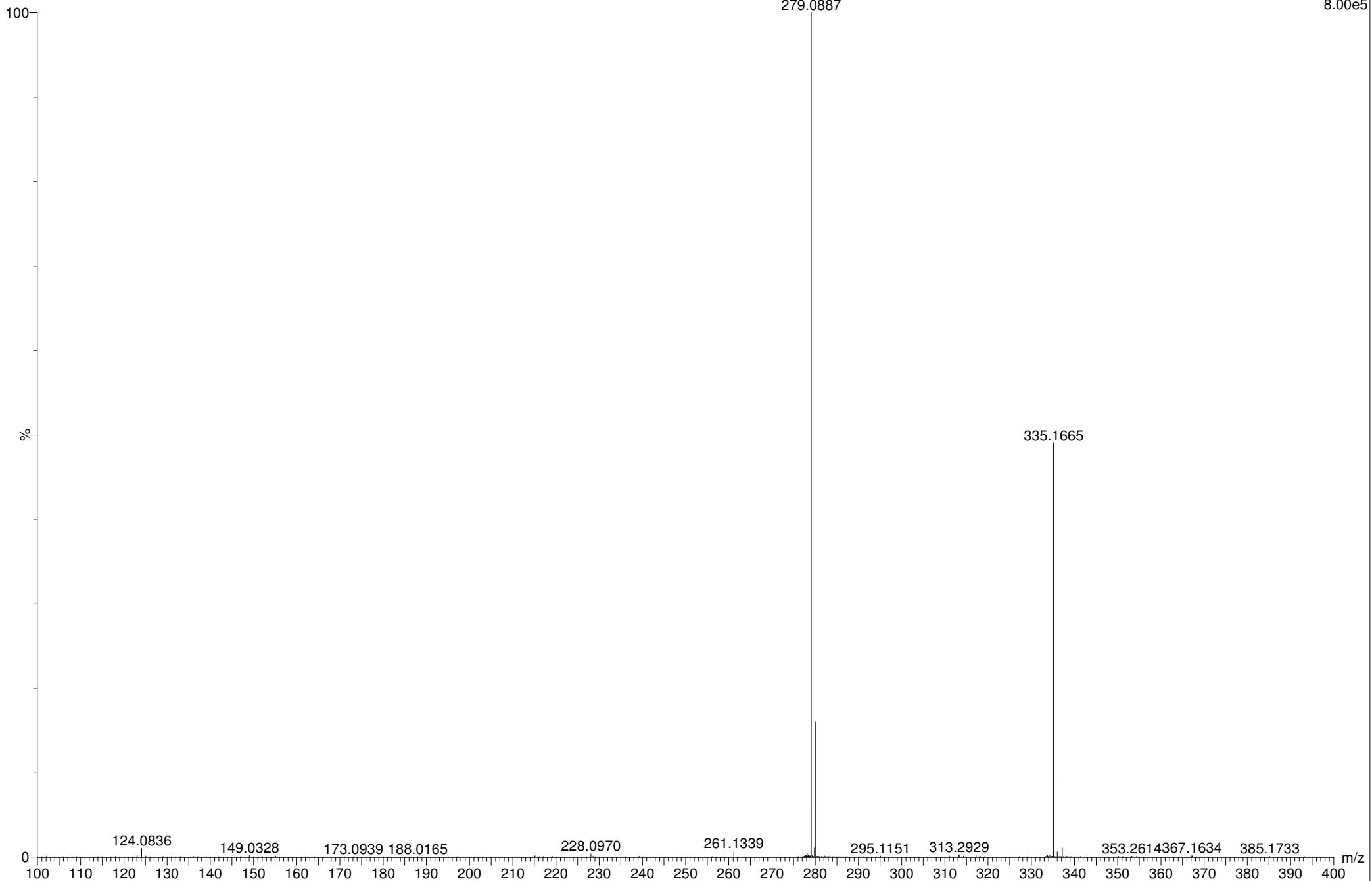
PROCESSED BY
PAWAN KUMAR

CSIR-IHBT
NPC&PD DIVISION

19-Jan-2017 14:57:04

MCR-A120-016_02 46 (0.852) AM (Cen,3, 80.00, Ar,5000.0,335.17,0.70); Sb (2,10.00); Cm (42:55)

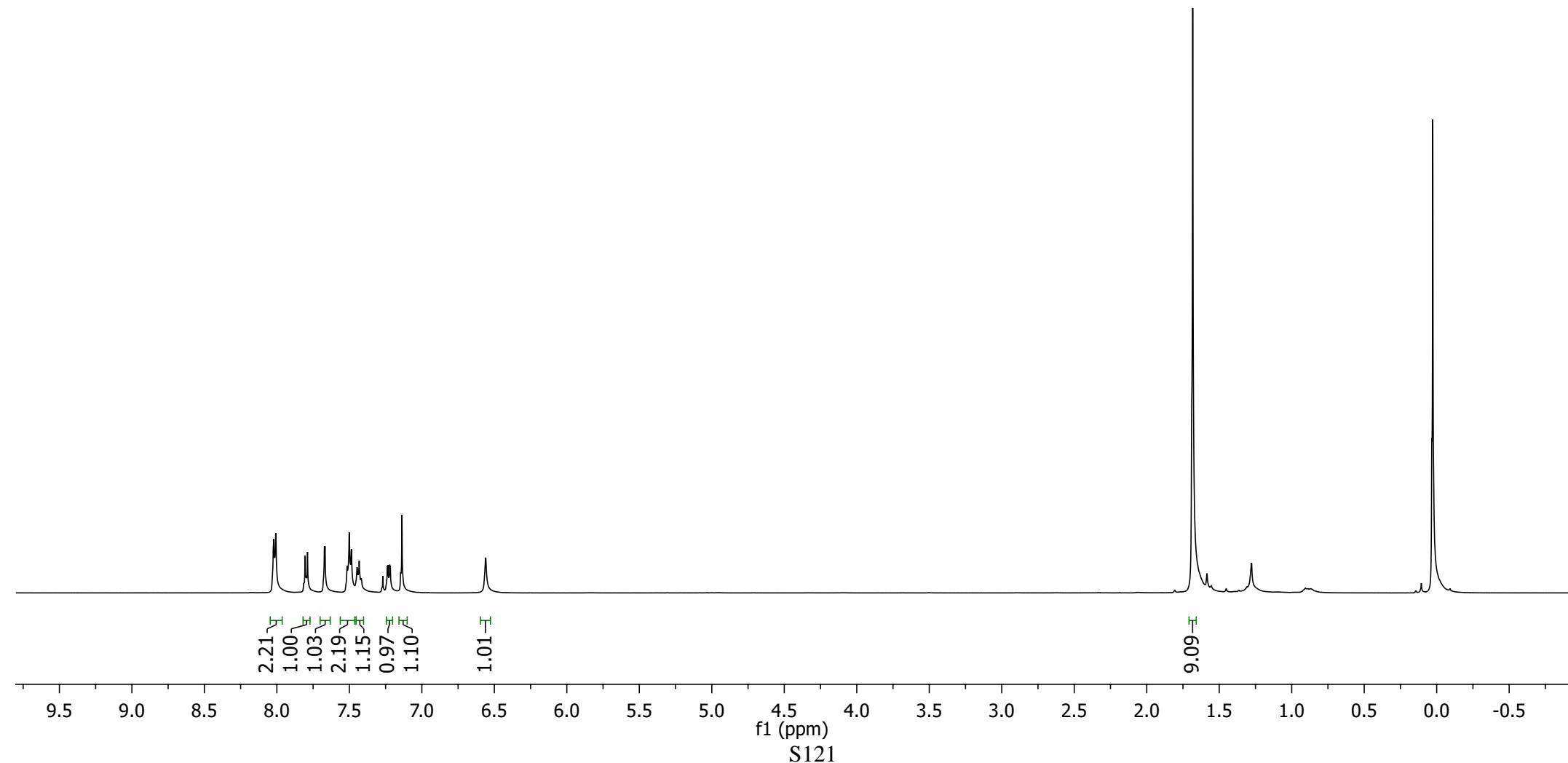
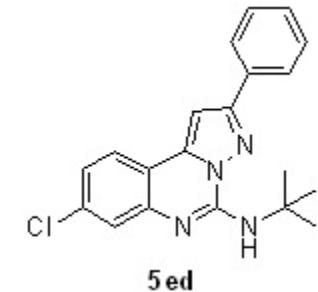
1: TOF MS ES+
8.00e5

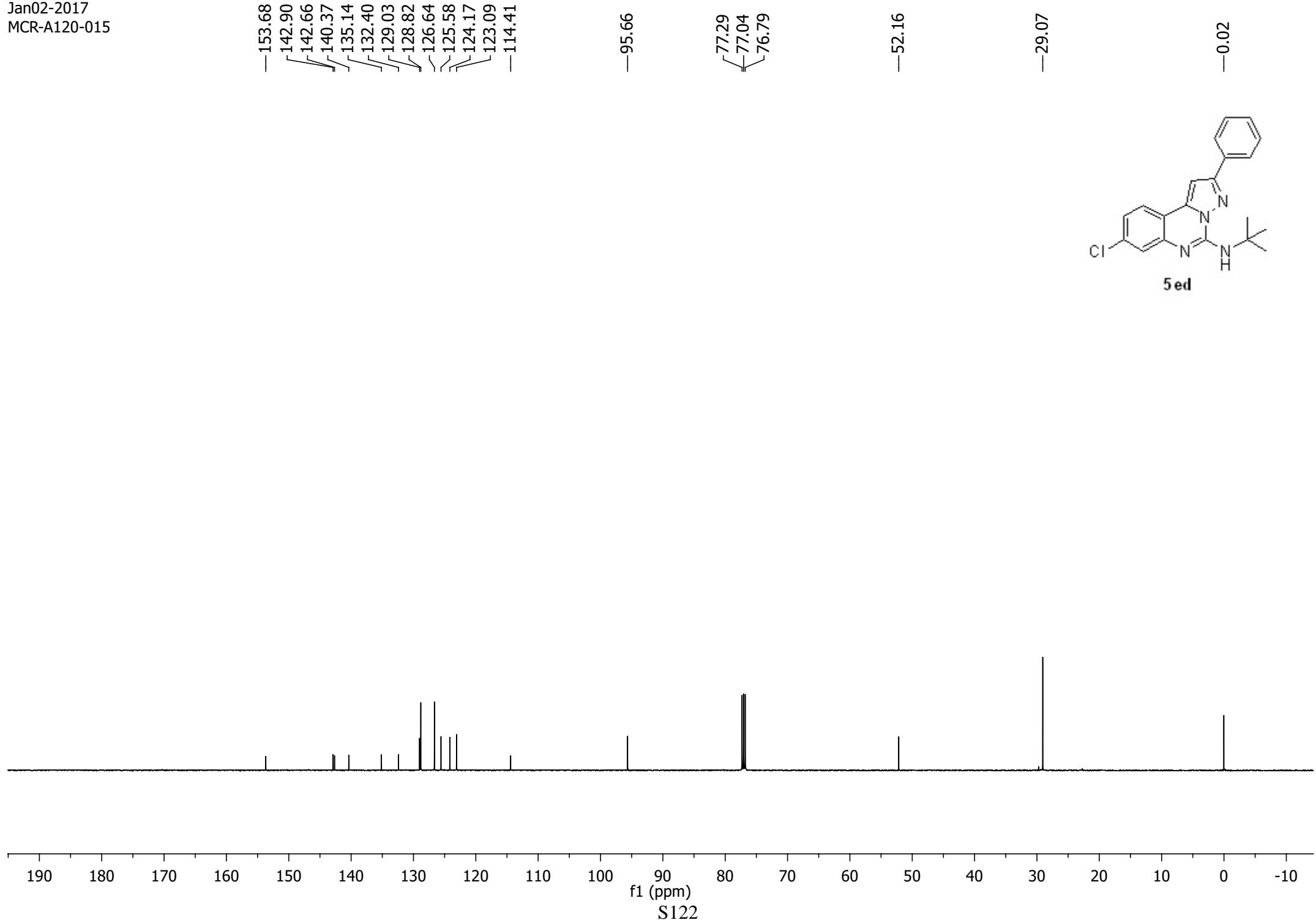


Jan02 2017
MCR-A170714

8.0120
8.0074
7.8054
7.7982
7.7887
7.6718
7.6679
7.5154
7.5009
7.4855
7.4466
7.4321
7.2382
7.2342
7.2215
7.2175
7.1460
6.5598
7.1374

-1.6827





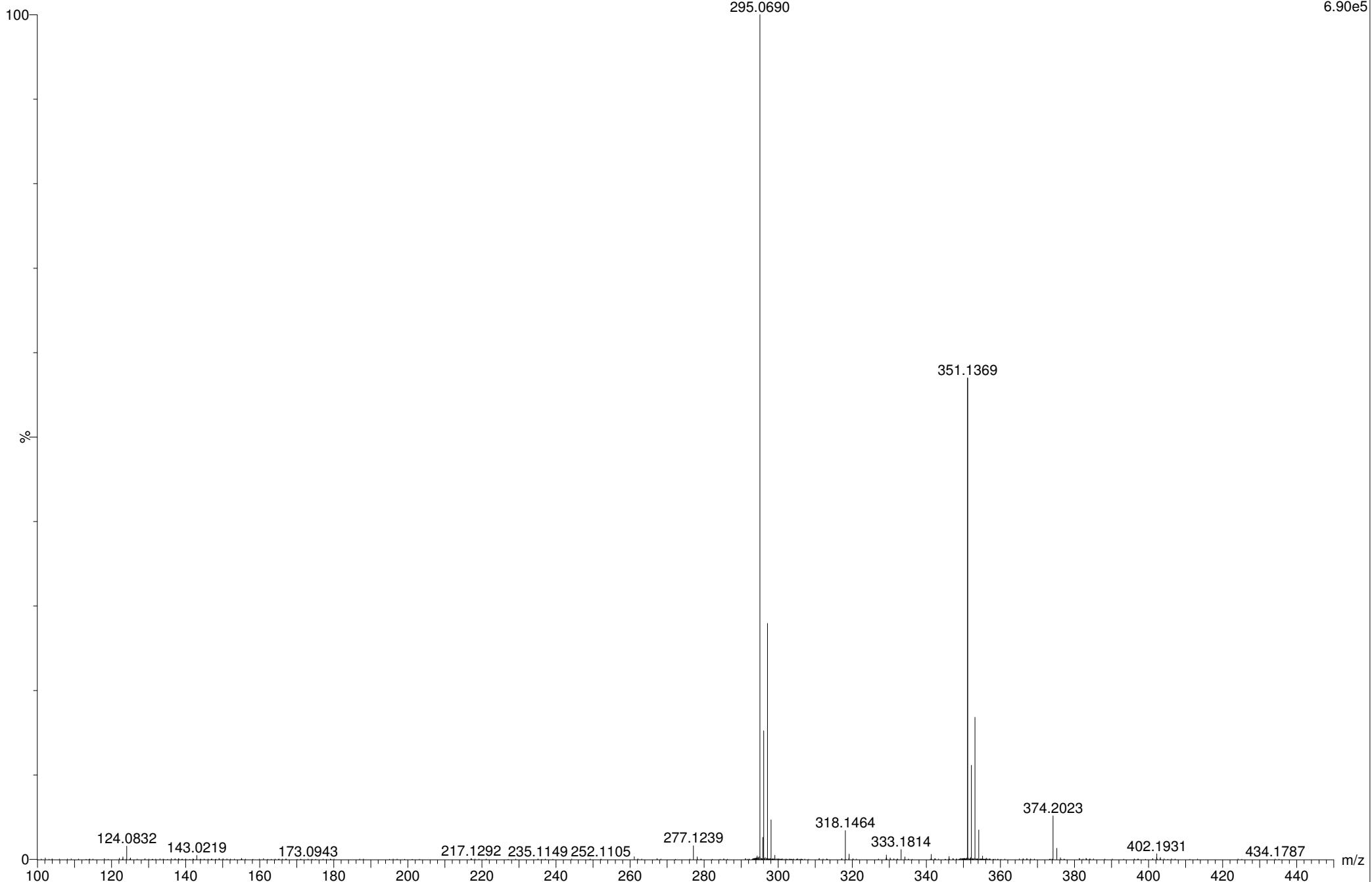
PROCESSED BY
PAWAN KUMAR

CSIR-IHBT
NPC&PD DIVISION

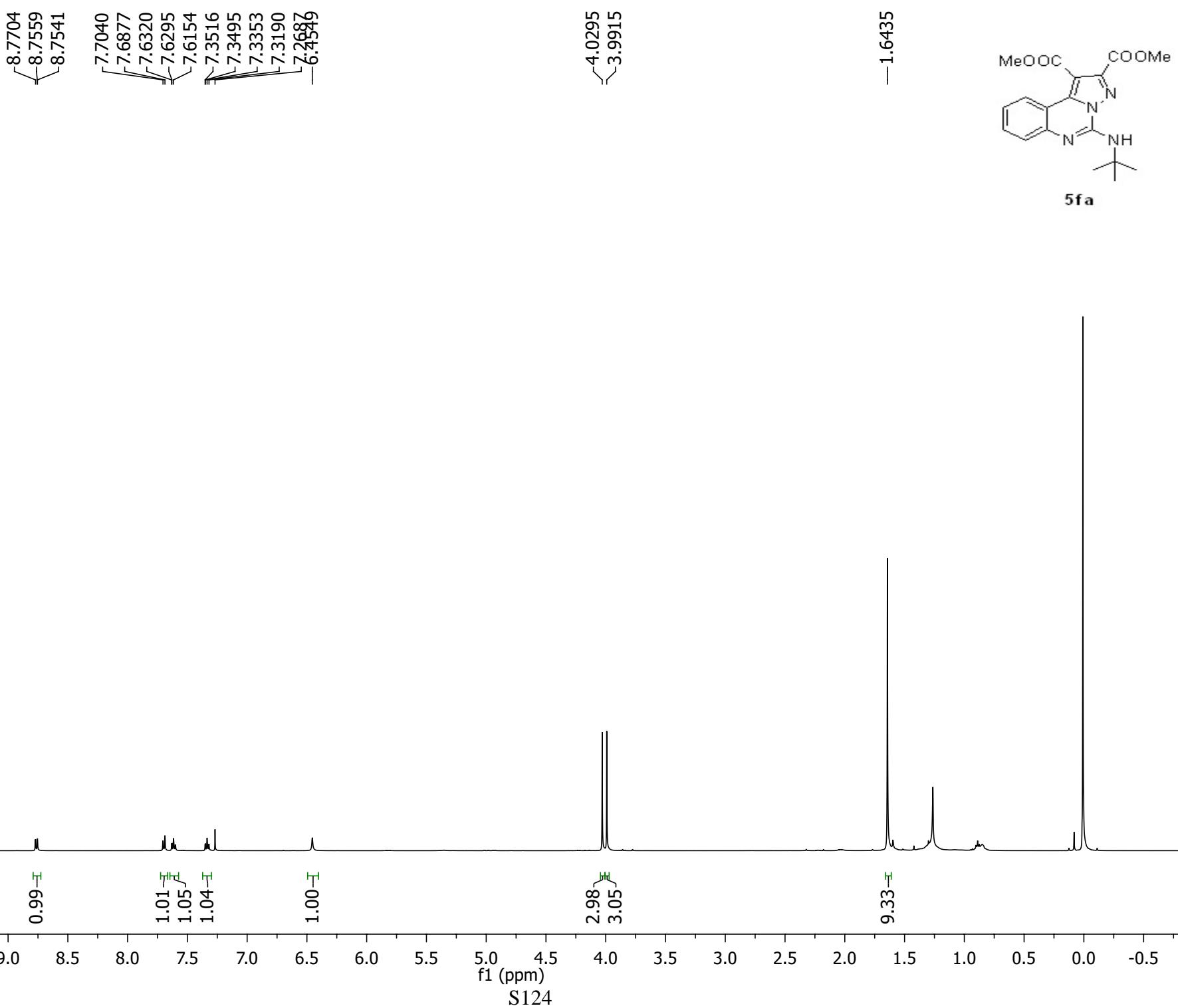
19-Jan-2017 13:33:48

MCR-A120-015_02 67 (1.242) AM (Cen,3, 80.00, Ar,5000.0,351.14,0.70); Sb (2,10.00); Cm (61:76)

1: TOF MS ES+
6.90e5



S123



Jan16-2017
SCA-PH005-224

✓163.73
✓163.18

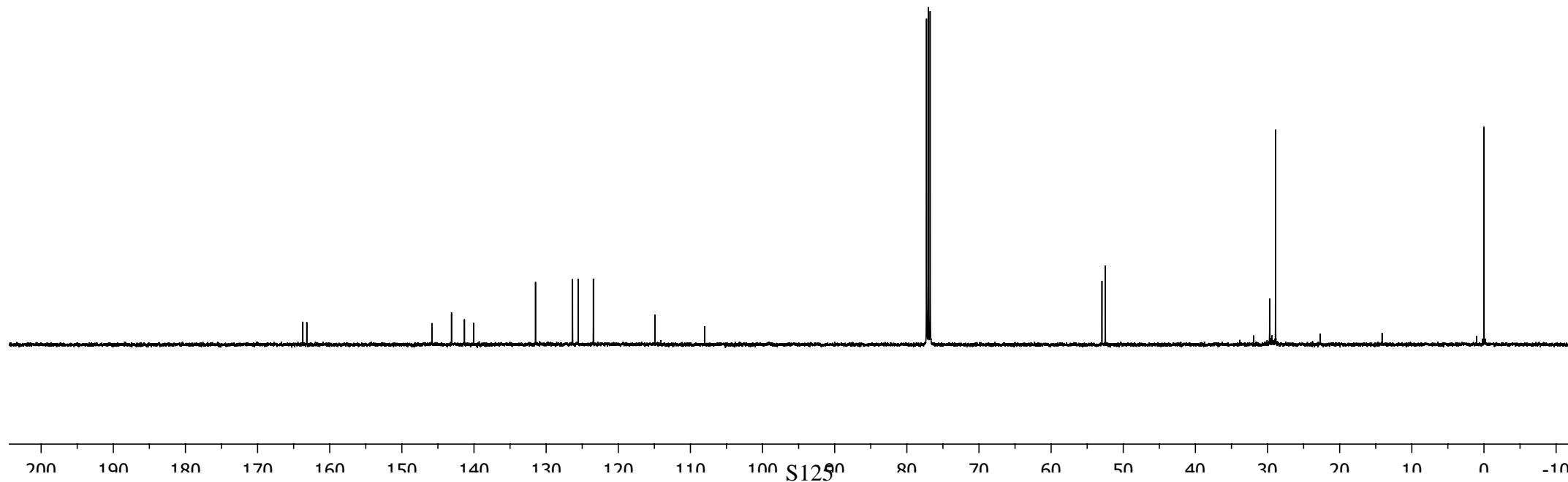
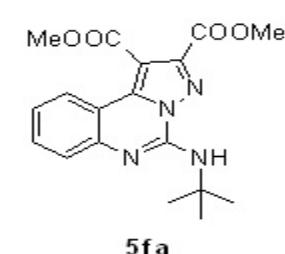
✓145.82
✓143.12
✓141.32
✓140.06
✓131.47
✓126.34
✓125.55
✓123.45

—114.92
—108.03

✓77.28
✓77.02
✓76.77

✓52.95
✓52.49
✓52.48

—28.89



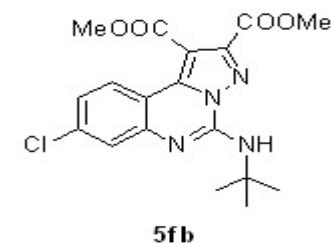
8.8290
8.8115

7.6952
7.6910
7.2904
7.2861
7.2728
7.2688

-6.5175

4.0277
3.9734

-1.6274



1.03

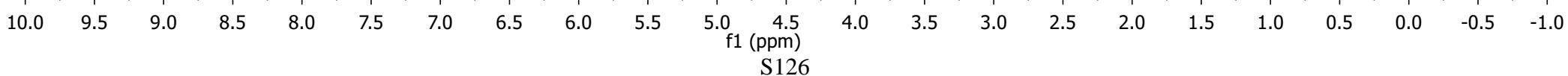
0.96

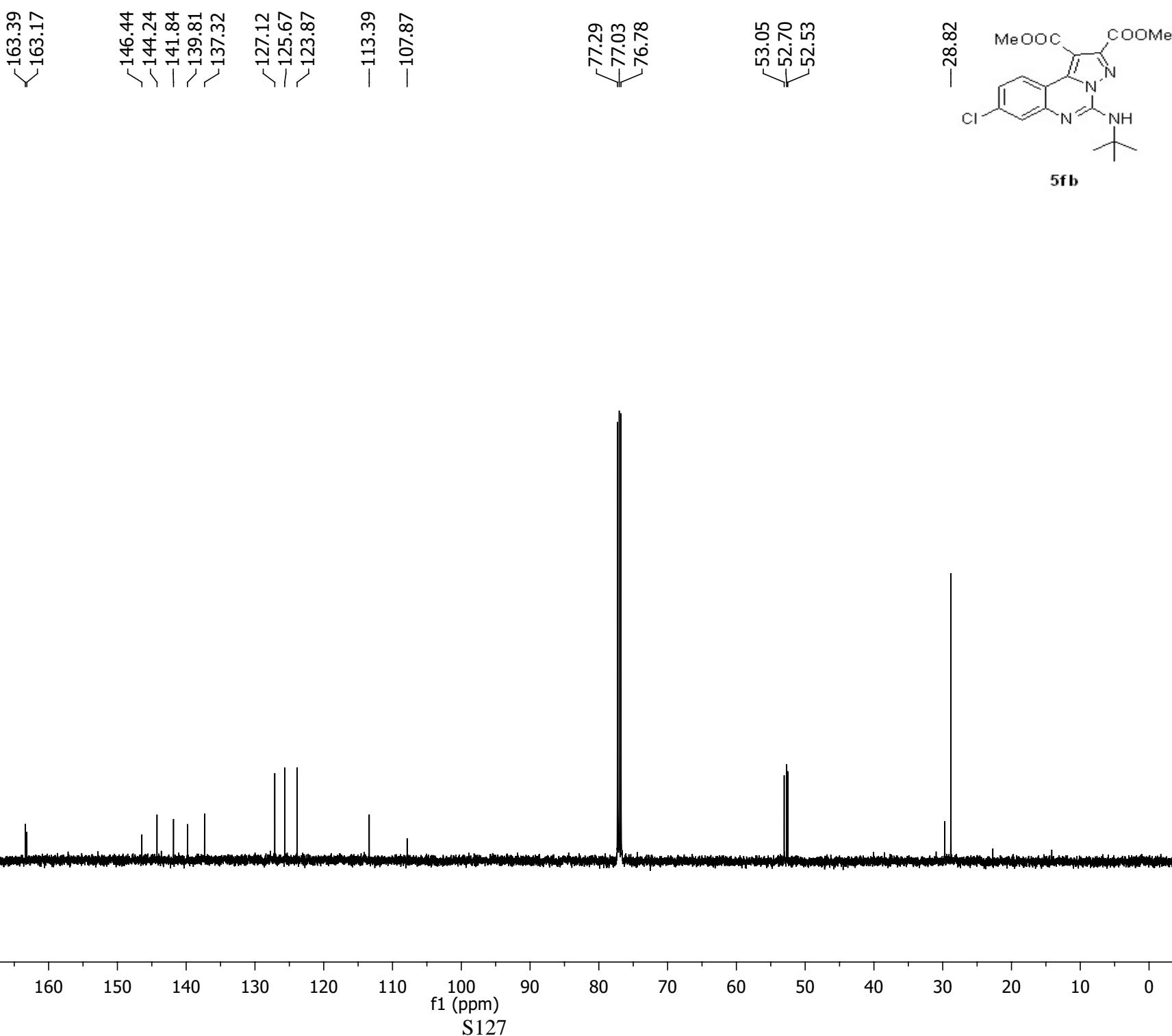
1.25

0.97

3.03
3.09

9.34





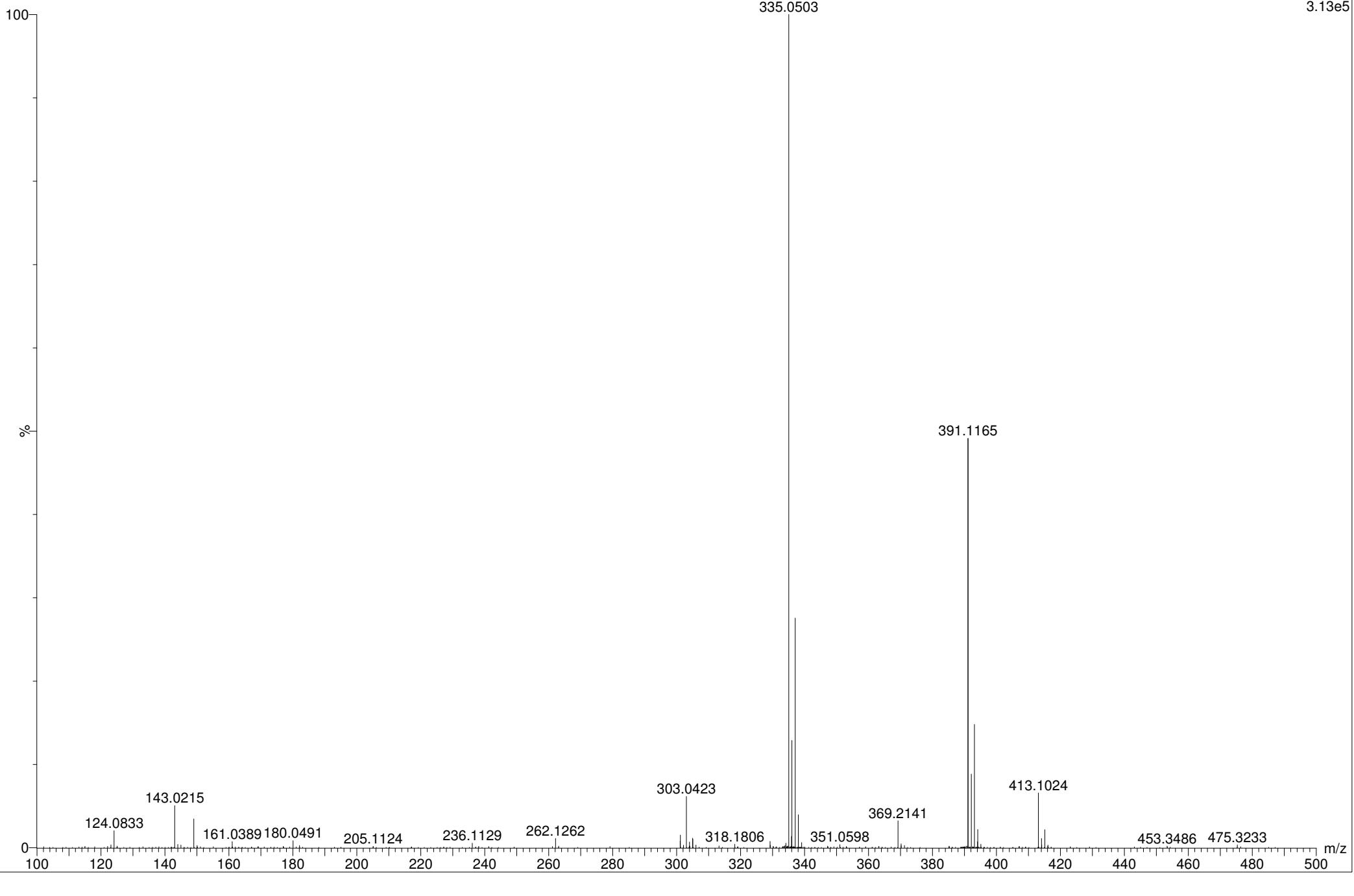
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19-Jan-2017 15:41:20

SCA-PH005-206_01 34 (0.631) AM (Cen,3, 80.00, Ar,5000.0,391.12,0.70); Sb (2,10.00); Cm (31:38)

1: TOF MS ES+
3.13e5



Feb03-2017
SCA-PH005-233

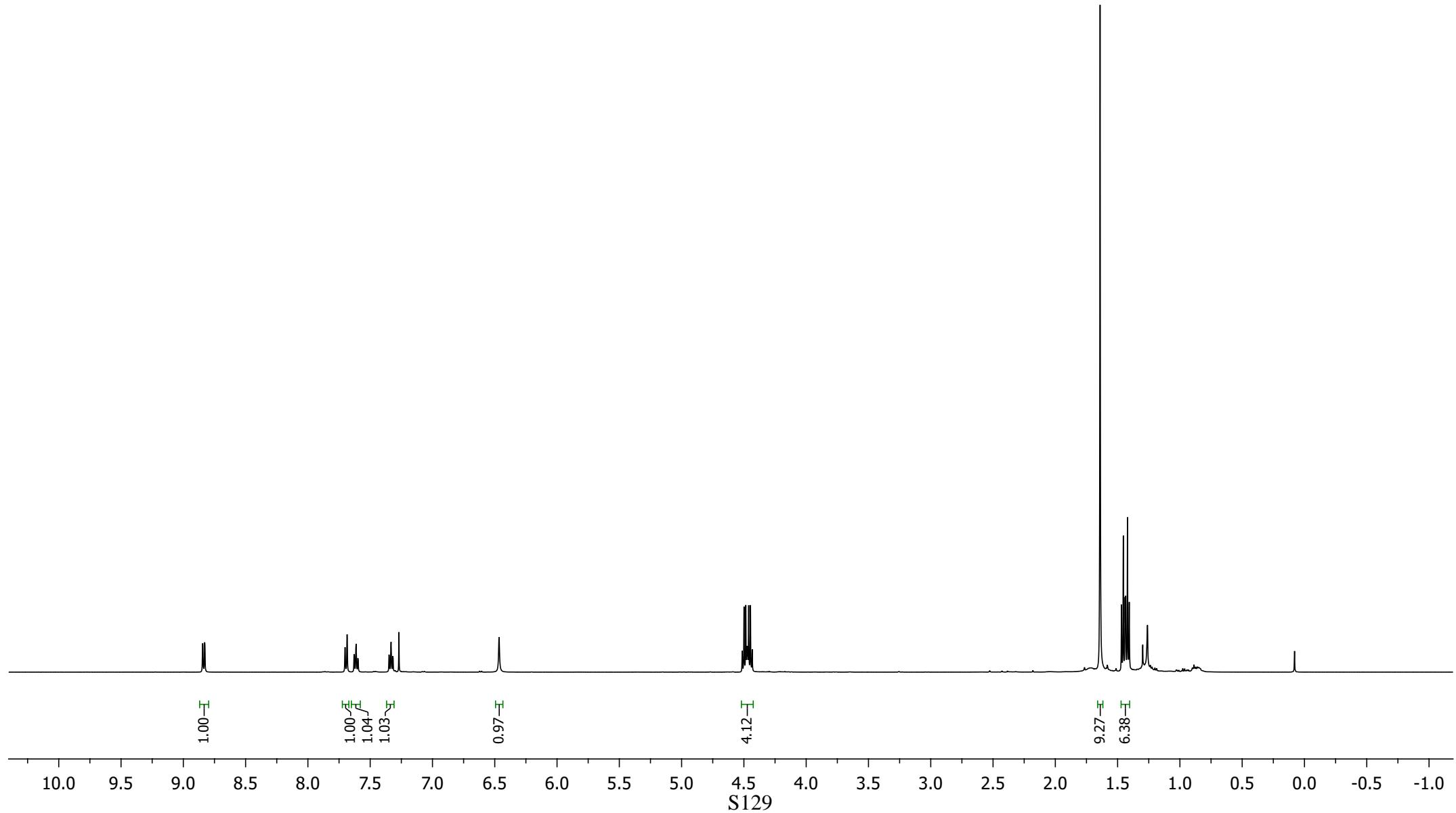
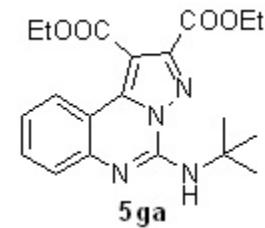
⁸8446
⁸8283

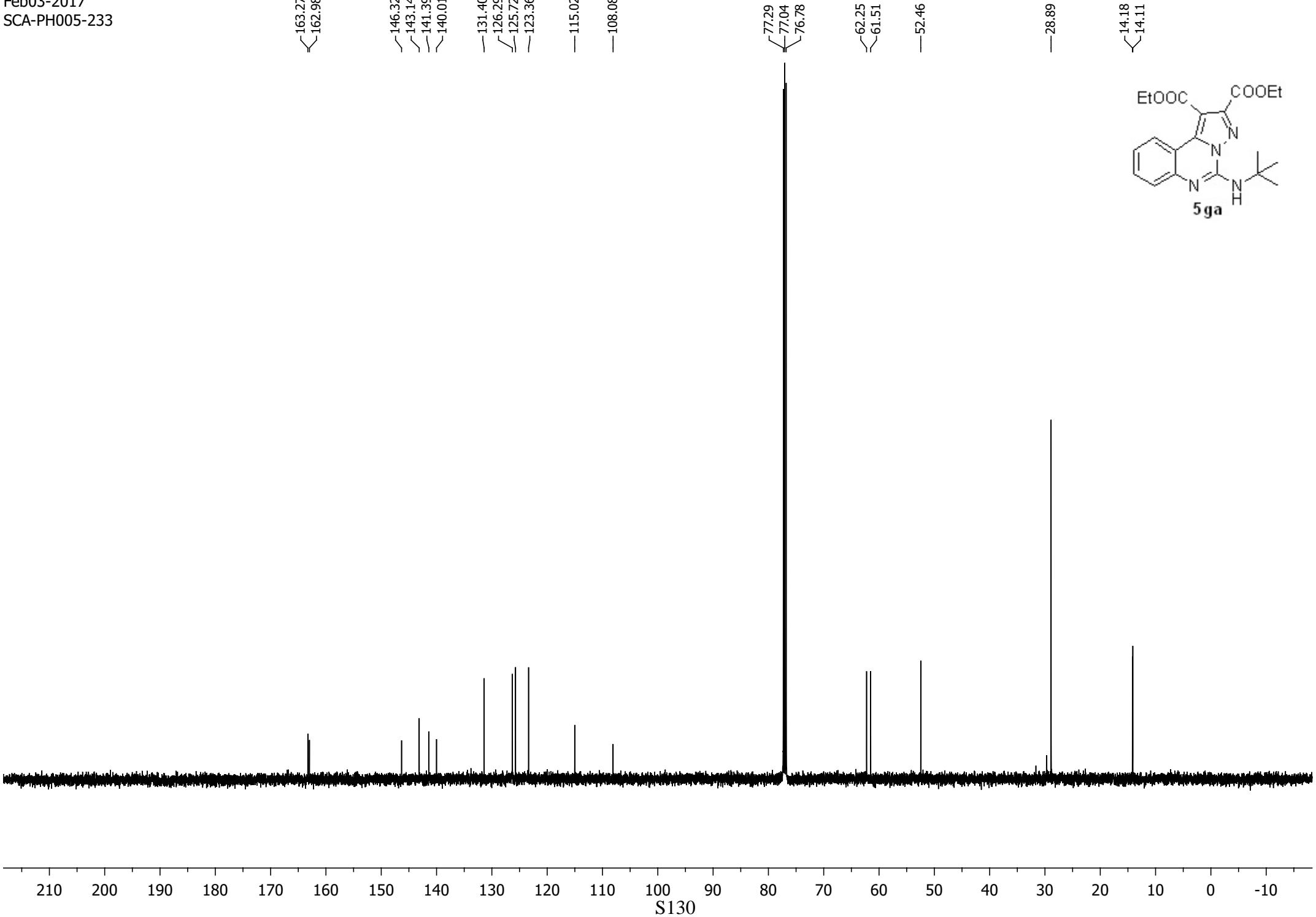
7.7016
7.6853
7.6290
7.6128
7.5985
7.3484
7.3324
7.3180
7.2689

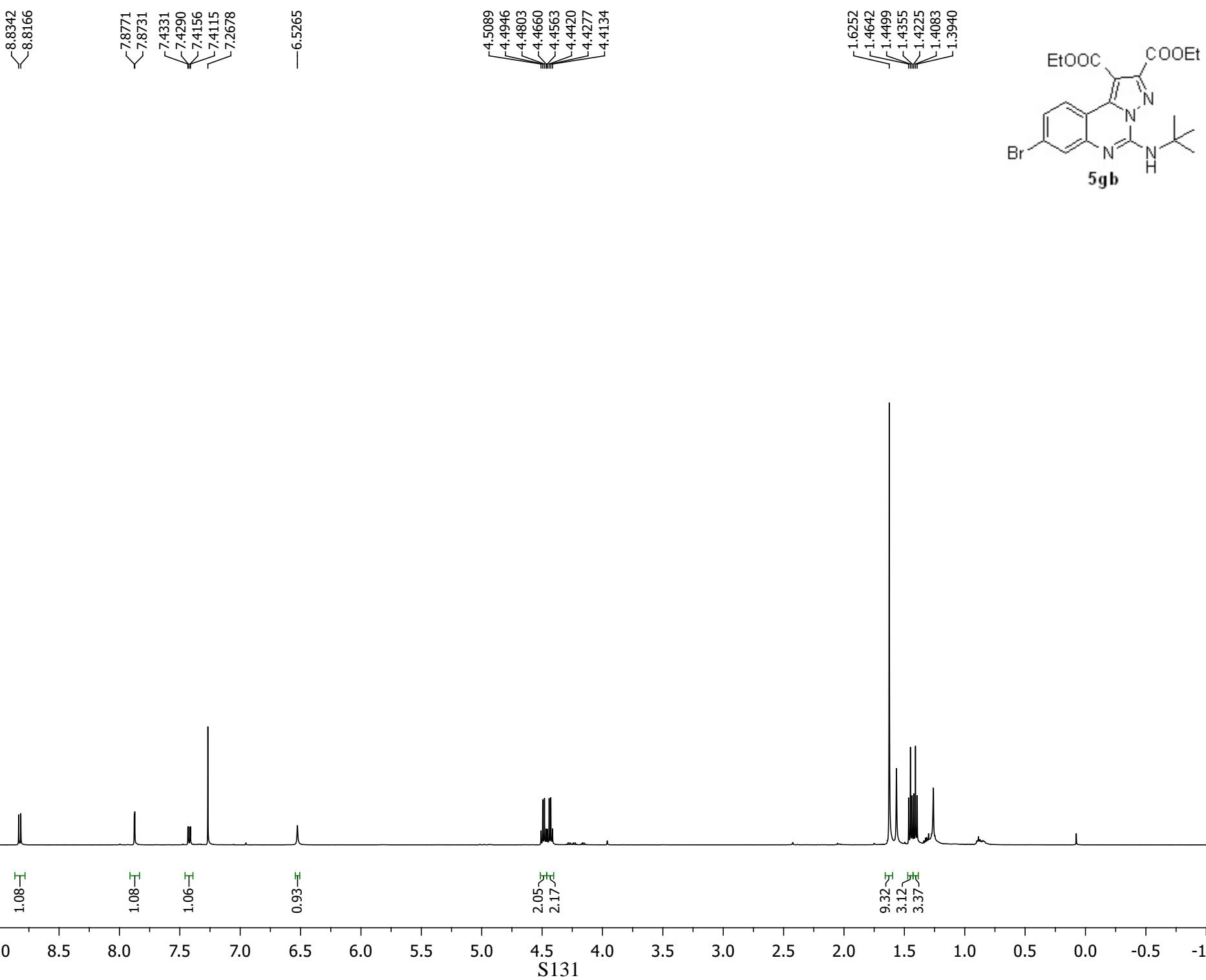
—6.4654

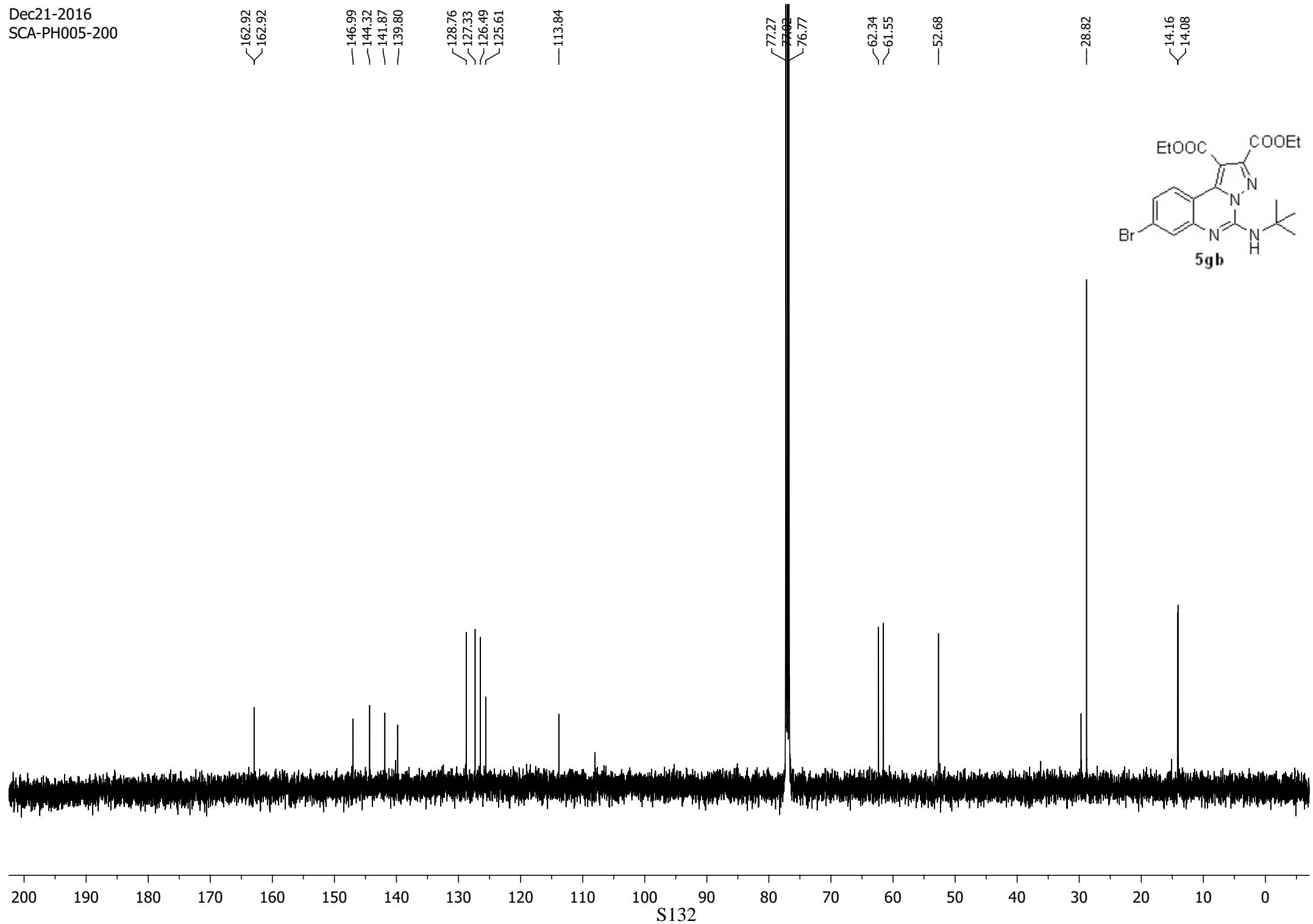
4.5131
4.4988
4.4845
4.4759
4.4702
4.4616
4.4473
4.4330

1.4686
1.4543
1.4399
1.4351
1.4206
1.4065









Jan06-2017
SCA-PH005-209

-9.4559

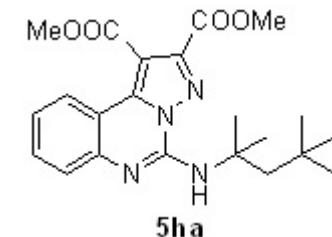
8.7678
8.7657
8.7514
8.7494
8.75641
7.7019
7.7004
7.6854
7.6839
7.6281
7.6168
7.6141
7.6118
7.3340
7.3318
7.3295
7.2689
6.5140

4.0263
3.9904

-2.0807

-1.6871

-1.0227



10.5 10.0 9.5 9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0 -0.5 -1.0

S133

1.00

1.02

1.15

1.13

1.00

2.96

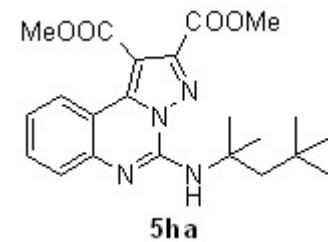
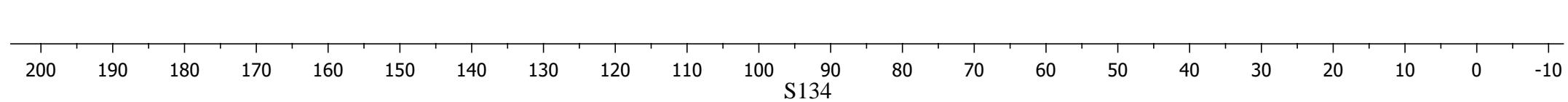
3.25

2.16

6.72

9.09

Jan07-2017
SCA-PH005-209



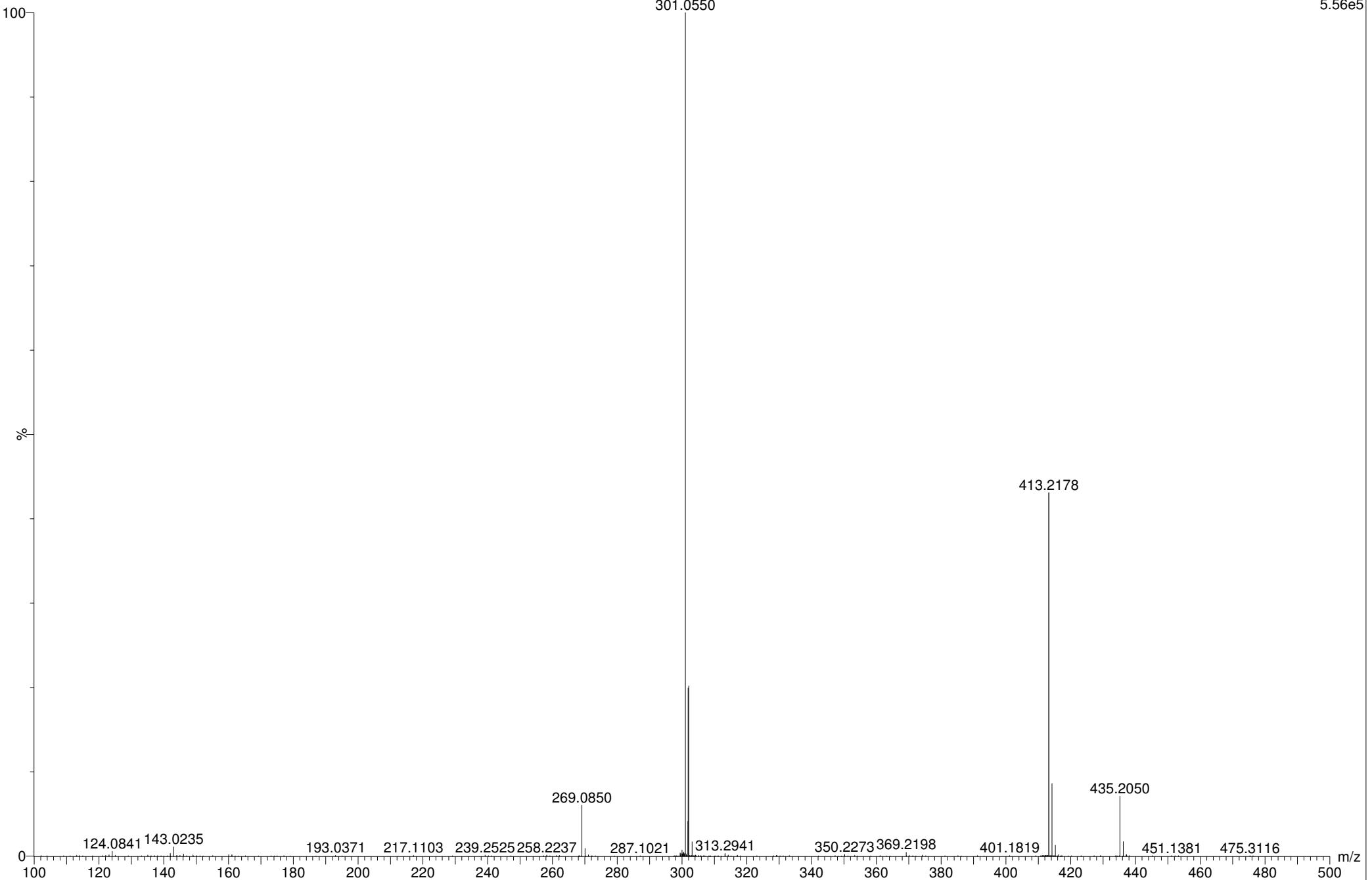
PROCESSED BY
PAWAN KUMAR

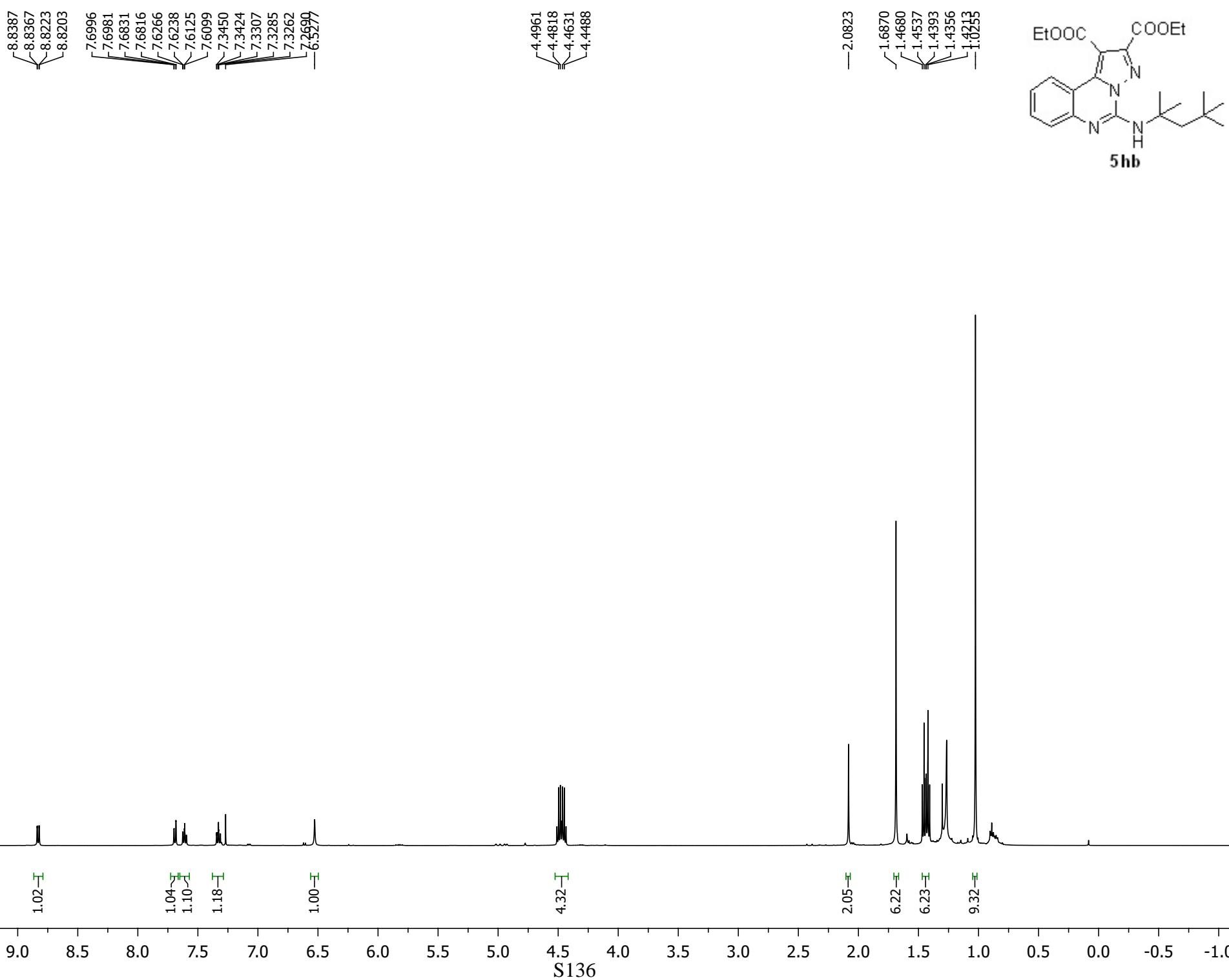
CSIR-IHBT
NPC&PD DIVISION

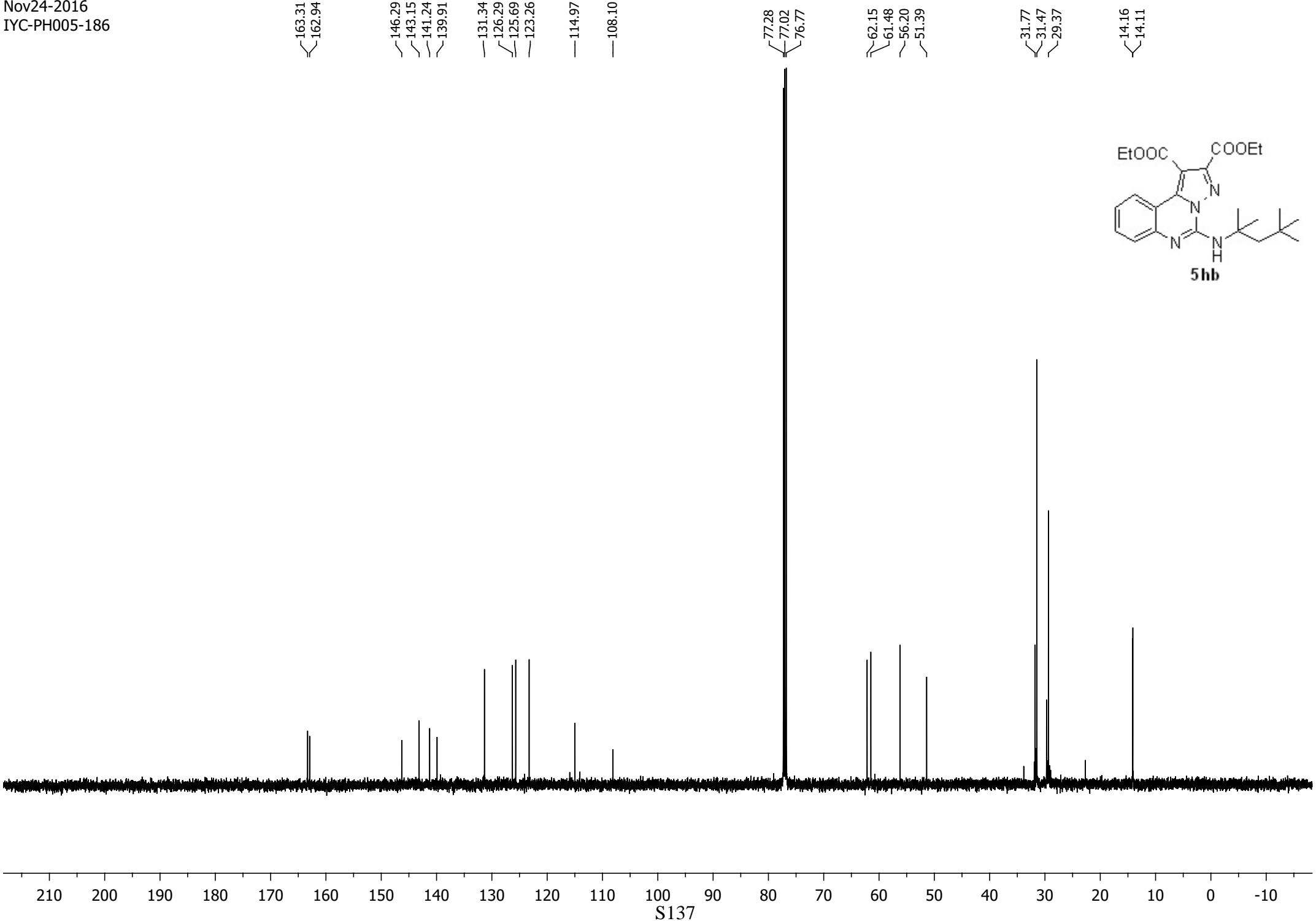
19-Jan-2017 15:21:11

SCA-PH005-209_02 39 (0.722) AM (Cen,3, 80.00, Ar,5000.0,413.22,0.70); Sb (2,10.00); Cm (37:44)

1: TOF MS ES+
5.56e5







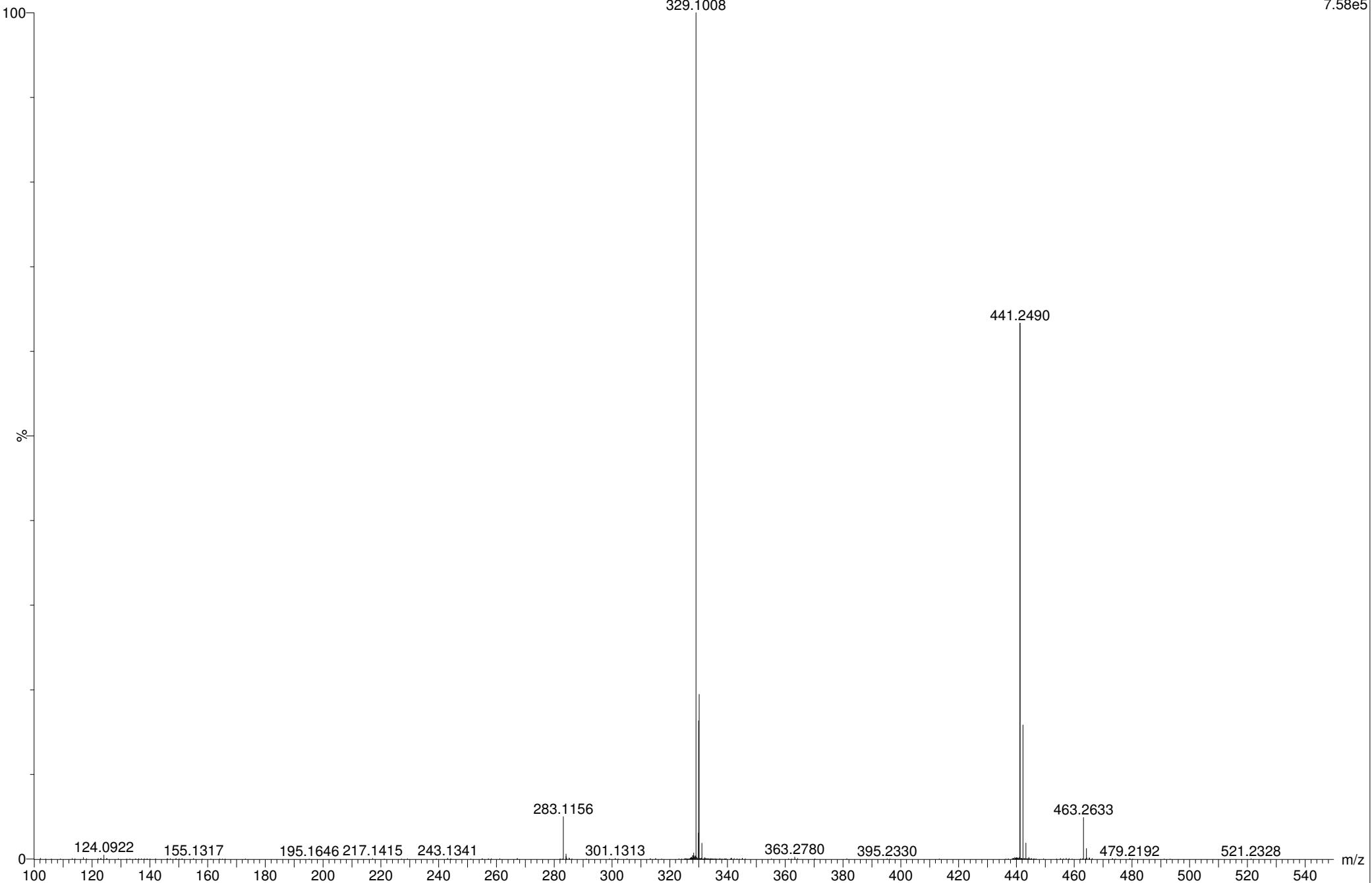
PROCESSED BY
PAWAN KUMAR

CSIR-IHBT
NPC&PD DIVISION

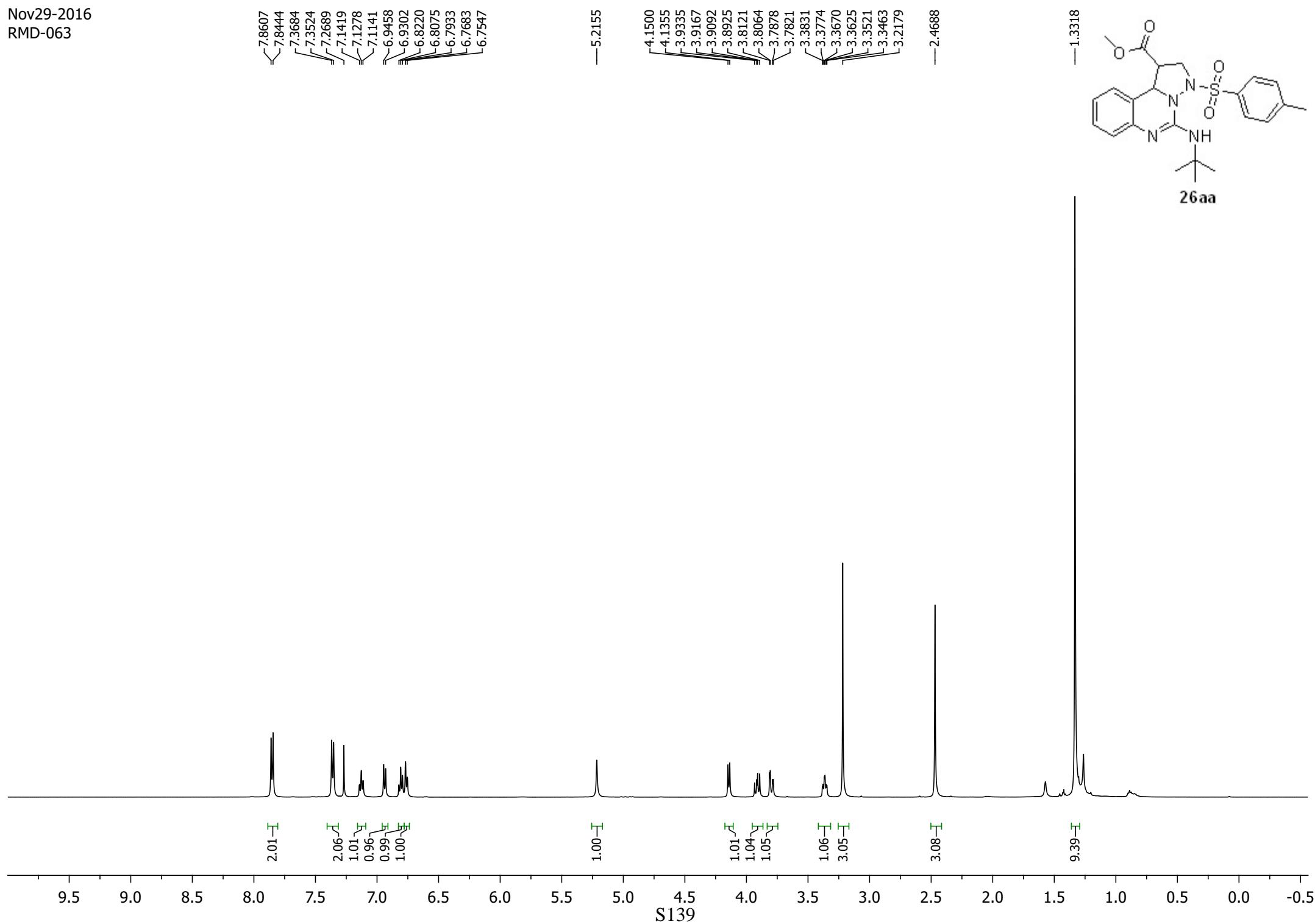
19-Jan-2017 14:12:58

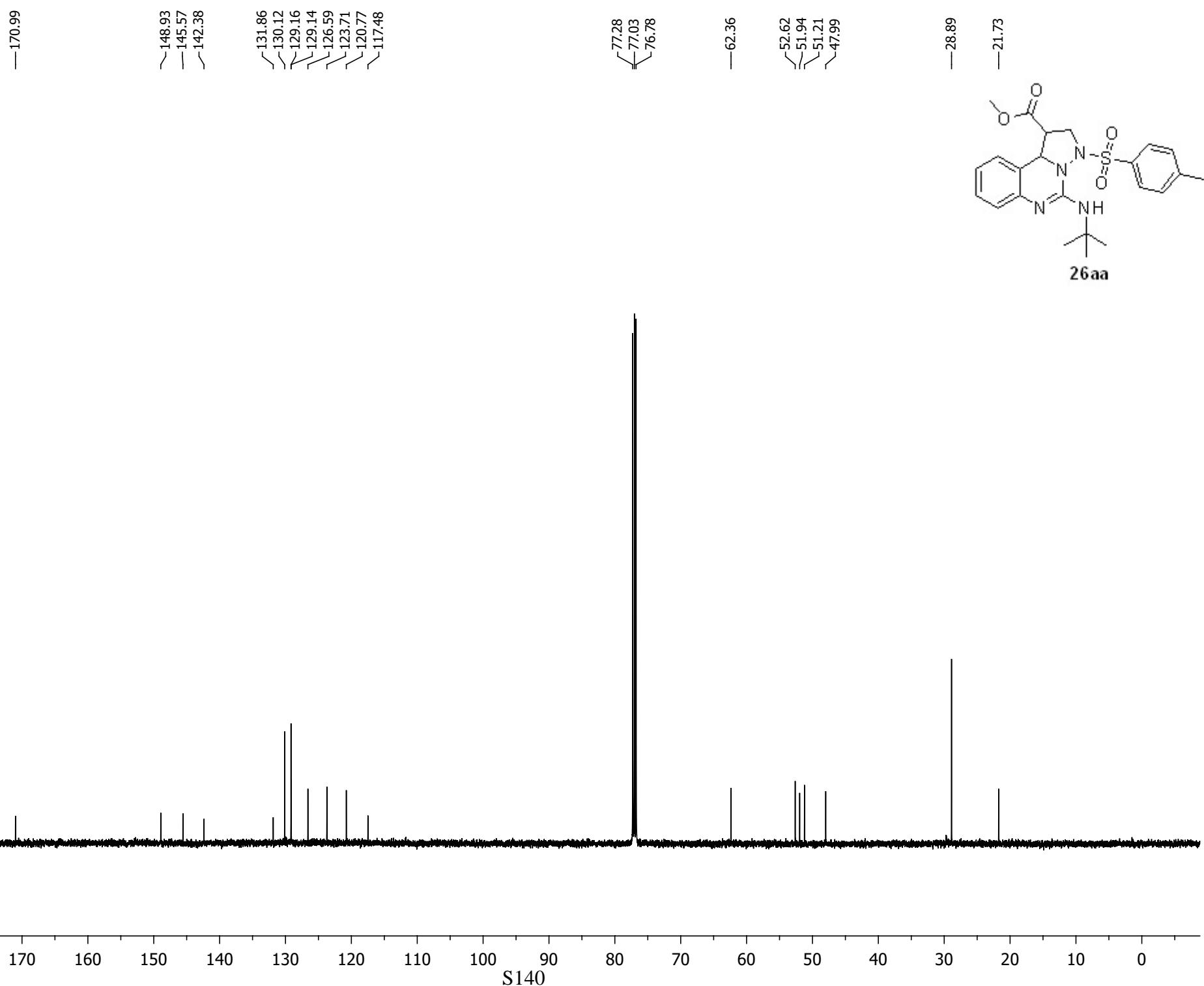
IYC-PH005-189_02 51 (0.946) AM (Cen,3, 80.00, Ar,5000.0,441.25,0.70); Sb (2,10.00); Cm (50:60)

1: TOF MS ES+
7.58e5



Nov29-2016
RMD-063





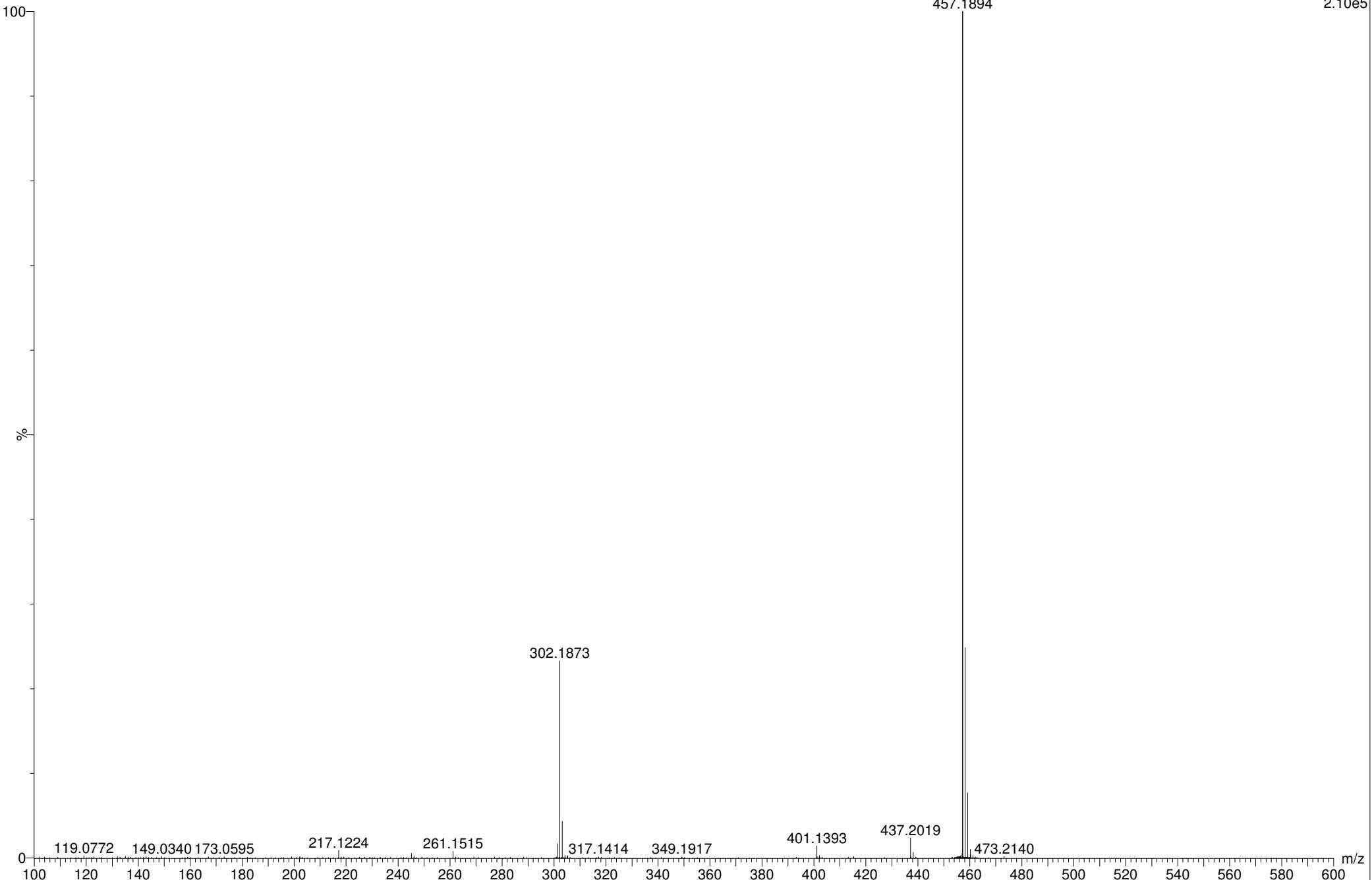
PROCESSED BY
PAWAN KUMAR

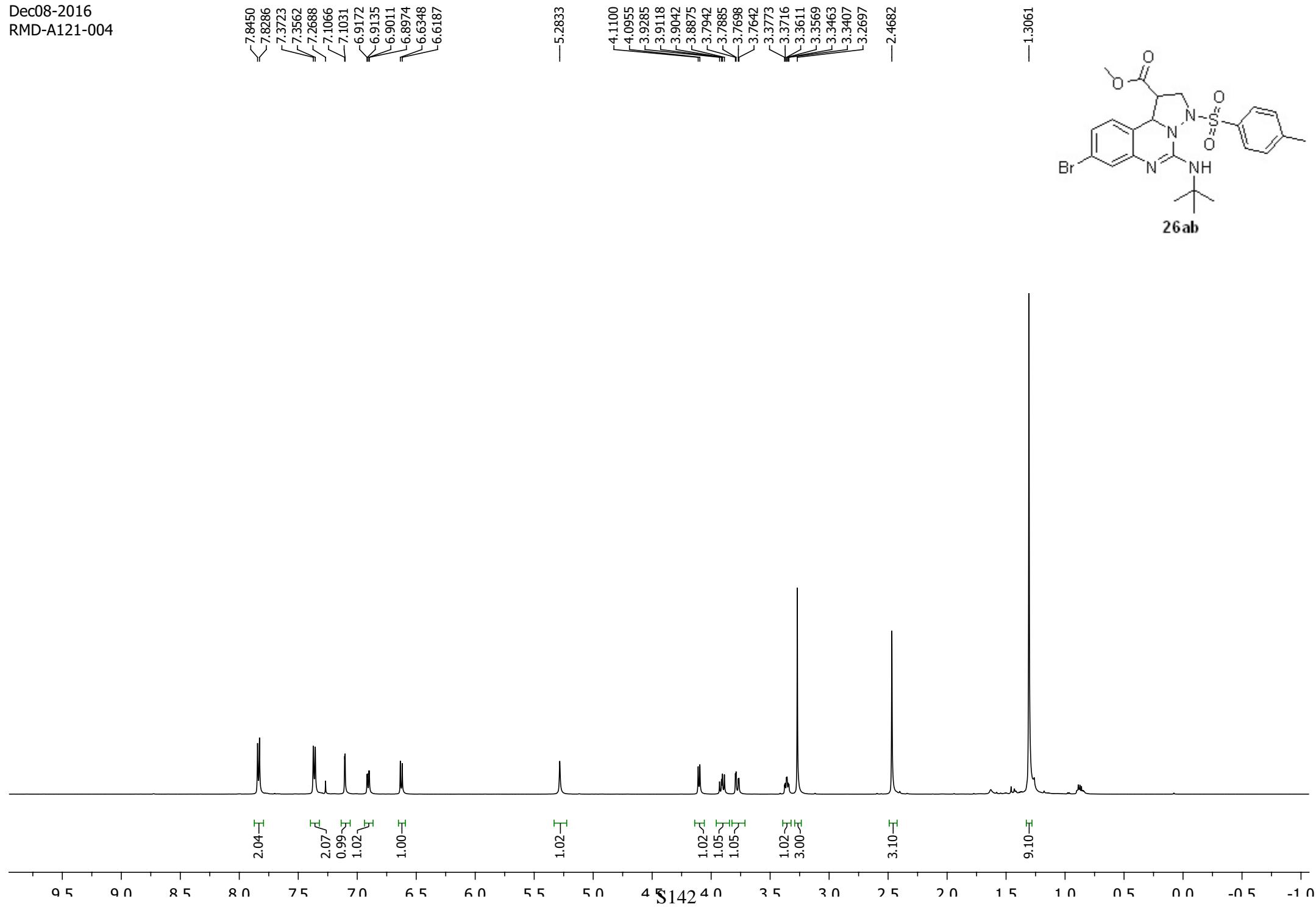
RMD063 20 (0.372) AM (Cen,3, 80.00, Ar,5000.0,457.19,0.70); Sb (2,10.00); Cm (17:27)

CSIR-IHBT
NPC&PD DIVISION

24-Nov-2016 16:18:44

1: TOF MS ES+
2.10e5





—170.81

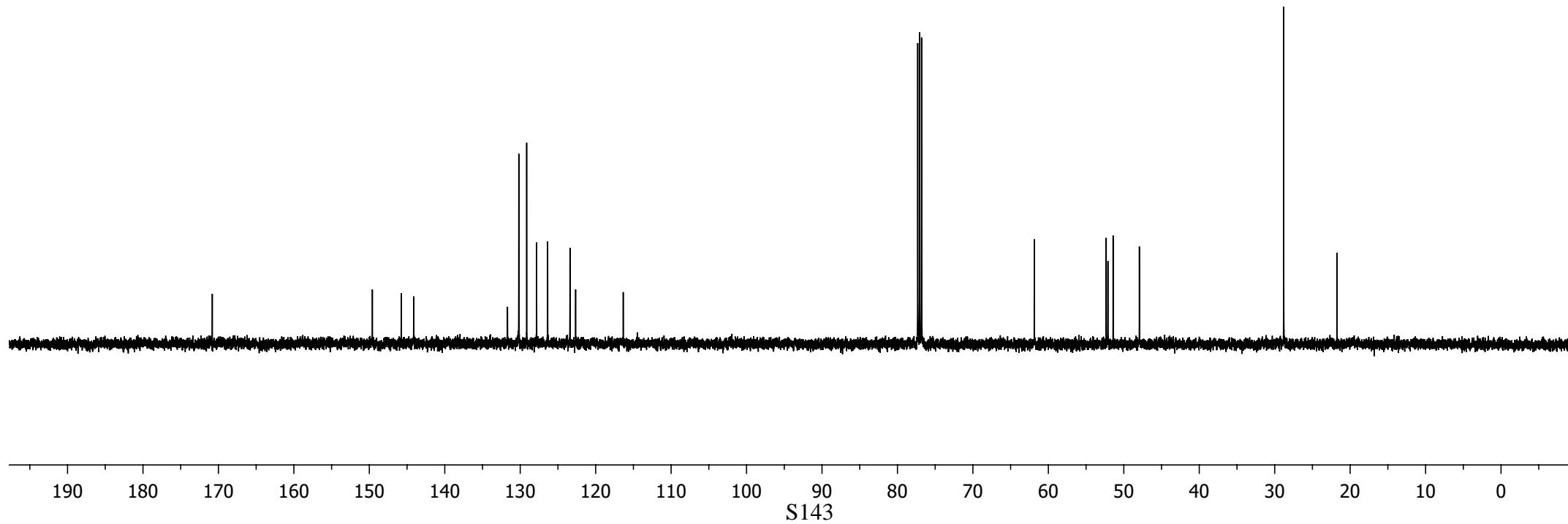
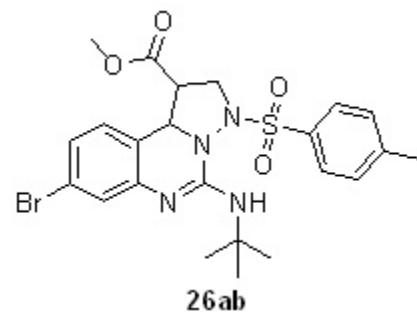
—149.60
—145.77
—144.13

131.68
130.19
129.14
127.83
126.36
123.40
122.66
—116.35

77.31
77.06
76.81

—61.86
52.33
52.10
51.42
47.94

—28.81
—21.75



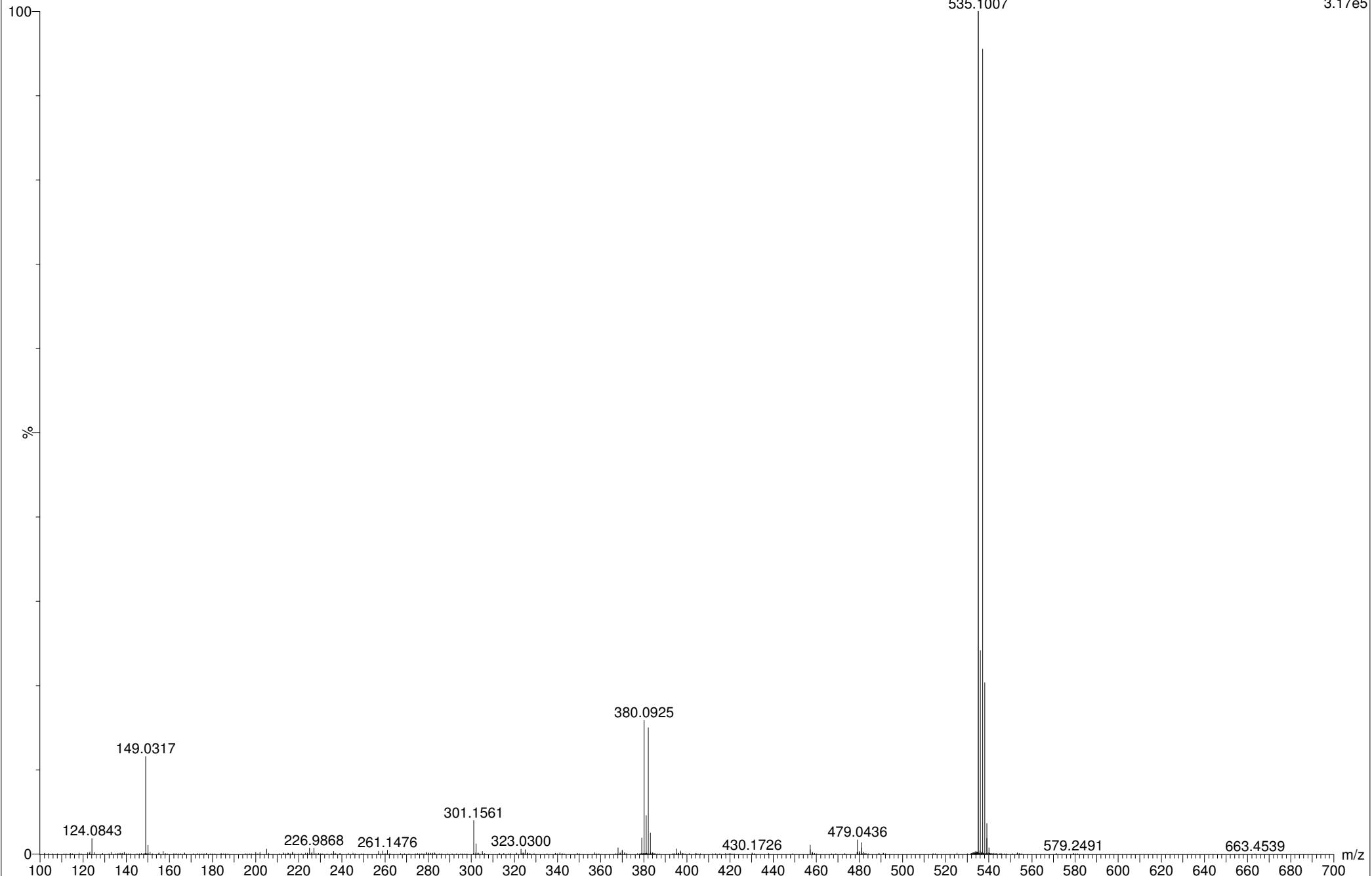
PROCESSED BY
PAWAN KUMAR

CSIR-IHBT
NPC&PD DIVISION

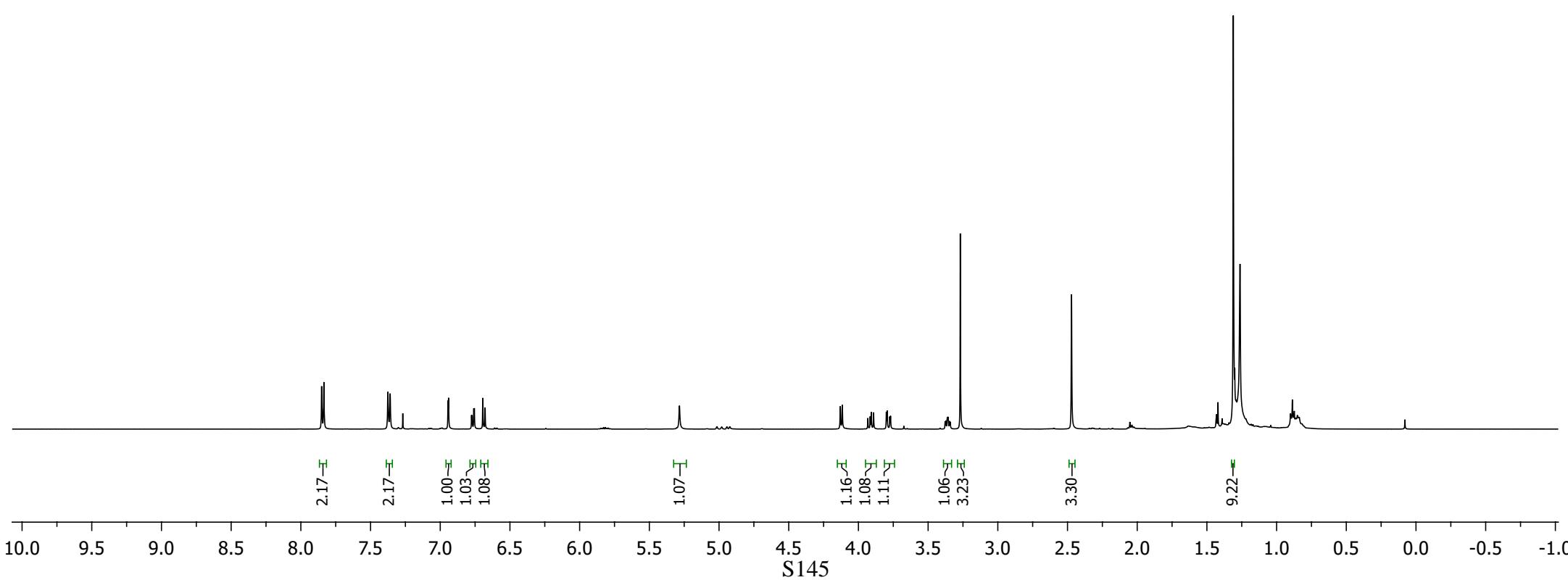
19-Jan-2017 15:05:04

MCR-A121-004_02 23 (0.427) AM (Cen,3, 80.00, Ar,5000.0,535.10,0.70); Sb (2,10.00); Cm (20:30)

1: TOF MS ES+
3.17e5



Dec29-2016
A121-RMD-024



<7.8347

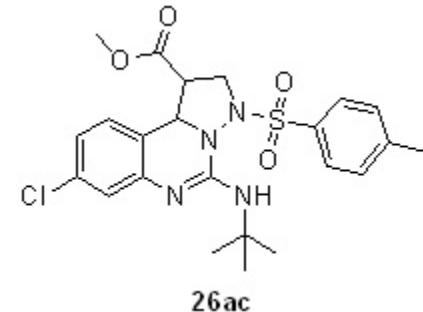
<7.3757
<7.3597
<7.2689
<6.9443
<6.9403
<6.7755
<6.7714
<6.7594
<6.7553
<6.6947
<6.6785

-5.2842

4.1301
<4.1157
<3.9324
<3.9157
<3.9080
<3.8913
<3.7996
<3.7938
<3.7752
<3.7694
<3.3781
<3.3723
<3.3617
<3.3576
<3.3470
<3.3412
<3.2693

-2.4712

-1.3105



—170.83

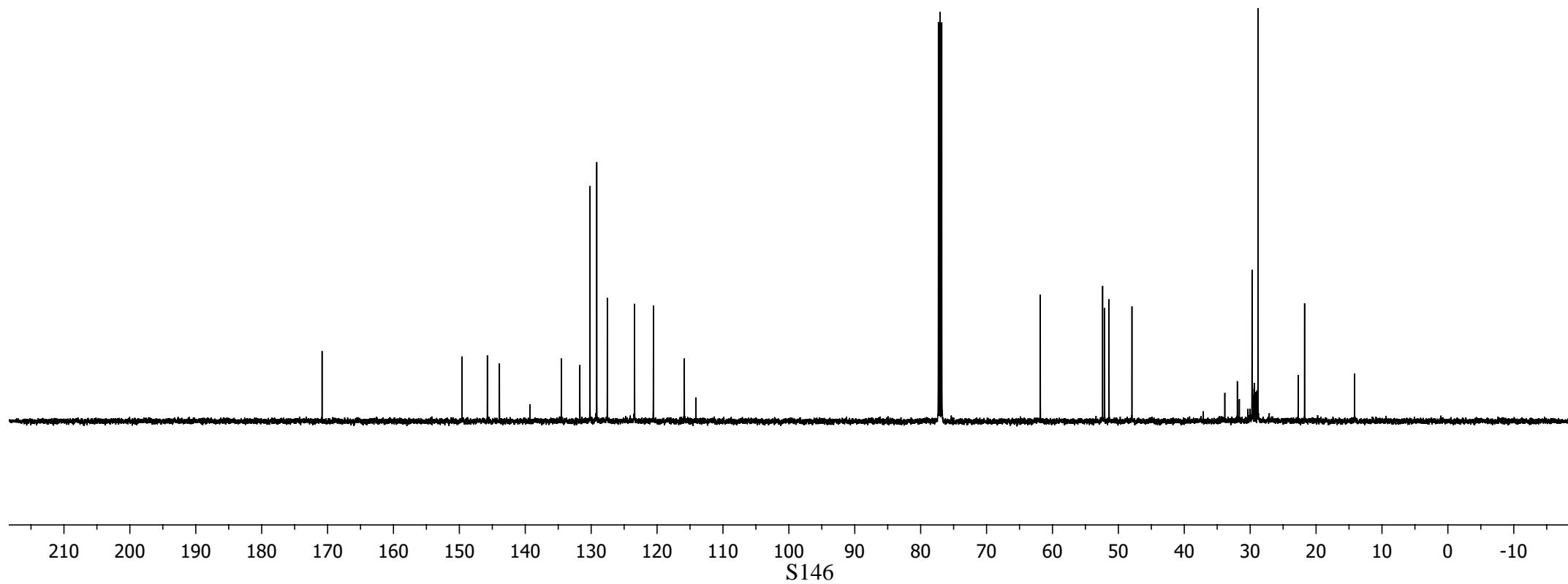
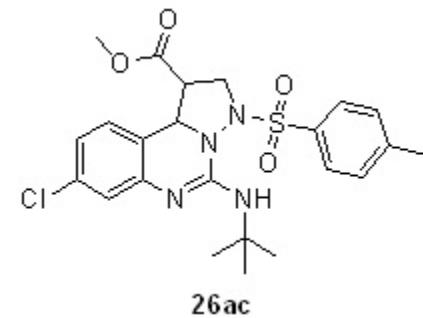
—149.60
—145.75
—143.92

—134.53
—131.71
—130.18
—129.15
—127.55
—123.41
—120.56
—115.86

77.31
77.05
76.80

—61.84
52.39
52.09
51.41
47.94

—23.81
—21.74



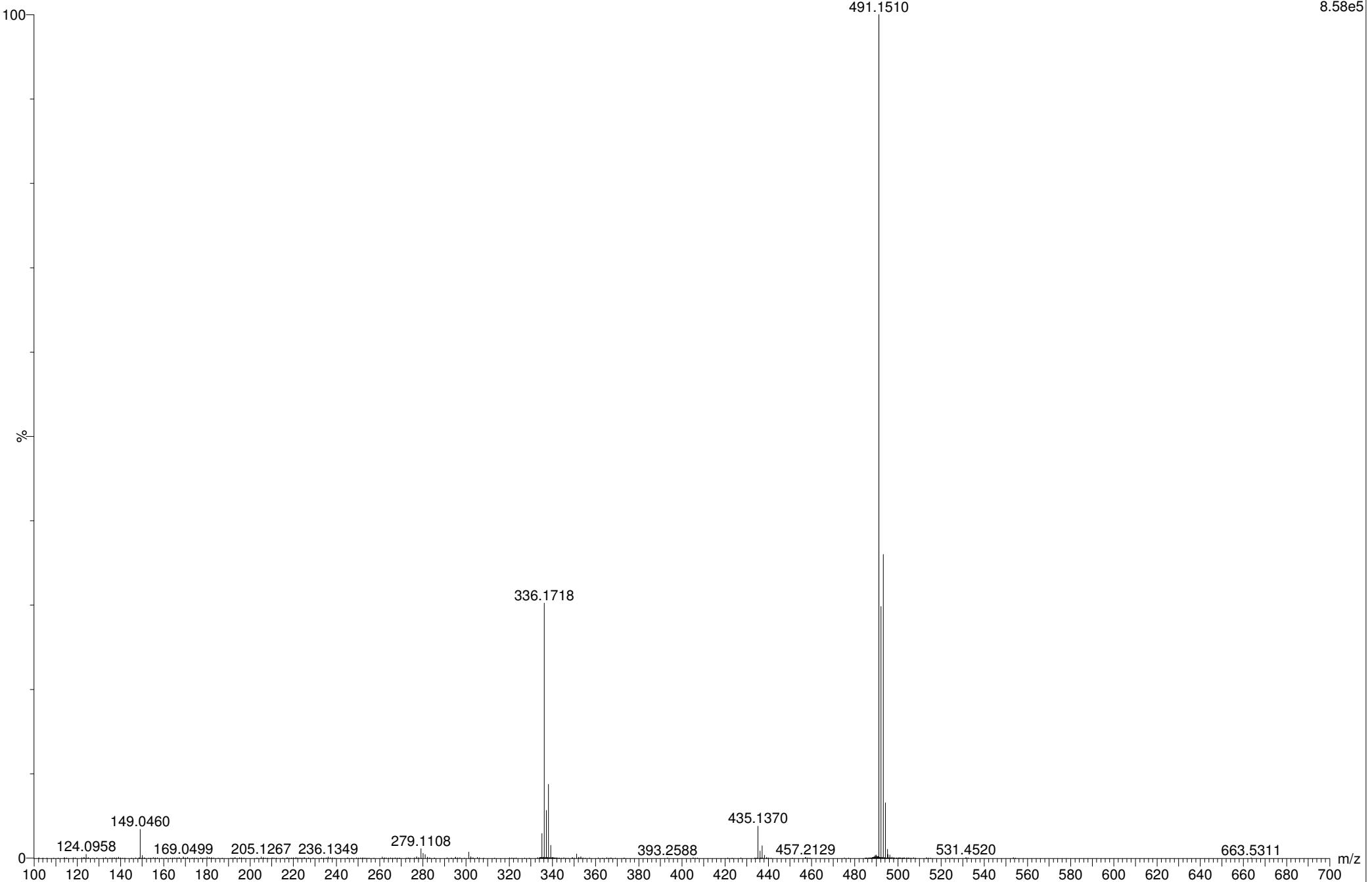
PROCESSED BY
PAWAN KUMAR

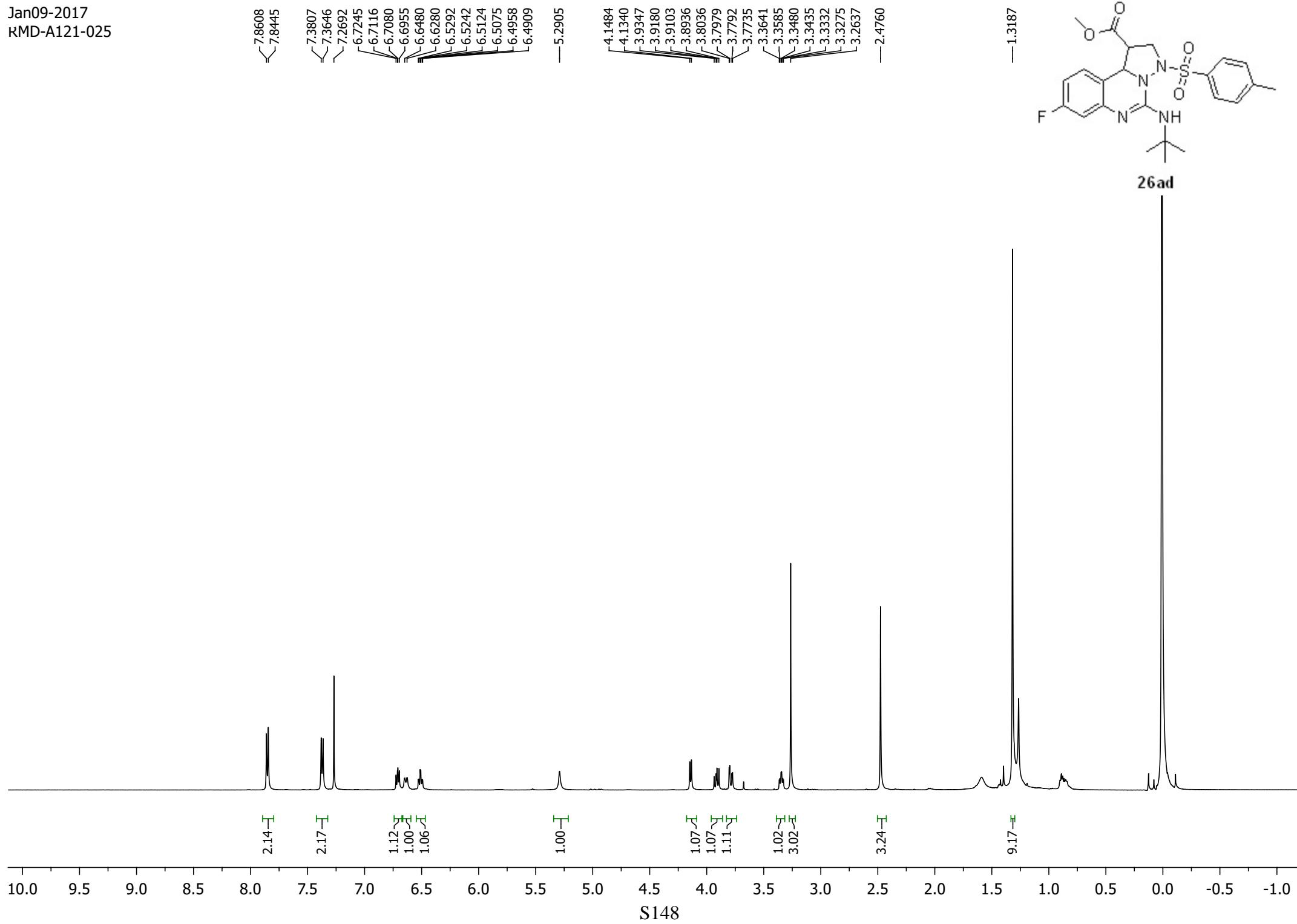
A121-024_02 22 (0.408) AM (Cen,3, 80.00, Ar,5000.0,491.15,0.70); Sb (2,10.00); Crn (19:31)

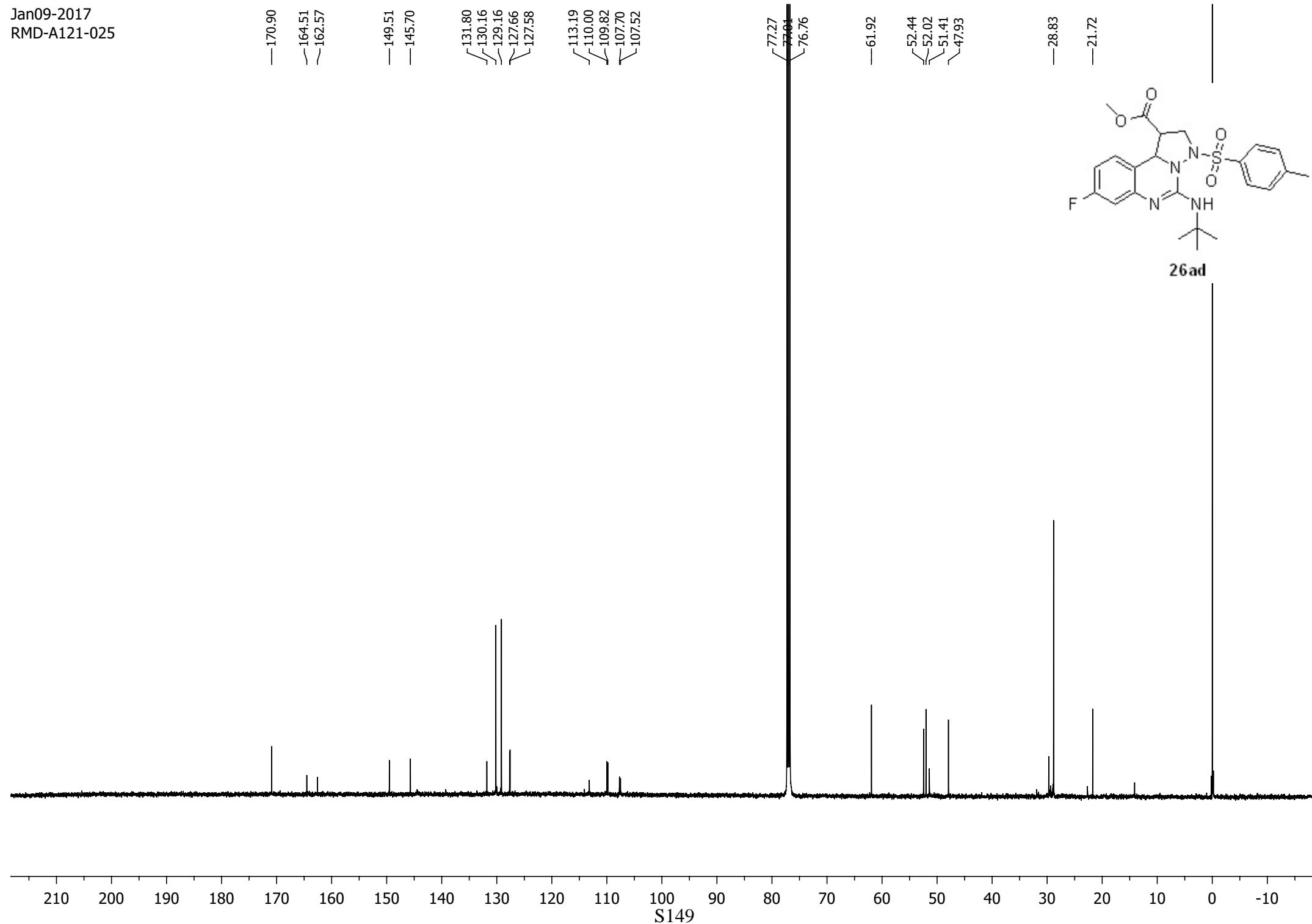
CSIR-IHBT
NPC&PD DIVISION

19-Jan-2017 14:09:03

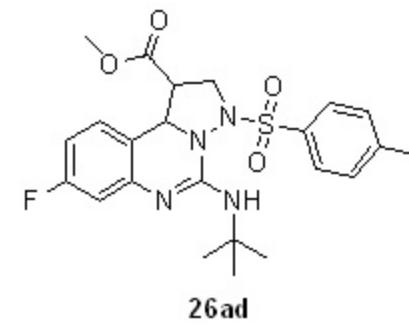
1: TOF MS ES+
8.58e5



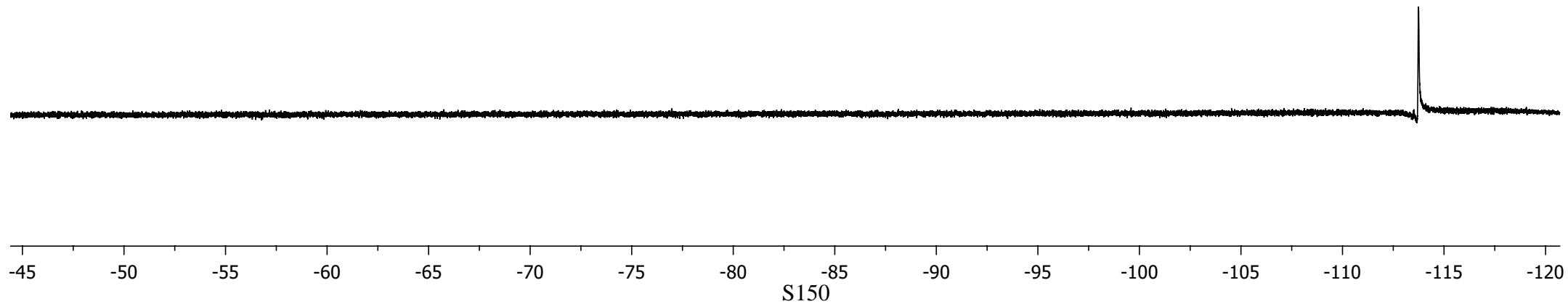




Jan09-2017
RMD-A121-025



-113.7427



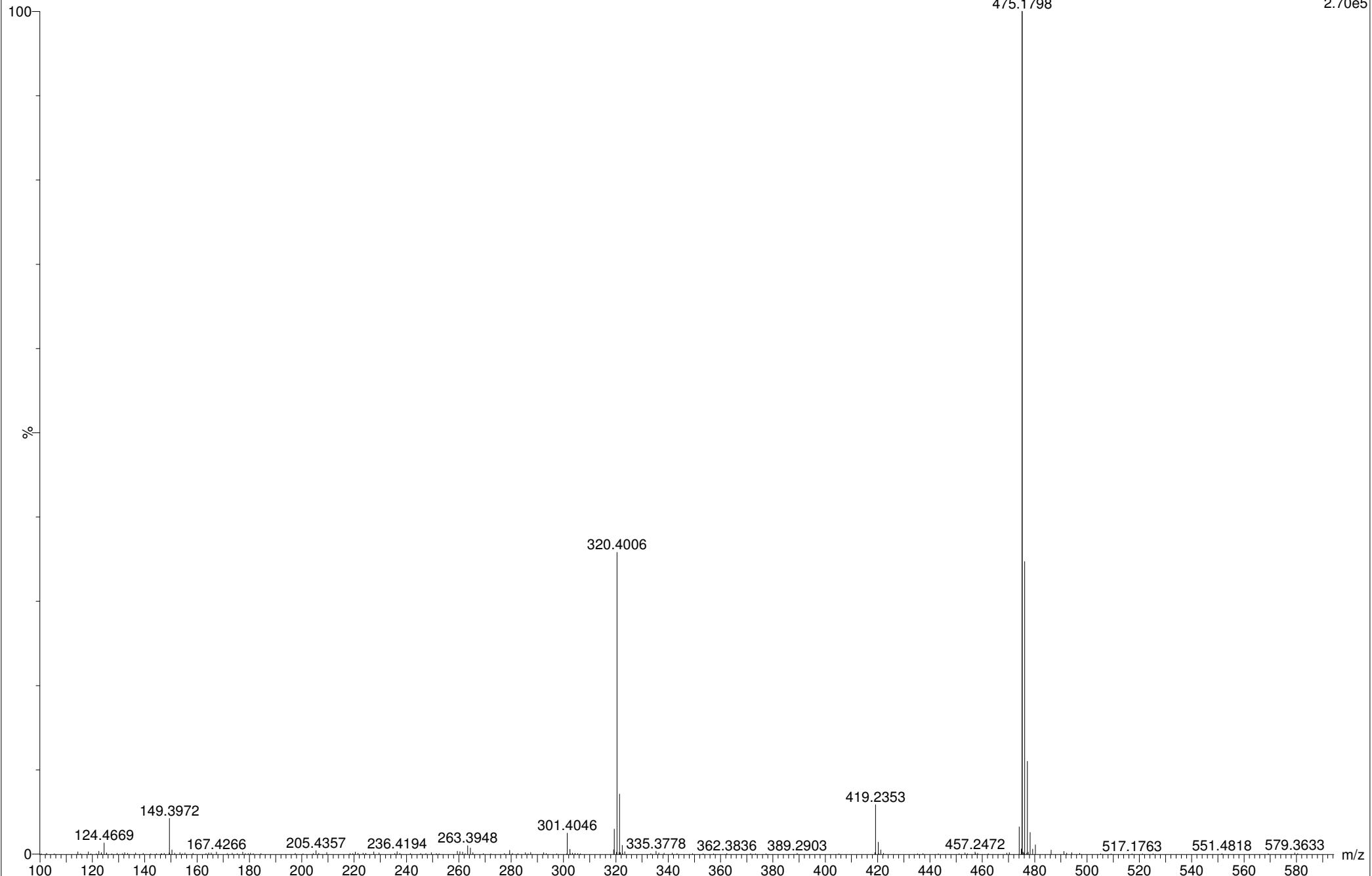
PROCESSED BY
PAWAN KUMAR

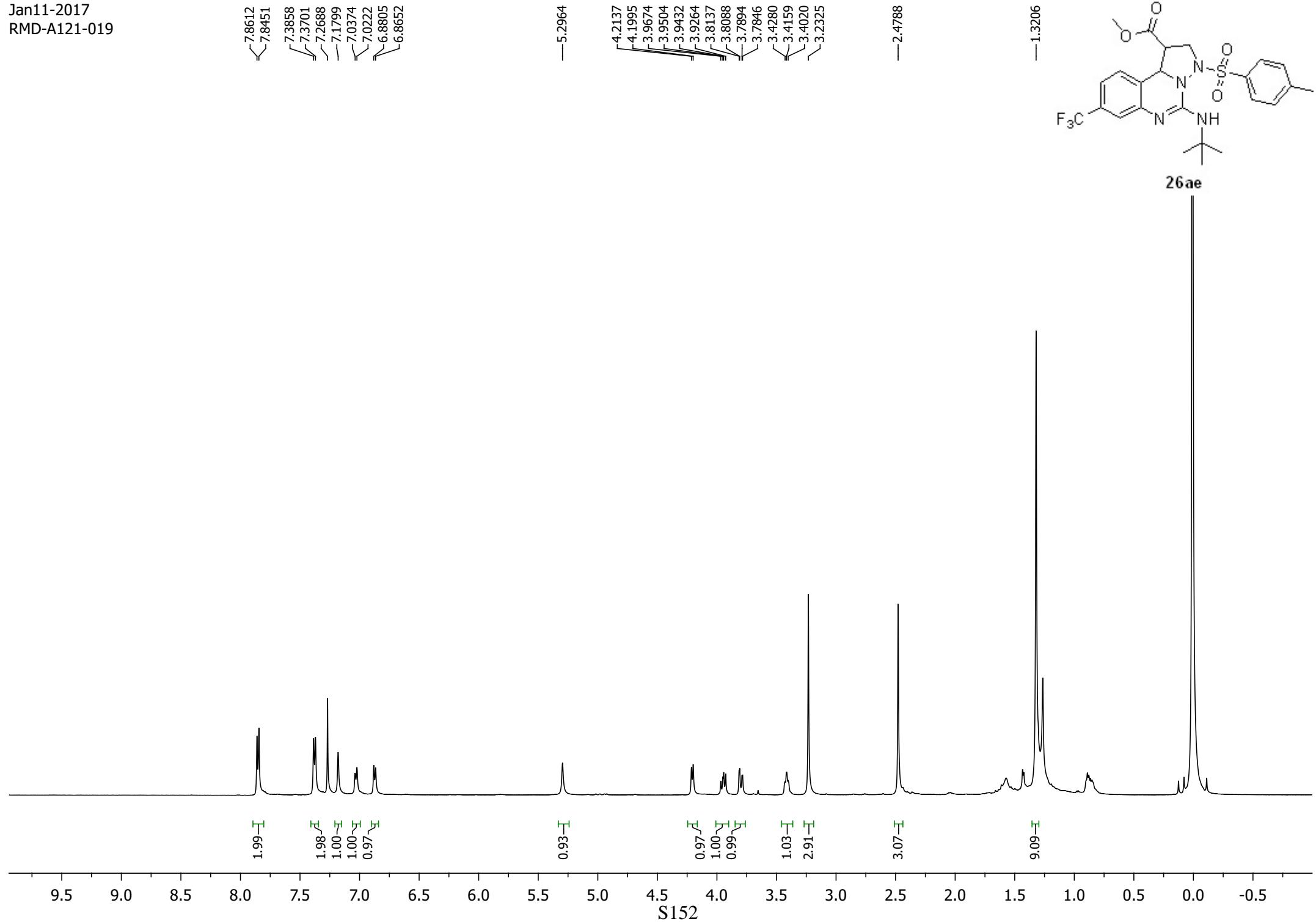
CSIR-IHBT
NPC&PD DIVISION

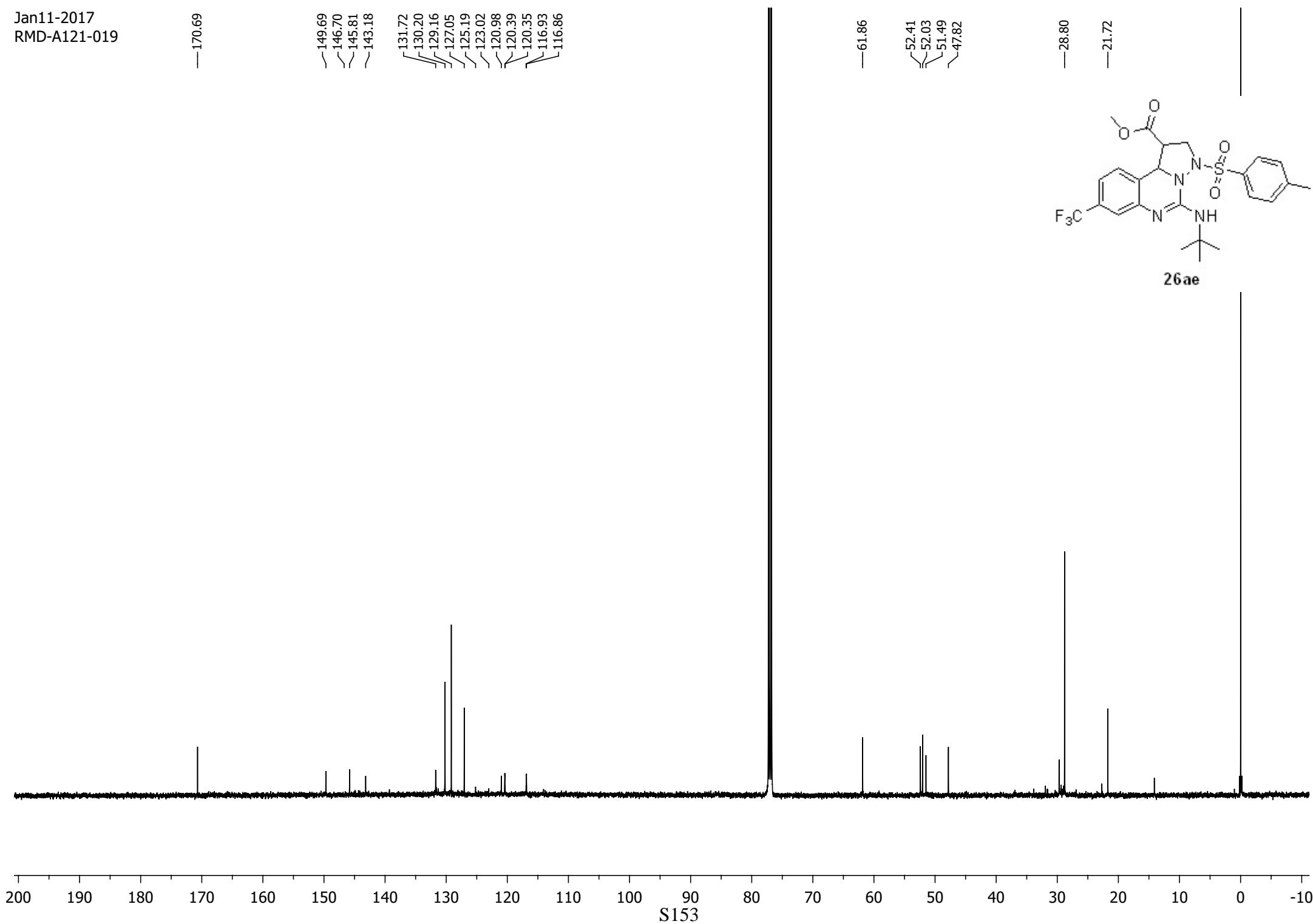
24-May-2017 12:49:34

A121-RMD-025_01 22 (0.408) AM (Top,10, Ar,5000.0,475.18,0.70); Sm (SG, 1x1.00); Cm (19:36)

1: TOF MS ES+
2.70e5

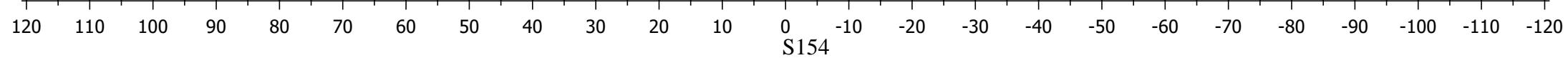
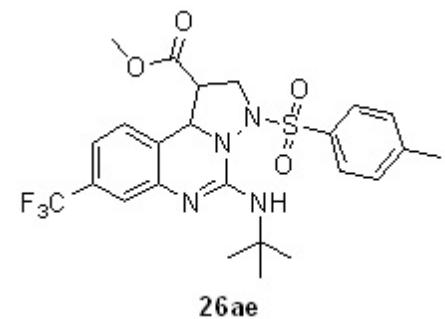






Jan11-2017
RMD-A121-019

-62.8836



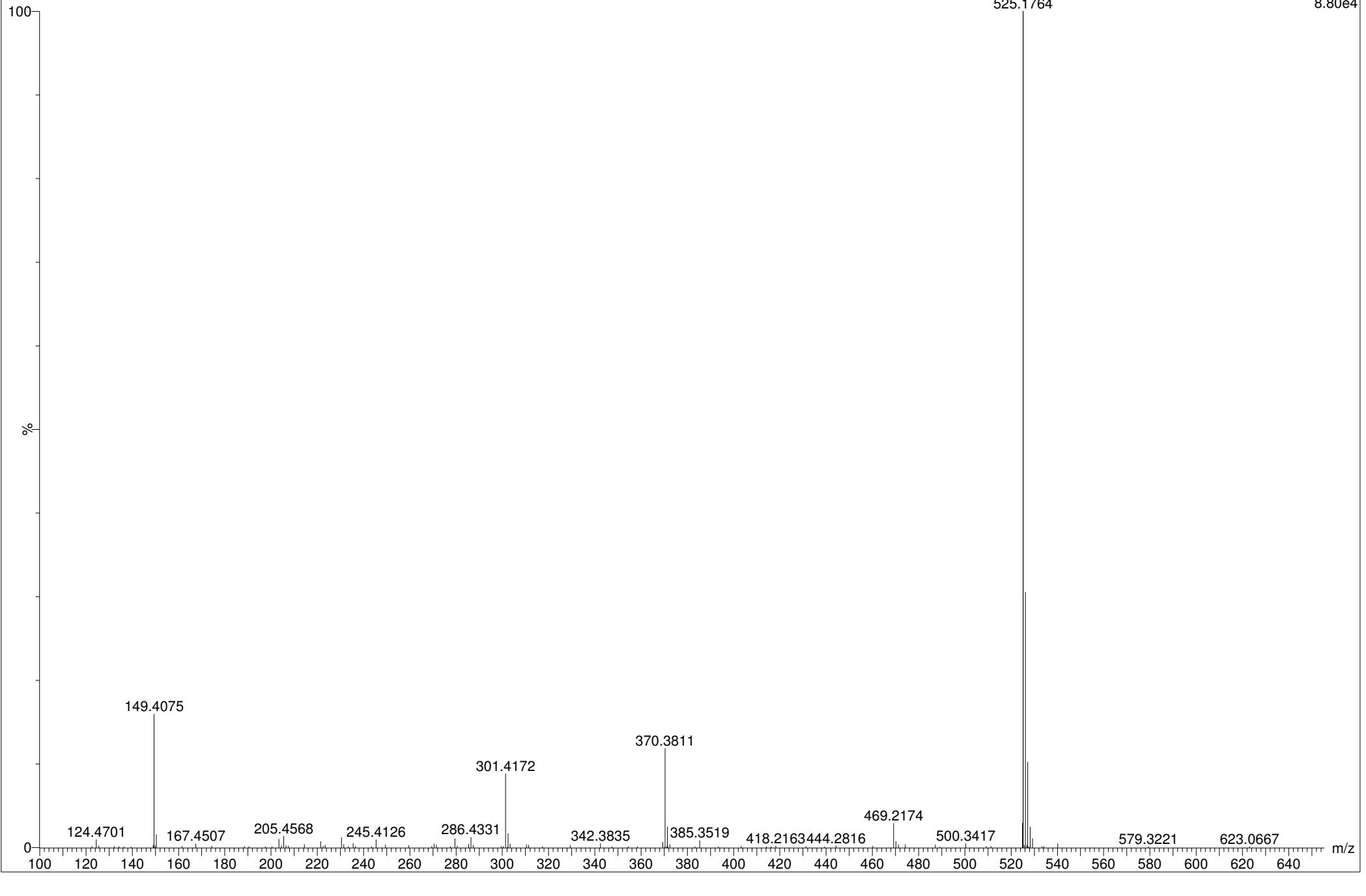
PROCESSED BY
PAWAN KUMAR

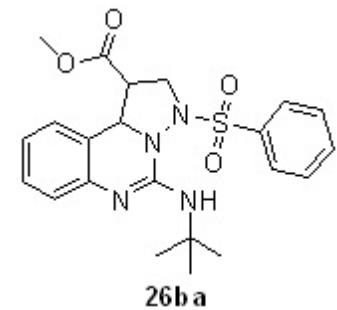
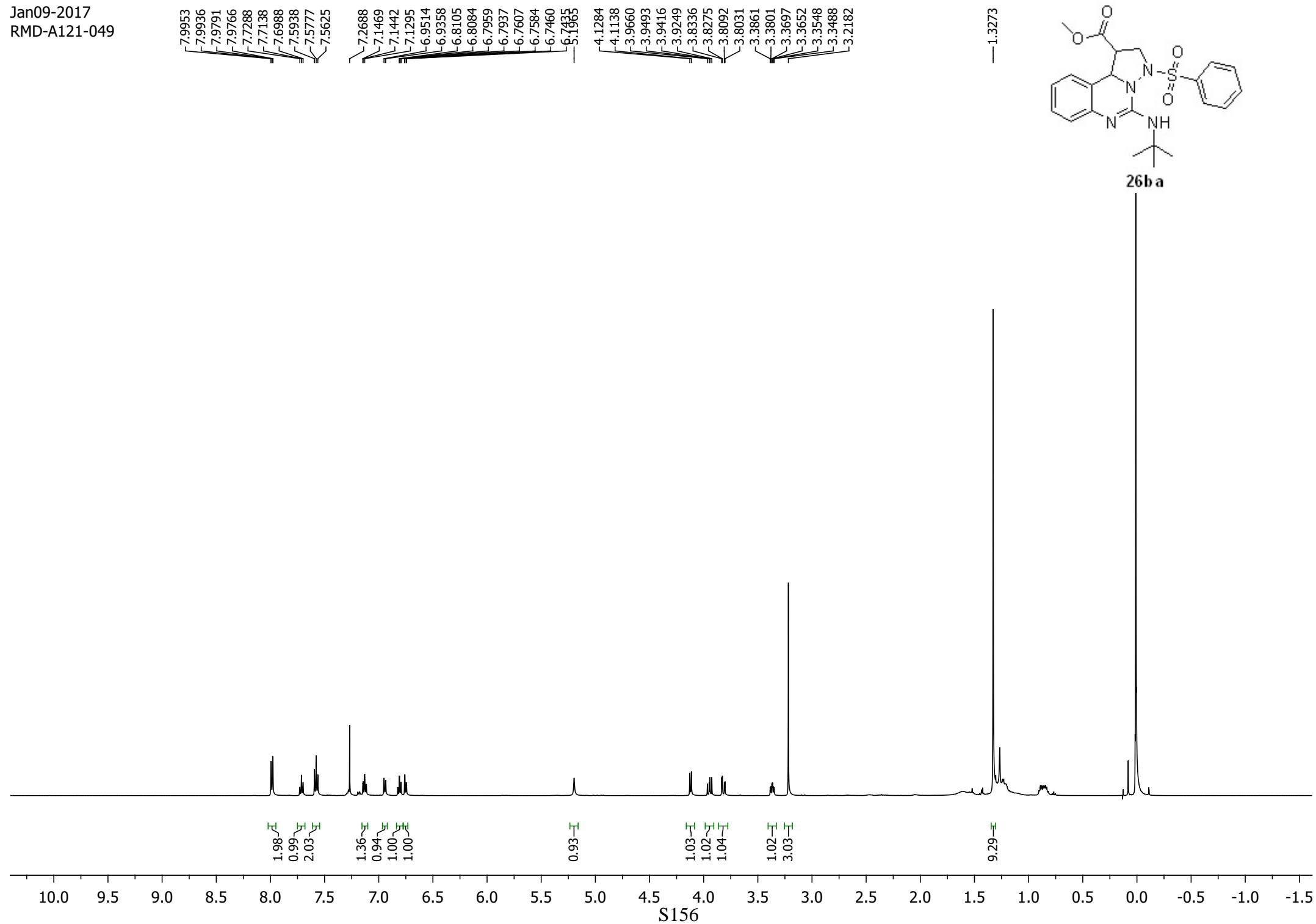
CSIR-IHBT
NPC&PD DIVISION

24-May-2017 12:53:29

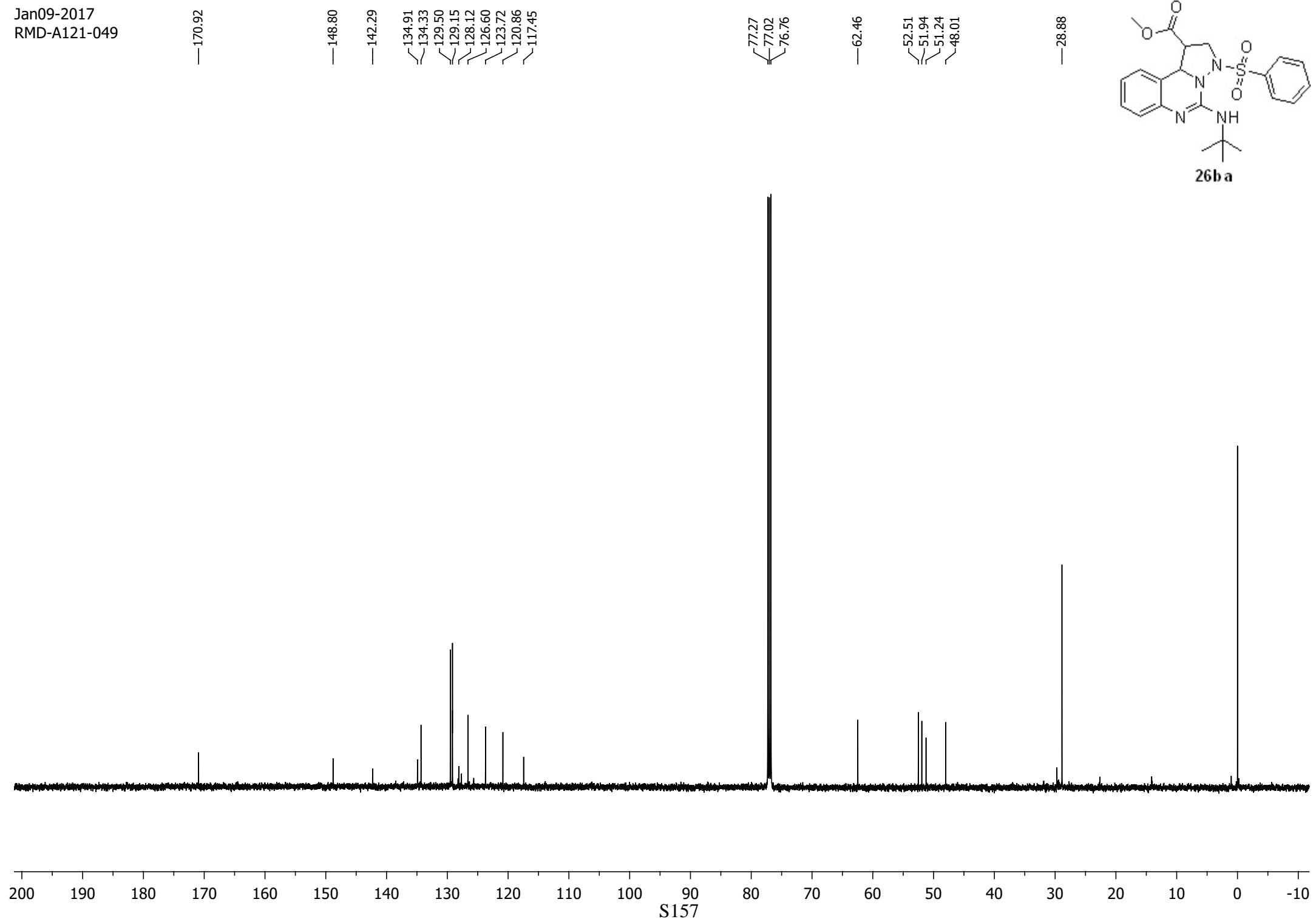
A121-RMD-019_01 29 (0.538) AM (Top,10, Ar,5000.0,525.18,0.70); Sm (SG, 1x1.00); Cm (26:38-13:22)

1: TOF MS ES+
8.80e4





Jan09-2017
RMD-A121-049



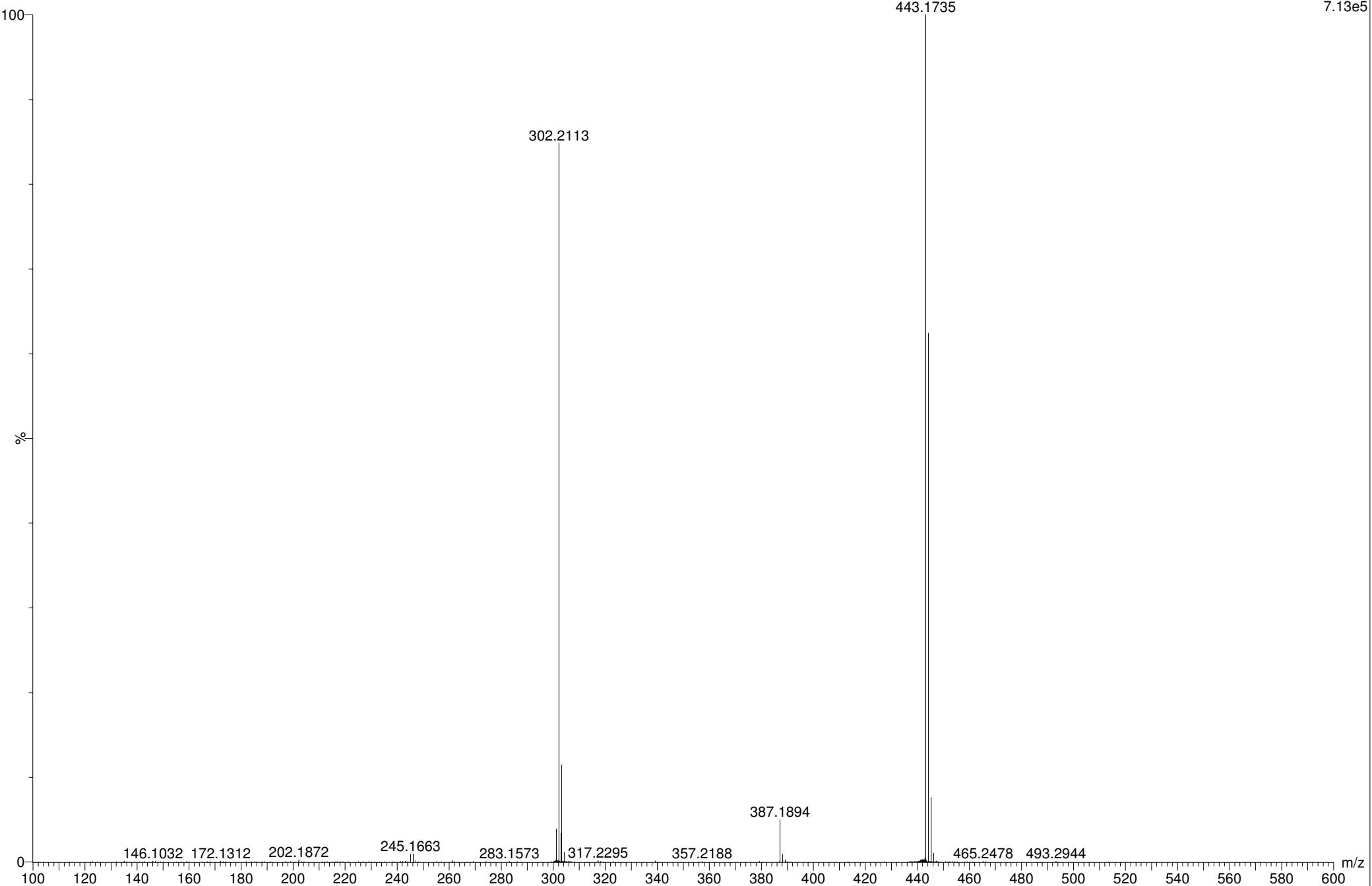
PROCESSED BY
PAWAN KUMAR

RMD-079 20 (0.371) AM (Cen,3, 80.00, Ar,5000.0,443.17,0.70); Sb (2,10.00); Cm (18:27)

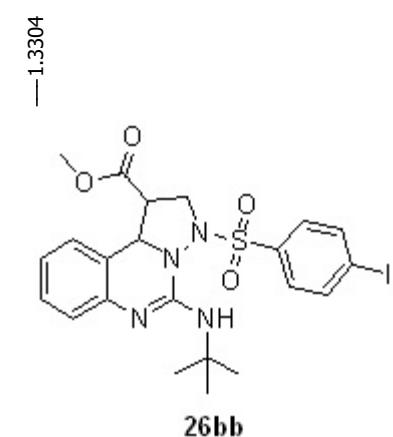
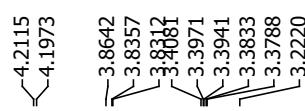
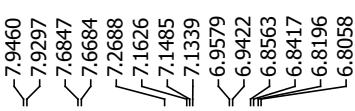
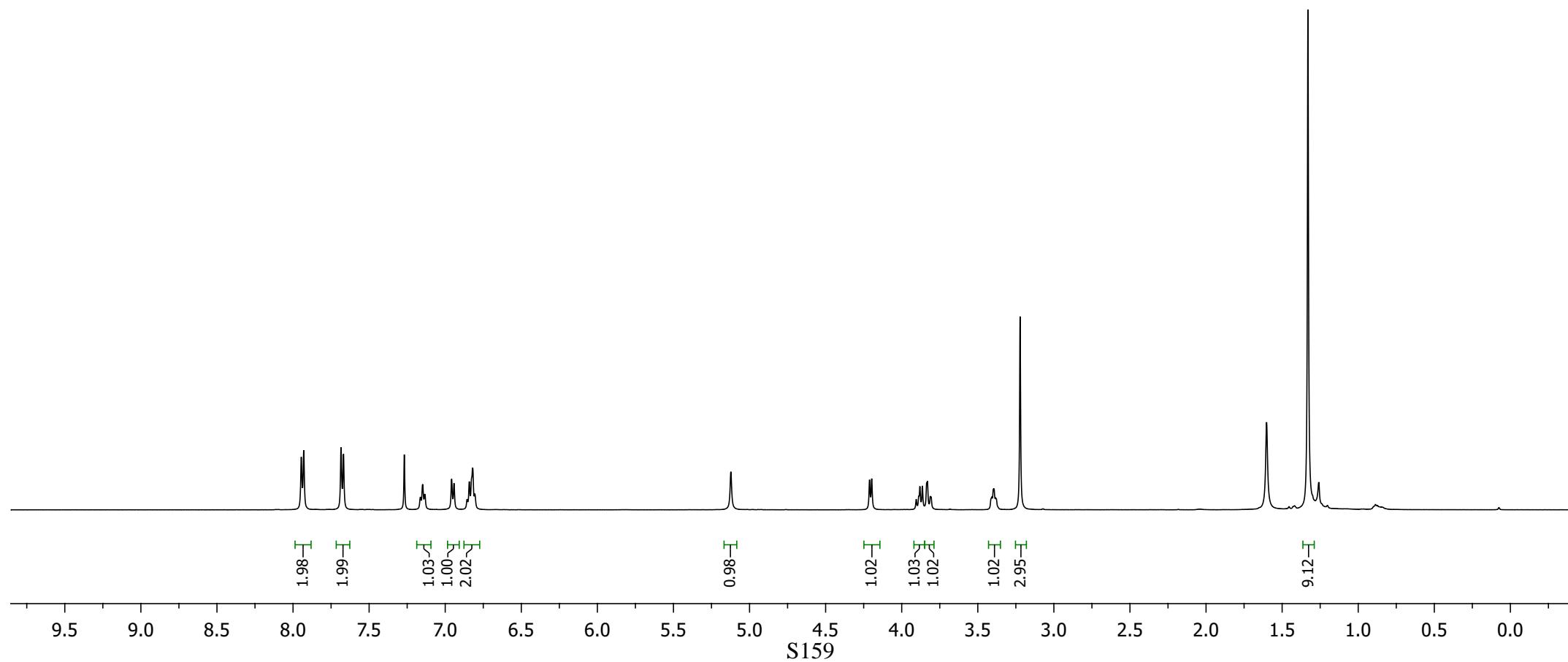
CSIR-IHBT
NPC&PD DIVISION

24-Nov-2016 16:43:17

1: TOF MS ES+
7.13e5



Jul05-2017
A124-022



Jul05-2017
A124-022

— 170.8439

— 148.5681

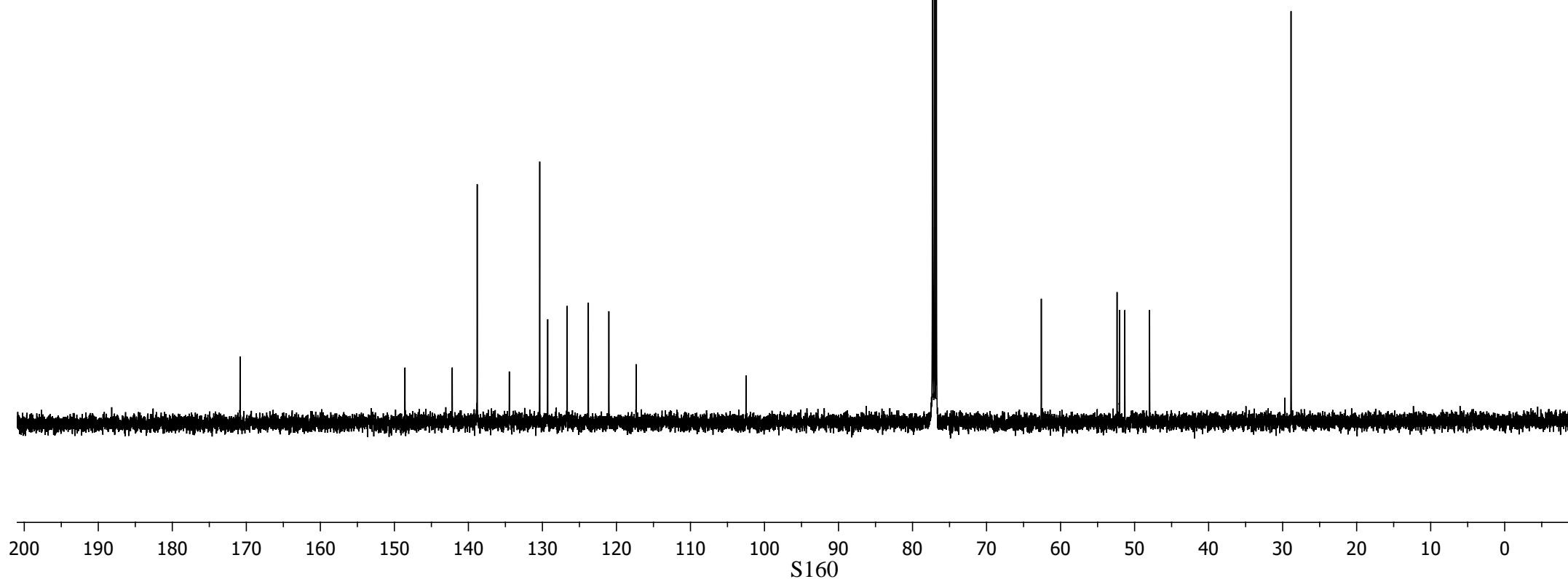
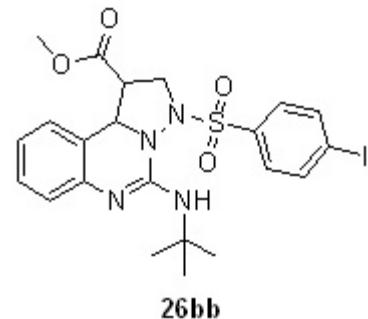
— 142.1835
— 138.8080
— 134.4715
— 130.3610
— 129.2846
— 126.6584
— 123.7935
— 121.0215
— 117.3402

— 102.4987

— 77.2943
— 77.0495
— 76.7864

— 52.3683
— 52.0431
— 51.3203
— 47.9788

— 28.8667



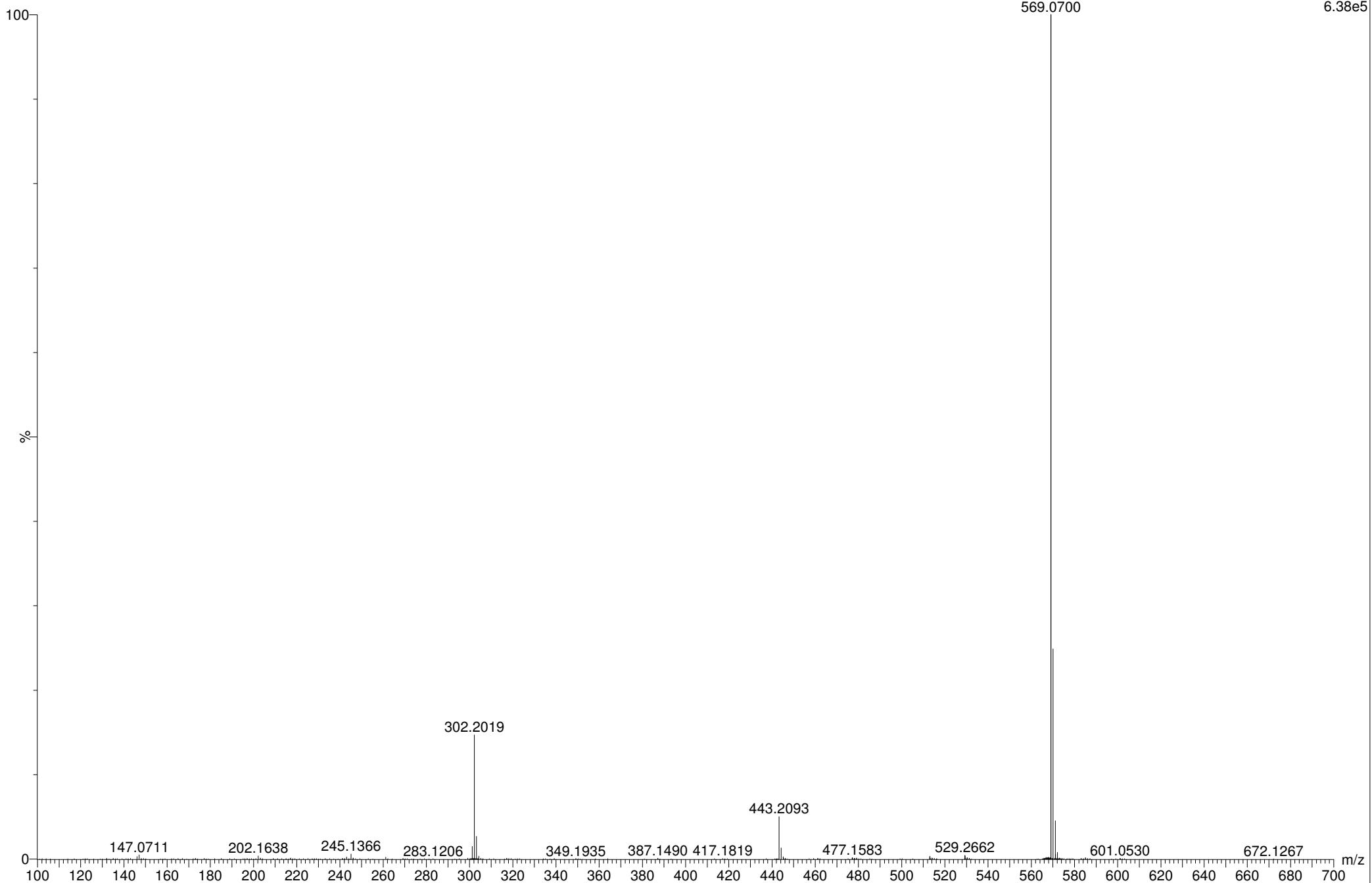
PROCESSED BY
PAWAN KUMAR

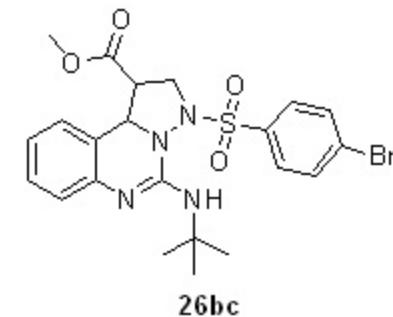
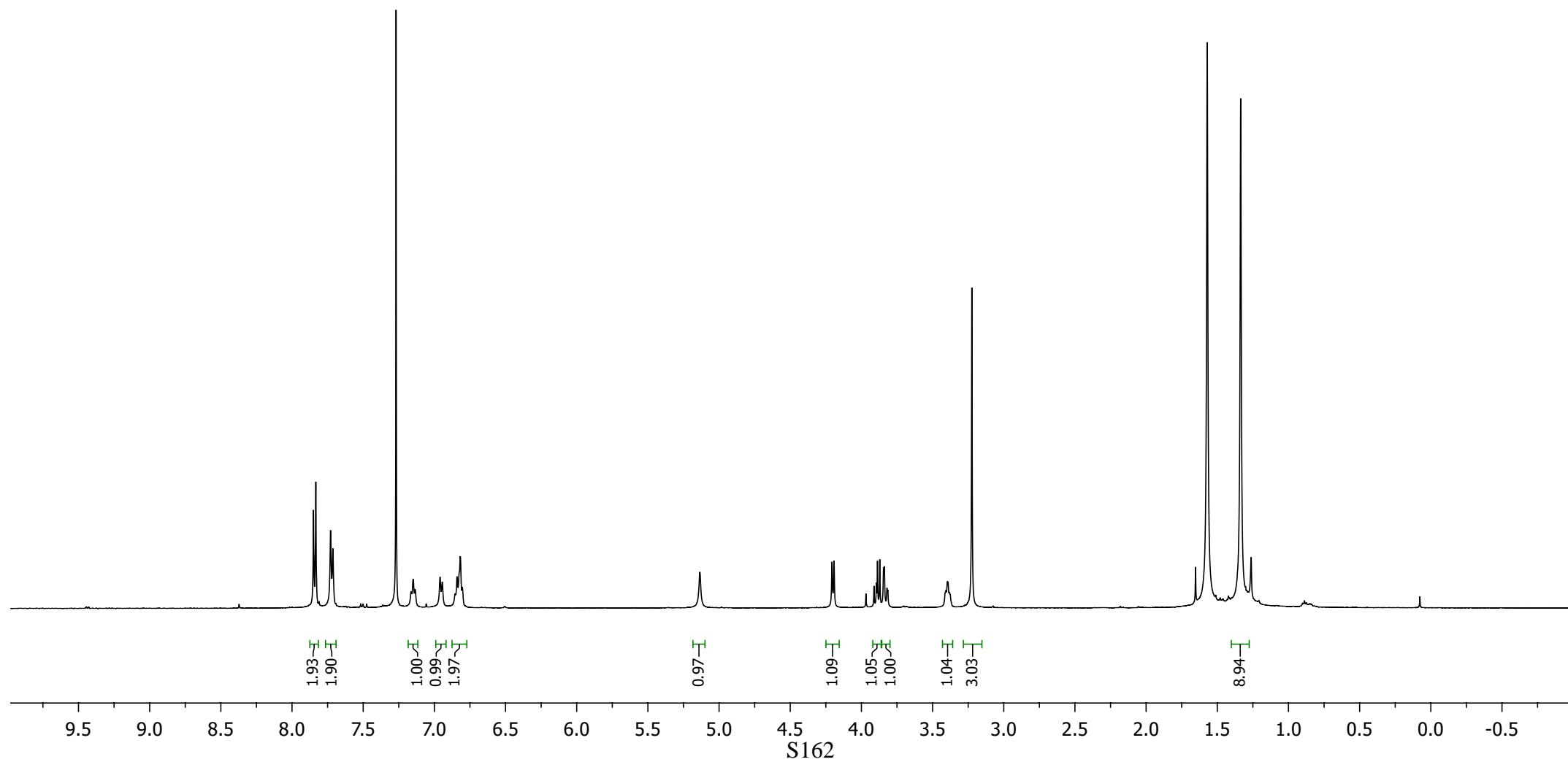
RMD-082 21 (0.390) AM (Cen,3, 80.00, Ar,5000.0,569.07,0.70); Sb (2,10.00); Cm (17:27)

CSIR-IHBT
NPC&PD DIVISION

24-Nov-2016 16:30:53

1: TOF MS ES+
6.38e5





Nov24-2016
RMD-101

-170.79

-148.57

-142.20

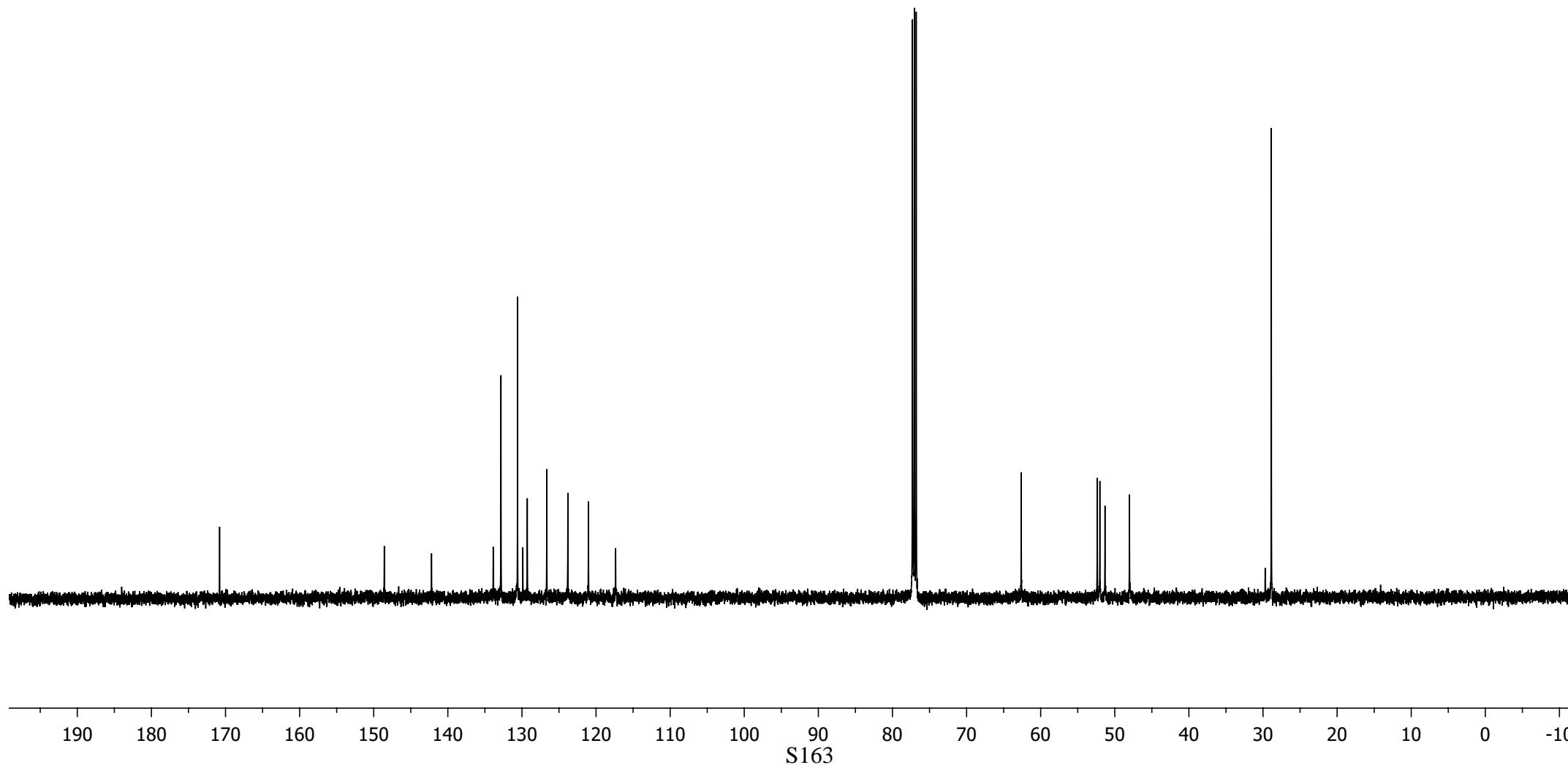
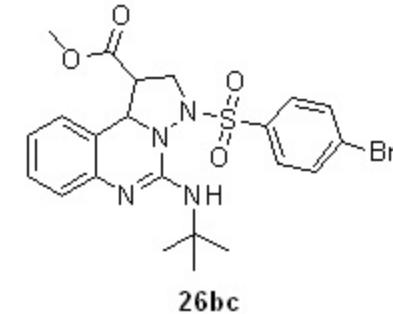
133.88
132.83
130.59
129.88
129.28
126.65
123.80
121.02
117.35

77.30
77.05
76.79

-62.62

52.37
52.01
51.32
48.02

-28.88



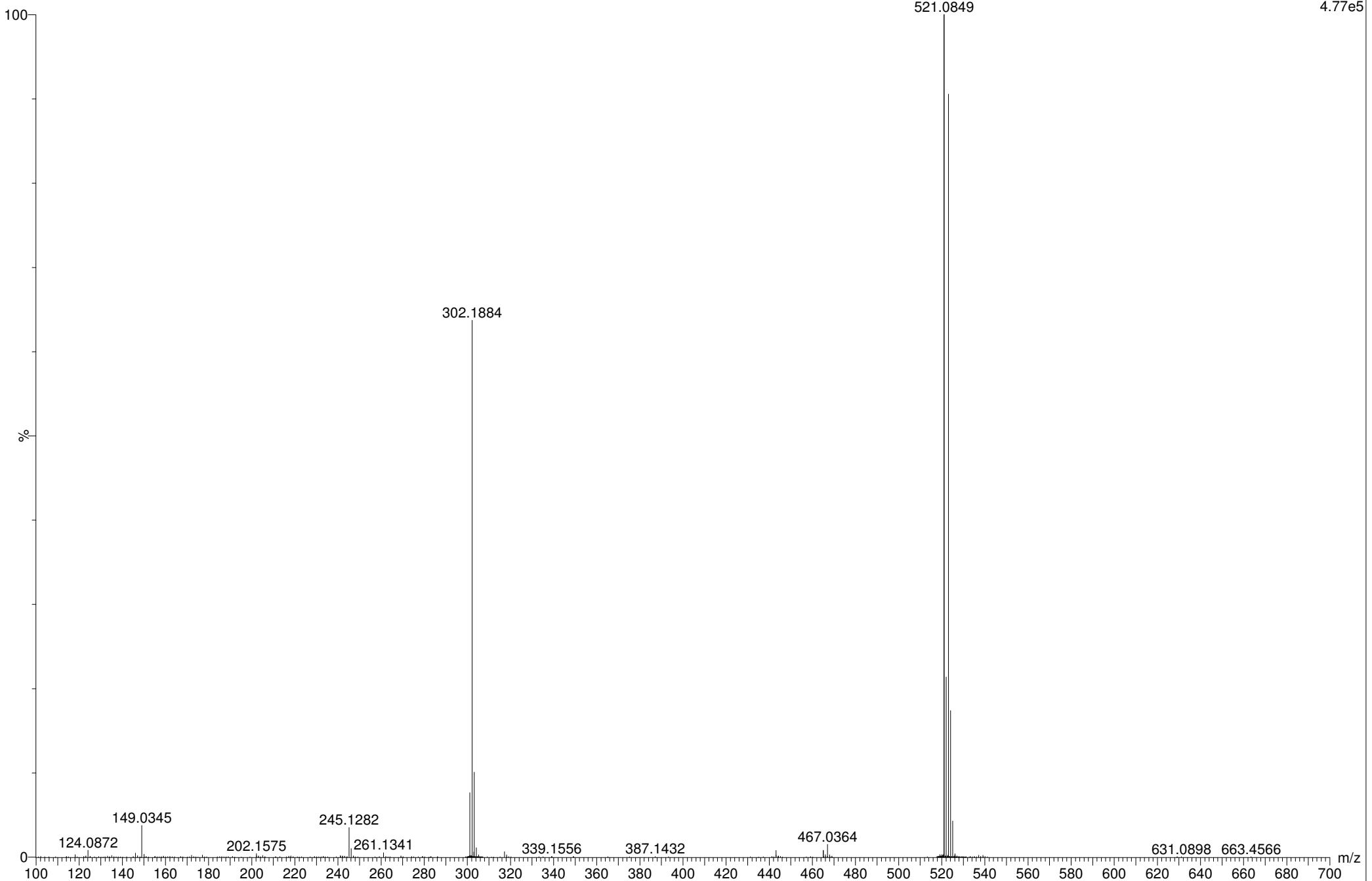
PROCESSED BY
PAWAN KUMAR

RMD-101_02 20 (0.371) AM (Cen,3, 80.00, Ar,5000.0,521.08,0.70); Sb (2,10.00); Cr (16:26)

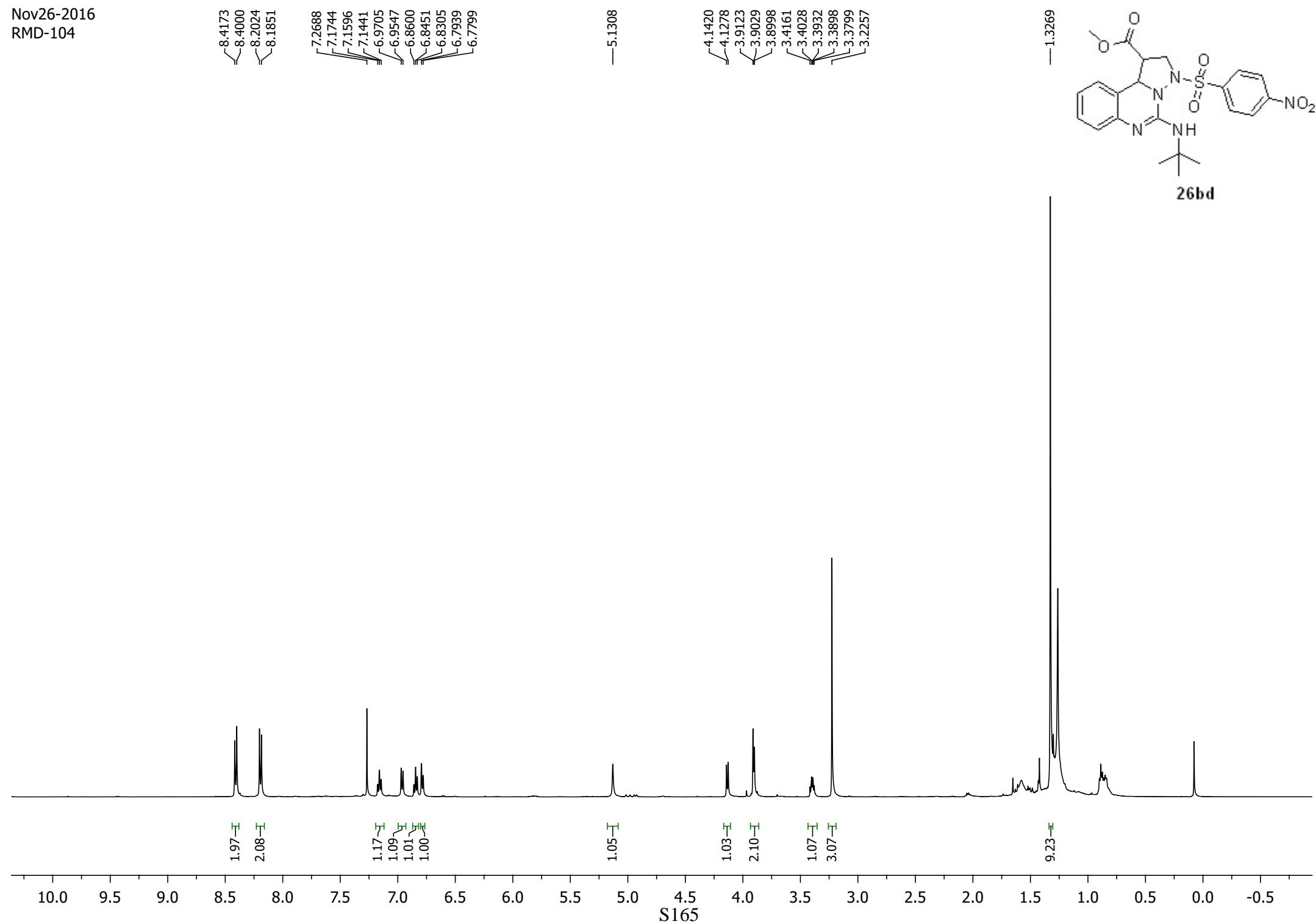
CSIR-IHBT
NPC&PD DIVISION

19-Jan-2017 13:57:16

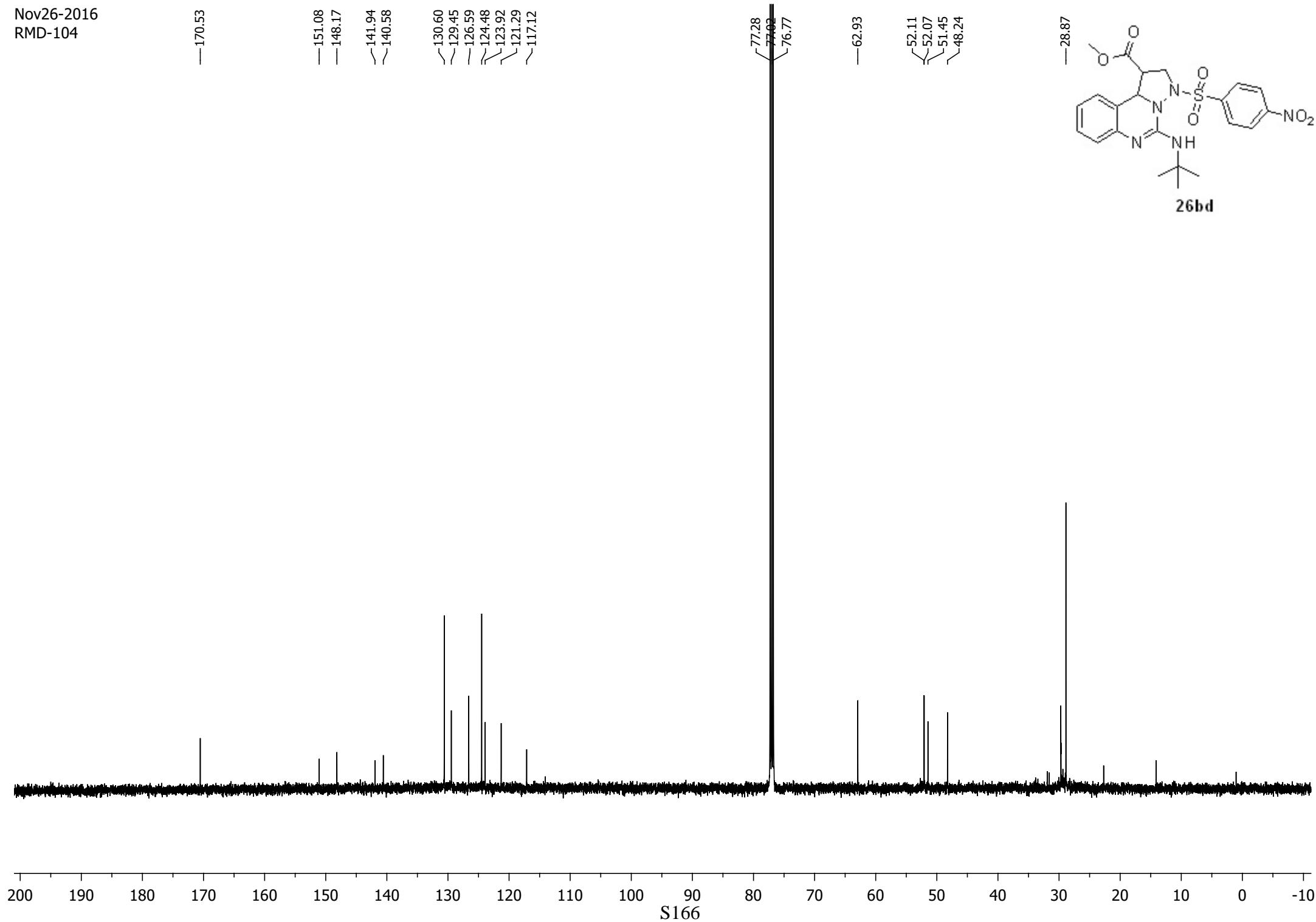
1: TOF MS ES+
4.77e5



Nov26-2016
RMD-104



Nov26-2016
RMD-104



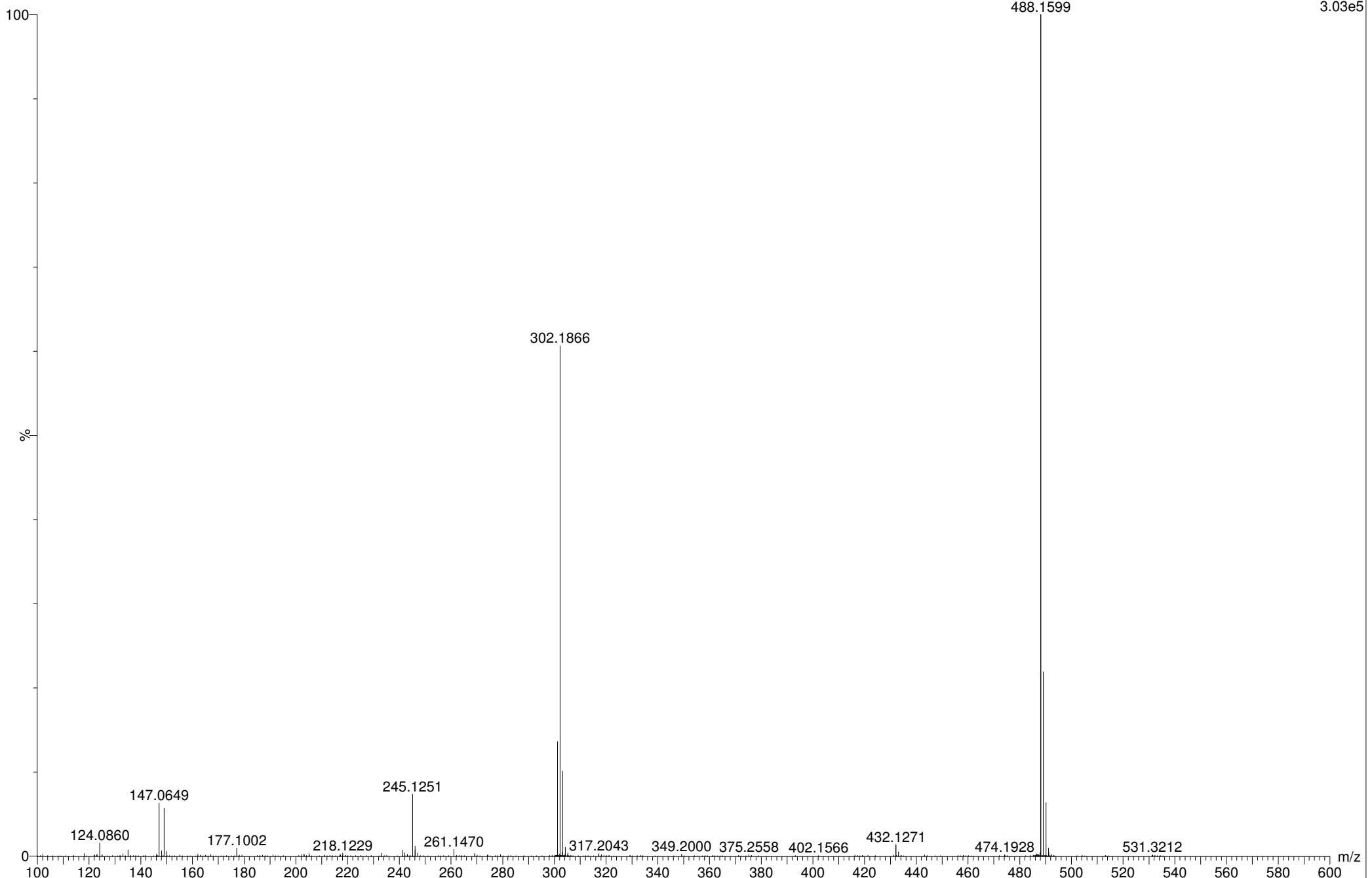
PROCESSED BY
PAWAN KUMAR

RMD-104_02 20 (0.371) AM (Cen,3, 80.00, Ar,5000.0,488.16,0.70); Sb (2,10.00); Crm (18:25)

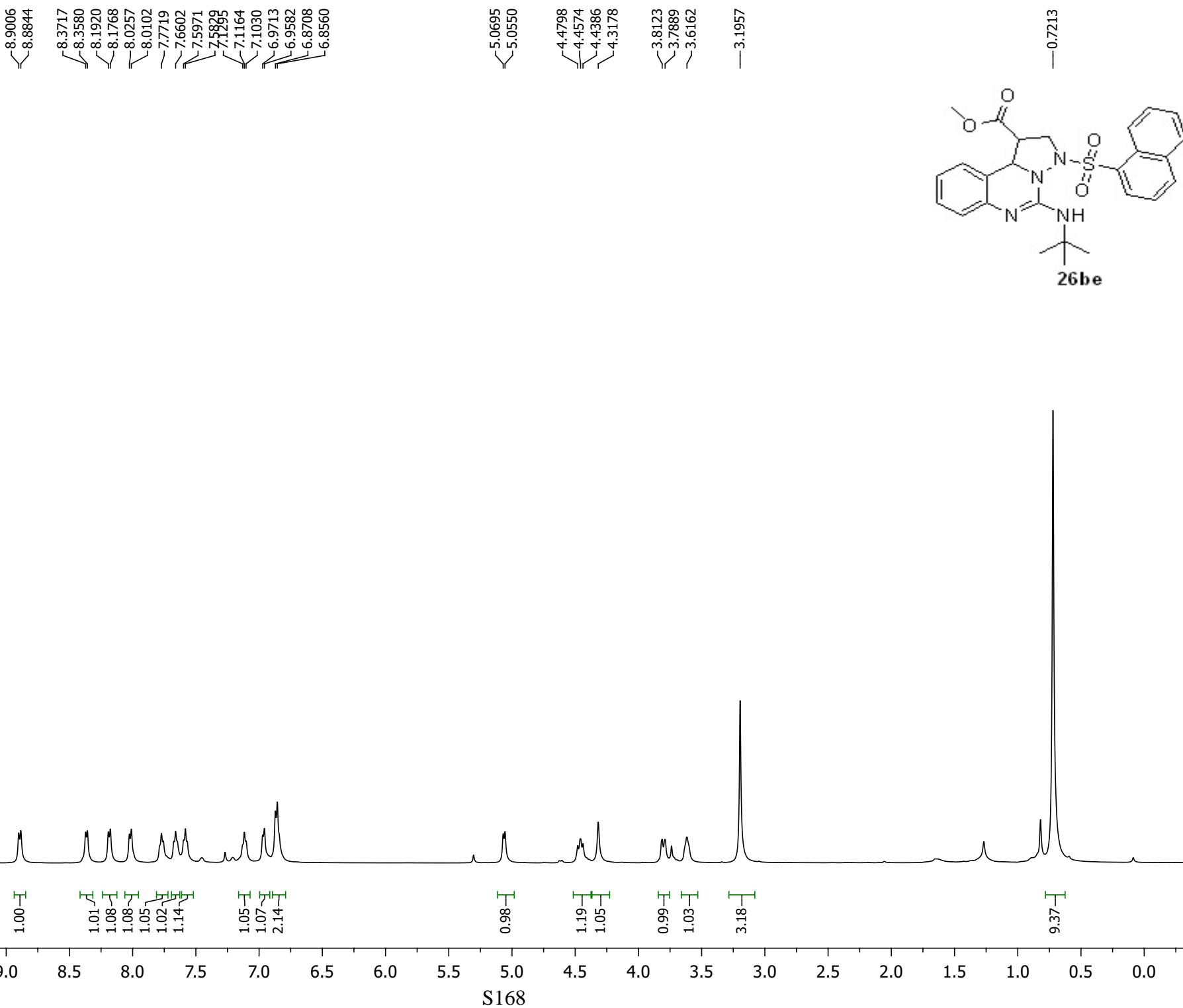
CSIR-IHBT
NPC&PD DIVISION

19-Jan-2017 13:06:16

1: TOF MS ES+
3.03e5



Nov15-2016
RMD-085



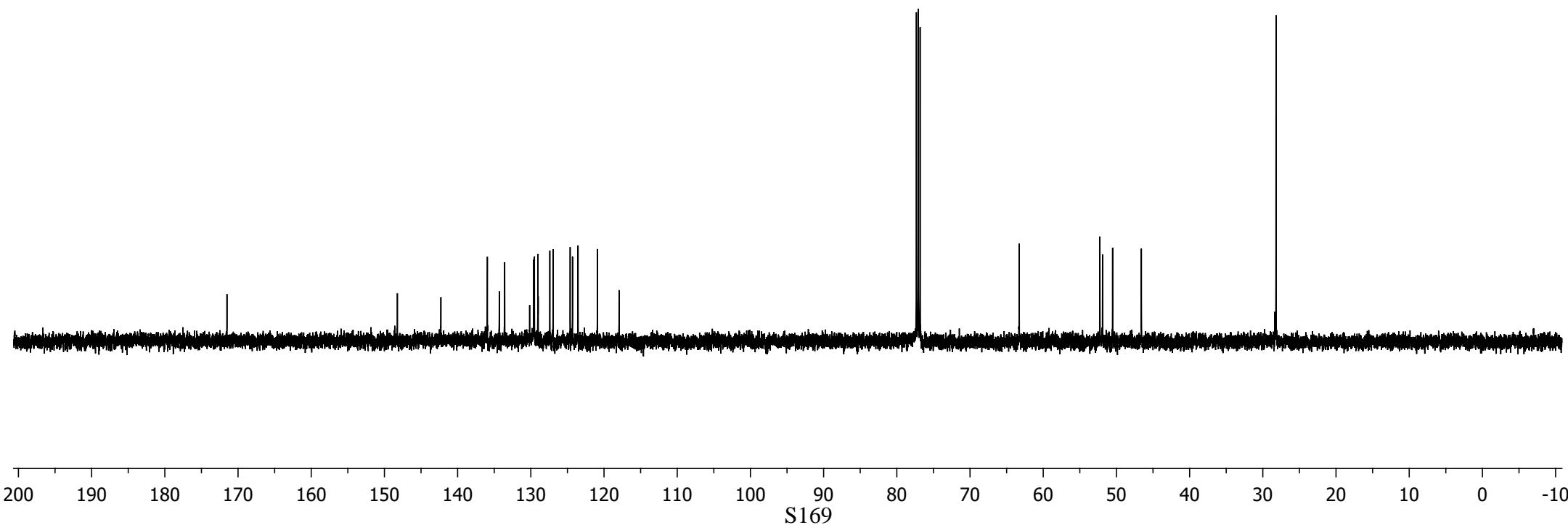
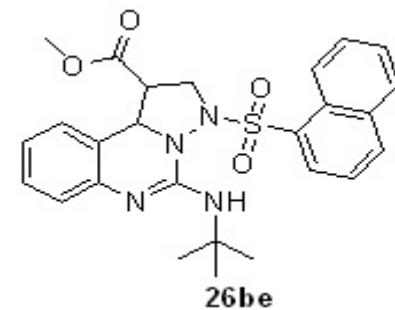
—171.51

—148.23
—142.28
—135.94
—134.31
—133.57
—130.16
—129.59
—129.51
—129.03
—128.99
—128.33
—127.40
—126.95
—124.63
—124.31
—123.55
—120.90
—117.94

—77.32
—77.07
—76.82

—63.28
—52.26
—51.89
—50.52
—46.59

—28.19



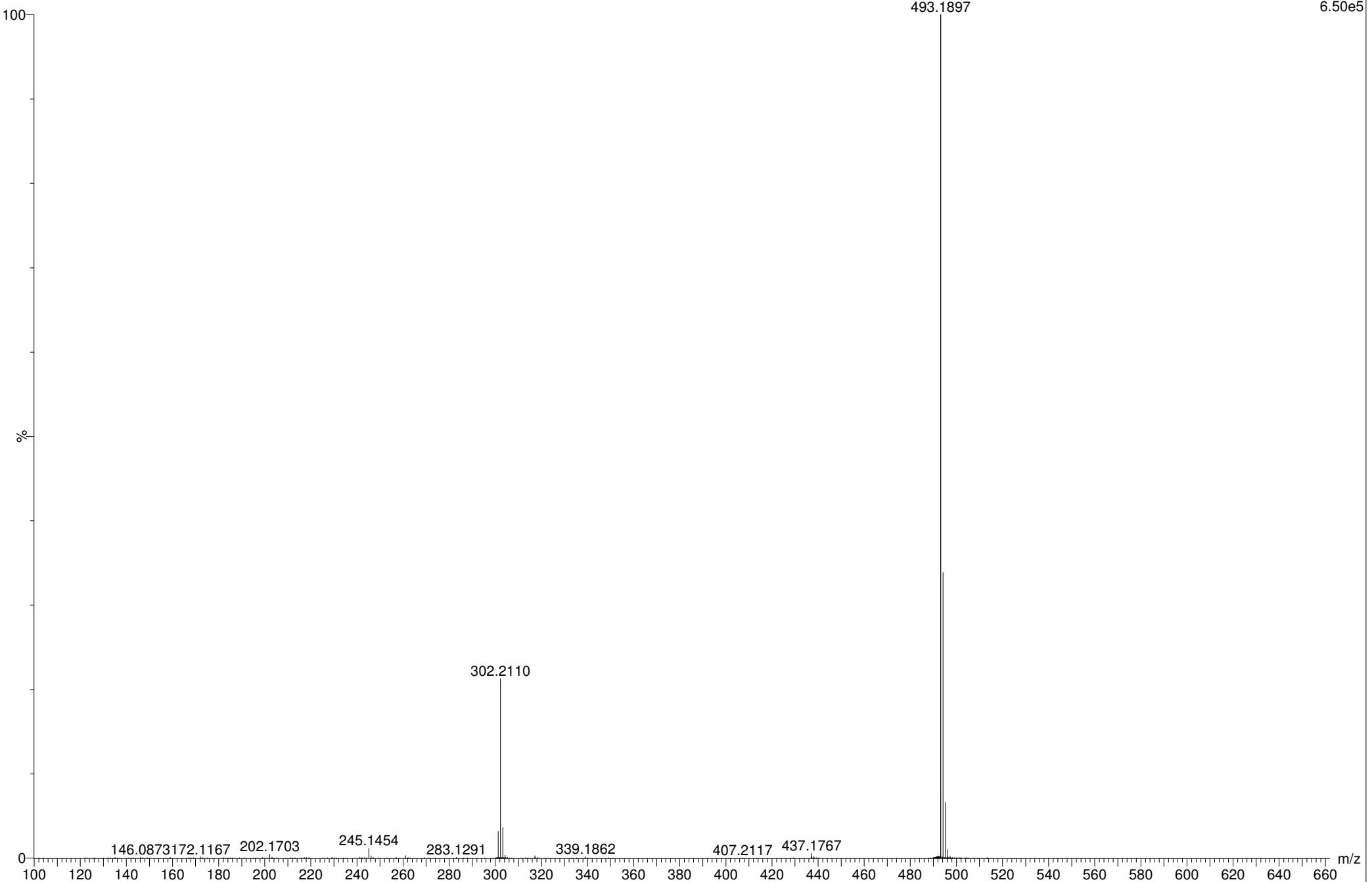
PROCESSED BY
PAWAN KUMAR

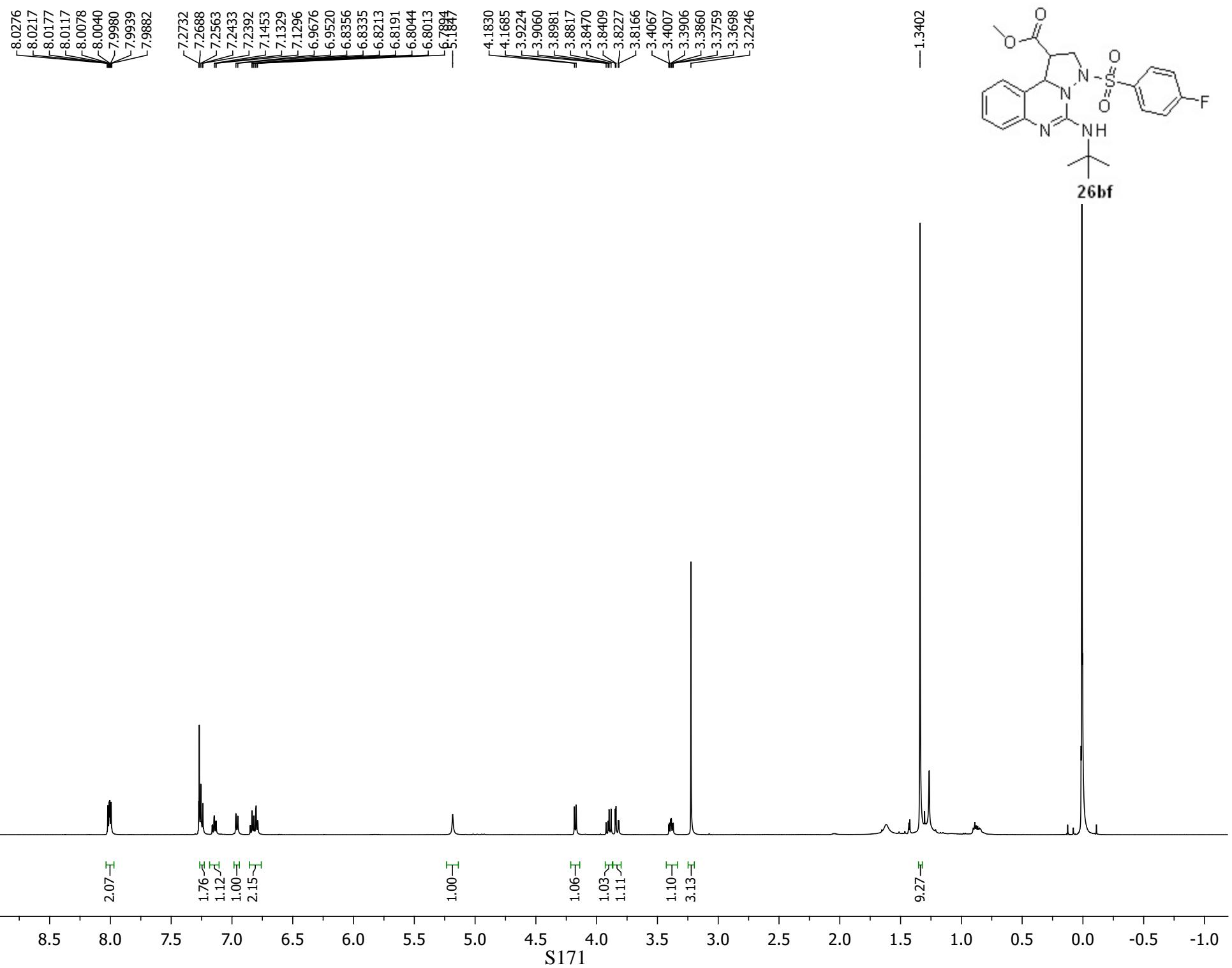
RMD-085 21 (0.390) AM (Cen,3, 80.00, Ar,5000.0,493.19,0.70); Sb (2,10.00); Cm (17:26)

CSIR-IHBT
NPC&PD DIVISION

24-Nov-2016 16:39:09

1: TOF MS ES+
6.50e5





Jan10-2017
RMD-A121-046

—170.82
—167.28
—165.23

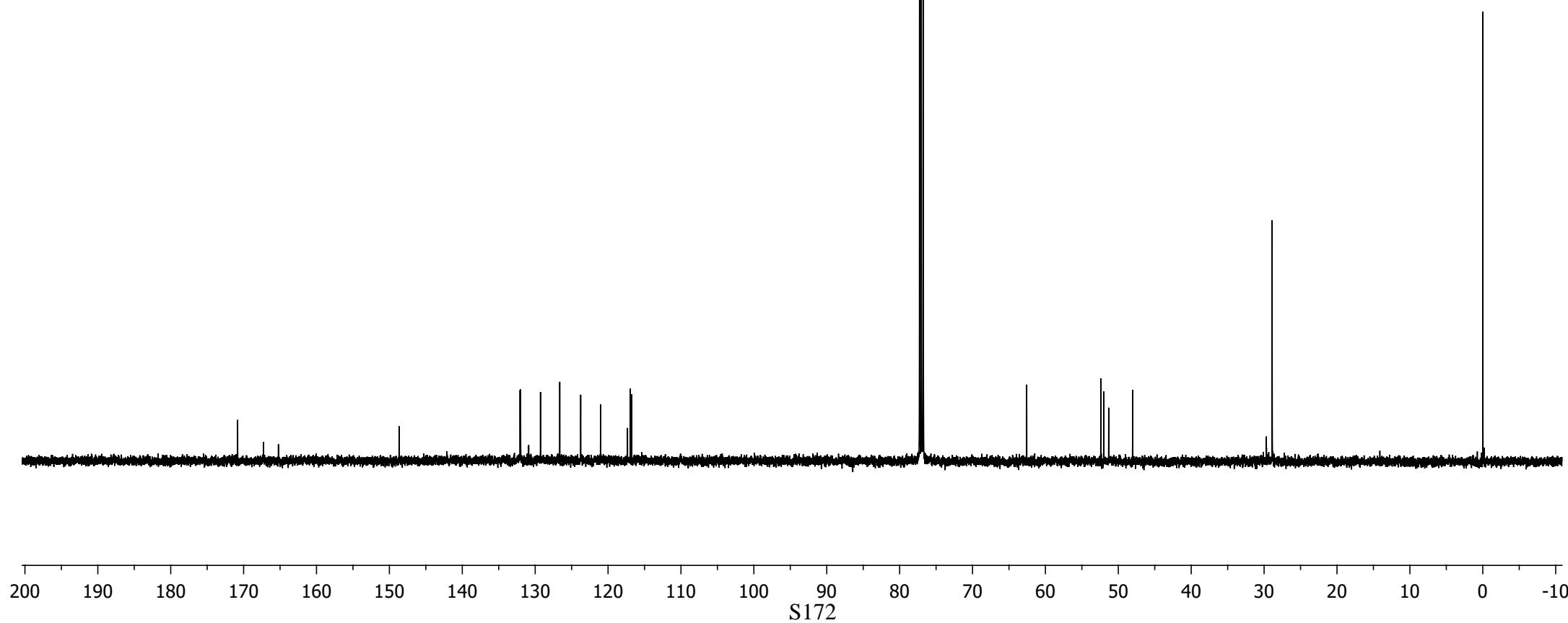
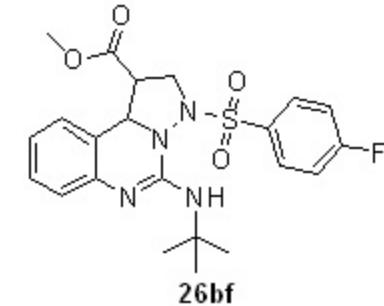
—148.67

—132.08
—132.00
—130.89
—129.27
—126.63
—123.75
—121.03
—117.33
—116.94
—116.76

—77.27
—77.02
—76.76

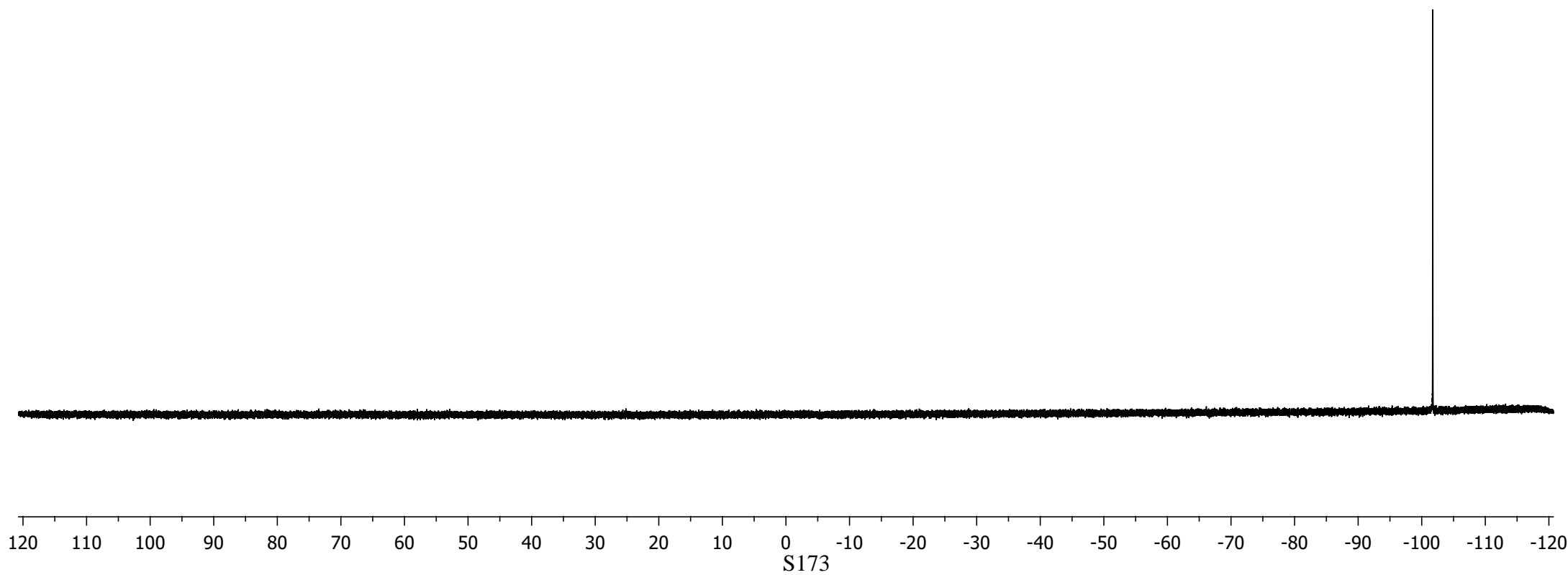
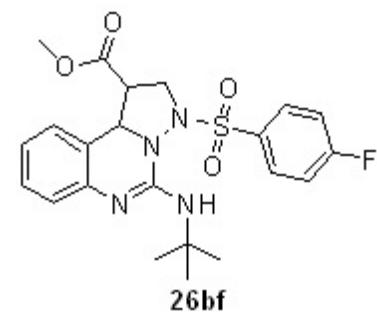
—62.57
—52.40
—52.00
—51.32
—48.04

—28.89



Jan10-2017
RMD-A121-046

-101.7311



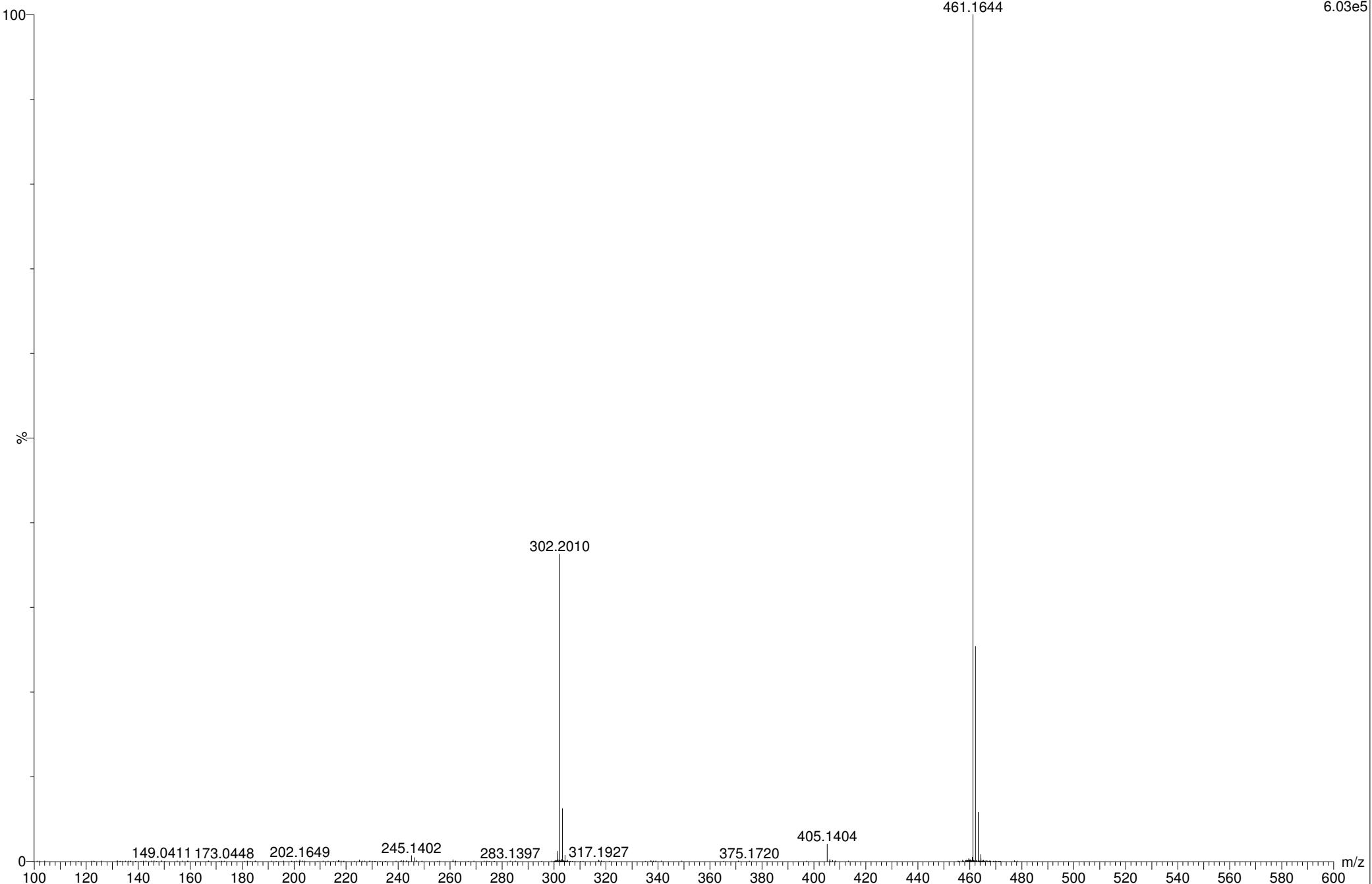
PROCESSED BY
PAWAN KUMAR

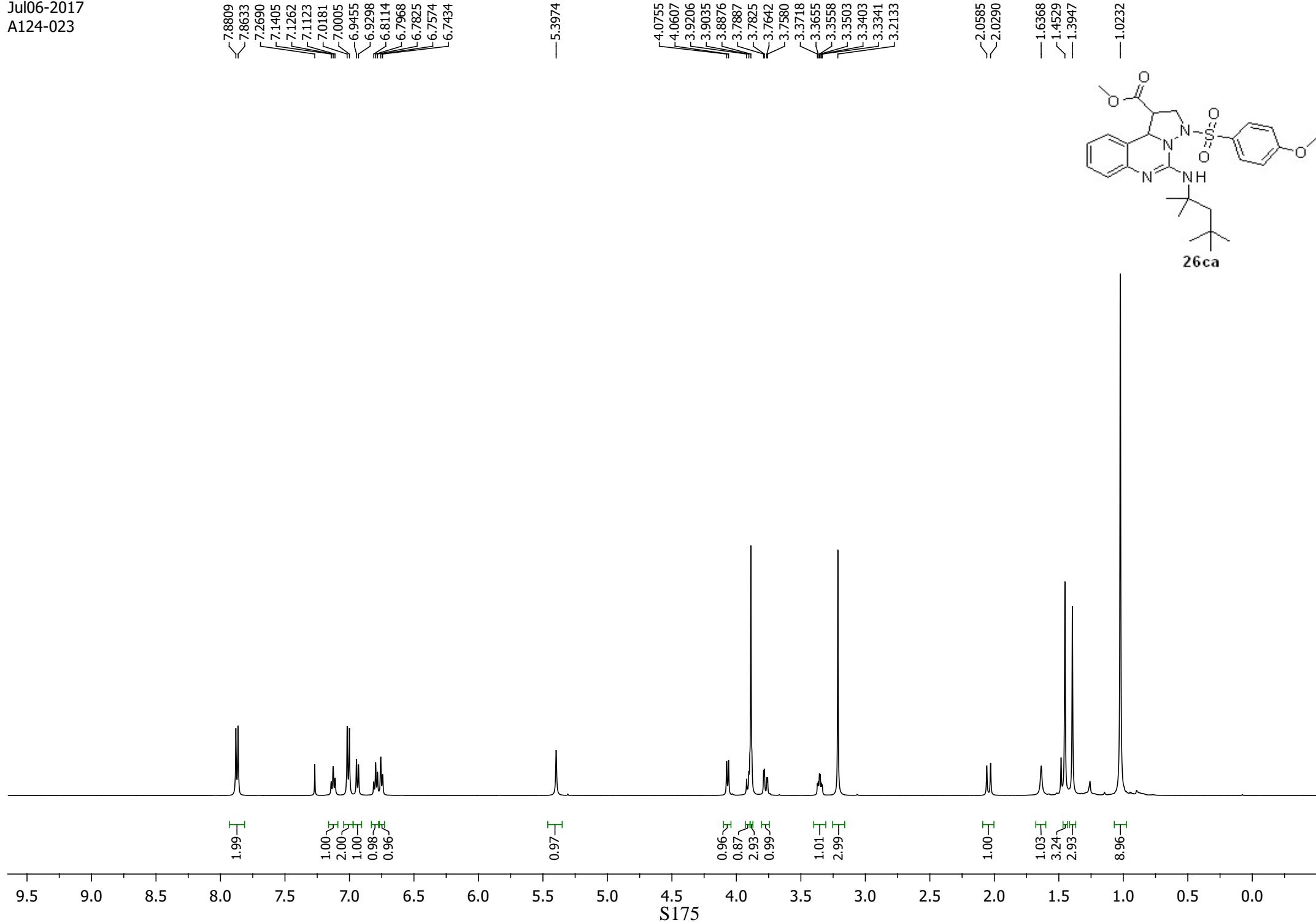
RMD-069 20 (0.371) AM (Cen,3, 80.00, Ar,5000.0,461.16,0.70); Sb (2,10.00); Cm (18:28)

CSIR-IHBT
NPC&PD DIVISION

24-Nov-2016 16:26:52

1: TOF MS ES+
6.03e5





Jul06-2017
A124-023

— 171.0031

— 164.2986

— 148.6239

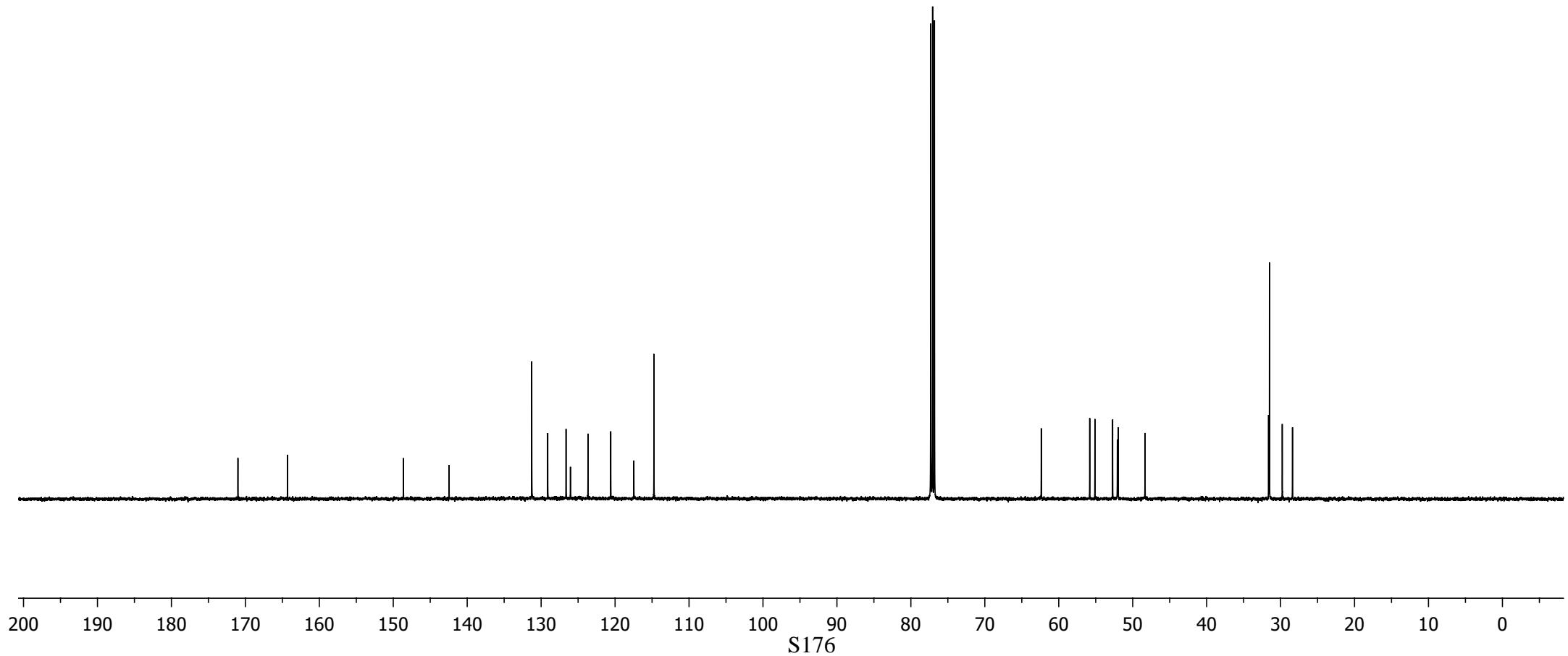
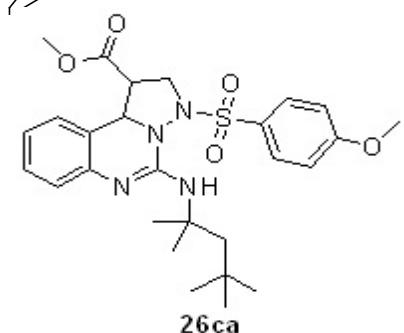
— 142.4466

— 131.2683
— 129.1093
— 126.6309
— 126.0403
— 123.6585
— 120.5853
— 117.4569
— 114.7464

— 77.3170
— 77.0630
— 76.8090

— 62.3665
— 55.8123
— 55.1107
— 52.7128
— 52.0754
— 51.9659
— 48.3321

— 31.6281
— 31.5060
— 29.7753
— 28.3921



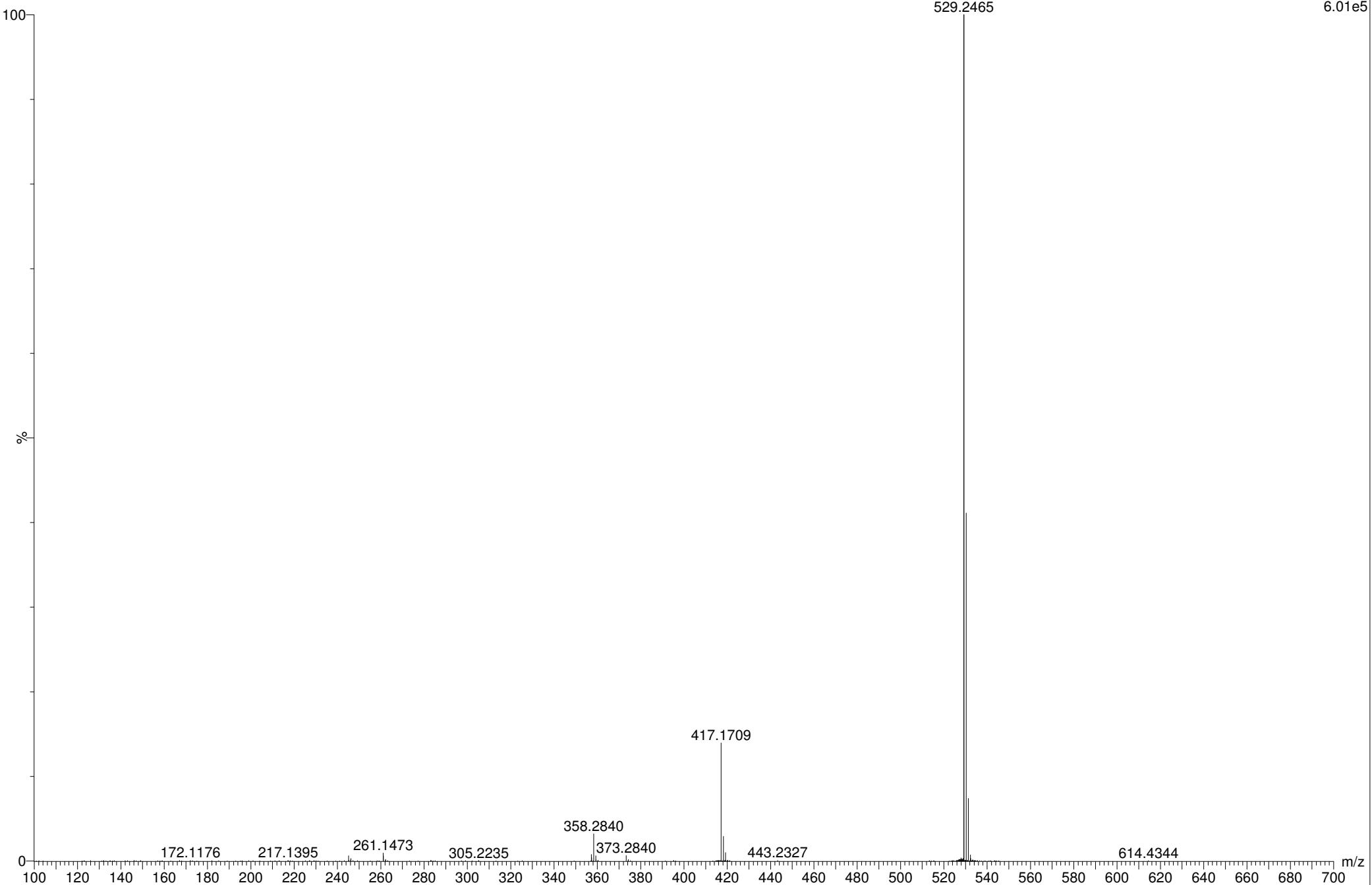
PROCESSED BY
PAWAN KUMAR

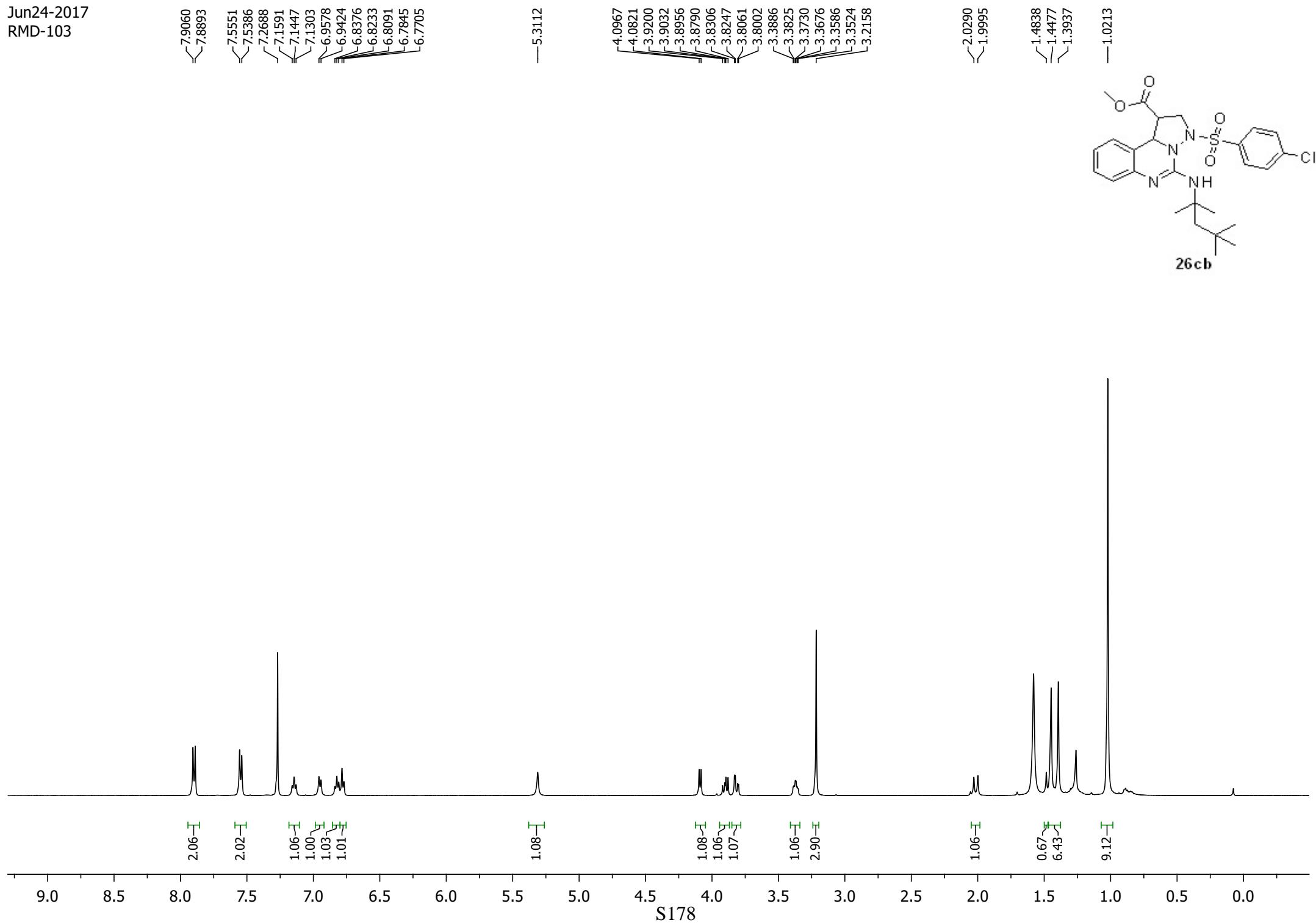
RMD-081 21 (0.389) AM (Cen,3, 80.00, Ar,5000.0,529.25,0.70); Sb (2,10.00); Cm (19:27)

CSIR-IHBT
NPC&PD DIVISION

24-Nov-2016 16:51:22

1: TOF MS ES+
6.01e5





—170.73

—148.20

—142.27

—141.31

—133.32

—130.50

—129.88

—129.23

—126.62

—123.76

—120.82

—117.32

—77.31

—77.05

—76.80

—62.66

—55.25

—52.40

—52.18

—51.99

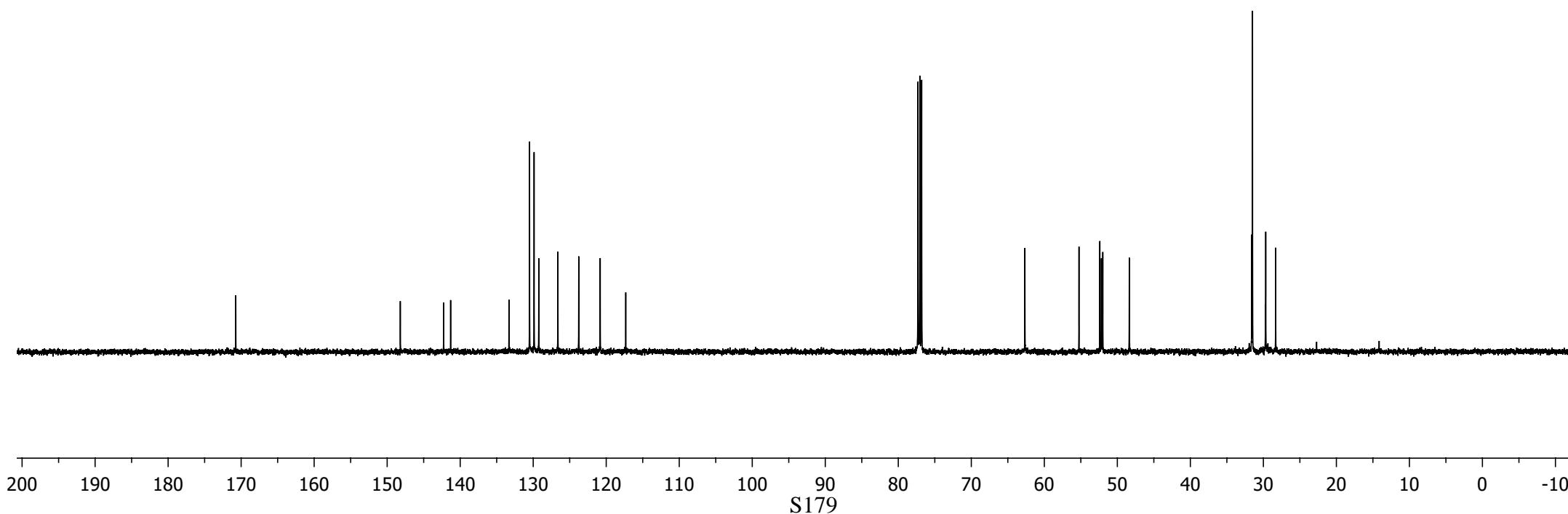
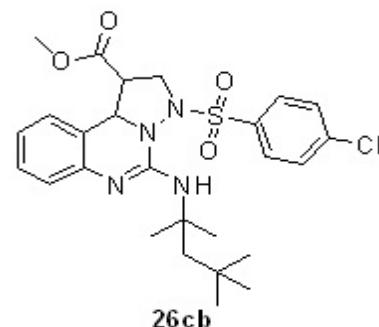
—48.36

—31.60

—31.48

—29.67

—28.31



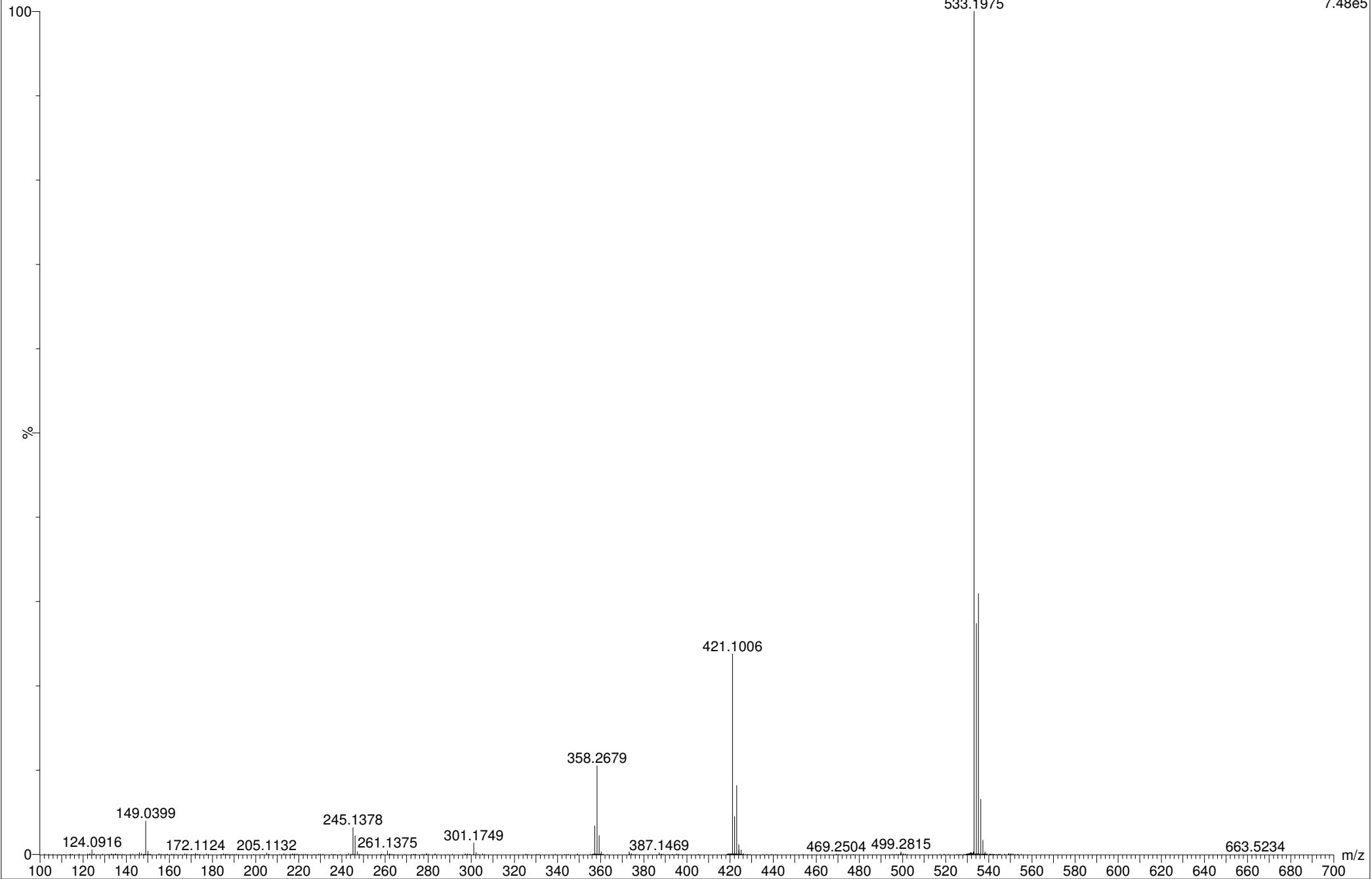
PROCESSED BY
PAWAN KUMAR

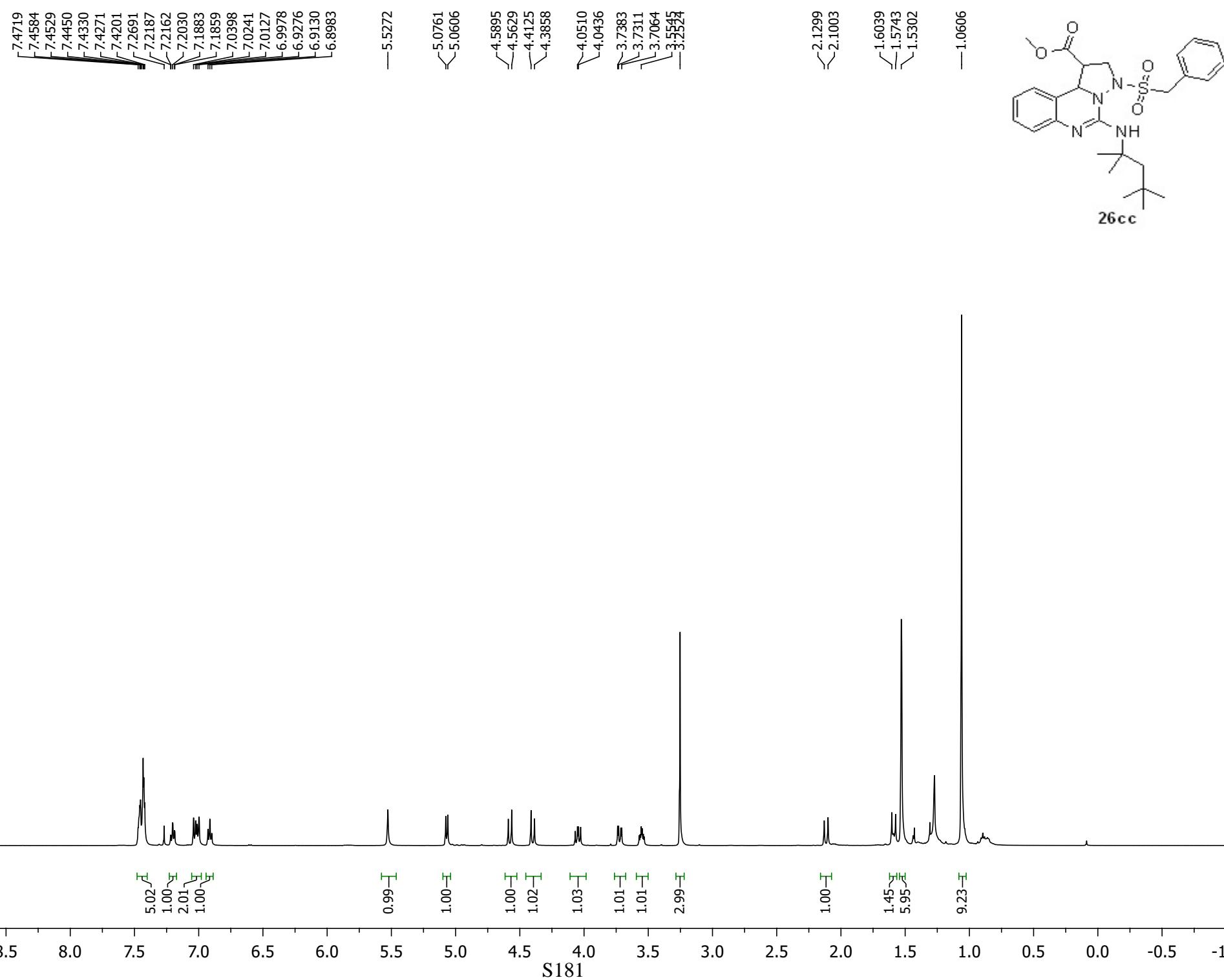
CSIR-IHBT
NPC&PD DIVISION

19-Jan-2017 13:14:09

RMD-103_02 20 (0.371) AM (Cen,3, 80.00, Ar,5000.0,533.20,0.70); Sb (2,10.00); Cr (17:28)

1: TOF MS ES+
7.48e5





Nov22-2016
RMD-094

—171.11

—148.23

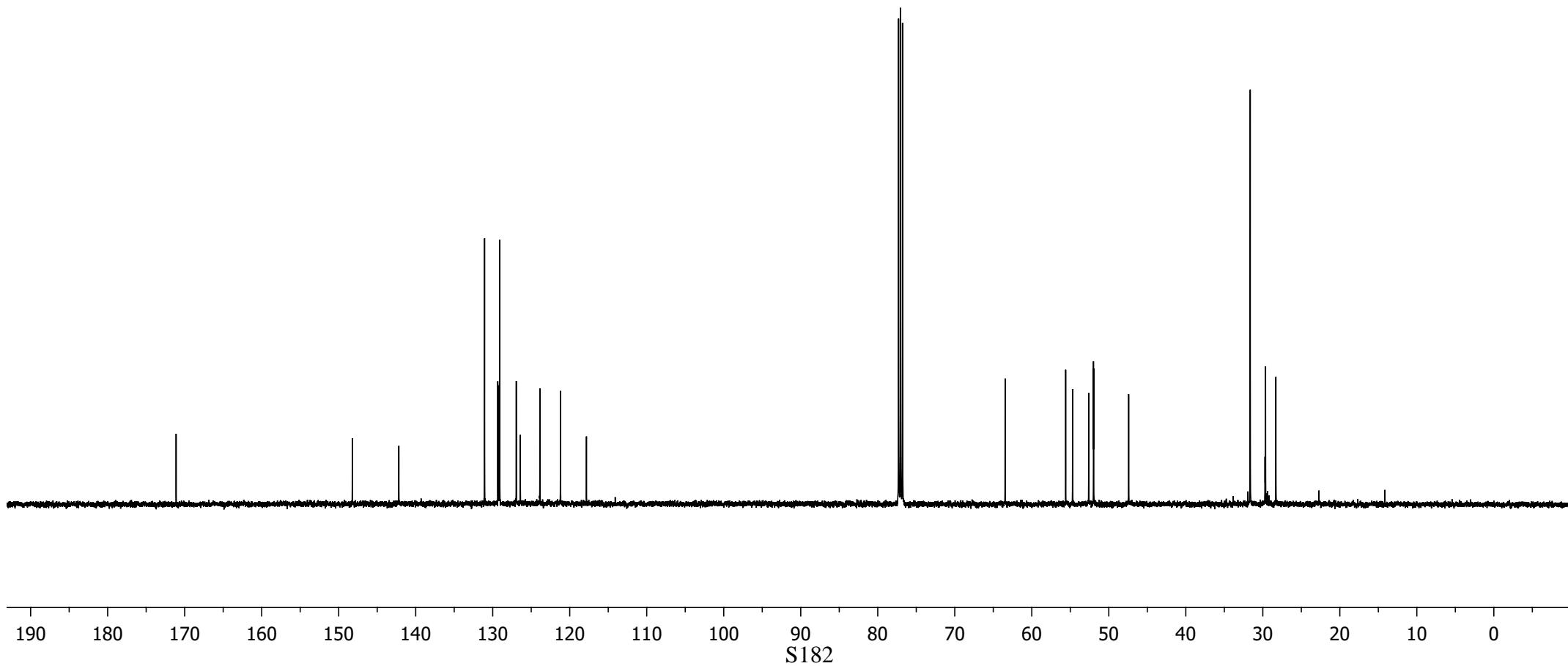
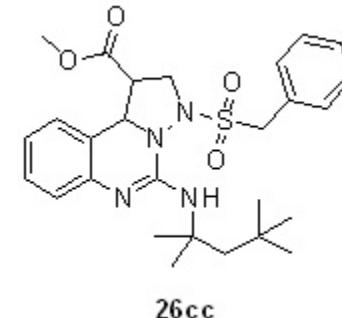
—142.19

131.06
129.34
129.27
129.10
126.94
126.41
123.87
121.20
117.83

77.30
77.04
76.79

—63.46
55.60
54.70
52.60
51.99
51.96
47.42

—31.67
31.63
29.64
28.30



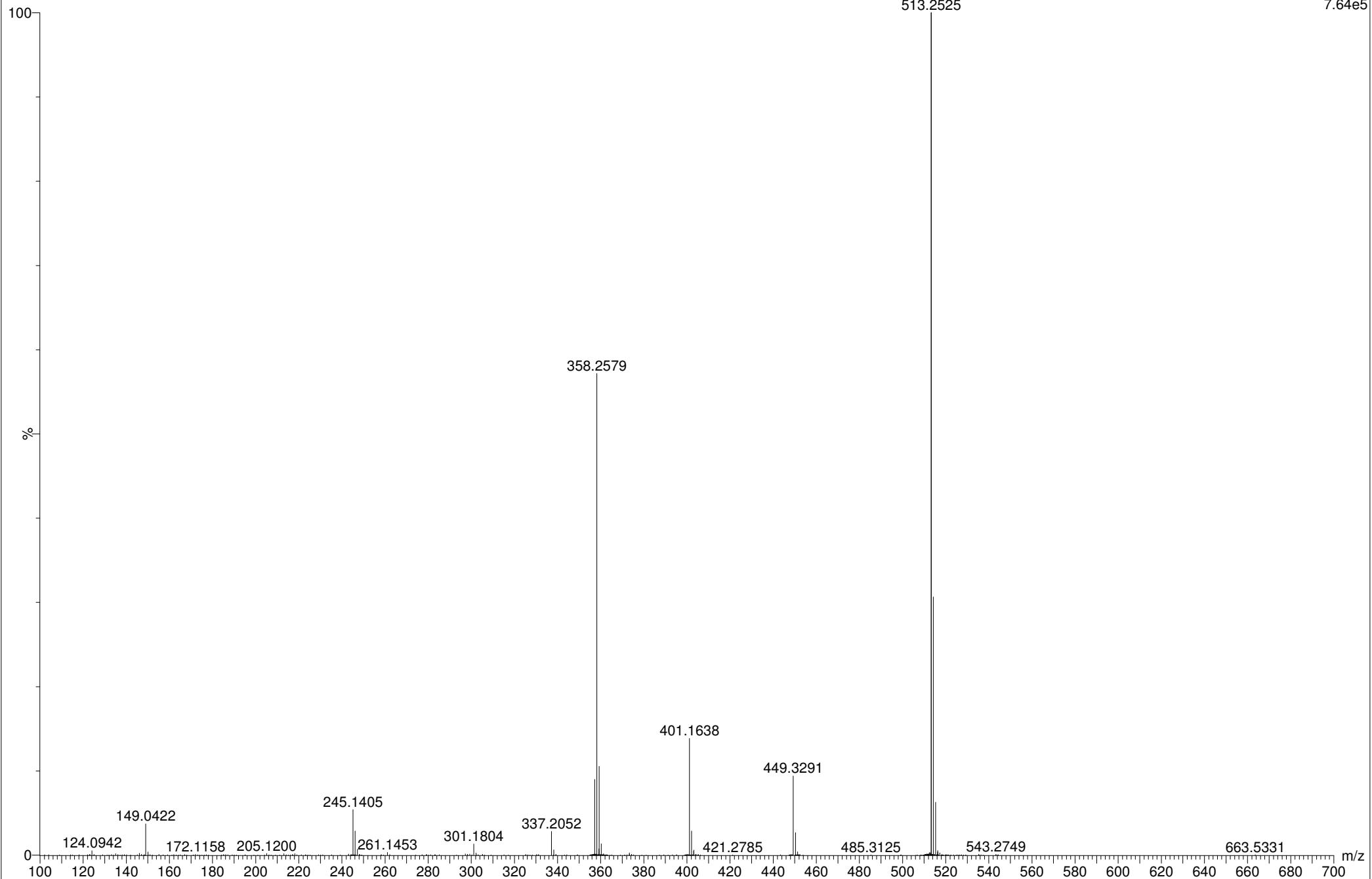
PROCESSED BY
PAWAN KUMAR

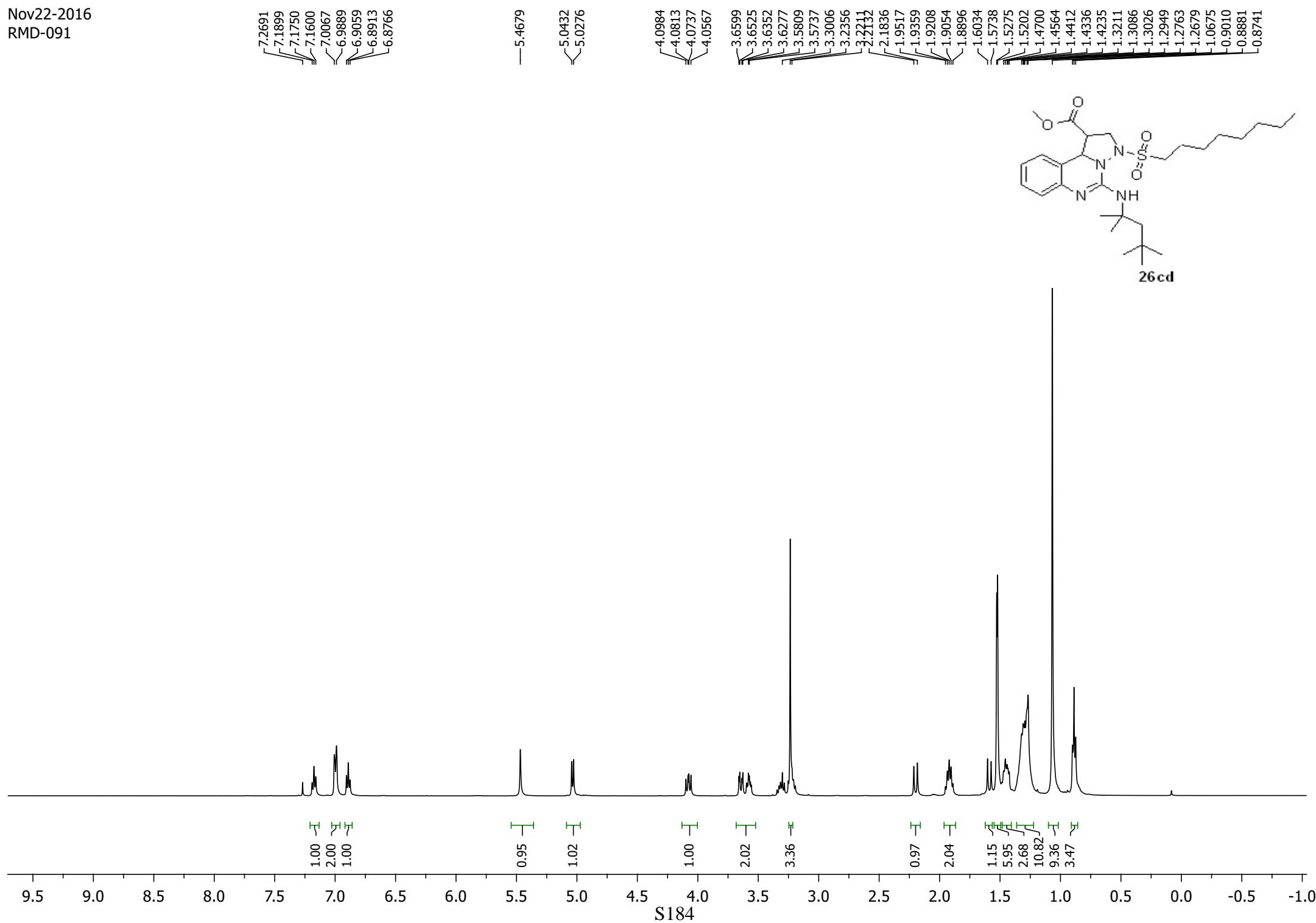
CSIR-IHBT
NPC&PD DIVISION

19-Jan-2017 15:29:19

RMD-094_02 20 (0.371) AM (Cen,3, 80.00, Ar,5000.0,513.25,0.70); Sb (2,10.00); Cr (17:28)

1: TOF MS ES+
7.64e5





Nov22-2016
RMD-091

—171.28

—148.19

—142.13

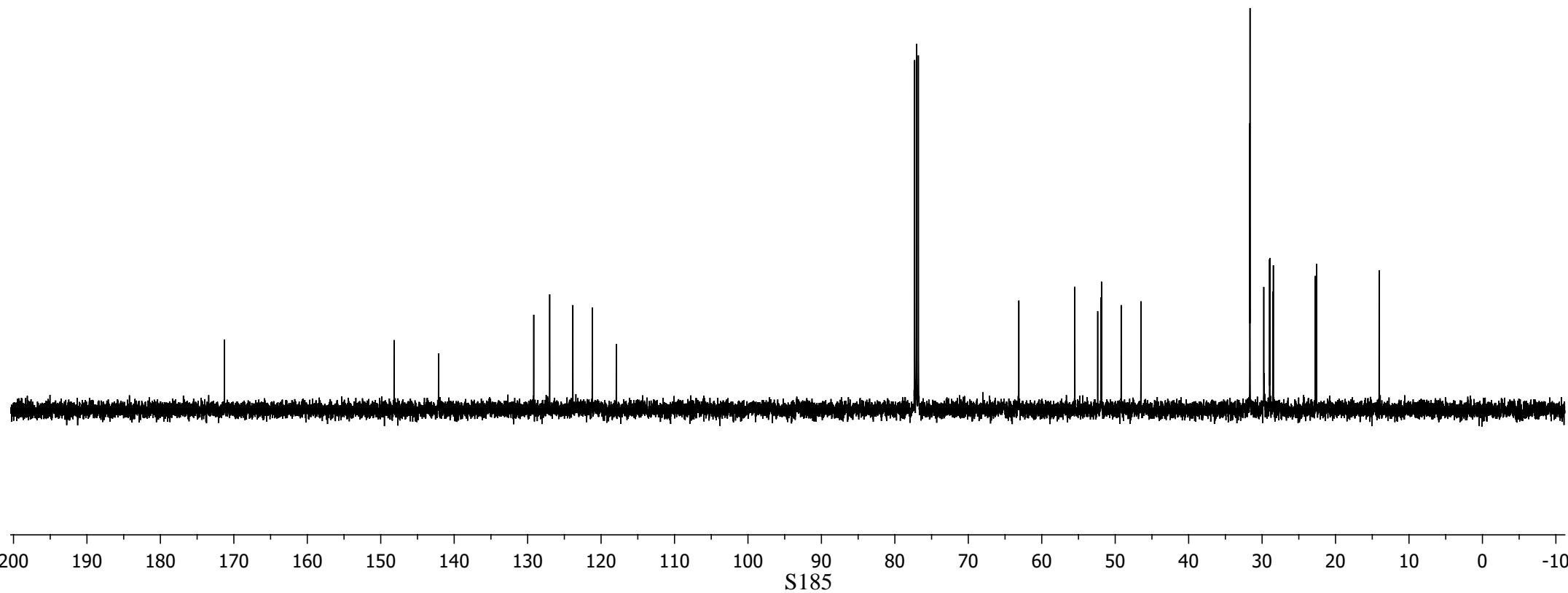
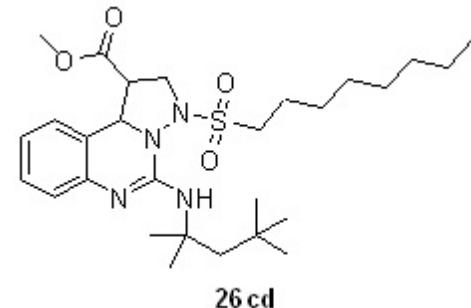
—129.17
—127.00
—123.84
—121.20
—117.93

77.31
77.05
76.80

—63.12

55.50
52.38
51.94
51.86
49.19
46.49
31.68
31.64
29.77
28.98
28.94
28.53
28.47
22.75
22.58

—14.07



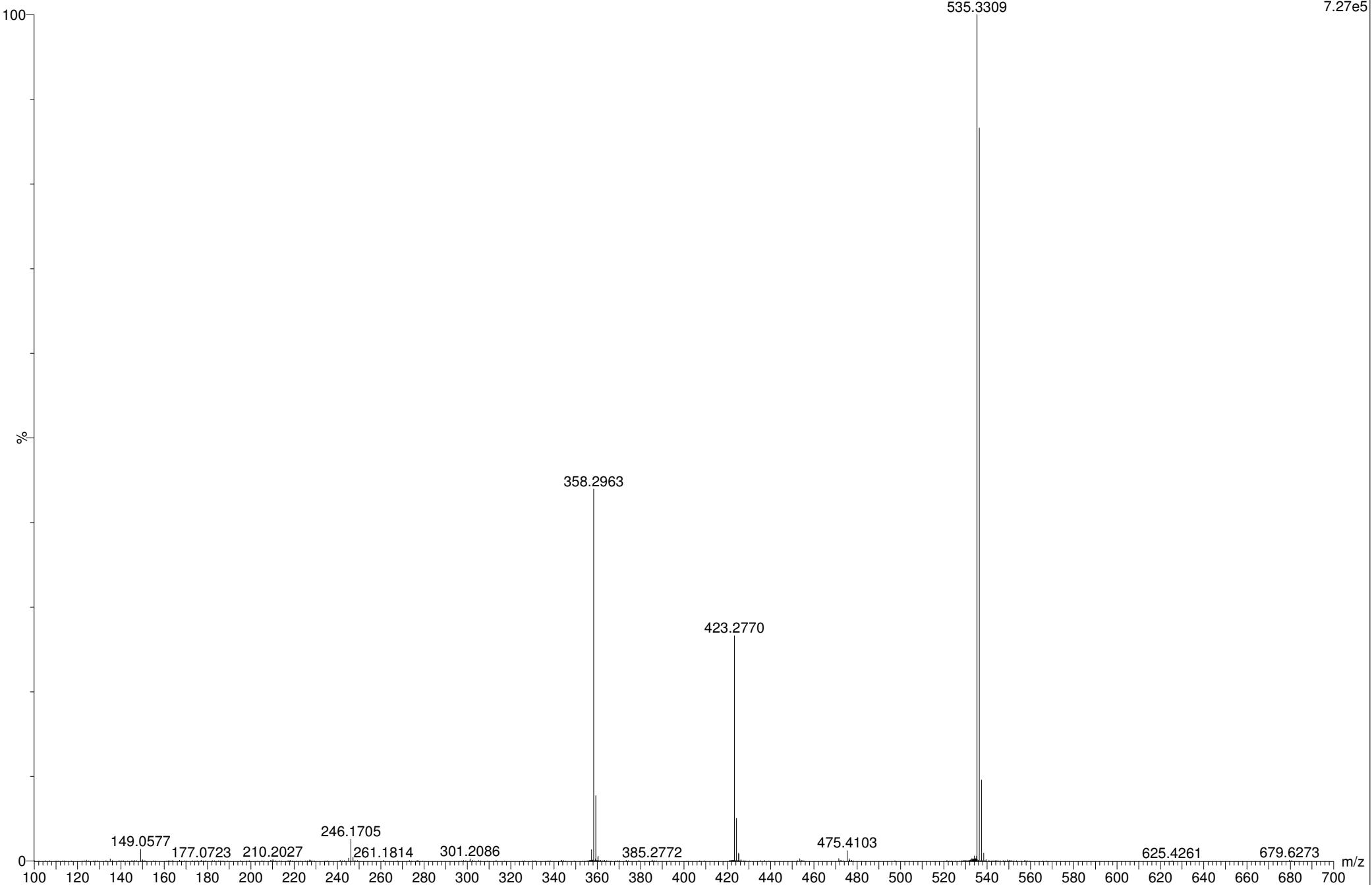
PROCESSED BY
PAWAN KUMAR

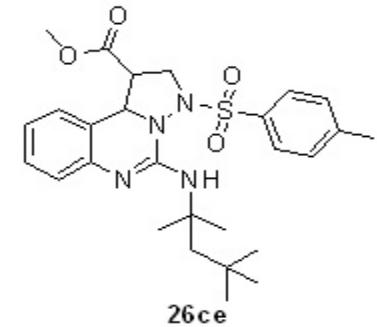
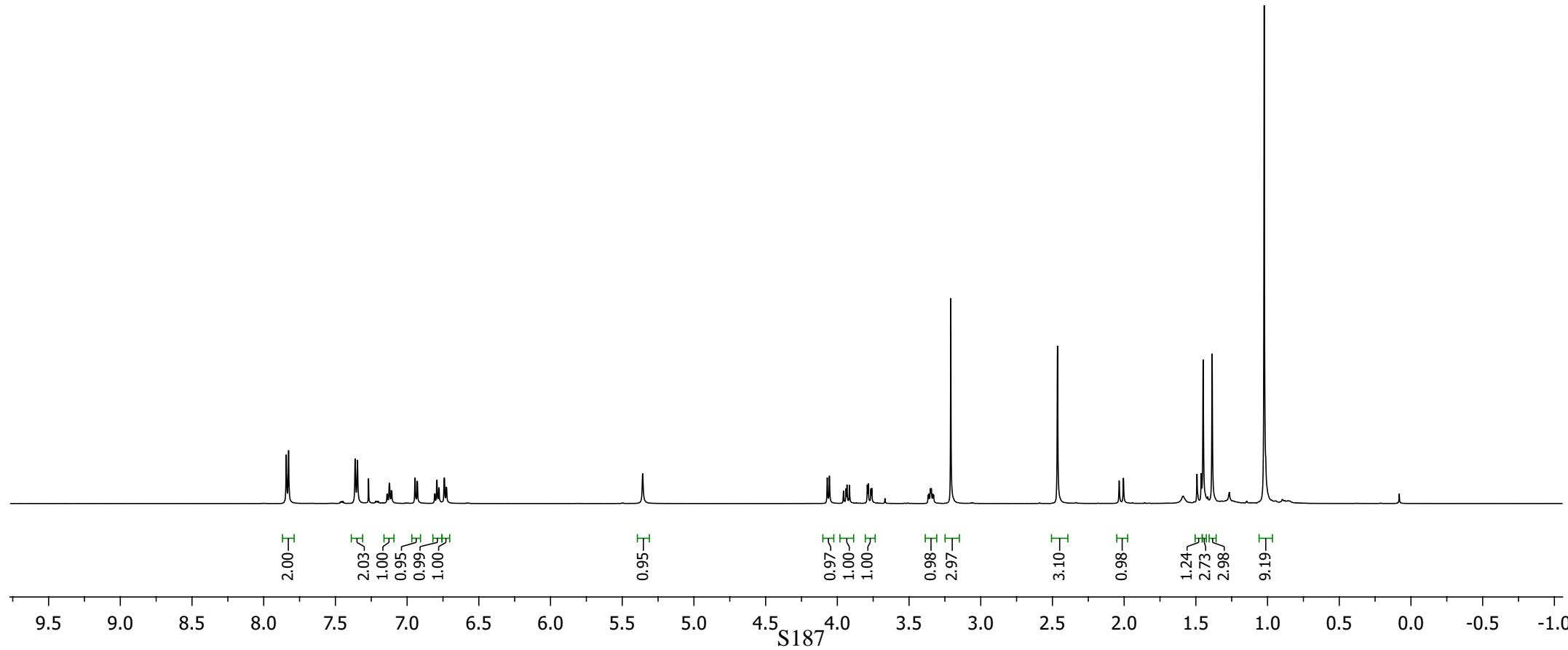
RMD-091_02 23 (0.426) AM (Cen,3, 80.00, Ar,5000.0,535.33,0.70); Sb (2,10.00); Cr (18:27)

CSIR-IHBT
NPC&PD DIVISION

19-Jan-2017 15:17:03

1: TOF MS ES+
7.27e5





-170.93

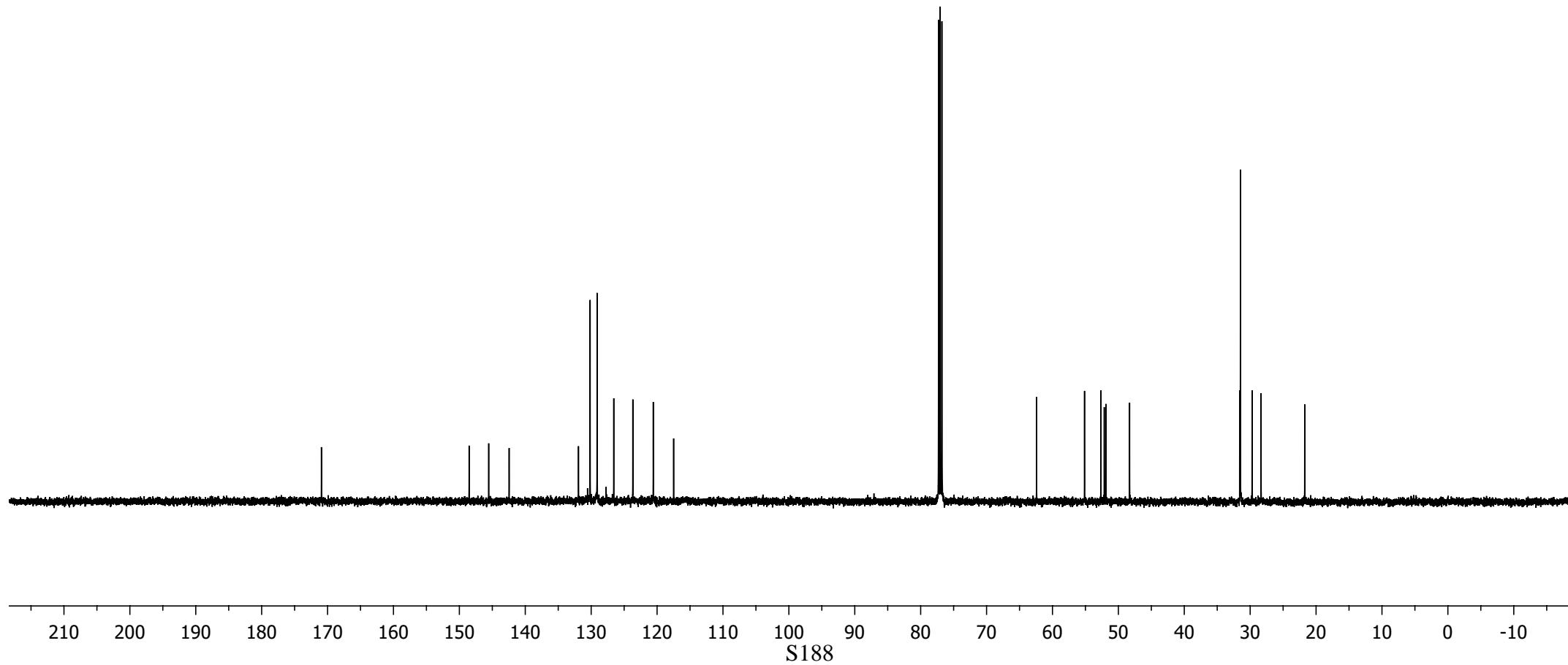
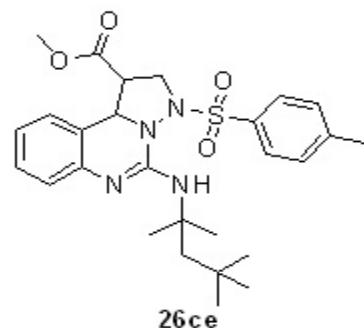
-148.52
-145.55
-142.45

131.92
130.16
129.10
129.08
126.57
123.68
120.58
117.48

77.28
77.03
76.78

-62.41
55.14
52.65
52.14
51.90
48.29

31.59
31.48
29.71
28.35
-21.72



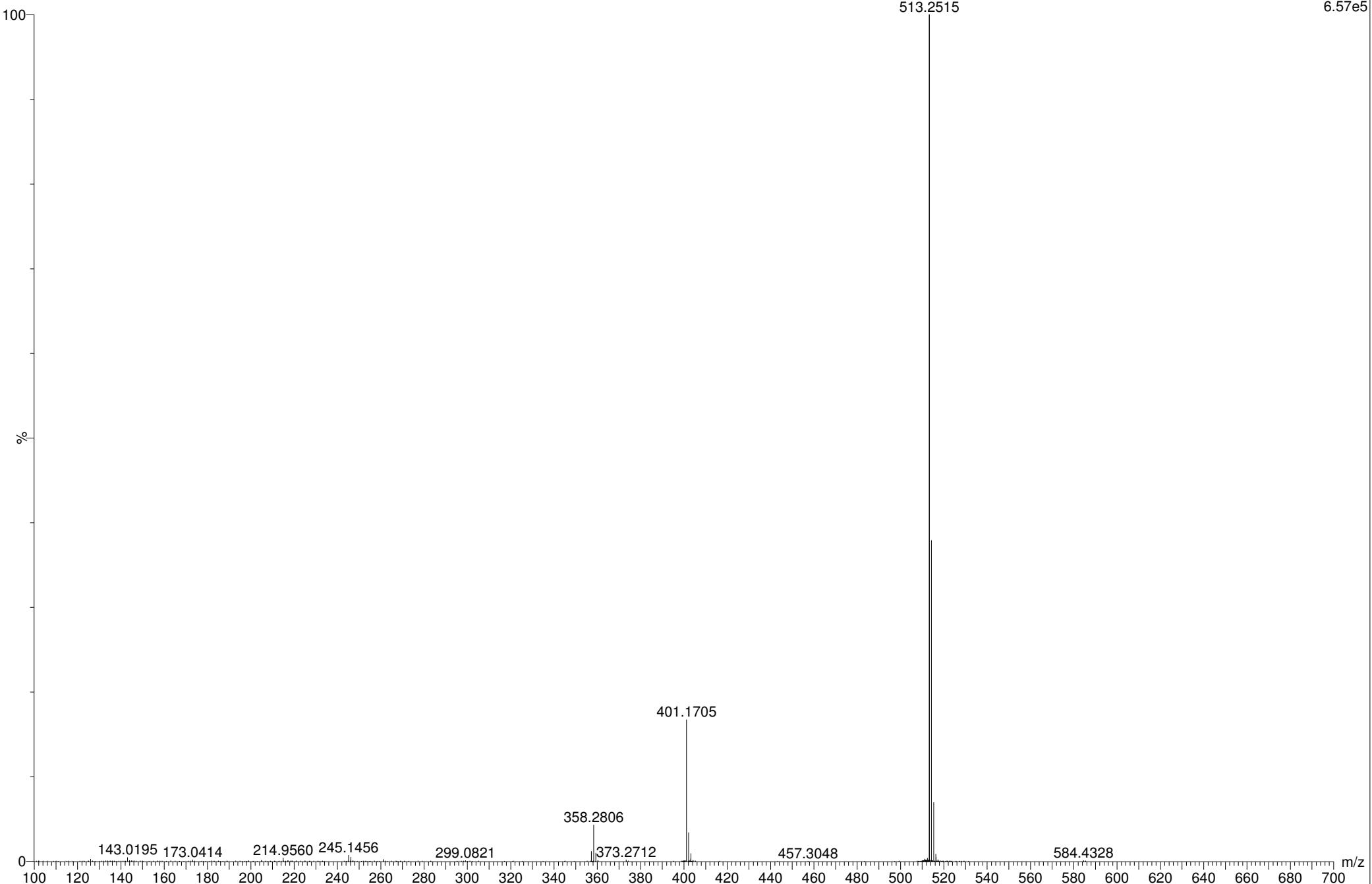
PROCESSED BY
PAWAN KUMAR

RMD-074 21 (0.390) AM (Cen,3, 80.00, Ar,5000.0,513.25,0.70); Sb (2,10.00); Cm (18:27)

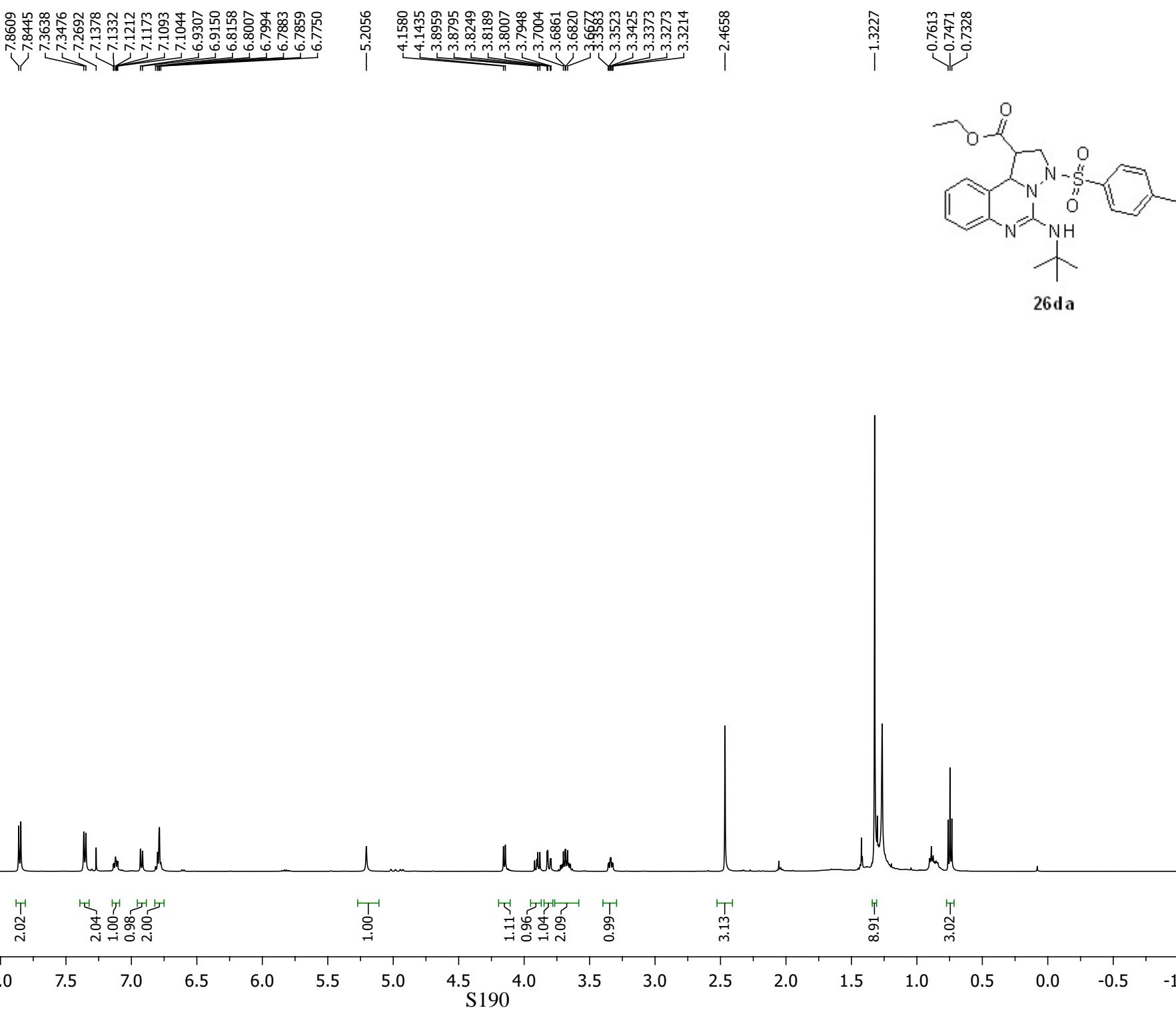
CSIR-IHBT
NPC&PD DIVISION

24-Nov-2016 17:25:13

1: TOF MS ES+
6.57e5



Nov25-2016
RMD-110



Nov25-2016
RMD-110

—170.68

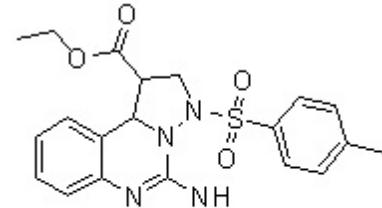
~148.93
—145.51
~142.66

131.94
130.09
129.17
129.04
126.73
123.72
120.66
117.73

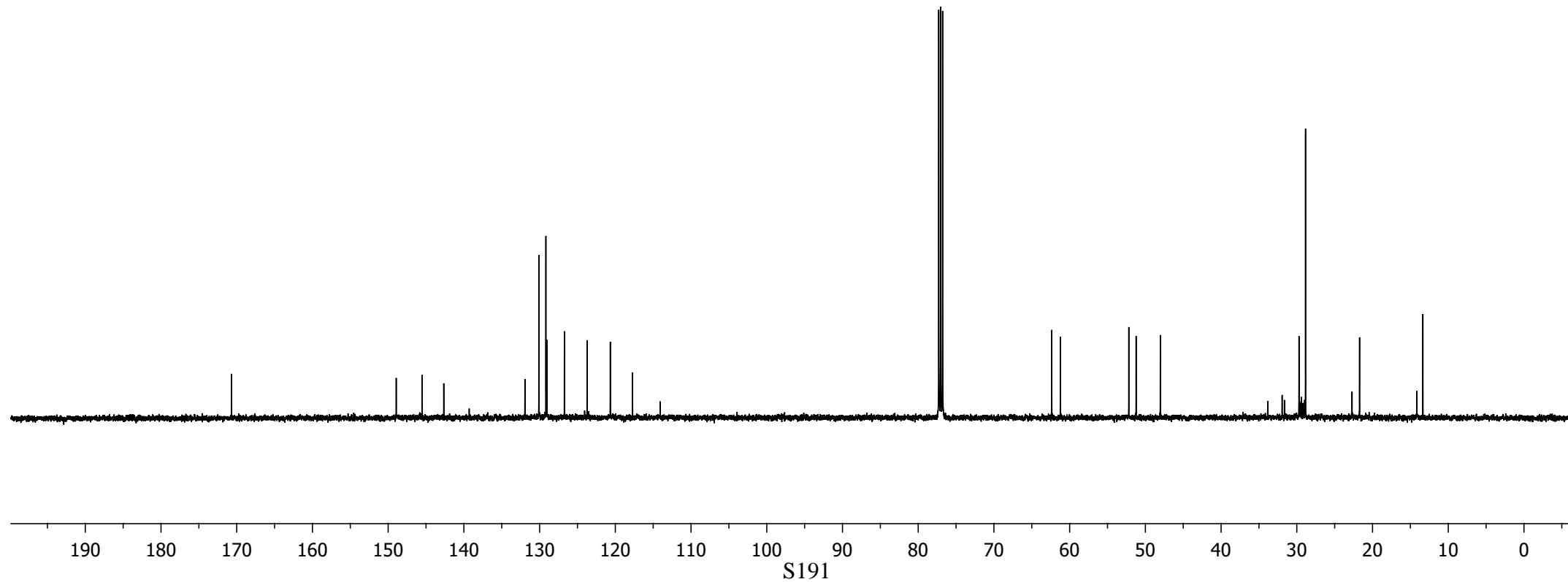
77.29
77.04
76.78

62.37
~61.22
52.19
~51.22
~48.02

—28.85
—21.71
—13.35



26da



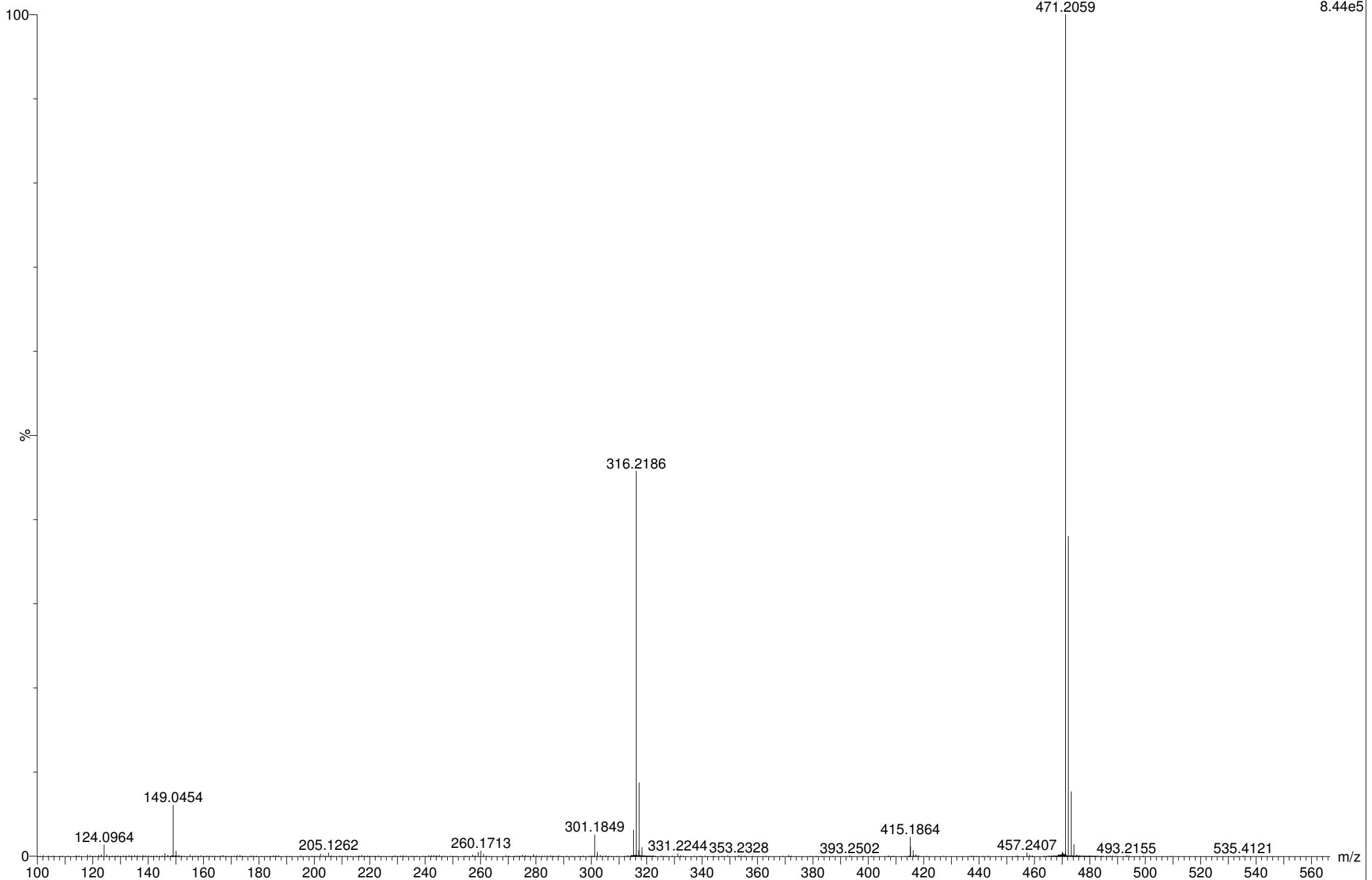
PROCESSED BY
PAWAN KUMAR

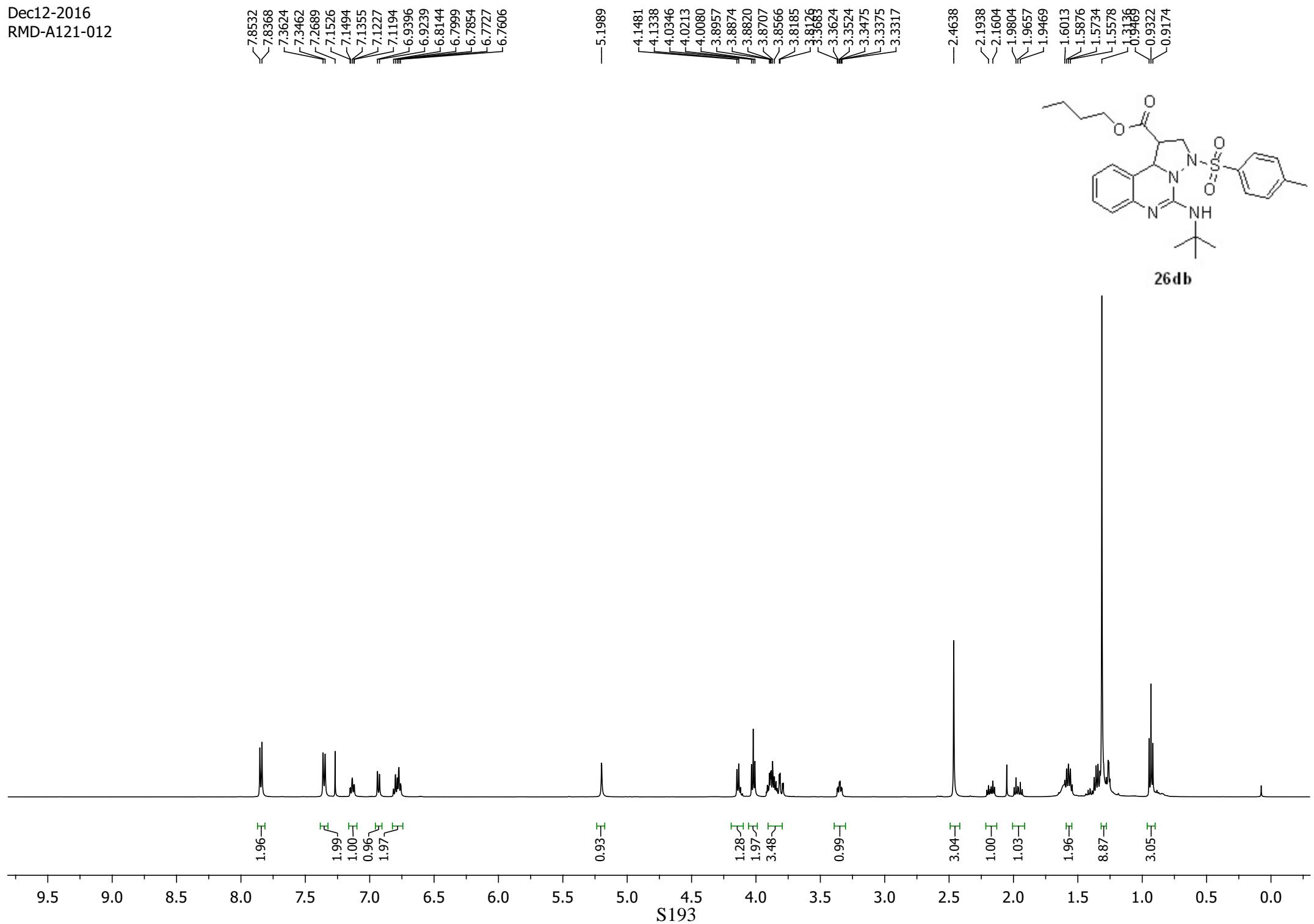
RMD-110_02 21 (0.389) AM (Cen,3, 80.00, Ar,5000.0,471.21,0.70); Sb (2,10.00); Cr (13:33)

CSIR-IHBT
NPC&PD DIVISION

19-Jan-2017 14:40:48

1: TOF MS ES+
8.44e5





-170.52

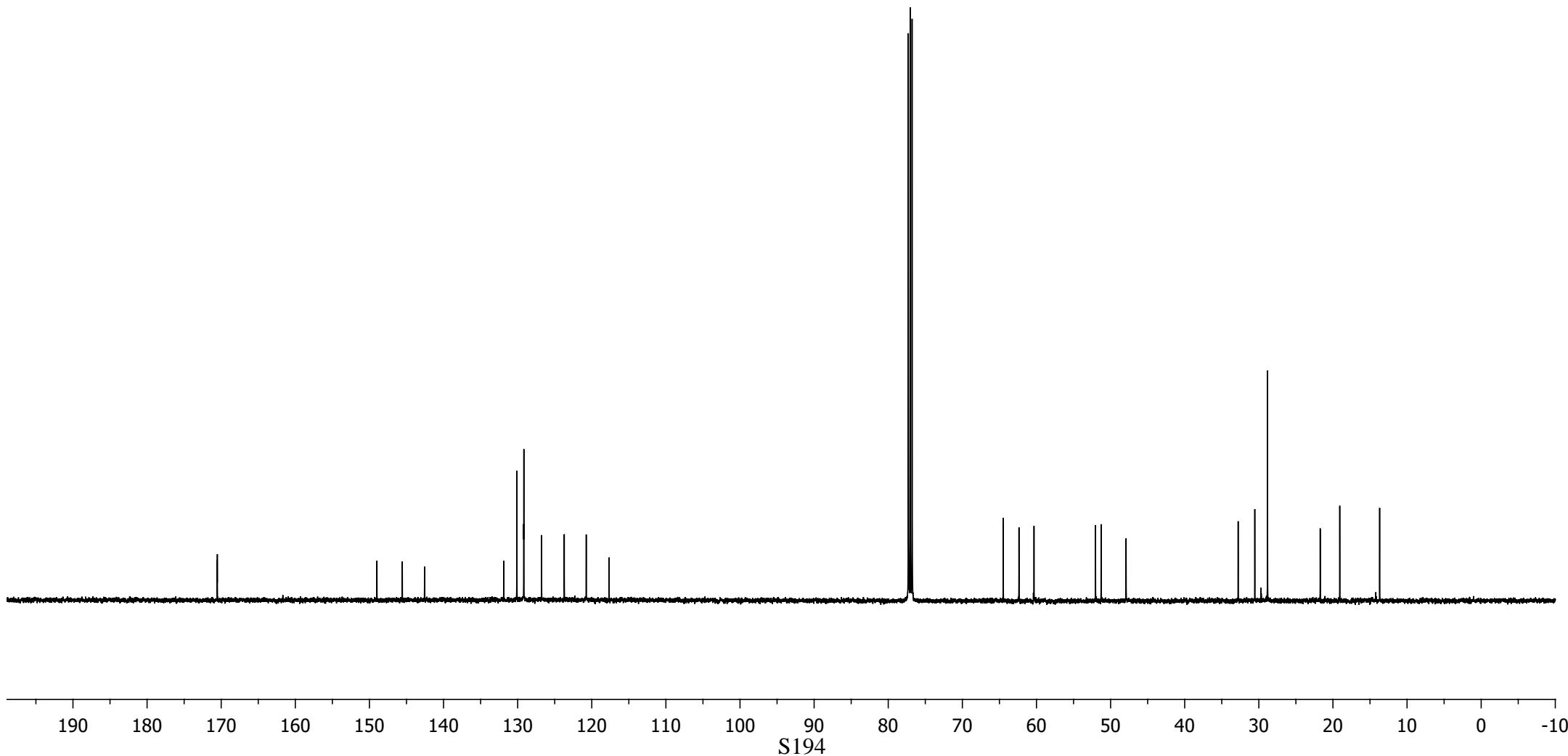
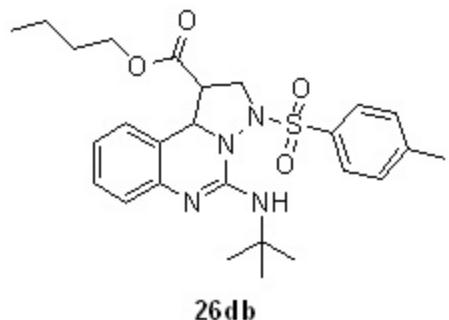
~148.98
-145.56
~142.56

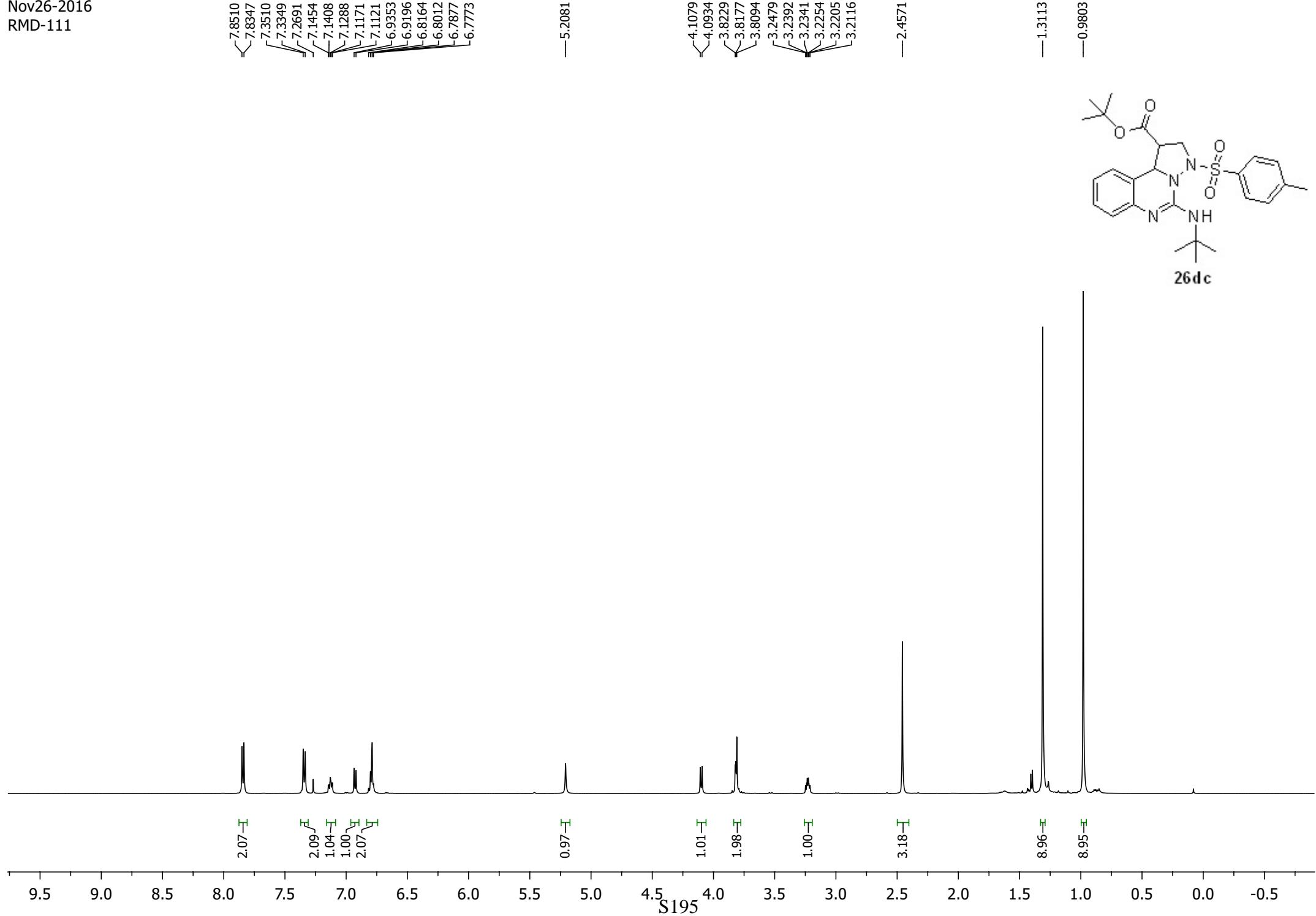
131.88
130.11
129.18
129.16
126.77
123.71
120.75
117.66

77.29
77.03
76.78

-62.36
-60.33
52.04
~51.28
~47.91

30.54
~28.83
-21.72
-19.08
-13.70





Nov26-2016
RMD-111

—169.54

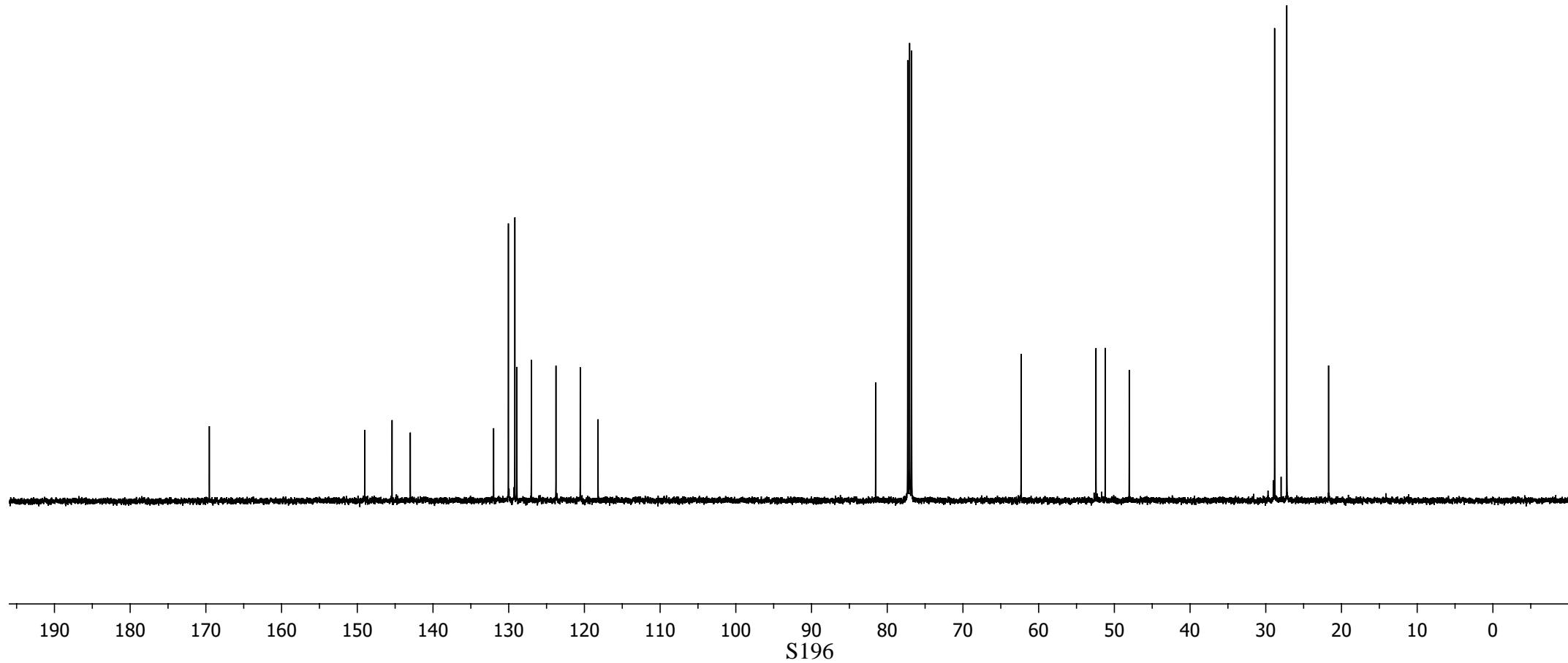
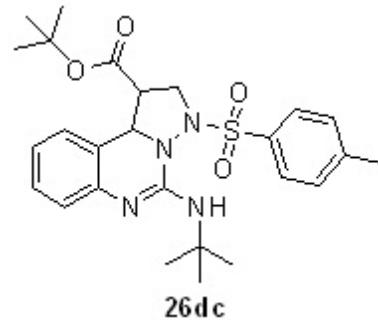
—149.02
—145.40
—142.99

—131.99
—130.04
—129.18
—128.92
—127.01
—123.75
—120.55
—118.19

—81.54
—77.30
—77.04
—76.79

—62.32
—52.43
—51.22
—48.04

—28.83
—27.22
—21.70



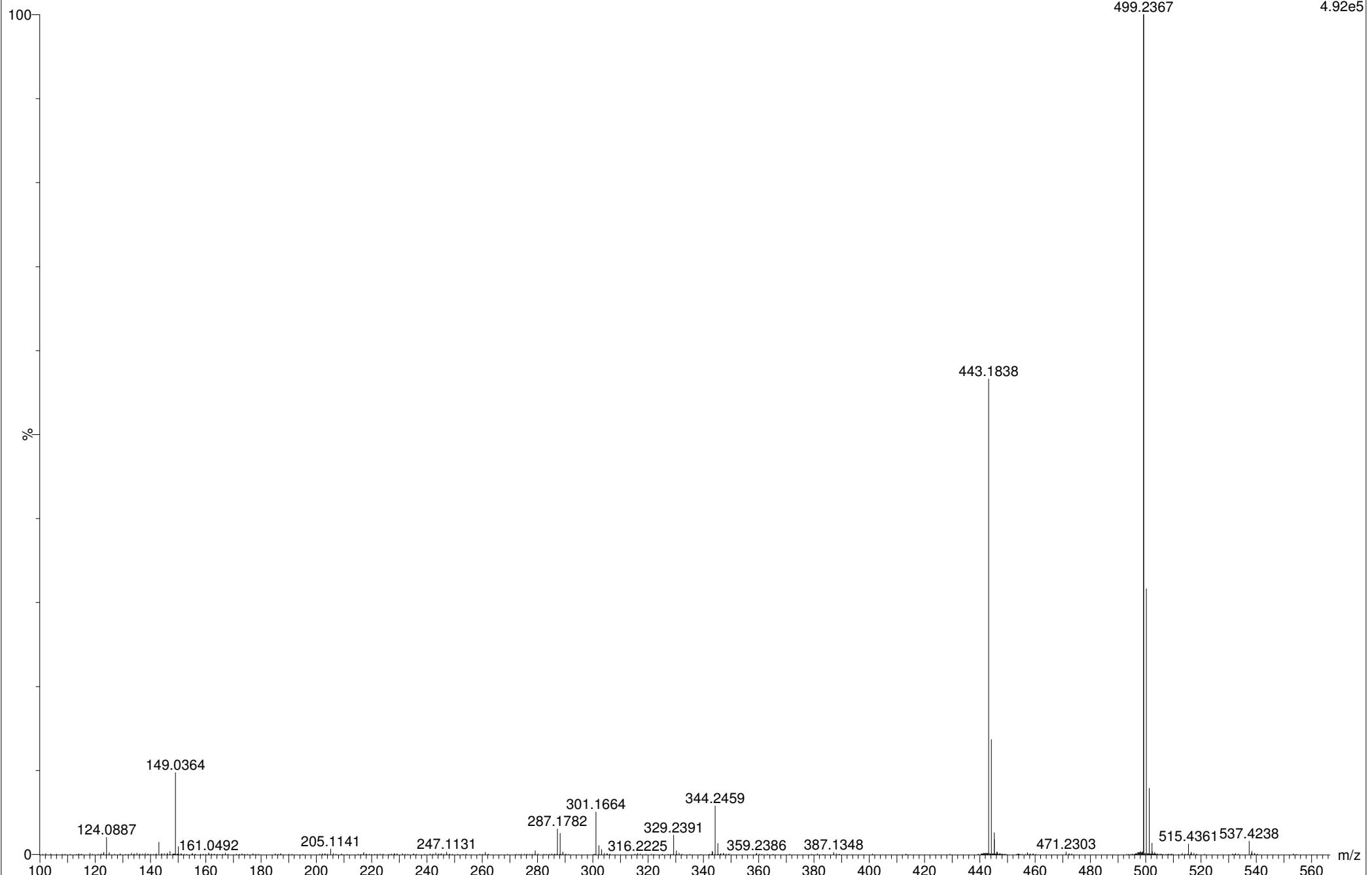
PROCESSED BY
PAWAN KUMAR

RMD-111_02 19 (0.353) AM (Cen,3, 80.00, Ar,5000.0,499.24,0.70); Sb (2,10.00); Crm (16:31)

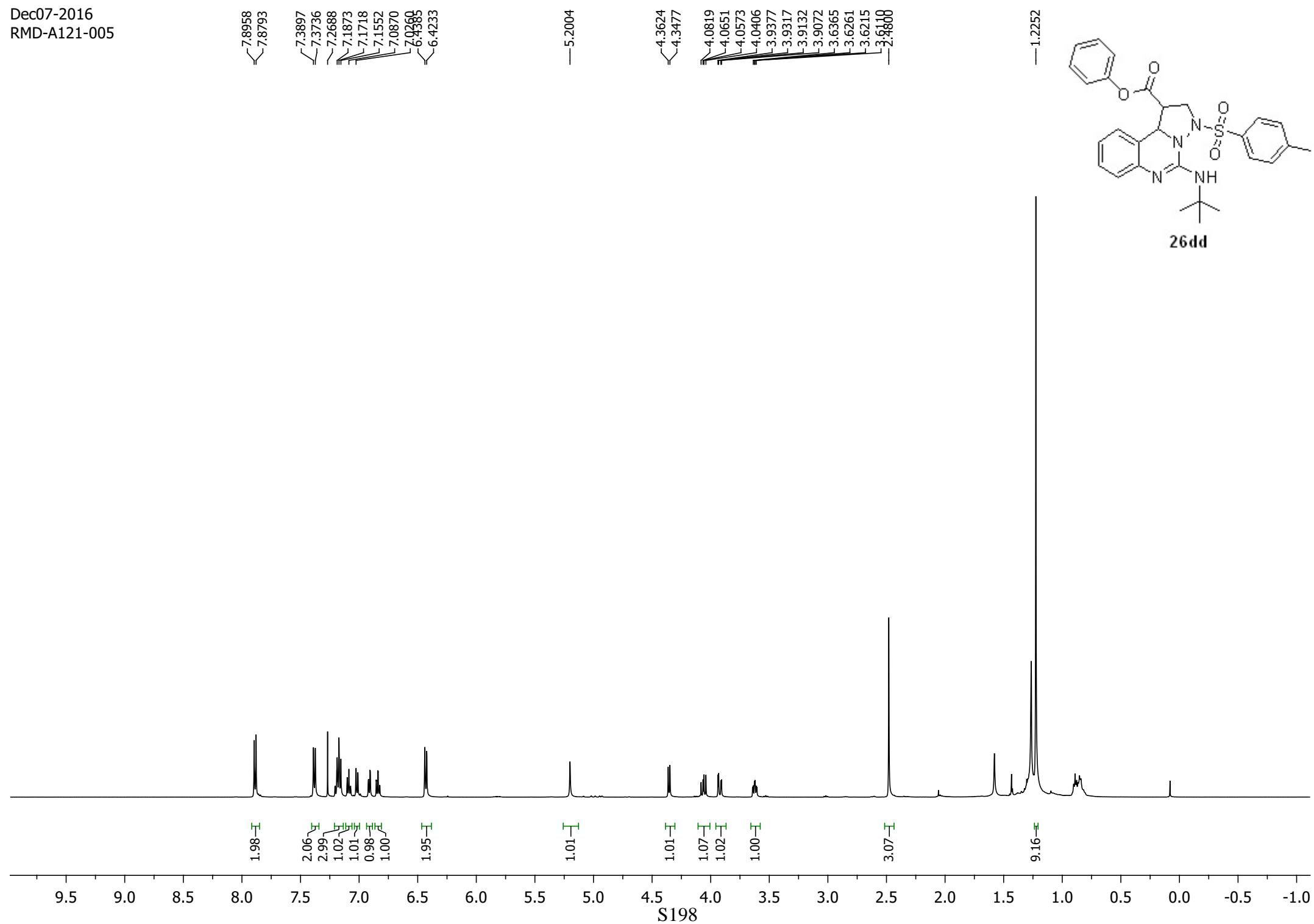
CSIR-IHBT
NPC&PD DIVISION

19-Jan-2017 14:44:56

1: TOF MS ES+
4.92e5



Dec07-2016
RMD-A121-005



Dec07-2016
RMD-A121-005

—169.61

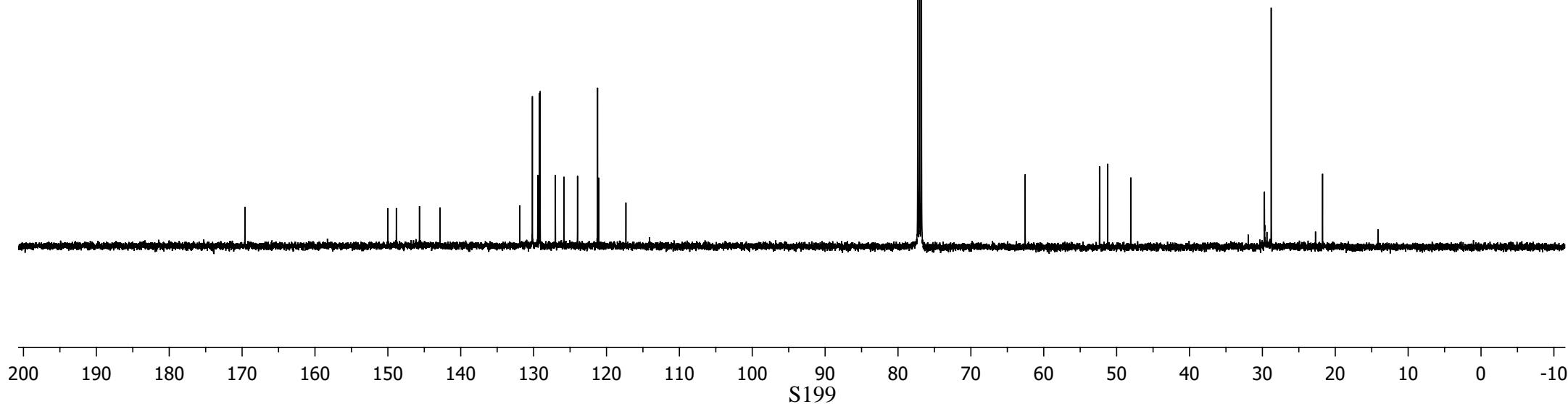
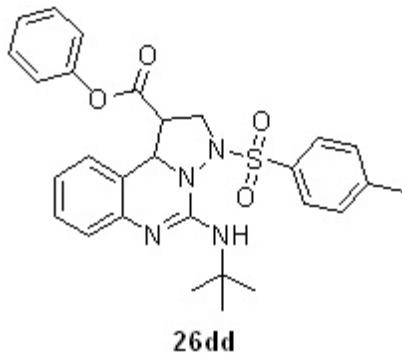
—150.00
~148.84
~145.66
—142.84

131.90
130.17
129.43
129.20
129.10
127.01
125.84
123.97
121.22
121.07
117.33

77.29
77.04
76.78

—62.55
—52.35
~51.24
~48.05

—28.79
—21.74



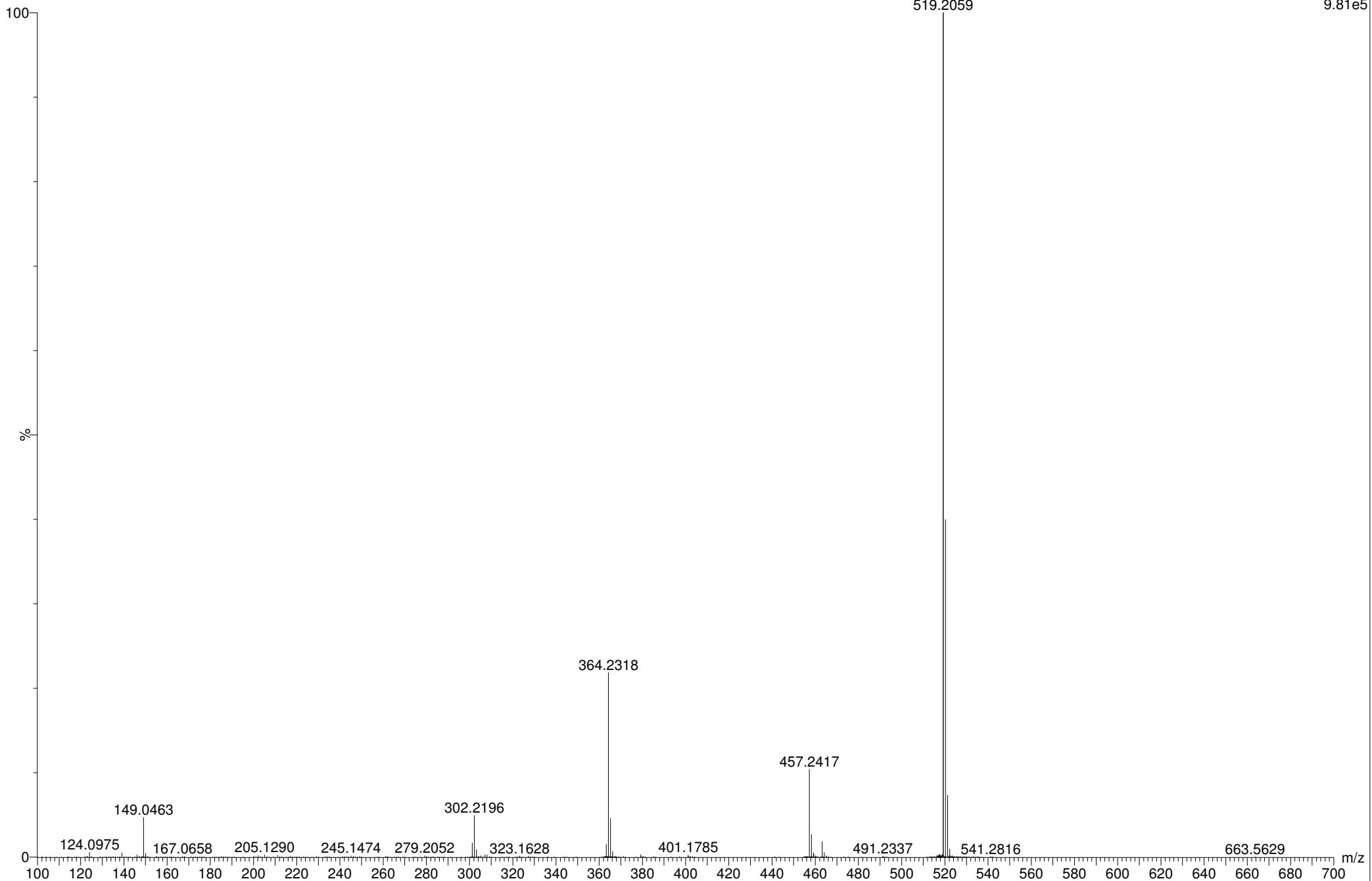
PROCESSED BY
PAWAN KUMAR

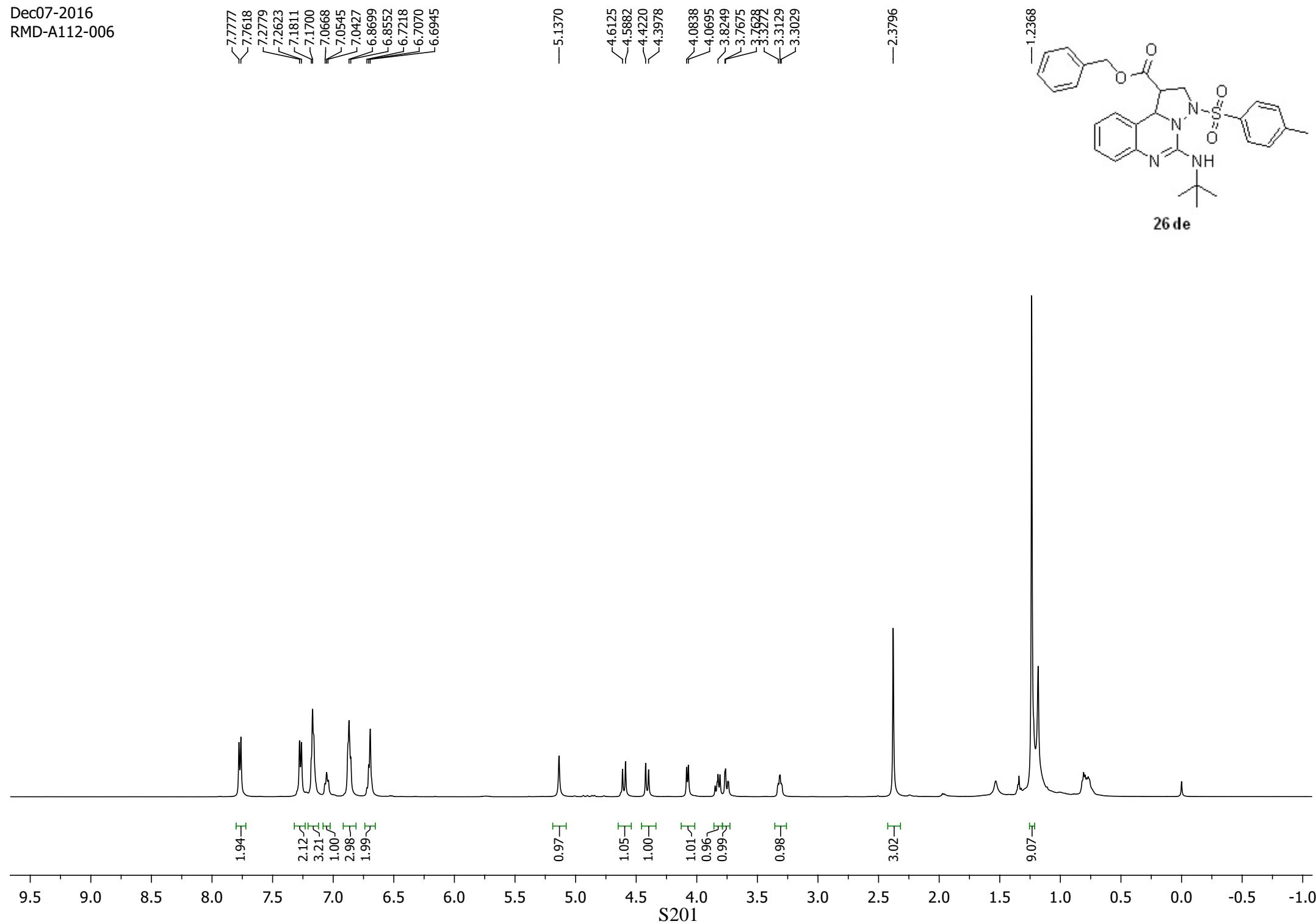
CSIR-IHBT
NPC&PD DIVISION

19-Jan-2017 14:16:53

RMD-A121-005_02 20 (0.371) AM (Cen,3, 80.00, Ar,5000.0,519.21,0.70); Sb (2,10.00); Cm (15:29)

1: TOF MS ES+
9.81e5





-170.71

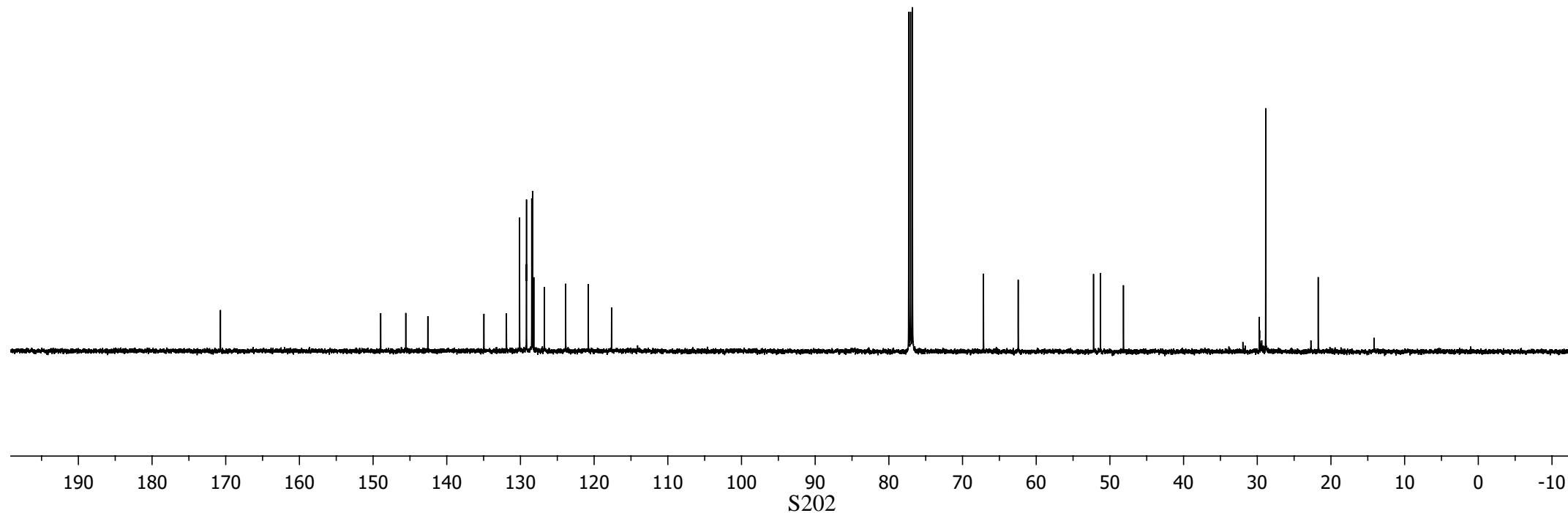
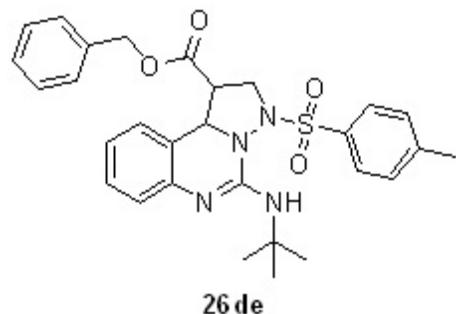
~148.96
~145.56
~142.57
134.96
131.90
130.12
129.20
~129.18
128.47
128.33
128.18
126.75
123.86
120.79
117.64

77.31
77.05
76.80

-67.16
-62.45

52.22
51.28
~48.16

-28.86
-21.73



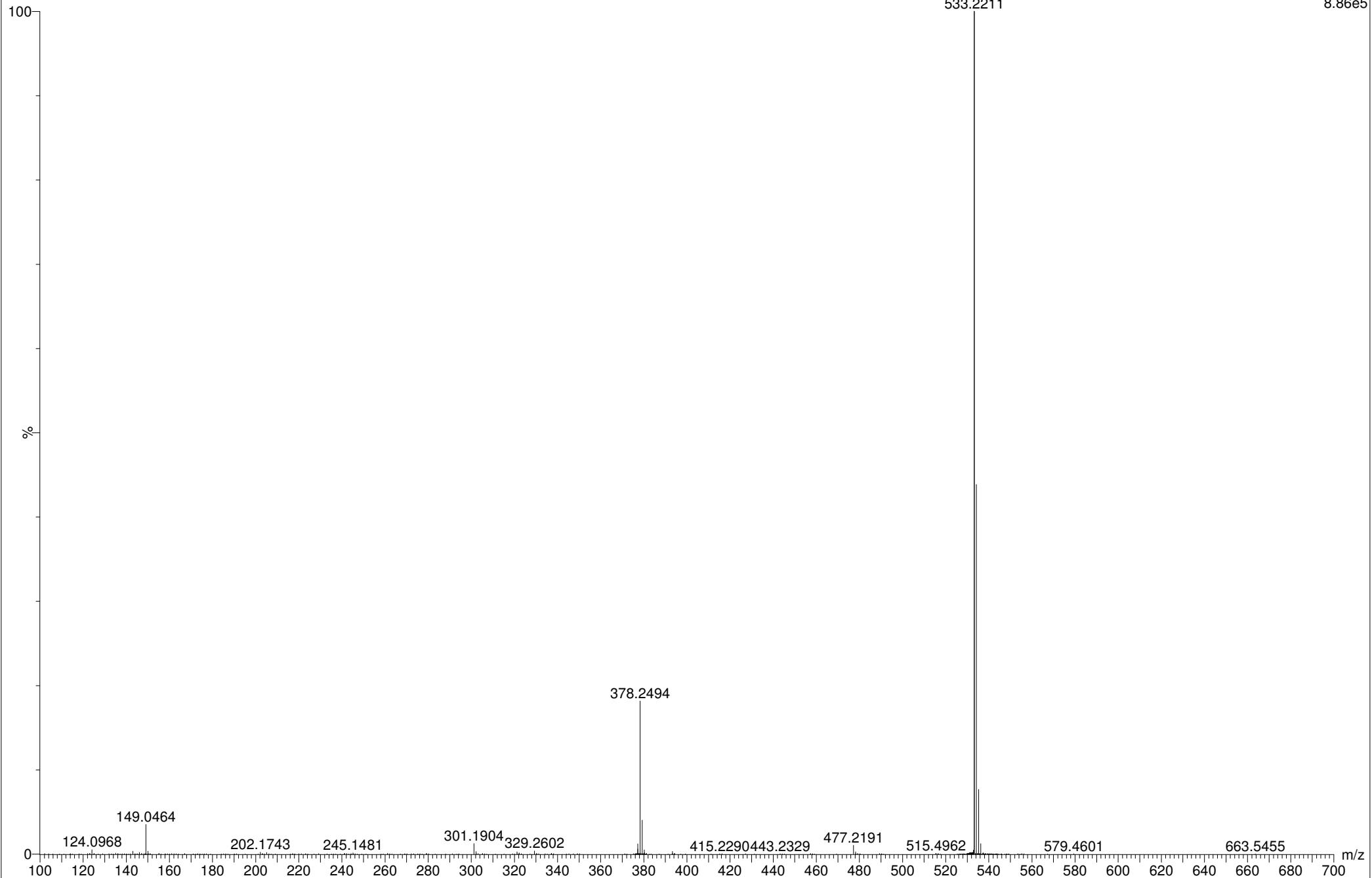
PROCESSED BY
PAWAN KUMAR

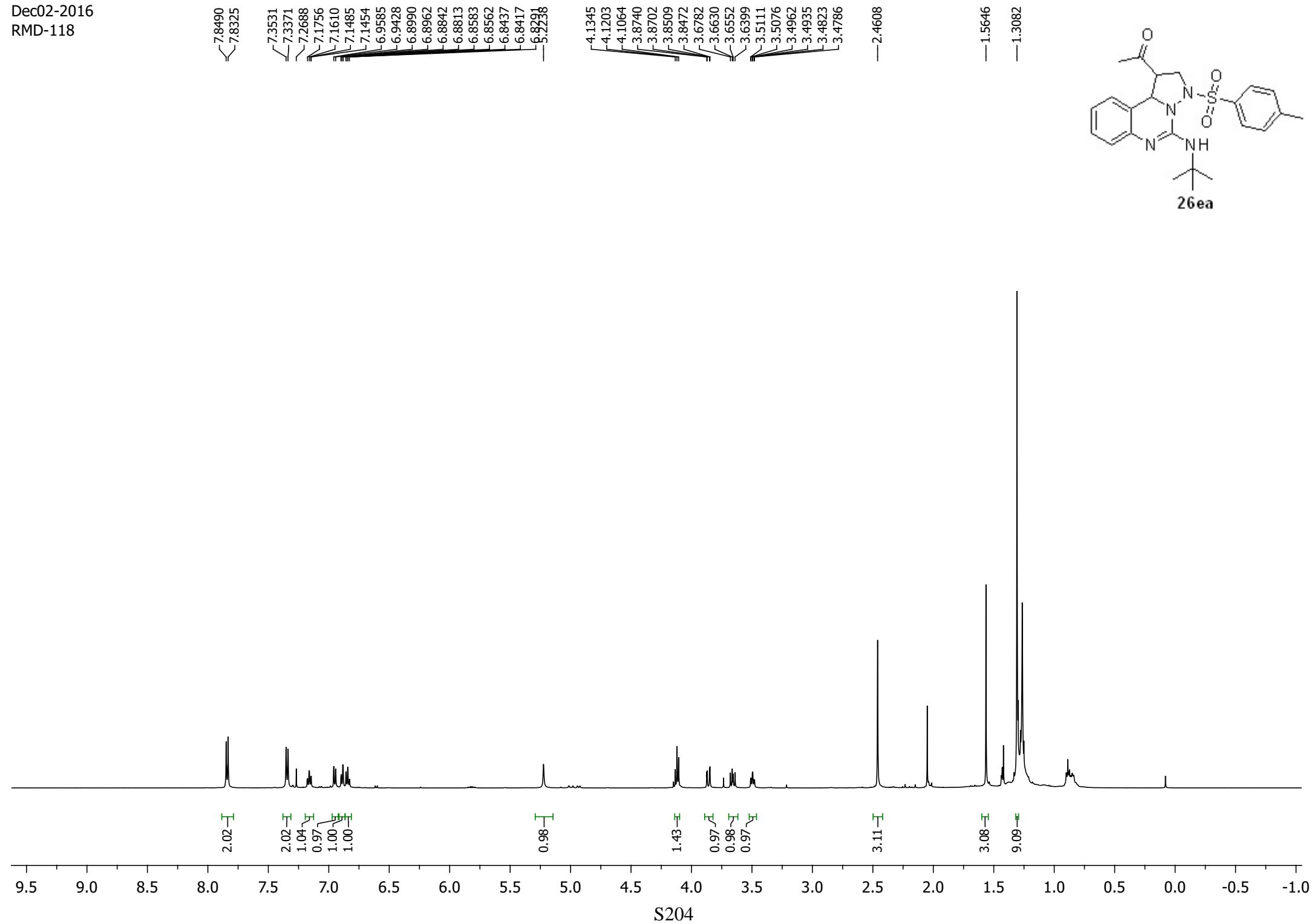
CSIR-IHBT
NPC&PD DIVISION

19-Jan-2017 13:25:56

RMD-A121-006_02 20 (0.371) AM (Cen,3, 80.00, Ar,5000.0,533.22,0.70); Sb (2,10.00); Cm (16:28)

1: TOF MS ES+
8.86e5





Dec02-2016

RMD-118¹³C

—205.16

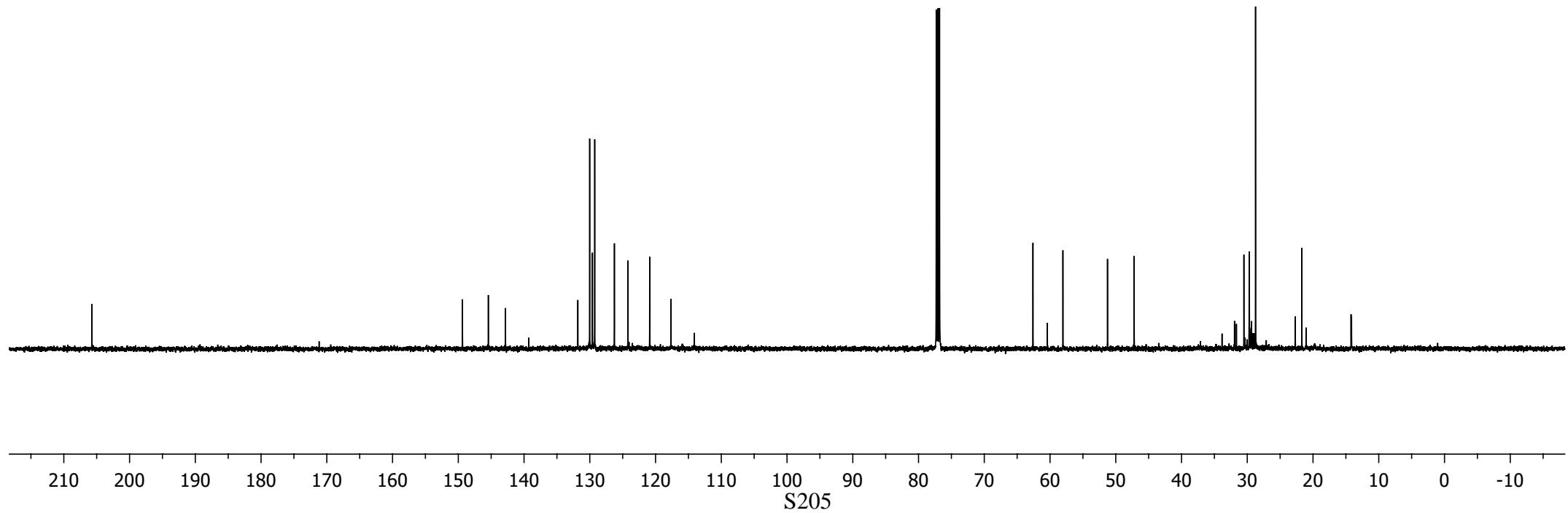
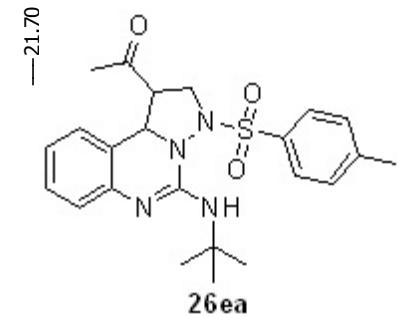
—149.36
—145.43
—142.83

131.82
130.02
129.60
129.25
126.26
124.20
120.89
117.66
114.09

77.31
77.05
76.80

—62.59
—60.41
—58.04
—51.26
—47.21

—30.51
—28.74



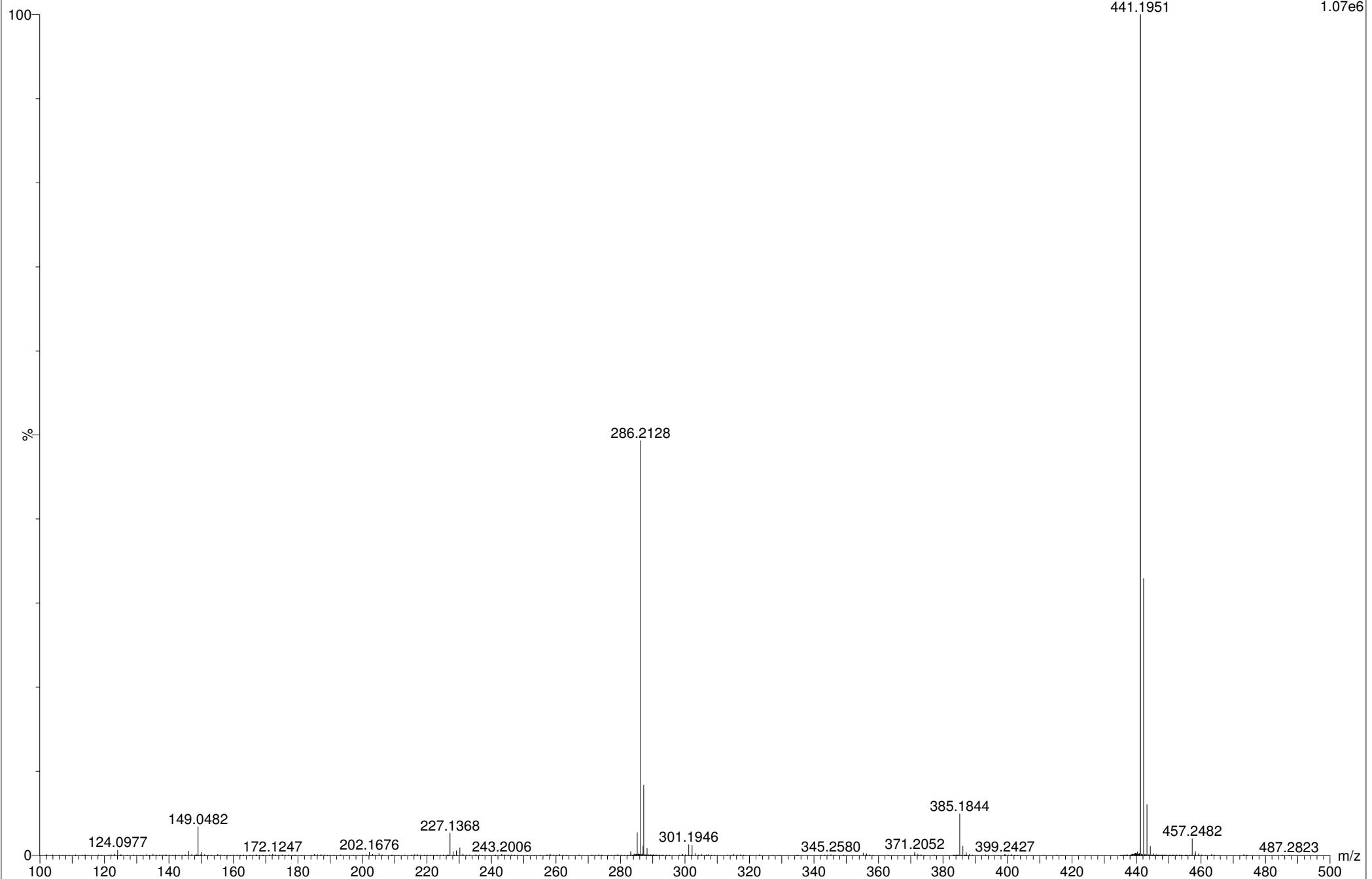
PROCESSED BY
PAWAN KUMAR

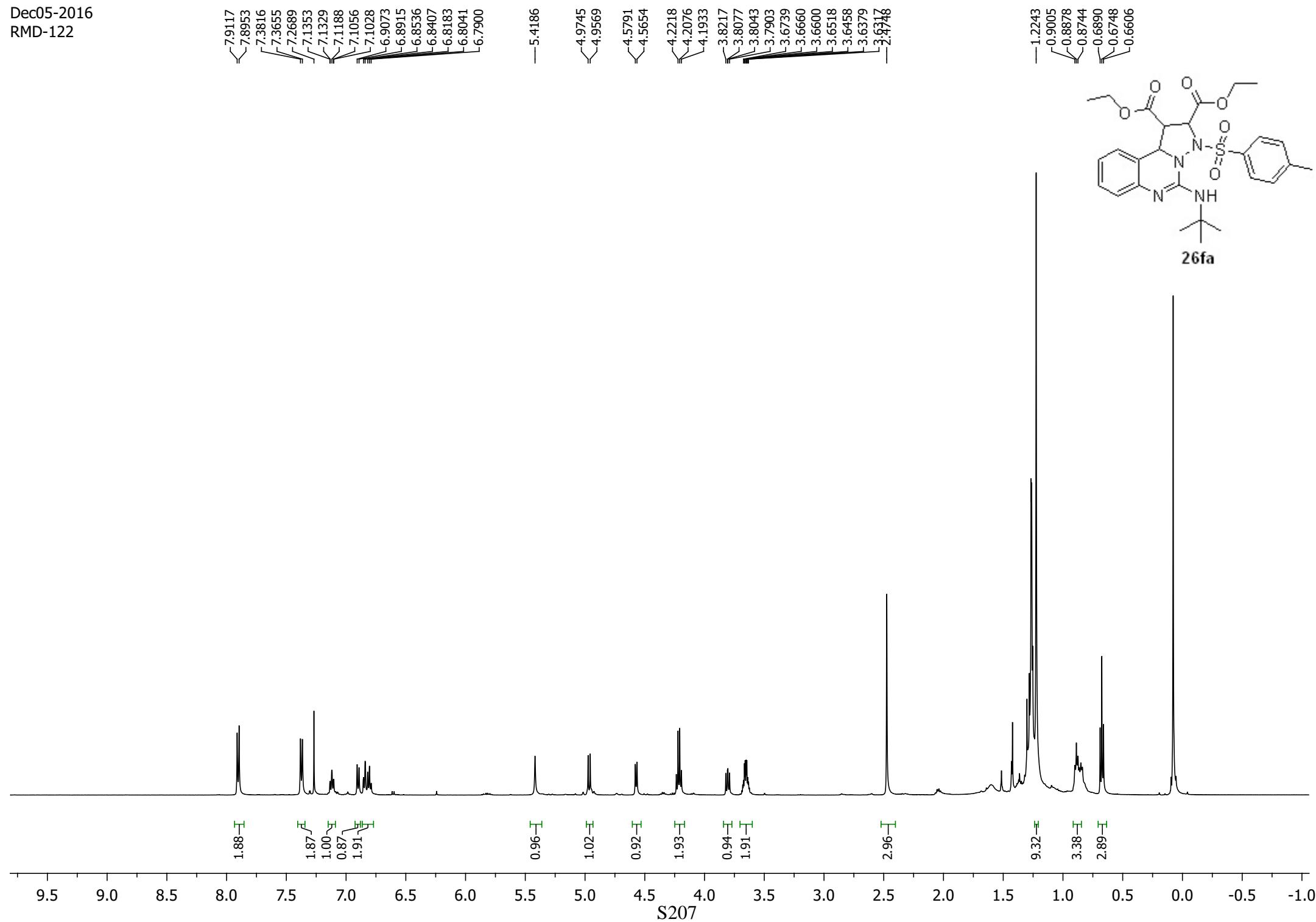
RMD-118_02 19 (0.352) AM (Cen,3, 80.00, Ar,5000.0,441.20,0.70); Sb (2,10.00); Cr (15:31)

CSIR-IHBT
NPC&PD DIVISION

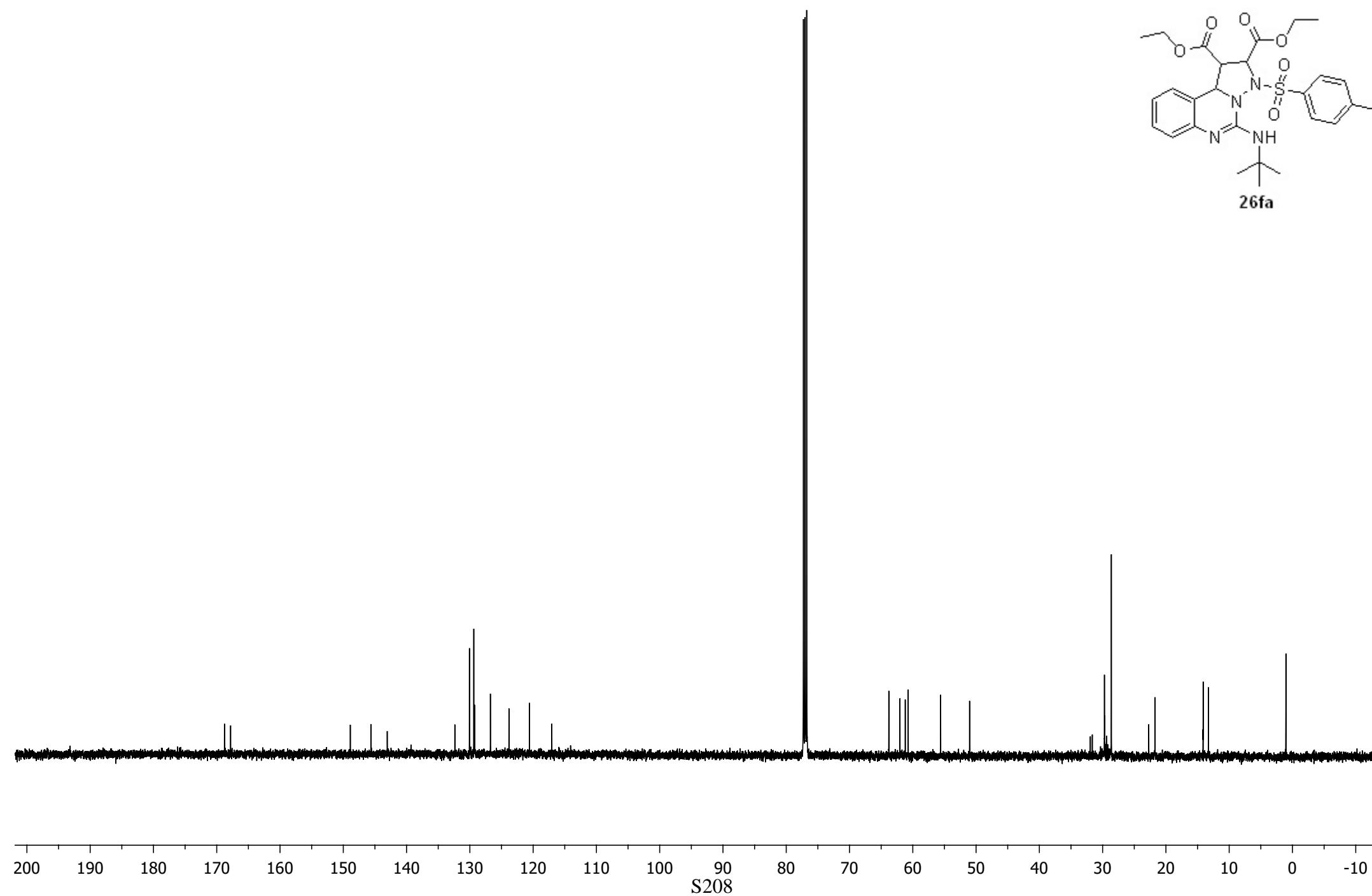
19-Jan-2017 13:49:26

1: TOF MS ES+
1.07e6





Dec05-2016
RMD-122



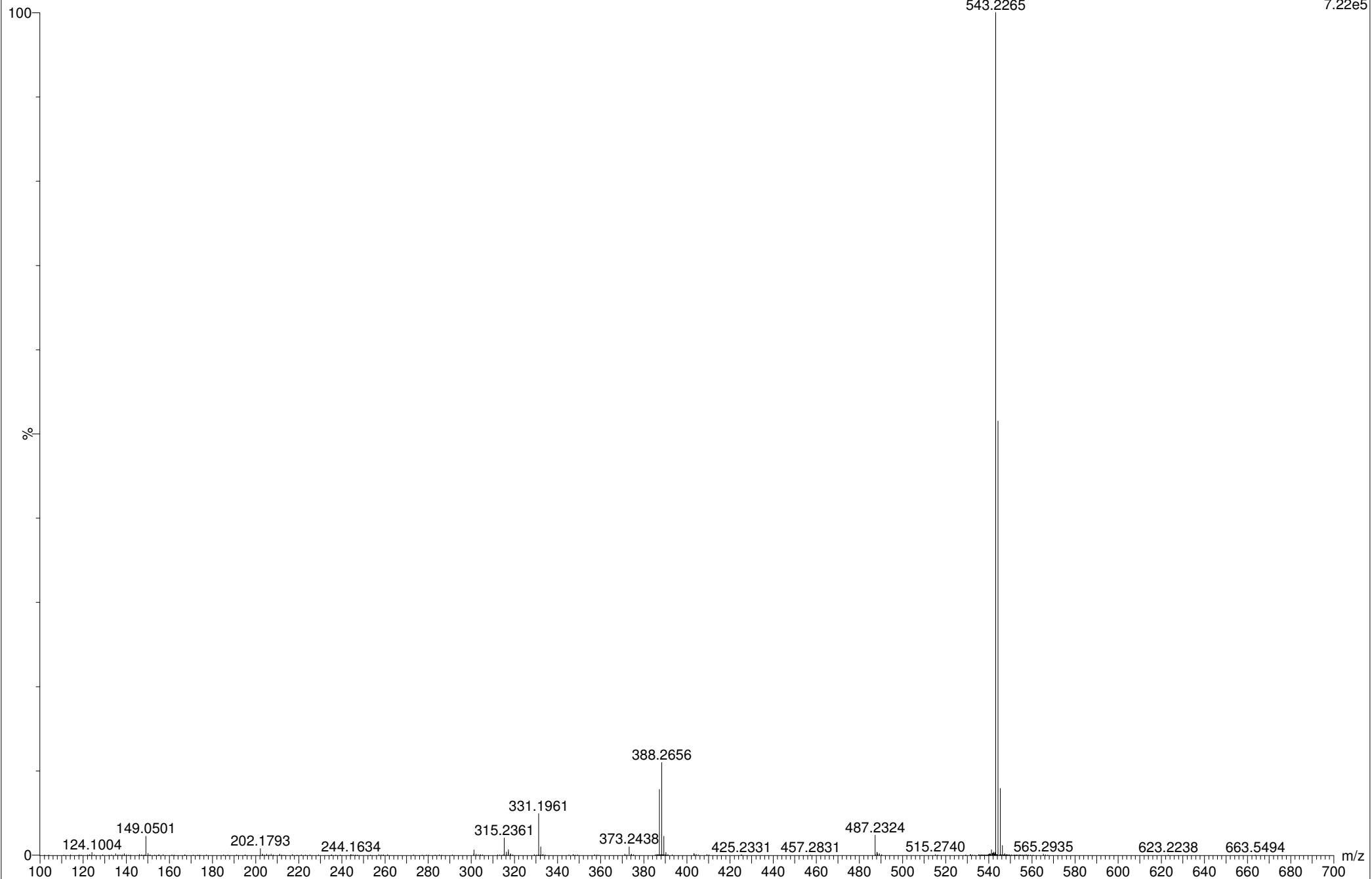
PROCESSED BY
PAWAN KUMAR

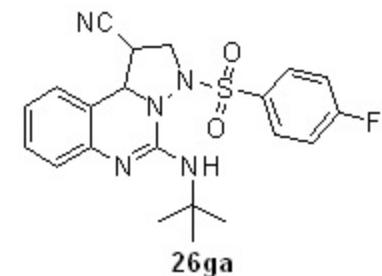
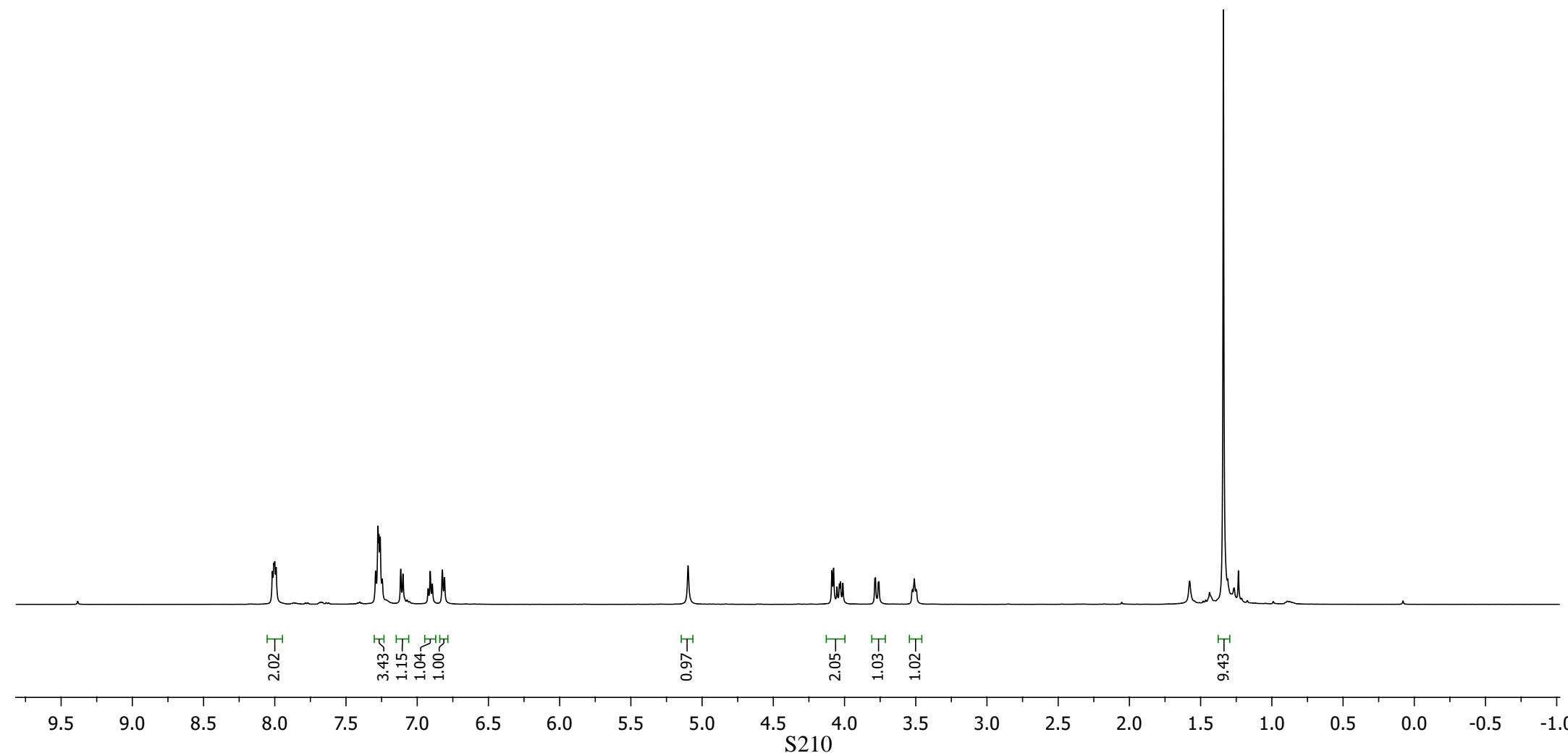
CSIR-IHBT
NPC&PD DIVISION

19-Jan-2017 15:25:19

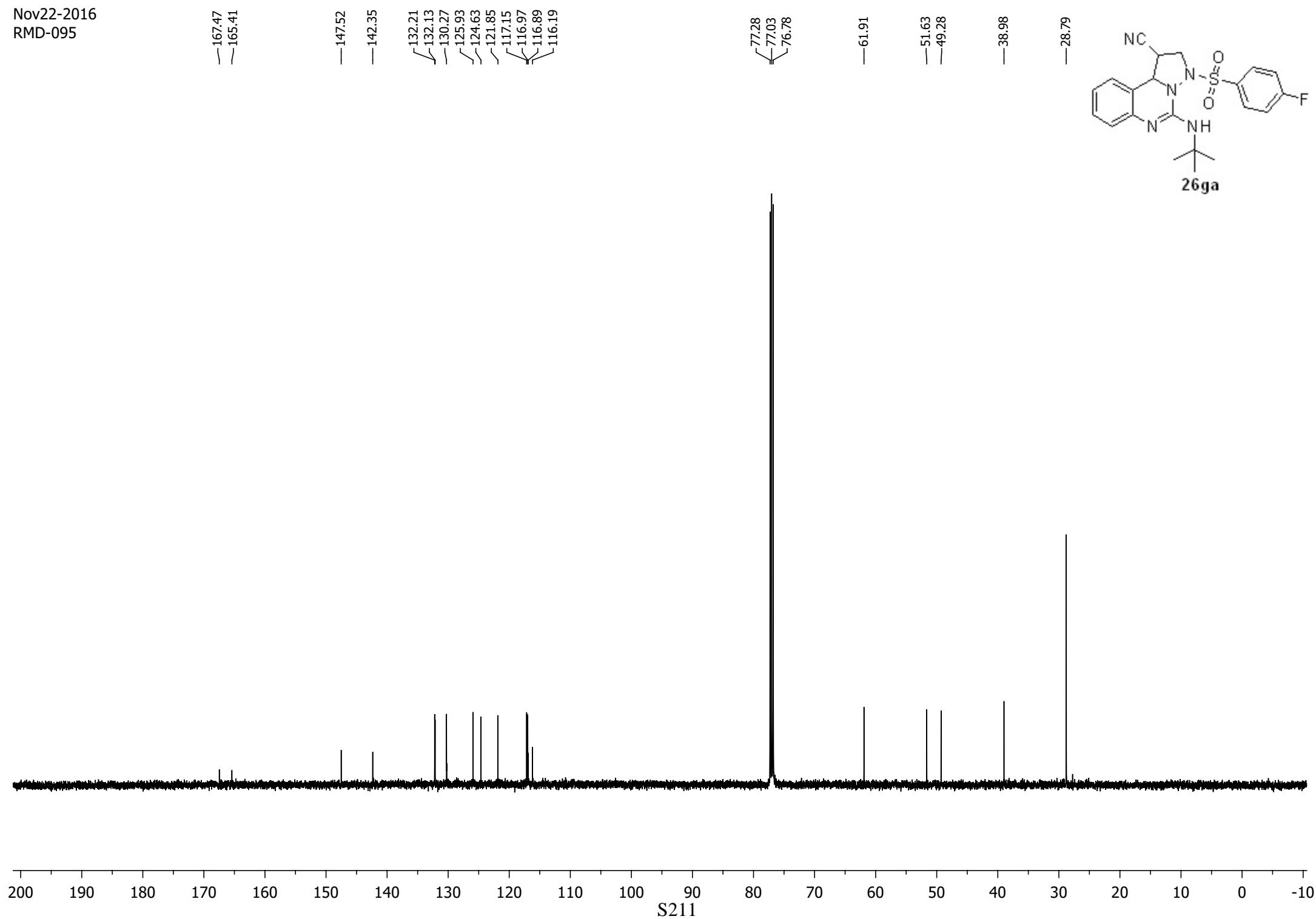
RMD-122_02 20 (0.371) AM (Cen,3, 80.00, Ar,5000.0,543.23,0.70); Sb (2,10.00); Cr (17:26)

1: TOF MS ES+
7.22e5

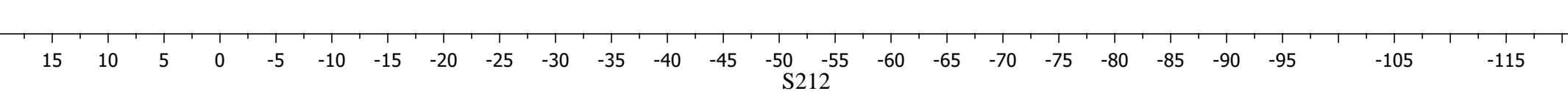
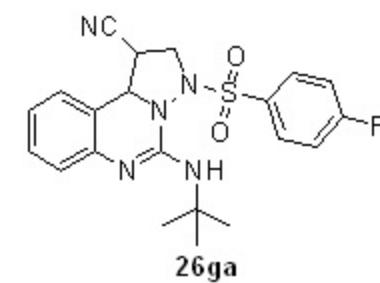




Nov22-2016
RMD-095



Nov24-2016
RMD-095



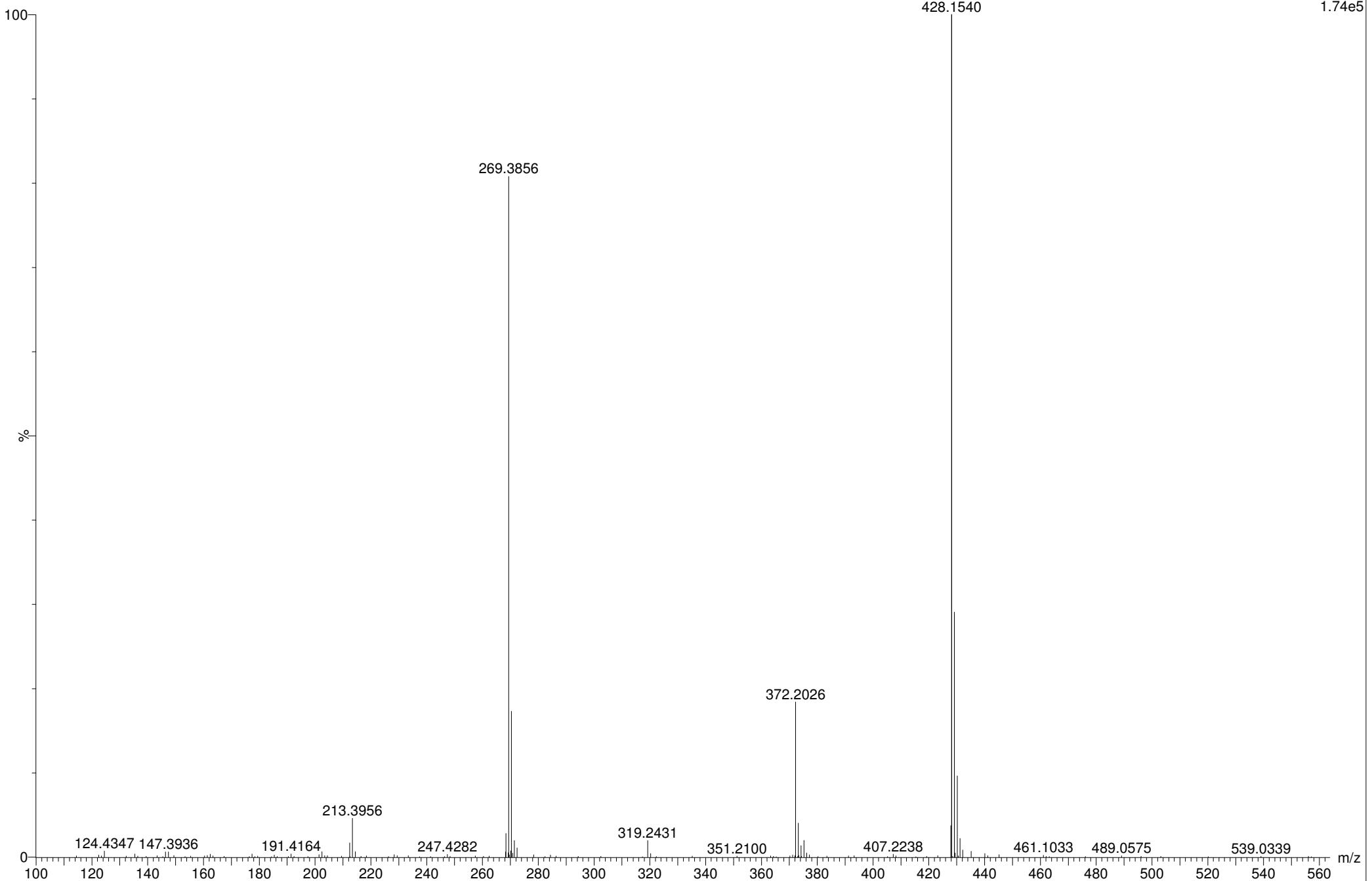
PROCESSED BY
PAWAN KUMAR

A121-RMD-095_01 22 (0.407) AM (Top,10, Ar,5000.0,428.15,0.70); Sm (SG, 1x1.00); Cm (17:31-45:52)

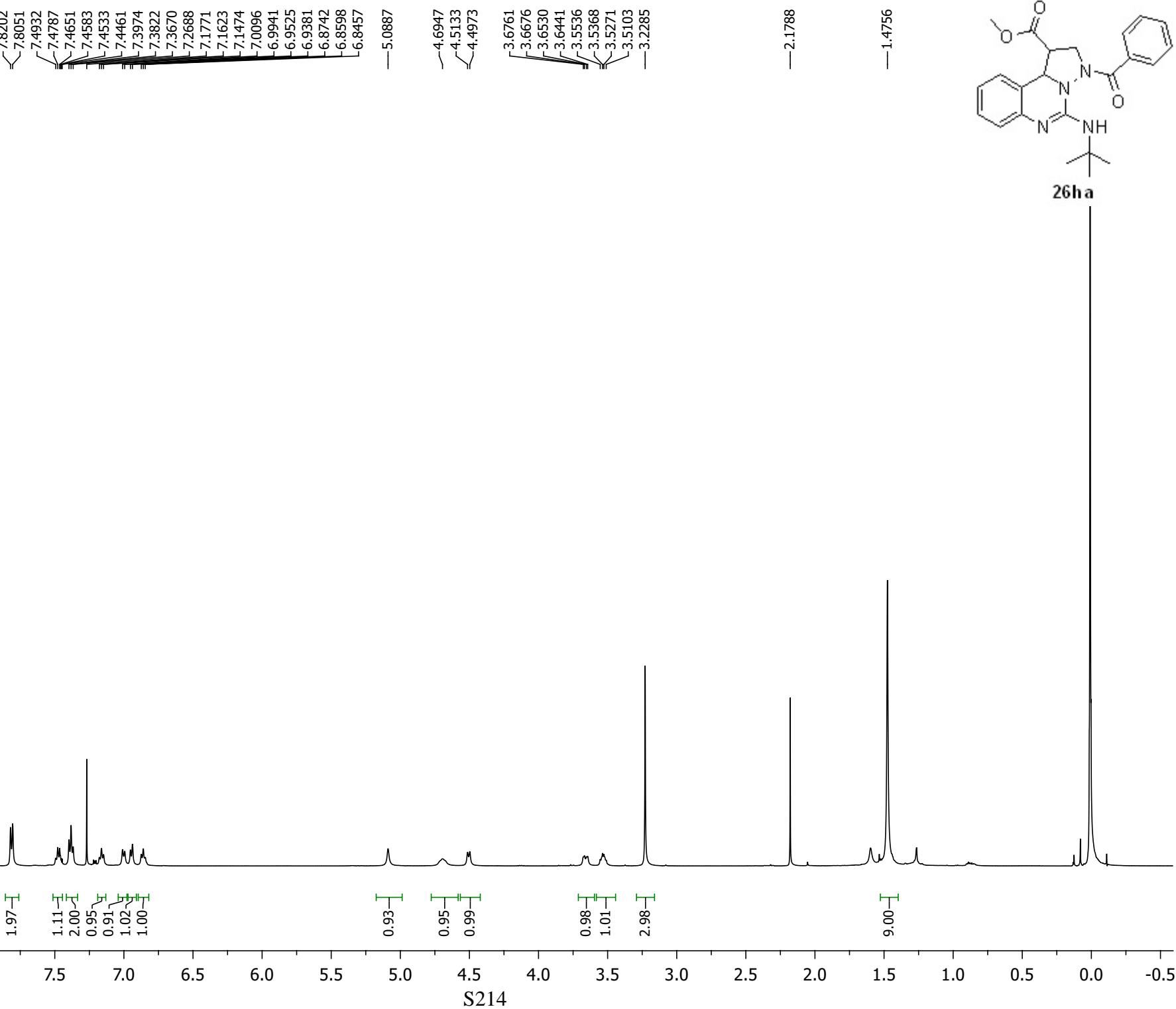
CSIR-IHBT
NPC&PD DIVISION

24-May-2017 12:57:28

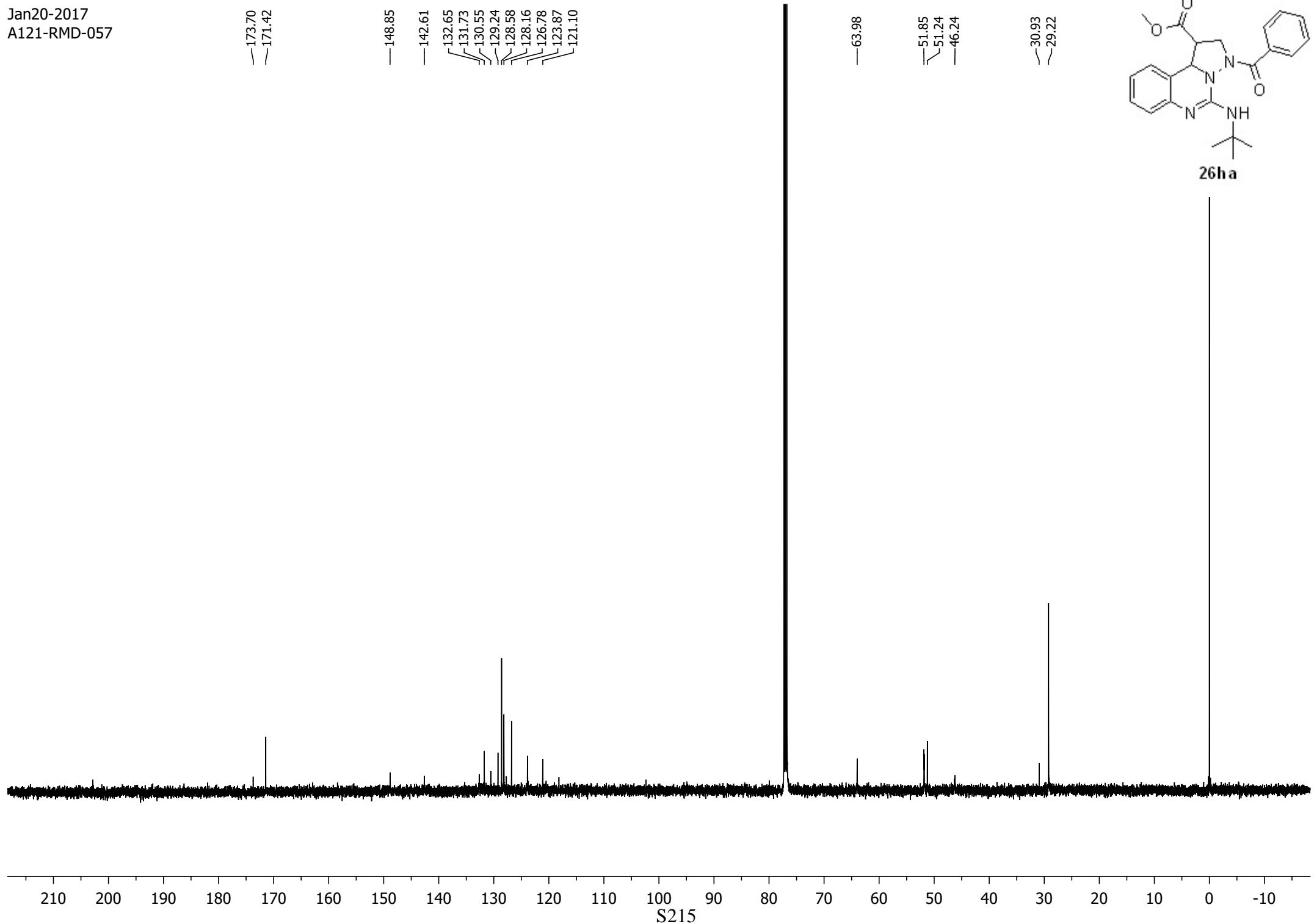
1: TOF MS ES+
1.74e5



Jan20-2017
A121-RMD-057



Jan20-2017
A121-RMD-057



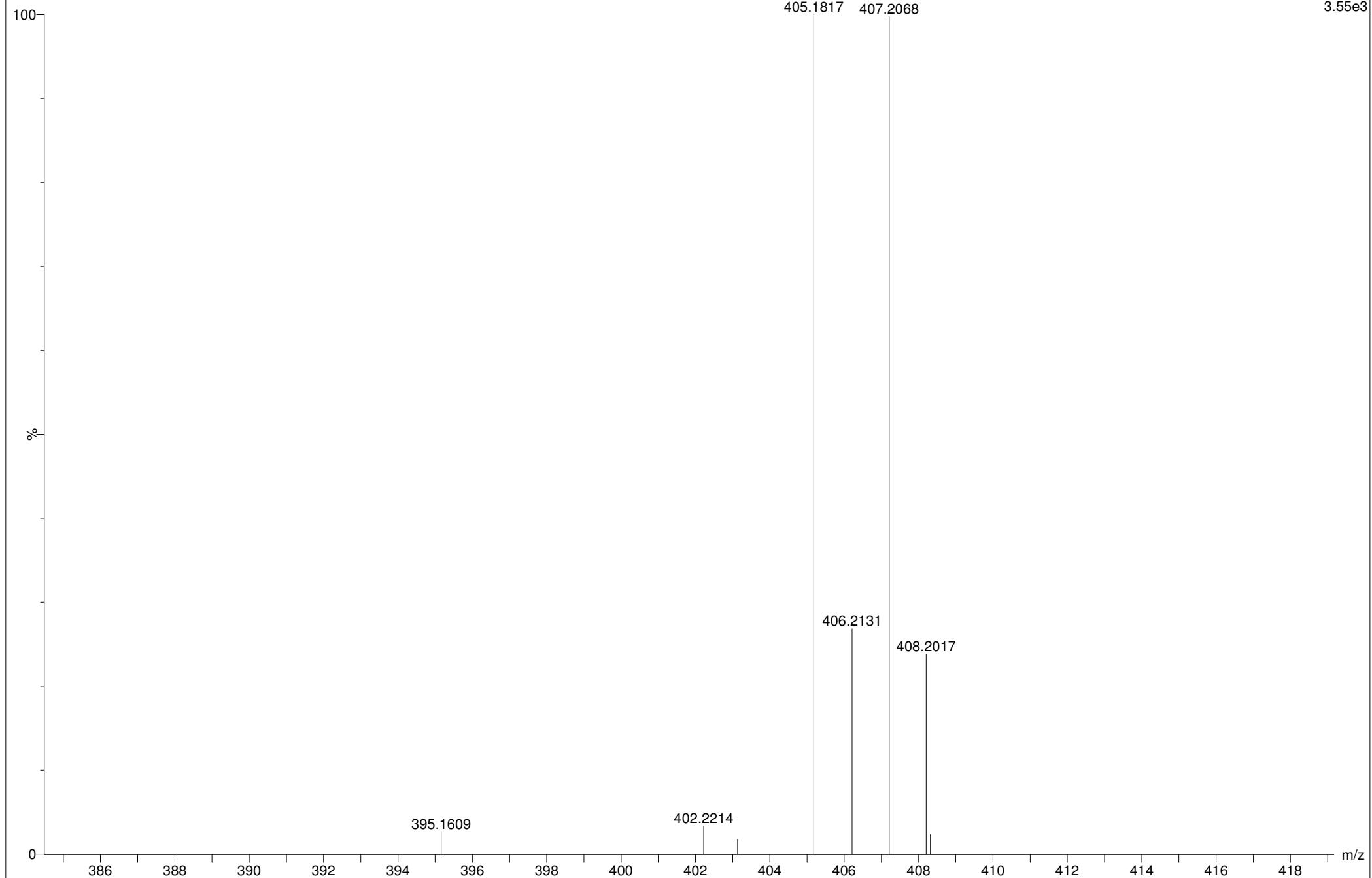
PROCESSED BY
PAWAN KUMAR

A121-RMD-057_01 20 (0.371) AM (Top,10, Ar,5000.0,407.21,0.70); Sm (SG, 1x1.00); Cm (16:31-44:57)

CSIR-IHBT
NPC&PD DIVISION

24-May-2017 13:01:28

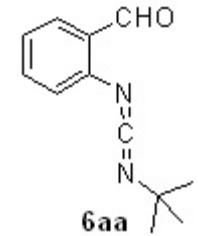
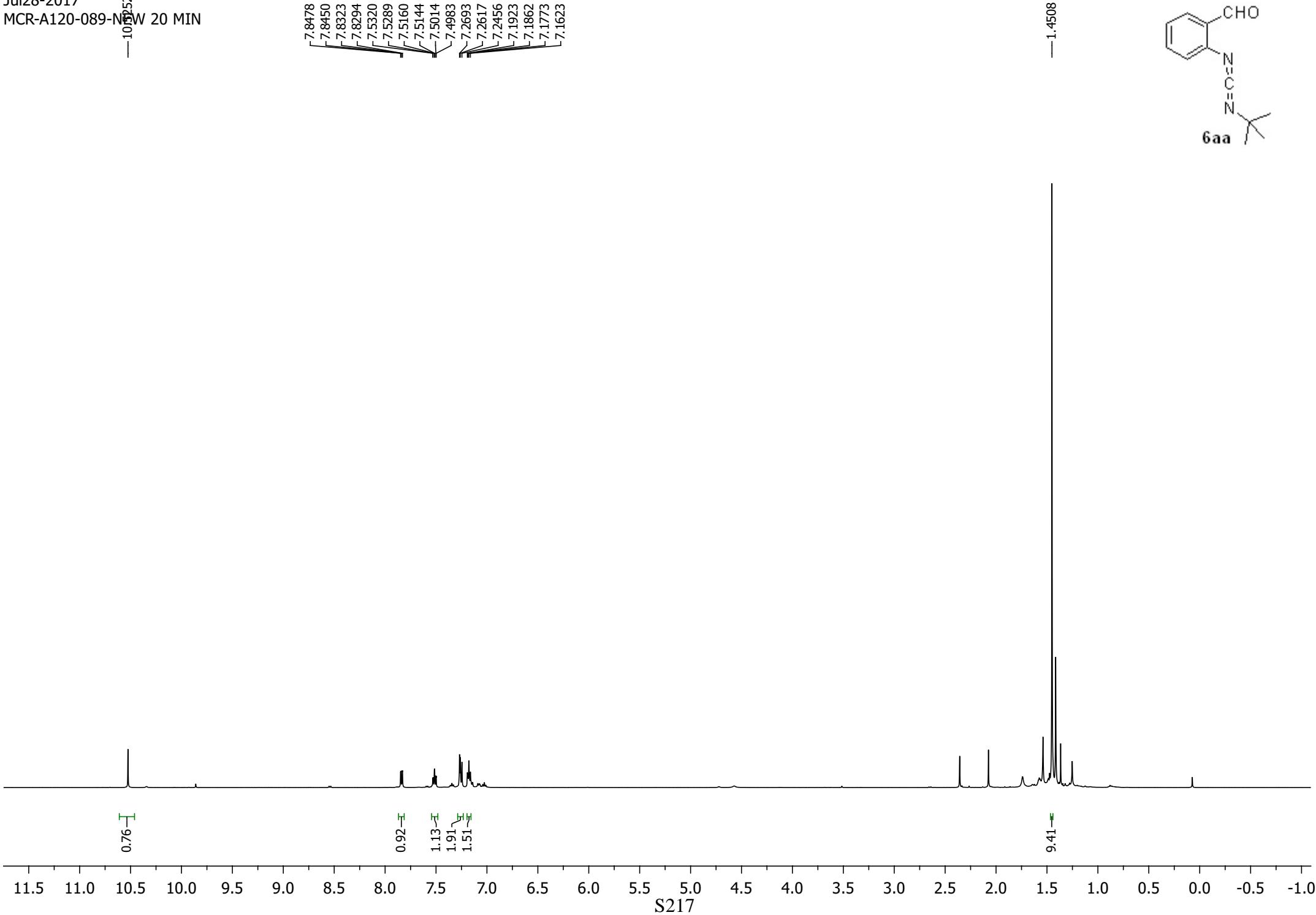
1: TOF MS ES+
3.55e3



Jul28-2017

MCR-A120-089-NEW 20 MIN

—105252



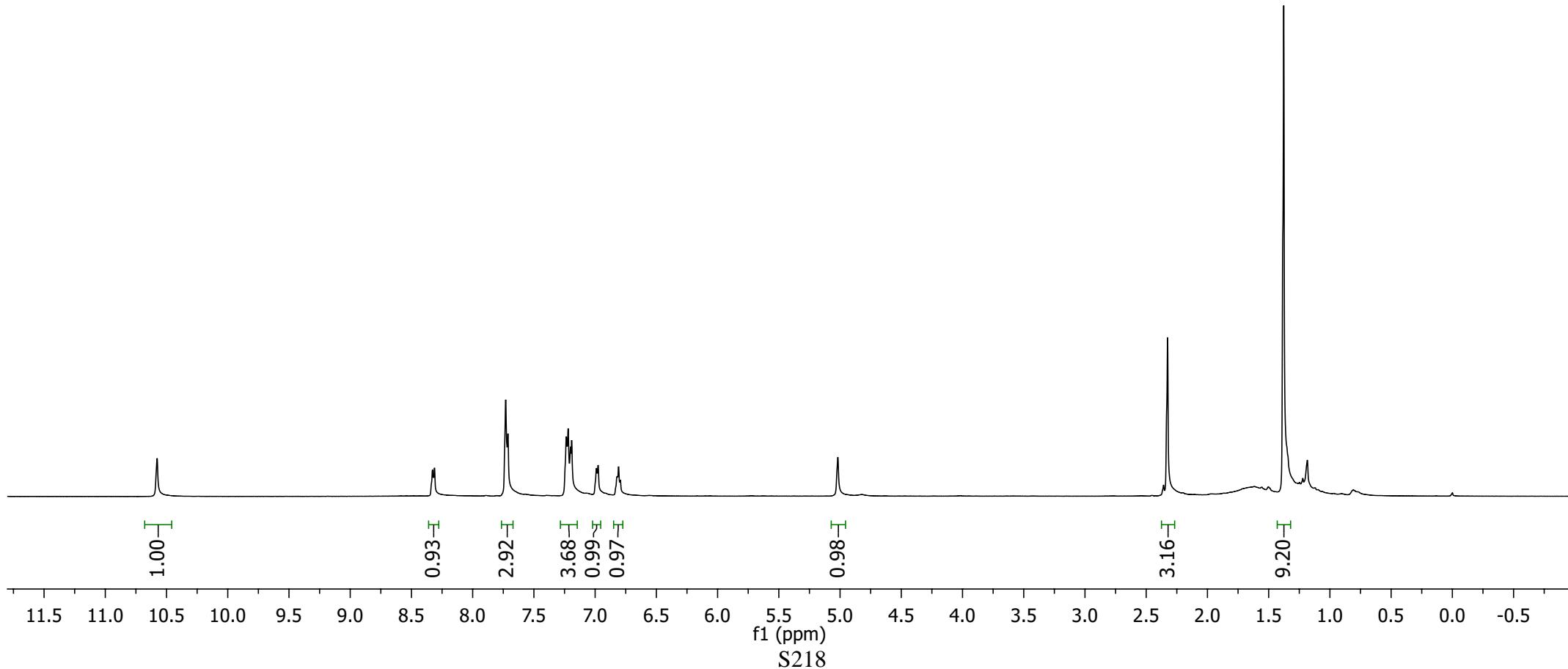
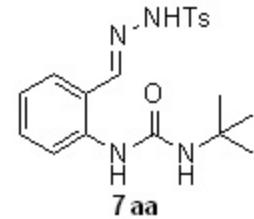
-10.5769

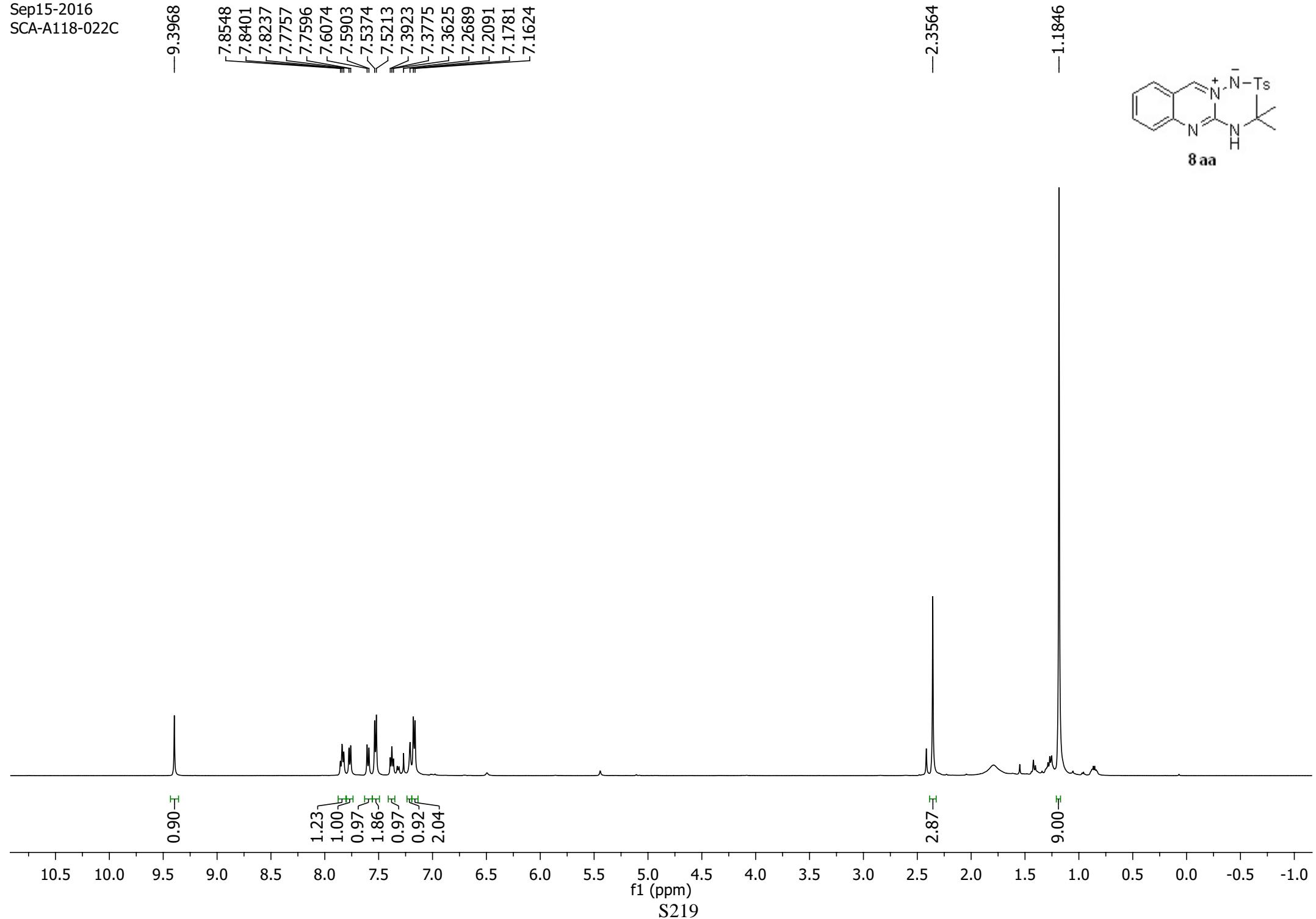
8.3281
8.3117
7.7303
7.7120
7.2355
7.2204
7.2009
7.1917
6.9896
6.9756
6.8200
6.8084
6.7947

-5.0178

-2.3254

-1.3711





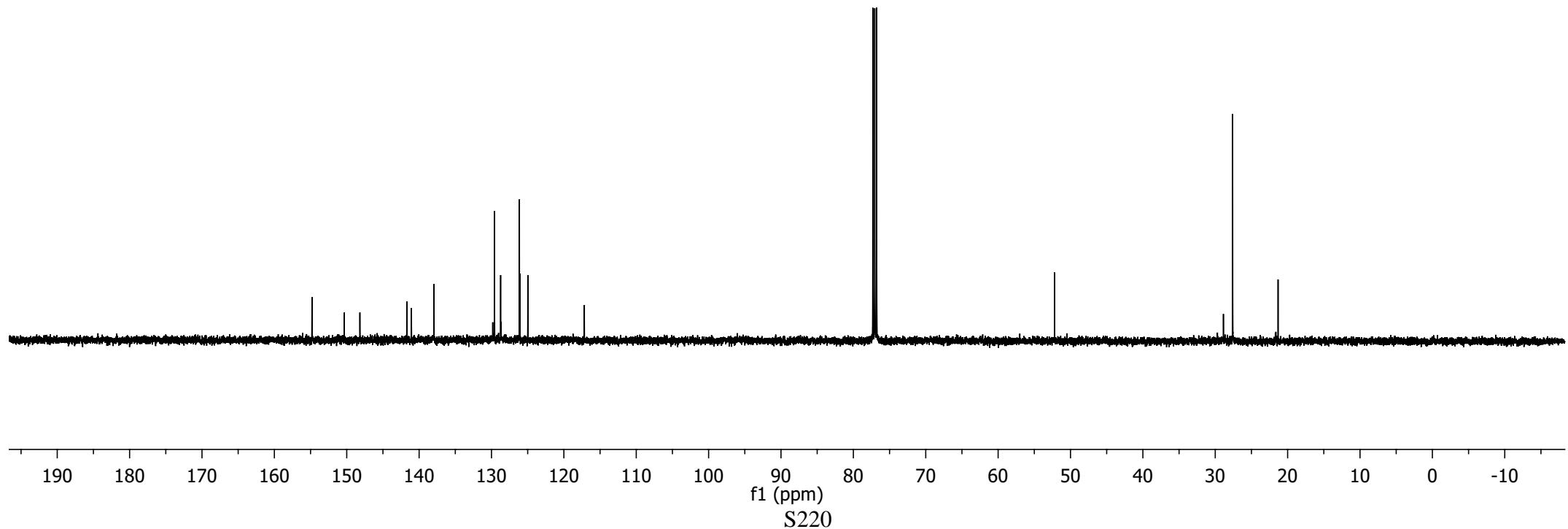
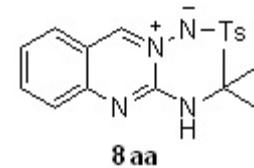
Sep15-2016
SCA-A118-022C

-154.80
-150.34
-148.18
-141.70
-141.06
-137.94
-129.58
-128.74
-126.14
-126.07
-124.98
-117.17

-52.21

-27.59

-21.32



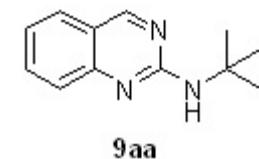
Apr08-2016
SCA-SUL-001

-8.9341

7.6710
7.6683
7.6543
7.6514
7.6407
7.6260
7.5786
7.5619
7.2687
7.2208
7.2190
7.2051
7.1912
7.1894

-5.2889

1.5353



0.96

2.03
1.01
1.00

1.05

9.35

9.5 9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0 -0.5 -1.0

S221

