Hyperpolarized Fumarate via Parahydrogen – Supporting Information

Barbara Ripka^{a,†}, James Eills^{b,†,*}, Hana Kouřilová^b, Markus Leutzsch^c, Malcolm H. Levitt^b, and Kerstin Münnemann^{a,d}

^a Max Planck Institute for Polymer Research, Mainz, Germany, ^b University of Southampton, Southampton, United Kingdom, ^c Department of Chemical Engineering and Biotechnology, University of Cambridge, Cambridge, United Kingdom, ^d Technical University of Kaiserslautern, Kaiserslautern, Germany

⁺ These authors contributed equally to this work.

*Corresponding Author: eills@soton.ac.uk

1. Thermal equilibrium ¹H spectra

Figure S1 shows thermal ¹H reference spectra after *trans*-hydrogenation for a sample composed of 100 mM acetylenedicarboxylic acid disodium salt and 6 mM [RuCp*(MeCN)₃]PF₆ in D₂O, both with and without 100 mM sodium sulphite present in solution. To react the sample, a high-pressure NMR tube containing the sample solution was pressurised with 5 bar of hydrogen gas and heated to 55°C. The sample tube was shaken vigorously by hand for 300 s. It can be observed that without sodium sulphite (Fig. S5a), the ratio of fumarate:maleate:succinate is 1:0.25:0.18. With 100 mM sodium sulphite in the sample solution (Fig. S5b), the ratio is 1:0:0.09 (i.e. there is no detectable maleate in the 32 transient ¹H NMR spectrum).



Figure S1: Thermal ¹H NMR spectra after trans-hydrogenation. (a) Reaction solution: 250mM acetylenedicarboxylic acid disodium salt, 6mM [RuCp*(MeCN)₃]PF₆ in D₂O. (b) Reaction solution: as in (a) but with 100mM ammonium sulphite.

2. Relaxation data

Proton and carbon relaxation times were measured on [1-13C]fumarate. The molecule is shown in Figure S2.



Figure S2: [1-13C]fumarate.

The ¹H T₁ was measured by inversion recovery, and the data is shown in Figure S3 along with the pulse sequence.



Figure S3: Inversion recovery pulse sequence (top) and the ${}^{1}H T_{1}$ data (bottom).

The ${}^{13}CT_1$ was measured by saturation recovery, and the data is shown in Figure S4 along with the pulse sequence.



The ¹H T_s was measured by the pulse sequence shown in Figure S5. Firstly, proton magnetization is converted into singlet order through the differential *J*-coupling to the ¹³C spin, using the M2S pulse sequence. Then, a variable delay allows the singlet order to relax. This is followed by a T₀₀ filter [1], which removes spin operators higher than rank-0. Finally, the S2hM sequence is applied to the ¹³C channel to convert proton singlet order into ¹³C magnetization for detection.

For the M2S and S2hM sequences, the values $\tau = 15.8$ ms and n = 7 were used.



Figure S5: Pulse sequence used to measure the ${}^{1}H$ T_s (top left) and the data (bottom left). The individual components of the pulse sequence are shown to the right. Pulse and receiver phases are included, and MA stands for magic angle.

The T_{00} filter parameters are given in the table below.

PFG	Shape	Strength / Gcm ⁻¹	Duration / ms
G_1	SINE.100	10	8.8
G ₂	SINE.100	-10	4.8
G₃	SINE.100	-15	4.0

[1] Tayler M. C. D.; Levitt M. H. J. Am. Chem. Soc. 2013, 135, 2120-2123