

Does Deamidation of Islet Amyloid Polypeptide Accelerate Amyloid Fibril Formation?

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Electronic Supplementary Information

Experimental Procedure

Sample preparation for deamidated hIAPP and fibrils. Wild-type hIAPP lyophilized powder (Sigma Aldrich Company Ltd, Dorset, England) was dissolved in Milli-Q (Direct-Q® 3 UV System, Millipore Corporation, US) H₂O (~pH 7.5) at a concentration of 500 μM and further diluted into 10 μM solution. The 10 μM hIAPP aqueous solution was incubated for 28 days (4 weeks). The incubated solution was then centrifuged at 14,000 rpm for one hour to separate the soluble hIAPP (supernatant) from the insoluble hIAPP fibril (fibrillary pellet). The supernatant solution containing soluble hIAPP was then diluted 20-fold with 49.5:49.5:1 water/acetonitrile/formic acid prior to MS analysis. hIAPP fibrillary pellets from 7-, 14-, 21-, and 28-day solutions were rinsed with 100 μL Milli-Q H₂O three times and then re-dissolved with 20 μL of 47.5:47.5:5 water/acetonitrile/formic acid, and sonicated in water bath at 37 °C for one hour. Re-dissolved samples were further diluted with 80 μL of 50:50 water/acetonitrile. The final concentration of formic acid in solutions was 1% (mol/mol).

Sample preparation for seeding mutant peptides. Wild-type hIAPP lyophilized powder and synthetic mutant hIAPPs (Pepscan Company Ltd, The Netherlands) were dissolved in Milli-Q H₂O to a concentration of 500 μM. The seeding experiments were performed by mixing wild-type hIAPP stock solution with 5%, 10%, 25%, or 50% mutant ((D)₃hIAPP) or ((isoD)₃hIAPP) solutions. Samples were then diluted to a final concentration of 10 μM wild-type hIAPP plus mutant ((D)₃hIAPP) or ((isoD)₃hIAPP), which were incubated for 1 week at 37°C. The supernatants and fibrillary pellets were separated and prepared as mentioned above.

FTICR MS analysis. Mass spectra were acquired on a 12 tesla solarix FTICR MS (Bruker Daltonik GmbH, Bremen, Germany). All samples were analyzed in positive ionization mode. For the detection of deamidated hIAPP and dissociated fibrils, an Apollo II electrospray ionization (ESI) source (Bruker Daltonik GmbH, Bremen, Germany) was used with a capillary voltage of 4-4.5 kV. The ESI flow rate was optimized to 100-150 $\mu\text{L}/\text{h}$ and the source temperature was set to 200°C. Ions were externally accumulated in a hexapole collision cell before transferred to an infinity cell (ICR cell) for excitation and detection.¹ Data obtained from FTICR-MS were analyzed using Bruker DataAnalysis 4.2 software (Bruker Daltonics, Bremen, Germany). For the CAD experiments, precursor ions were first isolated using the quadrupole mass filter, then collided with argon gas and accumulated in the collision cell. The collision energy was optimized to 2-18 V and the ion accumulation time to 1-3 seconds. Fragments were then transferred to the infinity cell for detection. The most intense isotopic peak from each fragment with signal-to-noise ratio (S/N) over 5 was manually matched with the theoretical m/z . All of the fragments were internally calibrated and then assigned with an uncertainty less than 1 part-per-million (ppm).

Quantification of deamidated/ mutant hIAPP in solutions and fibrillary pellets.

The monoisotopic peak area of non-deamidated and deamidated/mutant hIAPP peaks were measured using Bruker DataAnalysis 4.2 software. The percentage of deamidated hIAPP (%) was calculated as follows:

$$\text{Deamidated (\%)} = \frac{\text{Peak area of deamidated hIAPP}}{\text{Sum of Peak area of deamidated hIAPP and nondeamidated hIAPP}} \times 100\%$$

The same calculation method was applied to obtain the percentage of mutant hIAPP against wild-type hIAPP.

Transmission electron microscopy (TEM). The TEM images of the incubated solutions, including 10 μ M wild-type hIAPP, 10 μ M mutant ((D)₃hIAPP), and 10 μ M mutant ((isoD)₃hIAPP) solution were acquired on a Jeol 2010F TEM operated at 200 kV. 10 μ L of incubated solution was transferred onto a carbon-coated grid and settled for one minute, followed by removing the excess solution using filter paper. A 2% (w/v) uranyl acetate solution was used for the negative stain. Multiple images with magnification from x10,000 to x40,000 were acquired.

Thioflavin T (ThT) fluorescence reactivity. The fluorescence reactivity of the 10 μ M hIAPP, mutant ((D)₃hIAPP), mutant ((isoD)₃hIAPP), and hIAPP seeding solutions were measured using a GloMax®-Multi Detection System (Promega; Wisconsin, USA). All samples were placed in a black 96 well-plate and mixed with 10 μ M ThT aqueous solution. Fluorescence spectra of the samples were acquired every 45 minutes with excitation at 405 nm and emission measurement at 490 nm, in a similar fashion to Chan *et al.*^{2, 3} The intensities obtained from the fluorescence spectrometer were normalized to the signal intensity of the most mature fibril.

Deamidated (D)₃hIAPP: $\overset{\text{---}}{\text{KCNTATCATQRLANFLVHSS DDFGAILSST NVGSDTY-NH}_2}$

Deamidated (isoD)₃hIAPP: $\overset{\text{---}}{\text{KCNTATCATQRLANFLVHSS (isoD)(isoD)FGAILSST NVGS(isoD)TY-NH}_2}$

Figure S1. Sequences of synthetic mutant hIAPPs. Asn residues at position 21, 22, and 35 are replaced with aspartic acid – deamidated ((D)₃hIAPP) or iso-aspartic acid – deamidated ((isoD)₃hIAPP) in order to act as deamidation mimics.

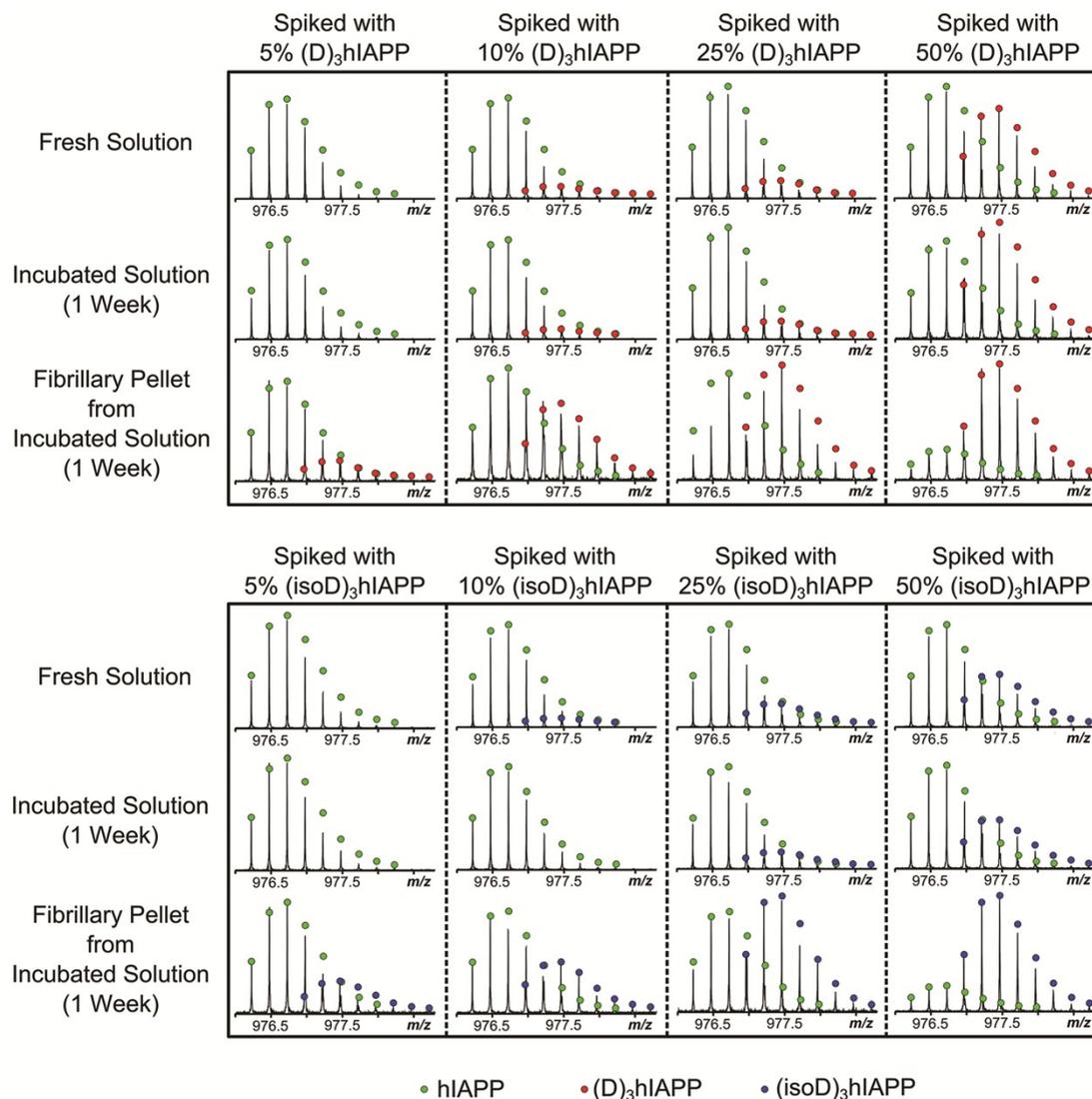


Figure S4. The nESI-MS spectra showing the fresh, the incubated solutions, and the incubated fibrillary pellets of hIAPP mixed with 5%, 10%, 25%, or 50% of mutant ((D)₃hIAPP) or ((isoD)₃hIAPP).

References

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2. F. T. S. Chan, G. S. K. Schierle, J. R. Kumita, C. W. Bertocini, C. M. Dobson and C. F. Kaminski, *Analyst*, 2013, **138**, 2156-2162.
3. T. Yang, X. H. Wang, C. L. Zhang, X. Ma, K. Wang, Y. Q. Wang, J. Luo, L. Yang, C. Yao and X. Y. Wang, *Chem. Commun.*, 2016, **52**, 2245-2248.