

Supporting Information

Synthesis of Chiral Chromanols via a RuPHOX-Ru Catalyzed Asymmetric Hydrogenation of Chromones

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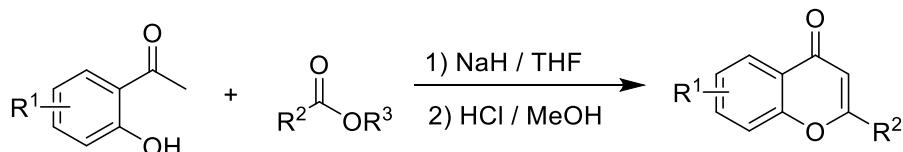
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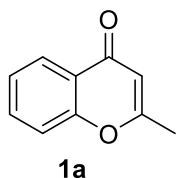
1. General Details

All reactions were performed in flame-dried glassware under an atmosphere of dry nitrogen, and the workup was carried out in air, unless otherwise noted. Solvents were dried and degassed by standard procedures. All commercially available reagents were used without further purification. ^1H NMR spectra were recorded on a Bruker AscendTM 400 (400 MHz), and Bruker AscendTM 500 (500 MHz). ^{13}C NMR spectra were recorded on a Bruker AscendTM 400 (100 MHz), and Bruker AscendTM 500 (125 MHz). Enantioselectivity was measured by high performance liquid chromatography (HPLC) using Daicel Chiralcel OD-H, AD-H and AS-H columns with *n*-hexane/*i*-PrOH as an eluent. Column chromatography was performed using 100 – 200 mesh silica gel.

2. Synthesis and Characterization of Chromones^[1]

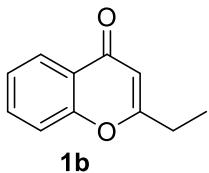


General procedure: Sodium hydride (60% dispersion in mineral oil, 3.53 g, 88.2 mmol) was rinsed three times with hexane and suspended in THF (9 mL). A mixture of 2-hydroxyacetophenone (22.0 mmol) and corresponding ester (55.1 mmol) in THF (2.5 mL) was added dropwise to the above suspension at room temperature. A vigorous reaction was observed, and the temperature rose to reflux. After complete addition, the reaction mixture was stirred for a further 5 min, quenched by pouring onto ice and further acidified to pH 6 with 6 M HCl (aq). The solution was extracted with EtOAc (10 mL×3) and the combined organic layers were washed with brine and dried over MgSO₄. The solvent was evaporated to yield the crude 1,3-dione. Without further purification, a solution of the crude 1,3-dione product (2.50 g, 14.0 mmol) in methanol (30 mL) was treated conc. HCl (1 mL), and the mixture was allowed to stirring at room temperature for 14 h. The mixture was concentrated under reduced pressure and the residue was diluted with ethyl acetate (50 mL), then washed successively with solutions of saturated NaHCO₃ (aq), water, and brine. The organic layer was then dried over MgSO₄ and concentrated under reduced pressure. The residue was purified via flash column chromatography (silica gel, ethyl acetate/hexanes 1:3 as eluent) to give the desired products **1**.

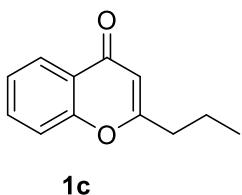


2-Methyl-4*H*-chromen-4-one (1a**):**^[2] A white solid (1.68 g, 79%). ^1H NMR (400 MHz, CDCl₃) δ 8.18

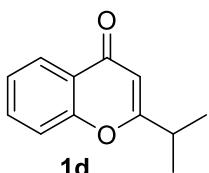
(d, $J = 7.6$ Hz, 1H), 7.64 (t, $J = 8.0$ Hz, 1H), 7.43–7.36 (m, 2H), 6.17 (s, 1H), 2.39 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 178.2, 166.1, 156.5, 133.4, 125.6, 124.9, 123.5, 117.7, 110.5, 20.5.



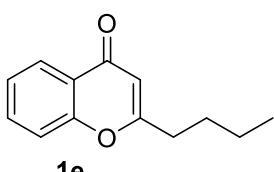
2-Ethyl-4H-chromen-4-one (1b):^[2] A white oil (1.4 g, 81%). ^1H NMR (400 MHz, CDCl_3) δ 8.17 (d, $J = 7.6$ Hz, 1H), 7.62 (t, $J = 7.6$ Hz, 1H), 7.41 (d, $J = 8.4$ Hz, 1H), 7.36 (t, $J = 7.6$ Hz, 1H), 6.17 (s, 1H), 2.65 (q, $J = 7.6$ Hz, 2H), 1.30 (t, $J = 7.6$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 178.4, 170.8, 156.5, 133.4, 125.6, 124.8, 123.7, 117.8, 108.9, 27.5, 10.9.



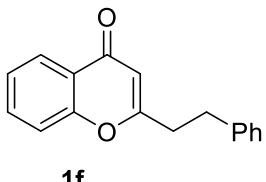
2-Propyl-4H-chromen-4-one (1c):^[3] A white oil (1.2 g, 66%). ^1H NMR (400 MHz, CDCl_3) δ 8.17 (d, $J = 8.0$ Hz, 1H), 7.63 (t, $J = 7.6$ Hz, 1H), 7.41 (d, $J = 8.4$ Hz, 1H), 7.36 (t, $J = 7.6$ Hz, 1H), 6.17 (s, 1H), 2.59 (t, $J = 7.6$ Hz, 2H), 1.82–1.72 (m, 2H), 1.02 (t, $J = 7.6$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 178.3, 169.5, 156.5, 133.4, 125.6, 124.8, 123.7, 117.8, 109.9, 36.2, 20.2, 13.5.



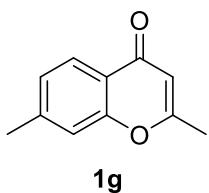
2-Isopropyl-4H-chromen-4-one (1d):^[2] A white oil (0.9 g, 69%). ^1H NMR (400 MHz, CDCl_3) δ 8.17 (d, $J = 8.0$ Hz, 1H), 7.65–7.61 (m, 1H), 7.43 (d, $J = 8.4$ Hz, 1H), 7.36 (t, $J = 7.6$ Hz, 1H), 6.19 (s, 1H), 2.91–2.81 (m, 1H), 1.32 (d, $J = 7.2$ Hz, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 178.7, 174.2, 156.5, 133.4, 125.6, 124.8, 123.7, 117.8, 107.5, 33.2, 20.1.



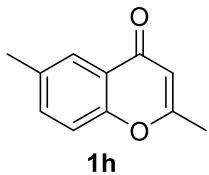
2-Butyl-4H-chromen-4-one (1e):^[2] A white oil (1.3 g, 78%). ^1H NMR (400 MHz, CDCl_3) δ 8.17 (dd, $J = 1.2, 8.0$ Hz, 1H), 7.65–7.60 (m, 1H), 7.41 (d, $J = 8.4$ Hz, 1H), 7.36 (t, $J = 7.2$ Hz, 1H) 6.17 (s, 1H), 2.61 (t, $J = 7.6$ Hz, 2H), 1.75–1.68 (m, 2H), 1.47–1.38 (m, 1H), 0.95 (t, $J = 7.6$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 178.3, 169.8, 156.5, 133.3, 125.6, 124.8, 123.7, 117.8, 109.7, 34.0, 28.8, 22.1, 13.7.



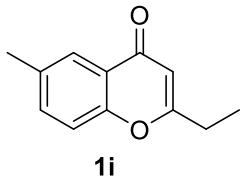
2-Phenethyl-4H-chromen-4-one (1f):^[2] A white solid (1.8 g, 88%). ¹H NMR (400 MHz, CDCl₃) δ 8.17 (d, *J* = 7.6 Hz, 1H), 7.65 (t, *J* = 7.2 Hz, 1H), 7.42 (d, *J* = 8.0 Hz, 1H), 7.37 (t, *J* = 7.6 Hz, 1H), 7.28 (q, *J* = 7.2 Hz, 2H), 7.21 (t, *J* = 6.8 Hz, 3H), 6.15 (s, 1H), 3.06 (t, *J* = 7.6 Hz, 2H), 2.93 (t, *J* = 8.0 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 178.2, 168.4, 156.4, 139.7, 133.5, 128.6, 128.2, 126.5, 125.7, 124.9, 123.7, 117.8, 110.2, 36.1, 33.0.



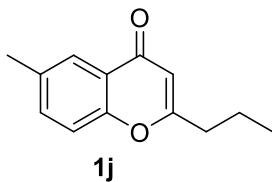
2,7-Dimethyl-4H-chromen-4-one (1g):^[2] A yellow solid (1.2 g, 66%). ¹H NMR (400 MHz, CDCl₃) δ 8.05 (d, *J* = 8.0 Hz, 1H), 7.20–7.17 (m, 2H), 6.13 (s, 1H), 2.47 (s, 3H), 2.36 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 178.5, 166.1, 156.8, 144.9, 126.6, 125.6, 121.5, 117.8, 110.7, 22.0, 20.8.



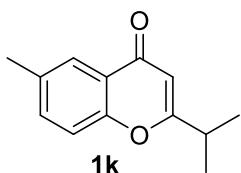
2,6-Dimethyl-4H-chromen-4-one (1h):^[2] A yellow solid (1.8 g, 76%). ¹H NMR (400 MHz, CDCl₃) δ 7.91 (s, 1H), 7.39 (d, *J* = 8.8 Hz, 1H), 7.25 (d, *J* = 8.4 Hz, 1H), 6.09 (s, 1H), 2.38 (s, 3H), 2.32 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 178.5, 166.2, 154.9, 135.0, 134.8, 125.1, 123.4, 117.7, 110.6, 21.1, 20.8.



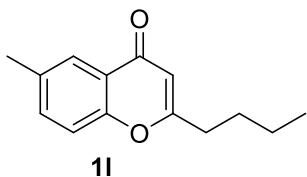
2-Ethyl-6-methyl-4H-chromen-4-one (1i): A white solid (1.4 g, 95%). Mp 44–45 °C; ¹H NMR (500 MHz, CDCl₃) δ 7.95 (d, *J* = 5.0 Hz, 1H), 7.43 (dd, *J* = 2.0, 8.5 Hz, 1H), 7.31 (d, *J* = 8.5 Hz, 1H), 6.16 (s, 1H), 2.64 (q, *J* = 7.5 Hz, 2H), 2.43 (s, 3H), 1.30 (t, *J* = 7.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 178.6, 170.7, 154.8, 134.8, 134.6, 125.0, 123.3, 117.6, 108.7, 27.5, 20.9, 11.0; IR (KBr) cm⁻¹: 3041, 2934, 1701, 1681, 1611, 1580, 1502, 1462, 813; HRMS (ESI): calcd for C₁₂H₁₂O₂ [M+H]⁺: 189.0916, found 189.0923.



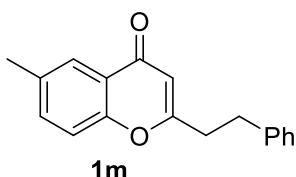
6-Methyl-2-propyl-4H-chromen-4-one (1j): Yellow oil (1.5 g, 79%). ^1H NMR (400 MHz, CDCl_3) δ 7.96 (d, $J = 1.2$ Hz, 1H), 7.44 (dd, $J = 6.0, 8.4$ Hz, 1H), 7.31 (d, $J = 8.8$ Hz, 1H), 6.15 (s, 1H), 2.58 (t, $J = 7.2$ Hz, 2H), 2.43 (s, 3H), 1.81–1.72 (m, 2H), 1.01 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 178.5, 169.4, 154.8, 134.8, 134.6, 125.0, 123.3, 117.6, 109.7, 36.2, 20.9, 20.2, 13.5; IR (KBr) cm^{-1} : 3022, 2944, 1721, 1681, 1601, 1580, 1501, 1464, 810; HRMS (ESI): calcd for $\text{C}_{13}\text{H}_{14}\text{O}_2$ [$\text{M}+\text{H}]^+$: 203.1072, found 203.1079.



2-Isopropyl-6-methyl-4H-chromen-4-one (1k): Yellow oil (3.0 g, 88%). ^1H NMR (400 MHz, CDCl_3) δ 7.99 (d, $J = 0.8$ Hz, 1H), 7.47 (dd, $J = 2.0, 8.4$ Hz, 1H), 7.36 (d, $J = 8.8$ Hz, 1H), 6.20 (s, 1H), 2.93–2.83 (m, 1H), 2.46 (s, 3H), 1.34 (d, $J = 7.2$ Hz, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 178.9, 174.0, 154.8, 134.7, 134.6, 125.0, 123.4, 117.6, 107.4, 33.2, 20.9, 20.1; IR (KBr) cm^{-1} : 3010, 2904, 1701, 1682, 1589, 1580, 1501, 1454, 1385, 1368, 800; HRMS (ESI): calcd for $\text{C}_{13}\text{H}_{14}\text{O}_2$ [$\text{M}+\text{H}]^+$: 203.1072, found 203.1070.

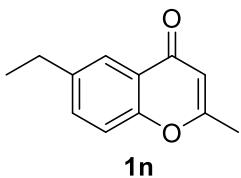


2-Butyl-6-methyl-4H-chromen-4-one (1l):^[4] Yellow oil (1.3 g, 72%). ^1H NMR (400 MHz, CDCl_3) δ 7.96 (d, $J = 1.2$ Hz, 1H), 7.44 (dd, $J = 2.4, 8.8$ Hz, 1H), 7.31 (d, $J = 8.4$ Hz, 1H), 6.15 (s, 1H), 2.61 (t, $J = 7.6$ Hz, 2H), 2.44 (s, 3H), 1.75–1.67 (m, 2H), 1.47–1.38 (m, 2H), 0.96 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 178.5, 169.7, 154.8, 134.8, 134.6, 125.0, 123.4, 117.6, 109.6, 34.0, 28.8, 22.1, 13.7.

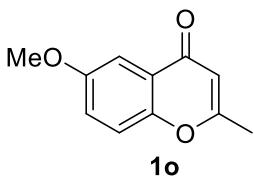


6-Methyl-2-phenethyl-4H-chromen-4-one (1m):^[5] A white powder (1.3 g, 68%). ^1H NMR (400 MHz, CDCl_3) δ 7.96 (d, $J = 1.2$ Hz, 1H), 7.46 (dd, $J = 2.4, 8.8$ Hz, 1H), 7.33 (d, $J = 8.4$ Hz, 1H), 7.29 (t, $J = 7.2$ Hz, 2H), 7.21 (t, $J = 7.6$ Hz, 3H), 6.13 (s, 1H), 3.06 (t, $J = 7.2$ Hz, 2H), 2.92 (t, $J = 7.2$ Hz, 2H), 2.44 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 178.4, 168.3, 154.7, 139.8, 134.9, 134.7, 128.6, 126.5, 123.4, 117.6,

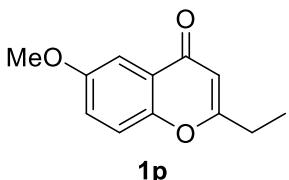
110.1, 36.1, 33.0, 20.9.



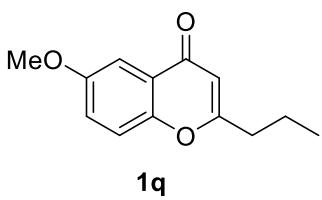
6-Ethyl-2-methyl-4H-chromen-4-one (1n): A white solid (1.6 g, 78%). Mp 63–65 °C; ^1H NMR (500 MHz, CDCl_3) δ 7.98 (s, 1H), 7.46 (dd, J = 2.0, 8.5 Hz, 1H), 7.32 (d, J = 8.5 Hz, 1H), 6.14 (s, 1H), 2.73 (q, J = 7.5 Hz, 2H), 2.36 (s, 3H), 1.26 (t, J = 7.5 Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 178.5, 166.0, 154.9, 141.2, 133.6, 123.8, 123.3, 117.6, 110.4, 28.3, 20.6, 15.5; IR (KBr) cm^{-1} : 3006, 2924, 1701, 1672, 1599, 1580, 1491, 1454, 1375, 820; HRMS (ESI): calcd for $\text{C}_{12}\text{H}_{12}\text{O}_2$ [$\text{M}+\text{H}]^+$: 189.0916, found 189.0906



6-Methoxy-2-methyl-4H-chromen-4-one (1o):^[6] A white solid (1.5 g, 72%). ^1H NMR (400 MHz, CDCl_3) δ 7.49 (d, J = 3.2 Hz, 1H), 7.29 (d, J = 9.2 Hz, 1H), 7.17 (dd, J = 3.2, 8.4 Hz, 1H), 6.11 (s, 1H), 3.84 (s, 3H), 2.33 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 178.3, 166.1, 156.9, 151.5, 124.3, 123.6, 119.4, 110.0, 105.0, 56.1, 20.7.

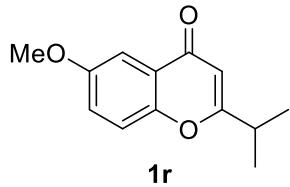


2-Ethyl-6-methoxy-4H-chromen-4-one (1p): A white solid (2.4 g, 90%). Mp 78–80 °C; ^1H NMR (400 MHz, CDCl_3) δ 7.53 (d, J = 2.8 Hz, 1H), 7.34 (d, J = 9.2 Hz, 1H), 7.21 (dd, J = 3.2, 9.2 Hz, 1H), 6.16 (s, 1H), 3.87 (s, 3H), 2.64 (q, J = 7.6 Hz, 2H), 1.29 (t, J = 7.6 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 178.3, 170.5, 156.7, 151.3, 124.2, 123.4, 119.2, 108.2, 104.8, 55.9, 27.5, 11.0; IR (KBr) cm^{-1} : 3001, 2944, 1721, 1652, 1619, 1580, 1501, 1454, 1371, 836; HRMS (ESI): calcd for $\text{C}_{12}\text{H}_{12}\text{O}_3$ [$\text{M}+\text{H}]^+$: 205.0865, found 205.0867.

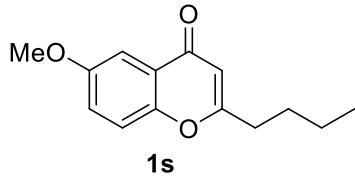


6-Methoxy-2-propyl-4H-chromen-4-one (1q): A white solid (1.9 g, 88%). Mp 60–61 °C; ^1H NMR (400 MHz, CDCl_3) δ 7.54 (d, J = 2.8 Hz, 1H), 7.35 (d, J = 9.2 Hz, 1H), 7.22 (dd, J = 2.8, 8.8 Hz, 1H), 6.16

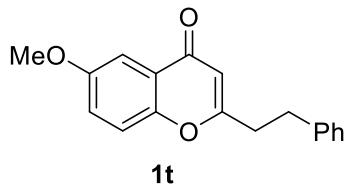
(s, 1H), 3.88 (s, 3H), 2.58 (t, J = 7.6 Hz, 2H), 1.81–1.72 (m, 2H), 1.01 (t, J = 7.2 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 178.3, 169.3, 156.7, 151.4, 124.3, 123.4, 119.2, 109.2, 104.8, 55.9, 36.2, 20.2, 13.5; IR (KBr) cm^{-1} : 3006, 2954, 1701, 1682, 1598, 1580, 1501, 1444, 1369, 812; HRMS (ESI): calcd for $\text{C}_{13}\text{H}_{14}\text{O}_3$ $[\text{M}+\text{H}]^+$: 219.1021, found 219.1023.



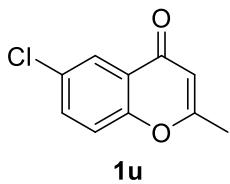
2-Isopropyl-6-methoxy-4H-chromen-4-one (1r): Yellow oil (0.8 g, 66%). ^1H NMR (400 MHz, CDCl_3) δ 7.54 (d, J = 3.2 Hz, 1H), 7.37 (d, J = 6.8 Hz, 1H), 7.22 (dd, J = 3.2, 9.2 Hz, 1H), 6.18 (s, 1H), 3.88 (s, 3H), 2.91–2.80 (m, 1H), 1.31 (t, J = 7.2 Hz, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 178.6, 173.9, 156.7, 151.3, 124.3, 123.5, 119.3, 106.8, 104.8, 55.9, 33.2, 20.2; IR (KBr) cm^{-1} : 3006, 2934, 1701, 1682, 1599, 1580, 1501, 1454, 1381, 1363, 828; HRMS (ESI): calcd for $\text{C}_{13}\text{H}_{14}\text{O}_3$ $[\text{M}+\text{H}]^+$: 219.1021, found 219.1026.



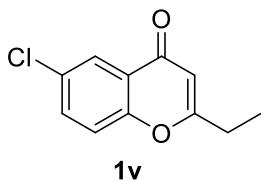
2-Butyl-6-methoxy-4H-chromen-4-one (1s):^[7] A white solid (1.7 g, 87%). ^1H NMR (400 MHz, CDCl_3) δ 7.56 (d, J = 2.8 Hz, 1H), 7.37 (d, J = 9.2 Hz, 1H), 7.24 (dd, J = 2.8, 8.8 Hz, 1H), 6.17 (s, 1H), 3.90 (s, 3H), 2.63 (t, J = 7.2 Hz, 2H), 1.77–1.69 (m, 2H), 1.49–1.39 (m, 2H), 0.97 (t, J = 7.6 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 178.2, 169.5, 156.7, 151.3, 124.3, 123.4, 119.2, 109.1, 104.8, 55.9, 34.0, 28.9, 22.1, 13.7.



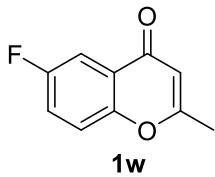
6-Methoxy-2-phenethyl-4H-chromen-4-one (1t):^[5] A white solid (1.8 g, 79%). ^1H NMR (400 MHz, CDCl_3) δ 7.56 (d, J = 3.2 Hz, 1H), 7.38 (d, J = 9.2 Hz, 1H), 7.33–7.21 (m, 6H), 6.16 (s, 1H), 3.90 (s, 3H), 3.08 (t, J = 7.2 Hz, 2H), 2.94 (t, J = 8.4 Hz, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 178.1, 168.1, 156.8, 151.3, 139.8, 128.6, 128.3, 126.5, 124.3, 123.5, 119.2, 109.5, 104.9, 55.9, 36.1, 33.0.



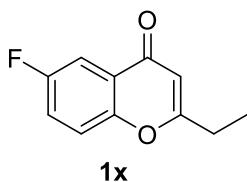
6-Chloro-2-methyl-4H-chromen-4-one (1u):^[8] A white solid (2.4 g, 96%). ¹H NMR (500 MHz, CDCl₃) δ 8.13 (d, *J* = 1.5 Hz, 1H), 7.57 (dd, *J* = 2.5, 9.0 Hz, 1H), 7.37 (d, *J* = 9.0 Hz, 1H), 6.17 (s, 1H), 2.38 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 177.0, 166.5, 154.8, 133.6, 130.9, 125.1, 124.5, 119.5, 110.5, 20.6.



6-Chloro-2-ethyl-4H-chromen-4-one (1v):^[8] A white solid (2.4 g, 88%). ¹H NMR (500 MHz, CDCl₃) δ 8.13 (d, *J* = 2.5 Hz, 1H), 7.57 (dd, *J* = 2.5, 9.0 Hz, 1H), 7.38 (d, *J* = 8.5 Hz, 1H), 6.18 (s, 1H), 2.66 (q, *J* = 7.5 Hz, 2H), 1.31 (t, *J* = 7.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 177.2, 171.1, 154.8, 133.6, 130.8, 125.1, 124.7, 119.6, 108.9, 27.5, 10.9.

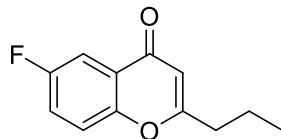


6-Fluoro-2-methyl-4H-chromen-4-one (1w): A white solid (1.3 g, 70%). Mp 95–96 °C; ¹H NMR (500 MHz, CDCl₃) δ 7.80 (dd, *J* = 2.0, 3.0 Hz, 1H), 7.41 (dd, *J* = 3.5, 8.5 Hz, 1H), 7.37–7.33 (m, 1H), 6.15 (s, 1H), 2.38 (s, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 177.4 (d, *J* = 2.1 Hz), 166.5, 159.4 (d, *J* = 244.8 Hz), 152.7 (d, *J* = 1.6 Hz), 124.7 (d, *J* = 7.3 Hz), 121.6 (d, *J* = 25.3 Hz), 119.8 (d, *J* = 8.0 Hz), 110.5 (d, *J* = 23.5 Hz), 109.9, 20.6; ¹⁹F NMR (470 MHz, CDCl₃) δ -115.7; IR (KBr) cm⁻¹: 3016, 2954, 1701, 1662, 1601, 1580, 1491, 1454, 1379, 1280, 812; HRMS (ESI): calcd for C₁₀H₇FO₂ [M+H]⁺: 179.0508, found 179.0516.



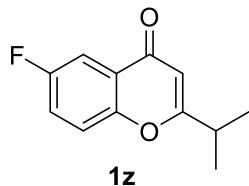
2-Ethyl-6-fluoro-4H-chromen-4-one (1x): A white solid (2.4 g, 85%). Mp 50–52 °C; ¹H NMR (500 MHz, CDCl₃) δ 7.84 (dd, *J* = 4.0, 10.5 Hz, 1H), 7.46 (dd, *J* = 5.5, 11.5 Hz, 1H), 7.41–7.36 (m, 1H), 6.21 (s, 1H), 2.69 (q, *J* = 9.0 Hz, 2H), 1.62 (s, 3H), 1.34 (t, *J* = 9.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 177.7, 171.1, 159.4 (d, *J* = 244.9 Hz), 152.7, 124.9 (d, *J* = 7.3 Hz), 121.6 (d, *J* = 25.3 Hz), 119.9 (d, *J* = 8.0 Hz),

110.5 (d, $J = 23.5$ Hz), 108.3, 27.5, 10.9; ^{19}F NMR (470 MHz, CDCl_3) δ -115.7; IR (KBr) cm^{-1} : 3001, 2954, 1701, 1652, 1598, 1580, 1491, 1460, 1375, 1280, 822; HRMS (ESI): calcd for $\text{C}_{11}\text{H}_9\text{FO}_2$ $[\text{M}+\text{H}]^+$: 193.0665, found 193.0667.

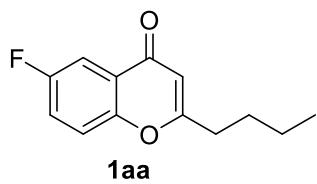


1y

6-Fluoro-2-propyl-4H-chromen-4-one (1y): A white solid (1.5 g, 80%). Mp 36–38 °C; ^1H NMR (400 MHz, CDCl_3) δ 7.82 (dd, $J = 3.2, 8.4$ Hz, 1H), 7.44 (dd, $J = 4.0, 8.8$ Hz, 1H), 7.39–7.34 (m, 1H), 6.18 (s, 1H), 2.61 (t, $J = 7.2$ Hz, 2H), 1.83–1.74 (m, 2H), 1.04 (t, $J = 7.6$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 177.5 (d, $J = 2.2$ Hz), 169.9, 159.4 (d, $J = 244.7$ Hz), 152.7 (d, $J = 1.7$ Hz), 124.9 (d, $J = 7.3$ Hz), 121.5 (d, $J = 25.2$ Hz), 119.9 (d, $J = 8.0$ Hz), 110.5 (d, $J = 23.4$ Hz), 109.2, 36.2, 20.2, 13.5; ^{19}F NMR (376 MHz, CDCl_3) δ -115.7; IR (KBr) cm^{-1} : 3012, 2944, 1711, 1658, 1601, 1580, 1501, 1454, 1380, 1280, 823; HRMS (ESI): calcd for $\text{C}_{12}\text{H}_{11}\text{FO}_2$ $[\text{M}+\text{H}]^+$: 207.0821, found 207.0826.

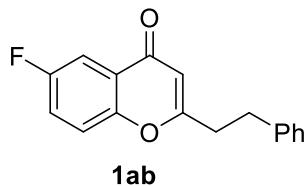


6-Fluoro-2-isopropyl-4H-chromen-4-one (1z): A white solid (1.4 g, 76%). Mp 51–52 °C; ^1H NMR (400 MHz, CDCl_3) δ 7.83 (dd, $J = 3.2, 8.4$ Hz, 1H), 7.46 (dd, $J = 0.4, 8.8$ Hz, 1H), 7.41–7.36 (m, 1H), 6.21 (s, 1H), 2.94–2.84 (m, 1H), 1.34 (d, $J = 7.2$ Hz, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 177.8 (d, $J = 2.2$ Hz), 174.4, 159.4 (d, $J = 244.7$ Hz), 152.7, 124.9 (d, $J = 7.3$ Hz), 121.5 (d, $J = 25.3$ Hz), 119.9 (d, $J = 8.0$ Hz), 110.5 (d, $J = 23.5$ Hz), 107.0, 33.3, 20.1; ^{19}F NMR (376 MHz, CDCl_3) δ -115.8; IR (KBr) cm^{-1} : 3007, 2924, 1699, 1668, 1601, 1580, 1500, 1454, 1385, 1369, 1280, 812; HRMS (ESI): calcd for $\text{C}_{12}\text{H}_{11}\text{FO}_2$ $[\text{M}+\text{H}]^+$: 207.0821, found 207.0818.

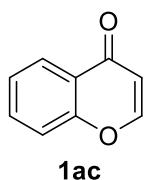


2-Butyl-6-fluoro-4H-chromen-4-one (1aa): Yellow oil (1.7 g, 87%). ^1H NMR (400 MHz, CDCl_3) δ 7.80 (dd, $J = 2.8, 8.0$ Hz, 1H), 7.41 (dd, $J = 4.0, 8.8$ Hz, 1H), 7.34 (td, $J = 3.2, 8.0$ Hz, 1H), 6.15 (s, 1H), 2.61 (t, $J = 7.6$ Hz, 2H), 1.75–1.67 (m, 2H), 1.47–1.37 (m, 2H), 0.95 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 177.5, 170.1, 159.4 (d, $J = 244.9$ Hz), 152.7, 124.9 (d, $J = 7.3$ Hz), 121.5 (d, $J = 25.3$ Hz),

119.9 (d, $J = 7.0$ Hz), 110.5 (d, $J = 23.5$ Hz), 109.1, 34.0, 28.8, 22.1, 13.7; ^{19}F NMR (376 MHz, CDCl_3) δ -115.7; IR (KBr) cm^{-1} : 3006, 2934, 1701, 1632, 1601, 1580, 1498, 1454, 1382, 1280, 832; HRMS (ESI): calcd for $\text{C}_{13}\text{H}_{13}\text{FO}_2$ [$\text{M}+\text{H}]^+$: 221.0978, found 221.0981.



6-Fluoro-2-phenethyl-4H-chromen-4-one (1ab): A white solid (1.7 g, 79%). Mp 65–66 °C; ^1H NMR (400 MHz, CDCl_3) δ 7.84 (dd, $J = 2.8, 8.0$ Hz, 1H), 7.46 (dd, $J = 4.0, 8.8$ Hz, 1H), 7.42–7.37 (m, 1H), 7.34–7.21 (m, 5H), 6.16 (s, 1H), 3.08 (t, $J = 6.8$ Hz, 2H), 2.96 (t, $J = 8.0$ Hz, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 177.4 (d, $J = 2.2$ Hz), 168.7, 159.4 (d, $J = 245.0$ Hz), 152.7, 139.6, 128.5 (d, $J = 43.3$ Hz), 126.6, 124.9 (d, $J = 7.3$ Hz), 121.6 (d, $J = 25.3$ Hz), 119.9 (d, $J = 8.0$ Hz), 110.6 (d, $J = 23.4$ Hz), 109.6, 36.1, 33.0; ^{19}F NMR (376 MHz, CDCl_3) δ -115.5; IR (KBr) cm^{-1} : 3008, 2914, 1711, 1652, 1601, 1580, 1491, 1454, 1384, 1280, 822; HRMS (ESI): calcd for $\text{C}_{17}\text{H}_{13}\text{FO}_2$ [$\text{M}+\text{H}]^+$: 269.0978, found 269.0986.



4H-Chromen-4-one (1ac):^[1b] A white solid (1.6 g, 73%). ^1H NMR (400 MHz, CDCl_3) δ 8.16 (dd, $J = 0.8, 8.0$ Hz, 1H), 7.82 (d, $J = 6.0$ Hz, 1H), 7.65–7.60 (m, 1H), 7.41 (d, $J = 8.4$ Hz, 1H), 7.38–7.34 (m, 1H), 6.30 (d, $J = 6.0$ Hz, 1H); ^{13}C NMR (100 MHz, CDCl_3) δ 177.6, 156.5, 155.3, 133.7, 125.7, 125.2, 124.8, 118.1, 112.9.

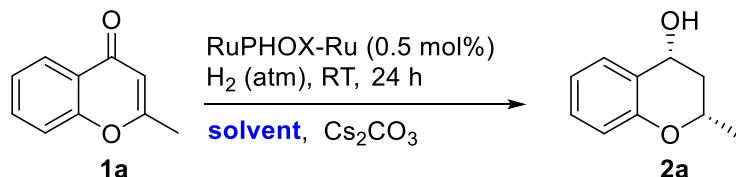
3. Asymmetric Synthesis of Chromanols 2

3.1 Solvent and Hydrogen Pressure Screening

As shown in Table 1, MeOH was first used as a solvent and the desired product (*2S,4R*)-2-methyl-4*H*-chroman-4-ol (chromanol **2a**) was obtained in full conversion and 99% ee (entry 1). The use of EtOH provided the same conversion but a slightly lower enantioselectivity than that of MeOH (entry 2). However, almost no reaction occurred when *i*-PrOH was used as a solvent (entry 3). As a comparison, aprotic solvents, such as DCM and toluene, were used and both provided the desired product **2a** with excellent enantioselectivities (entries 4 and 5). However, only 90% conversion was obtained when the reaction was conducted in DCM (entry 5). Next, the asymmetric hydrogenation was carried out under a low hydrogen pressure in either MeOH or toluene as a solvent. Similar conversion and enantioselectivities

were obtained when 20 bar hydrogen pressure was used (entries 6 and 7). However, only 94% conversion was observed in MeOH and only a trace amount of product was obtained in toluene when the reaction was carried out under a hydrogen pressure of 10 bar (entries 8 and 9).

Table S1 Solvent and hydrogen pressure screening.^a



Entry	Solvent	H ₂ (bar)	Conv (%) ^b	ee (%) ^c	dr ^d
1	MeOH	50	>99	99	>20:1
2	EtOH	50	>99	98	>20:1
3	i-PrOH	50	trace	-	-
4	toluene	50	>99	99.6	>20:1
5	DCM	50	90	99.9	>20:1
6	MeOH	20	>99	98	>20:1
7	toluene	20	>99	99.7	>20:1
8	MeOH	10	94	97	>20:1
9	toluene	10	trace	-	-

^a Conditions: **1a** (0.30 mmol), (S,Sp)-RuPHOX-Ru (0.5 mol %) and Cs₂CO₃ (0.5 equiv) in a suitable solvent (3 mL) under a certain hydrogen pressure at rt for 12 h.

^b By using ¹H NMR analysis of the crude product to determine the consumption of **1a**

and the formation of **2a** by comparison of ¹H NMR integrals corresponding to methyl groups.

^c Determined by chiral HPLC analysis of **2a** using an OD-H column; The absolute configuration of **2a** was determined by single crystal.

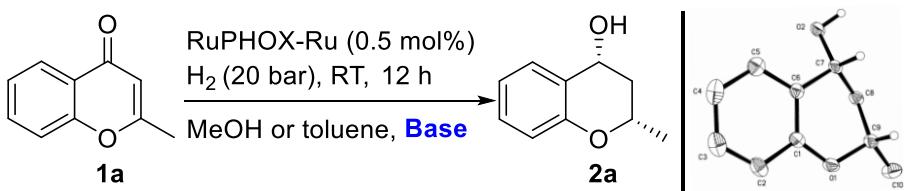
^d Determined by ¹H NMR.

3.2 Base Screening

Subsequently, we chose both MeOH and toluene as the solvents to examine the impact of different bases on the reaction when it was carried out at 20 bar hydrogen pressure over 12 h (Table S2). First, the reaction was carried out with MeOH as a solvent. Three frequently used sodium containing bases and salts were first examined with Na₂CO₃ giving the best results (entries 1-3). As a comparison, potassium containing bases and salts were also examined, and somewhat inferior results were obtained (entries 4-6). The organic base DIPEA was also examined and **2a** was obtained in 91% conversion and with 95% ee (entry 7). Next, the above reactions were carried out using toluene as a solvent instead of MeOH (entries 8-14). It was found that the reaction activity in toluene was affected significantly by base. Full conversions and excellent enantioselectivities were obtained when the reactions were carried out in the presence of KOH or K₂CO₃ (entries 11 and 12). Considering reaction activity, subsequent reactions were carried out

using Na_2CO_3 as a suitable base in MeOH under 20 bar hydrogen pressure at room temperature for 12 h (entry 2).

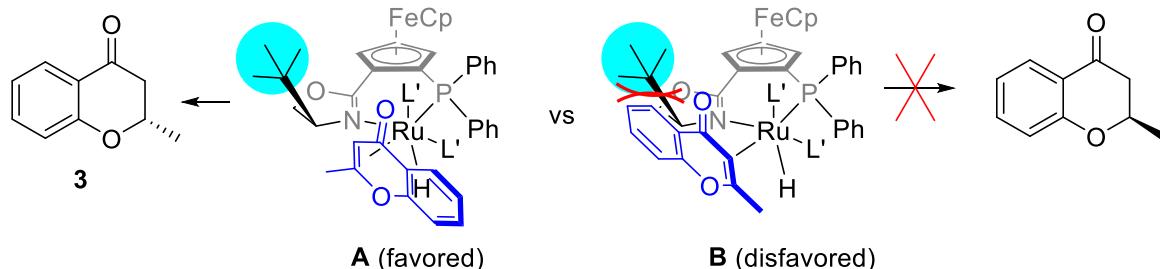
Table S2 Screening of base.^a



Entry	Solvent	Base	Conv (%) ^b	ee (%) ^c	<i>dr</i> ^d
1	MeOH	NaOH	>99	97	>20:1
2	MeOH	Na_2CO_3	>99	99.9	>20:1
3	MeOH	NaHCO_3	95	98	>20:1
4	MeOH	KOH	>99	97	>20:1
5	MeOH	K_2CO_3	>99	97	>20:1
6	MeOH	<i>t</i> -BuOK	89	95	>20:1
7	MeOH	DIPEA	91	95	>20:1
8	toluene	NaOH	48	99.6	>20:1
9	toluene	Na_2CO_3	NR	-	-
10	toluene	NaHCO_3	NR	-	-
11	toluene	KOH	>99	99.7	>20:1
12	toluene	K_2CO_3	>99	99.4	>20:1
13	toluene	<i>t</i> -BuOK	NR	-	-
14	toluene	DIPEA	NR	-	-

^a Conditions: **1a** (0.30 mmol), (*S*,*S*)-RuPHOX-Ru (0.5 mol%) and base (0.5 equiv) in MeOH or toluene (3 mL) under a 20 bar hydrogen pressure at rt for 12 h. ^b By using ¹H NMR analysis of the crude product to determine the consumption of **1a** and the formation of **2a** by comparison of ¹H NMR integrals corresponding to methyl groups. ^c Determined by chiral HPLC analysis of **2a** using an OD-H column; The absolute configuration of **2a** was determined by single crystal. ^d Determined by ¹H NMR.

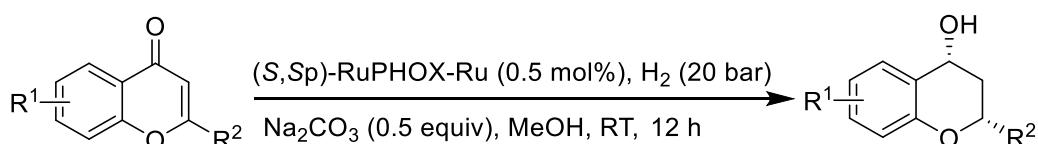
3.3 Proposed steric control pathway



According to the X-ray crystal structure of **2a** (Table S2), the chiral induction process in the asymmetric

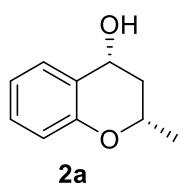
hydrogenation of C=C double bond of **1a** can be speculated. Due to the steric hindrance between the bulky *t*-Bu group of the chiral oxazoline ring and the phenyl ring of **1a**, the Ru-complex **A** is prone to form via the coordination of the Ru metal center to the C=C bond, thus leading to the major enantiomer (*S*)-**3**. As for Ru-complex **B**, the steric interaction is stronger than that in **A** and so is more difficult to form. Subsequently, **3** was further hydrogenated under the same reaction conditions, affording the terminal product **2a** with the indicated absolute configuration.

3.4 General procedure for the RuPHOX-Ru catalyzed asymmetric hydrogenation



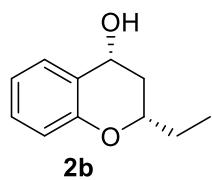
General Procedure: In a nitrogen-filled glovebox, a hydrogenation tube was charged with a stirring bar, substituted chromones (0.30 mmol), RuPHOX-Ru (2.6 mg, 0.5 mol%) and Na₂CO₃ (15.9 mg, 0.5 equiv, 0.15 mmol). MeOH (3 mL) was then injected into the hydrogenation tube by a syringe. The hydrogenation tube was then put into an autoclave. The system was evacuated and filled with hydrogen 3 times. The autoclave was then charged with hydrogen to 20 bar hydrogen pressure, and the reaction mixture was stirred at RT for 12 h before releasing the hydrogen. After the completion of reaction, the solvent was evaporated to afford the crude product. The conversion of substrate was determined by ¹H NMR analysis. HPLC with a chiral column was used to determine the ee value.

Gram scale synthesis of **2a**: To a 50 mL round-bottom flask was charged with a stir bar, compound **1a** (1.87 g, 11.7 mmol), RuPhOX-Ru (20 mg, 0.1 mol%), Na₂CO₃ (1.2 g, 5.8 mmol) in an argon-filled glovebox. Then, MeOH (20 mL) was injected into the flask by a syringe with stirring and the reaction flask was then put into an autoclave. The autoclave was evacuated and filled hydrogen for 3 times and then charged with hydrogen to 50 bar. After vigorous stirring at room temperature for 72 hours, the solvent was evaporated under reduced pressure to afford the crude product, which was determined by ¹H NMR analysis to determine the conversion of substrate. The crude product was passed through a short column of silicon (PE/EA = 20/1 as the eluent) to afford **2a** as a white solid (1.81 g, 94%, 96% ee).

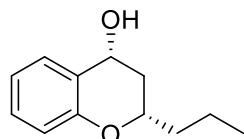


(2*S*,4*R*)-2-Methylchroman-4-ol (2a):^[2] A white solid (48.6 mg, 99%). ¹H NMR (400 MHz, CDCl₃) δ 7.46 (d, *J* = 7.6 Hz, 1H), 7.16 (t, *J* = 7.2 Hz, 1H), 6.93 (t, *J* = 7.2 Hz, 1H), 6.79 (d, *J* = 8.0 Hz, 1H),

4.95–4.89 (m, 1H), 4.29–4.21 (m, 1H), 2.86 (dd, J = 6.4, 12.8 Hz, 1H), 1.92 (d, J = 8.0 Hz, 1H), 1.78–1.69 (m, 1H), 1.42 (t, J = 6.4 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 154.5, 129.0, 126.9, 125.7, 120.6, 116.4, 71.2, 65.5, 39.8, 21.4; $[\alpha]_D^{26} = -131.82$ (c 0.50, CHCl_3); HPLC (Chiralcel OD-H, *n*-hexane/*i*-PrOH = 97/3, UV = 210 nm, flow rate = 0.5 mL/min) $t_{\text{R}1}$ = 21.982 min (minor) and $t_{\text{R}2}$ = 26.669 min (major), ee = 99.9%.

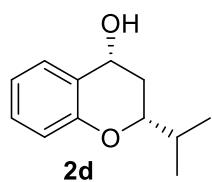


(2S,4R)-2-Ethylchroman-4-ol (2b):^[2] A white solid (52.78 mg, 99%). ^1H NMR (400 MHz, CDCl_3) δ 7.46 (d, J = 7.6 Hz, 1H), 7.16 (t, J = 8.0 Hz, 1H), 6.93 (t, J = 7.6 Hz, 1H), 6.80 (d, J = 8.0 Hz, 1H), 4.95–4.91 (m, 1H), 4.07–4.00 (m, 1H), 2.30 (ddd, J = 1.6, 6.4, 13.2 Hz, 1H), 1.84–1.69 (m, 4H), 1.05 (t, J = 7.6 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 154.8, 129.2, 127.1, 126.2, 120.7, 116.7, 76.3, 65.9, 37.7, 29.9, 28.7, 9.7; $[\alpha]_D^{26} = -130.94$ (c 0.54, CHCl_3); HPLC (Chiralcel OD-H, *n*-hexane/*i*-PrOH = 97/3, UV = 210 nm, flow rate = 0.5 mL/min) $t_{\text{R}1}$ = 20.598 min (minor) and $t_{\text{R}2}$ = 22.228 min (major), ee = 99%.



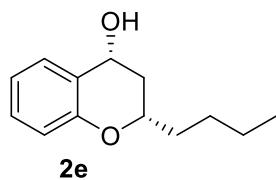
2c

(2S,4R)-2-Propylchroman-4-ol (2c): A white solid (54.8 mg, 96%). Mp 59–60 °C; ^1H NMR (400 MHz, CDCl_3) δ 7.46 (d, J = 7.6 Hz, 1H), 7.16 (t, J = 8.0 Hz, 1H), 6.93 (t, J = 7.6 Hz, 1H), 6.79 (d, J = 8.4 Hz, 1H), 4.95–4.91 (m, 1H), 4.15–4.08 (m, 1H), 2.30 (ddt, J = 1.2, 6.4, 12.8 Hz, 1H), 1.82–1.72 (m, 3H), 1.66–1.52 (m, 3H), 0.98 (t, J = 7.2 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 154.8, 129.2, 127.1, 126.2, 120.7, 116.7, 74.9, 65.9, 38.3, 37.9, 18.6, 14.2; HRMS (ESI): calcd for $\text{C}_{12}\text{H}_{16}\text{O}_2$ [$\text{M}+\text{Na}]^+$: 215.1048, found 215.1059; IR (KBr) cm^{-1} : 3580, 2914, 1601, 1580, 1491, 1465, 1384, 750; HRMS (ESI): calcd for $\text{C}_{12}\text{H}_{16}\text{O}_2$ [$\text{M}+\text{Na}]^+$: 215.1048, found 215.1057; $[\alpha]_D^{26} = -131.49$ (c 0.48, CHCl_3); HPLC (Chiralcel OD-H, *n*-hexane/*i*-PrOH = 99/1, UV = 210 nm, flow rate = 0.5 mL/min) $t_{\text{R}1}$ = 33.254 min (minor) and $t_{\text{R}2}$ = 35.837 min (major), ee = 99.7%.

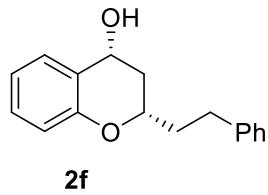


(2R,4R)-2-Isopropylchroman-4-ol (2d):^[2] A white solid (54.1 mg, 95%). ^1H NMR (400 MHz, CDCl_3) δ 7.46 (d, J = 7.2 Hz, 1H), 7.16 (t, J = 7.6 Hz, 1H), 6.93 (t, J = 7.2 Hz, 1H), 6.81 (d, J = 8.0 Hz, 1H),

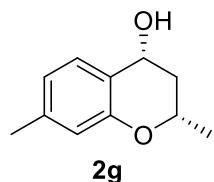
4.95–4.91 (m, 1H), 3.90–3.80 (m, 1H), 2.28 (dd, J = 5.6, 12.4 Hz, 1H), 2.00–1.91 (m, 1H), 1.72 (q, J = 11.6 Hz, 2H), 1.03 (t, J = 7.2 Hz, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 155.0, 129.1, 126.9, 126.3, 120.6, 116.7, 79.7, 66.3, 34.9, 32.6, 18.2, 18.1; $[\alpha]_D^{26} = -123.92$ (c 0.49, CHCl_3); HPLC (Chiralcel AS-H, *n*-hexane/*i*-PrOH = 99/1, UV = 210 nm, flow rate = 0.25 mL/min) $t_{R1} = 72.981$ min (minor) and $t_{R2} = 85.516$ min (major), ee = 99.1%.



(2S,4R)-2-Butylchroman-4-ol (2e):^[2] A white solid (59.3 mg, 97%). ^1H NMR (400 MHz, CDCl_3) δ 7.46 (d, J = 7.6 Hz, 1H), 7.16 (t, J = 7.2 Hz, 1H), 6.93 (t, J = 7.6 Hz, 1H), 6.79 (d, J = 7.6 Hz, 1H), 4.93–4.91 (m, 1H), 4.13–4.07 (m, 1H), 2.30 (dd, J = 10.4, 12.8 Hz, 1H), 1.83–1.73 (m, 3H), 1.70–1.60 (m, 2H), 1.45–1.34 (m, 3H), 0.95 (t, J = 6.8 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 154.8, 129.2, 127.1, 126.2, 120.7, 116.7, 75.2, 65.9, 38.2, 35.5, 27.5, 22.9, 14.3; $[\alpha]_D^{26} = -119.44$ (c 0.50, CHCl_3); HPLC (Chiralcel OD-H, *n*-hexane/*i*-PrOH = 98/2, UV = 210 nm, flow rate = 0.5 mL/min) $t_{R1} = 22.812$ min (minor) and $t_{R2} = 24.162$ min (major), ee = 99.6%.

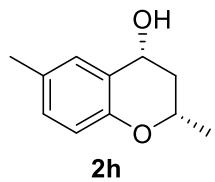


(2S,4R)-2-Phenethylchroman-4-ol (2f):^[2] A white solid (72.5 mg, 96%). ^1H NMR (400 MHz, CDCl_3) δ 7.49 (d, J = 6.0 Hz, 1H), 7.33 (t, J = 6.0 Hz, 2H), 7.28–7.20 (m, 4H), 6.98 (t, J = 6.0 Hz, 1H), 6.87 (d, J = 6.4 Hz, 1H), 4.99–4.88 (m, 1H), 4.15–4.10 (m, 1H), 2.96–2.83 (m, 2H), 2.31 (ddd, J = 1.2, 5.2, 10.4 Hz, 1H), 2.18–2.11 (m, 1H), 2.00–1.93 (m, 2H), 1.86–1.79 (m, 1H); ^{13}C NMR (100 MHz, CDCl_3) δ 154.5, 141.6, 129.0, 128.6, 128.5, 127.0, 126.0, 120.7, 116.6, 73.8, 65.5, 38.0, 37.2, 31.3; $[\alpha]_D^{26} = -133.55$ (c 0.51, CHCl_3); HPLC (Chiralcel OD-H, *n*-hexane/*i*-PrOH = 95/5, UV = 210 nm, flow rate = 0.7 mL/min) $t_{R1} = 25.283$ min (minor) and $t_{R2} = 35.795$ min (major), ee = 98%.

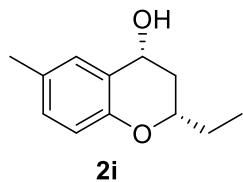


(2S,4R)-2,7-Dimethylchroman-4-ol (2g):^[2] A white solid (53.1 mg, 99%). ^1H NMR (400 MHz, CDCl_3) δ 7.33 (d, J = 8.0 Hz, 1H), 6.75 (d, J = 7.6 Hz, 1H), 6.61 (s, 1H), 4.93–4.84 (m, 1H), 4.26–4.18 (m, 1H), 2.28–2.24 (m, 4H), 1.75–1.66 (m, 2H), 1.40 (d, J = 6.4 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 154.6,

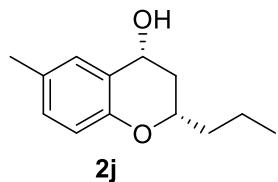
139.3, 127.0, 123.1, 121.8, 117.0, 71.4, 65.6, 40.2, 21.7, 21.4; $[\alpha]_D^{26} = -124.65$ (*c* 0.54, CHCl₃); HPLC (Chiralcel OD-H, *n*-hexane/*i*-PrOH = 97/3, UV = 210 nm, flow rate = 0.5 mL/min) t_{R1} = 20.492 min (minor) and t_{R2} = 24.786 min (major), ee = 99.7%.



(2S,4R)-2,6-Dimethylchroman-4-ol (2h):^[2] A white solid (52.8 mg, 99%). ¹H NMR (400 MHz, CDCl₃) δ 7.26 (s, 1H), 6.96 (d, *J* = 8.0 Hz, 1H), 6.69 (d, *J* = 8.4 Hz, 1H), 4.93–4.82 (m, 1H), 4.24–4.16 (m, 1H), 2.28–2.22 (m, 4H), 1.76–1.67 (m, 2H), 1.40 (dd, *J* = 1.6, 6.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 152.5, 130.0, 129.9, 127.4, 125.6, 116.4, 71.4, 65.7, 40.1, 21.7, 20.8; $[\alpha]_D^{26} = -130.98$ (*c* 0.52, CHCl₃); HPLC (Chiralcel OD-H, *n*-hexane/*i*-PrOH = 97/3, UV = 210 nm, flow rate = 0.5 mL/min) t_{R1} = 19.886 min (minor) and t_{R2} = 23.689 min (major), ee = 99.3%.

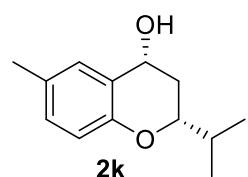


(2S,4R)-2-ethyl-6-methylchroman-4-ol (2i): A white solid (56.1 mg, 98%). Mp 64–65 °C; ¹H NMR (400 MHz, CDCl₃) δ 7.29 (s, 1H), 6.98 (dd, *J* = 1.6, 8.4 Hz, 1H), 6.72 (d, *J* = 8.4 Hz, 1H), 4.93–4.89 (m, 1H), 4.05–3.98 (m, 1H), 2.32–2.27 (m, 4H), 1.86–1.65 (m, 4H), 1.06 (t, *J* = 7.6 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 152.4, 129.7, 129.6, 127.1, 125.6, 116.2, 76.0, 65.7, 37.6, 28.4, 20.6, 9.5; IR (KBr) cm⁻¹: 3578, 2924, 1601, 1580, 1499, 1460, 1375, 810; HRMS (ESI): calcd for C₁₂H₁₆O₂ [M+Na]⁺: 215.1048, found 215.1039; $[\alpha]_D^{26} = -125.93$ (*c* 0.59, CHCl₃); HPLC (Chiralcel OD-H, *n*-hexane/*i*-PrOH = 97/3, UV = 210 nm, flow rate = 0.5 mL/min) t_{R1} = 17.891 min (minor) and t_{R2} = 19.170 min (major), ee = 94%.

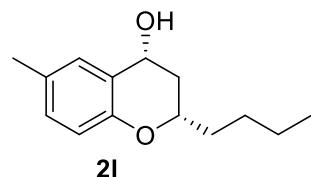


(2S,4R)-6-Methyl-2-propylchroman-4-ol (2j): A white solid (59.6 mg, 97%). Mp 79–81 °C; ¹H NMR (500 MHz, CDCl₃) δ 7.28 (s, 1H), 6.98 (dd, *J* = 2.0, 10.5 Hz, 1H), 6.72 (d, *J* = 10.0 Hz, 1H), 4.93–4.89 (m, 1H), 4.12–4.06 (m, 1H), 2.30–2.26 (m, 4H), 1.83–1.45 (m, 6H), 1.00 (t, *J* = 9.0 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 152.4, 129.7, 129.6, 127.2, 125.6, 116.3, 74.6, 65.7, 38.1, 37.6, 20.6, 18.4, 14.0; IR (KBr) cm⁻¹: 3565, 2934, 1599, 1579, 1491, 1460, 1380, 790; HRMS (ESI): calcd for C₁₃H₁₈O₂ [M+Na]⁺: 229.1204, found 229.1214; $[\alpha]_D^{26} = -118.21$ (*c* 0.49, CHCl₃); HPLC (Chiralcel AS-H, *n*-hexane/*i*-PrOH = 97/3, UV = 210 nm, flow rate = 0.5 mL/min) t_{R1} = 16.492 min (minor) and t_{R2} = 18.467 min (major), ee =

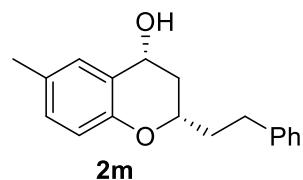
98%.



(2*R*,4*R*)-2-Isopropyl-6-methylchroman-4-ol (2k): A white solid (59.3 mg, 97%). Mp 72–73 °C; ^1H NMR (400 MHz, CDCl_3) δ 7.28 (s, 1H), 6.98 (dd, J = 1.6, 8.0 Hz, 1H), 6.73 (d, J = 8.0 Hz, 1H), 4.93–4.89 (m, 1H), 3.88–3.83 (m, 1H), 2.30–2.25 (m, 4H), 2.00–1.91 (m, 2H), 1.78–1.69 (m, 1H), 1.07–1.03 (m, 6H); ^{13}C NMR (100 MHz, CDCl_3) δ 152.6, 129.6, 126.9, 125.7, 116.2, 79.4, 66.1, 34.8, 32.4, 20.6, 18.0, 17.9; IR (KBr) cm^{-1} : 3588, 2904, 1601, 1580, 1501, 1465, 1384, 1368, 744; HRMS (ESI): calcd for $\text{C}_{13}\text{H}_{18}\text{O}_2$ [$\text{M}+\text{Na}]^+$: 229.1204, found 229.1210; $[\alpha]_D^{26} = -131.75$ (c 0.52, CHCl_3); HPLC (Chiralcel AS-H, *n*-hexane/*i*-PrOH = 99/1, UV = 210 nm, flow rate = 0.25 mL/min) $t_{R1} = 65.928$ min (minor) and $t_{R2} = 74.529$ min (major), ee = 99.8%.

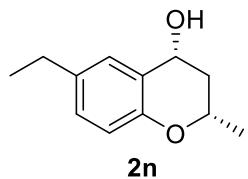


(2*S*,4*R*)-2-Butyl-6-methylchroman-4-ol (2l): A white solid (64.5 mg, 98%). Mp 74–75 °C; ^1H NMR (400 MHz, CDCl_3) δ 7.26 (s, 1H), 6.96 (d, J = 8.0 Hz, 1H), 6.69 (d, J = 8.4 Hz, 1H), 4.93–4.87 (m, 1H), 4.09–4.02 (m, 1H), 2.32–2.27 (m, 4H), 1.81–1.68 (m, 3H), 1.53–1.25 (m, 5H), 0.93 (t, J = 7.2 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 152.4, 129.7, 129.6, 127.1, 125.6, 116.3, 74.9, 65.7, 38.2, 35.2, 27.3, 22.6, 20.6, 14.0; IR (KBr) cm^{-1} : 3563, 2936, 1601, 1580, 1498, 1465, 1386, 721; HRMS (ESI): calcd for $\text{C}_{14}\text{H}_{20}\text{O}_2$ [$\text{M}+\text{Na}]^+$: 243.1364, found 249.1363; $[\alpha]_D^{26} = -113.42$ (c 0.28, CHCl_3); HPLC (Chiralcel AS-H, *n*-hexane/*i*-PrOH = 97/3, UV = 210 nm, flow rate = 0.3 mL/min) $t_{R1} = 24.493$ min (minor) and $t_{R2} = 26.637$ min (major), ee = 97%.

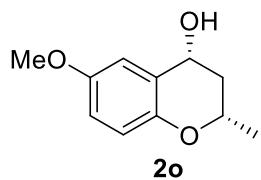


(2*S*,4*R*)-6-Methyl-2-phenethylchroman-4-ol (2m): A white solid (75.7 mg, 95%). Mp 88–90 °C; ^1H NMR (400 MHz, CDCl_3) δ 7.35–7.22 (m, 6H), 7.02 (dd, J = 1.6, 8.0 Hz, 1H), 6.79 (d, J = 8.4 Hz, 1H), 4.98–4.83 (m, 1H), 4.12–4.06 (m, 1H), 2.96–2.89 (m, 2H), 2.32–2.27 (m, 4H), 2.17–2.11 (m, 1H), 2.00–1.91 (m, 2H), 1.85–1.76 (m, 1H); ^{13}C NMR (100 MHz, CDCl_3) δ 152.3, 141.6, 129.9, 129.7, 128.6, 128.5, 127.2, 126.0, 125.6, 116.3, 73.7, 65.5, 38.2, 37.2, 31.4, 20.6; IR (KBr) cm^{-1} : 3579, 2914, 1601, 1579, 1491, 1465, 1384, 768; HRMS (ESI): calcd for $\text{C}_{18}\text{H}_{20}\text{O}_2$ [$\text{M}+\text{Na}]^+$: 291.1361, found 291.1360; $[\alpha]_D^{26} =$

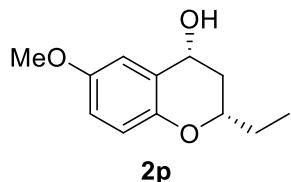
-70.65 (c 0.54, CHCl_3); HPLC (Chiralcel AD-H, n -hexane/ i -PrOH = 97/3, UV = 210 nm, flow rate = 0.5 mL/min) $t_{\text{R}1}$ = 42.124 min (minor) and $t_{\text{R}2}$ = 52.387 min (major), ee = 97%.



(2S,4R)-6-Ethyl-2-methylchroman-4-ol (2n): A white solid (57.4 mg, 99%). Mp 49–51 °C; ^1H NMR (400 MHz, CDCl_3) δ 7.30 (s, 1H), 7.01 (d, J = 10.0 Hz, 1H), 6.72 (d, J = 8.4 Hz, 1H), 4.93–4.89 (m, 1H), 4.26–4.18 (m, 1H), 2.58 (q, J = 7.6 Hz, 2H), 2.28 (ddd, J = 1.6, 6.4, 12.8 Hz, 1H), 1.77–1.66 (m, 2H), 1.41 (d, J = 6.0 Hz, 3H), 1.21 (t, J = 7.6 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 152.5, 136.4, 128.5, 126.0, 125.3, 116.3, 71.1, 65.6, 40.0, 28.1, 21.5, 15.9; IR (KBr) cm^{-1} : 3580, 2904, 1599, 1577, 1491, 1465, 1380, 767; HRMS (ESI): calcd for $\text{C}_{12}\text{H}_{16}\text{O}_2$ [$\text{M}+\text{Na}]^+$: 215.1048, found 215.1054; $[\alpha]_D^{26} = -112.25$ (c 0.50, CHCl_3); HPLC (Chiralcel OD-H, n -hexane/ i -PrOH = 97/3, UV = 210 nm, flow rate = 0.5 mL/min) $t_{\text{R}1}$ = 17.067 min (minor) and $t_{\text{R}2}$ = 20.204 min (major), ee = 96%.

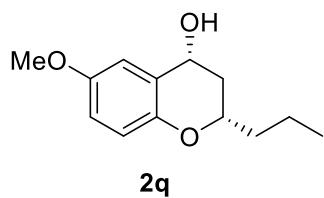


(2S,4R)-6-Methoxy-2-methylchroman-4-ol (2o): A white solid (57.5 mg, 99%). Mp 74–75 °C; ^1H NMR (400 MHz, CDCl_3) δ 7.00 (s, 1H), 6.75–6.70 (m, 2H), 4.92–4.86 (m, 1H), 4.22–4.15 (m, 1H), 3.75 (s, 3H), 2.27 (dd, J = 6.8, 13.2 Hz, 1H), 1.75–1.66 (m, 2H), 1.39 (d, J = 6.0 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 153.9, 148.8, 126.3, 117.4, 115.8, 111.3, 71.4, 65.9, 56.0, 40.1, 21.7; IR (KBr) cm^{-1} : 3580, 2934, 1601, 1580, 1501, 1465, 1384, 1215, 780; HRMS (ESI): calcd for $\text{C}_{11}\text{H}_{14}\text{O}_3$ [$\text{M}+\text{Na}]^+$: 217.0841, found 217.0850; $[\alpha]_D^{26} = -119.46$ (c 0.53, CHCl_3); HPLC (Chiralcel OD-H, n -hexane/ i -PrOH = 97/3, UV = 210 nm, flow rate = 0.5 mL/min) $t_{\text{R}1}$ = 32.079 min (minor) and $t_{\text{R}2}$ = 40.849 min (major), ee = 99.1%.

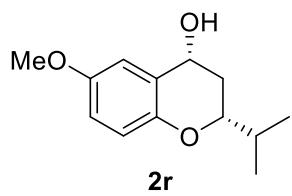


(2S,4R)-2-Ethyl-6-methoxychroman-4-ol (2p): A white solid (62.1 mg, 99%). Mp 56–58 °C; ^1H NMR (500 MHz, CDCl_3) δ 7.02–7.00 (m, 1H), 6.78–6.73 (m, 2H), 4.96–4.84 (m, 1H), 4.01–3.96 (m, 1H), 3.77 (s, 3H), 2.31–2.27 (m, 1H), 1.83–1.65 (m, 4H), 1.05 (t, J = 7.0 Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3) δ 153.6, 148.6, 126.4, 117.2, 115.5, 111.1, 76.1, 65.9, 55.8, 37.6, 28.4, 9.5; HRMS (ESI): calcd for $\text{C}_{12}\text{H}_{16}\text{O}_3$ [$\text{M}+\text{Na}]^+$: 231.0997, found 231.1005; IR (KBr) cm^{-1} : 3572, 2914, 1601, 1581, 1491, 1465, 1379, 1264, 750;

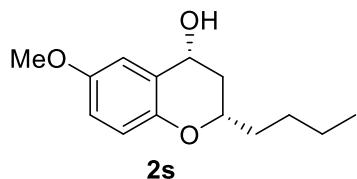
$[\alpha]_D^{26} = -107.06$ (*c* 0.50, CHCl₃); HPLC (Chiralcel OD-H, *n*-hexane/*i*-PrOH = 97/3, UV = 210 nm, flow rate = 0.5 mL/min) t_{R1} = 26.736 min (minor) and t_{R2} = 31.238 min (major), ee = 98%.



(2*S*,4*R*)-6-Methoxy-2-propylchroman-4-ol (2q): A white solid (63.9 mg, 97%). Mp 62–63 °C; ¹H NMR (500 MHz, CDCl₃) δ 7.01 (d, *J* = 2.5 Hz, 1H), 6.76–6.72 (m, 2H), 4.95–4.85 (m, 1H), 4.08–4.03 (m, 1H), 3.77 (s, 3H), 2.29–2.26 (m, 1H), 1.80–1.43 (m, 6H), 0.99 (t, *J* = 7.0 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 153.6, 148.7, 126.4, 117.2, 115.5, 111.1, 74.6, 65.8, 55.8, 38.1, 37.7, 18.4, 14.0; IR (KBr) cm⁻¹: 3580, 2914, 1601, 1580, 1491, 1465, 1384, 1266, 750; HRMS (ESI): calcd for C₁₃H₁₈O₃ [M+Na]⁺: 245.1154, found 245.1164; $[\alpha]_D^{26} = -108.58$ (*c* 0.55, CHCl₃); HPLC (Chiralcel AS-H, *n*-hexane/*i*-PrOH = 97/3, UV = 210 nm, flow rate = 0.5 mL/min) t_{R1} = 33.168 min (minor) and t_{R2} = 37.861 min (major), ee = 98%.

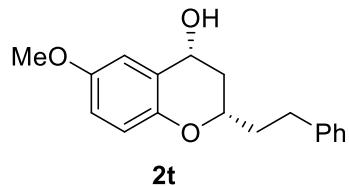


(2*R*,4*R*)-2-Isopropyl-6-methoxychroman-4-ol (2r):^[9] A white solid (64.1 mg, 97%). ¹H NMR (500 MHz, CDCl₃) δ 7.02 (s, 1H), 6.77–6.73 (m, 2H), 4.96–4.85 (m, 1H), 3.85–3.81 (m, 1H), 3.78 (s, 3H), 2.27 (ddd, *J* = 1.5, 6.5, 12.5 Hz, 1H), 1.98–1.92 (m, 1H), 1.71 (q, *J* = 12.0 Hz, 2H), 1.05 (d, *J* = 7.0 Hz, 3H), 1.03 (d, *J* = 6.5 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 153.5, 148.8, 126.5, 117.2, 115.5, 110.9, 79.5, 66.3, 55.8, 34.8, 32.4, 18.0, 17.9; $[\alpha]_D^{26} = -101.72$ (*c* 0.54, CHCl₃); HPLC (Chiralcel OD-H, *n*-hexane/*i*-PrOH = 97/3, UV = 210 nm, flow rate = 0.5 mL/min) t_{R1} = 22.162 min (minor) and t_{R2} = 25.229 min (major), ee = 97%.

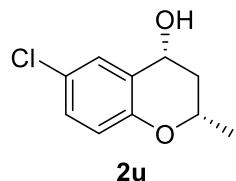


(2*S*,4*R*)-2-Butyl-6-methoxychroman-4-ol (2s): A white solid (68.8 mg, 98%). Mp 62–64 °C; ¹H NMR (500 MHz, CDCl₃) δ 7.02 (s, 1H), 6.77–6.73 (m, 2H), 4.95–4.85 (m, 1H), 4.07–4.04 (m, 1H), 3.78 (d, *J* = 2.4 Hz, 3H), 2.31–2.27 (m, 1H), 1.77–1.39 (m, 8H), 0.98–0.92 (m, 3H); ¹³C NMR (125 MHz, CDCl₃) δ 153.6, 148.7, 126.4, 117.2, 115.5, 111.1, 74.9, 65.9, 55.8, 38.2, 35.3, 27.3, 22.7, 14.0; IR (KBr) cm⁻¹: 3588,

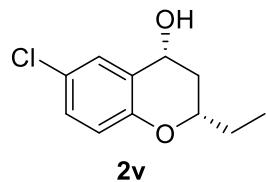
2934, 1611, 1580, 1491, 1465, 1384, 790; HRMS (ESI): calcd for $C_{14}H_{20}O_3$ [M+Na]⁺: 259.1310, found 259.1317; $[\alpha]_D^{26} = -106.11$ (*c* 0.48, CHCl₃); HPLC (Chiralcel OD-H, *n*-hexane/*i*-PrOH = 97/3, UV = 210 nm, flow rate = 0.5 mL/min) $t_{R1} = 24.062$ min (minor) and $t_{R2} = 26.916$ min (major), ee = 99%.



(2*S*,4*R*)-6-Methoxy-2-phenethylchroman-4-ol (2t): A white solid (80.4 mg, 95%). Mp 97–98 °C; ¹H NMR (500 MHz, CDCl₃) δ 7.33–7.20 (m, 5H), 7.03 (s, 1H), 6.81–6.76 (m, 2H), 4.96–4.87 (m, 1H), 4.09–4.03 (m, 1H), 3.79 (s, 3H), 2.94–2.80 (m, 2H), 2.31 (dd, *J* = 6.0, 12.5 Hz), 2.15–2.08 (m, 1H), 1.97–1.90 (m, 1H), 1.83–1.76 (m, 2H); ¹³C NMR (125 MHz, CDCl₃) δ 153.7, 148.5, 141.6, 128.5, 128.4, 126.3, 125.9, 117.3, 115.6, 111.1, 73.7, 65.8, 55.8, 38.2, 37.2, 31.3; IR (KBr) cm⁻¹: 3578, 2934, 1591, 1580, 1500, 1465, 1380, 790; HRMS (ESI): calcd for $C_{18}H_{20}O_3$ [M+Na]⁺: 307.1310, found 307.1306; $[\alpha]_D^{26} = -116.77$ (*c* 0.52, CHCl₃); HPLC (Chiralcel OD-H, *n*-hexane/*i*-PrOH = 95/5, UV = 210 nm, flow rate = 0.8 mL/min) $t_{R1} = 26.457$ min (minor) and $t_{R2} = 40.748$ min (major), ee = 98%.

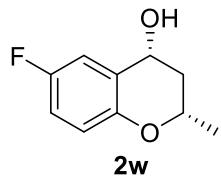


(2*S*,4*R*)-6-Chloro-2-methylchroman-4-ol (2u):^[9] A white solid (58.8 mg, 99%). ¹H NMR (400 MHz, CDCl₃) δ 7.44 (d, *J* = 1.2 Hz, 1H), 7.11–7.08 (m, 1H), 6.73–6.70 (m, 1H), 4.89–4.87 (m, 1H), 4.26–4.21 (m, 1H), 2.30–2.25 (m, 1H), 1.73–1.65 (m, 1H), 1.43–1.40 (m, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 153.1, 128.9, 127.1, 126.8, 125.4, 117.8, 71.5, 65.3, 39.4, 21.4; $[\alpha]_D^{26} = -131.68$ (*c* 0.54, CHCl₃); HPLC (Chiralcel OD-H, *n*-hexane/*i*-PrOH = 97/3, UV = 210 nm, flow rate = 0.5 mL/min) $t_{R1} = 21.424$ min (minor) and $t_{R2} = 25.758$ min (major), ee = 98%.

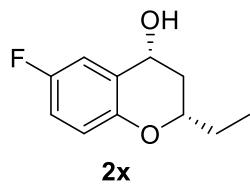


(2*S*,4*R*)-6-Chloro-2-ethylchroman-4-ol (2v): A white solid (60.9 mg, 96%). Mp 113–115 °C; ¹H NMR (400 MHz, CDCl₃) δ 7.46 (s, 1H), 7.12 (dd, *J* = 2.0, 7.2 Hz, 1H), 6.75 (d, *J* = 7.2 Hz, 1H), 4.91 (s, 1H), 4.07–4.03 (m, 1H), 2.31 (ddd, *J* = 1.2, 4.8, 10.4 Hz, 1H), 1.84–1.64 (m, 4H), 1.06 (t, *J* = 6.0 Hz, 3H); ¹³C

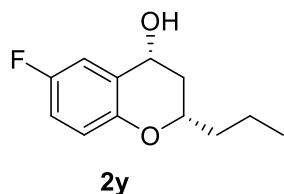
NMR (100 MHz, CDCl₃) δ 153.2, 128.9, 127.4, 126.7, 125.3, 117.9, 76.4, 65.4, 37.1, 28.3, 9.4; IR (KBr) cm⁻¹: 3588, 2930, 1598, 1580, 1500, 1465, 1382, 750; HRMS (ESI): calcd for C₁₁H₁₃ClO₂ [M+Na]⁺: 235.0502, found 235.0503; [α]_D²⁶ = -119.45 (c 0.51, CHCl₃); HPLC (Chiralcel OD-H, *n*-hexane/*i*-PrOH = 97/3, UV = 210 nm, flow rate = 0.5 mL/min) t_{R1} = 19.275 min (minor) and t_{R2} = 21.029 min (major), ee = 87%.



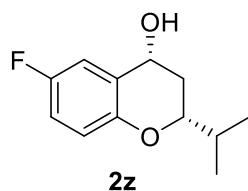
(2S,4R)-6-Fluoro-2-methylchroman-4-ol (2w): A white solid (54.2mg, 99%). Mp 94–96 °C; ¹H NMR (400 MHz, CDCl₃) δ 7.18 (dd, *J* = 2.4, 7.2 Hz, 1H), 6.87 (td, *J* = 2.0, 6.8 Hz, 1H), 6.74 (dd, *J* = 4.0, 7.2 Hz, 1H), 4.95–4.89 (m, 1H), 4.28–4.21 (m, 1H), 2.30 (dd, *J* = 4.8, 10.4 Hz, 1H), 1.74 (q, *J* = 9.2 Hz, 2H), 1.43 (d, *J* = 5.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 157.1 (d, *J* = 189.4 Hz), 150.5 (d, *J* = 1.5 Hz), 126.7 (d, *J* = 5.3 Hz), 117.4 (d, *J* = 6.3 Hz), 115.8 (d, *J* = 18.6 Hz), 113.1 (d, *J* = 18.7 Hz), 71.4, 65.4, 39.5, 21.4; ¹⁹F NMR (376 MHz, CDCl₃) δ -123.4; IR (KBr) cm⁻¹: 3588, 2934, 1596, 1584, 1500, 1465, 1380, 1217, 780; HRMS (ESI): calcd for C₁₀H₁₁FO₂ [M+Na]⁺: 205.0641, found 205.0650; [α]_D²⁶ = -97.07 (c 0.50, CHCl₃); HPLC (Chiralcel OD-H, *n*-hexane/*i*-PrOH = 97/3, UV = 210 nm, flow rate = 0.5 mL/min) t_{R1} = 22.519 min (minor) and t_{R2} = 27.109 min (major), ee = 99%.



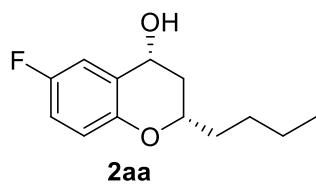
(2S,4R)-2-Ethyl-6-fluorochroman-4-ol (2x): A white solid (57.1mg, 98%). Mp 77–78 °C; ¹H NMR (400 MHz, CDCl₃) δ 7.18 (dd, *J* = 2.4, 8.8 Hz, 1H), 6.87 (td, *J* = 2.8, 8.4 Hz, 1H), 6.74 (dd, *J* = 4.8, 8.8 Hz, 1H), 4.98–4.85 (m, 1H), 4.06–4.00 (m, 1H), 2.30 (ddd, *J* = 1.2, 6.4, 12.8 Hz, 1H), 1.86–1.65 (m, 4H), 1.06 (t, *J* = 7.6 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 157.0 (d, *J* = 236.6 Hz), 150.6 (d, *J* = 1.9 Hz), 126.9 (d, *J* = 6.6 Hz), 117.4 (d, *J* = 7.7 Hz), 115.7 (d, *J* = 23.2 Hz), 113.0 (d, *J* = 23.4 Hz), 76.2, 65.6, 37.2, 28.4, 9.4; ¹⁹F NMR (376 MHz, CDCl₃) δ -124.0; IR (KBr) cm⁻¹: 3586, 2932, 1598, 1584, 1499, 1465, 1218, 1385, 780; HRMS (ESI): calcd for C₁₁H₁₃FO₂ [M+Na]⁺: 219.0797, found 219.0789; [α]_D²⁶ = -93.69 (c 0.55, CHCl₃); HPLC (Chiralcel OD-H, *n*-hexane/*i*-PrOH = 97/3, UV = 210 nm, flow rate = 0.5 mL/min) t_{R1} = 19.408 min (minor) and t_{R2} = 21.154 min (major), ee = 90%.



(2S,4R)-6-Fluoro-2-propylchroman-4-ol (2y): A white solid (62.6 mg, 99%). Mp 65–66 °C; ¹H NMR (400 MHz, CDCl₃) δ 7.17 (dd, *J* = 2.8, 8.8 Hz, 1H), 6.86 (td, *J* = 2.8, 8.4 Hz, 1H), 6.73 (dd, *J* = 4.4, 8.8 Hz, 1H), 4.97–4.83 (m, 1H), 4.12–4.06 (m, 1H), 2.28 (dd, *J* = 6.4, 12.8 Hz, 1H), 1.81–1.46 (m, 6H), 1.00 (t, *J* = 7.2 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 157.0 (d, *J* = 236.7 Hz), 150.6 (d, *J* = 1.9 Hz), 126.9 (d, *J* = 6.6 Hz), 117.4 (d, *J* = 7.8 Hz), 115.7 (d, *J* = 23.3 Hz), 113.0 (d, *J* = 23.3 Hz), 74.8, 65.6, 37.7, 37.6, 18.3, 14.0; ¹⁹F NMR (376 MHz, CDCl₃) δ -123.5; IR (KBr) cm⁻¹: 3571, 2934, 1600, 1584, 1498, 1465, 1380, 1218, 768; HRMS (ESI): calcd for C₁₂H₁₅FO₂ [M+Na]⁺: 233.0954, found 233.0961; [α]_D²⁶ = -109.08 (*c* 0.52, CHCl₃); HPLC (Chiralcel AD-H, *n*-hexane/*i*-PrOH = 97/3, UV = 210 nm, flow rate = 0.5 mL/min) t_{R1} = 24.338 min (minor) and t_{R2} = 29.615 min (major), ee = 98%.

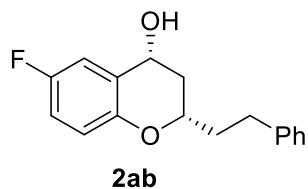


(2R,4R)-6-Fluoro-2-isopropylchroman-4-ol (2z): A white solid (60.3 mg, 96%). Mp 56–57 °C; ¹H NMR (400 MHz, CDCl₃) δ 7.16 (d, *J* = 8.8 Hz, 1H), 6.85 (t, *J* = 7.2 Hz, 1H), 6.73 (dd, *J* = 4.4, 8.8 Hz, 1H), 4.93–4.86 (m, 1H), 3.87–3.83 (m, 1H), 2.27 (dd, *J* = 6.0, 12.8 Hz, 1H), 1.98–1.90 (m, 1H), 1.80–1.67 (m, 2H), 1.02 (t, *J* = 7.2 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 157.0 (d, *J* = 236.5 Hz), 150.8 (d, *J* = 2.0 Hz), 127.0 (d, *J* = 6.0 Hz), 117.4 (d, *J* = 7.7 Hz), 115.6 (d, *J* = 23.2 Hz), 112.9 (d, *J* = 23.4 Hz), 79.7, 66.0, 34.4, 32.3, 18.0, 17.8; ¹⁹F NMR (376 MHz, CDCl₃) δ -123.7; HRMS (ESI): calcd for C₁₂H₁₅FO₂ [M+Na]⁺: 233.0954, found 233.0950; IR (KBr) cm⁻¹: 3589, 2934, 1596, 1584, 1500, 1465, 1380, 1368, 1217, 780; [α]_D²⁶ = -101.90 (*c* 0.59, CHCl₃); HPLC (Chiralcel OD-H, *n*-hexane/*i*-PrOH = 97/3, UV = 210 nm, flow rate = 0.5 mL/min) t_{R1} = 16.768 min (minor) and t_{R2} = 17.871 min (major), ee = 96%.

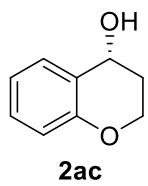


(2S,4R)-2-Butyl-6-fluorochroman-4-ol (2aa): A white solid (65.1 mg, 97%). Mp 76–77 °C; ¹H NMR (400 MHz, CDCl₃) δ 7.16 (dd, *J* = 2.4, 8.8 Hz, 1H), 6.85 (td, *J* = 2.4, 8.4 Hz, 1H), 6.72 (dd, *J* = 4.8, 8.8 Hz, 1H), 4.92–4.86 (m, 1H), 4.10–4.04 (m, 1H), 2.29 (dd, *J* = 6.4, 13.2 Hz, 1H), 1.78–1.62 (m, 4H), 1.53–1.33

(m, 4H), 0.94 (t, J = 6.8 Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 157.0 (d, J = 236.7 Hz), 150.6 (d, J = 1.9 Hz), 126.9 (d, J = 6.6 Hz), 117.4 (d, J = 7.8 Hz), 115.7 (d, J = 23.2 Hz), 113.0 (d, J = 23.4 Hz), 75.1, 65.6, 37.7, 35.2, 27.2, 22.6, 14.0; ^{19}F NMR (376 MHz, CDCl_3) δ -123.6; HRMS (ESI): calcd for $\text{C}_{13}\text{H}_{17}\text{FO}_2$ [$\text{M}+\text{Na}]^+$: 247.1110, found 247.1113; IR (KBr) cm^{-1} : 3581, 2942, 1600, 1584, 1500, 1465, 1383, 1217, 769; $[\alpha]_D^{26} = -91.31$ (c 0.49, CHCl_3); HPLC (Chiralcel AD-H, *n*-hexane/*i*-PrOH = 97/3, UV = 210 nm, flow rate = 0.5 mL/min) $t_{\text{R}1}$ = 23.747 min (minor) and $t_{\text{R}2}$ = 29.488 min (major), ee = 98%.



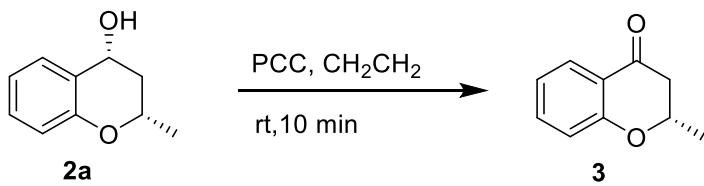
(2*S*,4*R*)-6-Fluoro-2-phenethylchroman-4-ol (2ab): A white solid (77.9 mg, 96%). Mp 91–92 °C; ^1H NMR (400 MHz, CDCl_3) δ 7.31–7.15 (m, 6H), 6.87 (td, J = 2.4, 8.8 Hz, 1H), 6.77 (dd, J = 4.8, 8.8 Hz, 1H), 4.90–4.84 (m, 1H), 4.10–4.04 (m, 1H), 2.92–2.76 (m, 2H), 2.29 (dd, J = 6.4, 13.2 Hz, 1H), 2.14–2.05 (m, 1H), 1.97–1.88 (m, 1H), 1.82–1.73 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 157.1 (d, J = 236.9 Hz), 150.5 (d, J = 2.0 Hz), 141.5, 128.5, 128.4, 126.9 (d, J = 6.6 Hz), 126.0, 117.5 (d, J = 7.8 Hz), 115.8 (d, J = 23.2 Hz), 113.1 (d, J = 23.3 Hz), 74.0, 65.4, 37.7, 37.1, 31.3; ^{19}F NMR (376 MHz, CDCl_3) δ -123.3; IR (KBr) cm^{-1} : 3589, 2934, 1599, 1580, 1500, 1465, 1384, 1216, 789; HRMS (ESI): calcd for $\text{C}_{17}\text{H}_{17}\text{FO}_2$ [$\text{M}+\text{Na}]^+$: 295.1110, found 295.1112; $[\alpha]_D^{26} = -124.32$ (c 0.58, CHCl_3); HPLC (Chiralcel OD-H, *n*-hexane/*i*-PrOH = 97/3, UV = 210 nm, flow rate = 0.5 mL/min) $t_{\text{R}1}$ = 84.433 min (minor) and $t_{\text{R}2}$ = 89.431 min (major), ee = 98%.



(*R*)-Chroman-4-ol (2ac):^[10] A white solid (44.8 mg, 99%). ^1H NMR (400 MHz, CDCl_3) δ 7.30 (d, J = 7.6 Hz, 1H), 7.21 (t, J = 6.8 Hz, 1H), 6.92 (t, J = 7.2 Hz, 1H), 6.84 (d, J = 8.4 Hz, 1H), 4.81–4.73 (m, 1H), 4.27–4.24 (m, 1H), 2.15–2.00 (m, 3H); ^{13}C NMR (100 MHz, CDCl_3) δ 154.8, 130.0, 129.9, 124.5, 120.8, 117.3, 63.5, 62.1, 31.0; $[\alpha]_D^{26} = +56.52$ (c 0.47, CHCl_3); HPLC (Chiralcel OD-H, *n*-hexane/*i*-PrOH = 97/3, UV = 210 nm, flow rate = 0.5 mL/min) $t_{\text{R}1}$ = 24.971 min (minor) and $t_{\text{R}2}$ = 29.406 min (major), ee = 99%.

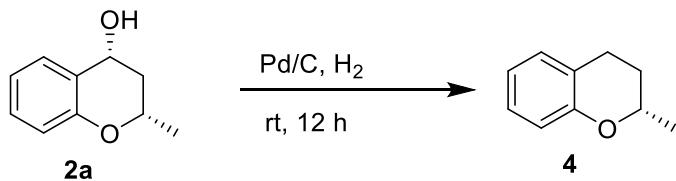
4. Transformation of Chromanol

4.1 Procedure for synthesis of 3^[2]



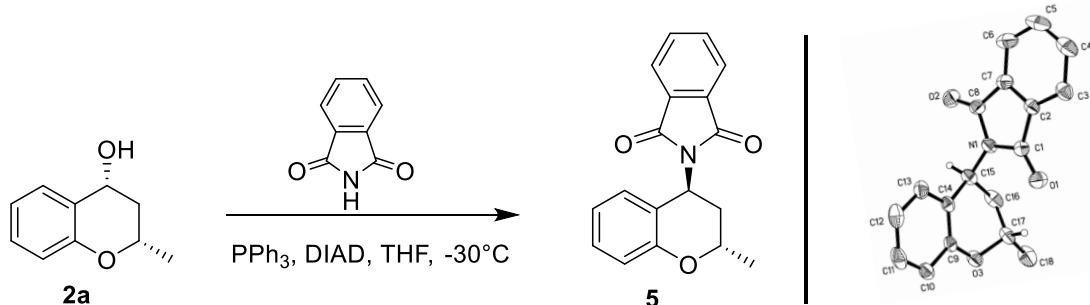
The **2a** (0.15 mmol) was treated with PCC (120 mg) in CH_2Cl_2 (2 mL) at room temperatures for 10 min. Evaporation of DCM gave a residue and then Et_2O was added. The solution was filtered through a plug of silica using Et_2O . The solvent was evaporated to give crude product. Then, it was purified by chromatography (PE:EtOAc = 20:1) to give pure product **3** as a colorless oil (46.3 mg, 95% yield). ^1H NMR (400 MHz, CDCl_3) δ 7.90 (dd, J = 1.6, 8.0 Hz, 1H), 7.49 (td, J = 1.6, 7.2 Hz, 1H), 7.04–6.98 (m, 2H), 4.67–4.57 (m, 1H), 2.70 (d, J = 7.6 Hz, 2H), 1.54 (d, J = 6.4 Hz, 3H); $[\alpha]_D^{26} = +26.56$ (c 0.47, CHCl_3); HPLC (Chiralcel OD-H, *n*-hexane/*i*-PrOH = 97/3, UV = 210 nm, flow rate = 0.5 mL/min) t_{R1} = 13.128 min (minor) and t_{R2} = 14.520 min (major), ee = 99.9%.

4.2 Procedure for synthesis of **4**^[11]



A hydrogenation tube was charged with a stirring bar, **2a** (0.30 mmol), Pd/C (20%). MeOH (3 mL) was then injected into the hydrogenation tube by a syringe. The hydrogenation tube was then put into an autoclave. The autoclave was then charged with hydrogen to 30 bar hydrogen pressure, and the reaction mixture was stirred at RT for 12 h. The solvent was evaporated to afford the crude product and it was purified by chromatography (PE:EtOAc = 10:1) to give pure product **4** as a colorless oil(44.1 mg, 99%). ^1H NMR (400 MHz, CDCl_3) δ 7.13–7.06 (m, 2H), 6.87–6.82 (m, 2H), 4.21–4.13 (m, 1H), 2.94–2.85 (m, 1H), 2.81–2.74 (m, 1H), 2.05–1.99 (m, 1H), 1.81–1.70 (m, 1H), 1.43 (d, J = 6.4 Hz, 3H); $[\alpha]_D^{26} = +84.20$ (c 0.47, CHCl_3); HPLC (Chiralcel OD-H, *n*-hexane/*i*-PrOH = 97/3, UV = 210 nm, flow rate = 0.5 mL/min) t_{R1} = 8.572 min (minor) and t_{R2} = 9.246 min (major), ee = 99.8%.

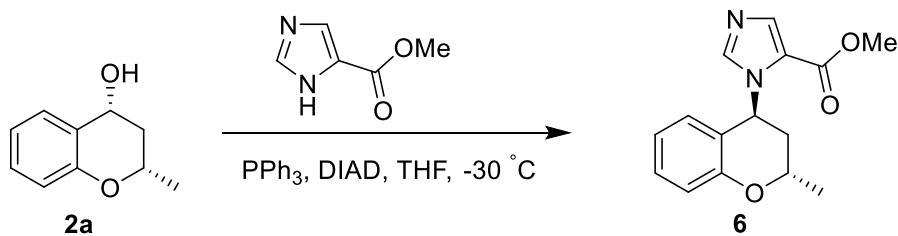
4.3 Procedure for synthesis of **5**



Diisopropyl azodicarboxylate (DIAD) (0.54 g, 2.7 mmol) in dry THF (5 ml) was added dropwise to a

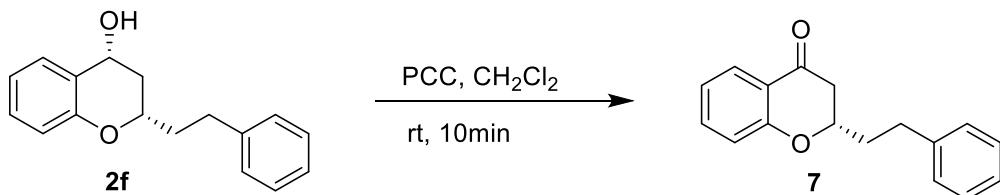
stirred solution of **2a** (0.36 g, 2.2 mmol), PPh₃, (0.68 g, 2.6 mmol) and phthalimide (0.33 g, 2.3 mmol) in dry THF (15 ml) at -30 °C before the mixture was warmed to room temperature and stirred for a further 1 h. Then the solvent was evaporated to provide crude product and it was purified by chromatography (PE:EA=10:1) to give pure product **5** as a white solid (0.6 g, 93%). Mp 145–146 °C ¹H NMR (400 MHz, CDCl₃) δ 7.87–7.83 (m, 2H), 7.78–7.74 (m, 2H), 7.23–7.19 (m, 1H), 7.02 (d, *J* = 7.2 Hz, 1H), 6.94 (dd, *J* = 0.8, 8.0 Hz, 1H), 6.84 (td, *J* = 0.8, 7.2 Hz, 1H), 5.55–5.53 (m, 1H), 4.76–4.68 (m, 1H), 2.34–2.29 (m, 1H), 2.19–2.09 (m, 1H), 1.44 (d, *J* = 6.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 168.1, 155.6, 134.1, 131.8, 129.2, 128.0, 123.3, 120.4, 118.8, 117.3, 69.1, 43.8, 35.8, 20.7; IR (KBr) cm⁻¹: 2934, 1651, 1601, 1580, 1500, 1465, 1380, 740; HRMS (ESI): calcd for C₁₈H₁₅NO₂ [M+H]⁺: 294.1130, found 294.1122; [α]_D²⁶ = +45.68 (*c* 0.52, CHCl₃); HPLC (Chiralcel AD-H, *n*-hexane/*i*-PrOH = 97/3, UV = 210 nm, flow rate = 0.3 mL/min) t_{R1} = 19.430 min (minor) and t_{R2} = 22.901 min (major), ee = 98%.

4.4 Procedure for synthesis of **6**



Diisopropyl azodicarboxylate (DIAD) (0.54 g, 2.7 mmol) in dry THF (5 ml) was added dropwise to a stirred solution of **1a** (0.36 g, 2.2 mmol), PPh₃, (0.68 g, 2.6 mmol) and methyl-5-imidazolecarboxylate (0.29 g, 2.3 mmol) in dry THF (15 ml) at -30°C before the mixture was warmed to room temperature and stirred for a further 1 h. Then the solvent was evaporated to provide crude product and it was purified by chromatography (PE:EA=10:1) to give pure product **6** as a white solid (0.6 g, 93%). Mp 115–117 °C ¹H NMR (400 MHz, CDCl₃) δ 7.81 (s, 1H), 7.30–7.26 (m, 1H), 7.09 (d, *J* = 7.6 Hz, 1H), 6.97–6.91 (m, 2H), 6.11 (s, 1H), 4.03–3.96 (m, 1H), 3.90 (s, 3H), 2.19–2.13 (m, 2H), 1.76 (s, 1H), 1.37 (d, *J* = 6.4 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 160.9, 155.9, 142.1, 138.8, 130.7, 130.6, 122.2, 121.1, 117.6, 117.1, 66.7, 51.6, 51.3, 36.8, 20.7; IR (KBr) cm⁻¹: 3098, 2934, 1725, 1601, 1580, 1500, 1465, 1380, 790; HRMS (ESI): calcd for C₁₈H₁₅NO₂ [M+H]⁺: 273.1239, found 273.1233; [α]_D²⁶ = +35.48 (*c* 0.52, CHCl₃); HPLC (Chiralcel OD-H, *n*-hexane/*i*-PrOH = 97/3, UV = 210 nm, flow rate = 0.5 mL/min) t_{R1} = 42.046 min (major) and t_{R2} = 49.029 min (minor), ee = 99.9%.

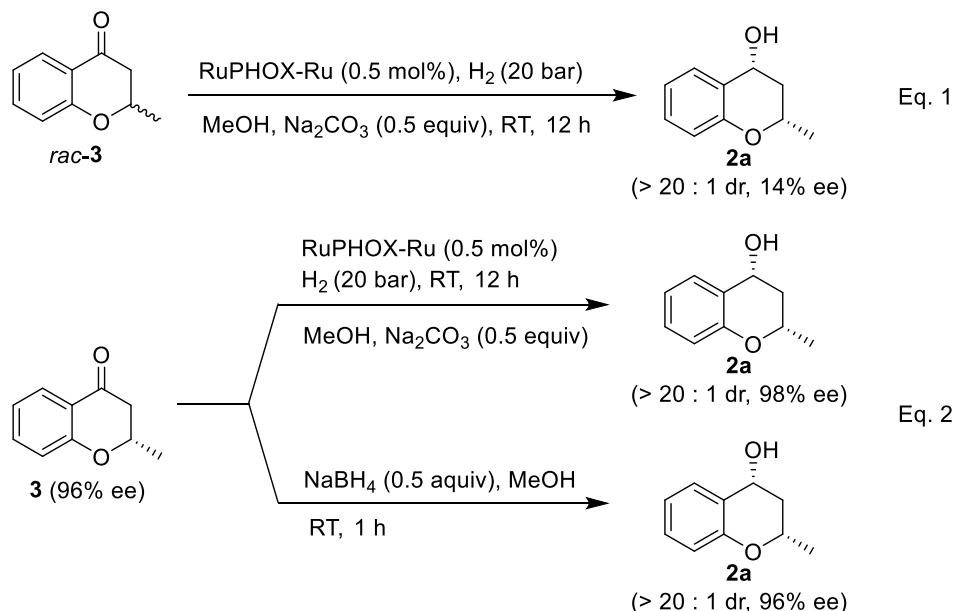
4.5 Procedure for synthesis of **7**^[2]



2f (0.15 mmol) was treated with PCC (120 mg) in CH₂Cl₂ (2 mL) at room temperatures for 10 min before evaporation of DCM. Then Et₂O was added to the residue. The resulting mixture was filtered

through a plug of silica using Et₂O. The solvent was evaporated to give crude product **7** as colorless oil (36.3 mg, 96%); ¹H NMR (400 MHz, CDCl₃) δ 7.91 (d, *J* = 7.6 Hz, 1H), 7.51 (t, *J* = 7.2 Hz, 1H), 7.34–7.25 (m, 5H), 7.04 (d, *J* = 7.6 Hz, 2H), 4.49–4.43 (m, 1H), 2.98–2.83 (m, 2H), 2.79–2.69 (m, 2H), 2.30–2.21 (m, 1H), 2.07–1.98 (m, 1H); [α]_D²⁶ = +59.89 (*c* 0.47, CHCl₃); HPLC (Chiralcel OD-H, *n*-hexane/*i*-PrOH = 90/10, UV = 210 nm, flow rate = 1.0 mL/min) t_{R1} = 15.848 min (minor) and t_{R2} = 28.636 min (major), ee = 98%.

5. Control Experiments to Verify Reaction Pathway



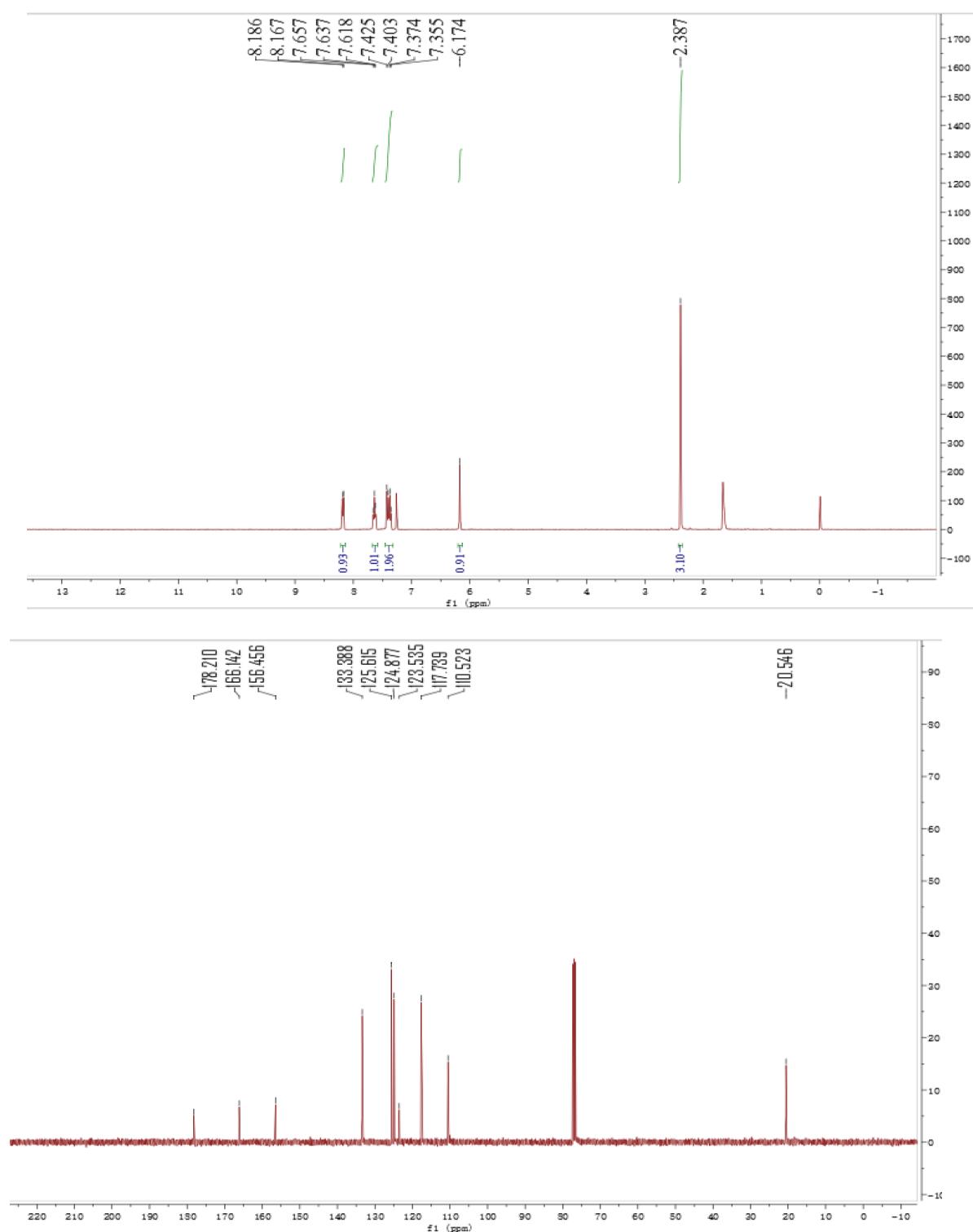
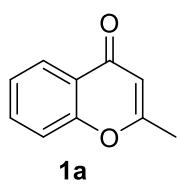
To further explore the reaction pathway, the asymmetric hydrogenation of racemic **3** was carried out under the optimal reaction conditions (Eq. 1). To our surprise, the desired product **2a** was obtained with up to 14% ee and > 20 : 1 dr. Subsequently, chiral intermediate **3** (96% ee), obtained via oxidation of chiral **2a** from the above mentioned gram-scale synthesis, was reduced under the same reaction conditions, affording **2a** in quantitative yield, 98% ee and > 20 : 1 dr (Eq. 1, top). These results indicated that the asymmetric catalytic hydrogenation of **3** proceeds together with a dynamic kinetic resolution process. Nevertheless, only a slight effect is observed in our case because the asymmetric catalytic hydrogenation was carried out using a weak base (Na₂CO₃) and the chiral center may be established in the first step. To verify this process, the chiral intermediate **3** (96% ee) used above was also reduced by NaBH₄ in MeOH, with **2a** being obtained in high yield, 96% ee and > 20 : 1 dr (Eq. 1, bottom). The results suggested that the subsequent hydrogenation of the C=O double bond can be directed by the previously formed carbon chiral center.

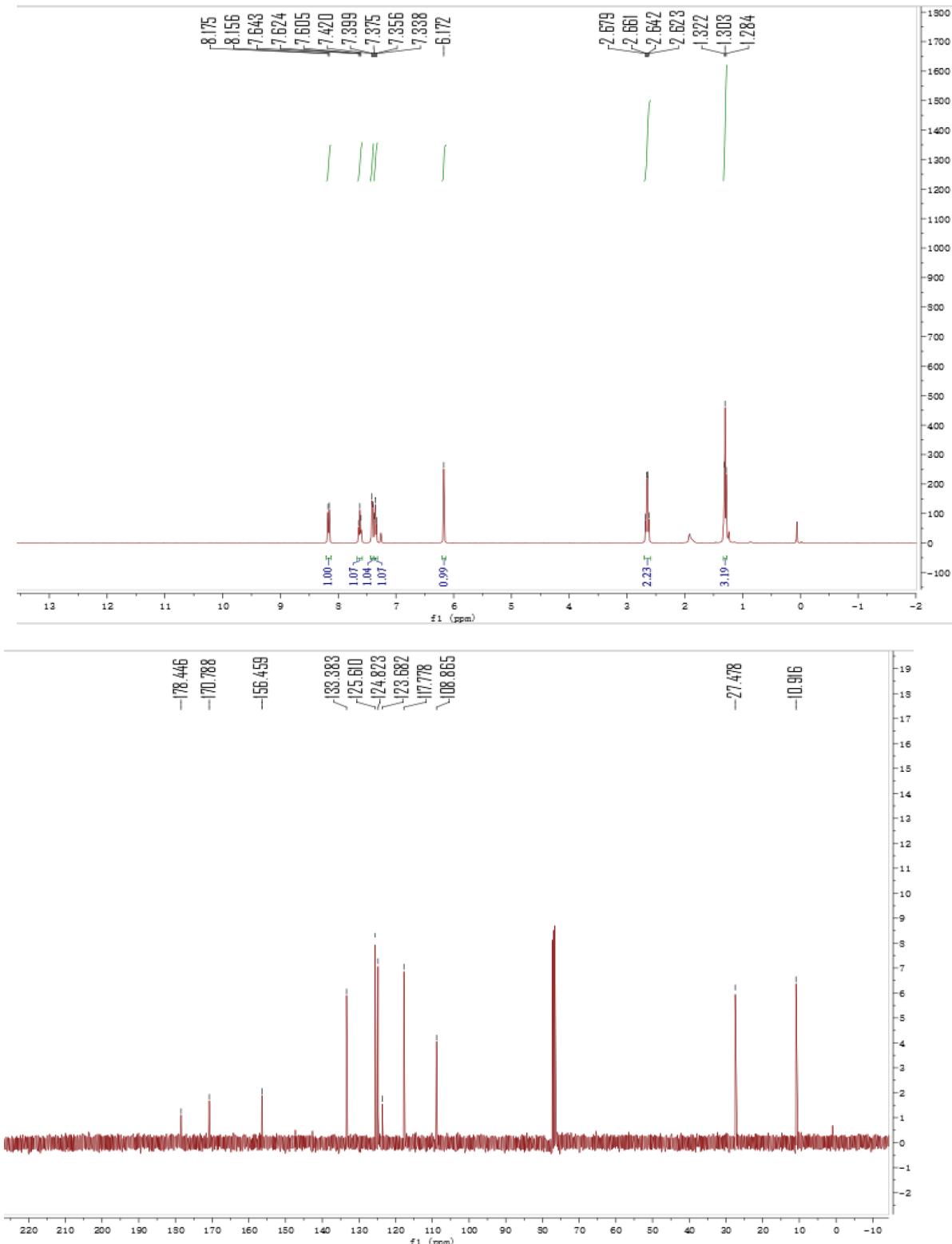
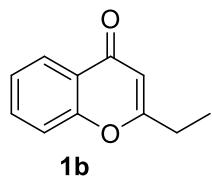
6. Reference

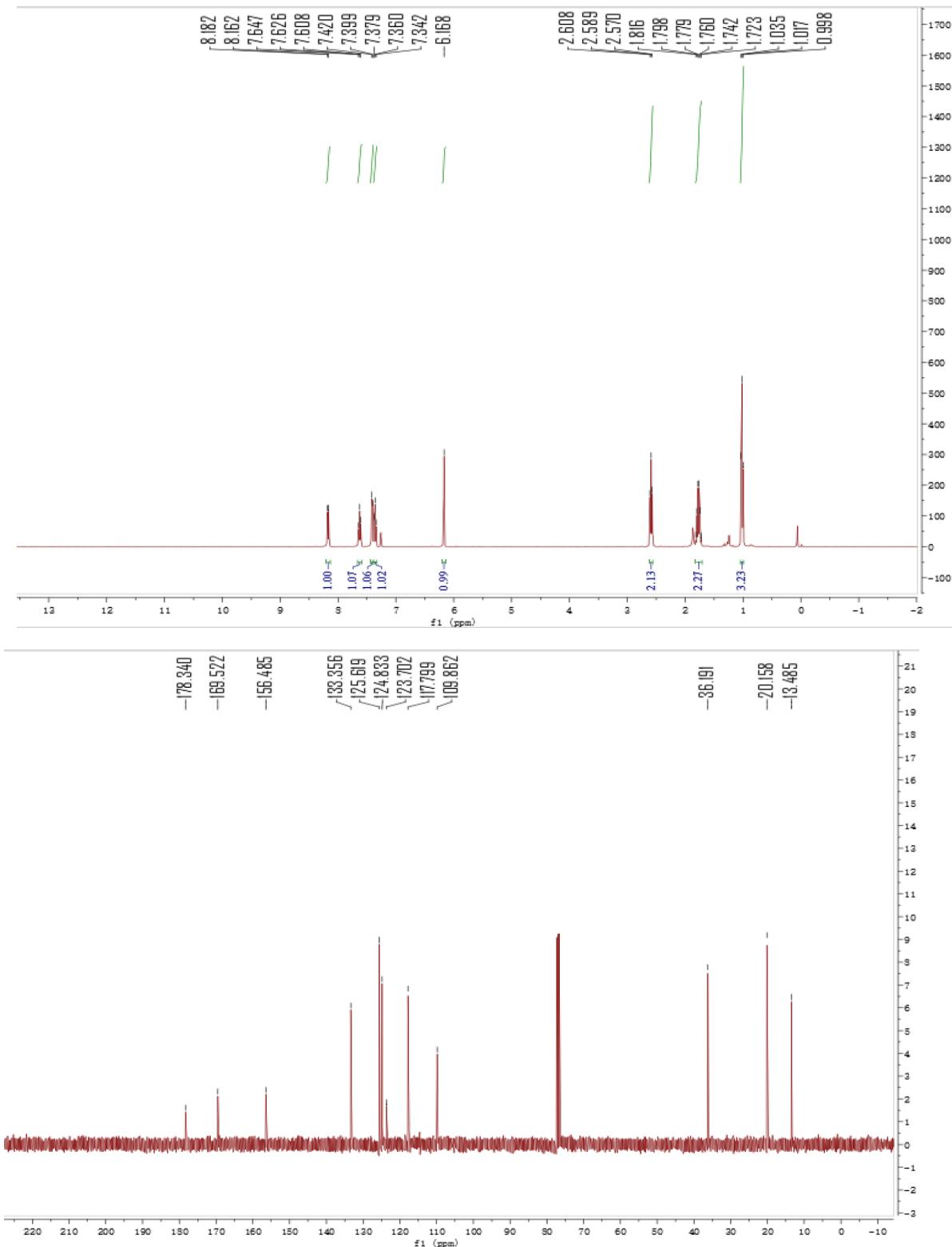
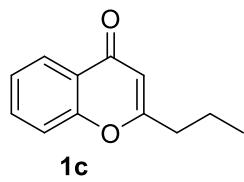
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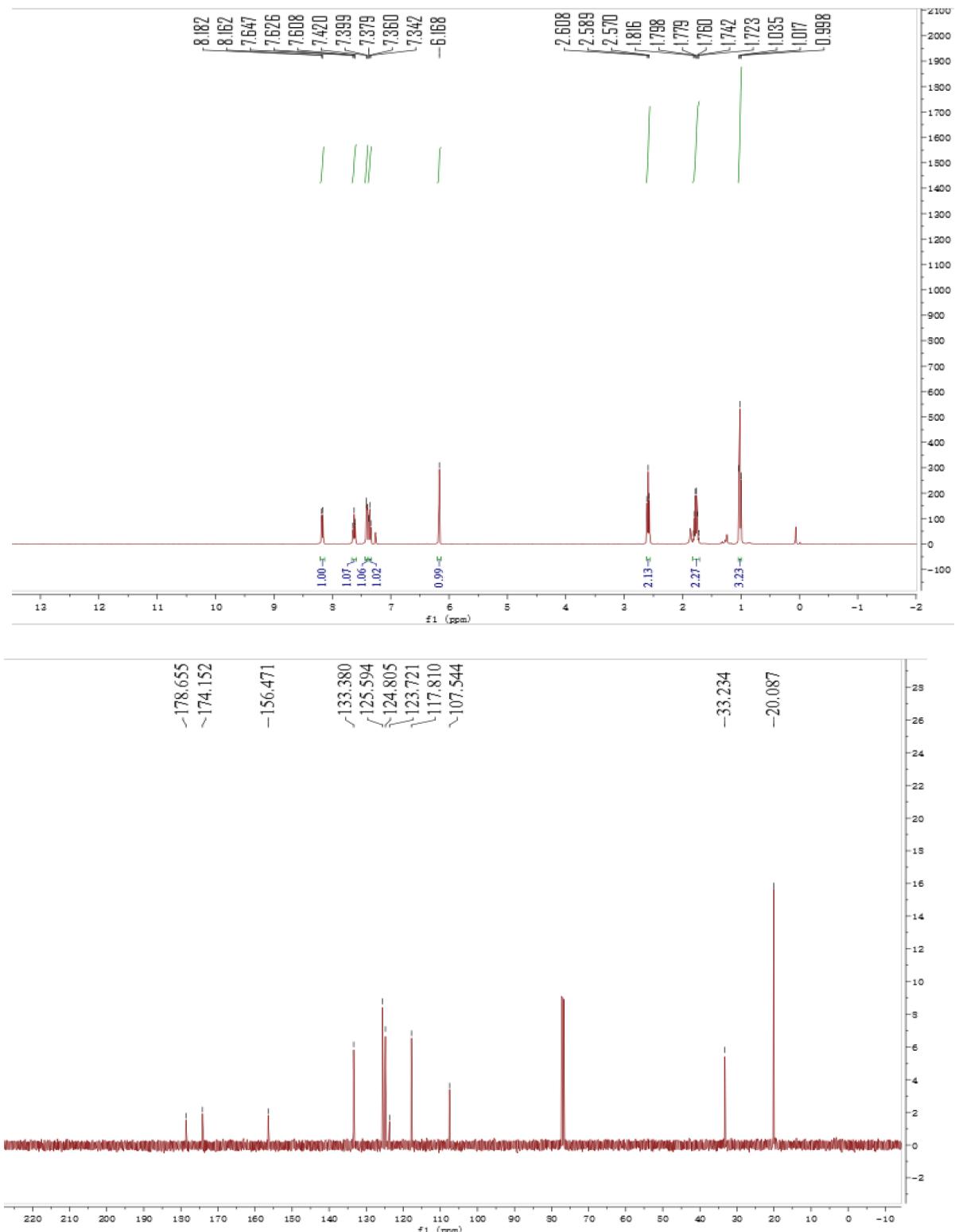
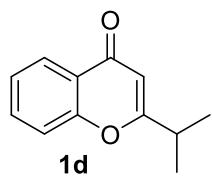
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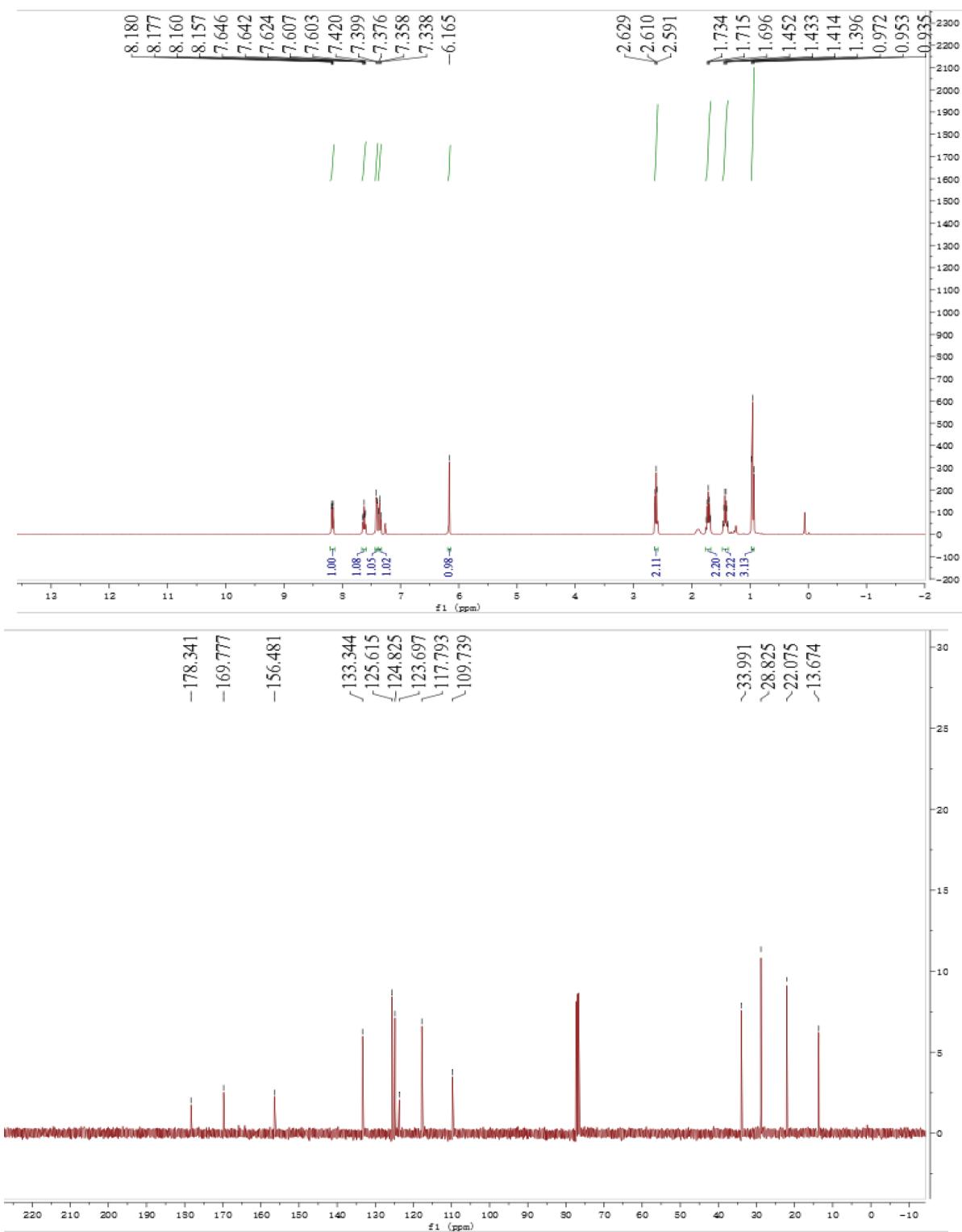
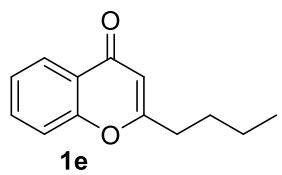
7. NMR spectra

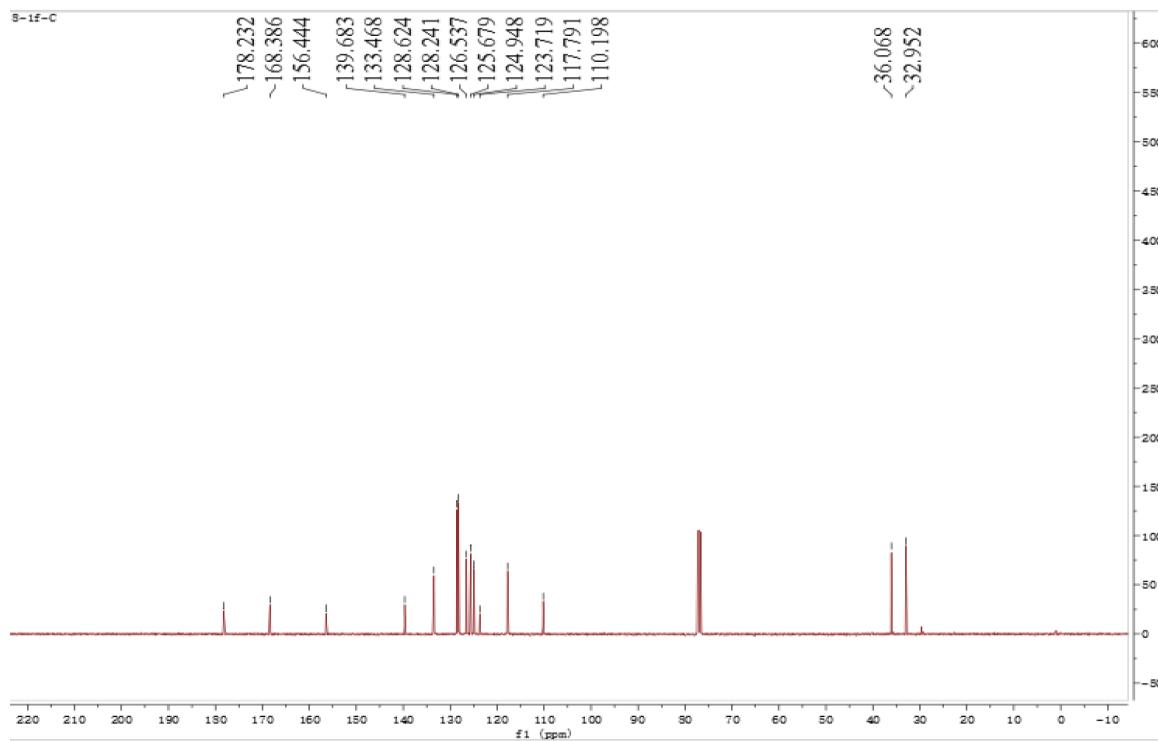
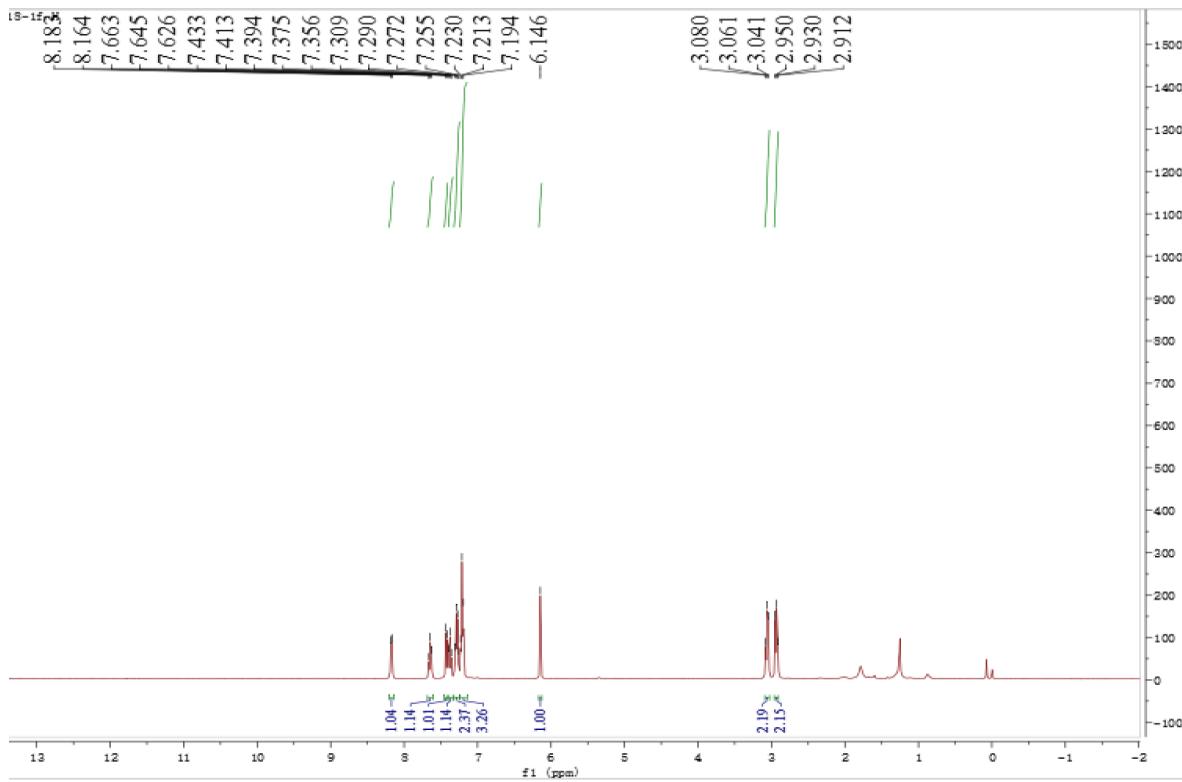
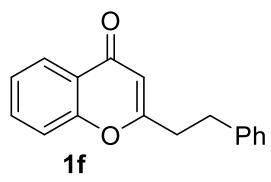


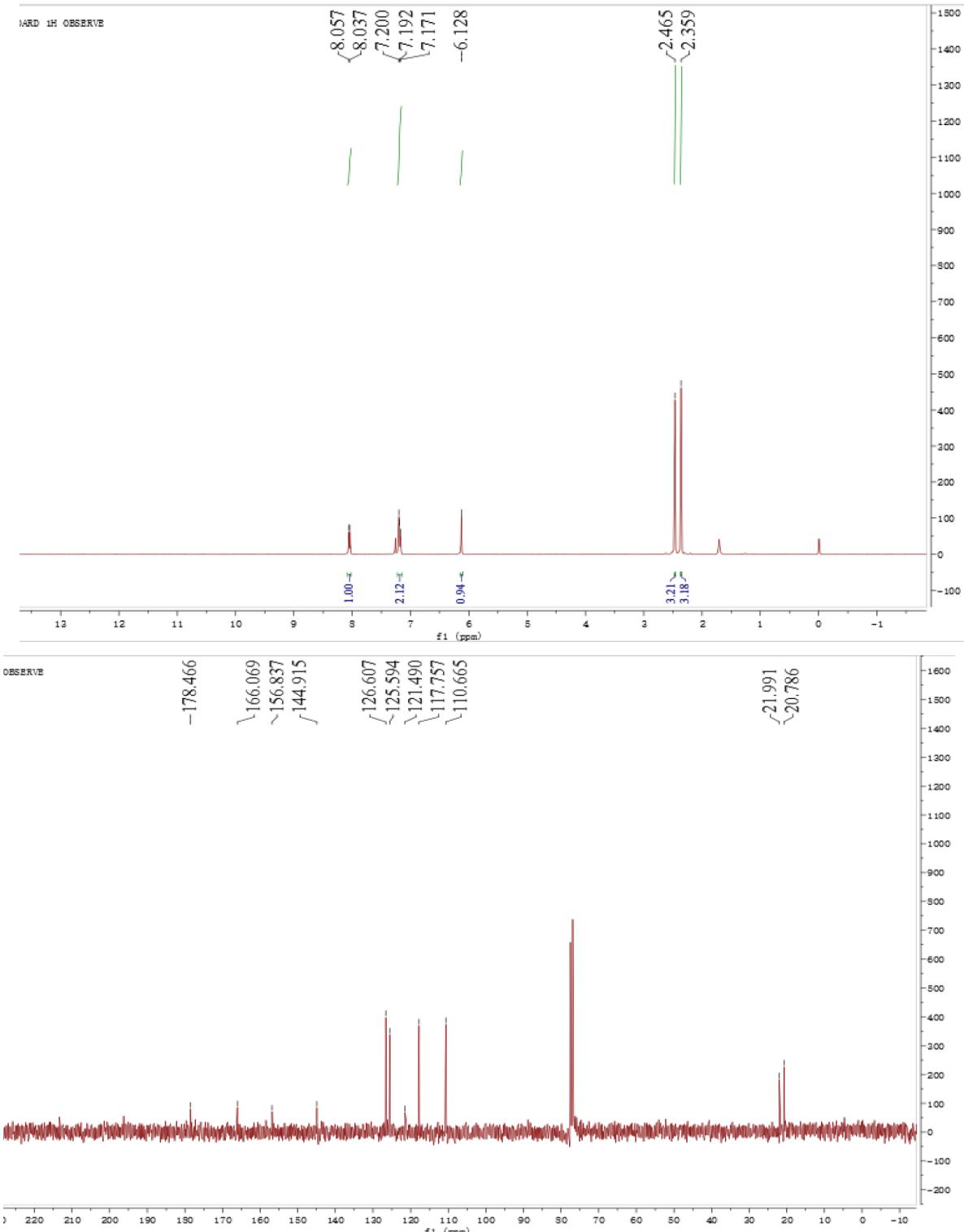
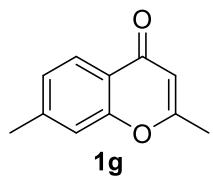


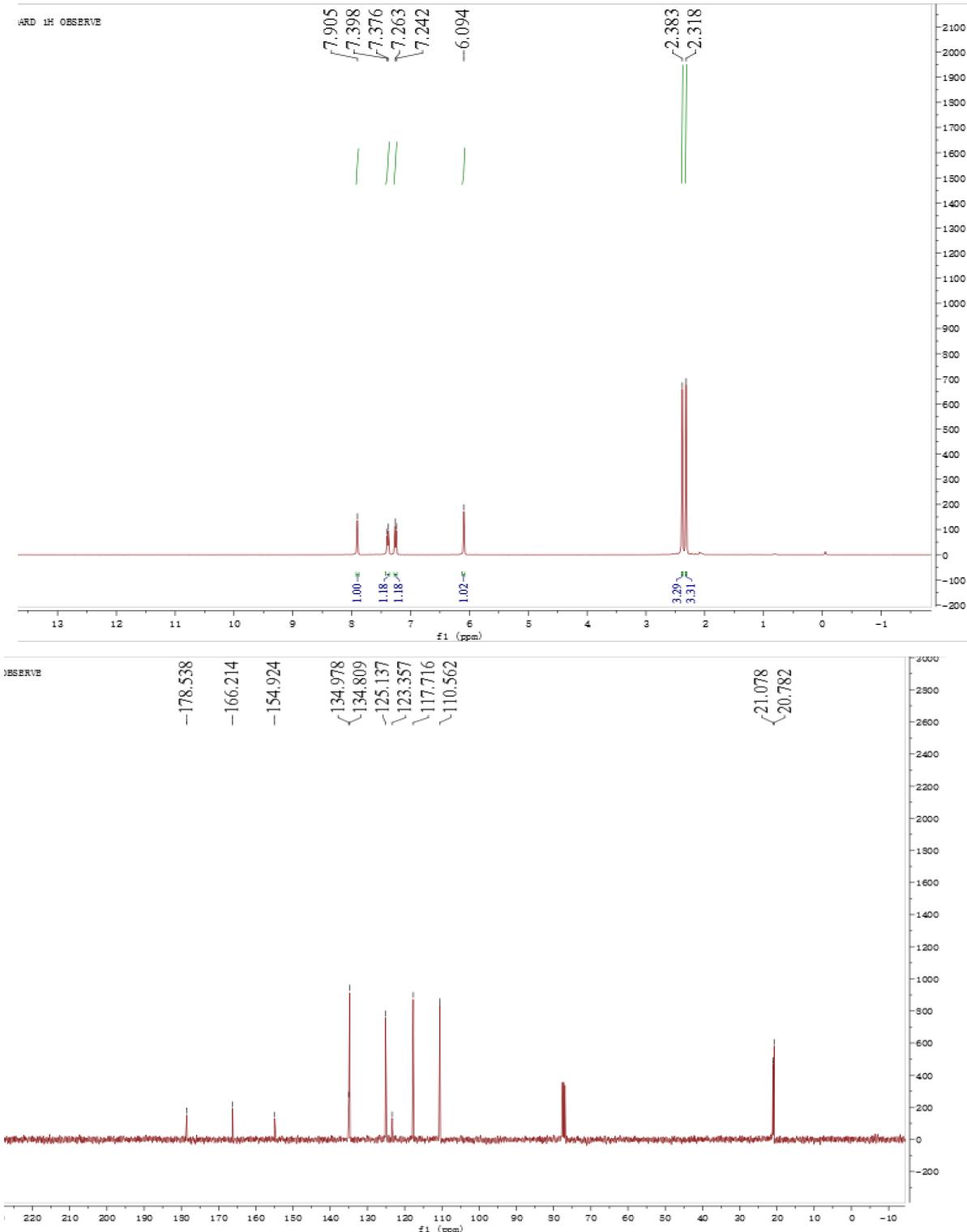
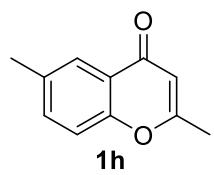


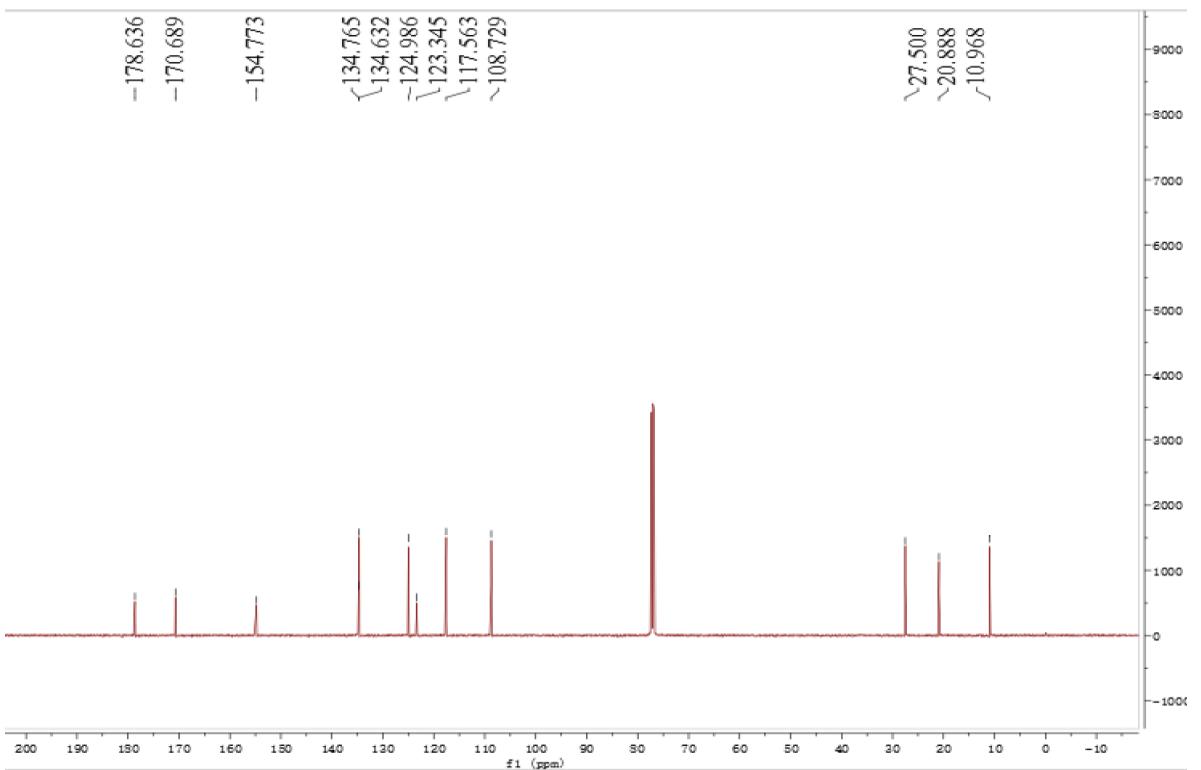
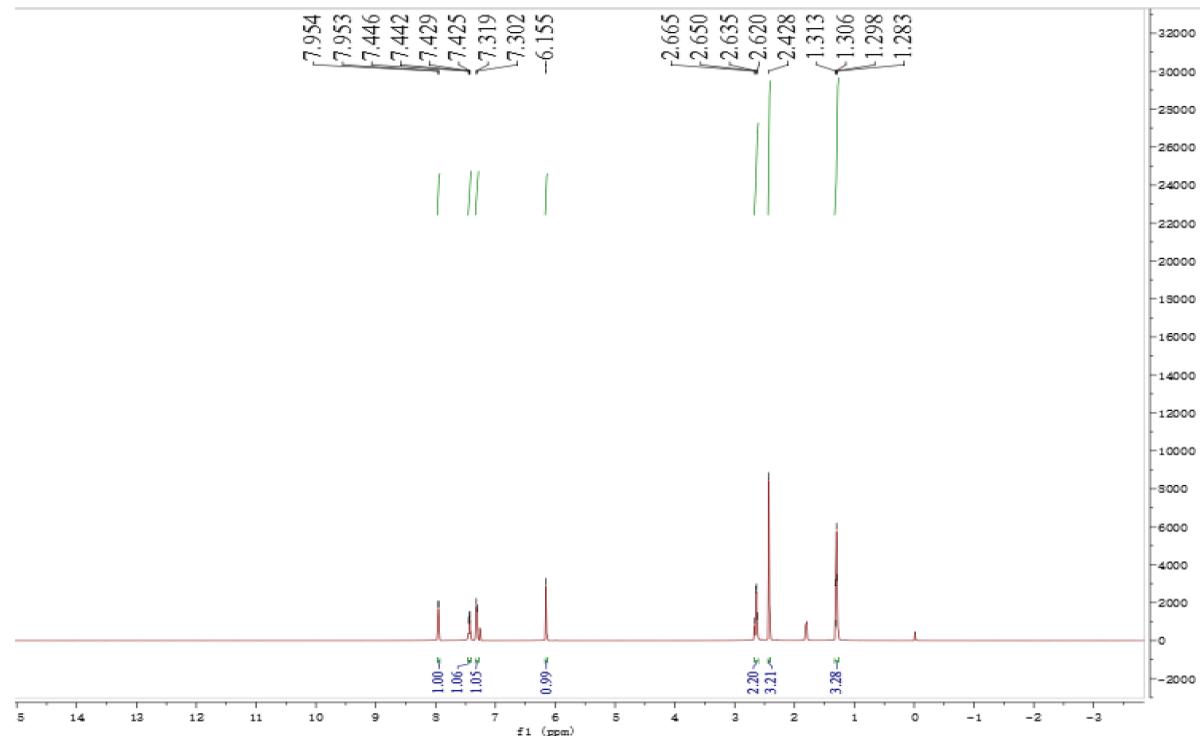
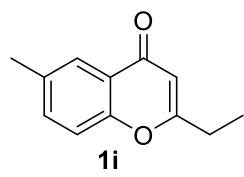


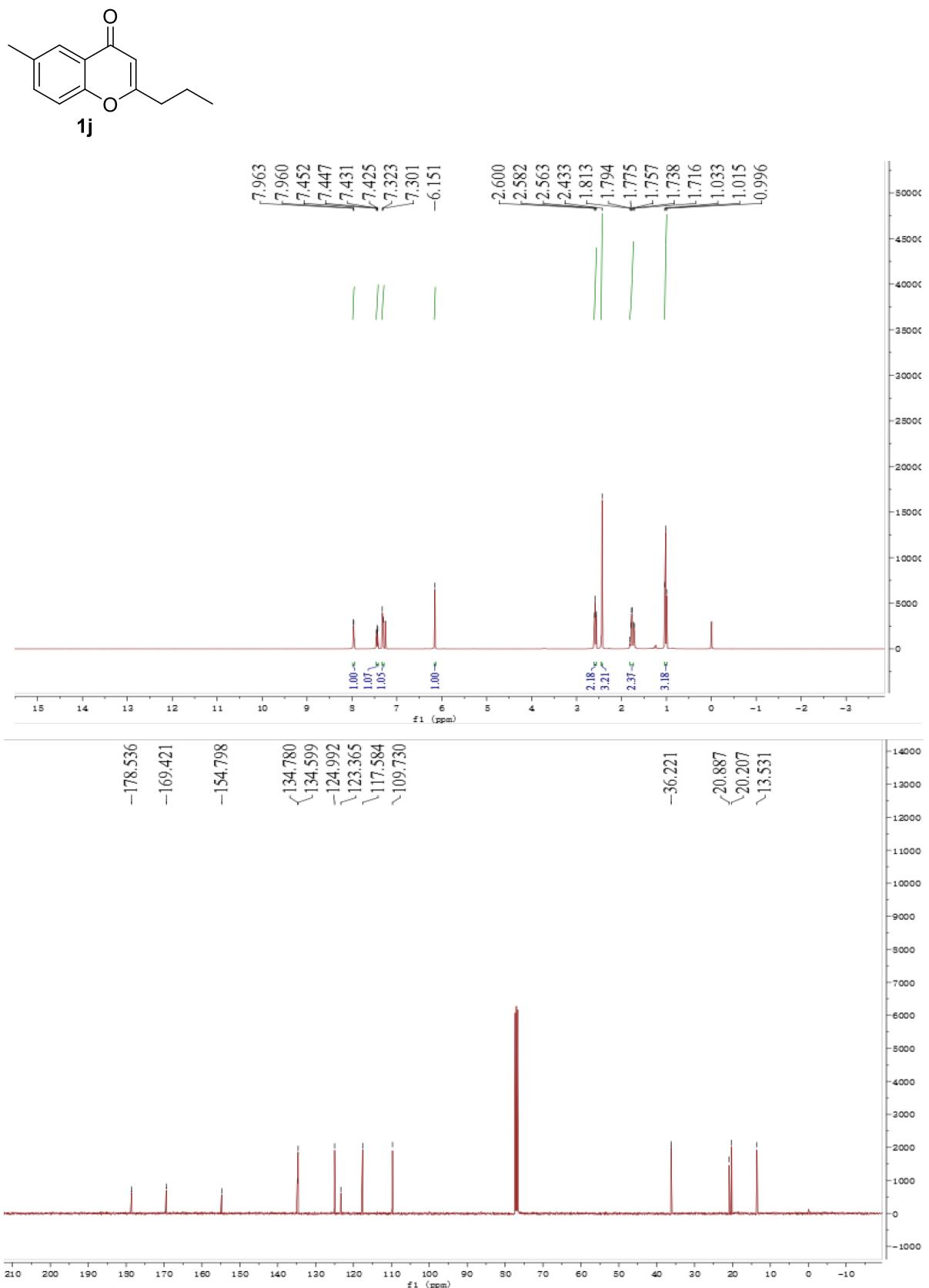


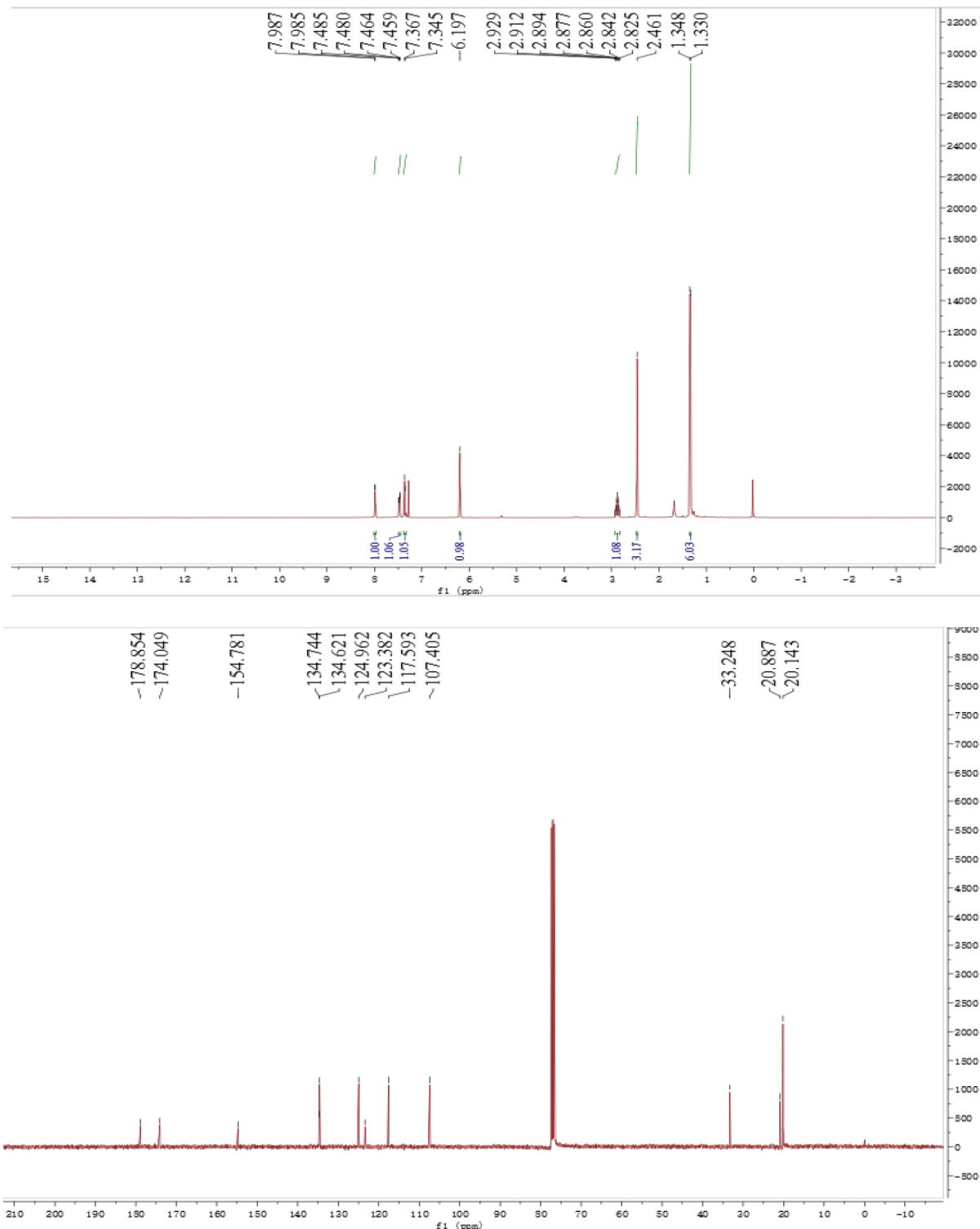
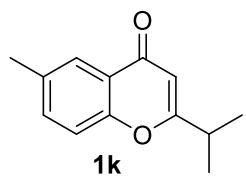


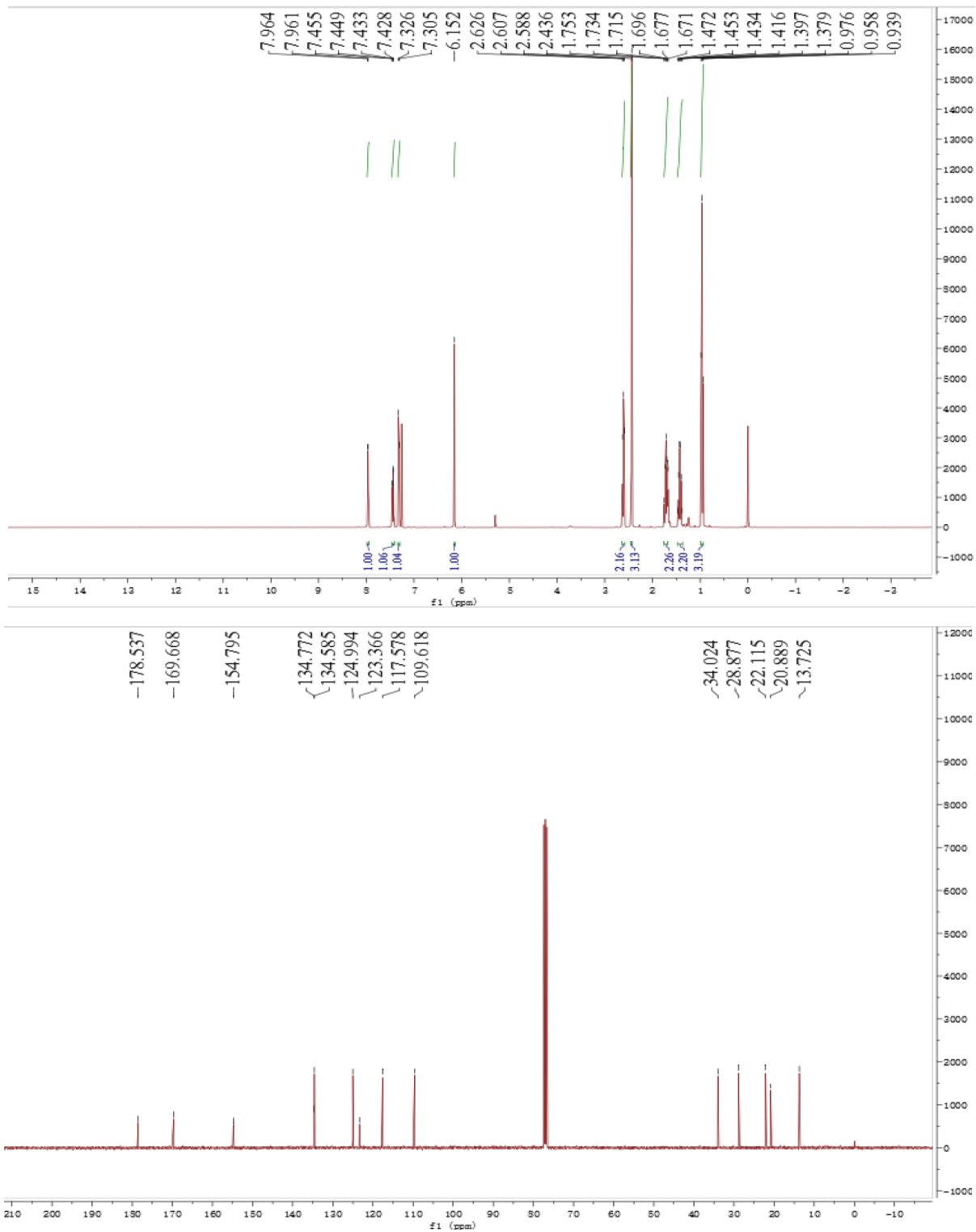
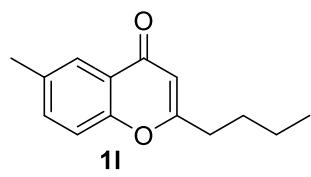


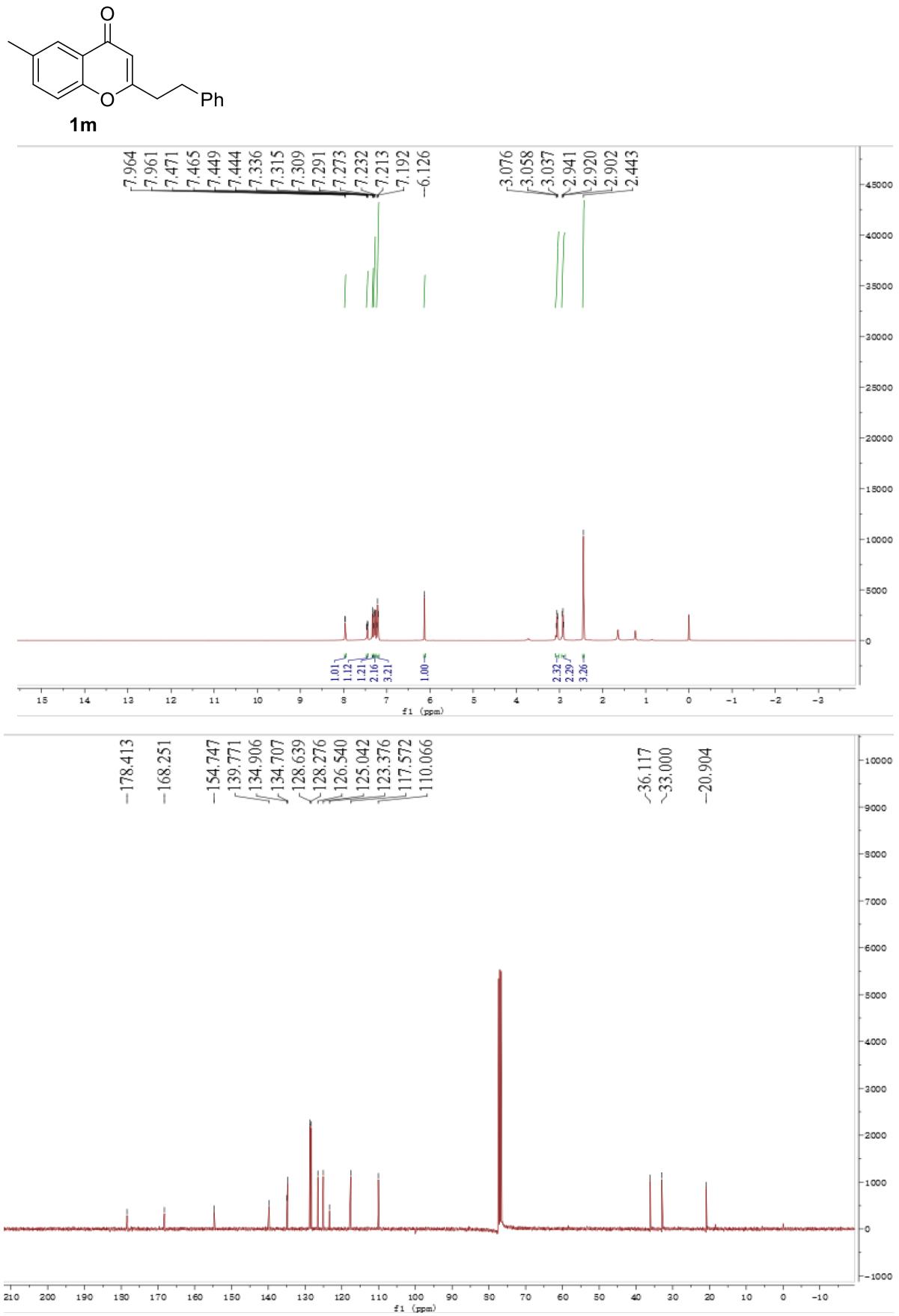


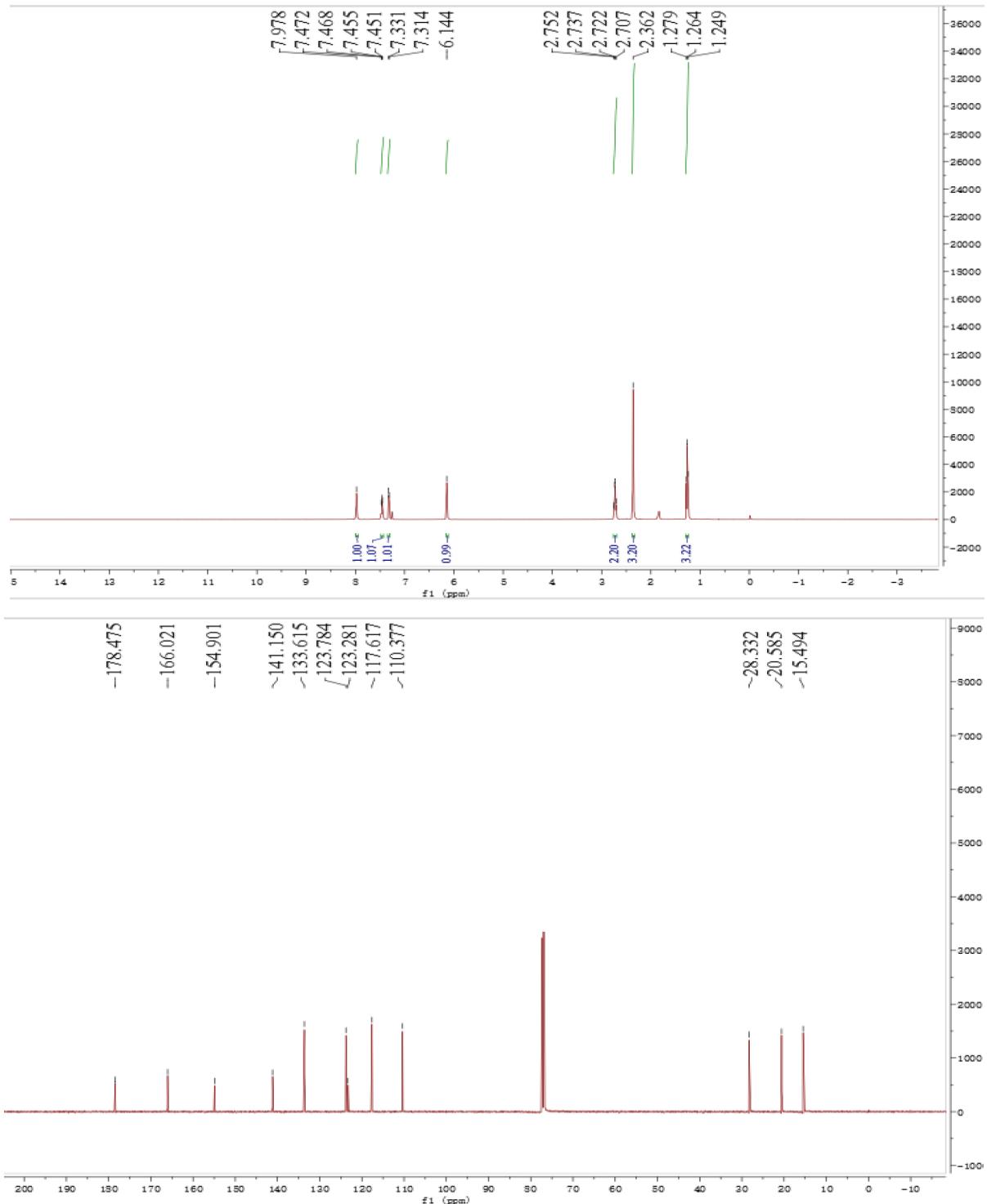
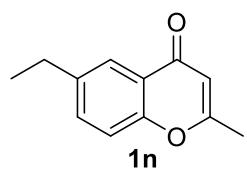


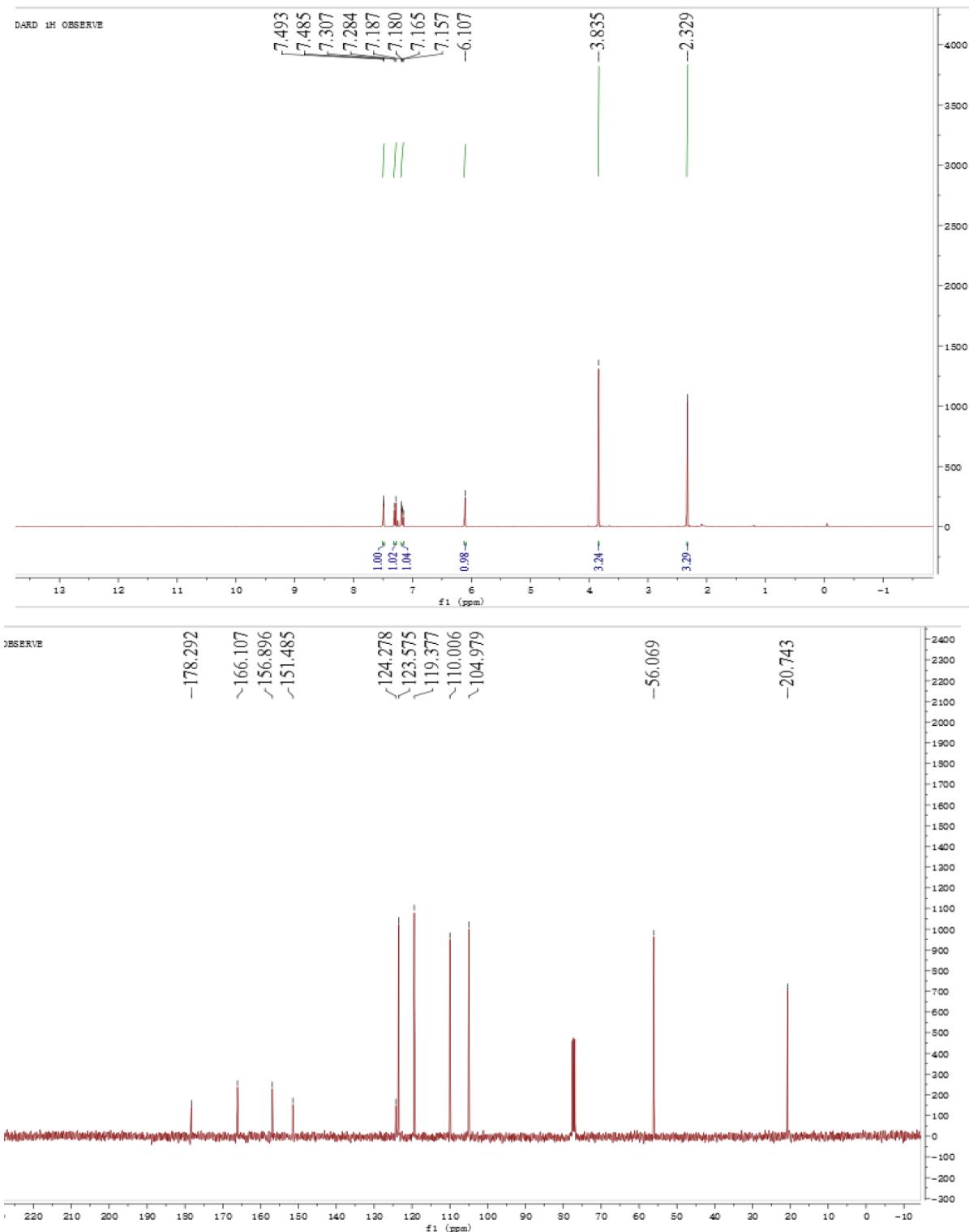
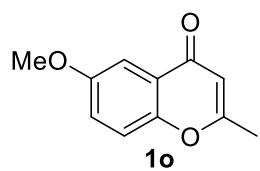


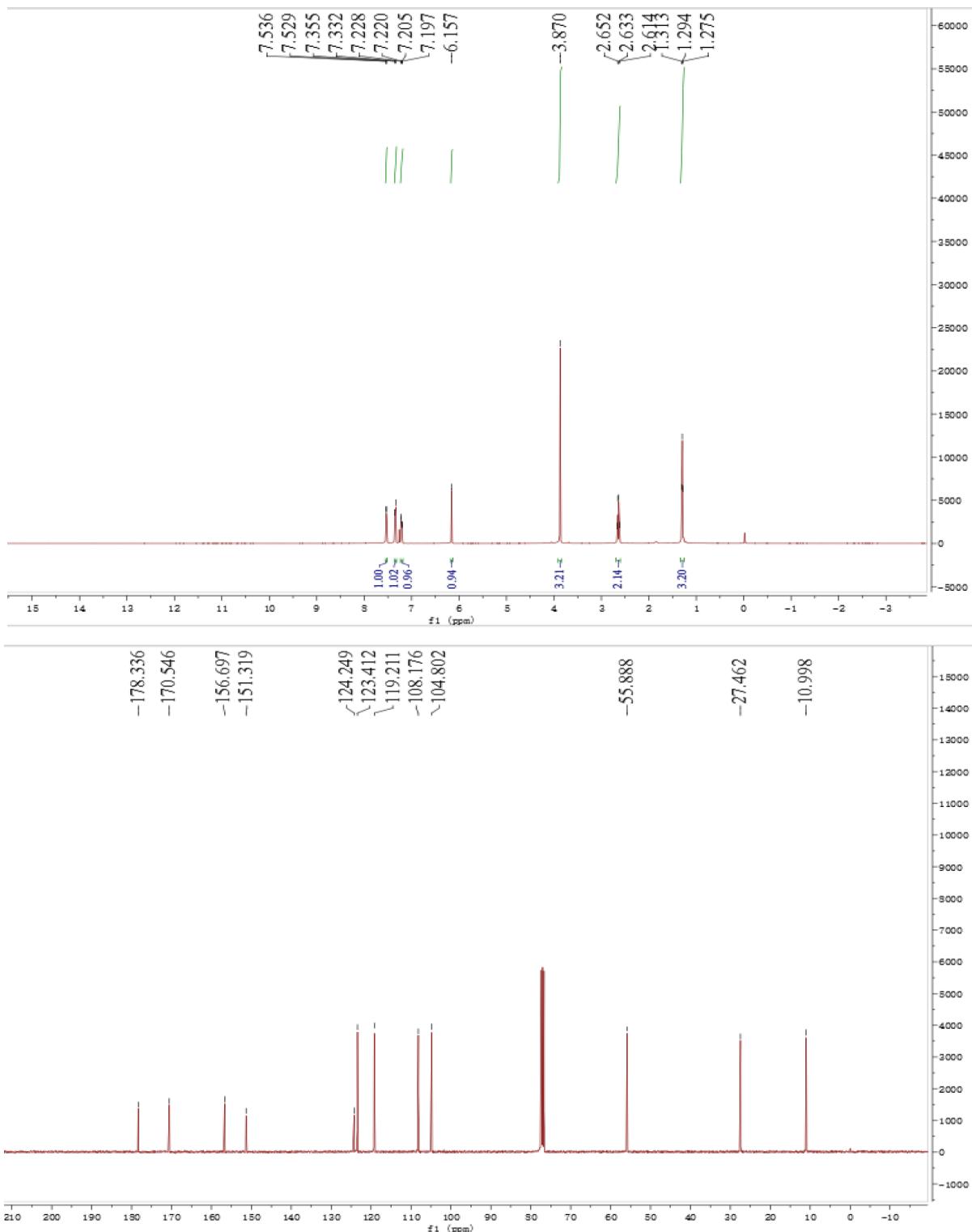
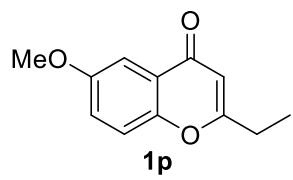


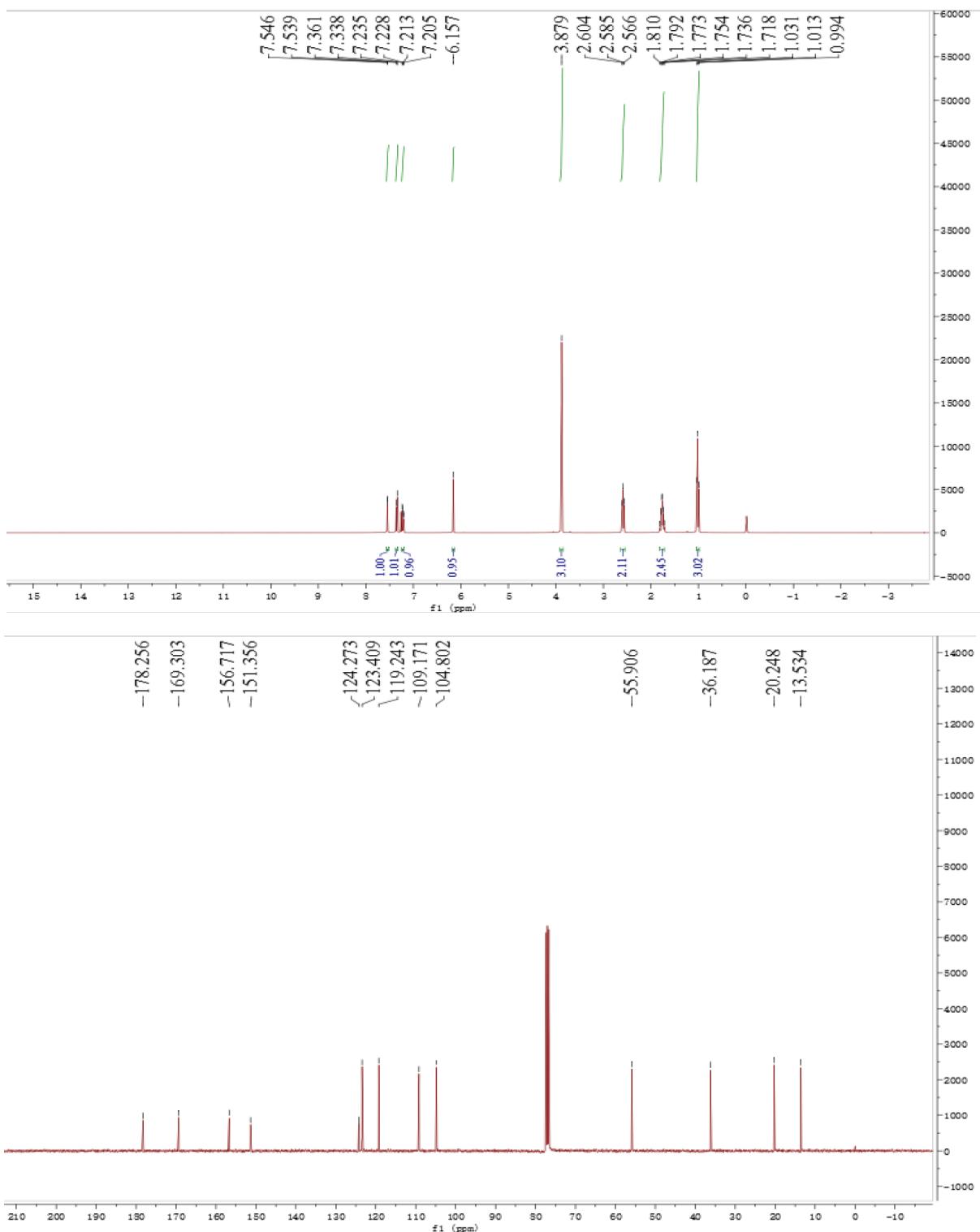
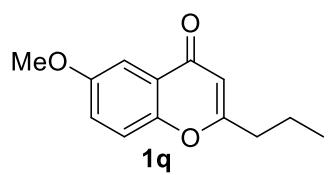


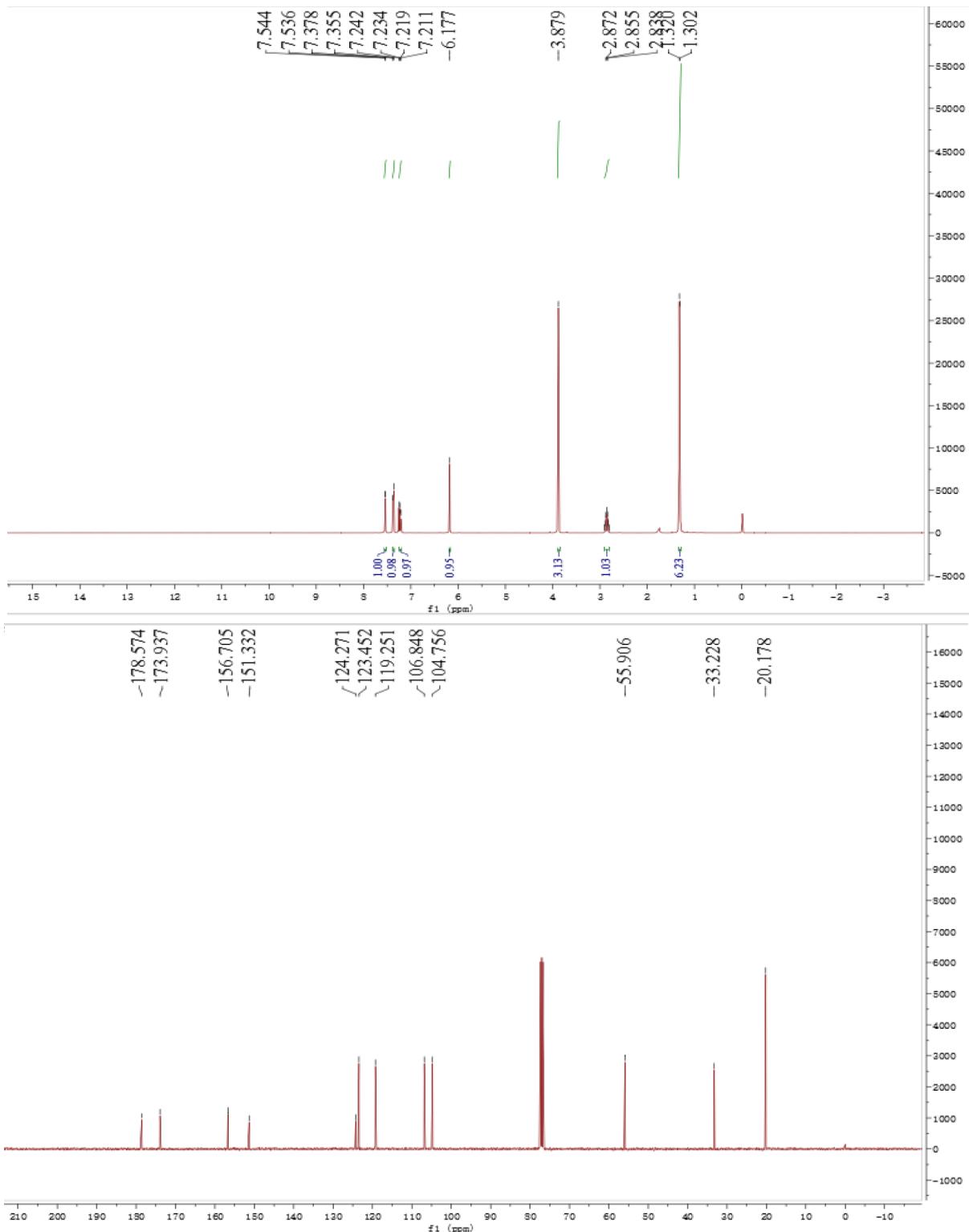
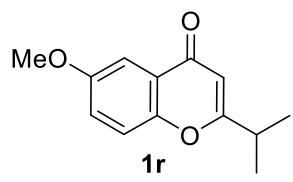


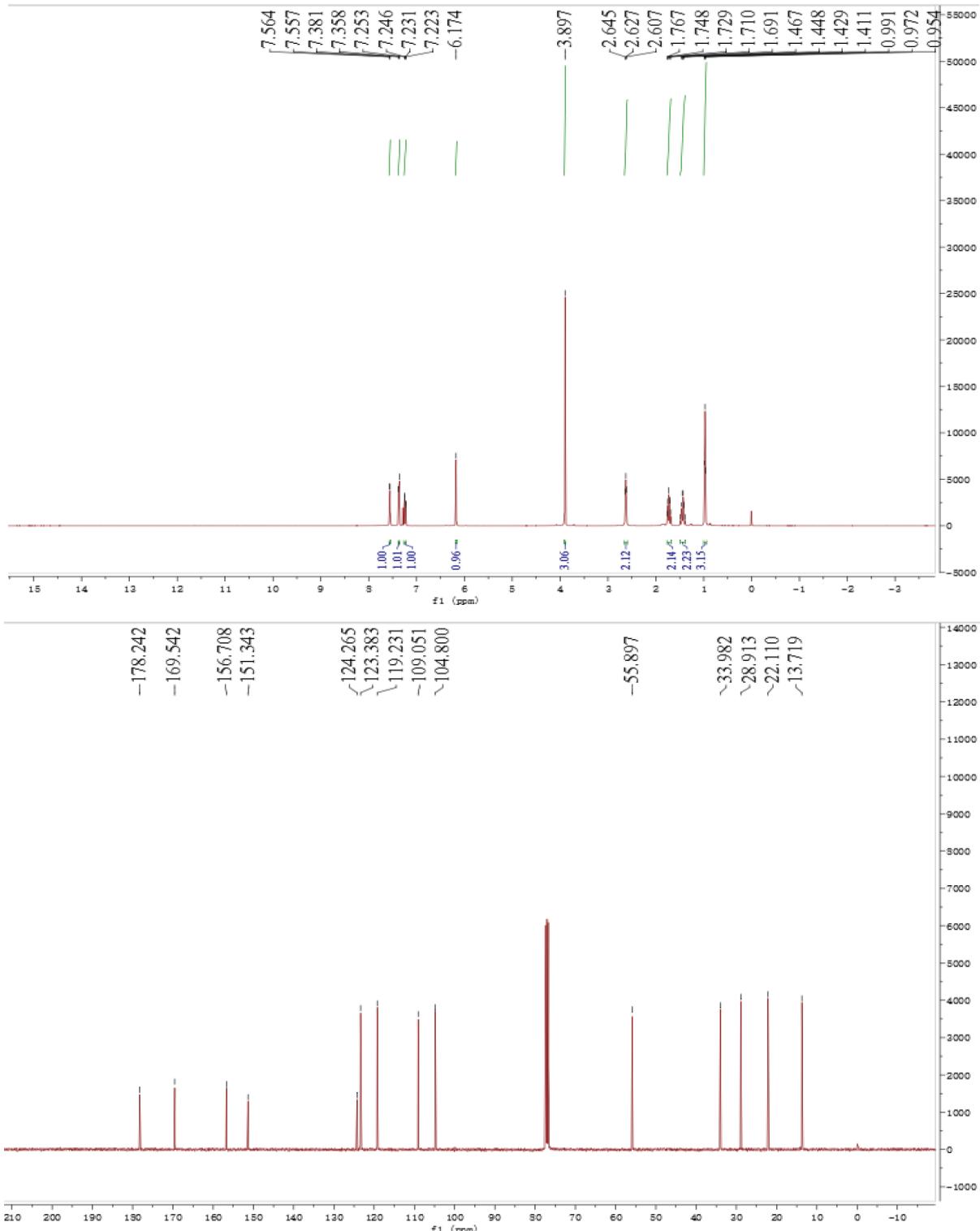
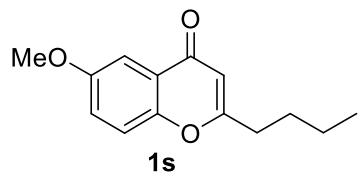


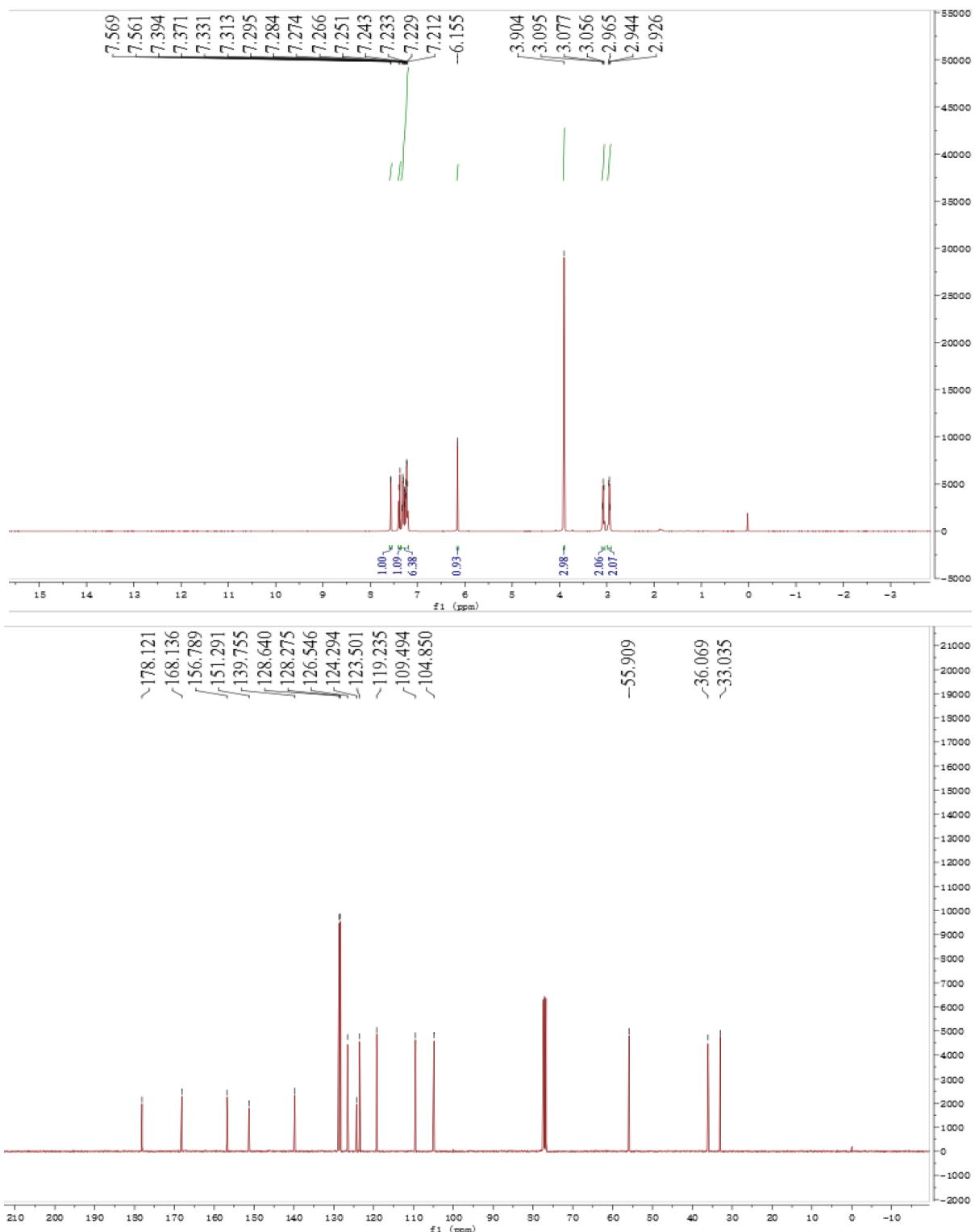
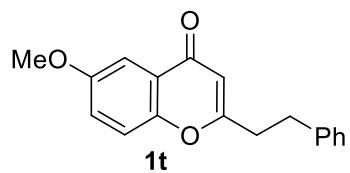


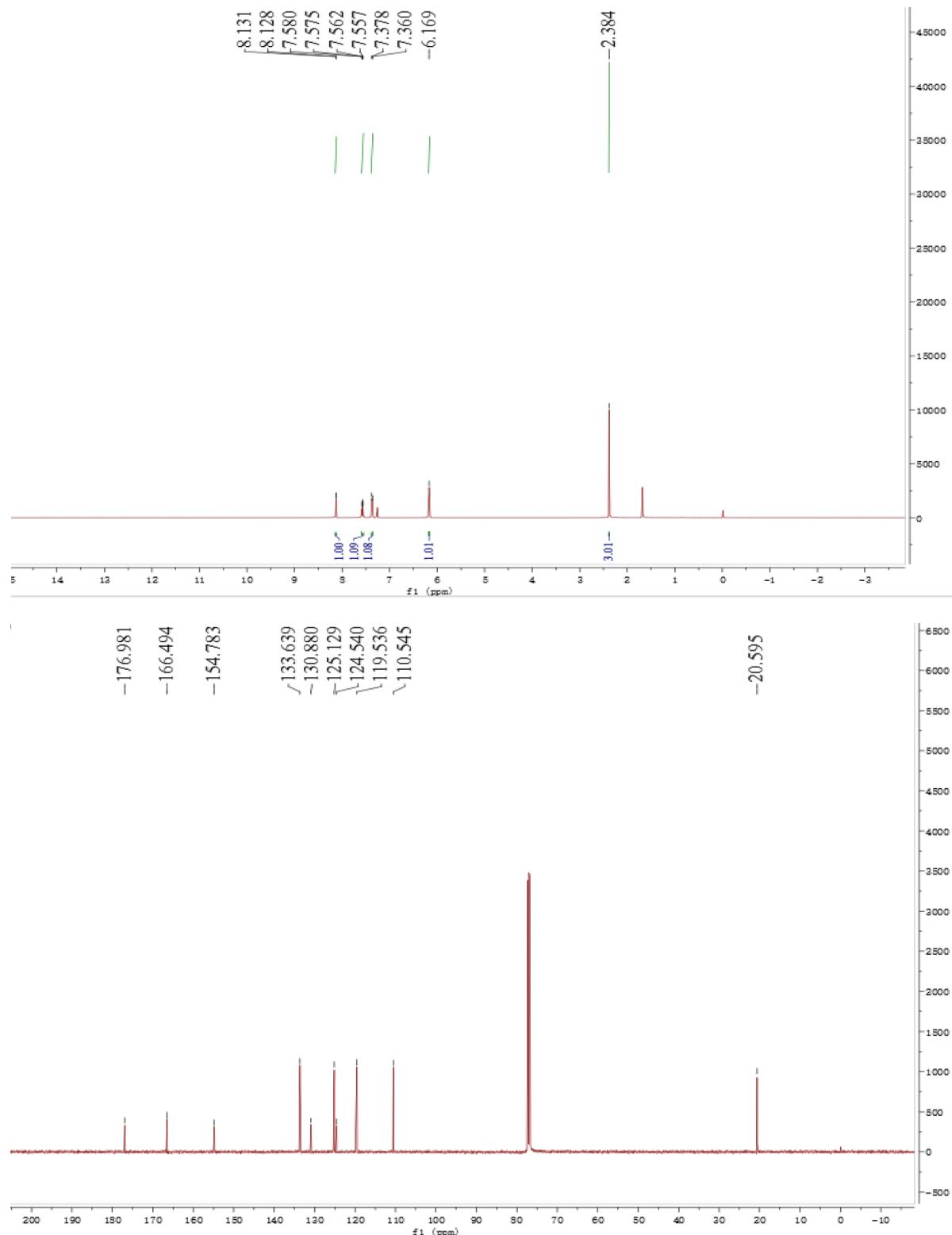
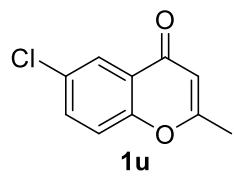


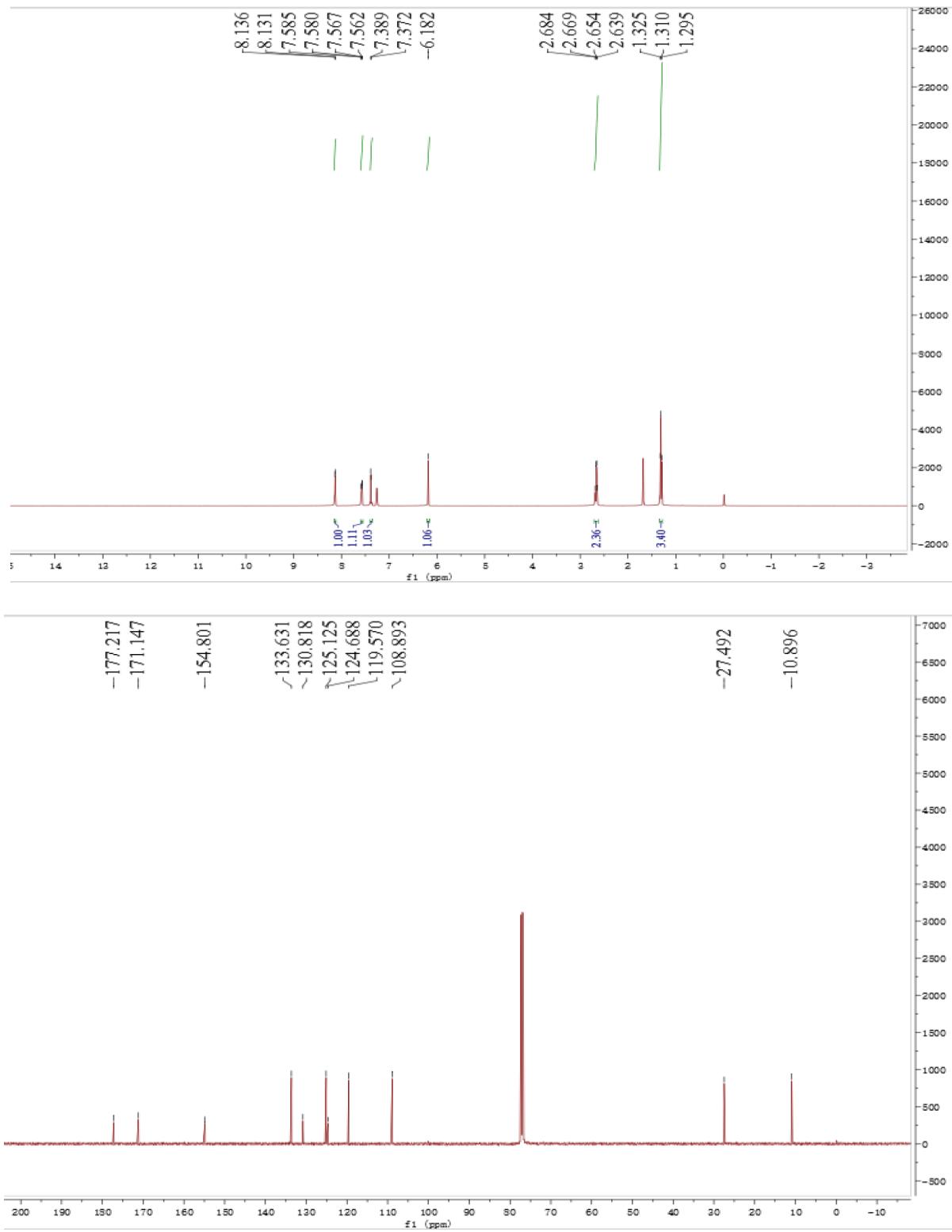
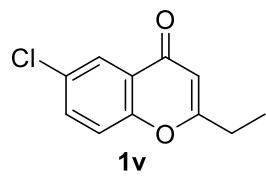


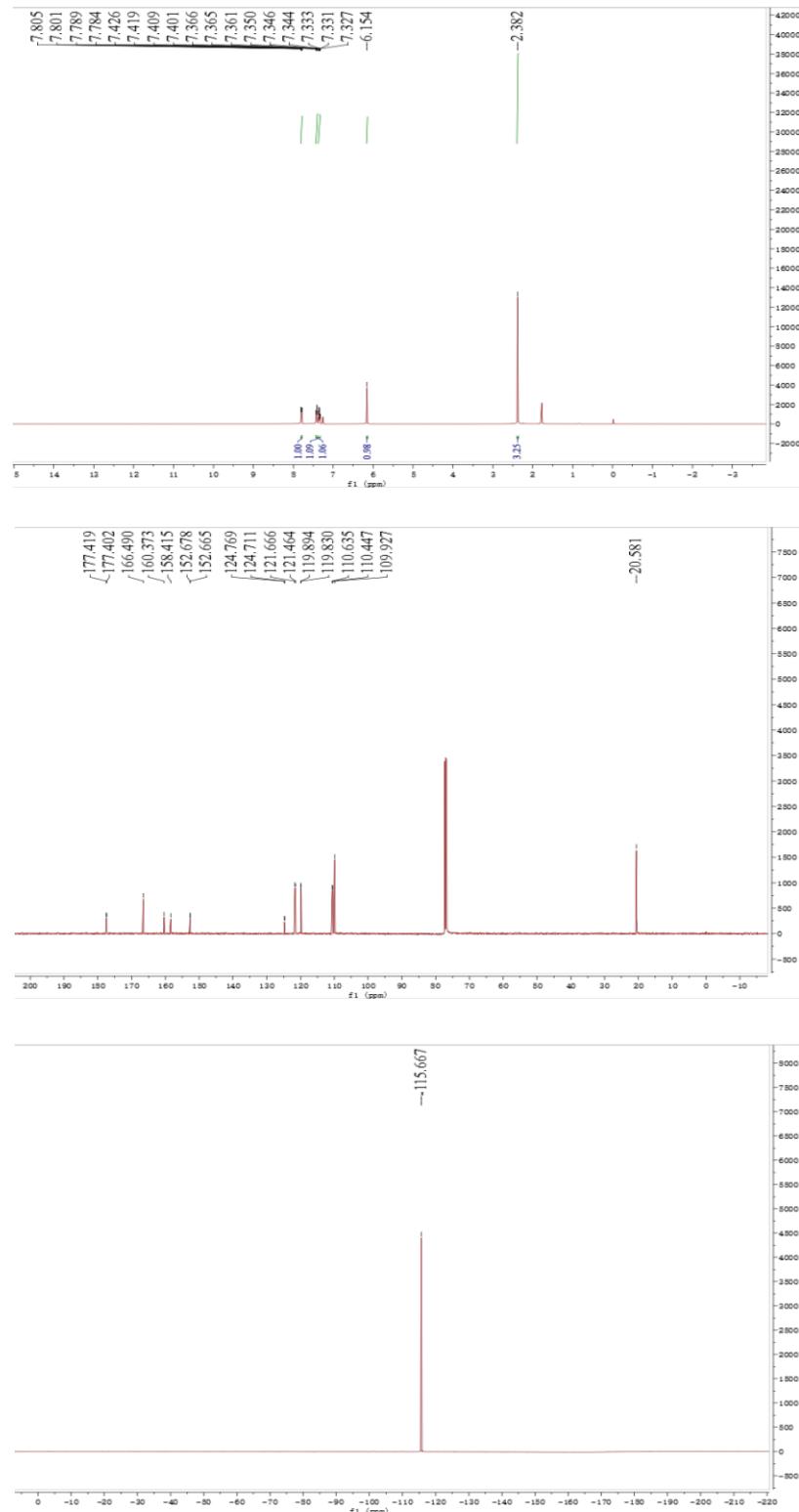
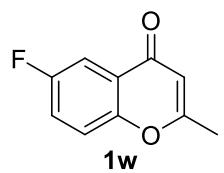


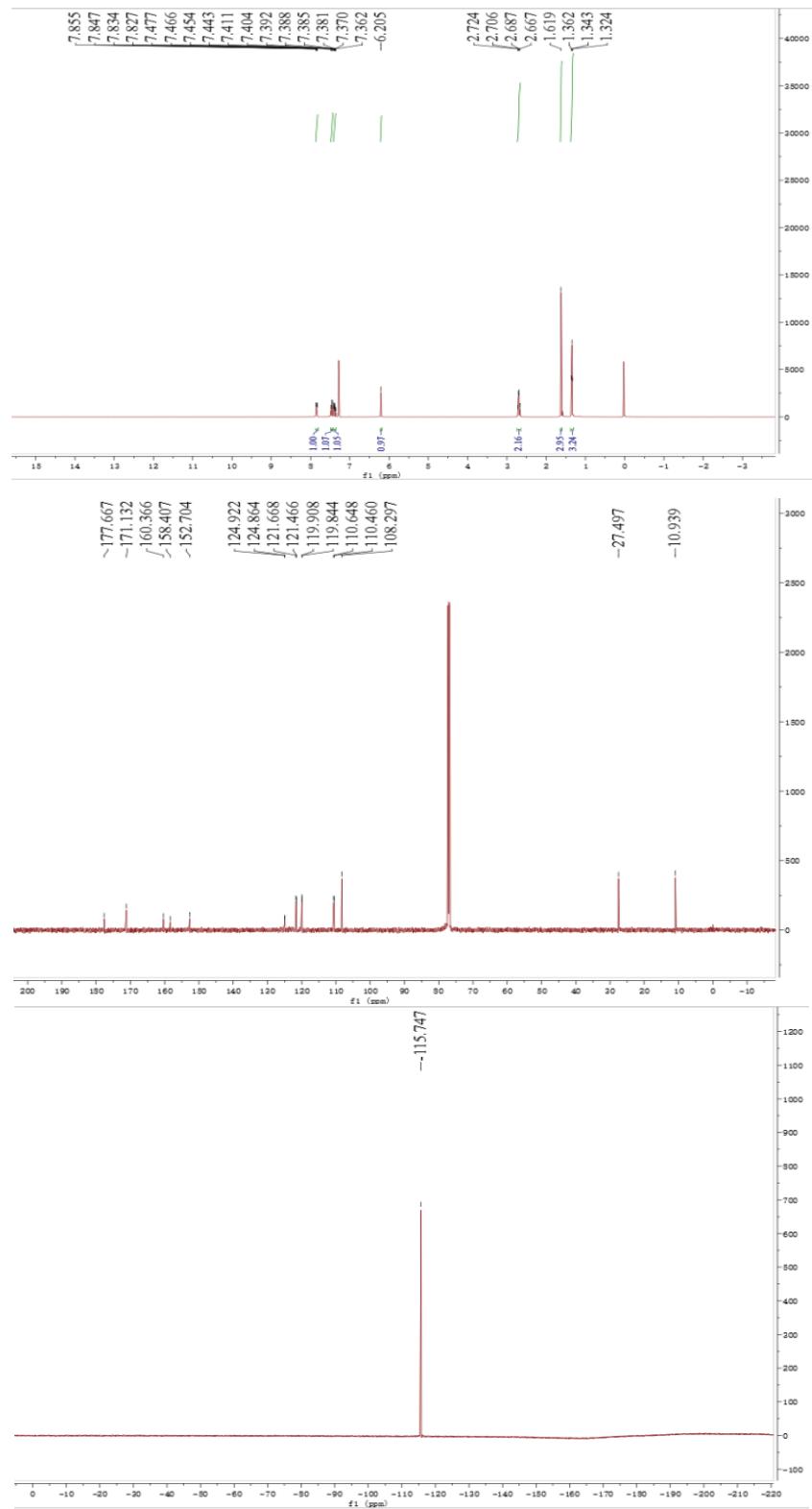
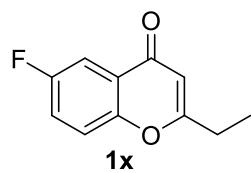


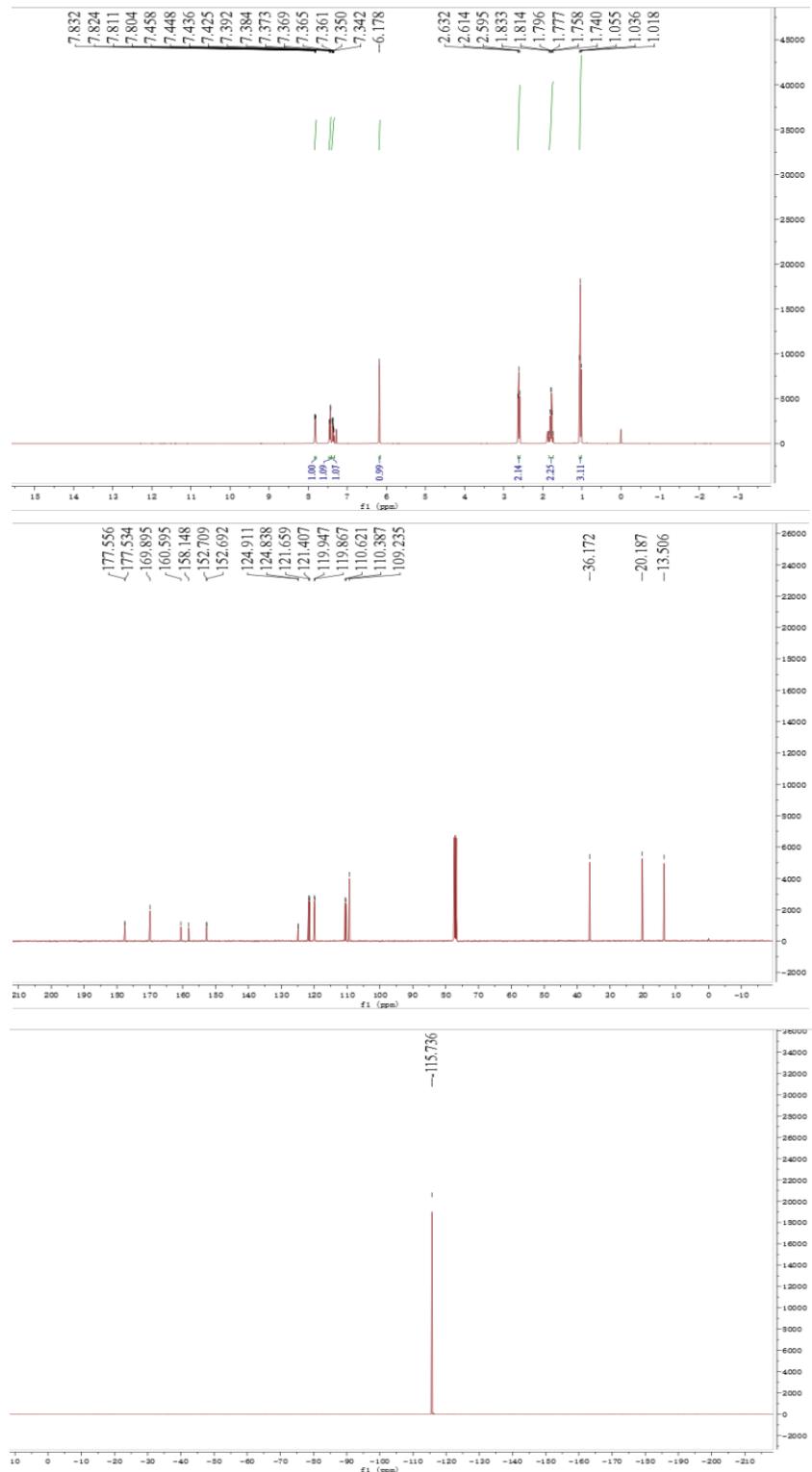
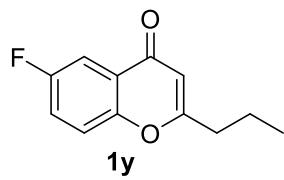


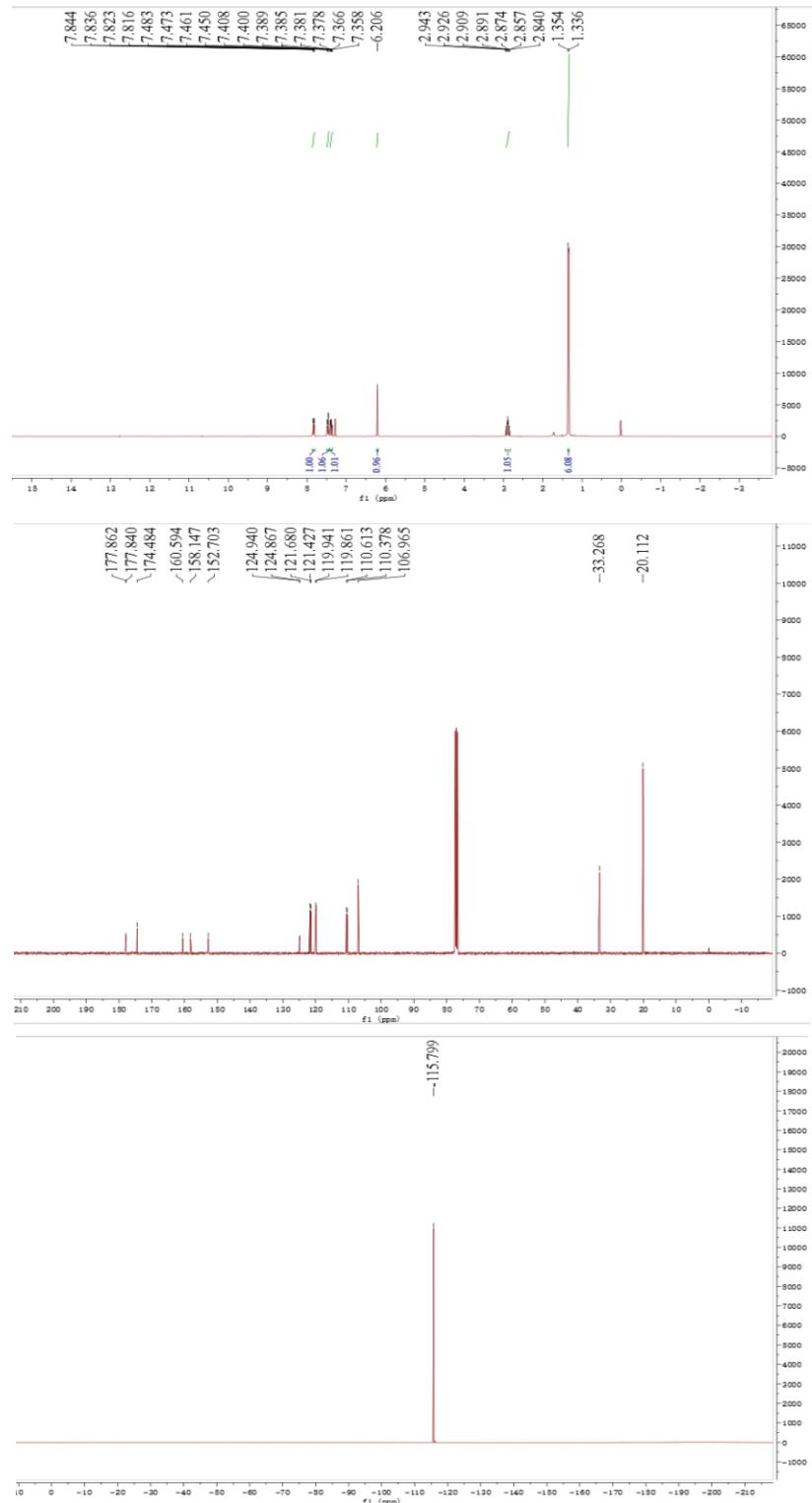
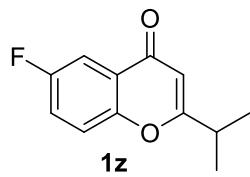


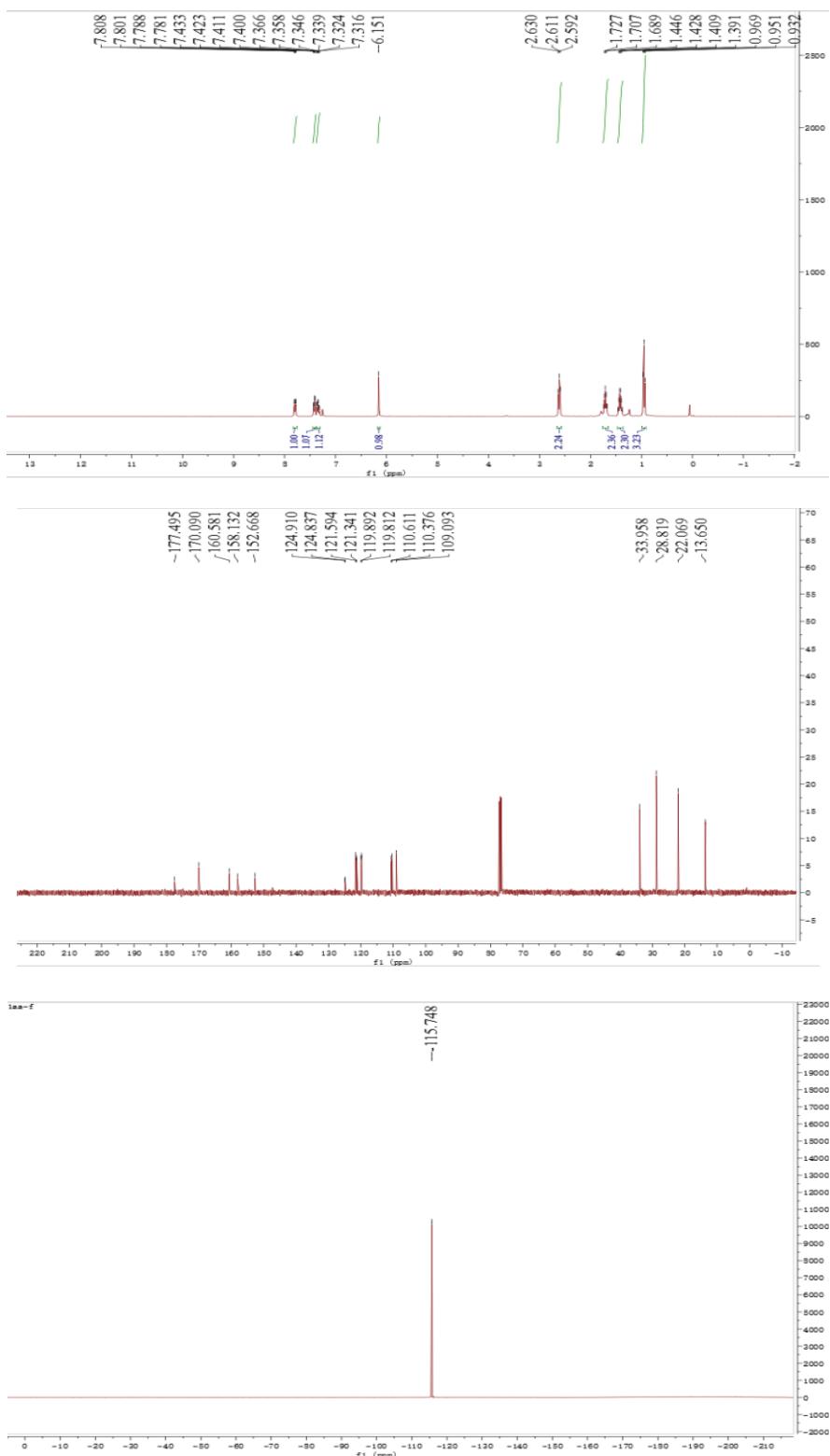
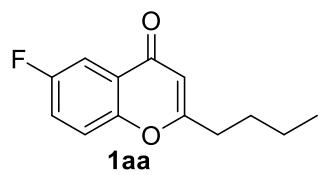


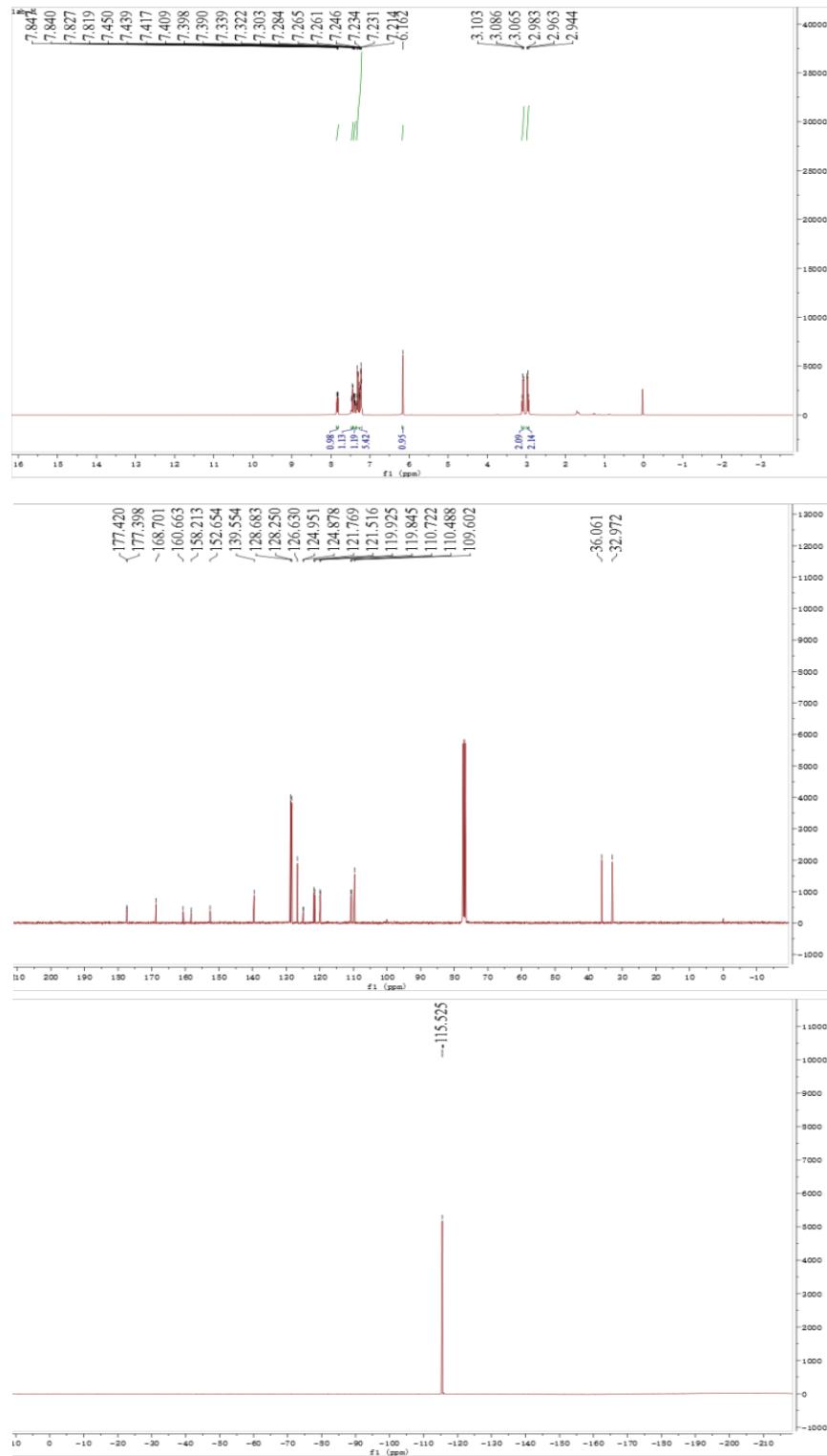
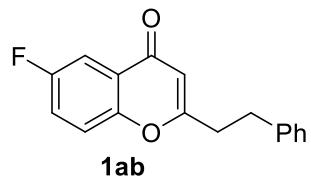


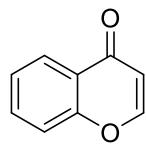




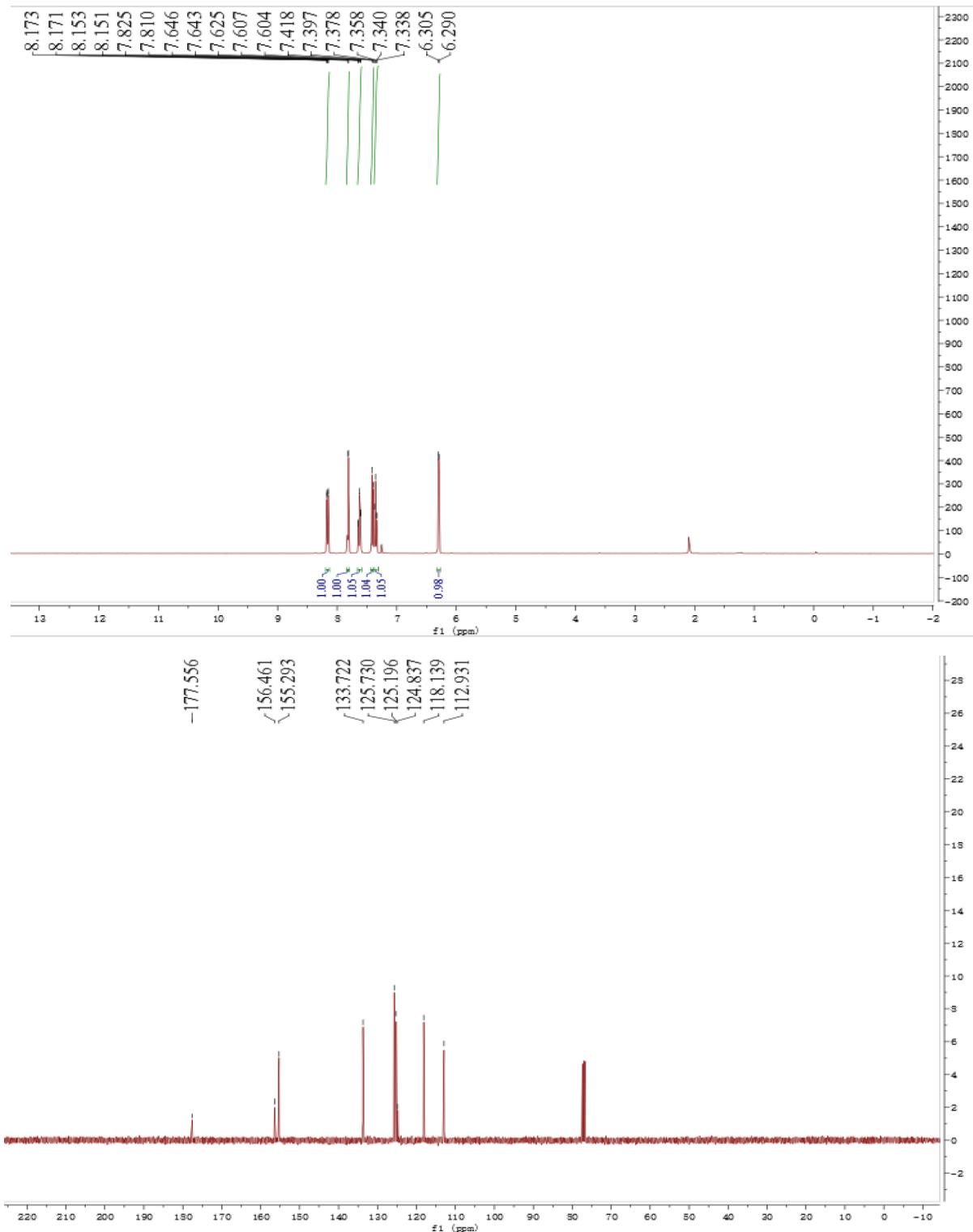


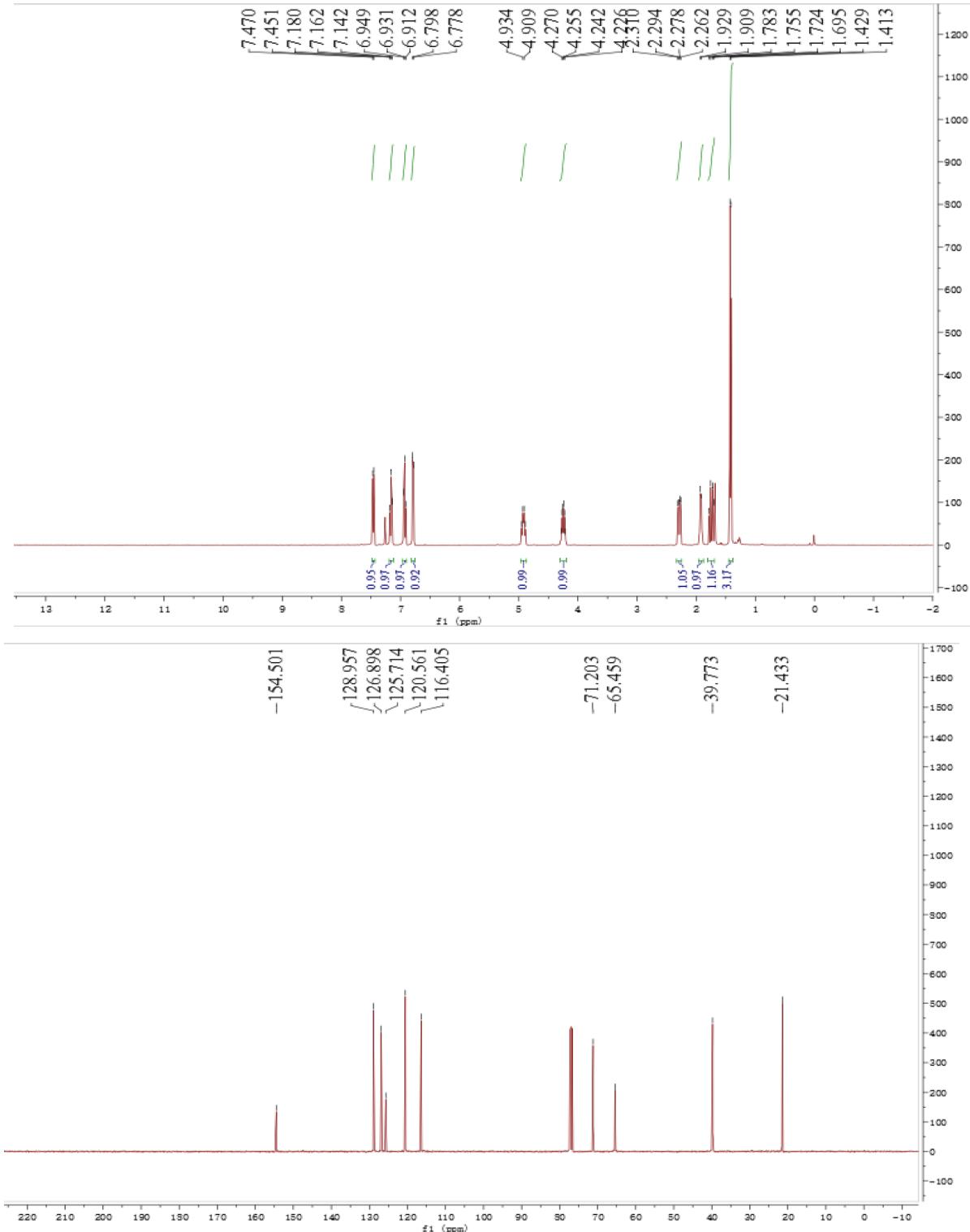
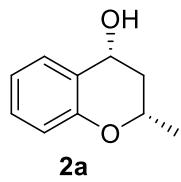


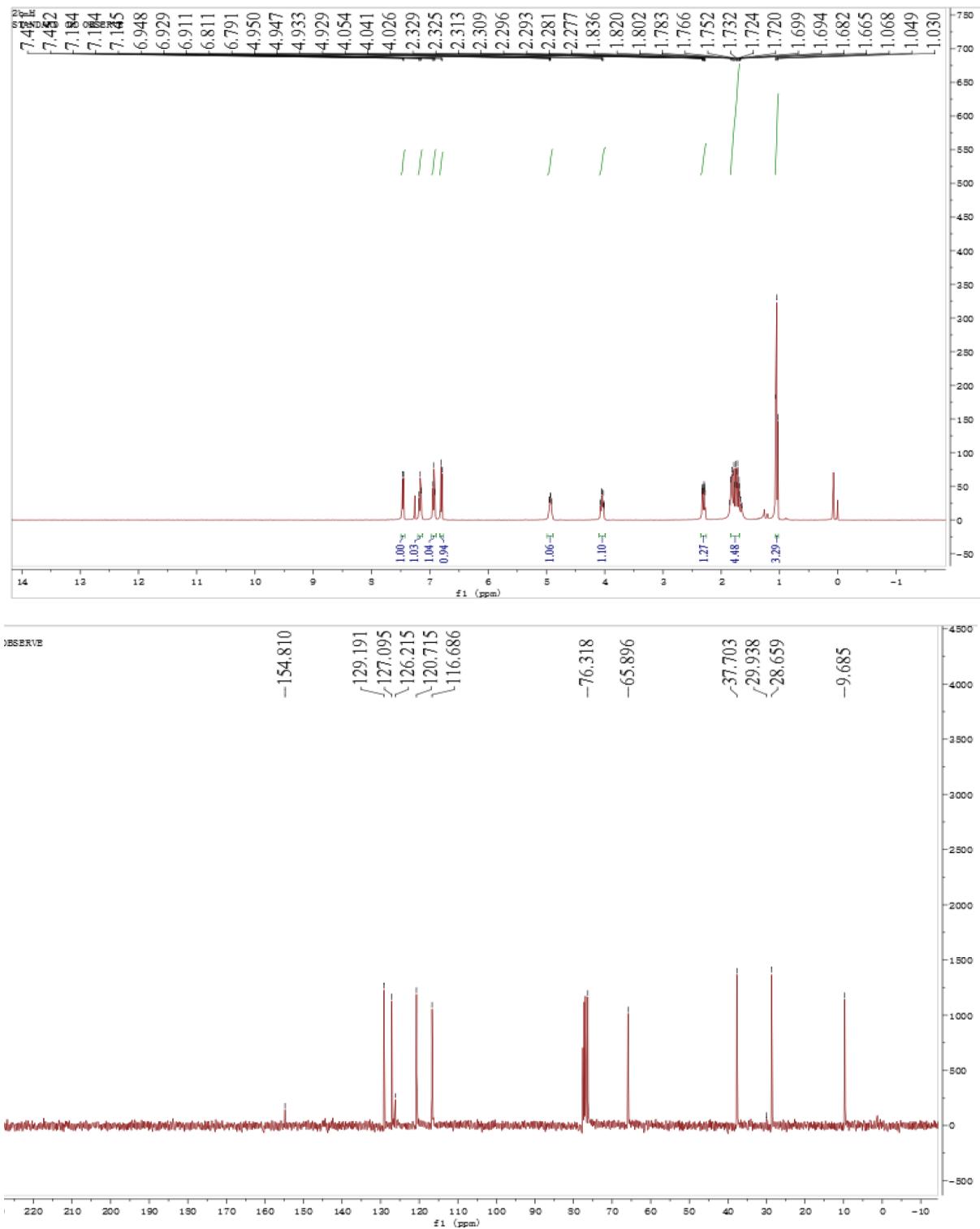
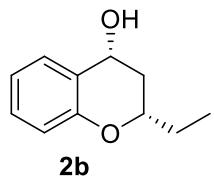


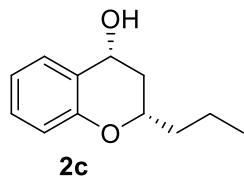


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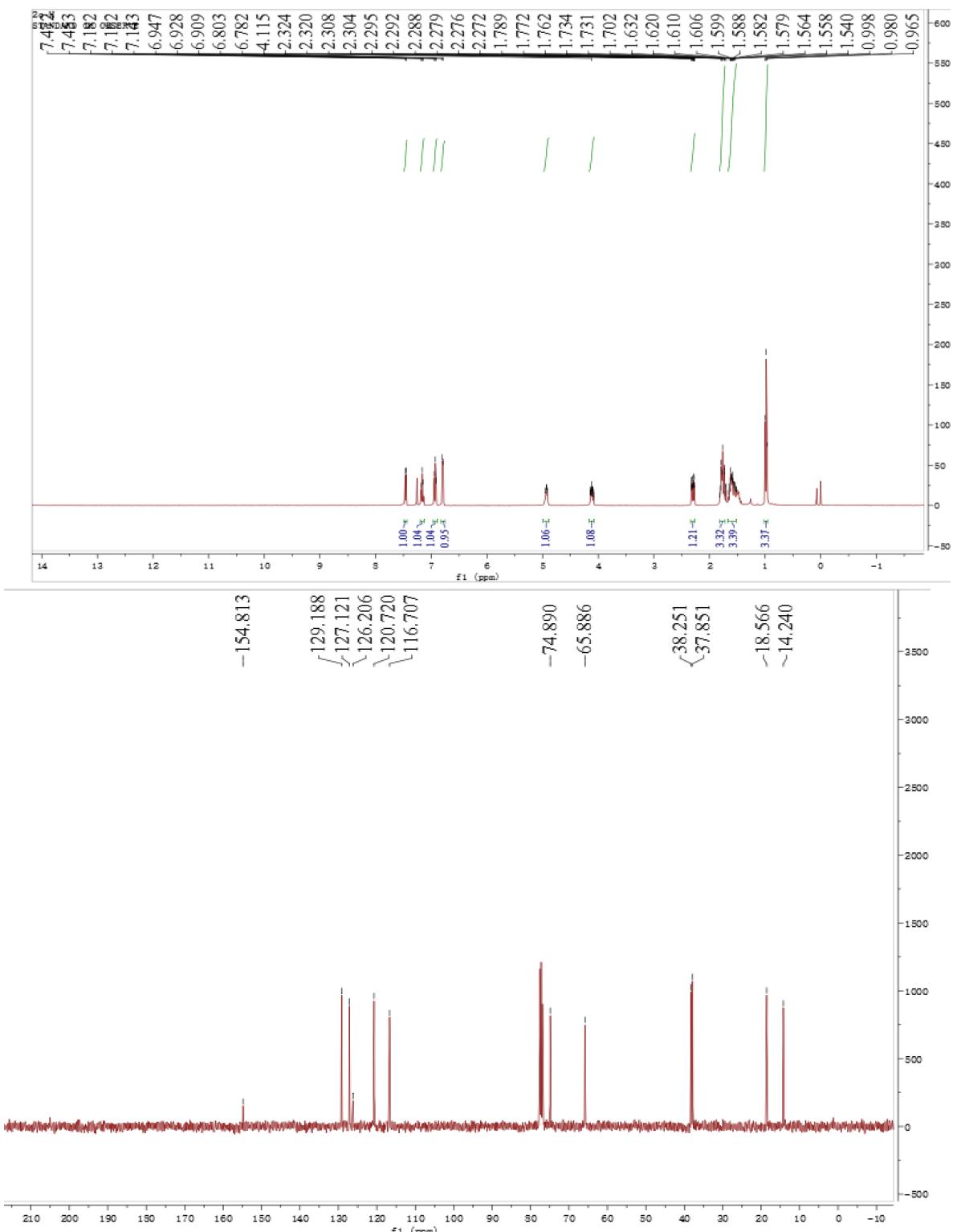


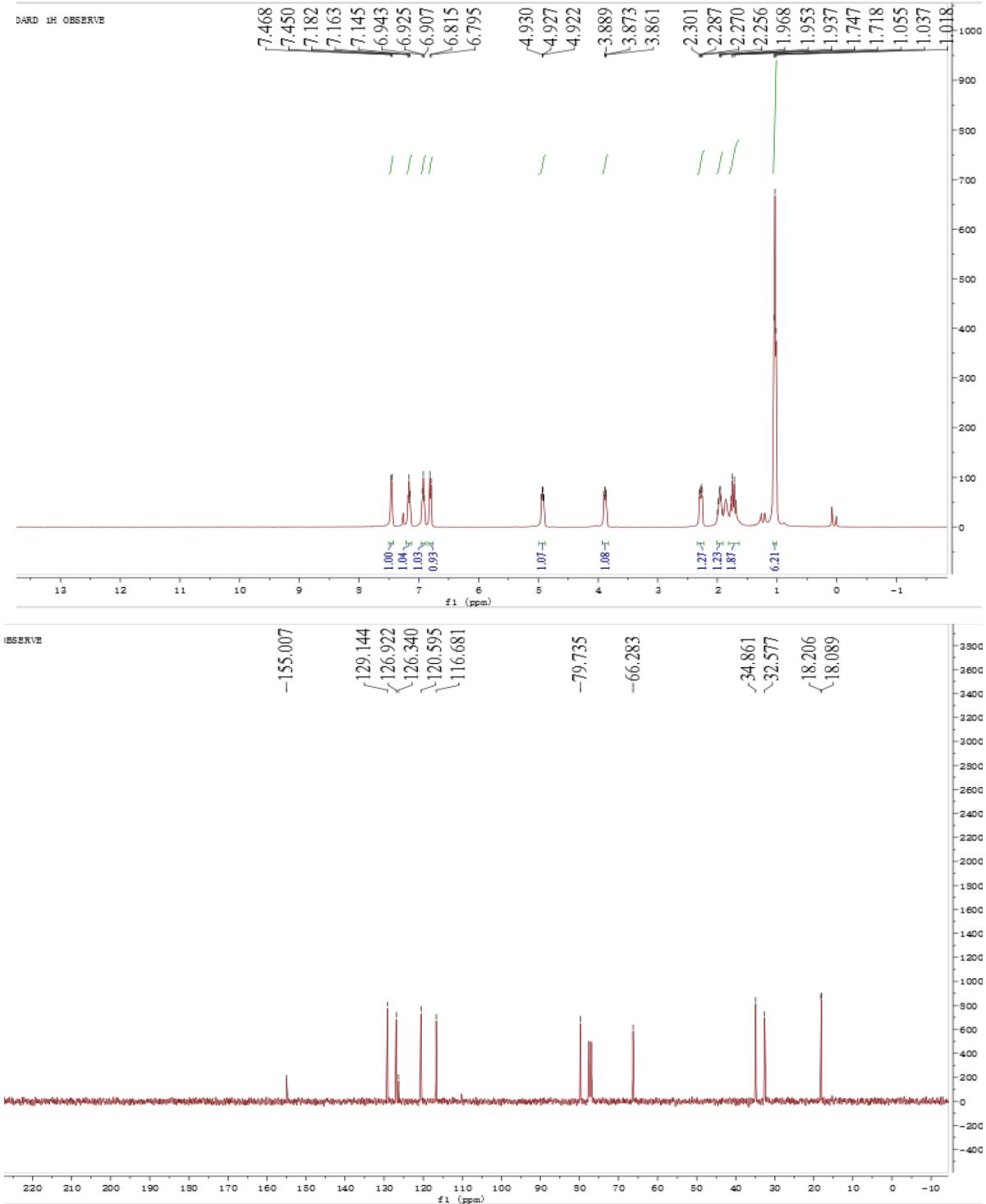
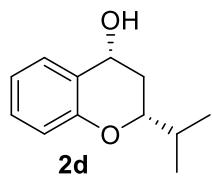


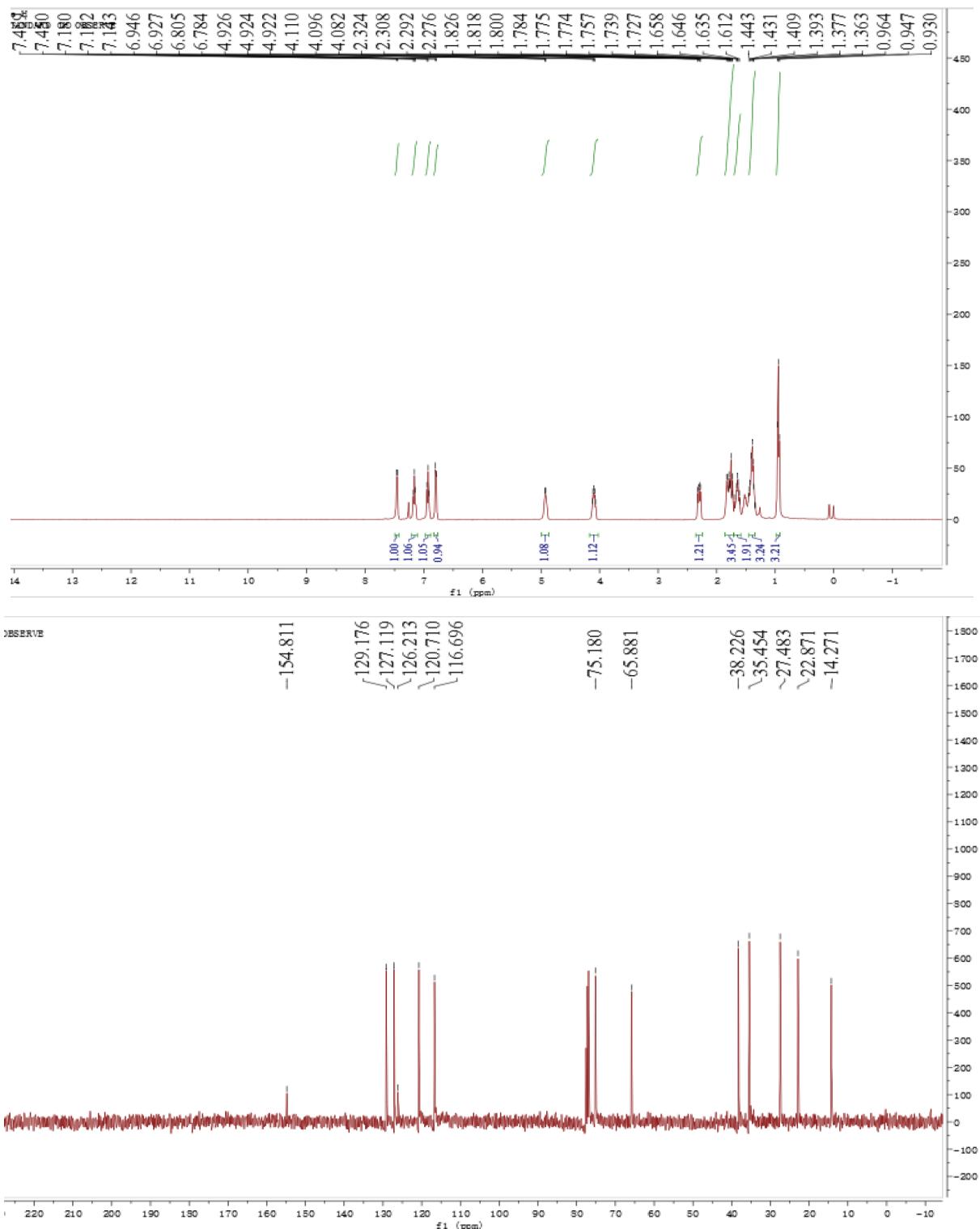
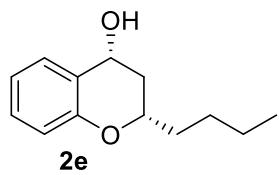


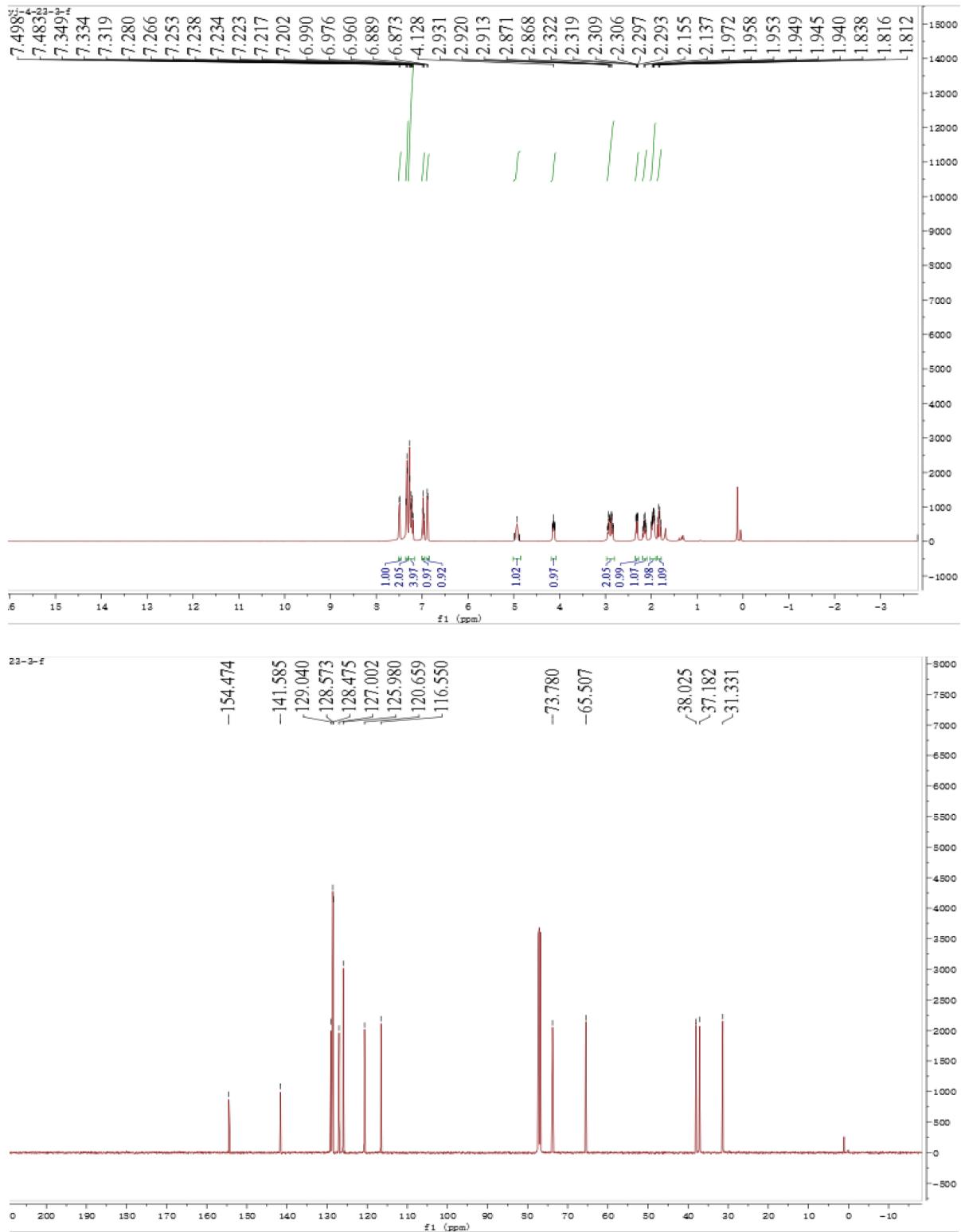
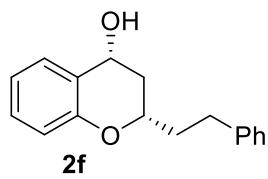


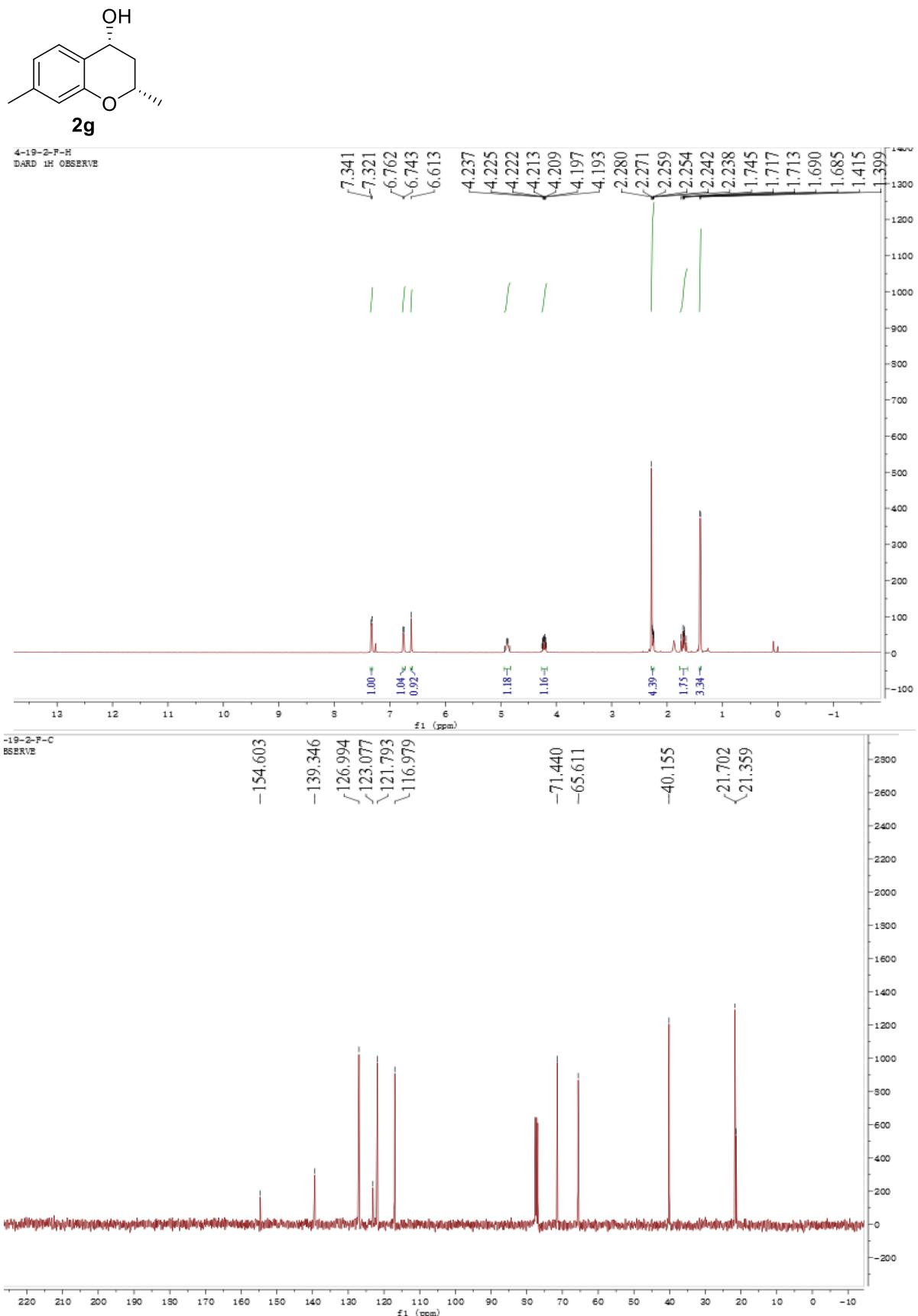
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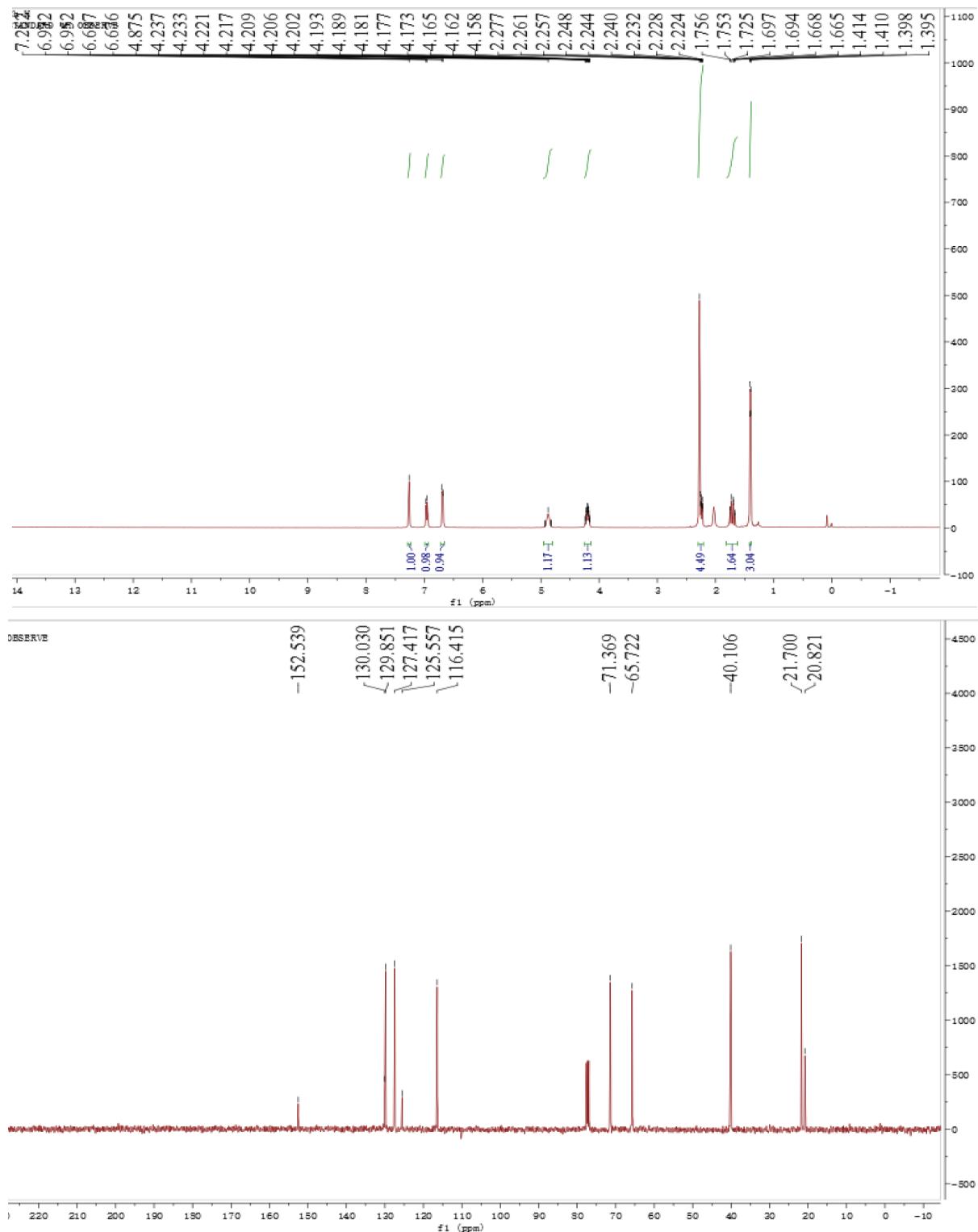
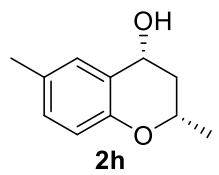


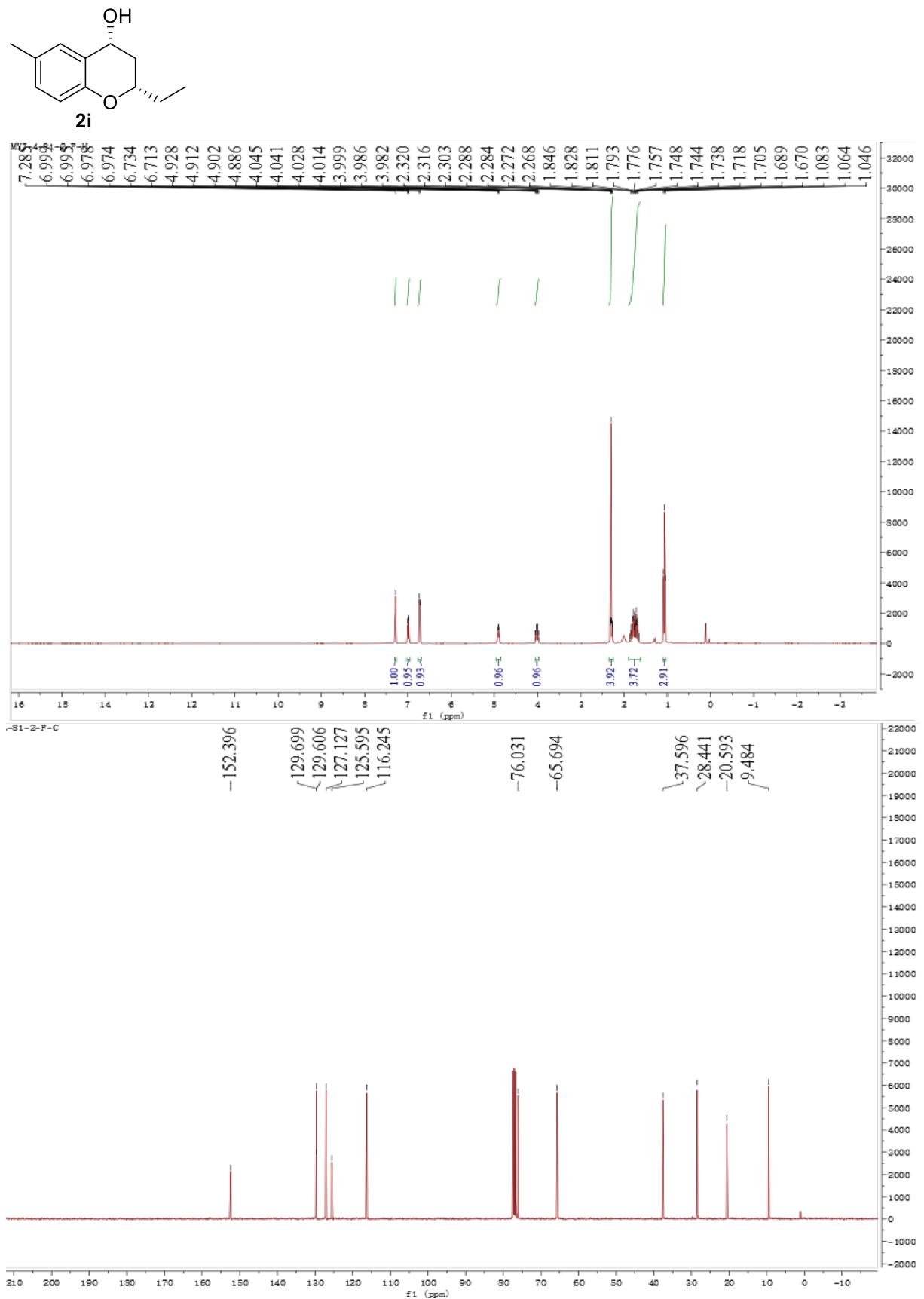


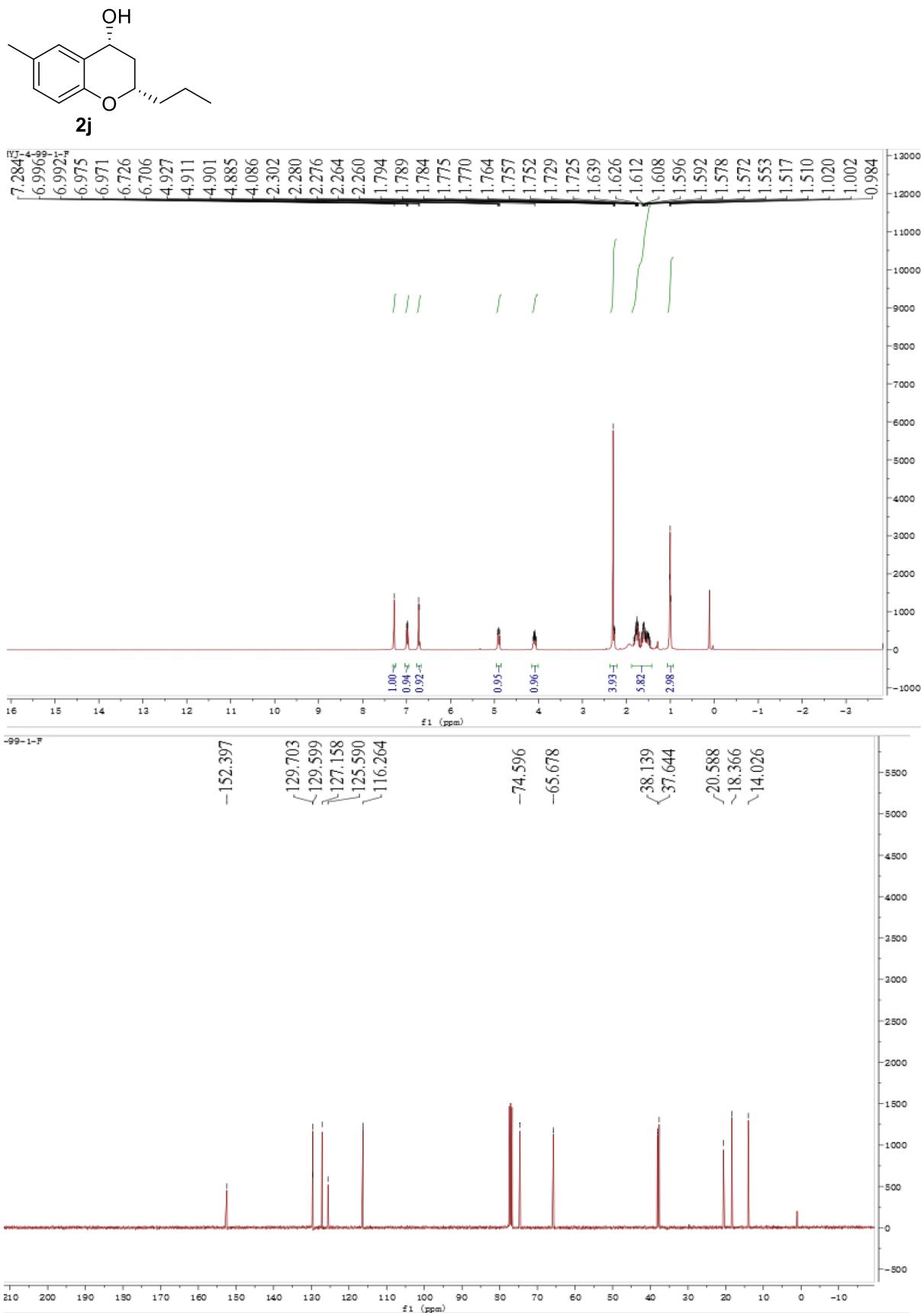


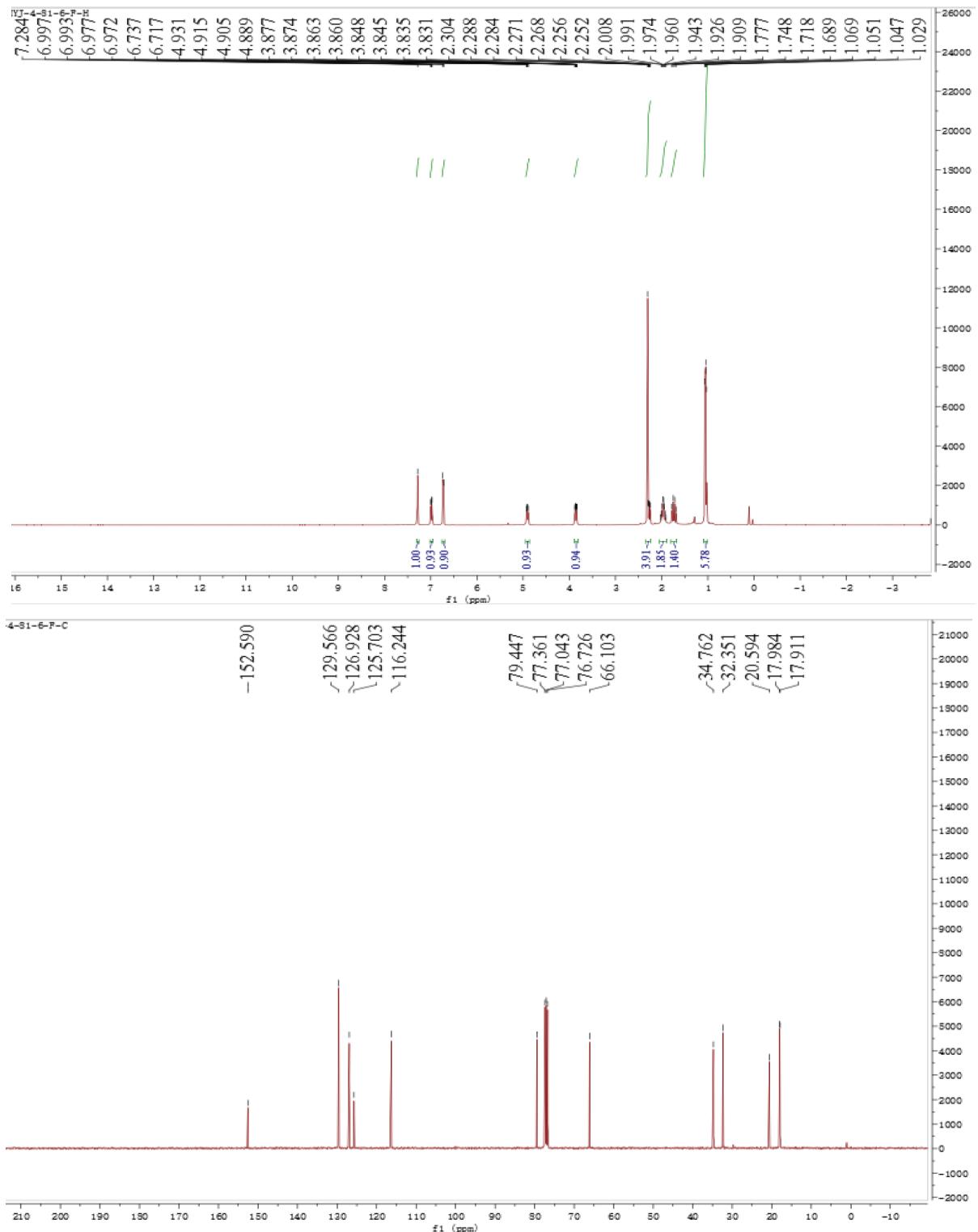
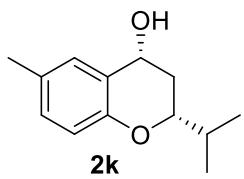


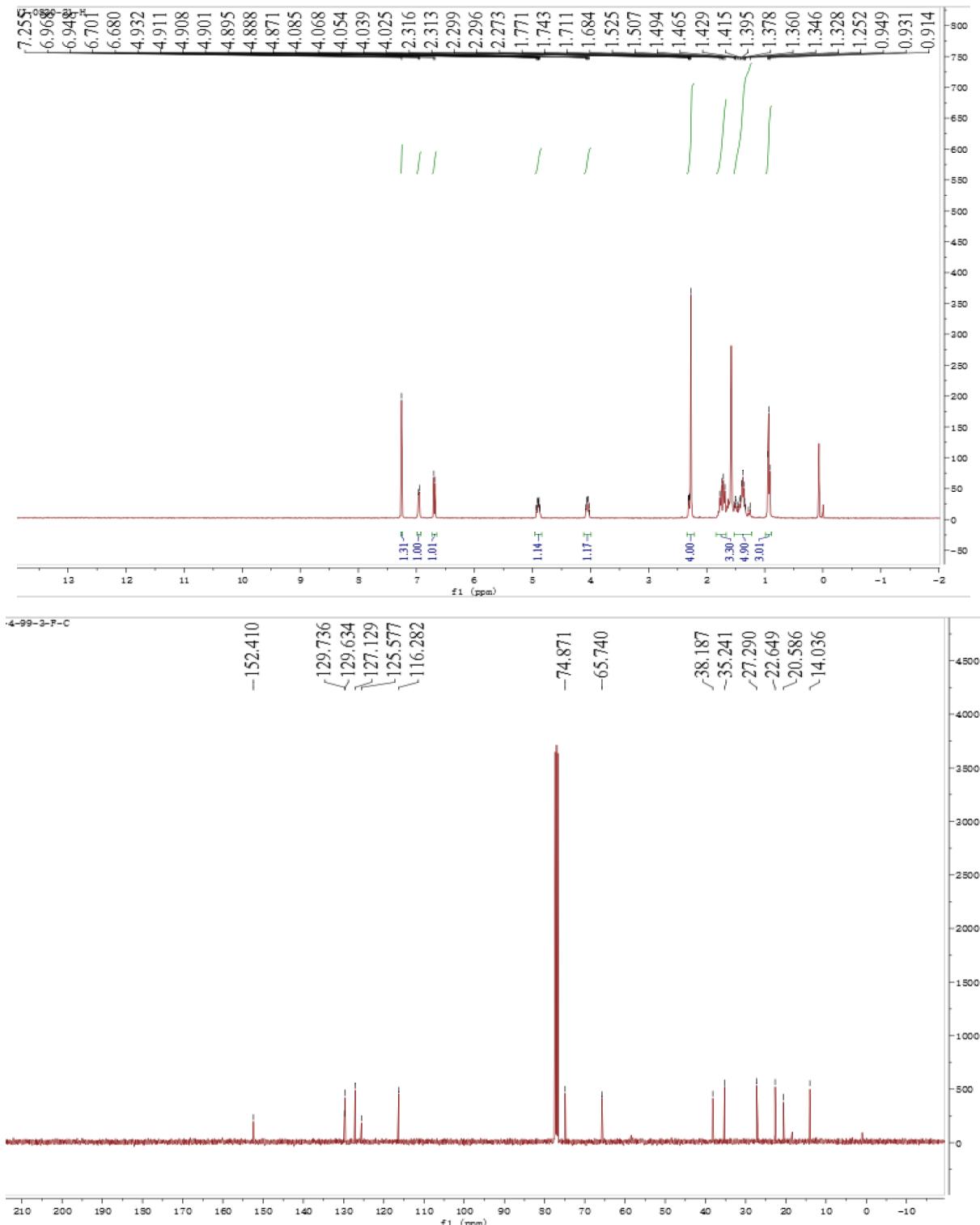
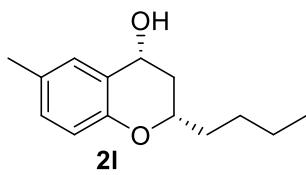


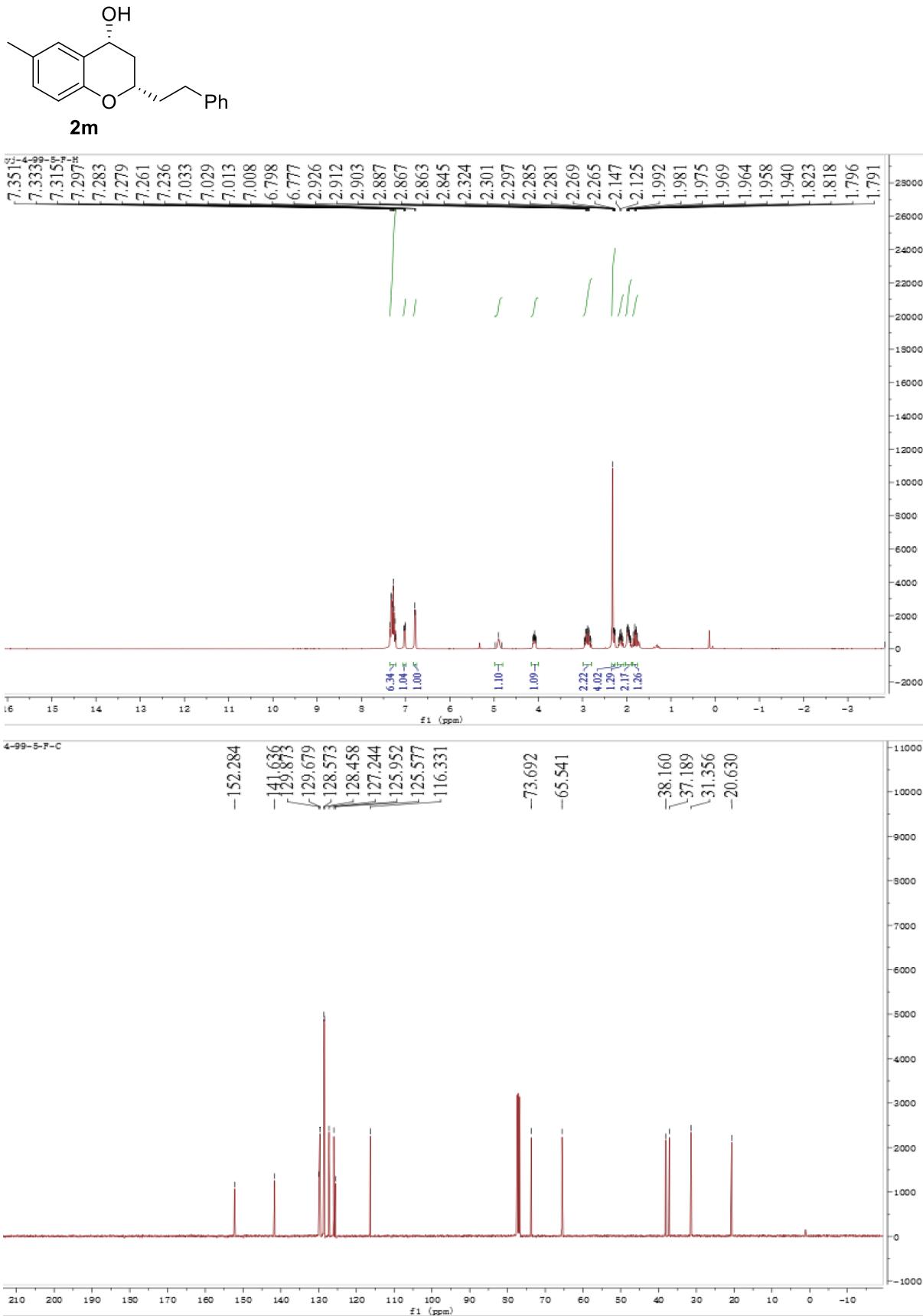


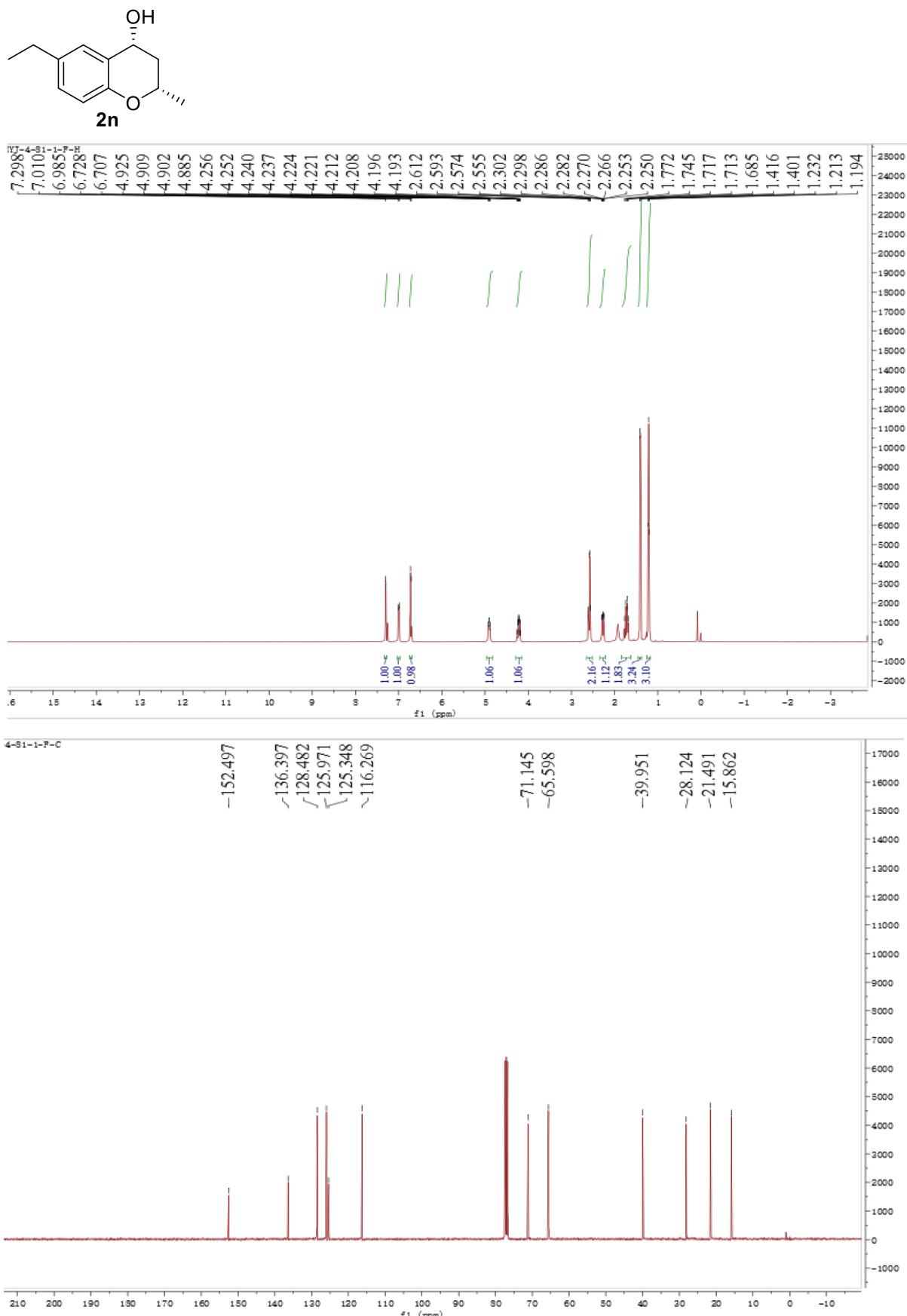


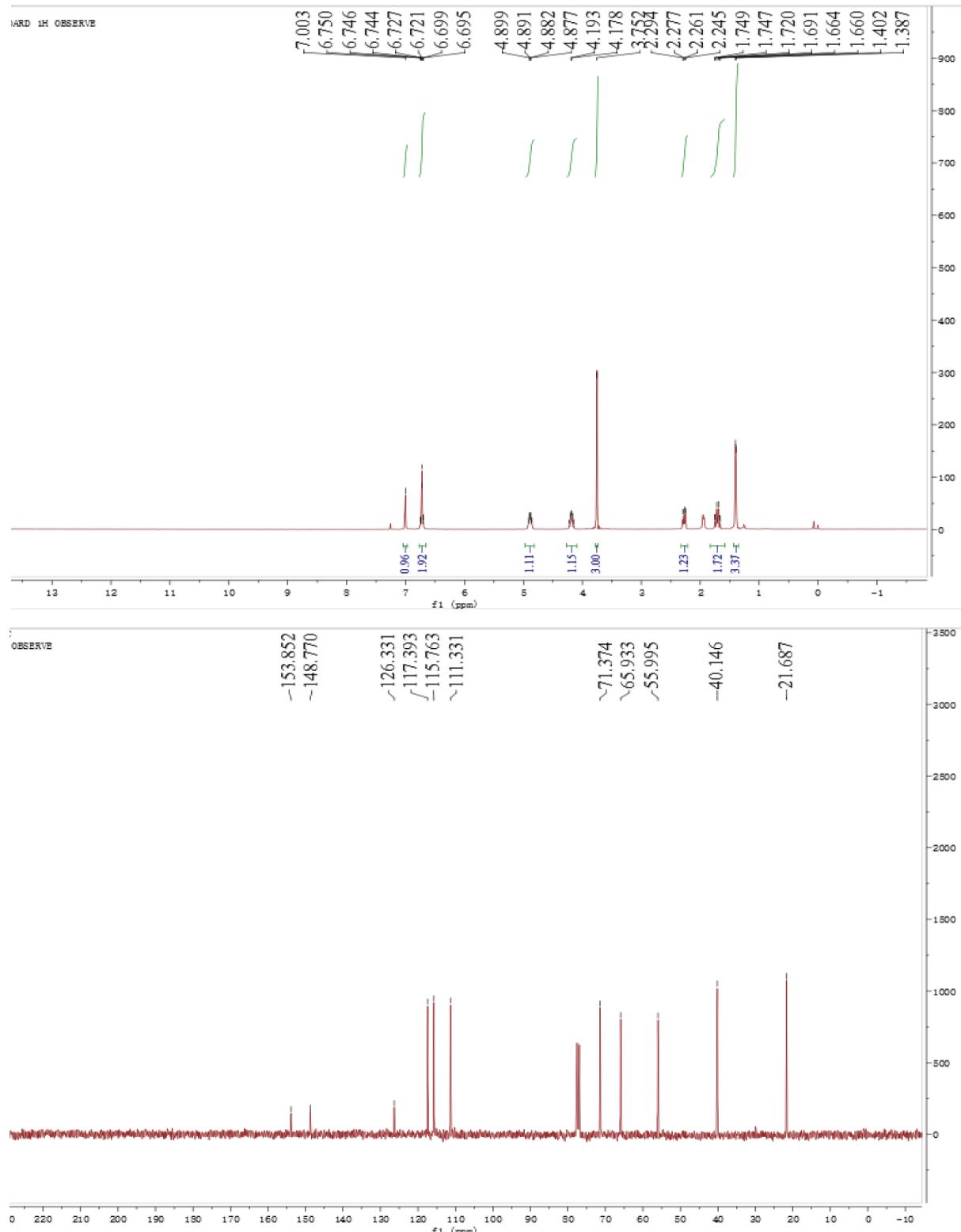
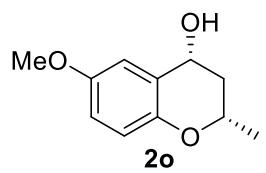


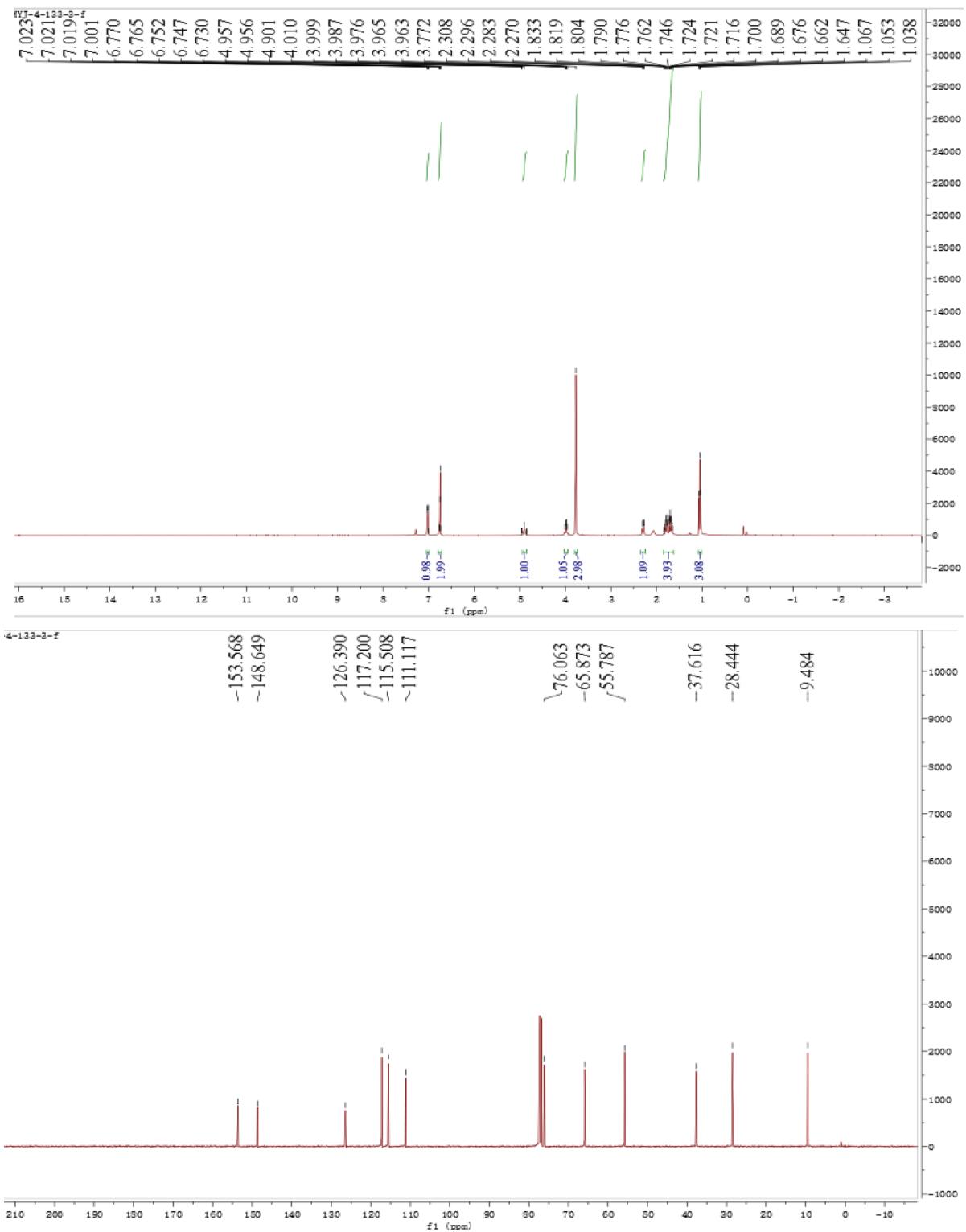
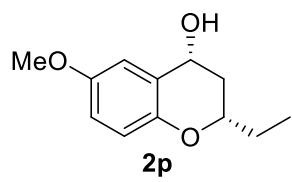


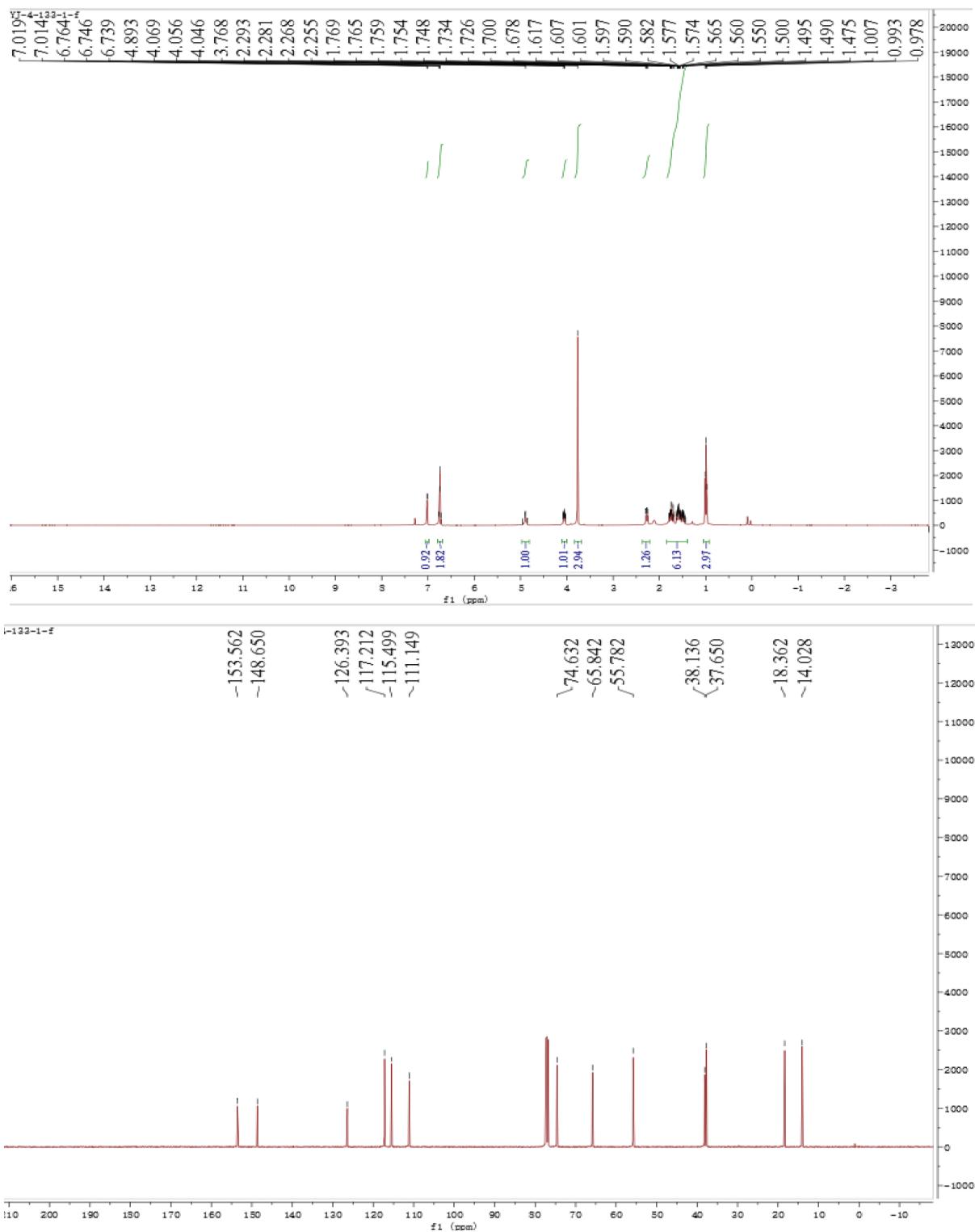
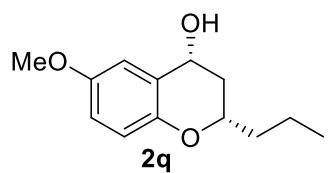


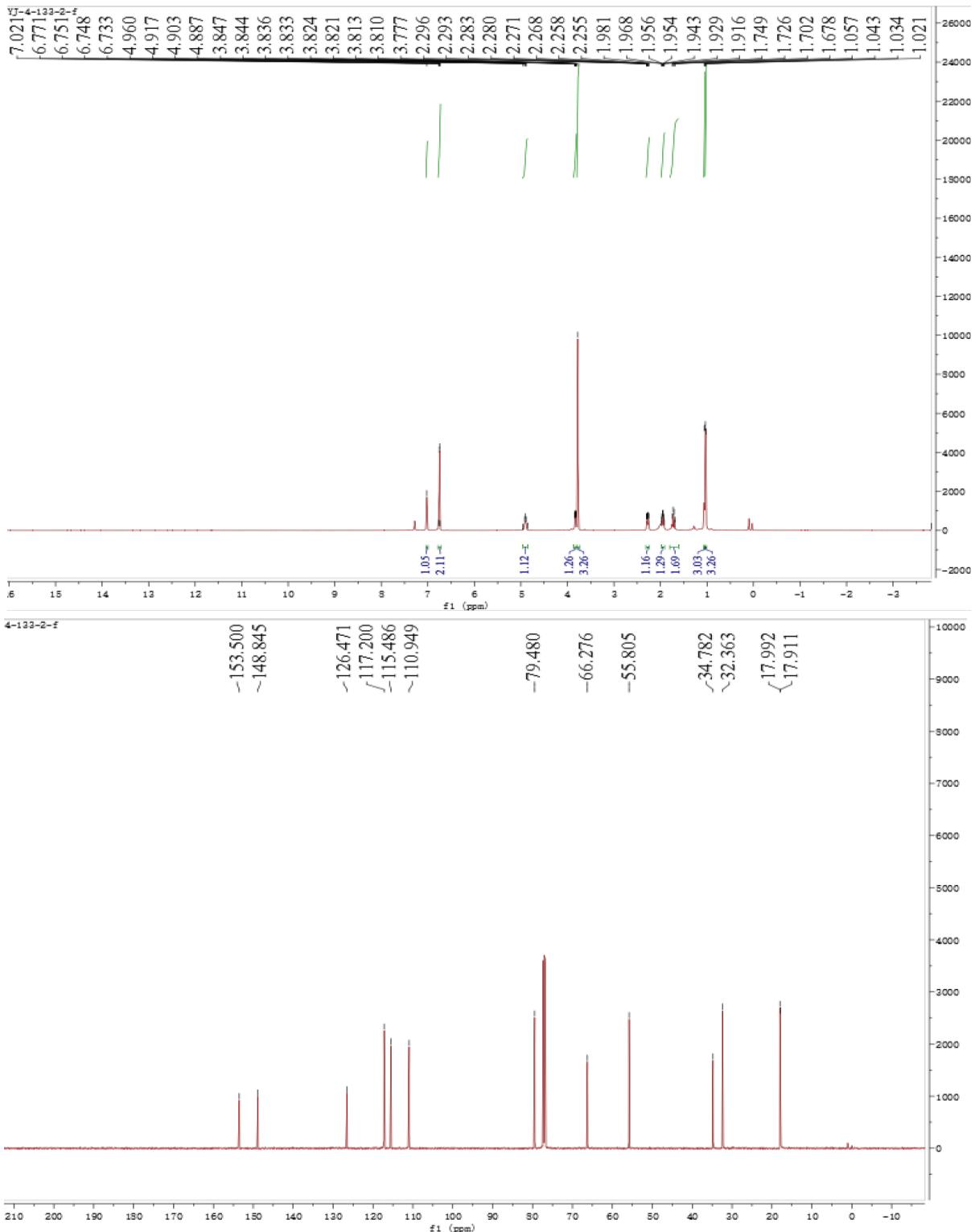
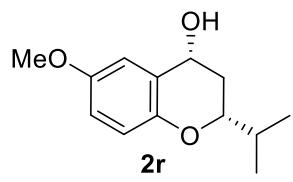


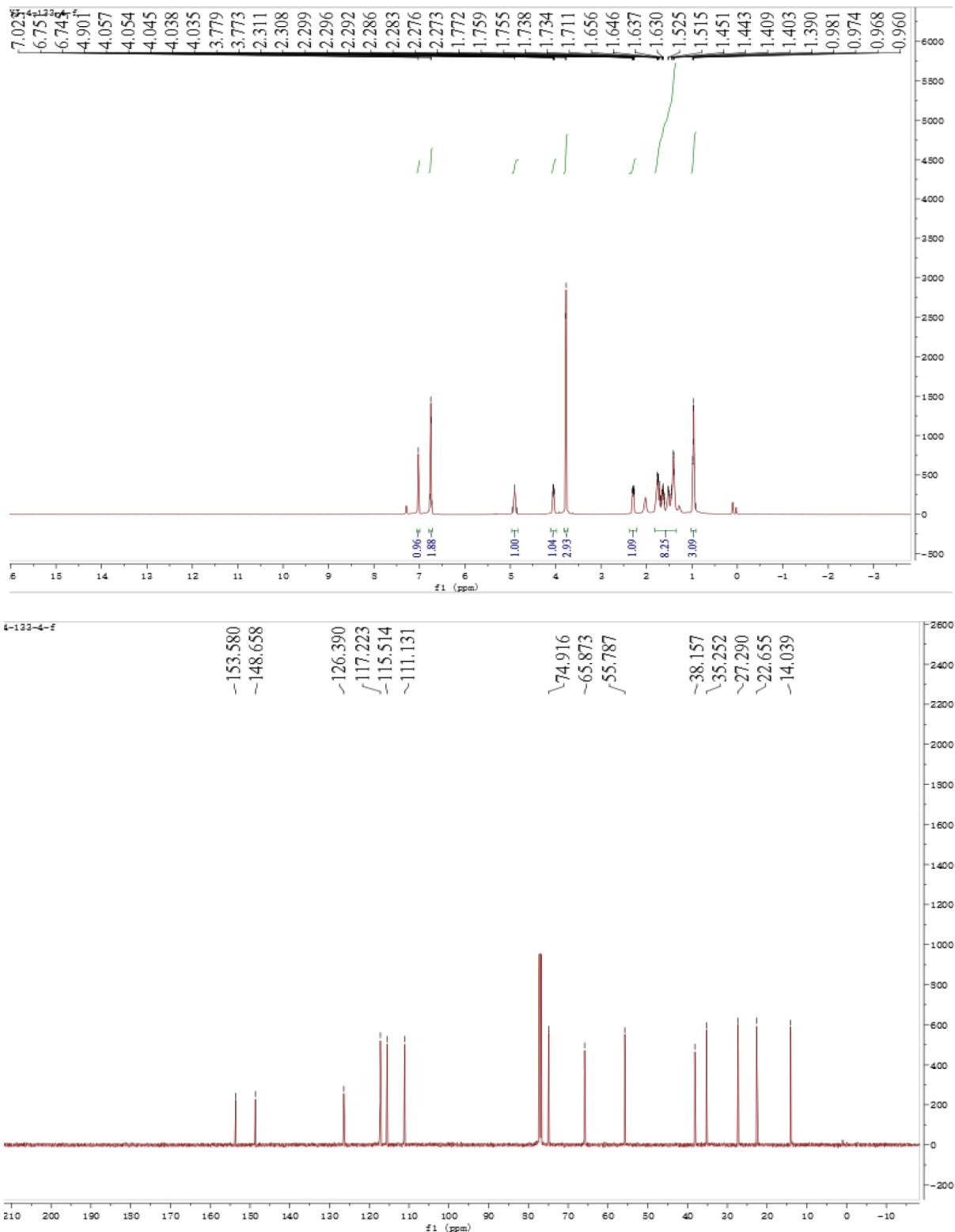
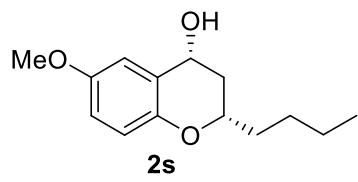


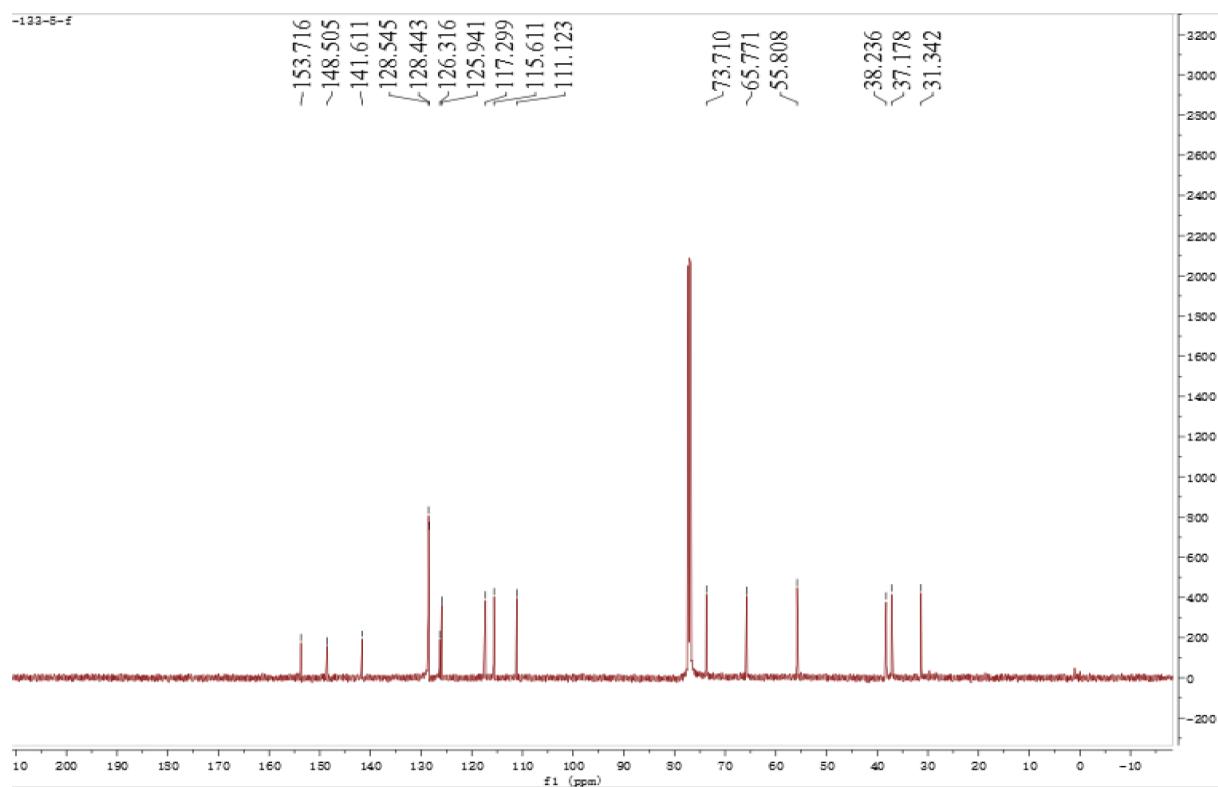
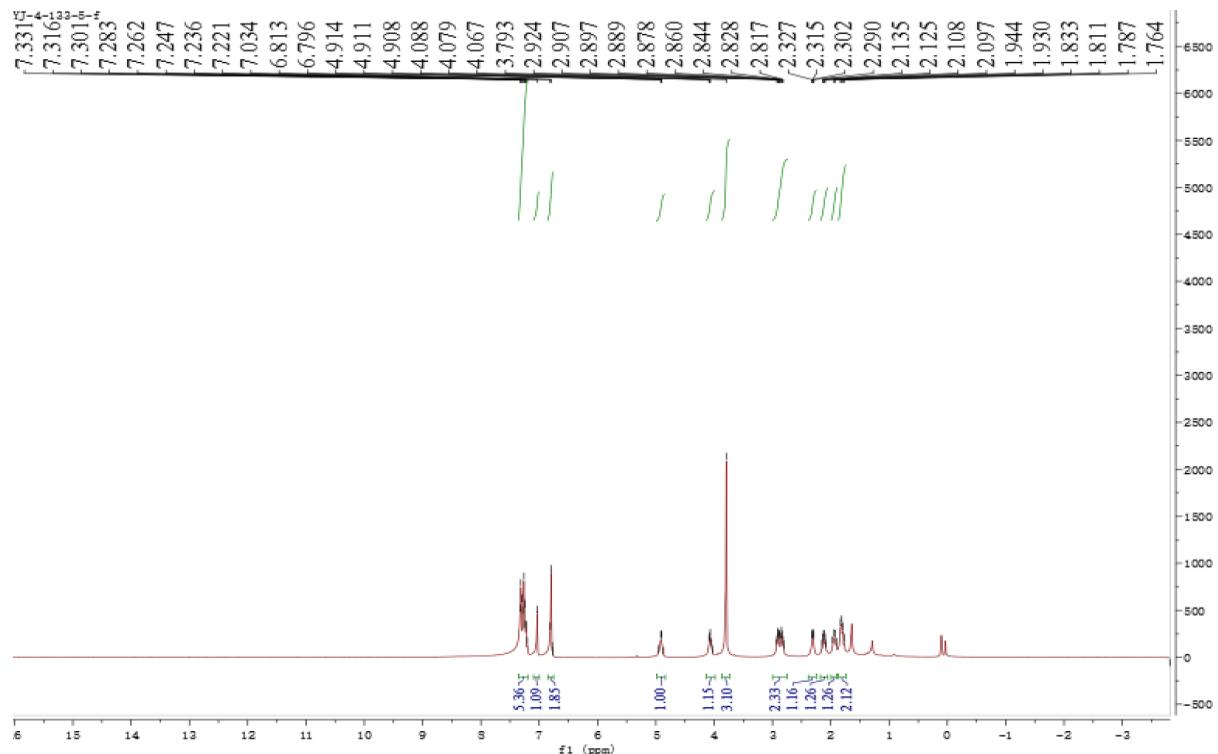
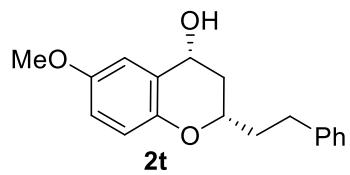


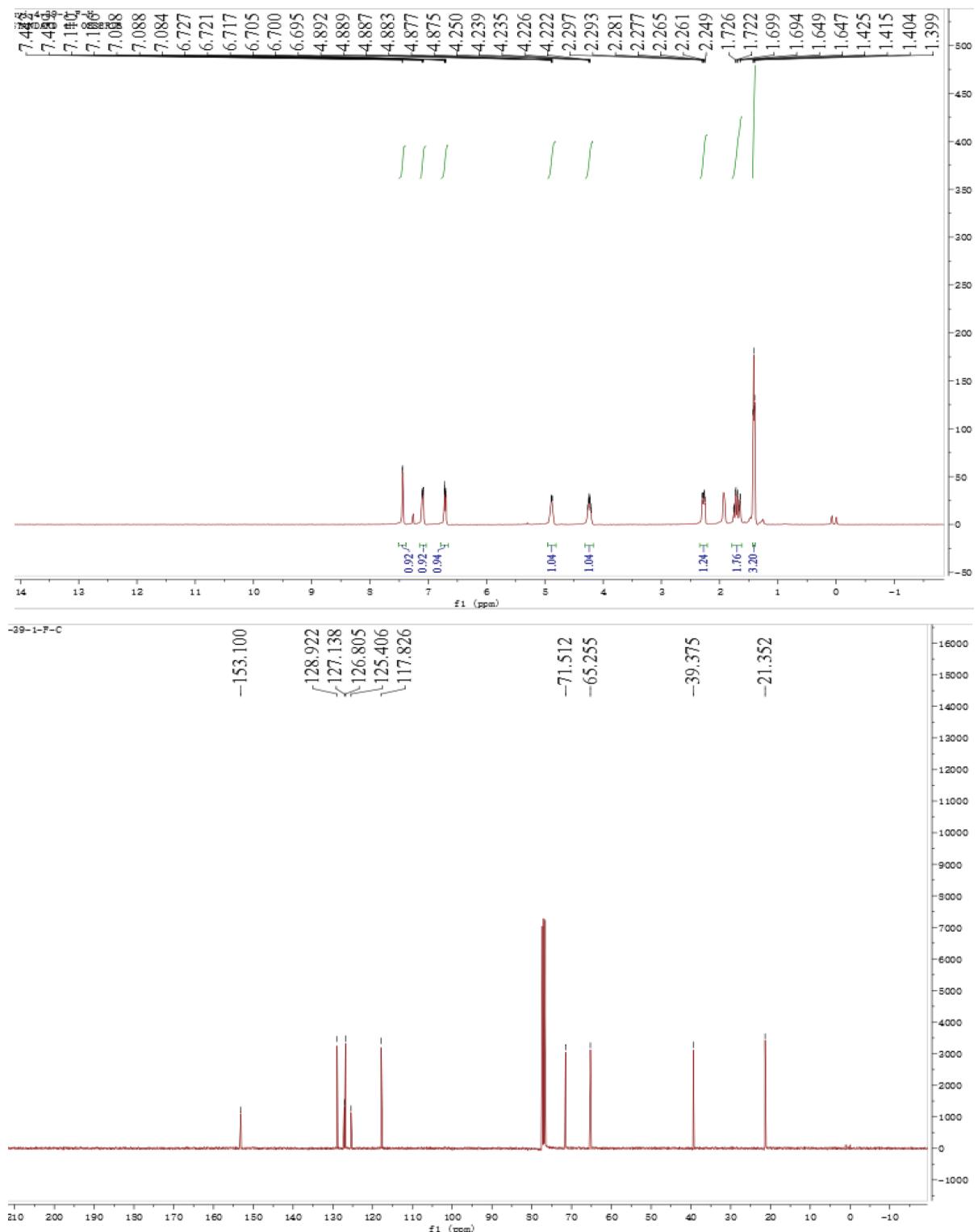
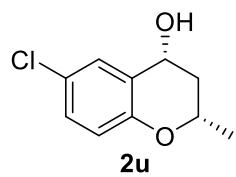


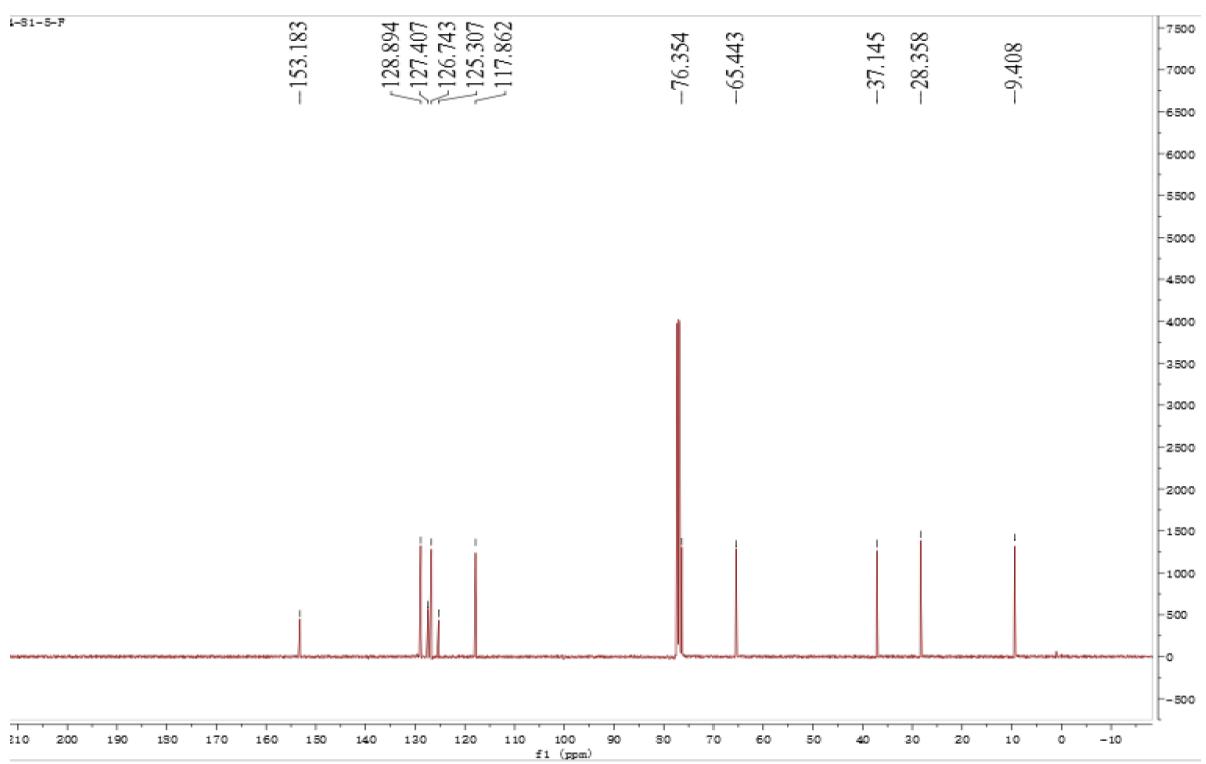
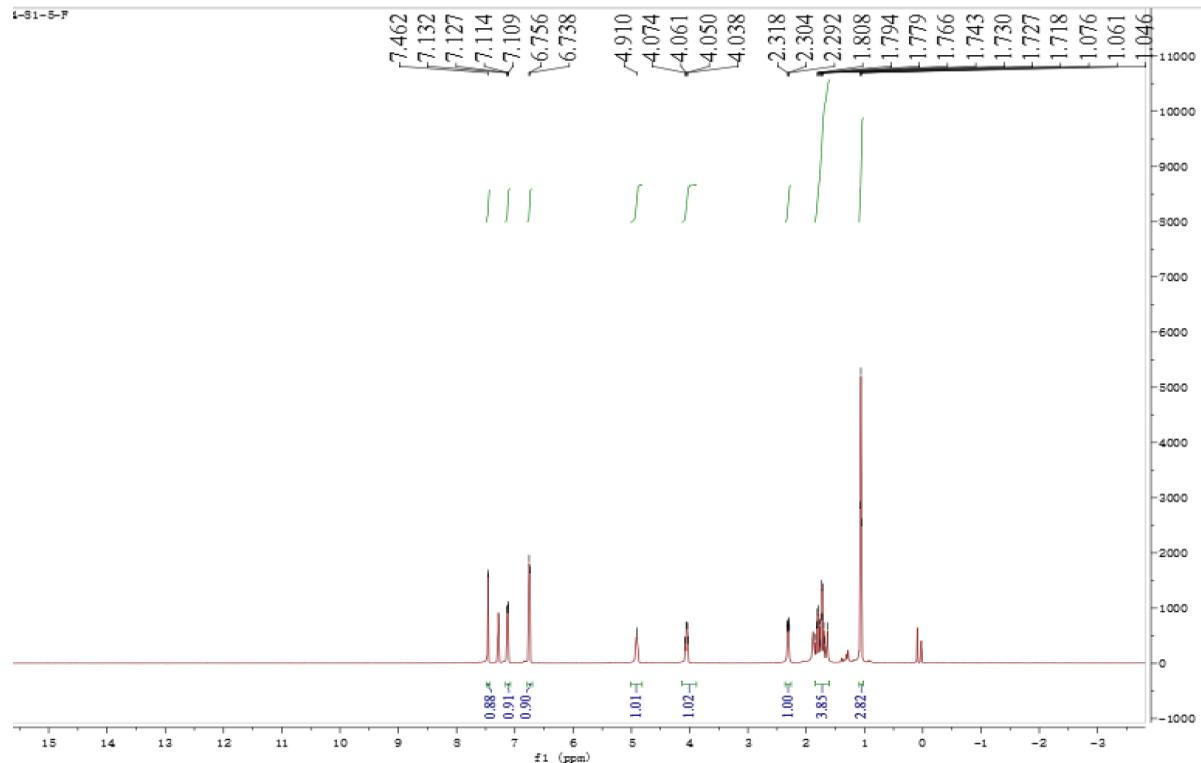
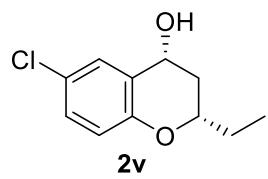


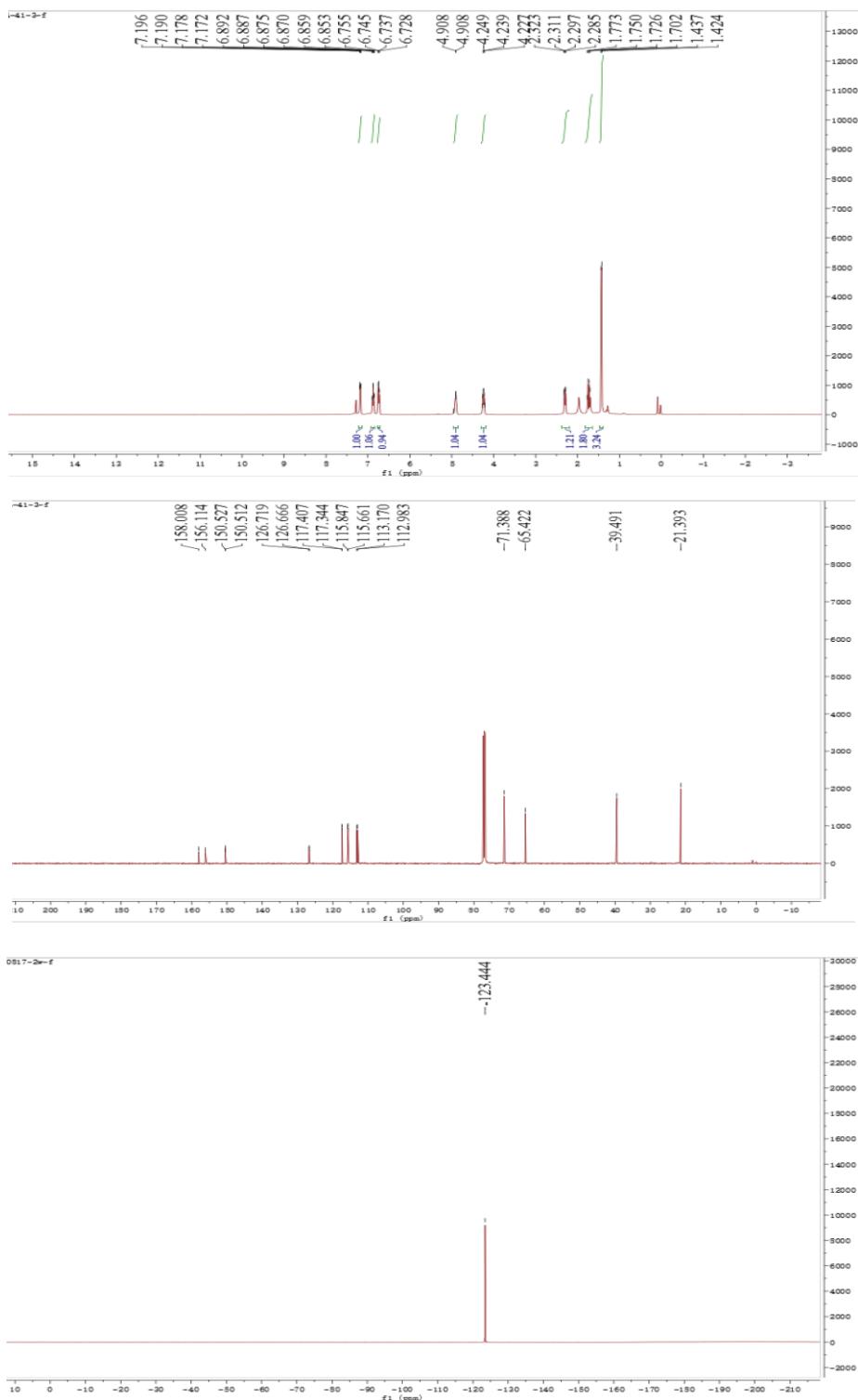
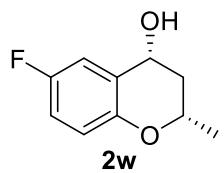


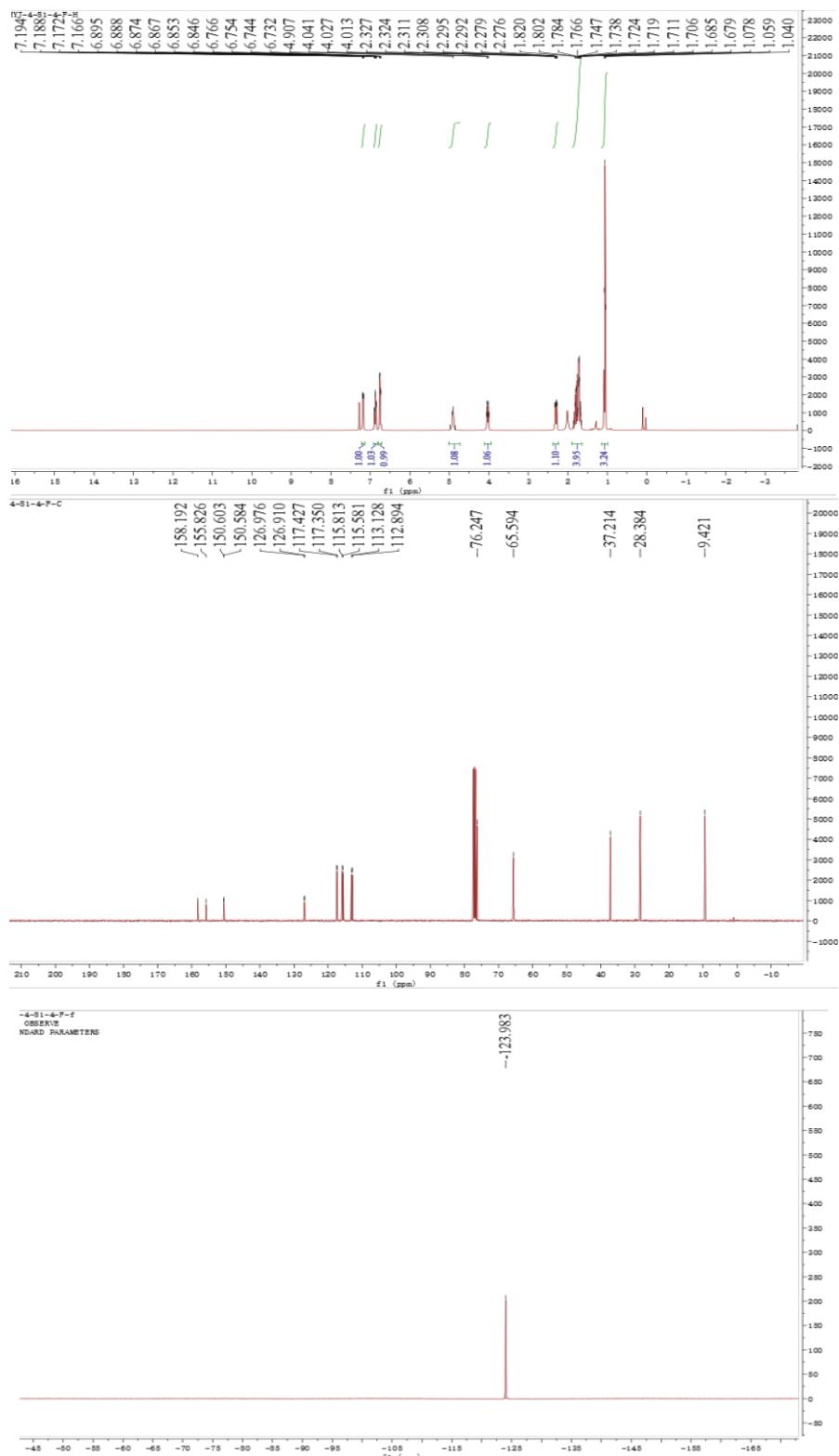
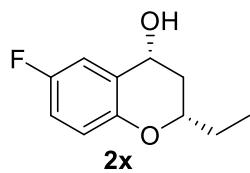


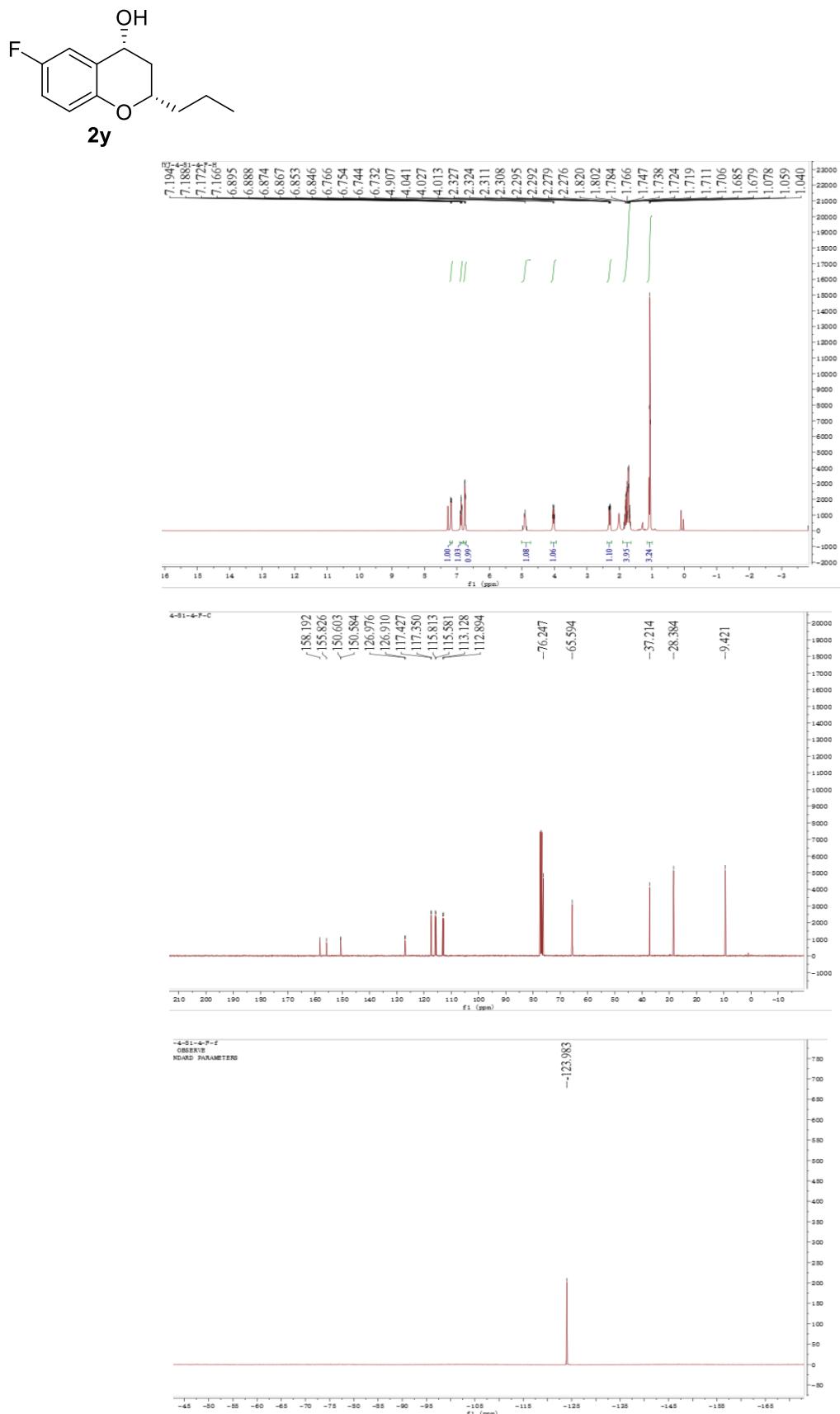


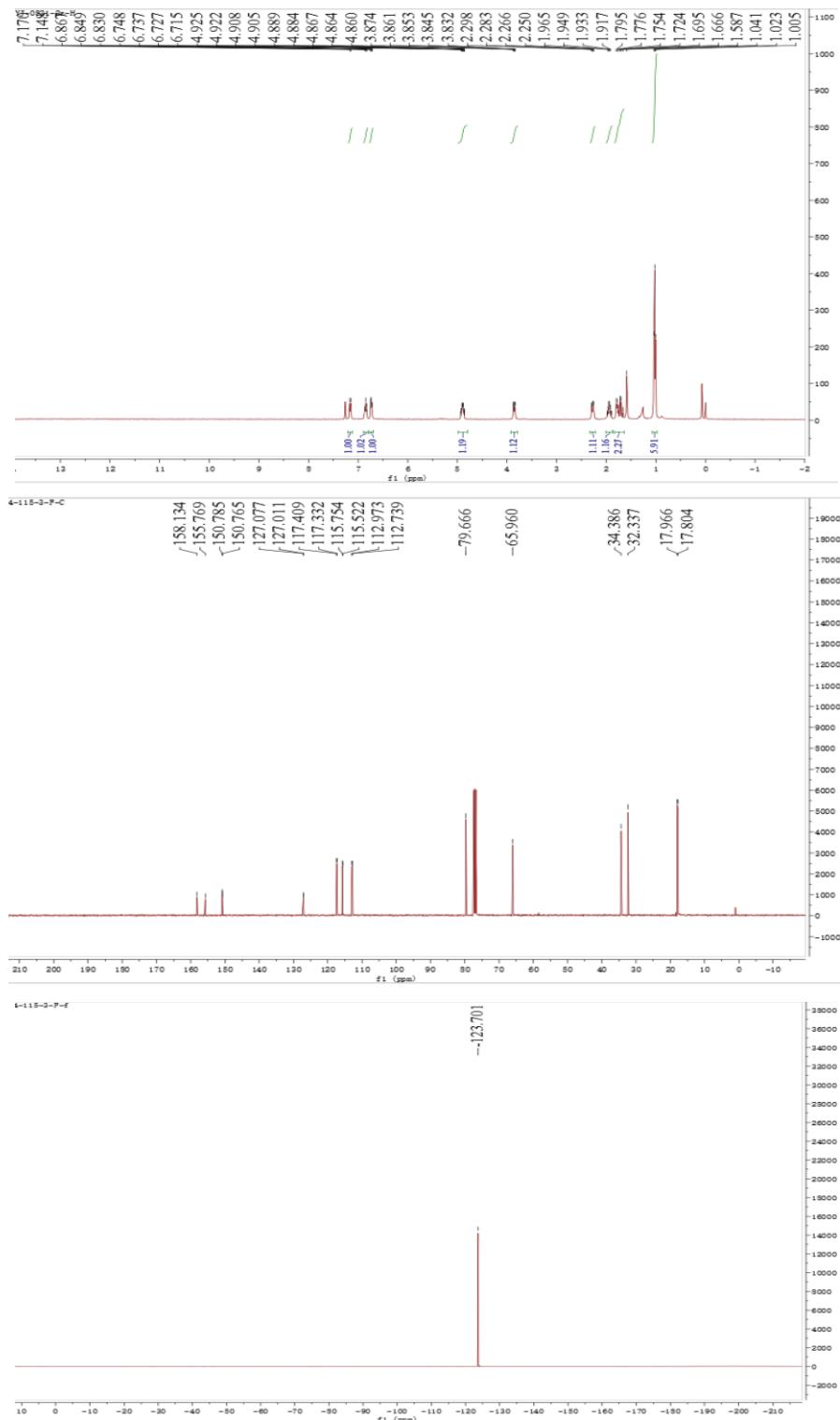
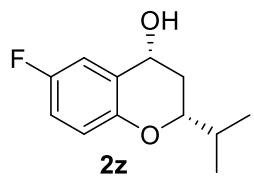


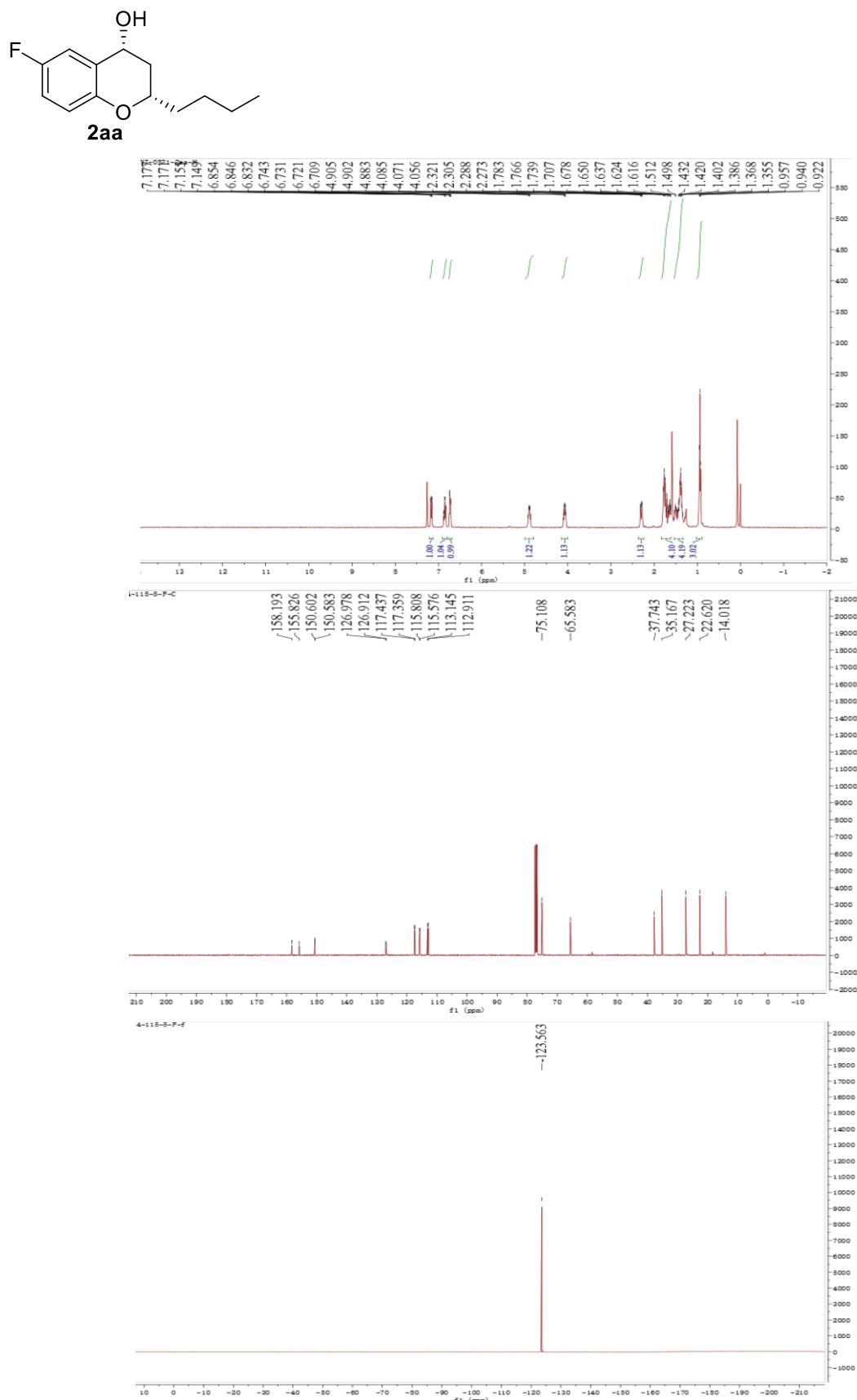


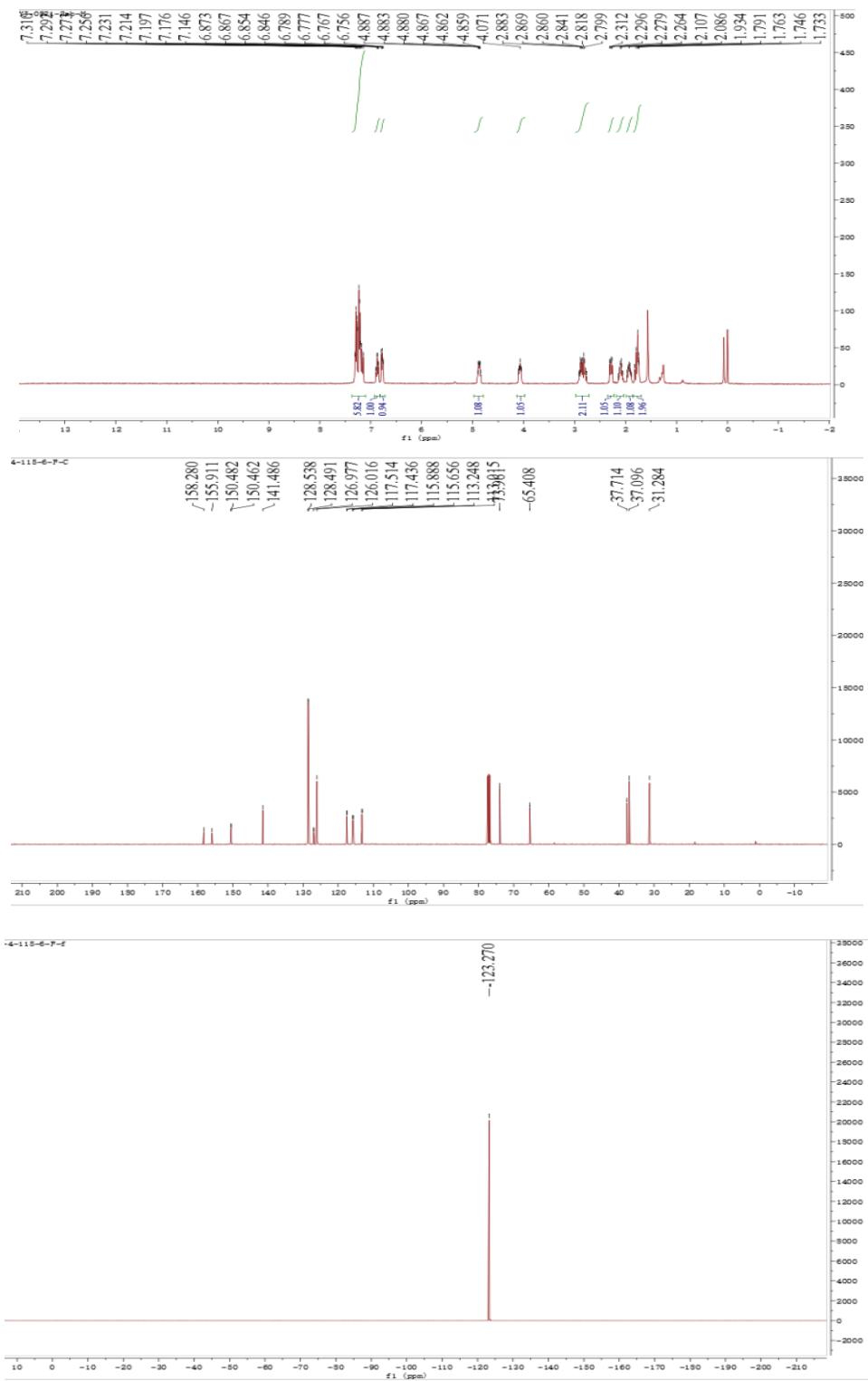
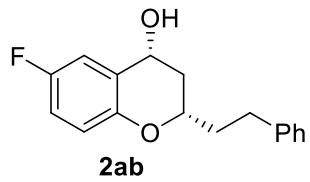


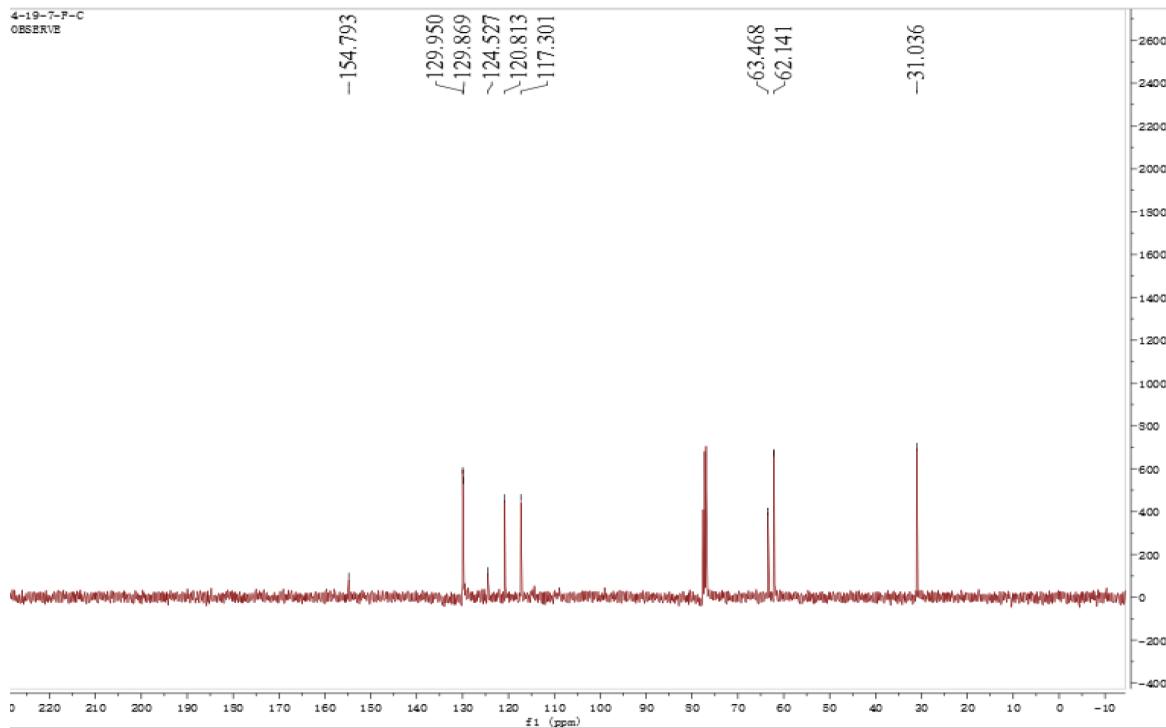
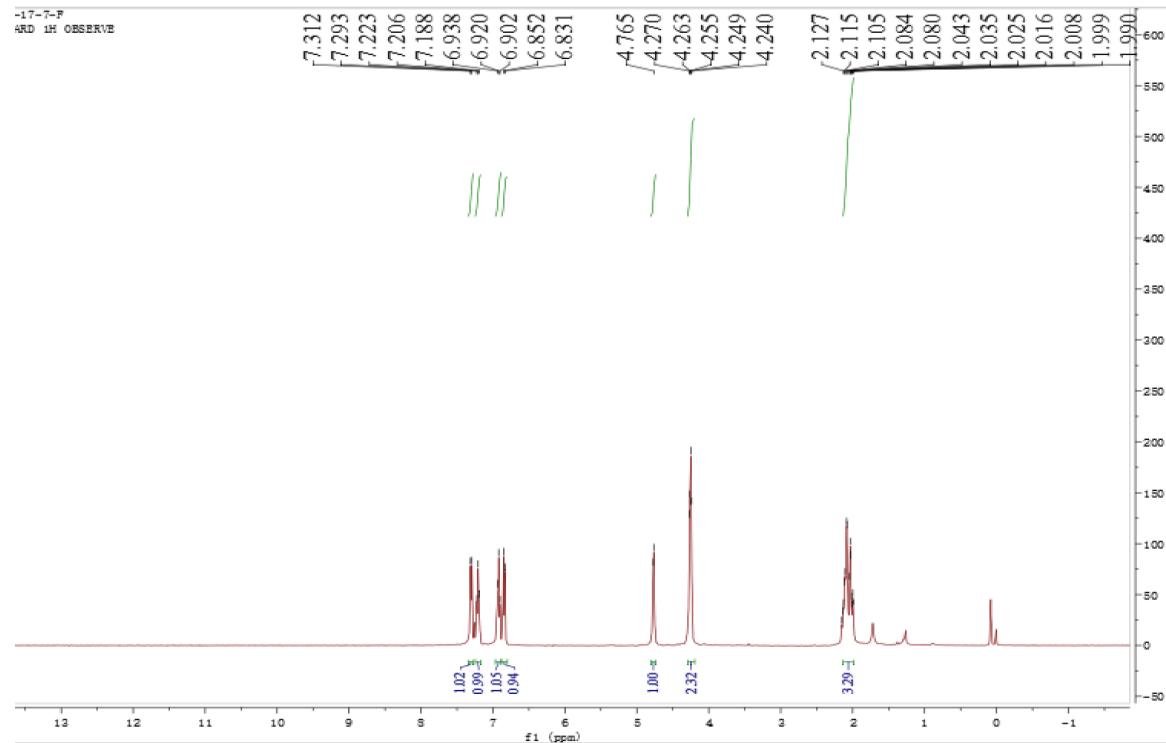
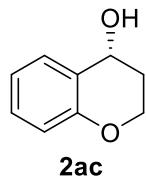


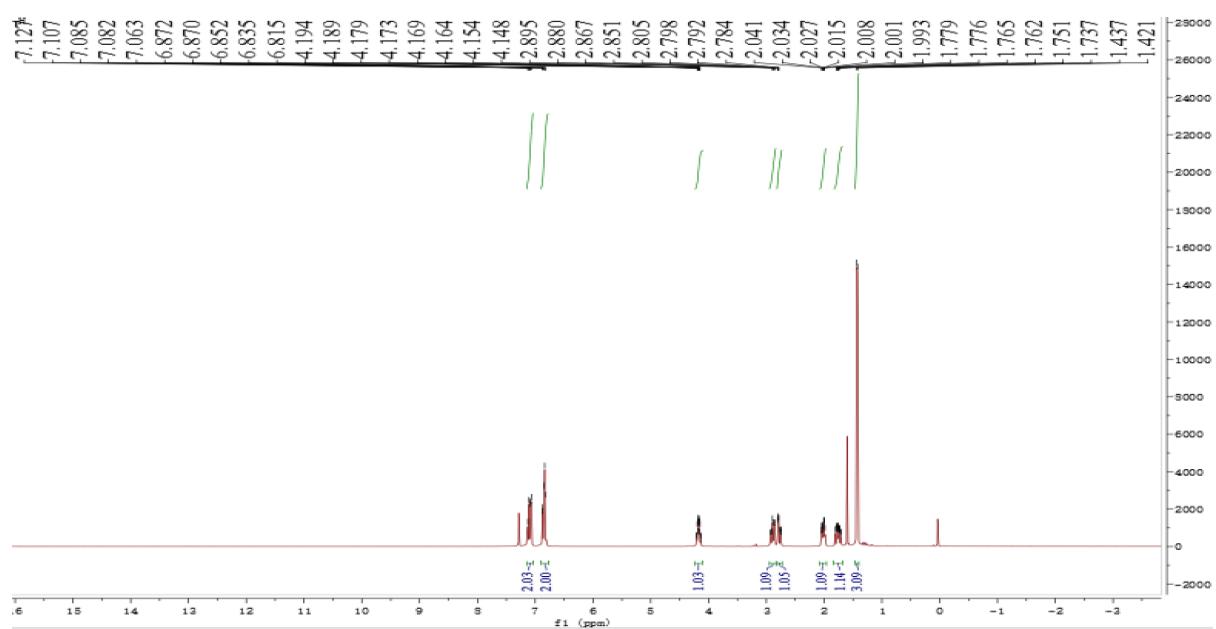
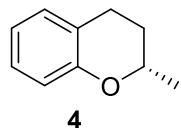
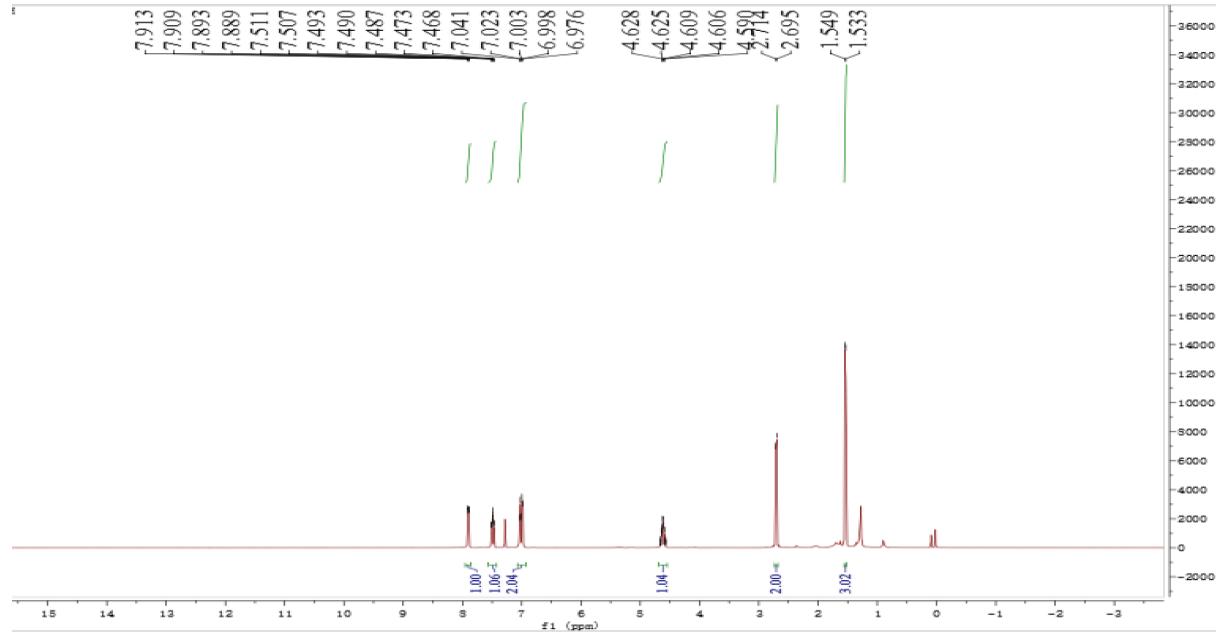
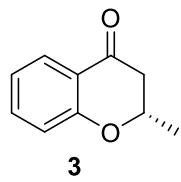


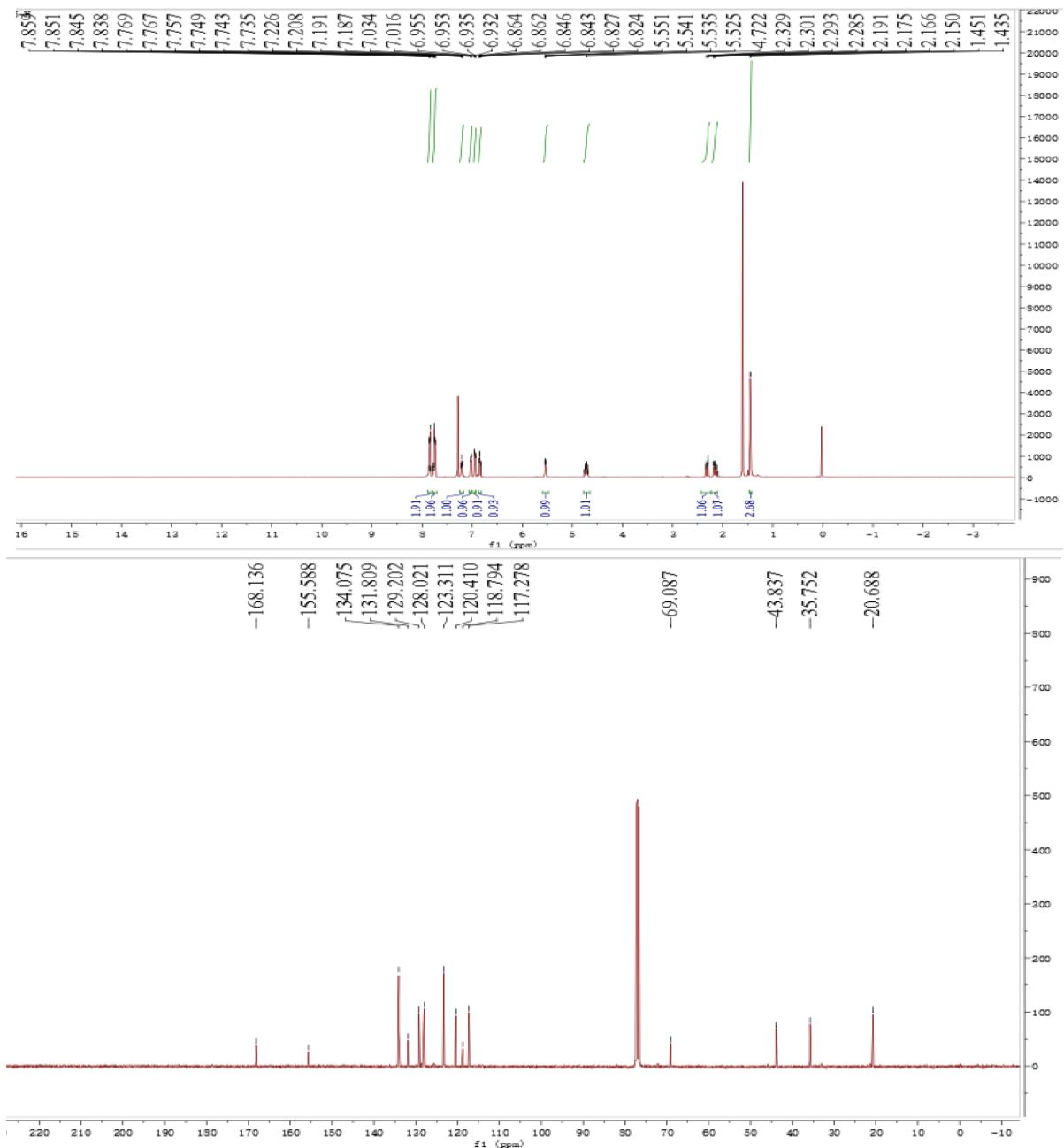
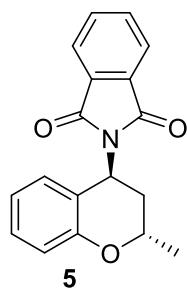


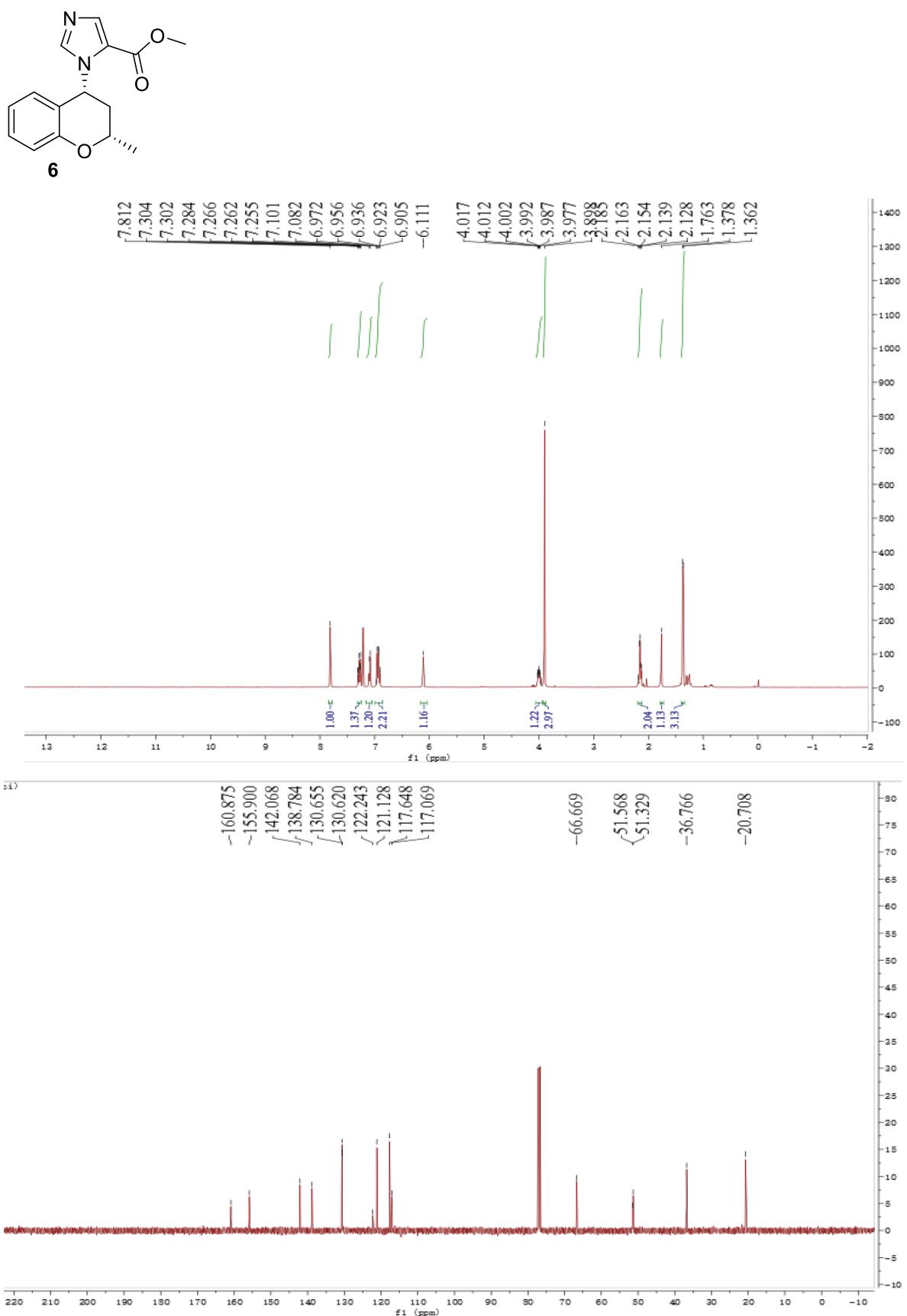


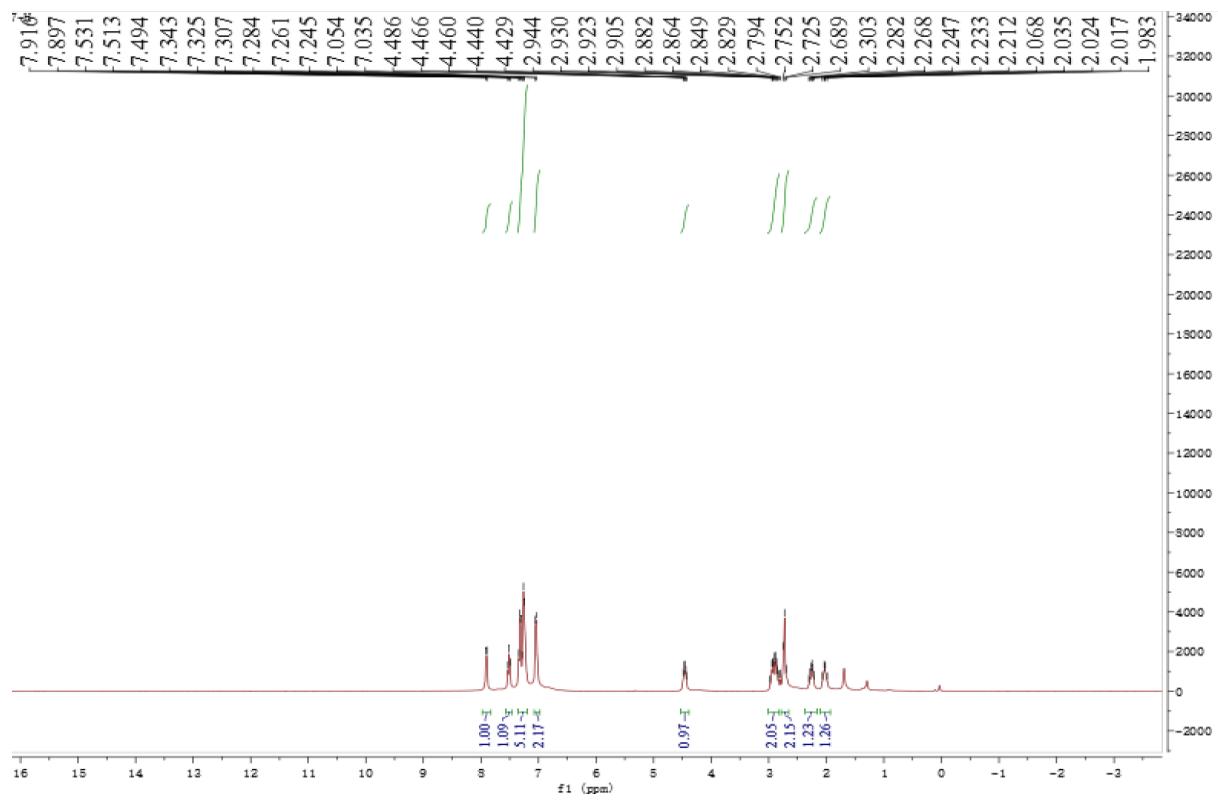
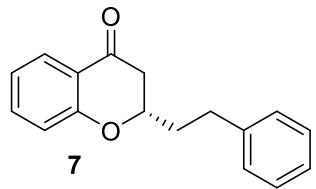




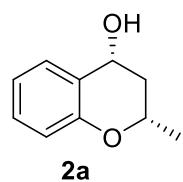






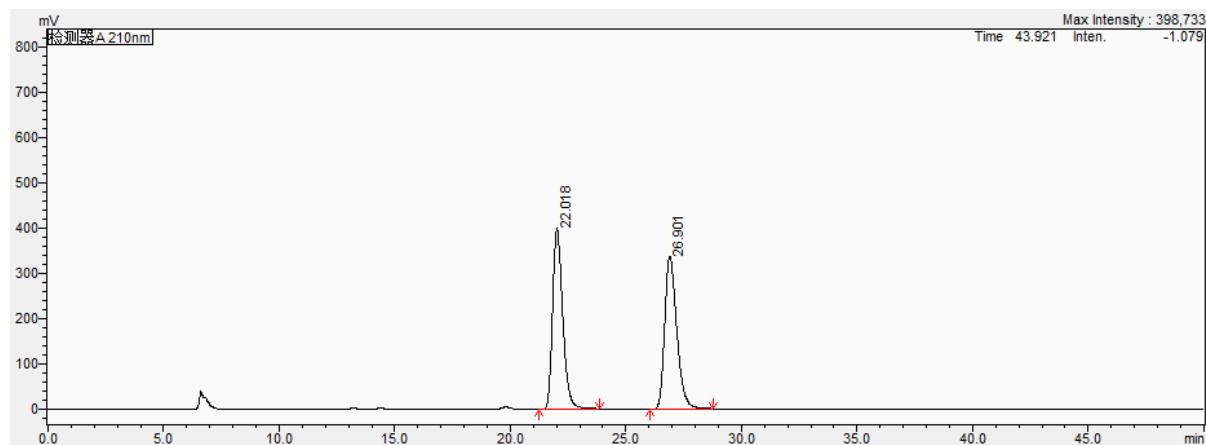


8. HPLC data

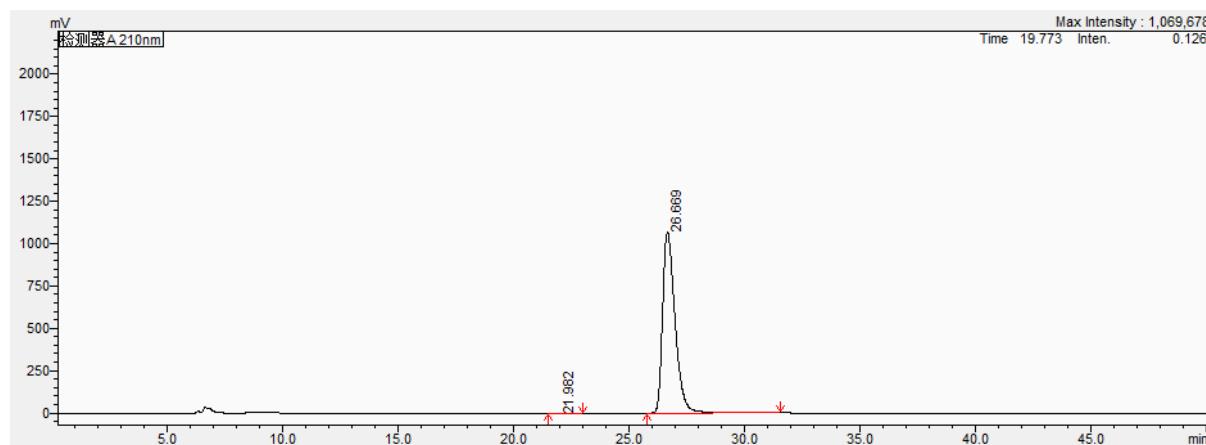


2a

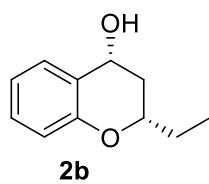
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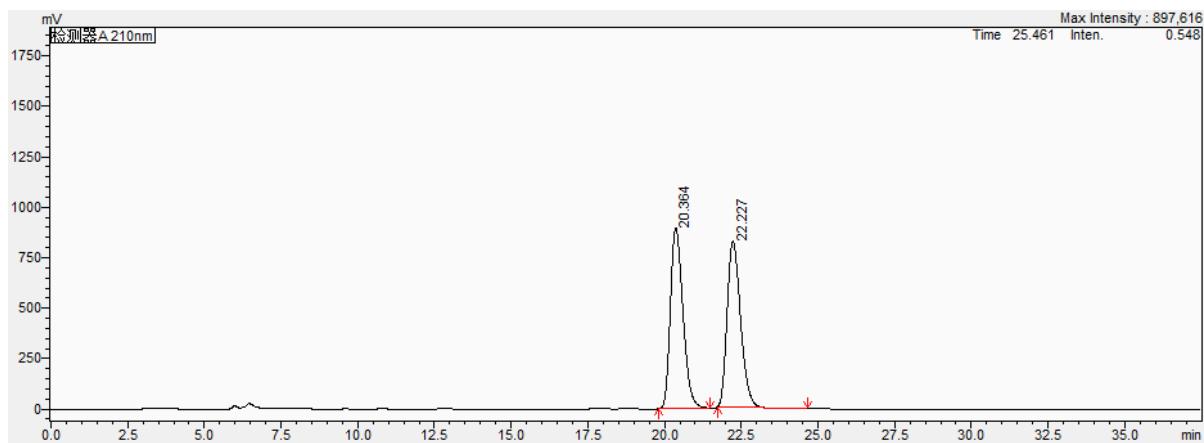
Chiral



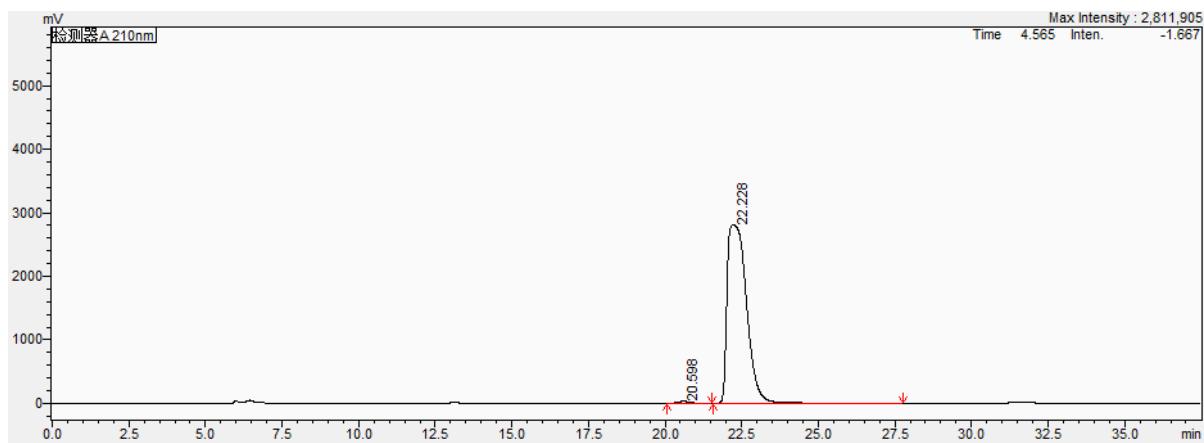
	Retention Time (min)	Area (%)	99.9% ee
Peak 1	21.982	0.054	
Peak 2	26.669	99.946	



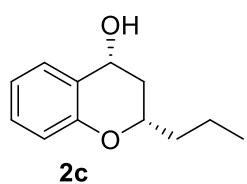
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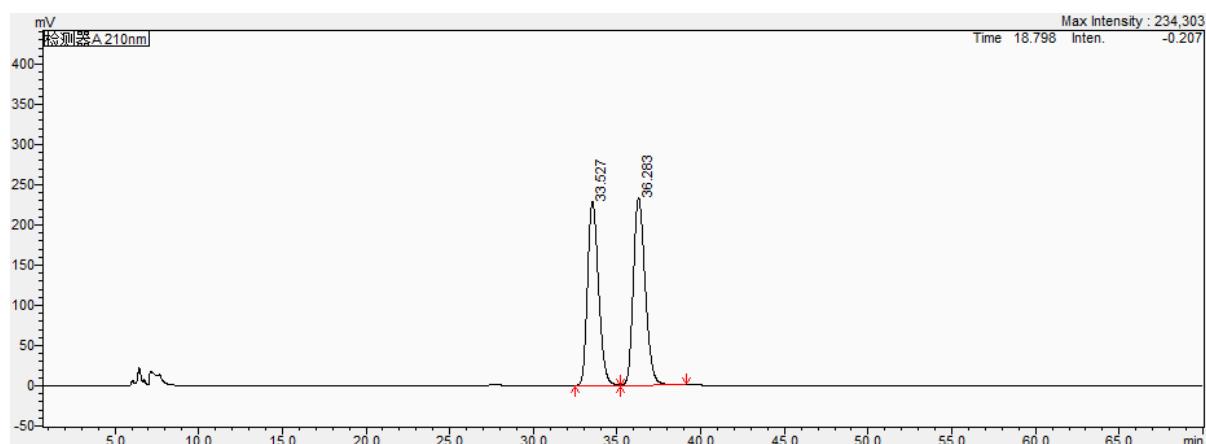
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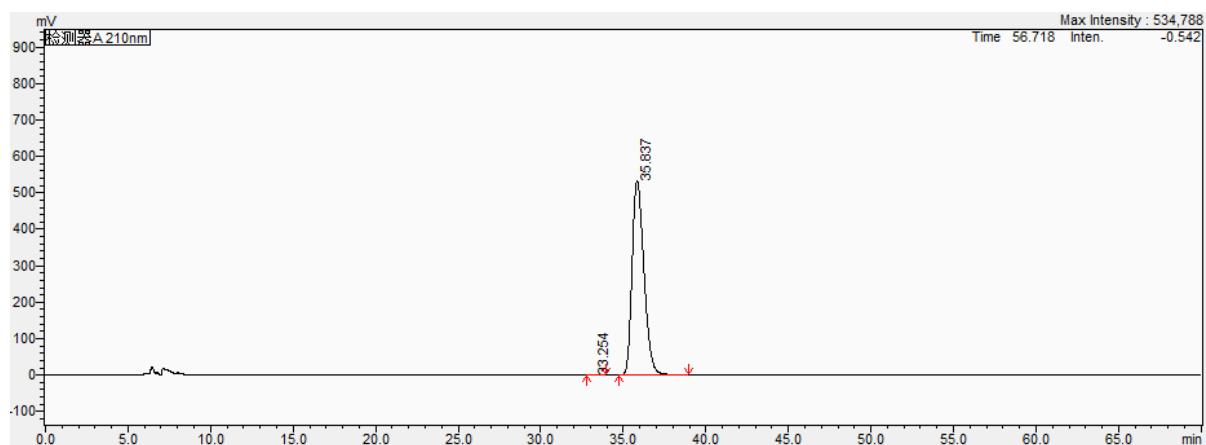
	Retention Time (min)	Area (%)	99% ee
Peak 1	20.598	0.679	
Peak 2	22.228	99.321	



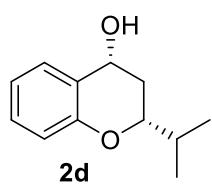
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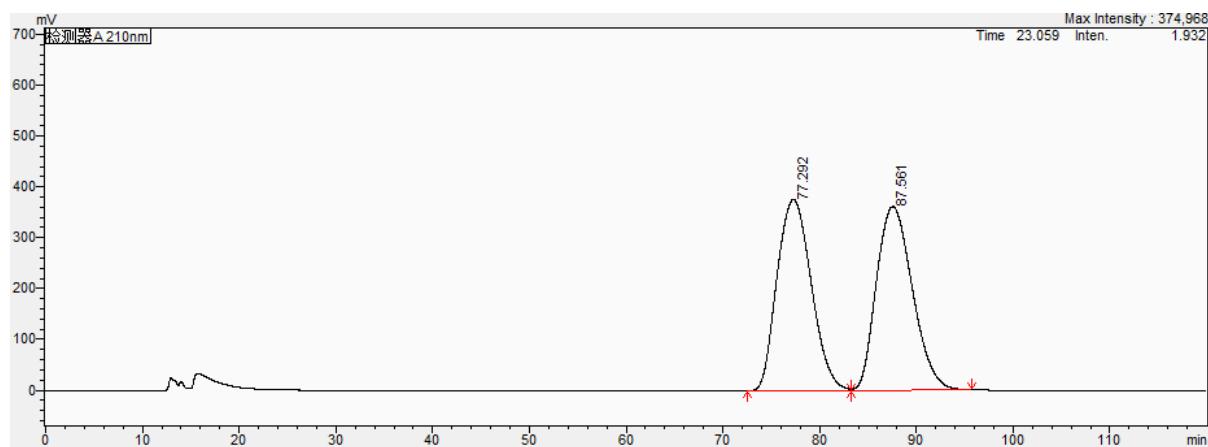
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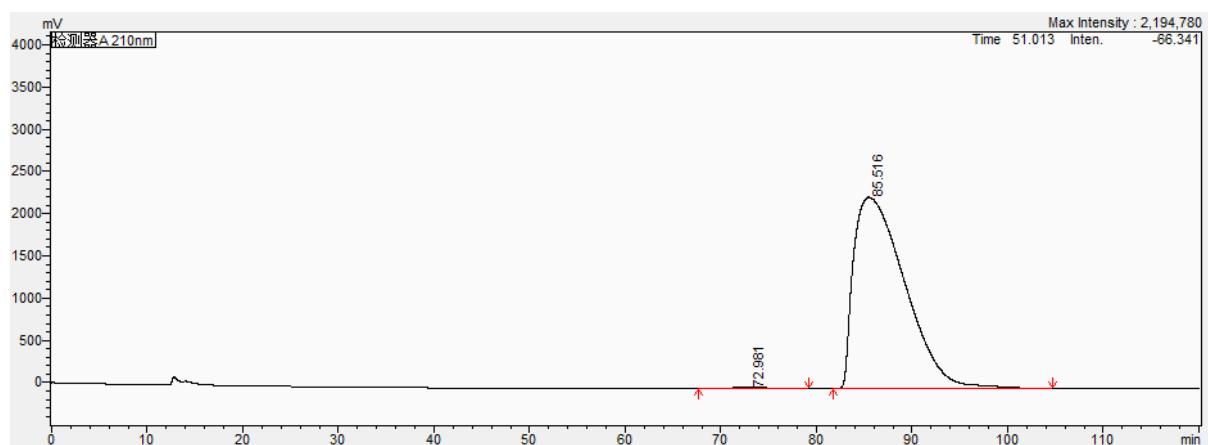
	Retention Time (min)	Area (%)	99.7% ee
Peak 1	33.254	0.134	
Peak 2	35.837	99.866	



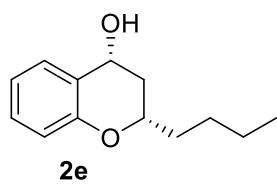
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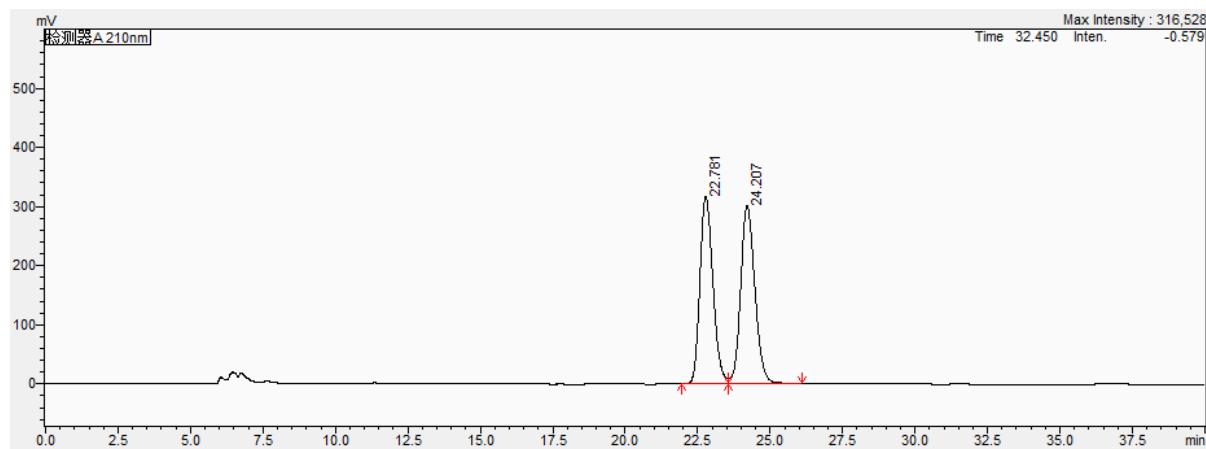
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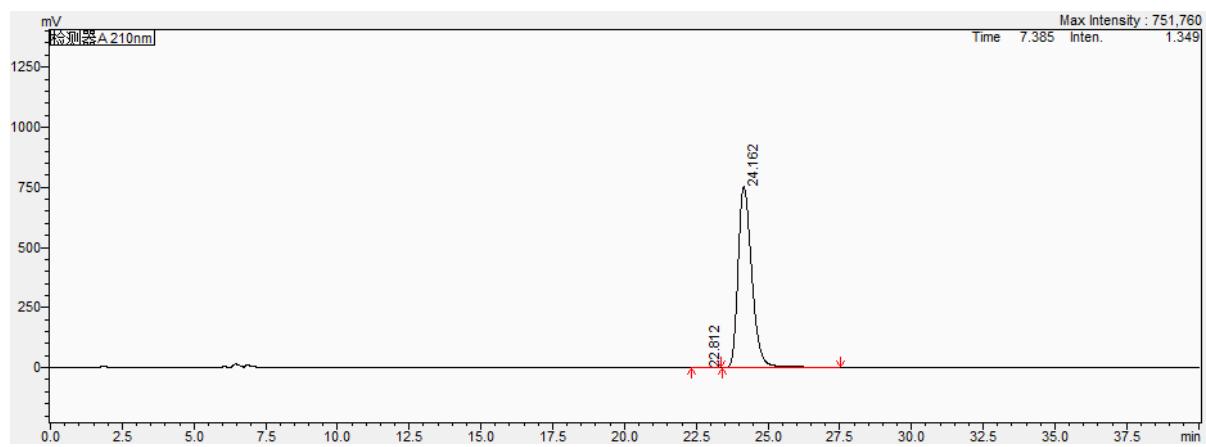
	Retention Time (min)	Area (%)	99.1% ee
Peak 1	72.981	0.433	
Peak 2	85.516	99.567	



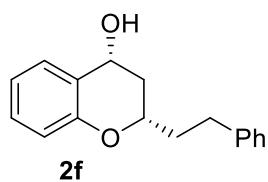
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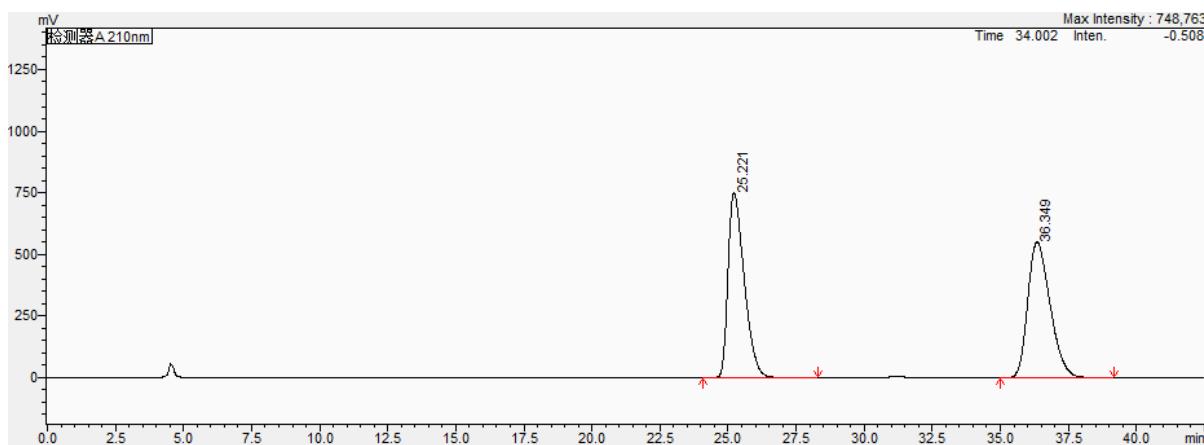
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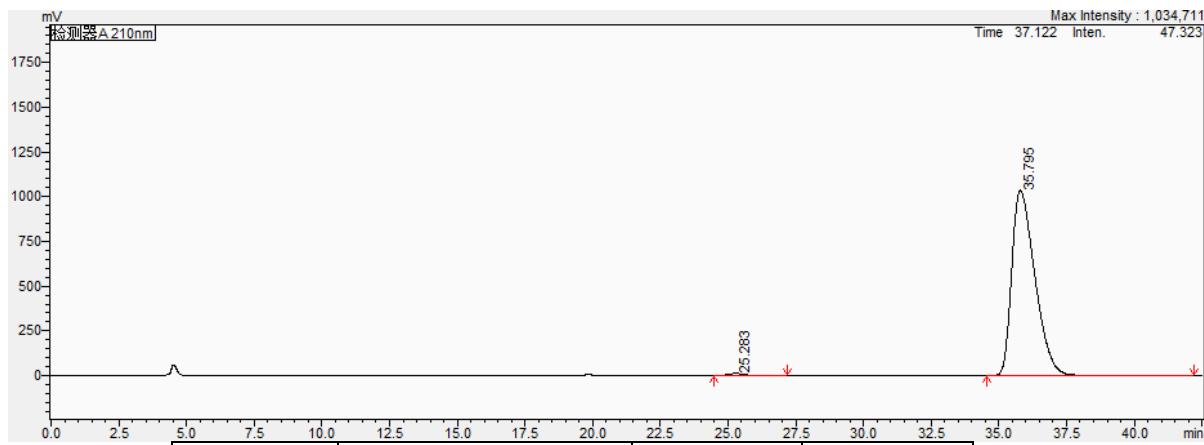
	Retention Time (min)	Area (%)	99.6 % ee
Peak 1	22.812	0.204	
Peak 2	24.162	99.796	



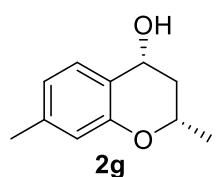
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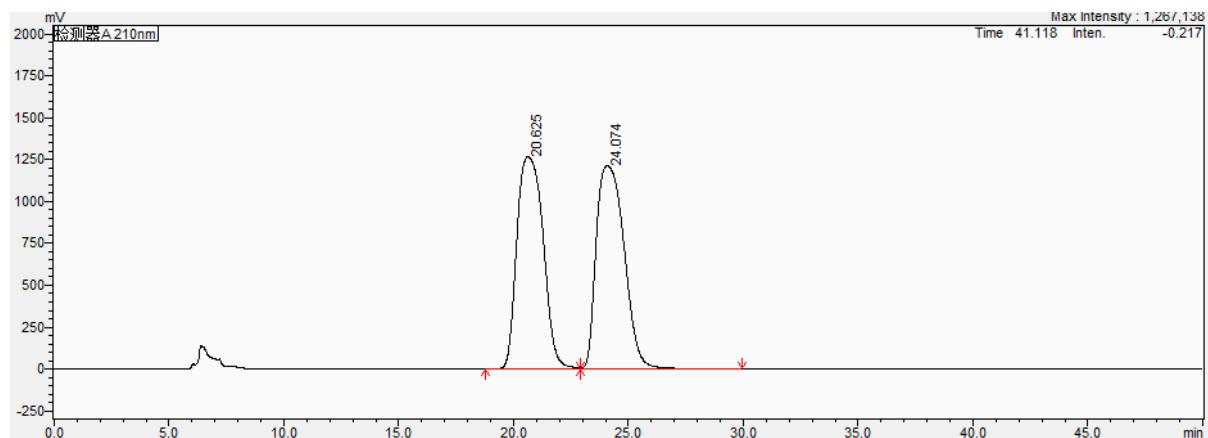
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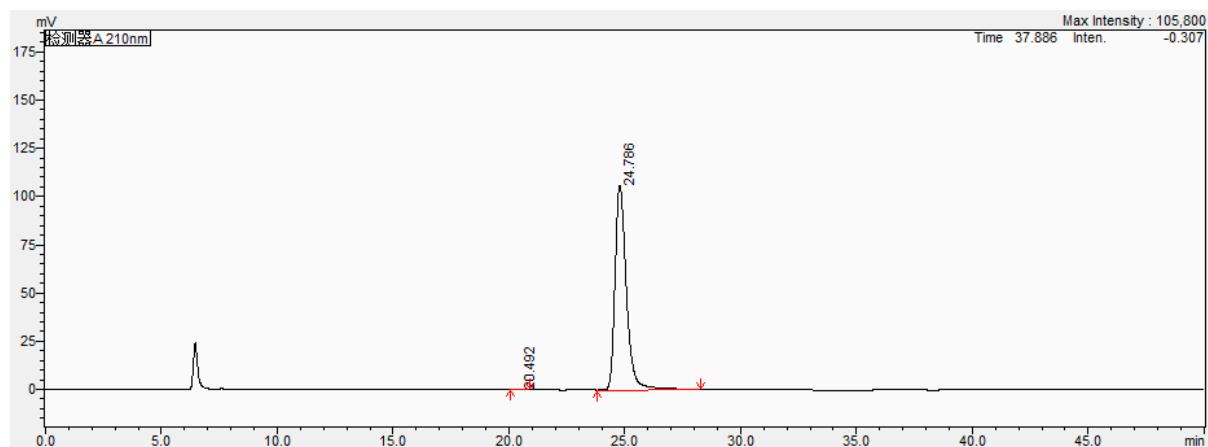
	Retention Time (min)	Area (%)	98% ee
Peak 1	25.283	0.786	
Peak 2	35.795	99.214	



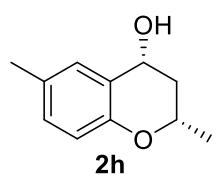
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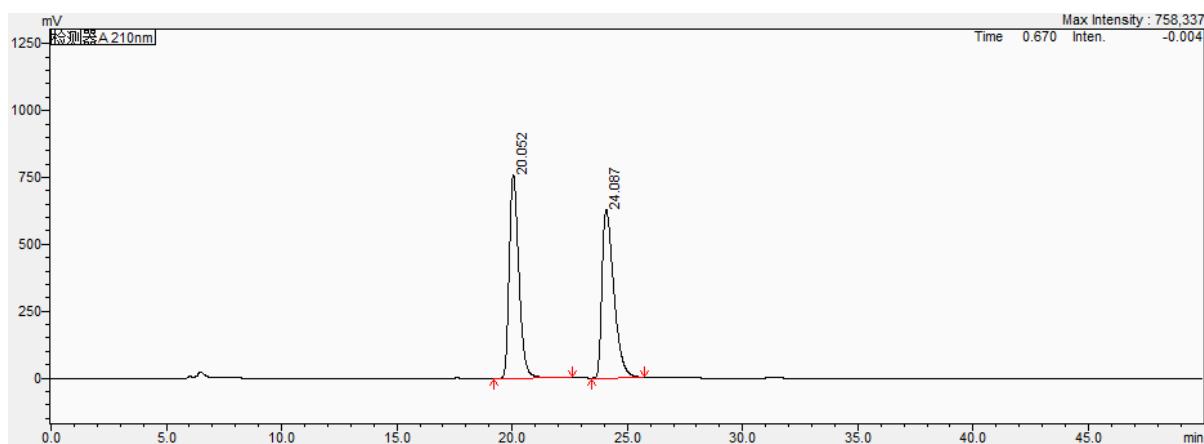
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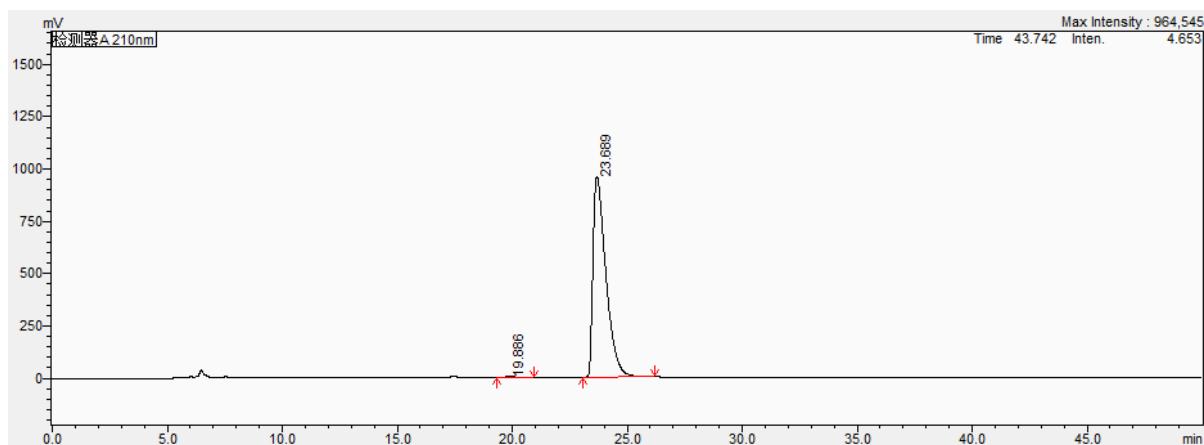
	Retention Time (min)	Area (%)	99.7% ee
Peak 1	20.492	0.168	
Peak 2	24.786	99.832	



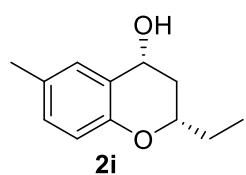
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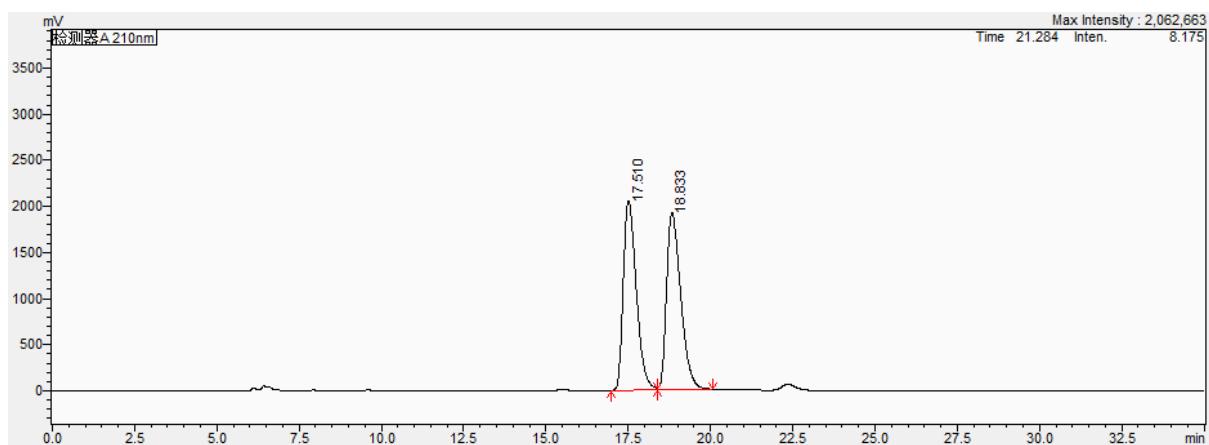
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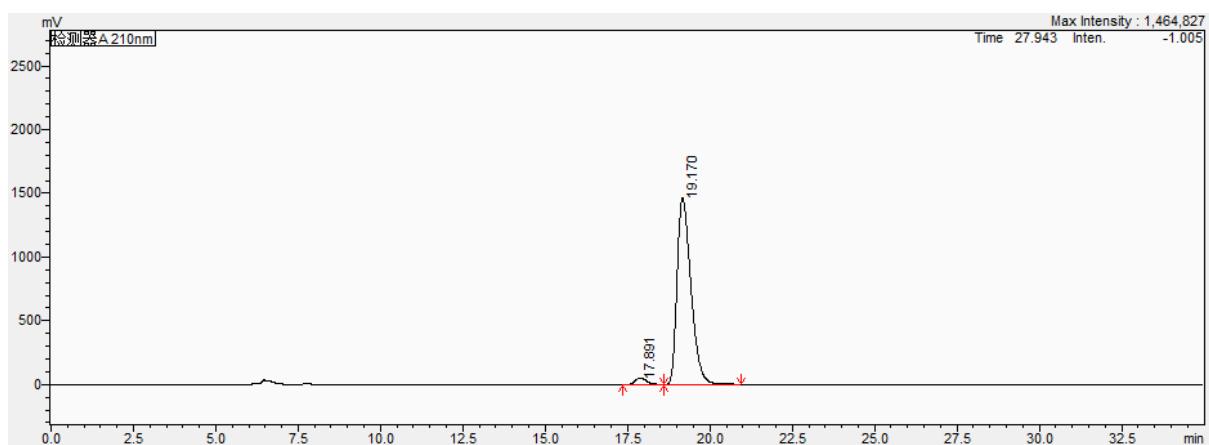
	Retention Time (min)	Area (%)	99.3% ee
Peak 1	19.886	0.323	
Peak 2	23.689	99.677	



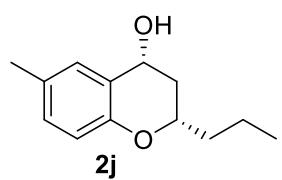
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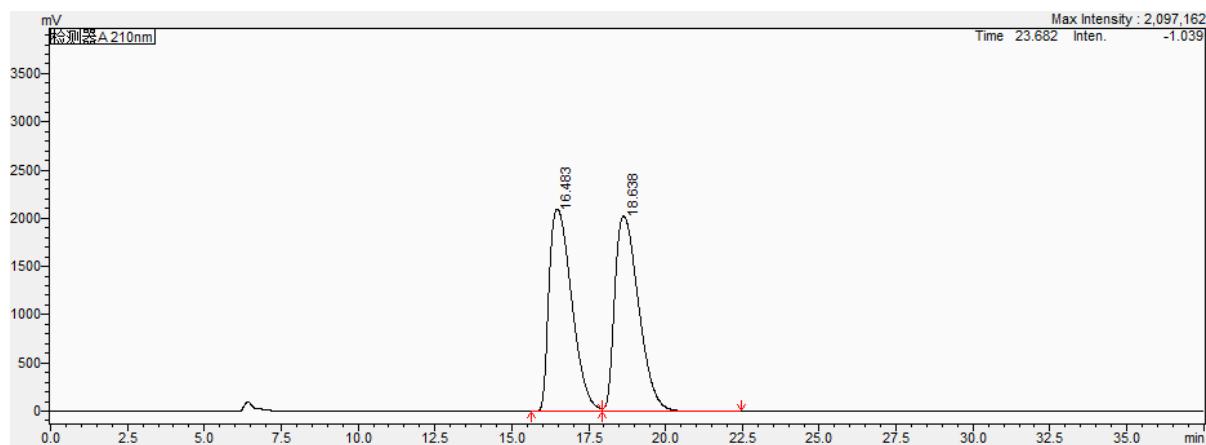
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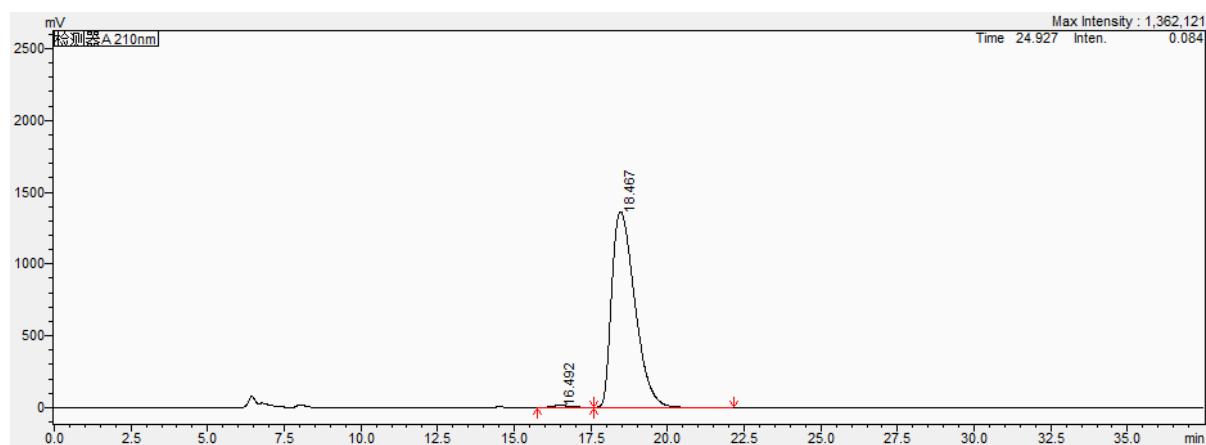
	Retention Time (min)	Area (%)	94% ee
Peak 1	17.891	2.867	
Peak 2	19.170	97.133	



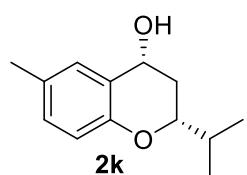
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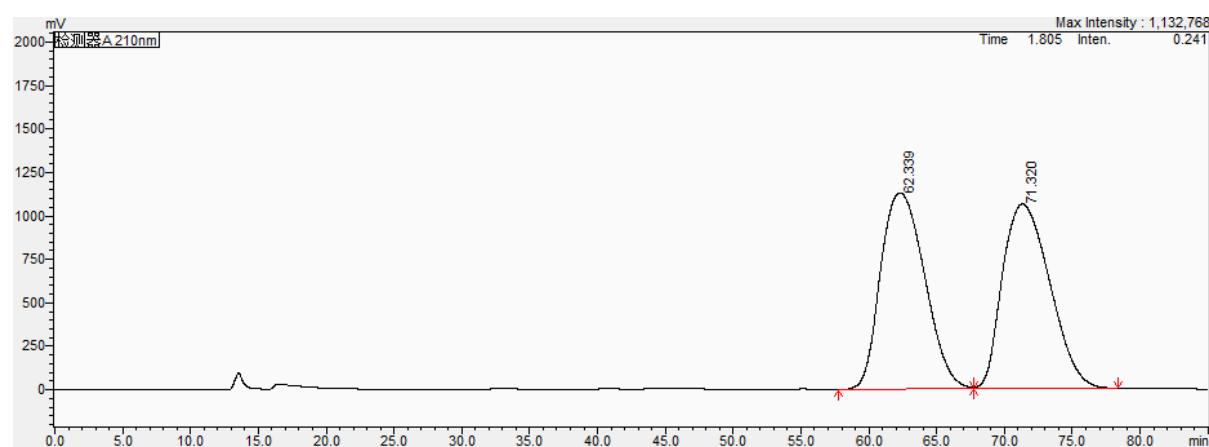
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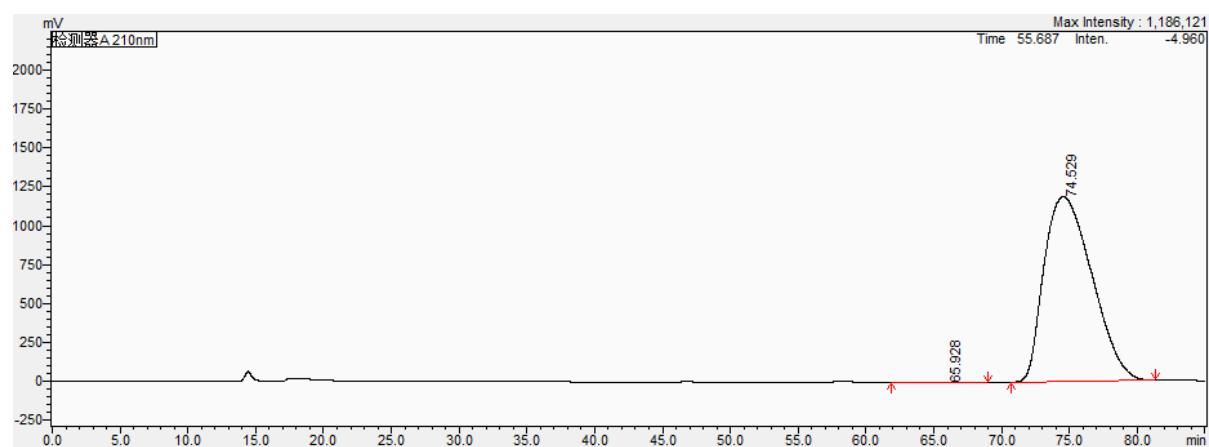
	Retention Time (min)	Area (%)	98% ee
Peak 1	16.492	0.857	
Peak 2	18.467	99.143	



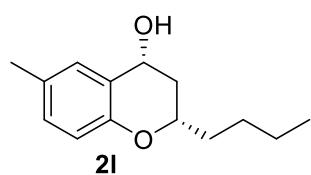
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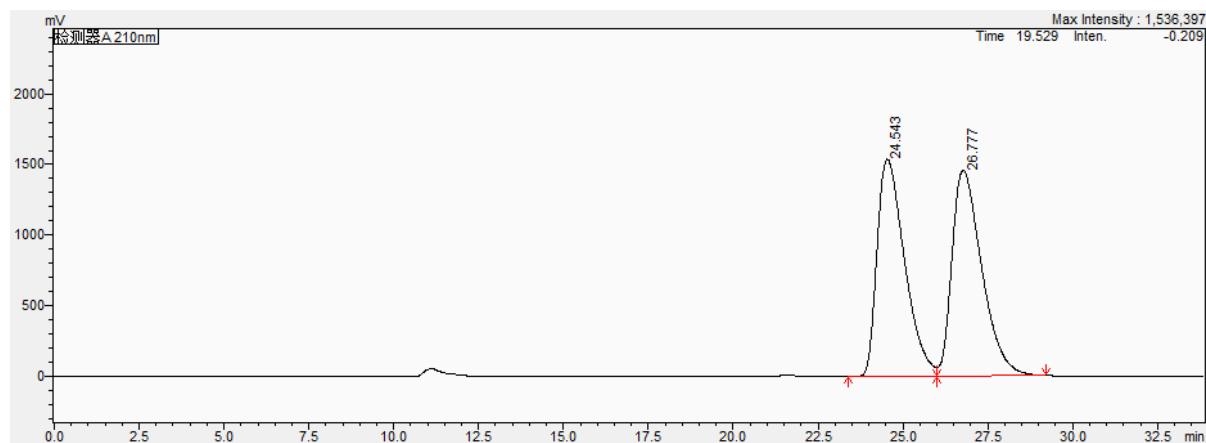
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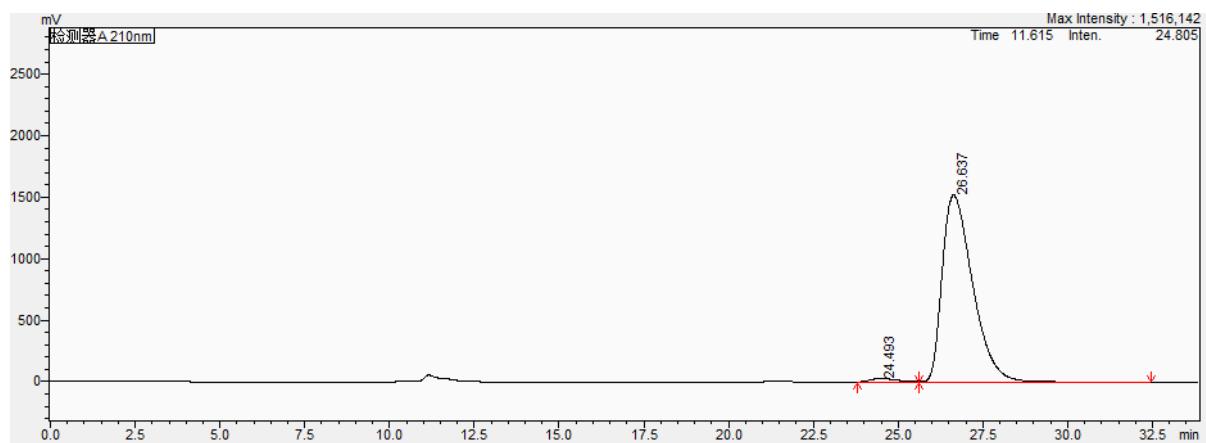
	Retention Time (min)	Area (%)	99.8% ee
Peak 1	65.928	0.099	
Peak 2	74.529	99.901	



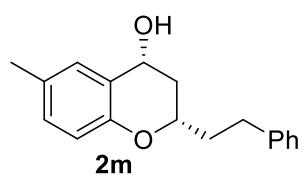
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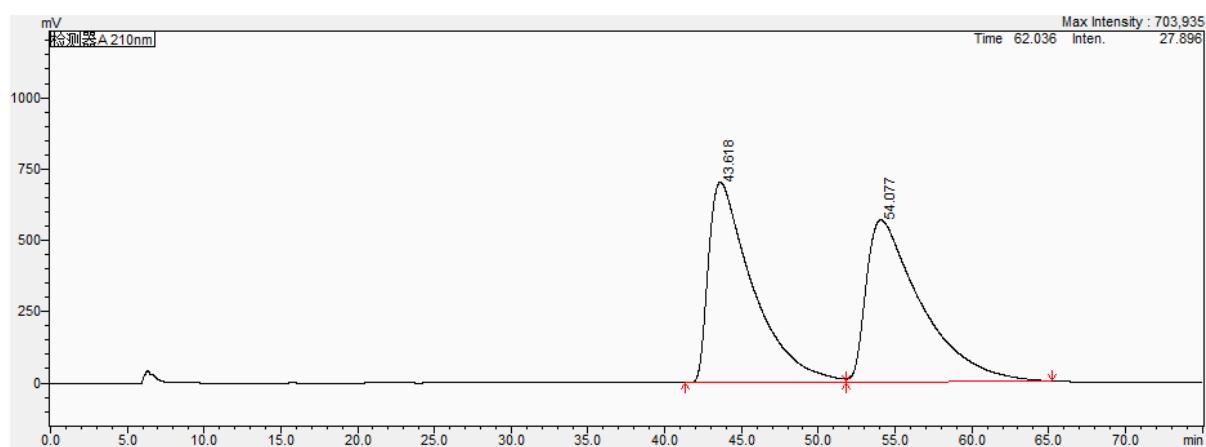
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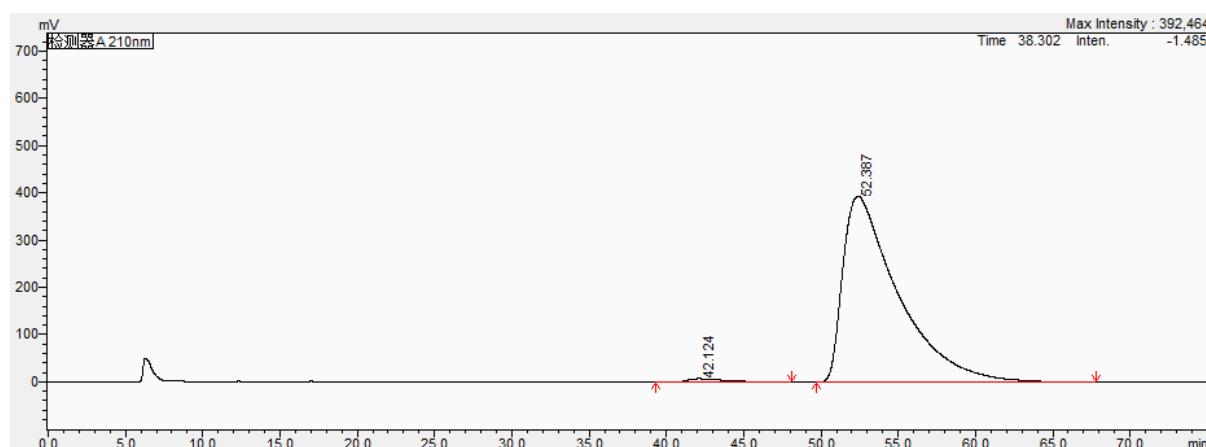
	Retention Time (min)	Area (%)	97% ee
Peak 1	24.493	1.523	
Peak 2	26.637	98.477	



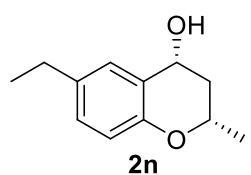
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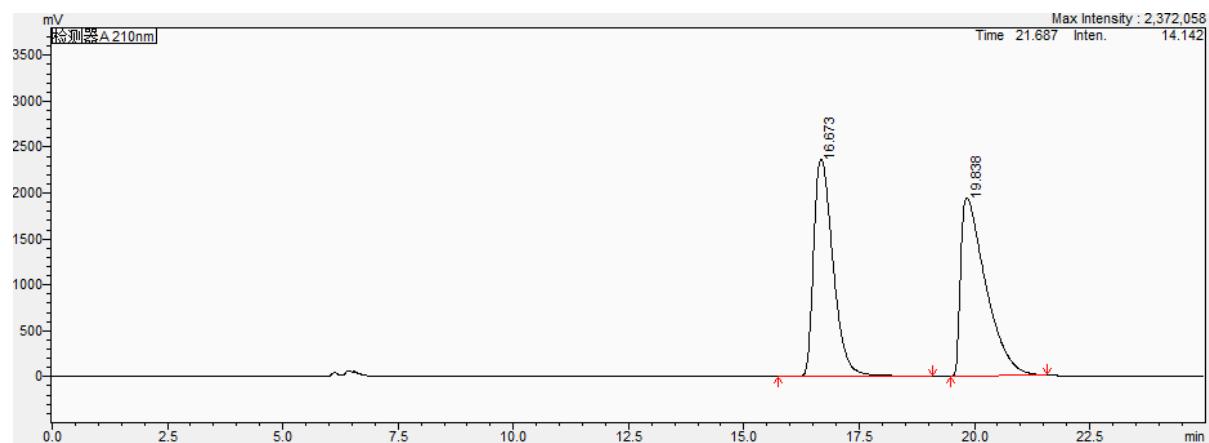
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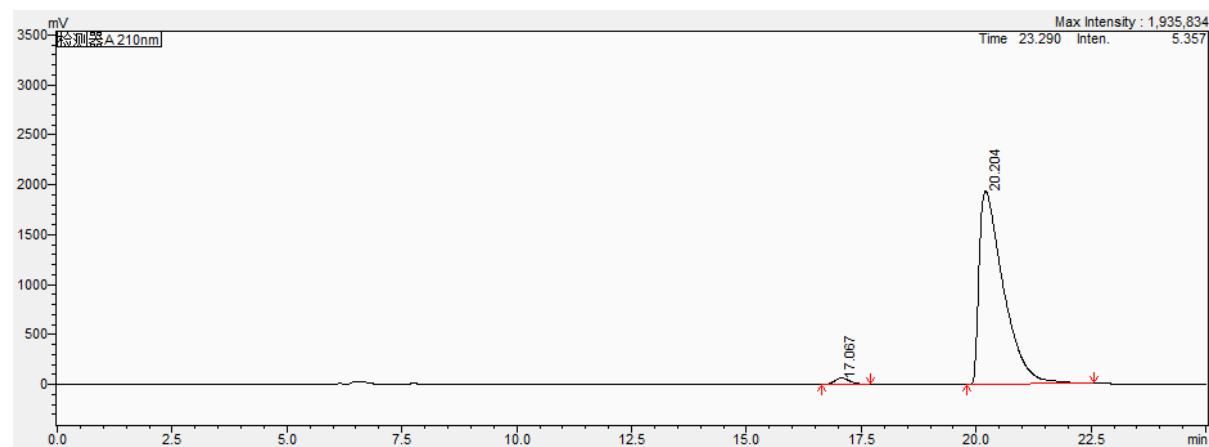
	Retention Time (min)	Area (%)	97% ee
Peak 1	42.124	1.434	
Peak 2	52.387	98.566	



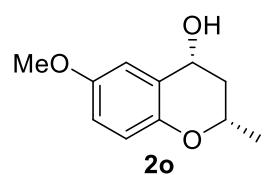
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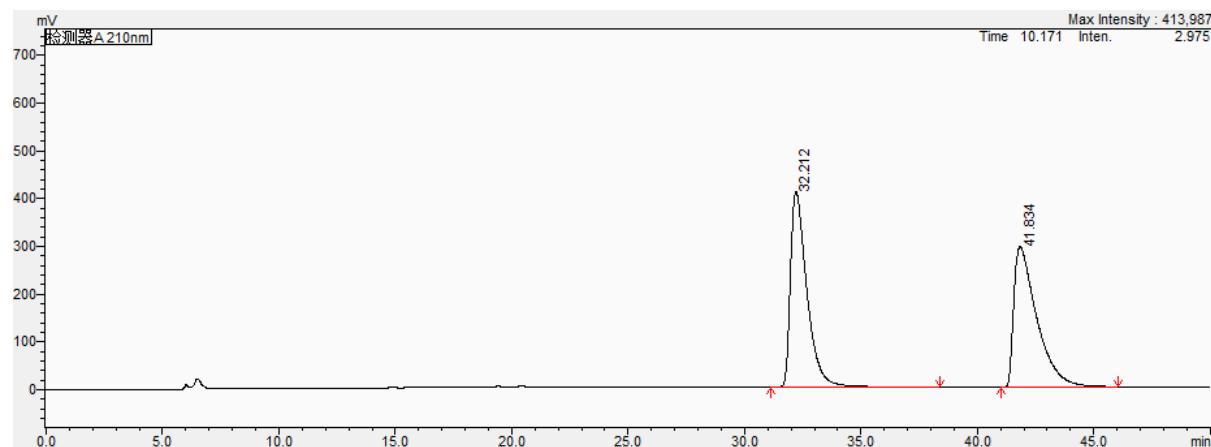
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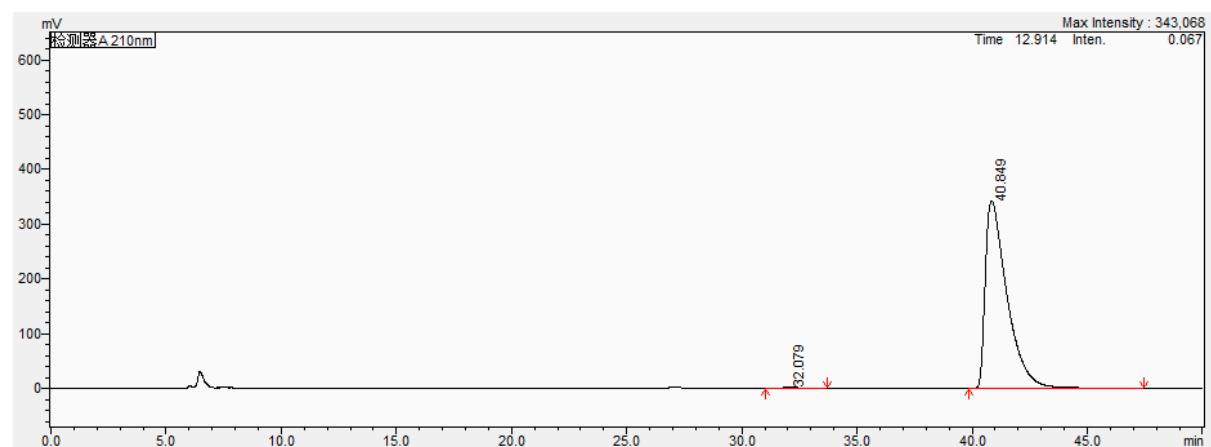
	Retention Time (min)	Area (%)	96% ee
Peak 1	17.067	1.914	
Peak 2	20.204	98.086	



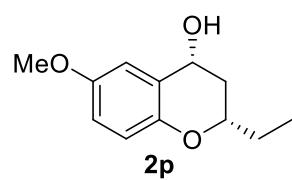
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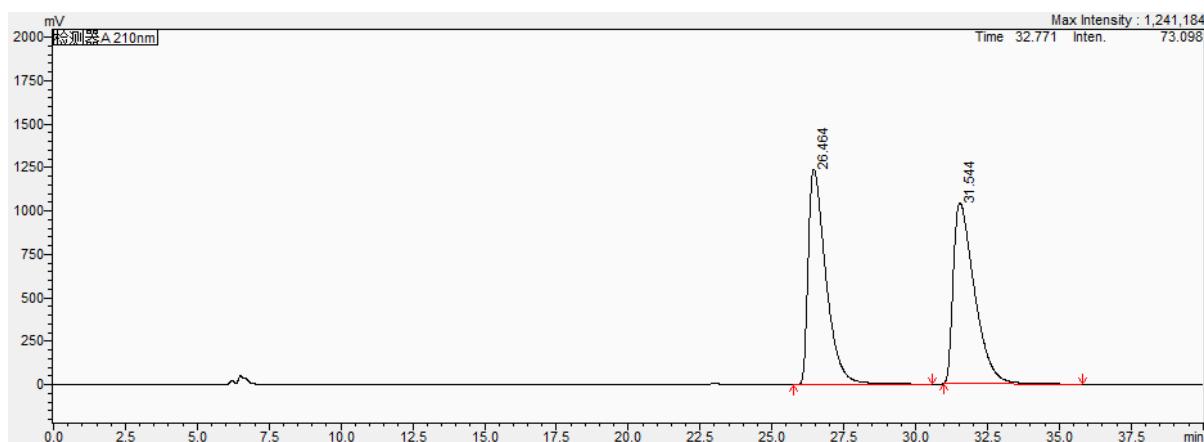
Chiral



	Retention Time (min)	Area (%)	99.1% ee
Peak 1	32.079	0.437	
Peak 2	40.849	99.563	



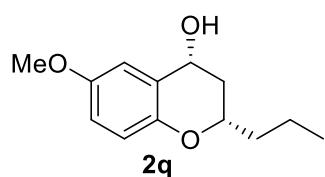
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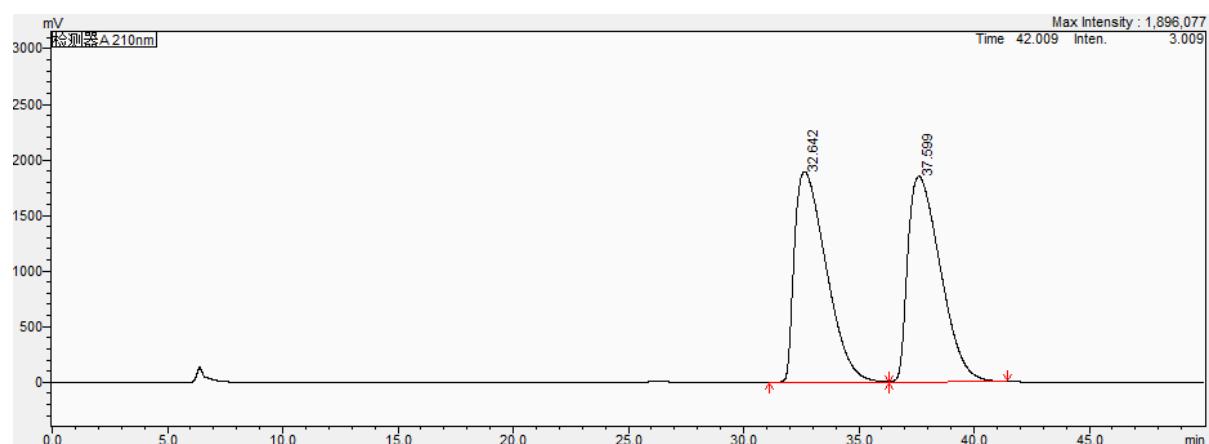
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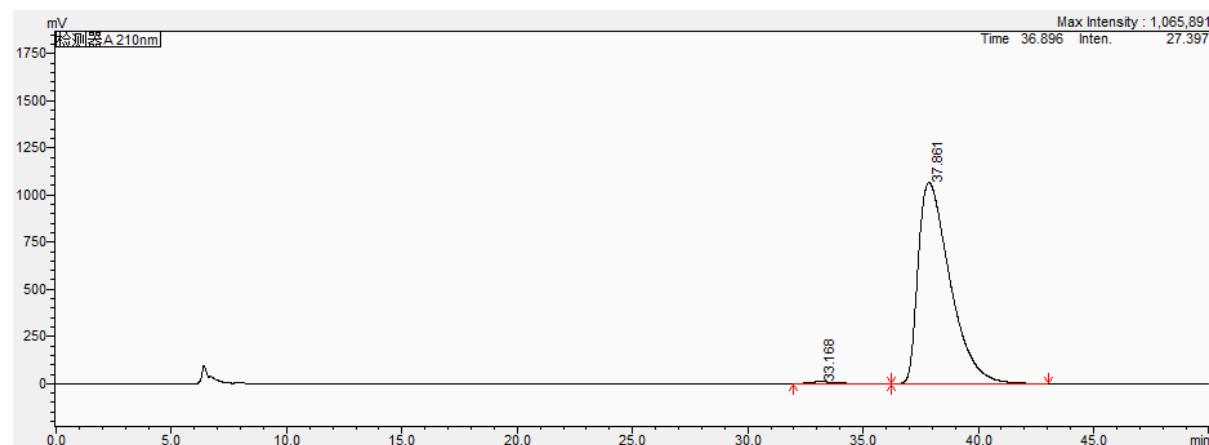
	Retention Time (min)	Area (%)	98% ee
Peak 1	26.736	0.942	
Peak 2	31.238	99.058	



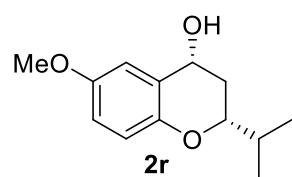
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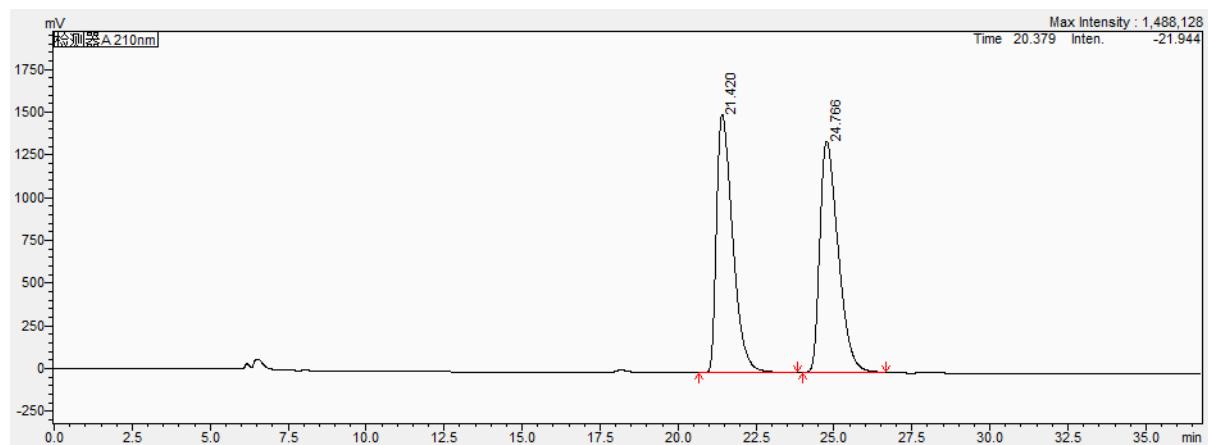
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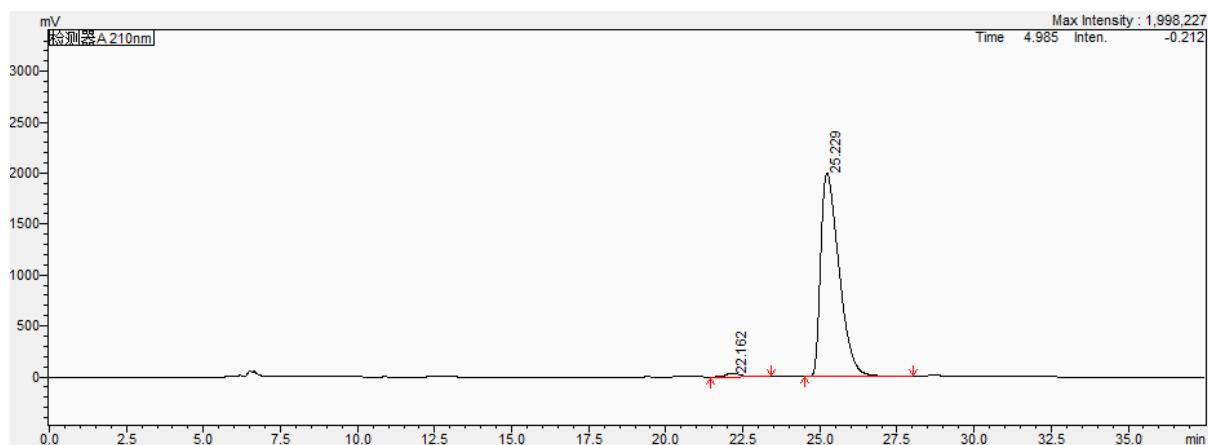
	Retention Time (min)	Area (%)	98% ee
Peak 1	33.168	0.856	
Peak 2	37.861	99.144	



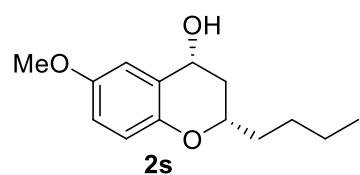
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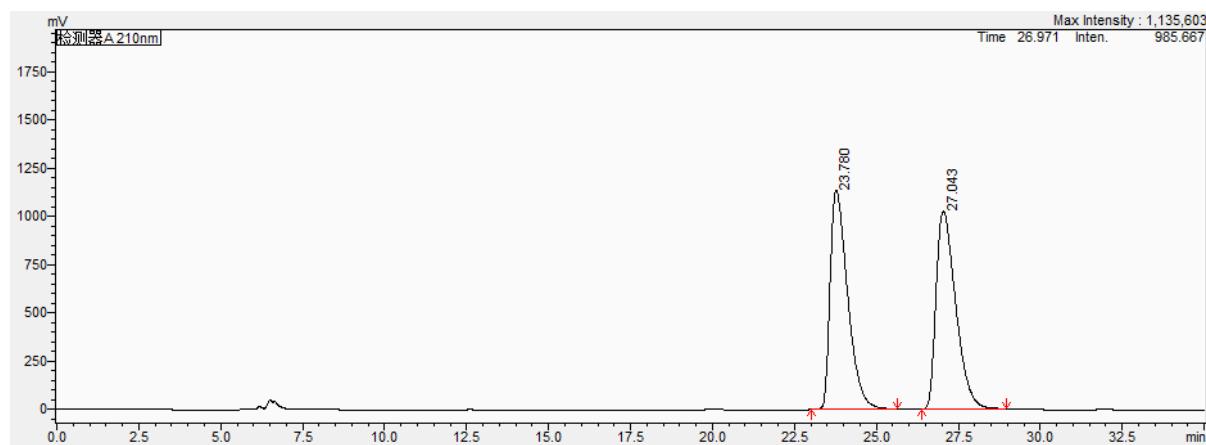
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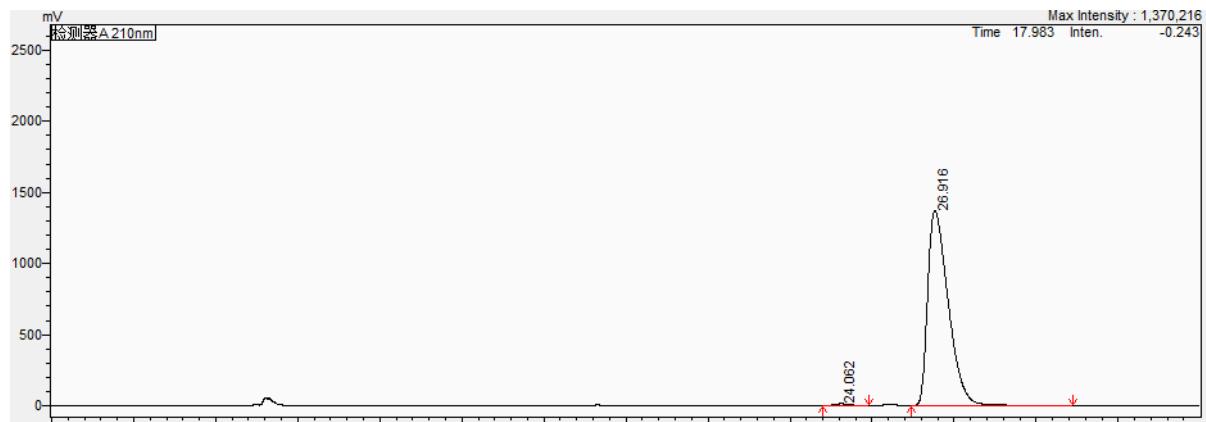
	Retention Time (min)	Area (%)	97% ee
Peak 1	22.162	1.344	
Peak 2	25.229	98.656	



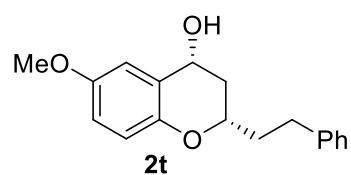
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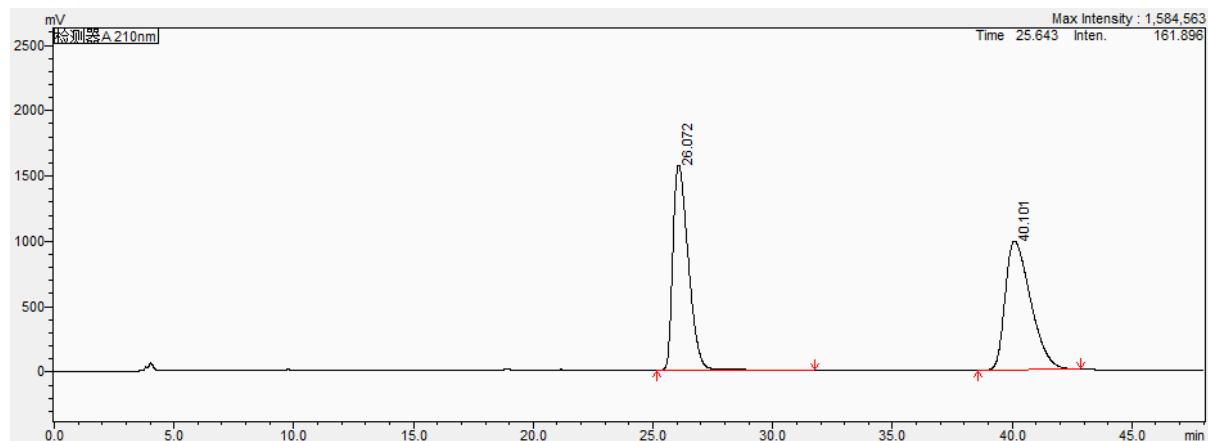
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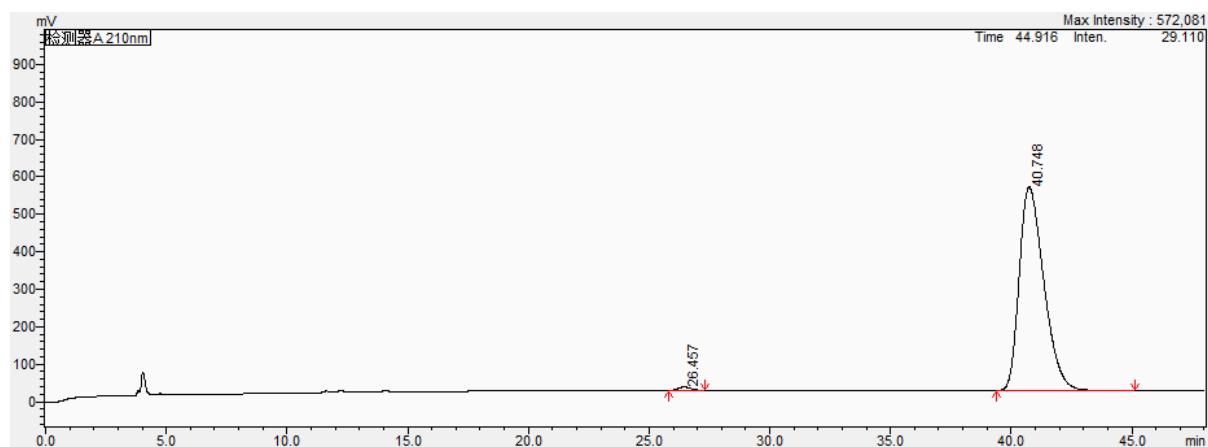
	Retention Time (min)	Area (%)	99% ee
Peak 1	24.062	0.710	
Peak 2	26.916	99.290	



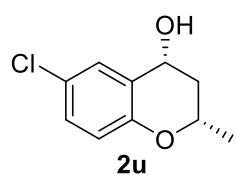
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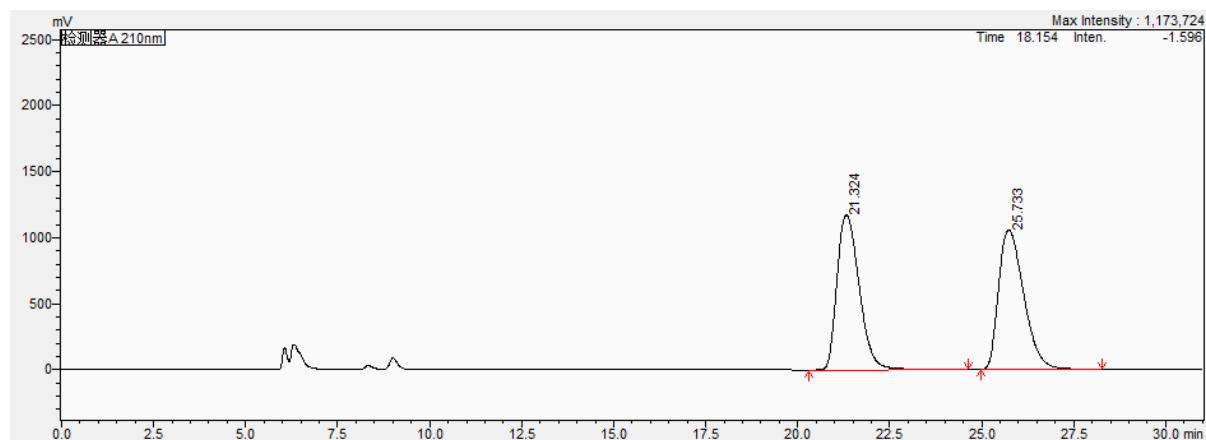
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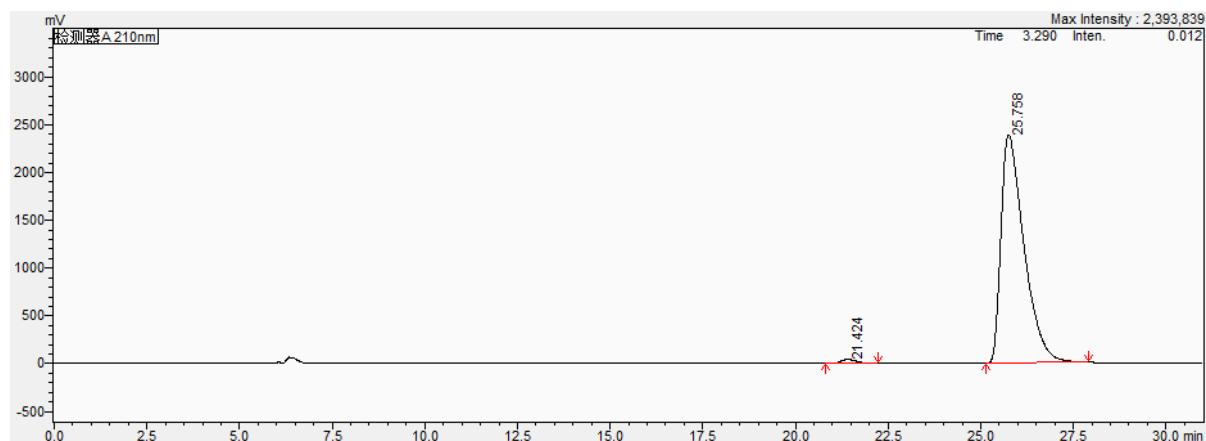
	Retention Time (min)	Area (%)	98% ee
Peak 1	26.457	1.023	
Peak 2	40.748	98.977	



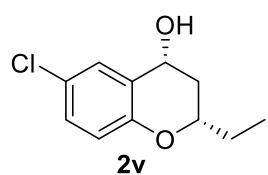
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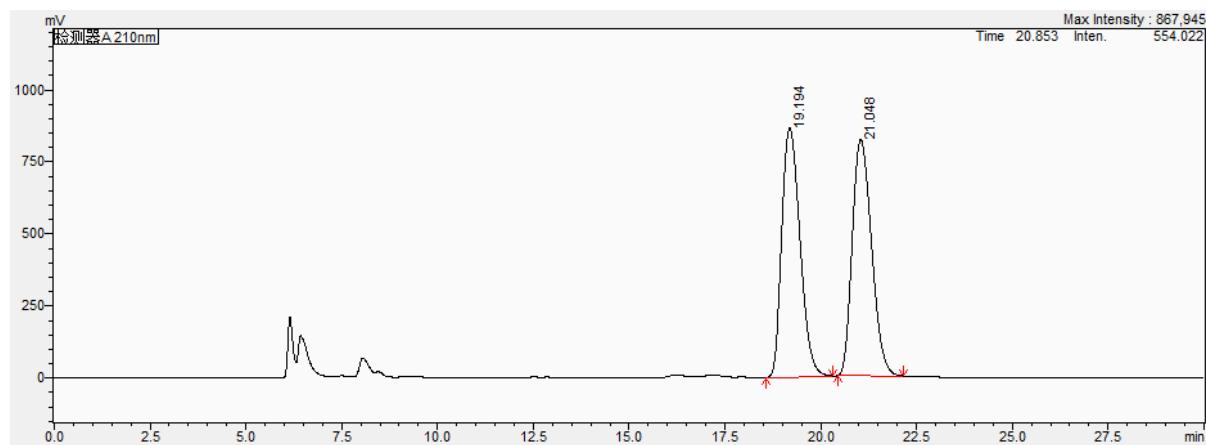
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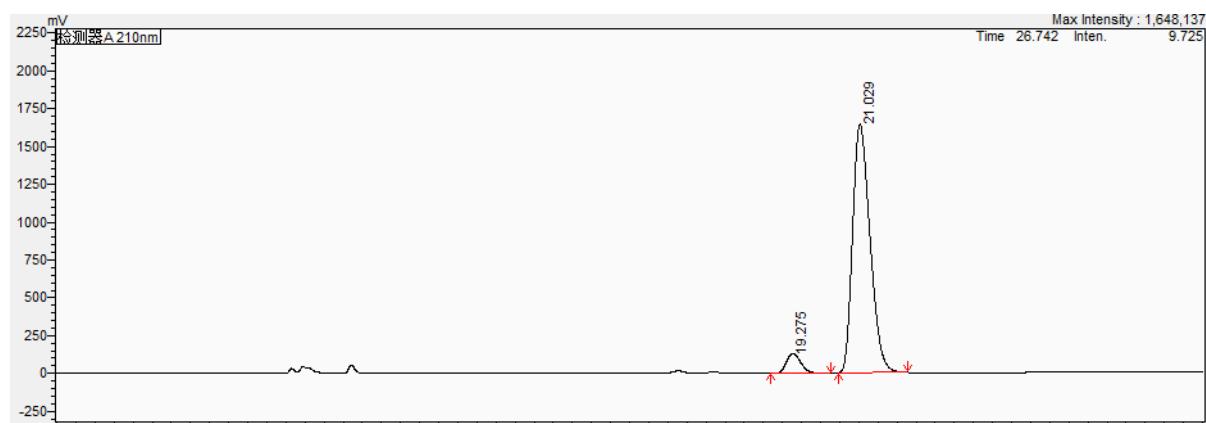
	Retention Time (min)	Area (%)	98% ee
Peak 1	21.424	1.230	
Peak 2	25.758	98.770	



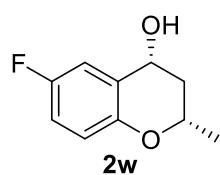
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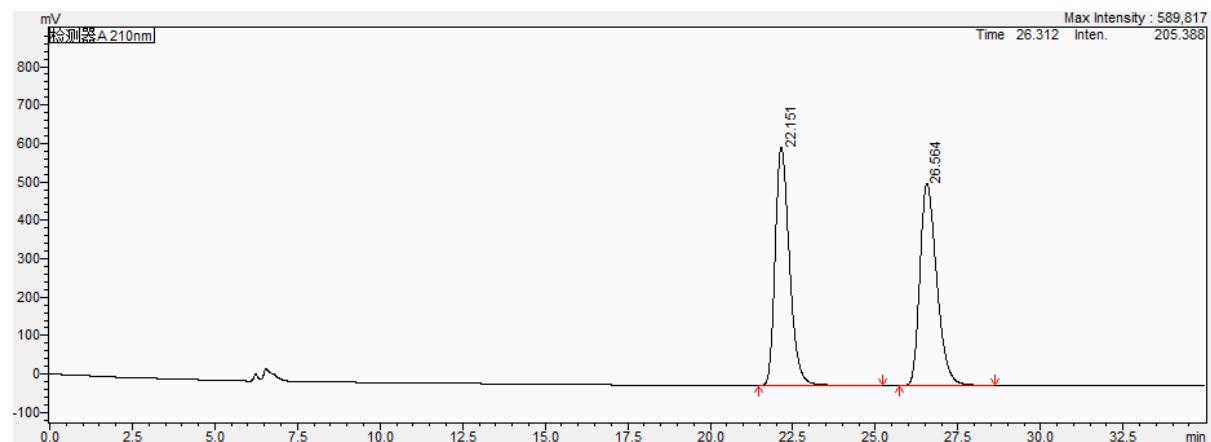
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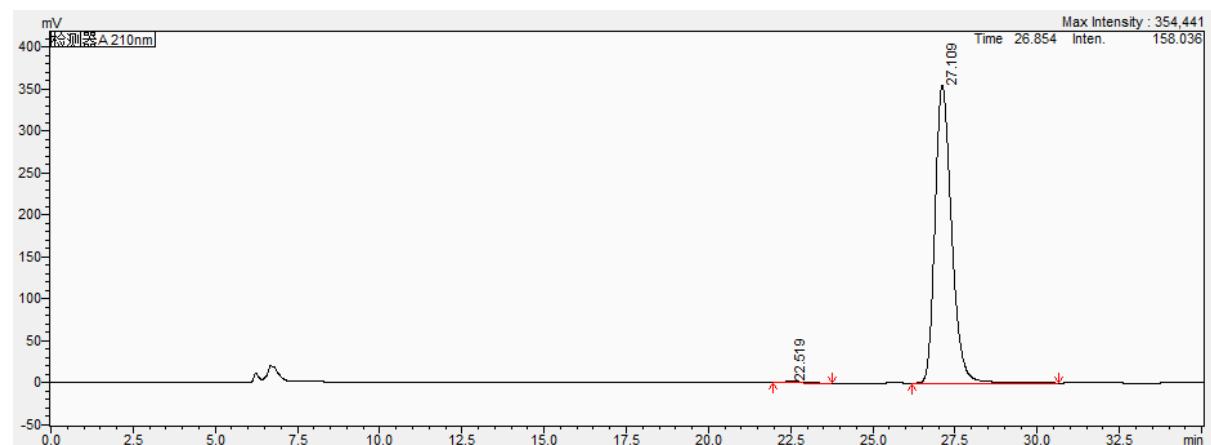
	Retention Time (min)	Area (%)	87% ee
Peak 1	19.275	6.311	
Peak 2	21.029	93.689	



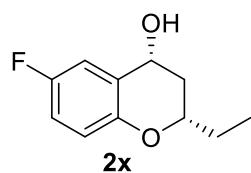
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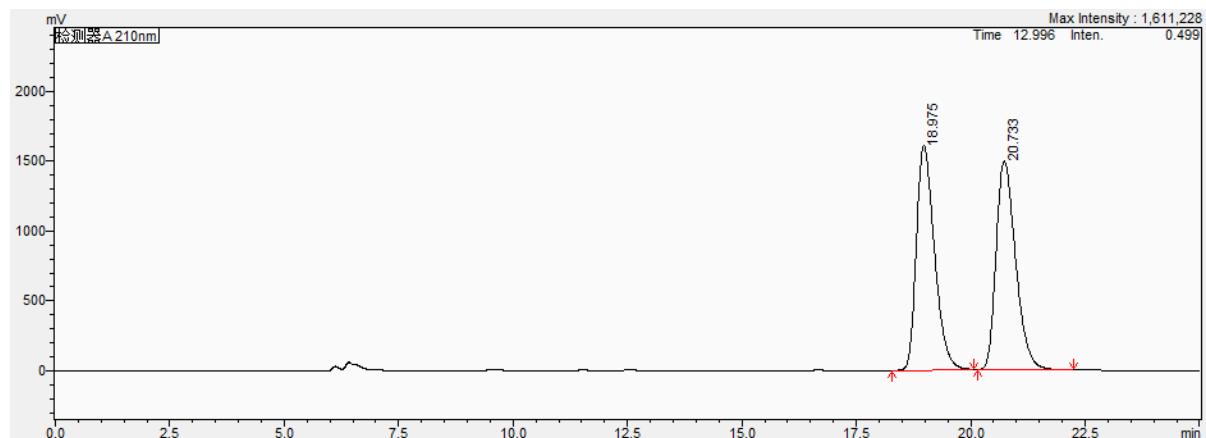
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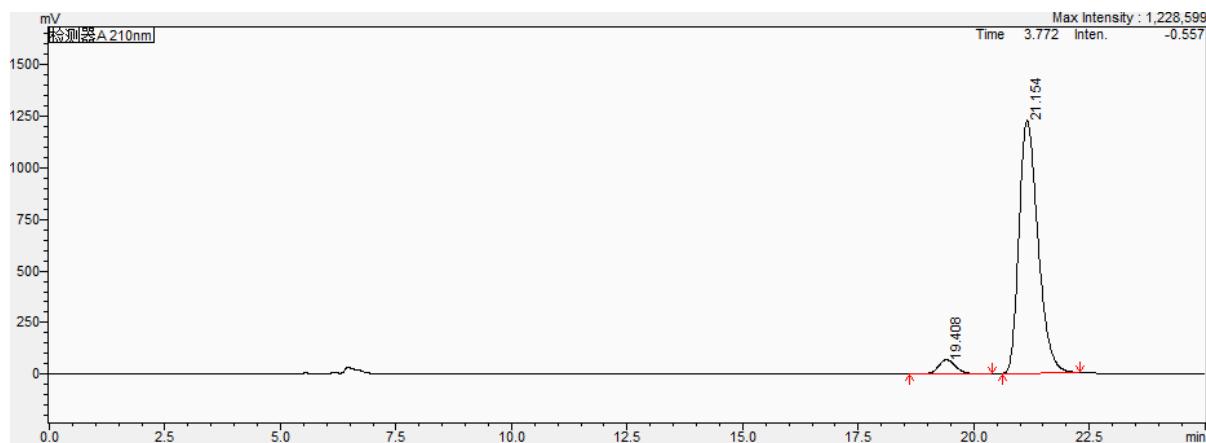
	Retention Time (min)	Area (%)	99% ee
Peak 1	22.519	0.683	
Peak 2	27.109	99.317	



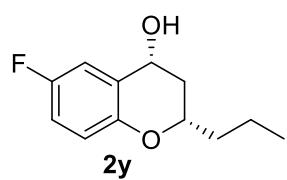
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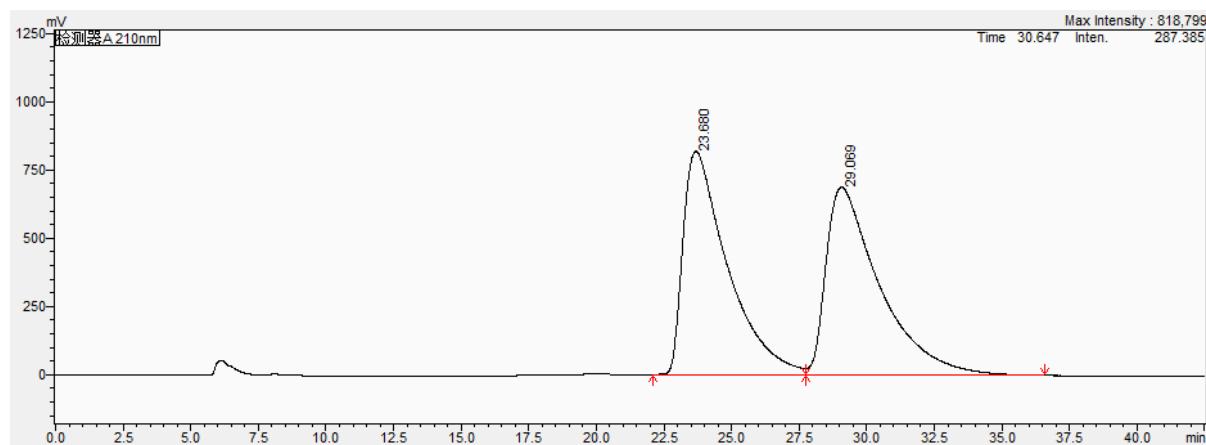
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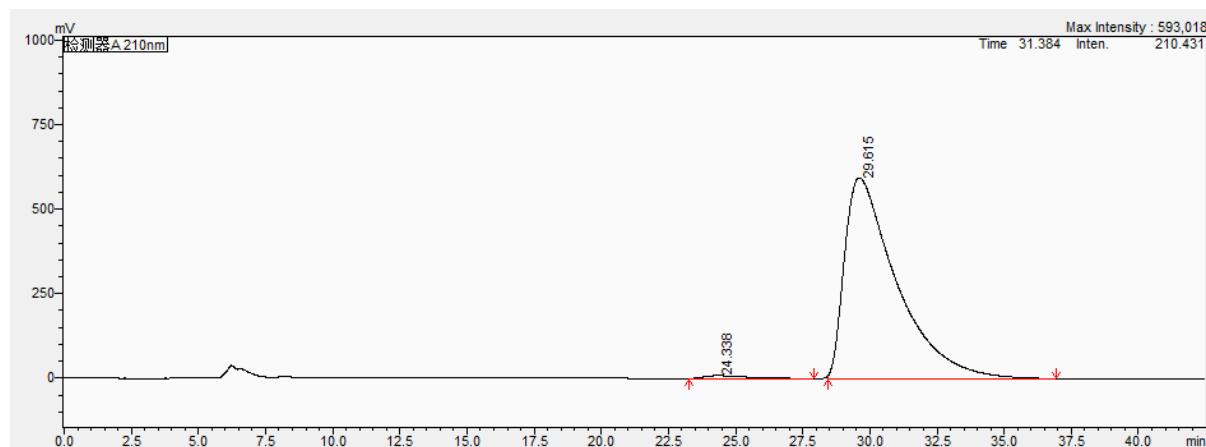
	Retention Time (min)	Area (%)	90% ee
Peak 1	19.408	4.853	
Peak 2	21.154	95.147	



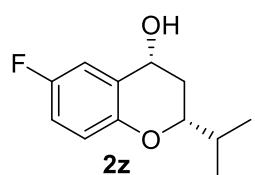
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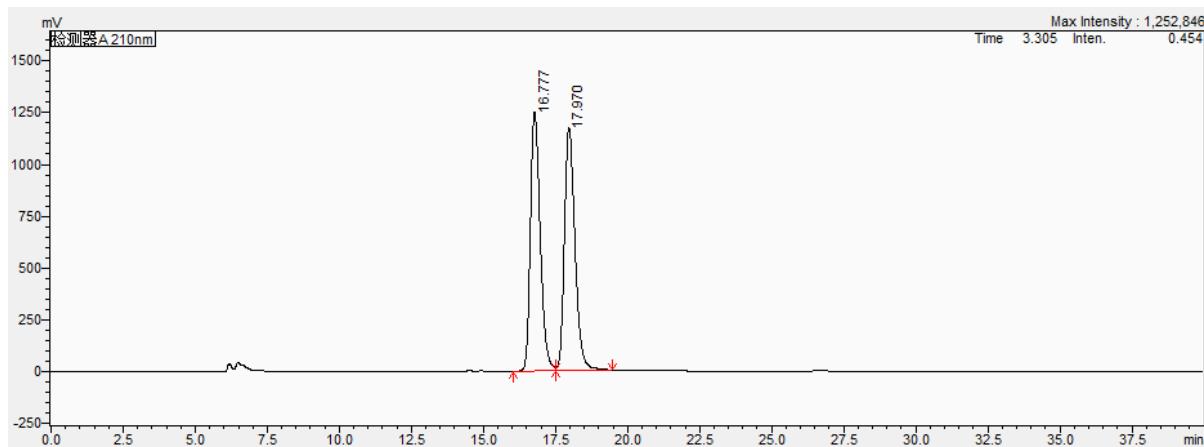
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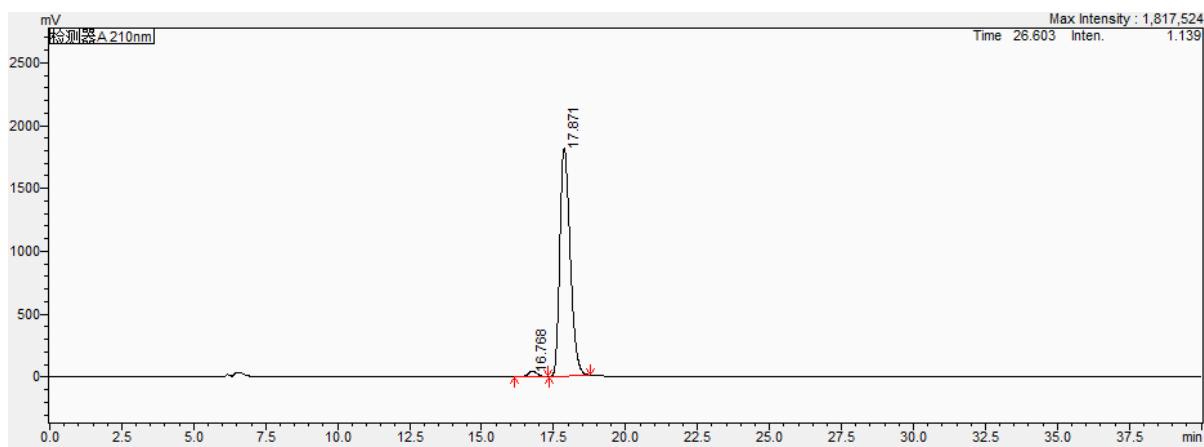
	Retention Time (min)	Area (%)	98% ee
Peak 1	24.338	1.179	
Peak 2	29.615	98.821	



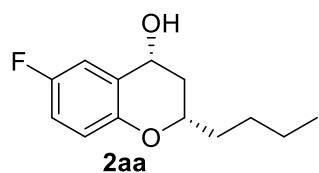
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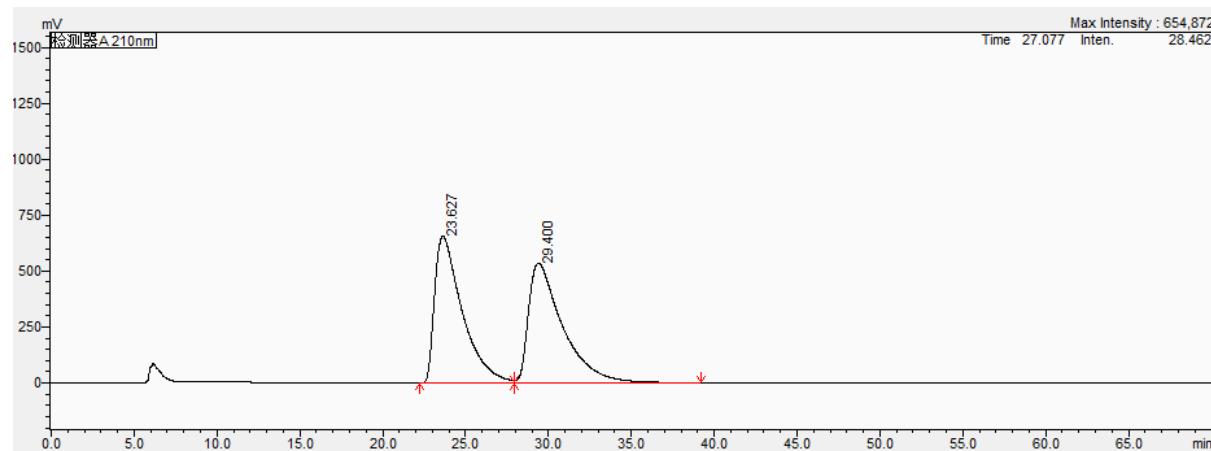
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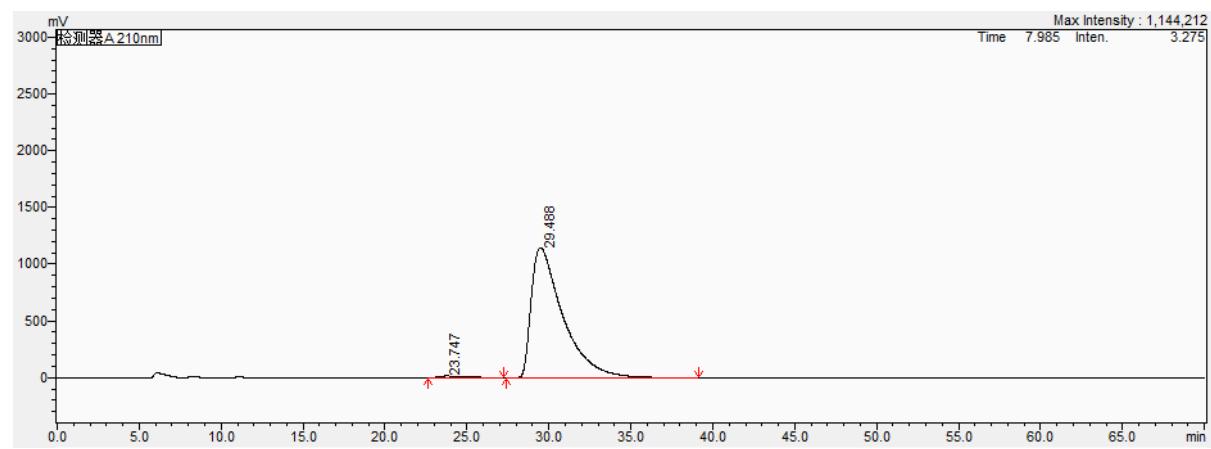
	Retention Time (min)	Area (%)	96% ee
Peak 1	16.768	2.065	
Peak 2	17.871	97.935	



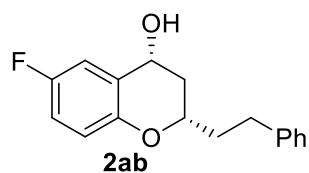
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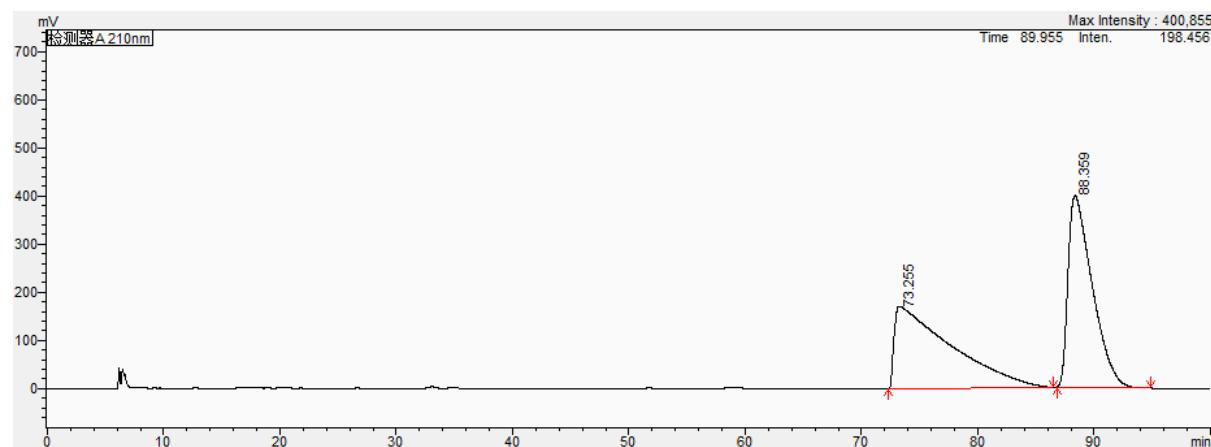
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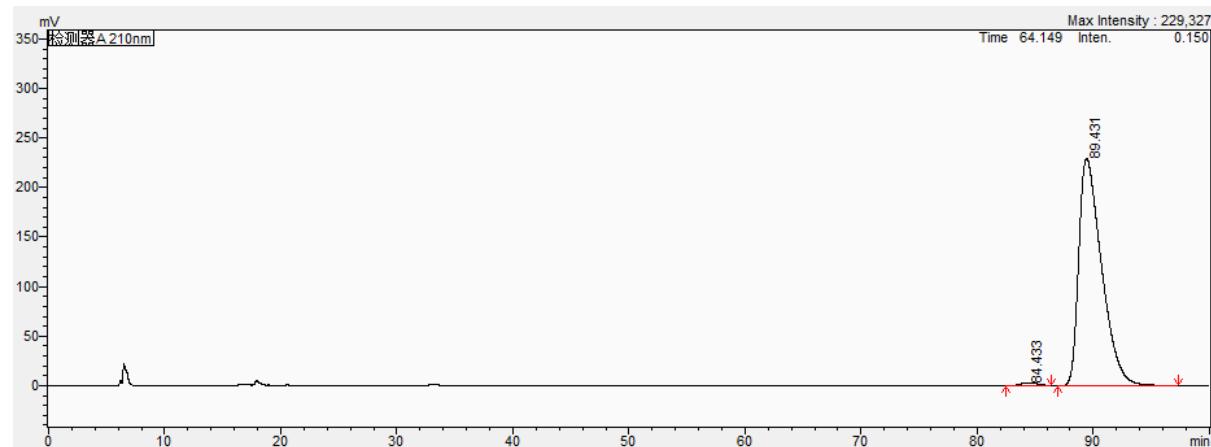
	Retention Time (min)	Area (%)	98% ee
Peak 1	23.747	0.875	
Peak 2	29.488	99.125	



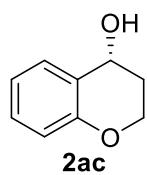
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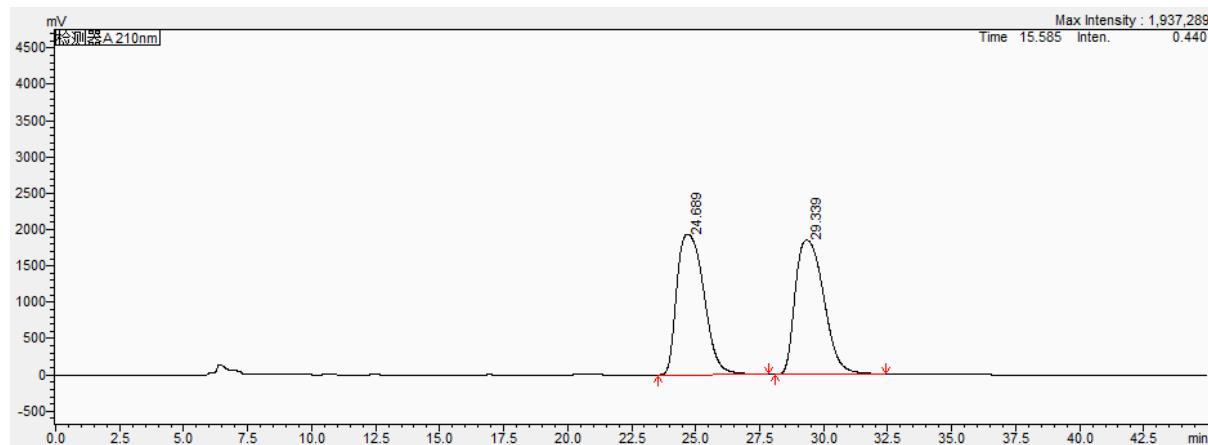
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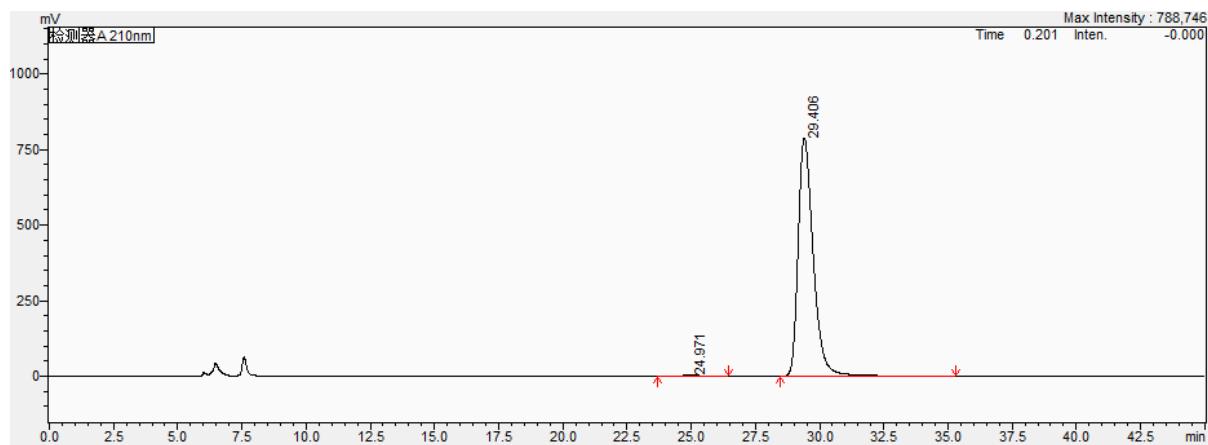
	Retention Time (min)	Area (%)	98% ee
Peak 1	84.433	0.783	
Peak 2	89.431	99.217	



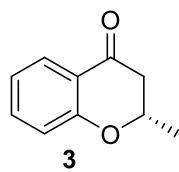
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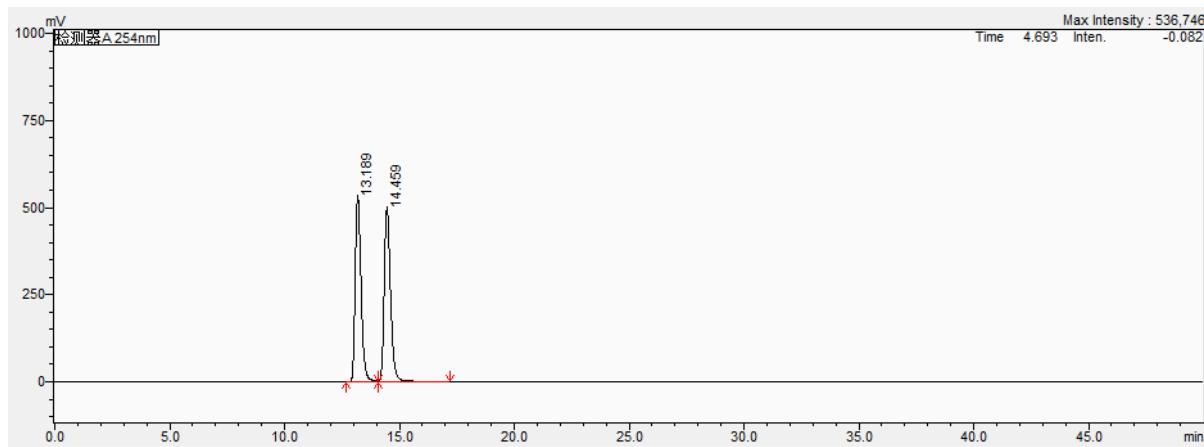
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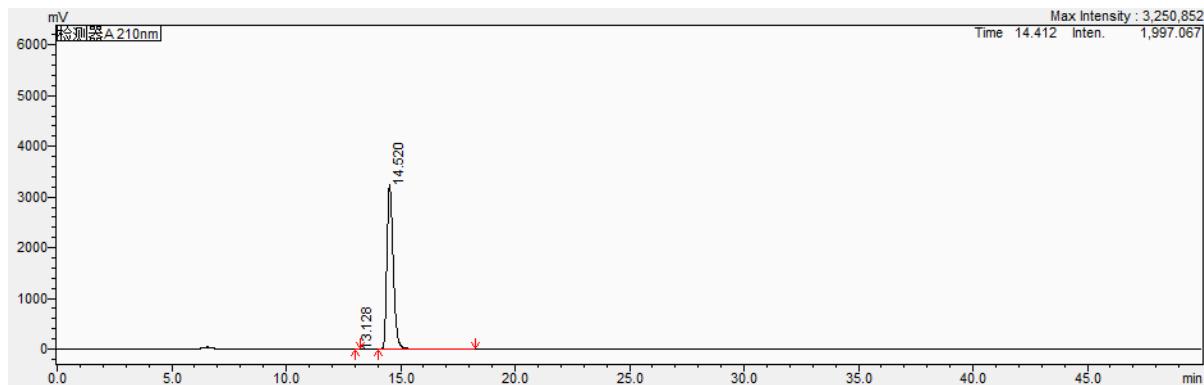
	Retention Time (min)	Area (%)	99% ee
Peak 1	24.971	0.690	
Peak 2	29.406	99.310	



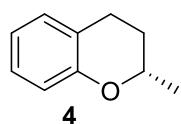
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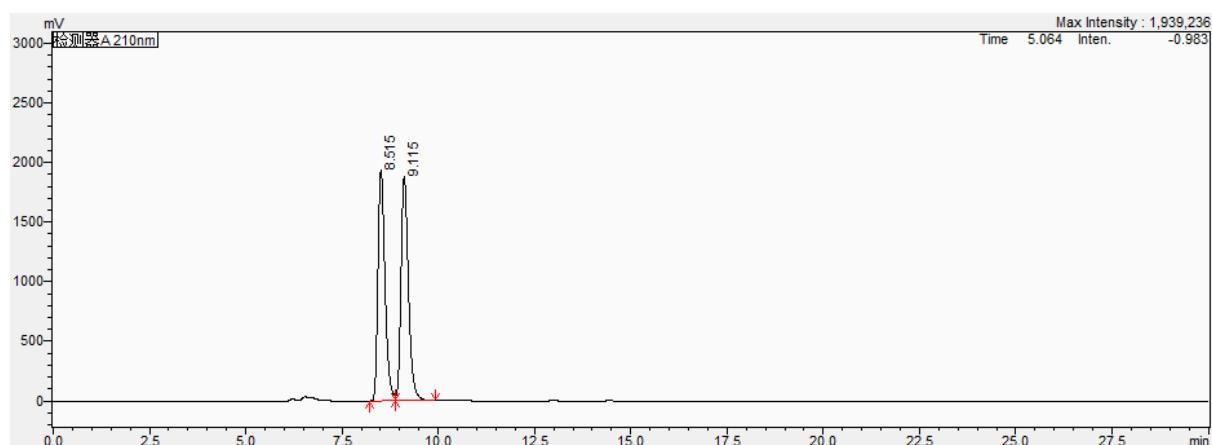
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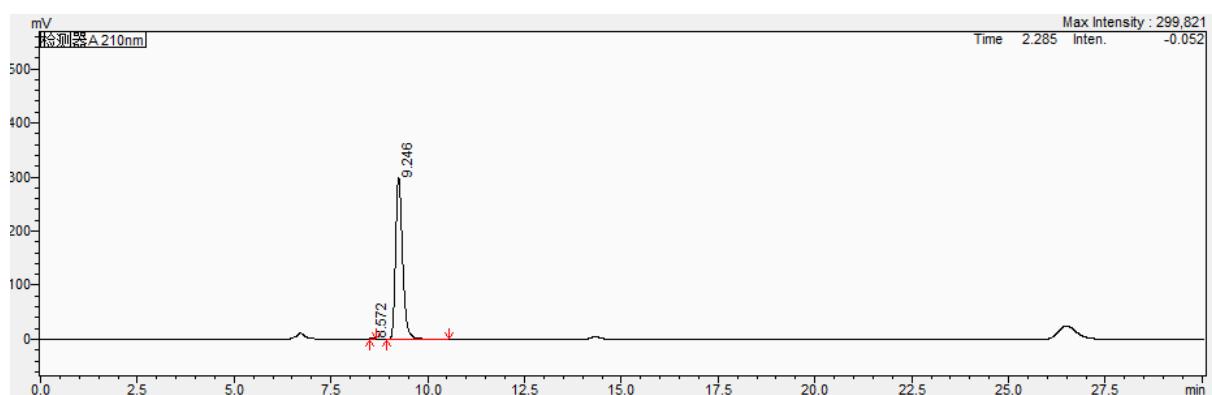
	Retention Time (min)	Area (%)	99.9% ee
Peak 1	13.128	0.065	
Peak 2	14.520	99.935	



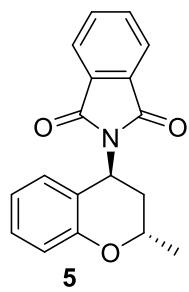
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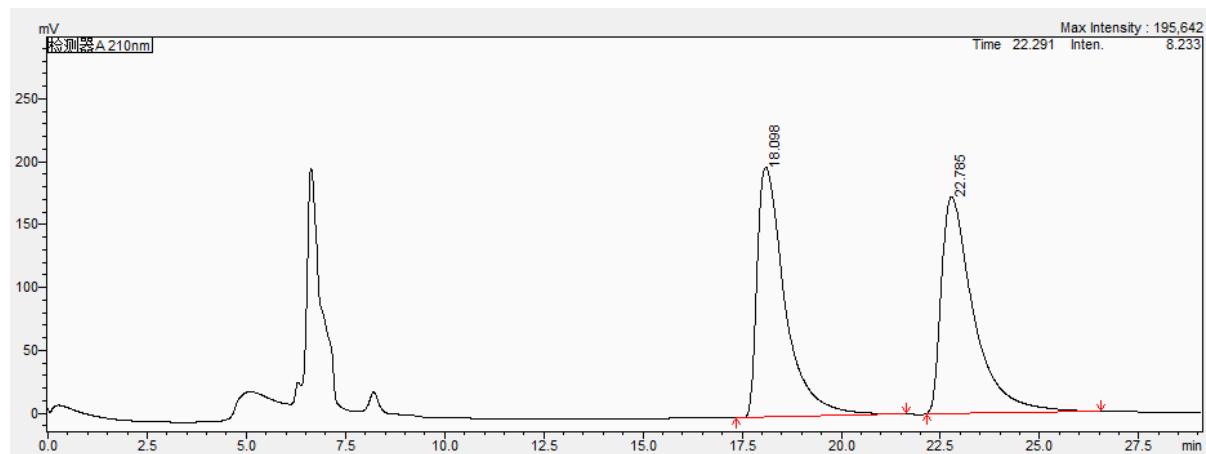
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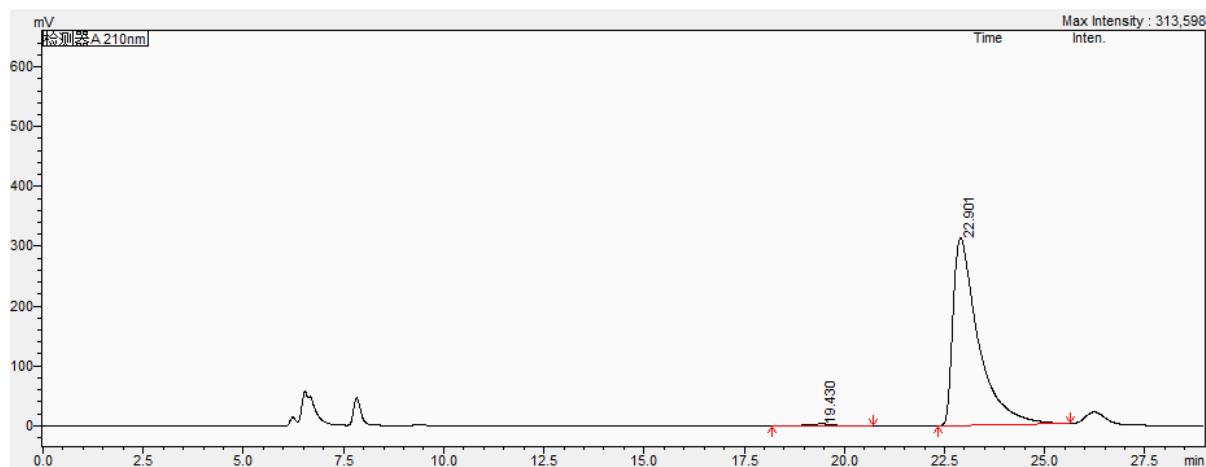
	Retention Time (min)	Area (%)	99.8% ee
Peak 1	8.572	0.121	
Peak 2	9.246	99.879	



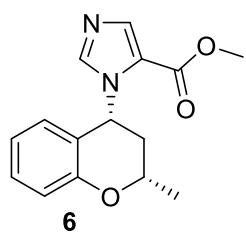
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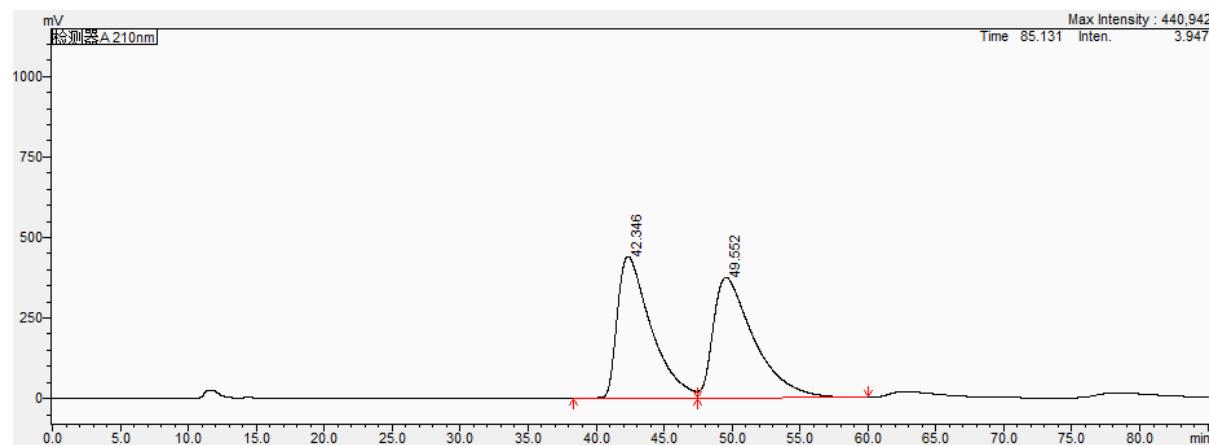
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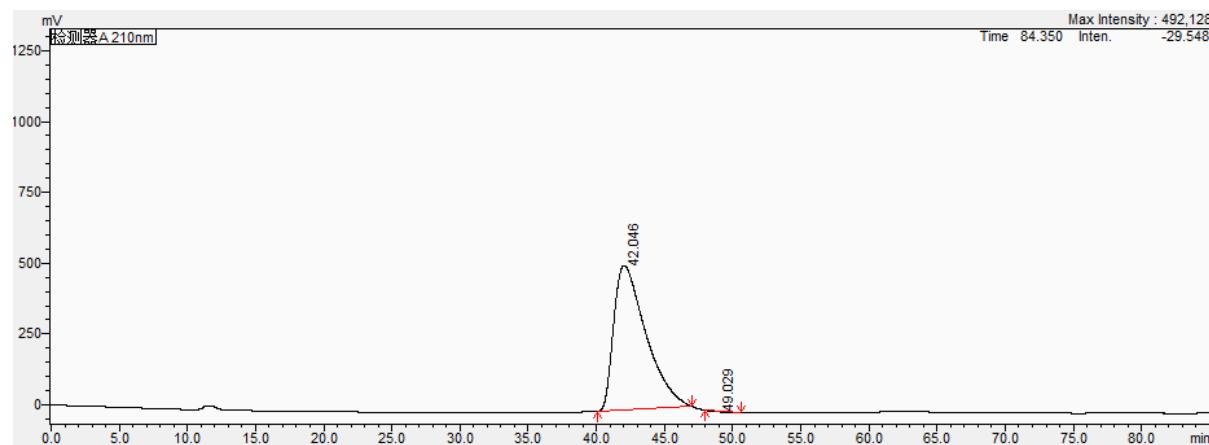
	Retention Time (min)	Area (%)	98% ee
Peak 1	19.430	1.172	
Peak 2	22.901	98.828	



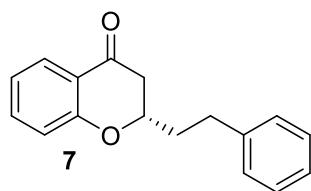
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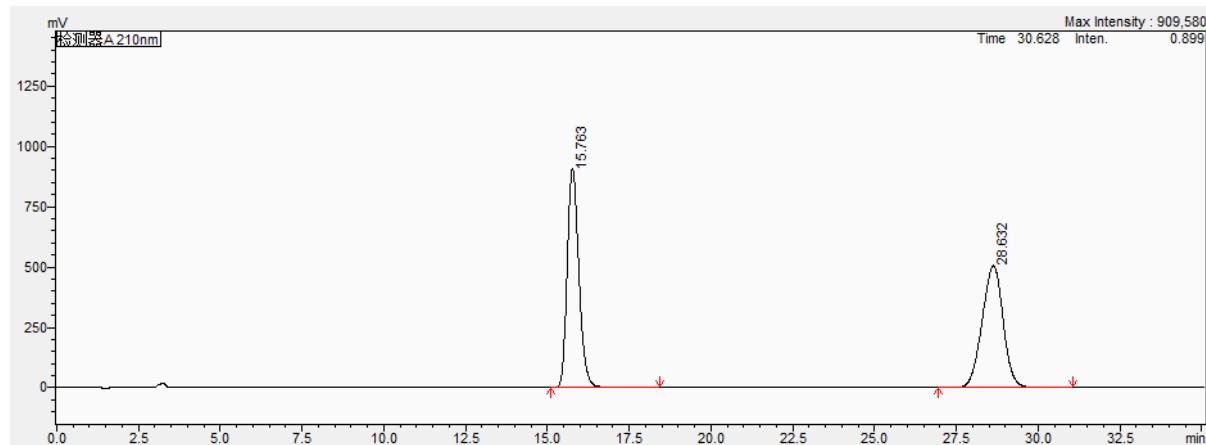
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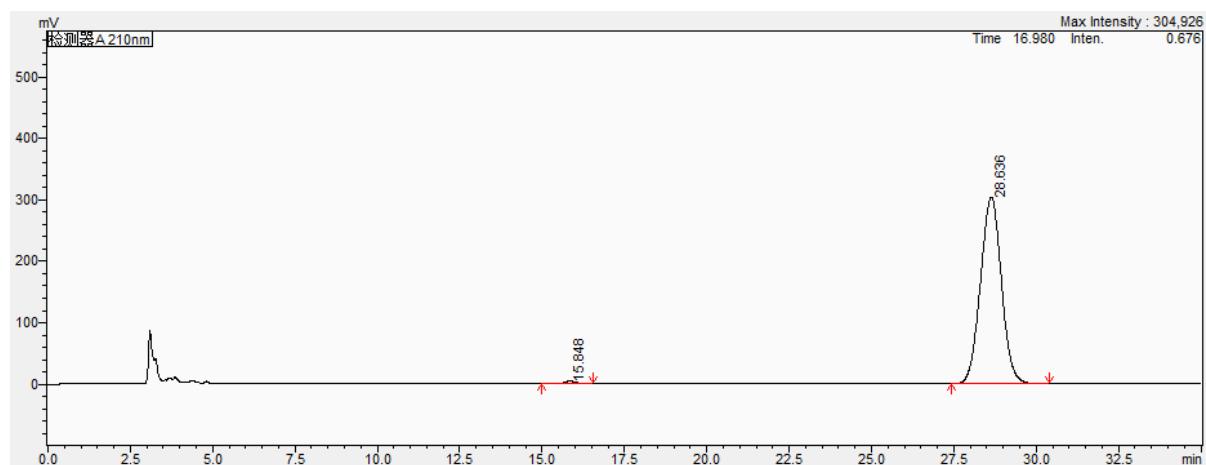
	Retention Time (min)	Area (%)	99.9% ee
Peak 1	42.046	99.972	
Peak 2	49.029	0.028	



Racemate



Chiral



	Retention Time (min)	Area (%)	98% ee
Peak 1	15.848	0.815	
Peak 2	28.636	99.185	