

**Supporting Information**

**Visible-Light Mediated Carbamoyl Radical Addition to Heteroarenes**

Ashique Hussain Jatoi, Govind Goroba Pawar, Frédéric Robert, and Yannick Landais\*

*University of Bordeaux, Institute of Molecular sciences (ISM), UMR-CNRS 5255, 351,*

*Cours de la Libération, 33405 Talence Cedex, France. E-mail : [yannick.landais@u-bordeaux.fr](mailto:yannick.landais@u-bordeaux.fr)*

**Supporting Information**

**Table of contents**

1.	General information -----	S2
2.	General procedure for the preparation of Oxamic Acids -----	S3-S6
3.	Synthesis of 1,2,3,5-tetrakis(carbazol-9-yl)-4,6-dicyanobenzene (4CzIPN) -----	S7
4.	Synthesis of Hypervalent Iodine Reagents (HIR) -----	S8
5.	General procedure for the Visible-Light Mediated Carbamoyl Radical Addition to Heteroarenes-----	S9
6.	$^1\text{H}$ and $^{13}\text{C}$ NMR data -----	S10-S20
7.	DFT calculations-----	S21-S25
8.	HPLC Data -----	S26-S27
9.	Fluorescence quenching experiments-----	S28-S30
10.	Copies of $^1\text{H}$ and $^{13}\text{C}$ spectrum -----	S31-S106

## Supporting Information

### 1. General Information

#### Reagents

All reagent-grade chemicals were obtained from commercial suppliers and were used as received unless otherwise noted. CH<sub>2</sub>Cl<sub>2</sub> and THF were dried over activated alumina columns on MBraun Solvent Purification System (SPS-800). DCE and MeCN were distilled from CaH<sub>2</sub> and anhydrous dimethylformamide and dimethylsulfoxide was purchased from Sigma Aldrich. Triethylamine (reagent grade, ≥98%, Sigma Aldrich) and DMSO (anhydrous, ≥99.9%, Aldrich).

#### Reactions

All reactions for the Visible-Light Photocatalyzed Oxidation of Oxamic Acids were set up on bench-top in the open air and carried out in re-sealable test tubes with Teflon septa under an argon atmosphere. Unless otherwise noted, the reaction test tubes were cooled to room temperature prior to other operations. Unless otherwise noted, the solvents and the solutions of reagents/reactants were transferred *via* micro-syringe or plastic syringe (fitted with metal needle) into the reaction test tubes under a positive argon pressure.

Photochemical reactions were performed with 455 nm (Castorama-blue LEDs ( $\lambda = 455$  nm ( $\pm 15$  nm), 12 V, 500 mA).

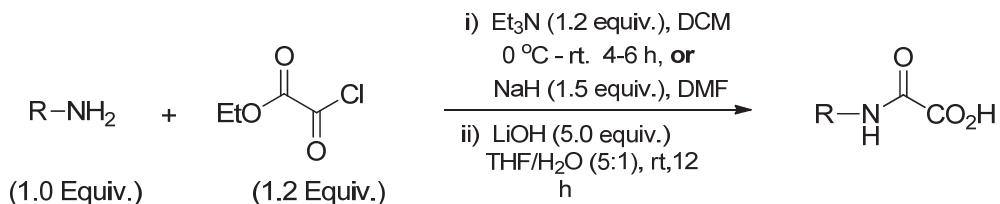
Analytical thin layer chromatography was performed using silica gel 60 F254 pre-coated plates (Merck) with visualization by ultraviolet light, Ceric Ammonium Molybdate and Ninhydrin. Flash chromatography was performed on silica gel (0.043-0.063 mm). Yields refer to chromatographically and spectroscopically (<sup>1</sup>H-NMR) homogeneous materials, unless otherwise stated.

#### Instruments

<sup>1</sup>H-NMR and <sup>13</sup>C-NMR were recorded on various spectrometers: a Brüker DPX 200 (<sup>1</sup>H: 200 MHz, <sup>13</sup>C: 50.25 MHz), a Brüker Avance 300 (<sup>1</sup>H: 300 MHz, <sup>13</sup>C: 75.46 MHz), a using CDCl<sub>3</sub> as internal reference unless otherwise indicated. The chemical shifts ( $\delta$ ) and coupling constants ( $J$ ) are expressed in ppm and Hz respectively. The following abbreviations were used to explain the multiplicities: bs = broad singlet, s = singlet, d = doublet, t = triplet, q = quartet, dd = doublet of doublets, m = multiplets. FTIR spectra were recorded on a Perkin-Elmer Spectrum 100 using a thin film between KBr plates. HRMS were recorded with a Waters Q-TOF 2 spectrometer in the electrospray ionization (ESI) and atmospheric pressure chemical ionization (APCI) mode.

## Supporting Information

### 2. A. General procedure for the preparation of Oxamic Acids



**Scheme 1.** Preparation of oxamic acids

To a solution of the corresponding aniline or amine (10 mmol) in  $\text{CH}_2\text{Cl}_2$  (0.3 M) was added  $\text{Et}_3\text{N}$  (11 mmol), oxalyl chloride (11 mmol) was then added to the solution slowly at  $0$   $^\circ\text{C}$ . The reaction mixture was warmed to room temperature and stirred for 4 - 6 h. The reaction mixture was then treated with 1 M HCl (20 mL) and extracted with dichloromethane ( $3 \times 20$  mL). The combined extracts were washed with brine, dried over  $\text{Na}_2\text{SO}_4$ , filtered, and concentrated in vacuo, directly subjected to hydrolysis.

The residue was dissolved in 15 mL THF and 5 mL  $\text{H}_2\text{O}$ , and LiOH (50 mmol) was added. After stirring for 6 - 8 h at room temperature, the basic reaction mixture was washed with dichloromethane ( $3 \times 30$  mL). The aqueous phase was separated and acidified with 1M aqueous HCl solution. The resulting mixture was extracted with ethyl acetate ( $3 \times 30$  mL) and the combined organic layers were washed with brine (30 mL) and dried over  $\text{Na}_2\text{SO}_4$ . The solvent was evaporated and the residue was recrystallized by  $\text{CH}_2\text{Cl}_2$ /hexanes.

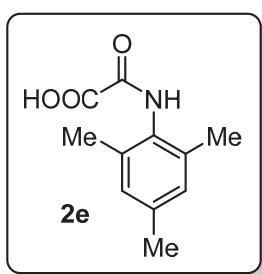
**Table 1.** Previously reported Oxamic acids

Oxamic Acids	References
$\text{H}_3\text{C}-\text{CH}_2-\text{CH}_2-\text{CH}_2-\text{CH}_2-\text{N}-\text{H}-\text{C}(=\text{O})-\text{C}(=\text{O})-\text{OH}$ <b>2a</b>	1. N. S. Vujicic, Z. Glasovac, N. Zweep, J. H. van Esch, M. Vinkovic, J. Popovic, M. Zinic <i>Chem. Eur. J.</i> <b>2013</b> , <i>19</i> , 8558-8572.

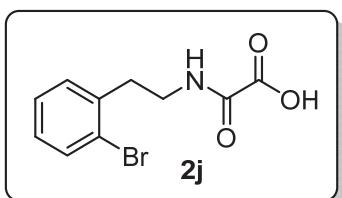
## Supporting Information

<p><b>2c</b></p> <p><b>2k</b></p> <p><b>2l</b></p> <p><b>2b</b></p>	<p>2. S. D. Linton et. al. <i>J. Med. Chem.</i> <b>2005</b>, <i>48</i>, 6779-6782.</p>
<p><b>2d</b></p>	<p>3. J. Bálint, G. Egri, M. Czugler, J. Schindler, V. Kiss, Z. Juvancz, E. Fogassy, <i>Tetrahedron: Asymmetry</i> <b>2001</b>, <i>12</i>, 1511–1518.</p>

**2-(Mesitylamino)-2-oxoacetic acid:** **2e** (1.2 g) was obtained through the general procedure A in 58 % yield as a white solid; mp 105-108 °C; **1H NMR** (**300 MHz, CDCl<sub>3</sub>**) δ (ppm) 8.49 (s, 1H), 6.94 (s, 2H), 2.29 (s, 3H), 2.19 (s, 6H). **13C NMR** (**75 MHz, CDCl<sub>3</sub>**) δ (ppm) 160.0, 138.5, 134.6, 129.4, 128.9, 21.1, 18.4. **IR (neat)**  $\nu_{\max}$  (cm<sup>-1</sup>) = 3250, 2922, 1883, 1668. **HRMS (ESI):** Calcd. For C<sub>11</sub>H<sub>11</sub>NO<sub>3</sub> [M-H]<sup>+</sup> 206.0817, found 206.0821.

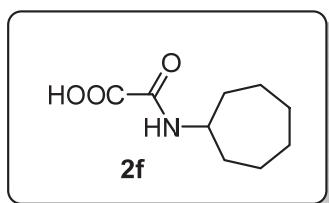


**2-((2-Bromophenethyl)amino)-2-oxoacetic acid:** **2j** (1.4 g) was obtained through the general procedure A in 52 % yield as a white solid; mp 137-140 °C; **1H NMR** (**300 MHz, CDCl<sub>3</sub>**) δ (ppm) 7.59 (dd, *J* = 8.0, 1.3 Hz, 1H), 7.45 (s, 1H), 7.35 – 7.20 (m, 2H), 7.19 – 7.10 (m, 1H), 3.68 (q, *J* = 7.2 Hz, 1H), 3.07 (t, *J* = 7.2 Hz, 1H). **13C NMR** (**75 MHz, CDCl<sub>3</sub>**) δ (ppm) 159.6, 157.5, 137.0, 133.2, 130.8, 128.8, 127.9, 124.5, 40.2, 35.3. **IR (neat)**  $\nu_{\max}$  (cm<sup>-1</sup>) = 3282, 2938, 1769, 17672. **HRMS (ESI):** Calcd. For C<sub>10</sub>H<sub>9</sub>NO<sub>3</sub>Br [M-H]<sup>+</sup> 269.9771, found 269.9770.



## Supporting Information

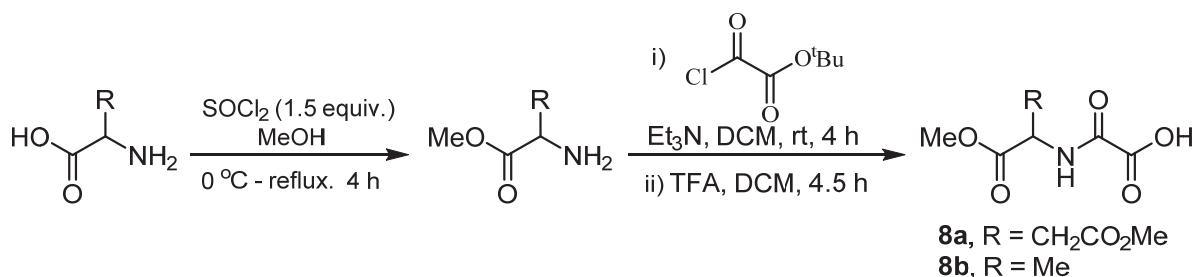
**2-(Cycloheptylamino)-2-oxoacetic acid: 2f** (1.0 g) was obtained through the general procedure A in 54



% yield as a white solid; mp 158–63 °C; **1H NMR** (300 MHz, CDCl<sub>3</sub>) δ (ppm) 7.26 (s, 1H), 4.07–3.67 (m, 1H), 2.08 – 1.86 (m, 2H), 1.81 – 1.41 (m, 10H). **13C NMR** (75 MHz, CDCl<sub>3</sub>) δ (ppm) 160.2, 156.3, 52.3, 34.6, 29.0, 24.0. **IR (neat) v<sub>max</sub> (cm<sup>-1</sup>)** = 3293, 2923, 1767, 1674. **HRMS (ESI)**: Calcd.

For C<sub>9</sub>H<sub>14</sub>NO<sub>3</sub> [M-H]<sup>+</sup>, 184.0979, found 184.0982.

## 2. B. General procedure for the preparation of Oxamic Acids<sup>1</sup>

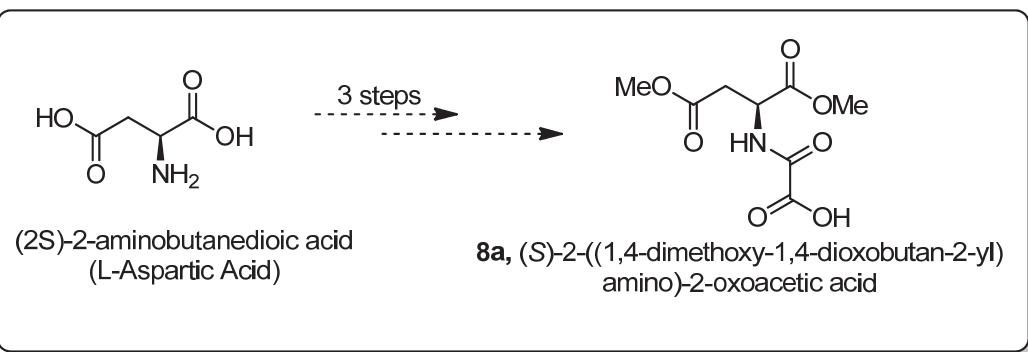


**Scheme 2.** Preparation of Oxamic acids

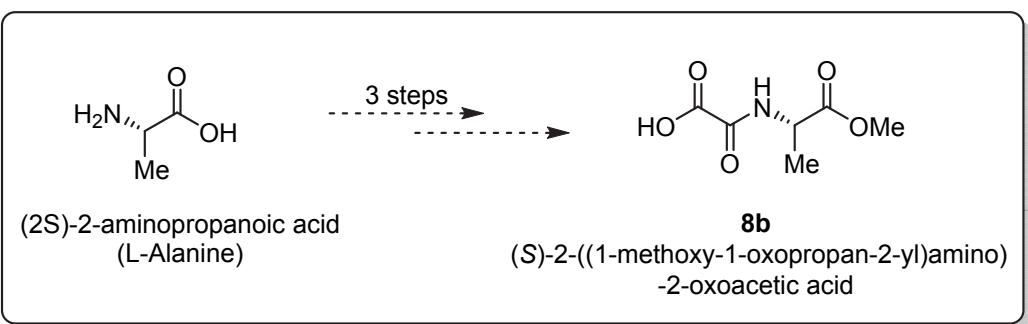
To a solution of the corresponding amino acid (30.07 mmol) in MeOH (25 mL) at 0 °C under nitrogen, thionyl chloride (3.2 mL, 45.11 mmol) was added dropwise over 15 mins. The reaction mixture was warmed to room temperature and then refluxed for 4 h. The solution was concentrated in vacuo to afford colorless oil. Hexane was added to this crude oil and was stirred for 10 min. Hexane was decanted and this procedure was repeated twice to obtain a solid compound.<sup>4a</sup> This solid compound was dissolved in DCM (60 mL) and *tert*-butyl 2-chloro-2-oxoacetate was added dropwise at 0 °C under nitrogen. The reaction mixture was stirred at room temperature for 4 h. The resulting mixture was washed with water (100 mL), and brine (100 mL) then dried over sodium sulfate, concentrated under reduced pressure to obtain a crude solid compound.<sup>4b</sup> The crude product was then treated with TFA in dichloromethane for 4 h at room temperature<sup>4b</sup> and then mixture was concentrated under reduced pressure to give the desired oxamic acid product as a colorless oil.

<sup>1</sup> (a) C. R. Reddy; M. D. Reddy; U. Dilipkumar. *Eur. J. Org. Chem.* **2014**, 6310–6313. (b) Y. Seki, K. Tanabe, D. Sasaki, Y. Sohma, K. Oisaki, M. Kanai, *Angew. Chem. Int. Ed.* **2014**, 53, 6501–6505.

## Supporting Information



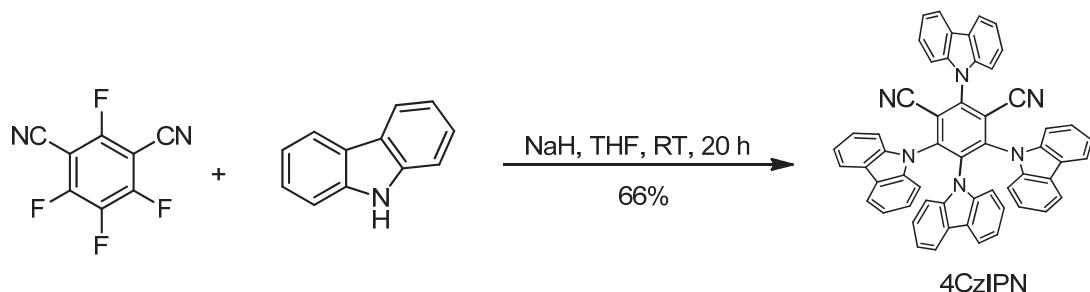
**(S)-2-((1,4-Ddimethoxy-1,4-dioxobutan-2-yl)amino)-2-oxoacetic acid: 8a** (4.5 g) was obtained through the general procedure B in 64 % yield as a colorless oil; **<sup>1</sup>H NMR** (300 MHz, CDCl<sub>3</sub>) δ (ppm) 8.48 (s, 1H), 8.24 (d, *J* = 8.4 Hz, 1H), 4.94 – 4.78 (m, 1H), 3.80 (s, 3H), 3.72 (s, 3H), 3.11 (dd, *J* = 17.4, 5.0 Hz, 1H), 2.92 (dd, *J* = 17.4, 4.5 Hz, 1H). **<sup>13</sup>C NMR** (76 MHz, CDCl<sub>3</sub>) δ (ppm) 171.0, 169.7, 159.5, 157.5, 53.4, 52.5, 49.5, 35.6. **IR (neat)  $\nu_{\max}$  (cm<sup>-1</sup>)** = 3355, 2959, 1739, 1696. **HRMS (ESI)**: Calcd. For C<sub>8</sub>H<sub>10</sub>NO<sub>7</sub> [M-H]<sup>+</sup> 232.0457, found 232.0462. [α]<sub>D</sub><sup>25</sup> +80.76 (c 1.8, CHCl<sub>3</sub>).



**(S)-2-((1-Methoxy-1-oxopropan-2-yl) amino)-2-oxoacetic acid: 8b** (3.1 g) was obtained through the general procedure B in 59 % yield as a colorless oil. **<sup>1</sup>H NMR** (300 MHz, CDCl<sub>3</sub>) δ (ppm) 9.69 (s, 1H), 8.06 (d, *J* = 7.6 Hz, 1H), 4.57 (p, *J* = 7.2 Hz, 1H), 3.75 (s, 3H), 1.47 (d, *J* = 7.2 Hz, 3H). **<sup>13</sup>C NMR** (75 MHz, CDCl<sub>3</sub>) δ (ppm) 172.0, 160.1, 157.5, 52.9, 49.0, 17.5. **IR (neat)  $\nu_{\max}$  (cm<sup>-1</sup>)** = 3290, 2958, 1739, 1692. **HRMS (ESI)**: Calcd. For C<sub>6</sub>H<sub>7</sub>NO<sub>5</sub> [M-H]<sup>+</sup> 174.0402, found 174.0406. [α]<sub>D</sub><sup>25</sup> +9.32 (c 4.0, CHCl<sub>3</sub>).

## Supporting Information

### 3. Synthesis of 1,2,3,5-tetrakis(carbazol-9-yl)-4,6-dicyanobenzene (4CzIPN):



**Scheme 3.** Synthesis of 4CzIPN

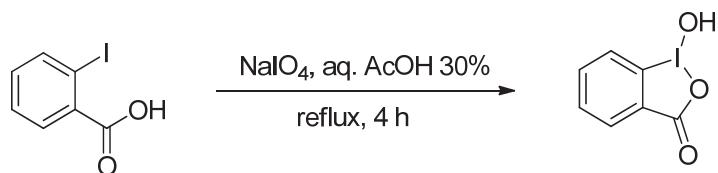
The 4CzIPN was synthesized according to the following reported procedure.<sup>5</sup> NaH (60% in oil, 1.4 g, 60 mmol) was added slowly to a stirred solution of carbazole (4.18 g, 25.0 mmol) in dry THF (100 mL) under a nitrogen atmosphere at room temperature. After 30 min, tetrafluoroisophthalonitrile (1.0 g, 5.0 mmol), was added. After stirring at room temperature for 12 h, 4-5 mL water was added to the reaction mixture to quench the excess NaH. The resulting mixture was then concentrated under reduced pressure and successively washed by water and EtOH to yield the crude yellow solid. The crude product was dissolved in a minimum quantity of CH<sub>2</sub>Cl<sub>2</sub> and crystallized by addition of pentane to give the pure 4CzIPN (2.21 g, 66%) as a yellow solid; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>) δ (ppm) 8.25 (dt, *J* = 7.8, 1.0 Hz, 2H), 7.80 – 7.66 (m, 8H), 7.57 – 7.47 (m, 2H), 7.36 (d, *J* = 7.6 Hz, 1H), 7.32 – 7.21 (m, 5H), 7.19 – 7.05 (m, 8H), 6.91 – 6.79 (m, 4H), 6.73 – 6.60 (m, 2H). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ (ppm) 145.2, 144.6, 140.0, 138.2, 137.0, 134.8, 127.0, 125.8, 125.0, 124.8, 124.5, 123.9, 122.4, 121.9, 121.4, 121.0, 120.4, 119.7, 116.4, 111.6, 110.0, 109.5, 109.4. Spectroscopic data were in good agreement with literature.<sup>2</sup>

<sup>2</sup> Uoyama, H.; Goushi, K.; Shizu, K.; Nomura, H.; Adachi, C. *Nature* **2012**, *492*, 234-238.

## Supporting Information

### 4. Hypervalent Iodine Reagents (HIR)

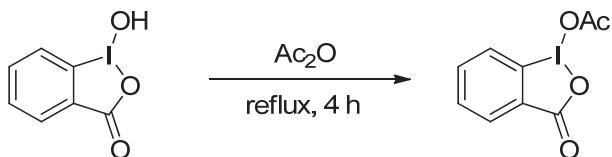
#### (a) 1-Hydroxy-1,2-benziodoxol-3(1H)-one (CAS: 131-62-4):



**Scheme 4.** Preparation of BI-OH

Following a reported procedure,<sup>3</sup> NaIO<sub>4</sub> (6.7 g, 31.0 mmol, 1.00 equiv) and 2-iodobenzoic acid (7.4 g, 30.0 mmol, 1.00 equiv.) were suspended in 30% (v:v) aqueous AcOH (45 mL) under air. The mixture was vigorously stirred and refluxed for 4 h, protected from light. Cold water (120 mL) was added and the reaction mixture was allowed to cool to room temperature. After 1 h, the crude product was collected by filtration, washed with ice water (3 x 30 mL) and cold acetone (3 x 30 mL). After air dried in the dark overnight to give the pure compound **BI-OH** (6.8 g, 86%) as a white solid; **1H NMR (300 MHz, DMSO-d<sub>6</sub>)** δ (ppm) 8.09 – 7.91 (m, 3H), 7.85 (dd, *J* = 8.2, 1.1 Hz, 1H), 7.70 (td, *J* = 7.3, 1.1 Hz, 1H). **13C NMR (75 MHz, DMSO-d<sub>6</sub>)** δ (ppm) 168.2, 134.9, 132.0, 131.6, 130.8, 126.7, 120.9. Spectroscopic data were in good agreement with literature.

#### (b) 1-Acetoxy-1,2-benziodoxol-3-(1H)-one (CAS:1829-25-0):



**Scheme 4.** Preparation of BI-OAc

Following a reported procedure,<sup>4</sup> BI-OH (6.00 g, 22.7 mmol) was heated in Ac<sub>2</sub>O (20 mL) to reflux until the solution turned clear (without suspension). The mixture was then left to cool down and white crystals started to form. The crystallization was continued at -20 °C. Crystals were then collected and dried overnight under high vacuum to give compound **BI-OAc** (6.1 g, 88%) as a white solid; **1H NMR (300 MHz, CDCl<sub>3</sub>)** δ (ppm) 8.28 (dd, *J* = 7.6, 1.6 Hz, 1H), 8.07 – 8.01 (m, 1H), 8.00 – 7.92 (m, 1H), 7.79 – 7.71 (m, 1H), 2.29 (s, 3H). **13C NMR (75 MHz, CDCl<sub>3</sub>)** δ (ppm) 176.4, 168.2, 136.1, 133.2, 131.3, 129.3, 129.0, 118.3, 20.3. Spectroscopic data were in good agreement with literature.

<sup>3</sup> Fernández González, D.; Brand, J. P.; Waser, J. *Chem. Eur. J.* **2010**, *16*, 9457-9461.

<sup>4</sup> Eisenberger, P.; Gischig, S.; Togni, A. *Chem. Eur. J.* **2006**, *12*, 2579-2586.

## Supporting Information

### 5. General procedure for the Visible-Light Mediated Carbamoyl Radical Addition to Heteroarenes

#### Reaction Setup:

Photochemical reactions were performed with 455 nm (Castorama-blue LEDs ( $\lambda = 455$  nm ( $\pm 15$  nm)), 12 V, 500 mA).

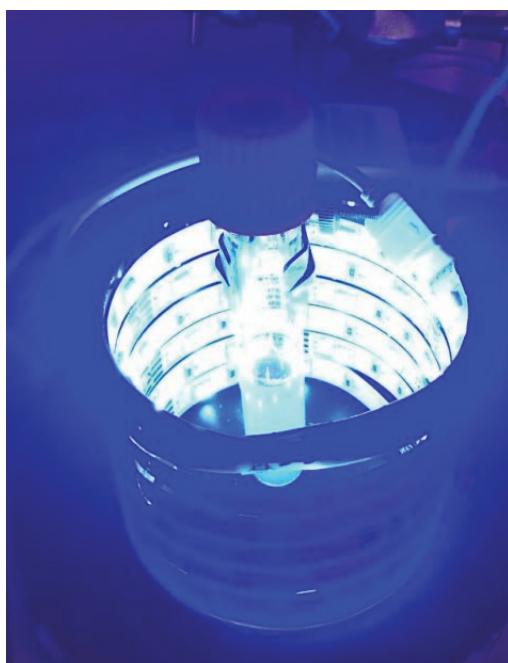
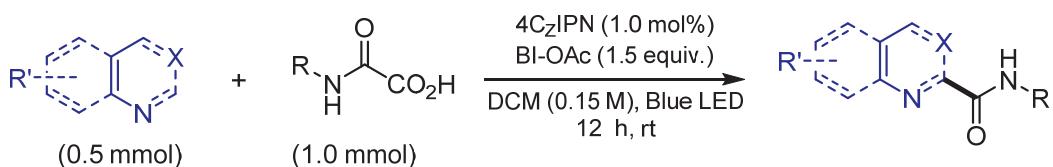


Figure 1. Reaction Setup



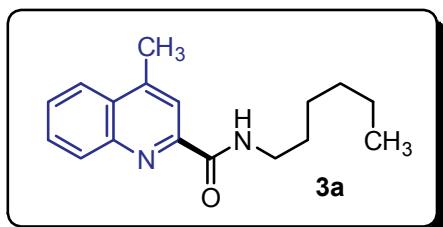
Scheme 5. Carbamoyl addition to Heteroarenes

The *N*-Heterocycle (1.0 equiv., 0.5 mmol), Oxamic Acid (1.5 equiv., 0.75 mmol), 4CzIPN (1.0 mol%, 0.005 mmol), and BIOAc (1.5 equiv., 0.75 mmol) were placed into a re-sealable test-tube with Teflon septa (10 mL) and a magnetic stir bar. Air was removed from the reaction vessel, which was then backfilled with argon three times, and DCM (0.2 M) was added afterwards (Note: for liquid substrates, they were added after the tube was backfilled with argon). The reaction mixture was stirred at room temperature under blue LED irradiation for 12 h. The reaction mixture was concentrated and purified directly by column chromatography to afford the product. (Eluting with ethyl acetate/hexanes).

## Supporting Information

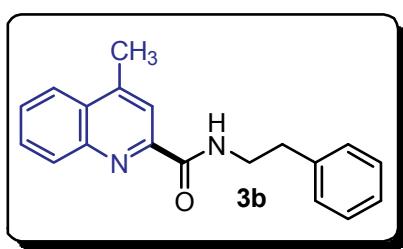
### 6. $^1\text{H}$ and $^{13}\text{C}$ NMR Data

**N-Hexyl-4-methylquinoline-3-carboxamide: 3a** (122 mg) was obtained through the general procedure



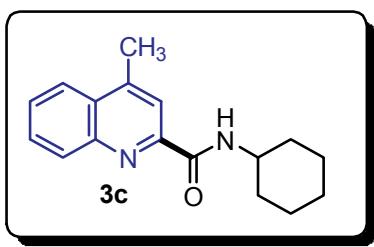
in 90 % yield as a white solid, m.p. 60–63 °C.  $R_f = 0.84$  (EtOAc–Hexane 10/90).  **$^1\text{H NMR}$  (300 MHz,  $\text{CDCl}_3$ )**  $\delta$  (ppm) 8.29 (s, 1H), 8.13 (d,  $J = 0.8$  Hz, 1H), 8.10–8.02 (m, 1H), 7.98 (dd,  $J = 8.4, 1.0$  Hz, 1H), 7.75–7.65 (m, 1H), 7.62–7.51 (m, 1H), 3.59–3.41 (m, 2H), 2.71 (d,  $J = 1.0$  Hz, 3H), 1.72–1.56 (m, 2H), 1.48–1.22 (m, 6H), 0.96–0.77 (m, 3H).  **$^{13}\text{C NMR}$  (75 MHz,  $\text{CDCl}_3$ )**  $\delta$  (ppm) 164.7, 149.6, 146.4, 146.0, 130.3, 129.6, 129.2, 127.5, 123.9, 119.5, 39.6, 31.6, 29.7, 26.8, 22.6, 18.9, 14.1. **IR (neat)  $\nu_{\max}$  (cm $^{-1}$ )** = 3373, 2928, 1672, 1528. **HRMS (ESI)**: Calcd. For  $\text{C}_{17}\text{H}_{22}\text{N}_2\text{O}$ ,  $[\text{M}+\text{H}]^+$  271.1804, found 271.1809.

**4-Methyl-N-phenethylquinoline-2-carboxamide: 3b** (121 mg) was obtained through the general



procedure in 83 % yield as a yellow gel.  $R_f = 0.62$  (EtOAc–Hexane 15/85).  **$^1\text{H NMR}$  (300 MHz,  $\text{CDCl}_3$ )**  $\delta$  (ppm) 8.42 (s, 1H), 8.19–8.12 (m, 1H), 8.09–7.95 (m, 3H), 7.73 (ddd,  $J = 8.4, 6.8, 1.4$  Hz, 1H), 7.61 (ddd,  $J = 8.2, 6.8, 1.2$  Hz, 1H), 7.42–7.26 (m, 5H), 3.88–3.73 (m, 2H), 3.01 (t,  $J = 7.3$  Hz, 2H), 2.75 (s, 3H).  **$^{13}\text{C NMR}$  (75 MHz,  $\text{CDCl}_3$ )**  $\delta$  (ppm) 164.7, 149.4, 146.3, 146.0, 139.1, 130.3, 129.7, 129.2, 128.9, 128.6, 127.6, 126.5, 123.9, 119.4, 40.9, 36.1, 18.9. **IR (neat)  $\nu_{\max}$  (cm $^{-1}$ )** = 3383, 2928, 1672, 1526. **HRMS (APCI)**: Calcd. For  $\text{C}_{19}\text{H}_{18}\text{N}_2\text{O}$ ,  $[\text{M}+\text{H}]^+$  291.1491, found 291.1493.

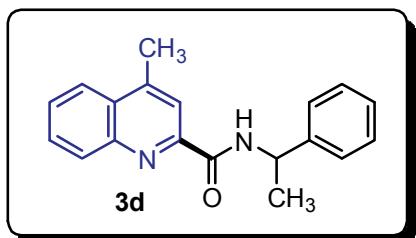
**N-Cyclohexyl-4-methylquinoline-2-carboxamide: 3c** (131 mg) was obtained through the general



procedure in 86 % yield as a yellow gel.  $R_f = 0.72$  (EtOAc–Hexane 15/85).  **$^1\text{H NMR}$  (300 MHz,  $\text{CDCl}_3$ )**  $\delta$  (ppm) 8.13 (d,  $J = 0.8$  Hz, 1H), 8.11–8.05 (m, 1H), 8.00–7.94 (m, 1H), 7.74–7.66 (m, 1H), 7.61–7.52 (m, 1H), 4.10–3.91 (m, 1H), 2.71 (d,  $J = 0.8$  Hz, 3H), 2.12–1.96 (m, 2H), 1.86–1.71 (m, 2H), 1.72–1.57 (m, 1H), 1.51–1.13 (m, 5H).  **$^{13}\text{C NMR}$  (75 MHz,  $\text{CDCl}_3$ )**  $\delta$  (ppm) 163.7, 149.7, 146.3, 146.0, 130.2, 129.6, 129.2, 127.5, 123.9, 119.5, 48.4, 33.2, 25.7, 25.0, 18.9. **IR (neat)  $\nu_{\max}$  (cm $^{-1}$ )** = 3378, 2930, 1669, 1550. **HRMS (APCI)**: Calcd. For  $\text{C}_{17}\text{H}_{20}\text{N}_2\text{O}$ ,  $[\text{M}+\text{H}]^+$  269.1648, found 269.1641.

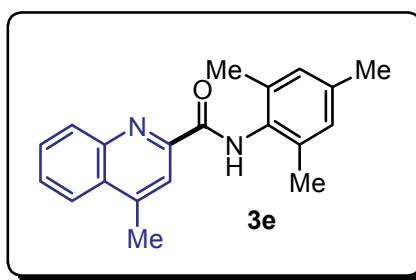
## Supporting Information

**4-Methyl-N-(1-phenylethyl) quinoline-2-carboxamide: 3d** (120 mg) was obtained through the general



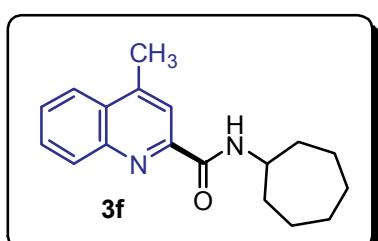
procedure in 83 % yield as a yellow gel.  $R_f = 0.86$  (EtOAc-Hexane 10/90).  **$^1\text{H NMR}$  (300 MHz,  $\text{CDCl}_3$ )**  $\delta$  (ppm) 8.62 (d,  $J = 8.2$  Hz, 1H), 8.24 – 8.17 (m, 1H), 8.17 – 8.09 (m, 1H), 8.07 – 7.97 (m, 1H), 7.79 – 7.69 (m, 1H), 7.67 – 7.57 (m, 1H), 7.54 – 7.45 (m, 2H), 7.44 – 7.36 (m, 2H), 7.34 – 7.30 (m, 1H), 5.44 (p,  $J = 7.0$  Hz, 1H), 2.75 (s, 3H), 1.71 (d,  $J = 6.8$  Hz, 2H).  **$^{13}\text{C NMR}$  (75 MHz,  $\text{CDCl}_3$ )**  $\delta$  (ppm) 163.9, 149.3, 146.3, 146.1, 143.4, 130.3, 129.7, 129.2, 128.7, 127.6, 127.3, 126.3, 123.9, 119.5, 48.9, 22.1, 18.9. **IR (neat)  $\nu_{\max}$  (cm<sup>-1</sup>)** = 3380, 2975, 1670, 1550. **HRMS (APCI)**: Calcd. For  $\text{C}_{19}\text{H}_{18}\text{N}_2\text{O}$ ,  $[\text{M}+\text{H}]^+$  291.1491, found, 291.1503.

**N-Mesitylquinoline-2-carboxamide: 3e** (65 mg) was obtained through the general procedure in 43 %



yield as a brown solid; m.p., 198–201 °C.  $R_f = 0.77$  (EtOAc-Hexane 10/90).  **$^1\text{H NMR}$  (300 MHz,  $\text{CDCl}_3$ )**  $\delta$  (ppm) 9.66 (s, 1H), 8.25 (d,  $J = 0.7$  Hz, 1H), 8.17 (s,  $J = 8.4$ , 0.7 Hz, 1H), 8.08 (m,  $J = 8.4$ , 0.9 Hz, 1H), 7.82 – 7.75 (m, 1H), 7.70 – 7.63 (m, 1H), 6.97 (s, 2H), 2.81 (d,  $J = 0.8$  Hz, 3H), 2.31 (d,  $J = 6.9$  Hz, 9H).  **$^{13}\text{C NMR}$  (75 MHz,  $\text{CDCl}_3$ )**  $\delta$  (ppm) 163.1, 149.4, 146.5, 136.9, 135.3, 131.4, 130.5, 129.9, 129.5, 129.1, 127.9, 124.1, 119.8, 21.1, 19.0, 18.7. **IR (neat)  $\nu_{\max}$  (cm<sup>-1</sup>)** = 3348, 2920, 1686, 1559. **HRMS (ESI)**: Calcd. For  $\text{C}_{20}\text{H}_{20}\text{N}_2\text{O}$   $[\text{M}+\text{Na}]^+$ , 327.1473, found 327.1469.

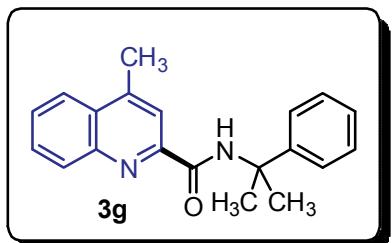
**N-Cycloheptyl-4-methylquinoline-2-carboxamide: 3f** (120 mg) was obtained through the general



procedure in 85 % yield as a white solid; m.p. 102–105 °C.  $R_f = 0.88$  (EtOAc-Hexane 20/80).  **$^1\text{H NMR}$  (300 MHz,  $\text{CDCl}_3$ )**  $\delta$  (ppm) 8.23 (d,  $J = 8.4$  Hz, 1H), 8.14 – 8.03 (m, 2H), 7.95 (dd,  $J = 1.0$ , 8.2 Hz, 1H), 7.68 (dt,  $J = 8.4$ , 6.8, 1.4 Hz, 1H), 7.55 (dt,  $J = 8.2$ , 6.8, 1.4 Hz, 1H), 4.29 – 4.04 (m, 1H), 2.70 (d,  $J = 1.0$  Hz, 3H), 2.14 – 1.93 (m, 2H), 1.80 – 1.40 (m, 10H).  **$^{13}\text{C NMR}$  (75 MHz,  $\text{CDCl}_3$ )**  $\delta$  (ppm) 163.4, 149.7, 146.3, 145.9, 130.2, 129.6, 129.1, 127.4, 123.8, 119.4, 50.6, 35.1, 28.1, 24.2, 18.8. **IR (neat)  $\nu_{\max}$  (cm<sup>-1</sup>)** = 3380, 2926, 1669, 1522. **HRMS (ESI)**: Calcd. For  $\text{C}_{18}\text{H}_{22}\text{N}_2\text{O}$   $[\text{M}+\text{Na}]^+$ , 305.1630, found 305.1624.

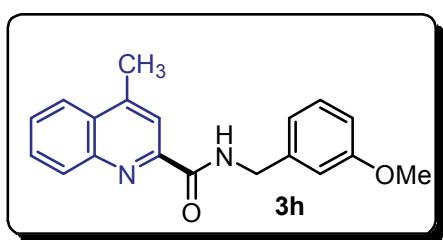
## Supporting Information

**4-Methyl-N-(2-phenylpropan-2-yl) quinoline-2-carboxamide: 3g** (131 mg) was obtained through the



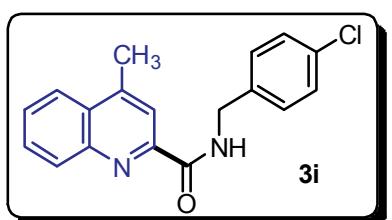
general procedure in 86 % yield as a yellow gel.  $R_f = 0.85$  (EtOAc-Hexane 10/90). **1H NMR (300 MHz, CDCl<sub>3</sub>)**  $\delta$  (ppm) 8.79 (s, 1H), 8.24 – 8.13 (m, 2H), 8.05 (dd,  $J = 8.2, 1.4$  Hz, 1H), 7.78 (ddd,  $J = 8.4, 6.8, 1.4$  Hz, 1H), 7.71 – 7.54 (m, 3H), 7.45 – 7.36 (m, 2H), 7.34 – 7.24 (m, 1H), 2.75 (d,  $J = 0.8$  Hz, 3H), 1.95 (s, 6H). **13C NMR (75 MHz, CDCl<sub>3</sub>)**  $\delta$  (ppm) 163.7, 149.9, 147.0, 146.3, 146.0, 130.3, 129.7, 129.2, 128.8, 128.5, 127.6, 126.7, 124.9, 123.9, 119.3, 55.8, 29.3, 18.9. **IR (neat)  $\nu_{max}$  (cm<sup>-1</sup>)** = 3372, 2976, 1677, 1503. **HRMS (ESI)**: Calcd. For C<sub>20</sub>H<sub>20</sub>N<sub>2</sub>O [M+Na]<sup>+</sup>, 327.1467, found 327.1474.

**N-(3-Methoxybenzyl)-4-methylquinoline-2-carboxamide: 3h** (115 mg) was obtained through the



general procedure in 66 % yield as a brown gel.  $R_f = 0.88$  (EtOAc-Hexane 10/90). **1H NMR (300 MHz, CDCl<sub>3</sub>)**  $\delta$  (ppm) 8.67 (s, 1H), 8.25 – 8.17 (m, 1H), 8.13 – 7.98 (m, 2H), 7.80 – 7.68 (m, 1H), 7.68 – 7.57 (m, 1H), 7.34 – 7.24 (m, 1H), 7.06 – 6.94 (m, 2H), 6.85 (dd,  $J = 8.0, 2.2$  Hz, 1H), 4.73 (d,  $J = 6.2$  Hz, 2H), 3.81 (s, 3H), 2.78 (d,  $J = 0.8$  Hz, 3H). **13C NMR (75 MHz, CDCl<sub>3</sub>)**  $\delta$  (ppm) 164.7, 160.0, 149.2, 146.3, 146.3, 140.0, 130.3, 129.8, 129.3, 127.7, 123.9, 120.2, 119.6, 113.5, 113.0, 55.3, 43.6, 19.0. **IR (neat)  $\nu_{max}$  (cm<sup>-1</sup>)** = 3381, 2936, 1672, 1527. **HRMS (ESI)**: Calcd. For C<sub>19</sub>H<sub>18</sub>N<sub>2</sub>O<sub>2</sub> [M+Na]<sup>+</sup>, 329.1260, found 329.1267.

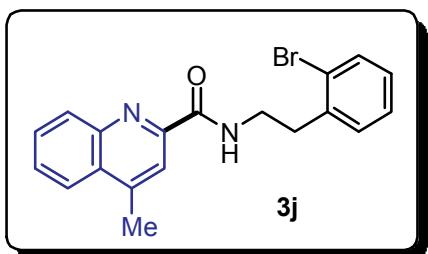
**N-(4-Chlorobenzyl)-4-methylquinoline-2-carboxamide: 3i** (132 mg) was obtained through the general



procedure in 77 % yield as a yellow gel.  $R_f = 0.61$  (EtOAc-Hexane 15/85). **1H NMR (300 MHz, CDCl<sub>3</sub>)**  $\delta$  (ppm) 8.68 (s, 1H), 8.24 – 8.16 (m, 1H), 8.13 – 8.01 (m, 1H), 7.82 – 7.59 (m, 3H), 7.39 – 7.30 (m, 4H), 7.16 – 7.05 (m, 1H), 4.71 (d,  $J = 6.2$  Hz, 1H), 2.80 (d,  $J = 0.8$  Hz, 3H). **13C NMR (75 MHz, CDCl<sub>3</sub>)**  $\delta$  (ppm) 164.8, 149.0, 146.3, 136.9, 133.1, 130.2, 129.7, 129.2, 128.7, 127.7, 123.9, 119.5, 42.8, 18.9. **IR (neat)  $\nu_{max}$  (cm<sup>-1</sup>)** = 3379, 2924, 1685, 1526. **HRMS (ESI)**: Calcd. For C<sub>18</sub>H<sub>15</sub>N<sub>2</sub>OCl, [M+H]<sup>+</sup>, 311.0945, found 311.0939.

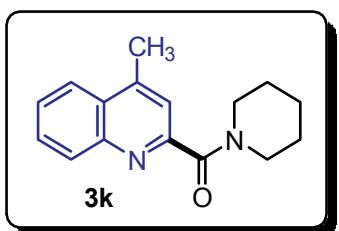
## Supporting Information

**N-(2-Bromophenethyl)-4-methylquinoline-2-carboxamide: 3j** (135 mg) was obtained through the general procedure in 73 % yield as a yellow solid; m.p. 245–249 °C.



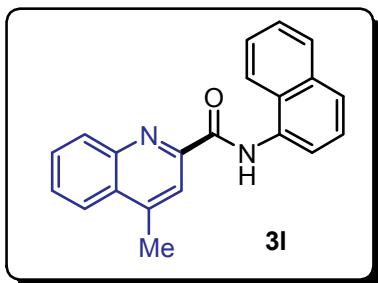
$R_f = 0.69$  (EtOAc-Hexane 15/85). **<sup>1</sup>H NMR** (300 MHz, CDCl<sub>3</sub>) δ (ppm) 8.45 (s, 1H), 8.19 (d, *J* = 0.8 Hz, 1H), 8.12 – 8.03 (m, 2H), 7.80 – 7.74 (m, 1H), 7.69 – 7.58 (m, 2H), 7.35 (dd, *J* = 7.6, 1.8 Hz, 1H), 7.31 – 7.28 (m, 1H), 7.17 – 7.11 (m, 1H), 3.84 (dt, *J* = 7.4, 6.5 Hz, 2H), 3.18 (t, *J* = 7.3 Hz, 2H), 2.79 (d, *J* = 0.9 Hz, 3H). **<sup>13</sup>C NMR** (75 MHz, CDCl<sub>3</sub>) δ (ppm) 164.9, 149.3, 146.4, 146.1, 138.4, 133.0, 131.1, 130.3, 129.7, 129.3, 128.3, 127.7, 124.7, 123.96, 119.4, 39.4, 36.2, 19.0. **IR (neat)**  $\nu_{\max}$  (cm<sup>-1</sup>) = 3379, 2930, 1672, 1525. **HRMS (ESI)**: Calcd. For C<sub>19</sub>H<sub>17</sub>N<sub>2</sub>OBr, [M+H]<sup>+</sup>, 369.0597, found 369.0594.

**(4-Methylquinoline-2-yl)(piperidin-1-yl)methanone: 3k** (103 mg) was obtained through the general



procedure in 81 % yield as a brown gel,  $R_f = 0.74$  (EtOAc-Hexane 10/90). **<sup>1</sup>H NMR** (300 MHz, CDCl<sub>3</sub>) δ (ppm) 8.16 – 8.09 (m, 1H), 8.00 (dd, *J* = 8.4, 1.2 Hz, 1H), 7.77–7.70 (m, 1H), 7.64–7.57 (m, 1H), 7.51 – 7.45 (m, 1H), 3.78 (t, *J* = 4.8 Hz, 2H), 3.55 – 3.39 (m, 2H), 2.73 (d, *J* = 0.8 Hz, 3H), 1.82 – 1.46 (m, 6H). **<sup>13</sup>C NMR** (75 MHz, CDCl<sub>3</sub>) δ (ppm) 167.5, 153.8, 146.6, 146.1, 130.1, 129.8, 128.1, 127.4, 123.8, 121.0, 48.4, 43.4, 26.6, 25.6, 24.6, 19.0. **IR (neat)**  $\nu_{\max}$  (cm<sup>-1</sup>) = 3344, 2937, 1669, 1557. **HRMS (APCI)**: Calcd. For C<sub>16</sub>H<sub>19</sub>N<sub>2</sub>O, [M+H]<sup>+</sup>, 255.1491, found 255.1492.

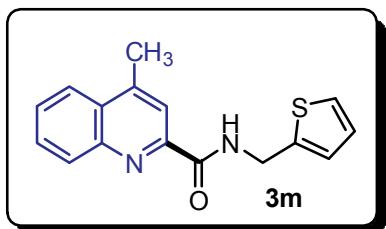
**4-Methyl-N-(naphthalen-1yl) quinolone-2-carboxamide: 3l** (91 mg) was obtained through the general



procedure in 58 % yield as a brown gel.  $R_f = 0.66$  (EtOAc-Hexane 15/85). **<sup>1</sup>H NMR** (300 MHz, CDCl<sub>3</sub>) δ (ppm) 11.01 (s, 1H), 8.47 (dd, *J* = 7.5, 1.0 Hz, 1H), 8.33 – 8.26 (m, 2H), 8.24 – 8.18 (m, 1H), 8.11 (dd, *J* = 8.4, 0.9 Hz, 1H), 7.93 (dd, *J* = 8.4, 1.0 Hz, 1H), 7.84 (ddd, *J* = 8.4, 6.9, 1.4 Hz, 1H), 7.77 – 7.53 (m, 6H), 2.85 (d, *J* = 0.9 Hz, 3H). **<sup>13</sup>C NMR** (75 MHz, CDCl<sub>3</sub>) δ (ppm) 162.8, 149.5, 146.8, 146.3, 134.3, 132.7, 130.6, 130.1, 129.6, 129.0, 128.0, 126.3, 125.1, 124.1, 120.7, 119.5, 118.76, 19.2. **IR (neat)**  $\nu_{\max}$  (cm<sup>-1</sup>) = 3331, 2926, 1691, 1542. **HRMS (ESI)**: Calcd. For C<sub>21</sub>H<sub>16</sub>N<sub>2</sub>O, [M+H]<sup>+</sup>, 313.1335, found 313.1345.

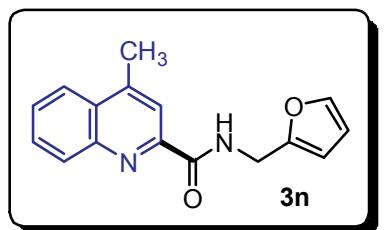
## Supporting Information

**4-Methyl-N-(thiophen-2-ylmethyl) quinoline-2-carboxamide: 3m** (113 mg) was obtained through the



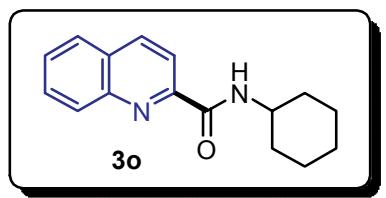
general procedure in 80 % yield as a white solid, m.p. 145-148 °C.  $R_f$  = 0.41 (EtOAc-Hexane 20/80).  **$^1\text{H NMR}$  (300 MHz,  $\text{CDCl}_3$ )**  $\delta$  (ppm) 8.71 (s, 1H), 8.19 (d,  $J$  = 0.9 Hz, 1H), 8.12 – 7.97 (m, 2H), 7.77 – 7.67 (m, 1H), 7.66 – 7.56 (m, 1H), 7.25 (dd,  $J$  = 5.2, 1.2 Hz, 1H), 7.14 – 7.05 (m, 1H), 6.99 (dd,  $J$  = 5.2, 3.4 Hz, 1H), 4.91 (dd,  $J$  = 6.0, 0.8 Hz, 2H), 2.75 (d,  $J$  = 0.8 Hz, 3H).  **$^{13}\text{C NMR}$  (75 MHz,  $\text{CDCl}_3$ )**  $\delta$  (ppm) 164.5, 149.0, 146.3, 146.2, 141.0, 130.2, 129.7, 129.3, 127.7, 126.9, 126.1, 125.2, 123.9, 119.5, 38.3, 18.9. **IR (neat)  $\nu_{\max}$  (cm<sup>-1</sup>)** = 3381, 2924, 1671, 1596. **HRMS (ESI)**: Calcd. For  $\text{C}_{16}\text{H}_{14}\text{N}_2\text{OS} [\text{M}+\text{Na}]^+$ , 305.0719, found 305.0714.

**N-(Furan-2-ylmethyl)-4-methylquinoline-2-carboxamide: 3n** (111 mg) was obtained through the



general procedure in 71 % yield as a white solid; m.p. 145-148 °C.  $R_f$  = 0.40 (EtOAc-Hexane 20/80).  **$^1\text{H NMR}$  (300 MHz,  $\text{CDCl}_3$ )**  $\delta$  (ppm) 8.59 (s, 1H), 8.18 (d,  $J$  = 1.0 Hz, 1H), 8.13 – 8.01 (m, 2H), 7.80-7.72 (m, 1H), 7.68-7.61 (m, 1H), 7.42 (dd,  $J$  = 1.8, 1.0 Hz, 1H), 6.41 – 6.32 (m, 2H), 4.74 (d,  $J$  = 6.0 Hz, 2H), 2.79 (d,  $J$  = 1.0 Hz, 3H).  **$^{13}\text{C NMR}$  (75 MHz,  $\text{CDCl}_3$ )**  $\delta$  (ppm) 164.6, 151.4, 149.0, 146.4, 146.1, 142.3, 130.3, 129.7, 127.7, 123.9, 119.5, 110.4, 107.5, 36.5, 18.9. **IR (neat)  $\nu_{\max}$  (cm<sup>-1</sup>)** = 3382, 2923, 1673, 1596.

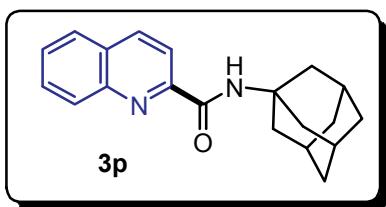
**N-Cyclohexylquinoline-2-carboxamide: 3o** (101 mg) was obtained through the general procedure in



80 % yield as a yellow solid; m.p. 92-94 °C.  $R_f$  = 0.57 (EtOAc-Hexane 15/85).  **$^1\text{H NMR}$  (300 MHz,  $\text{CDCl}_3$ )**  $\delta$  (ppm) 8.34 – 8.22 (m, 2H), 8.17 (d,  $J$  = 8.0 Hz, 1H), 8.12 – 8.05 (m, 1H), 7.83 (dd,  $J$  = 8.2, 1.0 Hz, 1H), 7.72 (ddd,  $J$  = 8.4, 6.8, 1.4 Hz, 1H), 7.57 (ddd,  $J$  = 8.0, 6.8, 1.2 Hz, 1H), 4.12 – 3.89 (m, 1H), 2.12 – 1.95 (m, 2H), 1.85 – 1.58 (m, 3H), 1.52 – 1.30 (m, 5H).  **$^{13}\text{C NMR}$  (75 MHz,  $\text{CDCl}_3$ )**  $\delta$  (ppm) 163.5, 150.2, 146.5, 137.4, 130.0, 129.7, 129.3, 127.8, 127.8, 118.9, 48.4, 33.2, 25.7, 25.0. **IR (neat)  $\nu_{\max}$  (cm<sup>-1</sup>)** = 3381, 2931, 1671, 1526. **HRMS (ESI)**: Calcd. For  $\text{C}_{16}\text{H}_{18}\text{N}_2\text{O}$ ,  $[\text{M}+\text{H}]^+$ , 255.1491, found 255.1499.

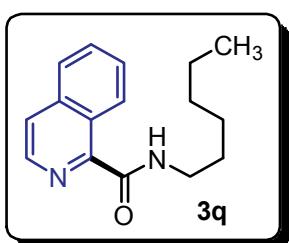
## Supporting Information

**N-((1s, 3s)-Adamantan-1-yl) quinoline-2-carboxamide: 3p** (111 mg) was obtained through the general



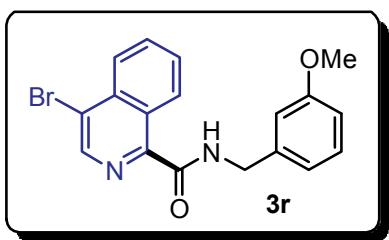
procedure in 72 % yield as a white gel.  $R_f = 0.56$  (EtOAc-Hexane 15/85). **1H NMR (300 MHz, CDCl<sub>3</sub>)** δ (ppm) 8.30 (s, 2H), 8.20 – 8.07 (m, 2H), 7.87 (dd,  $J = 8.2, 1.4$  Hz, 1H), 7.75 (dtd,  $J = 8.4, 6.8, 1.4$  Hz, 1H), 7.61 (ddd,  $J = 8.2, 6.8, 1.2$  Hz, 1H), 2.33 – 2.10 (m, 9H), 1.87 – 1.67 (m, 6H). **13C NMR (75 MHz, CDCl<sub>3</sub>)** δ (ppm) 163.4, 150.9, 146.4, 137.5, 130.0, 129.7, 129.2, 127.8, 127.7, 118.6, 51.8, 41.6, 36.6, 29.6. **IR (neat) v<sub>max</sub> (cm<sup>-1</sup>)** = 3364, 2908, 1676, 1527. **HRMS (ESI)**: Calcd. For C<sub>20</sub>H<sub>22</sub>N<sub>2</sub>O [M+Na]<sup>+</sup>, 329.1624, found 329.1625.

**N-Hexylisoquinoline-3-carboxamide: 3q** (99 mg) was obtained through the general procedure in 77 %



yield as a yellow gel.  $R_f = 0.60$  (EtOAc-Hexane 15/85). **1H NMR (300 MHz, CDCl<sub>3</sub>)** δ (ppm) 9.65 – 9.52 (m, 1H), 8.41 (d,  $J = 5.5$  Hz, 1H), 8.21 (s, 1H), 7.86 – 7.55 (m, 4H), 3.58 – 3.39 (m, 2H), 1.64 (q,  $J = 7.8$  Hz, 2H), 1.50 – 1.23 (m, 6H), 0.96 – 0.80 (m, 3H). **13C NMR (75 MHz, CDCl<sub>3</sub>)** δ (ppm) 166.1, 148.6, 140.2, 137.4, 130.5, 128.6, 128.0, 127.0, 126.8, 124.2, 39.6, 31.6, 29.7, 26.8, 22.6, 14.1. **IR (neat) v<sub>max</sub> (cm<sup>-1</sup>)** = 3381, 2929, 1666, 1518. **HRMS (ESI)**: Calcd. C<sub>16</sub>H<sub>21</sub>N<sub>2</sub>O, [M+H]<sup>+</sup>, 257.1648, found, 257.1651.

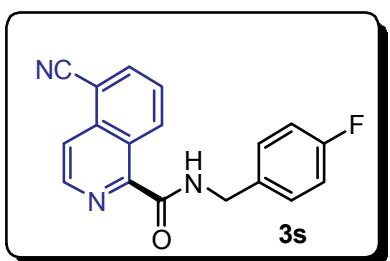
**4-Bromo-N-(3-methoxybenzyl) isoquinoline-1-carboxamide: 3r** (131 mg) was obtained through the



general procedure in 70 % yield as a yellow gel.  $R_f = 0.61$  (EtOAc-Hexane 15/85). **1H NMR (300 MHz, CDCl<sub>3</sub>)** δ (ppm) 9.68 (d,  $J = 8.4$  Hz, 1H), 8.61 (bs, 1H), 8.49 (bs, 1H), 8.19 (d,  $J = 8.4$  Hz, 1H), 7.90 – 7.66 (m, 2H), 7.27 (t,  $J = 7.0$  Hz, 1H), 7.07 – 6.92 (m, 2H), 6.84 (d,  $J = 8.2$  Hz, 1H), 4.70 (d,  $J = 6.0$  Hz, 2H), 3.81 (s, 3H). **13C NMR (75 MHz, CDCl<sub>3</sub>)** δ (ppm) 165.4, 159.9, 147.4, 142.1, 139.8, 135.9, 131.8, 129.8, 129.5, 128.4, 126.1, 123.5, 123.3, 120.1, 113.5, 113.0, 55.5, 55.3, 43.6. **IR (neat) v<sub>max</sub> (cm<sup>-1</sup>)** = 3378, 2929, 1668, 1516. **HRMS (ESI)**: Calcd. For, C<sub>18</sub>H<sub>16</sub>N<sub>2</sub>O<sub>2</sub>Br, [M+H]<sup>+</sup>, 371.0389, found 371.0392.

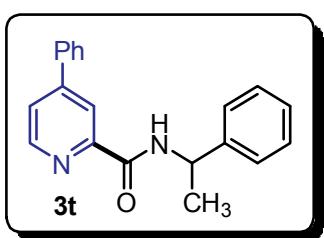
## Supporting Information

**Cyano-N-(4-fluorobenzyl)isoquinoline-1-carboxamide : 3s** (107 mg) was obtained through the general procedure in 70 % yield as a yellow gel.  $R_f = 0.30$  (EtOAc-Hexane 30/70).



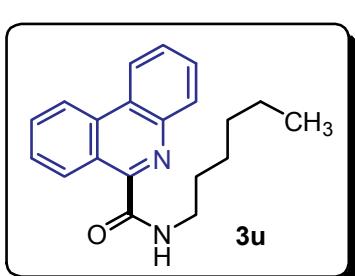
**1H NMR (300 MHz, CDCl<sub>3</sub>)**  $\delta$  (ppm) 10.06 (dt,  $J = 8.9, 1.1$  Hz, 1H), 8.78 – 8.55 (m, 2H), 8.29 – 8.11 (m, 2H), 7.79 (dd,  $J = 8.8, 7.2$  Hz, 1H), 7.46 – 7.36 (m, 2H), 7.15 – 7.00 (m, 2H), 4.71 (d,  $J = 6.1$  Hz, 2H). **13C NMR (75 MHz, CDCl<sub>3</sub>)**  $\delta$  (ppm) 165.2, 164.0, 148.6, 142.7, 137.0, 133.7, 129.7, 129.6, 127.9, 126.7, 121.6, 116.7, 115.9, 115.6, 109.9, 43.1. **IR (neat) v<sub>max</sub> (cm<sup>-1</sup>)** = 3381, 2929, 1666, 1518. **HRMS (ESI)**: Calcd. For C<sub>16</sub>H<sub>21</sub>N<sub>2</sub>O, [M+Na]<sup>+</sup>, 328.0856, found, 328.0855.

**4-Phenyl-N (1-phenylethyl) picolinamide: 3t** (86 mg) was obtained through the general procedure in



57 % yield as a yellow gel.  $R_f = 0.53$  (EtOAc-Hexane 20/80). **1H NMR (300 MHz, CDCl<sub>3</sub>)**  $\delta$  (ppm) 8.58 (dd,  $J = 5.1, 0.7$  Hz, 1H), 8.52 – 8.30 (m, 2H), 7.73 – 7.68 (m, 2H), 7.64 (dd,  $J = 5.1, 1.9$  Hz, 1H), 7.54 – 7.42 (m, 5H), 7.40 – 7.33 (m, 2H), 7.31 – 7.27 (m, 1H), 5.45 – 5.25 (m, 1H), 1.65 (d,  $J = 6.9$  Hz, 3H). **13C NMR (75 MHz, CDCl<sub>3</sub>)**  $\delta$  (ppm) 163.6, 150.6, 150.0, 148.7, 143.4, 137.5, 129.4, 128.8, 127.3, 126.4, 123.9, 120.3, 49.0, 22.2. **IR (neat) v<sub>max</sub> (cm<sup>-1</sup>)** = 3378, 2974, 1670, 1515. **HRMS (ESI)**: Calcd. For C<sub>20</sub>H<sub>18</sub>N<sub>2</sub>O [M+Na]<sup>+</sup>, 325.1311, found 325.1325.

**N-Hexylphenanthridine-6-carboxamide: 3u** (142 mg) was obtained through the general procedure in

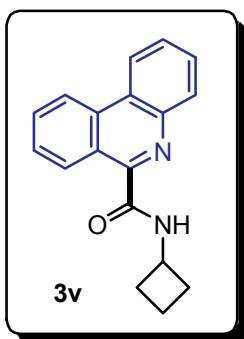


93 % yield as a yellow solid; m.p. 91–95 °C.  $R_f = 0.61$  (EtOAc-Hexane 10/90). **1H NMR (300 MHz, CDCl<sub>3</sub>)**  $\delta$  (ppm) 9.59 – 9.48 (m, 1H), 8.55 – 8.39 (m, 2H), 8.20 (s, 1H), 8.12 – 8.03 (m, 1H), 7.81 – 7.72 (m, 1H), 7.71 – 7.57 (m, 3H), 3.61 – 3.45 (m, 2H), 1.79 – 1.60 (m, 2H), 1.51 – 1.24 (m, 6H), 0.99 – 0.79 (m, 3H). **13C NMR (75 MHz, CDCl<sub>3</sub>)**  $\delta$  (ppm) 166.1, 149.8, 141.8, 133.6, 130.8, 130.3, 129.0, 128.7, 128.3, 127.8, 125.3, 124.2, 122.0, 121.7, 39.8, 31.6, 29.6, 26.8, 22.6, 14.1. **IR (neat) v<sub>max</sub> (cm<sup>-1</sup>)** = 3295, 2928, 1653, 1522.

**HRMS (ESI)**: Calcd. For C<sub>20</sub>H<sub>23</sub>N<sub>2</sub>O, [M+H]<sup>+</sup>, 307.1804, found 307.1810.

## Supporting Information

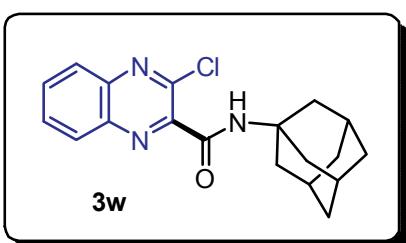
**N-Cyclobutylphenanthridine-6-carboxamide:** **3v** (110 mg) was obtained through the general



procedure in 79 % yield as a yellow gel.  $R_f = 0.53$  (EtOAc-Hexane 10/90).  **$^1H$  NMR (300 MHz, CDCl<sub>3</sub>)**  $\delta$  (ppm) 9.66 – 9.49 (m, 1H), 8.65 – 8.55 (m, 2H), 8.31 (d,  $J = 7.4$  Hz, 1H), 8.22 – 8.12 (m, 1H), 7.85 (td,  $J = 7.0, 3.5$  Hz, 1H), 7.80 – 7.69 (m, 3H), 4.67 (dd,  $J = 16.4, 8.1$  Hz, 1H), 2.59 – 2.43 (m, 2H), 2.19 – 2.09 (m, 2H), 1.89 – 1.79 (m, 2H).  **$^{13}C$  NMR (75 MHz, CDCl<sub>3</sub>)**  $\delta$  (ppm) 165.2, 149.5, 141.9, 133.8, 131.1, 130.5, 129.2, 128.9, 128.5, 128.0, 125.6, 124.4, 122.2, 121.9, 45.06, 31.3, 15.5, 8.8. **IR (neat)  $\nu_{max}$  (cm<sup>-1</sup>)** = 3372, 2976, 1678, 1520. **HRMS (ESI):**

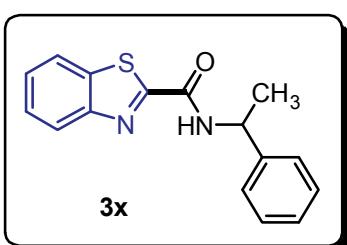
Calcd. For, C<sub>18</sub>H<sub>16</sub>N<sub>2</sub>O, [M+H]<sup>+</sup>, 277.1335, found 277.1340.

**N-((3s, 5s, 7s)-Adamantan-1-yl)-3-chloroquinoxaline-2-carboxamide:** **3w** (120 mg) was obtained



through the general procedure in 71 % yield as a yellow solid; m.p. 198-203 °C.  $R_f = 0.30$  (EtOAc-Hexane 30/70).  **$^1H$  NMR (300 MHz, CDCl<sub>3</sub>)**  $\delta$  (ppm) 8.11 – 7.96 (m, 2H), 7.89 – 7.72 (m, 2H), 7.15 (s, 1H), 2.28 – 2.07 (m, 9H), 1.73 (s, 6H).  **$^{13}C$  NMR (75 MHz, CDCl<sub>3</sub>)**  $\delta$  (ppm) 161.8, 145.1, 144.2, 142.4, 138.9, 132.4, 130.9, 129.2, 128.3, 52.8, 41.6, 41.4, 36.4, 29.7, 29.6. **IR (neat)  $\nu_{max}$  (cm<sup>-1</sup>)** = 3294, 2909, 1665, 1563. **HRMS (ESI):** Calcd. For, C<sub>19</sub>H<sub>21</sub>N<sub>3</sub>OCl, [M+H]<sup>+</sup>, 342.1367, found 342.1362.

**N-(1-Phenylethyl) benzo[d]thiazole-2-carboxamide:** **3x** (58 mg) was obtained through the general

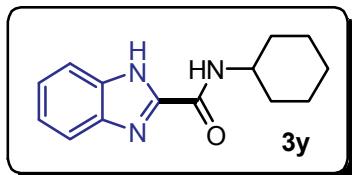


procedure in 41 % yield as a yellow solid; m.p. 145-148 °C.  $R_f = 0.50$  (EtOAc-Hexane 20/80).  **$^1H$  NMR (300 MHz, CDCl<sub>3</sub>)**  $\delta$  (ppm) 8.11 – 8.04 (m, 1H), 8.02 – 7.94 (m, 1H), 7.74 (d,  $J = 8.4$  Hz, 1H), 7.61 – 7.29 (m, 6H), 5.48 – 5.27 (m, 1H), 1.70 (d,  $J = 6.8$  Hz, 3H).  **$^{13}C$  NMR (75 MHz, CDCl<sub>3</sub>)**  $\delta$  (ppm) 164.1, 159.1, 152.9, 142.5, 137.2, 128.9, 127.7, 126.9, 126.8, 126.4, 124.3, 122.5, 49.6, 21.9. **IR (neat)  $\nu_{max}$  (cm<sup>-1</sup>)** = 3392, 2976, 1667, 1524. **HRMS (ESI):**

Calcd. For C<sub>16</sub>H<sub>14</sub>N<sub>2</sub>OS [M+Na]<sup>+</sup>, 305.0719, found 305.0717.

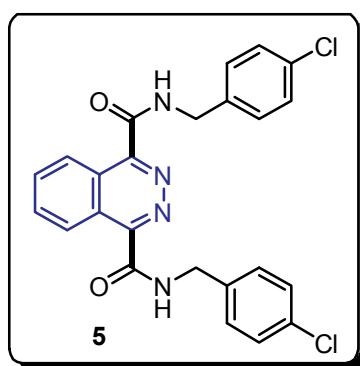
## Supporting Information

**N-Cyclohexyl-1*H*-benzo[d]thiazole-2-carboxamide: 3y** (71 mg) was obtained through the general



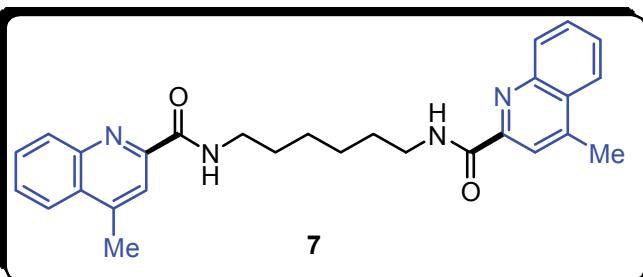
procedure in 59 % yield as a white solid; m.p. 153–156 °C.  $R_f = 0.50$  (EtOAc-Hexane 20/80). **1H NMR (300 MHz, CDCl<sub>3</sub>)** δ (ppm) 7.71 (s, 3H), 7.41 – 7.31 (m, 2H), 4.17 – 3.92 (m, 1H), 2.08 (d,  $J = 9.9$  Hz, 2H), 1.83 (dd,  $J = 9.2, 3.5$  Hz, 2H), 1.69 (d,  $J = 12.6$  Hz, 1H), 1.51 – 1.35 (m, 4H), 1.33 – 1.14 (m, 2H). **13C NMR (75 MHz, CDCl<sub>3</sub>)** δ (ppm) 159.1, 158.3, 145.3, 142.9, 134.1, 124.9, 123.3, 120.4, 112.2, 48.9, 32.9, 32.6, 25.4, 24.8. **IR (neat) v<sub>max</sub> (cm<sup>-1</sup>)** = 3379, 2971, 1658, 1541. **HRMS (ESI)**: Calcd. For C<sub>14</sub>H<sub>17</sub>N<sub>3</sub>O [M+Na]<sup>+</sup>, 266.1263, found 266.1270.

**N1,N4-Bis(4-chlorobenzyl)phthalazine-1,4-dicarboxamide: 5** (188 mg) was obtained through the



general procedure in 81 % yield as a yellow solid, m.p. 170–73 °C.  $R_f = 0.31$  (EtOAc-Hexane 30/70). **1H NMR (300 MHz, CDCl<sub>3</sub>)** δ (ppm) 9.65 – 9.42 (m, 2H), 8.46 (s, 2H), 8.14 – 7.91 (m, 2H), 7.40 – 7.28 (m, 8H), 4.71 (dd,  $J = 6.2, 1.8$  Hz, 4H). **13C NMR (75 MHz, CDCl<sub>3</sub>)** δ (ppm) 163.8, 150.4, 136.1, 134.0, 133.5, 129.2, 129.0, 126.8, 126.4, 43.1. **IR (neat) v<sub>max</sub> (cm<sup>-1</sup>)** = 3288, 2922, 1640, 1530. **HRMS (ESI)**: Calcd. For, C<sub>24</sub>H<sub>18</sub>N<sub>4</sub>O<sub>2</sub>Cl<sub>2</sub> [M+Na]<sup>+</sup>, 487.0699, found 487.0713.

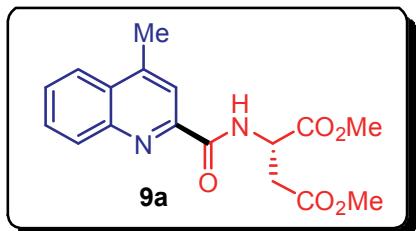
**N,N'-*(Hexane-1,6-diyl)*bis(4-methylquinoline-2-carboxamide) : 7** (115 mg) was obtained through the



general procedure, in 51 % yield as a yellow gel.  $R_f = 0.51$  (EtOAc-Hexane 30/70). **1H NMR (300 MHz, CDCl<sub>3</sub>)** δ (ppm) 8.31 (t,  $J = 5.2$  Hz, 2H), 8.15 (d,  $J = 0.8$  Hz, 2H), 8.09 (dd,  $J = 8.4, 0.8$  Hz, 2H), 8.02 (dd,  $J = 8.4, 0.8$  Hz, 2H), 7.77–7.70 (m, 2H), 7.65 – 7.56 (m, 2H), 3.58 – 3.49 (m, 4H), 2.76 (d,  $J = 0.8$  Hz, 6H), 1.78 – 1.67 (m, 4H), 1.57 – 1.48 (m, 4H). **13C NMR (75 MHz, CDCl<sub>3</sub>)** δ (ppm) 164.7, 149.5, 146.3, 146.0, 130.3, 129.6, 129.2, 127.5, 123.9, 119.4, 39.5, 29.7, 26.8, 18.9. **IR (neat) v<sub>max</sub> (cm<sup>-1</sup>)** = 3380, 2929, 1667, 1531. **HRMS (ESI)**: Calcd. For C<sub>28</sub>H<sub>30</sub>N<sub>4</sub>O<sub>2</sub> [M+Na]<sup>+</sup>, 477.2266, found 477.2260.

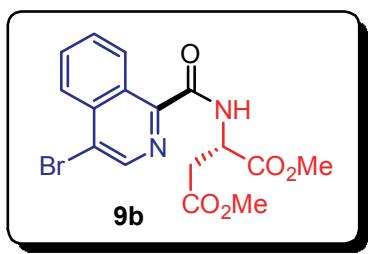
## Supporting Information

**(S)-Dimethyl 2-(4-methylquinoline-2-carboxamido) succinate: 9a** (144 mg) was obtained through the



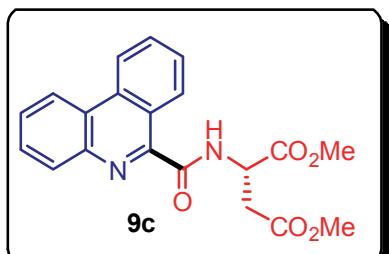
general procedure, in 87 % yield as a yellow gel.  $R_f = 0.43$  (EtOAc-Hexane 30/70).  **$^1\text{H NMR}$  (300 MHz,  $\text{CDCl}_3$ )**  $\delta$  (ppm) 9.04 (d,  $J = 8.4$  Hz, 1H), 8.14 – 8.06 (m, 2H), 7.97 (d,  $J = 8.4, 0.9$  Hz, 1H), 7.76 – 7.67 (m, 1H), 7.61 – 7.45 (m, 1H), 5.16 – 5.08 (m, 1H), 3.78 (s, 3H), 3.70 (s, 3H), 3.15 (dd,  $J = 17.0, 4.8$  Hz, 1H), 3.01 (dd,  $J = 17.0, 4.8$  Hz, 1H), 2.71 (d,  $J = 0.8$  Hz, 3H).  **$^{13}\text{C NMR}$  (75 MHz,  $\text{CDCl}_3$ )**  $\delta$  (ppm) 171.3, 164.7, 148.5, 146.4, 146.0, 130.6, 129.7, 129.3, 127.8, 123.8, 119.3, 52.9, 52.1, 48.8, 36.4, 18.9. **IR (neat)  $\nu_{\text{max}}$  (cm<sup>-1</sup>)** = 3383, 2954, 1677, 1504. **HRMS (ESI)**: Calcd. For  $\text{C}_{17}\text{H}_{18}\text{N}_2\text{O}_5$  [ $\text{M}+\text{Na}$ ]<sup>+</sup>, 353.1113, found 353.1111.  $[\alpha]_D^{25} +31.75$  ( $c$  1.8,  $\text{CHCl}_3$ ).

**(S)-Dimethyl 2-(4-bromoisoquinoline-1-carboxamido) succinate: 9b** (169 mg) was obtained through



the general procedure, in 86 % yield as a yellow gel.  $R_f = 0.41$  (EtOAc-Hexane 30/70).  **$^1\text{H NMR}$  (300 MHz,  $\text{CDCl}_3$ )**  $\delta$  (ppm) 9.56 – 9.51 (m, 1H), 8.95 (d,  $J = 8.2$  Hz, 1H), 8.65 (s, 1H), 8.21 – 8.08 (m, 1H), 7.81 – 7.74 (m, 1H), 7.72 – 7.65 (m, 1H), 5.14 – 5.06 (m, 1H), 3.79 (s, 3H), 3.69 (s, 3H), 3.16 (dd,  $J = 17.0, 4.8$  Hz, 1H), 3.02 (dd,  $J = 17.0, 4.8$  Hz, 1H).  **$^{13}\text{C NMR}$  (75 MHz,  $\text{CDCl}_3$ )**  $\delta$  (ppm) 171.2, 165.2, 146.5, 142.4, 135.8, 131.8, 129.6, 128.1, 126.1, 123.8, 53.0, 52.2, 48.8, 36.3. **IR (neat)  $\nu_{\text{max}}$  (cm<sup>-1</sup>)** = 3383, 2953, 1673, 1507. **HRMS (ESI)**: Calcd. For  $\text{C}_{16}\text{H}_{15}\text{BrN}_2\text{O}_5$  [ $\text{M}+\text{Na}$ ]<sup>+</sup>, 417.0062, found 417.0057.  $[\alpha]_D^{25} +29.38$  ( $c$  2.0,  $\text{CHCl}_3$ ).

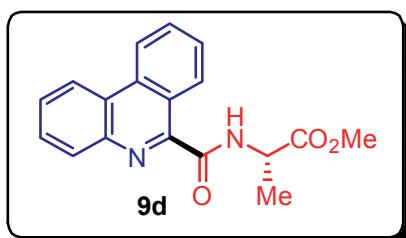
**(S)-Dimethyl 2-(4-phenanthridine-6-carboxamido) succinate: 9c** (163 mg) was obtained through the



general procedure, in 89 % yield as a yellow gel.  $R_f = 0.44$  (EtOAc-Hexane 30/70).  **$^1\text{H NMR}$  (300 MHz,  $\text{CDCl}_3$ )**  $\delta$  (ppm) 9.57 – 5.52 (m, 1H), 9.09 (d,  $J = 8.2$  Hz, 1H), 8.55 – 8.43 (m, 2H), 8.17 – 8.12 (m, 1H), 7.78 (m, 1H), 7.73 – 7.65 (s, 3H), 5.22 – 5.15 (m, 1H), 3.83 (s, 3H), 3.72 (s, 3H), 3.21 (dd,  $J = 17.0, 4.8$  Hz, 1H), 3.09 (dd,  $J = 17.0, 4.8$  Hz, 1H).  **$^{13}\text{C NMR}$  (75 MHz,  $\text{CDCl}_3$ )**  $\delta$  (ppm) 171.3, 165.8, 148.1, 141.7, 133.6, 130.8, 128.7, 127.9, 125.4, 124.2, 121.9, 52.9, 52.1, 48.9, 36.4. **IR (neat)  $\nu_{\text{max}}$  (cm<sup>-1</sup>)** = 3379, 2954, 1674, 1505.  $[\alpha]_D^{25} +71.80$  ( $c$  1.8,  $\text{CHCl}_3$ ).

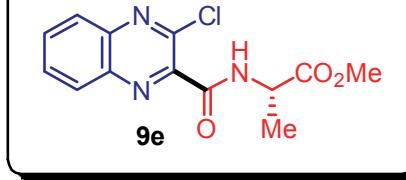
## Supporting Information

**(S)-Methyl 2-(phenanthridine-6-carboxamido) propanoate: 9d** (140 mg) was obtained through the general procedure, in 91 % yield as a white solid. m.p. 149–152 °C.

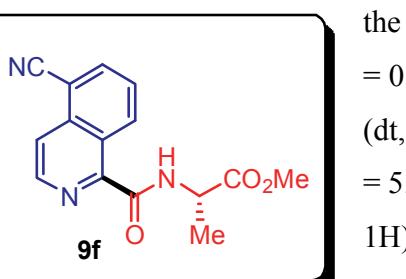


$R_f = 0.47$  (EtOAc-Hexane 30/70). **1H NMR (300 MHz, CDCl<sub>3</sub>)** δ (ppm) 9.54 (dd,  $J = 8.4, 0.8$  Hz, 1H), 8.73 (d,  $J = 7.4$  Hz, 1H), 8.60 – 8.37 (m, 2H), 8.22 – 8.06 (m, 1H), 7.86 – 7.59 (m, 4H), 4.92 – 4.81 (m, 1H), 3.84 (s, 3H), 1.66 (d,  $J = 7.2$  Hz, 3H). **13C NMR (75 MHz, CDCl<sub>3</sub>)** δ (ppm) 173.4, 165.6, 153.2, 148.5, 141.7, 133.6, 130.8, 130.5, 128.8, 128.5, 127.9, 125.4, 124.1, 122.0, 121.7, 52.5, 48.4, 18.3. **IR (neat) v<sub>max</sub> (cm<sup>-1</sup>)** = 3328, 2924, 1743, 1525. **HRMS (ESI)**: Calcd. For C<sub>18</sub>H<sub>16</sub>N<sub>2</sub>O<sub>3</sub> [M+Na]<sup>+</sup>, 331.1058, found 331.1054.  $[\alpha]_D^{25} +94.70$  (c 2.3, CHCl<sub>3</sub>).

**(S)-Methyl 2-(3-chloroquinoxaline-2-carboxamido) propanoate: 9e** (102 mg) was obtained through the general procedure, in 69 % yield as a yellow gel.  $R_f = 0.39$  (EtOAc-Hexane 30/70). **1H NMR (300 MHz, CDCl<sub>3</sub>)** δ (ppm) 8.22 – 8.16 (m, 1H), 8.13 – 8.08 (m, 1H), 7.98 – 7.83 (m, 2H), 4.94 – 4.80 (m, 1H), 3.85 (s, 3H), 1.64 (d,  $J = 7.2$  Hz, 3H). **13C NMR (75 MHz, CDCl<sub>3</sub>)** δ (ppm) 173.1, 161.9, 145.2, 142.7, 142.2, 138.9, 132.9, 131.0, 129.4, 128.3, 52.7, 48.5, 18.4. **IR (neat) v<sub>max</sub> (cm<sup>-1</sup>)** = 3331, 2924, 1741, 1680, 1530. **HRMS (ESI)**: Calcd. For C<sub>13</sub>H<sub>12</sub>ClN<sub>3</sub>O<sub>3</sub> [M+Na]<sup>+</sup>, 316.0464, found 316.0458.  $[\alpha]_D^{25} +39.33$  (c 1.7, CHCl<sub>3</sub>).



**(S)-Methyl 2-(5-cyanoisoquinoline-1-carboxamido) propanoate: 9f** (104 mg) was obtained through the general procedure, in 74 % yield as a white solid; m.p. 123–126 °C.  $R_f = 0.51$  (EtOAc-Hexane 30/70). **1H NMR (300 MHz, CDCl<sub>3</sub>)** δ (ppm) 9.94 (dt,  $J = 8.8, 1.2$  Hz, 1H), 8.74 (s, 1H), 8.72 (d,  $J = 5.6$  Hz, 2H), 8.23 (dd,  $J = 5.6, 1.2$  Hz, 1H), 8.16 (dd,  $J = 7.2, 1.2$  Hz, 1H), 7.75 (dd,  $J = 8.8, 7.2$  Hz, 1H), 4.90 – 4.77 (m, 1H), 3.83 (s, 3H), 1.62 (d,  $J = 7.2$  Hz, 3H). **13C NMR (75 MHz, CDCl<sub>3</sub>)** δ (ppm) 173.1, 164.7, 148.1, 142.7, 136.8, 133.3, 127.8, 126.5, 121.5, 116.5, 109.8, 52.6, 48.3, 27.7, 18.3. **IR (neat) v<sub>max</sub> (cm<sup>-1</sup>)** = 3377, 2929, 1743, 1671, 1513. **HRMS (ESI)**: Calcd. For C<sub>15</sub>H<sub>13</sub>N<sub>3</sub>O<sub>3</sub> [M+Na]<sup>+</sup>, 306.0854, found 306.0849.  $[\alpha]_D^{25} +167.66$  (c 1.4, CHCl<sub>3</sub>).



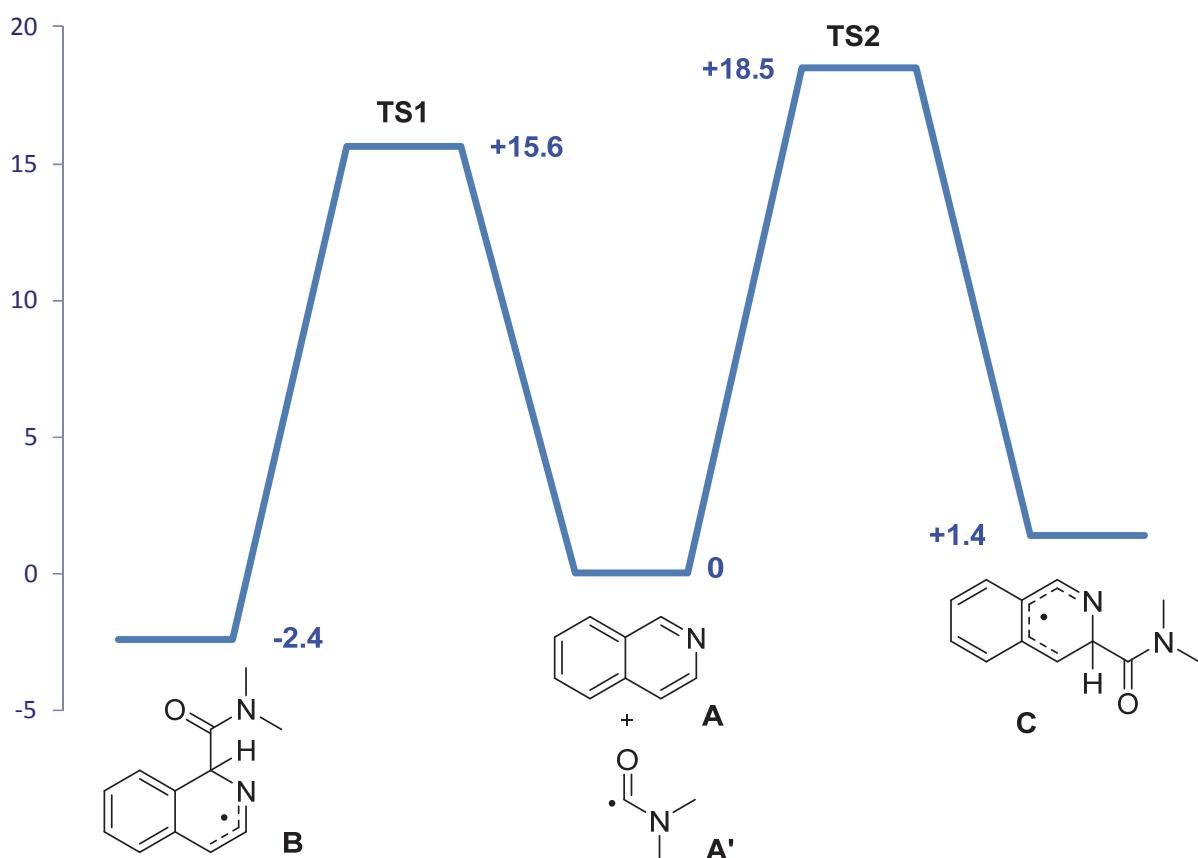
## Supporting Information

### 7. DFT calculations

All DFT calculations were performed with the Gaussian16 software package.<sup>5</sup> Frequency calculations were performed to insure that there is no imaginary frequencies for local minima and only one for transition states. All calculations were performed at the WB97XD/cc-pvtz level.

All these computed structures are available on request. This Supporting information contains free Gibbs energies and cartesian coordinates of all structures computed.

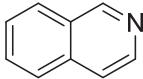
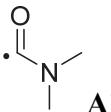
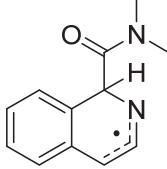
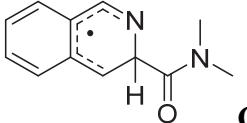
#### DFT WB97XD/cc-pvtz



<sup>5</sup> Gaussian 16, Revision A.03, M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, G. A. Petersson, H. Nakatsuji, X. Li, M. Caricato, A. V. Marenich, J. Bloino, B. G. Janesko, R. Gomperts, B. Mennucci, H. P. Hratchian, J. V. Ortiz, A. F. Izmaylov, J. L. Sonnenberg, D. Williams-Young, F. Ding, F. Lipparini, F. Egidi, J. Goings, B. Peng, A. Petrone, T. Henderson, D. Ranasinghe, V. G. Zakrzewski, J. Gao, N. Rega, G. Zheng, W. Liang, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, K. Throssell, J. A. Montgomery, Jr., J. E. Peralta, F. Ogliaro, M. J. Bearpark, J. J. Heyd, E. N. Brothers, K. N. Kudin, V. N. Staroverov, T. A. Keith, R. Kobayashi, J. Normand, K. Raghavachari, A. P. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, J. M. Millam, M. Klene, C. Adamo, R. Cammi, J. W. Ochterski, R. L. Martin, K. Morokuma, O. Farkas, J. B. Foresman, and D. J. Fox, Gaussian, Inc., Wallingford CT, 2016.

## Supporting Information

### Free Gibbs energies at WB97XD/cc-pvtz level

Compounds	Sum of electronic and thermal Free Energies (Hartrees)	Energy difference (kcal/mol)
 <b>A</b>	-401.807061	-
 <b>A'</b>	-247.799614	-
<b>A + A'</b>	-649.606675	0
<b>TS1</b>	-649.581789	+15.6
<b>TS2</b>	-649.577183	+18.5
 <b>B</b>	-649.610476	-2.4
 <b>C</b>	-649.604367	+1.4

#### Compound A

Center Number	Atomic Number	Atomic Type	Coordinates (Angstroms)		
			X	Y	Z
1	6	0	2.399307	-0.707492	0.000001
2	6	0	1.226812	-1.406049	-0.000009
3	6	0	-0.009814	-0.720189	-0.000001
4	6	0	-0.008492	0.691340	-0.000005
5	6	0	1.219334	1.390109	-0.000000
6	6	0	2.398703	0.704171	0.000007
7	1	0	-1.320845	-2.446562	-0.000012
8	1	0	3.342792	-1.236453	0.000006
9	1	0	1.228753	-2.488271	-0.000010
10	6	0	-1.266237	-1.366115	0.000002
11	6	0	-1.261649	1.350784	-0.000009
12	1	0	1.208577	2.472617	0.000001
13	1	0	3.339453	1.237162	0.000015
14	6	0	-2.404003	-0.615563	0.000019
15	1	0	-1.281442	2.437460	0.000007
16	1	0	-3.378255	-1.088314	-0.000004
17	7	0	-2.414684	0.740911	-0.000005

## Supporting Information

### Compound A'

Center Number	Atomic Number	Atomic Type	Coordinates (Angstroms)		
			X	Y	Z
1	6	0	-0.834256	-0.692394	-0.000016
2	8	0	-1.945125	-0.252429	-0.000049
3	7	0	0.327712	-0.041518	-0.000001
4	6	0	1.583793	-0.760055	0.000061
5	1	0	2.171240	-0.514683	-0.886768
6	1	0	2.171165	-0.514667	0.886935
7	1	0	1.375225	-1.826590	0.000062
8	6	0	0.337084	1.417898	-0.000009
9	1	0	-0.169089	1.804431	-0.884248
10	1	0	-0.168983	1.804448	0.884283
11	1	0	1.367725	1.764419	-0.000074

### TS1

Center Number	Atomic Number	Atomic Type	Coordinates (Angstroms)		
			X	Y	Z
1	6	0	0.068023	-0.041326	0.059340
2	6	0	1.436184	0.011345	0.072999
3	6	0	2.100819	1.252358	0.005286
4	6	0	1.336156	2.431374	-0.071739
5	6	0	-0.062577	2.355461	-0.079450
6	6	0	-0.690334	1.138107	-0.019911
7	1	0	4.140852	0.505604	0.046899
8	1	0	-0.435950	-0.997002	0.111651
9	1	0	2.021417	-0.897080	0.134941
10	6	0	3.512755	1.383738	-0.015593
11	6	0	2.039907	3.686021	-0.091869
12	1	0	-0.641408	3.269470	-0.123299
13	1	0	-1.770069	1.081987	-0.026515
14	6	0	4.070872	2.625767	-0.178111
15	1	0	1.474189	4.569762	-0.375678
16	1	0	5.146136	2.732391	-0.262525
17	7	0	3.367037	3.763069	-0.273140
18	6	0	1.810979	4.256010	2.001926
19	8	0	1.255428	3.449260	2.681197
20	7	0	2.316552	5.442761	2.338643
21	6	0	2.917161	6.368574	1.399048
22	1	0	2.226953	7.181872	1.160824
23	1	0	3.810102	6.800736	1.852721
24	1	0	3.218802	5.846556	0.496280
25	6	0	2.172259	5.916594	3.708477
26	1	0	1.690336	5.146951	4.303017
27	1	0	3.153755	6.140780	4.129307
28	1	0	1.566368	6.824816	3.731205

## Supporting Information

### TS2

Center Number	Atomic Number	Atomic Type	Coordinates (Angstroms)		
			X	Y	Z
1	6	0	-0.004828	0.011649	0.027405
2	6	0	1.361728	-0.019770	0.038488
3	6	0	2.105151	1.183180	-0.014345
4	6	0	1.401140	2.409616	-0.092329
5	6	0	-0.002648	2.413895	-0.106537
6	6	0	-0.698732	1.235156	-0.043624
7	1	0	4.084228	0.322459	0.049878
8	1	0	-0.564191	-0.913040	0.076211
9	1	0	1.891086	-0.961559	0.099419
10	6	0	3.506157	1.234230	-0.004177
11	6	0	2.170591	3.606473	-0.204242
12	1	0	-0.526331	3.359815	-0.168353
13	1	0	-1.779797	1.238604	-0.051226
14	6	0	4.149307	2.473240	0.052243
15	1	0	1.645421	4.547381	-0.349906
16	1	0	5.208033	2.519783	-0.178057
17	7	0	3.462056	3.656234	-0.173683
18	6	0	4.598721	2.661465	2.132463
19	8	0	4.121750	1.819902	2.830985
20	7	0	5.418578	3.662955	2.464600
21	6	0	5.935062	4.635070	1.522852
22	1	0	5.890528	5.625462	1.978300
23	1	0	6.977423	4.422026	1.271718
24	1	0	5.324531	4.652398	0.625332
25	6	0	5.888076	3.779084	3.837619
26	1	0	5.582871	4.738657	4.259068
27	1	0	5.461783	2.973989	4.427789
28	1	0	6.977759	3.714951	3.868586

### Compound B

Center Number	Atomic Number	Atomic Type	Coordinates (Angstroms)		
			X	Y	Z
1	6	0	0.019558	0.028165	0.020690
2	6	0	1.389207	0.058374	0.154736
3	6	0	2.081122	1.279176	0.106779
4	6	0	1.361722	2.471342	-0.068839
5	6	0	-0.014054	2.423910	-0.211957
6	6	0	-0.686442	1.214237	-0.168404
7	1	0	4.088027	0.470963	0.344986
8	1	0	-0.508511	-0.914618	0.059502
9	1	0	1.947658	-0.858349	0.294196
10	6	0	3.496692	1.362560	0.190476
11	6	0	2.098687	3.782325	-0.073887
12	1	0	-0.569890	3.344092	-0.343110
13	1	0	-1.762180	1.191196	-0.275440
14	6	0	4.136472	2.600660	-0.010391
15	1	0	1.723892	4.386996	-0.905672
16	1	0	5.222119	2.623252	-0.046366
17	7	0	3.541213	3.738614	-0.190750
18	6	0	1.792967	4.567658	1.240552
19	8	0	1.390174	3.978497	2.221885
20	7	0	2.008546	5.906079	1.210045
21	6	0	2.507984	6.638916	0.064233
22	1	0	1.693571	7.057431	-0.534824
23	1	0	3.120259	7.466480	0.422758
24	1	0	3.142615	6.009292	-0.551240
25	6	0	1.692254	6.694909	2.382398
26	1	0	1.263070	6.046772	3.138900
27	1	0	2.593778	7.165962	2.780688
28	1	0	0.976335	7.479483	2.126548

## Supporting Information

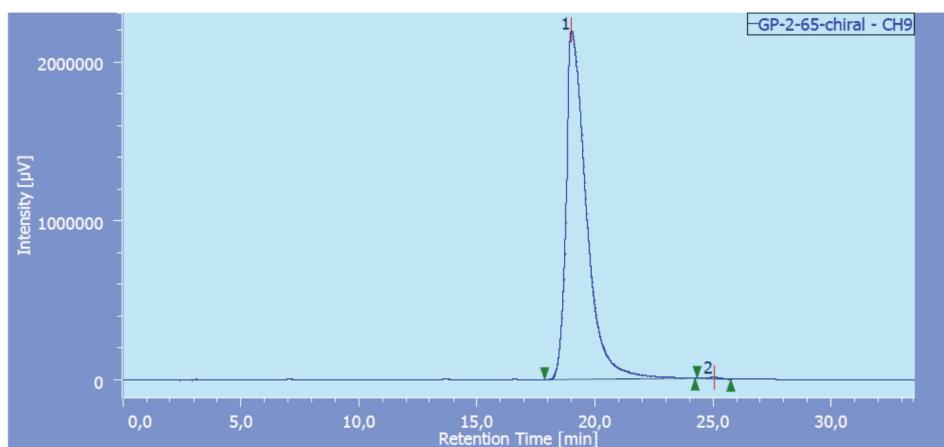
### Compound C

Center Number	Atomic Number	Atomic Type	Coordinates (Angstroms)		
			X	Y	Z
1	6	0	0.057260	0.003451	-0.035566
2	6	0	1.429668	0.012055	-0.079188
3	6	0	2.141163	1.236269	-0.041211
4	6	0	1.393715	2.441642	0.042353
5	6	0	0.009202	2.408904	0.085041
6	6	0	-0.665777	1.199456	0.046748
7	1	0	4.127114	0.422491	-0.100536
8	1	0	-0.473457	-0.938761	-0.062514
9	1	0	1.984289	-0.915206	-0.136925
10	6	0	3.526047	1.320084	-0.072764
11	6	0	2.133370	3.691642	0.071185
12	1	0	-0.542199	3.339017	0.150043
13	1	0	-1.745735	1.177509	0.081464
14	6	0	4.231087	2.630624	-0.016831
15	1	0	1.550210	4.610689	0.119950
16	1	0	4.863410	2.729551	-0.910521
17	7	0	3.397104	3.821505	0.031418
18	6	0	5.196566	2.650108	1.203107
19	8	0	5.087856	1.800099	2.065769
20	7	0	6.137287	3.625822	1.238644
21	6	0	6.348787	4.616327	0.203435
22	1	0	6.639353	5.556560	0.673402
23	1	0	7.151910	4.317204	-0.477238
24	1	0	5.434610	4.799331	-0.350369
25	6	0	7.090675	3.641456	2.328308
26	1	0	7.044294	4.596047	2.856556
27	1	0	6.855116	2.835978	3.015423
28	1	0	8.106194	3.506521	1.947171

## Supporting Information

### 8. HPLC Data for (*S*)-Dimethyl 2-(4-phenanthridine-6-carboxamido) succinate (9c)

Chromatogram



Chromatogram Information

User Name Administrator  
Date Modified 05/10/2018 15:53:15  
Description  
HPLC System Name Orga  
Injection Date 05/10/2018 15:19:40  
Volume 20.00 [ $\mu$ L]  
Sample Number 32  
Project Name Test  
Acquisition Time 59.0 [min]  
Acquisition Sequence GP-2-65-Chiral-85-Hx-15-EtOH 1mL-min 20° C IA  
Control Method 85A-Hx 15D-EtOH 60Min-1ml-min 20° C  
Peak ID Table  
Calibration Method  
Additional Information

Channel & Peak Information Table

Chromatogram Name GP-2-65-chiral-CH9

Sample Name  
Channel Name 252.5nm  
Sampling Interval 100 [msec]  
Peak Method IT (Manual)

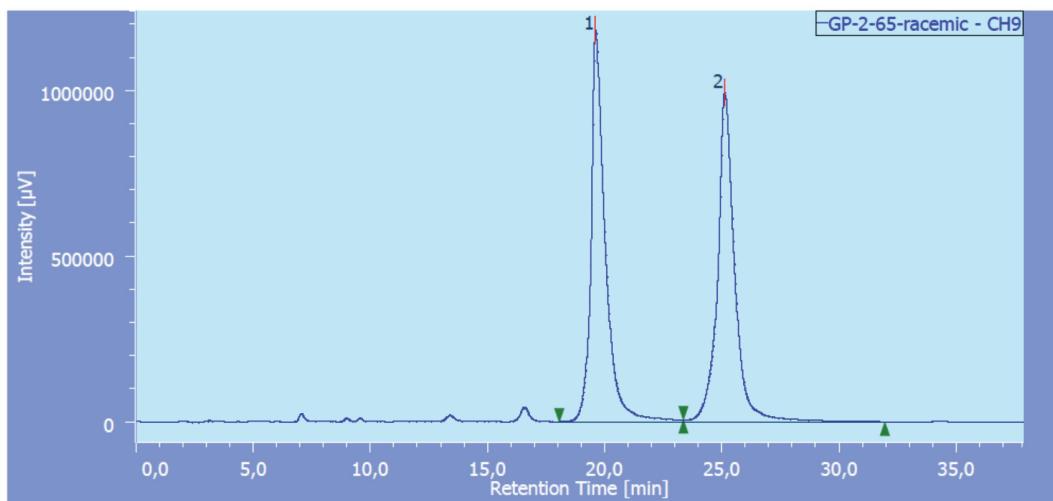
Formula  
Decision

#	Peak Name	CH	tR [min]	Area [ $\mu$ V·sec]	Height [ $\mu$ V]	Area%	Height%	Quantity	NTP	Resolution	Symmetry Factor	Warning
1	Unknown	9	18,993	131633855	2193708	99,762	99,599	N/A	2838	5,103	1,928	
2	Unknown	9	25,023	313987	8828	0,238	0,401	N/A	11256	N/A	0,945	

## Supporting Information

### HPLC Data for Dimethyl 2-(4-phenanthridine-6-carboxamido) succinate (rac-9c)

#### Chromatogram



#### Chromatogram Information

User Name Administrator  
Date Modified 05/10/2018 12:21:01  
Description  
HPLC System Name Orga  
Injection Date 05/10/2018 11:43:07  
Volume 20,00 [µL]  
Sample Number 31  
Project Name Test  
Acquisition Time 59,0 [min]  
Acquisition Sequence GP-2-65-Racemic-85-Hx-15-EtOH 1mL-min 20° C IA  
Control Method 85A-Hx 15D-EtOH 60Min-1ml-min 20° C  
Peak ID Table  
Calibration Method  
Additional Information

#### Channel & Peak Information Table

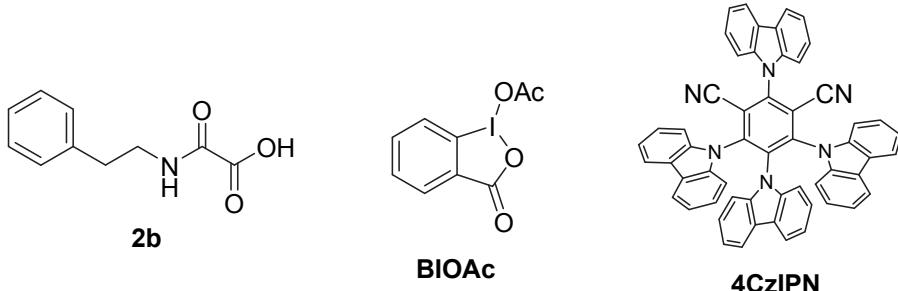
Chromatogram Name GP-2-65-racemic-CH9  
Sample Name  
Channel Name 252,5nm  
Sampling Interval 100 [msec]  
Peak Method IT (Manual)  
Formula  
Decision

#	Peak Name	CH	tR [min]	Area [µV·sec]	Height [µV]	Area%	Height%	Quantity	NTP	Resolution	Symmetry Factor	Warning
1	Unknown	9	19,608	49926380	1179801	49,688	54,319	N/A	7055	5,384	1,455	
2	Unknown	9	25,113	50554084	992200	50,312	45,681	N/A	8094	N/A	1,093	

## Supporting Information

### 9. Fluorescence quenching experiments

Fluorescence decay profiles were collected on aerated samples in dichloromethane solution at 530 nm using the single photon counting technique on a Horiba Jobin-Yvon Fluorolog 211 instrument using a 370-nm pulsed LED excitation source operated at 250 kHz and a cooled Hamamatsu 928P single photon counting photomultiplier. The data was deconvoluted from the IRF using a multi-exponential function and the goodness-of-fit judged by the  $\chi^2$  parameter, randomness of the residuals, and Durbin-Watson (D-W) test statistic. 4-CzIPN exhibited a bi-exponential decay in which the short component is assigned to the decay of the singlet excited state. The longer component is instead attributed to delayed fluorescence. The decay rates are only slightly shortened in the presence of oxamic acid **2b** or BIOAc (100 mM concentration) (**Figures 1-2**), indicating that they do not quench the catalyst in its excited singlet or triplet state. Instead, a 1:1 mixture of **2b** and BIOAc quenches more efficiently both the excited singlet and triplet state of 4-CzIPN (**Figures 1-4**). The rate constants can be calculated according to  $k_{\text{obs}} = \tau^{-1} - \tau_0^{-1}$ , where  $\tau_0$  and  $\tau$  are the decay parameters in the absence and presence of quencher, respectively (Table 1).



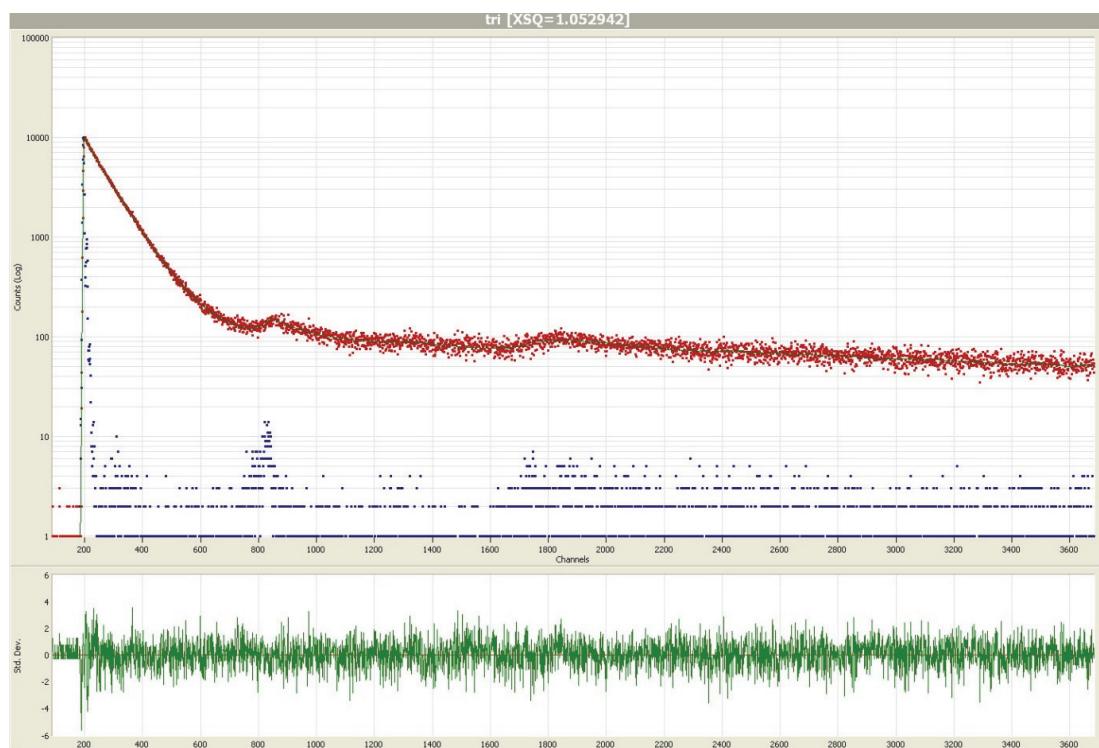
**Table 1.** Decay parameters and calculated quenching rates for 4-CzIPN in aerated DCM solutions.

Quencher <sup>a</sup>	$\tau_1$ (ns)	$\tau_2$ (ns)	$\chi^2$	$k_{\text{obs}}(\text{S}_1)$ (s <sup>-1</sup> )	$k_{\text{obs}}(\text{T}_1)$ (s <sup>-1</sup> )
None	20.4	742	1.19		
<b>2b</b>	19.7	634	1.05	$1.8 \times 10^6$	$2.3 \times 10^5$
BIOAc	19.3	647	1.22	$2.9 \times 10^6$	$2.0 \times 10^5$
<b>2b + BIOAc</b>	16.9	101	1.21	$1.02 \times 10^7$	$8.6 \times 10^6$

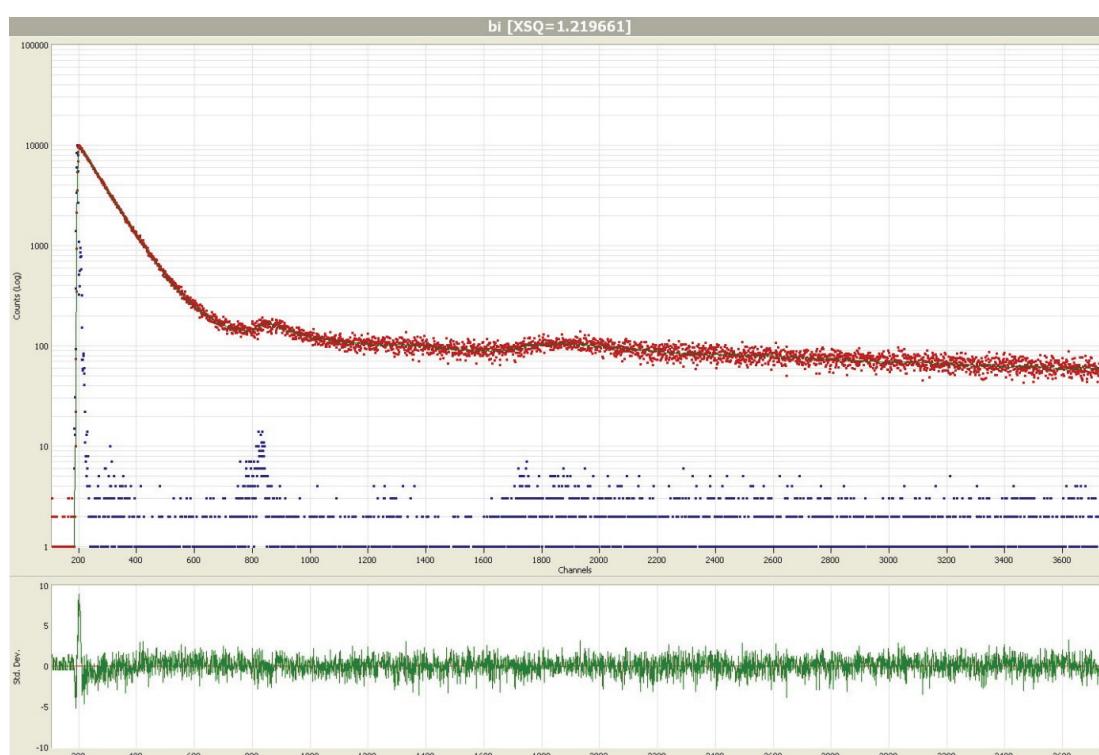
<sup>a</sup> 100m M concentration

## Supporting Information

**Figure 1.** Fluorescence decay of 4-CzIPN in the presence of oxamic acid **2b**.

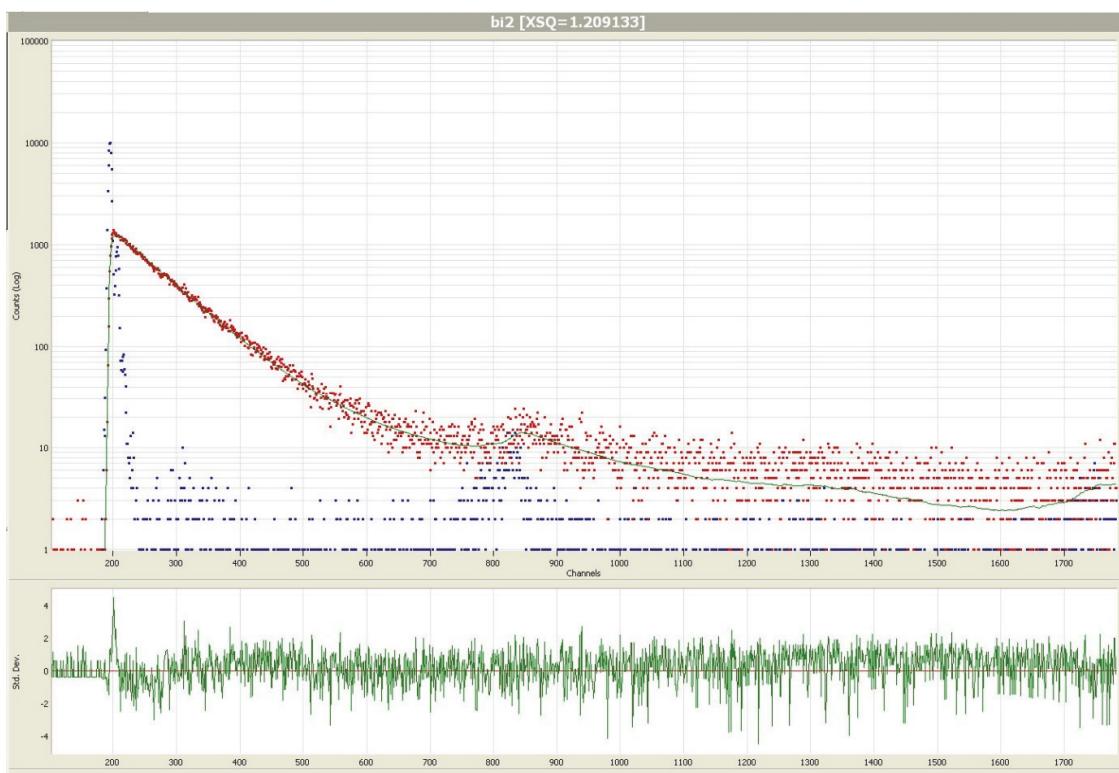


**Figure 2.** Fluorescence decay of 4-CzIPN in the presence of BIOAc.

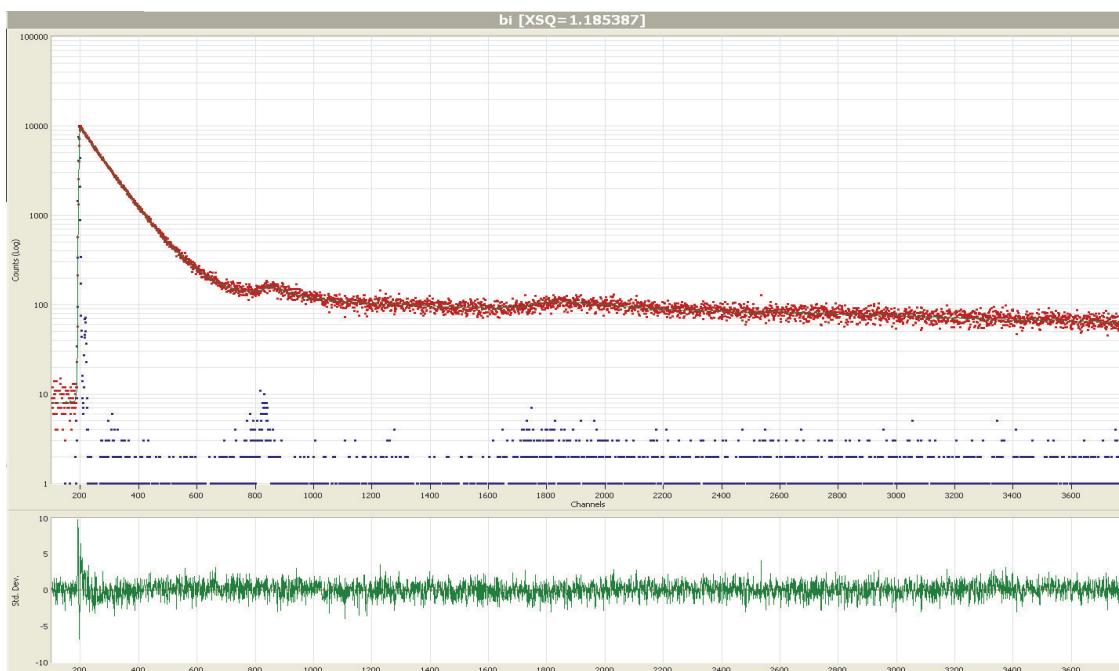


## Supporting Information

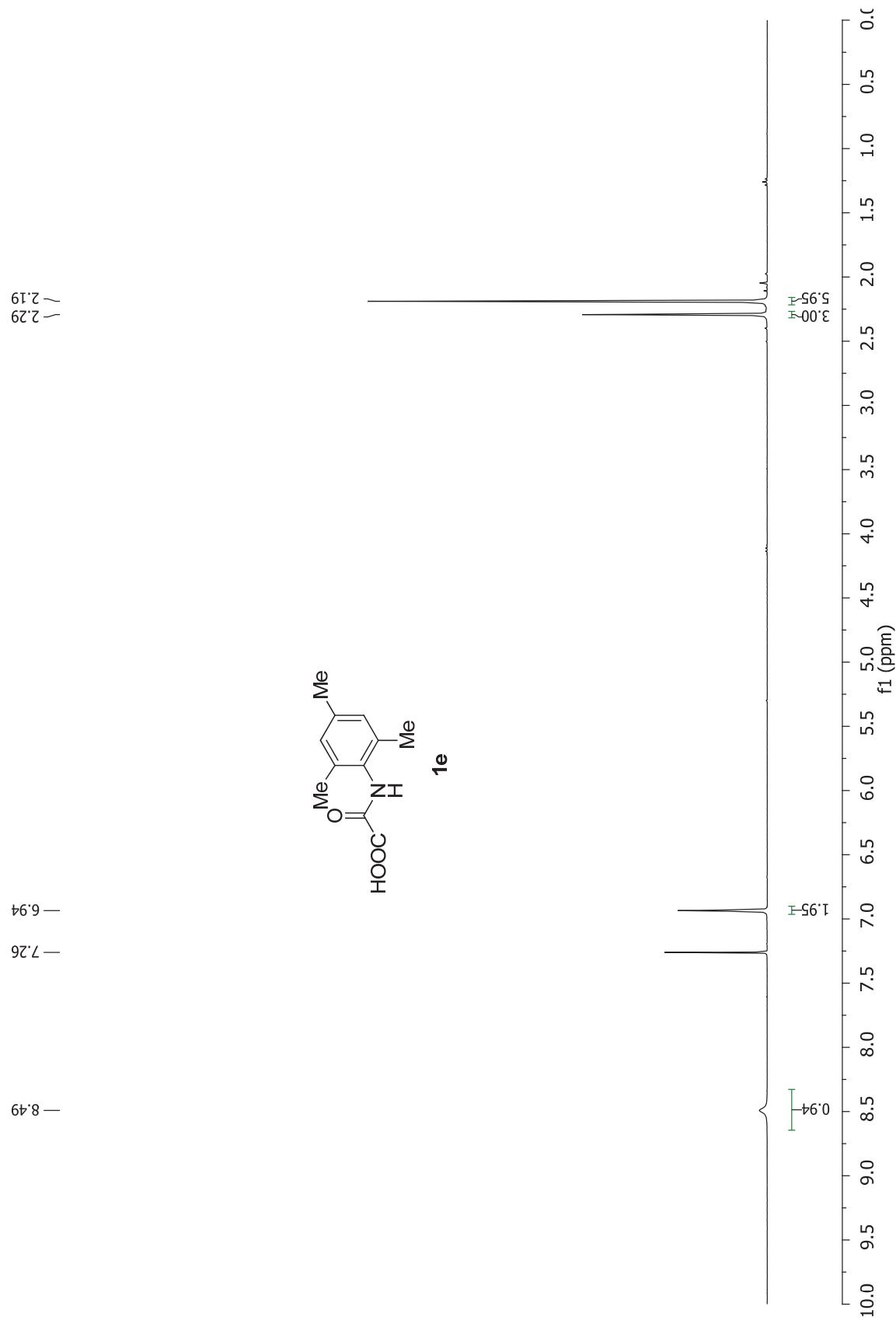
**Figure 3.** Fluorescence decay of 4-CzIPN in the presence of oxamic acid **2b** and BIOAc.



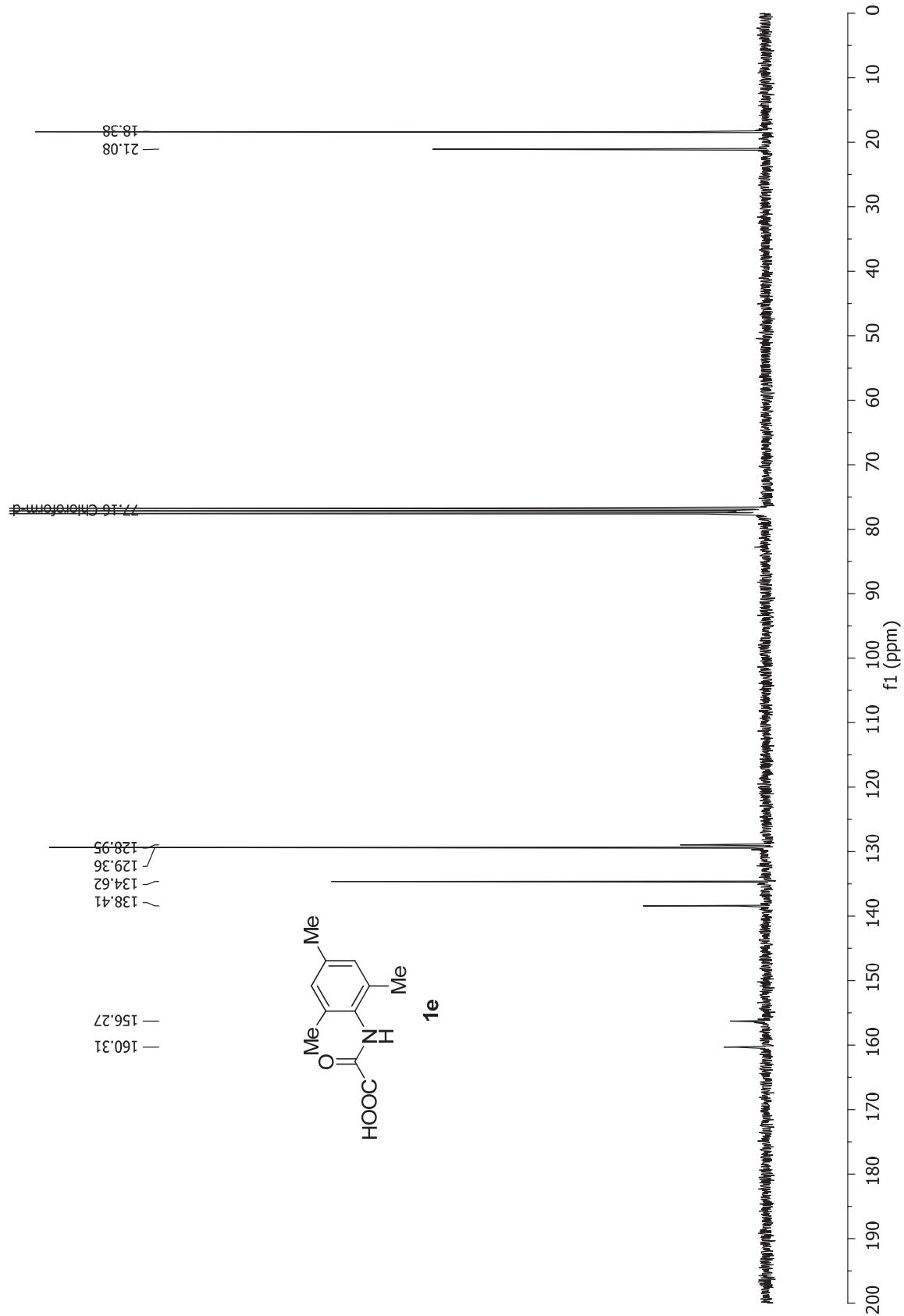
**Figure 4.** Fluorescence decay of 4-CzIPN.



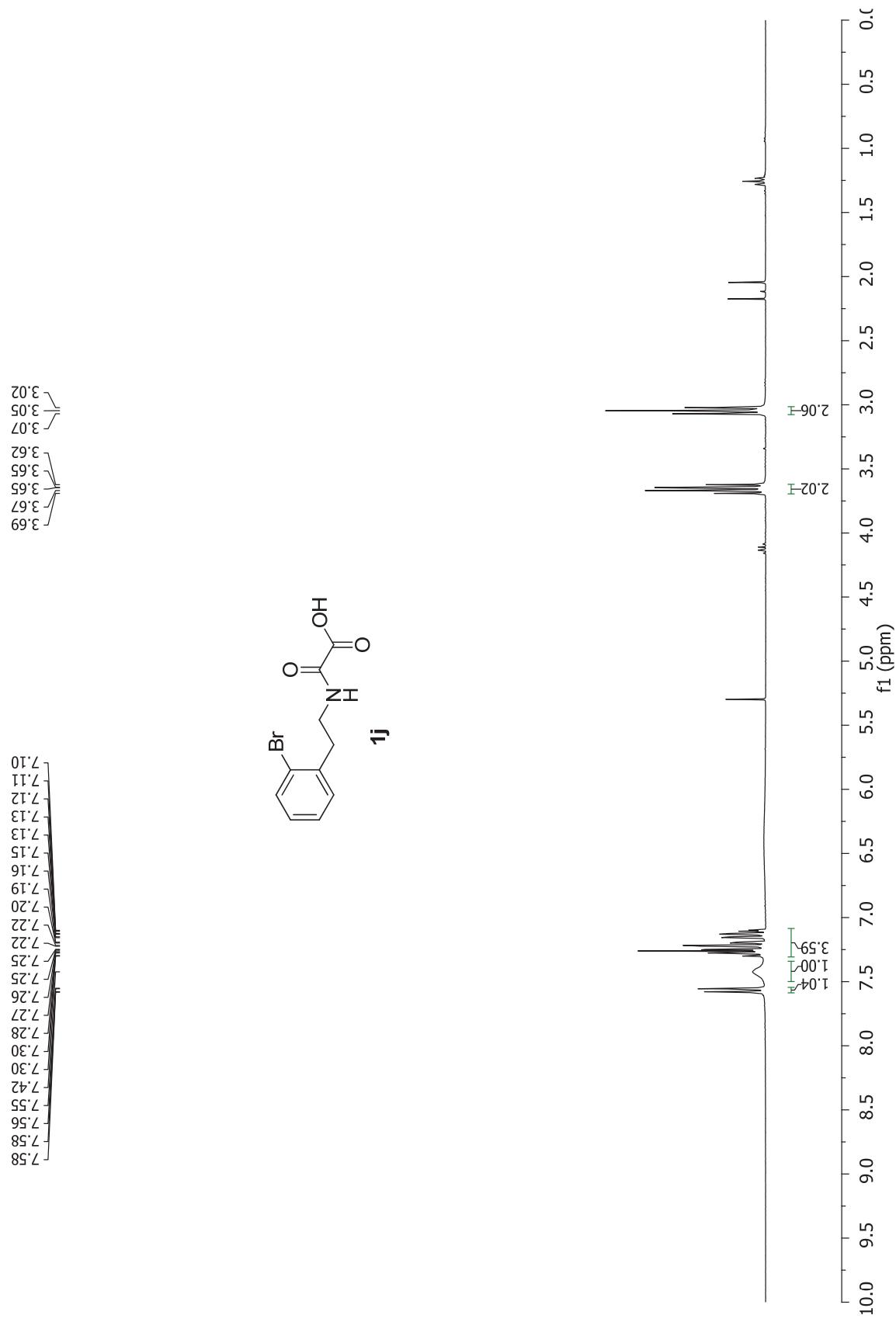
**Supporting Information**



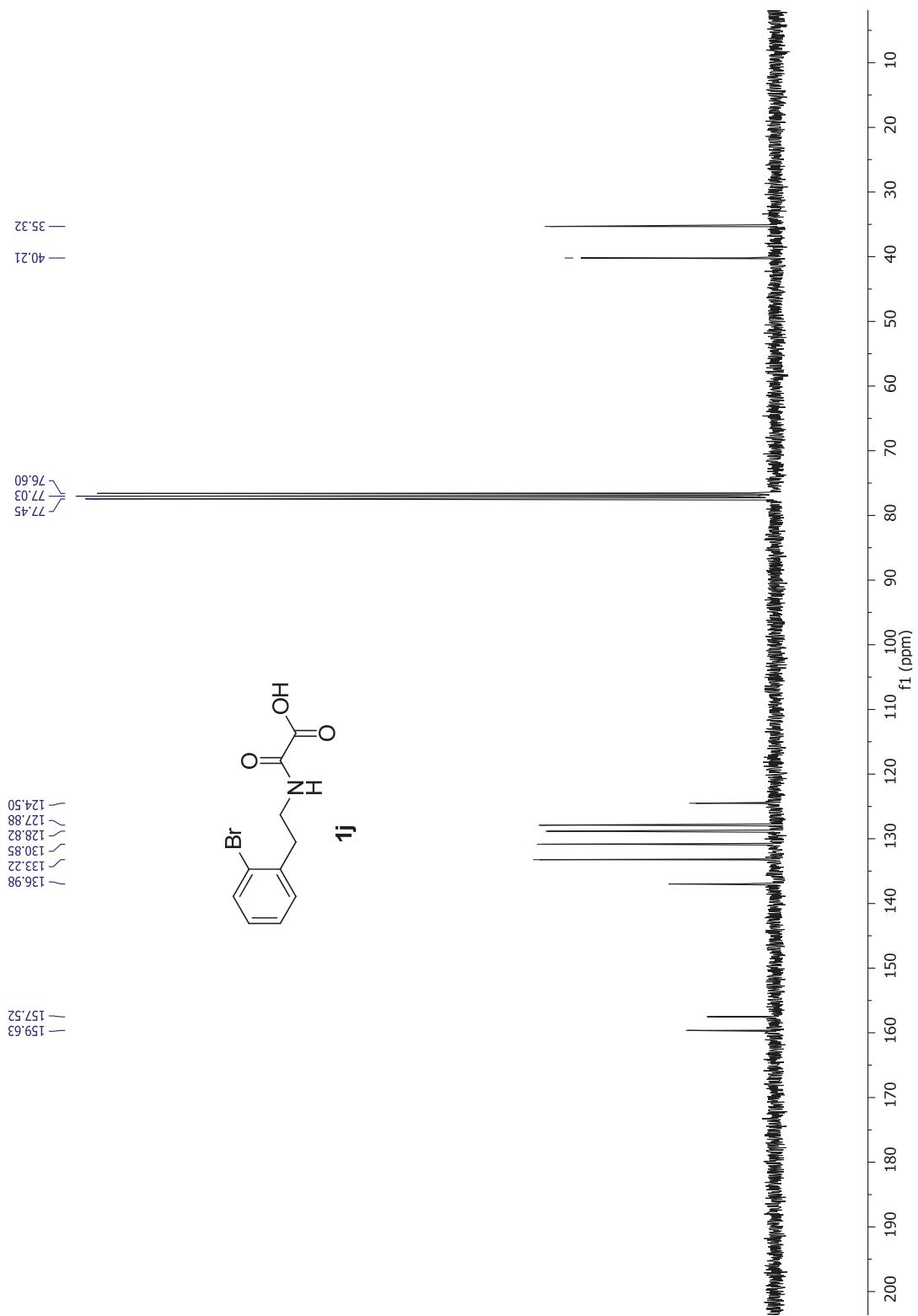
**Supporting Information**



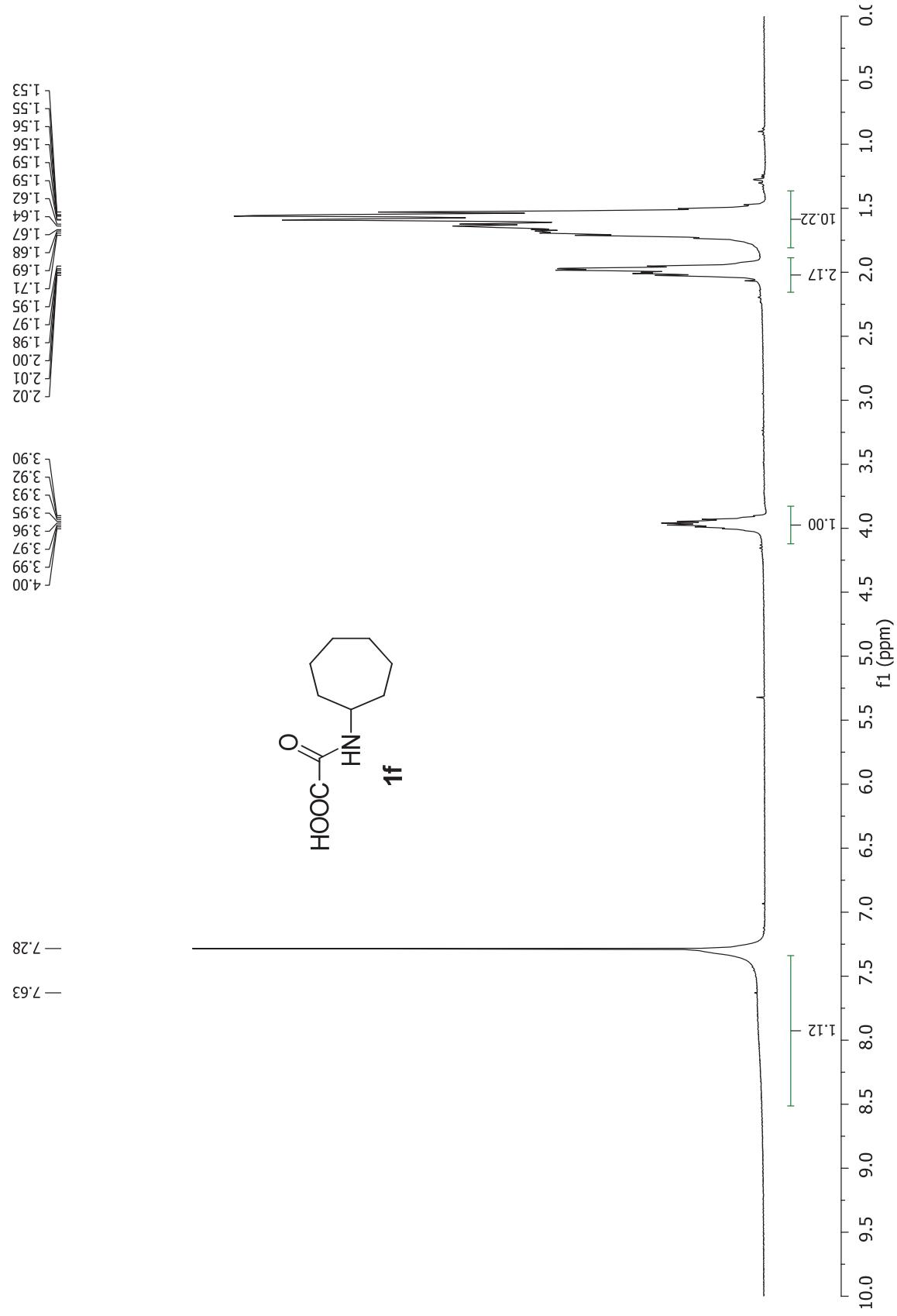
**Supporting Information**



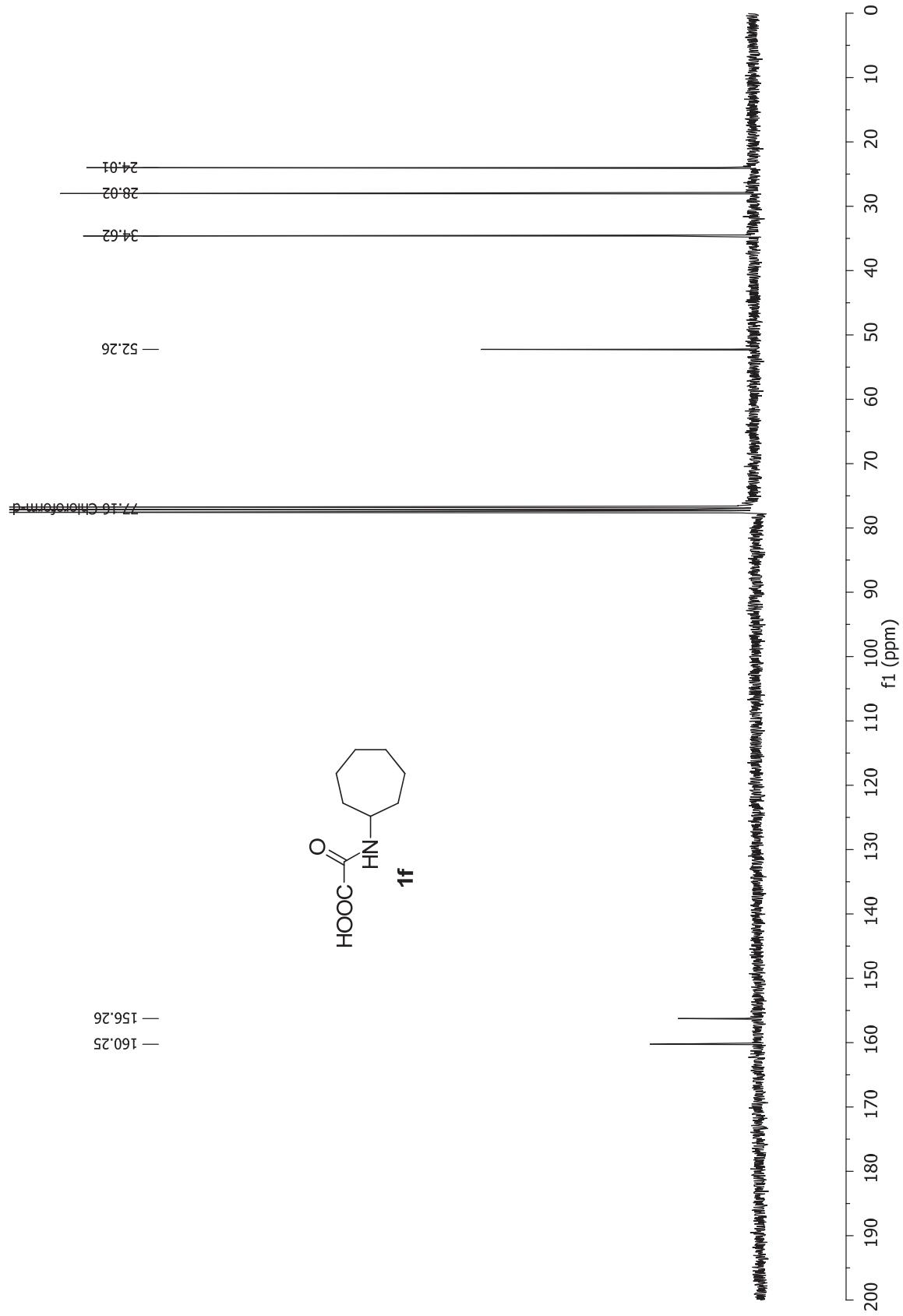
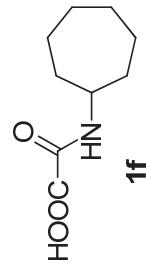
## Supporting Information



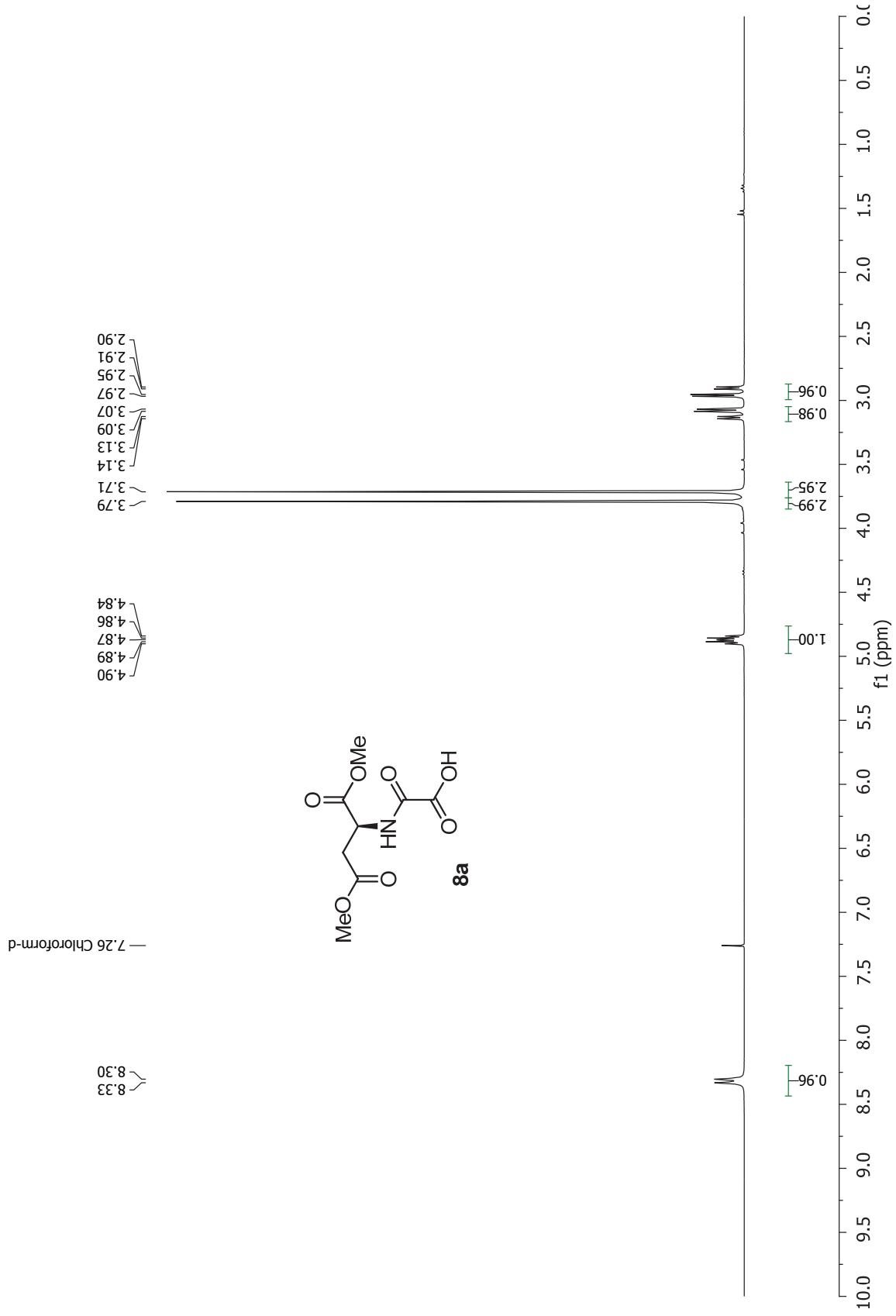
**Supporting Information**



## Supporting Information

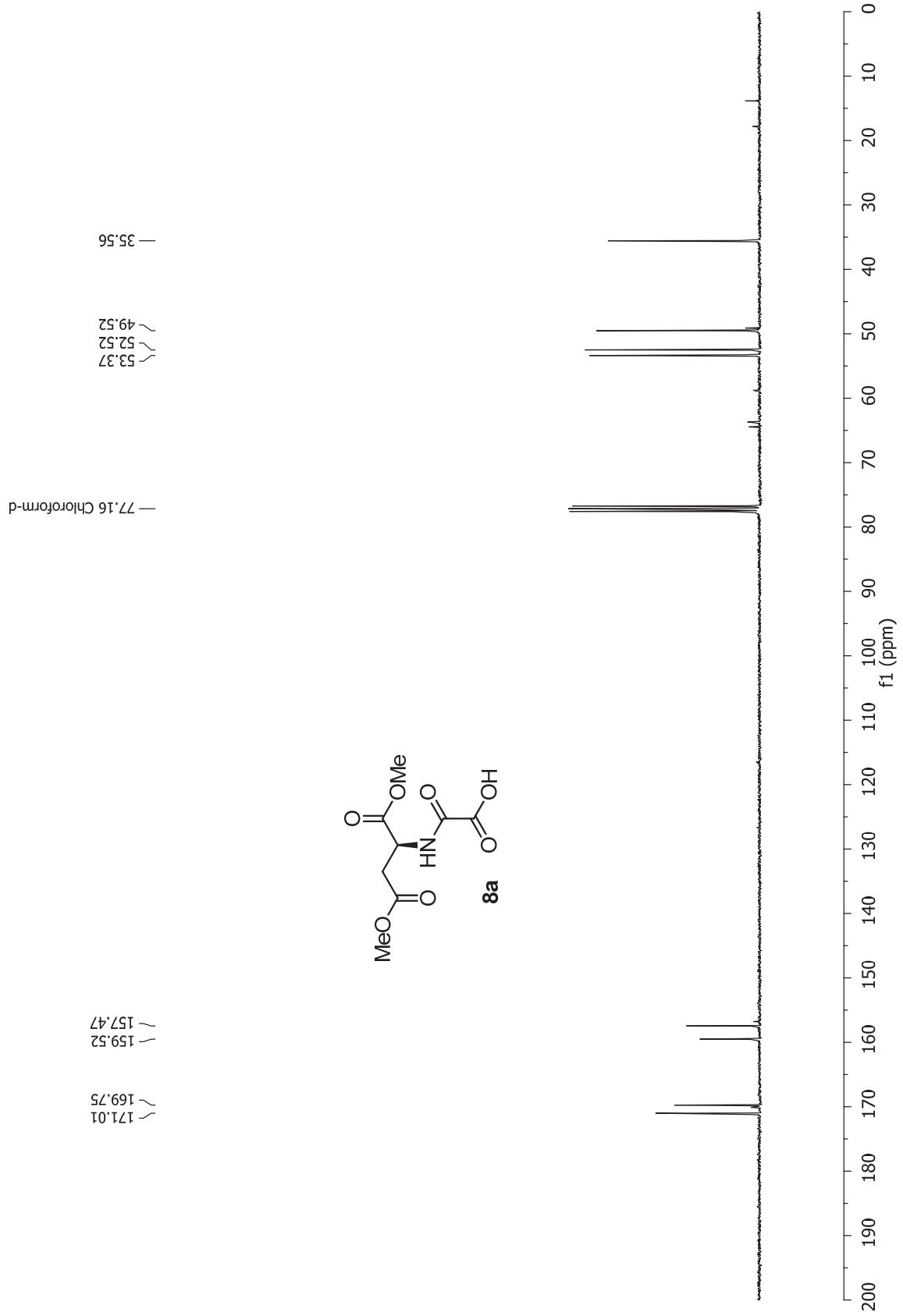


## Supporting Information

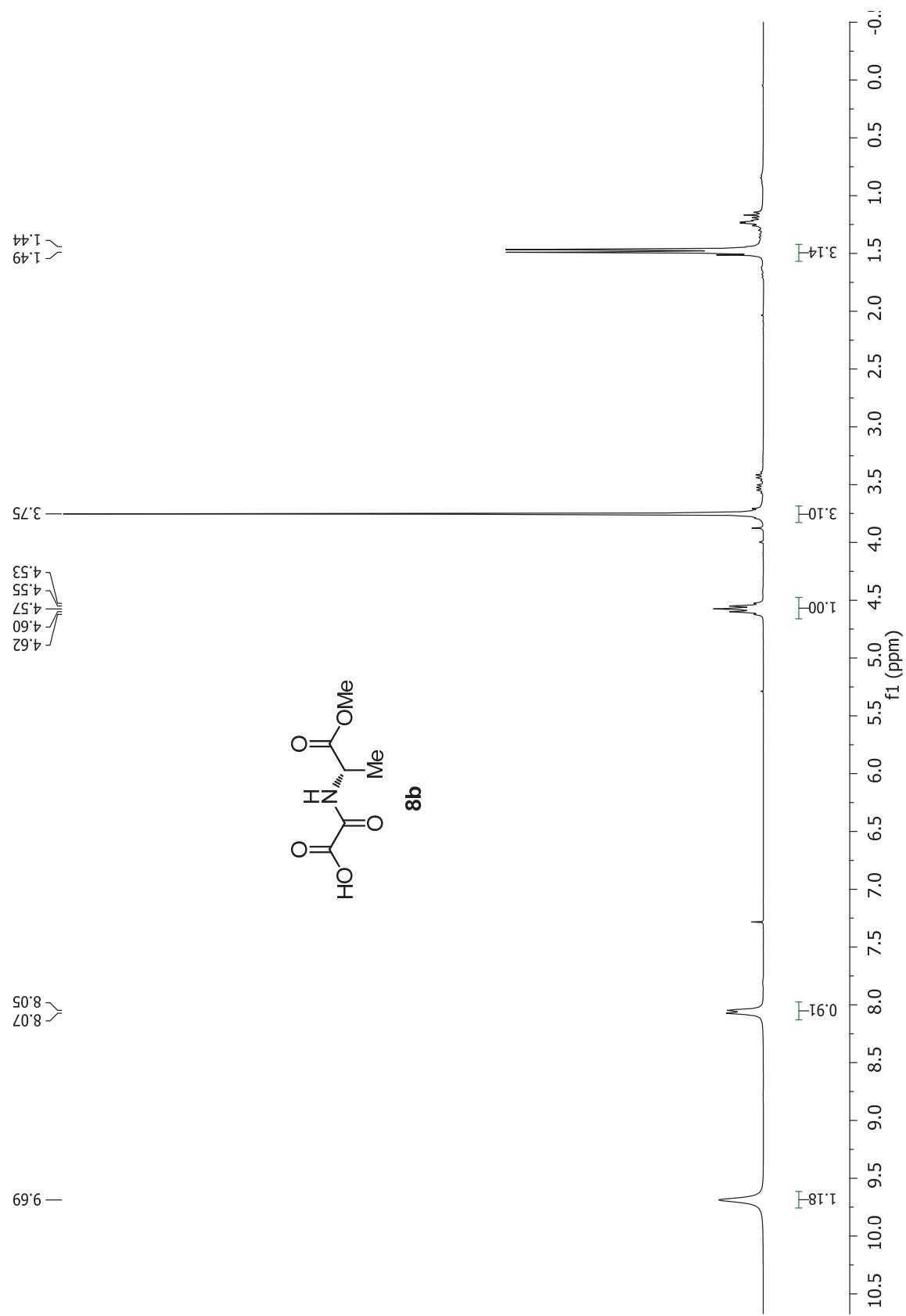


S37

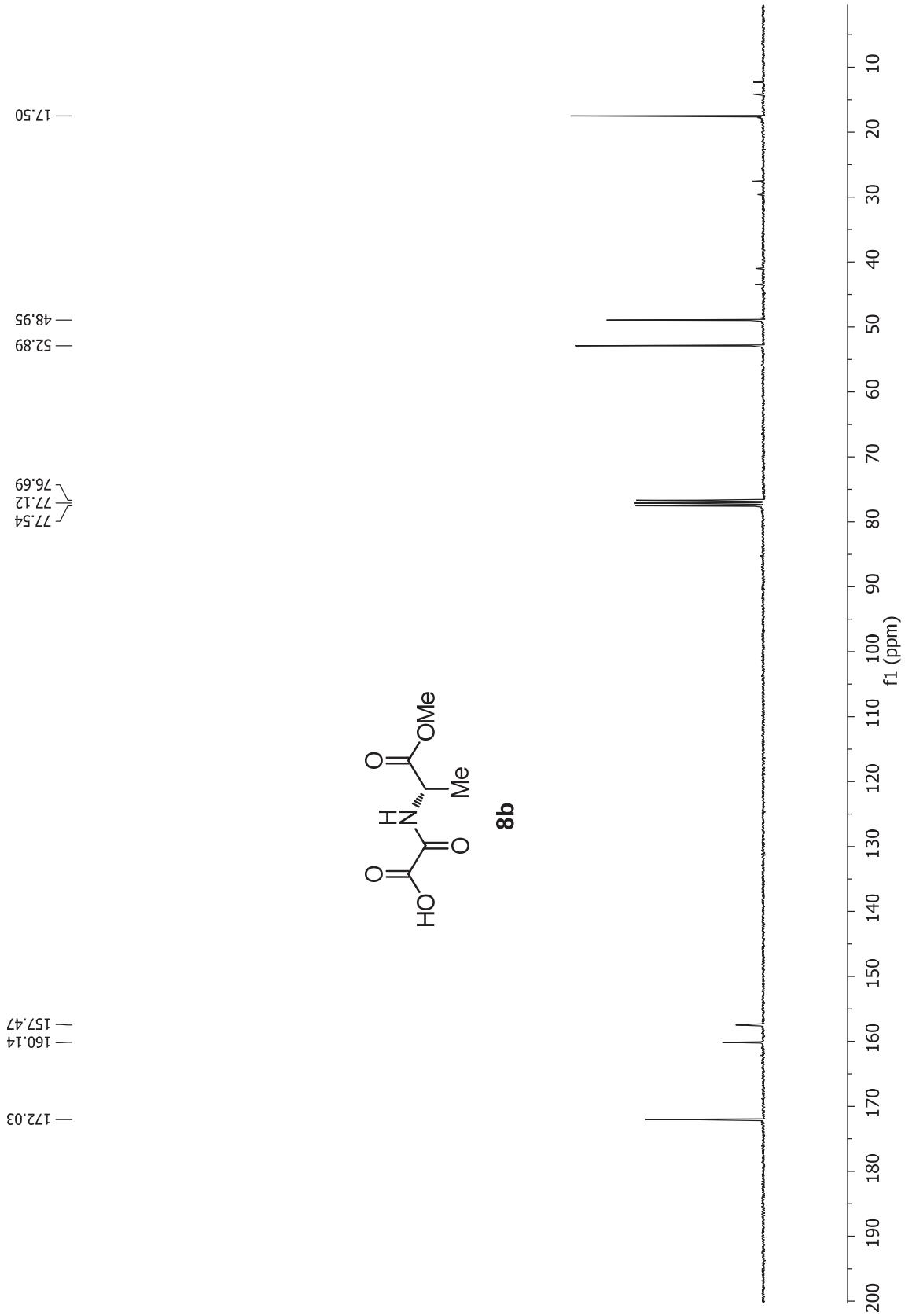
**Supporting Information**



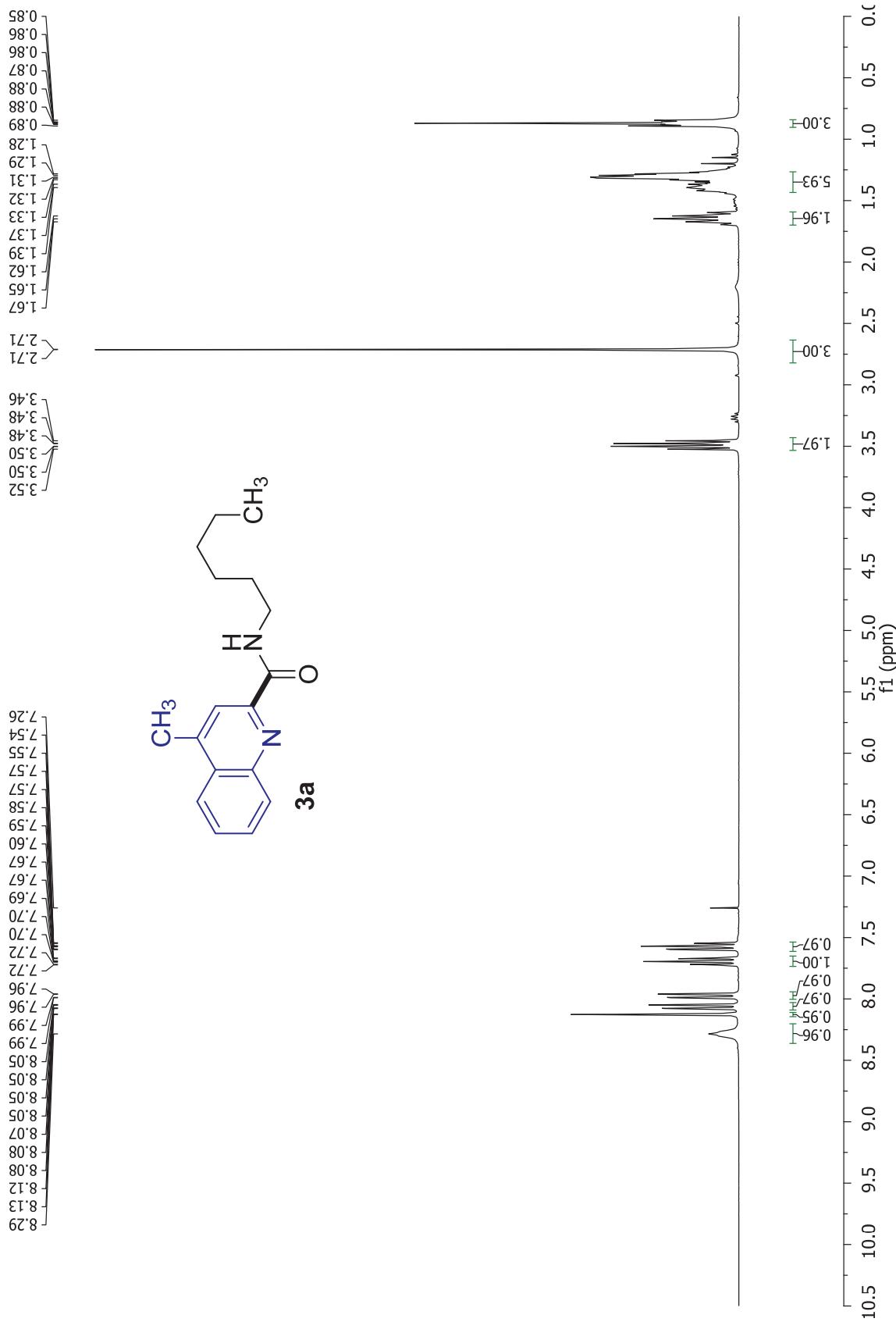
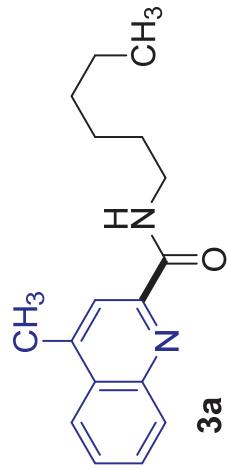
**Supporting Information**



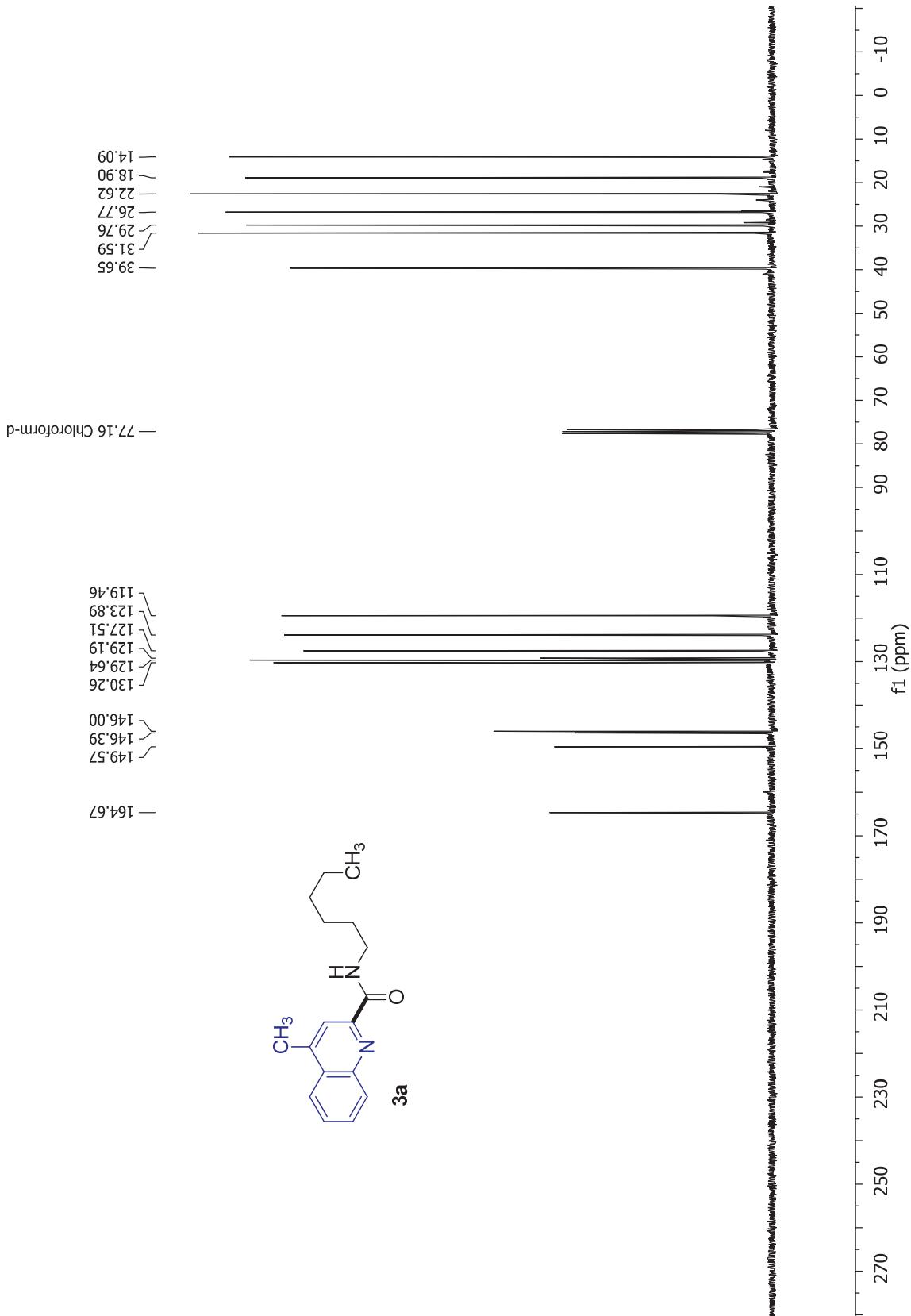
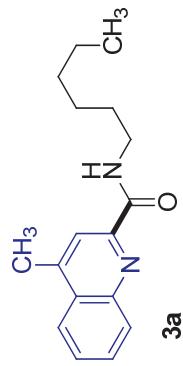
**Supporting Information**



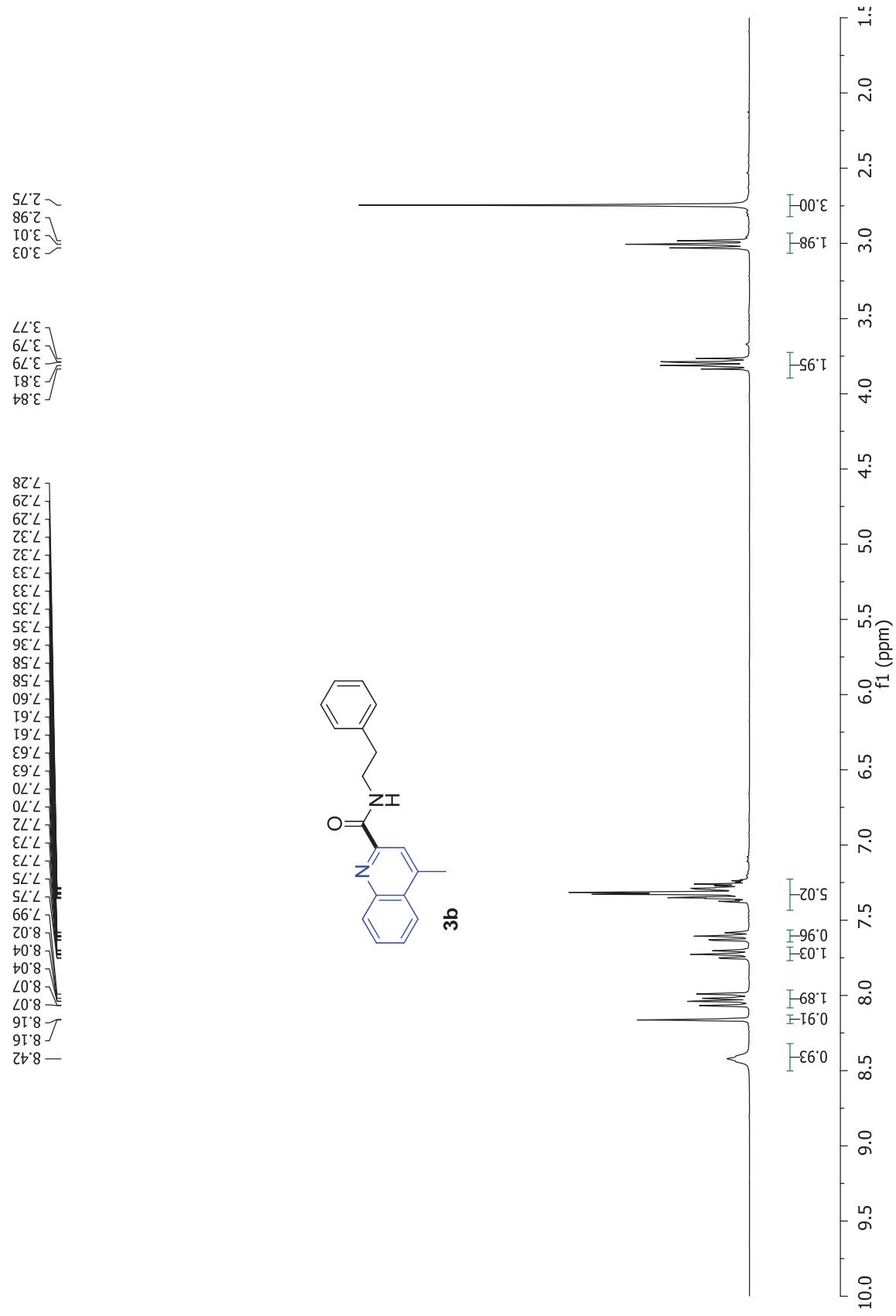
## Supporting Information



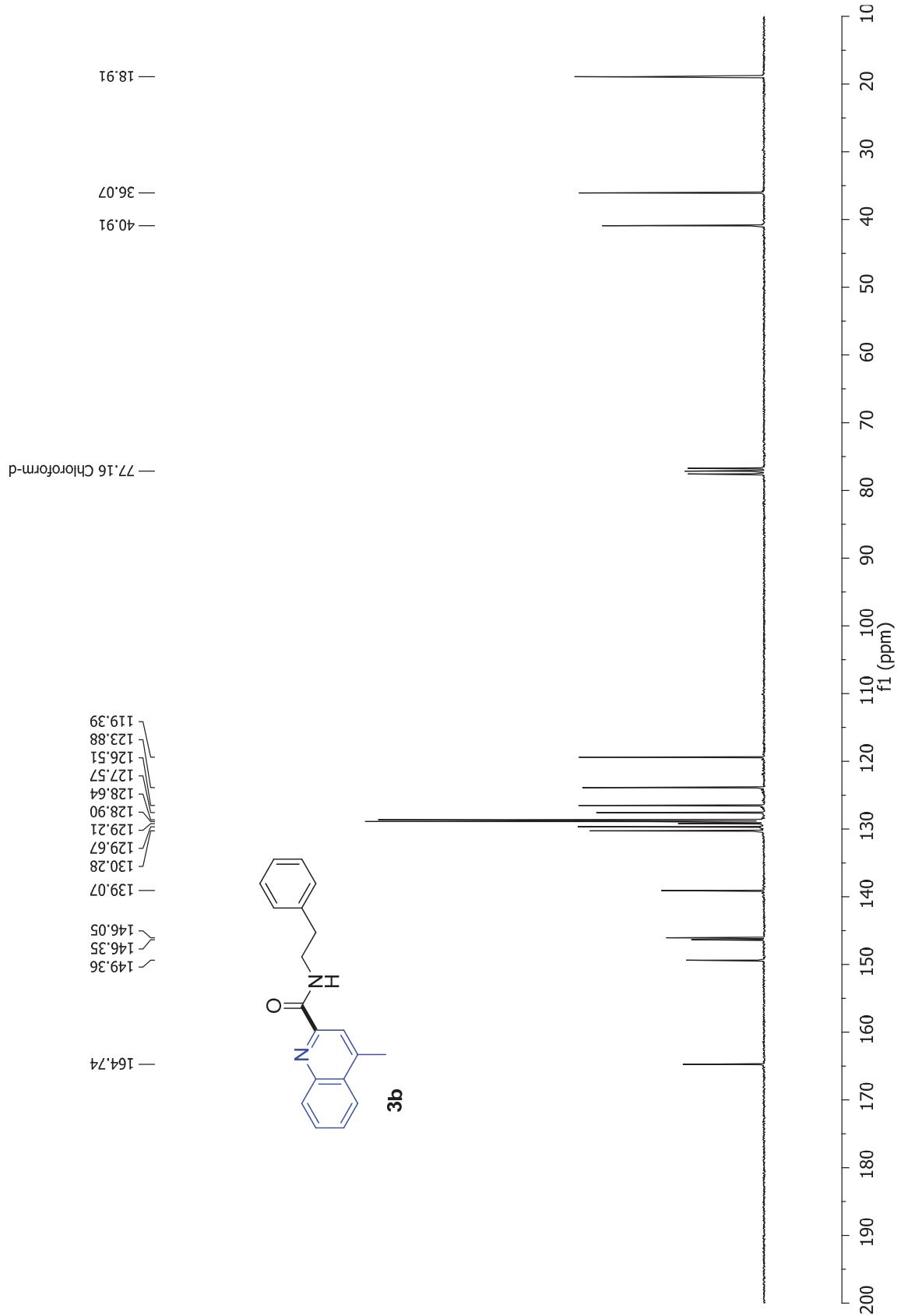
## Supporting Information



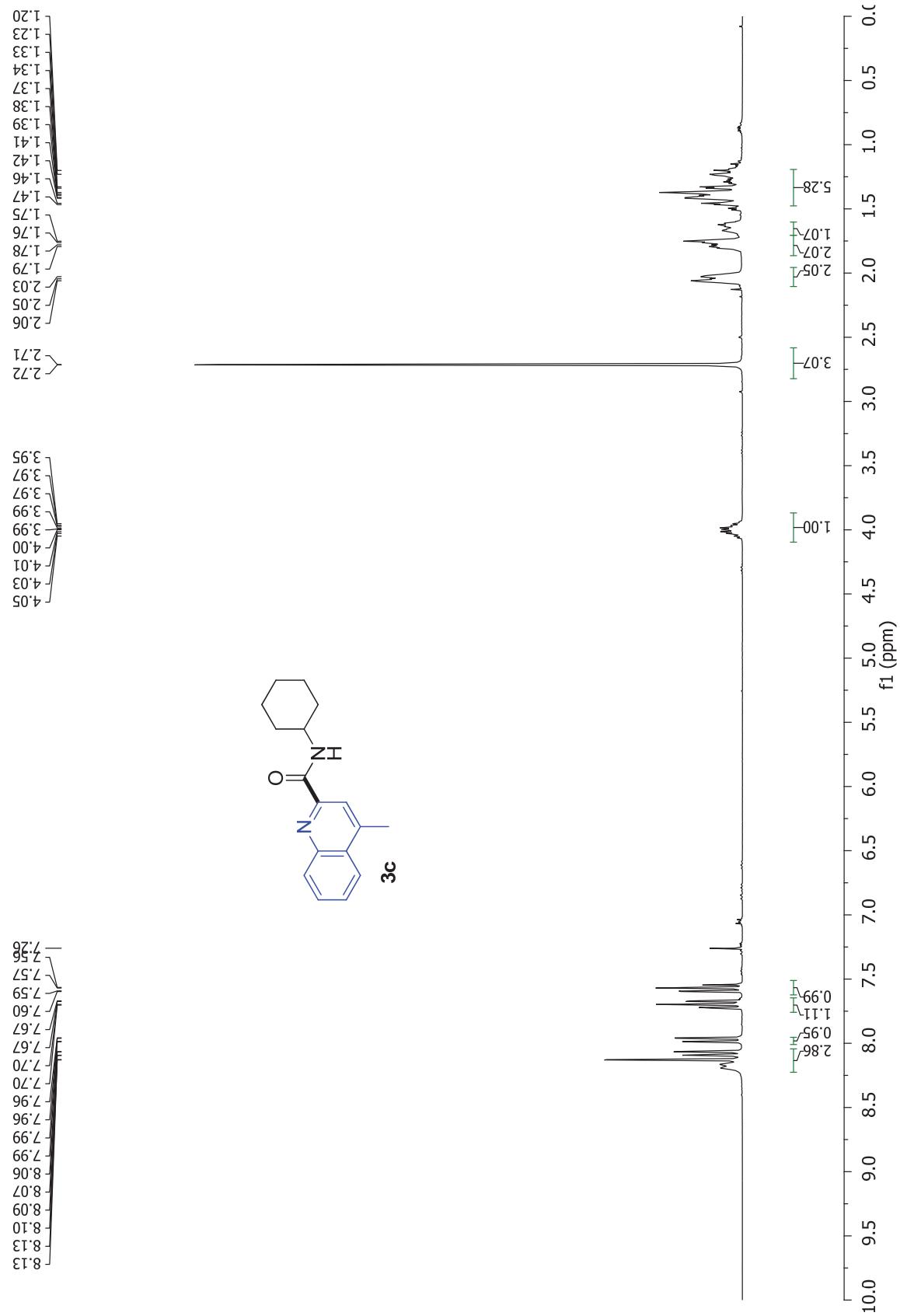
**Supporting Information**



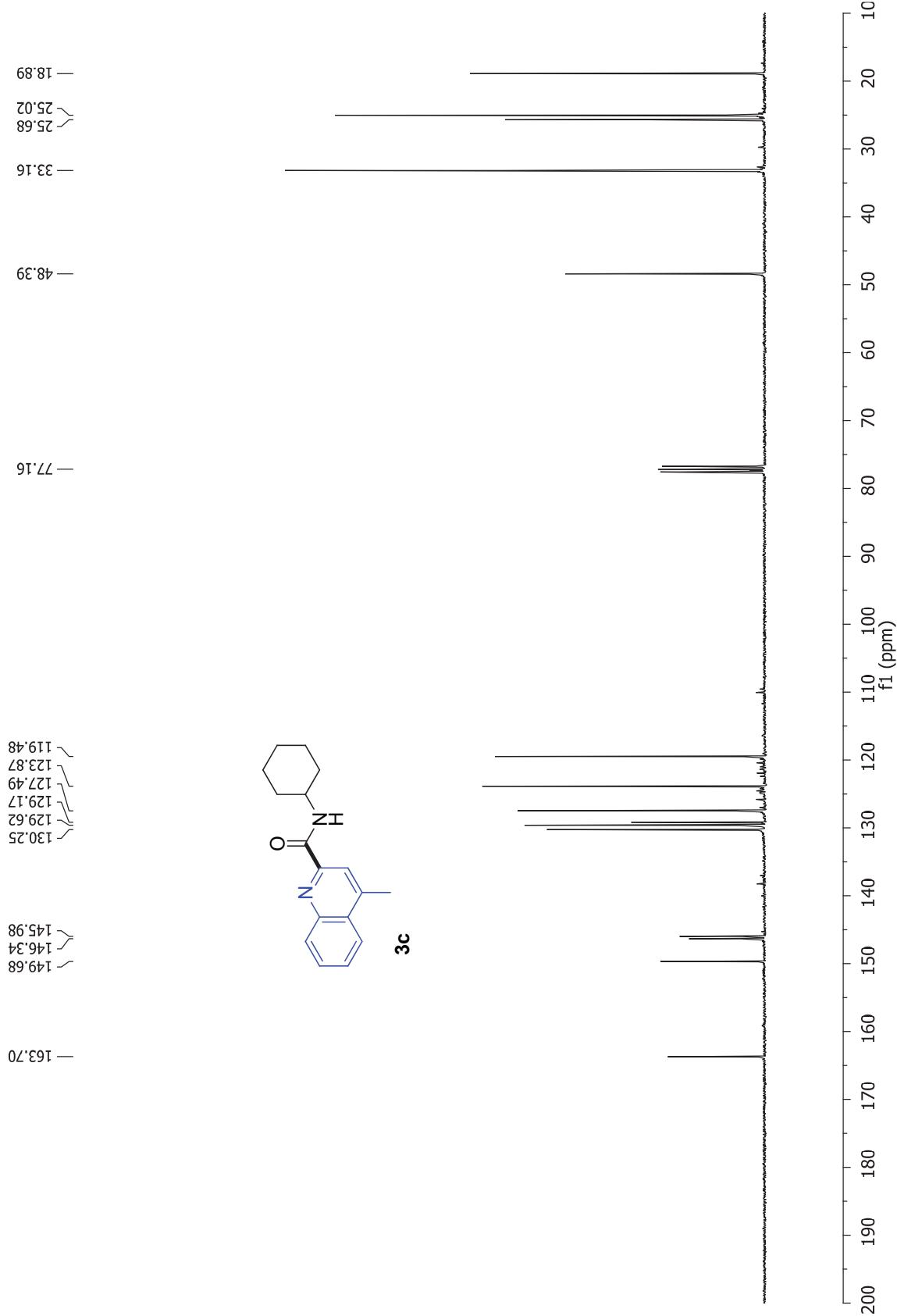
## Supporting Information



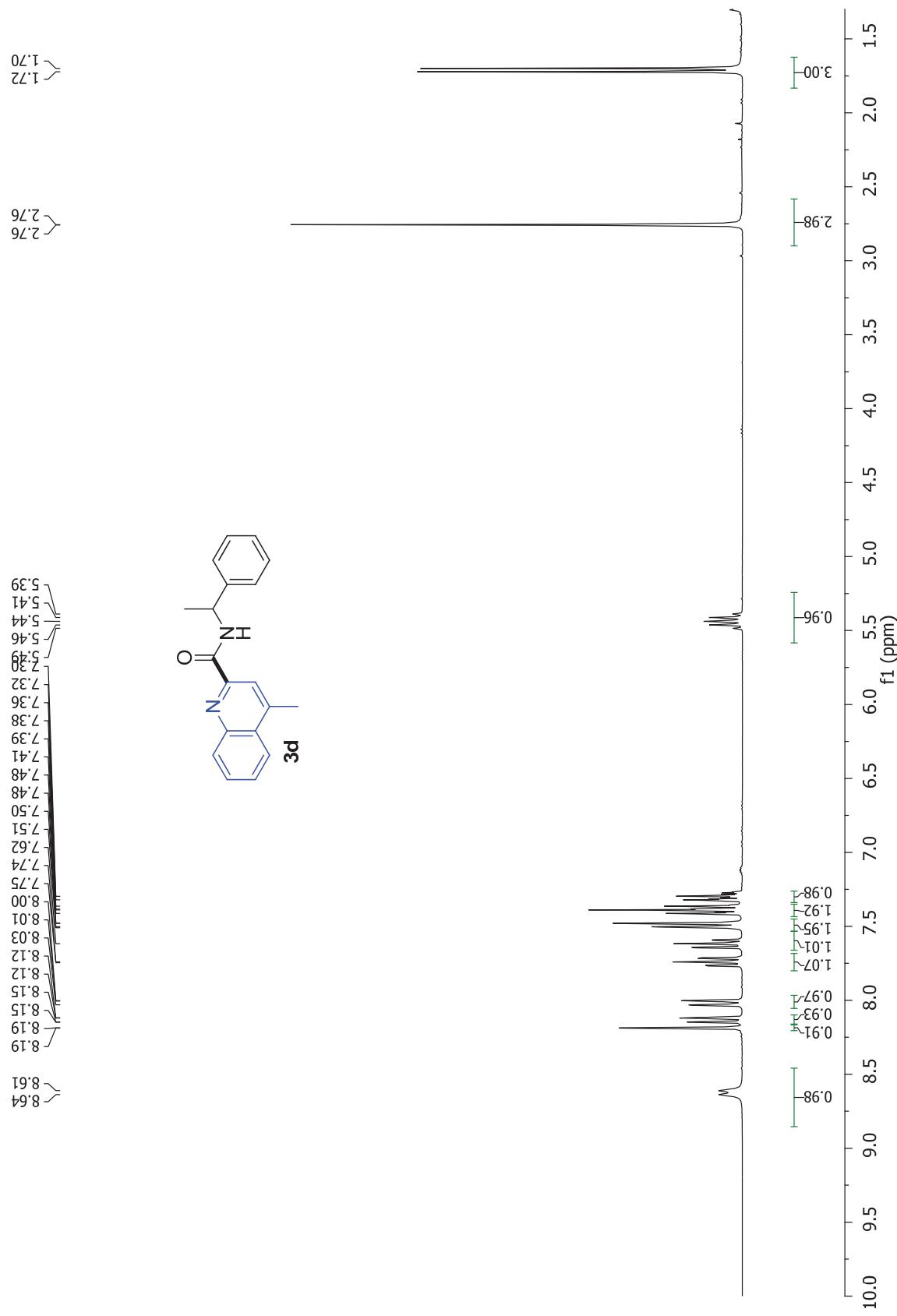
**Supporting Information**



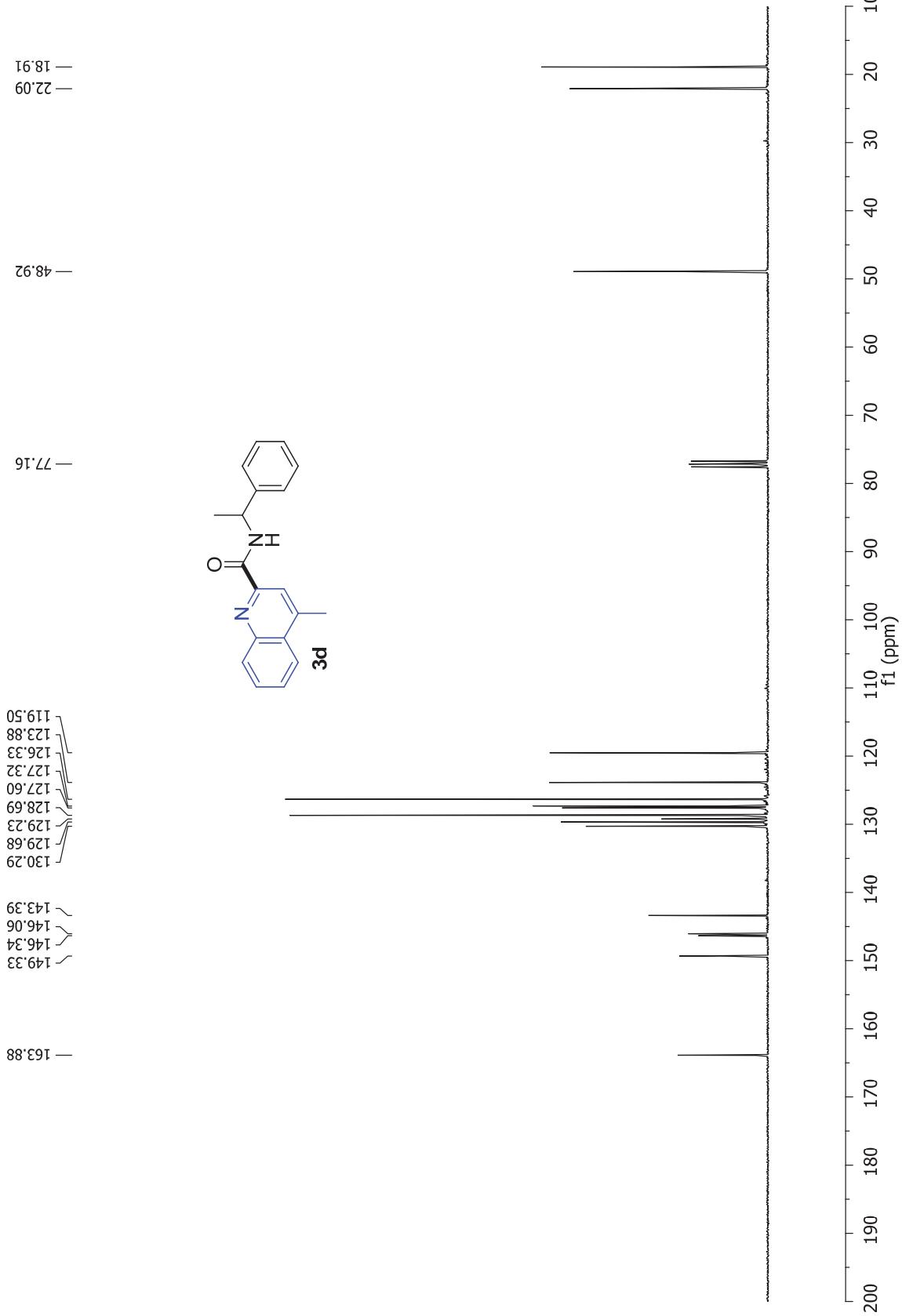
**Supporting Information**



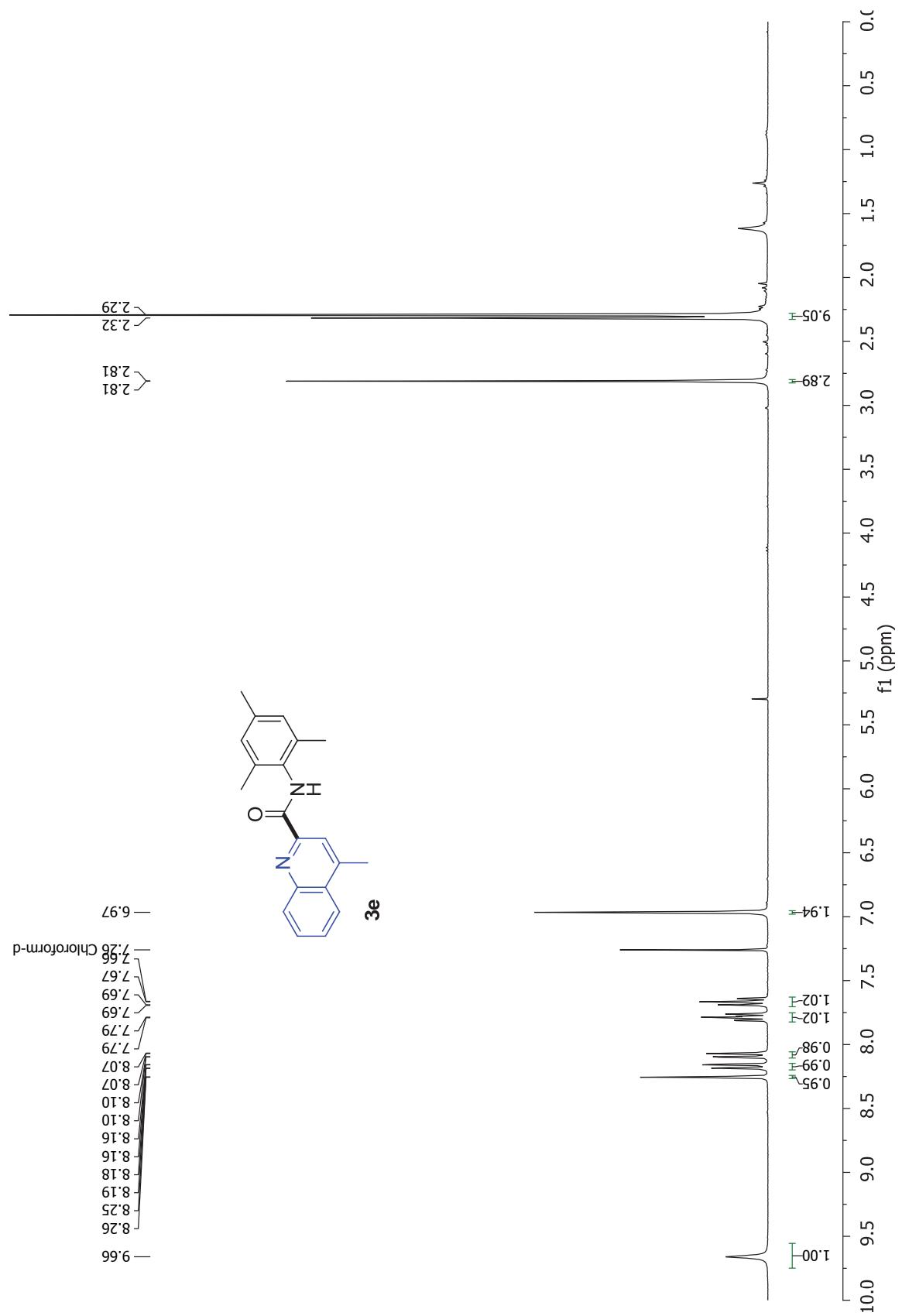
**Supporting Information**



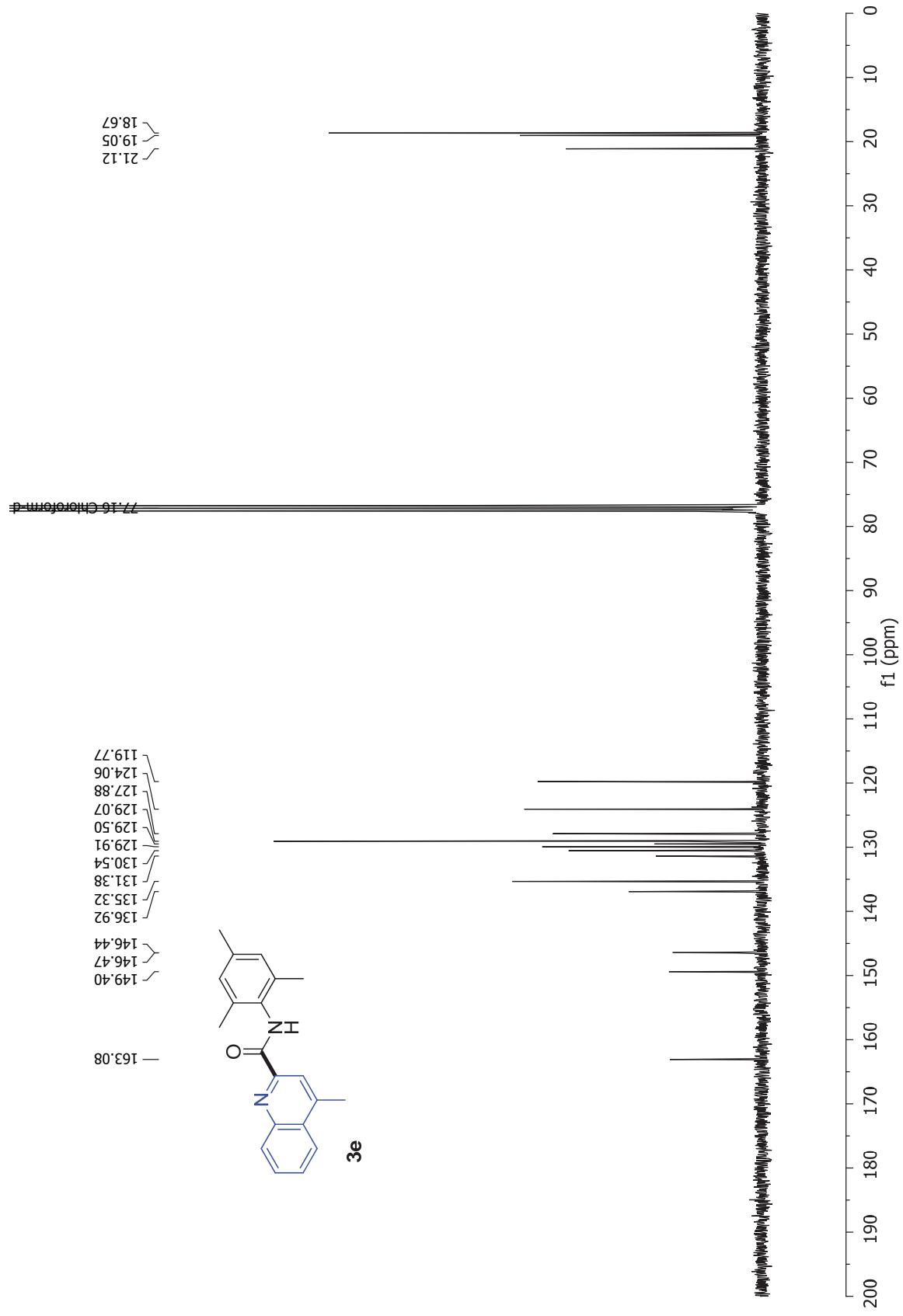
## Supporting Information



**Supporting Information**

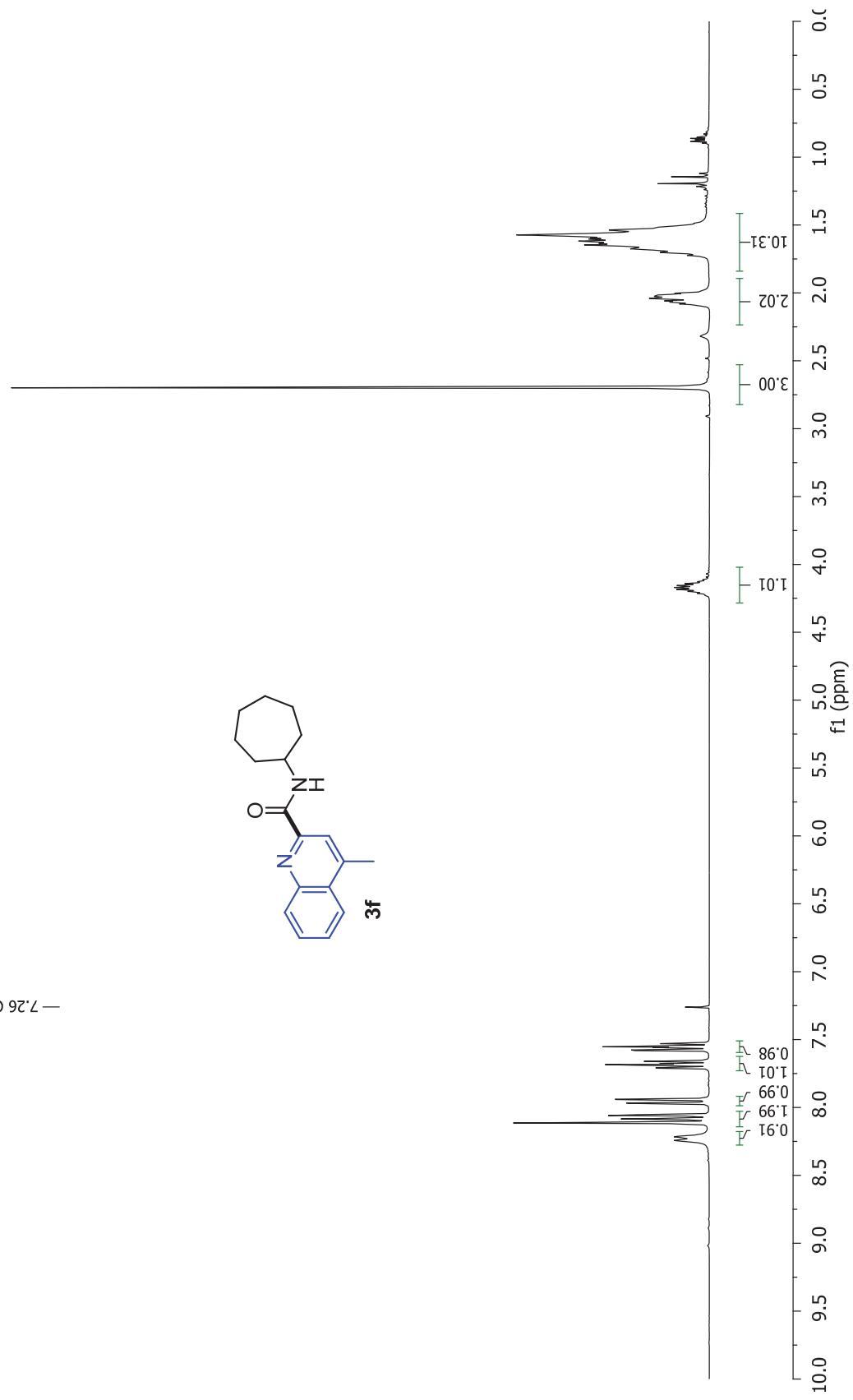
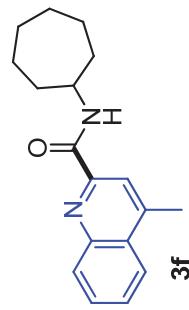


## Supporting Information

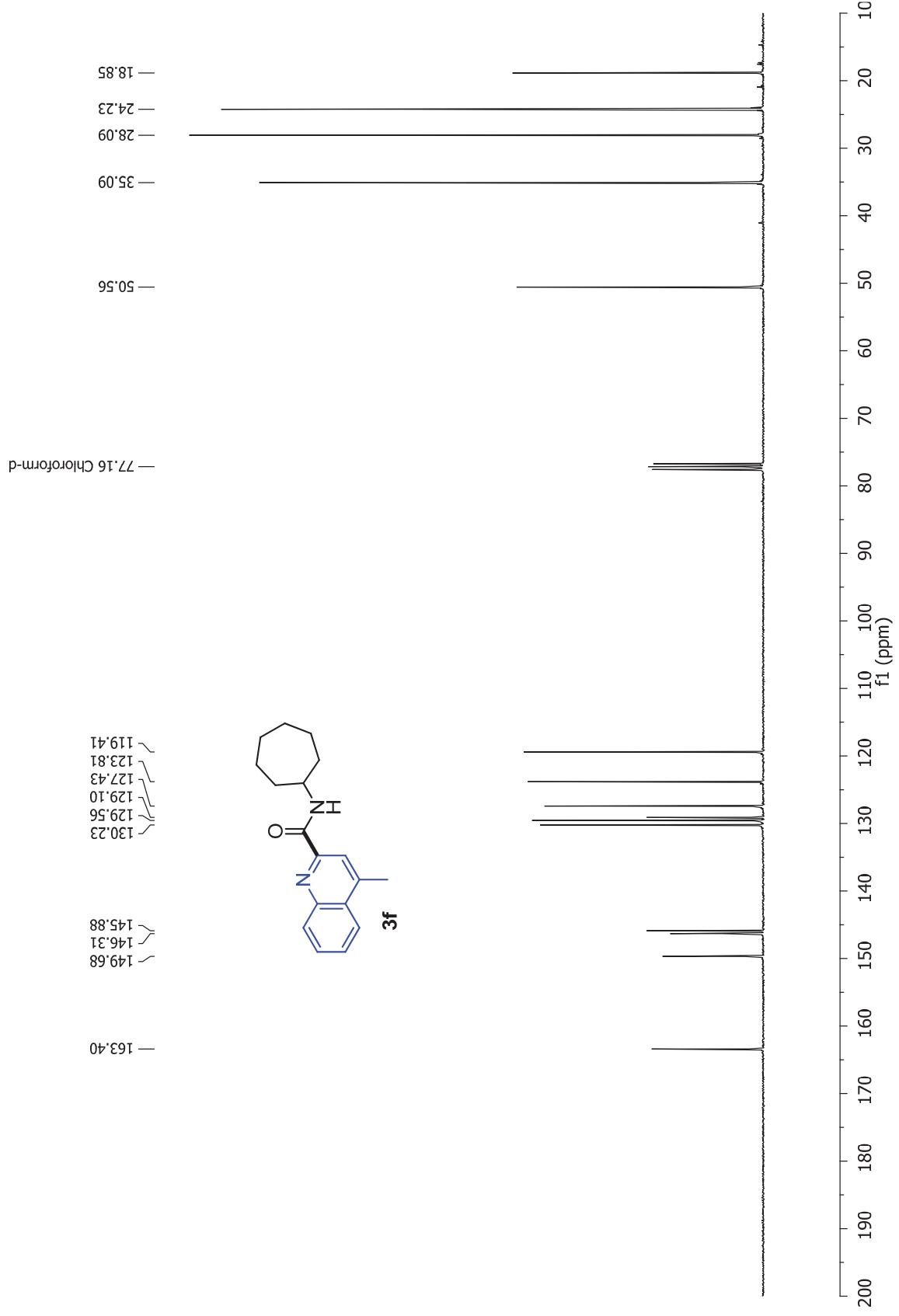


**Supporting Information**

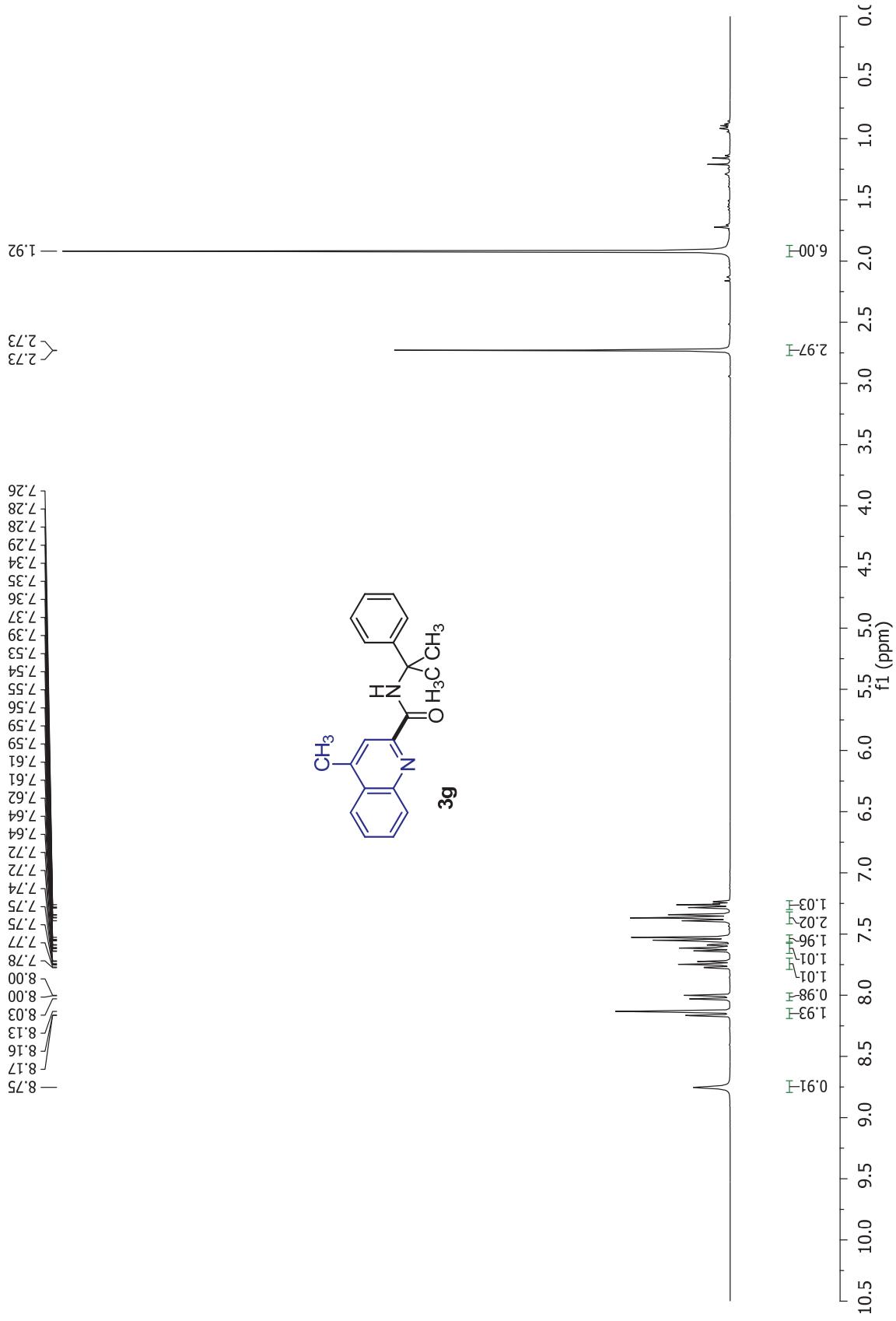
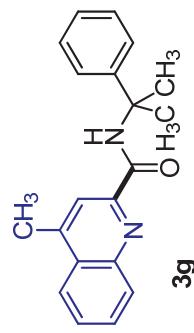
— 7.26 Chloroform-d



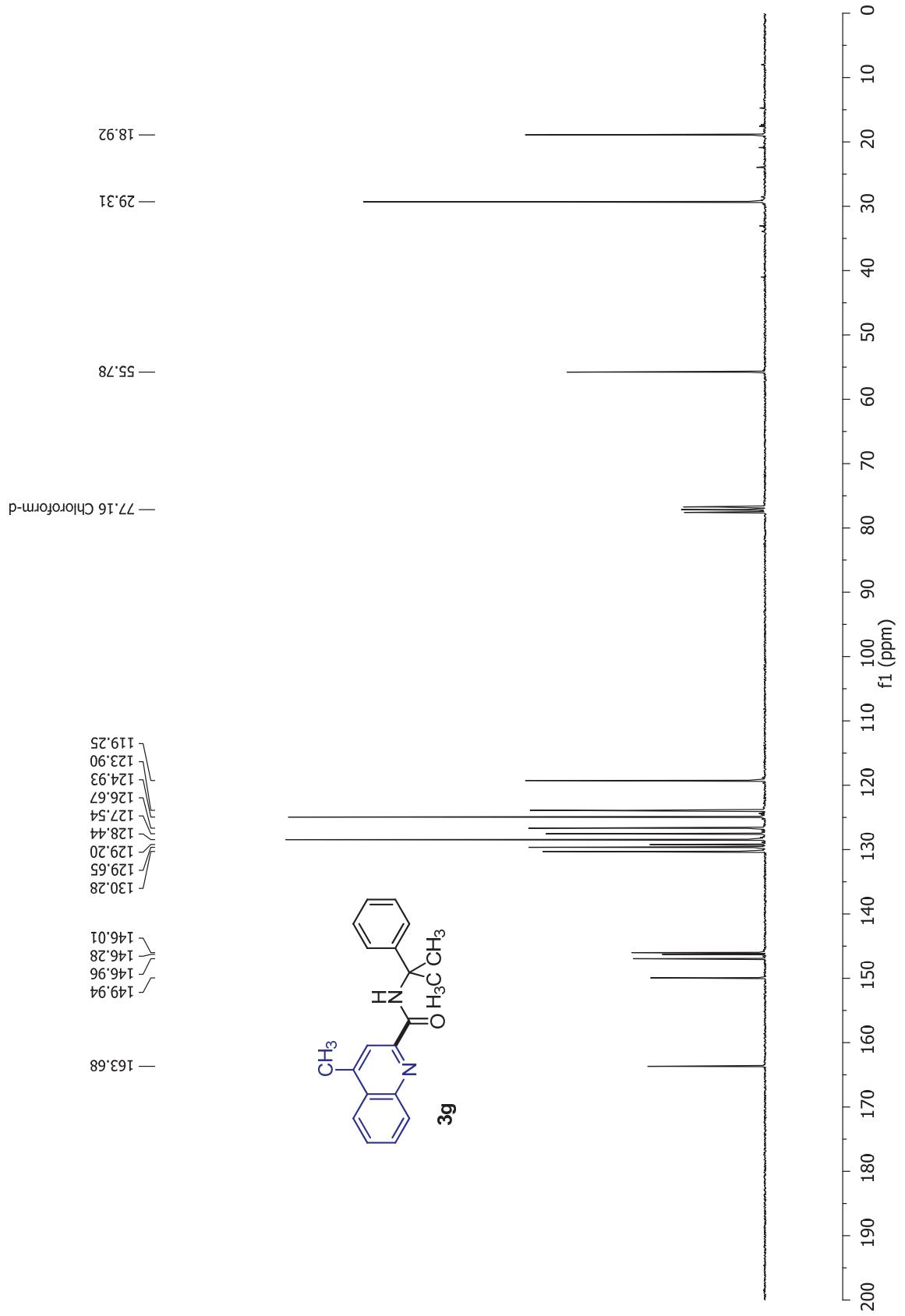
**Supporting Information**



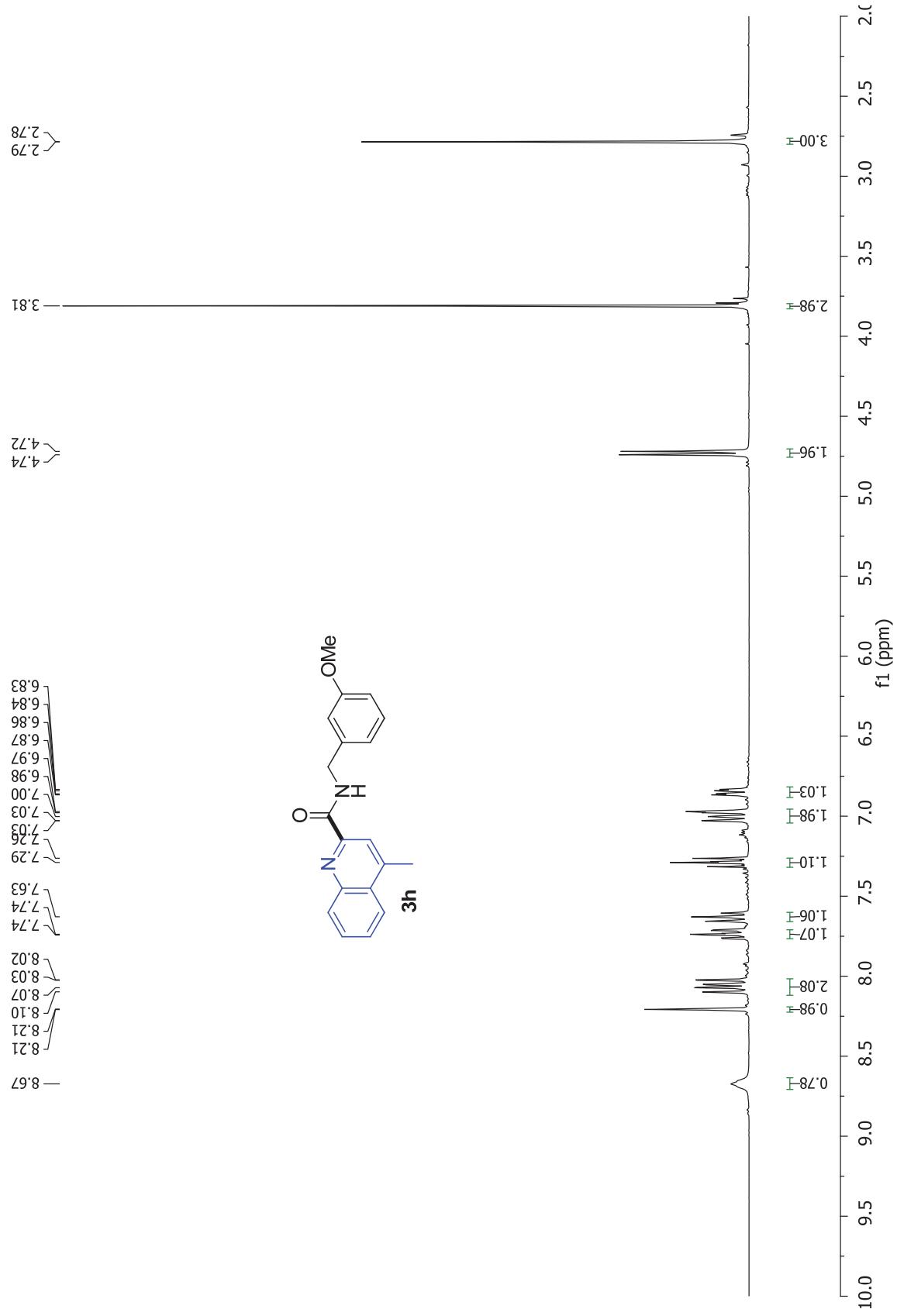
## Supporting Information



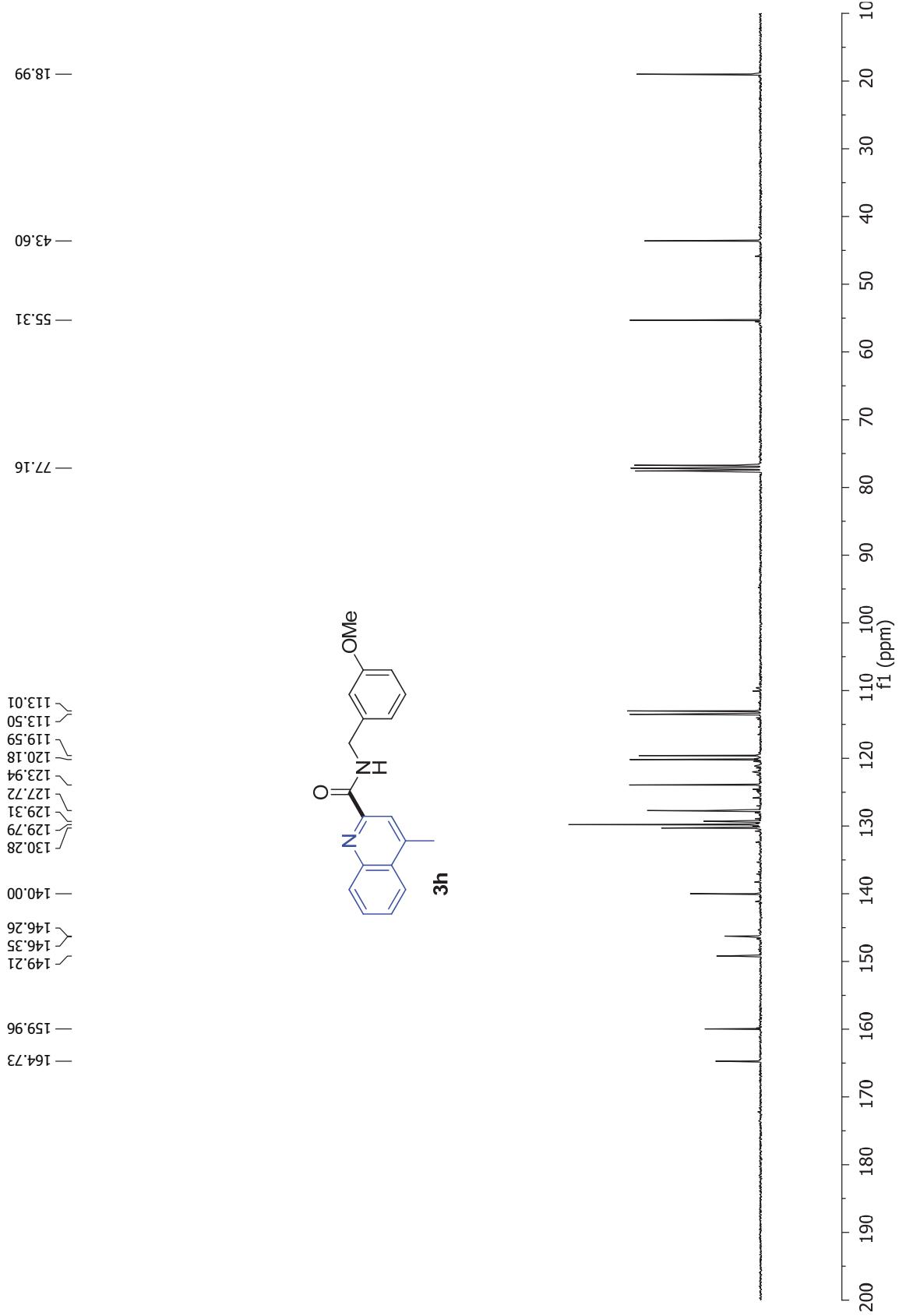
**Supporting Information**



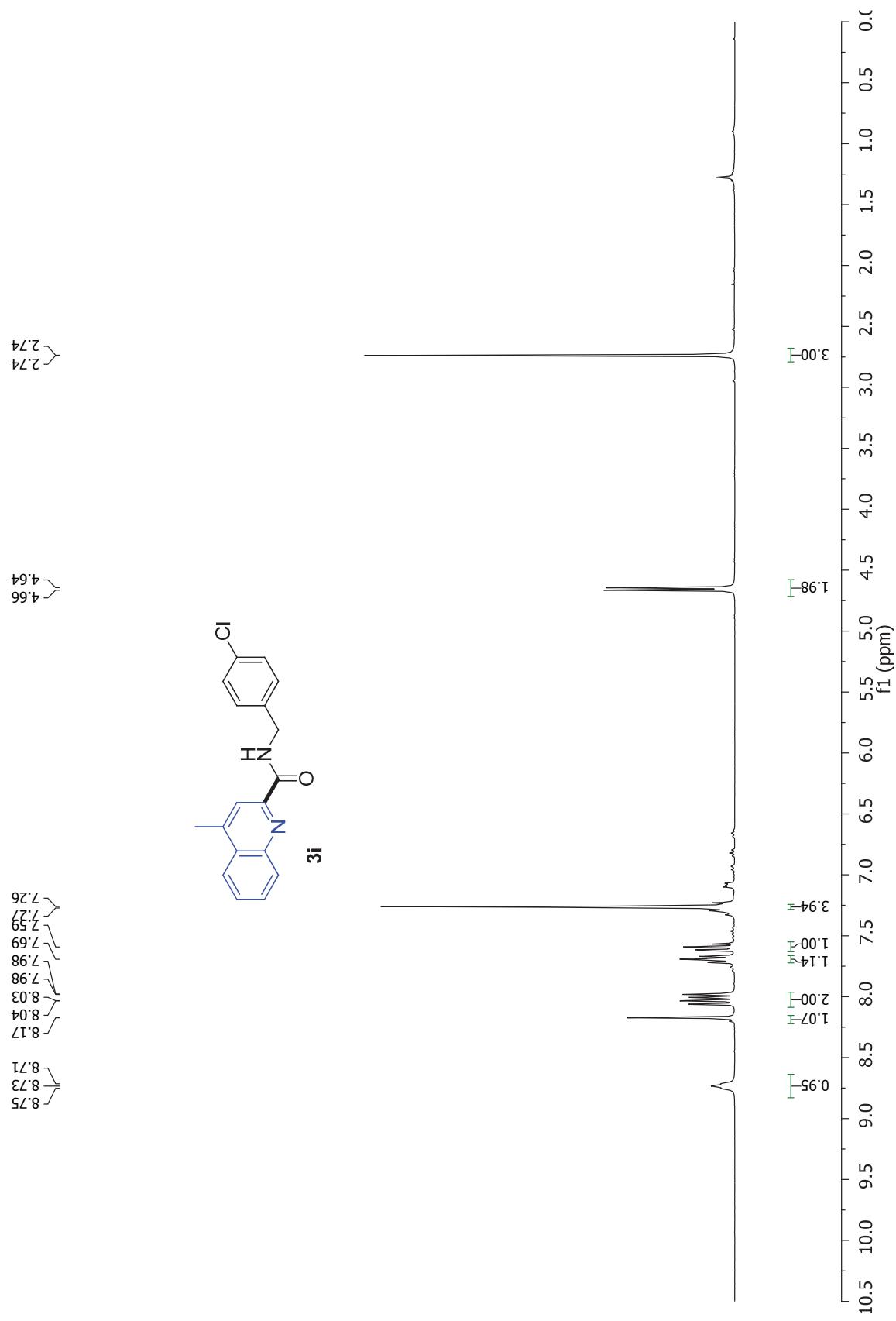
**Supporting Information**



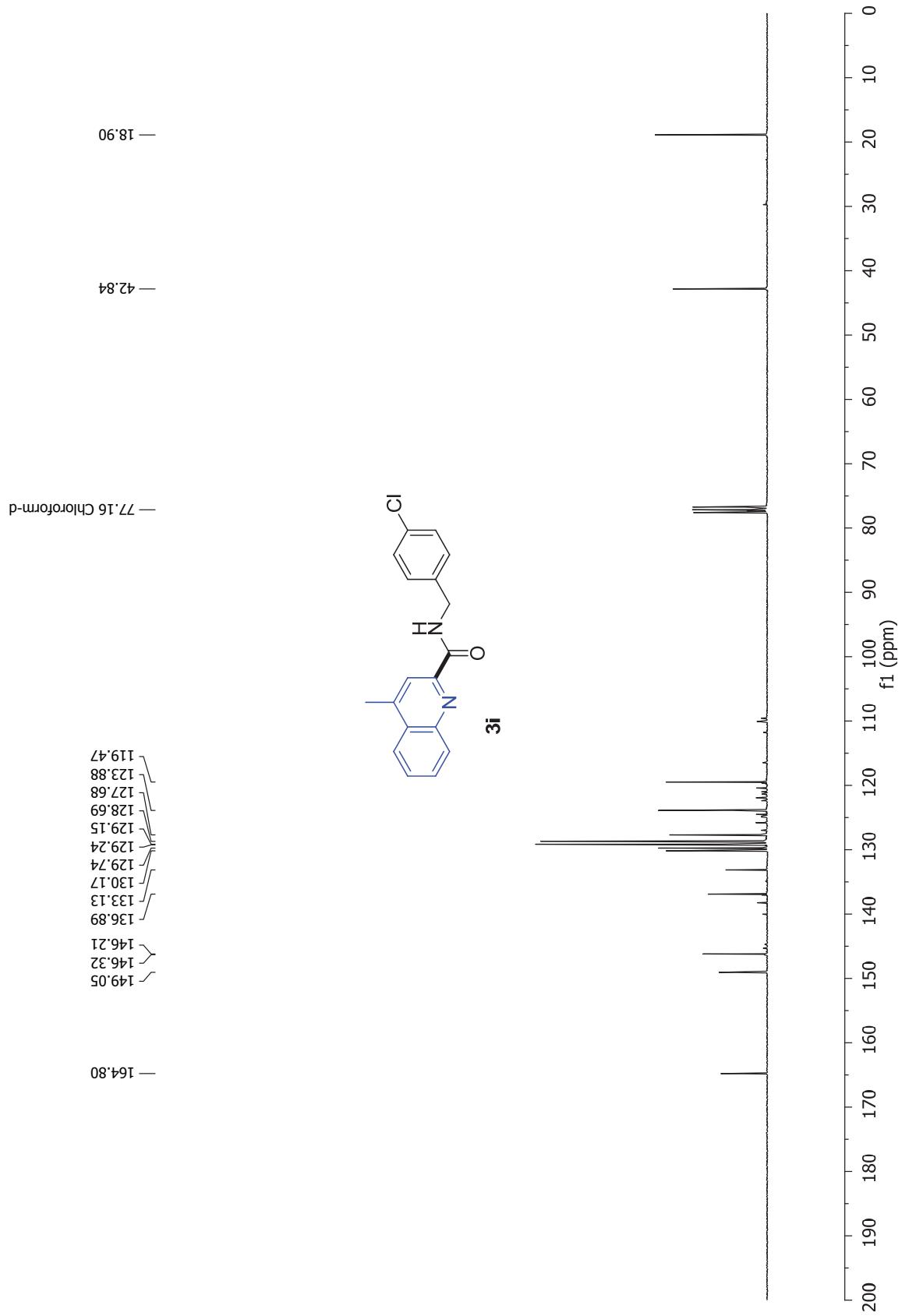
## Supporting Information



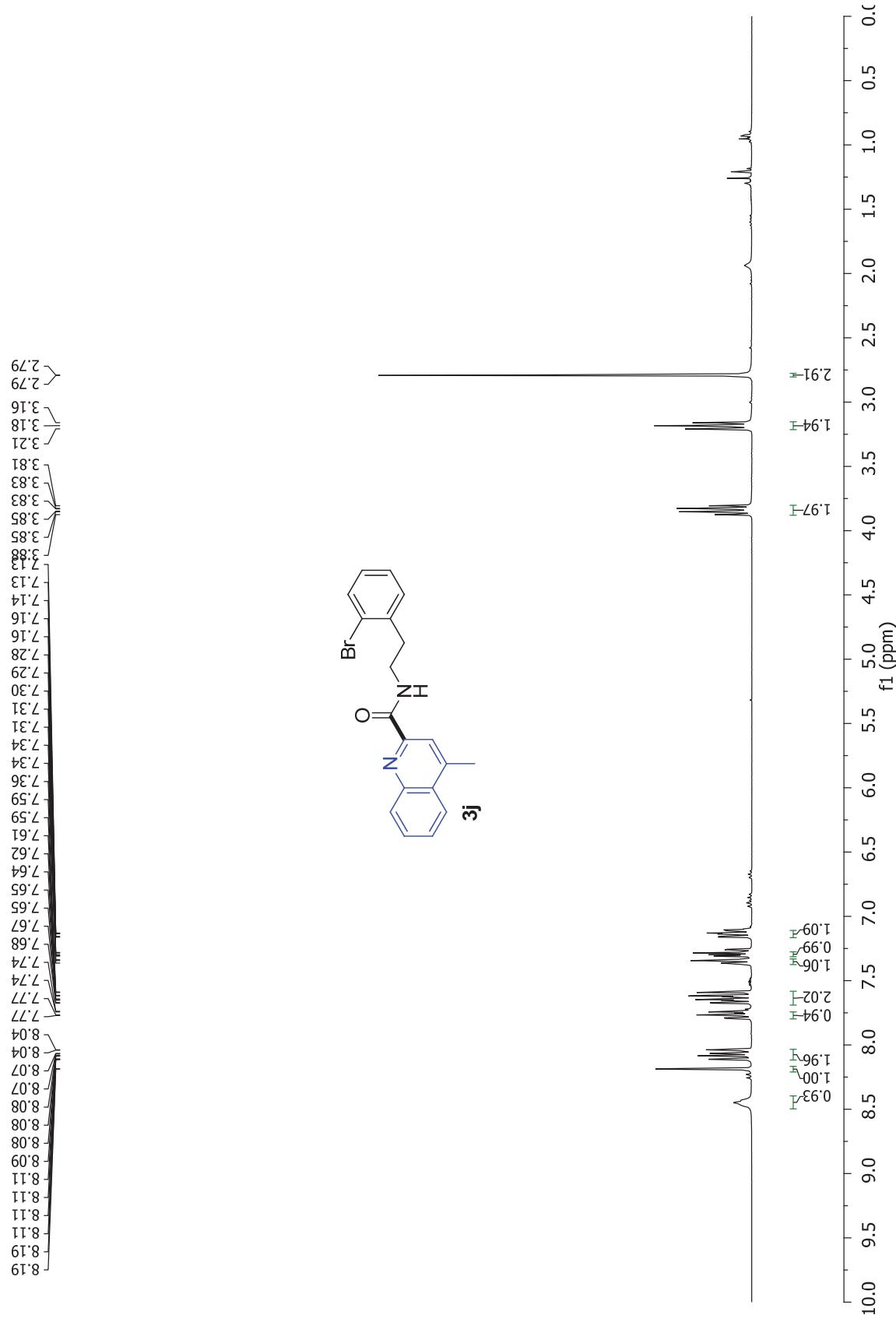
**Supporting Information**



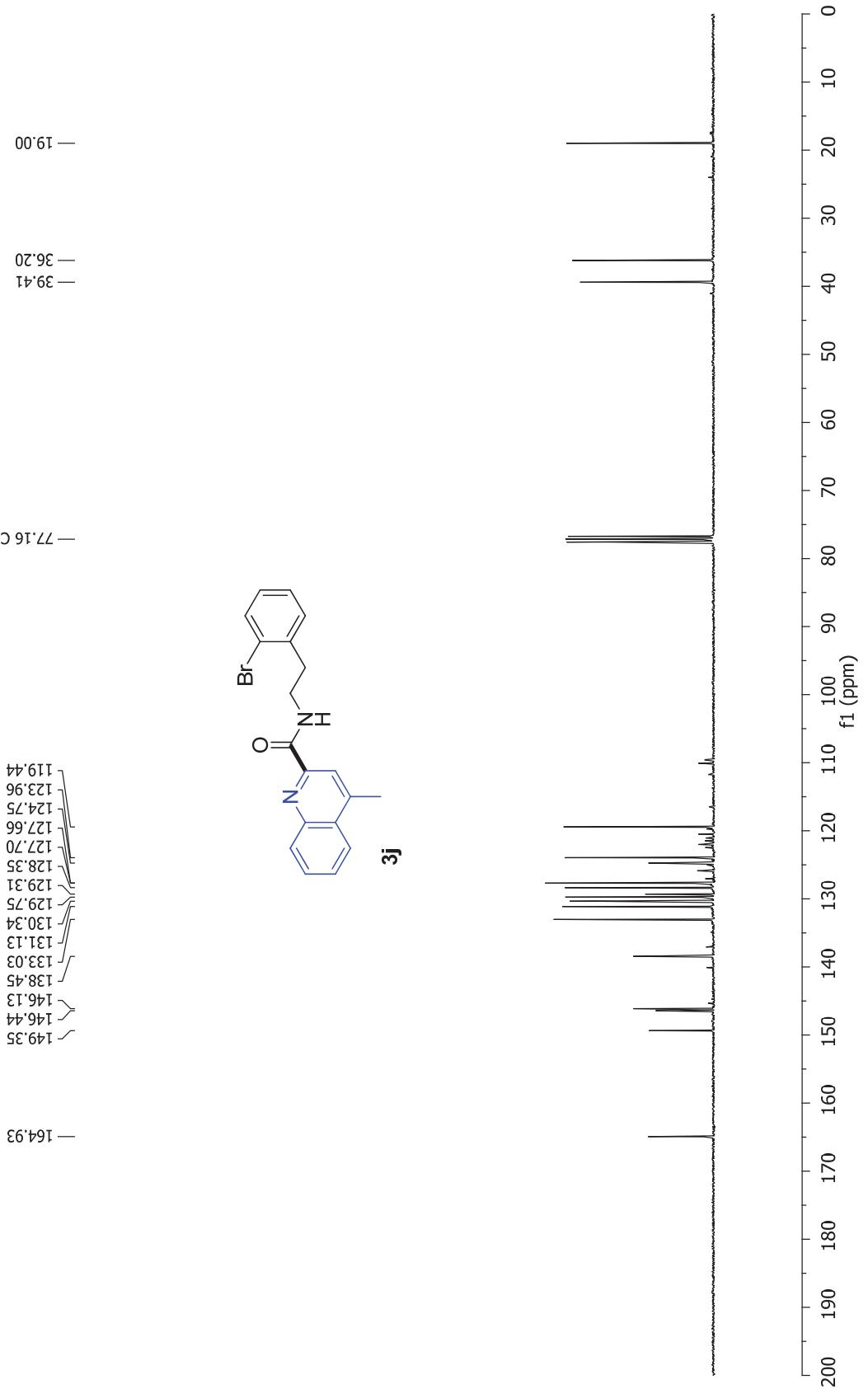
**Supporting Information**



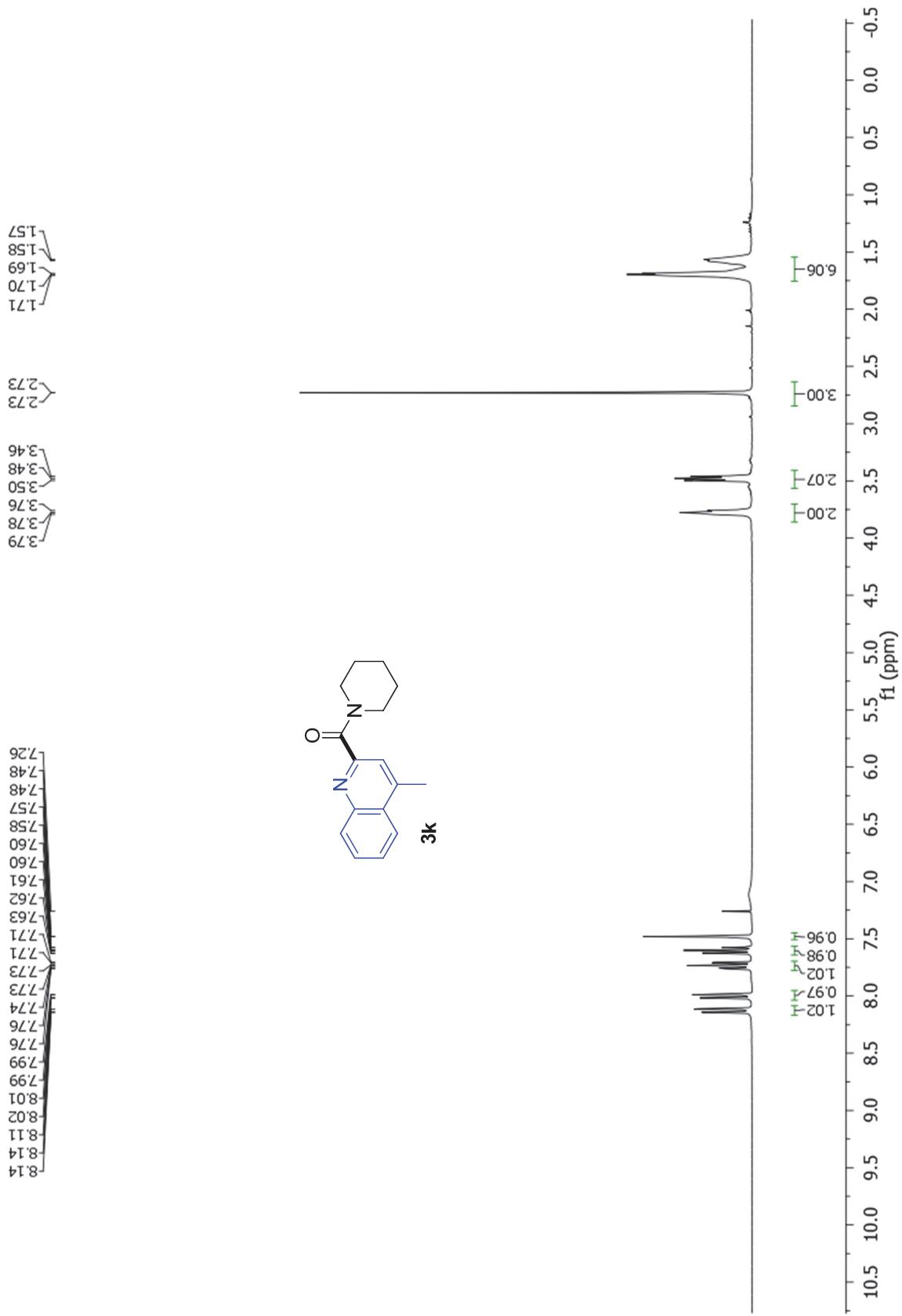
## Supporting Information



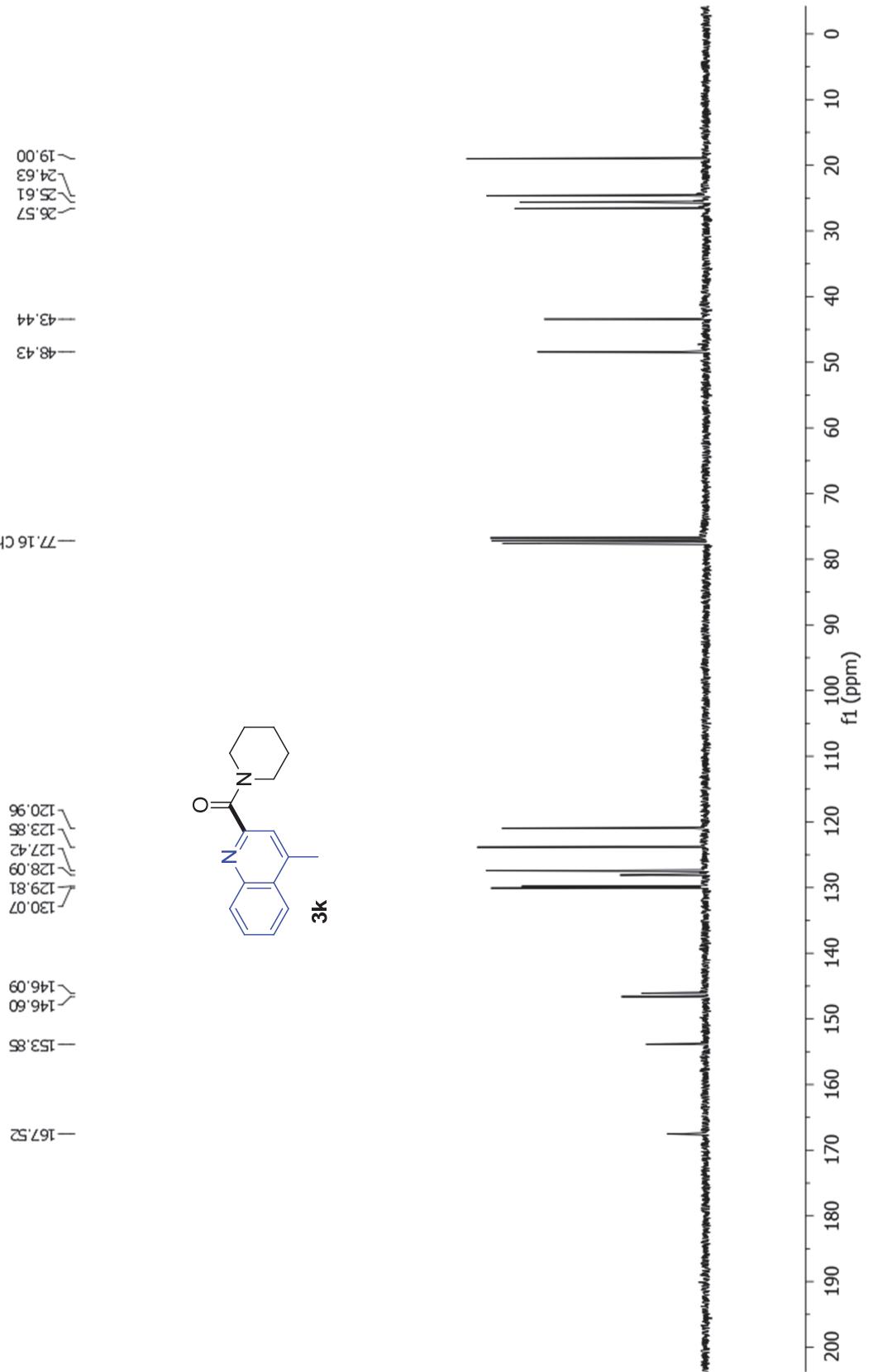
## Supporting Information



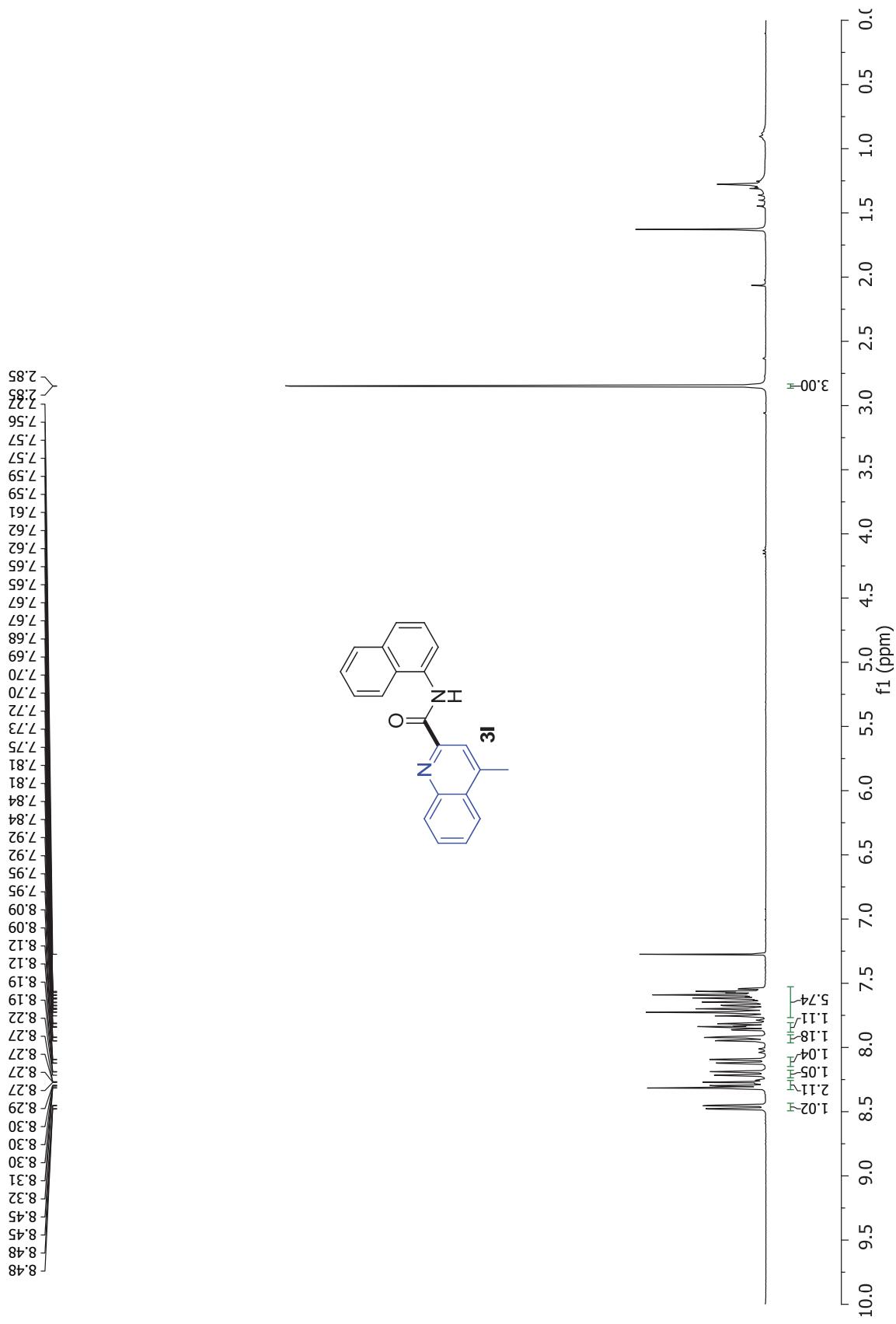
**Supporting Information**



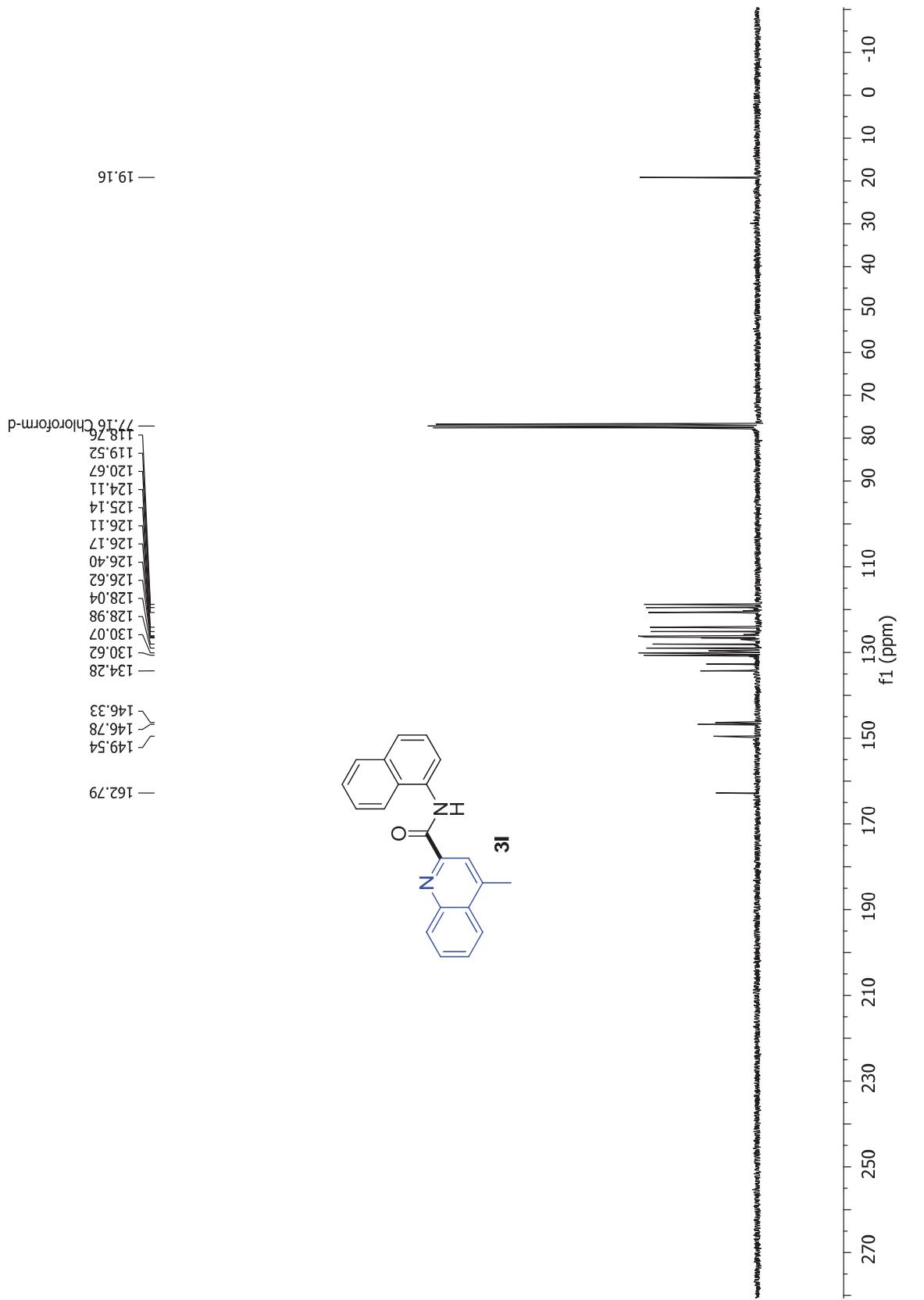
## Supporting Information



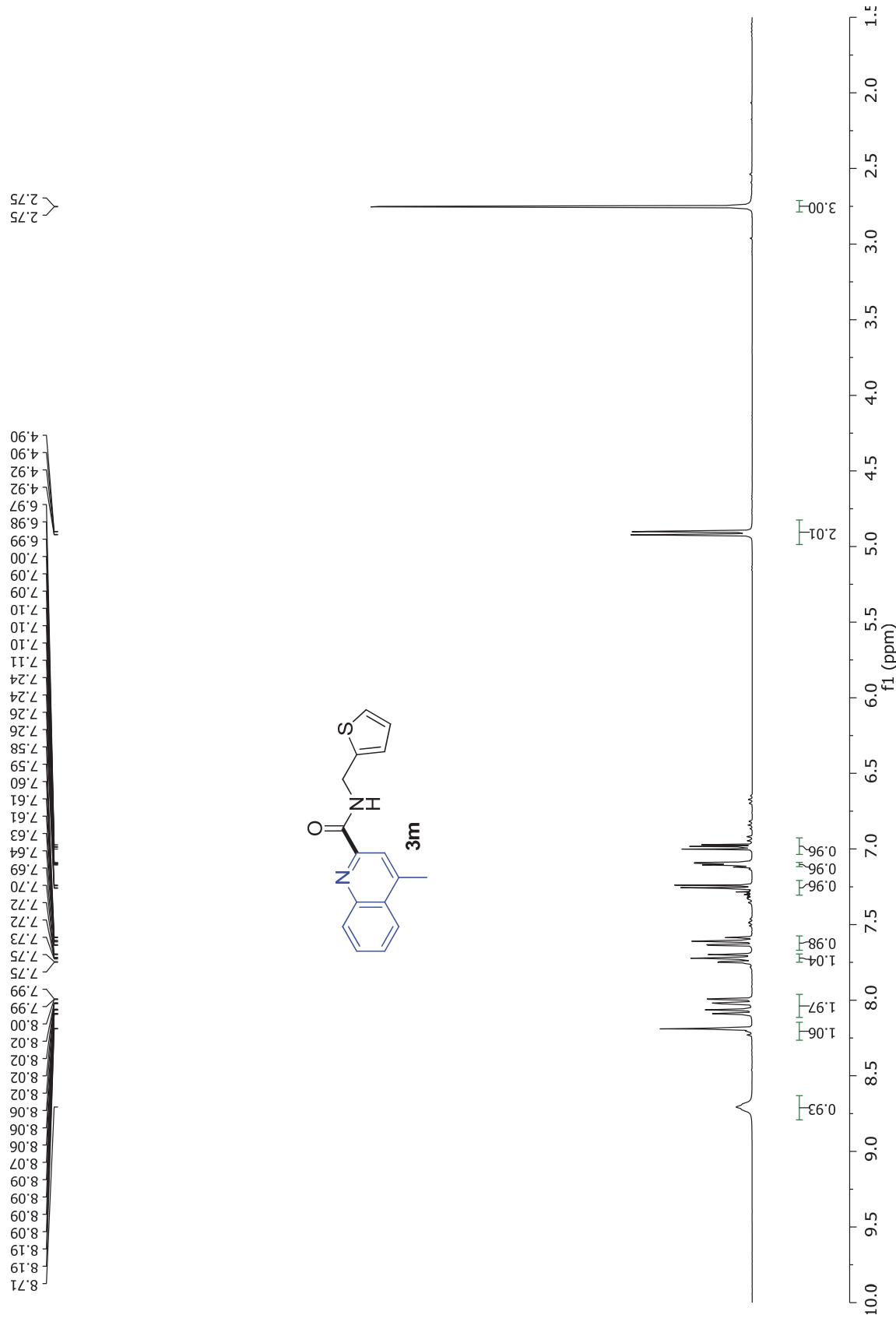
Supporting Information



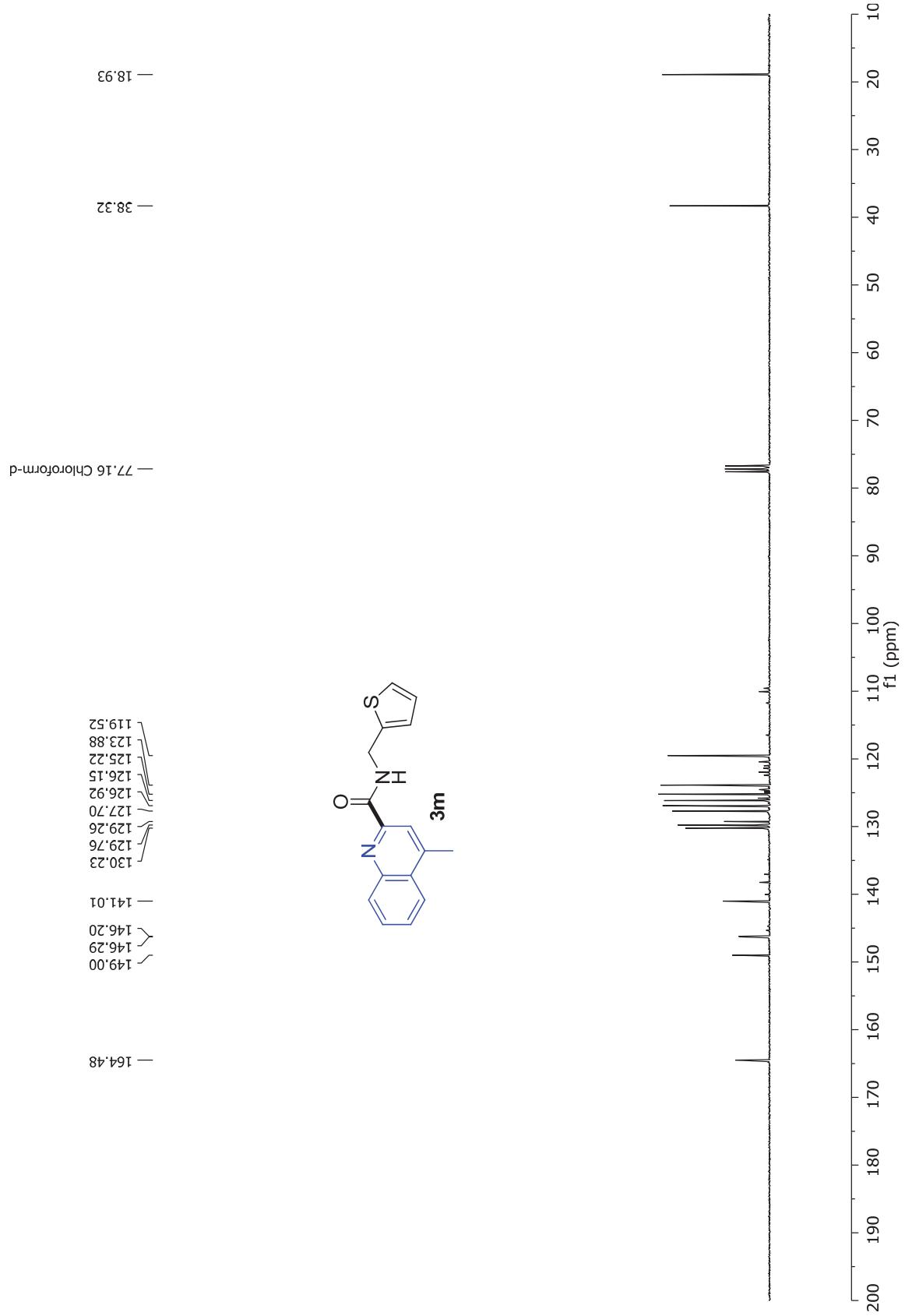
## Supporting Information



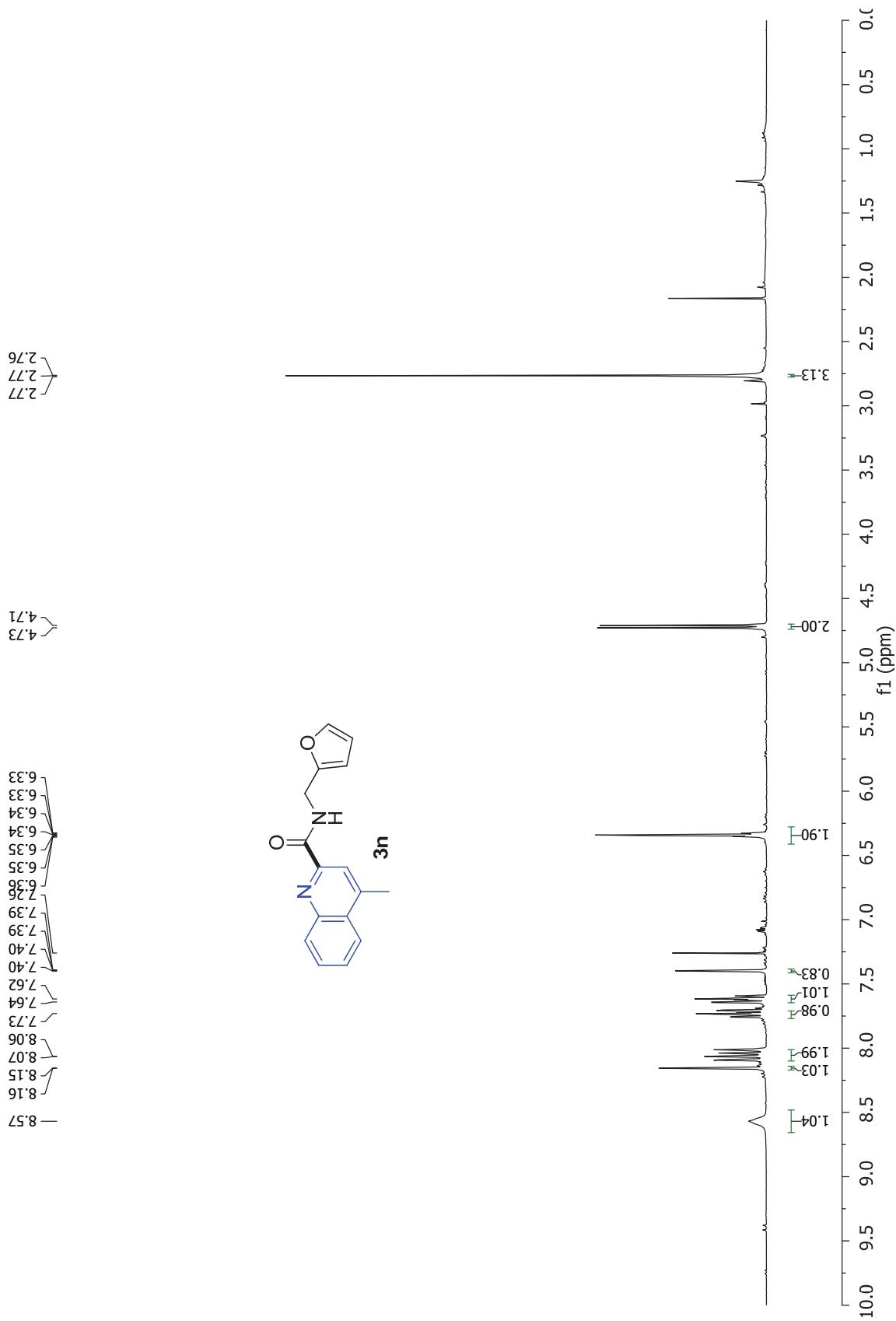
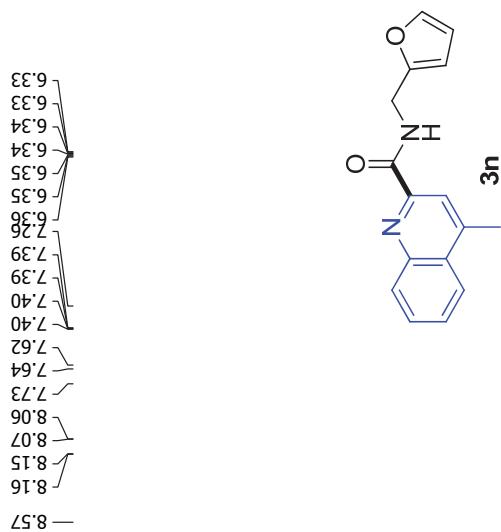
## Supporting Information



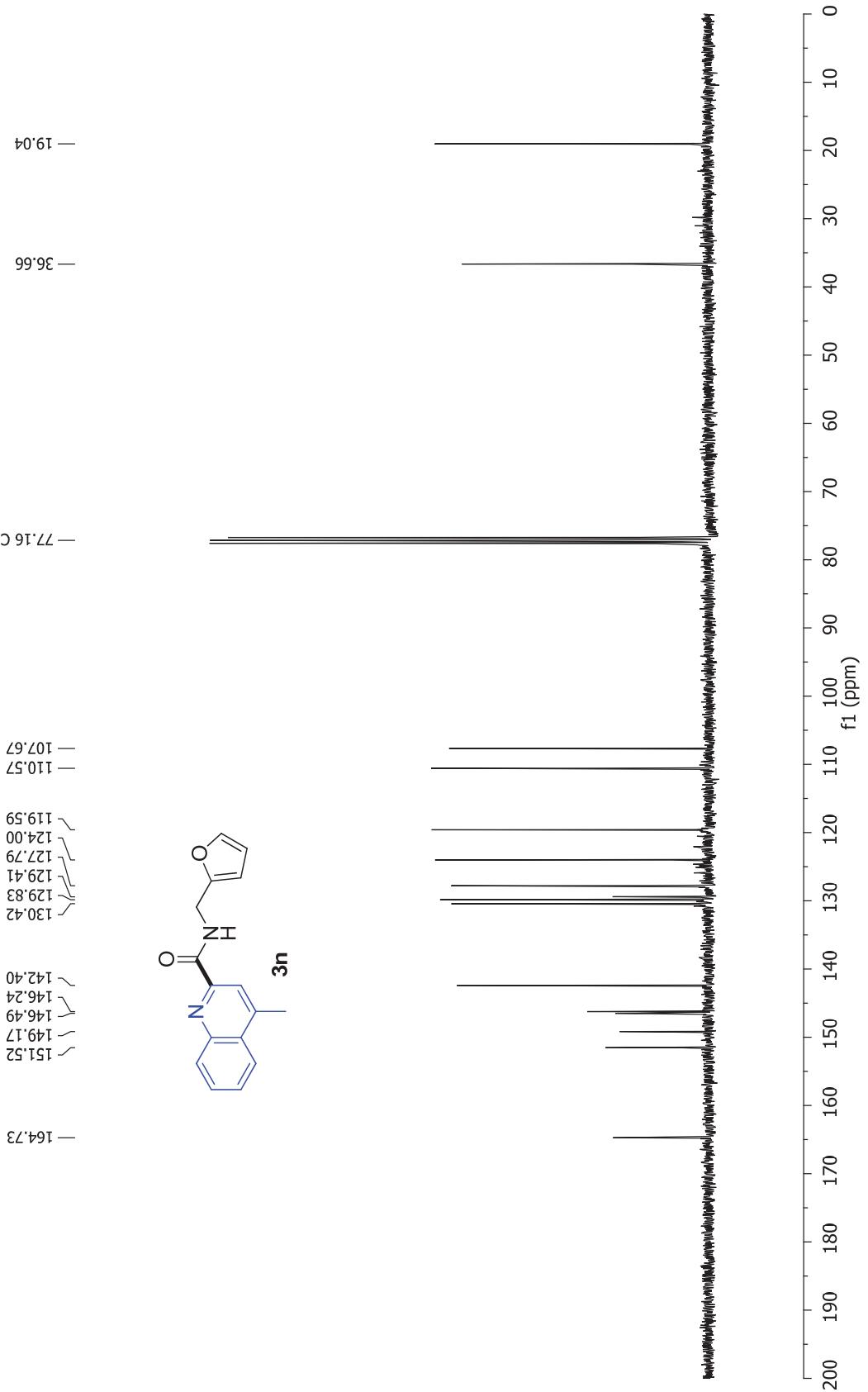
## Supporting Information



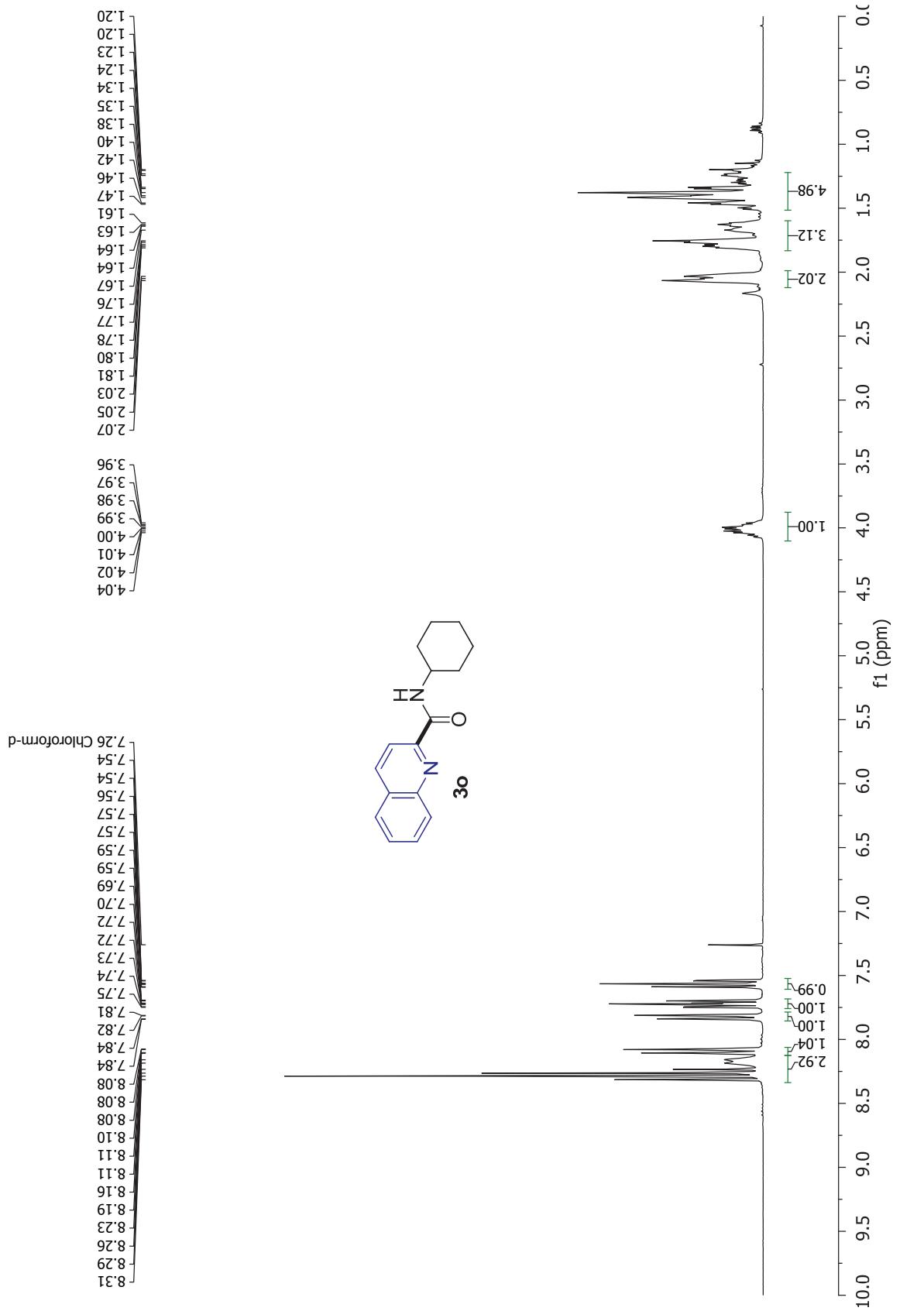
Supporting Information



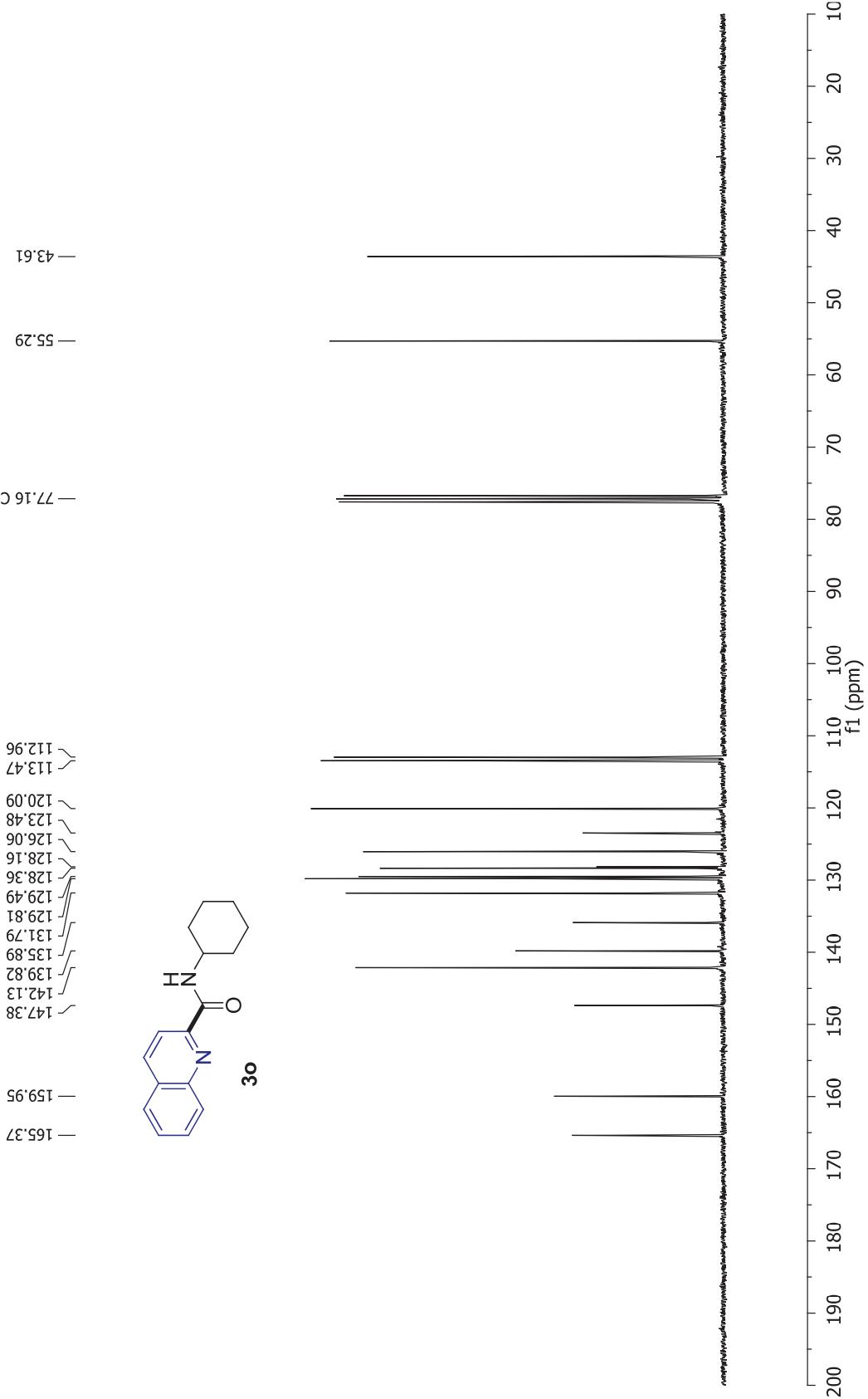
## Supporting Information



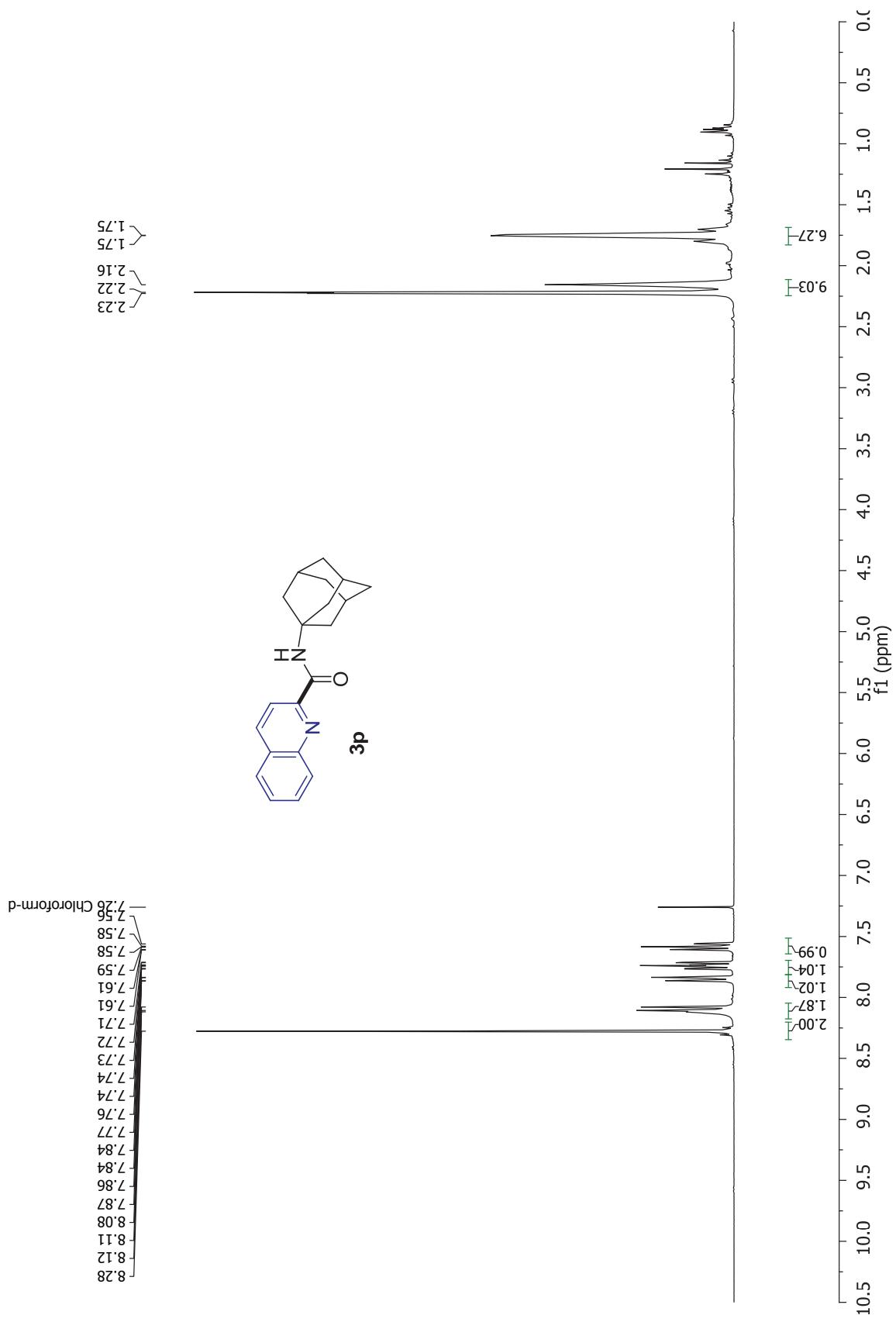
## Supporting Information



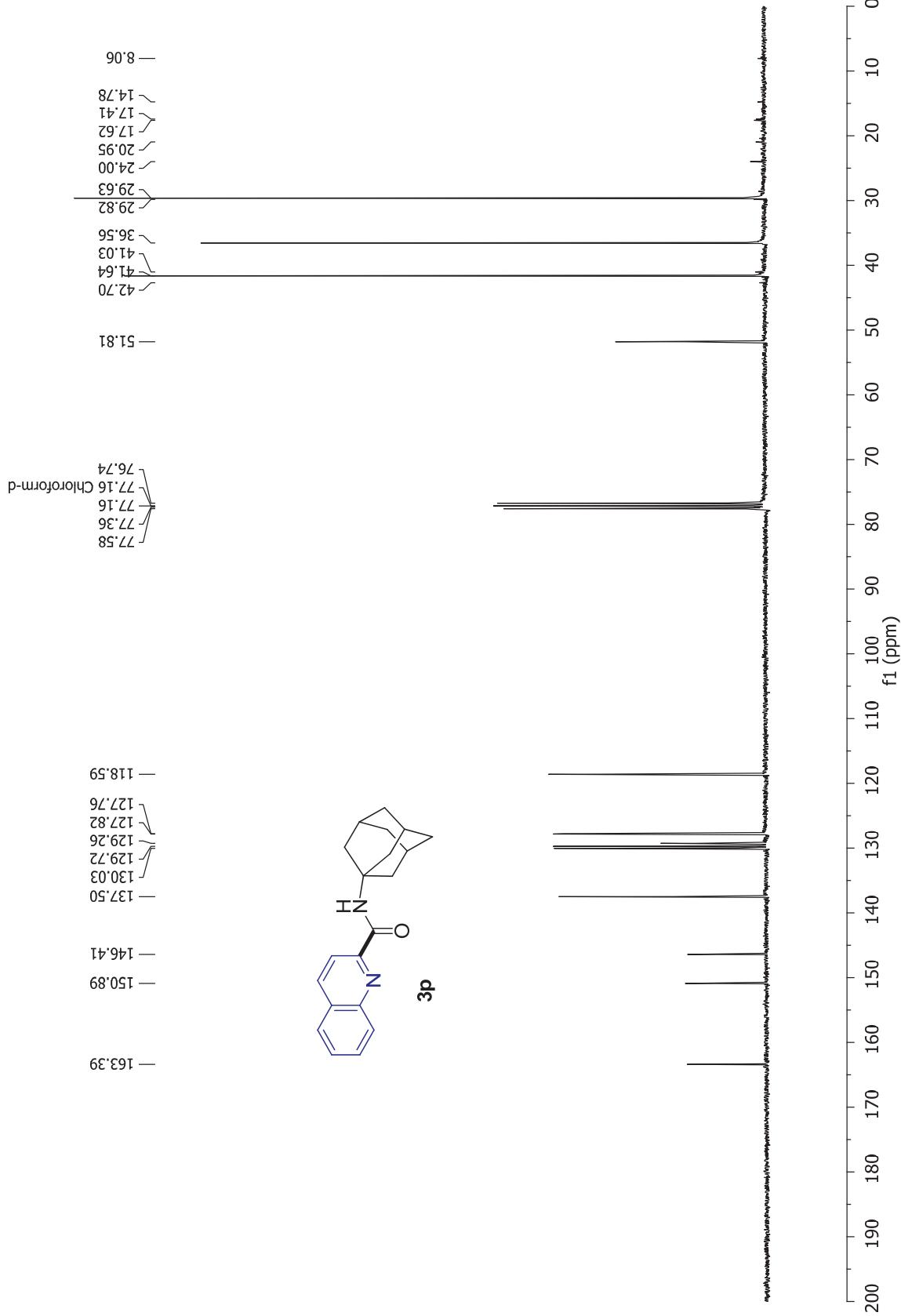
## Supporting Information



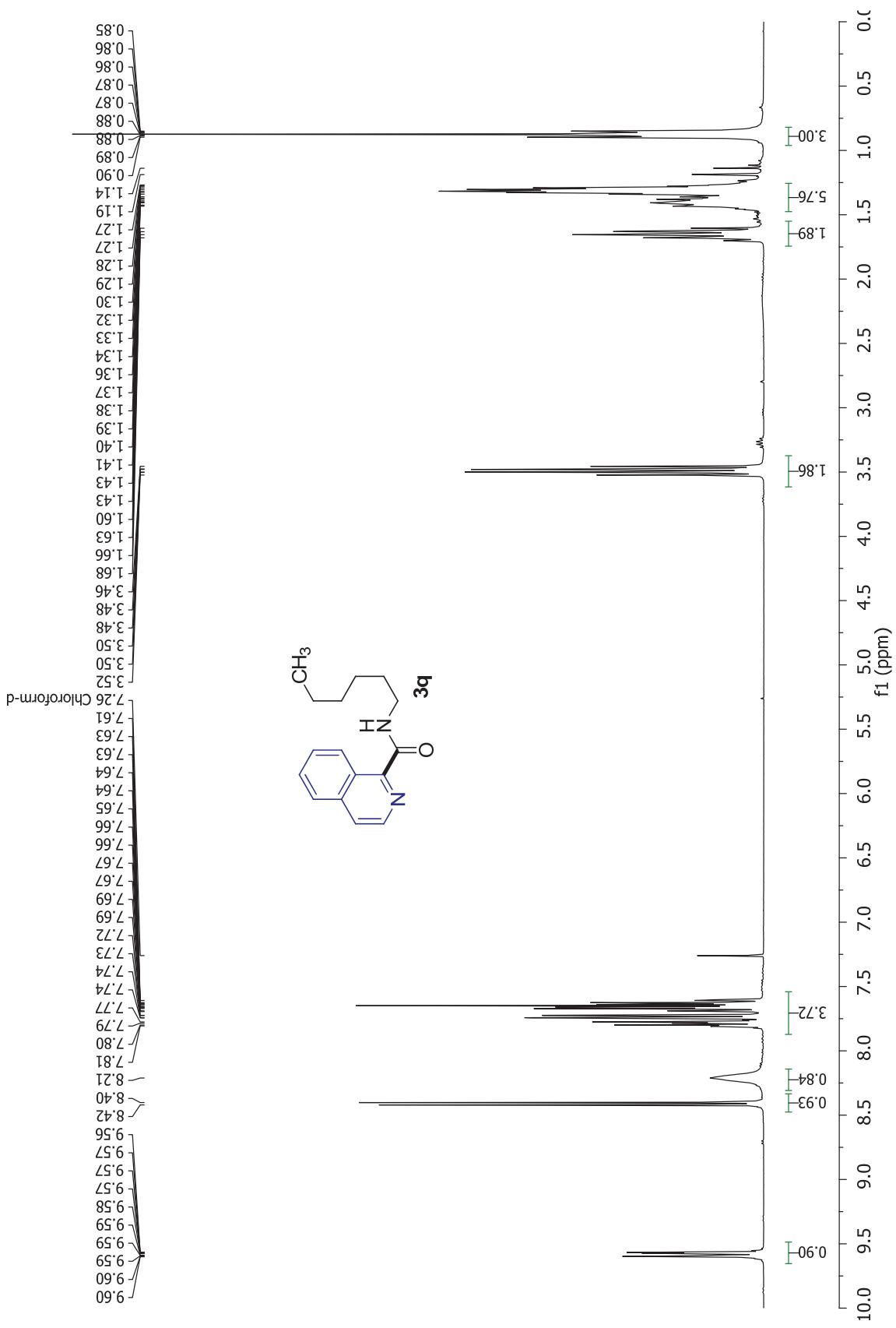
Supporting Information



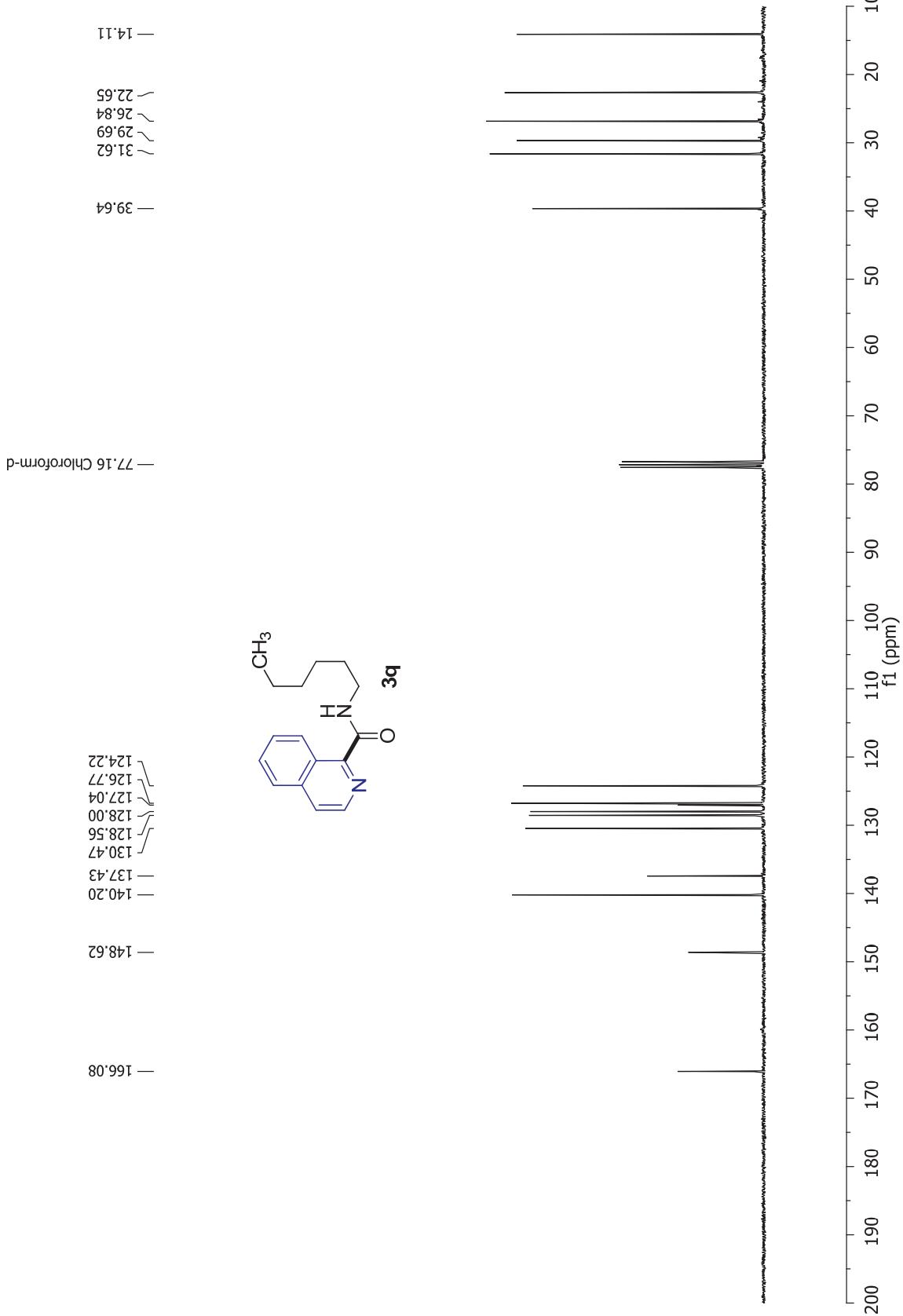
**Supporting Information**



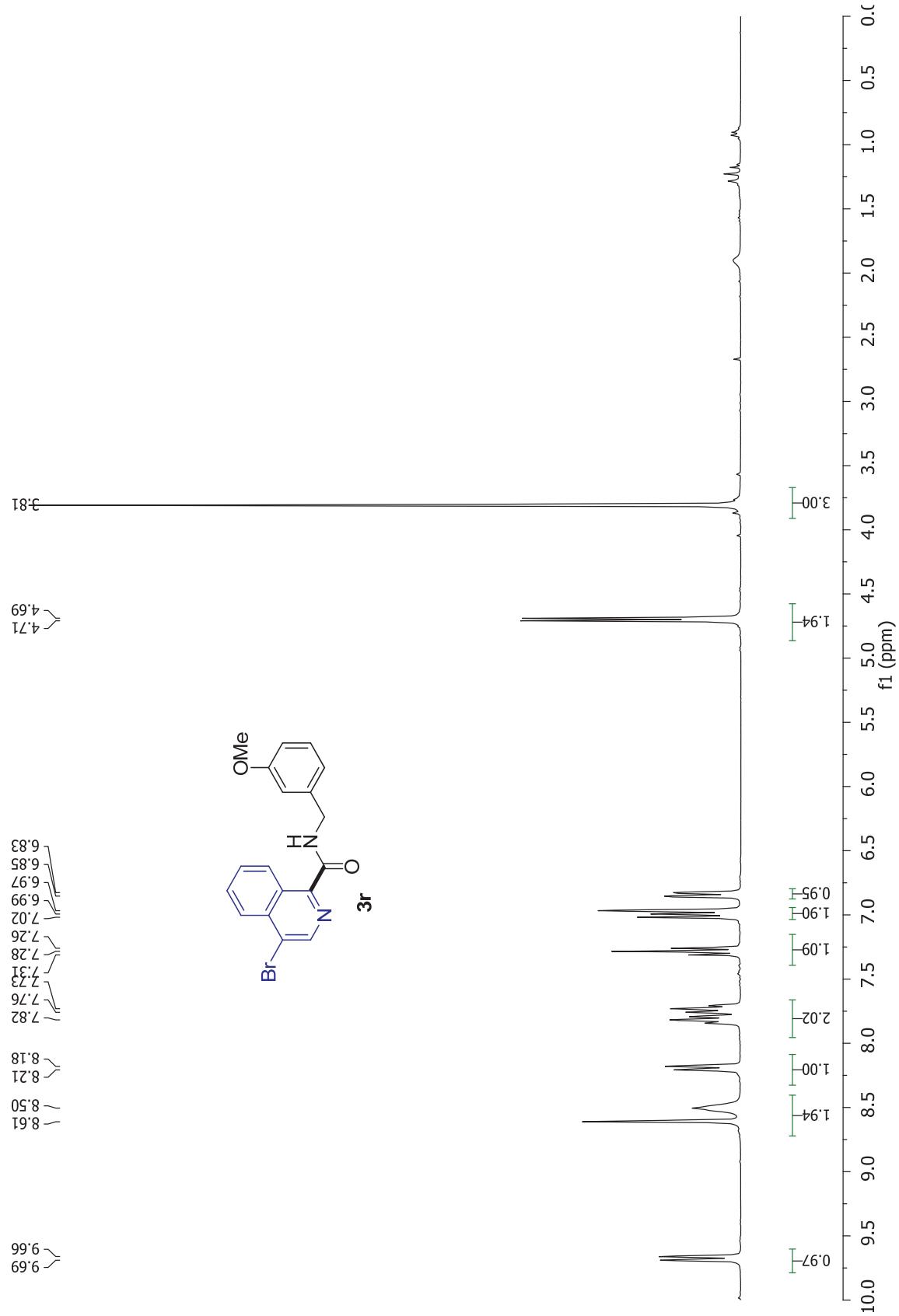
## Supporting Information



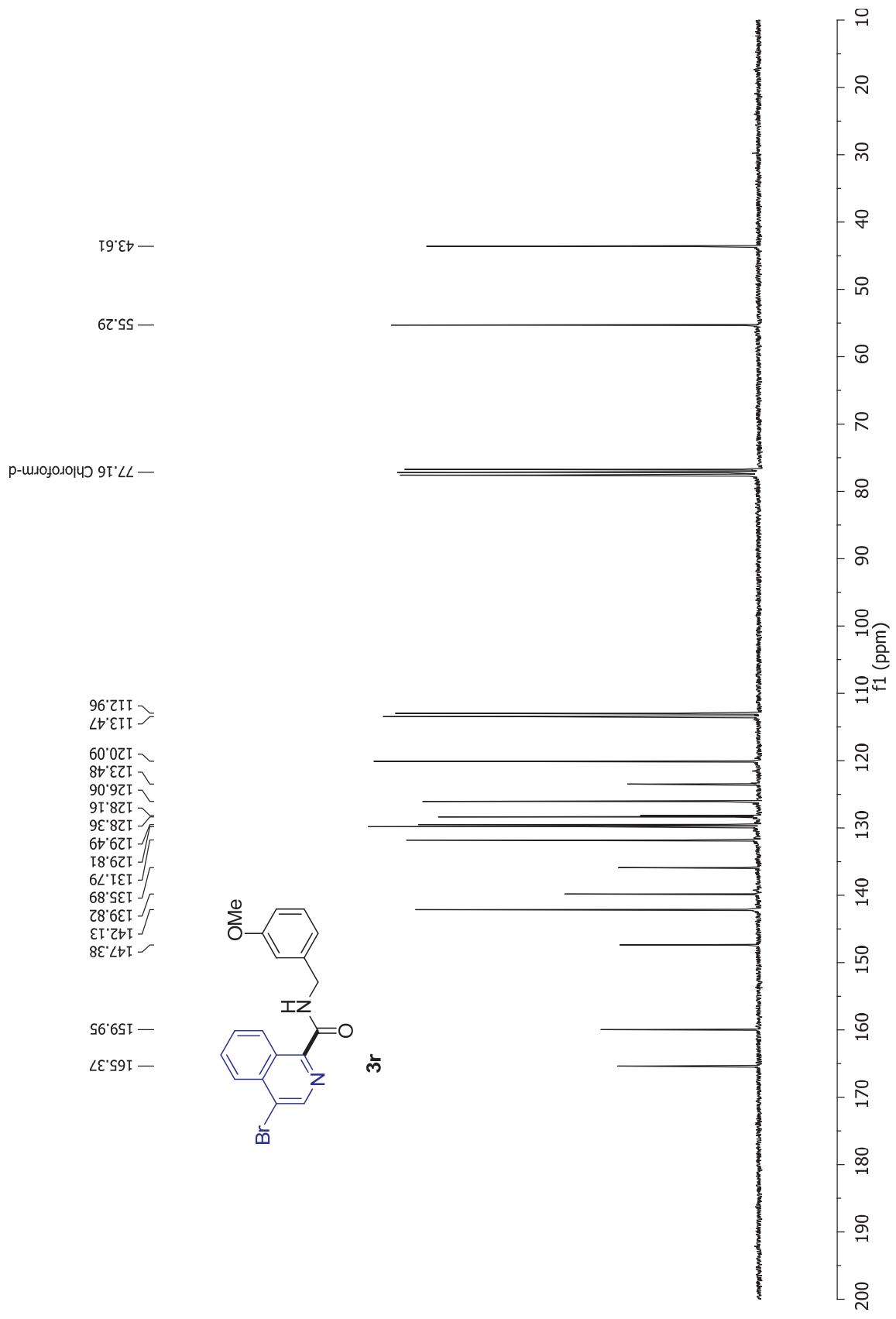
## Supporting Information



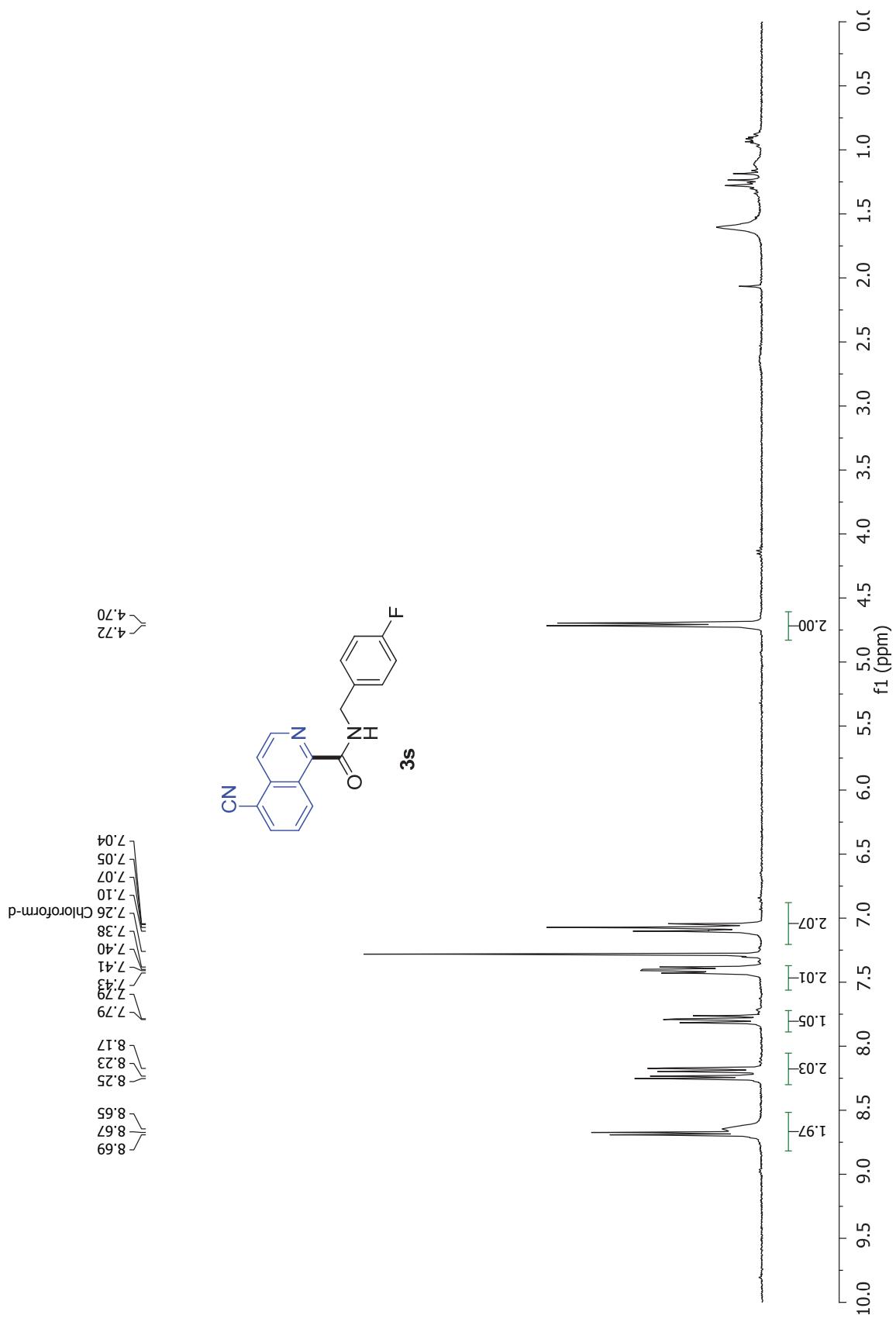
**Supporting Information**



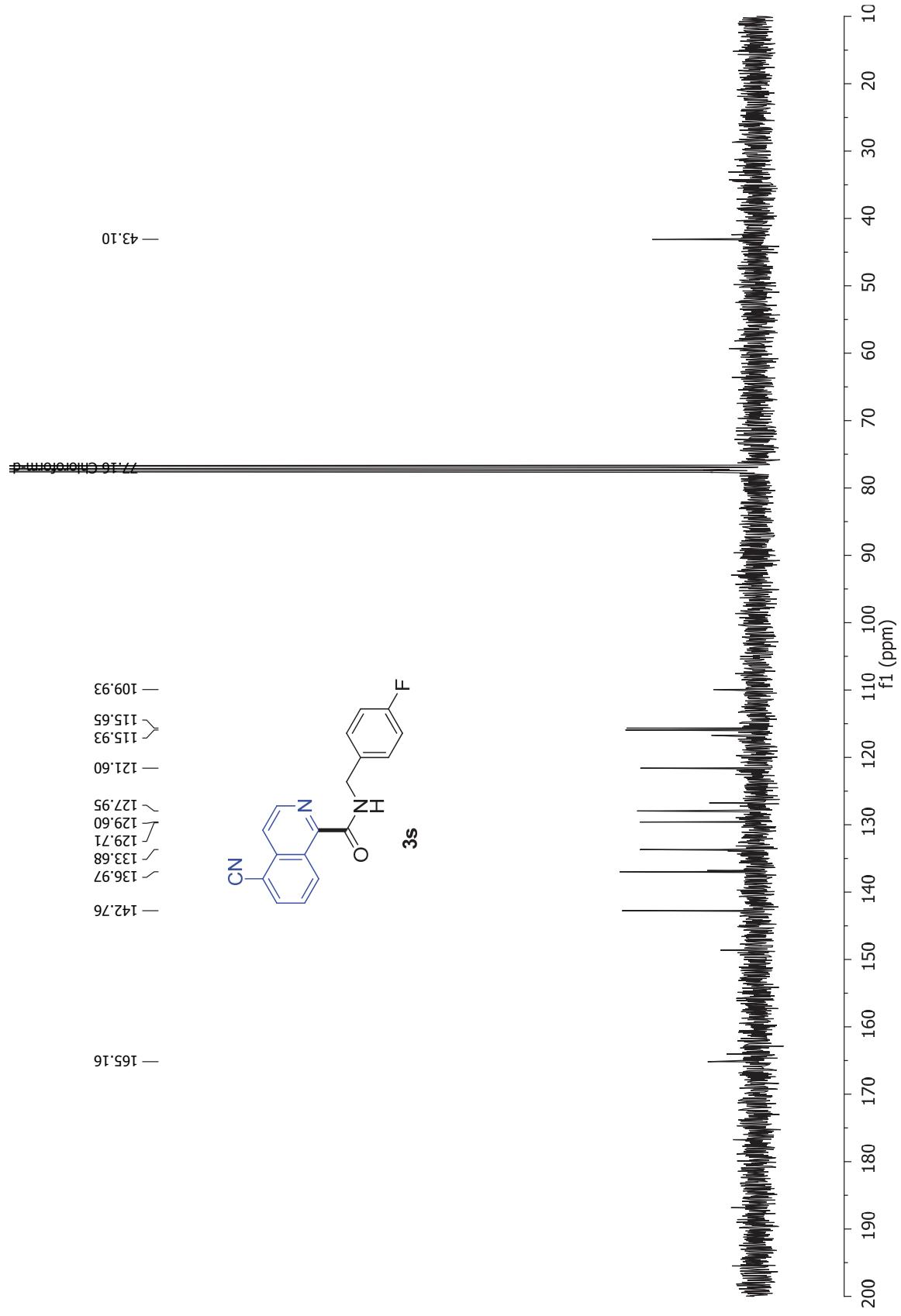
## Supporting Information



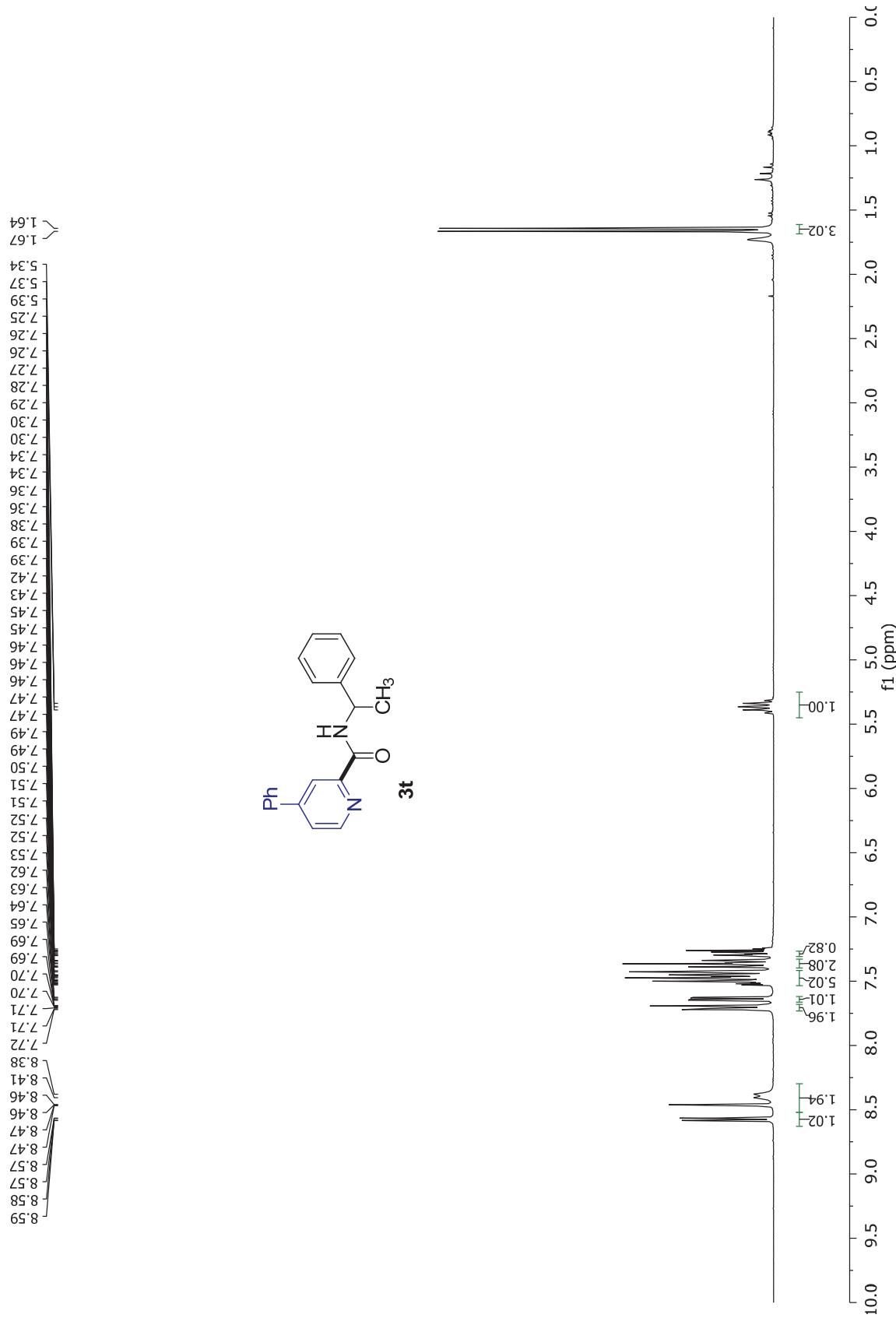
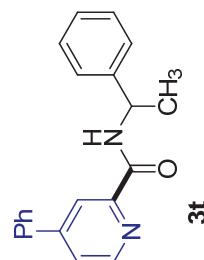
**Supporting Information**



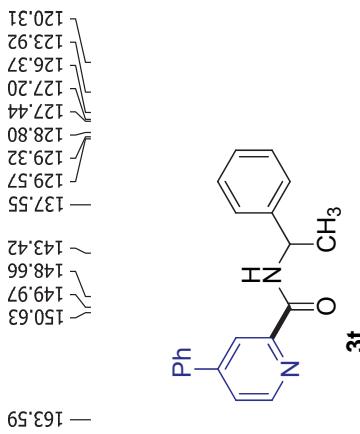
**Supporting Information**



## Supporting Information



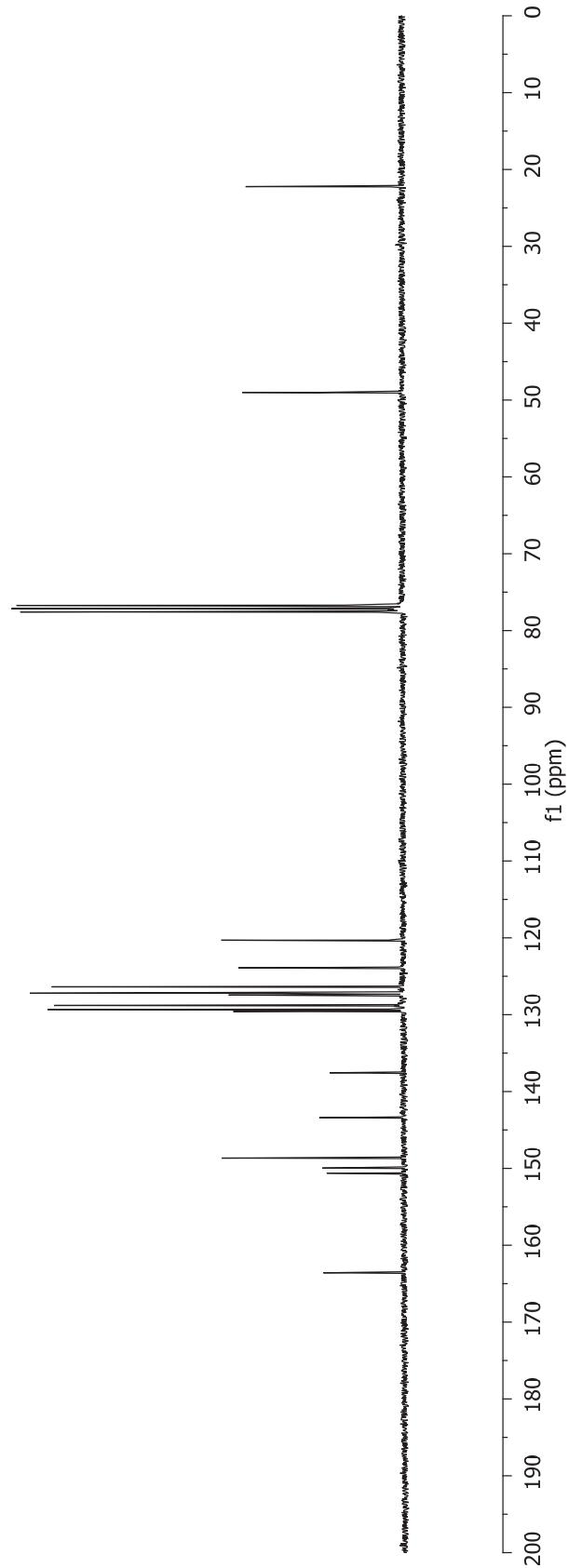
## Supporting Information



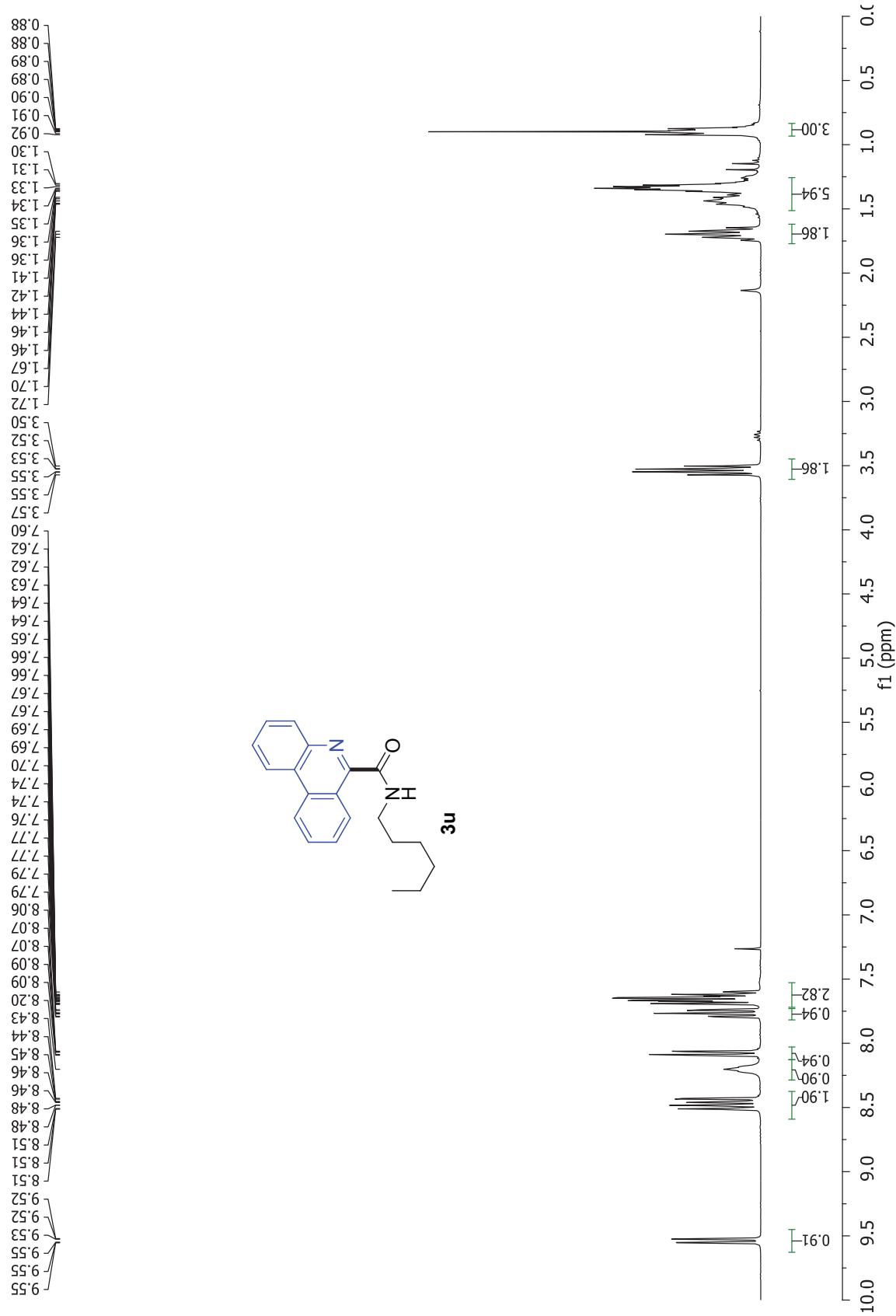
—77.16 Chloroform-d

—49.02

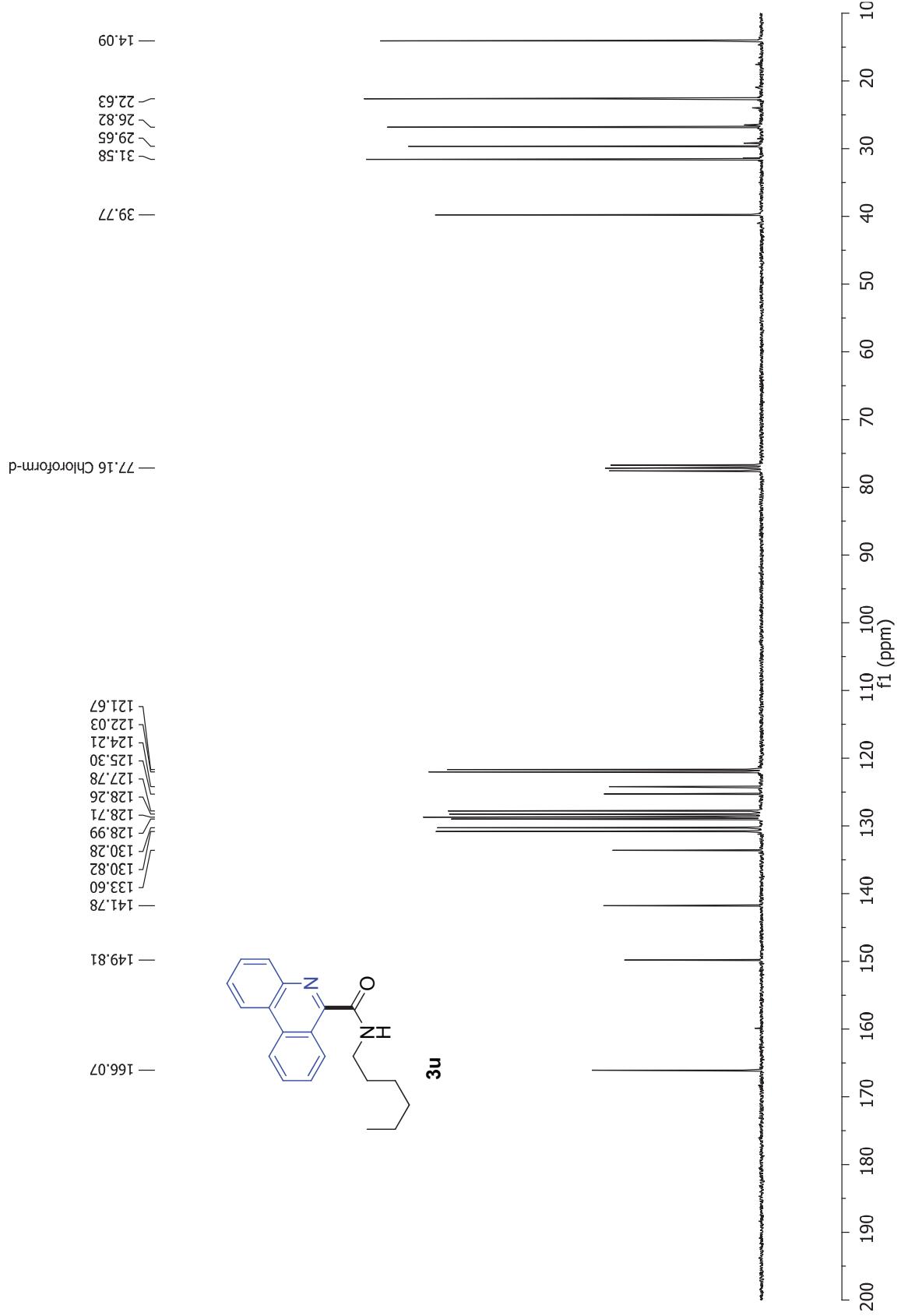
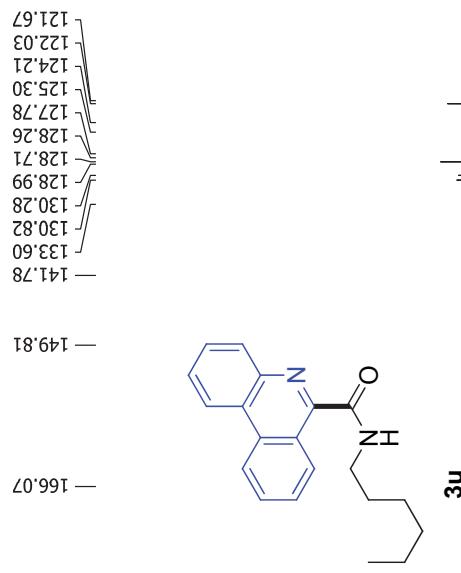
—22.23



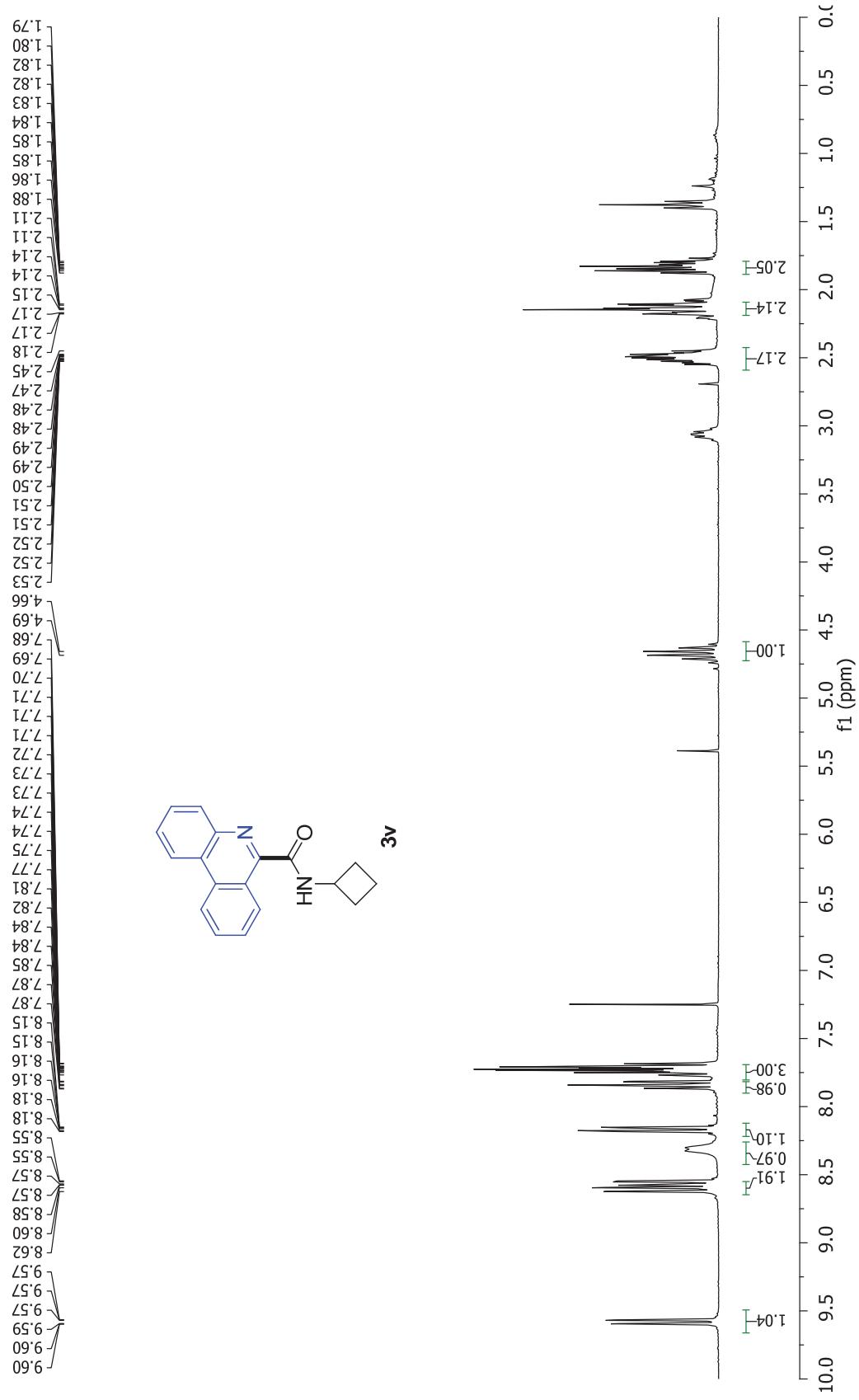
**Supporting Information**



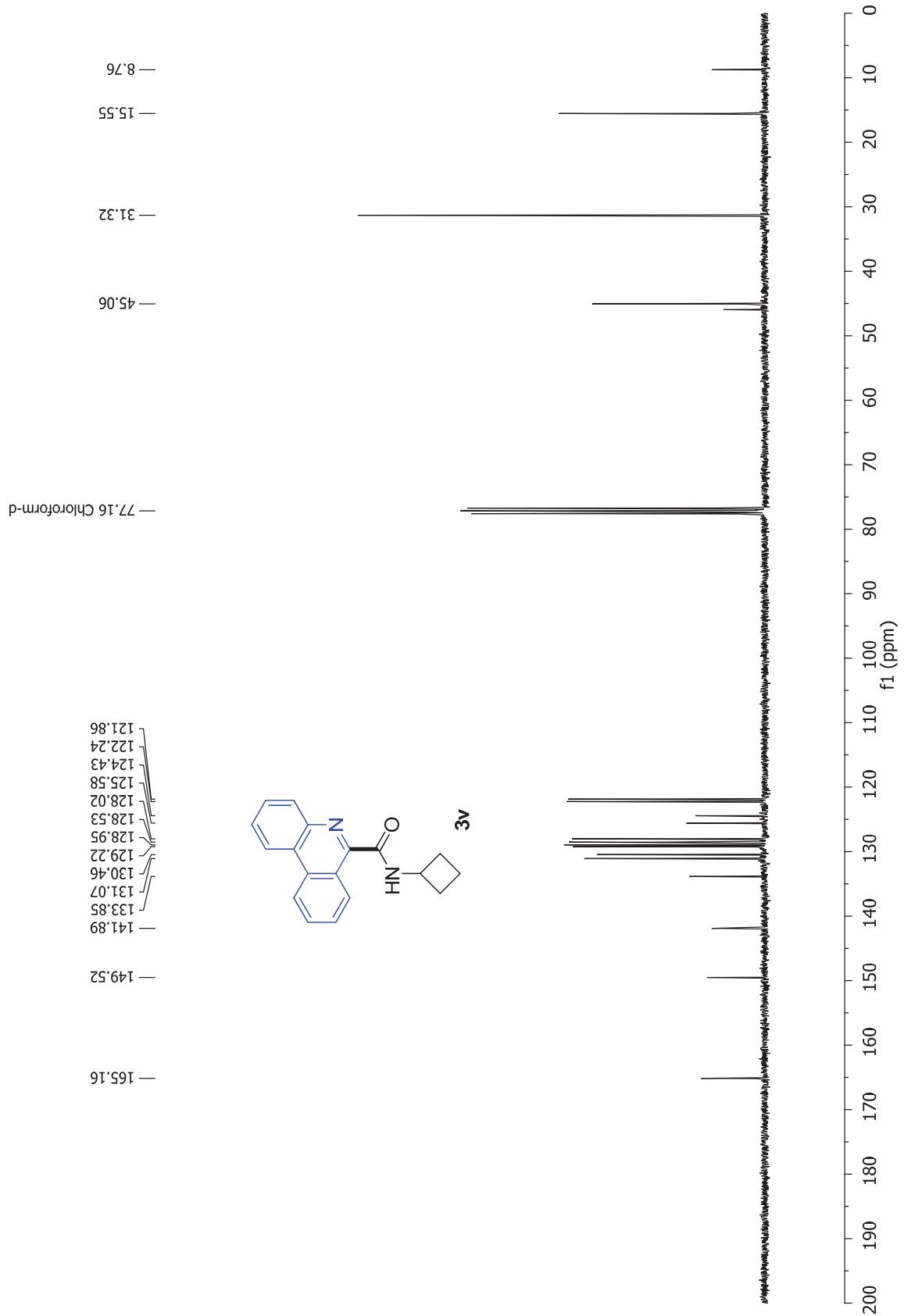
## Supporting Information



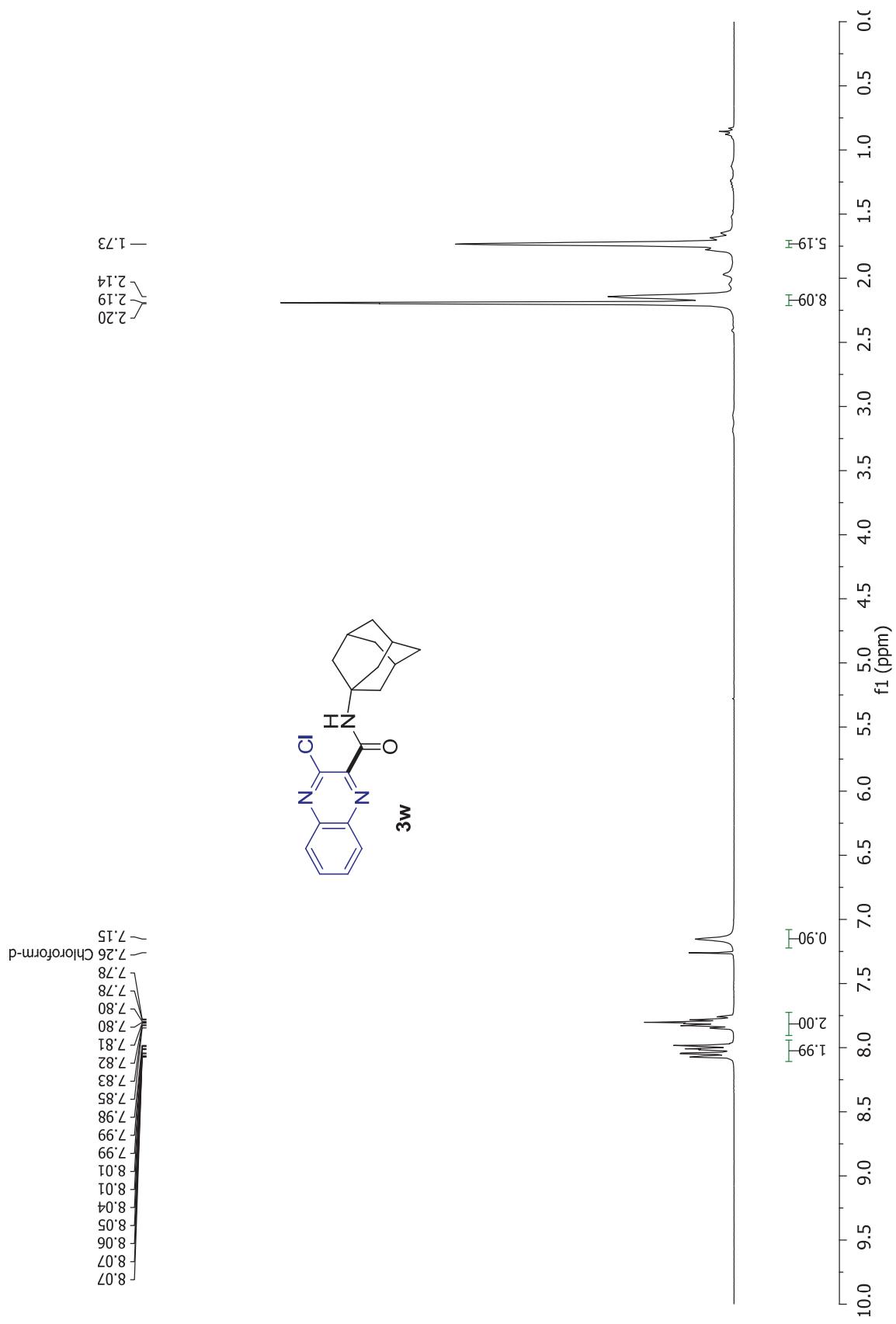
## Supporting Information



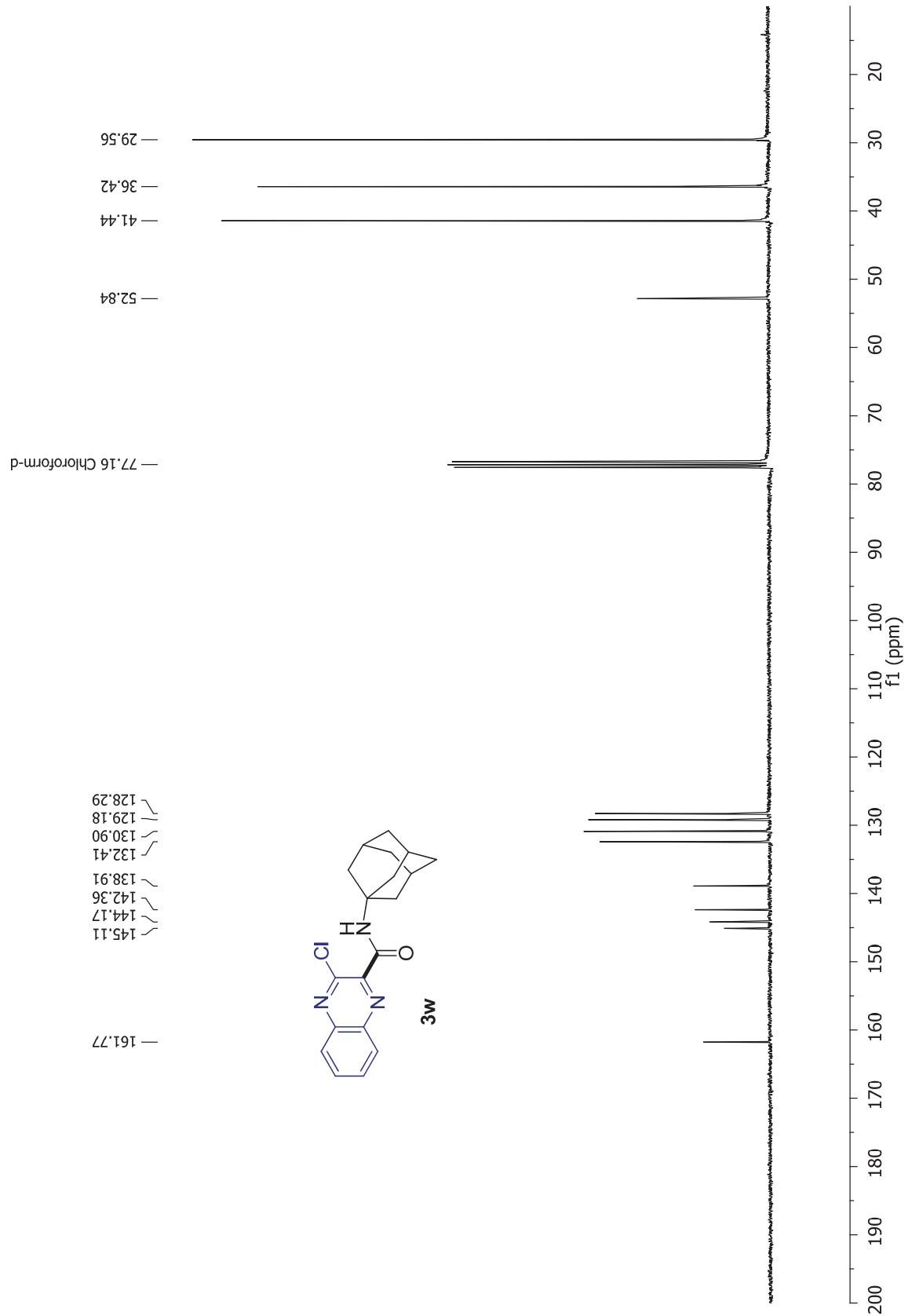
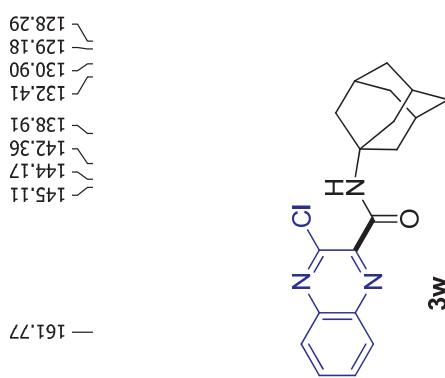
## Supporting Information



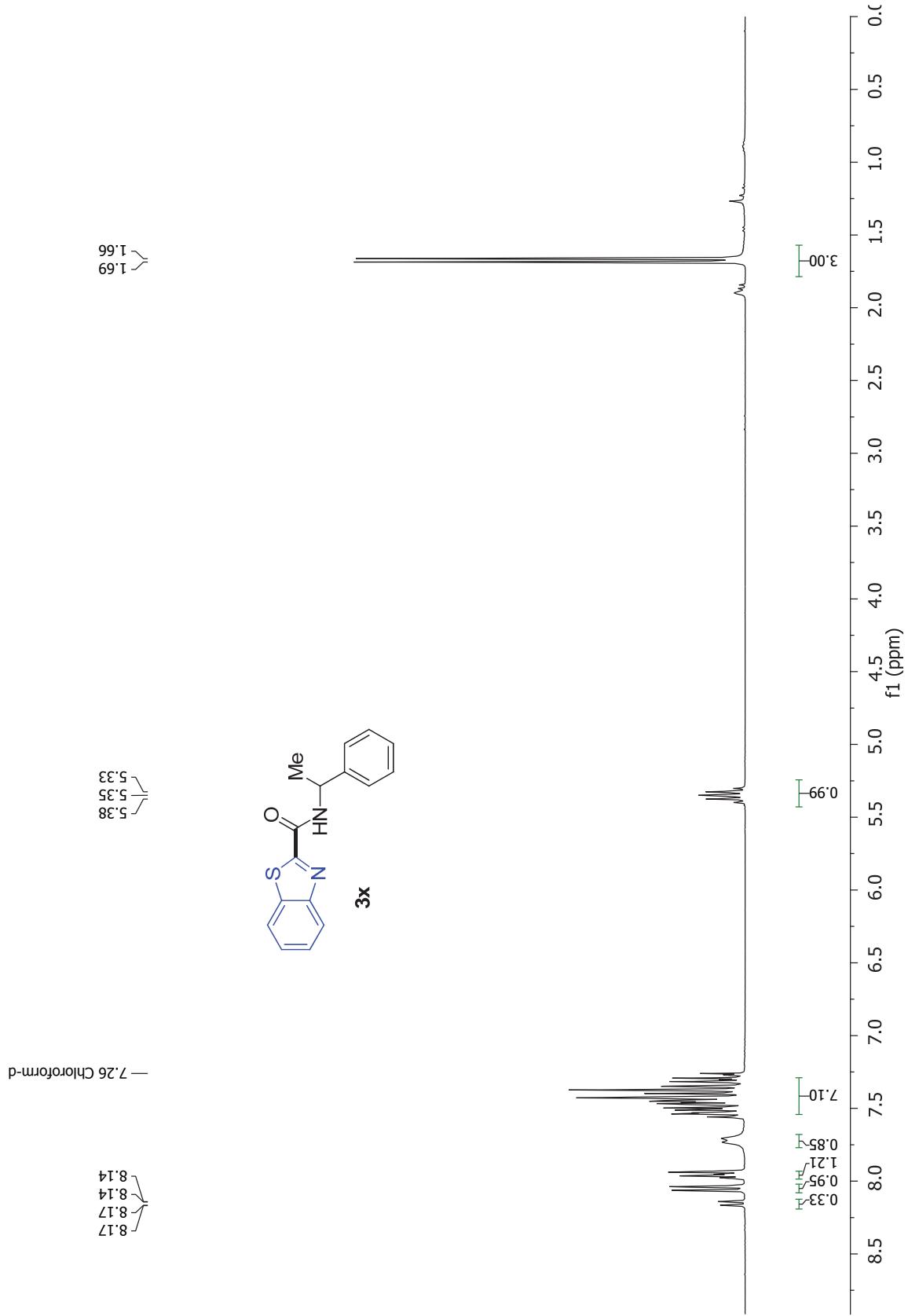
**Supporting Information**



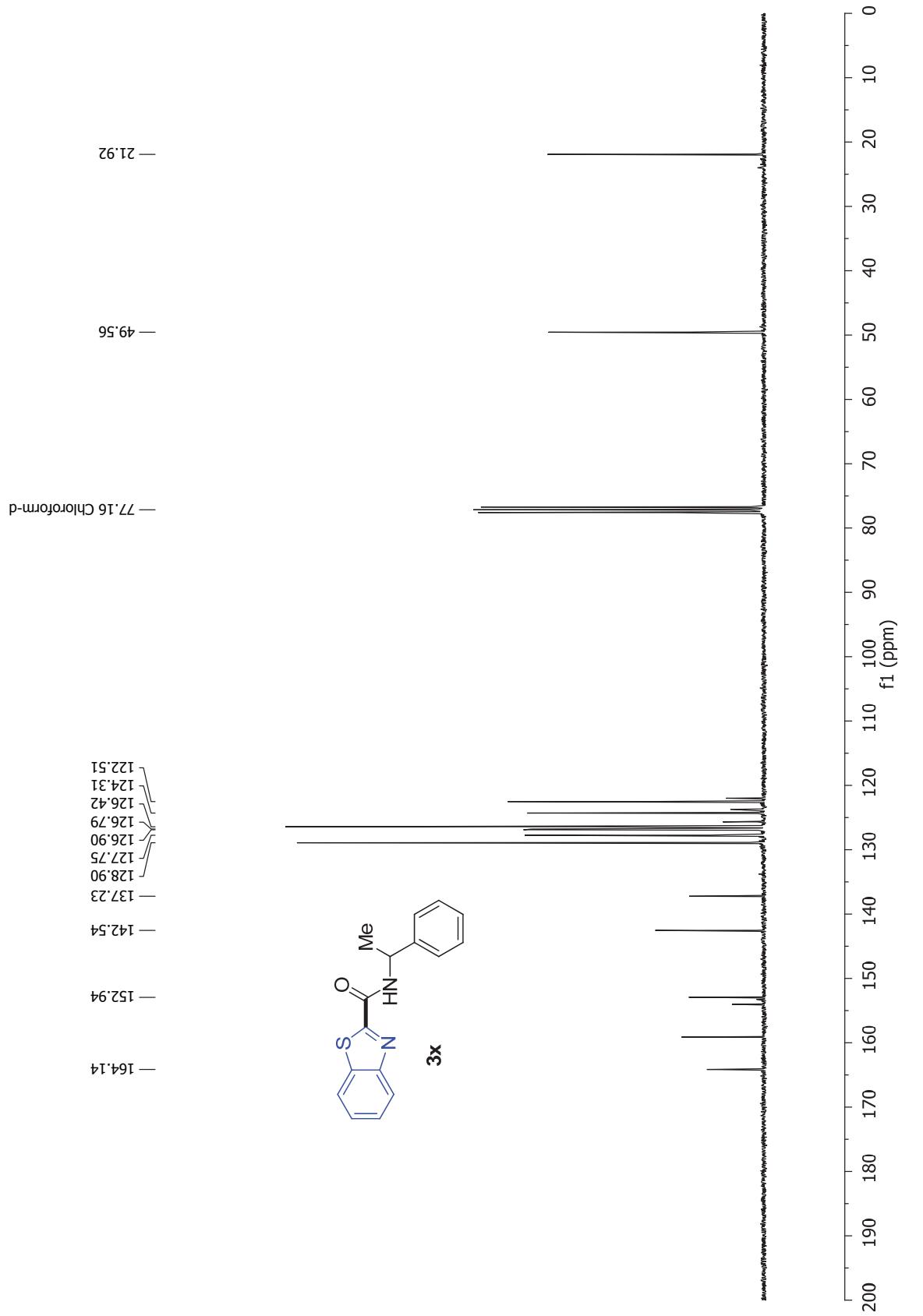
Supporting Information



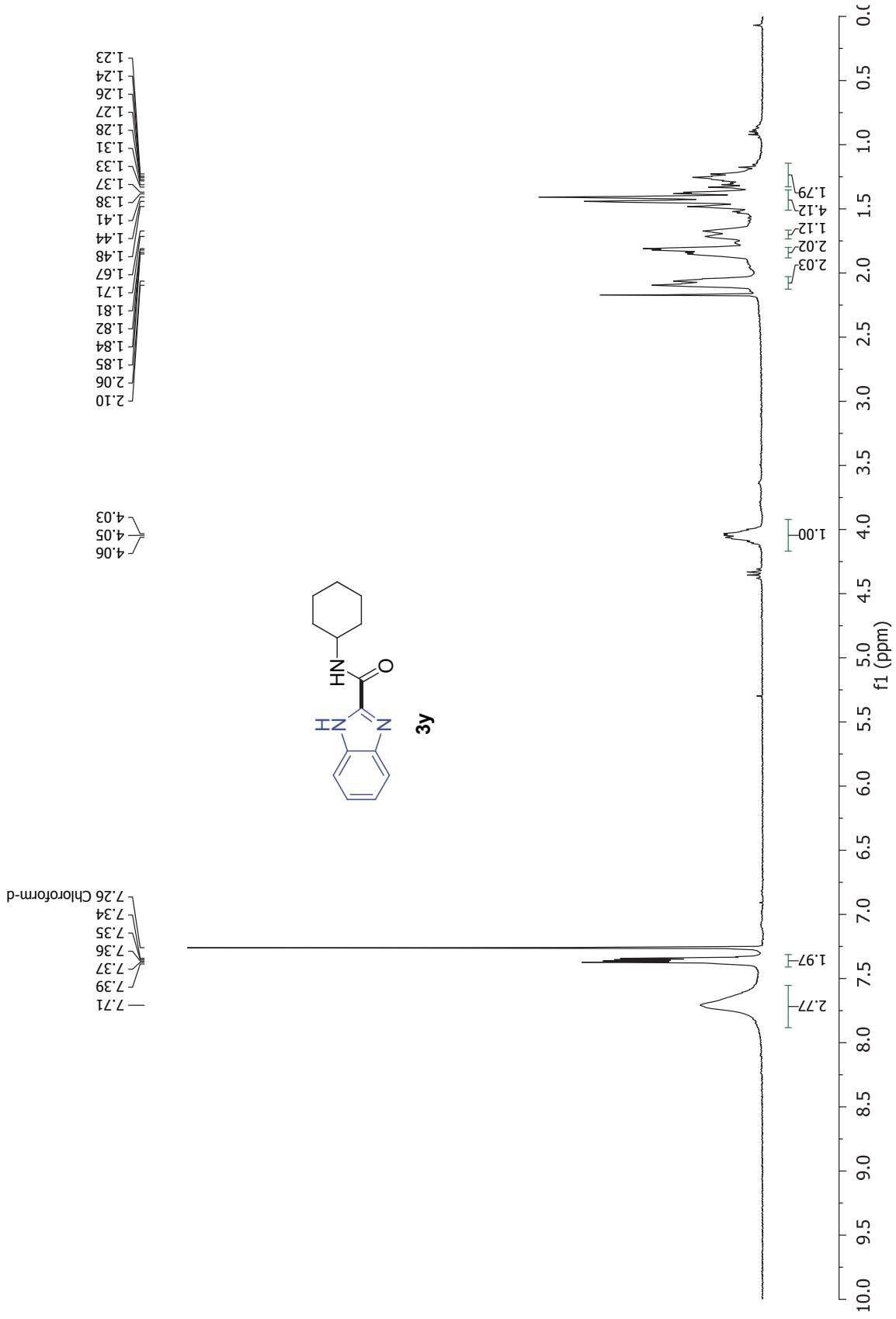
**Supporting Information**



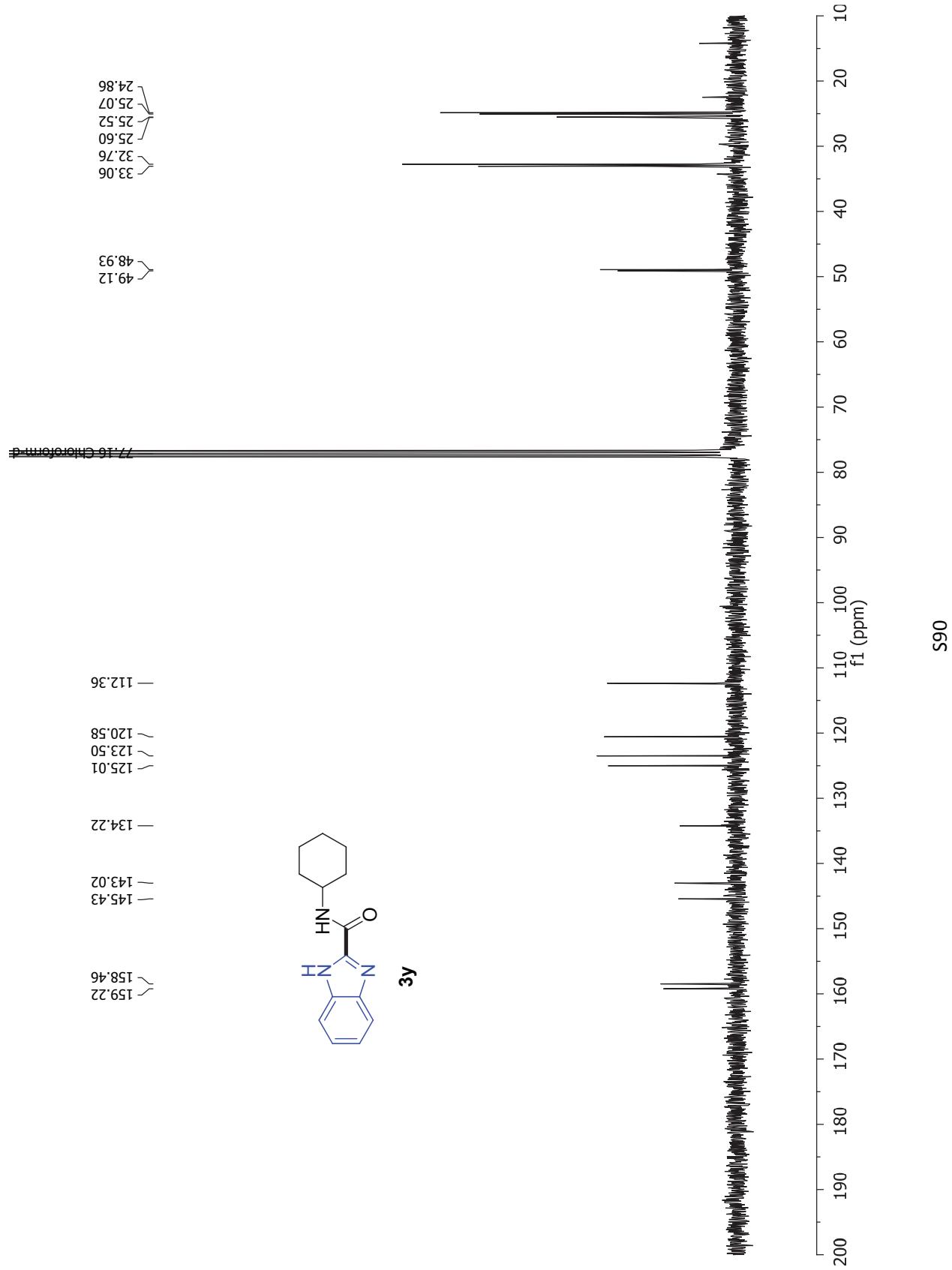
**Supporting Information**



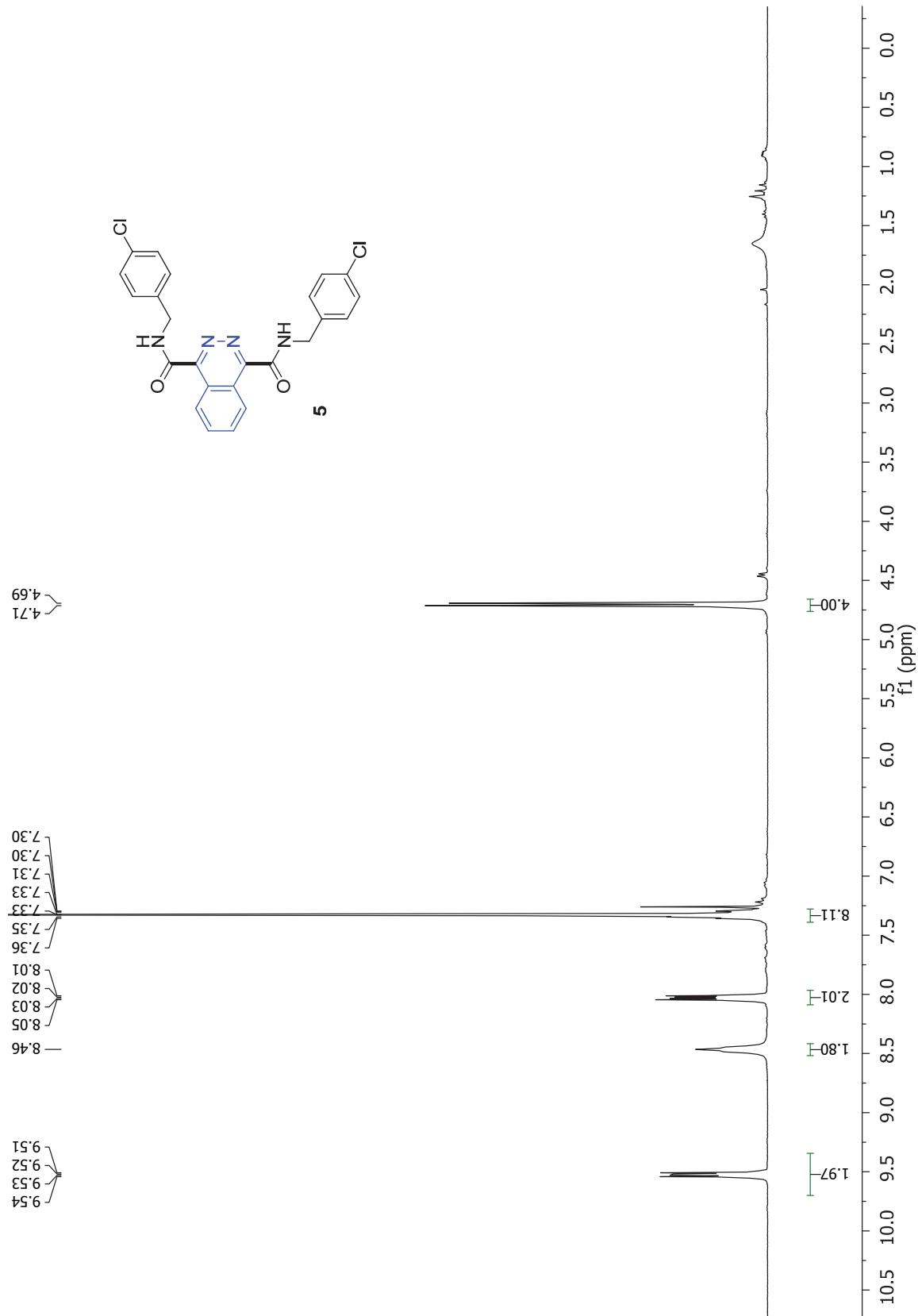
**Supporting Information**



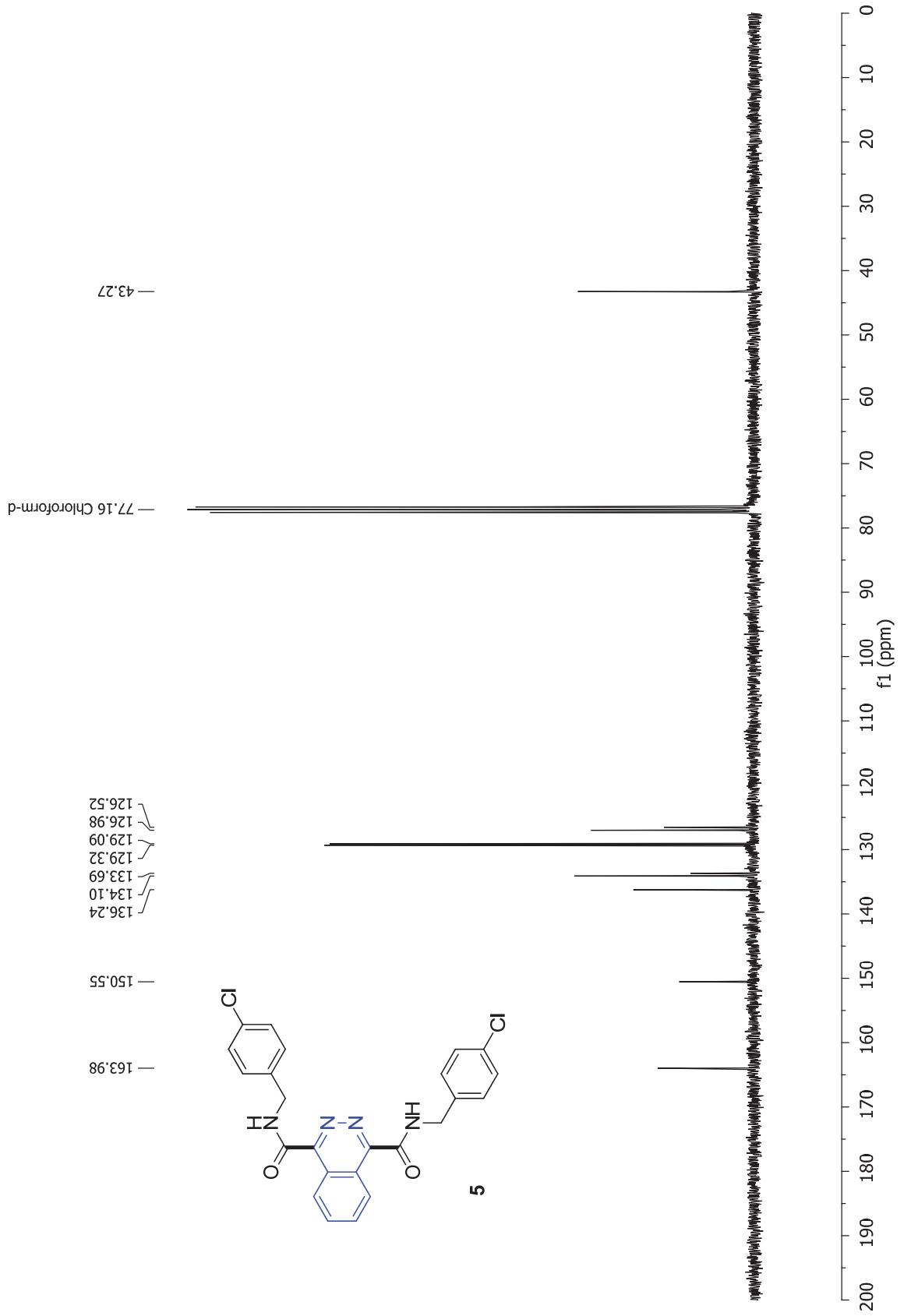
## Supporting Information



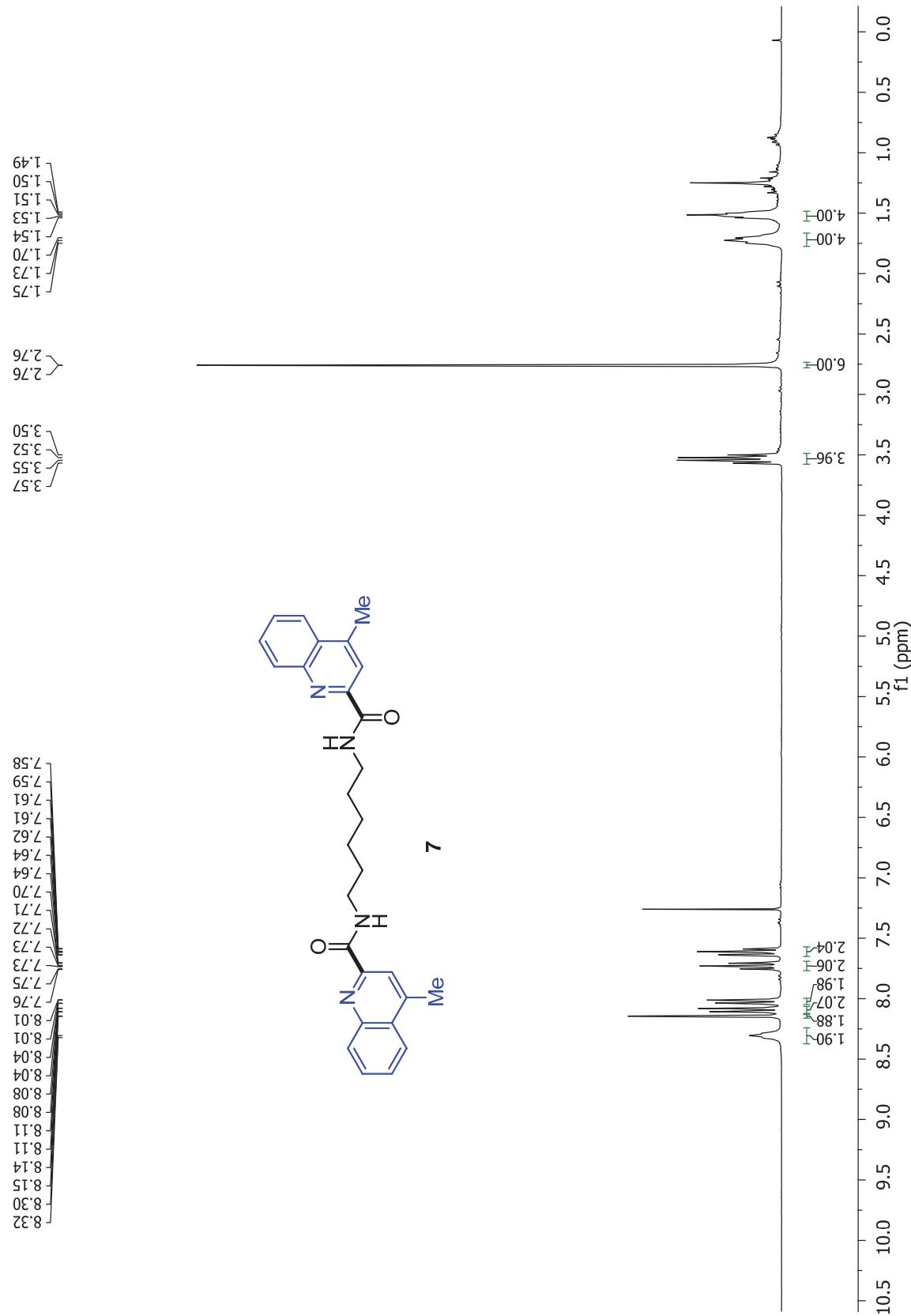
Supporting Information



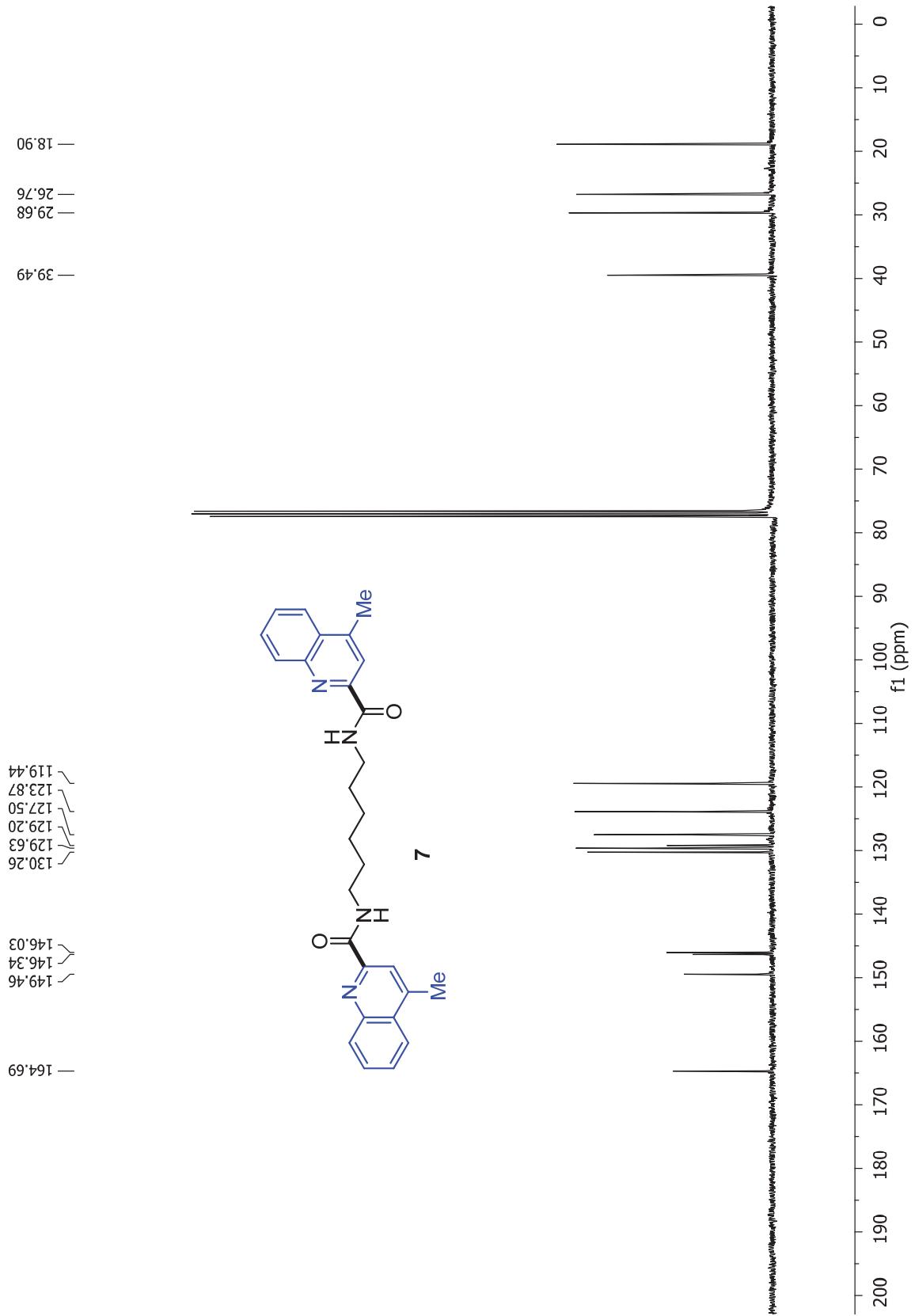
**Supporting Information**



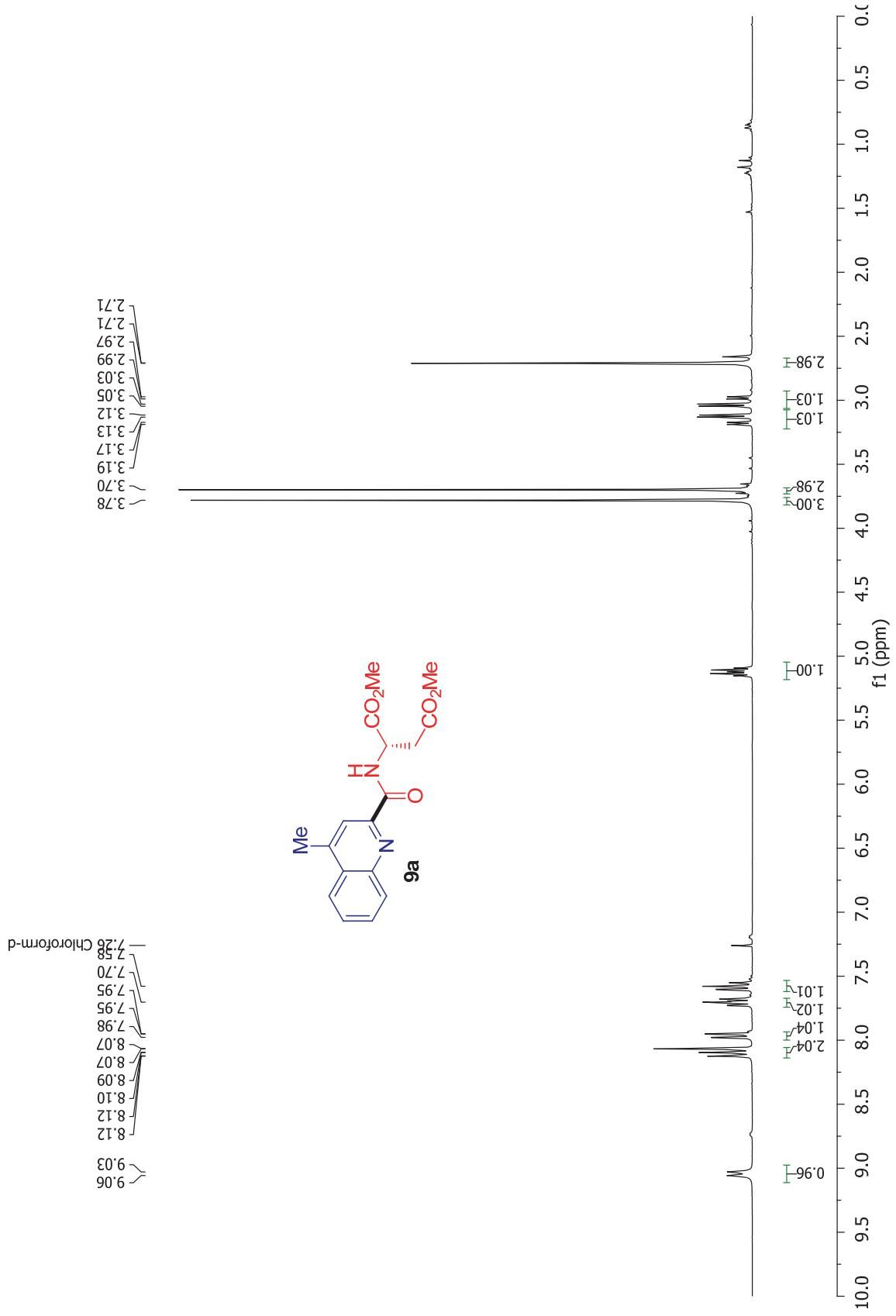
**Supporting Information**



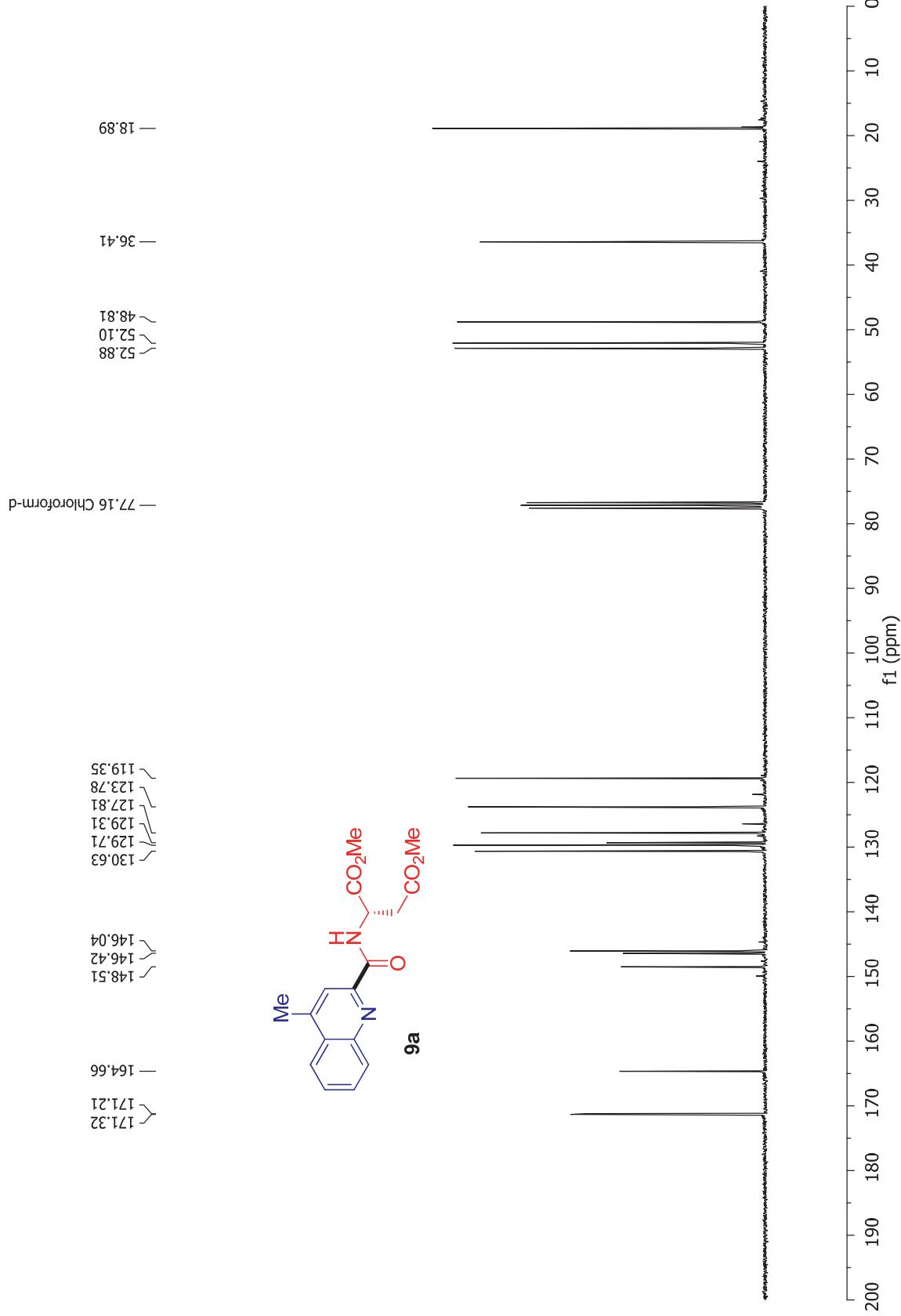
**Supporting Information**



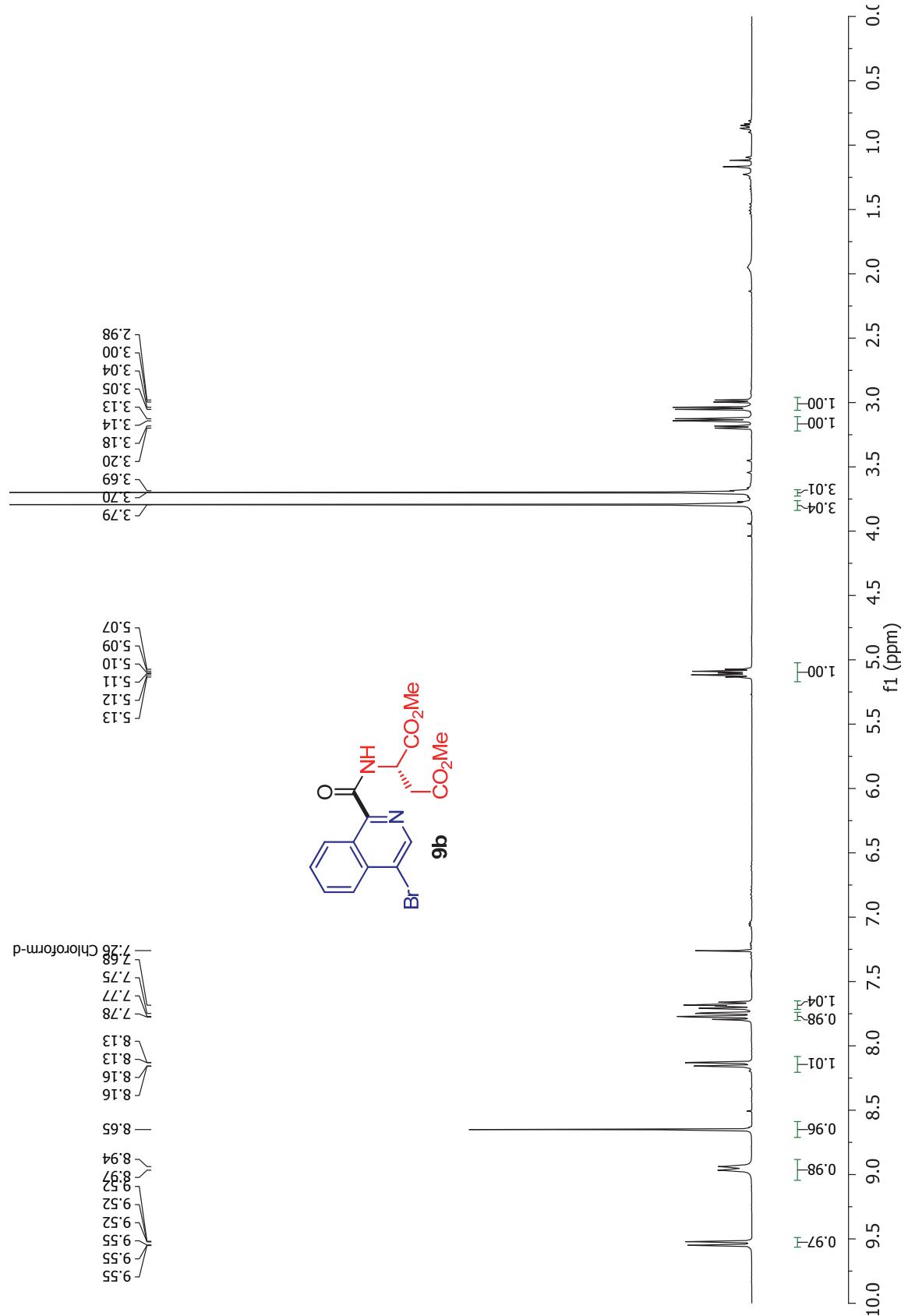
## Supporting Information



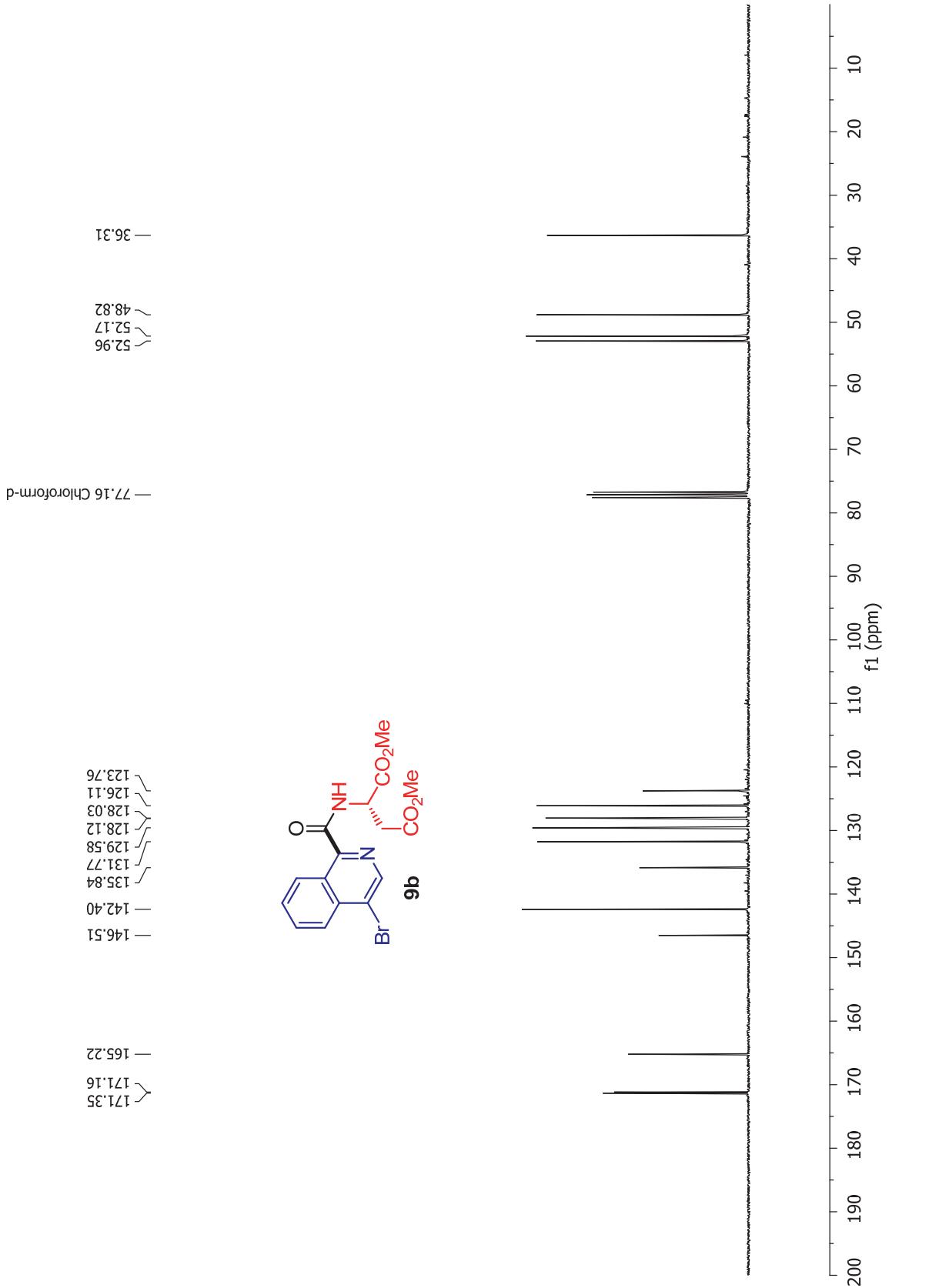
## Supporting Information



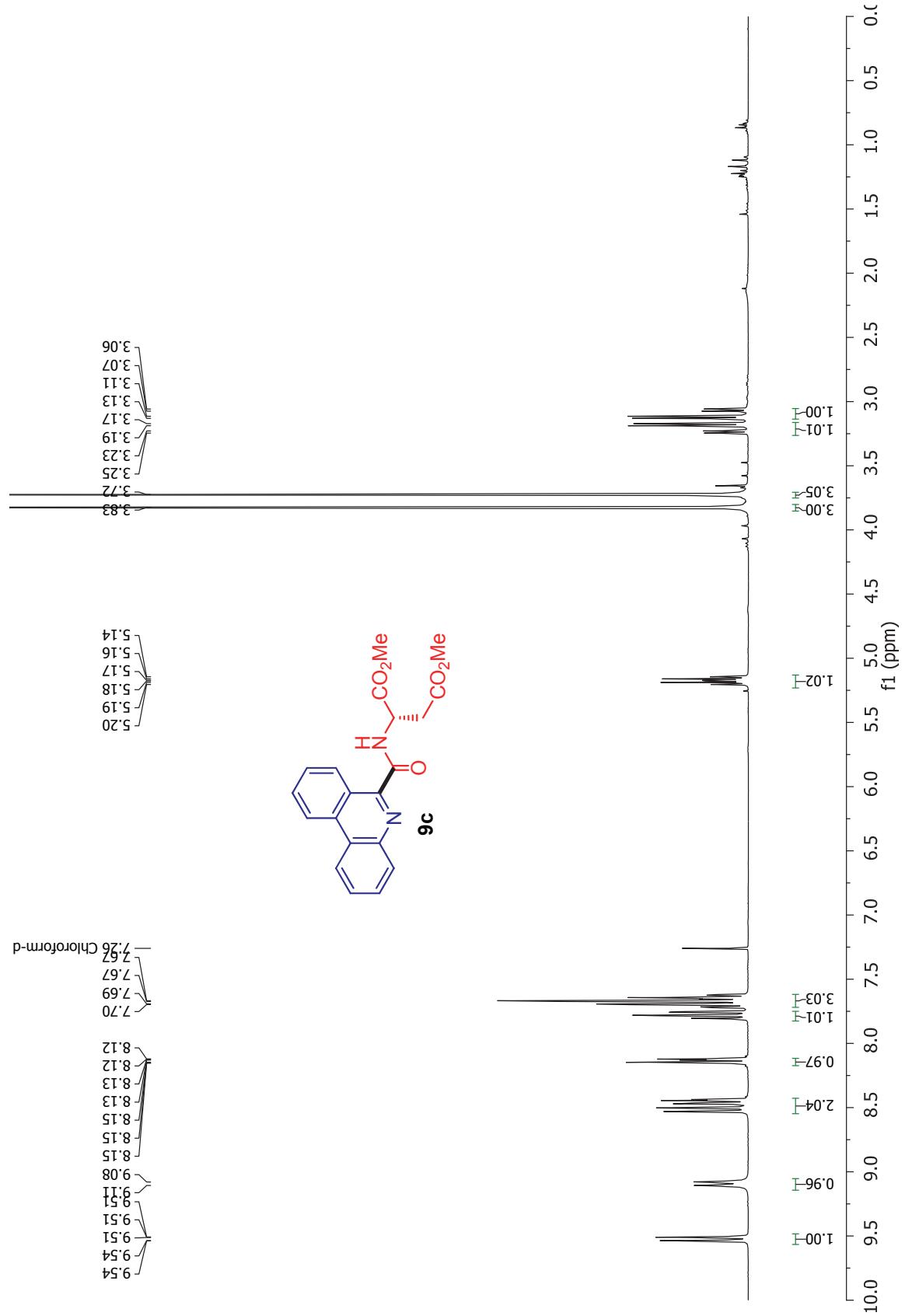
Supporting Information



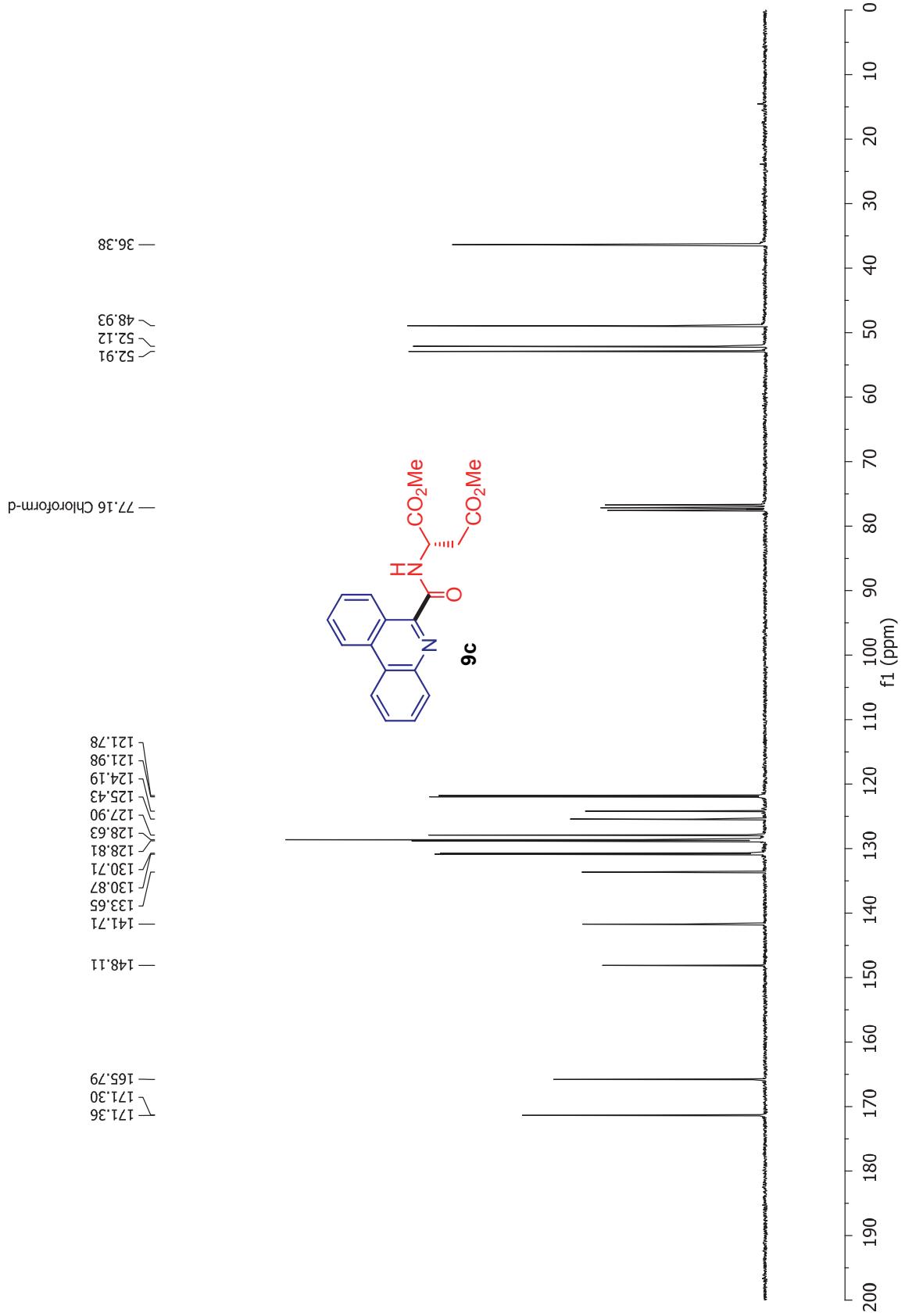
## Supporting Information



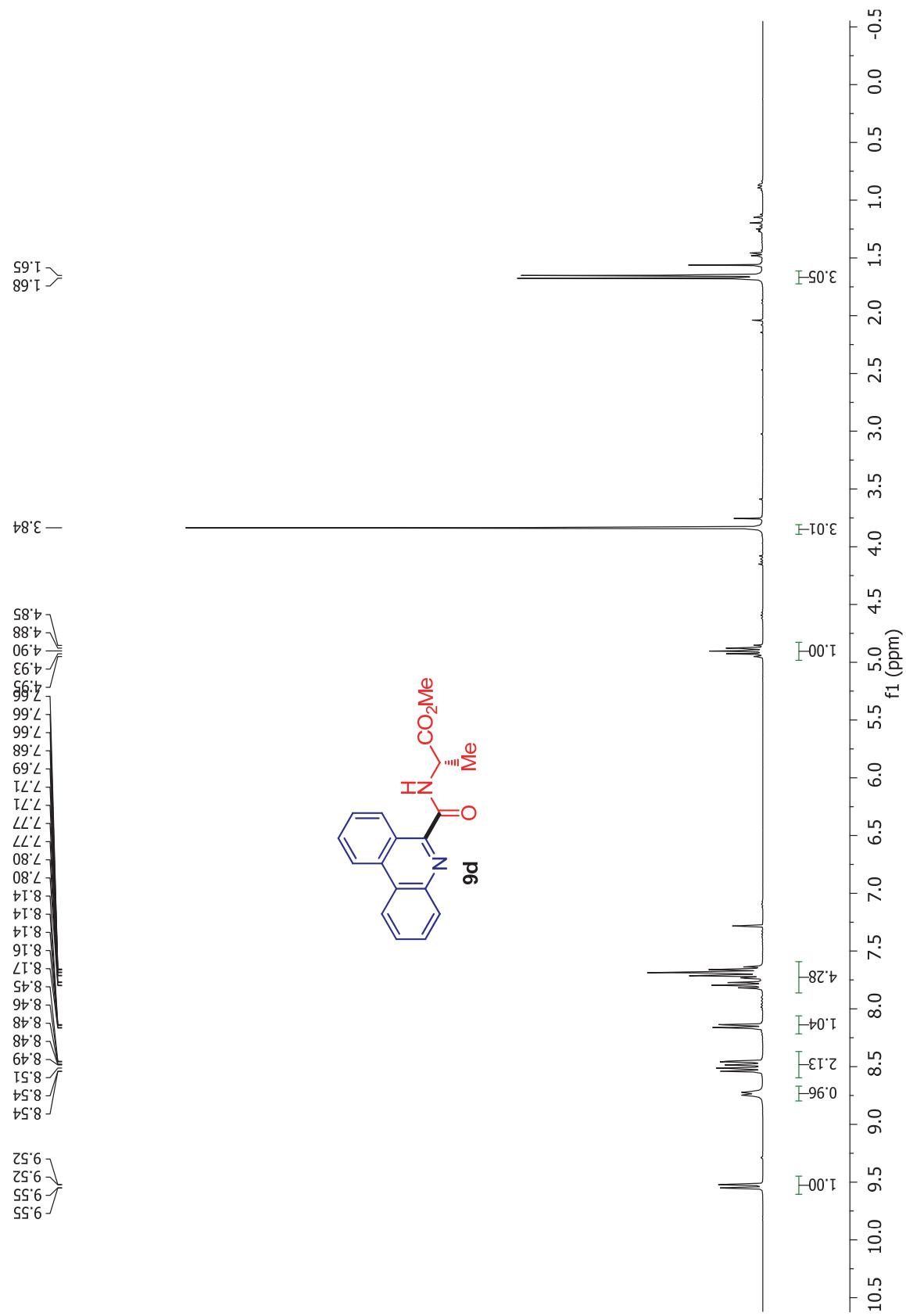
## Supporting Information



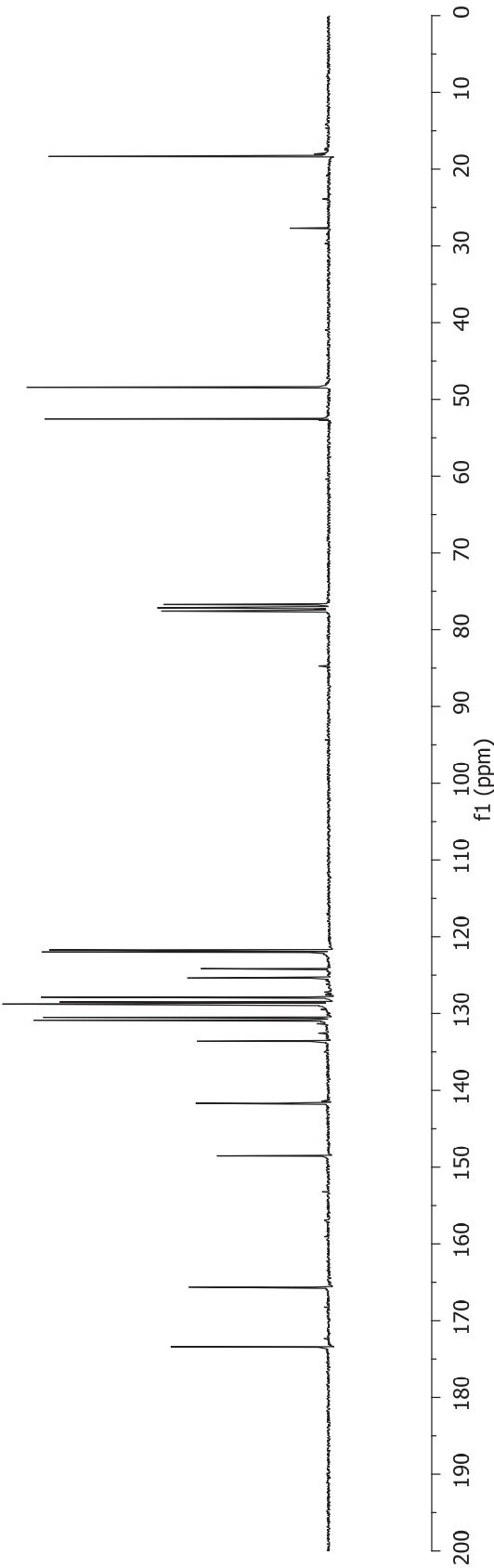
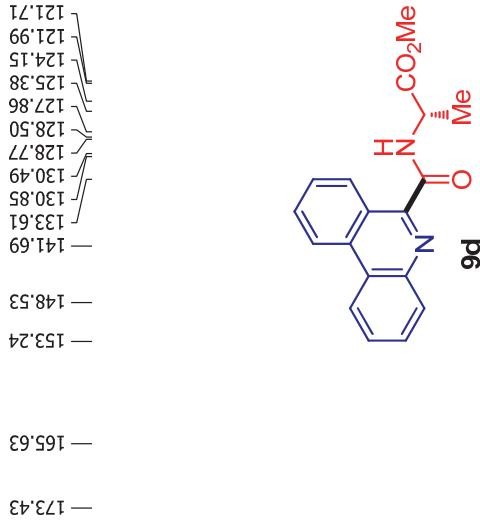
## Supporting Information



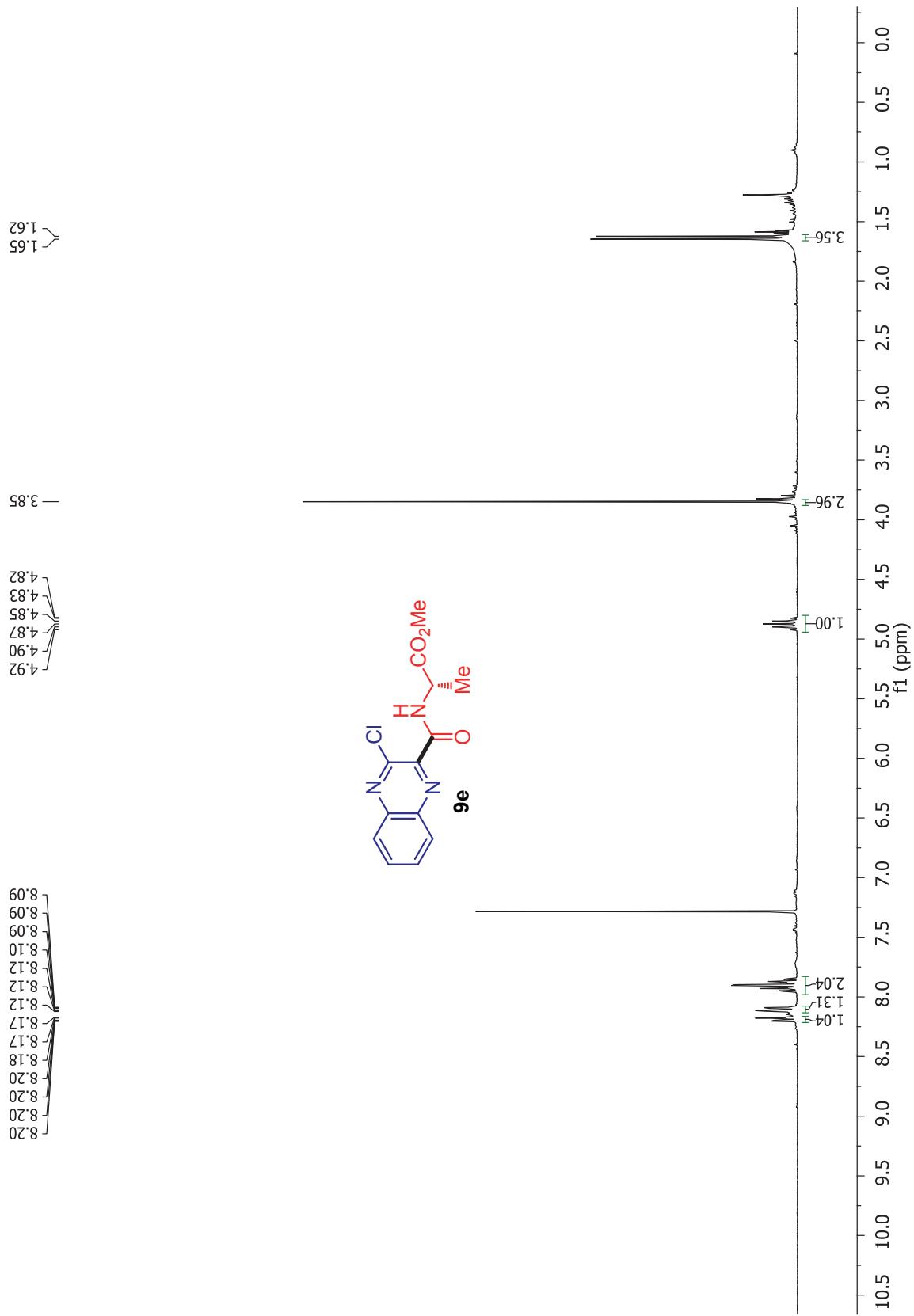
**Supporting Information**



## Supporting Information

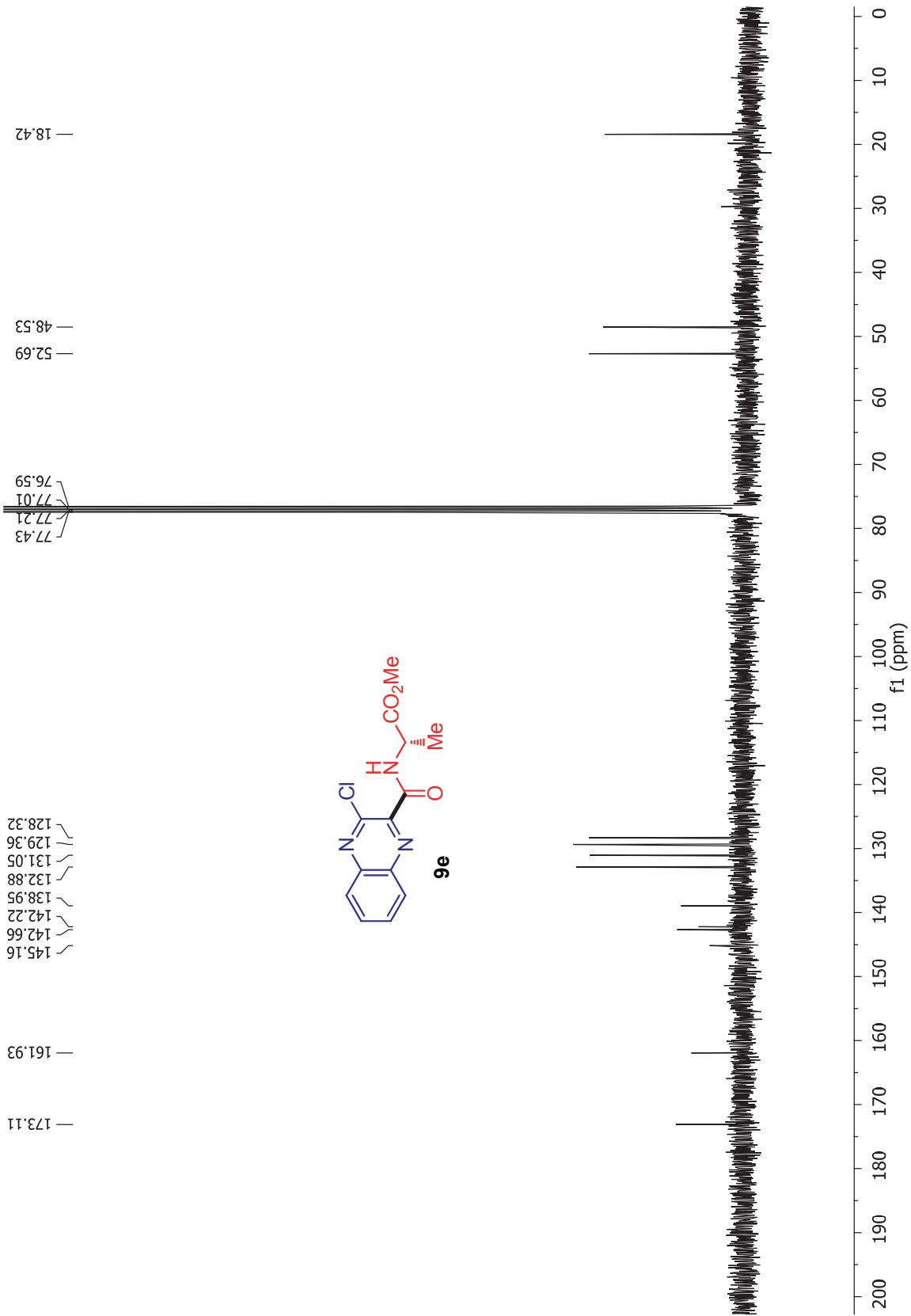


Supporting Information

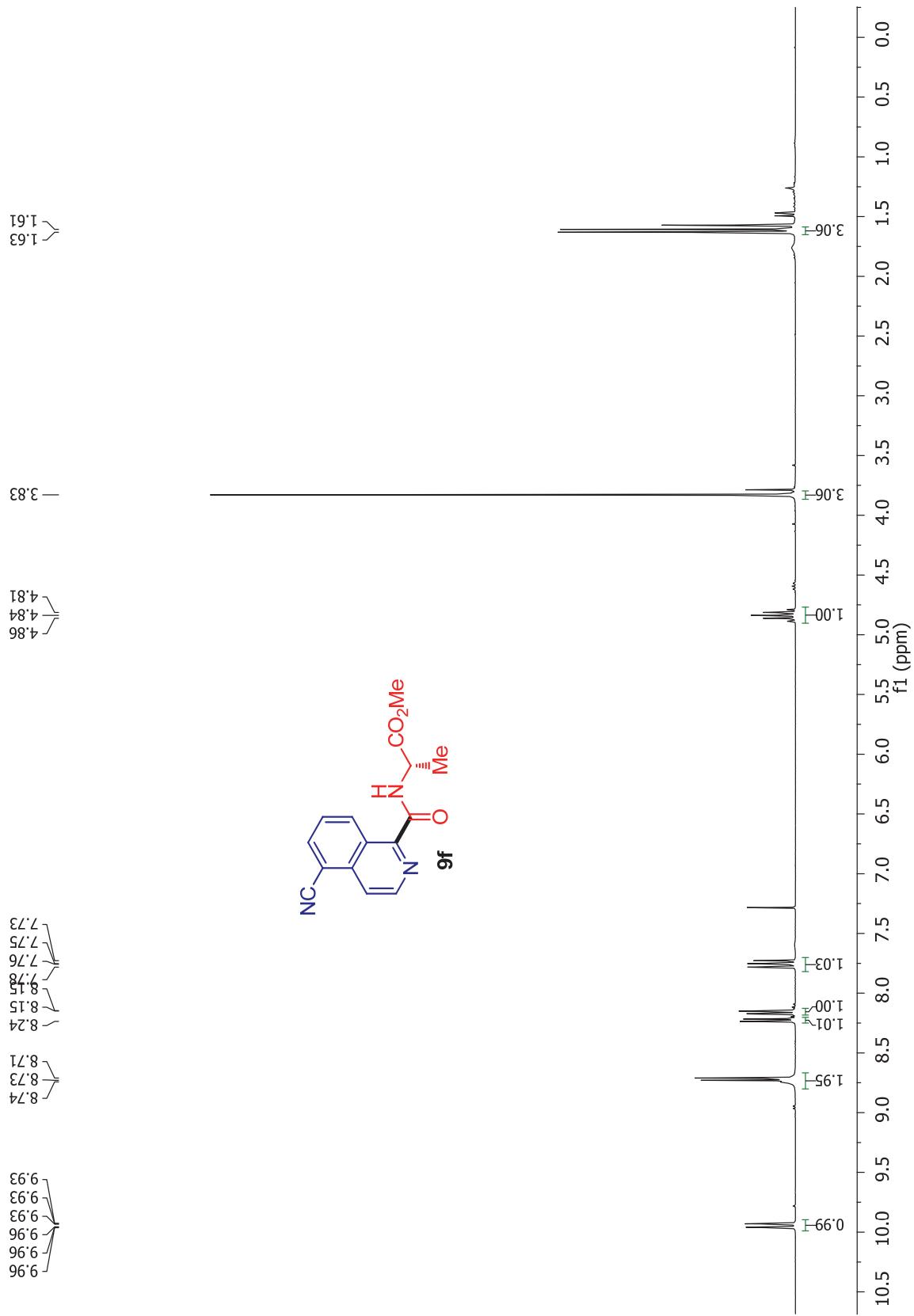


S103

**Supporting Information**



**Supporting Information**



**Supporting Information**

