

## Sulfo-click chemistry with $^{18}\text{F}$ -labeled thio acids

### SUPPLEMENTARY INFORMATION

Jenna Urkow, Cody Bergman, Frank Wuest\*

*\*Department of Oncology, University of Alberta, 11560 University Ave,  
Edmonton AB, T6G 1Z2, Canada*

**Material and methods.** All chemicals used obtained from Sigma-Aldrich<sup>®</sup>, with exception of thiolane-2,5-dione which was purchased from BOCSCI Inc..  $^1\text{H}$ -NMR and  $^{13}\text{C}$ -NMR spectra were recorded on an Agilent/Varian VNMRS three-channel 600 MHz spectrometer. Chemical shifts are given in ppm referenced to internal standards (s = singlet, bs = broad singlet, d = doublet, dd = doublet of doublet, t = triplet, m = multiplet). Mass spectra were recorded using a Micromass ZabSpec Hybrid Sector-TOF by positive mode electrospray ionization. Thin-layer chromatography (TLC) was monitored using HF254 silica gel. HPLC analyses were performed on a semi-preparative Luna C18 column (100 Å, 10 µm, 250 x 10 mm) or Jupiter C12 ((100 Å, 10 µm, 250 x 10 mm). Both columns were connected to their corresponding guard columns (Phenomenex Nucleosil LUNA (II) RP C18 pre-column (5 µm, 50x10mm) and Jupiter C12 pre-column (5 µm, 50x10mm)). UV detection was performed at 210 nm and 254 nm. Radioactivity detection was achieved using a well-scintillation NaI (TI) detector. [ $^{18}\text{F}$ ]Fluoride was produced by the  $^{18}\text{O}(\text{p},\text{n})^{18}\text{F}$  nuclear reaction through proton irradiation of enriched (98%)  $^{18}\text{O}$  water (3.0 ml, Rotem, Germany) using a TR19/9 cyclotron (Advanced Cyclotron Systems, Inc., Richmond, BC, Canada). [ $^{18}\text{F}$ ]SFB and 4- $^{18}\text{F}$ fluorobenzyl amine were prepared according to procedures published by our group.<sup>18,19</sup>

**4-Fluorobenzenecarbothioic S-acid (1).** A total of 0.71 mL (5.99 mmol) of 4-fluorobenzoyl chloride was added to a stirred solution of 300 mg (3.99 mmol) of thioacetamide in 3.0 mL of benzene. The mixture was heated to 30 °C and stirred for 3.5 h. After cooling to room temperature, 5 mL of 10% NaOH was added. The mixture was stirred for 30 min at room temperature. The mixture was then acidified using 10% HCl. The mixture was extracted twice with ethyl acetate. The organic layer was dried on  $\text{Na}_2\text{SO}_4$  and evaporated to give a yellow, crystal solid. The solid was washed with diethyl ether to extract the yellow impurity, giving a white solid. Yield: 73%;  $^1\text{H}$ -NMR (600 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.98-8.01 (m, 2H), 7.10-7.14 (m, 2H);  $m/z$  (ESI)  $\text{C}_7\text{H}_4\text{FOS}$  (M-H)<sup>-</sup> calcd. 155.2, found 154.8.

**4-[(4-Fluorobenzyl)amino]-4-oxobutanethioic S-acid (2).** 91  $\mu\text{L}$  (0.080 mmol) of 4-fluorobenzylamine was added to 3 mL of  $\text{CH}_2\text{Cl}_2$ . To this solution, 111  $\mu\text{L}$  (0.080 mmol) of triethylamine was added. In a separate vial, 278 mg (0.240 mmol) of thiolane-2,5-dione was dissolved in 1 mL of  $\text{CH}_2\text{Cl}_2$  and was then added to the reaction flask. The reaction was then set to reflux at 40  $^\circ\text{C}$  for 3 h. It was cooled to room temperature, and 20 mL of  $\text{CH}_2\text{Cl}_2$  was added. The mixture was then washed twice with 0.5 M citric acid and twice with water.

The organic layer was dried on  $\text{Na}_2\text{SO}_4$  and evaporated to dryness. The residue was dissolved in methanol, and filtered by vacuum filtration. The filtrate was evaporated, leaving a brown oil.

The oil was purified using column chromatography (50% EtOAc/hexane). Yield: 18%;  $^1\text{H-NMR}$  (600 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.30-7.16 (m, 2H), 6.96-6.91 (m, 2H), 4.34 (d, 2H,  $J=5.4$  Hz), 2.96 (t, 2H,  $J=6.6$  Hz), 2.52 (t, 2H,  $J=6.6$  Hz);  $m/z$  (ESI)  $\text{C}_{11}\text{H}_{11}\text{FNO}_2\text{S}$  (M-H) $^-$  calcd. 240.2, found 240.1.

**General procedure for sulfo-click reaction with 4-fluorobenzenecarbothioic S-acid (1) for the synthesis of reference compounds 3-6 and 12.** To a stirred solution of sulfonyl azide (1.0 equiv.) in methanol was added 2,6-lutidine (2.0 equiv.) and 4-fluorobenzenecarbothioic S-acid **1** (2.0 equiv.). The mixture was stirred at ambient temperature for 2-6 h. The solvent was removed in vacuo and purification of the residue by flash column chromatography gave the corresponding *N*-acylsulfonamide products.

**4-Fluoro-*N*-((2,4,6-triisopropylphenyl)sulfonyl)benzamide (3).** White solid; Yield: 41%;  $^1\text{H-NMR}$  (600 MHz,  $\text{CDCl}_3$ ):  $\delta$  8.65 (s, 1H), 7.69-7.72 (m, 2H), 7.11 (s, 2H), 7.03-7.05 (m, 2H), 4.21 (sept, 2H,  $J=6.6$  Hz), 2.81 (sept, 1H,  $J=6.6$  Hz), 1.20 (d, 12H,  $J=7.2$  Hz), 1.16 (d, 6H,  $J=7.2$  Hz);  $^{13}\text{C-NMR}$  (150.9 MHz,  $\text{CDCl}_3$ ):  $\delta$  165.8 (d,  $J=255.0$  Hz), 163.5, 154.2, 151.5, 131.1, 130.2 (d,  $J=10.56$ ), 127.6, 124.2, 116.3 (d,  $J=22.6$  Hz), 34.2, 29.7, 24.6, 23.5;  $m/z$  (ESI)  $\text{C}_{22}\text{H}_{29}\text{FNO}_3\text{S}$  (M+H) $^+$  calcd. 406.4, found 406.2.

**4-(*N*-(4-Fluorobenzoyl)sulfamoyl)benzoic acid (4).** White solid. Yield: 60%;  $^1\text{H-NMR}$  (600 MHz,  $\text{CD}_3\text{OD}$ ):  $\delta$  8.10-8.05 (m, 4H), 7.85-7.82 (m, 2H), 7.09-7.06 (m, 2H);  $^{13}\text{C-NMR}$  (150.9 MHz,  $\text{CD}_3\text{OD}$ ):  $\delta$  167.2, 166.9, 165.5 (d,  $J=252.00$  Hz), 154.7, 144.6, 134.8, 130.8 (d,  $J=9.05$  Hz), 129.5, 127.8, 115.0 (d,  $J=21.13$  Hz);  $m/z$  (ESI)  $\text{C}_{14}\text{H}_9\text{FNO}_5\text{S}$  (M-H) $^-$  calcd. 322.3, found 322.0.

***N*-((4-Acetamidophenyl)sulfonyl)-4-fluorobenzamide (5).** White solid. Yield: 76%;  $^1\text{H-NMR}$  (600 MHz,  $\text{DMSO-}d_6$ ):  $\delta$  12.48 (s, 1H), 10.40 (s, 1H), 7.92-7.96 (m, 4H), 7.79-7.81 (m, 2H), 7.31-7.34 (m, 2H), 2.09 (s, 3H);  $^{13}\text{C-NMR}$  (150.9 MHz,  $\text{DMSO-}d_6$ ):  $\delta$  169.6, 165.4 (d,  $J=252.00$  Hz), 164.7, 144.3, 133.2, 131.8 (d,  $J=10.56$  Hz), 129.6, 128.6, 118.8, 116.2 (d,  $J=22.6$  Hz), 24.6;  $m/z$  (ESI)  $\text{C}_{15}\text{H}_{14}\text{FN}_2\text{O}_4\text{S}$  (M+H) $^+$  calcd. 337.3, found 337.2.

**4-Fluoro-*N*-tosylbenzamide (6).** White solid. Yield: 76%; <sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>): δ 9.01 (s, 1H), 7.95 (d, 2H, J=8.4 Hz), 7.73-7.75 (m, 2H), 7.27 (d, 2H, J=8.4 Hz), 7.00-7.03 (m, 2H), 2.35 (s, 3H); <sup>13</sup>C-NMR (150.9 MHz, CDCl<sub>3</sub>): δ 165.9 (d, J=255.02 Hz), 163.0, 145.4, 135.3, 130.4 (d, J=22.56 Hz), 129.7, 128.7, 127.4, 116.2 (d, J=22.64 Hz), 21.7; *m/z* (ESI) C<sub>14</sub>H<sub>13</sub>FNO<sub>3</sub>S (M+H)<sup>+</sup> calcd. 294.3, found 294.1.

**4-Fluoro-*N*-((4-(5-(*p*-tolyl)-3-(trifluoromethyl)-1H-pyrazol-1-yl)phenyl)sulfonyl)benzamide (12).** White solid; Yield: 85%; <sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>): δ 8.52 (s, 1H), 8.04 (d, 2H, J=12.0 Hz), 7.67-7.70 (m, 2H), 7.42 (d, 2H, J=12.0 Hz), 7.03-7.08 (m, 6H), 6.65 (s, 1H), 2.30 (s, 3H); <sup>13</sup>C-NMR (150.9 MHz, CDCl<sub>3</sub>): δ 166.0 (d, J=256.53 Hz), 163.0, 145.4, 144.3 (q, J=39.23 Hz), 143.6, 139.9, 137.6, 130.5 (d, J=10.56 Hz), 129.9, 129.8, 128.7, 127.2 (d, J=3.02 Hz), 125.6, 125.1, 121.0 (q, J=268.60 Hz), 116.2 (d, J=22.5 Hz), 106.6, 21.4; *m/z* (ESI) C<sub>24</sub>H<sub>18</sub>F<sub>4</sub>N<sub>3</sub>O<sub>3</sub>S (M+H)<sup>+</sup> calcd. 504.5, found 504.3.

**General procedure for sulfo-click reaction with 4-[(4-Fluorobenzyl)amino]-4-oxobutanethioic *S*-acid (2) for the synthesis of reference compounds 7-10 and 13.** To a stirred solution of sulfonyl azide (1.0 equiv.) in methanol was added 2,6-lutidine (2.0 equiv.) and 4-[(4-fluorobenzyl)amino]-4-oxobutanethioic *S*-acid (2) (2.0 equiv.). The mixture was stirred at ambient temperature for 2-6 h. The solvent was removed in vacuo and purification of the residue by flash column chromatography gave the corresponding *N*-acylsulfonamide products.

***N*<sup>1</sup>-(4-Fluorobenzyl)-*N*<sup>4</sup>-((2,4,6-triisopropylphenyl)sulfonyl)succinamide (7).** Yellow-brown oil. Yield: 43%; <sup>1</sup>H-NMR (600 MHz, CD<sub>3</sub>OD): δ 7.20-7.16 (m, 2H) 7.08 (s, 2H), 6.91-6.87 (m, 2H), 4.31 (s, 2H), 4.21-4.27 (sep, 2H, J=7.2 Hz), 3.38 (t, 2H, J=6.6 Hz), 2.8 (sep, 1H, J=7.2 Hz), 2.67 (t, 2H, J=6.6 Hz), 1.15 (d, 6H, J=7.2 Hz), 1.09 (d, 12H, J=7.2 Hz); <sup>13</sup>C-NMR (150.9 MHz, CD<sub>3</sub>OD): δ 168.6, 151.9, 148.9, 136.8, 129.6 (d, J=9.05 Hz), 123.0, 114.8 (d, J=22.6 Hz), 44.1, 34.0, 30.3, 29.2, 28.4, 23.6, 22.7; *m/z* (ESI) C<sub>26</sub>H<sub>36</sub>FN<sub>2</sub>O<sub>4</sub>S (M+H)<sup>+</sup> calcd. 491.6, found 491.2.

**4-(*N*-(4-((4-Fluorobenzyl)amino)-4-oxobutanoyl)sulfamoyl)benzoic acid (8).** Yellow-brown oil. Yield 43%; <sup>1</sup>H-NMR (600 MHz, CD<sub>3</sub>OD): δ 8.17 (m, 4H) 7.94 (m, 2H), 7.18 (m, 2H), 4.28 (d, 2H, J=6.0 Hz), 3.52 (t, 2H, J=6.6 Hz), 2.74 (t, 2H, J=6.6 Hz); <sup>13</sup>C-NMR (150.9 MHz, CD<sub>3</sub>OD): δ 174.2, 168.2, 167.5, 163.2 (d, J=247.06), 143.9, 139.6, 130.8, 129.8, 129.0, 125.8, 115.3 (d, J=22.6), 46.3, 34.1, 33.7; *m/z* (ESI) C<sub>18</sub>H<sub>18</sub>FN<sub>2</sub>O<sub>6</sub>S (M+H)<sup>+</sup> calcd. 409.4, found 409.1.

***N*<sup>1</sup>-((4-Acetamidophenyl)sulfonyl)-*N*<sup>4</sup>-(4-fluorobenzyl)succinamide (9).** Yellow-brown oil. Yield 55%; <sup>1</sup>H-NMR (600 MHz, CD<sub>3</sub>OD): δ 7.72 (m, 4H) 7.32 (m, 2H), 7.05 (m, 2H), 4.35 (d, 2H, J=6.0 Hz), 3.58 (t, 2H, J=6.6 Hz), 2.79 (t, 2H, J=6.6 Hz), 2.56 (s, 3H); <sup>13</sup>C-NMR (150.9 MHz, CD<sub>3</sub>OD): δ 170.6, 166.4, 164.2, 163.4 (d, J=252.06), 144.4, 139.1, 135.4, 129.4, 128.8, 119.2, 115.0 (d, J=22.6), 43.8, 34.1, 33.9, 24.0, *m/z* (ESI) C<sub>19</sub>H<sub>21</sub>FN<sub>3</sub>O<sub>5</sub>S (M+H)<sup>+</sup> calcd. 422.4, found 422.2.

***N*<sup>1</sup>-(4-Fluorobenzyl)-*N*<sup>4</sup>-tosylsuccinamide (10).** Yellow oil. Yield 55%; <sup>1</sup>H-NMR (600 MHz, CD<sub>3</sub>OD): δ 7.55 (d, 2H, J=8.4 Hz) 7.19 (d, 2H, J=8.4 Hz), 7.13-7.15 (m, 2H), 6.87- 6.90 (m, 2H), 4.29 (s, 2H), 3.45 (t, 2H, J=6.6 Hz), 2.68 (t, 2H, J=6.6 Hz), 2.31 (s, 3H); <sup>13</sup>C-NMR (150.9 MHz, CD<sub>3</sub>OD): δ 175.5, 169.1, 162.2 (d, J=244.46), 142.4, 140.9, 129.5, 129.0 (d, J=9.05 Hz), 125.7, 114.7 (d, J=22.5 Hz), 44.3, 30.5, 30.2, 20.0; *m/z* (ESI) C<sub>18</sub>H<sub>20</sub>FN<sub>2</sub>O<sub>4</sub>S (M+H)<sup>+</sup> calcd. 379.4, found 379.3.

***N*<sup>1</sup>-(4-Fluorobenzyl)-*N*<sup>4</sup>-((4-(5-(*p*-tolyl)-3-(trifluoromethyl)-1H-pyrazol-1-yl)phenyl)sulfonyl)succinamide (13).** Yellow-brown oil. Yield: 60%; <sup>1</sup>H-NMR (600 MHz, CDCl<sub>3</sub>): δ 7.82 (m, 2H), 7.37-7.33 (m, 2H), 7.12-6.90 (m, 8H), 6.66 (s, 1H), 4.32 (d, 2H, J=5.4 Hz) 3.66 (t, 2H, J=5.4 Hz), 2.96 (t, 2H, J=6.6 Hz), 2.30 (s, 3H); <sup>13</sup>C-NMR (150.9 MHz, CDCl<sub>3</sub>): δ 174.4, 168.4, 163.3, 145.2, 143.9, 143.1, 141.7, 139.6, 129.8 (d, J=9.05 Hz), 129.7, 128.7, 127.4, 125.8, 125.2, 115.8 (d, J=21.13 Hz), 106.0, 45.5, 31.1, 29.3, 21.3; *m/z* (ESI) C<sub>28</sub>H<sub>23</sub>F<sub>4</sub>N<sub>4</sub>O<sub>4</sub>S (M-H)<sup>-</sup> calcd. 587.6, found 587.2.

**General procedure for one pot/three-component procedure for radiosynthesis of compounds [<sup>18</sup>F]3-<sup>[18F]</sup>6 and [<sup>18</sup>F]12.** In an eppendorf tube, 2.5 mg of NaSH was dissolved in 200 μL of deionized water. In a separate eppendorf tube, 5 mg of sulfonyl azide was dissolved in 200 μL of CH<sub>3</sub>CN. Next, 20-30 MBq of the [<sup>18</sup>F]SFB was added to the NaSH solution, and the sulfonyl azide solution was added to the reaction mixture. This was then stirred at 50 °C for 30 min. Product formation was monitored with radio-TLC analysis. Radiochemical conversions (RCCs) refer to the percentage of radiolabeled products [<sup>18</sup>F]3-<sup>[18F]</sup>6 [<sup>18</sup>F]12 and present in the reaction mixture as determined by radio-TLC analysis (10% MeOH/CH<sub>2</sub>Cl<sub>2</sub>).

**4-<sup>[18F]</sup>Fluoro-*N*-((2,4,6-triisopropylphenyl)sulfonyl)benzamide ([<sup>18</sup>F]3).** RCC: 77%. Radio-TLC (10% MeOH/CH<sub>2</sub>Cl<sub>2</sub>): R<sub>f</sub> = 0.75.

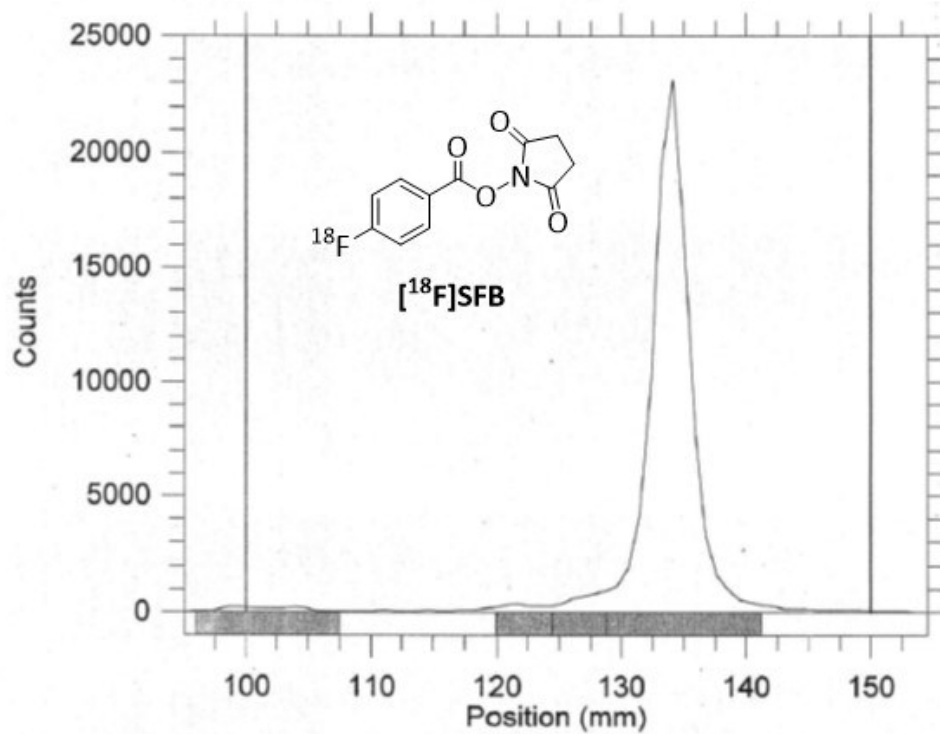
**4-(*N*-(4-<sup>[18F]</sup>Fluorobenzoyl)sulfamoyl)benzoic acid ([<sup>18</sup>F]4).** RCC: 76%. Radio-TLC (10% MeOH/CH<sub>2</sub>Cl<sub>2</sub>): R<sub>f</sub> = 0.5.

***N*-((4-Acetamidophenyl)sulfonyl)-4-<sup>[18F]</sup>fluorobenzamide ([<sup>18</sup>F]5).** RCC: 70%. Radio-TLC (10% MeOH/CH<sub>2</sub>Cl<sub>2</sub>): R<sub>f</sub> = 0.6.

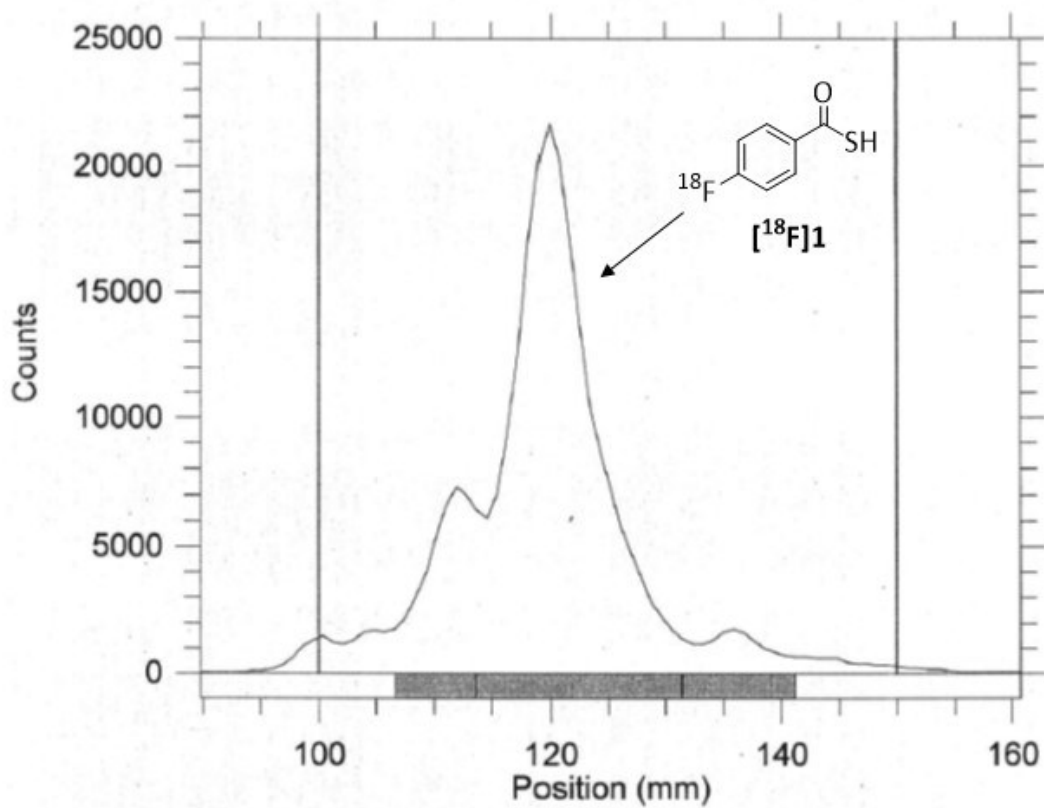
**4-<sup>[18F]</sup>Fluoro-*N*-tosylbenzamide ([<sup>18</sup>F]6).** RCC: 99%. Radio-TLC (10% MeOH/CH<sub>2</sub>Cl<sub>2</sub>): R<sub>f</sub> = 0.7.

**4-<sup>[18F]</sup>Fluoro-*N*-((4-(5-(*p*-tolyl)-3-(trifluoromethyl)-1H-pyrazol-1-yl)phenyl)sulfonyl)benzamide ([<sup>18</sup>F]12).** RCC: 67%. Radio-TLC (10% MeOH/CH<sub>2</sub>Cl<sub>2</sub>): R<sub>f</sub> = 0.3.

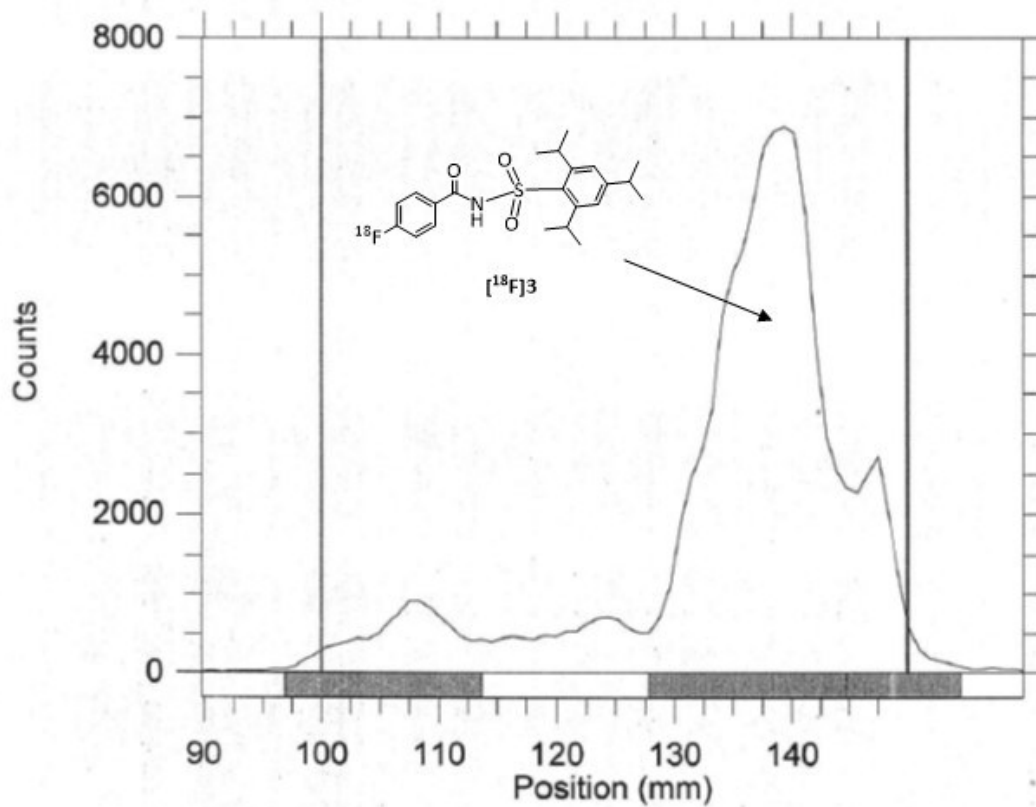
Reg	(mm) Start	(mm) Stop	(mm) Centroid	RF	Region Counts	Region CPM	% of Total	% of ROI
Rgn 1	96.0	107.5	101.3	0.027	1805.0	1805.0	1.79	1.82
Rgn 2	119.9	125.2	122.2	0.444	1795.0	1795.0	1.78	1.81
Rgn 3	124.4	129.7	127.1	0.542	3788.0	3788.0	3.76	3.82
Rgn 4	128.8	141.2	134.0	0.680	91801.0	91801.0	91.05	92.55
4 Peaks					99189.0	99189.0	98.37	100.00



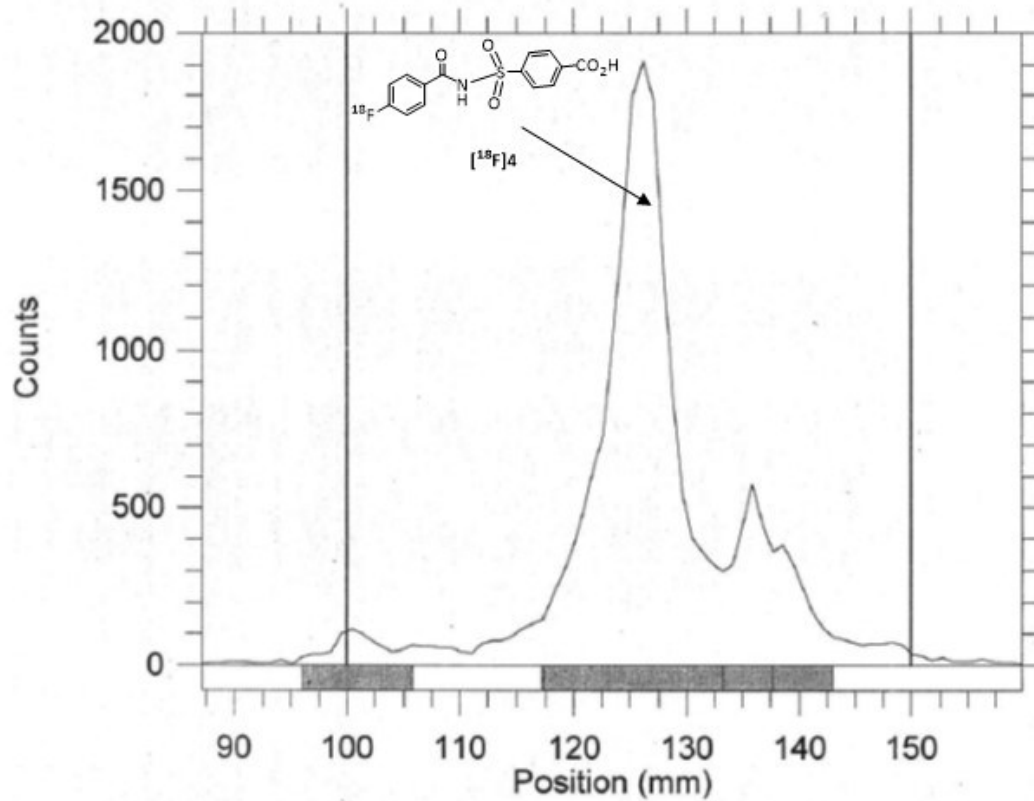
Reg	(mm) Start	(mm) Stop	(mm) Centroid	RF	Region Counts	Region CPM	% of Total	% of ROI
Rgn 1	106.6	114.6	111.1	0.221	43475.0	43475.0	16.28	17.05
Rgn 2	113.7	131.5	120.6	0.413	197787.0	197787.0	74.07	77.57
Rgn 3	131.5	141.2	135.5	0.711	13715.0	13715.0	5.14	5.38
3 Peaks					254977.0	254977.0	95.48	100.00



Reg	(mm) Start	(mm) Stop	(mm) Centroid	RF	Region Counts	Region CPM	% of Total	% of ROI
Rgn 1	96.9	113.7	106.8	0.135	9079.0	9079.0	8.36	8.92
Rgn 2	127.9	145.6	137.7	0.754	78326.0	78326.0	72.12	76.99
Rgn 3	144.8	154.5	147.1	0.943	14325.0	14325.0	13.19	14.08
3 Peaks					101730.0	101730.0	93.67	100.00

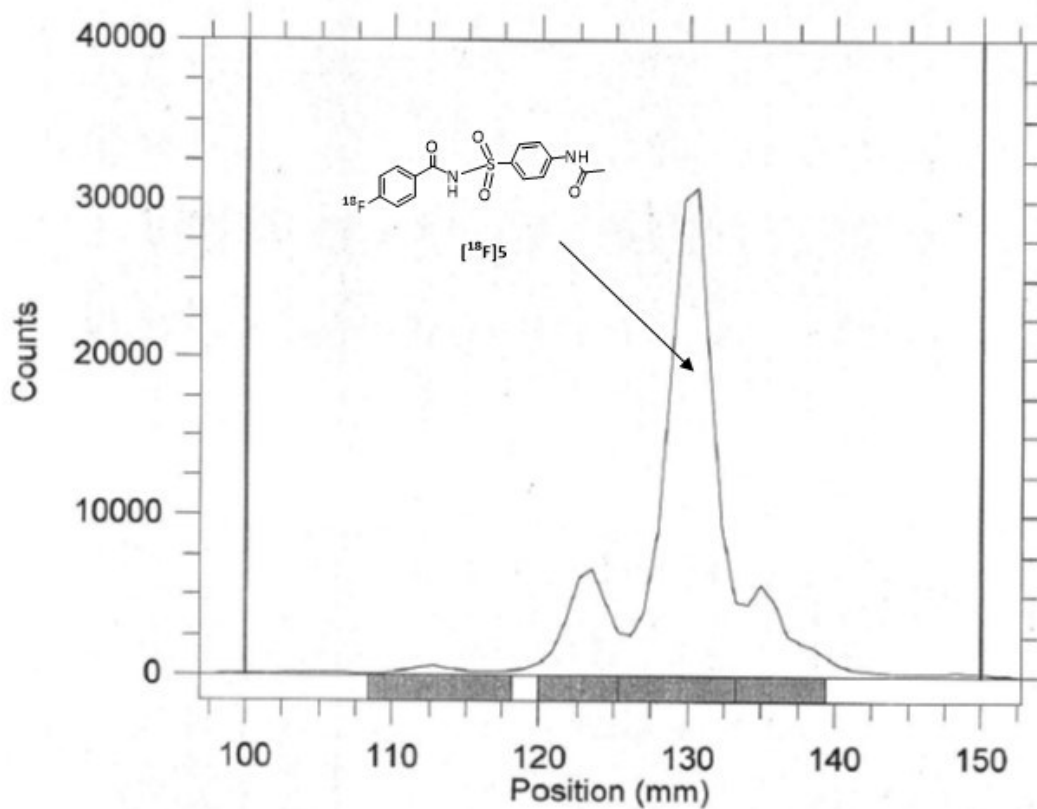


Reg	(mm) Start	(mm) Stop	(mm) Centroid	RF	Region Counts	Region CPM	% of Total	% of ROI
Rgn 1	96.0	105.7	100.8	0.016	687.0	687.0	3.25	3.51
Rgn 2	117.3	134.1	125.7	0.514	14844.0	14844.0	70.20	75.81
Rgn 3	133.2	138.5	135.6	0.712	2439.0	2439.0	11.53	12.46
Rgn 4	137.7	143.0	139.3	0.787	1610.0	1610.0	7.61	8.22
4 Peaks					19580.0	19580.0	92.59	100.00

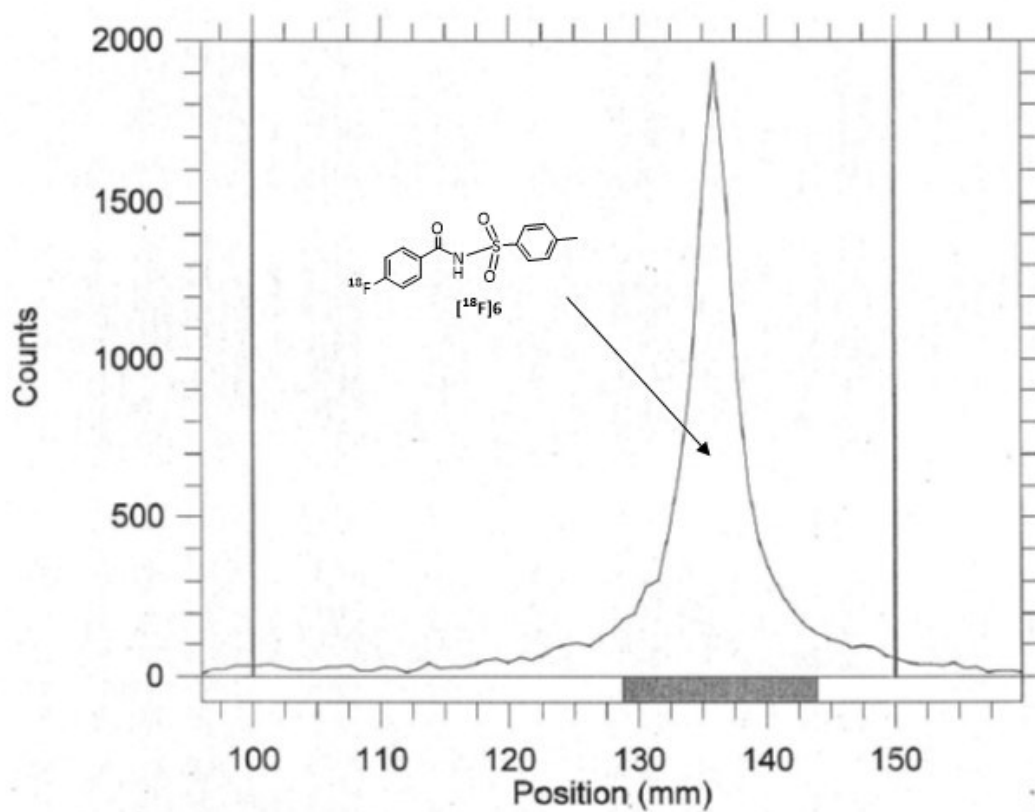




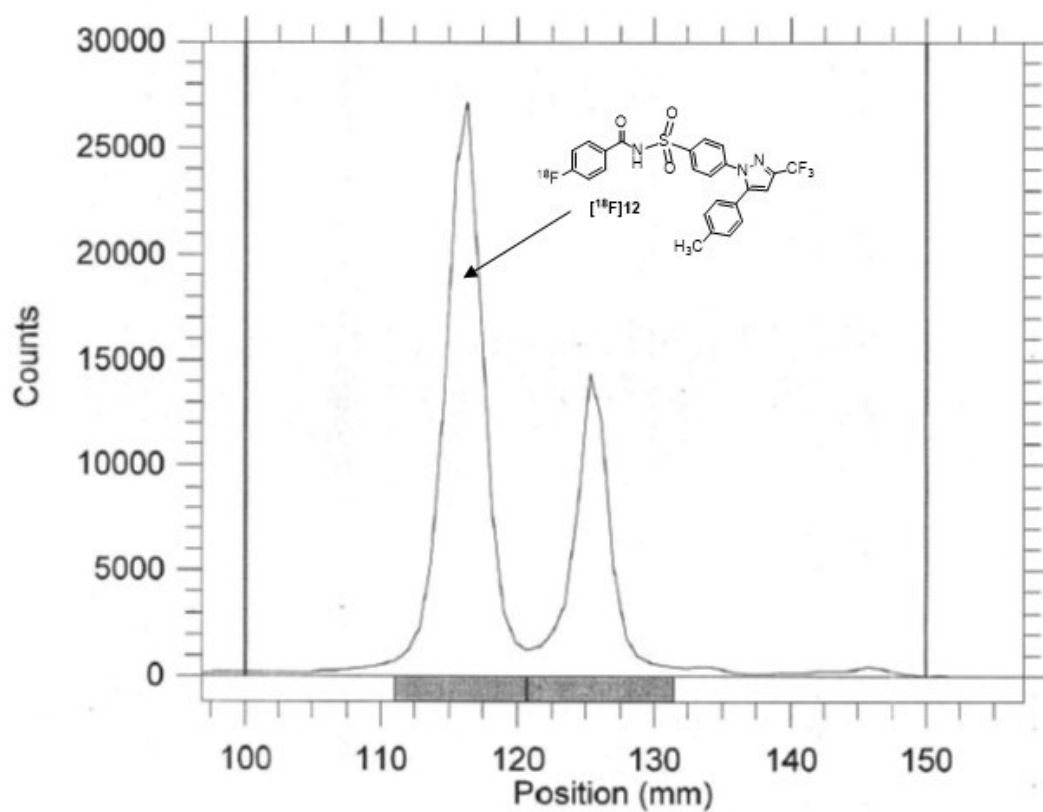
Reg	(mm) Start	(mm) Stop	(mm) Centroid	RF	Region Counts	Region CPM	% of Total	% of ROI
Rgn 1	108.4	118.1	113.2	0.263	3627.0	3627.0	1.94	1.94
Rgn 2	119.9	126.1	123.1	0.462	26199.0	26199.0	13.98	14.00
Rgn 3	125.2	134.1	130.0	0.599	131548.0	131548.0	70.19	70.28
Rgn 4	133.2	139.4	135.3	0.706	25799.0	25799.0	13.77	13.78
4 Peaks					187173.0	187173.0	99.88	100.00



Reg	(mm) Start	(mm) Stop	(mm) Centroid	RF	Region Counts	Region CPM	% of Total	% of ROI
Rgn 1	128.8	143.9	135.8	0.717	10933.0	10933.0	76.74	100.00
1 Peaks					10933.0	10933.0	76.74	100.00



Reg	(mm) Start	(mm) Stop	(mm) Centroid	RF	Region Counts	Region CPM	% of Total	% of ROI
Rgn 1	111.1	120.8	116.1	0.321	104523.0	104523.0	62.86	66.80
Rgn 2	120.8	131.5	125.3	0.507	51939.0	51939.0	31.24	33.20
2 Peaks					156462.0	156462.0	94.10	100.00



**General procedure for one pot/three-component procedure for radiosynthesis of compounds [<sup>18</sup>F]7-<sup>[18F]10</sup> and [<sup>18</sup>F]13.** In an eppendorf tube, 2 mg of thiolane-2,5-dione was dissolved in 250 μL of DMF and 30-40 MBq of 4-<sup>[18F]</sup>fluorobenzylamine in THF was added. The reaction was heated to 80 °C for 15 min. Then, 3-5 mg of sulfonyl dissolved in 200 μL of THF was added. The reaction mixture was stirred at 50 °C for 30 min. Product formation was monitored with radio-TLC analysis. Radiochemical conversions (RCCs) refer to the percentage of radiolabeled products [<sup>18</sup>F]7-<sup>[18F]10</sup> and [<sup>18</sup>F]13 present in the reaction mixture as determined by radio-TLC analysis (10% MeOH/CH<sub>2</sub>Cl<sub>2</sub>).

***N*<sup>1</sup>-(4-<sup>[18</sup>F]Fluorobenzyl)-*N*<sup>4</sup>-((2,4,6-triisopropylphenyl)sulfonyl)succinamide ([<sup>18</sup>F]7).** RCC: 38%. Radio-TLC (50% EtOAc/hexane): *R*<sub>f</sub> = 0.6.

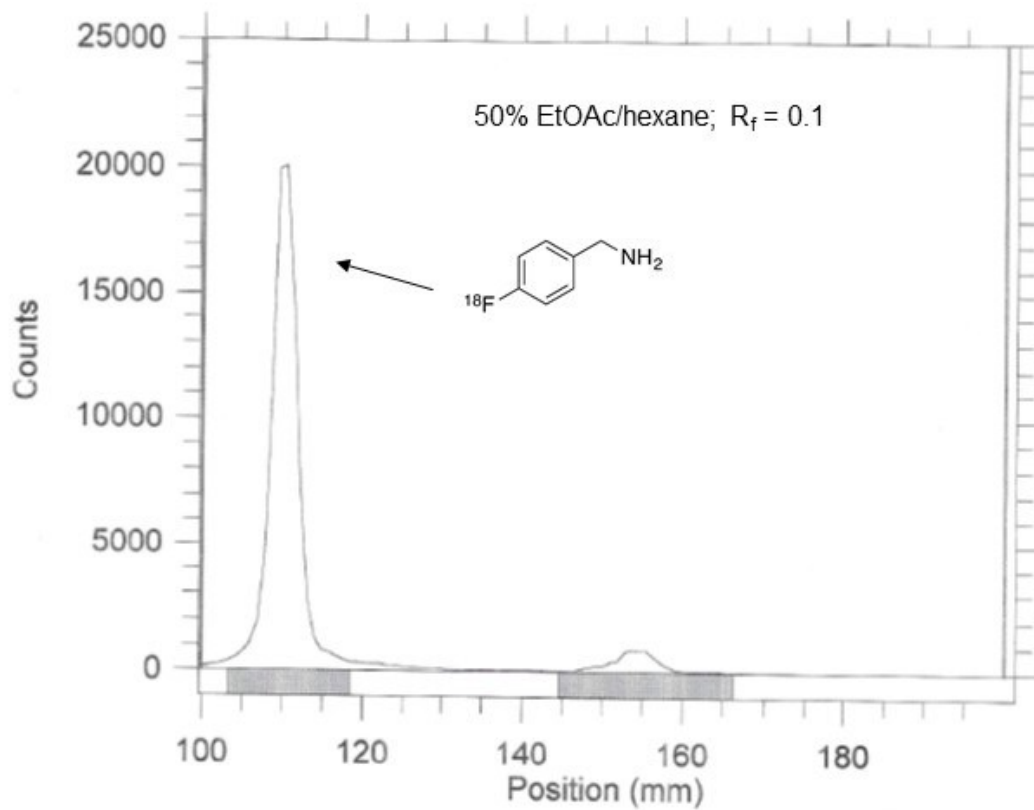
**4-(*N*-(4-((4-<sup>[18</sup>F]Fluorobenzyl)amino)-4-oxobutanoyl)sulfamoyl)benzoic acid ([<sup>18</sup>F]8).** RCC: 78%. Radio-TLC (50% EtOAc/hexane): *R*<sub>f</sub> = 0.16.

***N*<sup>1</sup>-((4-Acetamidophenyl)sulfonyl)-*N*<sup>4</sup>-(4-<sup>[18</sup>F]fluorobenzyl)succinamide ([<sup>18</sup>F]9).** RCC: 70%. Radio-TLC (50% EtOAc/hexane): *R*<sub>f</sub> = 0.54.

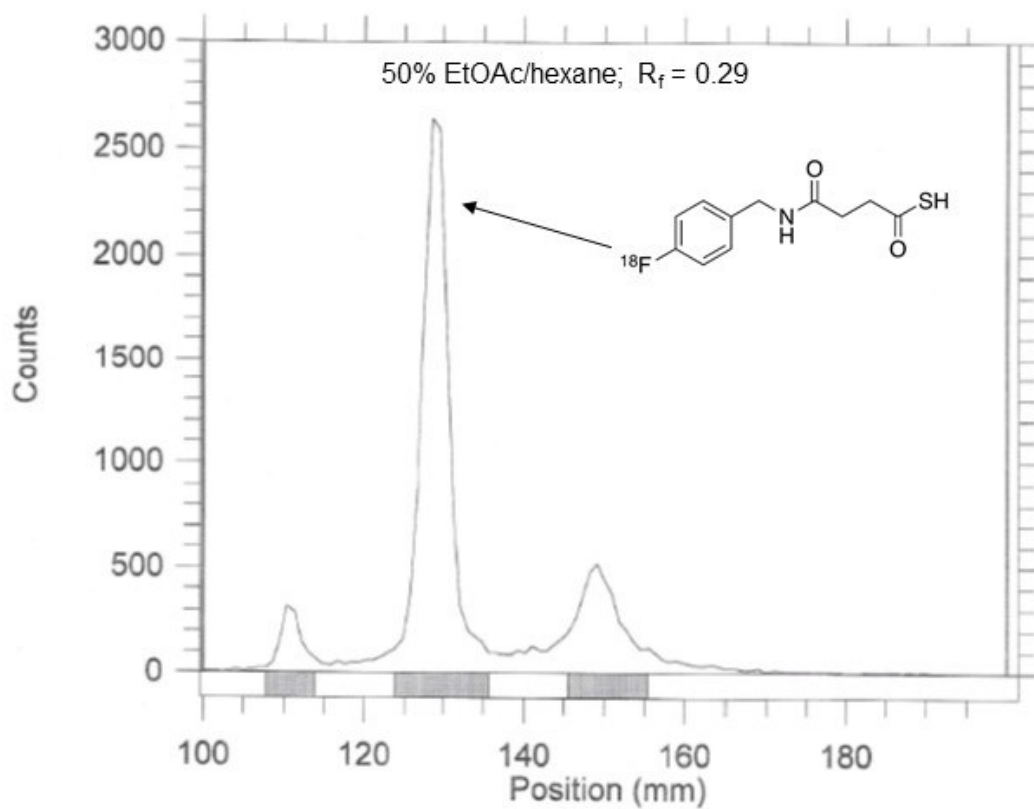
***N*<sup>1</sup>-(4-<sup>[18</sup>F]Fluorobenzyl)-*N*<sup>4</sup>-tosylsuccinamide ([<sup>18</sup>F]10).** RCC: 30%. Radio-TLC (50% EtOAc/hexane): *R*<sub>f</sub> = 0.46.

***N*<sup>1</sup>-(4-<sup>[18</sup>F]Fluorobenzyl)-*N*<sup>4</sup>-((4-(5-(*p*-tolyl)-3-(trifluoromethyl)-1H-pyrazol-1-yl)phenyl)sulfonyl)succinamide ([<sup>18</sup>F]13).** RCC: 55%. Radio-TLC (50% EtOAc/hexane): *R*<sub>f</sub> = 0.37.

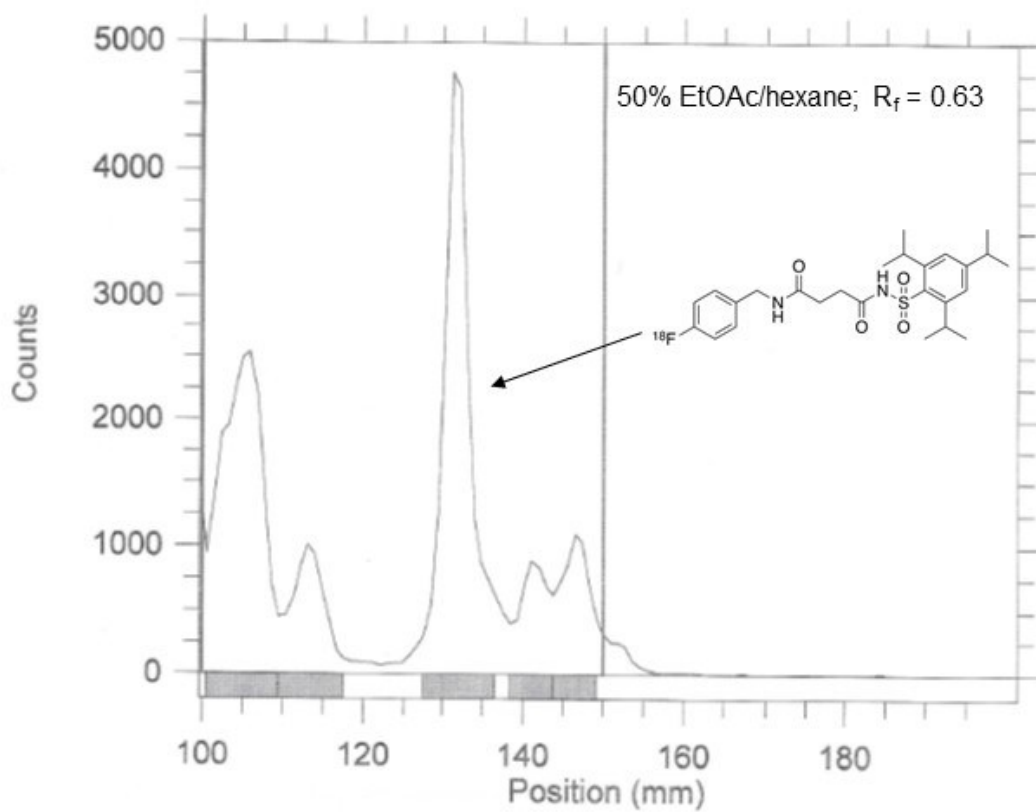
Reg	(mm) Start	(mm) Stop	(mm) Centroid	RF	Region Counts	Region CPM	% of Total	% of ROI
Rgn 1	103.2	118.5	110.2	0.102	87850.0	87850.0	88.71	92.97
Rgn 2	144.7	166.3	153.9	0.539	6646.0	6646.0	6.71	7.03
2 Peaks					94496.0	94496.0	95.42	100.00



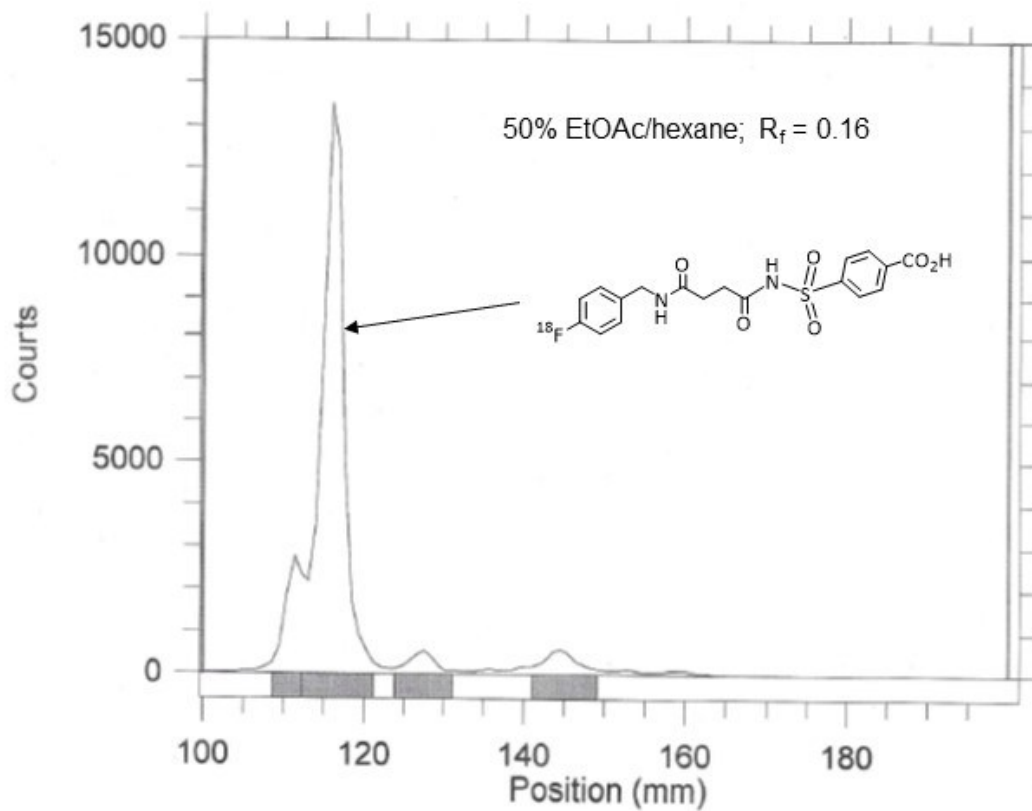
Reg	(mm) Start	(mm) Stop	(mm) Centroid	RF	Region Counts	Region CPM	% of Total	% of ROI
Rgn 1	107.7	114.0	110.9	0.109	1048.0	1048.0	5.54	6.46
Rgn 2	123.9	135.7	129.0	0.290	11814.0	11814.0	62.46	72.84
Rgn 3	145.6	155.5	149.5	0.495	3358.0	3358.0	17.75	20.70
3 Peaks					16220.0	16220.0	85.75	100.00



Reg	(mm) Start	(mm) Stop	(mm) Centroid	RF	Region Counts	Region CPM	% of Total	% of ROI
Rgn 1	100.5	109.5	104.6	0.093	17700.0	17700.0	30.64	34.04
Rgn 2	109.5	117.6	113.0	0.260	5605.0	5605.0	9.70	10.78
Rgn 3	127.5	136.6	131.7	0.634	19731.0	19731.0	34.15	37.94
Rgn 4	138.4	143.8	140.9	0.818	3953.0	3953.0	6.84	7.60
Rgn 5	143.8	149.2	146.1	0.923	5016.0	5016.0	8.68	9.65
5 Peaks					52005.0	52005.0	90.02	100.00

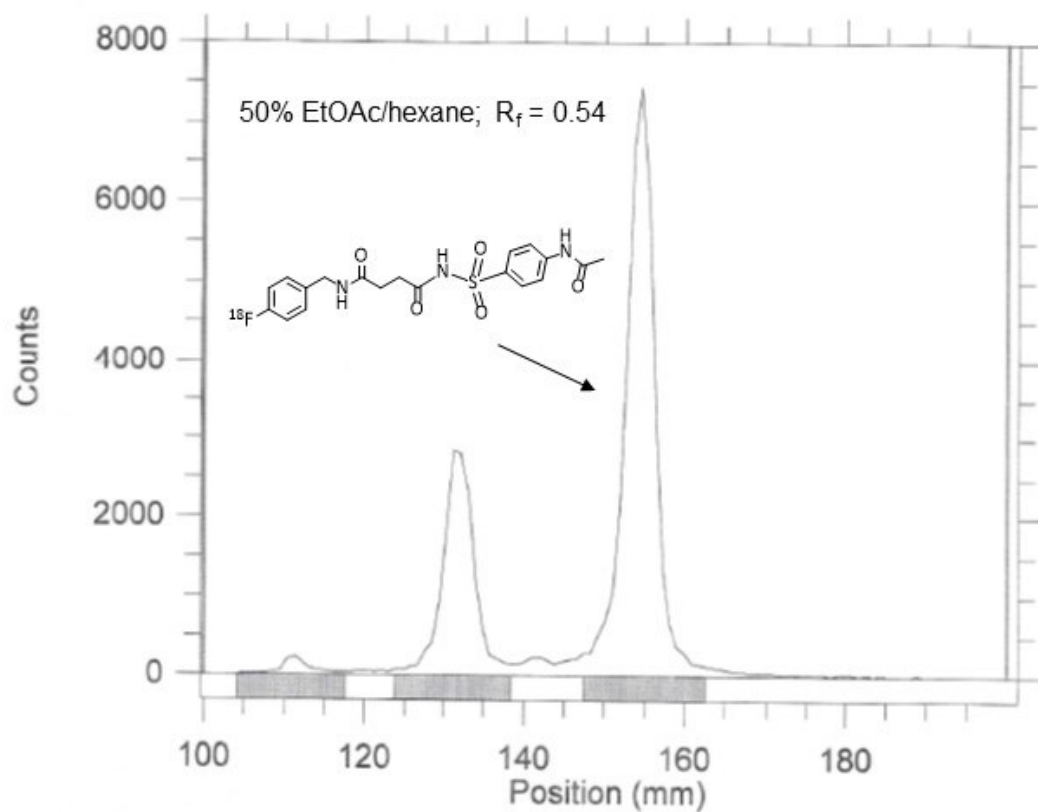


Reg	(mm) Start	(mm) Stop	(mm) Centroid	RF	Region Counts	Region CPM	% of Total	% of ROI
Rgn 1	108.6	113.1	111.2	0.112	7883.0	7883.0	12.21	12.48
Rgn 2	112.2	121.2	115.9	0.159	49655.0	49655.0	76.92	78.60
Rgn 3	123.9	131.2	127.1	0.271	2393.0	2393.0	3.71	3.79
Rgn 4	141.1	149.2	144.5	0.445	3240.0	3240.0	5.02	5.13
4 Peaks					63171.0	63171.0	97.86	100.00

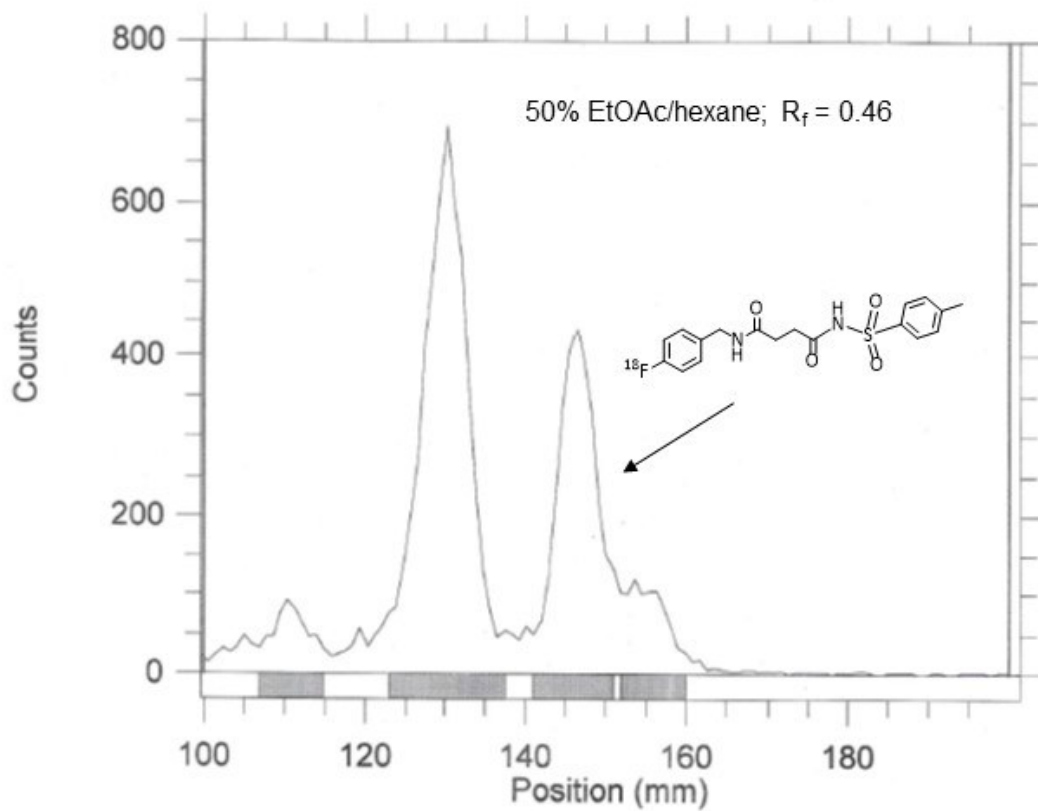




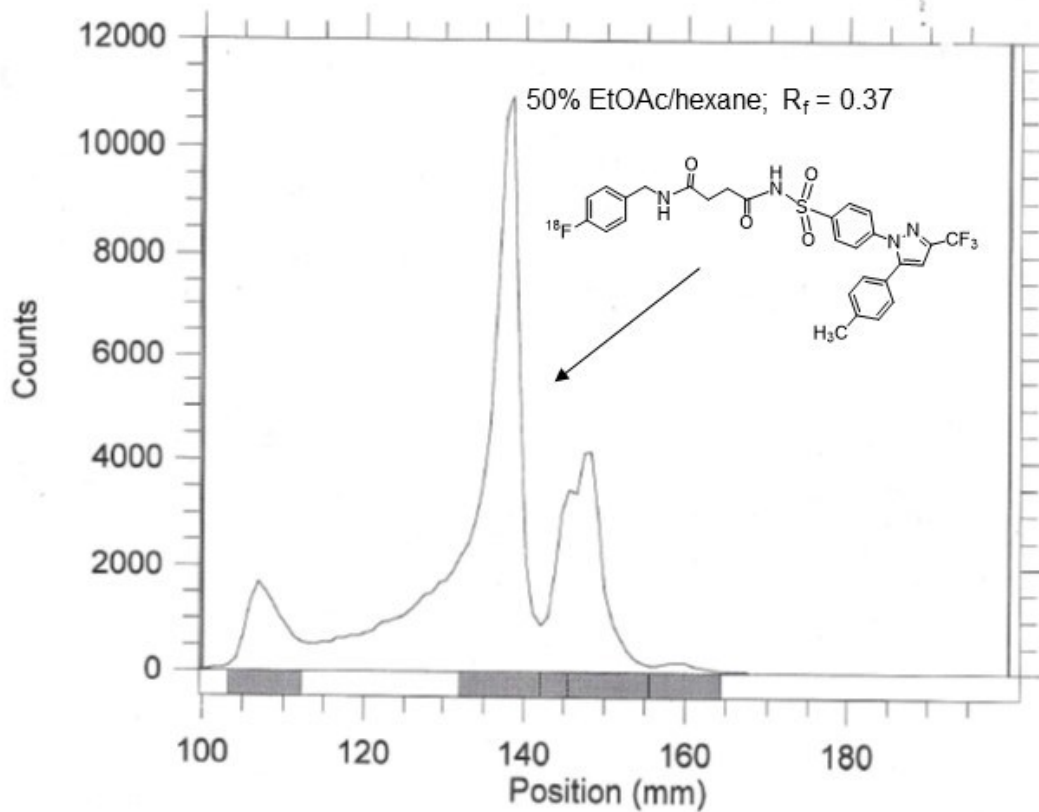
Reg	(mm) Start	(mm) Stop	(mm) Centroid	RF	Region Counts	Region CPM	% of Total	% of ROI
Rgn 1	104.1	117.6	111.7	0.117	1032.0	1032.0	1.92	2.03
Rgn 2	123.9	138.4	131.7	0.317	14256.0	14256.0	26.51	28.10
Rgn 3	147.4	162.7	154.3	0.543	35437.0	35437.0	65.90	69.86
3 Peaks					50725.0	50725.0	94.33	100.00



Reg	(mm) Start	(mm) Stop	(mm) Centroid	RF	Region Counts	Region CPM	% of Total	% of ROI
Rgn 1	106.8	114.9	110.6	0.106	523.0	523.0	5.17	5.76
Rgn 2	123.0	137.5	129.9	0.299	5007.0	5007.0	49.47	55.19
Rgn 3	141.1	151.0	146.3	0.463	2737.0	2737.0	27.04	30.17
Rgn 4	151.9	160.0	154.9	0.549	805.0	805.0	7.95	8.87
4 Peaks					9072.0	9072.0	89.63	100.00



Reg	(mm) Start	(mm) Stop	(mm) Centroid	RF	Region Counts	Region CPM	% of Total	% of ROI
Rgn 1	103.2	112.2	107.8	0.078	9393.0	9393.0	8.18	9.76
Rgn 2	132.1	142.0	136.9	0.369	52972.0	52972.0	46.15	55.04
Rgn 3	142.0	146.5	144.4	0.444	10338.0	10338.0	9.01	10.74
Rgn 4	145.6	155.5	148.1	0.481	22168.0	22168.0	19.31	23.03
Rgn 5	155.5	164.5	159.1	0.591	1370.0	1370.0	1.19	1.42
5 Peaks					96241.0	96241.0	83.84	100.00

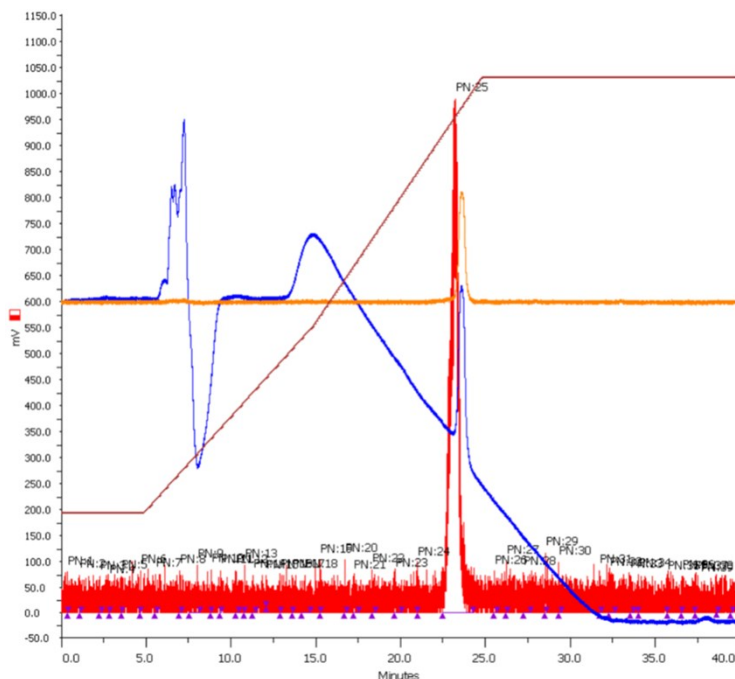


**Peptide synthesis and radiolabeling.** Sulfonyl azide-decorated tetrapeptide Gly-Leu-Ser-Phe **14** was prepared using a 4-benzyloxybenzyl alcohol (Wang) resin, preloaded with a C-terminal Fmoc-protected phenylalanine (loading 0.67 mmol/g), provided the solid support for the peptide synthesis. The peptide was synthesised by solid phase peptide synthesis (SPPS) using a fully automated peptide synthesizer (Syrol, MultisynTech/Biotage). The method used consisted of swelling the pre-loaded Wang resin in 2 mL of *N,N*-dimethylformamide (DMF) for 15 min. Fmoc-deprotection was achieved by treatment with 40% piperidine/DMF for 5 min and followed by a second incubation with 20% piperidine/DMF for 15 min. Fmoc-protected amino acids and the 4-(azidosulfonyl)benzoic acid (5 equiv.) were activated and coupled using 5 eq. of O-benzotriazole-*N,N,N',N'*-tetramethyl-uronium-hexafluoro-phosphate (HBTU), 5 equiv. of ethyl 2-cyano-2-(hydroxyimino)acetate (Oxyma), and 10 equiv. *N,N*-diisopropylethylamine (DIPEA) over a 60 min time period followed by wash steps with DMF. The cleavage of the protection groups from the peptide and the peptide from the resin was done in 95% trifluoroacetic acid (TFA) and 5% water for 2 hours at room temperature. Due to the sensitivity of the sulfonyl azide group to reduction, no scavengers were added. The peptide was separated from the resin through a syringe filter and precipitated by the addition of ice-cold diethyl ether. The crude peptides were obtained by removing the residual ether using a syringe filter and dried under vacuum. Purification by HPLC with semi-preparative HPLC at 2 mL/min using a gradient of water/0.2% TFA (A) and MeCN (B): 10% eluent B for 5 min, increased to 30% eluent B over an additional 5 min, then to 50% eluent B over 15 min, then 90% eluent B over 5 minutes, holding at that point for a final 10 min. Subsequent lyophilization gave sufficiently pure (>98% purity by HPLC) peptide as a white powder (10.9 mg, 34.4% yield). Analysis:  $t_R = 34.6$  min. MW  $C_{27}H_{33}N_7O_9S$  calculated 631.21 g/mol, found LC-MS (ESI, positive)  $m/z$  632.2  $[M + H]^+$ , HR-MS (ESI, negative)  $m/z$  630.2  $[M - H]^-$ .

**Synthesis of reference compound 15.** 2 mg (3.17  $\mu$ mol) of peptide **14** was mixed with 1 mg (6.4  $\mu$ mol) of 4-fluorobenzothioic *S*-acid **1** in the presence of 0.74  $\mu$ L (6.4  $\mu$ mol) of 2,6-lutidine in 1 mL of methanol. The resulting mixture was allowed to react for 4 hours at room temperature. Purification by HPLC with semi-preparative HPLC at 2 mL/min using a gradient of water/0.2% TFA (A) and MeCN (B): 10% eluent B for 5 min, increased to 30% eluent B over an additional 5 min, then to 50% eluent B over 15 min, then 90% eluent B over 5 min, holding at that point for a final 10 min. Subsequent lyophilization gave sufficiently pure (>98% purity by HPLC) peptide as a white powder (1.6 mg, 70.0% yield). Analysis:  $t_R = 35.0$  min; MW  $C_{34}H_{38}FN_5O_{10}S$  calculated 727.23 g/mol, found LC-MS (ESI, positive)  $m/z$  728.2  $[M + H]^+$ .

**Radioynthesis of [ $^{18}F$ ]15.** 100-150 MBq of [ $^{18}F$ ]SFB in 250  $\mu$ L  $CH_3CN$  was added to 1 mg (17.8  $\mu$ mol) of NaSH and left to react at 50  $^\circ C$  for 5 min. 1.1 mg (1.7  $\mu$ mol) of peptide **14** and 0.2  $\mu$ L (2.0  $\mu$ mol) of 2,6-lutidine was added and reacted at 50  $^\circ C$  for 30 min. Purification by HPLC with semi-preparative HPLC at 2 mL/min using a gradient of water/0.2% TFA (A) and MeCN (B): 20%

eluent B for 5 min, increased to 50% eluent B over an additional 10 min, then to 90% eluent B over 10 min, holding at that point for a final 10 min. The desired peak was isolated and had a decay-corrected radiochemical yield of 25% and a final radiochemical purity >99%. Analysis:  $t_R = 23.2$  min (radioactivity trace), 23.6 min (UV trace) (**Figure S1**).

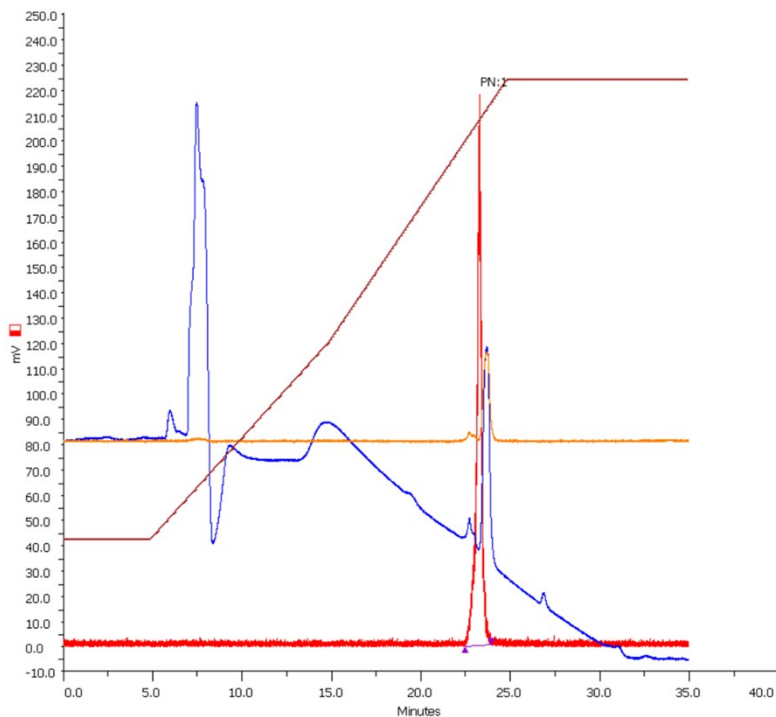


**Figure S1.** Radio-HPLC trace (red) of purified [ $^{18}\text{F}$ ]**15** co-injected with reference compound **15** (blue; 210 nm; orange 254 nm).

**Synthesis of reference compound 16.** 0.34  $\mu\text{L}$  (2.5  $\mu\text{mol}$ ) of 4-fluorobenzylamine was added to 0.28  $\mu\text{L}$  (1.6  $\mu\text{mol}$ ) of 2,6-lutidine and 0.34 mg (3.0  $\mu\text{mol}$ ) of thiolane-2,5-dione in 0.5 mL of DMF and reacted at room temperature for 30 min yielding a clear, lightly yellow liquid. 1.6 mg (2.5  $\mu\text{mol}$ ) of peptide **14** was added and incubated at 50  $^{\circ}\text{C}$  for 30 min. Purification by HPLC with semi-preparative HPLC at 2 mL/min using a gradient of water/0.2% TFA (A) and MeCN (B): 10% eluent B for 5 min, increased to 30% eluent B over an additional 5 min, then to 50% eluent B over 15 min, then 90% eluent B over 5 min, holding at that point for a final 10 min. Subsequent lyophilization gave sufficiently pure (>99% purity by HPLC) peptide as a white powder (1.6 mg, 74.5% yield). Analysis:  $t_R = 35.0$  min; MW  $\text{C}_{38}\text{H}_{45}\text{FN}_6\text{O}_{11}\text{S}$  calculated 812.29 g/mol, found LC-MS (ESI, positive)  $m/z$  813.2  $[\text{M} + \text{H}]^+$ .

**Radioynthesis of [ $^{18}\text{F}$ ]**16.** 100-150 MBq of 4- $^{18}\text{F}$ fluorobenzyl amine in 500  $\mu\text{L}$  THF was added to 0.2 mg (1.7  $\mu\text{mol}$ ) of thiolane-2,5-dione and allowed to react for 10 min at 50  $^{\circ}\text{C}$ . Then, 1.1 mg (1.7  $\mu\text{mol}$ ) of peptide **14** and 0.2  $\mu\text{L}$  (1.8  $\mu\text{mol}$ ) of 2,6-lutidine was added and allowed to**

react for 30 min at 50 °C. Purification by HPLC with semi-preparative HPLC at 2 mL/min using a gradient of water/0.2% TFA (A) and MeCN (B): 20% eluent B for 5 min, increased to 50% eluent B over an additional 10 min, then to 90% eluent B over 10 min, holding at that point for a final 10 min. The desired peak was isolated and had a decay-corrected radiochemical yield of 20% and a final radiochemical purity >99%. Analysis:  $t_R = 23.2$  min (radioactivity trace), 23.6 min (UV trace) (**Figure S2**).



**Figure S2.** Radio-HPLC trace (red) of purified [ $^{18}\text{F}$ ]**16** co-injected with reference compound **16** (blue; 210 nm; orange 254 nm).