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Supporting Information

Synthesis and Nitric Oxide Releasing Properties of Novel

Fluoro S-Nitrosothiols

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1. General information

All commercial organic solvents and chemicals were directly used in all synthetic reactions. SNAP was purchased from PharmaBlock Sciences Inc (Nanjing, China). Column chromatography was carried out using silica gel (60 Å, 40 - 63 µm) with ACS grade solvents of MeOH and CH₂Cl₂. All novel synthetic compounds were characterized by ¹H/¹³C/¹⁹F NMR spectroscopy and high-resolution mass spectrometry (HRMS) with an electrospray ionization source. ¹H/¹³C/¹⁹F NMR spectra were recorded using Varian 400/500 MHz spectrometers. CFCl₃ was used in CDCl₃ as an internal reference in ¹⁹F NMR spectroscopy. The ¹⁹F NMR spectra were obtained without proton and carbon decoupled mode. HRMS data was collected using an Agilent 6520 Accurate-Mass Quadrupole Time-of-Flight (Q-TOF) LC/MS in the electrospray positive-ion mode with a ZORBAX RRHD Eclipse Plus C18 reversed-phase column (2.1 x 50 mm, 1.8 Mm). Phosphate-buffered saline (PBS, 10.0 mM, pH 7.40, with 100 MM EDTA) was prepared using Na₂HPO₄, KH₂PO₄, NaCl, KCl and EDTA. The pH of PBS was adjusted by a diluted HCl solution.

2. Syntheses

2.1 Synthesis of 3(D)-acetamido-4,4-dimethylthietan-2-one



To a stirred solution of *D*-penicillamine (2.001 g, 13.41 mmol) in pyridine (25 mL) at room temperature was added acetic anhydride (3.55 mL, 37.52 mmol) in one portion. After stirring for 20 h at room temperature, the reaction solution was washed by HCl solution (1.0 M, 80 mL), and extracted by chloroform (3 x 50 mL). All the organic solution was dried by NaSO₄, and taken to dryness to obtain a crude yellow solid. The resulting yellow solid was triturated by petroleum ether to give a pale yellow solid, which was further dried under vacuum overnight to give the desired product (782.8 mg, 34%). ¹H NMR (400 MHz, CDCl₃): δ 6.12 (s, 1 H), 5.66 (d, *J* = 7.6 Hz, 1 H), 2.05 (s, 3 H), 1.86 (s, 3 H), 1.62 (s, 3 H).

2.2 Synthesis and characterization of free thiols 2a-2n



All **2a-2n** thiol intermediates were prepared using the same procedures as described as follows. To a solution of lactone (600.0 mg, 3.464 mmol) in CHCl₃ (25 mL) was added various commercially available amines (4.157 mmol, 1.2 eq. of lactone) in one portion. After stirring at room temperature (**2a-2i**, **2m**, **2n** for 24 h; **2j-2l** for 48 h), the reaction solution was then taken to dryness. The crude products were purified by column chromatography (silica gel/3:97 MeOH-CH₂Cl₂) to yield the desired products **2a-2n** as pale white powers. The corresponding yields for **2a-2n** were listed in Table S1. All desired products were characterized by ¹H/¹³C/¹⁹F NMR and HRMS.

Compound #	Yields (%)
2a	92
2b	93
2c	71
2d	83
2e	90
2f	92
2g	82
2h	94
2i	91
2j	89
2k	93
21	85
2m	74
<u>2n</u>	82

Table S1. Yields for the synthesis of free thiols 2a-2n.

Characterization of thiol **2a**: ¹H NMR (500 MHz, CDCl₃): δ 7.52 (t, J = 5.5 Hz, 1 H), 7.30 (t, J = 7.5 Hz, 1 H), 7.25 (q, J = 7.5 Hz, 1 H), 7.07 (t, J = 7.5 Hz, 1 H), 7.02 (t, J = 8.5 Hz, 1 H), 6.72 (d, J = 9.5 Hz, 1 H), 4.64 (d, J = 9.0 Hz, 1 H), 4.52 (dd, J = 14.75, 6.5 Hz, 1 H), 4.35 (dd, J = 15.0, 6.5 Hz, 1 H), 2.58 (s, 1 H), 1.94 (s, 3 H), 1.46 (s, 3 H), 1.28 (s, 3 H). ¹³C NMR (126 MHz, CDCl₃): δ 170.31, 169.77, 160.91 (d, J = 247.09 Hz), 130.23 (d, J = 4.16 Hz), 129.36 (d, J = 8.19 Hz), 124.71 (d, J = 14.89 Hz), 124.22 (d, J = 3.53 Hz), 115.39 (d, J = 21.29 Hz), 60.09,

46.20, 37.40 (d, J = 4.03 Hz), 30.95, 28.58, 23.10. ¹⁹F NMR (377 MHz, CDCl₃): - 119.01 (m). HRMS m/z (ESI): calculated for MH⁺ 299.1224, found 299.1228.

Characterization of thiol **2b**: ¹H NMR (500 MHz, CDCl₃): δ 7.30-7.28 (m, 1 H), 7.17 (t, *J* = 5.5 Hz, 1 H), 7.03 (d, *J* = 7.5 Hz, 1 H), 6.97-6.94 (m, 2 H), 6.66 (d, *J* = 9.0 Hz 1 H), 4.51 (d, *J* = 9.0 Hz, 1 H), 4.46 (dd, *J* = 15.0, 6.0 Hz, 1 H), 4.35 (dd, *J* = 15.0, 6.0 Hz, 1 H), 2.73 (s, 1 H), 1.99 (s, 3 H), 1.51 (s, 3 H), 1.30 (s, 3 H). ¹³C NMR (126 MHz, CDCl₃): δ 170.42, 169.84, 162.95 (d, *J* = 246.96 Hz), 140.28 (d, *J* = 6.93 Hz), 130.22 (d, *J* = 8.19 Hz), 123.17 (d, *J* = 3.02 Hz), 114.58 (d, *J* = 21.80 Hz), 114.41 (d, *J* = 21.17 Hz), 60.18, 45.76, 42.96 (d, *J* = 6.5 Hz), 31.13, 28.53, 23.17. ¹⁹F NMR (377 MHz, CDCl₃): -113.90 (m). HRMS m/z (ESI): calculated for MH⁺ 299.1224, found 299.1285.

Characterization of thiol **2c**: ¹H NMR (500 MHz, CDCl₃): δ 7.24 (d, *J* = 5.5 Hz, 1 H), 7.22 (d, *J* = 5.5 Hz, 1 H), 7.02 (d, *J* = 8.5 Hz, 1 H), 7.00 (d, *J* = 8.5 Hz, 1 H), 6.79 (s, 1 H), 6.61 (d, *J* = 9.0 Hz, 1H), 4.44 (dd, *J* = 15.0, 6.0 Hz, 1 H), 4.41 (d, *J* = 8.5 Hz, 1 H), 4.33 (dd, *J* = 15.0, 6.0 Hz, 1 H), 2.73 (s, 1 H), 2.02 (s, 3 H), 1.50 (s, 3 H), 1.28 (s, 3 H). ¹³C NMR (126 MHz, CDCl₃): 170.35, 169.67, 162.21 (d, *J* = 246.58 Hz), 133.42 (d, *J* = 3.02 Hz), 129.44 (d, *J* = 8.19 Hz), 115.61 (d, *J* = 21.67 Hz), 60.25, 45.66, 42.86, 31.13, 28.45, 23.22. ¹⁹F NMR (377 MHz, CDCl₃): -115.28 (m). HRMS m/z (ESI): calculated for MH⁺ 299.1224, found 299.1265.

Characterization of thiol **2d**: ¹H NMR (500 MHz, CDCl₃): δ 7.15 (d, *J* = 5.5 Hz, 1 H), 7.14 (d, *J* = 5.5 Hz, 1 H), 6.99 (t, *J* = 8.5 Hz, 2 H), 6.53 (d, *J* = 7.5 Hz, 1 H), 6.31 (s, 1 H), 4.27 (d, *J* = 9.0 Hz, 1 H), 3.55 (app. sextet, *J* = 6.5 Hz, 1 H), 3.46 (app. sextet, *J* = 7.0 Hz, 1 H), 2.79 (t, *J* = 7.0 Hz, 2 H), 2.68 (s, 1 H), 2.03 (s, 3 H), 1.44 (s, 3 H), 1.25 (s, 3 H). ¹³C NMR (126 MHz, CDCl₃): δ 170.21, 169.70, 161.71 (d, *J* = 245.32 Hz), 134.08 (d, *J* = 3.02 Hz), 130.15 (d, *J* = 7.94 Hz), 115.46 (d, *J* = 21.29 Hz), 60.27, 45.50, 40.57, 34.71, 31.02, 28.40, 23.25. ¹⁹F NMR (377 MHz, CDCl₃): -116.91 (m). HRMS m/z (ESI): calculated for MH⁺ 313.1381, found 313.1476.

Characterization of thiol **2e**: ¹H NMR (500 MHz, CDCl₃): δ 7.59 (s, 1 H), 7.24 (app. pentet, *J* = 7.5 Hz, 1 H), 6.87 (t, *J* = 8.0 Hz, 2 H), 6.74 (d, *J* = 9.0 Hz, 1 H), 4.65 (d, *J* = 9.5 Hz, 1 H), 4.59, (dd, *J* = 14.0, 6.0 Hz, 1 H), 4.37 (dd, *J* = 14.5, 5.0 Hz, 1 H), 2.54 (s, 1 H), 1.92 (s, 3 H), 1.44 (s, 3 H), 1.26 (s, 3 H). ¹³C NMR (126 MHz, CDCl₃): δ 170.19, 169.50, 161.54 (dd, *J* = 250.11, 7.94 Hz), 129.60 (t, *J* = 10.21 Hz), 113.43 (t, *J* = 19.40 Hz), 111.42 (d, *J* = 5.80 Hz), 111.26 (d, *J* = 5.54 Hz), 59.94, 46.38, 31.11 (t, *J* = 4.03 Hz), 30.77, 28.52, 23.03. ¹⁹F NMR (377 MHz, CDCl₃): -115.07 (q, *J* = 5.28 Hz). HRMS m/z (ESI): calculated for MH⁺ 317.1130, found 317.1134.

Characterization of thiol **2f**: ¹H NMR (500 MHz, CDCl₃): δ 7.52 (s, 1 H), 6.78 (d, *J* = 6.5 Hz, 2 H), 6.73-6.67 (m, 2 H), 4.56 (d, *J* = 9.5 Hz, 1 H), 4.42 (dd, *J* = 15.50, 6.0 Hz, 1 H), 4.33 (dd, *J* = 15.50, 6.0 Hz, 1 H), 2.72 (s, 1 H), 2.01 (s, 3 H), 1.51 (s, 3 H), 1.31 (s, 3 H). ¹³C NMR (126 MHz, CDCl₃): δ 170.55, 170.04, 163.16 (dd, *J* = 249.61, 12.73 Hz), 141.87 (t, *J* = 8.82 Hz), 110.31 (d, *J* = 6.05 Hz), 110.15 (d, *J* = 5.92 Hz), 102.84 (t, *J* = 25.33 Hz), 60.12, 45.71, 42.60, 31.12, 28.56, 23.16. ¹⁹F NMR (377 MHz, CDCl₃): -109.84 (t, *J* = 7.92 Hz). HRMS m/z (ESI): calculated for MH⁺ 317.1130, found 317.1205.

Characterization of thiol **2g**: ¹H NMR (500 MHz, CDCl₃): δ 7.54-7.50 (m, 3 H), 7.45-7.41 (m, 2 H), 6.70 (d, *J* = 9.0 Hz, 1 H), 4.57 (d, *J* = 9.0 Hz, 1 H), 4.52 (dd, *J* = 15.0, 6.0 Hz, 1 H), 4.38 (dd, *J* = 15.0, 6.0 Hz, 1 H), 2.71 (s, 1 H), 1.96 (s, 3 H), 1.50 (s, 3 H), 1.30 (s, 3 H). ¹³C NMR (126 MHz, CDCl₃): 170.47, 169.97, 138.90, 130.97 (q, *J* = 32.26 Hz), 130.93, 129.12, 124.26, 123.97(q, *J* = 272.92 Hz), 60.11, 45.80, 42.87, 31.08, 28.52, 23.08. ¹⁹F NMR (377 MHz, CDCl₃): -63.22 (s). HRMS m/z (ESI): calculated for MH⁺ 349.1192, found 349.1267.

Characterization of thiol **2h**: ¹H NMR (400 MHz, CDCl₃): δ 7.50-7.38 (m, 4 H), 6.57-6.52 (m, 2 H), 4.31 (d, *J* = 8.8 Hz, 1 H), 3.62 (app. sextet, *J* = 6.8 Hz, 1 H), 3.46 (app. sextet, *J* = 6.8 Hz, 1 H), 2.90 (t, *J* = 6.8 Hz, 2 H), 2.67 (s, 1 H), 2.02 (s, 3 H), 1.43 (s, 3 H), 1.24 (s, 3 H). ¹³C NMR (126 MHz, CDCl₃): δ 170.25, 169.83, 139.49, 132.14, 131.47 (q, *J* = 32.36 Hz), 129.08, 125.48 (q, *J* = 3.78 Hz), 123.53 (q, *J* = 3.78 Hz), 124.06 (q, *J* = 272.79 Hz), 60.21, 45.52, 40.32, 35.31, 31.00, 28.41, 23.22. ¹⁹F NMR (377 MHz, CDCl₃): -63.13 (s). HRMS m/z (ESI): calculated for MH⁺ 363.1349, found 363.1442.

Characterization of thiol **2i**: ¹H NMR (500 MHz, CDCl₃): δ 7.47 (s, 1 H), 6.89 (t, *J* = 7.0 Hz, 2 H), 6.89 (d, *J* = 9.0 Hz, 1 H), 4.52 (d, *J* = 9.0 Hz, 1 H), 4.37 (dd, *J* = 15.5, 6.0 Hz, 1 H), 4.30 (dd, *J* = 15.5, 6.0 Hz, 1 H), 2.72 (s, 1 H), 2.03 (s, 3 H), 1.51 (s, 3 H), 1.31 (s, 3 H). ¹³C NMR (126 MHz, CDCl₃): δ 170.59, 170.08, 151.28 (ddd, *J* = 251.12, 10.08, 3.91 Hz), 139.96 (t, *J* = 15.37 Hz), 137.96 (t, *J* = 15.75 Hz), 134. 34 (m), 111.52 (d, *J* = 5.04 Hz), 111.38 (d, *J* = 5.04 Hz), 60.17, 45.61, 42.27, 31.13, 28.57, 23.20. ¹⁹F NMR (377 MHz, CDCl₃): -134.07 (m), -162.26 (m). HRMS m/z (ESI): calculated for MH⁺ 335.1036, found 335.1081.

Characterization of thiol **2j**: ¹H NMR (500 MHz, CDCl₃): δ 7.73 (s, 1H), 7.53 (t, *J* = 7.5 Hz, 1 H), 7.11 (t, *J* = 7.5 Hz, 2 H), 6.75 (d, *J* = 9.0 Hz, 1 H), 4.60 (d, *J* = 9.5 Hz, 1 H), 4.48 (dd, *J* = 15.5, 6.0 Hz, 1 H), 4.38 (dd, *J* = 15.5, 6.0 Hz, 1 H), 2.68 (s, 1), 1.99 (s, 3 H), 1.51 (s, 3 H), 1.32 (s, 3 H). ¹³C NMR (126 MHz, CDCl₃): δ 170.60, 170.17, 159.86 (dd, *J* = 257.29, 2.27 Hz), 145.13 (d, *J* = 7.43 Hz), 127.40 (qd, *J* = 4.54, 1.64 Hz), 122.85 (d, *J* = 3.40 Hz), 122.48 (q, *J* = 272.54 Hz), 117.29 (qd, *J* = 33.14, 12.47 Hz), 115.67 (d, *J* = 21.29 Hz), 60.22, 45.73, 42.51,

31.13, 28.61, 23.16. ¹⁹F NMR (377 MHz, CDCl₃): -61.82 (d, J = 11.31 Hz), -114.42 (m). HRMS m/z (ESI): calculated for MH⁺ 367.1098, found 367.1102.

Characterization of thiol **2k**: ¹H NMR (500 MHz, CDCl₃): δ 7.50 (s, 1 H), 7.31 (s, 1 H), 7.21 (d, *J* = 8.5 Hz, 1 H), 7.17 (d, *J* = 8.5 Hz, 1 H), 6.68 (d, *J* = 8.0 Hz, 1 H), 4.53 (dd, *J* = 15.5, 6.0 Hz, 1 H), 4.48 (d, *J* = 6.0 Hz, 1 H), 4.42 (dd, *J* = 15.5, 6.0 Hz, 1 H), 2.75 (s, 1 H), 2.01 (s, 3 H), 1.52 (s, 3 H), 1.31 (s, 3 H). ¹³C NMR (126 MHz, CDCl₃): δ 170.61, 170.16, 162.58 (d, *J* = 249.98 Hz), 141.98 (d, *J* = 7.18 Hz), 132.77 (qd, *J* = 33.01, 8.06 Hz), 123.15 (qd, *J* = 273.17, 3.02 Hz), 119.81 (m), 117.85 (d, *J* = 22.05 Hz), 111.83 (dq, *J* = 24.57, 3.78 Hz), 60.13, 45.55, 42.46, 31.10, 28.54, 23.11. ¹⁹F NMR (377 MHz, CDCl₃): -61.39 (d, *J* = 2.52 Hz), -110.78 (t, *J* = 2.52 Hz). HRMS m/z (ESI): calculated for MH⁺ 367.1098, found 367.1101.

Characterization of thiol **2I**: ¹H NMR (500 MHz, CDCl₃): δ 7.78 (m, 2 H), 7.72 (s, 1 H), 6.71 (d, *J* = 9.0 Hz, 1 H), 4.58 (d, *J* = 9.5 Hz, 1 H), 4.57 (dd, *J* = 16.0, 6.0 Hz, 1 H), 2.73 (s, 1 H), 2.00 (s, 3 H), 1.51 (s, 3 H), 1.31 (s, 3 H). ¹³C NMR (126 MHz, CDCl₃): δ 170.66, 170.28, 140.78, 131.89 (q, *J* = 33.39 Hz), 127.45 (d, *J* = 2.77 Hz), 123.19 (q, *J* = 273.29 Hz), 121.31 (pentet, *J* = 3.78 Hz), 60.12, 45.56, 42.38, 31.05, 28.52, 23.05. ¹⁹F NMR (377 MHz, CDCl₃): -63.44 (s). HRMS m/z (ESI): calculated for MH⁺ 417.1066, found 417.1099.

Characterization of thiol **2m**: ¹H NMR (500 MHz, CDCl₃): δ 7.34-7.31 (m, 2 H), 7.29-7.25 (m, 3 H), 6.82 (s, 1 H), 6.61 (d, *J* = 9.0 Hz, 1 H), 4.48 (dd, *J* = 15.0, 6.0 Hz, 1 H), 4.44 (d, *J* = 9.5 Hz, 1 H), 4.36 (dd, *J* = 15.0, 6.0 Hz, 1 H), 2.73 (s, 1 H), 2.00 (s, 3 H), 1.51 (s, 3 H), 1.29 (s, 3 H). ¹³C NMR (126 MHz, CDCl₃): δ 170.31, 169.63, 137.54, 128.75, 127.75, 127.62, 60.27, 45.79, 43.61, 31.13, 28.47, 23.20. HRMS m/z (ESI): calculated for MH⁺ 281.1318, found 281.1357.

Characterization of thiol **2n**: ¹H NMR (500 MHz, CDCl₃): δ 6.93 (s, 1 H), 6.65 (d, J = 9.0 Hz, 1 H), 6.49 (s, 2 H), 4.47 (d, J = 9.0 Hz, 1 H), 4.46 (dd, J = 14.5, 5.0 Hz, 1 H), 4.23 (dd, J = 14.5, 5.0 Hz, 1 H), 3.83 (s, 6 H), 3.82 (s, 3 H), 2.72 (s, 1 H), 1.52 (s, 3 H), 1.31 (s, 3 H). ¹³C NMR (126 MHz, CDCl₃): δ 170.30, 169.68, 153.42, 137.30, 133.34, 104.59, 60.85, 60.39, 56.11, 45.81, 43.73, 31.20, 28.46, 23.21. HRMS m/z (ESI): calculated for MH⁺ 371.1635, found 371.1664.

2.3 Synthesis and characterization of NO donors 3a-3n



R = 2-FPh (**a**), 3-FPh (**b**), 4-FPh (**c**), 4-FPhCH₂ (**d**), 2,6-diFPh (**e**), 3,5-diFPh (**f**),3-CF₃Ph (**g**), 3-CF₃PhCH₂ (**h**), 3,4,5-triFPh (**i**), 3-F,4-CF₃Ph (**j**), 3-F,5-CF₃Ph (**k**), 3,5-diCF₃Ph (**I**), Ph (**m**), and 3,4,5-triMeOPh (**n**)

Due to RSNOs are light sensitive, the synthetic procedure and the workup were performed under dark conditions. To a solution of free thiols **2a-2n** (200 mg) in MeOH (10 mL) was added HCI (10 mL, 1.0 M). Then the mixture solution was cooled in ice-water bath. Concentrated H_2SO_4 (2 mL) was added into reaction mixture. When the temperature of reaction solution was below 5 °C, NaNO₂ (3.0 eq.) dissolved in water (2 mL) was added into the reaction in one portion, and then ice-water bath was removed. After string for 1 h, the reaction mixture was extracted by CH_2CI_2 (5 mL × 3). All the organic layer was dried by $NaSO_4$, and taken to dryness. The product was dried under vacuum overnight under dark conditions to obtain yields (see Table S2). All desired products were characterized by ${}^{1}H/{}^{13}C/{}^{19}F$ NMR and HRMS.

Compound #	Yields (%)
3a	90
3b	89
3c	85
3d	93
3e	93
3f	90
3g	91
3h	91
3i	92
Зј	90
3k	95
31	99
3m	92
<u>3n</u>	94

Table S2. Yields for the synthesis of NO donors 3a-3n.

All the donors **3a-3n** were re-characterized by ¹H NMR spectroscopy after 7 days stored at -20 °C under dark conditions in the refrigerator, and all of them were stable without any decomposition based on ¹H NMR results (data not showed). The stability studies of NO donor **3I** during -20 °C storage are summarized in Figure S1. During the storage at -20 °C under dark conditions, no disulfide of **3I**

was observed after being stored for 1, 7 and 40 days based on the ¹H NMR results. Clean ¹H NMR spectra for **3I** in DMSO- d_6 were exactly the same after the storage for 1, 7 and 40 days. Therefore, no decomposition of NO donor **3I** was observed after being stored at -20 °C under dark conditions for more than one month.



Figure S1. ¹H NMR spectra for NO donor **3I** stored at -20 °C under dark conditions after (1) day 1, (2) day 7, (3) day 40 (spectrum 1: DMSO- d_6 (2.48 ppm) and H₂O (3.30 ppm); spectrum 2: DMSO- d_6 (2.48 ppm) and H₂O (3.49 ppm); spectrum 3: DMSO- d_6 (2.48 ppm) and H₂O (3.65 ppm)).

Characterization of target **3a**: ¹H NMR (500 MHz, DMSO-*d*₆): δ 8.96 (t, *J* = 5.5 Hz, 1 H), 8.43 (d, *J* = 10.0 Hz, 1 H), 7.34-7.30 (m, 2 H), 7.19-7.15 (m, 2 H), 5.27 (d, *J* = 9.5 Hz, 1 H), 4.37 (dd, *J* = 15.0, 6.0 Hz, 1 H), 4.30 (dd, *J* = 15.0, 6.0 Hz, 1 H), 1.96 (s, 3 H), 1.92 (s, 3 H), 1.85 (s, 3 H). ¹³C NMR (126 MHz, DMSO-*d*₆): δ 169.80, 168.78, 160.53 (d, *J* = 245.32 Hz), 130.37 (d, *J* = 4.41 Hz), 129.58 (d, *J* = 8.06 Hz), 125.80 (d, *J* = 14.99 Hz), 124.70 (d, *J* = 3.40 Hz), 115.57 (d, *J* = 21.80 Hz), 60.06, 59.35, 36.56 (d, *J* = 4.41 Hz), 27.08, 25.30, 22.75. ¹⁹F NMR (377 MHz, DMSO-*d*₆): -118.66 (m). HRMS m/z (ESI): calculated for MH⁺ 328.1126, found 328.1171.

Characterization of target **3b**: ¹H NMR (500 MHz, DMSO-*d*₆): δ 9.00 (t, *J* = 5.5 Hz, 1 H), 8.45 (d, *J* = 9.5 Hz, 1 H), 7.36 (q, *J* = 7.0 Hz, 1 H), 7.10-7.05 (m, 3 H), 5.26 (d, *J* = 9.5 Hz, 1 H), 4.32 (dd, *J* = 15.0, 6.0 Hz, 1 H), 4.28 (dd, *J* = 15.0, 6.0 Hz, 1 H), 1.96 (s, 3 H), 1.94 (s, 3 H), 1.86 (s, 3 H). ¹³C NMR (126 MHz, DMSO-*d*₆): δ 169.87, 168.92, 162.59 (d, *J* = 243.18 Hz), 142.35 (d, *J* = 6.3 Hz), 130.67 (d, *J* = 7.56 Hz), 123.79 (d, *J* = 2.52 Hz), 114.45 (d, *J* = 21.42 Hz), 114.07 (d, *J* = 20.16

Hz), 59.92, 59.48, 42.17, 27.00, 25.45, 22.75. ¹⁹F NMR (377 MHz, DMSO- d_6): - 113.52 (m). HRMS m/z (ESI): calculated for MH⁺ 328.1126, found 328.1144.

Characterization of target **3c**: ¹H NMR (500 MHz, DMSO-*d*₆): δ 8.98 (t, *J* = 5.5 Hz, 1 H), 8.44 (d, *J* = 10.0 Hz, 1 H), 7.29 (d, *J* = 8.5 Hz, 1 H), 7.28 (d, *J* = 8.5 Hz, 1 H), 7.16 (d, *J* = 8.5 Hz, 1 H), 7.14 (d, *J* = 8.5 Hz, 1 H), 5.25 (d, *J* = 9.5 Hz, 1 H), 4.30 (dd, *J* = 15.0, 6.0 Hz, 1 H), 4.24 (dd, *J* = 15.0, 6.0 Hz, 1 H), 1.95 (s, 3 H), 1.92 (s, 3 H), 1.85 (s, 3 H). ¹³C NMR (126 MHz, DMSO-*d*₆): δ 169.82, 168.71, 162.64 (d, *J* = 243.05 Hz), 135.55 (d, *J* = 2.77 Hz), 129.88 (d, *J* = 8.32 Hz), 115.46 (d, *J* = 21.29 Hz), 60.04, 59.41, 41.98, 27.09, 25.37, 22.75. ¹⁹F NMR (377 MHz, DMSO-*d*₆): -115.99 (m). HRMS m/z (ESI): calculated for MH⁺ 328.1126, found 328.1131.

Characterization of target **3d**: ¹H NMR (500 MHz, DMSO-*d*₆): δ 8.50 (t, *J* = 5.0 Hz, 1 H), 8.35 (d, *J* = 9.5 Hz, 1 H), 7.24 (d, *J* = 8.5 Hz, 1 H), 7.23 (d, *J* = 8.5 Hz, 1 H), 7.09 (d, *J* = 8.5 Hz, 1 H), 7.07 (d, *J* = 8.5 Hz, 1 H), 5.15 (d, *J* = 10.0 Hz, 1 H), 3.36 (app septet, *J* = 7.0 Hz, 1 H), 3.28 (app septet, *J* = 7.0 Hz, 1 H), 2.76-2.66 (m, 2 H), 1.88 (s, 3 H), 1.86 (s, 3 H), 1.83(s, 3 H). ¹³C NMR (126 MHz, DMSO-*d*₆): δ 169.67, 168.56, 161.28 (d, *J* = 241.92 Hz), 135.76 (d, *J* = 3.15 Hz), 130.89 (d, *J* = 7.94 Hz), 115.31 (d, *J* = 2.67 Hz), 60.03, 59.37, 40.47, 34.17, 27.06, 25.23, 22.72. ¹⁹F NMR (377 MHz, DMSO-*d*₆): -117.28 (m). HRMS m/z (ESI): calculated for MH⁺ 342.1282, found 342.1323.

Characterization of target **3e**: ¹H NMR (500 MHz, DMSO-*d*₆): δ 8.90 (t, *J* = 5.0 Hz, 1 H), 8.38 (d, *J* = 9.5 Hz, 1 H), 7.40 (app pentet, *J* = 9.5 Hz, 1 H), 7.09 (t, *J* = 8.0 Hz, 2 H), 5.22 (d, *J* = 9.5 Hz, 1 H), 4.39 (dd, *J* = 14.5, 5.5 Hz, 1 H), 4.28 (dd, *J* = 14.5, 5.5 Hz, 1 H), 1.91 (s, 3 H), 1.88 (s, 3 H), 1.82 (s, 3 H). ¹³C NMR (126 MHz, DMSO-*d*₆): δ 169.68, 168.34, 161.50 (dd, *J* = 248.47, 8.32 Hz), 130.50 (t, *J* = 10.46 Hz), 113.95 (t, *J* = 19.40 Hz), 111.88 (dd, *J* = 19.91, 5.67 Hz), 60.20, 59.06, 30.91 (t, *J* = 3.65 Hz), 27.07, 25.06, 22.73. ¹⁹F NMR (377 MHz, DMSO-*d*₆): - 114.73 (t, *J* = 5.66 Hz). HRMS m/z (ESI): calculated for MH⁺ 346.1031, found 346.1069.

Characterization of target **3f**: ¹H NMR (500 MHz, DMSO-*d*₆): δ 9.03 (t, *J* = 5.5 Hz, 1 H), 8.47 (d, *J* = 10.0 Hz, 1 H), 7.11 (t, *J* = 9.5 Hz, 1 H), 6.95 (d, *J* = 7.0 Hz, 2 H), 5.26 (d, *J* = 9.5 Hz, 1 H), 4.30 (app d, *J* = 6.0 Hz, 2 H), 1.97 (s, 3 H), 1.94 (s, 3 H), 1.87 (s, 3 H). ¹³C NMR (126 MHz, DMSO-*d*₆): δ 169.96, 169.14, 162.79 (dd, *J* = 246.58, 13.36 Hz), 144.18 (t, *J* = 8.95 Hz), 110.67 (dd, *J* = 19.78, 5.80 Hz), 102.70 (t, *J* = 25.70 Hz), 59.72, 59.55, 41.94, 26.88, 25.52, 22.73. ¹⁹F NMR (377 MHz, DMSO-*d*₆): -105.39 (t, *J* = 6.79 Hz). HRMS m/z (ESI): calculated for MH⁺ 346.1031, found 346.1038.

Characterization of target **3g**: ¹H NMR (500 MHz, DMSO-*d*₆): δ 9.08 (t, *J* = 5.5 Hz, 1 H), 8.49 (d, *J* = 9.5 Hz, 1 H), 7.61-7.56 (m, 4 H), 5.28 (d, *J* = 9.5 Hz, 1 H), 4.41 (dd, *J* = 15.5, 6.0 Hz, 1 H), 4.35 (dd, *J* = 15.5, 6.0 Hz, 1 H), 1.96 (s, 3 H), 1.93 (s, 3 H), 1.86 (s, 3 H). ¹³C NMR (126 MHz, DMSO-*d*₆): δ 169.87, 169.06, 140.90, 131.91, 129.77, 129.48 (q, *J* = 31.50 Hz), 124.67 (q, *J* = 272.54 Hz), 124.19 (q, *J* = 3.78 Hz), 124.03 (q, *J* = 3.78 Hz), 59.80, 59.46, 42.16, 26.87, 25.45, 22.70. ¹⁹F NMR (377 MHz, DMSO-*d*₆): -61.19 (s). HRMS m/z (ESI): calculated for MH⁺ 378.1094, found 378.1124.

Characterization of target **3h**: ¹H NMR (500 MHz, DMSO-*d*₆): δ 8.54 (t, *J* = 5.0 Hz, 1 H), 8.37 (d, *J* = 9.5 Hz, 1 H), 7.58-7.50 (m, 4 H), 5.13 (d, *J* = 9.5 Hz, 1 H) 3.47 (app septet, *J* = 7.0 Hz, 1 H), 3.32 (app septet, *J* = 7.0 Hz, 1 H), 2.85 (t, *J* = 6.5 Hz, 2 H), 1.82(s, 9 H). ¹³C NMR (126 MHz, DMSO-*d*₆): δ 169.65, 168.62, 141.16, 133.40, 129.60, 129.41 (q, *J* = 31.50 Hz), 125.72 (q, *J* = 3.78 Hz), 124.75 (q, *J* = 272.79 Hz), 123.35 (q, *J* = 3.65 Hz), 59.97, 59.32, 40.06, 34.55, 27.01, 25.09, 22.67. ¹⁹F NMR (377 MHz, DMSO-*d*₆): -61.30 (s). HRMS m/z (ESI): calculated for MH⁺ 392.1250, found 392.1341.

Characterization of target **3i**: ¹H NMR (500 MHz, DMSO-*d*₆): δ 9.02 (t, *J* = 4.0 Hz, 1 H), 8.48 (d, *J* = 6.5 Hz, 1 H), 7.16 (t, *J* = 5.0 Hz, 1 H), 5.25 (d, *J* = 6.5 Hz, 1 H), 4.28 (app d, *J* = 4.5 Hz, 2 H), 1.97 (s, 3 H), 1.92 (s, 3 H), 1.87 (s, 3 H). ¹³C NMR (126 MHz, DMSO-*d*₆): δ 170.01, 169.19, 150.51 (ddd, *J* = 177.03, 6.93, 2.52 Hz), 138.70 (t, *J* = 11.09 Hz), 137.29 (t, *J* = 11.09 Hz), 136.90 (m), 112.10 (dd, *J* = 12.47, 2.65 Hz), 59.62, 41.58, 26.83, 25.54, 22.71. ¹⁹F NMR (377 MHz, DMSO-*d*₆): -135.50 (dd, *J* = 21.49, 9.05 Hz), -164.08 (tt, *J* = 21.87, 6.79 Hz). HRMS m/z (ESI): calculated for MH⁺ 364.0937, found 364.0963.

Characterization of target **3j**: ¹H NMR (500 MHz, DMSO-*d*₆): δ 9.09 (t, *J* = 5.5 Hz, 1 H), 8.48 (d, *J* = 9.5 Hz, 1 H), 7.74 (t, *J* = 8.0 Hz, 1 H), 7.33 (d, *J* = 12.0 Hz, 1 H), 7.28 (d, *J* = 8.0 Hz, 1 H), 5.28 (d, *J* = 9.5 Hz, 1 H), 4.39 (app d, *J* = 5.5 Hz, 2 H), 1.98 (s, 3 H), 1.94 (s, 3 H), 1.88 (s, 3 H). ¹³C NMR (126 MHz, DMSO-*d*₆): δ 169.98, 169.22, 159.27 (d, *J* = 253.76 Hz), 147.96 (d, *J* = 7.69 Hz), 127.71 (dd, *J* = 16.13, 4.03 Hz), 124.04 (d, *J* = 18.77 Hz), 123.12 (q, *J* = 272.03 Hz), 115.99 (t, *J* = 20.79 Hz), 115.44 (qd, *J* = 32.38, 12.22 Hz), 59.69, 59.57 (d, *J* = 17.51 Hz), 41.99, 26.89 (d, *J* = 4.16 Hz), 25.49 (d, *J* = 3.02 Hz), 22.73. ¹⁹F NMR (377 MHz, DMSO-*d*₆): -59.83 (d, *J* = 12.06 Hz), -116.29 (m). HRMS m/z (ESI): calculated for MH⁺ 396.1000, found 396.1009.

Characterization of target **3k**: ¹H NMR (500 MHz, DMSO- d_6): δ 9.09 (t, J = 5.5 Hz, 1 H), 8.49 (d, J = 9.5 Hz, 1 H), 7.55 (d, J = 9.0 Hz, 1 H), 7.47 (s, 1 H), 7.41 (d, J = 9.5 Hz, 1 H), 5.28 (d, J = 9.5 Hz, 1 H), 4.41 (dd, J = 15.5, 6.5 Hz, 1 H), 4.37 (dd, J = 15.5, 6.5 Hz, 1 H), 1.96 (s, 3 H), 1.94 (s, 3 H), 1.87 (s, 3 H). ¹³C NMR (126)

MHz, DMSO- d_6): δ 169.96, 169.27, 162.32 (d, J = 246.83 Hz), 144.29 (d, J = 7.56 Hz), 131.28 (qd, J = 32.83, 8.57 Hz) 123.75 (qd, J = 273.17, 3.15 Hz), 120.41 (t, J = 3.28 Hz), 118.75 (d, J = 21.80 Hz), 111.61 (dd, J = 24.95, 3.65 Hz), 59.60, 59.56, 41.84, 26.76, 25.55, 22.69. ¹⁹F NMR (377 MHz, DMSO- d_6): -61.36 (s), -111.07 (t, J = 9.05 Hz). HRMS m/z (ESI): calculated for MH⁺ 396.1000, found 396.1009.

Characterization of target **3I**: ¹H NMR (500 MHz, DMSO-*d*₆): δ 9.15 (t, *J* = 7.0 Hz, 1 H), 8.51 (d, *J* = 12.0 Hz, 1 H), 7.99 (s, 1 H), 7.93 (s, 2 H), 5.28 (d, *J* = 12.0 Hz, 1 H), 4.51 (dd, *J* = 20.0, 7.5 Hz, 1 H), 4.44 (dd, *J* = 20.0, 7.5 Hz, 1 H), 1.96 (s, 3 H), 1.94 (s, 3 H), 1.87 (s, 3 H). ¹³C NMR (126 MHz, DMSO-*d*₆): δ 169.97, 169.40, 143.07, 130.57 (q, *J* = 32.89 Hz), 128.43 (d, *J* = 2.77 Hz), 123.78 (q, *J* = 273.29 Hz), 121.00 (septet, *J* = 3.78 Hz), 59.57, 59.46, 41.80, 26.62, 25.58, 22.64. ¹⁹F NMR (377 MHz, DMSO-*d*₆): -61.48 (s). HRMS m/z (ESI): calculated for MH⁺ 446.0968, found 446.0981.

Characterization of target **3m**: ¹H NMR (500 MHz, DMSO-*d*₆): δ 8.96 (t, *J* = 5.5 Hz, 1 H), 8.43 (d, *J* = 9.5 Hz, 1 H), 7.34-7.31 (m, 2 H), 7.26-7.23 (m, 3 H), 5.26 (d, *J* = 9.5 Hz, 1 H), 4.32 (dd, *J* = 15.0, 6.0 Hz, 1 H), 4.27 (dd, *J* = 15.0, 6.0 Hz, 1 H), 1.96 (s, 3 H), 1.93 (s, 3 H), 1.85 (s, 3 H). ¹³C NMR (126 MHz, DMSO-*d*₆): δ 169.80, 168.70, 139.32, 128.72, 127.87, 127.33, 60.11, 59.44, 42.71, 27.12, 25.40, 22.76. HRMS m/z (ESI): calculated for MH⁺ 310.1220, found 310.1243.

Characterization of target **3n**: ¹H NMR (500 MHz, DMSO-*d*₆): δ 8.94 (t, J = 6.0 Hz, 1 H), 8.45 (d, J = 9.5 Hz, 1 H), 6.56 (s, 2 H), 5.27 (d, J = 10.0 Hz, 1 H), 4.28 (dd, J = 15.0, 6.0 Hz, 1 H), 4.21 (dd, J = 15.0, 6.0 Hz, 1 H), 3.74 (s, 6 H), 3.62 (s, 3 H), 1.99 (s, 3 H), 1.97 (s, 3 H), 1.86 (s, 3 H). ¹³C NMR (126 MHz, DMSO-*d*₆): δ 169.86, 168.79, 153.19, 136.65, 135.02, 104.73, 60.42, 59.90, 59.61, 56.16, 42.63, 27.06, 25.56, 22.74. HRMS m/z (ESI): calculated for MH⁺ 400.1537, found 400.1732.

3. Decompostion kinetic studies of NO donors

All the donors were dissolved in DMSO as the stock solution for kinetic studies. Both aerobic kinetics studies of photolytic and thermal decomposition of NO donors in a mixture of PBS and DMSO (50:50, v/v) were performed using either a PerkinElmer Lambda 35 or Shimadzu UV-1601 UV-Vis spectrophotometer. The photolytic decomposition of fluorinated NO donors in a mixture of PBS and DMSO (50:50, v/v) at room temperature ($23 \,^{\circ}$ C) was irradiated under a Rayonet photochemical reactor (RMR-600), which was equipped with eight 350 nm lamps (4 W). The photolysis was manually scanned by the UV-Vis spectrophotometer at room temperature ($23 \,^{\circ}$ C) after each irradiation time of 0, 15, 30, 45, 60, 90, 120, 180, 240, 360, 480, 600 and 780 s. The corresponding kinetic data was analyzed

with a first-order rate equation. Kinetics of **3I** were monitored at 282 nm, and the others' kinetics were analyzed at 342 nm. The thermal decomposition kinetic studies in a mixture of PBS and DMSO (50:50, v/v) at 37 °C were monitored at 342 nm overnight with a time period of 10 min. The kinetic profiles were analyzed using either a zero-order or a first-order rate equation. All UV-Vis data were proceeded using the Origin 8.0 software.

4. Analysis of decomposition products from NO donor 3k

A Sievers 280i chemiluminescence Nitric Oxide Analyzer (NOA) was utilized to determine the NO gas released from NO donor **3k** (1.50×10^{-7} mol). To have a rapid NO release, photolysis of NO donor **3k** in a mixture of PBS and DMSO (3 mL, 50:50, v/v) at room temperature (23 °C) was performed under a Rayonet photochemical reactor (RMR-600) with 350 nm lamps (4 W, 8 lamps). NO detection signals over time were integrated to calculate the total moles of NO released in each photolysis from the NO donor (1.50×10^{-7} mol). To determine the stoichiometric ratio, the photolysis of NO donor **3k** (1.50×10^{-7} mol) was independently carried out three times.

The disulfide derived from NO donor **3k** during the NO release in pure DMSO- d_6 was characterized by ¹H NMR spectroscopy. Before the NO release, NO donor **3k** (2.98 mol) in pure DMSO- d_6 was recorded by ¹H NMR spectroscopy. Then two separate sample of NO donor **3k** (2.98 mM) in pure DMSO- d_6 was prepared with trace amount of 3-trimethylsilylpropionic-2,2,3,3- d_4 acid sodium salt (chemical shift: 0 ppm). One sample was photolyzed at room temperature (23 °C) under a Rayonet photochemical reactor (RMR-600) with 350 nm lamps (4 W, 8 lamps), and the corresponding products was characterized by ¹H NMR spectroscopy and HRMS (ESI). The other sample covered by foil paper was kept in the oven set at 37 °C for 4 day, which was further recorded by ¹H NMR spectroscopy.

5. Theoretical calculations

All calculations were performed using Gaussian16.^[1] Geometry optimizations were done using UB3LYP^[2] with 6-311G(d,p) basis set. NO donors **1**, **3e**, **3I** and **3m** were selected as model compounds in our computational calculations to simulate this system. The trans conformation of RSNO was obtained as the most stable structure. In addition, it is found that the triplet state energy of the reactants is higher than that of the corresponding singlet state by 23-25 kcal/mol. Therefore, singlet state was assigned as the ground state. During the singlet transition state search process, no saddle point was observed, indicating that the S-N bond may not have a transition state in the homolysis process with the formation of NO. The optimized structure for the singlet ground-state of NO donors SNAP (**1**), **3e**, **3I** and **3m** were shown in Figure 1S, and the calculated S-N bond dissociation enthalpies (BDE) were listed in Table S3.



Figure S2. The optimized structures of NO donors (1, 3e, 3I and 3m).

Table S3. The calculated S-N bond dissociation enthalpies (BDE) of NO donors (1, 3e, 3l and3m).

NO donors	BDE (S-N) (kcal/mol)
1	25.11
3e	21.52
31	21.30
3m	19.01

6. References

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7. NMR spectra









-90 ppm

-110

-130

-150

-170

-190

-70

20

0 -10

-30

-50

-8000 -7000 -5000 -5000 -3000 -2000 -1000 -0 -1000











































¹⁹F NMR (377 MHz, CDCI₃)



ppm

 ż

 Ó

-1

-2









































13C NMR (126 MHz, DMSO-d a)





















































¹H NMR (500 MHz, DMSO-d₆)





















