Supporting Information for

Regio- and Stereo-Selective [4+4] Photodimerization of Angular-shaped Dialkyltetracenedithiophene

Tze-Gang Hsu,^a Hsiao-Chieh Chou,^a Min-Ju Liang,^a Yu-Ying Lai^{*b} and Yen-Ju Cheng^{*a}

^aDepartment of Applied Chemistry, National Chiao Tung University, 1001 University Road, Hsinchu, 30010, Taiwan (R.O.C) E-mail: yjcheng@mail.nctu.edu.tw

^bInstitute of Polymer Science and Engineering, National Taiwan University, No.1, Sec. 4, Roosevelt Rd., Taipei 10617, Taiwan (R.O.C.) E-mail: yuyinglai@ntu.edu.tw

Table of contents

| 1. | General remarks |
|------|---|
| 2. | Synthetic details |
| 3 | Time-dependent ¹ H NMR spectra of reversibility of <i>syn</i> -[2,2]-daTDT1S10 |
| 4. | Structural characterization of <i>syn</i> -[2,2]-daTDT by NMR spectroscopy |
| 4.1. | Possible isomers |
| 4.2. | Heteronuclear single quantum coherence (HSQC) spectroscopy to establish the one-bond C–H relations in daTDT1 |
| 4.3. | Heteronuclear multiple bond correlation (HMBC) spectroscopy to establish the long-range C–H relations in daTDT1 |
| 4.4. | Rotating-frame nuclear Overhauser effect correlation spectroscopy (ROESY) of aTDT1 and daTDT2 |
| 5. | Concentration dependent emission spectra of aTDT1 |
| 6. | Powder x-ray diffraction (XRD) pattern of aTDT1 and the simulated pattern |
| 7. | Photodimerization of aTDT2 thin film after irradiation of 15 min |
| 8. | X-ray diffraction (XRD) pattern of aTDT2 powder and thin film |
| 9. | Single crystal of aTDT1 |
| 10. | ¹ H and ¹³ C NMR spectra |
| 11. | Reference |

1. General remarks

All chemicals were obtained from commercial sources and used as received unless otherwise specified. Anhydrous toluene was obtained from a SD-300 solvent purification system (AsiaWong Enterprise). ¹H and ¹³C NMR spectra were measured using Agilent 400-MR DD2 (400MHz) or Agilent VNMRS600 (600 MHz) instrument spectrometers. Chemical shifts (δ values) are reported in ppm with respect to CHCl₃ (δ = 7.26 ppm for ¹H NMR and δ = 77 ppm for ¹³C NMR). Coupling constants (J) are given in Hz. ¹³C NMR was proton broad-band-decoupled. Multiplicities of peaks are denoted by the following abbreviations: s, singlet; d, doublet; t, triplet; m, multiplet. High resolution molecular weight was determined by a high resolution gas chromatograph mass spectrometer (type: AccuTOF GCX, JEOL) at the center of advanced instrumentation, National Chiao Tung university. Heteronuclear single quantum coherence (HSQC) spectroscopy, heteronuclear multiple bond correlation (HMBC) spectroscopy, rotating-frame nuclear Overhauser effect correlation spectroscopy (ROESY), National Chiao Tung university with Agilent VNMRS600 spectrometers. Absorption spectra were collected with a Hitachi U-4100 UV/Vis spectrophotometer. Fluorescence spectra were recorded with a Horiba FluoroMax 4 spectrometer. Mass spectra are collected with JEOL AccuTOF GCX High Resolution Gas Chromatograph Mass Spectrometer. X-ray crystal structure of aTDT1 was determined by the instrumentation center, National Tsing Hua University with a single-crystal diffractometer (type: Bruker APEX DUO).

2. Synthetic details

Scheme S1 Synthetic route to syn-[2,2]daTDT



1,3,5,7-tetrabromo-2,6-dihydroxynaphthalene, 1,5-dibromo-2,6-dihydroxynaphthalene, 3,7dibromo-2,6-bis(trifluoromethanesulfonyloxy)naphthalene were synthesized based on the reported procedures.¹

Synthesis of 3,7-di(thiophen-2-yl)naphthalene-2,6-diyl bis(trifluoromethanesulfonate)



A solution of 3,7-dibromonaphthalene-2,6-diyl bis(trifluoromethanesulfonate) (1.75 g, 3.0 mmol), tributyl(thiophen-2-yl)stannane (2.24 g, 6.0 mmol) in toluene (50 mL) was degassed under nitrogen for 1 h. Pd(PPh₃)₄ (174 mg, 0.15 mmol) and tri-*tert*-butylphosphine (31 mg, 0.15 mmol) was added and the mixture was stirred at 120 °C for 12 h, quenched with water (50 mL), and extracted with ethyl acetate (100 mL). The combined organic layer was washed with brine solution (100 mL) and dried over anhydrous MgSO₄. After filtration, the solvent was removed under

vacuum and the residue was washed by MeOH. Recrystallization from MeOH furnished 3,7di(thiophen-2-yl)naphthalene-2,6-diyl bis(trifluoromethanesulfonate) **2** as a white solid (1.09 g, yield: 61%). ¹H NMR (CDCl₃, 400 MHz): δ 8.09 (s, 2 H), 7.91 (s, 2 H), 7.51 (d, *J* = 4.8 Hz, 2 H), 7.48 (d, *J* = 3.2 Hz, 2 H), 7.19 (dd, *J* = 4.8 Hz, *J* = 3.2 Hz, 2 H); ¹³C NMR (CDCl₃, 100 MHz): ¹³C NMR (100 MHz, CDCl₃) δ 145.77, 135.66, 131.23, 130.38, 128.74, 128.34, 128.10, 127.77, 123.26, 120.22, 120.07, 116.88, 113.68; HRMS (FD, C₂₀H₁₀O₆F₆S₄⁺⁻): calcd, 587.9258; found, 587.9256.

Synthesis of 2,2'-(3,7-di(dodec-1-yn-1-yl)naphthalene-2,6-diyl)dithiophene



To a mixture of 3,7-di(thiophen-2-yl)naphthalene-2,6-diyl bis(trifluoromethanesulfonate) 2 (450 mg, 0.76 mmol), dodec-1-yne (350 mg, 1.6 mmol), Pd(PPh₃)₂Cl₂ (140 mg, 0.15 mmol), CuI (40 mg, 0.15 mmol), and triphenylphosphine (53 mg, 0.15 mmol) was added THF/diisopropylamine (20 mL/20 mL) degassed by nitrogen. The mixture was then stirred for 8 h, quenched with water (30 mL), and extracted with ethyl acetate (100 mL). The combined organic layer was washed with brine solution (100 mL) and dried over anhydrous MgSO₄. After filtration, the solvent was removed under vacuum and the residue was purified by column chromatography (hexane:ethyl acetate=30:1). Recrystallization from MeOH furnished 2,2'-(3,7-di(dodec-1-yn-1vl)naphthalene-2,6-divl)dithiophene **3** as a white solid (410 mg, yield: 86%). ¹H NMR (CDCl₃, 400 MHz): δ 7.97 (s, 2 H), 7.86 (s, 2 H), 7.64-7.65 (m, 2 H), 7.37 -7.38 (m, 2 H), 7.11-7.13 (m, 2 H), 2.45 (t, J = 7.2 Hz, 4 H), 1.58-1.64 (m, 4 H), 1.41-1.45 (m, 4 H), 1.26–1.28 (m, 24 H), 0.88 (t, J =6.8 Hz, 6 H); ¹³C NMR (CDCl₃, 100 MHz): 142.0, 133.4, 133.2, 131.1, 127.1, 127.0, 126.8, 125.7, 121.0, 95.9, 80.3, 31.9, 29.6, 29.55, 29.3, 29.2, 29.0, 28.4, 22.7, 19.8, 14.1; HRMS (EI, C₄₂H₅₂S₂⁺): calcd, 620.3505; found, 620.3491.

Synthesis of 2,2'-(3,7-bis(4-octyltetradec-1-yn-1-yl)naphthalene-2,6-diyl)dithiophene



To a mixture of 3,7-di(thiophen-2-yl)naphthalene-2,6-diyl bis(trifluoromethanesulfonate) **2** (225 mg, 0.38 mmol), 9-(prop-2-yn-1-yl)nonadecane (338 mg, 0.84 mmol), Pd(PPh₃)₂Cl₂ (71 mg, 0.1 mmol), CuI (20 mg, 0.76 mmol), and triphenylphosphine (27 mg 0.76 mmol) was added THF/diisopropylamine (13 mL/13 mL) degassed by nitrogen. The mixture was then stirred for 8 h, quenched with water (30 mL), and extracted with ethyl acetate (100 mL). The combined organic layer was washed with brine solution (100 mL) and dried over anhydrous MgSO₄. After filtration, the solvent was removed under vacuum and the residue was purified by column chromatography (hexane:ethyl acetate=30:1). Recrystallization from MeOH furnished 2,2'-(3,7-bis(4-octyltetradec-1-yn-1-yl)naphthalene-2,6-diyl)dithiophene **4** as a white solid (327 mg, yield: 95%). ¹H NMR (CDCl₃, 400 MHz): δ 7.98 (s, 2 H), 7.87 (s, 2 H), 7.65 (d, *J* = 3.6 Hz, 2 H), 7.37 (d, *J* = 5.2 Hz, 2 H), 7.12 (dd, *J* = 5.2 Hz, 2 H), 7.87 (s, 2 H), 7.65 (d, *J* = 3.6 Hz, 2 H), 7.37 (d, *J* = 5.2 Hz, 2 H), 7.12 (dd, *J* = 6.4 Hz, 12 H); ¹³C NMR (CDCl₃, 100 MHz): δ 142.1, 133.4, 133.3, 131.1, 127.2, 127.0, 126.9, 125.7, 121.2, 94.7, 81.1, 37.23, 33.7, 32.0, 31.9, 30.0, 29.7, 29.7, 29.7, 29.4, 26.9, 24.1, 22.7, 14.1.; HRMS (FD, C₆₂H₉₂S₂⁺): caled, 900.6635; found, 900.6622.

Synthesis of aTDT1



To a solution of 2,2'-(3,7-di(dodec-1-yn-1-yl)naphthalene-2,6-diyl)dithiophene **3** (200 mg, 0.322 mmol) in toluene (10 mL) was added PtCl₂ (8.6 mg, 0.0322 mmol). The mixture was stirred for 12 h at 110 °C, filtered with celite, and extracted by toluene. The solvent was removed under vacuum and the residue was recrystallized from toluene to give **aTDT1** as an orange solid (156 mg, yield: 78%). ¹H NMR (CDCl₃, 400 MHz): δ 8.81 (s, 2 H), 8.61 (s, 2 H), 7.60 (s, 2 H), 7.56 (s, 4 H), 3.04 (t, *J* = 8.0 Hz, 4 H), 1.82-1.86 (m, 4 H), 1.27–1.49 (m, 28 H), 0.88 (s, *J* = 7.0 Hz, 6 H); HRMS (EI, C₄₂H₅₂S₂): calcd, 620.3505; found, 620.3478. ¹³C NMR is not detected due to low solubility.

Synthesis of aTDT2



To a solution of 2,2'-(3,7-bis(4-octyltetradec-1-yn-1-yl)naphthalene-2,6-diyl)dithiophene 4 (200 mg, 0.22 mmol) in toluene (7 mL) was added PtCl₂ (5.9 mg, 0.022 mmol). The mixture was stirred for 12 h at 110 °C, filtered with celite, and extracted by toluene. The solvent was removed under vacuum and the residue was purified by column chromatography (hexane:ethyl acetate=30:1). Recrystallized from MeOH to give **aTDT2** as a yellow solid (98 mg, yield: 49%). ¹H NMR (CDCl₃, 400 MHz): δ 8.81 (s, 2 H), 8.61 (s, 2 H), 7.54 (s, 6 H), 2.94 (d, *J* = 6.4 Hz, 4 H), 1.93-1.85 (m, 2 H), 1.44-1.18 (m, 64 H), 0.86-0.80 (m, 12 H); ¹³C NMR (100 MHz, CDCl₃) δ 137.49, 137.25, 134.78,

130.28, 129.99, 126.69, 126.04, 124.83, 124.46, 124.08, 120.78, 39.57, 38.18, 33.57, 31.90, 30.05, 29.65, 29.33, 26.59, 22.67, 14.10; HRMS (FD, C₆₂H₉₂S₂⁺⁻): calcd, 900.6635; found, 900.6642.

Synthesis of syn-[2,2]-daTDT1



A solution of **aTDT1** (10 mg, 0.016 mmol) in CDCl₃ (15 mL) was irradiated by a daylight lamp (3M, DL5000 OSRAM DULUX, 20W/800. 50/60Hz. 6000K) with a filter (Company: ROCOES; Product: P490/585, pink: $T_{ave}>88\%$ @ 610-700nm, $T_{ave}>80\%$ @ 420-460nm, $T_{ave}<10\%$ @ 520-550nm) and the mixture was stirred for 1 h at room temperature. The solvent was removed under vacuum to afford quantitatively *syn*-[2,2]-daTDT1 as a brown oil (10 mg, yield: 99 %).¹H NMR (CDCl₃, 400 MHz): δ 8.11 (s, 2 H), 7.93 (s, 2 H), 7.53 (s, 4 H), 7.49 (s, 2 H), 7.46 (s, 4 H), 7.32 (s, 2 H), 6.42 (s, 2 H), 6.29 (s, 2 H), 3.01 (t, *J* = 7.8 Hz, 4 H), 2.94 (t, *J* = 7.6 Hz, 4 H), 1.74-1.77 (m, 4 H), 1.67-1.71 (m, 4 H), 1.25–1.38 (m, 56 H), 0.87 (t, *J* = 6.8 Hz, 12 H); ¹³C NMR (CDCl₃, 125 MHz): δ 139.1, 138.0, 137.8, 136.9, 135.4, 134.9, 134.4, 134.3, 130.4, 130.2, 126.9, 125.8, 125.1, 123.8, 123.2, 122.7, 122.6, 120.2, 118.1, 79.8, 78.0, 34.2, 34.1, 31.9, 30.9, 30.3, 29.61, 29.59, 29.54, 29.51, 29.3, 22.7, 14.1; HRMS (FD, C₈₄H₁₀₄S₄⁺): calcd, 1240.7015; found, 1240.6997. Synthesis of syn-[2,2]-daTDT2



A solution of **aTDT2** (10 mg, 0.011 mmol) in CDCl₃ (5 mL) was irradiated by a daylight lamp (3M, DL5000 OSRAM DULUX, 20W/800. 50/60Hz. 6000K) with a filter (Company: ROCOES; Product: P490/585, pink: $T_{ave} > 88\%$ @ 610-700nm, $T_{ave} > 80\%$ @ 420-460nm, $T_{ave} < 10\%$ @ 520-550nm) and the mixture was stirred for 15 minutes at room temperature. The solvent was removed under vacuum to afford quantitatively *syn*-[2,2]-daTDT2 as a brown oil (10 mg, yield: 99 %). ¹H NMR (CDCl₃, 400 MHz): $\delta 8.12$ (s, 2 H), 7.93 (s, 2 H), 7.52 (s, 4 H), 7.45 (s, 6 H), 7.28 (s, 2 H), 6.43 (s, 2 H), 6.29 (s, 2 H), 2.95-2.89 (m, 4 H), 2.89-2.82 (m, 4 H), 1.82-1.77 (m, 2 H), 1.75-1.69 (m, 2 H), 1.33-1.16 (m, 128 H) 0.89-0.81 (m, 24 H); HRMS (FD, C₁₂₄H₁₈₄S₄⁺⁻): calcd, 1801.3275; found, 1801.3260.



3. Time-dependent ¹H NMR spectra of reversibility of *syn*-[2,2]-daTDT1

Figure S1 Time-dependent ¹H NMR spectra of reversibility of syn-[2,2]-daTDT1

4. Structural characterization of syn-[2,2]-daTDT by NMR spectroscopy

4.1. Possible isomers



Scheme S2 Six possible stereoisomers and corresponding symmetry point groups for daTDT

4.2. Heteronuclear single quantum coherence (HSQC) spectroscopy to establish the one-bond C-H relations in daTDT1.

The HSQC spectrum of **daTDT1** is shown in **Figure S2**, in which 14 C–H correlations were found. Correlations- a", b", and e" arise from the phenyl and naphthalenyl moieties. Correlations- c" and d" result from the outmost thienyl groups. Correlations- f" and g" belong to the bridgehead C–H pairs. The rest are the C–H couplings related to the decanyl aliphatic side chains.





4.3. Heteronuclear multiple bond correlation (HMBC) spectroscopy to establish the longrange C-H relations in daTDT1.

Magnified HMBC spectra of various regions are listed in **Figure S3**, **Figure S4**, **Figure S5**, **Figure S6**, and **Figure S7** respectively. In theory, C_{20} would couple with H_{22} and H_{18} in HMBC spectroscopy and C_{10} correlates with H_8 . Based on this principle, C_{10} and C_{20} can be assigned accordingly (**Figure S3**, correlation A, B, and C). Moreover, the characterization of H_{18} and H_{22} is assisted by the statement that $C_{\alpha L}$ would couple with H_{18} rather than H_{22} (**Figure S7**, correlation Bx). In conjunction with the HSQC spectrum (**Figure S2**, correlation f'', g'', b'', e'', and a''), the NMR absorption peaks of C_{10}/H_{10} , C_{20}/H_{20} , C_8/H_8 , C_{18}/H_{18} , and C_{22}/H_{22} are determined.

HMBC correlation



Figure S3 Magnified HMBC spectrum of daTDT1

Subsequently, C_{21} would couple with H_{20} and H_{22} . The peak position of C_{21} can thus be confirmed (**Figure S4**, correlation D and E). Similarly, C_{19} would couple with H_{18} and H_{20} and C_{19} is then located (**Figure S4**, correlation F and G). C_9 would couple with H_8 and H_{10} and the chemical shift of C_9 can be verified (**Figure S4**, correlation H and I). H_{10} would couple with C_9 and C_{11} . Since the position of C_9 is identified, the other correlation should be H_{10} – C_{11} (**Figure S4**, correlation J). The recognition of C_{11} is accomplished.

⇐⇒ HMBC correlation



Figure S4 Magnified HMBC spectrum of daTDT1.

As shown in **Figure S5**, searching along the vertical direction of correlation H (H₈–C₉), correlation K and L are found. They should correspond to H₈–C₇ and H₈–C₆. Correlation M (H_{α R}–C₆) and L are linked to the same carbon, which is assigned as C₆. Therefore, one can deduce that K is H₈–C₇ and L is H₈–C₆. Following a similar strategy, correlation N and O are detected along the perpendicular line of E (H₂₂–C₂₁). N and O are H₂₂–C₂₃ and H₂₂–C₂₄ couplings. Correlation P (H₆–C₂₃) and N are associated with the same carbon, which is designated C₂₃. Since N is the H₂₂–C₂₃ signal, C₂₄ can thus be identified.

HMBC correlation



Figure S5 Magnified HMBC spectrum of daTDT1.



Figure S6 Magnified HMBC spectrum of daTDT1.

In **Figure S6**, one can use $H_{\alpha R}$ to label C_5 via correlation Q ($H_{\alpha R}$ – C_5). Q, R, and S are located at the same horizontal line, suggesting that the protons all relate to C_5 . Through connecting P (H_6 – C_{23}) and R (H_6 – C_5), the identity of S should be H_3 – C_5 . In association with the HSQC spectrum (**Figure S2**, correlation c''), one can then infer that V is H_3 - C_2 or H_2 - C_3 and W is H_3 - C_4 . Furthermore, T ($H_{\alpha L}$ – C_{17}) is used to identify C_{17} , which can then be employed to find correlation U and X relating to C_{17} . The signal G (H_{18} – C_{19}) can differentiate U (H_{18} – C_{17}) and X (H_{15} – C_{17}). The identification of H_{15} together with the HSQC spectrum (**Figure S2**, correlation d"), Z (H_{15} – C_{14} or H_{14} – C_{15}) and Y (H_{15} – C_{16}) are recognized. There is still one aromatic carbon not being characterized. By eliminating all other possibilities, it should be C_{12} at 137 ppm.

The proton and carbon ($H_{\alpha R}$, $C_{\alpha R}$, $H_{\beta R}$, $C_{\beta R}$, $H_{\alpha L}$, $C_{\alpha L}$, $H_{\beta L}$, $C_{\beta L}$, $H_{internal}$, $C_{internal}$, $H_{terminal}$, and $C_{terminal}$) absorptions in the aliphatic region can be easily assigned. The assignment is summarized in **Figure S7**.



Figure S7 Magnified HMBC spectrum of daTDT1

4.4. Rotating-frame nuclear Overhauser effect correlation spectroscopy (ROESY) of aTDT1 and daTDT2



Figure S8 ROESY spectrum of aTDT1. (H_{α} - H_{β} coupling only)



Figure S9 Magnified ROESY spectrum of daTDT2.

5. Concentration dependent emission spectra of aTDT1



Figure S10 Absorption spectrum (dash line) and concentration dependent emission spectra of aTDT1.

6. The powder x-ray diffraction (XRD) pattern of aTDT1 and the simulated pattern



Figure S11. The powder x-ray diffraction (XRD) pattern of aTDT1 ($\lambda = 1.320898$ Å) and the simulated pattern derived from the aTDT1 single crystal structure ($\lambda = 1.54056$ Å).

7. Photodimerization of aTDT2 thin film after irradiation of 15 min



Figure S12 Photodimerization of aTDT2 thin film, (a) before (b) after irradiation of 15 min.

8. X-ray diffraction (XRD) pattern of aTDT2 powder and thin film



Figure S13 The x-ray diffraction (XRD) pattern of aTDT2 powder and thin film.

9. Single crystal data of aTDT1

 $Crystal \ data \ and \ structure \ refinement \ for \ cu_141239lt_0m_a.$

| Identification code | cu_141239lt_0m_a | | |
|--|---|-------------------------|--|
| Empirical formula | C42 H52 S2 | | |
| Formula weight | 620.95 | | |
| Temperature | 100(2) K | | |
| Wavelength | 1.54178 Å | | |
| Crystal system | Monoclinic | | |
| Space group | P 21/c | | |
| Unit cell dimensions | a = 4.8832(3) Å | α= 90°. | |
| | b = 7.6289(4) Å | β= 90.819(3)°. | |
| | c = 45.134(3) Å | $\gamma = 90^{\circ}$. | |
| Volume | 1681.21(17) Å ³ | | |
| Ζ | 2 | | |
| Density (calculated) | 1.227 Mg/m ³ | | |
| Absorption coefficient | 1.636 mm ⁻¹ | | |
| F(000) | 672 | | |
| Crystal size | 0.20 x 0.02 x 0.01 mm ³ | | |
| Theta range for data collection | 0.979 to 66.614°. | | |
| Index ranges | -5<=h<=3, -8<=k<=9, -53<=l<=50 | | |
| Reflections collected | 11508 | | |
| Independent reflections | 2958 [R(int) = 0.0379] | | |
| Completeness to theta = 67.679° | 97.6 % | | |
| Absorption correction | Semi-empirical from equivalents | | |
| Max. and min. transmission | 0.9492 and 0.7898 | | |
| Refinement method | Full-matrix least-squares on F ² | | |
| Data / restraints / parameters | 2958 / 0 / 201 | | |

| Goodness-of-fit on F ² | 1.100 | |
|-----------------------------------|------------------------------------|--|
| Final R indices [I>2sigma(I)] | R1 = 0.0832, wR2 = 0.2677 | |
| R indices (all data) | R1 = 0.0857, wR2 = 0.2736 | |
| Extinction coefficient | n/a | |
| Largest diff. peak and hole | 0.514 and -0.492 e.Å ⁻³ | |

10. ¹H and ¹³C NMR spectra



























11. Reference

 S. Shinamura, I. Osaka, E. Miyazaki, A. Nakao, M. Yamagishi, J. Takeya, K. Takimiya, J. Am. Chem. Soc. 2011, 133, 5024.