

## Electronic supporting information

### Chiral hydrogen-bonded supramolecular capsules: Synthesis, characterization and complexation of C<sub>70</sub>

Martha Kohlhaas,<sup>[a]</sup> Manfred Zähres,<sup>[b]</sup> Christian Mayer,<sup>[b]</sup> Marianne Engeser,<sup>[c]</sup> Christian Merten<sup>[d]</sup> and Jochen Niemeyer\*<sup>[a]</sup>

*[a] Institute of Organic Chemistry and Center for Nanointegration Duisburg- Essen (CENIDE),  
University of Duisburg-Essen, Universitätsstrasse 7, 45141 Essen, Germany,  
E-mail: jochen.niemeyer@uni-due.de*

*[b] Department of Physical Chemistry and Center for Nanointegration Duisburg-Essen  
(CENIDE), University of Duisburg-Essen, 45141 Essen, Germany.*

*[c] Kekulé-Institute for Organic Chemistry and Biochemistry, University of Bonn, Gerhard-  
Domagk-Str. 1, 53121 Bonn, Germany*

*[d] Ruhr-Universität Bochum, Organische Chemie II, Universitätsstrasse 150,  
44801 Bochum, Germany*

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# 1. General information

## 1.1. Analytical methods

Melting points were measured with a Büchi Melting-Point B-540 apparatus with open end glass capillary tubes.

IR spectra were measured on a Jasco FT/IR-430 spectrometer.

NMR spectra were recorded with a Bruker DMX 300 spectrometer [ $^1\text{H}$ : 300 MHz,  $^{13}\text{C}$ : 75.5 MHz,  $^{31}\text{P}$ : 121.5 MHz] or with a Bruker DMX 600 spectrometer [ $^1\text{H}$ : 600 MHz,  $^{13}\text{C}$ : 151 MHz,  $^{31}\text{P}$ : 243 MHz]. All measurements were performed at room temperature, using  $[\text{D}_1]$ -chloroform,  $[\text{D}_6]$ -dimethylsulfoxide or  $[\text{D}_4]$ -methanol as solvents. The chemical shifts are referenced relative to the residual proton signals of the solvents in the  $^1\text{H}$ -NMR ( $[\text{D}_1]$ -chloroform:  $\delta = 7.24$  ppm,  $[\text{D}_4]$ -methanol:  $\delta = 3.31$  ppm,  $[\text{D}_6]$ -dimethylsulfoxide:  $\delta = 2.50$  ppm) or relative to the solvent signal in the  $^{13}\text{C}$ -NMR ( $[\text{D}_1]$ -chloroform:  $\delta = 77.16$  ppm,  $[\text{D}_4]$ -methanol:  $\delta = 49.15$  ppm,  $[\text{D}_6]$ -dimethylsulfoxide:  $\delta = 39.51$  ppm). In case of the  $[\text{D}_1]$ -chloroform/ $[\text{D}_4]$ -methanol mixtures, the  $[\text{D}_1]$ -chloroform signals were used for referencing. The apparent coupling constants are given in Hertz. The description of the fine structure means: s = singlet, bs = broad singlet, d = doublet, ps d = pseudo doublet, dd = doublet of doublets, dt = doublet of triplets, t = triplet, m = multiplet.

DOSY-spectra were recorded on a Bruker DRX 500 or on a Bruker Avance Neo 500 spectrometer equipped with a gradient unit (maximum z-gradient of 1200 G/cm) and a DIFF30 probe equipped with a  $^1\text{H}/^2\text{H}$  coil. Analysis of the data was performed using the Stejskal-Tanner-equation for relevant integral areas. Hydrodynamic radii were calculated using the Stokes-Einstein equation using the correction for cylindrical particles. Diameters and heights of the capsules **1** and **2** were estimated from their DFT-calculated structures.

Low resolution ESI mass spectra were recorded on a Bruker Amazon SL spectrometer. High resolution ESI mass spectra were recorded on a Bruker Maxis 4G spectrometer.

Mass spectra of the capsule (*all-R*)-**2** were measured with an ESI-Q/TOF instrument (Bruker micrOTOF-Q) in positive and negative mode. Samples in varying concentrations in the range of  $10^{-5}$  -  $5 \cdot 10^{-4}$  M in a mixture  $\text{CHCl}_3/\text{CH}_3\text{OH}$  8/2 were introduced into the ESI source with flow rates of 3-7  $\mu\text{L}/\text{min}$ . Other solvents (acetonitrile, acetone) did not lead to better results. Ionizing parameters were tuned from normal to very soft ionization conditions (source temperature 30 - 200  $^\circ\text{C}$ , quadrupole voltage 1-4 V, collision energy voltage 1-6 V).

The UV-VIS and CD spectra were recorded on a JASCO J-815 spectropolarimeter at 20  $^\circ\text{C}$ . The quartz cuvettes were from Hellma@Analytics type 100-QS (1 mm light path) for UV-VIS and CD. All solvents used were of spectrometric grade.

The fluorescence spectra were recorded on a Shimadzu RF-6000 spectro fluorometer at 25 °C. The quartz cuvettes were from Hellma®Analytics type 101-QS (10 mm light path) for fluorescence. All solvents used were of spectrometric grade.

## 1.2. Materials and Methods

### *Materials*

For thin layer chromatography (TLC) analysis throughout this work, Polygram® SIL G/UV254 TLC plates (silica gel 0.2 mm, 40 × 80 mm) were used. Visualization of the spots was carried under a 254 nm UV light source. The products were purified by flash column chromatography on silica gel 60M (40-63 µm) which was purchased from MACHEREY-NAGEL GmbH & Co. KG. Eppendorf Research® Plus pipettes were used with epT.I.P.S. pipette tips for exact volumes.

### *Solvents*

Dry pyridine was distilled from KOH and stored over molecular sieves under argon. Aqueous sodium carbonate solution (2 M) was degassed by bubbling with argon for 15 minutes. Phosphoryl chloride (POCl<sub>3</sub>) was distilled under vacuum and stored in a Schlenk flask under argon. Aqueous work-ups and column chromatographies were carried out using technical grade solvents. Ethyl acetate was distilled prior to use. Dry tetrahydrofuran was distilled freshly from Na/benzophenone prior to use. Dimethoxyethane (DME) was distilled and stored under argon prior to use.

### *Chemicals*

Copper(I) iodide, tetrabutylammonium fluoride and Pd(PPh<sub>3</sub>)<sub>4</sub> were purchased from Sigma-Aldrich and used without further purification.

Sodium hydride (60% dispersion in mineral oil), triisopropylborate, terephthaloyl dichloride and bromoethane were purchased from TCI and used without further purification.

Bromine, boron tribromide, *n*-butyllithium (2.5 M in hexane), 1-bromo-4-iodobenzene, sodium iodide, ethylamine solution (70 % in water) and [1,1'-Bis(diphenylphosphino)ferrocene]palladium(II) dichloride [(dppf)PdCl<sub>2</sub>] were purchased from ACROS Organics and used without further purification.

Sodium chloride, potassium chloride and hydrochloric acid (12 M) were purchased from Bernd Kraft GmbH and used without further purification.

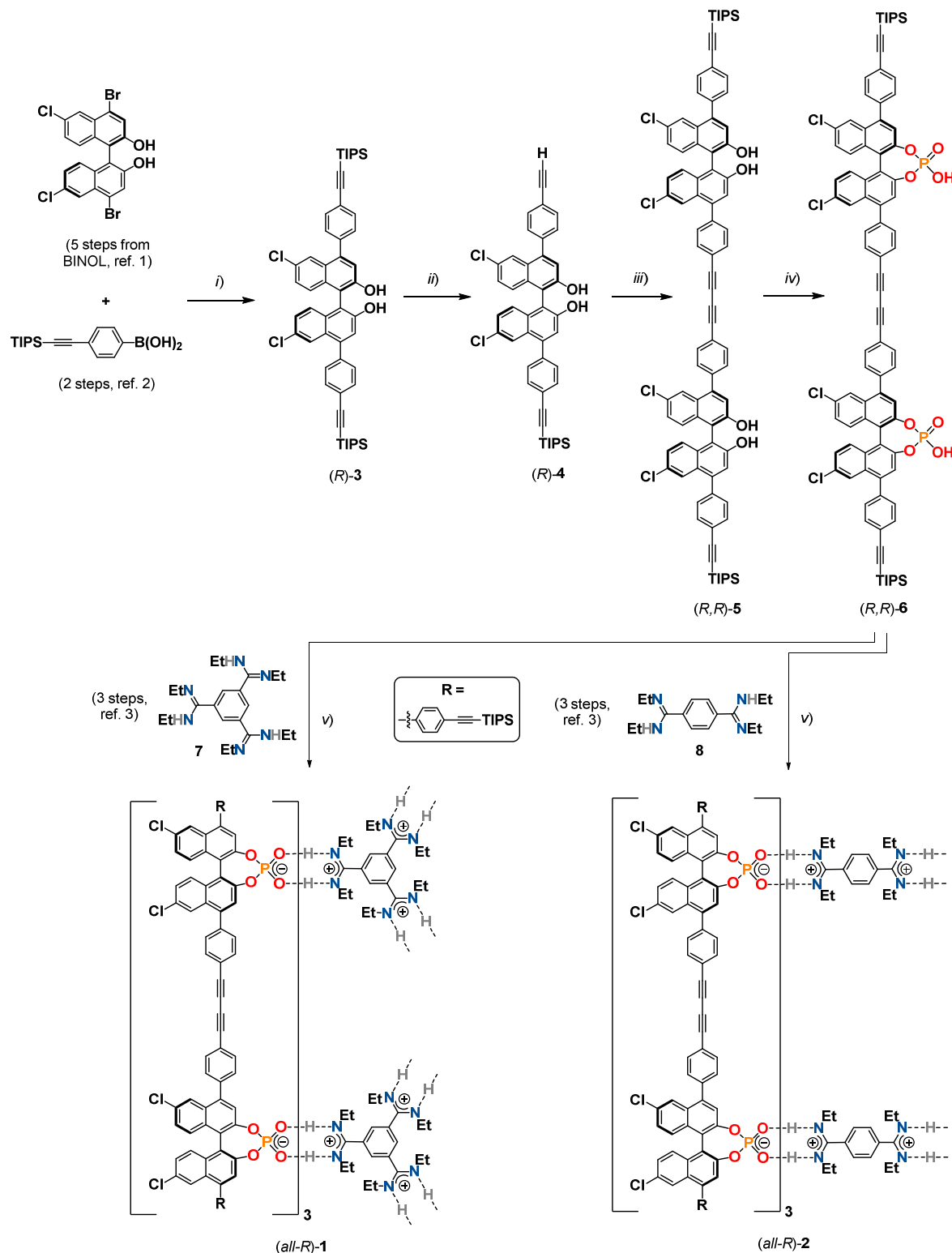
Thionyl chloride, ethylamine solution (2M in tetrahydrofuran) and hexachloroethane were purchased from Alfa Aesar and used without further purification.

1,1'-Bis(diphenylphosphino)palladium(II) dichloride was purchased from Fluorochem and used without further purification.

(*S*)- and (*R*)-1,1'-Binaphthyl-2,2'-diol (>99.9% ee) were purchased from RCA Separations and used without further purification.

## 2. Synthetic procedures

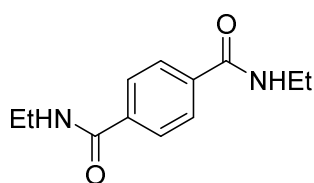
### 2.1. Overview



**Figure S 1:** Synthesis of the capsules **1** and **2**. Reagents and conditions: *i*) 2.4 equiv. of boronic acid, (dppf)PdCl<sub>2</sub>, Na<sub>2</sub>CO<sub>3</sub>, 85 °C, DME/Na<sub>2</sub>CO<sub>3</sub> (2 M), 91%; *ii*) TBAF (0.8 equiv.), 25 °C, RT, THF, 34% (alongside with 57% reisolated (R)-3); *iii*) (PPh<sub>3</sub>)<sub>2</sub>PdCl<sub>2</sub>, CuI, 25 °C, THF/CHCl<sub>3</sub>, 98%; *iv*) POCl<sub>3</sub>, pyridine, 60°C, then H<sub>2</sub>O, 26%; *v*) [D<sub>1</sub>]-chloroform / [D<sub>4</sub>]-methanol = 8 / 2 (v/v), 25 °C, 5 min.

## 2.2. Synthesis of the amidines

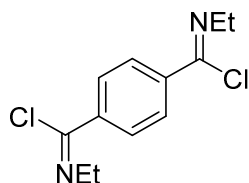
### 2.2.1. *N,N'*-diethylterephthalamide<sup>[1]</sup>



Terephthaloyl dichloride (3.00 g, 14.8 mmol, 1 eq.) was added to ethylamine (70% in water, 17.8 ml, 222 mmol, 15 eq.) at 0° C and stirred at 0° C for 30 minutes. After the reaction was completed, water (2 ml) were added alongside with concentrated hydrochloric acid (3 ml), ensuring that the pH stays basic. The product was filtered off and washed with water (2 x 15 ml) and methanol (2 x 15 ml), then dried under vacuum. This work up yielded analytically pure product (2.92 g, 13.3 mmol, 90.0 %).

C<sub>12</sub>H<sub>16</sub>N<sub>2</sub>O<sub>2</sub> (220.27 g/mol)

<sup>1</sup>H NMR (300 MHz, [D<sub>6</sub>]-dimethylsulfoxide) δ = 8.55 (t, *J* = 5.5 Hz, 2H), 7.88 (s, 2H), 3.29 (d, *J* = 7.3 Hz, 4H), 1.12 (t, *J* = 7.2 Hz, 6H).



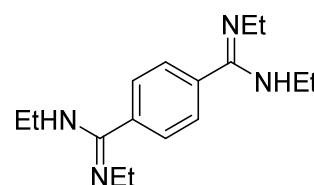
### 2.2.2. *N,N'*-diethylterephthalimidoyl dichloride<sup>[1]</sup>

*N,N'*-diethylterephthalamide (2.90 g, 13.2 mmol, 1 eq.) was placed under argon, and thionyl chloride (19.1 ml, 263 mmol, 20 eq.) was added dropwise. The reaction mixture was heated under reflux for three hours and stirred at room temperature overnight. After the reaction was completed, the excess thionyl chloride was removed under reduced pressure, then the crude oil was extracted with *n*-hexane (3 x 30 ml) and the solvent removed under reduced pressure. The work up yielded analytically pure product as a yellow oil (3.38 g, 11.3 mmol, 85.6 %).

C<sub>12</sub>H<sub>14</sub>Cl<sub>2</sub>N<sub>2</sub> (257.16 g/mol)

<sup>1</sup>H NMR (300 MHz, [D<sub>1</sub>]-chloroform, 298 K) δ = 8.04 (q, *J* = 0.8 Hz, 4H), 3.78 (q, *J* = 7.3 Hz, 4H), 1.36 (t, *J* = 7.3 Hz, 6H).

### 2.2.3. *N,N,N',N'*-tetraethyl-terephthalimidamide<sup>[1]</sup> (8)



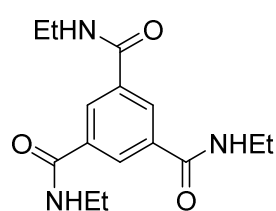
*N,N'*-diethylterephthalimidoyl dichloride (3.25 g, 12.6 mmol, 1 eq.) was dissolved in dichloromethane (20 ml) and added dropwise to a solution of ethylamine in tetrahydrofuran (2.0 M, 34.8 ml, 69.5 mmol, 5.5 eq.) at -10° C. The resulting solution was stirred at -10° C for an hour and overnight at room temperature. After the reaction was completed, the solvent was removed under reduced pressure and the remaining precipitate dissolved in water (30 ml). Sodium hydroxide (5

g, 125 mmol, 10 eq.) was added and the solution extracted with ethyl acetate (3 x 40 ml). The combined organic phases were dried over sodium sulfate, filtered and the solvent removed under reduced pressure. The crude product was purified using *Kugelrohr* distillation at 0.15 mbar and 220° C to give the analytically pure product **9** as a white solid (1.63 g, 5.95 mmol, 47.2 %).

C<sub>16</sub>H<sub>26</sub>N<sub>4</sub> (274.41 g/mol)

<sup>1</sup>H NMR (600 MHz, [D<sub>1</sub>]-chloroform, [D<sub>4</sub>]-methanol, 298 K) δ = 7.16 (s, 4H, Ar-H), 2.96 (bs, 8H, CH<sub>2</sub>), 0.98 (bs, 12H, CH<sub>3</sub>).

#### 2.2.4. *N,N',N''*-triethylbenzene-1,3,5-tricarboxamide<sup>[1]</sup>

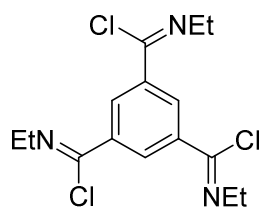


1,3,5-tricarboxybenzene (3.00 g, 14.3 mmol, 1 eq.) was dissolved in thionyl chloride (51.8 ml, 714 mmol, 50 eq.) alongside with dimethylformamide (one drop). The resulting mixture was heated under reflux for three hours and then the solvent removed under reduced pressure. A yellow oil was obtained, which was slowly added to ethylamine (70 % in water, 17.2 ml, 214 mmol, 15 eq.) at 0° C. The solution was stirred for 30 minutes, then diluted with water (30 ml). Concentrated hydrochloric acid (3 ml) was added and the mixture filtrated, washing the filtrate with water (2 x 5 ml) and methanol (2 x 5 ml). The workup yielded analytically pure product (3.07 g, 10.5 mmol, 73.4 %).

C<sub>15</sub>H<sub>21</sub>N<sub>3</sub>O<sub>3</sub> (291.35 g/mol)

<sup>1</sup>H NMR (300 MHz, [D<sub>6</sub>]-dimethylsulfoxide) δ = 8.66 (t, *J* = 5.5 Hz, 1H), 8.36 (s, 1H), 3.30 (qd, *J* = 7.2, 5.4 Hz, 2H), 1.13 (t, *J* = 7.2 Hz, 3H).

#### 2.2.5. *N,N',N''*-triethylbenzene-1,3,5-tris(carbonimidoyl trichloride)<sup>[1]</sup>



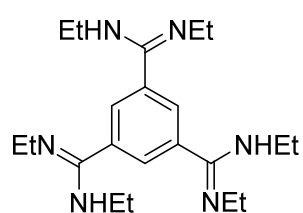
*N,N',N''*-Triethylbenzene-1,3,5-tricarboxamide (2.72 g, 9.32 mmol, 1 eq.) was dissolved in thionyl chloride (13.5 ml, 187 mmol, 20 eq.) and heated for three hours under reflux, then overnight at room temperature. After the reaction was completed, the excess thionyl chloride was removed under reduced pressure and the remaining solid extracted with *n*-hexane (3 x 10 ml). The combined organic phases were dried over magnesium sulfate, filtered and the solvent removed under reduced pressure. The workup yielded analytically pure solid (1.23 g, 3.57 mmol, 38.3 %).

C<sub>15</sub>H<sub>18</sub>Cl<sub>3</sub>N<sub>3</sub> (346.68 g/mol)

<sup>1</sup>H NMR (300 MHz, [D<sub>1</sub>]-chloroform, 298 K) δ = 8.66 (s, 3H), 3.79 (q, *J* = 7.3 Hz, 6H), 1.36 (t, *J* = 7.3 Hz, 9H).



### 2.2.6. *N,N,N',N',N'',N''*-hexaethylbenzene-1,3,5-tris(carboximidamide)<sup>[1]</sup> (**7**)

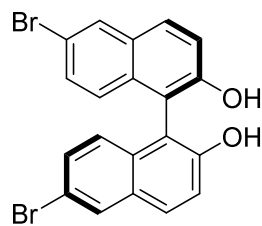


*N,N,N'*-triethylbenzene-1,3,5-tris(carboximidoyl trichloride) (1.23 g, 3.55 mmol, 1 eq.) was dissolved in dichloromethane (20 ml) and added dropwise to ethylamine solution (2.0 M in tetrahydrofuran, 21.3 ml, 42.5 mmol, 12 eq.) at  $-10^{\circ}$  C. The solution was stirred at  $-10^{\circ}$  C for an hour and overnight at room temperature. After the reaction went to completion, the solvent was removed under reduced pressure and the remaining solid dissolved in water (30 ml). Sodium hydroxide (1.4 g, 35.0 mmol, 10 eq.) was added and the solution extracted with ethyl acetate (3 x 50 ml), dried over sodium sulfate, filtered and the solvent removed under reduced pressure. The crude product was purified *via Kugelrohr* distillation at 0.35 mbar and  $295^{\circ}$  C. This workup yielded analytically pure product **8** as a white solid (1.06 g, 2.85 mmol, 80.3 %).

$C_{21}H_{36}N_6$  (372.56 g/mol)

$^1H$  NMR (300 MHz,  $[D_1]$ -chloroform,  $[D_4]$ -methanol, 298 K)  $\delta$  = 7.11 (s, 3H, Ar-H), 2.99 (bs, 8H,  $CH_2$ ), 0.99 (bs, 12H,  $CH_3$ ).

## 2.3. Synthesis of BINOL-precursors



### 2.3.1. (*R*)-6,6'-Dibromo-1,1'-binaphthalene-2,2'-diol<sup>[2]</sup>

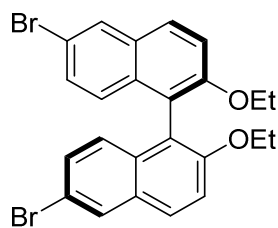
1,1'-Binaphthyl-2,2'-diol (20.0 g, 69.9 mmol, 1 eq.) was suspended in dichloromethane (400 ml) and cooled to  $-78^{\circ}\text{C}$ . Bromine (9.72 ml, 30.1 g, 189 mmol, 2.7 eq.) was dissolved in dichloromethane (100 ml) and was added dropwise to the suspension. The mixture was stirred for two hours at  $-78^{\circ}\text{C}$ , then overnight at room temperature.

After the reaction was completed, sodium bisulfite (3.65 M, 37 %, 5 eq., 349 mmol, 95.6 ml) was added at  $0^{\circ}\text{C}$  and the biphasic mixture was stirred for two hours. Dichloromethane (100 ml) and water (100 ml) were added and the phases separated. The aqueous phase was extracted with dichloromethane (100 ml), the organic phase washed with water (100 ml), then the organic phase was dried over magnesium sulfate, filtered and the solvent removed under reduced pressure. This workup yielded analytically pure product (30.3 g, 68.1 mmol, 97.5 %).

$\text{C}_{20}\text{H}_{12}\text{Br}_2\text{O}_2$  (444.12 g/mol)

$^1\text{H NMR}$  (300 MHz,  $[\text{D}_1]$ -Chloroform, 298 K)  $\delta = 7.90$  (d,  $J = 2.0$  Hz, 2H), 7.73 (dt,  $J = 8.9$ , 0.6 Hz, 2H), 7.27 – 7.18 (m, 4H), 6.82 (dt,  $J = 8.9$ , 0.6 Hz, 2H), 4.87 (bs, 2H).

### 2.3.2. (*R*)-6,6'-dibromo-2,2'-diethoxy-1,1'-binaphthalene<sup>[2]</sup>



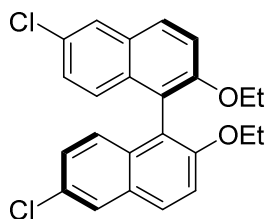
(*R*)-6,6'-Dibromo-1,1'-binaphthalene-2,2'-diol (30.3 g, 68.0 mmol, 1 eq.) was suspended in acetone (300 ml) alongside with sodium iodide (510 mg, 3.41 mmol, 0.05 eq.) and potassium carbonate (33.5 g, 241 mmol, 4 eq.). The suspension was refluxed for one hour, then bromoethane (31.5 ml, 408 mmol, 6 eq.) was added in four parts over a four hour period. The resulting solution was refluxed overnight.

After the reaction was completed, the solution was filtered and the solvent was removed under reduced pressure. The remaining solid was taken up in *n*-hexane (100 ml) and stirred for two hours at room temperature, then the supernatant was filtered off and the filtrate washed with *n*-hexane (2 x 10 ml), then dried under high vacuum. This workup yielded analytically pure product. (25.9 g, 51.7 mmol, 76.0 %).

$\text{C}_{24}\text{H}_{20}\text{Br}_2\text{O}_2$  (500.23 g/mol)

**<sup>1</sup>H NMR (300 MHz, [D<sub>1</sub>]-chloroform, 298 K)** δ = 7.99 (d, *J* = 2.1 Hz, 2H, Ar-H), 7.83 (d, *J* = 9.0 Hz, 2H, Ar-H), 7.40 (d, *J* = 9.0 Hz, 2H, Ar-H), 7.25 (dd, *J* = 9.0, 2.1 Hz, 2H, Ar-H), 6.95 (dt, *J* = 9.1, 0.6 Hz, 2H, Ar-H), 4.09-3.95 (m, 4H, CH<sub>2</sub>), 1.05 (t, *J* = 7.0 Hz, 6H, CH<sub>3</sub>).

### 2.3.3. (*R*)-6,6'-dichloro-2,2'-diethoxy-1,1'-binaphthalene<sup>[2]</sup>



(*R*)-6,6'-dibromo-2,2'-diethoxy-1,1'-binaphthalene (25.9 g, 51.7 mol, 1 eq.) was dissolved in dry tetrahydrofuran (160 ml) in a Schlenk flask and cooled to -78° C. *N*-butyllithium (2.5 M in hexane, 62.0 ml, 155 mmol, 3 eq.) was added to the solution and the mixture was stirred for 30 minutes. Hexachloroethane (24.5 g, 103 mmol, 2 eq.) was dissolved in dry tetrahydrofuran (50 ml) and added dropwise at -78° C. The solution was stirred for two hours at -78° C, then warmed to room temperature overnight.

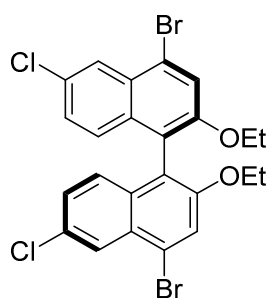
After the reaction was completed, ammonium chloride (100 ml) was added and the phases separated. The aqueous phase was extracted with ethyl acetate (2 x 200 ml), the combined organic phases were dried over magnesium sulfate, filtered and the solvent removed under reduced pressure.

The remaining solid was suspended in *n*-hexane (50 ml) and stirred for one hour. The supernatant was filtered off and the filtrate was washed with *n*-hexane (3 x 10 ml) and dried under high vacuum. This workup yielded analytically pure product (11.3 g, 27.5 mmol, 53.3 %).

C<sub>24</sub>H<sub>20</sub>Cl<sub>2</sub>O<sub>2</sub> (411.32 g/mol)

**<sup>1</sup>H NMR (300 MHz, [D<sub>1</sub>]-chloroform, 298 K)** δ = 7.83 (d, *J* = 9.0 Hz, 2H, Ar-H), 1H), 7.81 (d, *J* = 2.2 Hz, 2H, Ar-H), 7.41 (d, *J* = 9.0 Hz, 2H, Ar-H), 7.12 (dd, *J* = 9.1, 2.2 Hz, 2H, Ar-H), 7.00 (d, *J* = 9.0 Hz, 2H, Ar-H), 4.07-3.97 (m, 4H, CH<sub>2</sub>), 1.04 (t, *J* = 7.0 Hz, 6H, CH<sub>3</sub>).

### 2.3.4. (*R*)-4,4'-dibromo-6,6'-dichloro-2,2'-diethoxy-1,1'- binaphthalene<sup>[2]</sup>



(*R*)-6,6'-dichloro-2,2'-diethoxy-1,1'-binaphthalene (11.3 g, 27.6 mmol, 1 eq.) was dissolved in dichloromethane (400 ml) and cooled to -78° C. Bromine (21.3 ml, 66.0 g, 413 mmol, 15 eq.) was dissolved in dichloromethane (50 ml) and added dropwise to the solution over the course of an hour, then stirred at room temperature overnight.

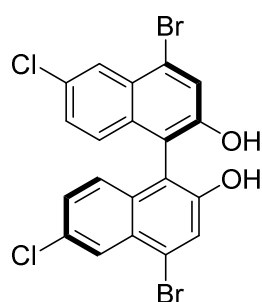
After the reaction was completed, sodium bisulfite (3.65 M, 37 %, 151 ml, 20 eq.) was added and the solution stirred for two hours at room temperature. The phases were then separated and the aqueous phase was extracted with dichloromethane (2 x 100 ml) and the organic phase washed with sodium chloride solution (2 x 100 ml), then the combined organic phases were dried over magnesium

sulfate, filtered, and the solvent removed under reduced pressure. The remaining solid was taken up in acetone (10 ml) and precipitated with *n*-hexane (75 ml). The solvent was removed, the solid suspended in *n*-hexane (50 ml) and stirred for one hour. After stirring, the supernatant was filtered off and the filtrate dried under high vacuum. This workup yielded analytically pure product (11.6 g, 20.4 mmol, 73.9 %).

C<sub>24</sub>H<sub>18</sub>Br<sub>2</sub>Cl<sub>2</sub>O<sub>2</sub> (569.11 g/mol)

<sup>1</sup>H NMR (300 MHz, [D<sub>1</sub>]-chloroform, 298 K) δ = 8.21 (d, *J* = 2.1 Hz, 2H, Ar-H), 7.71 (s, 2H, Ar-H), 7.16 (dd, *J* = 9.0, 2.1 Hz, 2H, Ar-H), 6.99 (d, *J* = 9.0 Hz, 2H, Ar-H), 4.07-3.97 (m, 4H, CH<sub>2</sub>), 1.06 (t, *J* = 7.0 Hz, 6H, CH<sub>3</sub>).

### 2.3.5. (*R*)-4,4'-dibromo-6,6'-dichloro-[1,1'-binaphthalene]-2,2'-diol<sup>[2]</sup>



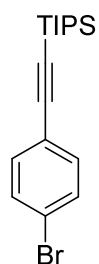
(*R*)-4,4'-dibromo-6,6'-dichloro-2,2'-diethoxy-1,1'-binaphthalene (11.6 g, 20.4 mmol, 1 eq.) was dissolved in dichloromethane (150 ml) and cooled to -78° C. Boron tribromide (4.83 ml, 12.5 g, 50.9 mmol, 2.5 eq.) was added and the solution was stirred at -78° C for two hours, then warmed up to room temperature overnight.

After the reaction was completed, sodium hydrogen carbonate (50 ml) was added, the phases were separated and the organic phase washed with sodium chloride solution (2 x 50 ml). The combined organic phases were dried over magnesium sulfate, filtered and the solvent removed under reduced pressure. This workup yielded analytically pure product (*R*)-**3** (9.65 g, 18.8 mmol, 92.2%).

C<sub>20</sub>H<sub>10</sub>Br<sub>2</sub>Cl<sub>2</sub>O<sub>2</sub> (513.01 g/mol)

<sup>1</sup>H NMR (300 MHz, [D<sub>1</sub>]-chloroform, 298 K) δ = 8.28 (d, *J* = 2.1 Hz, 1H, Ar-H), 7.74 (s, 1H, Ar-H), 7.28 (dd, *J* = 8.9, 2.1 Hz, 1H, Ar-H), 7.02 (d, *J* = 9.0 Hz, 1H, Ar-H), 4.96 (s, 2H, OH).

### 2.3.6. ((4-bromophenyl)ethynyl)triisopropylsilane<sup>[3]</sup>



Commercially available 1-bromo-4-iodobenzene (12.9 g, 45.7 mmol, 1 eq) and (triisopropylsilyl)acetylene (10.0 g, 12.3 ml, 54.8 mmol, 1.2 eq) were dissolved in piperidine (60 ml) in a Schlenk flask flushed with argon. The solution was degassed by bubbling argon through it for 15 minutes. The solution was cooled to 0° C and copper(I)iodide (0.17 g, 0.91 mmol, 0.02 eq) and bis(triphenylphosphine)palladium(II) dichloride (0.32 g, 0.46 mmol, 0.01 eq) were added. The solution was stirred under argon for two hours at 0° C and then for twelve hours at room temperature.

After the reaction was completed, the solution was poured into cold 2 M hydrochloric acid (300 ml). The resulting mixture was extracted with diethyl ether (300 ml), and the resulting organic phases were washed with 2 M hydrochloric acid (2 x 50 ml). The

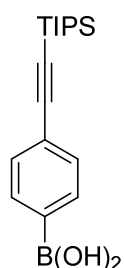
combined aqueous phases were extracted with diethyl ether (3 x 150 ml). The combined organic phases were dried over magnesium sulfate, filtered and the solvent was removed under vacuum. The resulting black oil was distilled using *via* Kugelrohr at 2.5 mbar and 150-170° C. The resulting yellow oil was obtained analytically pure (9.65 g, 28.6 mmol, 62.6 %).

Comment: The product contains a small amount (10%) of the disubstituted product (1,4-Bis(triisopropylsilylethynyl)benzene) as an impurity (<sup>1</sup>H NMR: δ = 7.37 ppm). This can be removed in the next step because it cannot be transformed into the more polar boronic acid and elutes more quickly during column chromatography.

C<sub>17</sub>H<sub>25</sub>BrSi (337.38 g/mol)

<sup>1</sup>H NMR (300 MHz, [D<sub>1</sub>]-chloroform, 298 K) δ = 7.41 (ps d, *J* = 8.6 Hz, 2H), 7.31 (ps d, *J* = 8.6 Hz, 2H, 2H), 1.11 (s, 3 H, TIPS-CH), 1.10 (s, 18 H, TIPS-CH<sub>3</sub>).

### 2.3.7. (4-((triisopropylsilyl)ethynyl)phenyl) boronic acid<sup>[3]</sup>



The synthesized ((4-bromophenyl)ethynyl)triisopropylsilane (9.65 g, 28.6 mmol, 1 eq.) was dissolved in dry tetrahydrofuran (300 ml) and cooled to -78° C in a multinecked flask under argon. After cooling, *n*-butyllithium was added (2.5 M in hexane, 14.9 ml, 37.2 mmol, 1.3 eq) and the solution stirred for 30 minutes. It was then transferred into a solution of triisopropylborate (19.8 ml, 85.8 mmol, 3 eq.) in dry tetrahydrofuran (50 ml) at -78° C in a multinecked flask under argon. Following the transfer, the resulting solution

was stirred at room temperature overnight.

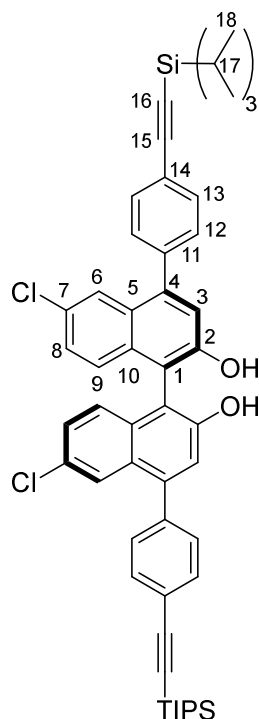
After the reaction was completed, hydrochloric acid (2 M, 200 ml) was added alongside with tetrahydrofuran (400 ml), and the reaction stirred for an additional hour. The phases were separated, sodium chloride solution (150 ml) was added, and the organic phase washed with sodium chloride solution (2 x 50 ml). The combined organic phases were dried over magnesium sulfate, filtered and the solvent removed under reduced pressure. The resulting white powder was purified using column chromatography (Cy: Et<sub>2</sub>O: MeOH 1 : 2 : 1) and obtained as analytically pure product (6.79 g, 22.5 mmol, 78.5 %).

C<sub>17</sub>H<sub>27</sub>BO<sub>2</sub>Si (302.30 g/mol)

<sup>1</sup>H NMR (300 MHz, [D<sub>6</sub>]- dimethylsulfoxide) δ = 8.17 (s, 2H, OH), 7.78 (ps d, *J* = 8.1 Hz, 2H, Ar-H), 7.41 (ps d, *J* = 8.1 Hz, 2H, Ar-H), 1.10 (s, 21 H, TIPS).

## 2.4. Synthesis of the bisphosphoric acid (*R,R*)-6

### 2.4.1. Synthesis of (*R*)-3



(*R*)-4,4'-dibromo-6,6'-dichloro-[1,1'-binaphthalene]-2,2'-diol (4.00 g, 7.80 mmol, 1 eq.) was dissolved in distilled dimethoxyethane (80 ml) alongside with (4-((triisopropylsilyl)ethynyl)phenyl)boronic acid (5.66 g, 18.7 mmol, 2.4 eq.) and [1,1'-Bis(diphenylphosphino)ferrocene]palladium(II) dichloride (318 mg, 0.390 mmol, 0.05 eq). Degassed sodium carbonate solution (2 M, 25.7 ml, 51.5 mmol, 6.6 eq.) was added and the reaction heated to reflux under argon overnight.

After the reaction was completed, saturated ammonium chloride solution (100 ml) and tetrahydrofuran (300 ml) were added. The phases were separated and the aqueous phase was extracted with tetrahydrofuran (2 x 100 ml). The combined organic phases were dried over magnesium sulfate, filtered and the solvent removed under reduced pressure. The resulting black solid was purified using column chromatography (Cy : EA = 4 : 1) and the product (*R*)-3 was obtained as a yellowish solid (6.77 g, 7.80 mmol, 90.8%).

Comment: For some batches of the starting dibromide, we observed that the coupling only worked at higher catalyst loadings (i.e. 20%-50%). Repurification of the dibromide by column chromatography did not alleviate this problem, only the synthesis of a new batch of the dibromide starting from BINOL was found to be helpful.

C<sub>54</sub>H<sub>60</sub>Cl<sub>2</sub>O<sub>2</sub>Si<sub>2</sub> (868.14 g/mol)

**<sup>1</sup>H NMR (600 MHz, [D<sub>1</sub>]-chloroform, 298 K)** δ = 7.85 (d, <sup>4</sup>J = 2.2 Hz, 2H, H-6), 7.68 (ps d, <sup>3</sup>J = 8.4 Hz, 4H, H-13), 7.51 (ps d, <sup>3</sup>J = 8.4 Hz, 4H, H-12), 7.35 (s, 2H, H-3), 7.27 (dd, <sup>3</sup>J = 9.0 Hz, <sup>4</sup>J = 2.2 Hz, 2H, H-8), 7.17 (dd, <sup>3</sup>J = 9.0 Hz, <sup>5</sup>J = 0.5 Hz, 2H, H-9), 5.11 (s, 2H, OH), 1.17 (s, 36H, H-18), 1.16 (s, 6H, H-17).

**<sup>13</sup>C NMR (151 MHz, [D<sub>1</sub>]-chloroform, 298 K)** δ = 152.4 (C-2), 142.8 (C-4), 139.1 (C-11), 132.5 (C-13), 132.4 (C-10), 130.7 (C-7), 129.8 (C-12), 128.7 (C-5), 128.6 (C-8), 126.4 (C-9), 125.7 (C-6), 123.6 (C-14), 119.7 (C-3), 110.6 (C-1), 106.7 (C-15), 92.1 (C-16), 18.9 (C-18), 11.5 (C-17).

**COSY (600 MHz, [D<sub>1</sub>]-chloroform, 298 K)** δ (<sup>1</sup>H) / δ (<sup>1</sup>H) = 7.85/7.27 (H-6/H-8), 7.68/7.51 (H-13/H-12), 7.51/7.68 (H-12/H-13), 7.27/7.85, 7.17 (H-8/H-6, H-9), 7.17/7.27 (H-9/H-8).

**HSQC (600/151 MHz, [D<sub>1</sub>]-chloroform, 298 K)**  $\delta$  (<sup>1</sup>H) /  $\delta$  (<sup>13</sup>C) = 7.85/125.7 (H-6/C-6), 7.68/132.5 (H-13/C-13), 7.51/129.8 (H-12/C-12), 7.35/119.7 (H-3/C-3), 7.27/128.6 (H-8/C-8), 7.17/126.4 (H-9/C-9), 1.17/18.9 (TIPS-CH<sub>3</sub>/TIPS-CH<sub>3</sub>), 1.16/11.5 (TIPS-CH/TIPS-CH).

**HMBC (600/151 MHz, [D<sub>1</sub>]-chloroform, 298 K)**  $\delta$  (<sup>1</sup>H) /  $\delta$  (<sup>13</sup>C) = 7.85/142.8, 132.4, 130.7, 128.6 (H-6/C-4, C-10, C-7, C-8), 7.68/139.1, 132.5, 129.8, 106.7 (H-13/C-11, C-13, C-12, C-15), 7.51/142.8, 132.5, 129.8, 123.6 (H-12/C-4, C-13, C-12, C-14), 7.35/152.4, 142.8, 139.1, 110.6 (H-3/C-2, C-4, C-11, C-1), 7.27/132.4; 130.7, 125.7 (H-8/C-10, C-7, C-6), 7.17/132.4, 130.7, 128.7, 125.7 (H-9/C-10, C-7, C-5, C-6), 5.11/152.4, 142.8, 119.7, 110.6 (OH/C-2, C-4, C-3, C-1), 1.17, 1.16/92.1, 18.9, 11.5 (H-18, H-17/C-16, C-18, C-17).

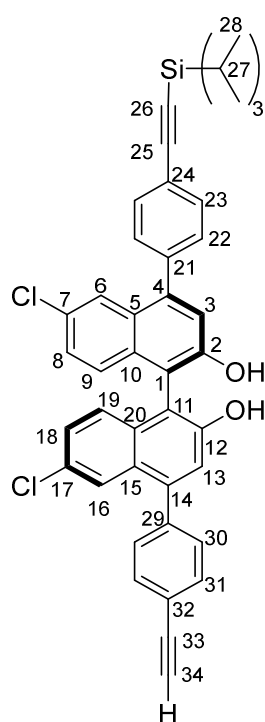
**Melting point:** 168°C

**MS (ESI, -Mode):**  $m/z$  = 865.3404 ([M-H]<sup>-</sup>, calculated:  $m/z$  = 865.3436 for [C<sub>54</sub>H<sub>60</sub>Cl<sub>2</sub>O<sub>2</sub>Si<sub>2</sub>]).

**IR (ATR)  $\nu$  bar:** 3526, 2941, 2863, 2360, 2156, 1588, 1496, 1461, 1379, 1140 cm<sup>-1</sup>.

**Elemental analysis** calcd. for C<sub>54</sub>H<sub>60</sub>Cl<sub>2</sub>O<sub>2</sub>Si<sub>2</sub>: C, 74.71; H, 6.97. Found: C, 74.5; H, 7.14.

#### 2.4.2. Synthesis of (*R*)-4



The Suzuki-coupling product (*R*)-3 (925 mg, 1.07 mmol, 1 eq.) was dissolved in tetrahydrofuran (30 ml). A solution of tetrabutylammonium fluoride (209 mg, 0.799 mmol, 0.7 eq.) in tetrahydrofuran (20 ml) was added dropwise *via* an addition funnel, then the solution was stirred overnight at room temperature.

The reaction was followed by TLC, and stopped after ca. 50% conversion. The solvent was removed under reduced pressure and the resulting yellow oil was purified *via* column chromatography (Cy : EA = 10 : 1). As a first fraction, reisolated starting material (*R*)-4 was obtained (529 mg, 0.609 mmol, 56.9%). As a second fraction, the desired product (*R*)-5 was obtained as a yellow solid (255 mg, 0.359 mmol, 33.6 %).

C<sub>45</sub>H<sub>40</sub>Cl<sub>2</sub>O<sub>2</sub>Si (711.80 g/mol)

**<sup>1</sup>H NMR (600 MHz, [D<sub>1</sub>]-chloroform, 298 K)**  $\delta$  = 7.85, 7.84 (each d, each <sup>4</sup>J = 2.1 Hz, each 1H, H-6 and H-16), 7.69 (ps d, <sup>3</sup>J = 8.2 Hz, 2H, H-31), 7.68 (ps d, <sup>3</sup>J = 8.2 Hz, 2H, H-23), 7.52 (ps d, <sup>3</sup>J = 8.2 Hz, 2H, H-30), 7.51 (ps d, <sup>3</sup>J = 8.2 Hz, 2H, H-22), 7.353 (s, 1H, H-13), 7.347 (s, 1H, H-3), 7.275, 7.271 (each dd, each <sup>3</sup>J = 9.0 Hz, <sup>4</sup>J = 2.1 Hz, each 1H, H-8 and H-18), 7.17, 7.16 (each d, each <sup>3</sup>J = 9.0 Hz,

each 1H, H-9 and H-19), 5.12 (bs, 2H, OH), 3.18 (s, 1H, H-34), 1.164 (s, 18H, H-28), 1.162 (s, 3H, H-27).

**<sup>13</sup>C NMR (151 MHz, [D<sub>1</sub>]-chloroform, 298 K)**  $\delta$  = 152.43, 152.42 (C-2, C-12), 142.8, 142.6 (C-4, C-14), 139.7 (C-29), 139.1 (C-21), 132.6, 132.5 (C-23, C-31), 132.40 132.37 (C-10, C-20), 130.78, 130.76 (C-7, C-17), 129.9, 129.8 (C-22, C-30), 128.7, 128.6 (C-8, C-18), 128.62, 128.60 (C-5, C-15), 126.4, 126.3 (C-9, C-19), 125.7, 125.6 (C-6, C-16), 123.6 (C-24), 122.2 (C-32), 119.84, 119.75 (C-3, C-13), 110.7, 110.6 (C-1, C-11), 106.7 (C-25), 92.1 (C-26), 83.4 (C-33), 78.0 (C-34), 18.9 (C-28), 11.5 (C-27).

**COSY (600 MHz, [D<sub>1</sub>]-chloroform, 298 K)**  $\delta$  (<sup>1</sup>H) /  $\delta$  (<sup>1</sup>H) = 7.85, 7.84 / 7.275, 7.271 (H-6, H-16 / H-8, H-18), 7.69, 7.68 / 7.52, 7.51 (H-31, H-23 / H-22, H-30), 7.52, 7.51 / 7.69, 7.68 (H-30, H-22 / H-23, H-31), 7.275, 7.271 / 7.85, 7.84, 7.17, 7.16 (H-8, H-18 / H-6, H-16, H-9, H-19), 7.17, 7.16 / 7.85, 7.84 (H-9, H-19 / H-8, H-18).

**HSQC (600/151 MHz, [D<sub>1</sub>]-chloroform, 298 K)**  $\delta$  (<sup>1</sup>H) /  $\delta$  (<sup>13</sup>C) = 7.85, 7.84 / 125.7, 125.6 (H-6, H-16 / C-6, C-16), 7.69, 7.68 / 132.6, 132.5 (H-23, H-31 / C-23, C-31), 7.52, 7.51 / 129.9, 129.8 (H-22, H-30 / C-22, C-30), 7.353, 7.347 / 119.84, 119.75 (H-3, H-13 / C-3, C-13), 7.275, 7.271 / 128.7, 128.6 (H-8, H-18 / C-8, C-18), 7.17, 7.16 / 126.4, 126.3 (H-9, H-19 / C-9, C-19), 3.18 / 78.0 (H-34 / C-34), 1.164 / 18.9 (H-28 / C-28), 1.162 / 11.5 (H-27 / C-27).

**HMBC (600/151 MHz, [D<sub>1</sub>]-chloroform, 298 K)**  $\delta$  (<sup>1</sup>H) /  $\delta$  (<sup>13</sup>C) = 7.85, 7.84 / 142.8, 142.6, 132.40, 132.37, 130.78, 130.76, 128.7, 128.6 (H-6, H-16 / C-4, C-14, C-10, C-20, C-7, C-17, C-8, C-18), 7.69 / 139.7, 132.6, 132.5, 129.9, 129.8, 83.4 (H-31 / C-29, C-31, C-30, C-33), 7.68 / 139.1, 132.6, 132.5, 129.9, 129.8, 106.7 (H-23 / C-21, C-23, C-22, C-25), 7.52 / 142.8, 142.6, 132.6, 132.5, 129.9, 129.8, 122.2 (H-30 / C-4, C-14, C-23, C-31, C-22, C-30, C-32), 7.51 / 142.8, 142.6 132.6, 132.5 129.9, 129.8 123.6 (H-22 / C-4, C-14, C-23, C-31, C-22, C-30, C-24), 7.353 / 152.43, 152.42, 139.7, 128.62, 128.60, 110.7, 110.6 (H-13 / C-12, C-29, C-15, C-11), 7.347 / 152.43, 152.42, 139.1, 128.62, 128.60, 110.7, 110.6 (H-3 / C-2, C-21, C-5, C-1), 7.275, 7.271 / 132.40, 132.37, 130.78, 130.76, 125.7, 125.6 (H-8, H-18 / C-10, C-20, C-7, C-17, C-6, C-16), 7.17, 7.16 / 130.78, 130.76, 128.62, 128.60, 110.7, 110.6 (H-9, H-19 / C-7, C-17, C-5, C-15, C-1, C-11), 3.18 / 132.6, 132.5, 122.2 (H-34 / C-31, C-32), 1.164, 1.162 / 18.9, 11.5 (H-28, H-27 / C-28, C-27).

**MS (ESI, -Mode):** m/z = 709.2091 ([M-H]<sup>-</sup>, calculated: m/z = 709.2102)

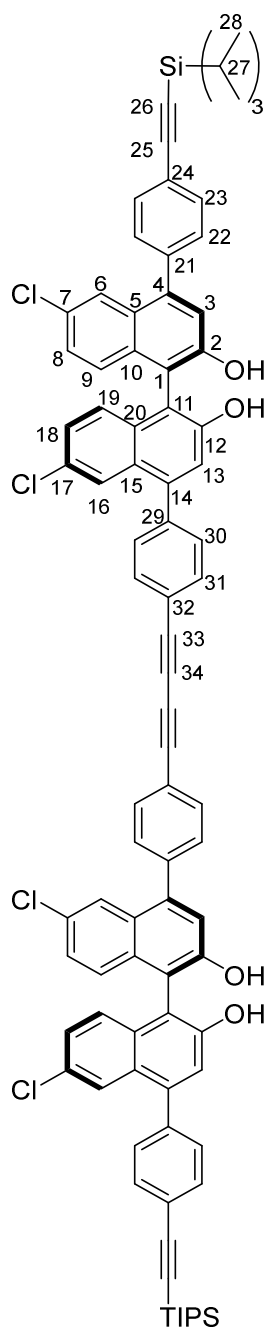
**IR (ATR)  $\nu$  bar:** 2940, 2863, 2360, 2156, 1587, 1496, 1461, 1379, 1263, 1140 cm<sup>-1</sup>

**Melting point:** 189°C

**Elemental analysis:** expected C = 75.93 %, H = 5.66 %, found C = 76.9 %, H = 6.22 %



### 2.4.3. Synthesis of (*R,R*)-5



The monodeprotected product (*R*)-4 (866 mg, 1.22 mmol, 1 eq.) was dissolved in a chloroform : tetrahydrofuran 1:2 mixture (135 ml) alongside with bis(triphenylphosphine)palladium(II) dichloride (42.8 mg, 0.06 mmol, 0.05 eq.) and copper(I)iodide (11.6 mg, 0.06 mmol, 0.05 eq.). To this was added triethylamine (1.25 ml, 9.76 mmol, 8 eq.) and the resulting solution was stirred at room temperature overnight.

After the reaction was completed, the solvent was removed under reduced pressure, then the resulting solid was taken up in dichloromethane (400 ml) and washed with hydrochloric acid (1 M, 3 x 100 ml). The combined organic phases were dried over sodium sulfate, filtered, and the solvent removed under reduced pressure. This workup yielded (*R*)-5 as a brown solid, containing minor impurities stemming from the catalysts (850 mg, 0.60 mmol, 98.0%). An analytically pure sample can be obtained by column chromatography (Cy : EA = 4 : 1).

$C_{90}H_{78}Cl_4O_4Si_2$  (1421.58 g/mol)

**$^1H$  NMR (600 MHz,  $[D_1]$ -chloroform, 298 K)**  $\delta$  = 7.87, 7.86 (each d, each  $^4J$  = 2.1 Hz, each 2H, H-6 and H-16), 7.77 (ps d,  $^3J$  = 8.2 Hz, 4H, H-31), 7.69 (ps d,  $^3J$  = 8.2 Hz, 4H, H-23), 7.58 (ps d,  $^3J$  = 8.2 Hz, 4H, H-30), 7.52 (ps d,  $^3J$  = 8.2 Hz, 4H, H-22), 7.38 (s, 2H, H-13), 7.36 (s, 2H, H-3), 7.29, 7.28 (each dd, each  $^3J$  = 9.0 Hz,  $^4J$  = 2.1 Hz, each 2H, H-8 and H-18), 7.19 (d,  $^3J$  = 8.7 Hz, 2H, H-19), 7.18 (d,  $^3J$  = 8.7 Hz, 2H, H-9), 5.14 (bs, 4H, OH), 1.167 (s, 36H, H-28), 1.165 (s, 6H, H-27).

**$^{13}C$  NMR (151 MHz,  $[D_1]$ -chloroform, 298 K)**  $\delta$  = 152.45, 152.43 (C-2, C-12), 142.9 (C-4) 142.4 (C-14), 140.2 (C-29), 139.1 (C-21), 133.0 (C-31), 132.5 (C-23), 132.44 132.37 (C-10, C-20), 130.9, 130.8 (C-7, C-17), 130.1 (C-30), 129.8 (C-22), 128.73,

128.68, 128.63, 128.56 (C-8, C-18, C-5, C-15), 126.5, 126.3 (C-9, C-19), 125.7, 125.5 (C-6, C-16), 123.7 (C-24), 121.9 (C-32), 119.9, 119.8 (C-3, C-13), 110.9 (C-11), 110.5 (C-1), 106.7 (C-25), 92.2 (C-26), 81.8 (C-33), 75.1 (C-34), 18.9 (C-28), 11.5 (C-27).

**COSY (600 MHz,  $[D_1]$ -chloroform, 298 K)**  $\delta$  ( $^1H$ ) /  $\delta$  ( $^1H$ ) = 7.87, 7.86 / 7.29, 7.28 (H-6, H-16 / H-8, H-18), 7.77 / 7.58 (H-31 / H-30), 7.69 / 7.52 (H-23 / H-22), 7.58 / 7.77 (H-30 / H-31), 7.52 / 7.69 (H-22 / H-23), 7.29, 7.28 / 7.87, 7.86, 7.19, 7.18 (H-8, H-18 / H-6, H-16, H-9, H-19), 7.19, 7.18 / 7.87, 7.86 (H-9, H-19 / H-8, H-18).

**HSQC (600/151 MHz, [D<sub>1</sub>]-chloroform, 298 K)**  $\delta$  (<sup>1</sup>H) /  $\delta$  (<sup>13</sup>C) = 7.87, 7.86 / 125.7, 125.5 (H-6, H-16 / C-6, C-16), 7.77 / 133.0 (H-31 / C-31), 7.69 / 132.5 (H-23 / C-23), 7.52 / 130.1 (H-30 / C-30), 7.38 / 129.8 (H-22 / C-22), 7.38, 7.36 / 119.9, 119.8 (H-3, H-13 / C-3, C-13), 7.29, 7.28 / 128.73, 128.68, 128.63, 128.56 (H-8, H-18 / C-8, C-18), 7.19, 7.18 / 126.5, 126.3 (H-9, H-19 / C-9, C-19), 1.167 / 18.9 (H-28 / C-28), 1.165 / 11.5 (H-27 / C-27).

**HMBC (600/151 MHz, [D<sub>1</sub>]-chloroform, 298 K)**  $\delta$  (<sup>1</sup>H) /  $\delta$  (<sup>13</sup>C) = 7.87, 7.86 / 142.9, 142.4, 132.44, 132.37, 130.9, 130.8, 128.73, 128.68, 128.63, 128.56 (H-6, H-16, C-4, C-14, C-10, C-20, C-7, C-17, C-8, C-18, C-5, C-15), 7.77 / 140.2, 133.0, 81.8 (H-31 / C-29, C-31, C-33), 7.69 / 139.1, 132.5, 106.7 (H-23 / C-21, C-23, C-25), 7.58 / 142.4, 130.1, 121.9 (H-30 / C-14, C-30, C-32), 7.52 / 142.9, 129.8, 123.7 (H-22 / C-4, C-22, C-24), 7.38 / 152.45, 152.43, 140.2, 128.73, 128.68, 128.63, 128.56, 110.9 (H-13 / C-12, C-29, C-15, C-11), 7.36 / 152.45, 152.43, 139.1, 128.73, 128.68, 128.63, 128.56, 110.5 (H-3 / C-2, C-21, C-5, C-1), 7.29, 7.28 / 132.44, 132.37, 125.7, 125.5 (H-8, H-18 / C-10, C-20, C-6, C-16), 7.19 / 130.9, 130.8, 128.73, 128.68, 128.63, 128.56, 110.9 (H-19 / C-17, C-15, C-11), 7.18 / 130.9, 130.8, 128.73, 128.68, 128.63, 128.56, 110.5 (H-9 / C-7, C-5, C-1), 1.167, 1.165 / 92.2, 18.9, 11.5 (H-28, H-27 / C-26, C-28, C-27).

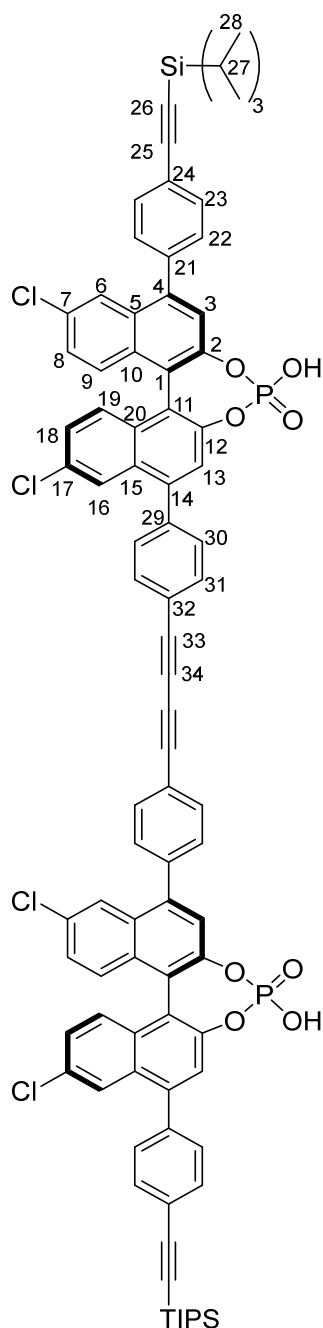
**MS (ESI, -Mode):** m/z = 1417.4151 ([M-H]<sup>-</sup>, calculated: m/z = 1417.4120)

**IR (ATR)  $\nu$  bar:** 2924, 2862, 2360, 2341, 2157, 1717, 1588, 1495, 1460, 1379, 1263, 1140 cm<sup>-1</sup>

**Melting point:** 172°C

**Elemental analysis:** expected C = 76.04 %, H = 5.53 %, found C = 75.4 %, H = 6.06 %

#### 2.4.4. Synthesis of (*R,R*)-6



The Glaser-coupling product (*R,R*)-5 (274 mg, 0.193 mmol, 1 eq.) was dissolved in dry pyridine (4 ml). Phosphorus oxychloride (141  $\mu$ l, 237 mg, 1.54 mmol, 8 eq.) was added to the solution and the reaction stirred at 60° C overnight.

After the reaction was completed, water (1.5 ml) was added to the solution and the reaction stirred for two hours at 60° C. The solvent was then removed under reduced pressure and the remaining solid taken up in dichloromethane (100 ml), then washed with hydrochloric acid (1 M, 5 x 10 ml). The combined organic phases were dried over sodium sulfate, filtered and the solvent removed under reduced pressure. The resulting solid was purified *via* column chromatography (DCM : MeOH = 10:1 to 1:1). The solvent was removed under reduced pressure, the solid taken up in a 10:1 mixture of dichloromethane : methanol (30 ml) and washed with hydrochloric acid (1 M, 3 x 10 ml). This yielded analytically pure product (*R,R*)-6 (77.9 mg, 50.4  $\mu$ mol, 26.1 %).

C<sub>90</sub>H<sub>76</sub>Cl<sub>4</sub>O<sub>8</sub>P<sub>2</sub>Si<sub>2</sub> (1545.51 g/mol)

**<sup>1</sup>H NMR (600 MHz, [D<sub>1</sub>]-chloroform, [D<sub>4</sub>]-methanol, 298 K)**  $\delta$  = 7.81, 7.80 (each d, each <sup>4</sup>*J* = 2.1 Hz, each 2H, H-6 and H-16), 7.65 (ps d, <sup>3</sup>*J* = 8.2 Hz, 4H, H-31), 7.56 (ps d, <sup>3</sup>*J* = 8.2 Hz, 4H, H-23), 7.48 (ps d, <sup>3</sup>*J* = 8.2 Hz, 4H, H-30), 7.43 (s, 2H, H-13), 7.41 (ps d, <sup>3</sup>*J* = 8.2 Hz, 4H, H-22), 7.40 (s, 2H, H-3), 7.30 (d, <sup>3</sup>*J* = 9.0 Hz, 2H, H-19), 7.28 (d, <sup>3</sup>*J* = 9.0 Hz, 2H, H-9), 7.18, 7.17 (each dd, each <sup>3</sup>*J* = 9.0 Hz, <sup>4</sup>*J* = 2.1 Hz, each 2H, H-8 and H-18), 1.05 (s, 36H, H-28), 1.04 (s, 6H, H-27).

**<sup>13</sup>C NMR (151 MHz, [D<sub>1</sub>]-chloroform, [D<sub>4</sub>]-methanol, 298 K)**  $\delta$  = 146.95, 146.94 (each d, each <sup>2</sup>*J*<sub>C-P</sub> = 9.2 Hz, C-2 and C-12), 142.3 (C-4), 141.9 (C-14), 139.6 (C-29), 138.5 (C-21), 132.8 (C-31), 132.26 (C-23), 132.28, 132.19 (C-7, C-17), 131.05, 131.00 (C-10, C-20), 130.7 (C-5), 130.5 (C-15), 130.0 (C-30), 129.7 (C-22), 128.9, 128.8 (C-9, C-19), 127.75, 127.71 (C-8, C-18), 125.3, 125.2 (C-6, C-16), 123.5 (C-24), 122.5 (d, <sup>3</sup>*J*<sub>C-P</sub> = 2.0 Hz, C-13), 122.4 (d, <sup>3</sup>*J*<sub>C-P</sub> = 2.0 Hz, C-3), 121.7 (C-32), 121.0 (d, <sup>4</sup>*J*<sub>C-P</sub> = 1.7 Hz, C-11), 120.7 (d, <sup>4</sup>*J*<sub>C-P</sub> = 1.7 Hz, C-1), 106.5 (C-25), 92.0 (C-26), 81.5 (C-33), 74.9 (C-34), 18.5 (C-28), 11.3 (C-27).

**<sup>31</sup>P NMR (243 MHz, [D<sub>1</sub>]-chloroform, [D<sub>4</sub>]-methanol, 298 K)**  $\delta$  = 3.6 (*v*<sub>1/2</sub> = 19.5 Hz).

**COSY (600 MHz, [D<sub>1</sub>]-chloroform, [D<sub>4</sub>]-methanol, 298 K)**  $\delta$  (<sup>1</sup>H) /  $\delta$  (<sup>1</sup>H) = 7.65 / 7.48 (H-31 / H-30), 7.56 / 7.41 (H-23 / H-22), 7.48 / 7.65 (H-30 / H-31), 7.41 / 7.56 (H-22 / H-23), 7.30, 7.28 / 7.18, 7.17 (H-9, H-19 / H-8, H-18), 7.18, 7.17 / 7.30, 7.28 (H-8, H-18 / H-9, H-19).

**HSQC (600/151 MHz, [D<sub>1</sub>]-chloroform, [D<sub>4</sub>]-methanol, 298 K)**  $\delta$  (<sup>1</sup>H) /  $\delta$  (<sup>13</sup>C) = 7.81, 7.80 / 125.3, 125.2 (H-6, H-16 / C-6, C-16), 7.65 / 132.8 (H-31 / C-31), 7.56 / 132.26 (H-23 / C-23), 7.48 / 130.0 (H-30 / C-30), 7.43 / 122.5 (H-13 / C-13), 7.41 / 129.7 (H-22 / C-22), 7.40 / 122.4 (H-3 / C-3), 7.30, 7.28 / 128.9, 128.8 (H-9, H-19 / C-9, C-19), 7.18, 7.17 / 127.75, 127.71 (H-8, H-18 / C-8, C-18), 1.05 / 18.5 (H-28 / C-28), 1.04 / 11.3 (H-27 / C-27).

**HMBC (600/151 MHz, [D<sub>1</sub>]-chloroform, [D<sub>4</sub>]-methanol, 298 K)**  $\delta$  (<sup>1</sup>H) /  $\delta$  (<sup>13</sup>C) = 7.81, 7.80 / 142.3, 141.9, 132.28, 132.19, 131.05, 131.00, 127.75, 127.71 (H-6, H-16 / C-4, C-14, C-7, C-17, C-10, C-20, C-8, C-18), 7.65 / 139.6, 132.8, 81.5 (H-31 / C-29, C-31, C-33), 7.56 / 138.5, 132.26, 129.7, 106.5 (H-23 / C-21, C-23, C-22, C-25), 7.48 / 141.9, 130.0, 121.7 (H-30 / C-14, C-30, C-32), 7.43 / 146.95, 146.94, 139.6, 130.7, 121.0 (H-13 / C-2, C-12, C-29, C-15, C-11), 7.41 / 142.3, 132.26, 129.7, 123.5 (H-22 / C-4, C-23, C-22, C-24), 7.40 / 146.95, 146.94, 138.5, 130.5, 120.7 (H-3 / C-2, C-12, C-21, C-5, C-1), 7.30, 7.28 / 132.28, 132.19, 130.7, 130.5, 121.0, 120.7 (H-9, H-19 / C-7, C-17, C-15, C-5, C-11, C-1), 7.18, 7.17 / 131.05, 131.00, 125.3, 125.2 (H-8, H-18 / C-10, C-20, C-6, C-16), 1.05, 1.04 / 92.0, 18.5, 11.3 (H-28, H-27 / C-26, C-28, C-27).

**IR (ATR)  $\nu$  bar:** 2940, 2863, 2360, 2156, 1583, 1493, 1320, 1259, 1180, 1149 cm<sup>-1</sup>

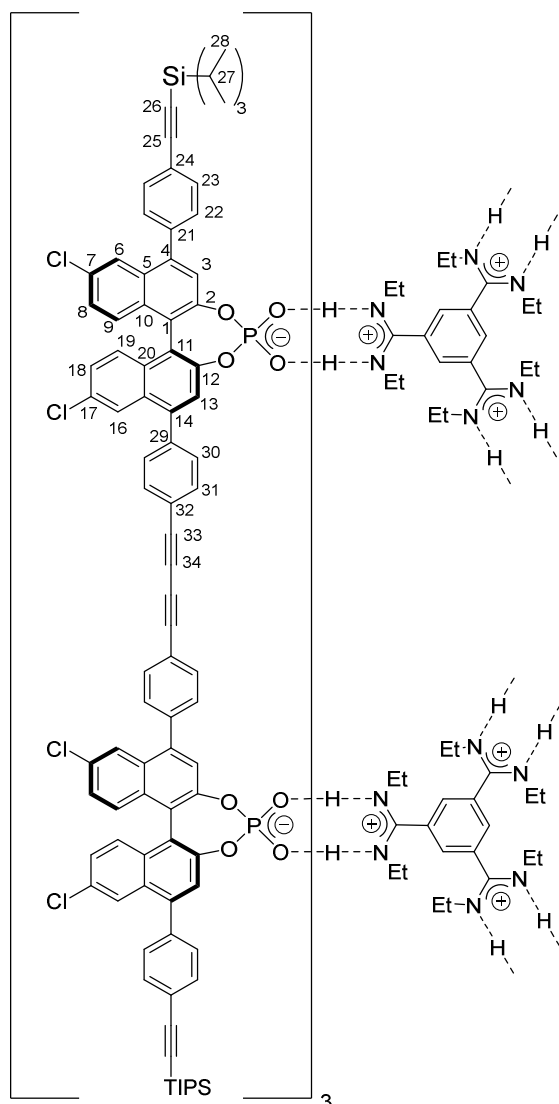
**MS (ESI, -Mode):** m/z = 770.1535 ([M-H]<sup>2-</sup>, calculated: m/z = 770.1581)

**Melting point:** 153-154°C

**Elemental analysis:** expected C = 69.94 %, H = 4.96 %, found C = 69.3 %, H = 5.33 %

## 2.5. Synthesis of the supramolecular capsules

### 2.5.1. Capsule (*all-R*)-1



The bisphosphoric acid (*R,R*)-**7** (9.27 mg, 6.00  $\mu\text{mol}$ , 3 eq.) was weighed into a vial. A stock solution of trisamidine **7** (500  $\mu\text{L}$  of an 8 mM solution in  $[\text{D}_1]$ -chloroform :  $[\text{D}_4]$ -methanol (8/2 v/v), containing 4.00  $\mu\text{mol}$ , 2 eq). The resulting solution (4 mM in capsule **1**) was analyzed by NMR.

$\text{C}_{312}\text{H}_{300}\text{Cl}_{12}\text{N}_{12}\text{O}_{24}\text{P}_6\text{Si}_6$  (5381.65 g/mol)

**$^1\text{H}$  NMR (600 MHz,  $[\text{D}_1]$ -chloroform,  $[\text{D}_4]$ -methanol, 298 K)**  $\delta$  = 7.78 (bs, 6H, H-35), 7.74, 7.72 (each d, each  $^4J$  = 1.8 Hz, each 6H, H-6 and H-16), 7.56 (ps d,  $^3J$  = 7.8 Hz, 12H, H-31), 7.49 (ps d,  $^3J$  = 7.8 Hz, 12H, H-23), 7.42 (ps d,  $^3J$  = 7.8 Hz, 12H, H-30), 7.33 (ps d,  $^3J$  = 7.8 Hz, 12H, H-22), 7.27 (s, 6H, H-13), 7.25 (s, 6H, H-3), 7.23, 7.21 (each d, each  $^3J$  = 9.0 Hz, each 6H, H-9, H-19), 7.09, 7.07 (each dd, each  $^3J$  = 9.0 Hz,  $^4J$  = 1.8 Hz, each 6H, H-8 and H-18), 3.13, 3.10 (each bs, each 12H, H-38, H-40), 1.05 (s, 126H, H-28, H-27), 0.95, 0.90 (each bs, each 18H, H-39, H-41).

**$^{13}\text{C}$  NMR (151 MHz,  $[\text{D}_1]$ -chloroform,  $[\text{D}_4]$ -methanol, 298 K)**  $\delta$  = 160.7 (C-37), 148.6 (d,  $^2J_{\text{C-P}}$  = 8.9 Hz, C-2, C-12 (isochronous signals)), 141.3 (C-4), 140.8 (C-14), 140.0 (C-29), 138.9 (C-21), 132.7 (C-31), 132.3 (C-23), 131.5, 131.4 (C-7, C-17), 131.2, 131.1 (C-10, C-20), 130.6 (C-35), 130.0, 129.8 (C-5, C-15), 130.0 (C-30), 129.6 (C-22), 128.8, 128.7 (C-9, C-19), 127.24, 127.20 (C-8, C-18), 125.0, 124.9 (C-6, C-16), 123.4 (C-13), 123.3 (C-3), 123.2 (C-24), 121.7 (C-11), 121.39 (C-1), 121.36 (C-32), 106.5 (C-25), 91.8 (C-26), 81.5 (C-33), 74.9 (C-34), 40.8, 38.2 (C-38, C-40), 18.5 (C-28), 14.9, 12.2 (C-39, C-41), 11.2 (C-27). C-36 was not observed.

**$^{31}\text{P}$  NMR (243 MHz,  $[\text{D}_1]$ -chloroform,  $[\text{D}_4]$ -methanol, 298 K)**  $\delta$  = 4.7 ( $\nu_{1/2}$  = 15 Hz).

**COSY (600 MHz,  $[\text{D}_1]$ -chloroform,  $[\text{D}_4]$ -methanol, 298 K)**  $\delta$  ( $^1\text{H}$ ) /  $\delta$  ( $^1\text{H}$ ) = 7.74, 7.72 / 7.09, 7.07 (H-6, H-16 / H-8, H-18), 7.56 / 7.42 (H-31 / H-30), 7.49 / 7.33 (H-23 / H-22), 7.42 / 7.56 (H-30 / H-31), 7.33 / 7.49 (H-22 / H-23), 7.23, 7.21 / 7.09, 7.07 (H-9, H-19 / H-8, H-18), 7.09, 7.07 / 7.74, 7.72, 7.23, 7.21 (H-8, H-18 / H-6, H-16, H-9, H-19), 3.13,

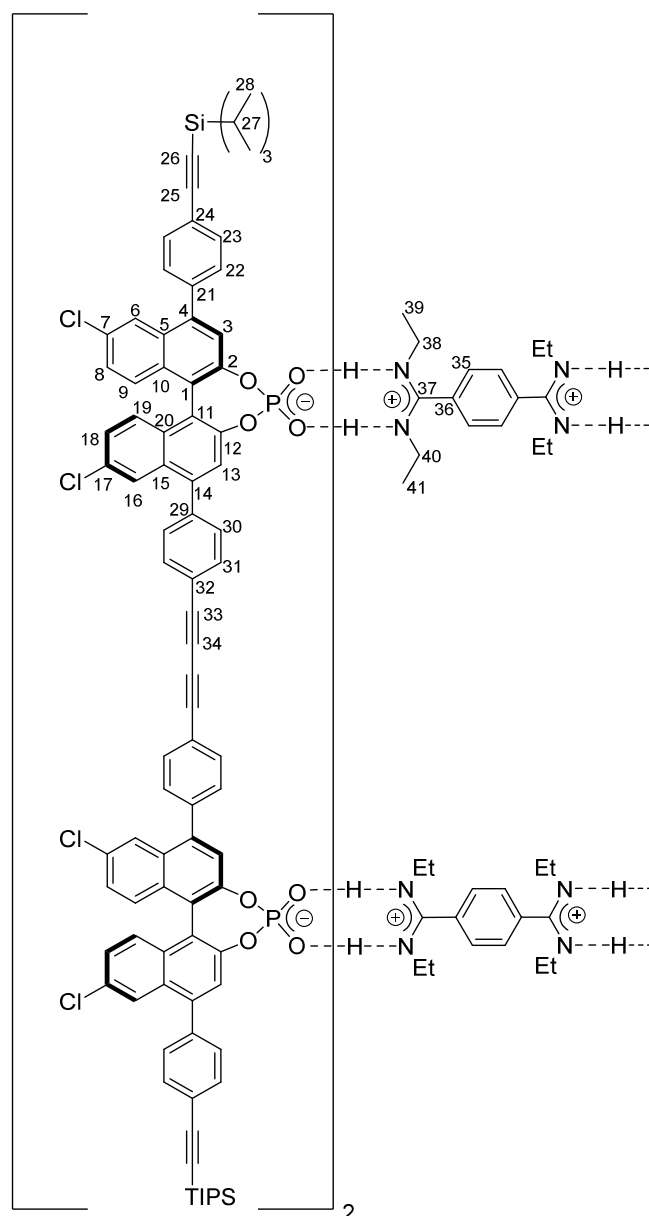
3.10 / 0.95, 0.90 (H-38, H-40 / H-39, H-41), 0.95, 0.90 / 3.13, 3.10 (H-39, H-41 / H-38, H-40).

**HSQC (600/151 MHz, [D<sub>1</sub>]-chloroform, [D<sub>4</sub>]-methanol, 298 K)**  $\delta$  (<sup>1</sup>H) /  $\delta$  (<sup>13</sup>C) = 7.78 / 130.6 (H-35 / C-35), 7.74, 7.72 / 125.0, 124.9 (H-6, H-16 / C-6, C-16), 7.56 / 132.7 (H-31 / C-31), 7.49 / 132.3 (H-23 / C-23), 7.42 / 130.0 (H-30 / C-30), 7.33 / 129.6 (H-22 / C-22), 7.27 / 123.4 (H-13 / C-13), 7.25 / 123.3 (H-3 / C-3), 7.23, 7.21 / 128.8, 128.7 (H-9, H-19 / C-9, C-19), 7.09, 7.07 / 127.24, 127.20 (H-8, H-18 / C-8, C-18), 3.13, 3.10 / 40.8, 38.2 (H-38, H-40 / C-38, C-40), 1.05 / 18.5, 11.2 (H-28, H-27 / C-28, C-27), 0.95, 0.90 / 14.9, 12.2 (H-39, H-41 / C-39, C-41).

**HMBC (600/151 MHz, [D<sub>1</sub>]-chloroform, [D<sub>4</sub>]-methanol, 298 K)**  $\delta$  (<sup>1</sup>H) /  $\delta$  (<sup>13</sup>C) = 7.74, 7.72 / 141.3, 140.8, 131.5, 131.4, 131.2, 131.1, 127.24, 127.20 (H-6, H-16 / C-4, C-14, C-7, C-17, C-10, C-20, C-8, C-18), 7.56 / 140.0, 132.7, 130.0, 81.5 (H-31 / C-29, C-31, C-30, C-33), 7.49 / 138.9, 132.3, 129.6, 106.5 (H-23 / C-21, C-23, C-22, C-25), 7.42 / 140.8, 132.7, 130.0, 121.36 (H-30 / C-14, C-31, C-30, C-32), 7.33 / 141.3, 132.3, 129.6, 123.2 (H-22 / C-4, C-23, C-22, C-24), 7.27 / 148.6, 140.0, 130.0, 129.8, 121.7 (H-13 / C-12, C-29, C-15, C-11), 7.25 / 148.6, 138.9, 130.0, 129.8, 121.39 (H-3 / C-2, C-21, C-5, C-1), 7.23, 7.21 / 131.5, 131.4, 130.0, 129.8, 121.7, 121.39 (H-9, H-19 / C-7, C-17, C-5, C-15, C-1, C-11), 7.09, 7.07 / 131.2, 131.1, 125.0, 124.9 (H-8, H-18 / C-10, C-20, C-6, C-16), 1.05 / 91.8, 18.5, 11.2 (H-28, H-27 / C-26, C-28, C-27).

**NOESY (600 MHz, [D<sub>1</sub>]-chloroform, [D<sub>4</sub>]-methanol, 298 K)**  $\delta$  (<sup>1</sup>H) /  $\delta$  (<sup>1</sup>H) = 7.78 / 3.13, 3.10, 0.95, 0.90 (H-35 / H-38, H-40, H-39, H-41), 7.74, 7.72 / 7.42, 7.33 (H-6, H-16 / H-30, H-22), 7.56 / 7.42, 0.95, 0.90 (H-31 / H-30, H-39, H-41), 7.49 / 7.33, 0.95, 0.90 (H-23 / H-22, H-39, H-41), 7.42 / 7.74, 7.72, 7.56, 7.27, 7.25, 0.95, 0.90 (H-30 / H-16, H-31, H-13, H-39, H-41), 7.33 / 7.74, 7.72, 7.49, 0.95, 0.90 (H-22 / H-6, H-23, H-39, H-41), 7.27, 7.25 / 7.42, 3.13, 3.10, 0.95, 0.90 (H-3, H-13 / H-30, H-38, H-40, H-39, H-41), 7.23, 7.21 / 7.09, 7.07 (H-9, H-19 / H-8, H-18), 7.09, 7.07 / 7.23, 7.21 (H-8, H-18 / H-9, H-19), 3.13, 3.10 / 7.78, 7.27, 7.25, 0.95, 0.90 (H-38, H-40 / H-35, H-3, H-13, H-39, H-41), 0.95, 0.90 / 7.56, 7.49, 7.42, 7.33, 7.27, 7.25, 3.13, 3.10 (H-39, H-41 / H-31, H-23, H-30, H-22, H-13, H-3, H-38, H-40).

## 2.5.2. Capsule (*all-R*)-2



The bisphosphoric acid (*R,R*)-**6** (2.16 mg, 1.40  $\mu\text{mol}$ , 2 eq.) was weighed into a vial. A stock solution of bisamidine **8** (175  $\mu\text{L}$  of an 8 mM solution in  $[\text{D}_1]$ -chloroform :  $[\text{D}_4]$ -methanol (8/2 v/v), containing 1.40  $\mu\text{mol}$ , 2 eq.). The resulting solution (4 mM in capsule **2**) was analyzed by NMR.

$\text{C}_{212}\text{H}_{204}\text{Cl}_8\text{N}_8\text{O}_{16}\text{P}_4\text{Si}_4$  (3639.84 g/mol)

**$^1\text{H}$  NMR (600 MHz,  $[\text{D}_1]$ -chloroform, 298 K)**  $\delta$  = 7.70, 7.69 (each d, each  $^4J$  = 1.8 Hz, each 4H, H-6 and H-16), 7.55 (ps d,  $^3J$  = 7.8 Hz, 8H, H-31), 7.47 (ps d,  $^3J$  = 7.8 Hz, 8H, H-23), 7.44 (s, 8H, H-35), 7.41 (ps d,  $^3J$  = 7.8 Hz, 8H, H-30), 7.38 (ps d,  $^3J$  = 7.8 Hz, 8H, H-22), 7.30, 7.28 (each s, each 4H, H-3 and H-13), 7.22, 7.20 (each d, each  $^3J$  = 9.0 Hz, each 4H, H-9, H-19), 7.05, 7.04 (each dd, each  $^3J$  = 9.0 Hz,  $^4J$  = 1.8 Hz, each 4H, H-8 and H-18), 3.15, 3.02 (each q, each  $^3J$  = 7.5 Hz, each 8H, H-38, H-40), 0.98 (s, 84H, H-28, H-27), 0.95, 0.94 (each t, each  $^3J$  = 7.5 Hz, each 12H, H-39, H-41).

**$^{13}\text{C}$  NMR (151 MHz,  $[\text{D}_1]$ -chloroform,  $[\text{D}_4]$ -methanol, 298 K)**  $\delta$  = 162.3 (C-37), 148.6 (d,  $^2J_{\text{C-P}}$  = 9.1 Hz, C-2, C-12 (isochronous signals)), 141.3 (C-4), 140.9 (C-14), 140.0 (C-29), 138.9 (C-21), 132.6 (C-31), 132.1 (C-23), 131.39, 131.35 (C-7, C-17), 131.29 (C-36), 131.13, 131.08 (C-10, C-20), 129.96, 129.8 (C-5, C-15), 129.93 (C-30), 129.6 (C-22), 128.8, 128.6 (C-9, C-19), 128.7 (C-35), 127.11, 127.07 (C-8, C-18), 125.0, 124.8 (C-6, C-16), 123.5, 123.3 (C-3, C-13), 123.1 (C-24), 121.6, 121.35 (C-1, C-11), 121.27 (C-32), 106.5 (C-25), 91.6 (C-26), 81.4 (C-33), 74.7 (C-34), 40.6, 38.0 (C-38, C-40), 18.4 (C-28), 15.0, 12.1 (C-39, C-41), 11.2 (C-27).

**$^{31}\text{P}$  NMR (243 MHz,  $[\text{D}_1]$ -chloroform,  $[\text{D}_4]$ -methanol, 298 K)**  $\delta$  = 5.4 ( $\nu_{1/2}$  = 99 Hz).

**COSY (600 MHz,  $[\text{D}_1]$ -chloroform,  $[\text{D}_4]$ -methanol, 298 K)**  $\delta$  ( $^1\text{H}$ ) /  $\delta$  ( $^1\text{H}$ ) = 7.70, 7.69 / 7.05, 7.04 (H-6, H-16 / H-8, H-18), 7.47 / 7.38 (H-23 / H-22), 7.38 / 7.47 (H-22 / H-23),

7.22, 7.20 / 7.05, 7.04 (H-9, H-19 / H-8, H-18), 7.05, 7.04 / 7.70, 7.69, 7.22, 7.20 (H-8, H-18 / H-6, H-16, H-9, H-19).

**HSQC (600/151 MHz, [D<sub>1</sub>]-chloroform, 298 K)**  $\delta$  (<sup>1</sup>H) /  $\delta$  (<sup>13</sup>C) = 7.70, 7.69 / 125.0, 124.8 (H-6, H-16 / C-6, C-16), 7.55 / 132.6 (H-31 / C-31), 7.47 / 132.1 (H-23 / C-23), 7.44 / 128.7 (H-35 / C-35), 7.41 / 129.93 (H-30 / C-30), 7.38 / 129.6 (H-22 / C-22), 7.30, 7.28 / 123.5, 123.3 (H-3, H-13 / C-3, C-13), 7.22, 7.20 / 128.8, 128.6 (H-9, H-19 / C-9, C-19), 7.05, 7.04 / 127.11, 127.07 (H-8, H-18 / C-8, C-18), 3.15, 3.02 / 40.6, 38.0 (H-38, H-40 / C-38, C-40), 0.98 / 18.4, 11.2 (H-28, H-27 / C-28, C-27), 0.95, 0.94 / 15.0, 12.1 (H-39, H-41 / C-39, C-41).

**HMBC (600/151 MHz, [D<sub>1</sub>]-chloroform, [D<sub>4</sub>]-methanol, 298 K)**  $\delta$  (<sup>1</sup>H) /  $\delta$  (<sup>13</sup>C) = 7.70, 7.69 / 141.3, 140.9, 131.39, 131.35, 131.13, 131.08, 127.11, 127.07 (H-6, H-16 / C-4, C-14, C-7, C-17, C-10, C-20, C-8, C-18), 7.55 / 140.0, 132.6, 81.4 (H-31 / C-29, C-31, C-33), 7.47 / 138.9, 132.1, 106.5 (H-23 / C-21, C-23, C-25), 7.44 / 131.29 (H-35 / C-36), 7.41 / 129.93, 121.27 (H-30 / C-30, C-32), 7.38 / 129.6, 123.1 (H-22 / C-22, C-24), 7.30, 7.28 / 129.96, 129.8 (H-3, H-13 / C-5, C-15), 7.22, 7.20 / 131.39, 131.35, 129.96, 129.8, 121.6, 121.35 (H-9, H-19 / C-7, C-17, C-5, C-15, C-1, C-11), 7.05, 7.04 / 131.39, 131.35, 131.13, 131.08, 125.0, 124.8 (H-8, H-18 / C-7, C-17, C-10, C-20, C-6, C-16), 0.98 / 18.4, 11.2 (H-28, H-27 / C-28, C-27), 0.95, 0.94 / 40.6, 38.0 (H-39, H-41 / C-38, C-40).

**NOESY(600 MHz, [D<sub>1</sub>]-chloroform, [D<sub>4</sub>]-methanol, 298 K)**  $\delta$  (<sup>1</sup>H) /  $\delta$  (<sup>1</sup>H) = 7.70, 7.69 / 7.41, 7.38 (H-6, H-16 / H-30, H-22), 7.55 / 7.41 (H-31 / H-30), 7.47 / 7.38, 0.95, 0.94 (H-23 / H-22, H-39, H-41), 7.41 / 7.55, 7.70, 7.69 (H-30 / H-31, H-16), 7.38 / 7.47, 7.70, 7.69 (H-22 / H-23, H-6), 7.22, 7.20 / 7.05, 7.04 (H-9, H-19 / H-8, H-18), 7.05, 7.04 / 7.22, 7.20 (H-8, H-18 / H-9, H-19), 0.95, 0.94 / 7.47 (H-39, H-41 / H-23).



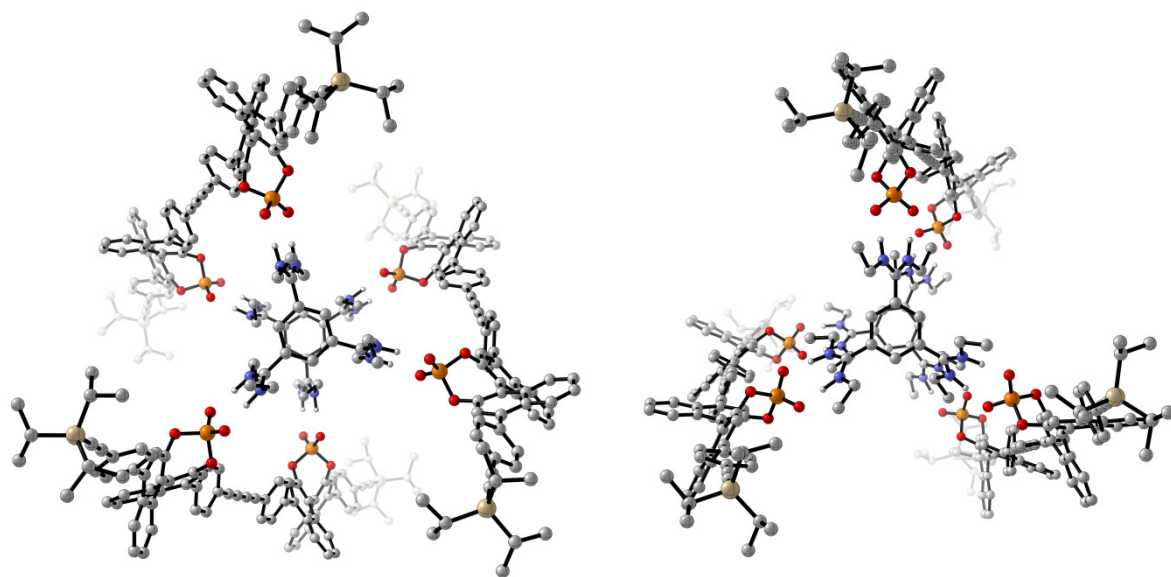
### 3. Characterization of the capsules

#### 3.1. Structure calculations

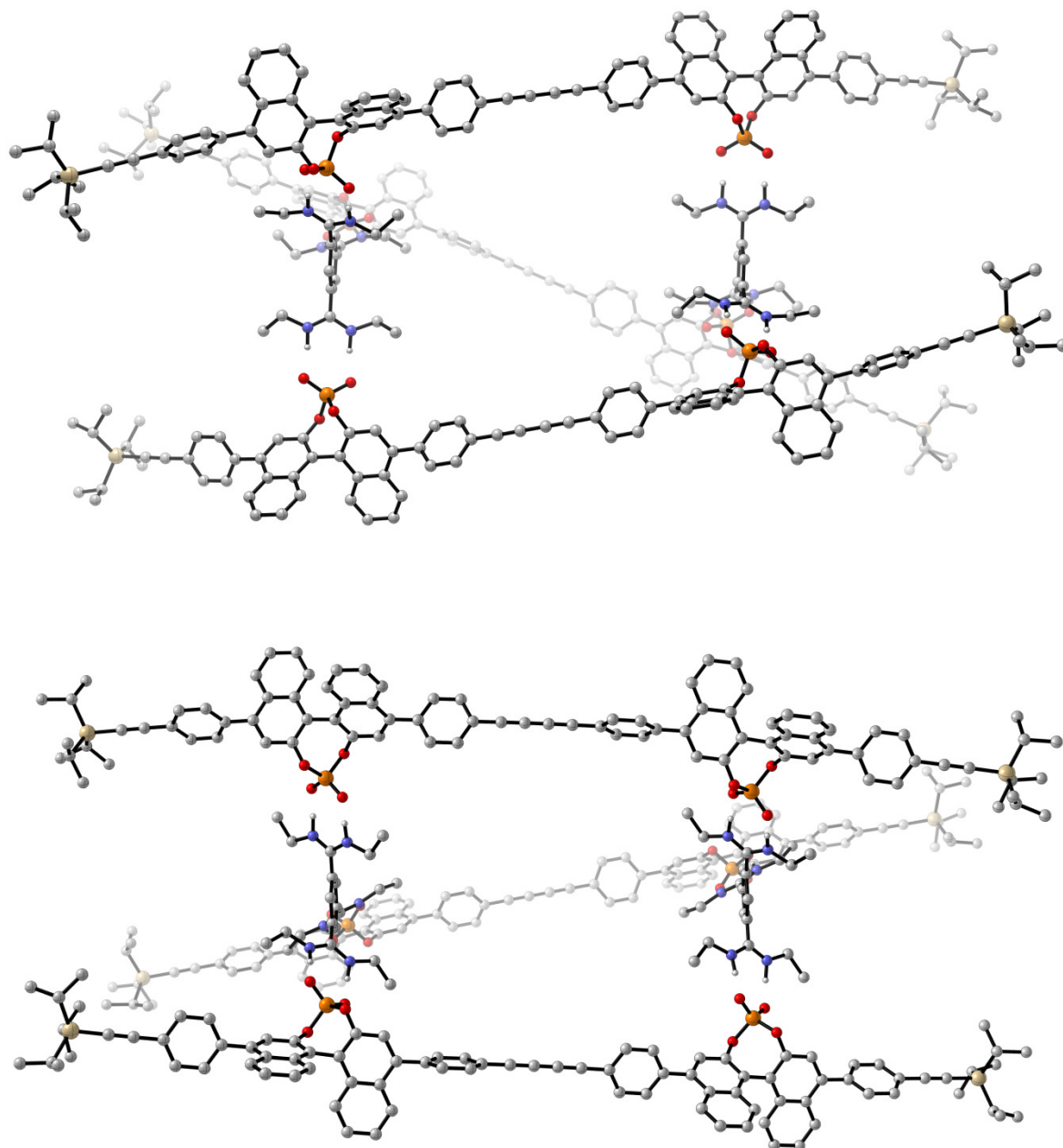
C<sub>3</sub>- and C<sub>2</sub>-symmetric input structures of the (*all-R*)-**1** and (*all-R*)-**2** were generated and pre-optimized on force field level (Schrödinger MacroModel 10.1, OPLS 2005 force field, chloroform solvent) to remove any large structural constraints. Afterwards, two FF-optimized structures of (*all-R*)-**1** were obtained, which represent a right- and a left-handed helical assembly (cf. Figures S2 and S3). The left-handed helical form was found to be about 3 kcal/mol more stable than the right-handed structure. For (*all-R*)-**2**, a single almost planar and rectangular structure was obtained (Fig. S4).

In order to further refine the structures, the FF-optimized geometries were subject to density functional theory (DFT) calculations at the B3LYP/6-31G(d,p)/IEFPCM(chloroform) level of theory.<sup>[4]</sup> During the course of the DFT-based geometry optimization, the almost planar structure of (*all-R*)-**2** changed to a notably twisted structure with the  $\equiv\text{C}-\underline{\text{C}}-\underline{\text{C}}-\text{C}\equiv$  bonds on the opposing sides featuring a tilt angle of  $\sim 39^\circ$ . Interestingly, the left-handed helical structure of (*all-R*)-**1** was found to be unfavorable on DFT level. In fact, it converted to the right-handed structure in any of the attempts to optimize it. This is particularly important to note, as DFT therefore suggests the former higher-energy conformer to actually be the only structure that is adopted by (*all-R*)-**1**. It moreover suggests that there might be structural or steric effects that are not accurately captured by the force field, which render the left-handed structure energetically so unfavorable that it is no longer a minimum on the potential energy surface. As the structural change occurred within the first ten steps of the geometry optimization, this observation leads to the suggestion that force-field optimized structures should always be further refined on low DFT level in order to confirm the stability of the structure and to exclude large destabilizing effects that the force field may not capture accurately.

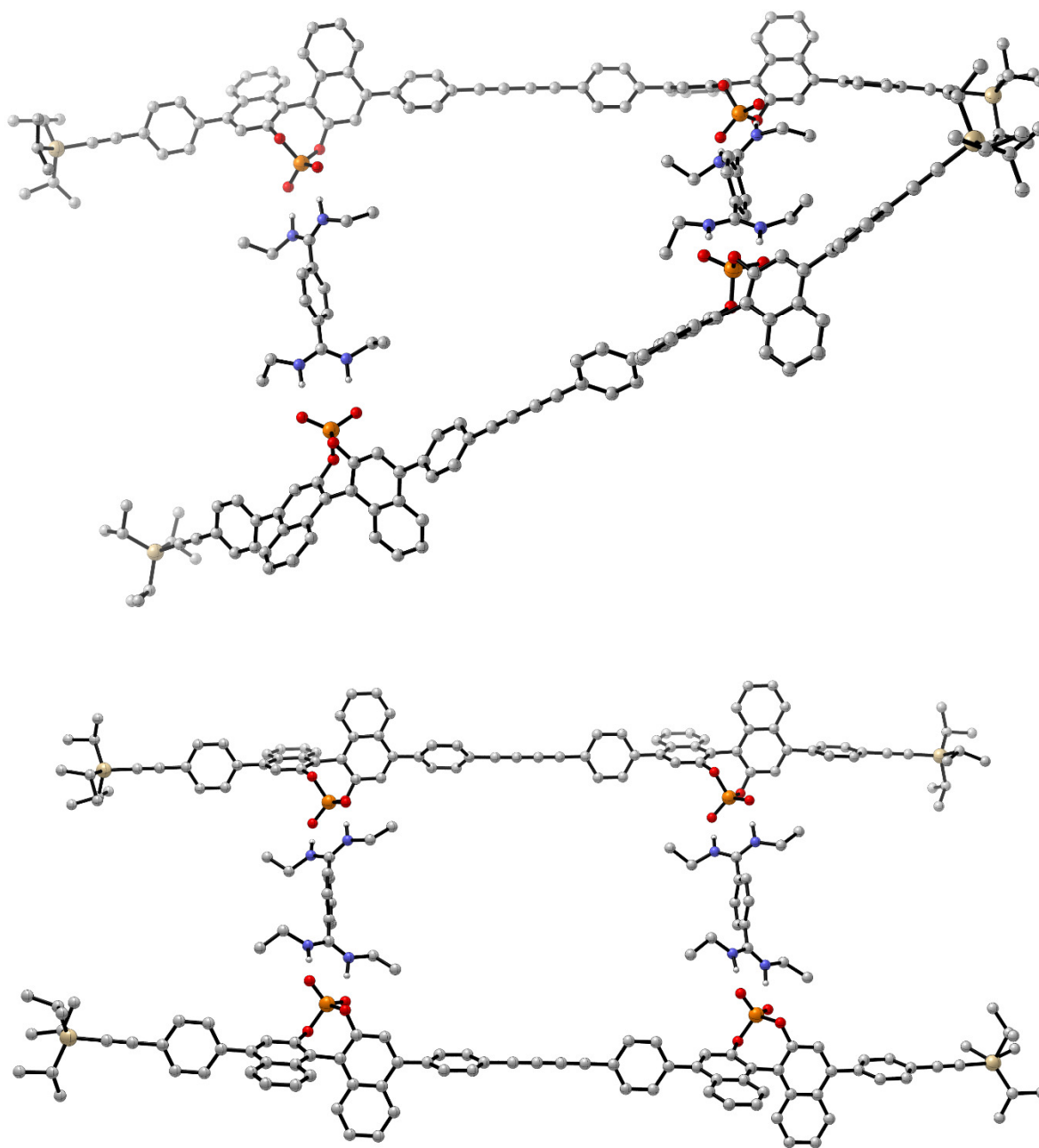
Cartesian coordinates and figures of the structures are provided below. It should be noted that the final DFT structure of (*all-R*)-**1** was not optimized to full convergence of the electronic structure, but geometrical changes between subsequent optimization steps were very small and only involving the exact position of the TIPS groups.



**Figure S 2:** View along the C3-axis of the DFT-optimized (*all-R*)-**1** in its right-handed helical structure (left) and of the OPLS-force field optimized left-handed structure (right) which is not stable on DFT level.



**Figure S 3:** Side-view of (*all-R*)-**1** in its right-handed helical structure obtained from DFT calculations (top) and in its left-handed structure obtained on force field level (bottom).



**Figure S 4:** DFT-optimized (top) and force-field optimized (bottom) structure of (*all-R*)-2

















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H	-1.43686500	-13.06477500	-2.69683800	H	13.88060000	-0.98700000	2.11240000
H	-2.23128300	-13.13190000	-4.27016500	H	13.04150000	-2.45200000	1.66180000
H	-1.77444700	-8.36707200	-1.34236200	H	8.31160000	-0.09510000	2.49480000
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H	3.67902200	-13.04768500	-2.58466000	H	12.92030000	-0.08390000	-3.30410000
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H	-0.36385700	-7.83451700	2.53071400	H	7.55380000	4.42790000	-2.31940000
H	1.10301700	-7.72912700	3.52257100	H	8.39400000	3.95340000	-3.77620000

<i>(all-R)</i> -2, DFT optimized twisted structure				<i>(all-R)</i> -2, OPLS-optimized flat structure			
C	10.60850000	2.02480000	0.55940000	C	11.04695900	1.42560200	0.12211500
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H	-10.71090000	1.13170000	-2.73000000	H	-10.99280400	1.10212500	-2.23792300
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N	-9.24700000	-3.85450000	-2.53070000	N	-10.09781100	-3.57965100	-0.24539900
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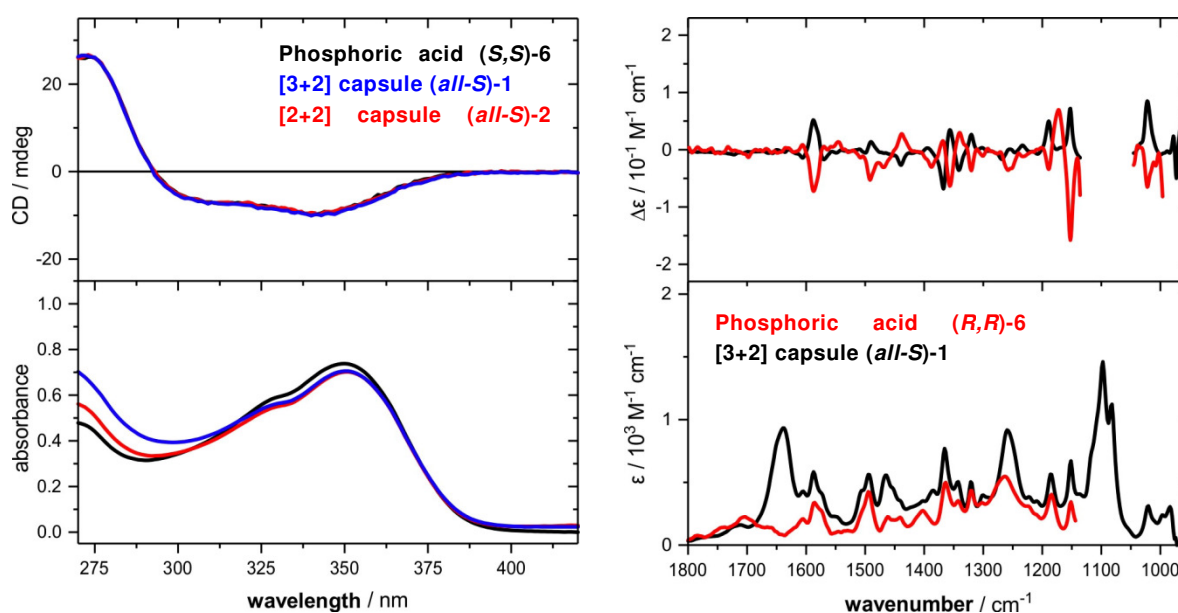




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H	23.73740000	11.10010000	-2.71940000	H	24.13833900	10.74734700	4.08712900
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C	-6.98120000	-3.89470000	-1.54060000	C	-7.70326200	-4.08212900	-0.61634500
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H	7.90020000	-2.97320000	0.45040000	H	7.48105700	-4.31148000	-0.02661900
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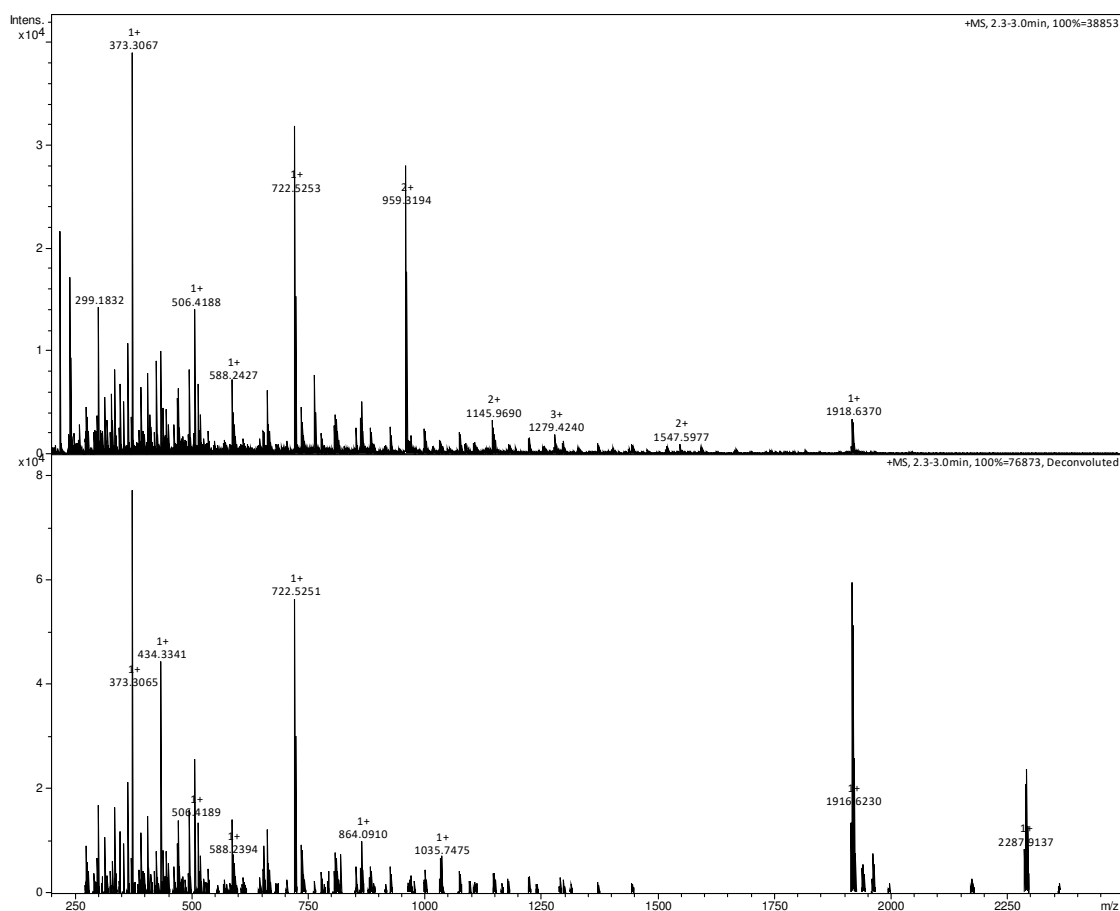
### 3.2. CD- and VCD-measurements

In order to further characterize the structures of **1** and **2**, we carried out ECD and VCD measurements. The results of these experiments are summarized in Figure S5. In comparison to the free bisphosphoric acid **6**, neither ECD or VCD show any additional spectral signatures. While one may argue that the UV chromophore of the phosphoric acid must dominate the UV/ECD spectra and thus overlap with any small contributions of the amidines, the lack of any additional VCD signatures is unexpected. In particular the strong amidine band at around  $1650\text{ cm}^{-1}$  is a nicely isolated chromophore which we expected to feature a VCD signature due to the chiral twist or tilt of the trisamidine core. Due to the size of the capsules and the lack of additional spectral signatures, we have not attempted any VCD spectra calculations. It is, however, possible that the structural flexibility that was indicated by the geometry optimizations and the resulting conformational changes between the right- and left-handed forms of **1** may lead to a cancelation of the amidine VCD signatures.

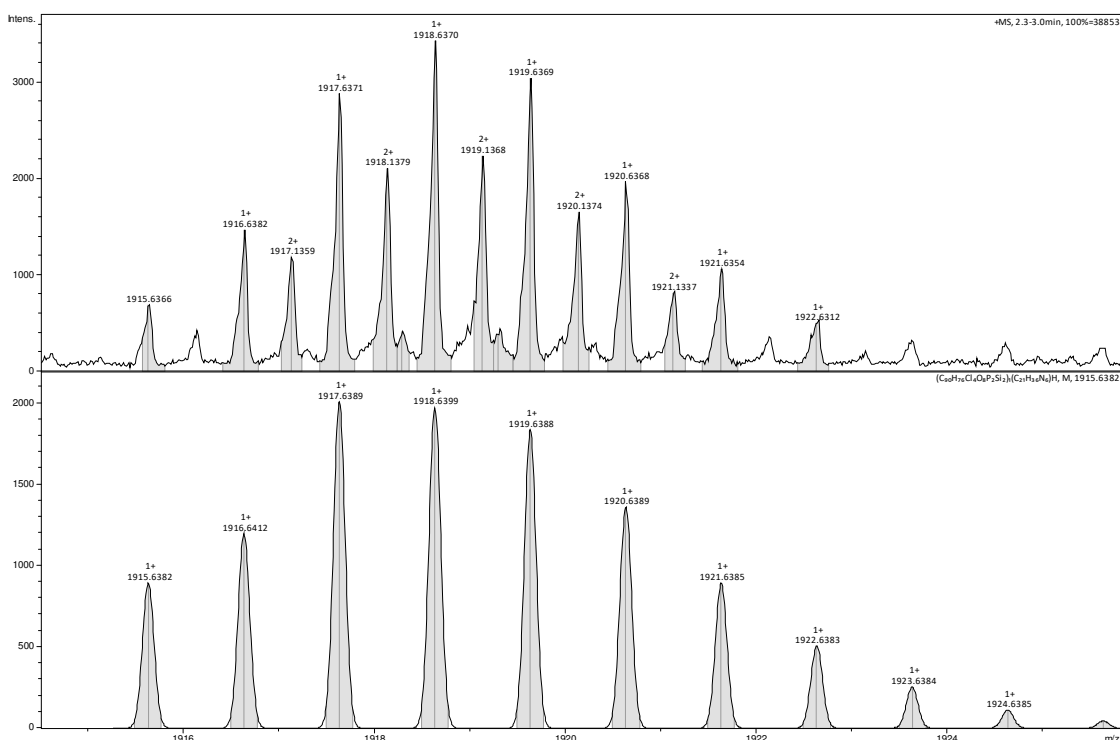


**Figure S 5:** Experimental UV/ECD spectra of phosphoric acid (*R,R*)-**6** and capsules (*all-R*)-**1/2** (left, each  $100\ \mu\text{M}$  in  $[\text{D}_1]$ -chloroform/ $[\text{D}_4]$ -methanol = 8/2) and IR/VCD spectra (right) of phosphoric acid (*S,S*)-**6** and capsule (*all-R*)-**1** ( $30\ \text{mM}$ , chloroform/ $[\text{D}_4]$ -methanol = 8/2,  $100\ \mu\text{m}$  path length).

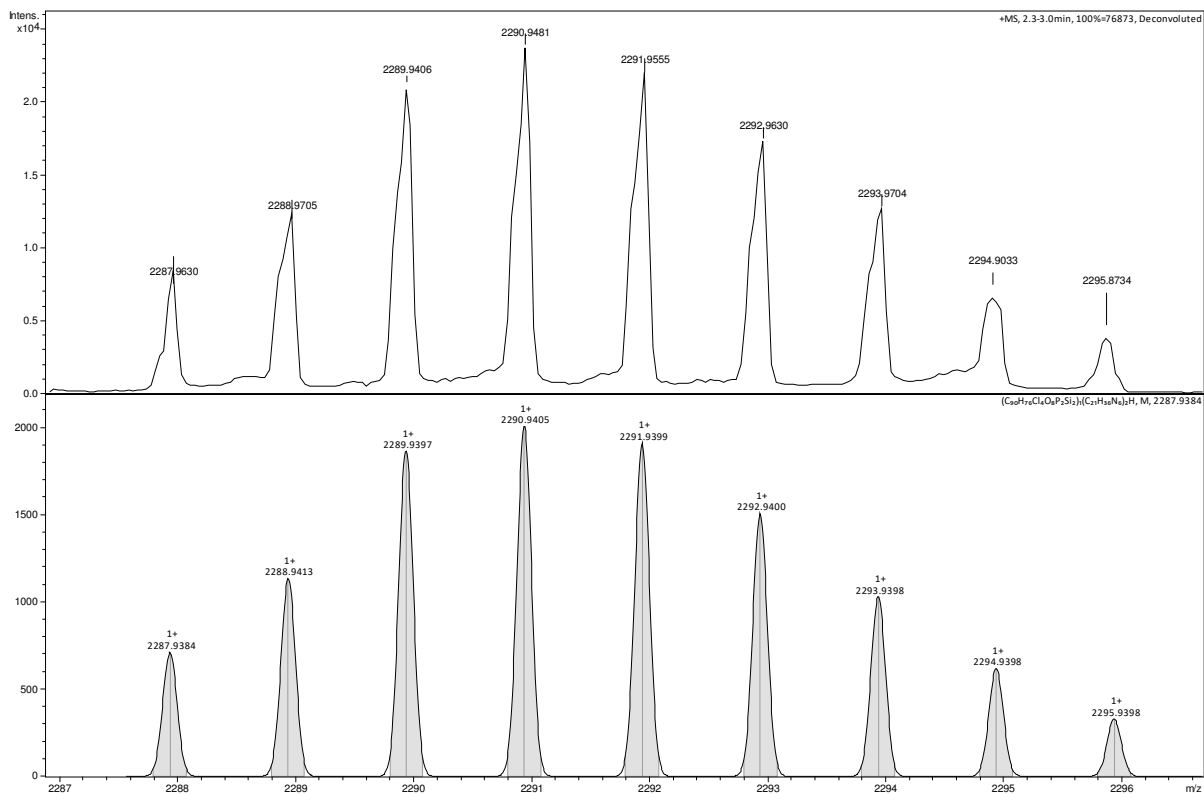
### 3.3. MS-Measurements



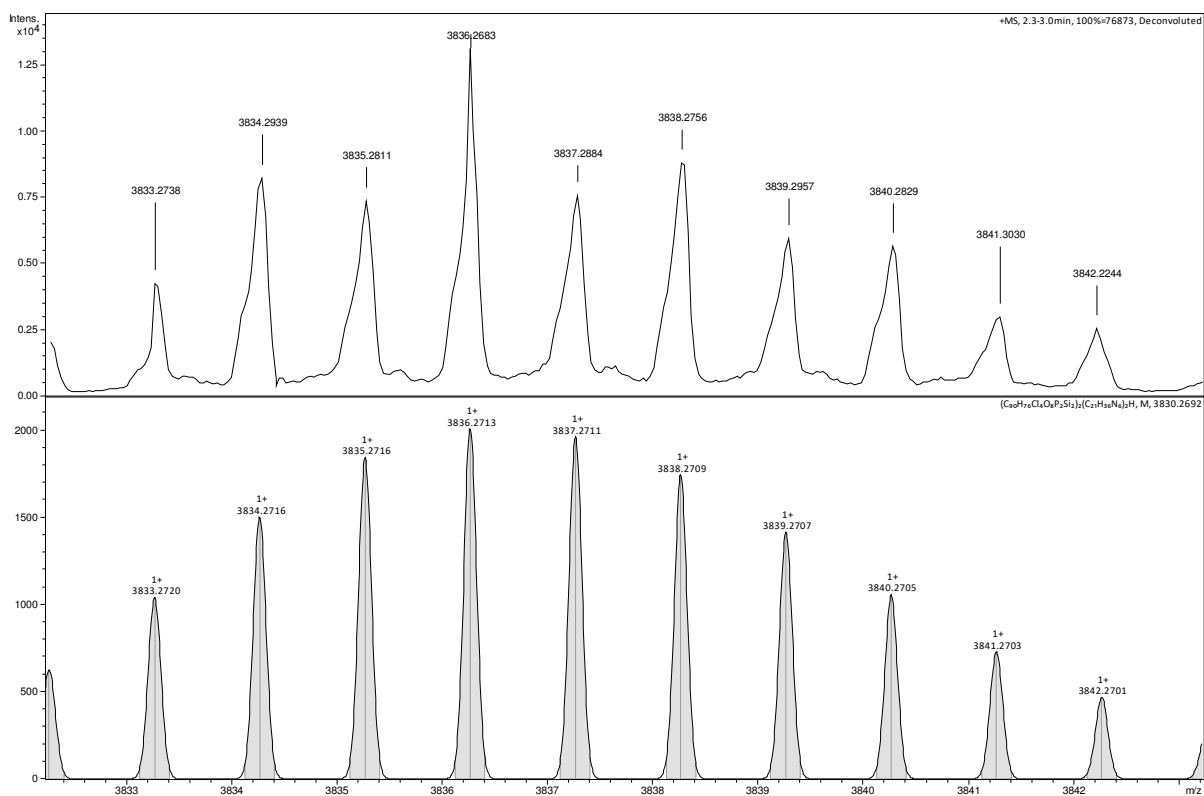
**Figure S 6:** ESI-MS spectrum of the [3+2]-capsule (*all-R*)-1 (full range, top: as recorded, bottom: deconvoluted).



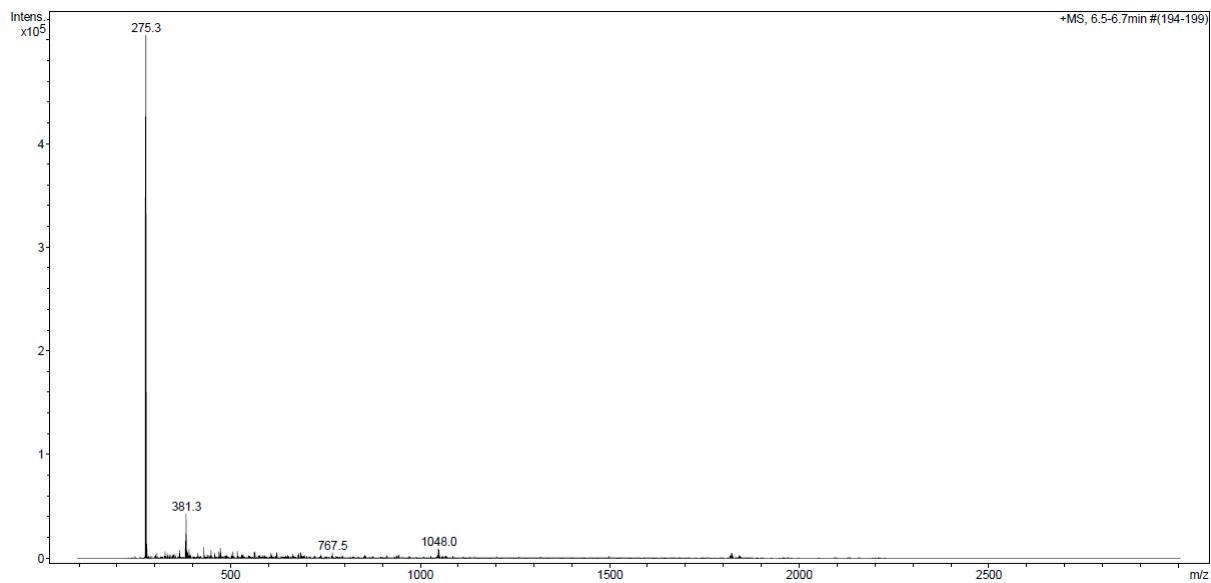
**Figure S 7:** ESI-MS spectrum of the [3+2]-capsule (*all-R*)-1, showing the aggregates  $[6+7+H]^+$  and  $[(6)_2+(7)_2+2H]^{2+}$  (top: measured spectrum, bottom: simulated spectrum of  $[6+7+H]^+$ ).



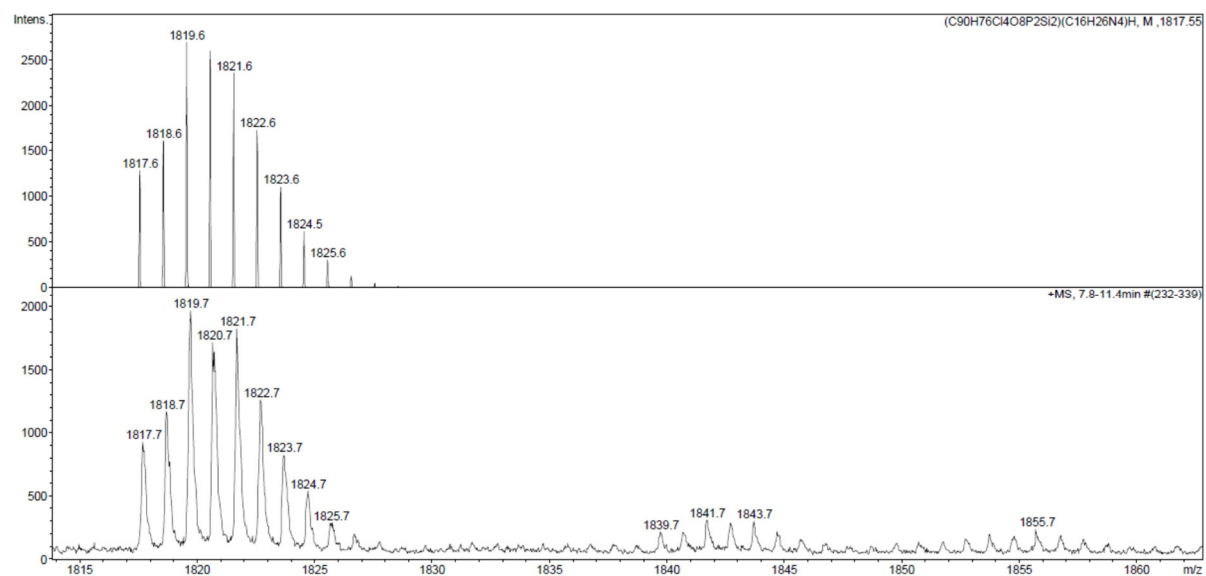
**Figure S 8:** ESI-MS spectrum of the [3+2]-capsule (*all-R*)-1, showing the aggregate  $[6+(7)_2+H]^+$  (top: measured spectrum (deconvoluted), bottom: simulated spectrum).



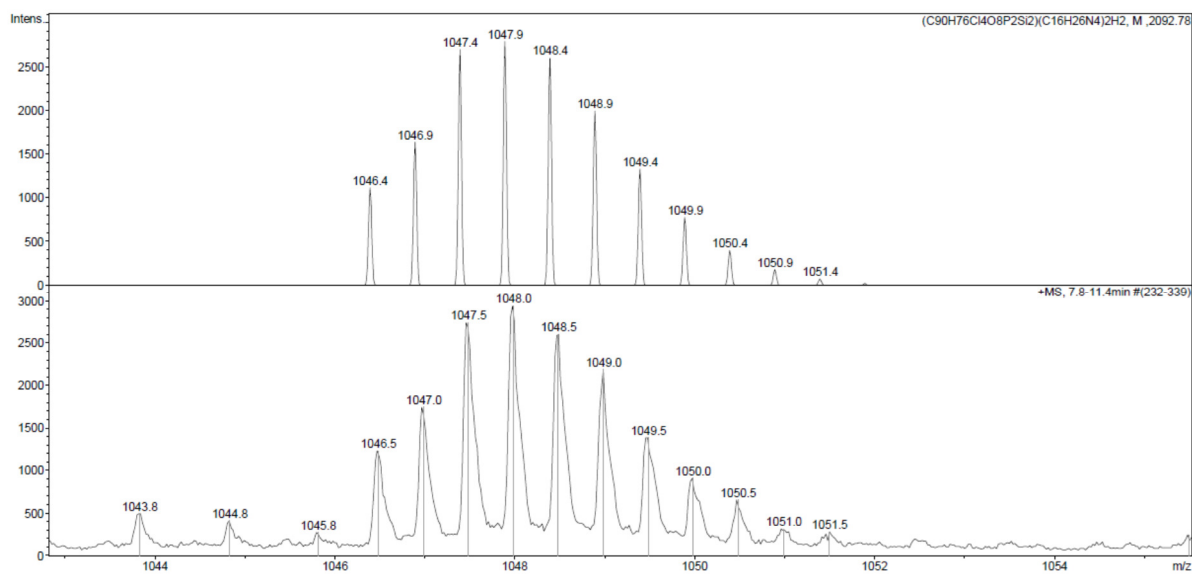
**Figure S 9:** ESI-MS spectrum of the [3+2]-capsule (*all-R*)-1, showing the aggregate  $[(6)_2+(7)_2+H]^+$  (top: measured spectrum (deconvoluted), bottom: simulated spectrum).



**Figure S 10:** ESI-MS spectrum of the [2+2]-capsule (*all-R*)-2 (full range).

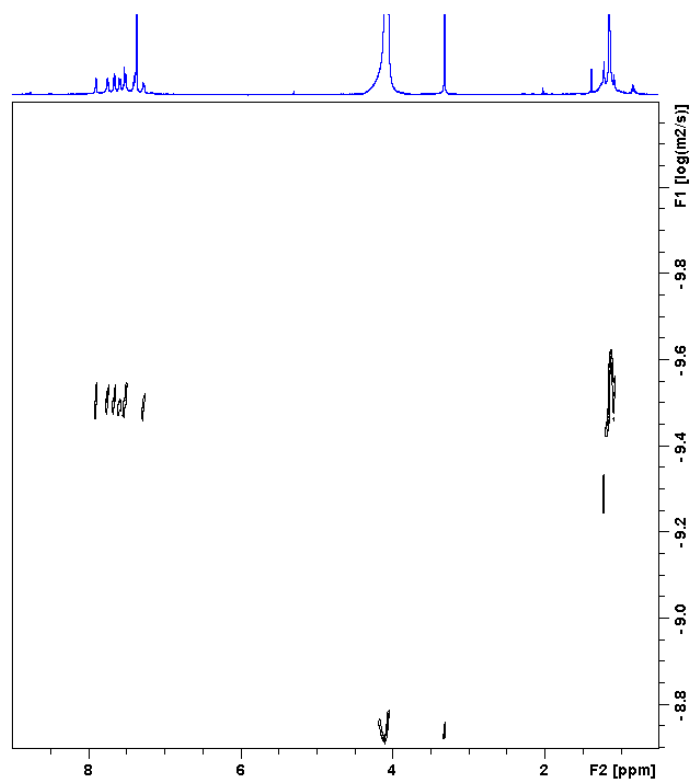


**Figure S 11:** ESI-MS spectrum of the [2+2]-capsule (*all-R*)-2, showing the aggregates  $[6+8+H]^+$  and  $[6+8+Na]^+$  (top: simulated spectrum, bottom: measured spectrum).

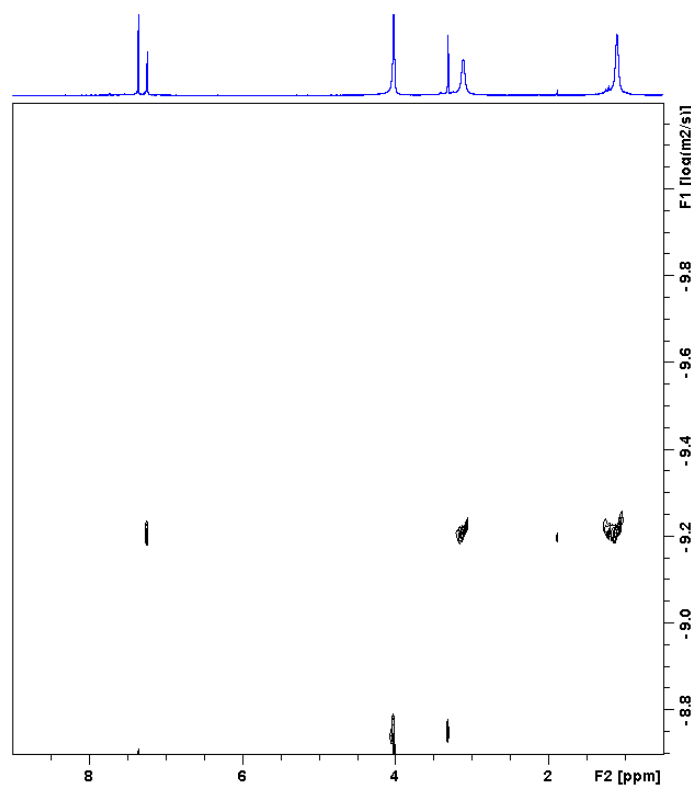


**Figure S 12:** ESI-MS spectrum of the [2+2]-capsule (*all-R*)-2, showing the aggregate  $[6+(8)_2+2H]^{2+}$  (top: simulated spectrum, bottom: measured spectrum).

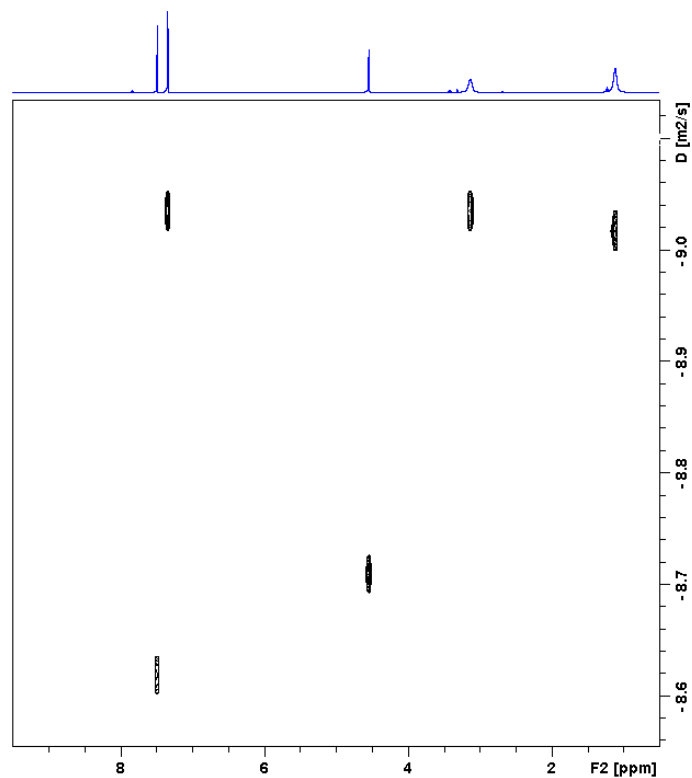
### 3.4. DOSY-Measurements



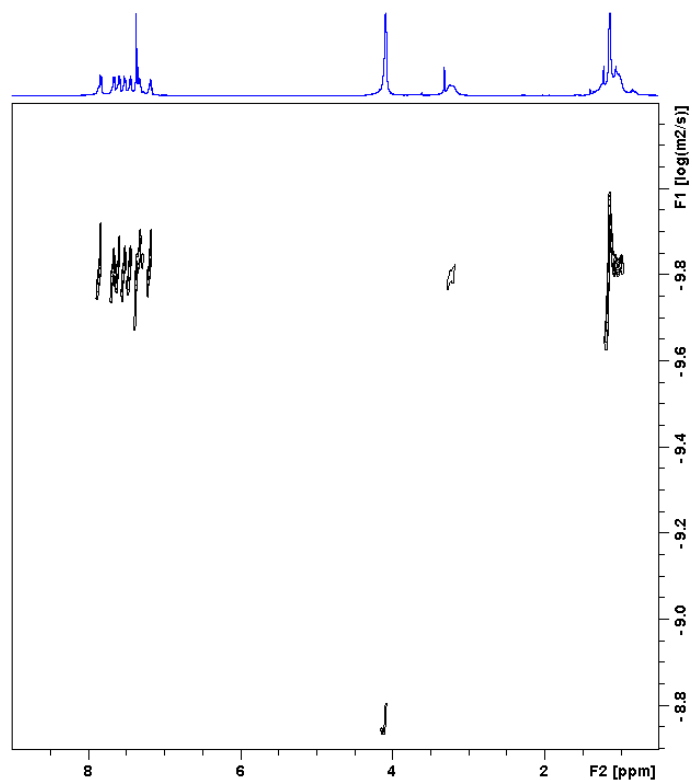
**Figure S 13:** DOSY-NMR spectrum of bisphosphoric acid (*R,R*)-**6** (12 mM) (500 MHz, [D<sub>1</sub>]-chloroform/[D<sub>4</sub>]-methanol = 8/2 (v/v), 298 K).



**Figure S 14:** DOSY-NMR spectrum of trisamidine **7** (500 MHz, [D<sub>1</sub>]-chloroform/[D<sub>4</sub>]-methanol = 8/2 (v/v), 298 K).

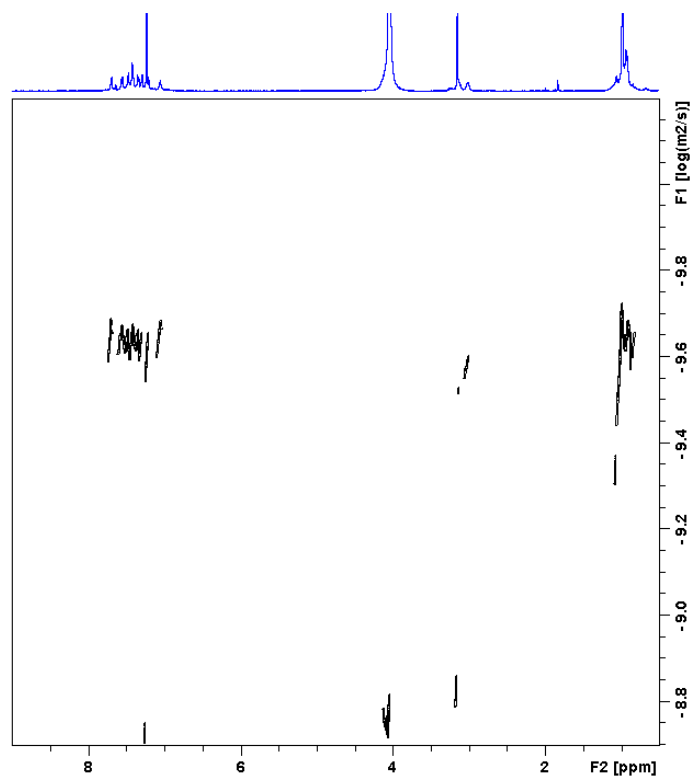


**Figure S 15:** DOSY-NMR spectrum of bisamidine **8** (500 MHz, [D<sub>1</sub>]-chloroform/[D<sub>4</sub>]-methanol = 8/2 (v/v), 298 K).



**Figure S 16:** DOSY-NMR spectrum of [3+2]-capsule (*all-R*)-**1** (4 mM) (500 MHz, [D<sub>1</sub>]-chloroform/[D<sub>4</sub>]-methanol = 8/2 (v/v), 298 K).





**Figure S 17:** DOSY-NMR spectrum of [2+2]-capsule (*all-R*)-**2** (4 mM) (500 MHz, [D<sub>1</sub>]-chloroform/[D<sub>4</sub>]-methanol = 8/2 (v/v), 298 K).

**Table S 1:** Diffusion coefficients as determined per DOSY-NMR and calculated molecular diffusion coefficients. All measurements were performed in [D<sub>1</sub>]-chloroform/[D<sub>4</sub>]-methanol (8/2 v/v) at 298 K. Theoretical values were calculated based on the DFT-optimized structures using the Stokes-Einstein equation using the approximation for cylindrical particles.

<b>Trisamidine 7</b>									
Integral region (ppm)	7.2	3.2	1.1				Mean value	Calculated value	
D	6.10	5.99	5.97				<b>6.02</b>	-	
SD (%)	0.23	0.23	0.21				<b>0.22</b>	-	

<b>Bisamidine 8</b>									
Integral region (ppm)	7.3	3.2	1.1				Mean value	Calculated value	
D	8.81	8.79	8.90				<b>8.83</b>	-	
SD (%)	0.02	0.01	0.01				<b>0.01</b>	-	

<b>Bisphosphoric acid (<i>R,R</i>)-6</b>									
Integral region (ppm)	7.9	7.8	7.7	7.6	7.5	7.3	1.1	Mean value	Calculated value
D	3.17	3.18	3.17	3.28	3.21	3.26	3.19	<b>3.21</b>	-
SD (%)	0.05	0.05	0.05	0.07	0.06	0.06	0.06	<b>0.06</b>	-

<b>[3+2]-capsule (<i>all-R</i>)-1</b>										
Integral region (ppm)	7.9	7.7	7.6	7.5	7.4	7.2	3.2	1.1	Mean value	Calculated value
D	1.57	1.59	1.55	1.58	1.56	1.54	1.55	1.61	<b>1.57</b>	<b>1.58</b>
SD (%)	0.07	0.09	0.07	0.09	0.11	0.07	0.07	0.10	<b>0.08</b>	-

<b>[2+2]-capsule (<i>all-R</i>)-2</b>									
Integral region (ppm)	7.7	7.6	7.5	7.4	7.3	3.2	1.1	Mean value	Calculated value
D	2.30	2.28	2.30	2.35	2.34	2.68	2.29	<b>2.36</b>	<b>1.84</b>
SD (%)	0.02	0.02	0.02	0.02	0.03	0.05	0.02	<b>0.03</b>	-

## 4. Encapsulation of C<sub>70</sub>-IPH

### 4.1. UV-/CD-and fluorescence measurements

#### 4.1.1. Sample preparation for 1:3 mixtures

Mixtures with a fixed 1:3 host:guest ratio were prepared as follows: Bisphosphoric acid (*R,R*)-**6**, [3+2] capsule (*all-R*)-**1** or [2+2] capsule (*all-R*)-**2** were dissolved in [D<sub>1</sub>]-chloroform/[D<sub>4</sub>]-methanol = 8/2 (2 ml, each 100 μM final concentration). C<sub>70</sub>-IPH (2.39 mg, 300 μM, 3 eq for each substrate) was added into the solution and the mixture was stirred for 72 hours. The suspension was then centrifuged for one hour at 1200 rpm before being filtered through a syringe filter (0.2 μm) and measured directly.

#### 4.1.2. Fluorescence titrations

Fluorescence titrations were performed as follows: Bisphosphoric acid (*R,R*)-**6**, [3+2] capsule (*all-R*)-**1** or [2+2] capsule (*all-R*)-**2** were dissolved in [D<sub>1</sub>]-chloroform/[D<sub>4</sub>]-methanol = 8/2 (2 ml, each 100 μM final concentration). C<sub>70</sub>-IPH (5 mM, in [D<sub>1</sub>]-chloroform/[D<sub>4</sub>]-methanol = 8/2) was titrated into the solution in a stepwise fashion and the corresponding fluorescence spectra were recorded.

Fitting of the fluorescence data was performed as follows:

Software: The data was fitted using Microsoft Excel. The RGP-function was used for linear curve fitting and statistical values. The SOLVER plugin was used for nonlinear curve fitting. The sum of square residues for the calculated F<sub>0</sub>/F was minimized by changing K<sub>11</sub>. The cubic equation for the 1:2 binding was solved using the CUBIC macro.<sup>[4]</sup> Statistical values were obtained from the SOLVSTAT Macro written by E. J. Billo.<sup>[5]</sup>

Quenching components: Both dynamic and static quenching components were considered. For the dynamic quenching component, the F<sub>0</sub>/F-values for low C<sub>70</sub>-IPH concentrations [≤ 0.2 eq for (*all-R*)-**1**, ≤ 1 eq for (*all-R*)-**2** and (*R,R*)-**6**] were fitted with a linear fit according to the Stern-Volmer equation to determine K<sub>SV</sub>.

#### Definitions:

---

H: Host ( <b>1/2/6</b> )	[X]: Equilibrium conc. of X	F <sub>0</sub> : Fluorescence in absence of G
G: Guest (C <sub>70</sub> -IPH)	[X <sub>0</sub> ]: Total (added) conc. of X	F: Fluorescence for each [G]
HG: 1:1 complex		
HGG: 1:2 complex		

**Dynamic quenching** ( $K_{SV}$  determined as the slope  
in an  $F_0/F$  vs.  $[G]$  plot for low  $C_{70}$ -IPH concentrations):

$$F_0/F = 1 + K_{SV}*[G_0]$$

The fluorescence data was then fitted using both dynamic and static quenching in a  $F_0/F$  vs.  $[C_{70}$ -IPH] plot, assuming a nonfluorescent complex between  $C_{70}$ -IPH and (*all-R*)-**1/(all-R)-2/(R,R)-6**.<sup>[6]</sup>

**Total fluorescence**

$$F/F_0 = [F/F_0]_{\text{dyn}} * [H]/[H_0]$$

$$\text{with } [F/F_0]_{\text{dyn}} = 1/(1 + K_{SV}*[G_0])$$

Since only small excesses of  $C_{70}$ -IPH are used, the static quenching component cannot be abbreviated as  $F_0/F = 1 + K_a*[G_0]$  (because  $[G]$  is not equal  $[G_0]$ ). Instead, the equilibrium concentrations of the components have to be expressed via the full equations. For this, 1:1 and 1:2 equilibria were used as follows:

#### 1:1 case

---

**Host:** H

**Guest:** G

**1:1-complex:** HG

**Reaction:**  $H + G \Rightarrow HG$

**Equilibrium constant K:**  $K = [HG]/([H]*[G])$

**Total conc. of H:**  $[H_0] = [H] + [HG]$

**Total conc. of G:**  $[G_0] = [G] + [HG]$

**Equilibrium conc. of HG:**  $[HG] = 0.5 * ([G_0] + [H_0] + 1/K) - (([G_0] + [H_0] + 1/K)^2 - 4 * [H_0] * [G_0])^{0.5}$

#### 1:2 case

---

**Host:** H

**Guest:** G

**Complex:** HGG

**Reaction step 1:**  $H + G \Rightarrow HG$

**Reaction step 2:**  $HG + G \Rightarrow HGG$

**Total reaction:**  $H + 2 G \Rightarrow HGG$

**Equilibrium constant K1:**  $K1 = [HG]/([G]*[H])$

**Equilibrium constant K1:**  $K2 = [HGG]/([HG]*[H])$

**Total equilibrium constant:**  $K1*K2 = [HGG]/([H]*[G]^2)$

**Total conc. of H:**  $[H_0] = [H] + [HG] + [HGG]$

**Total conc. of G:**  $[G_0] = [G] + [HG] + 0.5[HGG]$

**Equilibrium conc. of G:**  $K11*K21*[G]^3 + (2*K11*K21*[H_0] - K11*K21*[G_0] + K11)*[G]^2 + (1 - K11*[G_0] + K11[H_0])*[G] - [G_0] = 0$

(G is obtained by solving the cubic equation)

$$A*[G]^3 + B*[G]^2 + C*[G] + D = 0$$

$$\begin{aligned}
A &= K_{11} * K_{21} \\
B &= 2 * K_{11} * K_{21} * [H_0] - K_{11} * K_{21} * [G_0] + K_{11} \\
C &= 1 - K_{11} * [G_0] + K_{11} [H_0] \\
D &= -[G_0] \\
&K_{11} * K_{21} * [G]^3 + (2 * K_{11} * K_{21} * [H_0] - K_{11} * K_{21} * [G_0] + K_{11}) * [G]^2 + (1 - \\
&K_{11} * [G_0] + K_{11} [H_0]) * [G] - [G_0] = 0
\end{aligned}$$

**Equilibrium conc. of HG:**  $[HG] = (K_{11} * [H_0] * [G]) / (1 + K_{11} * [G] + K_{11} * K_{21} * [G]^2)$

**Equilibrium conc. of HGG:**  $[HGG] = K_{21} * [HG] * [G] = (K_{11} * K_{21} * [H_0] * [G]^2) / (1 + K_{11} * [G] + K_{11} * K_{21} * [G]^2)$

This results in the following equations for fitting of the total fluorescence change:

### 1:1 case

---

**Total fluorescence**  $F/F_0 = [F/F_0]_{\text{dyn}} * [H]/[H_0]$

**Conc. of HG:**  $[HG] = 0.5 * ([G_0] + [H_0] + 1/K) - (([G_0] + [H_0] + 1/K)^2 - 4 * [H_0] * [G_0])^{0.5}$

**Conc. of H:**  $[H] = [H_0] - [HG]$

$$[H] = [H_0] - 0.5 * ([G_0] + [H_0] + 1/K) - (([G_0] + [H_0] + 1/K)^2 - 4 * [H_0] * [G_0])^{0.5}$$

**Stern-Volmer plot**

(Fitting carried out by variation of K in least-square analysis (SOLVER))

$$F_0/F = [F_0/F]_{\text{dyn}} * [H_0]/[H]$$

$$F_0/F = (1 + K_{\text{sv}}[G]) * [H_0]/[H]$$

$$F_0/F = (1 + K_{\text{sv}}[G]) * [H_0] / ([H_0] - 0.5 * ([G_0] + [H_0] + 1/K) - (([G_0] + [H_0] + 1/K)^2 - 4 * [H_0] * [G_0])^{0.5})$$

### 1:2 case

---

**Total fluorescence**  $F/F_0 = [F/F_0]_{\text{dyn}} * [H]/[H_0]$

**Conc. of HG:**  $[HG] = (K_{11} * [H_0] * [G]) / (1 + K_{11} * [G] + K_{11} * K_{21} * [G]^2)$

**Conc. of HGG:**  $[HGG] = K_{21} * [HG] * [G] = (K_{11} * K_{21} * [H_0] * [G]^2) / (1 + K_{11} * [G] + K_{11} * K_{21} * [G]^2)$

**Conc. of H:**  $[H] = [H_0] - [HG] - [HGG]$

**Stern-Volmer plot**

(Fitting carried out by variation of K<sub>11</sub> and K<sub>12</sub> in least-square analysis (SOLVER))

$$F_0/F = [F_0/F]_{\text{dyn}} * [H_0]/[H]$$

$$F_0/F = (1 + K_{\text{sv}}[G]) * [H_0] / ([H_0] - [HG] - [HGG])$$

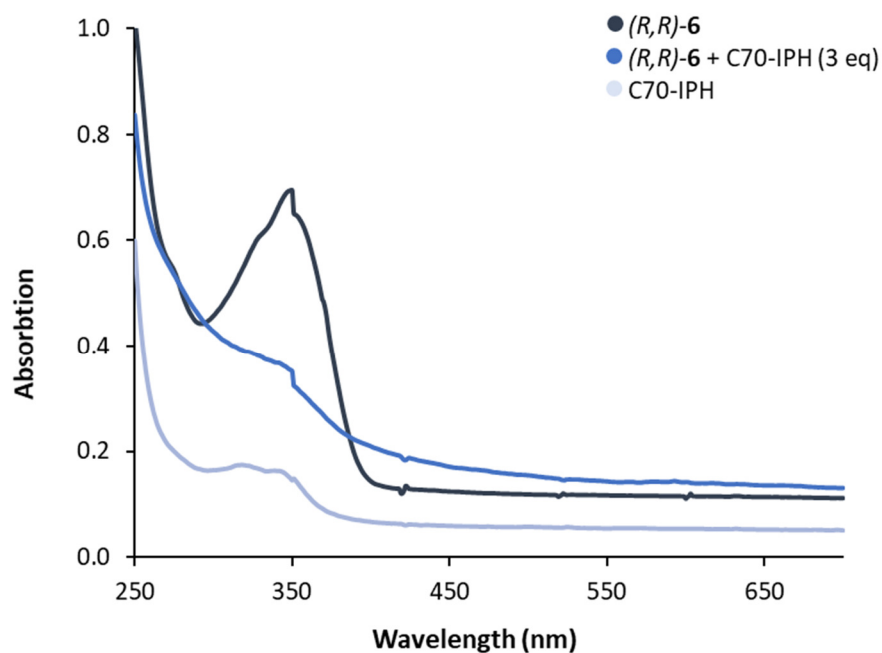
Fitting was carried out both for the 1:1 case and for the 1:2 case and the results were compared. For the 1:2 case, different fitting routines were used (independent K<sub>11</sub> and K<sub>12</sub>, equal K<sub>11</sub> and K<sub>12</sub>, noncooperative case with K<sub>11</sub> = 4 \* K<sub>12</sub>).

The best results were obtained for the noncooperative 1:2 case, which was thus applied for all cases.

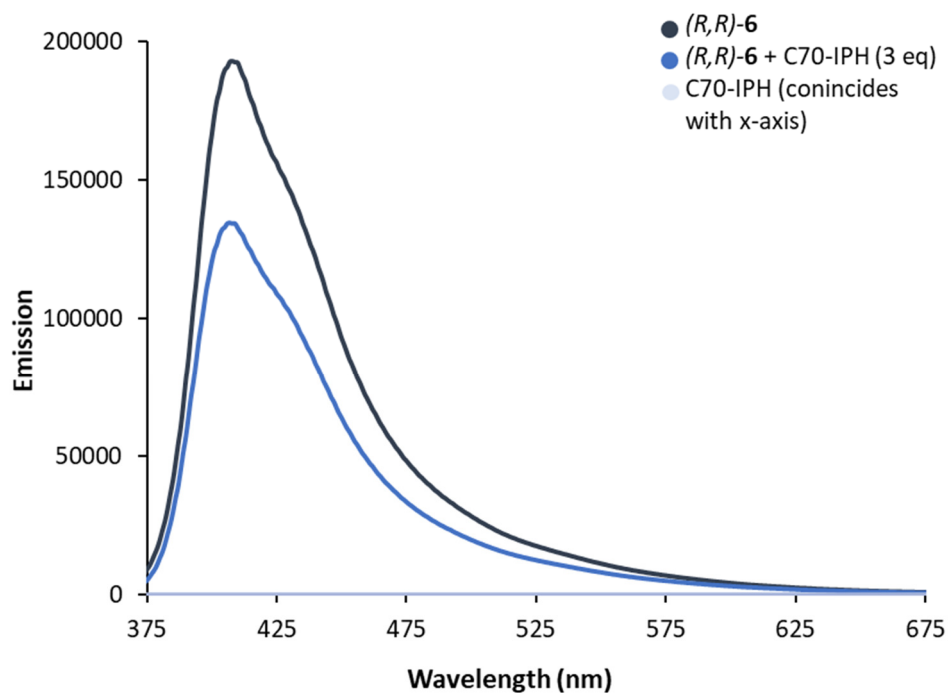
**Table S 2:** Fitting results for fluorescence quenching of the different host-molecules by C<sub>70</sub>-IPH [based on dynamic and static quenching, static quenching based on 1:2 complex stoichiometry in a noncooperative binding mode ( $K_{11} = 4 * K_{12}$ , only  $K_{11}$  reported)].

	Phosphoric acid ( <i>R,R</i> )- <b>6</b>	[3+2]capsule ( <i>all-R</i> )- <b>1</b>	[2+2]capsule ( <i>all-R</i> )- <b>2</b>
$K_{SV} [M^{-1}]$	4700 ± 770	20 300 ± 690	5000 ± 60
$R^2$	0.974	0.999	0.999
$K_{11} [M^{-1}]$	2100 ± 1500	20200 ± 4500	4600 ± 2100
$R^2$	0.893	0.917	0.936

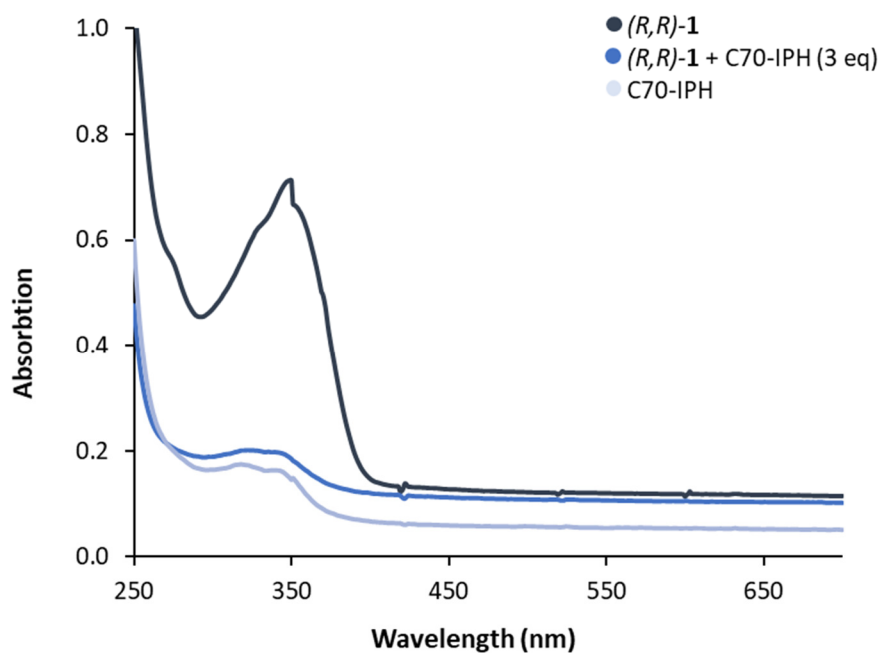
### 4.1.3. UV- and fluorescence spectra for the 1:3 mixtures



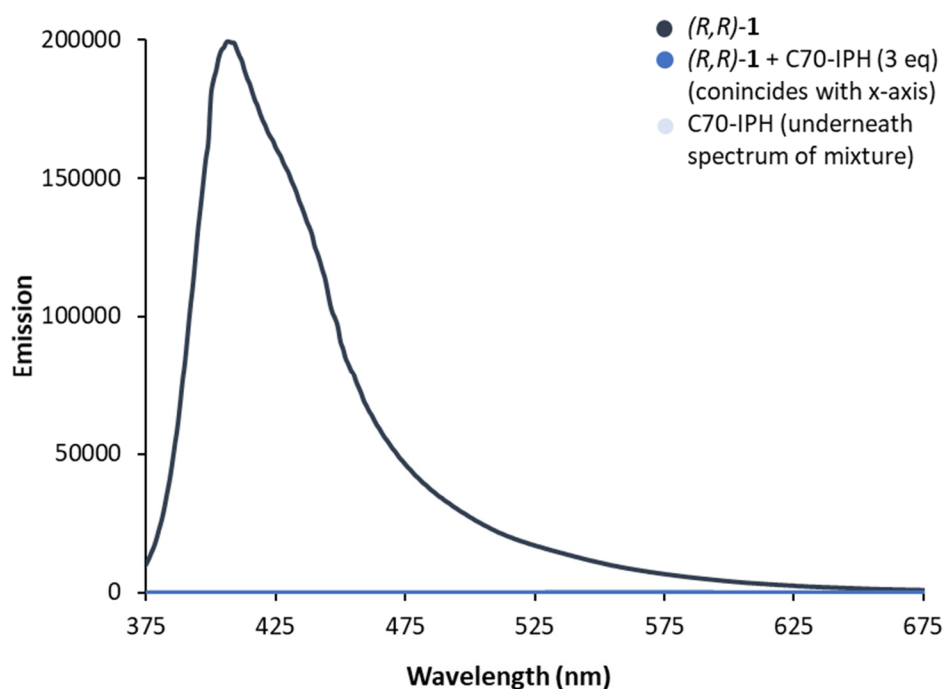
**Figure S 18:** UV absorption spectra of (*R,R*)-**6** (100  $\mu$ M), C<sub>70</sub>-IPH (300  $\mu$ M), and a mixture of (*R,R*)-**6** (100  $\mu$ M) and C<sub>70</sub>-IPH (300  $\mu$ M) (all: [D<sub>1</sub>]-chloroform/[D<sub>4</sub>]-methanol = 8/2).



**Figure S 19:** Fluorescence emission spectra of (*R,R*)-**6** (100  $\mu$ M), C<sub>70</sub>-IPH (300  $\mu$ M), and a mixture of (*R,R*)-**6** (100  $\mu$ M) and C<sub>70</sub>-IPH (300  $\mu$ M) (all: [D<sub>1</sub>]-chloroform/[D<sub>4</sub>]-methanol = 8/2,  $\lambda_{exc}$  = 350 nm).  
S55

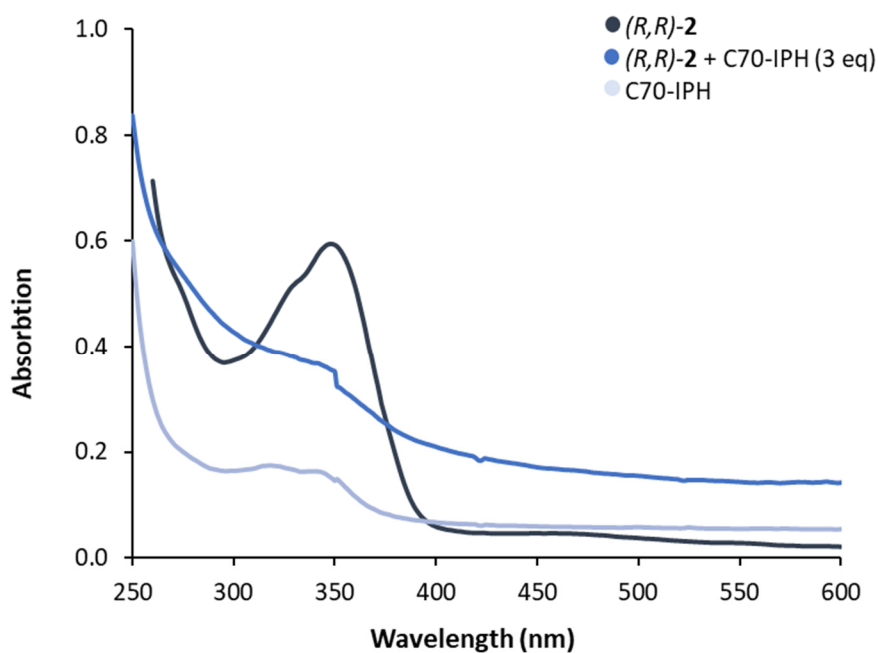


**Figure S 20:** UV absorption spectra of *(R,R)*-1 (100  $\mu\text{M}$ ),  $\text{C}_{70}$ -IPH (300  $\mu\text{M}$ ), and a mixture of *(R,R)*-1 (100  $\mu\text{M}$ ) and  $\text{C}_{70}$ -IPH (300  $\mu\text{M}$ ) (all:  $[\text{D}_1]$ -chloroform/ $[\text{D}_4]$ -methanol = 8/2).

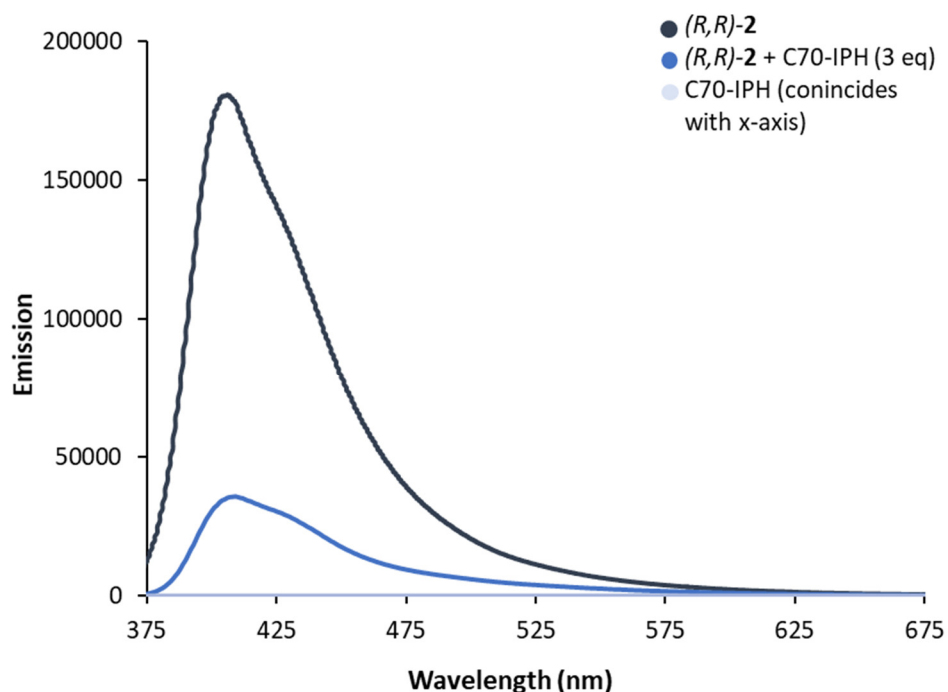


**Figure S 21:** Fluorescence emission spectra of *(R,R)*-1 (100  $\mu\text{M}$ ),  $\text{C}_{70}$ -IPH (300  $\mu\text{M}$ ), and a mixture of *(R,R)*-1 (100  $\mu\text{M}$ ) and  $\text{C}_{70}$ -IPH (300  $\mu\text{M}$ ) (all:  $[\text{D}_1]$ -chloroform/ $[\text{D}_4]$ -methanol = 8/2,  $\lambda_{\text{exc}} = 350 \text{ nm}$ ).

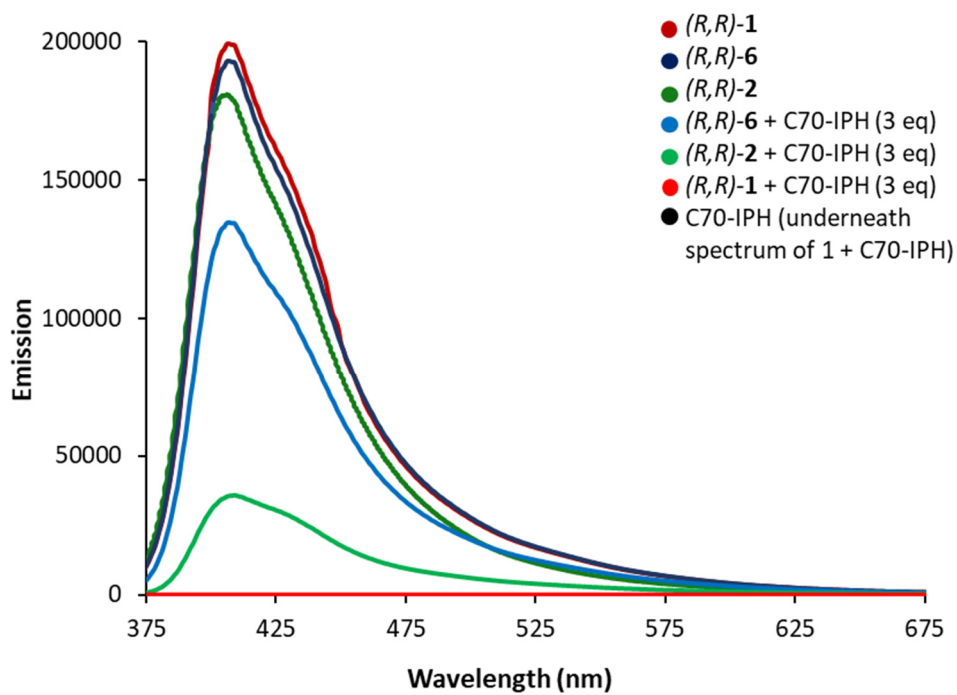




**Figure S 22:** UV absorption spectra of (*R,R*)-**2** (100  $\mu$ M), C<sub>70</sub>-IPH (300  $\mu$ M), and a mixture of (*R,R*)-**2** (100  $\mu$ M) and C<sub>70</sub>-IPH (300  $\mu$ M) (all: [D<sub>1</sub>]-chloroform/[D<sub>4</sub>]-methanol = 8/2).

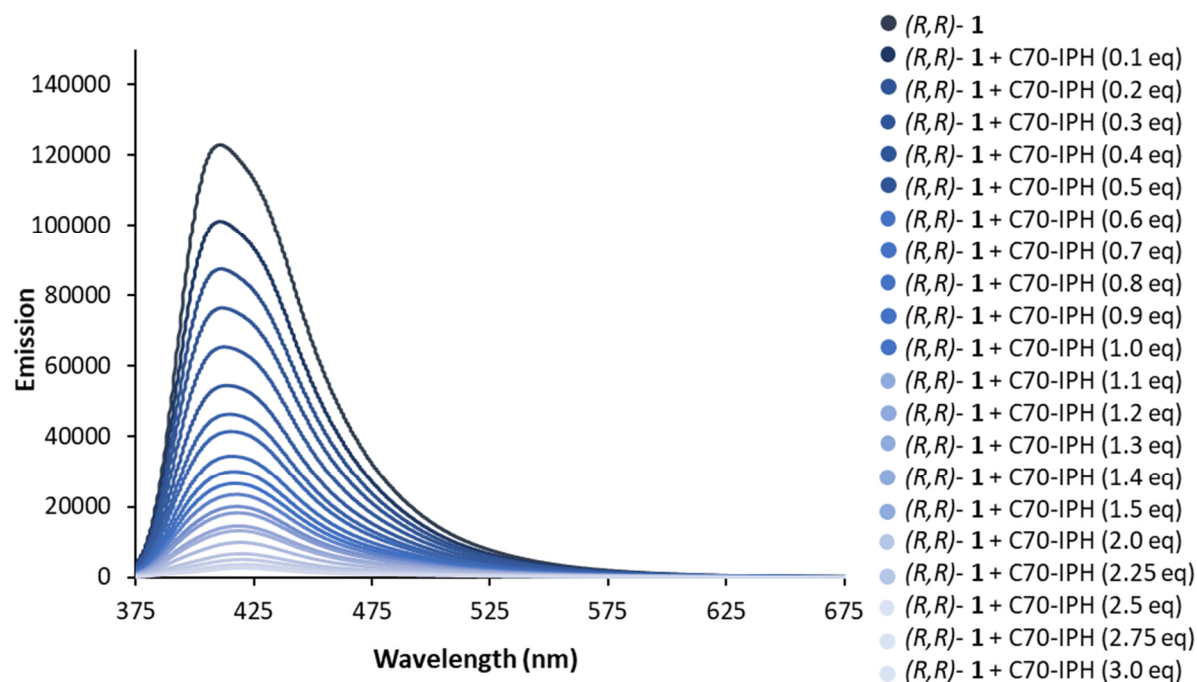


**Figure S 23:** Fluorescence emission spectra of (*R,R*)-**2** (100  $\mu$ M), C<sub>70</sub>-IPH (300  $\mu$ M), and a mixture of (*R,R*)-**1** (100  $\mu$ M) and C<sub>70</sub>-IPH (300  $\mu$ M) (all: [D<sub>1</sub>]-chloroform/[D<sub>4</sub>]-methanol = 8/2,  $\lambda_{exc}$  = 350 nm).

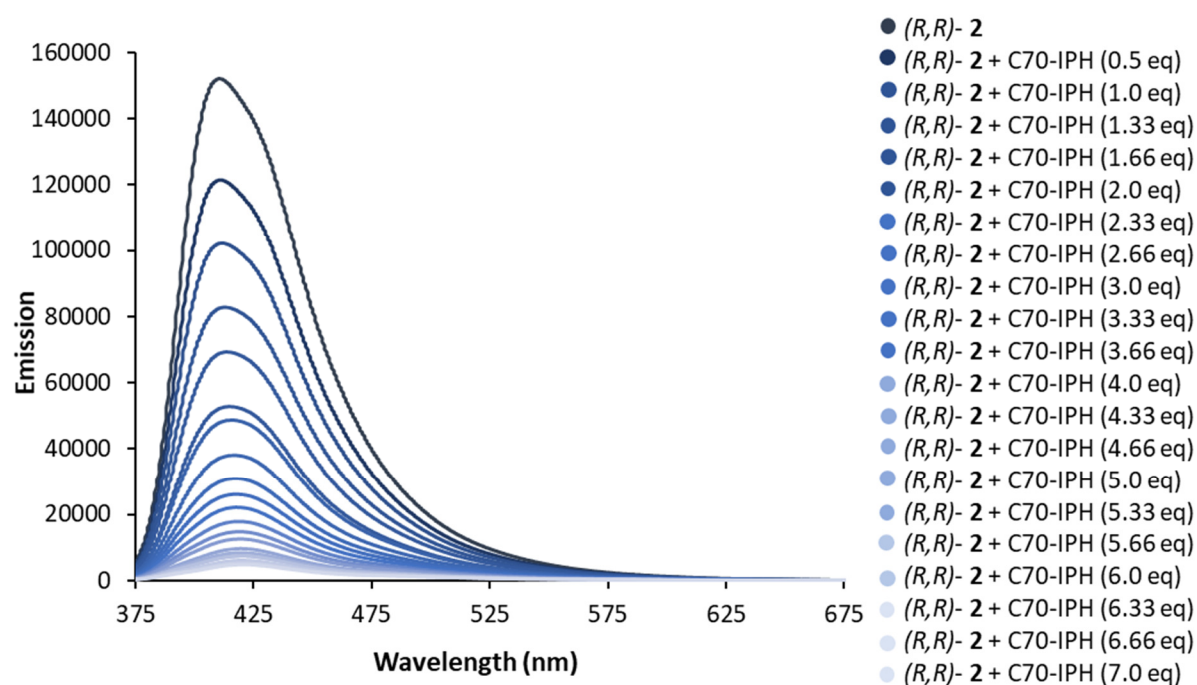


**Figure S 24:** Fluorescence emission spectra of (R,R)-6/(R,R)-1/(R,R)-2 (each 100  $\mu\text{M}$ ), C<sub>70</sub>-IPH (300  $\mu\text{M}$ ), and of mixtures of (R,R)-6/(R,R)-1/(R,R)-2 (each 100  $\mu\text{M}$ ) and C<sub>70</sub>-IPH (300  $\mu\text{M}$ ) (all: [D<sub>1</sub>]-chloroform/[D<sub>4</sub>]-methanol = 8/2,  $\lambda_{\text{exc}}$  = 350 nm).

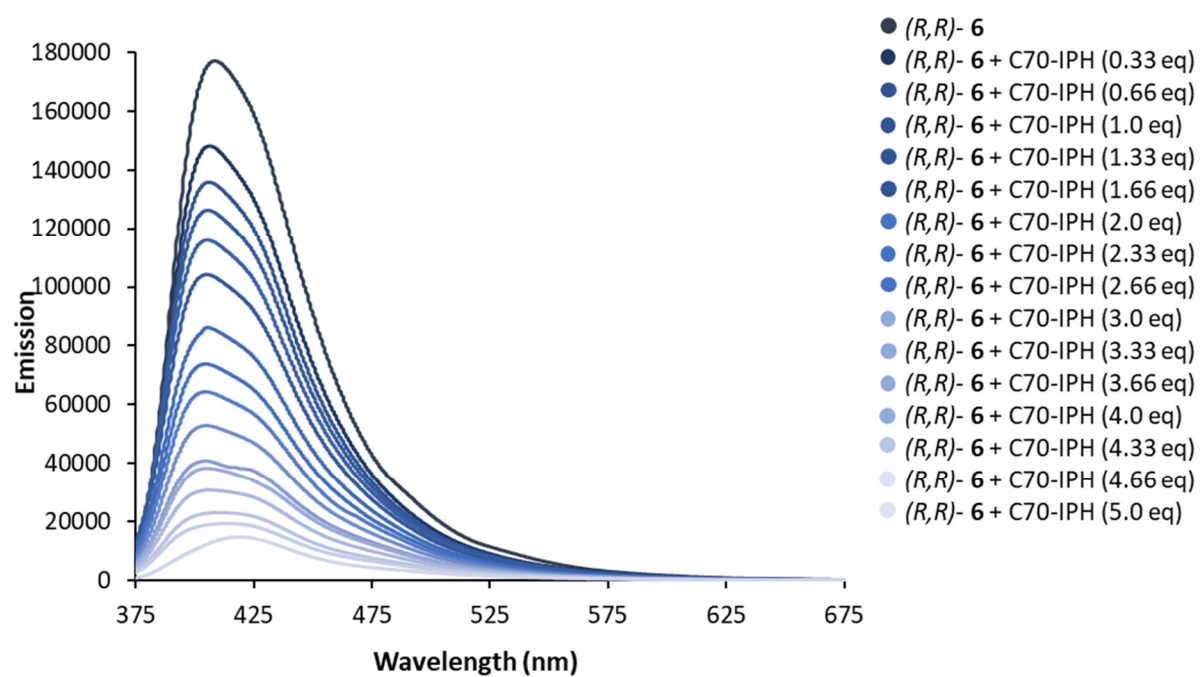
#### 4.1.4. Fluorescence spectra and fitting data for the titrations



**Figure S 25:** Fluorescence emission spectra of a titration of (R,R)-1 (100 μM), to which C<sub>70</sub>-IPH (5 mM) was added in a stepwise fashion (both: [D<sub>1</sub>]-chloroform/[D<sub>4</sub>]-methanol = 8/2, λ<sub>exc</sub> = 350 nm).

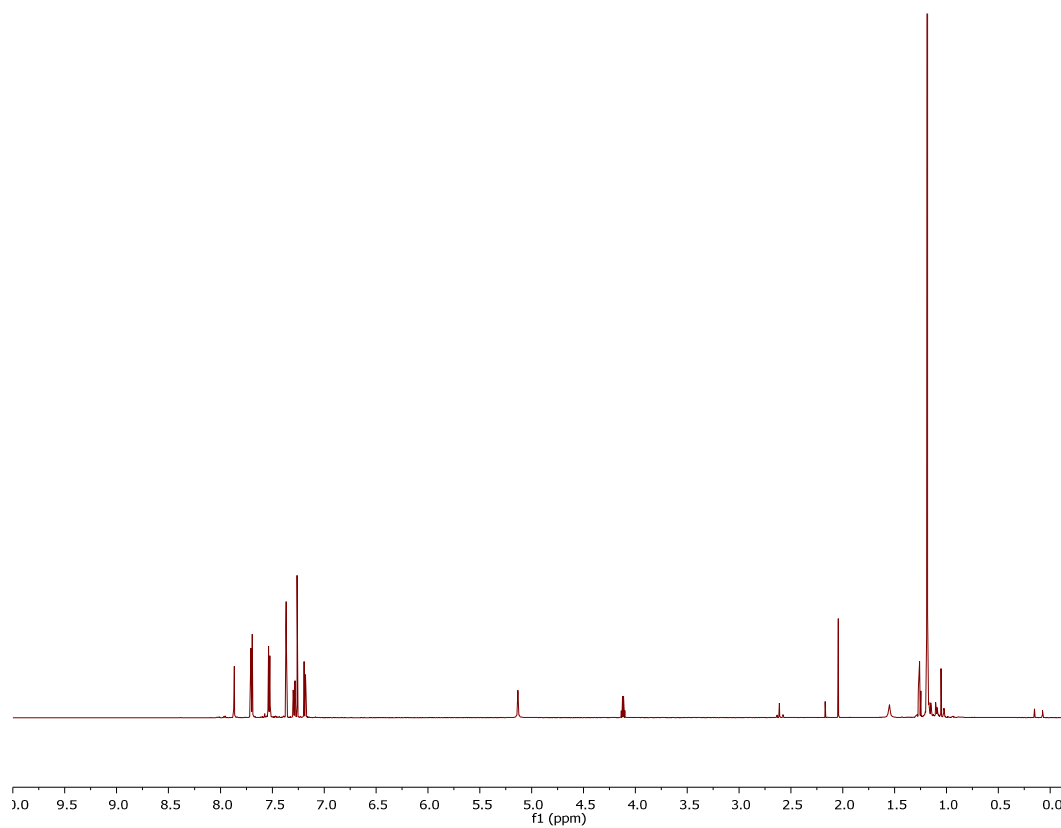


**Figure S 26:** Fluorescence emission spectra of a titration of (R,R)-2 (100 μM), to which C<sub>70</sub>-IPH (5 mM) was added in a stepwise fashion (both: [D<sub>1</sub>]-chloroform/[D<sub>4</sub>]-methanol = 8/2, λ<sub>exc</sub> = 350 nm).

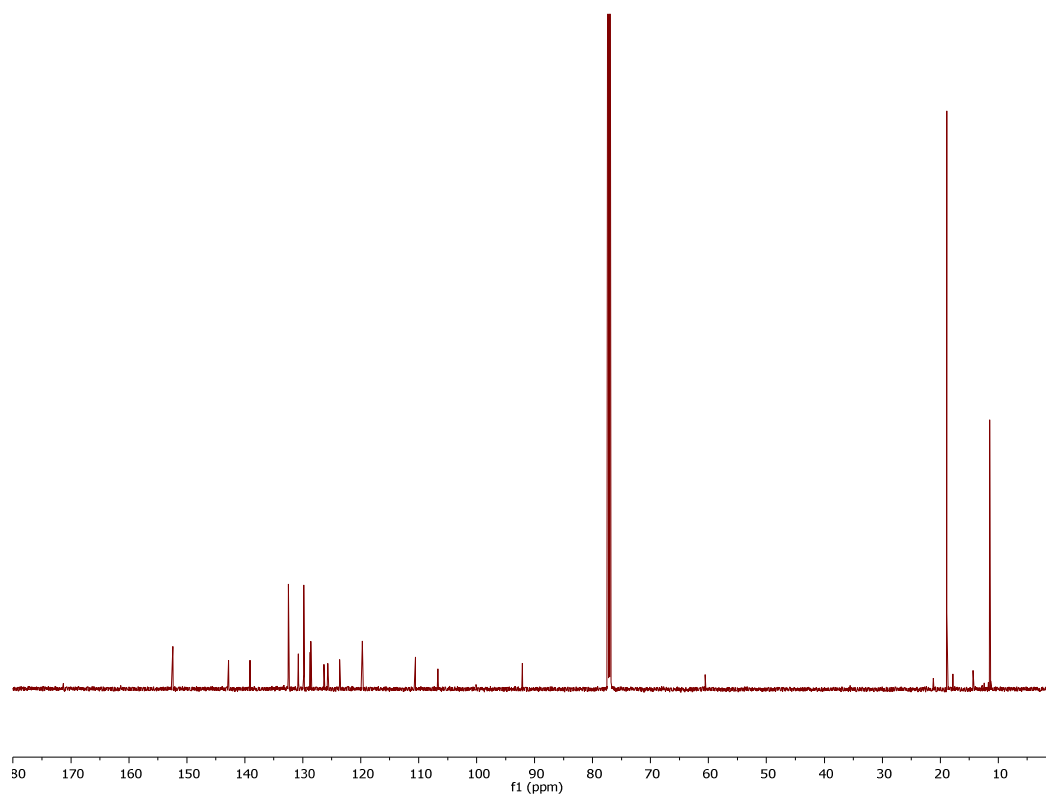


**Figure S 27:** Fluorescence emission spectra of a titration of *(R,R)*-**6** (100  $\mu$ M), to which C<sub>70</sub>-IPH (5 mM) was added in a stepwise fashion (both: [D<sub>1</sub>]-chloroform/[D<sub>4</sub>]-methanol = 8/2,  $\lambda_{\text{exc}}$  = 350 nm).

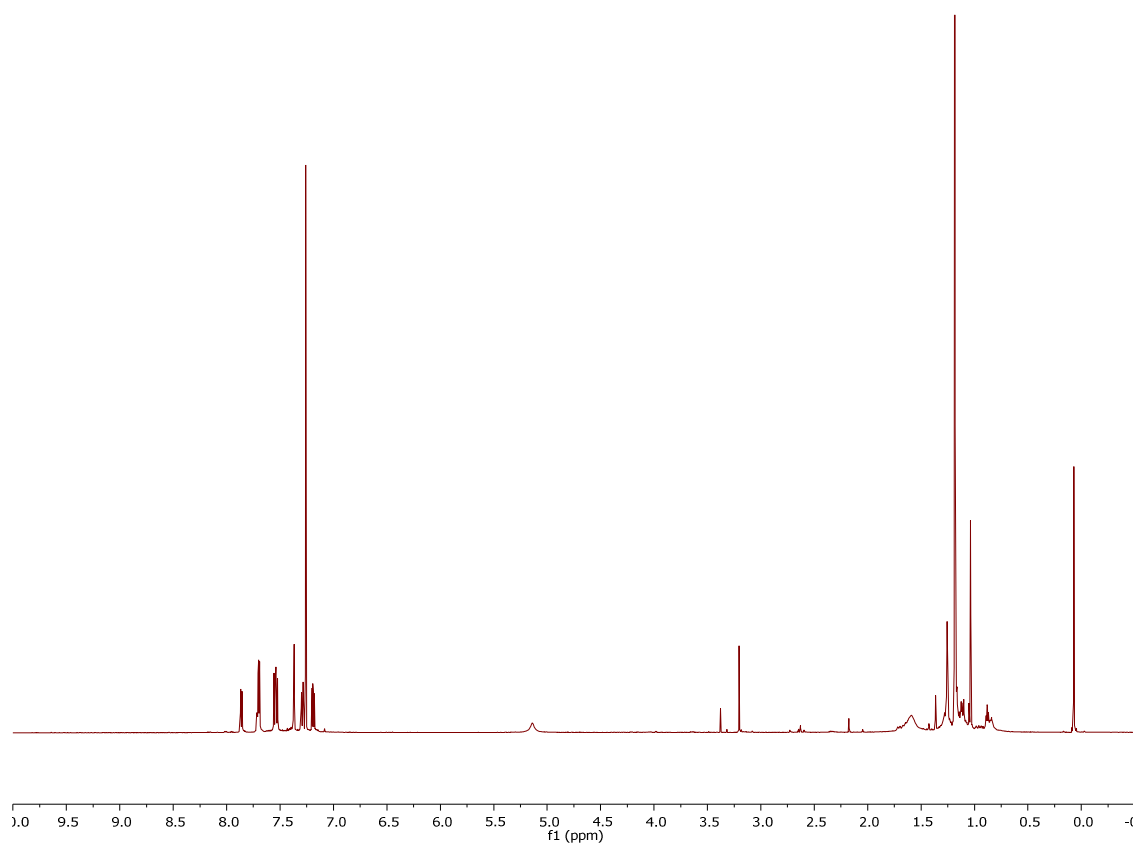
## 5. NMR-Spectra



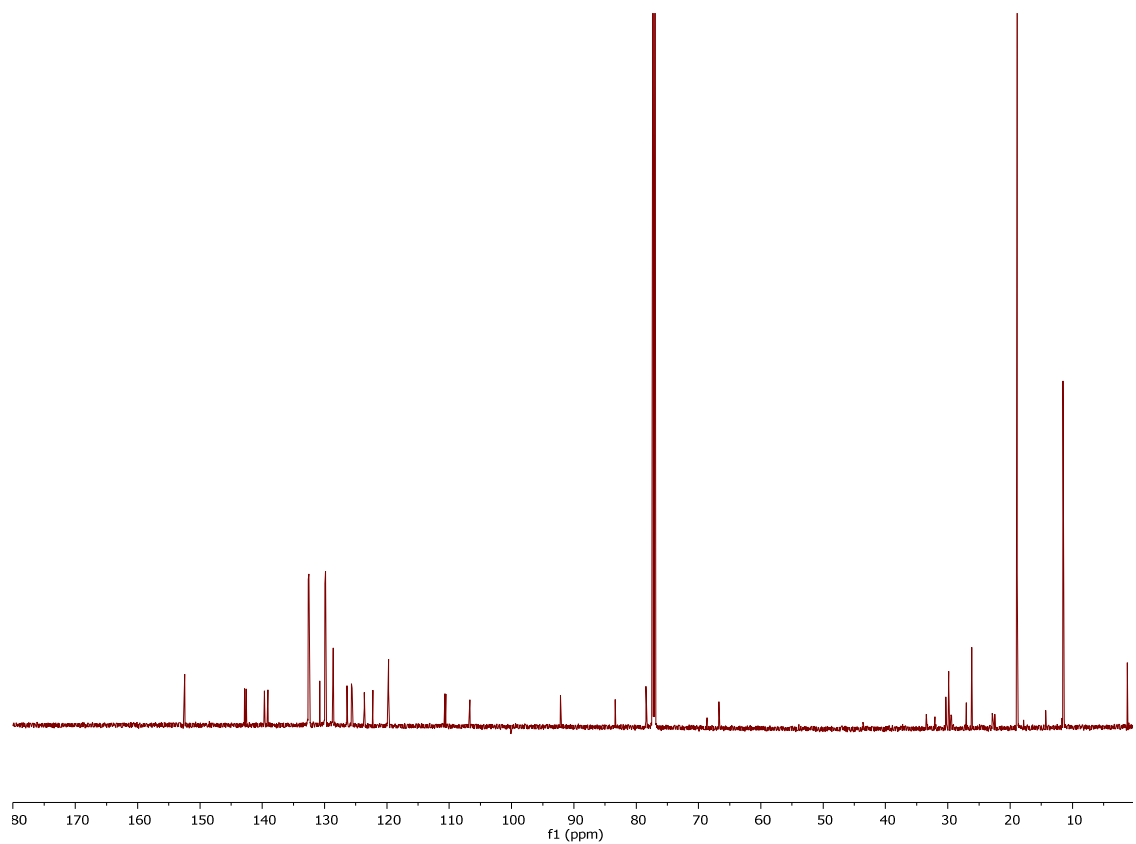
**Figure S 28:**  $^1\text{H}$ -NMR spectrum of (*R*)-3 (600MHz,  $[\text{D}_1]$ -chloroform, 298 K).



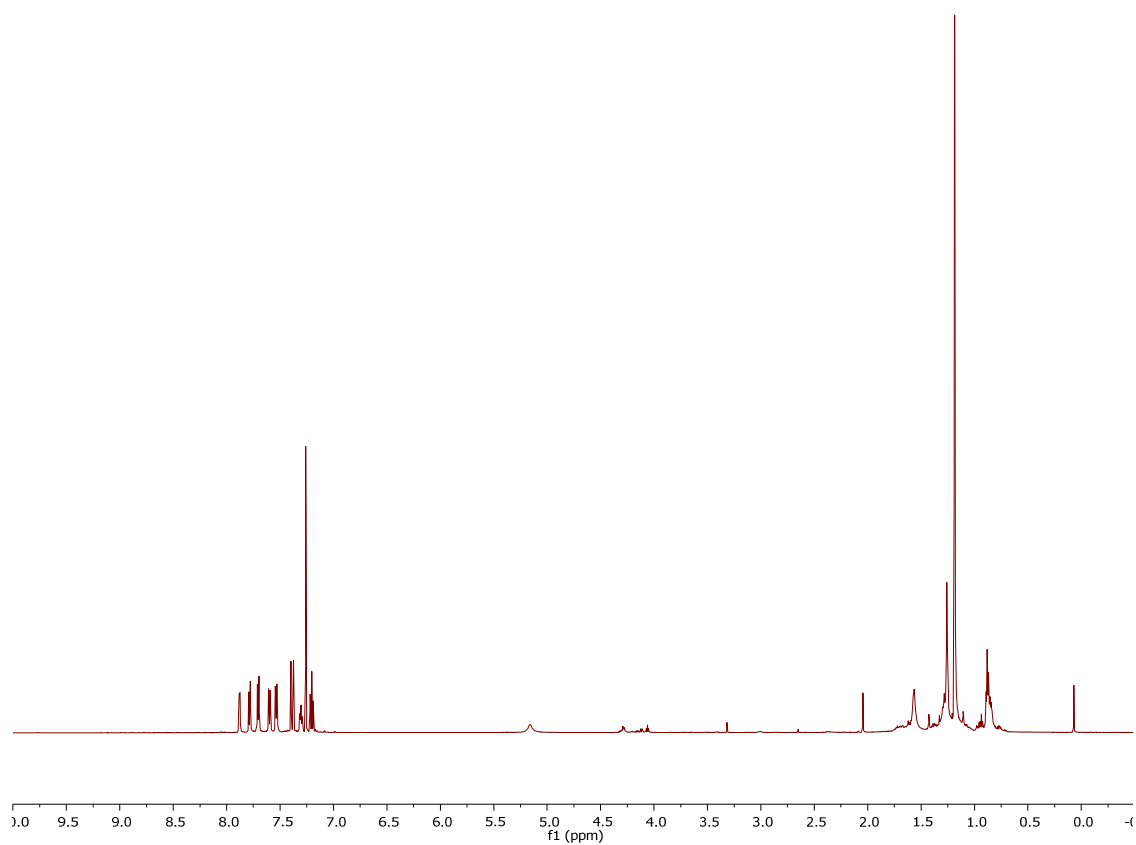
**Figure S 29:**  $^{13}\text{C}$ -NMR Spectrum of (*R*)-3 (150 MHz,  $[\text{D}_1]$ -chloroform, 298 K).



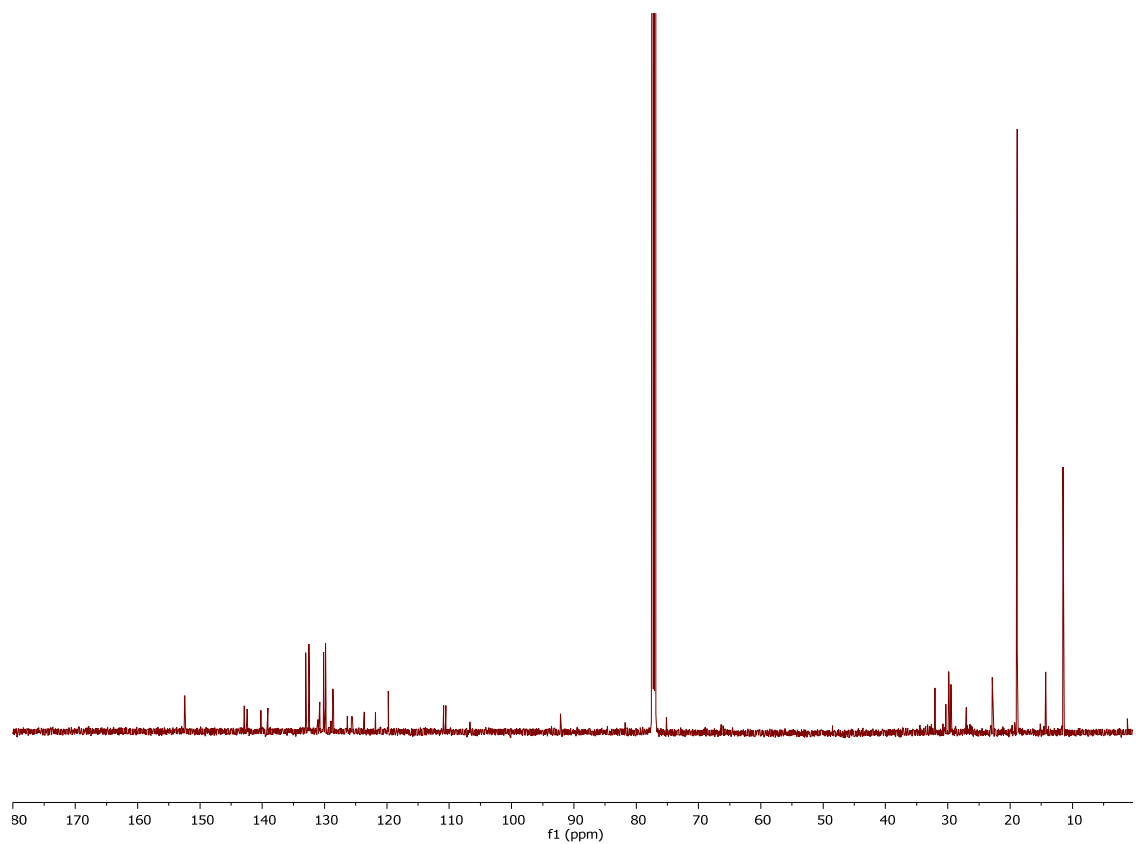
**Figure S 30:**  $^1\text{H}$ - NMR Spectrum of (*R*)-**4** (600MHz,  $[\text{D}_1]$ -chloroform, 298 K).



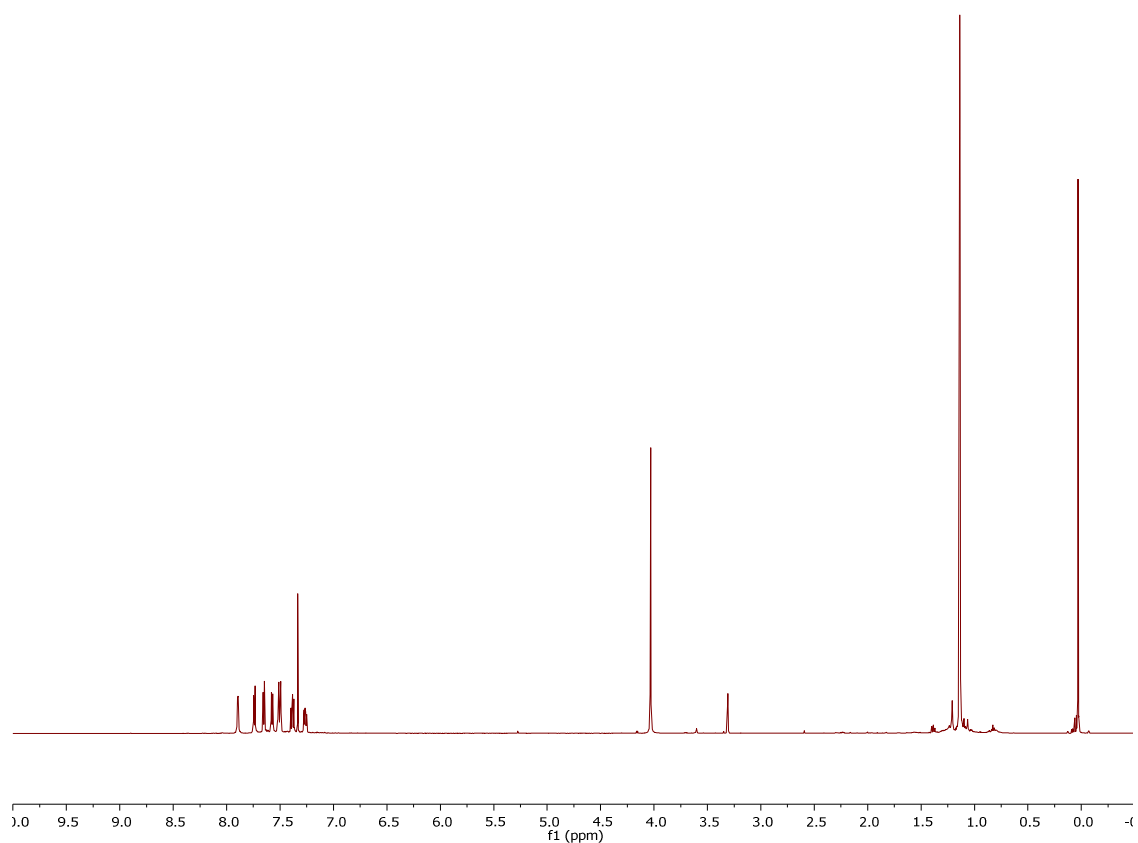
**Figure S 31:**  $^{13}\text{C}$ -NMR Spectrum of (*R*)-**4** (150 MHz,  $[\text{D}_1]$ -chloroform, 298 K).



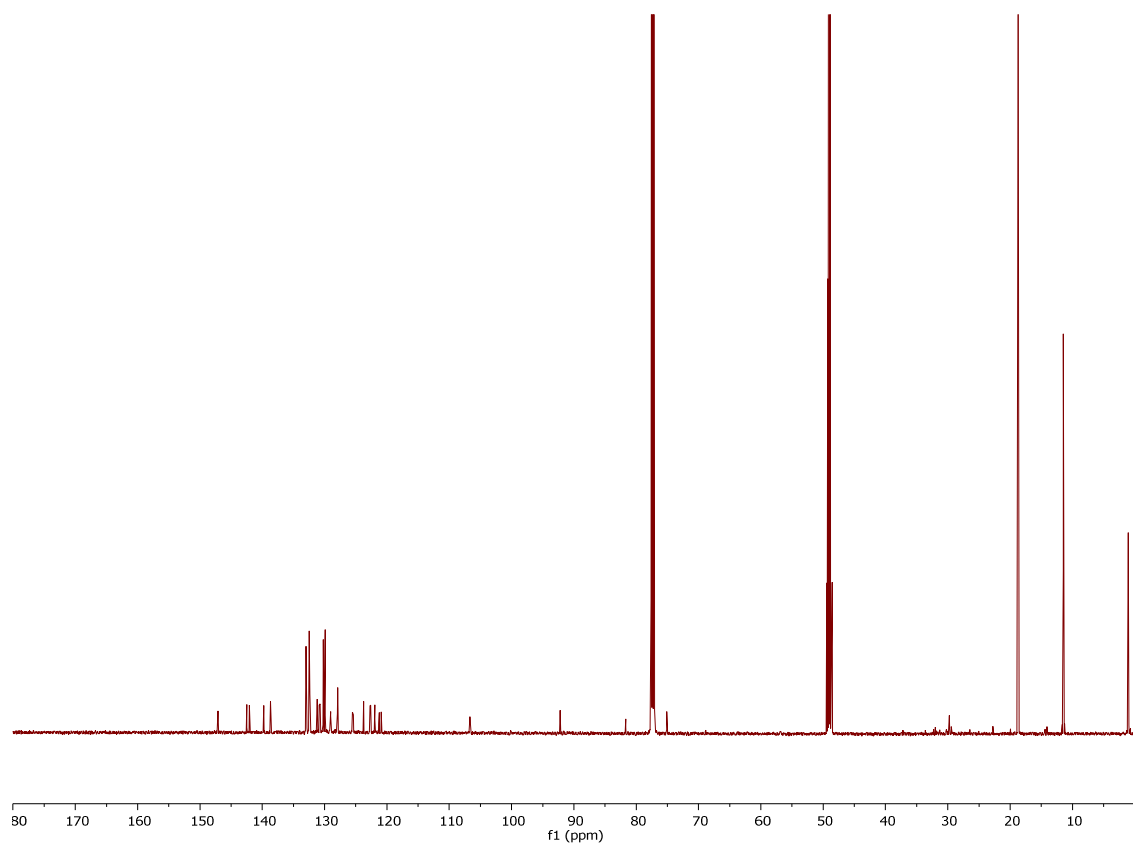
**Figure S 32:** <sup>1</sup>H-NMR spectrum of (*R*)-**5** (600MHz, [D<sub>1</sub>]-chloroform, 298 K).



**Figure S 33:** <sup>13</sup>C-NMR Spectrum of (*R*)-**5** (150 MHz, [D<sub>1</sub>]-chloroform, 298 K).

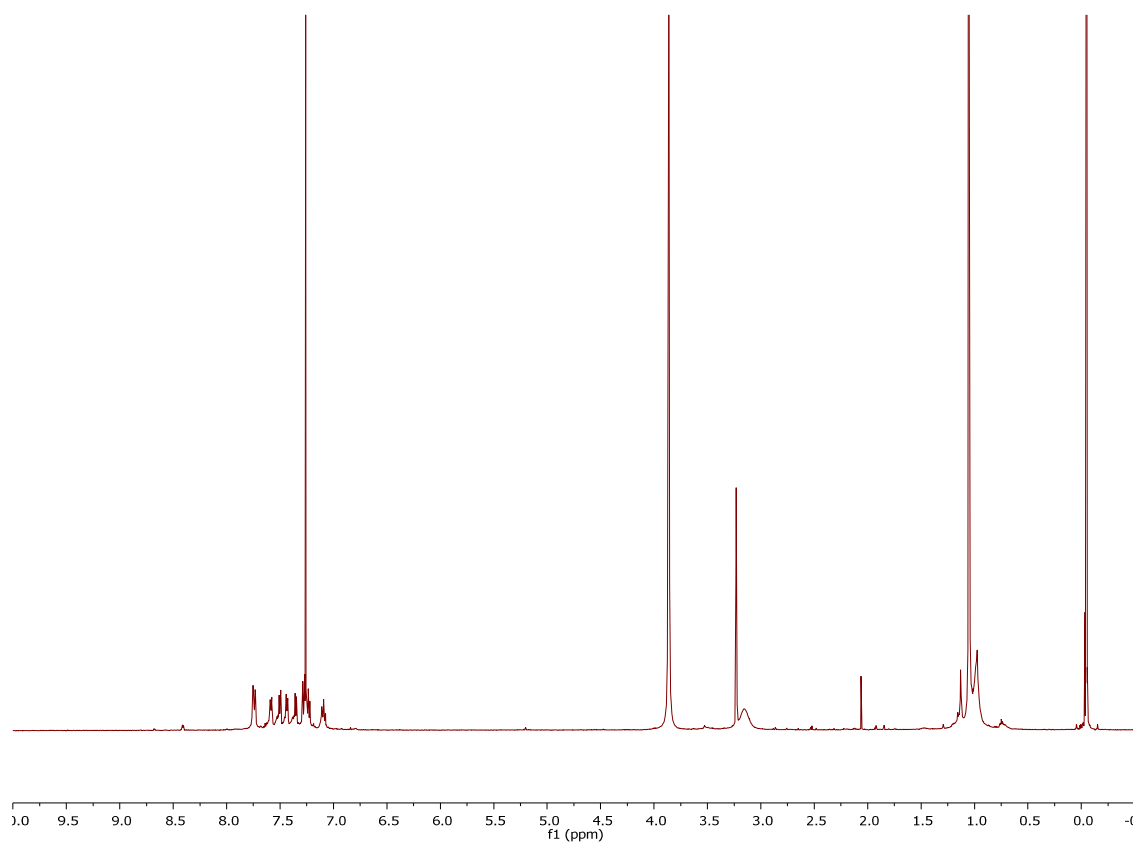


**Figure S 34:** <sup>1</sup>H-NMR Spectrum of (*R,R*)-**6** (600 MHz, [D<sub>1</sub>]-chloroform/[D<sub>4</sub>]-methanol = 8/2 (v/v), 298 K).

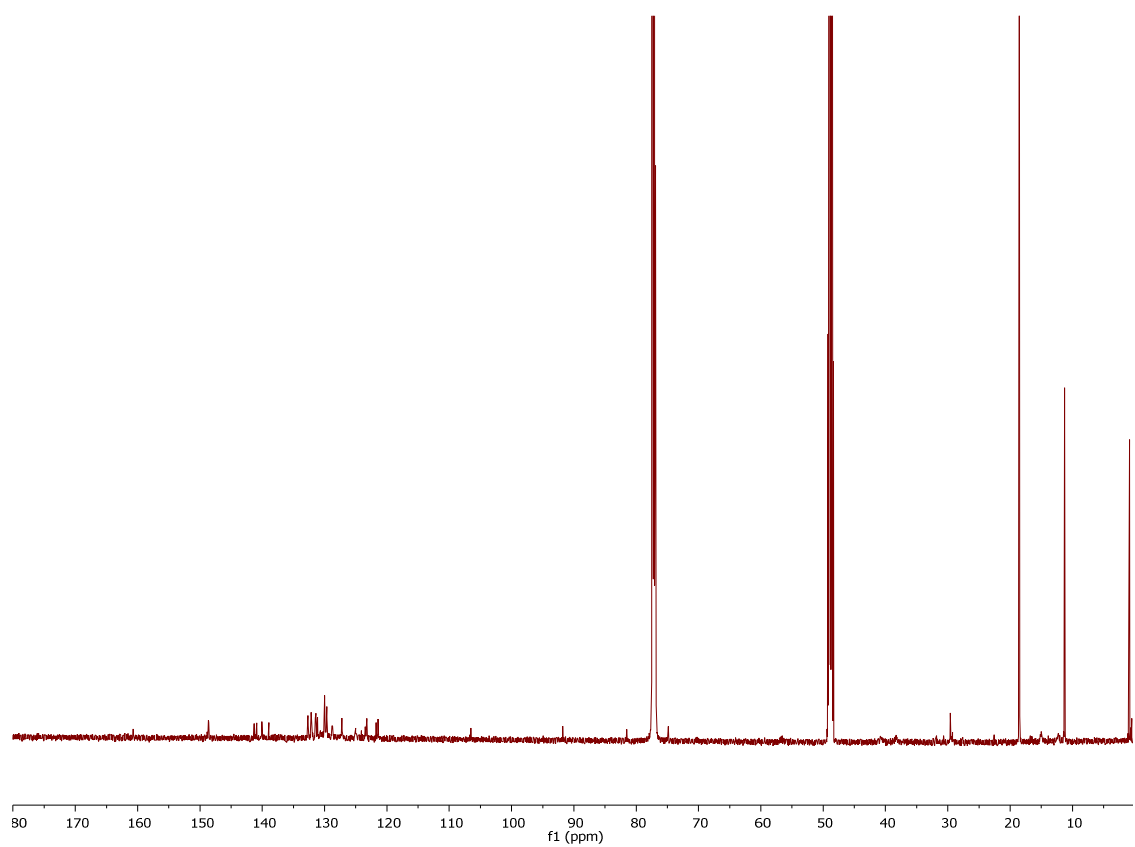


**Figure S 35:** <sup>13</sup>C-NMR Spectrum of (*R,R*)-**6** (150 MHz, [D<sub>1</sub>]-chloroform/[D<sub>4</sub>]-methanol = 8/2 (v/v), 298 K).

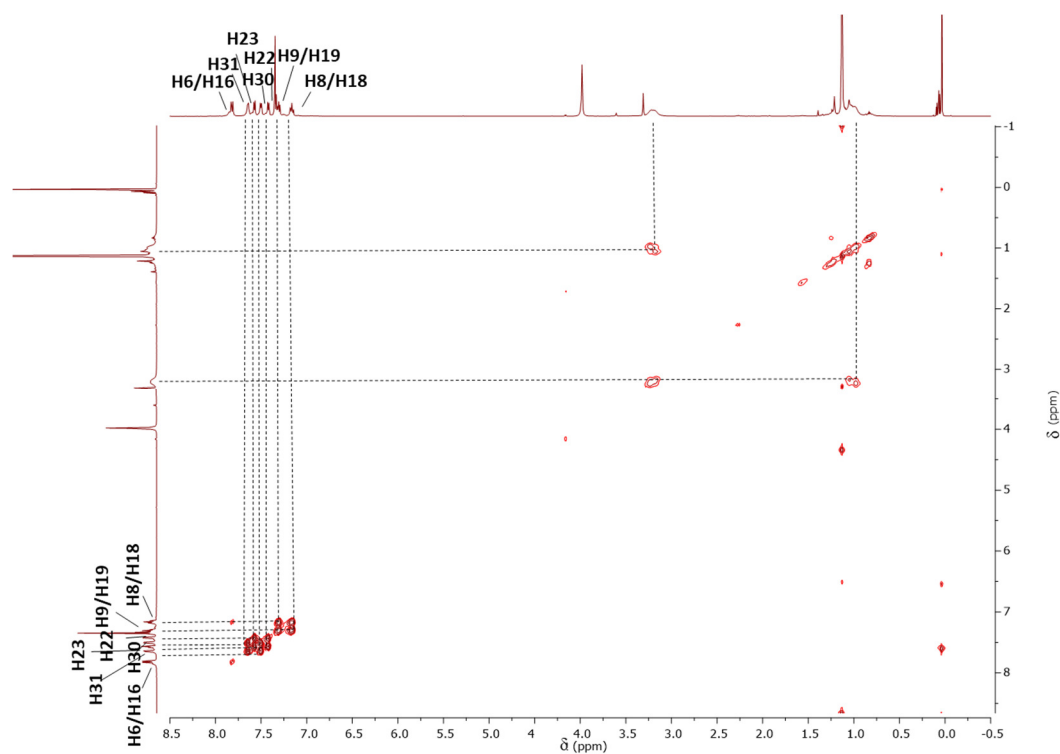




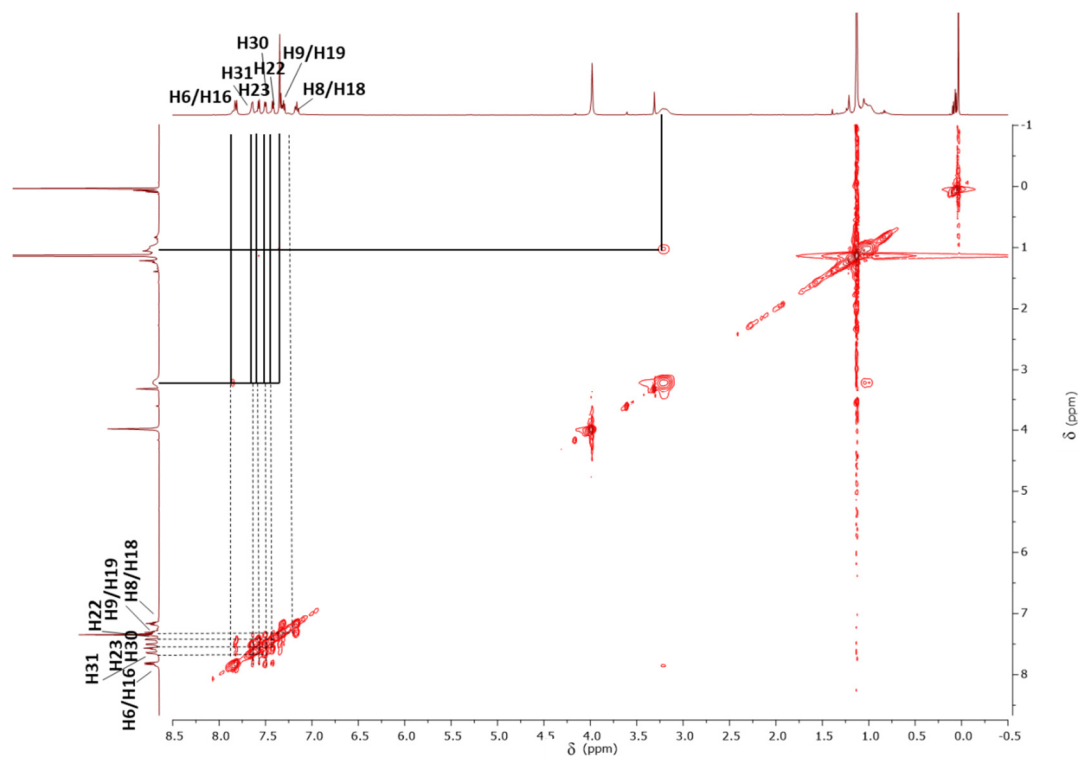
**Figure S 36:** <sup>1</sup>H-NMR spectrum of (*all-R*)-**1** (600 MHz, [D<sub>1</sub>]-chloroform/[D<sub>4</sub>]-methanol = 8/2 (v/v), 298 K).



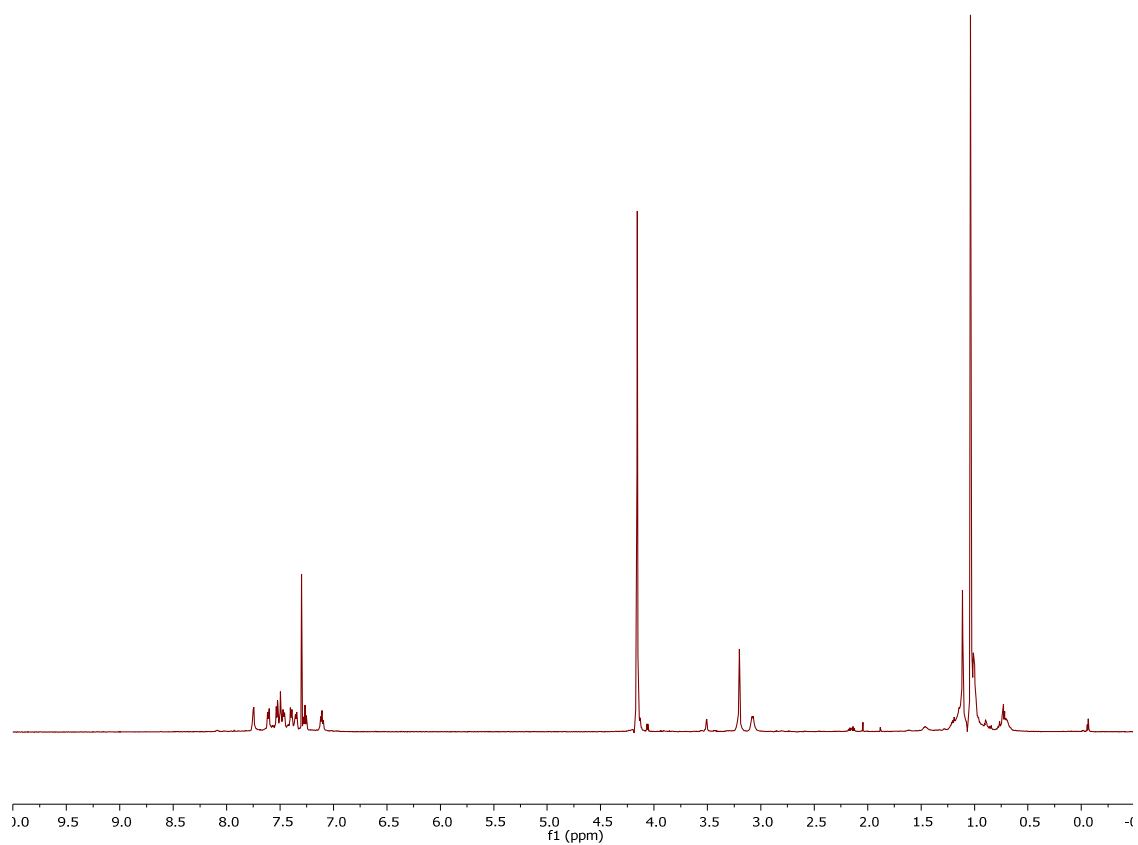
**Figure S 37:** <sup>13</sup>C-NMR spectrum of (*all-R*)-**1** (150 MHz, [D<sub>1</sub>]-chloroform/[D<sub>4</sub>]-methanol = 8/2 (v/v), 298 K).



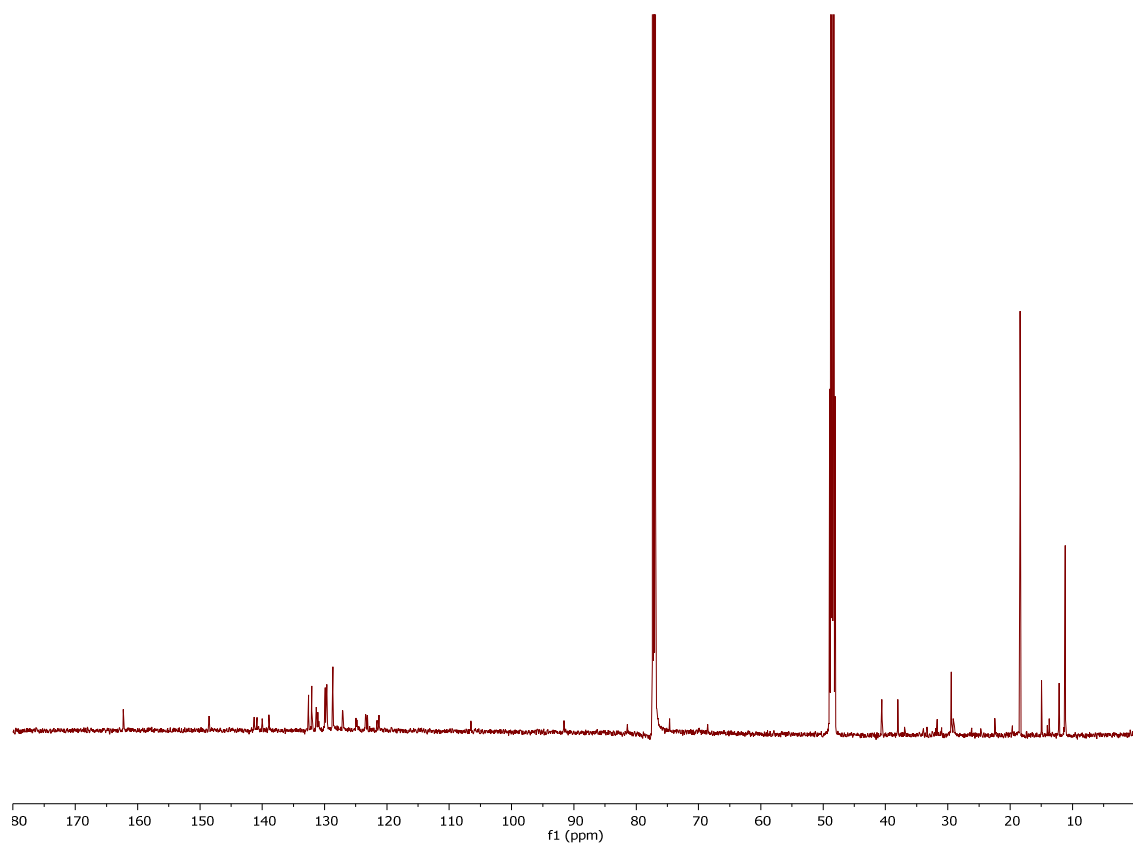
**Figure S 38:**  $^1\text{H}$ -COSY NMR spectrum of (*all-R*)-**1** (600/600 MHz,  $[\text{D}_1]$ -chloroform/ $[\text{D}_4]$ -methanol = 8/2 (v/v), 298 K). Dashed lines indicate intramolecular interactions, while solid lines represent intermolecular interactions.



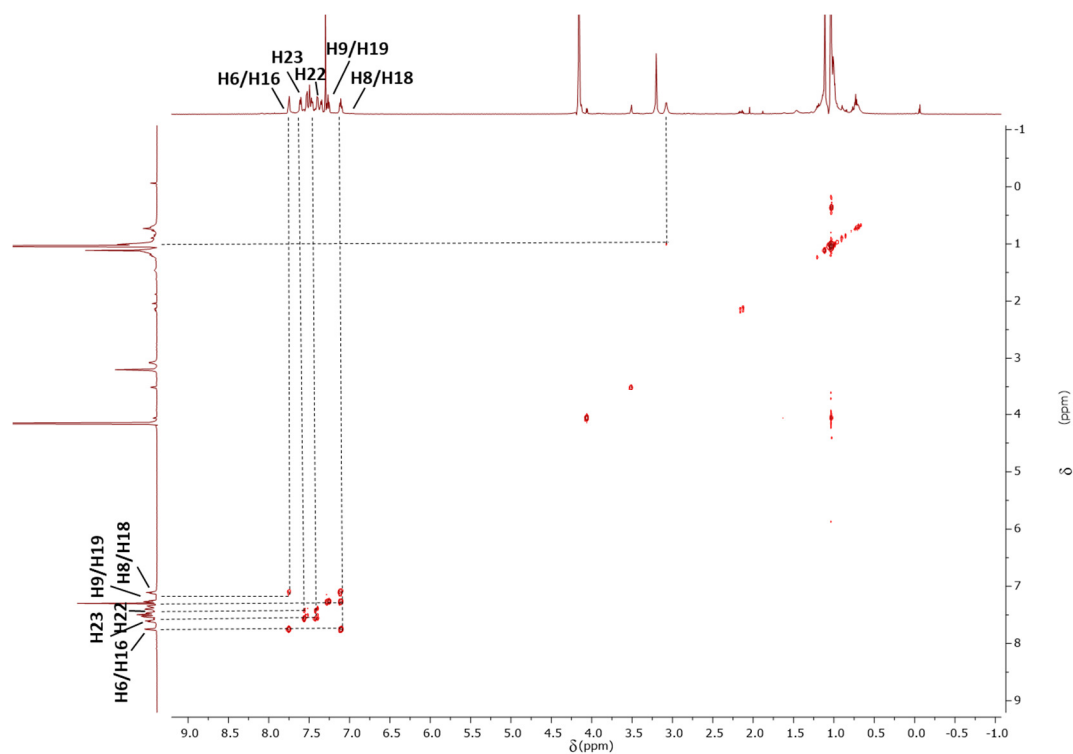
**Figure S 39:**  $^1\text{H}$ -NOESY NMR spectrum of (*all-R*)-**1** (600/600 MHz,  $[\text{D}_1]$ -chloroform/ $[\text{D}_4]$ -methanol = 8/2 (v/v), 298 K). Dashed lines indicate intramolecular interactions, while solid lines represent intermolecular interactions. For enlargement see the main paper.



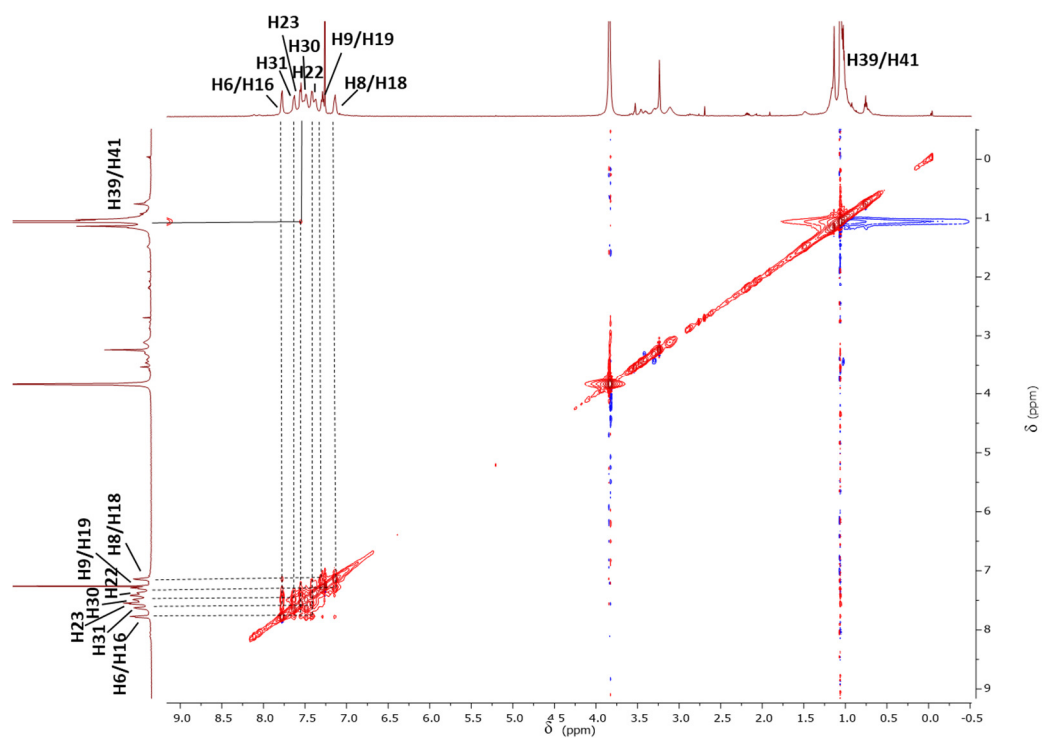
**Figure S 40:**  $^1\text{H}$ -NMR Spectrum of (*all-R*)-**2** (600 MHz,  $[\text{D}_1]$ -chloroform/ $[\text{D}_4]$ -methanol = 8/2 (v/v), 298 K).



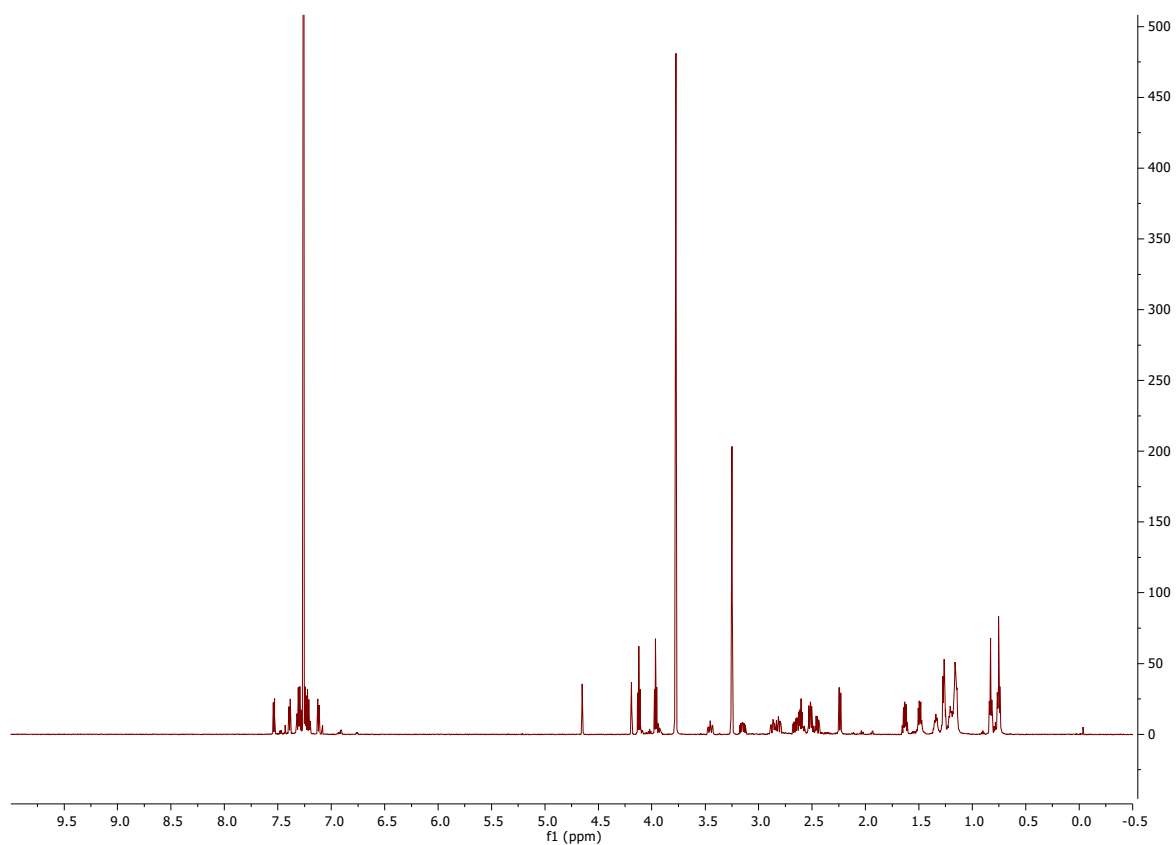
**Figure S 41:**  $^{13}\text{C}$ -NMR Spectrum of (*all-R*)-**2** (150 MHz,  $[\text{D}_1]$ -chloroform/ $[\text{D}_4]$ -methanol = 8/2 (v/v), 298 K).



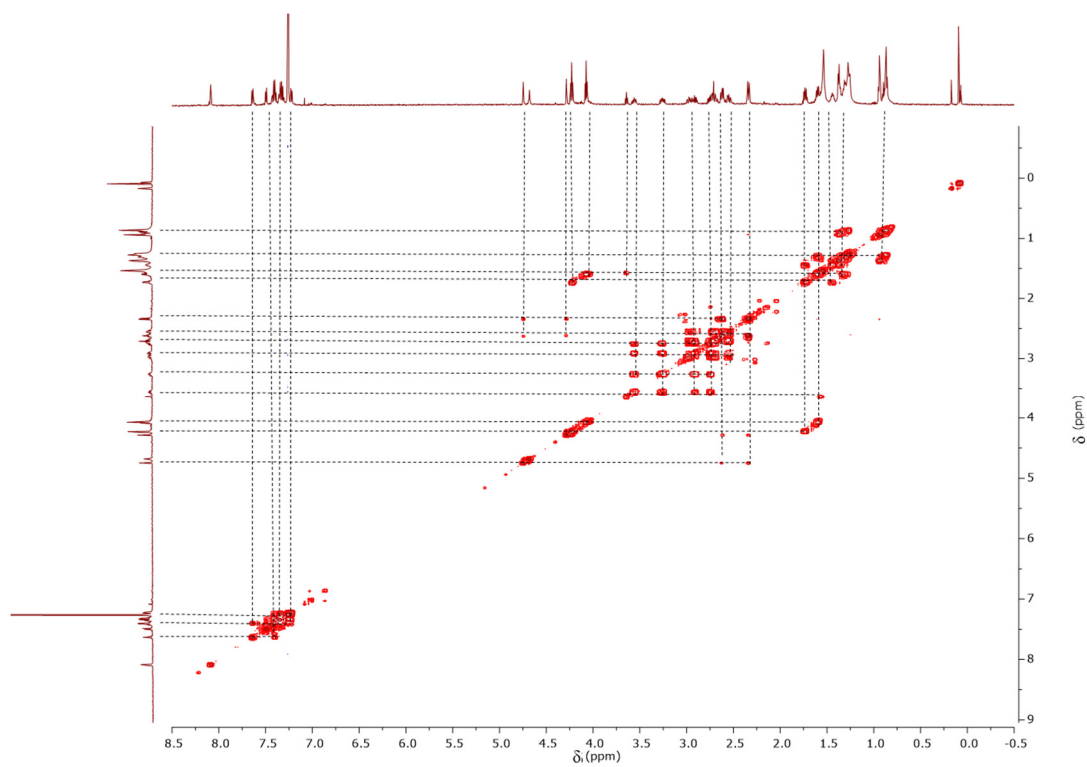
**Figure S 42:**  $^1\text{H}$ -COSY NMR spectrum of (*all-R*)-**2** (600/600 MHz,  $[\text{D}_1]$ -chloroform/ $[\text{D}_4]$ -methanol = 8/2 (v/v), 298 K). Dashed lines indicate intramolecular interactions, while solid lines represent intermolecular interactions.



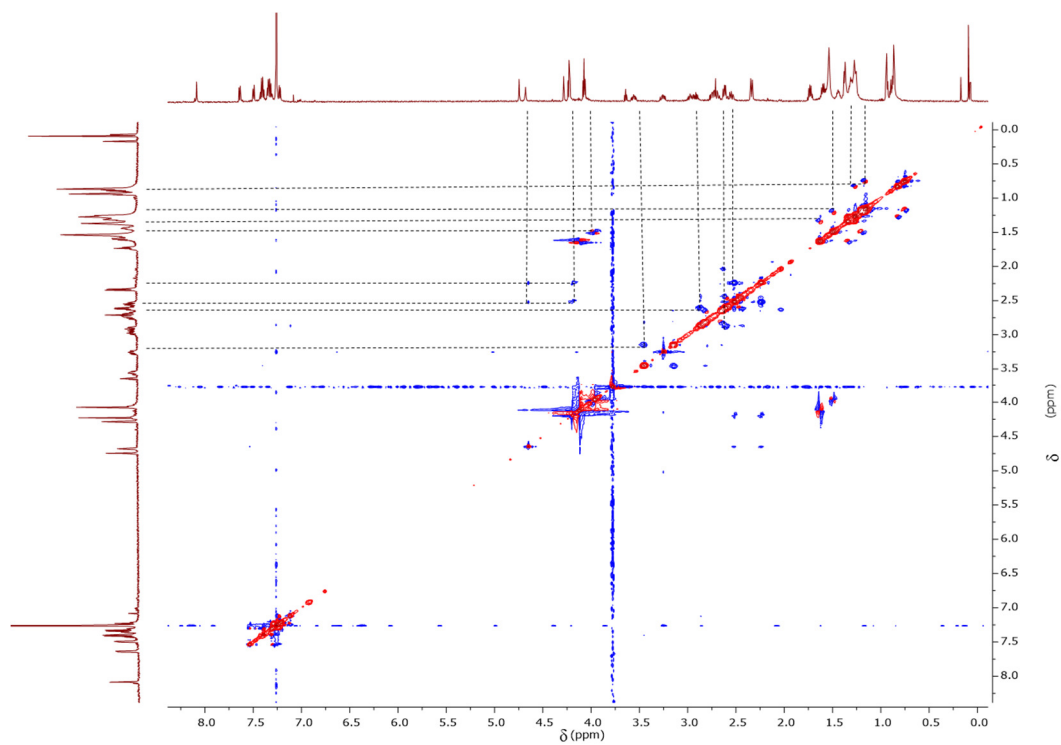
**Figure S 43:**  $^1\text{H}$ -NOESY NMR spectrum of (*all-R*)-**2** (600/600 MHz,  $[\text{D}_1]$ -chloroform/ $[\text{D}_4]$ -methanol = 8/2 (v/v), 298 K). Dashed lines indicate intramolecular interactions, while solid lines represent intermolecular interactions.



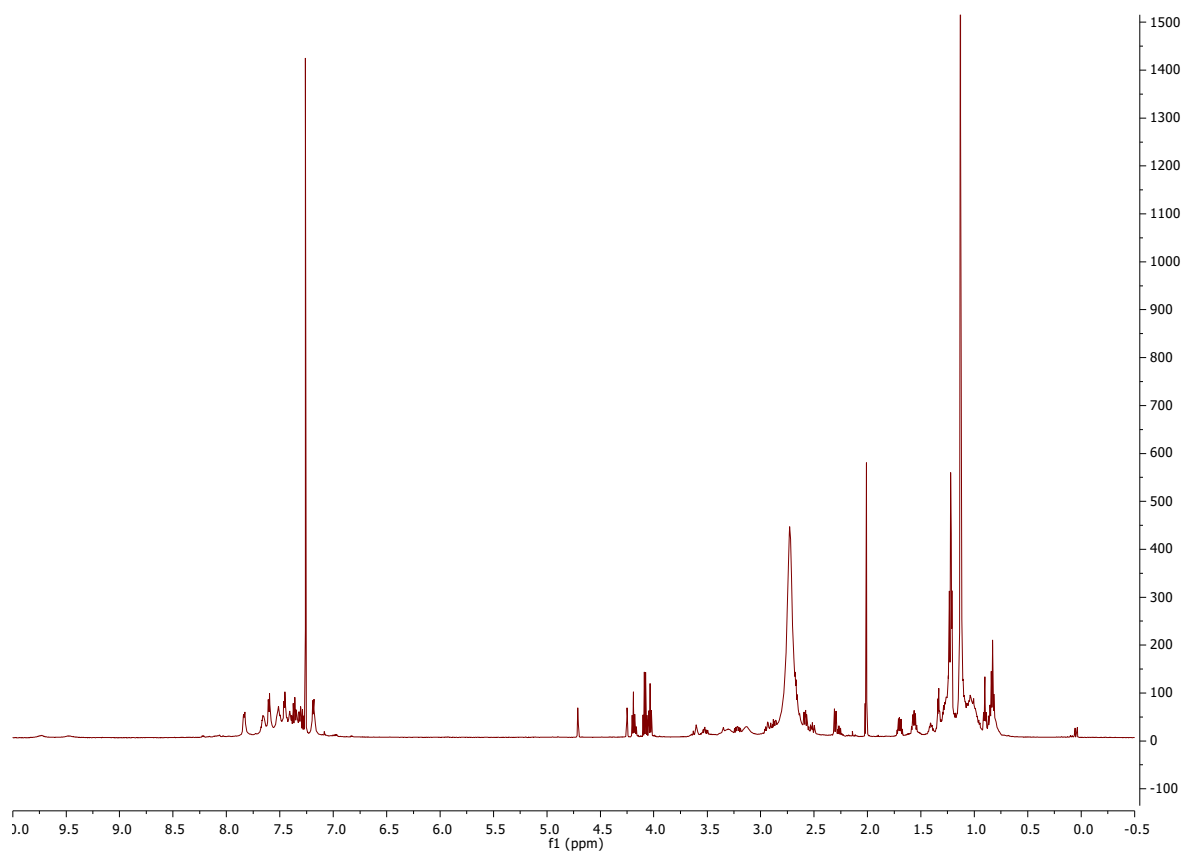
**Figure S 44:**  $^1\text{H}$ -NMR spectrum of  $\text{C}_{70}$ -IPH (600 MHz,  $[\text{D}_1]$ -chloroform/ $[\text{D}_4]$ -methanol = 8/2 (v/v), 298 K).



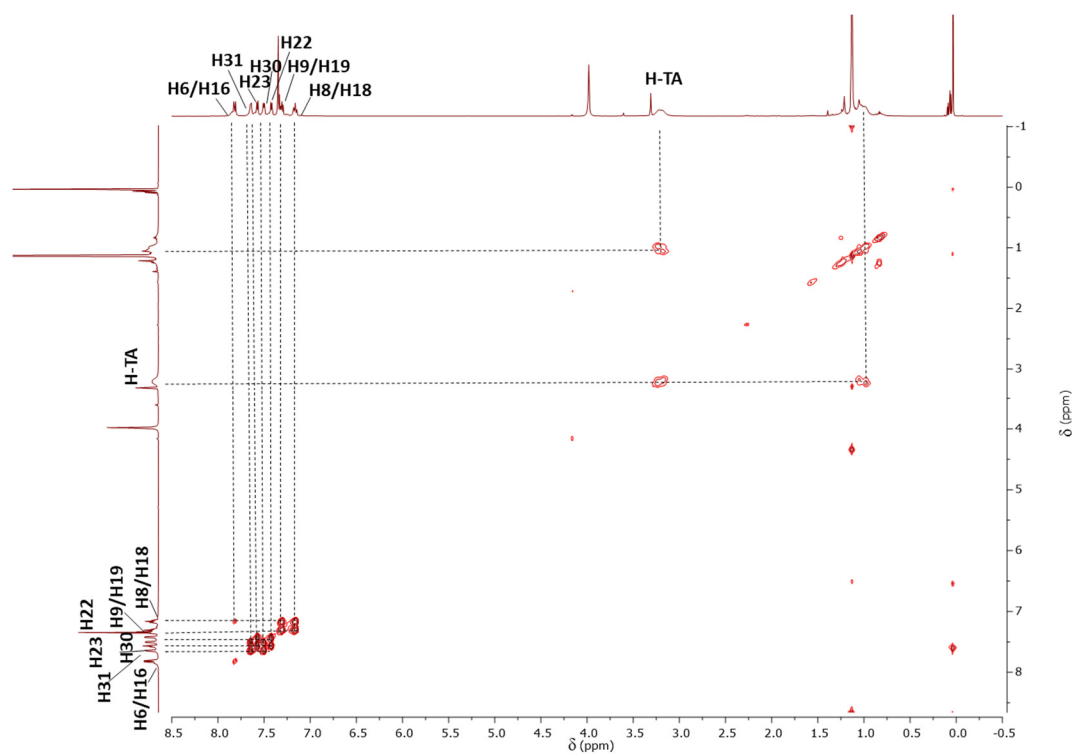
**Figure S 45:**  $^1\text{H}$ -COSY NMR spectrum of  $\text{C}_{70}$ -IPH (600/600 MHz,  $[\text{D}_1]$ -chloroform/ $[\text{D}_4]$ -methanol = 8/2 (v/v), 298 K). Dashed lines indicate intramolecular interactions, while solid lines represent intermolecular interactions.



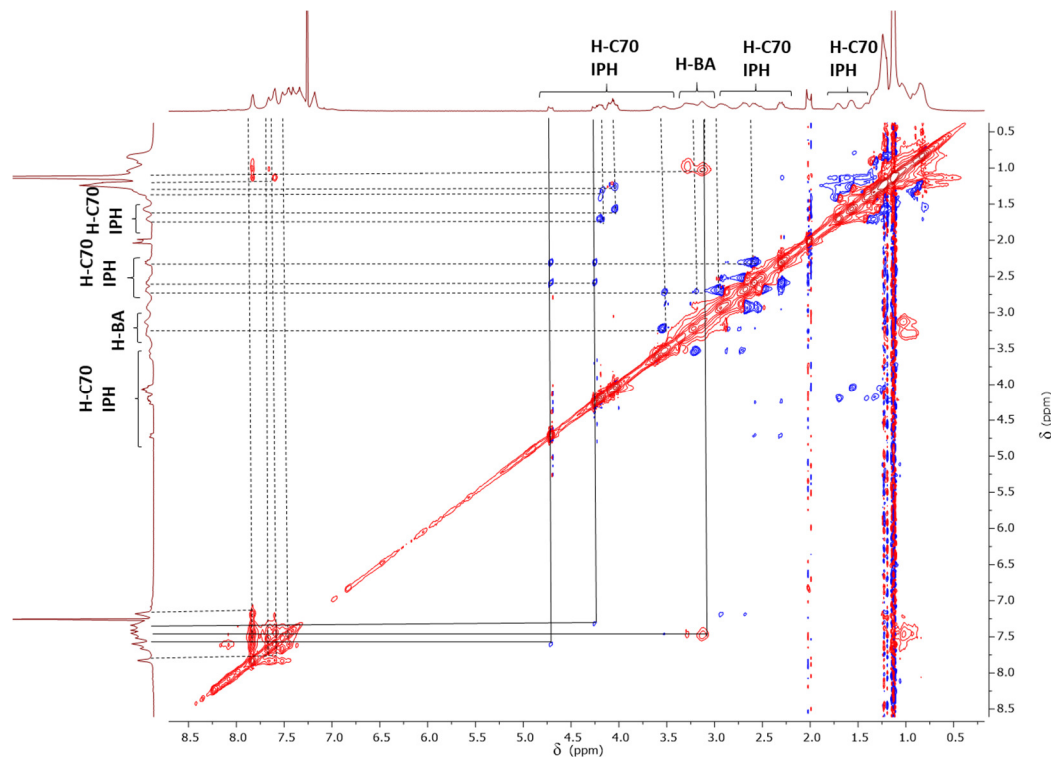
**Figure S 46:**  $^1\text{H}$ -NOESY NMR spectrum of  $\text{C}_{70}$ -IPH (600/600 MHz,  $[\text{D}_1]$ -chloroform/ $[\text{D}_4]$ -methanol = 8/2 (v/v), 298 K). Dashed lines indicate intramolecular interactions, while solid lines represent intermolecular interactions.



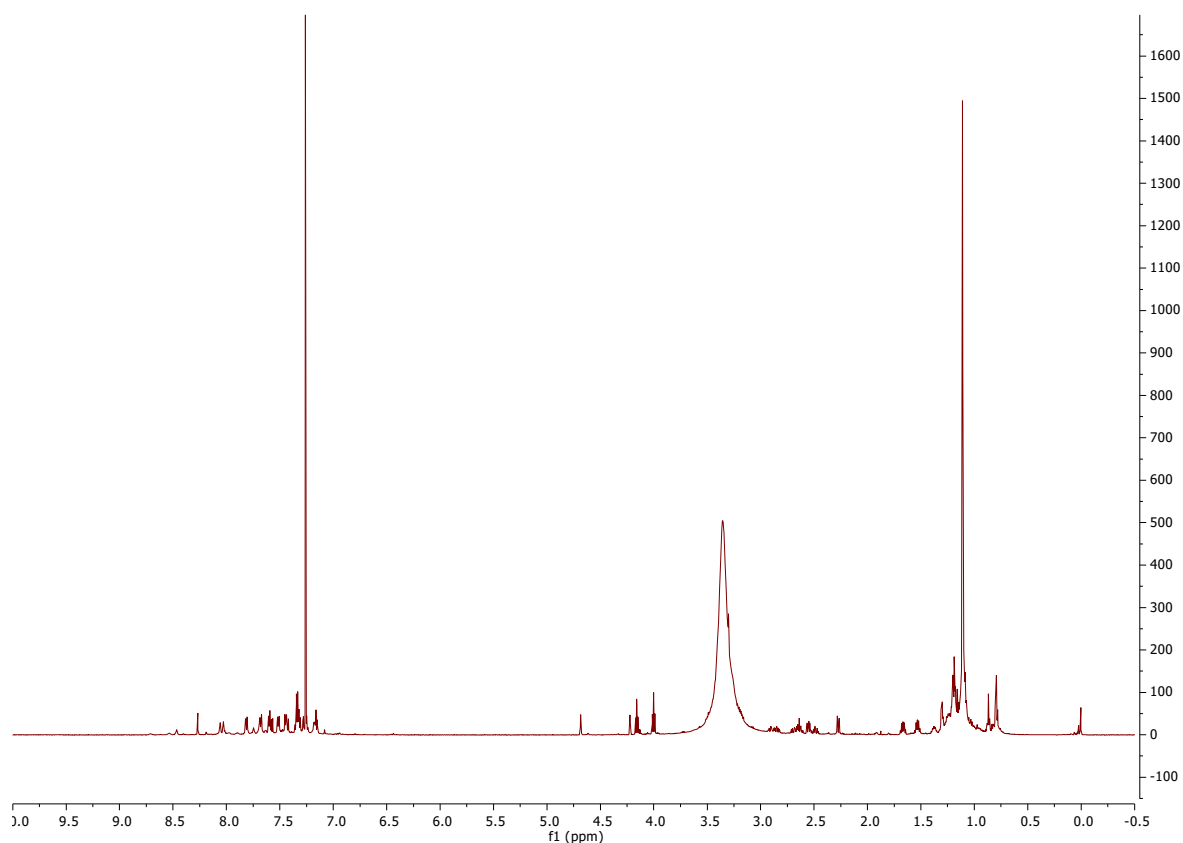
**Figure S 47:**  $^1\text{H}$ -NMR spectrum of a mixture of the [3+2]-capsule (*all-R*)-1 (4 mM) and  $\text{C}_{70}$ -IPH (ca. 10.6 mM by integration) (600 MHz,  $[\text{D}_1]$ -chloroform/ $[\text{D}_4]$ -methanol = 8/2 (v/v), 298 K).



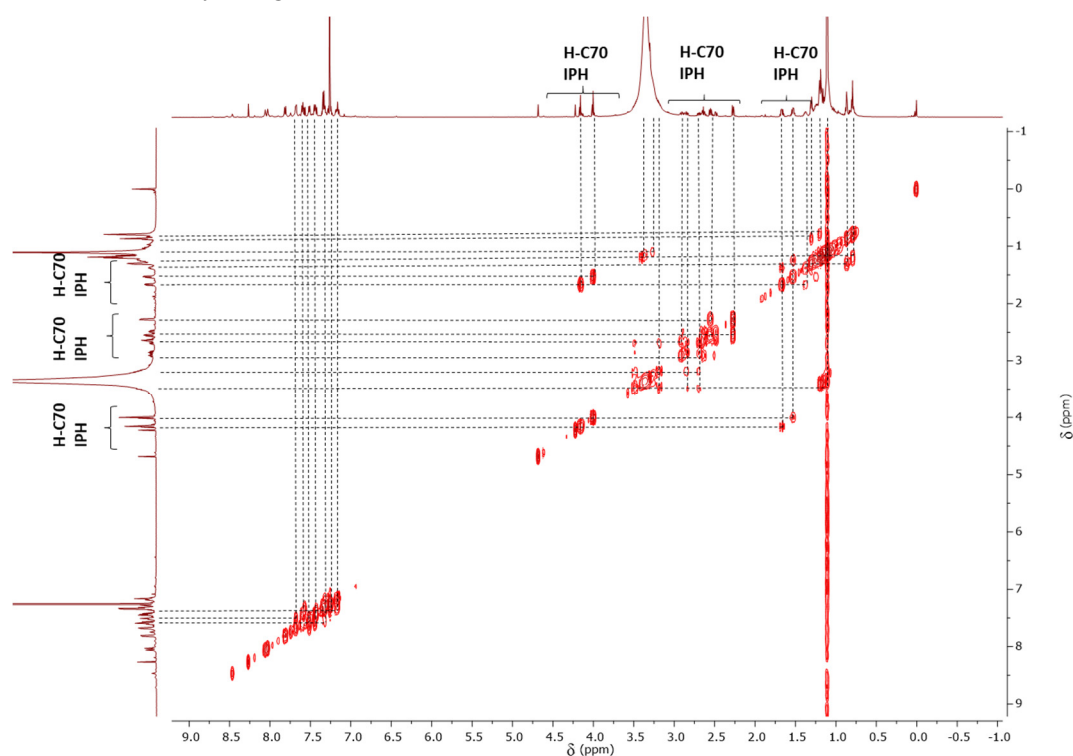
**Figure S 48:** <sup>1</sup>H-COSY NMR spectrum of a mixture of the [3+2]-capsule (*all-R*)-1 (4 mM) and C<sub>70</sub>-IPH (ca. 10.6 mM by integration) (600/600 MHz, [D<sub>1</sub>]-chloroform/[D<sub>4</sub>]-methanol = 8/2 (v/v), 298 K). Dashed lines indicate intramolecular interactions, while solid lines represent intermolecular interactions.



**Figure S 49:** <sup>1</sup>H-NOESY NMR spectrum of a mixture of the [3+2]-capsule (*all-R*)-1 (4 mM) and C<sub>70</sub>-IPH (ca. 10.6 mM by integration) (600/600 MHz, [D<sub>1</sub>]-chloroform/[D<sub>4</sub>]-methanol = 8/2 (v/v), 298 K). Dashed lines indicate intramolecular interactions, while solid lines represent intermolecular interactions.

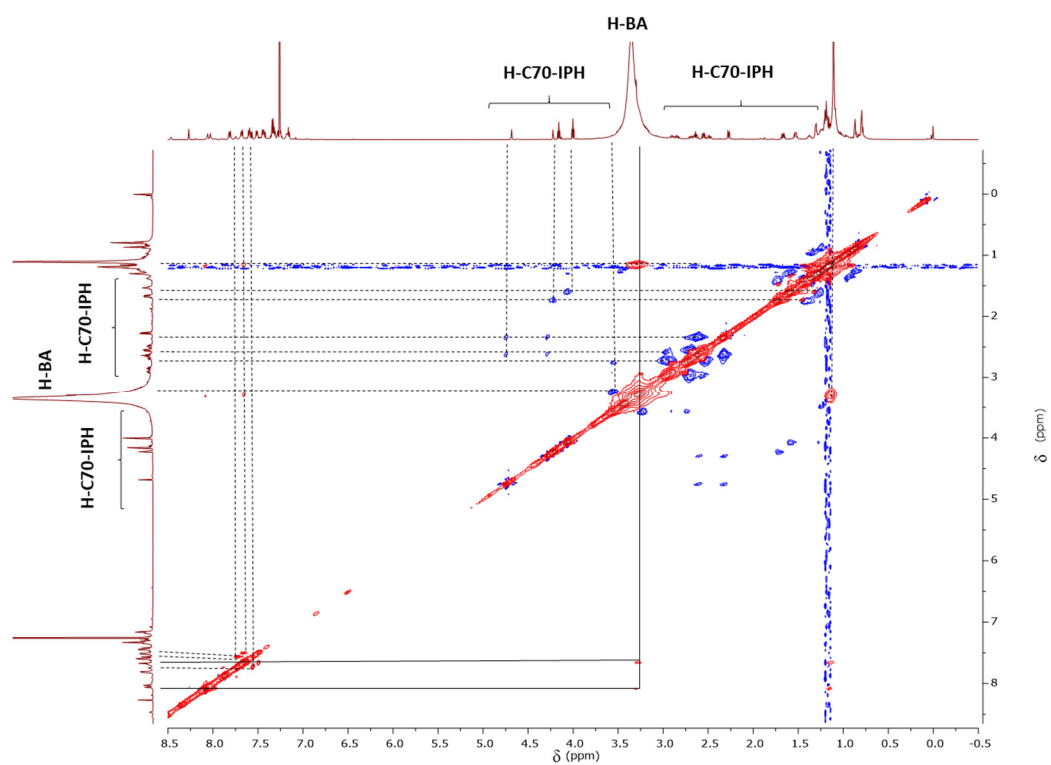


**Figure S 50:**  $^1\text{H}$ -NMR spectrum of a mixture of the [2+2]-capsule (*all-R*)-**2** (4 mM) and  $\text{C}_{70}$ -IPH (ca. 10.2 mM by integration) (600 MHz,  $[\text{D}_1]$ -chloroform/ $[\text{D}_4]$ -methanol = 8/2 (v/v), 298 K).



**Figure S 51:**  $^1\text{H}$ -COSY NMR spectrum a mixture of the [2+2]-capsule (*all-R*)-**2** (4 mM) and  $\text{C}_{70}$ -IPH (ca. 10.2 mM by integration) (600/600 MHz,  $[\text{D}_1]$ -chloroform/ $[\text{D}_4]$ -methanol = 8/2 (v/v), 298 K). Dashed lines indicate intramolecular interactions, while solid lines represent intermolecular interactions.





**Figure S 52:** <sup>1</sup>H-NOESY NMR spectrum a mixture of the [2+2]-capsule (*all-R*)-**2** (4 mM) and C<sub>70</sub>-IPH (ca. 10.2 mM by integration) (600/600 MHz, [D<sub>1</sub>]-chloroform/[D<sub>4</sub>]-methanol = 8/2 (v/v), 298 K). Dashed lines indicate intramolecular interactions, while solid lines represent intermolecular interactions.

## 6. References

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