

Supporting Information

Experimental section

Materials

Molybdenum sulfide (MoS_2) powder was purchased from Shanghai Aladdin Biochemical Technology Co., Ltd. Tungsten disulfide (WS_2) powder was purchased from Nanjing Xianfeng Nano Material Technology Co., Ltd. Adenosine, adenosine diphosphate (ADP), adenosine triphosphate (ATP), uridine, guanosine, thymidine and cytidine were purchased from Bioengineering (Shanghai) Co., Ltd. carboxyfluorescein (FAM)-labelled anti-adenosine aptamer (5'- FAM- AAC CTG GGG GAG TAT TGC GGA GGA AGGT- 3') were synthesized and purified by Sangon Biotech (Shanghai) Co., Ltd. Other chemicals were used as received without further purification. All experimental water used was ultrapure water (18.2 $\text{M}\Omega$ CM, Millipore).

Preparation of MoS_2 nanosheets

Experimentally, 200 mg MoS_2 powder was added into 40 mL aqueous solution containing 40 mg ATP. The mixed suspension was ultrasonic exfoliation for 30 h. After centrifugation at 6000 rpm for 20 min to remove large particles, the supernatant was collected. The collected supernatant was further centrifuged at 12000 rpm for 10 min, the supernatant was removed, and the sediment was re-dispersed in water. Similarly, ADP and adenosine were used to prepare MoS_2 nanosheets under the same procedure respectively. In addition, ATP was also used to exfoliate WS_2 powder by the same method.

Characterization

Transmission electron microscopy (TEM) image, selected area electron diffraction (SAED) pattern and high-resolution TEM image of as-exfoliated MoS_2 nanosheets were characterized by an FEI Tecnai G2F20 transmission electron microscope with the accelerating voltage of 200 kV. Before measurement, the dispersion of as-exfoliated MoS_2 nanosheets was diluted with ethanol, dropped onto the carbon film supported by the copper mesh, and naturally dried by air. Atomic

force microscopy (AFM) image of as-exfoliated MoS₂ nanosheets was characterized by a Bruker Multimode III scanning probe microscope. Before measurement, the dispersion solution of as-exfoliated MoS₂ nanosheets was diluted with ethanol, dropped onto the mica plate and dried by N₂. X-ray photon spectroscopy (XPS) spectra of MoS₂ nanosheets were characterized by a Thermo Scientific ESCALAB 250 X-ray photoelectron spectrometer. Before measurement, the dispersion solution of as-exfoliated MoS₂ nanosheets was concentrated by centrifugation, and then added dropwisely to the surface of the silicon wafer and air-dried. The X-ray diffraction (XRD) pattern of MoS₂ nanosheets was measured by an Ultima IV X-ray diffractometer. Before measurement, the dispersion solution of MoS₂ nanosheets was concentrated by centrifugation, dropped into the groove of a glass slide, and air-dried until the groove of the glass slide was filled with a thin layer of MoS₂ nanosheets. The particle size distribution and zeta potential of MoS₂ nanosheets were measured by dynamic light scattering (DLS) by using a Malvern zetasize nano-zs90 nanoparticle size potential analyzer. Before measurement, the dispersion solution of MoS₂ nanosheets was diluted with water.

Fluorescent biosensor for adenosine

40 μL FAM-labelled anti-adenosine aptamer (100 nmol L^{-1}) was mixed with 325 μL different concentrations of adenosine in Tris-HCl buffer (20 mmol L^{-1} , pH 7.4, 100 mmol L^{-1} NaCl, 5 mmol L^{-1} KCl, 5 mmol L^{-1} MgCl₂). The mixture was incubated in a metal bath at 25 °C for 20 min with a slight shaking (300 rpm). Then, 35 μL dispersion solution of MoS₂ nanosheets (0.24 mg mL^{-1}) was added, the mixture was further incubated for 5 min in a metal bath at 25 °C with a slight shaking (300 rpm). Finally, the fluorescent spectrum was recorded by a F4600 fluorospectrometer.

The pretreatment for human serum samples was as follows: 50 μL serum sample was mixed with 500 μL Ba(OH)₂ solution (0.08 mmol L^{-1}) and 500 μL ZnSO₄ solution (0.10 mmol L^{-1}), followed by vortexing for 10 s; the mixture was centrifuged at 8000 rpm for 10 min, and the supernatant was 10-fold diluted with tris-HCl buffer (20 mmol L^{-1} , pH 7.4, 100 mmol L^{-1} NaCl, 5 mmol L^{-1} KCl, 5 mmol L^{-1} MgCl₂) and spiked with different concentration of adenosine.

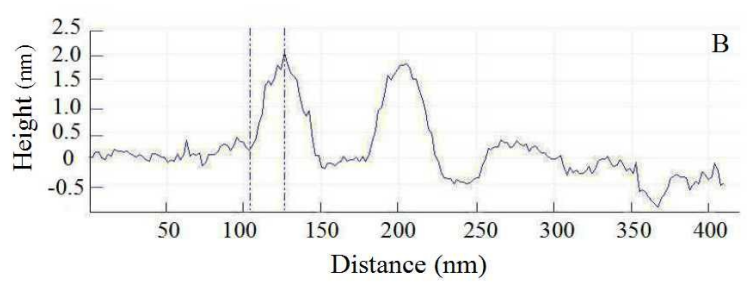
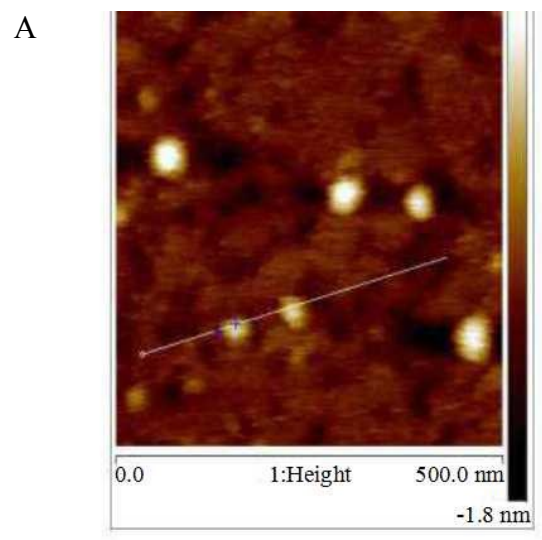


Figure S1. AFM image (A) and height profile (B) of as-exfoliated MoS₂ nanosheets

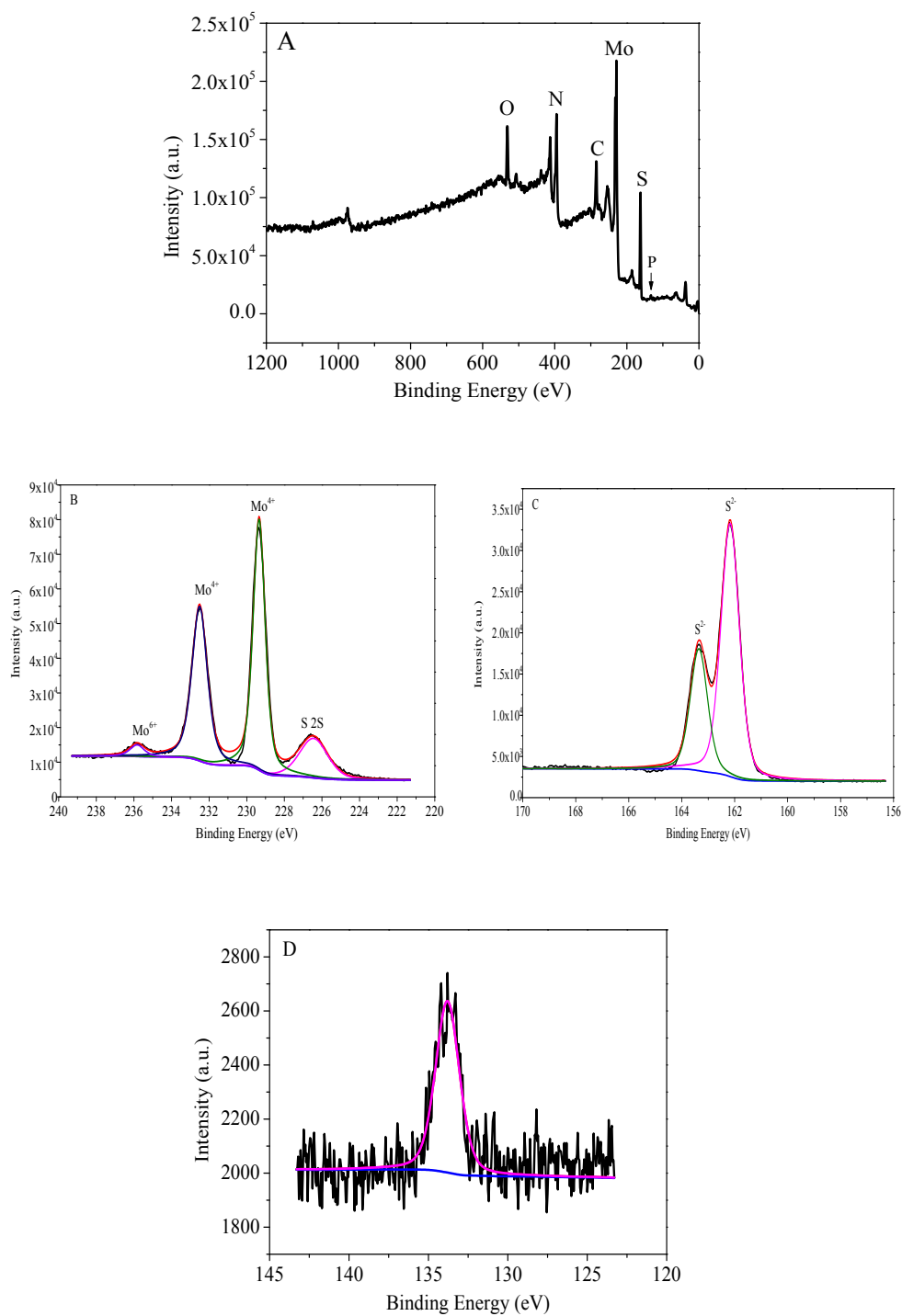


Figure S2. The survey XPS spectrum (A), Mo 3d (B), S 2P (C) and P 2P (D) core-level XPS of as-exfoliated MoS₂ nanosheets.

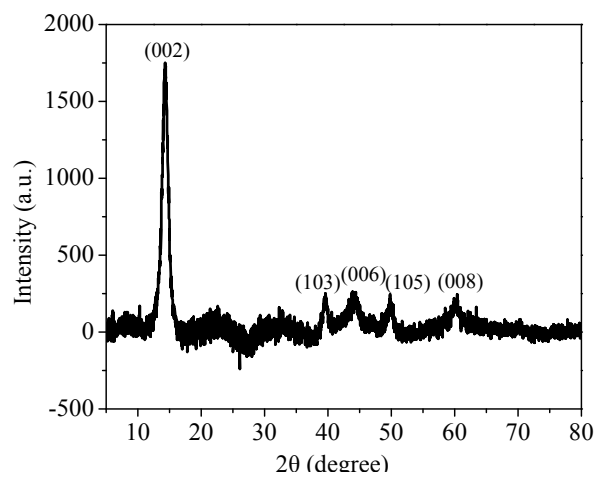


Figure S3. XRD pattern of as-exfoliated MoS₂ nanosheets

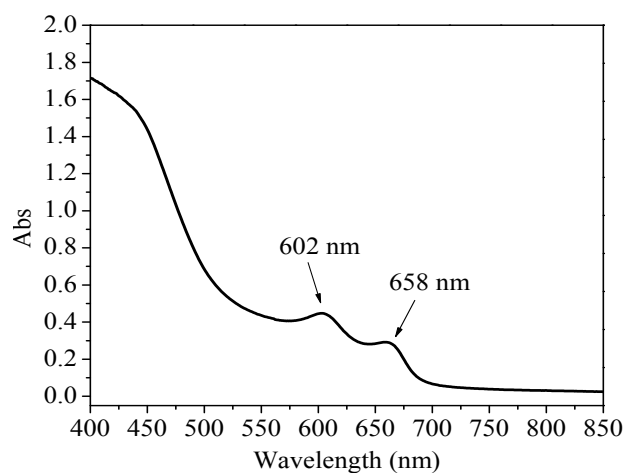


Figure S4. Absorption spectrum of dispersion solution of as-exfoliated MoS₂ nanosheets (0.28 mg mL⁻¹)

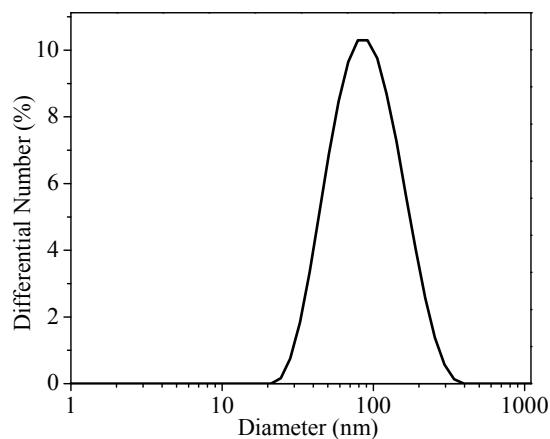


Figure S5. The size distribution of as-exfoliated MoS₂ nanosheets

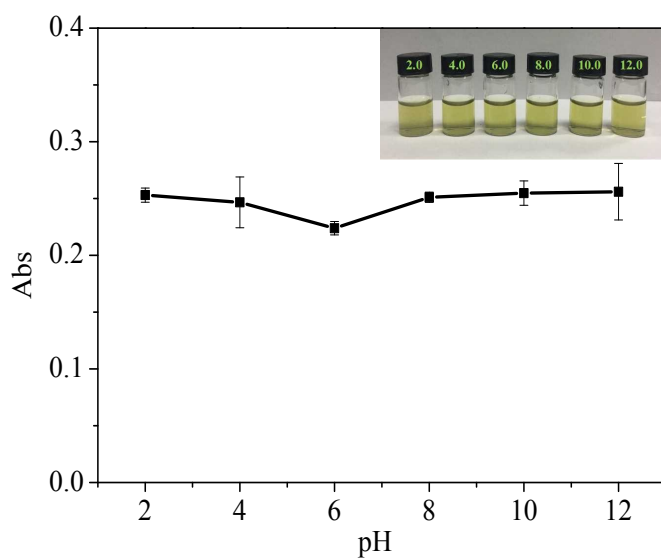


Figure S6. Absorption intensities at 658 nm of as-exfoliated MoS₂ nanosheets (0.2 mg mL⁻¹) in various pH phosphate buffer (10 mmol L⁻¹). Insert were the corresponding photos of as-exfoliated MoS₂ nanosheets in pH 2.0, pH 4.0, pH 6.0, pH 8.0, pH 10.0 and pH 12.0 phosphate buffers (10 mmol L⁻¹).

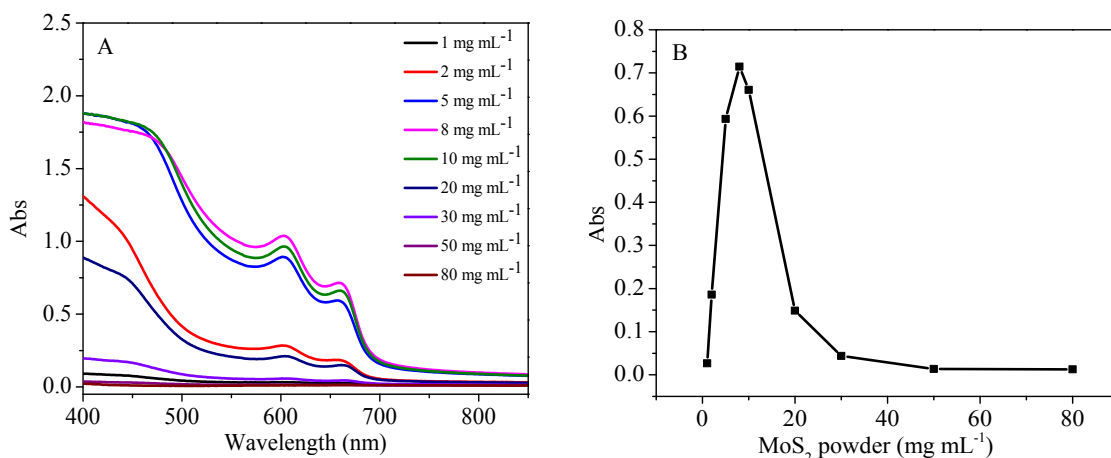


Figure S7. Absorption spectra of as-exfoliated MoS₂ nanosheets with different mass concentration of MoS₂ powder (A) and the relationship between absorption intensity of as-exfoliated MoS₂ nanosheets at 658 nm and mass concentration of MoS₂ powder (B). (ATP concentration was 1 mg mL⁻¹, ultrasonic time was 30 h.)

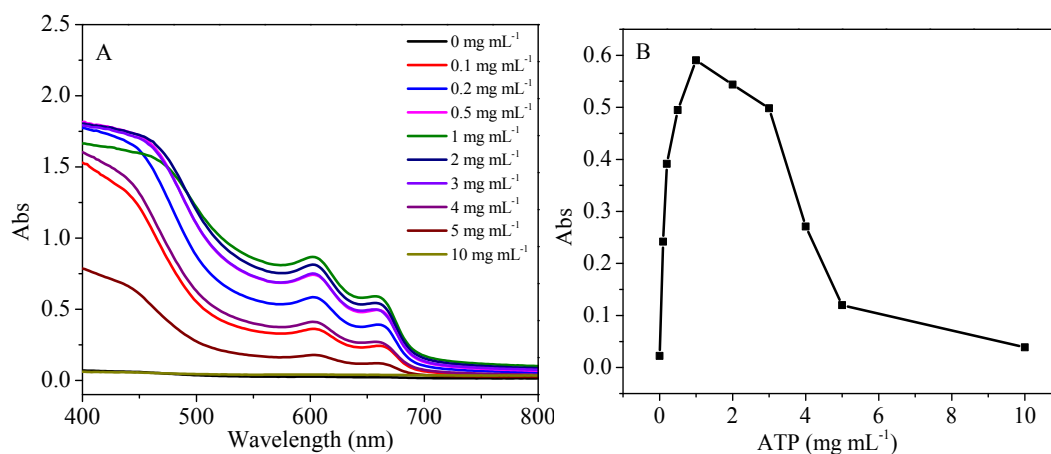


Figure S8. Absorption spectra of as-exfoliated MoS₂ nanosheets with different concentrations of ATP (A) and the relationship between absorption intensity of as-exfoliated MoS₂ nanosheets at 658 nm and ATP concentration. (B) (MoS₂ mass was 5 mg mL⁻¹, ultrasonic time was 30 h.)

Table S1. Comparison of exfoliation yield of MoS₂ nanosheets by direct ultrasonication in solvent

Solvent	Time	Yield	References
N-methyl-pyrrolidone	140 h	40%	S3
Ethanol/water	8 h	0.6%	S4
Ethanol	48 h	21%	S5
Water	48	6%	S5
Chloroform/acetonitrile	1 h	13.3%	S6
Water	30 h	2.3%	This work

Table S2. Comparison of aqueous ultrasonic exfoliation of TMDCs with the assistant reagents

Assistant reagents	Time	Yield	Application	References
High-molecular weight ssDNA	3 h	80%-90% (WS ₂ /WSe ₂)	antibacterial application	S7
Alkali lignin	80 h	17.5% (MoS ₂)	lithium ion battery	S8
Nanocellulose	4 h	18% (MoS ₂)	sodium ion battery	S9
Chitosan	5 h	25.5% (MoS ₂)	composites	S10
Sodium cholate	16 h	10% (MoS ₂)	lithium ion batteries	S11
Bovine serum albumin	35 h	27.2 % (MoS ₂)	adsorption and capacitance	S12
Gelatin	8 h	1.6% (MoS ₂)	composites	S13
Gelatin	8 h	1.8% (MoS ₂)	composites	S13
Chitin nanofibrils	48 h	13.2% (WS ₂)	supercapacitors and photo-thermal converter	S14
Thermoresponsive polymeric ionic liquid	18 h	24.8% (MoS ₂)	highly thermally conductive films	S15
Tannic acid	2 h	60.5% (MoS ₂)	photothermal nanocarriers	S16
Nafion	8 h	8.8% (MoS ₂)	memory membrane	S17
Poly(3,4-ethylenedioxythiophene):poly(styrenesulfonate)	50 h	1.6% (MoS ₂)	organic solar cells	S18
Bovine serum albumin	10	15.4% (WSe ₂)	photodynamic/photothermal therapy	S19
ATP	30 h	23.6% (MoS ₂)	biosensor	This work

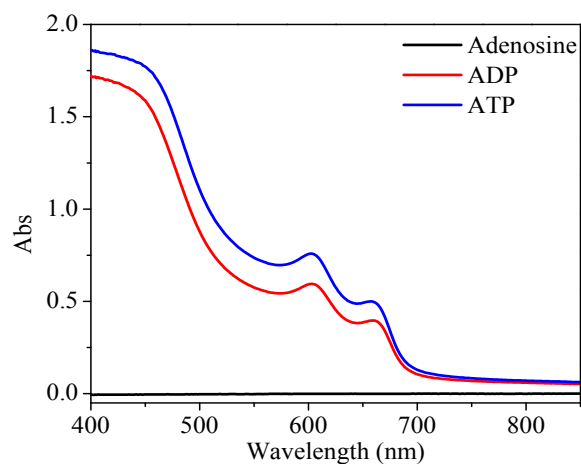
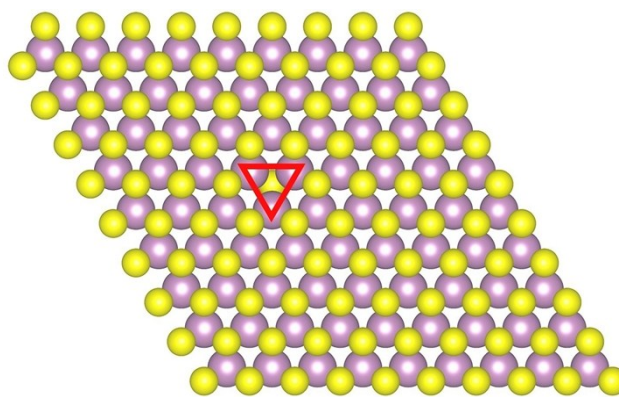


Figure S9. Absorption spectra of dispersion solutions of as-exfoliated MoS₂ nanosheets obtained from ultrasonic exfoliation with adenosine, ADP and ATP as the exfoliation reagent, respectively. 200 mg MoS₂ powder was added into 40 mL aqueous solution containing 40 mg ATP, ADP or adenosine, respectively and was ultrasonically exfoliated for 30 h.

Theoretical calculation of MoS₂-ATP

To study the adsorption of ATP on MoS₂ nanosheets, first-principles calculations based on density functional theory (DFT) were carried out by using the Vienna ab initio simulation package^[S1] (VASP) with the projected augmented wave (PAW) method. The generalized gradient approximation Perdew-Burke-Ernzerhof (PBE) exchange-correlation functions were employed. For the calculations of binding energy, the first Brillouin zone was sampled with a 1×1×1 Monkhorst-Pack grid. All the structures were fully relaxed with residual forces smaller than 0.5 eV/Å. In addition, effect of vdW interaction was accounted for by using the dispersion corrected DFT (vdW-DF2 functional).^[S2] The vacuum length was longer than 20 Å to decouple interaction between adjacent layers. The adsorption energy ($E(a)$) of ATP molecule on defective MoS₂ layer was calculated as $E(a) = E(\text{MoS}_x\text{-molecule}) - E(\text{MoS}_x) - E(\text{molecule})$, where $E(\text{molecule})$, $E(\text{MoS}_x)$, and $E(\text{MoS}_x\text{-molecule})$ were the energies of ATP molecule, the defective MoS₂ layer, and the complex of defective MoS₂ and ATP molecule, respectively. In this work, the defective MoS₂ monolayer was constructed by S-Mo-S three atom layers with a large rhombohedral supercell that contains 81 Mo and 161 S atoms. On a side of S layers, one S atom was defected (see Figure S10 (a)). To simulate the ATP-doped MoS₂ synthesized experimentally, an ATP molecule was introduced in the above supercell, and terminal O atom of terminal phosphate in the ATP was set in the S defect in the MoS₂ monolayer, the corresponding stoichiometry was Mo₁₆₁S₈₁C₁₀H₁₆N₅O₁₃P₃ (see Figure S10 (b-c)).



(a)

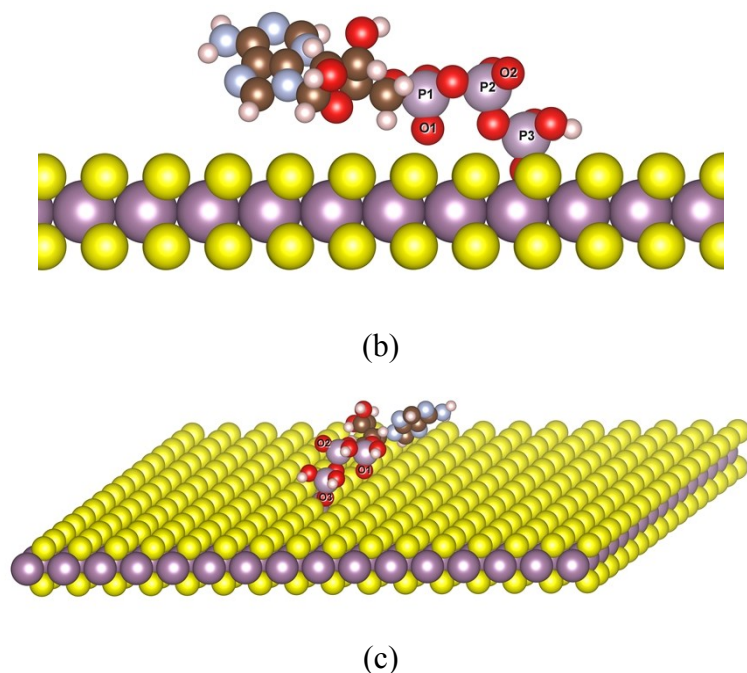


Figure S10. The top views of defective MoS₂ (a) and the side views (b) (c) of ATP on the defective MoS₂ monolayer

Results of theoretical calculations

To study the adsorption of ATP on MoS₂ nanosheets, we simulated the adsorption energy of ATP molecule on MoS₂ monolayer by using the density of function (DFT). The results of structural optimizations show that ATP molecule can be attached to MoS₂ monolayer through the formation of three Mo-O bonds (one strong binding within 2.40 Å bond length and two vdW binding with 2.85 and 2.89 Å). The adsorption energy of ATP on MoS₂ monolayer is -0.34 eV. Namely, it shows that ATP molecule can stably adsorb on the defect MoS₂ nanosheets, which are agreement with experiment. As displayed in Figure S10, it is clear that the terminal O atom of terminal phosphate in the ATP molecule tends to take up the S defect position, then bonds to Mo atoms. After adsorption, the double bond of P-O is broken. The length of new formed P-O bond is about 1.50 Å, 0.04 Å larger than the former double bond. Besides, two other terminal O atoms in the ATP molecule behave different. One has no changes due to far from S atoms layer after adsorbing and the other is repelled by S atoms layer, which results from perpendicularly closer to S atoms layer after

adsorbing. This can be proved by the bond lengths of P-O in Table S2. Thus, it shows that ATP will be easy to adsorb on the S atomic vacancy defect of MoS₂ monolayer, and it may be hard to adsorb in the pure MoS₂ monolayer.

Table S3 The bond length of P-O in the ATP molecule.

Bond length (Å)	P1-O1	P2-O2	P3-O3
ATP molecule	1.47	1.46	1.46
ATP on MoS ₂ monolayer	1.40	1.46	1.50

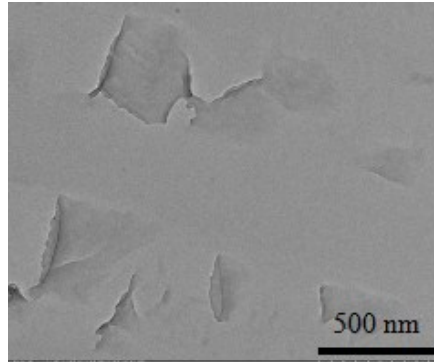


Figure S11. TEM image of as-exfoliated WS₂ nanosheets

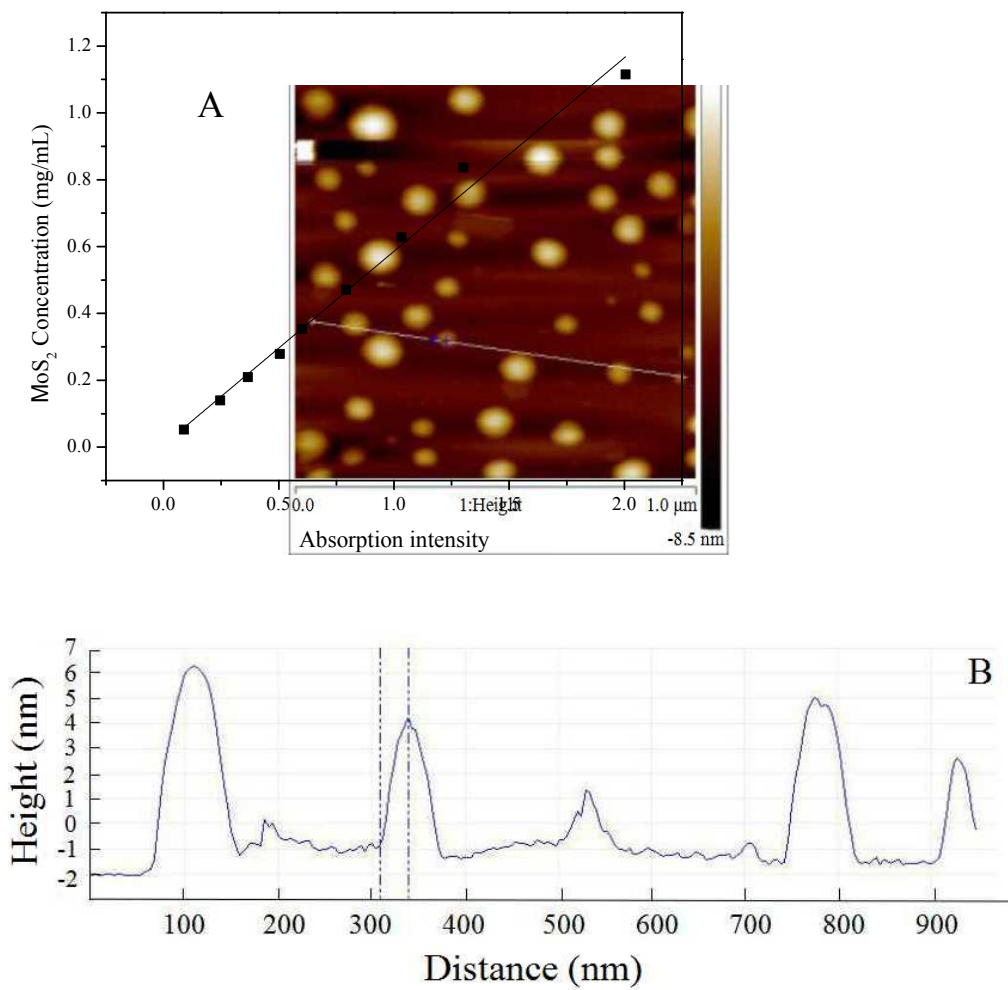


Figure S12. AFM image (A) and height profile (B) of the as-exfoliated WS₂ nanosheets

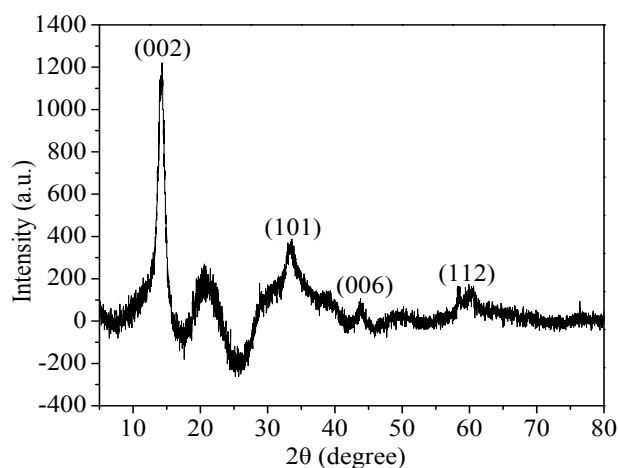


Figure S13. XRD image of as-exfoliated WS₂ nanosheets

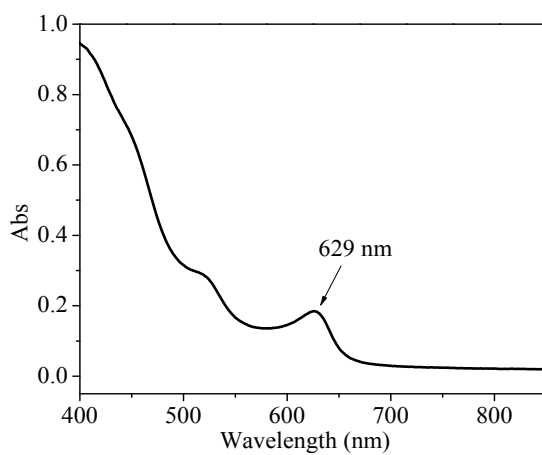


Figure S14. Absorption spectrum of as-exfoliated WS₂ nanosheets dispersion

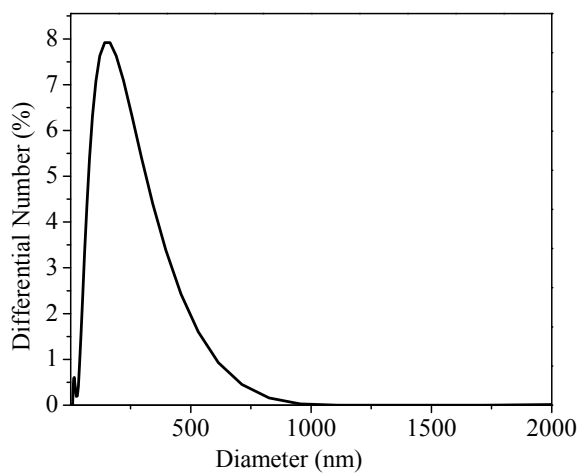
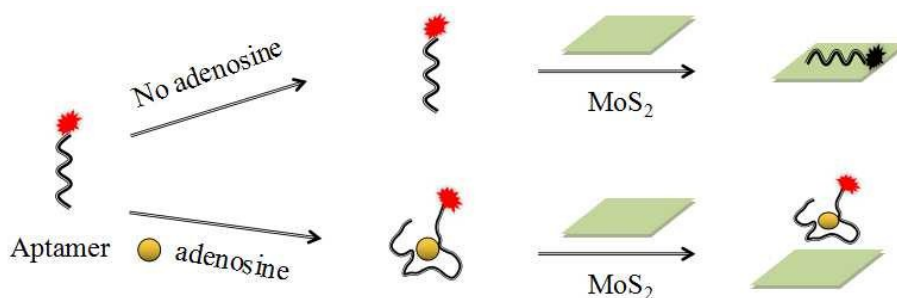


Figure S15. The size distribution of as-exfoliated WS₂ nanosheets



Scheme S1. The fluorescent biosensor for adenosine based on as-exfoliated MoS₂ nanosheets as the quenching platform.

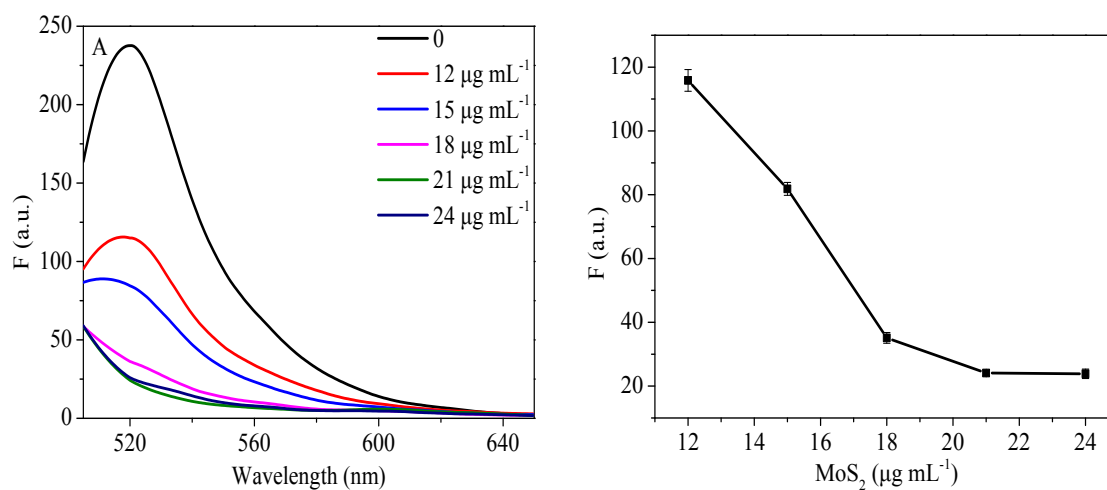


Figure S16. Fluorescence spectra (A) of FAM-labelled anti-adenosine aptamer (100 nmol L⁻¹) in the presence of different concentrations of as-exfoliated MoS₂ nanosheets; (B) Effect of as-exfoliated MoS₂ nanosheets concentration on the fluorescence intensity of FAM-labelled anti-adenosine aptamer. Excitation and emission wavelengths were 495 and 520 nm, respectively.

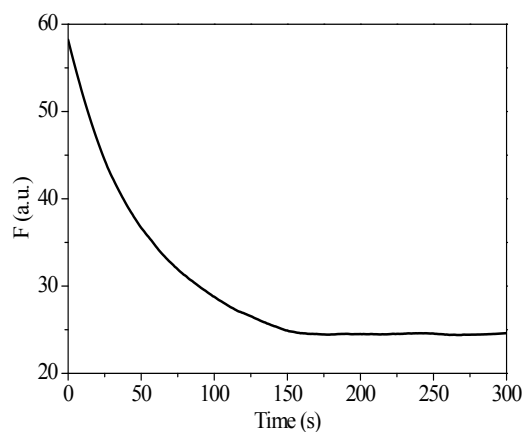


Figure S17. Fluorescence kinetic curve of FAM-labelled anti-adenosine aptamer (100 nmol L^{-1}) on addition of $21 \text{ }\mu\text{g mL}^{-1}$ as-exfoliated MoS_2 nanosheets. Excitation and emission wavelengths were 495 nm and 520 nm, respectively.

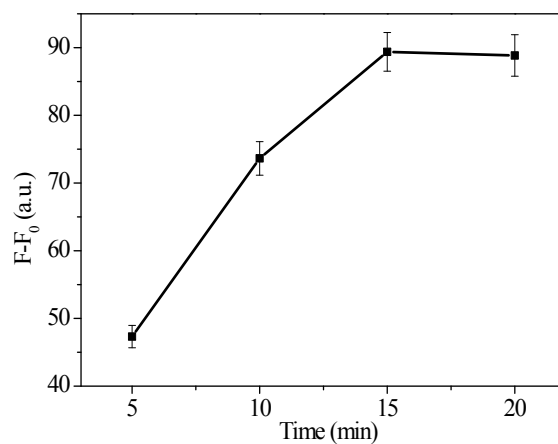


Figure S18. Effect of incubation time on the fluorescent enhancement upon addition of 1 mmol L^{-1} adenosine into the mixture of 100 nmol L^{-1} FAM-labelled anti-adenosine aptamer with $21 \text{ }\mu\text{g mL}^{-1}$ as-exfoliated MoS_2 nanosheets. Excitation and emission wavelengths were 495 nm and 520 nm, respectively.

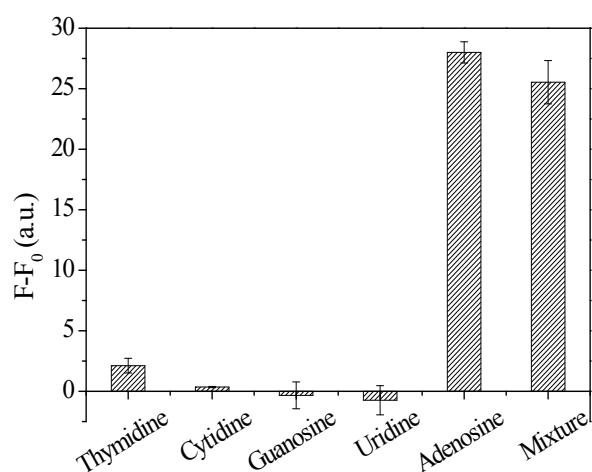


Figure S19. The fluorescent enhancement of this biosensor on addition of $50 \mu\text{mol L}^{-1}$ adenosine, $250 \mu\text{mol L}^{-1}$ thymidine, $250 \mu\text{mol L}^{-1}$ cytidine, $250 \mu\text{mol L}^{-1}$ guanosine, $250 \mu\text{mol L}^{-1}$ uridine, and the mixture of $50 \mu\text{mol L}^{-1}$ thymidine, $50 \mu\text{mol L}^{-1}$ cytidine, $50 \mu\text{mol L}^{-1}$ guanosine, $50 \mu\text{mol L}^{-1}$ uridine, and $50 \mu\text{mol L}^{-1}$ adenosine.

Table S4. Recoveries for the detection of spiked adenosine in human serum samples

Samples	Added ($\mu\text{mol L}^{-1}$)	Found mean \pm SD ($\mu\text{mol L}^{-1}$, n=3)	Recovery (%)
1	10.0	8.6 ± 2.6	86.0
2	25.0	24.3 ± 7.2	97.2
3	40.0	36.8 ± 5.9	92.0

References

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