Supplementary material

Significantly increased catalytic activity of Candida antarctica Lipase B for the

resolution of *cis*-(±)-dimethyl 1-acetylpiperidine-2,3-dicarboxylate

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Library	Oligonucleotide sequences
A-f:	CTTATGGCCTTTGCTCCT NDT TACAAAGGT NDT GTGTTGGCTGGTCCA
B-f:	CCGTGTTGGCTGGTCCANDTGACGCCTTGGCANDTTCCGCTCCTTCCGTC
C-f:	CAGTGTCCGCTCCTTCCNDTTGGCAANDTACCACGGGTTCAGCT
D-f:	TACTCAGCTACAGACGAA NDTNDT CAGCCTCAAGTTAGTA
A-D-r:	GAGAACTGTGATGTCAAAGATCCTGCATGATCGATAAC
E-f:	GCAGCCCTGCTGGCTCCANDTNDTGCCGCTATCGTTGCTG
E-r:	GTTCTAGATCAATGATGATGATGGTGGTGAGGAGTAA
L278-f:	AAGGTCGCTGCCGCAGCCCTGNNKGCTCCAGCAGCTGCCGCTATCG
L278-r:	CGATAGCGGCAGCTGCTGGAGCMNNCAGGGCTGCGGCAGCGACCTT
I285-f:	CTGGCTCCAGCAGCTGCCGCTNNKGTTGCTGGTCCAAAACAAAATT
I285-r:	AATTTTGTTTTGGACCAGCAACMNNAGCGGCAGCTGCTGGAGCCAG

Table S1 Primers for the combinatorial active-site saturation test and site-saturation

mutagenesis

Table S2 Comparison of mut-I189K with other reported counterparts for resolution of

cis-(±)-1.

Enzyme	Substrate conc. (g L ⁻¹)	Catalyst load (g L ⁻¹)	Time (h)	Conv (%)	Ε	S/C (g/g)	Reference
Immobilized CALB	80	40 ^a	140	>49.9%	>2000	2	Riccardo et al., 2014
Addzyme CALB	80	40 ^b	16	>49.9%	>80	2	Ramesh et al., 2014
mut-I189K	243.26	0.1°	5	>49.9%	>200	2432.6	This study
mut-I189K	486.52	0.8°	8	>49.9%	>200	608.15	This study

^a The Immobilized CALB used as catalyst.

^b The CALB enzyme solution (activity 5000 TBU, obtained from Advanced Enzyme Technologies Ltd.,) used as catalyst.

^c The purified enzyme used as catalyst.

Reference

M. Riccardo, A. Giancarlo, B. Elisabetta, C. Andrea, F. Stefano and G. Marco, US

Pat., 8680276B2, 2014.

P. Ramesh, T. Harini and N. W. Fadnavis, Org Process Res & Dev., 2014, 19, 296-

301.

Fig. S1. CALB mediated resolution of $cis-(\pm)$ -dimethyl 1-acetylpiperidine-2,3-dicarboxylate.



Fig. S2. SDS-PAGE analysis of expressed and purified proteins. (a) lane M: protein marker; lane 1, 2: supernatant extract and insoluble fraction of *E. coli* Rosetta (DE3)/ pET22b-CALB without induced; lane 3, 4: supernatant extract and insoluble fraction of *E. coli* Rosetta (DE3)/ pET22b-CALB induced by 0.1 mM IPTG. (b) lane M: protein marker; lane 1-7: the whole cell of wild-type CALB, mut-I189K, mut-I189R, mut-I189A, mut-I189N/V190L, mut-I189Y/V190L and mut-I189H/V190L. (c) lane M: protein marker; lane1: the whole cell of wild-type CALB; lane 2-7: the whole cell



Fig. S3. The colorimetric high-throughput method for the screening of positive mutants. The 96-well plate with different colors in the wells containing different mutants, the wells of A1-A3 which containing wild-type CALB was used as the control. The wells with the color closer to yellow indicate the corresponding mutants with higher hydrolytic activity.

Fig. S4. Effects of temperatures on the thermostabilities of wild-type CALB and mut-I189K.

Fig. S5. Enzymatic resolution of *cis*-(\pm)-1 at 2M, employing mut-I189K (0.8 g L⁻¹). The reaction was conducted at 35 °C, and the pH was maintained at 6.0 by automatically titrating 4 M NaOH.

Fig. S6. Enzymatic resolution of *cis*-(\pm)-**1** at 1 M, adding product 0 M (black squares), 0.5 M (red dots) and 1.0 M (blue triangles) at a catalyst load of 0.1 g L⁻¹ of purified mut-I189K. The reaction was conducted at 35 °C, and the pH was maintained at 6.0 by automatically titrating 4 M NaOH.

Fig. S7. (2R,3S)-1 in the active site of wild-type CALB (a) and mutants mut-I189K, mut-I189A, mut-I189N/V190L (b, c, d), as predicted from the docking analysis. H bonds for interaction and the stabilization of the oxyanion indicated by green dotted lines. The distances for effective nucleophilic attack of serine in the catalytic triad at the carbonyl function of (2R,3S)-1 are labeled as yellow dotted lines.

