

Supplementary Information

Table.S1 all of the PCR primers used for construction of mutants

Round	Primer name	Nucleotide sequence
First-round	L127-F	GTGTTAC <u>CNNNT</u> GGACCTCTTCTCTGG
	L127-R	GAGGTCC <u>ANNNG</u> GTAACACCAGCAAC
	T129-F	CCCTGTGG <u>NNNT</u> CTTCTCTGGTTA
	T129-R	CAGAGAAG <u>ANNN</u> CCACAGGGTAAC
	L244-F	CTGTCTCGT <u>NNN</u> CTGAAACCGG
	L244-R	GTTTCAG <u>NNN</u> ACGAGACAGGTCAG
	A250-F	GAAAAC <u>NNNG</u> GTAACCACTACCCG
	A250-R	GTTAC <u>CNNNG</u> TTTTCCGGTTTCAG
	L245-F	CGTGGT <u>NNN</u> AAACCGGAAAAC
	L245-R	CGGTTT <u>NNN</u> ACCACGAGACAG
Second-round	L127I-F	GGTGTAC <u>CATTT</u> GGACCTCTTCTCTGG
	L127I-R	GAGGTCC <u>AAATG</u> GTAACACCAGCAAC
	L244G-F	GTCTCGT <u>GGT</u> CTGAAACCGG
	L244G-R	GTTTCAG <u>ACC</u> ACGAGACAGGTC
	A250G-F	GGAAAAC <u>CGGT</u> GGTAACCACTAC
	A250G-R	GTTACC <u>ACCG</u> TTTTCCGGTTTC
Third-round	L245-F	CGTGGT <u>NNN</u> AAACCGGAAAAC
	L245-R	CGGTTT <u>NNN</u> ACCACGAGACAG
	L245R-F	CGTGGT <u>CCG</u> AAACCGGAAAAC
	L245R-R	CGGTTT <u>CGG</u> ACCACGAGACAG

Table S2 Comparison of Asymmetric reduction of OPBA to (R)-HPBA through bio-reduction with the reported works

Enzyme	Product (mM)	Productivity (mM·h ⁻¹)	<i>ee_p</i> (%)	references
D-LDH from <i>Staphylococcus epiderimidis</i> and FDH	182	38.2	>99	[3]
D-LDH (EC 1.1.1.28) and FDH	56.7	49.9	No report	[26]
YiaE from <i>E. Coli</i> K12 and GDH	100	4.2	98	[27]
LDH from <i>Lactobacillus bulgaricus</i> and FDH	71.8	47.9	>99	[28]
D-MDH from <i>Enterococcus faecali</i> and GDH	224	85.7	>99	This study

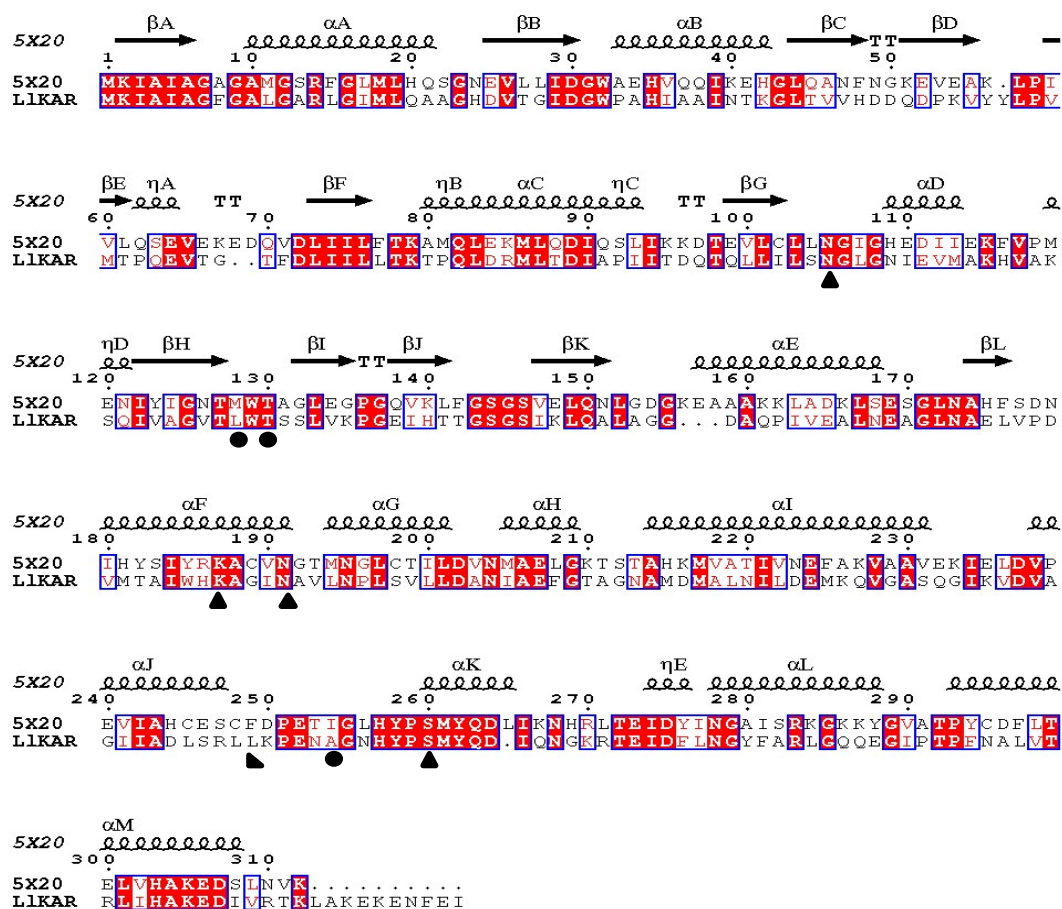


Figure S1. Structure-related sequence alignment between *L/KAR* and its close homologue: D-mandelate dehydrogenases (D-MDH) from *Leuconostoc lactis*. The homologue of *L/KAR*, D-mandelate dehydrogenases (D-MDH) with a PDB code of 5X20, 3WFI and 3WFJ, was identified by performing a BLASTP search, and the alignment was carried out using the program ClustalW. The sequence alignment was subsequently visualized using ESPrpt 3.2. Above the alignments are elements of the secondary structure of 5X20. The numbering shown is from 5X20. Triangles, critical coenzyme binding residues; stars, putative catalytic residues located at the corresponding positions of the conserved catalytic residues Asn104, Asn183, and Glu263. Strictly conserved residues are highlighted with red boxes.

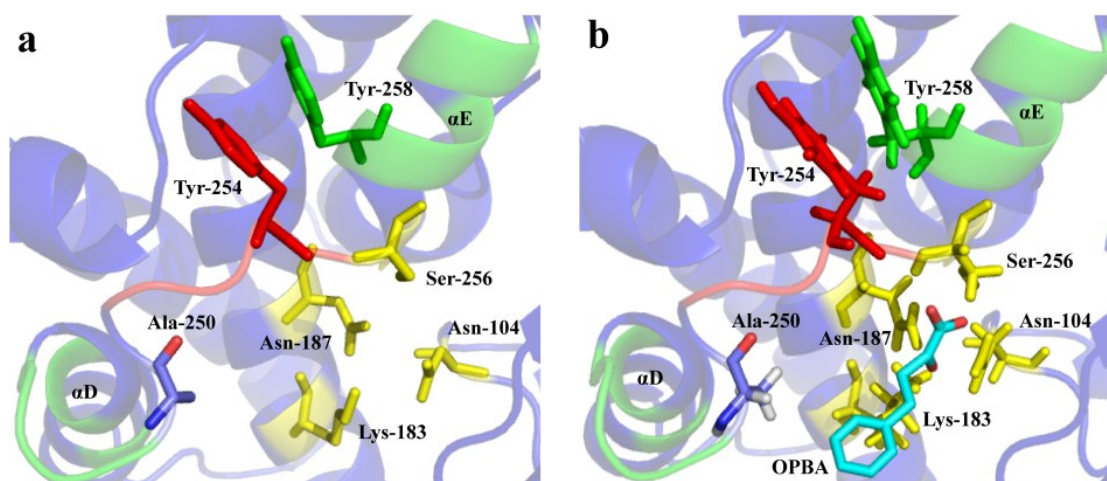


Figure S2. The clear spatial structure of the *L/KAR* catalytic and binding pocket. (a) The catalytic tetrad, Asn187, Lys183, and Ser256 are shown in yellow sticks. And the hydrophilic domains are shown in green and red cartoon; (b) the clear Spatial structure-activity relationship between α -helices and loop α D- α E. OPBA are shown as cyan sticks. All of the α -helices, β -sheets and loops are shown in blue cartoon.

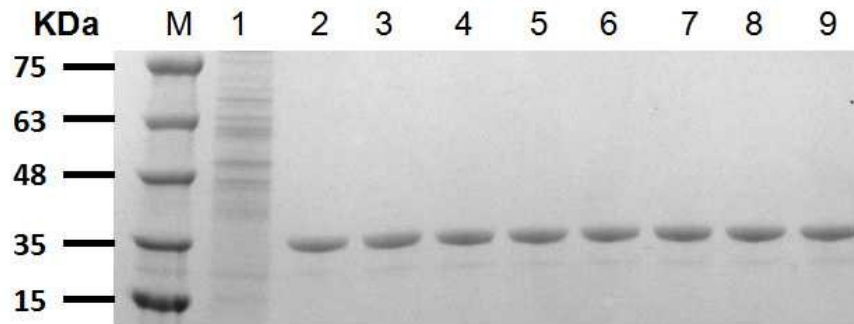


Figure S3 SDS-PAGE analysis of the wild-type *L/KAR* and its mutants. 10 μ L sample which contained approximately 40 μ g protein was loaded for each lane. Lane M: protein marker; Lane 1: the supernatant of the recombination *E.coli* BL21 (harboring pET-28b); Lane 2: the purified wild-type *L/KAR*; Lane 3: the purified mut-L127I; Lane 4: the purified mut-L244G; Lane 5: the purified mut-A250G; Lane 6: the purified mut-L127I/L244G; Lane 7: the purified mut-L127I/A250G; Lane 8: the purified mut-L244G/A250G; Lane 9: the purified mut-L244G/A250G/L245R.

References

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27. H. Yun, H. L. Choi, N. W. Fadnavis and B. G. Kim, *Biotechnology Progress*, 2005, **21**, 366-371.
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