Supporting Information

Optical Readout of Controlled Monomer-Dimer Self-Assembly

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General Information

UV-vis spectra were recorded on a Hitachi U-2900 spectrophotometer using quartz cuvettes (10×10 mm) with the samples dissolved in pyridine, DMSO, *o*-DCB, unless otherwise specified. ¹H and ¹³C NMR spectra were recorded on Bruker Avance 500 (500 and 125 MHz, respectively) with the samples dissolved in pyridine-*d*₅ or DMSO-*d*₆. Chemical shifts are given in parts per million (ppm) relative to TMS. Matrix-assisted laser desorption/ionization time-of-flight (MALDI-TOF) mass spectra were acquired on a VISION-2000 mass spectrometer with α -cyano-4-hydroxycinnamic acid (CHCA) as the matrix. Size exclusion column chromatography was carried out using Bio-Beads S-X1 resin (BIORAD) eluted with pyridine. All other reagents and solvents were obtained or distilled according to standard procedures. All reactions were controlled by TLC and UV-vis until complete disappearance of the starting materials, unless otherwise specified.

Electrochemical measurements were carried out in pyridine using a conventional three-electrode cell and Autolab/PGSTAT101 (Metrohm Autolab B.V., the Netherlands). A double junction Ag/AgCl reference electrode (Metrohm, Switzerland) was filled with 2 M LiCl in ethanol as the reference (inner) electrolyte and 0.1 M [TBA][BF₄] in pyridine as the bridge electrolyte. Before making any measurements, a platinum disk working electrode (2 mm in diameter, BASi, USA) was regenerated by polishing with an abrasive paper (grit 400), washed with ultrapure water and drying. Blank voltammograms were recorded in pure pyridine containing 0.1 M [TBA][BF₄] and then approximately 1 mM of a sample was introduced into the cell. After a series of measurements, ferrocene was added as internal reference, and additional voltammograms were recorded. Before the measurements the solutions were purged with N₂ for at least 30 min. All measurements were performed at room temperature ($22 \pm 2^{\circ}C$).

Synthesis

General procedure for the preparation of complexes 2a,b. Mg metal (50 mg) was boiled in isoamyl alcohol (50 mL) in the presence of a catalytic amount of I_2 until complete dissolution was observed (ca. 3 h). The solution of the resulting Mg alkoxide was cooled to room temperature, 1,4-diazepine-2,3-dicarbonitrile 1a or 1b (1 mmol) was added, and the mixture was heated at reflux for 16 h. The course of the reaction was monitored by UV-vis spectroscopy. Then, the reaction mixture was cooled to room temperature and the solvent was evaporated under reduced pressure. The resulting dry residue was successively washed with 50% aqueous acetic acid (4 × 50 mL), a 5% aqueous solution of sodium bicarbonate (2 × 50 mL), distilled water (4 × 50 mL) and finally with MeOH (50 mL) followed by drying *in vacuo* at 50 °C. The resulting solid was subjected to gel permeation chromatography (Bio-Beads S-X1, pyridine). This yielded 2a and 2b in the form of dark-green solids.

Tetrakis{**5,7-bis**[**2'-(4-bromophenyl**)**ethenyl**]-**6H-1,4-diazepino**}[**2,3-***b*,*g*,*l*,*q*]**porphyrazinato magnesium** (**II**) (**2a**). Yield 0.44 g (86%). UV-vis (DMSO) λ_{max} /nm (log ε): 387 (4.77), 659 (4.59), 700 (4.41). ¹H NMR (500 MHz; DMSO-*d*₆) δ/ppm: 8.20 (8H, d, ³*J* = 15.9 Hz, 2'-H), 7.7–7.65 (24H, br m, 1'-H; H^{o-Ar}), 7.47 (16H, br s, H^{m-Ar}), 6.11 (4H, br s, H^{eq}), 4.77 (4H, br s, H^{ax}). ¹³C NMR (125 MHz; Py-d₅) δ/ppm: 154.99, 149.19, 143.05, 137.46, 132.50, 131.68, 129.82, 37.75. MS (MALDI-TOF; HCCA): *m/z* 2050.87 [*M*+H]⁺, 4101.46 [*M*₂+H]⁺; calculated for C₉₂H₅₇Br₈MgN₁₆: 2050.82.

$Tetrakis \{5,7-bis [2'-(4-methoxyphenyl) ethenyl] - 6H - 1, 4-diazepino \} [2,3-b,g,l,q] porphyrazina to the statemethold of t$

magnesium(**II**) (**2b**). Yield 0.39 g (94%). UV-vis (DMSO) λ_{max}/nm (log ε): 386 (5.05), 661 (4.87), 698 (4.78). ¹H NMR (500 MHz; pyridine- d_5) δ/ppm: 8.49 (8H, d, ${}^{3}J = 16.53$ Hz, 2'-H), 8.13 (8H, d, ${}^{3}J = 16.06$ Hz, 1'-H), 7.69 (16H, d, ${}^{3}J = 7.79$ Hz, H^{o-Ar}), 6.89 (16H, d, ${}^{3}J = 8.15$ Hz, H^{m-Ar}), 6.67 (4H, br s, H^{eq}), 5.94 (4H, d, ${}^{2}J = 12.64$ Hz, H^{ax}), 3.81 (24H, s, -OMe). ¹³C NMR (125 MHz; Py-d₅) δ/ppm: 161.12, 154.86, 143.05, 138.65, 130.15, 130.03, 128.95, 114.99, 55.62, 38.48. MS (MALDI-TOF/TOF; HCCA): m/z 1658.60 [M+H]⁺, 3317.07 [M_2 +H]⁺; calculated for C₁₀₀H₈₁MgN₁₆O₈: 1658.63.

Tetrakis(5,7-bis(4-*tert*-butylphenyl)-6*H*-1,4-diazepino)[2,3-*b*,*g*,*l*,*q*]porphyrazinato magnesium(II) (2c) was synthesized according to a previously published procedure.¹

NMR Spectra



Figure S2. 1 H- 1 H COSY NMR spectrum of **2a** (DMSO- d_{6} , 293 K).



Figure S3. ¹³C NMR spectrum of 2a (pyridine- d_5 , 293 K).



Figure S4. 1 H- 13 C HSQC spectrum of 2a (DMSO- d_{6} , 293 K).



Figure S6. ¹H NMR spectrum of 2b (pyridine-*d*₅, 293 K).







Figure S9. 1 H- 13 C HSQC spectrum of **2b** (pyridine- d_5 , 293 K).



Figure S10. ¹H-¹H NOESY spectrum of **2b** (pyridine- d_5 , 293 K).

X-ray Diffraction Analysis

The crystal of **2a** (C₉₄H₆₂N₁₆OSMgBr₈, M = 2127.16) used for study was triclinic, space group P-1, at T = 100K: a = 15.325(3) Å, b = 27.895(6) Å, c = 33.306(7) Å, $\alpha = 113.35(3)^{\circ}$, $\beta = 100.91(3)^{\circ}$, $\gamma = 95.58(3)^{\circ}$, $V = 100.91(3)^{\circ}$, $\gamma = 100.91($ 12599(6) Å³, Z = 4, $d_{calc} = 1.121$ g/cm³, F(000) = 4216, $\mu = 0.973$ mm⁻¹. The X-ray diffraction study was carried out on the 'Belok' beamline ($\lambda = 0.96990$ Å) of the National Research Center "Kurchatov Institute" (Moscow, Russian Federation) using a Rayonix SX-165 CCD detector. A total of 720 images (117852 reflections, 40519 independent reflections, $R_{int} = 0.0851$) were collected using an oscillation range of 1.0° (ϕ scan mode, $2\theta_{\text{max}} = 76.44^{\circ}$) and corrected for absorption using the Scala program ($T_{\text{min}} = 0.848$; $T_{\text{max}} =$ 0.898).² The data were indexed, integrated and scaled using the utility *iMOSFLM* in CCP4 program.³ The structure was determined by direct methods and refined by full-matrix least squares technique on F^2 with anisotropic displacement parameters for non-hydrogen atoms. Twelve of the sixteen para-bromophenyl substituents were disordered over two sites each with equal occupancies. The independent part of the unit cell of 2a contained several dimethylsulfoxide and water solvate molecules, which were strongly disordered. All attempts to model and refine their positions were unsuccessful. Therefore, the contribution to the scattering by the solvent molecules was removed by use of the utility SQUEEZE in PLATON06.⁴ The hydrogen atoms were placed in calculated positions and refined within a riding model with fixed isotropic displacement parameters $[U_{iso}(H) = 1.5U_{eq}(C)$ for the methyl groups and $1.2U_{eq}(C)$ for the other groups]. The final divergence factors were $R_1 = 0.1361$ for 15689 independent reflections with $I > 2\sigma(I)$ and $wR_2 = 0.2672$ for all independent reflections, S = 1.080. The calculations were performed using the SHELXTL program.⁵ Full crystallographic data for the investigated compounds have been deposited with the Cambridge Crystallographic Data Center, CCDC 1489687. Copies of this information may be obtained free of charge from the Director, CCDC, 12 Union Road, Cambridge CB2 1EZ, UK (fax: +44 1223 336033; e-mail: deposit@ccdc.cam.ac.uk or www.ccdc.cam.ac.uk).



Figure S11. X-ray analysis data for 2a. Show packing of 2a in the crystal cell (hydrogen atoms are omitted for clarity).



Figure S12. Views of the X-ray structure of **2a**. Ellipsoid (left) and space filling (right) representations, respectively (hydrogen atoms and (aryl)ethenyl substituents are omitted for clarity).



Figure S13. The extent of dissociation (α) as function of concentration (mol L⁻¹) of **2c** in DMSO (red line) and in pyridine (black line).



Figure S14. The extent of dissociation (α) as function of concentration of **2b** (mol L⁻¹) in DMSO (red line) and in pyridine (black line).

Studies of the Influence of Fluoride Ion Concentration on the Dimer-Monomer Equilibria

Complex 2c in DMSO



Figure S15. UV-vis spectroscopic changes seen upon addition of tetrabutylammonium fluoride to a solution of 2c in DMSO.



Figure S16. Change in absorbance at 640 nm seen upon titration of 2c with tetrabutylammonium fluoride in DMSO. Concentrations are in mol L⁻¹.



Figure S17. Plot of the indicator concentration ratio for **2c** at 640 nm *vs*. the negative log of the tetrabutylammonium fluoride concentration.



Figure S18. UV-vis spectroscopic changes seen upon addition of tetrabutylammonium fluoride to a solution of 2c in pyridine.



Figure S19. Change in absorbance at 679 and 641 nm seen upon titration of 2c with tetrabutylammonium fluoride in pyridine. Concentrations are in mol L⁻¹.



Figure S20. Plots of the indicator concentration ratio for **2c** at 679 and 641 nm *vs*. the negative log of the tetrabutylammonium fluoride concentration.



Figure S21. UV-vis spectroscopic changes during dissociation of dimeric form of **2c** in pyridine seen upon addition of tetrabutylammonium fluoride (**A**) and the following reverse dimerization process upon addition of FeBr₃ (**B**).



Figure S22. Change in absorbance at 680 and 642 nm seen upon titration of **2c** with tetrabutylammonium fluoride in pyridine and upon titration of the resulting solution with FeBr₃. Concentrations are in mol L^{-1} .



Figure S23. Plots of the indicator concentration ratio for **2c** at 680 and 642 nm *vs*. the negative log of the tetrabutylammonium fluoride or FeBr₃ concentration.



Figure S24. UV-vis spectroscopic changes seen upon addition of tetrabutylammonium fluoride to a solution of 2b in DMSO.



Figure S25. Change in absorbance at 702 and 660 nm seen upon titration of 2b with tetrabutylammonium fluoride in DMSO. Concentrations are in mol L⁻¹.



Figure S26. Plots of the indicator concentration ratio for **2b** at 702 and 660 nm *vs*. the negative log of the tetrabutylammonium fluoride concentration.



Figure S27. UV-vis spectroscopic changes during dissociation of dimeric form of **2b** in DMSO seen upon addition of tetrabutylammonium fluoride (**A**) and the following reverse dimerization process upon addition of FeBr₃ (**B**).



Figure S28. Change in absorbance at 702 and 660 nm seen upon titration of **2b** with tetrabutylammonium fluoride in DMSO and upon titration of the resulting solution with FeBr₃. Concentrations are in mol L^{-1} .



Figure S29. Plots of the indicator concentration ratio for **2b** at 702 and 660 nm *vs.* the negative log of the tetrabutylammonium fluoride or FeBr₃ concentration.



Figure S30. UV-vis spectroscopic changes seen upon addition of tetrabutylammonium fluoride to a solution of **2b** in pyridine.



Figure S31. Change in absorbance at 706 and 666 nm seen upon titration of 2b with tetrabutylammonium fluoride in pyridine. Concentrations are in mol L⁻¹.



Figure S32. Plots of the indicator concentration ratio for **2b** at 706 and 666 nm *vs.* the negative log of the tetrabutylammonium fluoride concentration.



Figure S33. UV-vis spectroscopic changes during dissociation of dimeric form of **2b** in pyridine seen upon addition of tetrabutylammonium fluoride (**A**) and the following reverse dimerization process upon addition of FeBr₃ (**B**).



Figure S34. Change in absorbance at 706 and 666 nm seen upon titration of **2b** with tetrabutylammonium fluoride in pyridine and upon titration of the resulting solution with FeBr₃. Concentrations are in mol L^{-1} .



Figure S35. Plots of the indicator concentration ratio for **2b** at 706 and 666 nm *vs.* the negative log of the tetrabutylammonium fluoride or FeBr₃ concentration.

Electrochemistry

Electrochemical analyses of **2a** and **2b** were carried out in pyridine containing 0.1 M [TBA][BF₄] as the supporting electrolyte. Under these conditions the complexes exist in their dimeric forms as illustrated in Figure S36. One broad irreversible oxidation peak was observed at a potential of 0.804 and 0.777 V (*vs.* SCE) for **2a** and **2b**, respectively. Reductive scans revealed more complex electrochemistry; up to five reversible redox processes were observed (Table S1, Figure S37). Such behavior is consistent with the proposed dimeric nature of the complexes, which can lead to a splitting of redox transitions similar to what is seen in the case of double-decker sandwich complexes stabilized by a central lanthanide cation.⁶⁻⁸ The single broad oxidation peak could result from two closely located irreversible redox transitions. Complexes **2a** and **2b** appear to be noticeably more stable towards oxidation than monomeric phthalocyanine complexes⁹ and sandwich-type double-decker systems.^{7, 8} The $\Delta R_{21} (R_2 - R_1)$ and $\Delta R_{43} (R_4 - R_3)$ values are larger for **2a** than for **2b**, which is as expected given the greater Q-band splitting seen in the UV-vis spectrum (cf. Figures 2 and S36). The reduction potentials R₁, R₂, and R₃ for **2b** were slightly shifted to more negative values relative to **2a**, meaning that the reduction of complex **2b** is more difficult. Such a finding is as expected given the stronger electron-donating effect of the substituents present in **2b**.



Figure S36. UV-vis spectra recorded for samples of 2a and 2b in pyridine in the presence of supporting electrolyte. The concentrations of 2a and 2b were both ca. 0.1 mM; the cuvette path length was 1 mm.





Figure S37. SWVA of complexes **2a** and **2b** in pyridine containing 0.1 M [TBA][BF₄]. Amplitude, 50 mV; frequency, 10 Hz; step potential, 5 mV; $E_{1/2}$ (ferrocene⁺/ ferrocene) = 0.49 V under these experimental conditions.

Table S1. Reduction potentials for 2a and 2b in pyridine (values vs. SCE^{*a*})

	Ox_1^{b}	Red ₁	Red ₂	Red ₃	Red ₄	Red ₅
2a	1.294	-0.790	-1.016	-1.356	-1.646	-
2b	1.267	-0.883	-1.073	-1.433	-1.593	-1.793

 ${}^{a}E_{1/2}$ (Fc⁺/Fc) = 0.49 V. b Irreversible process; the value of the peak potential is given.

In Vitro and in Vivo Toxicity Tests

Compounds **2a**,**b** were converted into water-soluble forms by co-solvation with polyvinylpyrrolidone (PVP) for biological testing.

Cell viability was recorded as dehydrogenase activity as inferred from 3-(4,5-dimethylthiazol-2-yl)-2,5diphenyltetrazolium bromide (MTT) assays.¹⁰ Briefly, the SH-SY5Y cells or primary culture of rat cerebellar granule cells were incubated with the test compounds or equivalent volume of vehicle (less than 1% of the whole volume of medium under the layer of cells) for 24 h. Then, 20 μ L of MTT (2 mg mol L⁻¹ in PBS) were added to each well and the cells were incubated at 37°C for 2 h. The supernatants were aspirated carefully, 200 μ L of DMSO were added to each well to dissolve the precipitate, and the absorbance was measured at 570 nm using a microplate reader Victor (Perkin Elmer).

Acute intraperitoneal toxicity of complexes 2a and 2b was determined using male hybrid mice BDF1 (weighing 20–22 g) by injection of single doses of 25, 50, 100, and 150 mg kg⁻¹ body weight, respectively. Mice were kept under specific pathogen-free conditions; water and food were provided *ad libitum*. All animals were observed for clinical signs of toxicity within the first 30 min following dose administration, and then – with an interval of 1–4 h and further – once daily for 14 days. No deaths were registered during the 14 day study periods associated with either complex. The data obtained were plotted as a dose–effect curve, from which the acute toxicity values (MTD, LD₅₀ and LD₁₀₀) of complexes **2a** and **2b** were determined.

Compound	Injected dose, mg	Number of animals	Number of deaths at
Compound	kg^{-1}	per group	day 14 after injection
	100	6	0
	250	6	0
PVP	500	6	0
	750	6	0
	1500	6	0
	3750	6	0
	7500	6	0
	10	6	0
20	50	6	0
2 a	100	6	0
	150	6	0
	10	6	0
2 h	50	6	0
20	100	6	0
	150	6	0

Table S2. Acute toxicity studies of compounds 2a and 2b as determined using mal hybrid mice BDF1.

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