Supporting Information

(32 pages including the cover page)

A Bromo-Capped Diruthenium(I,I) N-Heterocyclic Carbene Compound for *in situ* Bromine Generation with NBS: Catalytic Olefin Aziridination Reactions⁵

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 ξ Dedicated to Professor Kim R. Dunbar on the occasion of her 60th Birthday

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1. Synthesis and characterization

Synthesis of PIN HI



Scheme S1. Synthesis of PIN.HI.

¹H NMR (500 MHz, DMSO-D6, 294 K): δ 10.28 (s, 1H, Im), 8.95 (d, *J* = 9.1 Hz, 1H, NP), 8.69 (s, 1H, NP), 8.22 (d, *J* = 9.1 Hz, 1H, NP), 8.19 (s, 1H, Im), 7.52 (s, 1H, Im), 4.80 (m, 1H, CH-ⁱPr), 2.73 (s, 3H, CH₃-NP), 2.68 (s, 3H, CH₃-NP), 1.58 (d, 6H, J = 6.8 Hz, ⁱPr). ¹³C NMR (125.8 MHz, DMSO-D6, 296.2 K): δ 164.7 (NCNNP), 153.5 (CCNNP), 147.8 (CCNNP), 139.9 (CCCNP), 135.3 (CCCNP), 124.9 (CCCNP), 122.4 (NCCIm), 121.5 (CCCNP), 120.1 (CCCNP), 112.7 (NImCC), 53.9 (CHⁱPr), 25.3 (CH₃ NP), 22.7 (CH₃ ⁱPr), 18.4 (CH₃ NP). ESI–MS, *m/z*: 267.1611, (z=1), [M]⁺

Synthesis of 2



¹H NMR (500 MHz, CD₃CN, 294 K): δ 8.86 - 8.69 (m, 1H, NP), 8.14 (s, 1H, Im), 8.02 -7.88 (m, 1H, NP), 7.52 - 7.50 (m, 1H, NP), 7.40 (s, 1H, Im), 5.86 – 5.81 and 5.52 – 5.48 (m, 1H, CH-ⁱPr), 2.83 – 2.74 (m, 6H, CH₃-NP), 1.68 – 1.47 (m, 6H, ⁱPr). ¹³C NMR (125.8 MHz, CD₃CN, 296.2 K): δ 202.9 (CO), 191.3 (CO), 173.2 (NCNIm), 166.4 (NCNNP), 155.2 (NNPCNIm), 147.8 (CCNNP), 140.3 (CCCNP), 126.3 (CCCNP), 125.6 (CCCNP), 122.7 (NCCIm), 120.8 (CCCNP),112.5 (NImCC), 111.5 (NImCC), 55.4 (CHⁱPr), 25.7 (CH3 NP), 23.0 (CH3 ⁱPr), 22.8 (CH3 ⁱPr), 18.3 (CH3 NP). IR (KBr, cm⁻¹): v(CO) 2009, 1991, 1968, 1924. Anal. Calcd for C₃₆H₃₆N₈O₄I₂Ru₂.CH₃CN: C, 39.89; H, 3.43; N, 11.03. Found: C, 39.34; H, 3.37; N, 11.08. ESI-MS, m/z 947.0106 corresponding to [M–I–CO]⁺, where M is Ru₂(CO)₄(PIN)₂I₂.



Figure S1. ¹H NMR (top) and ¹³C NMR (below) of PIN.HI.





Figure S3. Simulated (red line) and experimental mass distributions (black line) for **2** [M-I-CO]⁺ at m/z (z = 1) M = Ru₂(CO)₄(PIN)₂I₂.

2. X–Ray Data Collection and Refinements

Single-crystal X-ray studies were performed on a CCD Bruker SMART APEX diffractometer equipped with an Oxford Instruments low-temperature attachment. All the data were collected at 100(2) K using graphite–monochromated Mo–K α radiation ($\lambda \alpha$ = 0.71073 Å). The frames were indexed, integrated, and scaled using the SMART and SAINT software packages,¹ and the data were corrected for absorption using the SADABS program.² The structure was solved and refined with the SHELX suite of programs as implemented in X-seed.³ All hydrogen atoms were included in the final stages of the refinement and were refined with a typical riding model. The crystal was brittle and poorly diffracting. Number of datasets was collected on single crystals from different batches, whereof one of the best data is reported herein. Despite our best efforts, the Rint value is on the higher side, which generated a B level alert. Some of the lattice solvent molecules in the crystal lattice of 2 cannot be modelled satisfactorily due to the presence of severe disorders. Therefore, the Olex-2 mask program has been used to discard those disordered solvents molecules. A total electron densities of 254 were removed which can be tentatively ascribed for four CH_3CN , one Et_2O and one H_2O molecules. Pertinent crystallographic data for compound 2 is summarized in Table S1. The crystallographic figure used in this manuscript has been generated using Diamond 3.1e software.⁴ CCDC number 1841806 contains the supplementary crystallographic data for compound 2. This data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data request/cif.



Figure S4. Molecular structure of **2** with important atoms labelled. All hydrogen atoms have been omitted for the sake of clarity. Thermal ellipsoids are drawn at the 40% probability level. Selected bond lengths (Å) and angles (deg): Ru1–Ru1' 2.8448(16), Ru1–C2 2.049(10), Ru1–N3 2.172(7), Ru1–I1 2.7560(11), Ru1–C17 1.829(14), C17–O1 1.174(13). Ru1'–Ru1–I1 177.00(4), Ru1'–Ru1–C2 89.5(3), Ru1'–Ru1–N3 91.96(17), Ru1'–Ru1–C17 89.9(3), C2–Ru1–C17 98.4(4). Dihedral angles (deg): N3–Ru1–Ru1'–N3' 37.1(6), C2–Ru1–Ru1'–C2' 170.3(6).

for 2 .			
	2		
Empirical Formula	$C_{36}H_{36}N_8O_4I_2Ru_2$		
Formula Weight	1100.67		
Crystal System	Trigonal		
Space Group	P -3c1		
a (Å)	21.118(2)		
b (Å)	21.118(2)		
c (Å)	17.870(3)		
α (deg)	90		
β (deg)	90		
γ (deg)	120		
V (Å ³)	6901.7(18)		
Z	6		
ρ _{calcd} (g cm⁻³)	1.589		
μ (mm ⁻¹)	2.040		
F(000)	3204		
Reflections Collected	57918		
Independent	4063		
Observed [$I > 2\sigma$ (I)]	2358		
No. of parameters	239		
GooF	1.027		
R _{int}	0.2467		
Final R indices $[I > 2\sigma(I)]^a$	R1 = 0.0720		
	wR2 = 0.1506		
R indices (all data) ^a	R1 = 0.1292		
	wR2 = 0.1733		
${}^{a}R_{1} = \Sigma F_{o} - F_{c} /\Sigma F_{o} $ with $F_{o}^{2} > 2\sigma(F_{o}^{2})$. w $R_{2} = [\Sigma w(F_{o}^{2} - 1)]$			
$ F_{c}^{2})^{2}/\Sigma F_{o}^{2} ^{2}]^{1/2}$			

 Table S1. Crystallographic data and refinement parameters

3. HPLC method for the separation of d,l (rac) and meso-1,2-dibromo1,2-diphenylethane compound

The *d*,*I* and meso-1,2-dibromo1,2-diphenylethane compounds were dissolved in ⁱPrOH and subjected to normal phase chiral HPLC analysis (Chiral AD-H column, 5:95 ⁱPrOH:Hexane, flow rate 0.5 ml/min, detection at 254 nm).







Figure S6. HPLC chromatogram of *d*,*l*-1,2-dibromo1,2-diphenylethane.

4. In situ generation of lodine from an lodide analogue of 1

A mixture of complex **2** (20 mg) and NIS (3.5 mg) was dissolved in 2 mL of acetonitrile solution and stirred for 20 minutes. After that freshly prepared starch solution was added to the reaction mixture. Immediately the color of solution changed from brownish yellow to dark blue. Similar reaction was repeated with tetrabutylammonium iodide and NIS, however, with addition of starch solution no color change was observed.



Figure S7a. Bromination of phenol in NBS and K₂CO₃ (GC-MS conversion).



Figure S7b. Bromination of phenol in liquid Br₂ (GC-MS conversion).





Figure S7c. Bromination of phenol in catalyst 1, NBS, K₂CO₃ (GC-MS conversion).



Figure S7d. Bromination of phenol in catalyst 1, NBS (GC-MS conversion).

5. Control experiment for olefin aziridination reaction

Table S2. Catalyst screening for aziridination recation.			
	Catalyst, TsNH ₂ NBS, K ₂ CO ₃ rt, 24 hrs	► N Ts	
Entry	Catalyst (mol%)	Yield ^a (%)	
1.	1 (5)	90	
2.	No catalyst	17	
3. ^{<i>b</i>}	1 and TIBF₄ (5)	18	
4.	3 (5)	20	
5.	4 (5)	76	
6. ^c	5 (1.0, 0.1)	77, 62	
[a] lealeted violate. [b] establish 4 and TIDE vises atims of in a starity in			

[a] Isolated yields, [b] catalyst **1** and TIBF₄ was stirred in acetonitrile solution and the residue obtained after workup was used as a catalyst, [c] A. J. Catino, J. M. Nichols, R. E. Forslund and M. P. Doyle, *Org. Lett.*, 2005, **7**, 2787.

6. ¹H NMR study



Scheme. S3. Formation of β -bromoamide product.



Figure S8. Spectrum A: ¹H NMR study for reaction mixture of styrene and NBS. Spectrum B: ¹H NMR study on the reaction mixture of styrene and NBS with TsNH₂, 5 mol% **1**.



Figure S9. ¹H NMR (top) and ¹³C NMR (below) of *trans*1,2-dibromocyclohexane.

7. Spectral data of the aziridines

¹H NMR (CDCl₃, 400 MHz): δ 7.86 (d, J = 8.7 Hz, 2H), 7.32 (d, J = 8.3 Hz, 2H), 7.28-7.19 (m, 5H), 3.76 (dd, J = 4.6 Hz, 7.3 Hz, 1H), 2.97 (d, J = 7.3 Hz, 1H), 2.42 (s, 3H) 2.37 (d, J = 4.1 Hz, 1H); ¹³C NMR (CDCl₃, 100 MHz): δ 144.8, 135.1, 135.0, 129.9, 128.7, 128.4, 128.0, 126.6, 41.1, 36.0, 21.8.

¹H NMR (CDCl₃, 500 MHz): δ 7.86 (d, J = 8.0 Hz, 2H), 7.27 (d, J = 7.9 Hz, 2H), 7.04 (s, 4H), 3.68 (q, J = 4.3 Hz, 1H), 2.92 (d, J = 7.3 Hz, 1H), 2.37 (s, 3H), 2.32 (d, J = 4.9 Hz, 1H), 2.25 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz): δ 144.7, 138.3, 135.2, 132.1, 129.8, 129.3, 128.0, 126.6, 41.1, 35.8, 21.7, 21.2.

¹H NMR (CDCl₃, 500 MHz): δ 7.81 (d, J = 7.9 Hz, 2H), 7.32 (d, J = 7.9 Hz, 2H), 7.25-7.01 (m, 4H), 3.72 (br, 1H), 2.95 (d, J = 7.3 Hz, 1H), 2.43 (s, 3H), 2.36 (m, 1H), 2.17 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz): δ 144.6, 138.4, 134.9, 134.6, 129.8, 129.1, 128.5, 128.0, 127.2, 123.7, 41.1, 35.9, 29.7, 21.7, 21.3.

¹H NMR (CDCl₃, 500 MHz): δ 7.85 (d, J = 8.0 Hz, 2H), 7.33 (d, J = 8.0 Hz, 2H), 7.17 (q, J = 8.6 Hz, 2H), 6.97 (t, J = 8.6 Hz, 2H) 3.75 (dd, J = 4.0 Hz, 7.4 Hz, 1H), 2.96 (d, J = 6.9 Hz, 1H), 2.43 (s, 3H), 2.34 (d, J = 4.6 Hz, 1H); ¹³C NMR (CDCl₃, 125 MHz): δ 144.9, 134.9, 129.9, 128.4, 128.3, 128.0, 115.7, 115.5, 40.4, 36.1, 21.3.

¹H NMR (CDCl₃, 400 MHz): δ 7.86 (d, J = 8.2 Hz, 2H), 7.32 (d, J = 8.2 Hz, 2H), 7.25-6.97 (m, 4H), 3.96 (dd, J = 4.3 Hz, 7.3 Hz, 1H), 2.99 (d, J = 7.1 Hz, 1H), 2.42 (s, 3H), 2.38 (d, J = 4.4 Hz, 1H); ¹³C NMR (CDCl₃, 100 MHz): δ 160.5, 144.9, 134.7, 129.9, 129.8, 128.2, 127.5, 124.4, 115.3, 35.6, 35.2, 21.8.

¹H NMR (CDCl₃, 500 MHz): δ 7.82 (d, J = 8.2 Hz, 2H), 7.26-7.24 (m, 5H), 7.14 (d, J = 6.1 Hz, 2H), 3.79 (d, J = 4.3 Hz, 1H), 2.90 (dq, J = 6.1, 4.6 Hz, 1H), 2.38 (s, 3H), 1.83 (d, J = 6.1 Hz, 1H); ¹³C NMR (CDCl₃, 125 MHz): δ 144.0, 137.9, 135.6, 129.8, 128.3, 128.1, 127.3, 126.3, 49.3, 49.2, 21.7, 14.2.













N-Ts

¹H NMR (CDCl₃, 400 MHz): δ 7.86 (d, *J* = 8.2 Hz, 2H), 7.40 (d, *J* = 7.4 Hz, 1H), 7.32 (d, *J* = 8.2 Hz, 2H), 7.23-7.22 (m, 1H), 7.19-7.15 (m, 2H), 4.30 (d, *J* = 5.2 Hz, 1H), 3.95 (m, 1H), 3.02 (d, *J* = 3.8 Hz, 2H), 2.43 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz): δ 144.6, 138.4, 129.8, 129.0, 128.0, 126.9, 125.7, 125.2, 50.3, 45.1, 34.9, 21.8.

¹H NMR (CDCl₃, 500 MHz): δ 7.82 (d, *J* = 7.9 Hz, 2H), 7.33 (d, *J* = 7.9 Hz, 2H), 2.74-2.69 (m, 1H), 2.62 (d, *J* = 7.3 Hz, 1H,) 2.43 (s, 3H), 2.04 (d, *J* = 4.8 Hz, 1H,) 1.55-1.49 (m, 1H), 1.27-1.20 (m, 5H), 0.80 (t, *J* = 7.3 Hz, 3H); ¹³C NMR (CDCl₃, 125 MHz): δ 144.4, 135.3, 129.7, 128.0, 40.5, 33.9, 31.0, 29.8, 22.2, 21.7, 13.9.

¹H NMR (CDCl₃, 400 MHz): δ 7.78 (d, *J* = 8.5 Hz, 2H), 7.29 (d, *J* = 7.9 Hz, 2H), 2.66-2.63 (m, 1H), 2.59 (d, *J* = 6.7 Hz, 1H,) 2.39 (s, 3H), 2.02 (d, *J* = 4.6 Hz, 1H,) 1.53-1.45 (m, 1H), 1.20-1.10 (m, 9H), 0.80 (t, *J* = 7.3 Hz, 3H); ¹³C NMR (CDCl₃, 100 MHz): δ 144.4, 135.2, 129.6, 128.0, 40.5, 33.8, 31.6, 31.4, 28.7, 26.7, 22.5, 21.6, 14.0.

¹H NMR (CDCl₃, 500 MHz): δ 7.81 (d, *J* = 8.3 Hz, 2H), 7.32 (d, *J* = 8.0 Hz, 2H), 2.73-2.67 (m, 1H), 2.62 (d, *J* = 7.0 Hz, 1H,) 2.42 (s, 3H), 2.04 (d, *J* = 4.6 Hz, 1H,) 1.25-1.19 (br, m, 14H), 0.86 (t, *J* = 7.9 Hz, 3H); ¹³C NMR (CDCl₃, 125 MHz): δ 144.3, 135.2, 129.5, 127.9, 40.5, 33.7, 31.7, 31.2, 29.3, 29.1, 29.0, 22.5, 21.5, 14.0.

¹H NMR (CDCl₃, 500 MHz): δ 7.69 (d, *J* = 8.0 Hz, 2H), 7.22 (d, *J* = 8.0 Hz, 2H), 2.85 (s, 2H), 2.55 (s, 3H), 1.68 – 1.66 (m, 4H,) 1.30-1.26 (m, 2H), 1.14-1.10 (m, 2H) ; ¹³C NMR (CDCl₃, 125 MHz): δ 143.8, 135.3, 129.2, 127.3, 39.6, 29.3, 22.4, 21.2, 21.1, 19.1.

¹H NMR (CDCl₃, 500 MHz): δ 7.80 (d, *J* = 8.3 Hz, 2H), 7.31 (d, *J* = 8.0 Hz, 2H), 2 77 (br, 2H), 2.43 (s, 3H), 2.02 -1.99 (m, 2H), 1.57-1.50 (m, 6H), 1.43-1.24 (m, 4H); ¹³C NMR (CDCl₃, 500 MHz): δ 144.1, 135.9, 129.7, 127.6, 44.0, 26.5, 26.2, 25.3, 21.7.

¹H NMR (CDCl₃, 400 MHz): δ 7.71 (d, *J* = 7.9 Hz, 2H), 7.29 (d, *J* = 7.9 Hz, 2H), 5.68 - 5.64 (m, 1H), 5.54 - 5.47 (m, 1H), 4.33 (t, *J* = 6.7 Hz, 1H), 3.65 - 3.59 (m, 1H), 2.42 (s, 3H), 2.07 - 1.94 (m, 2H), 1.88 - 1.80 (m, 2H), 1.54 - 1.47 (m, 2H), 1.23 (s, 2H,); ¹³C NMR (CDCl₃, 100 MHz): δ 143.6, 137.8, 130.6, 129.9, 126.9, 35.6, 29.7, 25.7, 21.6.





















⊃N—Ts 4.19 3.09 2.13 2.25 2.08 3.0 8.0 7.0 2.0 4.0 6.0 5.0 7.2307 7.6899 2.5515 0.0000 77.2575 77.0000 76.7425 30.9 140.0 130₁0 120.0 70.0 60.0 10.0 110.0 100.0 90.0 50.0 22.4694 21.2867 21.1817 21.1817 19.1215 40,0 0 39.6575 143.8827 135.3936 129.2891 127.3051



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