Synthesis, cytotoxic activity and DNA interaction studies of new dinuclear platinum(II) complexes with aromatic 1,5-naphthyridine bridging ligand: DNA binding mode of polynuclear platinum(II) complexes in relation to the complex structure

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Abstract

The synthesis, spectroscopic characterization, cytotoxic activity and DNA binding evaluation of seven new dinuclear platinum(II) complexes Pt1-Pt7, with general formula $[{Pt(L)Cl}_2(\mu-1,5-nphe)](ClO_4)_2$ (1,5-nphe is 1,5-naphthyridine; while L is two ammines (Pt1) or one bidentate coordinated diamine: ethylenediamine (Pt2), (\pm) -1,2propylenediamine (Pt3), trans-(±)-1,2-diaminocyclohexane (Pt4), 1,3-propylenediamine (Pt5), 2,2-dimethyl-1,3-propylenediamine (Pt6), and 1,3-pentanediamine (Pt7)), were reported. In vitro cytotoxic activity of these complexes was evaluated against three tumor cell lines, murine colon carcinoma (CT26), murine mammary carcinoma (4T1) and murine lung cancer (LLC1) and two normal cell lines, murine mesenchymal stem cells (MSC) and human fibroblasts (MRC-5) cells. The results of MTT assay indicate that all investigated complexes have almost no cytotoxic effects on 4T1 and very low cytotoxicity toward LLC1 cell lines. In contrast to the effects on LLC1 and 4T1 cells, complexes Pt1 and Pt2 had significant cytotoxic activity toward CT26 cells. Complex Pt1 had much lower IC₅₀ value for activity on CT26 cells compared with cisplatin. In comparison to cisplatin, all dinuclear Pt1-Pt7 complexes showed lower cytotoxicity toward normal MSC and MRC-5 cells. In order to measure the amount of the platinum(II) complexes taken up by the cells, we quantified the cellular platinum content using inductively coupled plasma mass spectrometry (ICP-QMS). Molecular docking study, performed to evaluate the potential binding mode of dinuclear platinum(II) complexes Pt1-Pt7 and their aqua derivatives W1-W7, respectively, at double stranded DNA was shown that groove spanning and backbone tracking are the most stable binding modes.

Keywords: Dinuclear platinum(II) complexes; 1,5-naphthyridine; Cytotoxicity; Molecular docking; DNA interaction

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Table S1 NMR (¹H and ¹³C) chemical shifts (δ , ppm), together with multiplicities **S6** and coupling constants (J_{H-H}, Hz), for the free 1,5-naphthyridine and the corresponding 1,5-naphthyridine-bridged Pt(II) complexes **Pt1-Pt7** in D₂O as solvent with TSP as the internal standard.

Table S2 The amounts (μ g/g) of platinum taken up by the LLC1 cells after 2 h of**S7**treatment at 37 °C with dinuclear **Pt1–Pt7** complexes and cisplatin at 2 μ M inDMSO medium determined by using inductively coupled plasma massspectrometry (ICP-QMS).The results represent the mean value of six replicatemeasurements. The correlation coefficient of the regression line was 1.0000.**Table S3** Optimized operating conditions of ICP-QMS.**S7**

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Fig. S3 Illustrations of backbone tracking of ligands on DNA in crystal structures, **S10** extracted from PDB. The ligand is shown by ball&stick style.

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orientation of these complexes and iron-free (apo) form of the recombinant N-lobe of human serum transferrin (ApoTfN), as assessed by molecular docking.

Fig. S8 Absorption spectra of Pt2 and Pt5 complexes in the absence and in S15 the presence of increasing amounts of CT-DNA measured in 0.01 M phosphate buffer solution (PBS) at pH 7.40 and 37 °C. c(Pt(II) complex) = 8 μ M (r = c(DNA)/c(Pt(II) complex) = 0, 0.1, 0.6, 1.0, 1.6, 2.0). Arrow shows the absorption intensity changes upon increasing of CT-DNA concentration. Fig. S9 Emission spectra of the CT-DNA-EtBr system in the absence and in S16 the presence of increasing amounts of Pt2 and Pt5 complexes. The spectra were measured in 0.01 M in phosphate buffer solution at pH 7.40 and room temperature. c(EtBr) = 8 μ M; c(DNA) = 8 μ M; c(Pt(II) complex) = (0-8) μ M; λ_{ex} = 527 nm. Arrow shows the emission intensity changes upon increasing of the complex concentration.

Fig. S10 Binding environments for the most stable binding of aqua complexes W1 S17 and W2 to DNA, as assessed by molecular docking (W1 and W2 represent aqua derivatives of the corresponding chloride platinum(II) complexes Pt1 and Pt2, respectively).

Fig. S11 Binding environments for the most stable binding of aqua complexes W3 S17 and W4 to DNA, as assessed by molecular docking (W3 and W4 represent aqua derivatives of the corresponding chloride platinum(II) complexes Pt3 and Pt4, respectively).

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respectively).

Fig. S13 Two the most stable binding modes and binding environment for the most **S18** stable binding mode of platinum(II)-aqua complex **W7** to DNA (backbone tracking), as assessed by molecular docking (**W7** represents aqua derivative of the corresponding chloride platinum(II) complex **Pt7**).

Fig. S14 The structures of the most stable binding modes of dinuclear platinum(II) **S19** complexes **Pt1–Pt7** to DNA, as assessed by molecular docking.

Scheme S1 Reaction pathway for preparation of dinuclear platinum(II) complexes S20

Pt1-Pt7 (L is 2NH₃ or bidentate coordinated diamine ligand: en, 1,2-pn, dach, 1,3-

pd, 2,2-diMe-1,3-pd and 1,3-pnd; 1,5-nphe is 1,5-naphthyridine).

Scheme S2 Molecular structures of some polynuclear platinum(II) complexes, S20 mentioned in the main body of the manuscript.

Table 1 NMR (¹H and ¹³C) chemical shifts (δ , ppm), together with multiplicities and coupling constants (J_{H-H} , Hz), for the free 1,5-naphthyridine and the corresponding 1,5-naphthyridine-bridged Pt(II) complexes **Pt1–Pt7** in D₂O as solvent with TSP as the internal standard.

Ligand/ Complex	NMR assignments							
	1 ¹ H			¹³ C				
	H2, H6	H4, H8	H3, H7	C2, C6	C4, C8	C3, C7	C4a, C8a	
1,5-nphe	8.57 <i>dd;</i> J= 4.3, 1.4	7.91 <i>d</i> ; J=7.4	7.48 <i>dd;</i> J= 8.6, 4.3	153.6	137.7	127.8	142.8	
Pt1	10.38 <i>d</i> ; J= 8.6	9.69 <i>d</i> ; J=5.3	8.24 <i>dd;</i> J= 9.0, 5.4	161.4	147.4	130.5	147.5	
Pt2	10.25 <i>d</i> J=8.7	9.61 <i>d</i> ; J=5.1	8.17 <i>m</i>	159.6	141.4	129.0	147.6	
Pt3	10.22 <i>d</i> J=8.6	9.58 m	8.16 <i>d;</i> J=7.6	161.0	142.9	130.3	147.5	
Pt4	10.23 <i>m</i>	9.58 m	8.16 <i>m</i>	161.7	142.2	130.2	147.2	
Pt5	10.27 <i>d;</i> J=9.1	9.58 <i>d</i> ; J=5.7)	8.18 m	161.2	142.3	130.4	148.0	
Pt6	10.27 <i>d;</i> J=8.9	9.57 <i>d</i> ; J=5.1	8.16 <i>dd;</i> J=9.0, 5.1	161.0	142.3	130.5	148.4	
Pt7	10.23 <i>m</i>	9.58 m	8.17 <i>m</i>	161.1	142.6	130.4	147.6	

s = singlet; d = doublet; dd = doublet of doublets; m = multiplet

Table S2 The amounts (μ g/g) of platinum taken up by the LLC1 cells after 2 h of treatment at 37 °C with dinuclear **Pt1–Pt7** complexes and cisplatin at 2 μ M in DMSO medium determined by using inductively coupled plasma mass spectrometry (ICP-QMS).The results represent the mean value of six replicate measurements. The correlation coefficient of the regression line was 1.0000.

Compley	Concentration of platinum (µg/g)				
Complex	Mean	St. Dev.			
Pt1	4.23	0.07			
Pt2	6.52	0.10			
Pt3	1.51	0.03			
Pt4	1.51	0.04			
Pt5	4.94	0.07			
Pt6	1.20	0.03			
Pt7	1.05	0.02			
cisplatin	0.72	0.02			
untreated	0.01	0.01			

S3

operating

ICP-QMS.

Table

Instrument parameter	Operating condition
Forward power (W)	1550
Ar gas flow rates (L/min)	
Coolant	0.80
Auxiliary	1.13
Transport gas	14
Spray chamber	Glass cyclonic
Nebulizer	PFA-ST MicroFlow
Dwell time(s)	0.01
Number of Channels	1
Spacing	0.1
Resolutin	Resolutin
Replicates	6

Optimized conditions of



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Fig. S6 The structures of the most stable binding modes of dinuclear platinum(II) complexes (**Pt1–Pt7**) to iron-free (apo) form of the recombinant N-lobe of human serum transferrin (ApoTfN), as assessed by molecular docking.



Fig. S7 Amino-acid environment of **Pt1–Pt7** complexes in the most stable orientation of these complexes and iron-free (apo) form of the recombinant N-lobe of human serum transferrin (ApoTfN), as assessed by molecular docking.



Fig. S8 Absorption spectra of **Pt2** and **Pt5** complexes in the absence and in the presence of increasing amounts of CT-DNA measured in 0.01 M phosphate buffer solution (PBS) at pH 7.40 and 37 °C. $c(Pt(II) \text{ complex}) = 8 \mu M (r = c(DNA)/c(Pt(II) \text{ complex}) = 0, 0.1, 0.6, 1.0, 1.6, 2.0)$. Arrow shows the absorption intensity changes upon increasing of CT-DNA concentration.



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Fig. S11 Binding environments for the most stable binding of aqua complexes **W3** and **W4** to DNA, as assessed by molecular docking (**W3** and **W4** represent aqua derivatives of the corresponding chloride platinum(II) complexes **Pt3** and **Pt4**, respectively).



Fig. S12 Binding environments for the most stable binding of aqua complexes **W5** and **W6** to DNA, as assessed by molecular docking (**W5** and **W6** represent aqua derivatives of the corresponding chloride platinum(II) complexes **Pt5** and **Pt6**, respectively).



Fig. S13 Two the most stable binding modes and binding environment for the most stable binding mode of platinum(II)-aqua complex **W7** to DNA (backbone tracking), as assessed by molecular docking (**W7** represents aqua derivative of the corresponding chloride platinum(II) complex **Pt7**).



Fig. S14 The structures of the most stable binding modes of dinuclear platinum(II) complexes **Pt1–Pt7** to DNA, as assessed by molecular docking.

$$[Pt(L)Cl_{2}] + AgNO_{3} \xrightarrow{in DMF} [Pt(L)Cl(DMF)]^{+} + 0.5 \text{ mol } 1,5\text{-nphe}$$

$$\downarrow in DMF$$

$$12 \text{ h}$$

$$[\{Pt(L)Cl\}_{2}(\mu\text{-}1,5\text{-nphe})](ClO_{4})_{2} \xrightarrow{in \text{ excess of } LiClO_{4}}{-DMF} [\{Pt(L)Cl\}_{2}(\mu\text{-}1,5\text{-nphe})]^{2+}$$

Scheme S1 Reaction pathway for preparation of dinuclear platinum(II) complexes Pt1–Pt7 (L is 2NH₃ or bidentate coordinated diamine ligand: en, 1,2-pn, dach, 1,3-pd, 2,2-diMe-1,3-pd and 1,3-pnd; 1,5-nphe is 1,5-naphthyridine).



Scheme S2 Molecular structures of some polynuclear platinum(II) complexes, mentioned in the main body of the manuscript.