

# Synthesis of heterobimetallic gold(I) ferrocenyl-substituted 1,2,3-triazol-5-ylidene complexes as potential anticancer agents

## Electronic Supplementary Information

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#### I. General considerations

Nuclear magnetic resonance (NMR) spectra were obtained using either a Bruker AVANCE-III-300 operating at 300.13 MHz for <sup>1</sup>H, 75.47 MHz for <sup>13</sup>C, 121.49 MHz for <sup>31</sup>P and 282.40 MHz for <sup>19</sup>F; or AVANCE-III-400 operating at 400.21 MHz for <sup>1</sup>H, 100.64 MHz for <sup>13</sup>C, 162.01 MHz for <sup>31</sup>P and 376.57 MHz for <sup>19</sup>F. <sup>1</sup>H Chemical shifts are reported as δ (ppm) values downfield from Me<sub>4</sub>Si and chemical shifts were referenced to residual non-deuterated solvents peaks (CDCl<sub>3</sub>, 7.26 ppm). <sup>13</sup>C{<sup>1</sup>H} chemical shifts are also reported as δ (ppm) values downfield from Me<sub>4</sub>Si and chemical shifts were referenced to residual non-deuterated solvents peaks (CDCl<sub>3</sub>, 77.16 ppm). The chemical shifts are given in ppm and the proton coupling constants (*J*) are given in Hz. The spectral coupling patterns are designated as follows: s - singlet; d - doublet; t - triplet; q - quartet; sept-septet; m - multiplet; br - broad signal. The assignment of the

NMR for each complex follows the numbering scheme individually assigned for each compound illustrated on the respective NMR spectra below. An asterisk (\*) denotes solvent contaminant in the NMR spectra. Chemical shift assignment in the  $^1\text{H}$  NMR spectra is based on first-order analysis and when required were confirmed by two-dimensional (2D) ( $^1\text{H}$ - $^1\text{H}$ ) homonuclear chemical shift correlation (COSY) experiments. The  $^{13}\text{C}$  shifts were obtained from proton-decoupled  $^{13}\text{C}$  NMR spectra. Where necessary, the multiplicities of the  $^{13}\text{C}$  signals were deduced from proton-decoupled DEPT-135 spectra. The resonances of the proton-bearing carbon atoms were correlated with specific proton resonances using 2D ( $^{13}\text{C}$ - $^1\text{H}$ ) heteronuclear single-quantum coherence (HSQC) experiments. Standard Bruker pulse programs were used in the experiments. Single crystal X-ray diffraction data were collected on a Bruker D8 Venture with Apex3 and a Photon 100 CMOS detector (**2 – 5**) or a Bruker Apex II with a CCD detector (**6**) using Mo- $K_{\alpha}$  radiation ( $\lambda = 0.71073 \text{ \AA}$ ) ( $l\mu\text{s}$ , **2 – 5**; sealed tube, **6**). Crystals were selected under paratone oil, mounted on nylon loops then immediately placed in a cold stream of  $\text{N}_2$  at 150 K (at 173 K for **6**). Data were reduced using Bruker SAINT and SADABS, solved using Bruker SHELXTS and refined using SHELXTL (**2 – 5**) and Olex2 (**6**).<sup>1</sup> Mass spectral analyses were performed on a Waters Synapt G2 HDMS by direct infusion at 5  $\mu\text{L}/\text{min}$  with positive electron spray as the ionization technique. The  $m/z$  values were measured in the range of 400-1500 with acetonitrile as solvent. Prior to analysis, a 5 mM sodium formate solution was used to calibrate the instrument in resolution mode. Elemental analyses were carried out using a Thermo Flash 1112 Series CHNS-O Analyzer.

## II. NMR Spectra of complexes 1–6

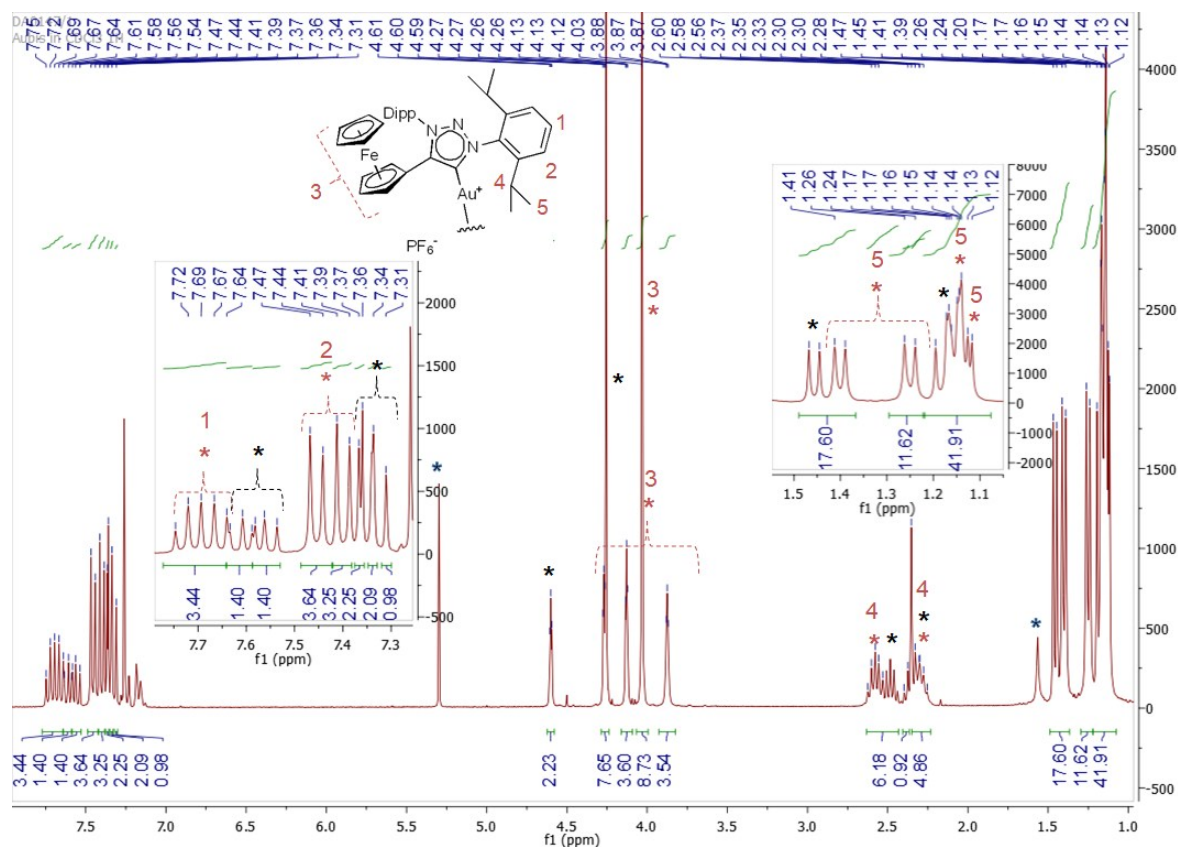


Figure S1. The  $^1\text{H}$  NMR spectrum showing the presence of two gold(I) complexes, where (\*) denotes the peaks belonging to the cationic bis(carbene) gold(I) complex (1), and (\*) denotes neutral complex 2 in solvent  $\text{CDCl}_3$ .

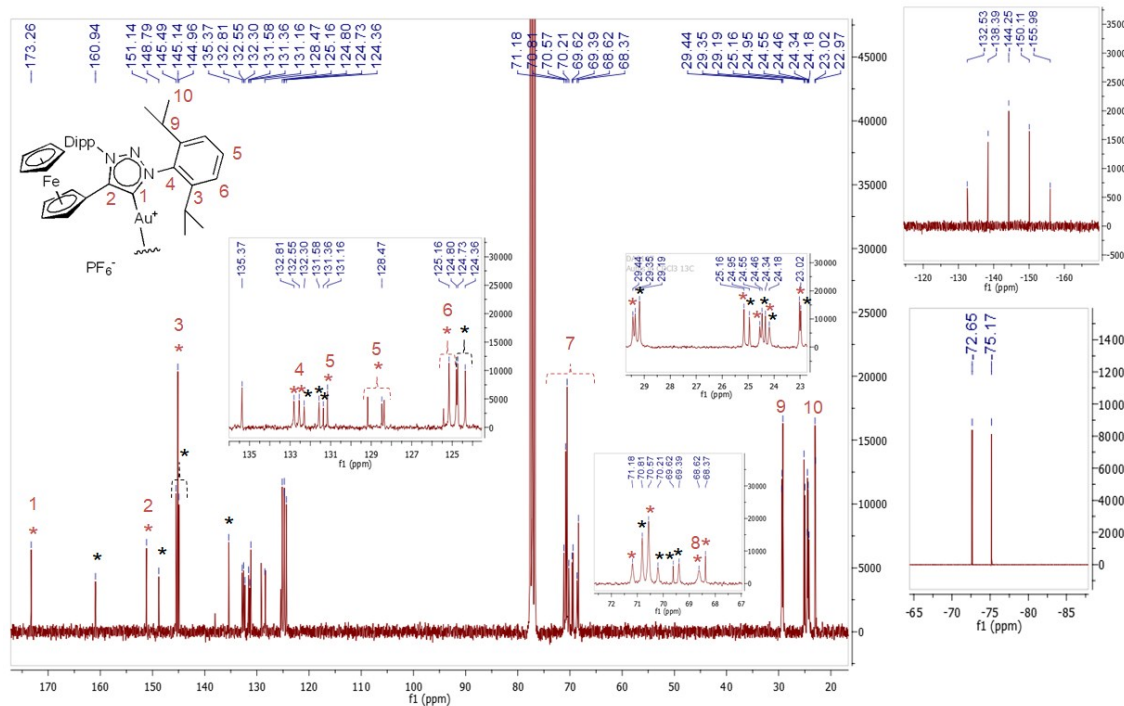


Figure S2. The  $^{13}\text{C}$  NMR spectrum showing the presence of two gold(I) complexes, where (\*) denotes the peaks belonging to the cationic biscarbene gold(I) complex (**1**) and (\*) denotes complex **2** in solvent  $\text{CDCl}_3$ . The  $^{31}\text{P}$  NMR spectrum is shown top right, and the  $^{19}\text{F}$  NMR spectrum bottom right.

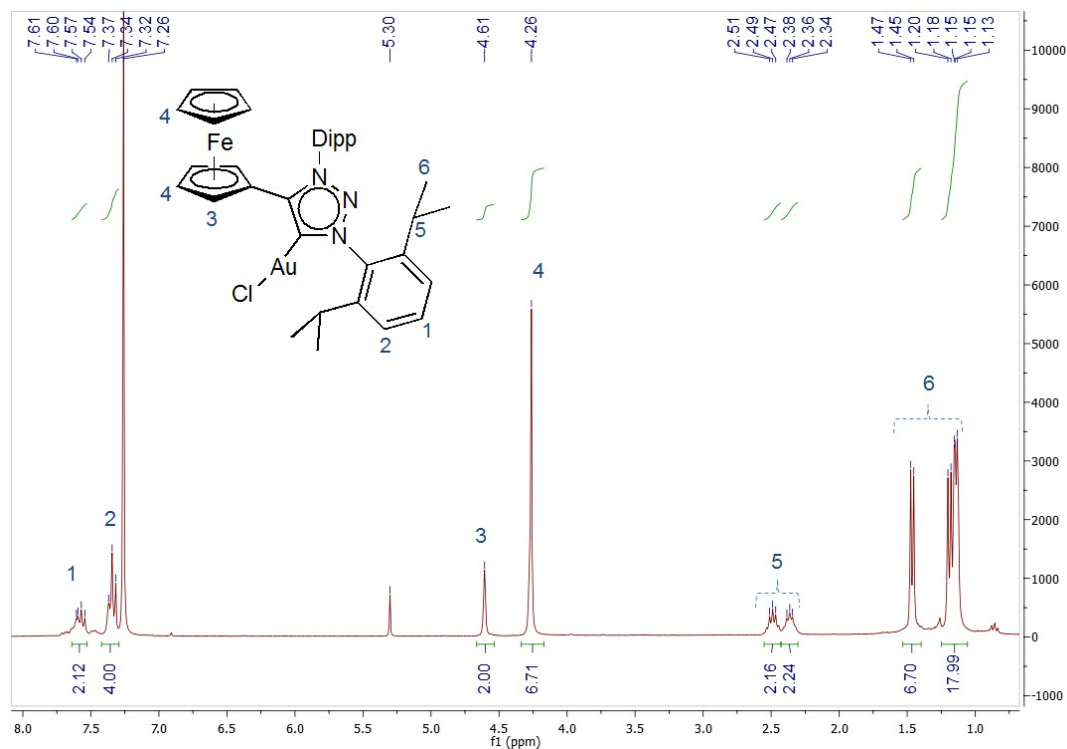


Figure S3. The  $^1\text{H}$  NMR spectrum of the triazolylidene chlorido gold(I) complex **2** in solvent  $\text{CDCl}_3$ .

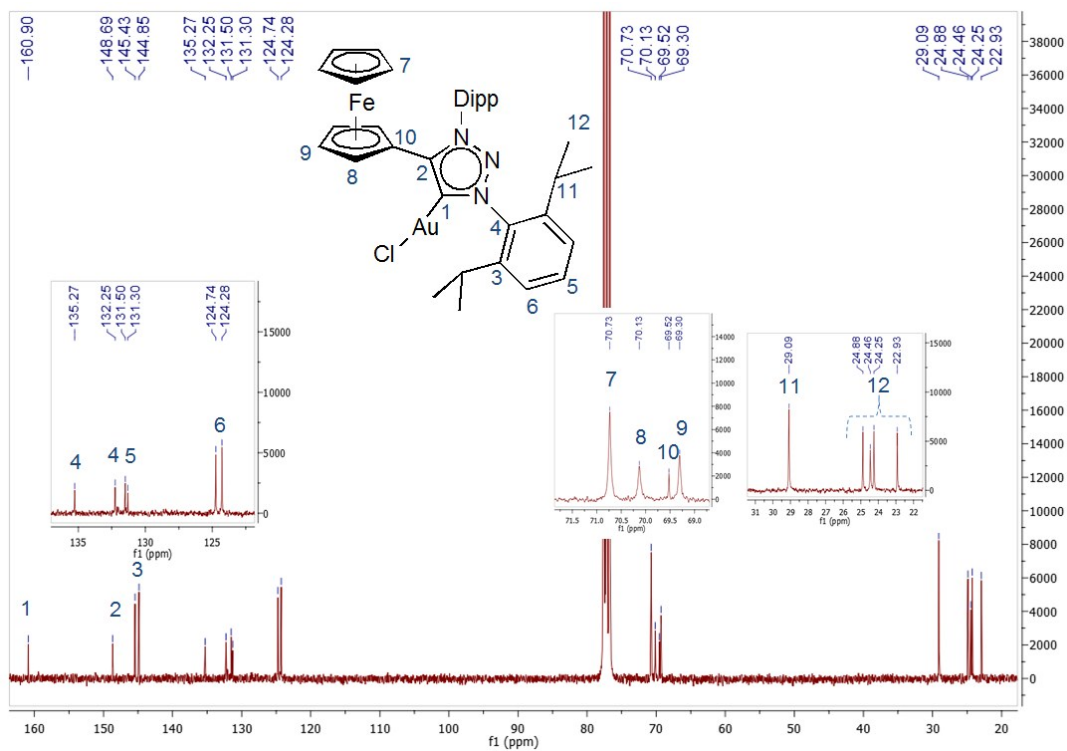


Figure S4. The  $^{13}\text{C}$  NMR spectrum of the triazolylidene chlorido gold(I) complex **2** in solvent  $\text{CDCl}_3$ .

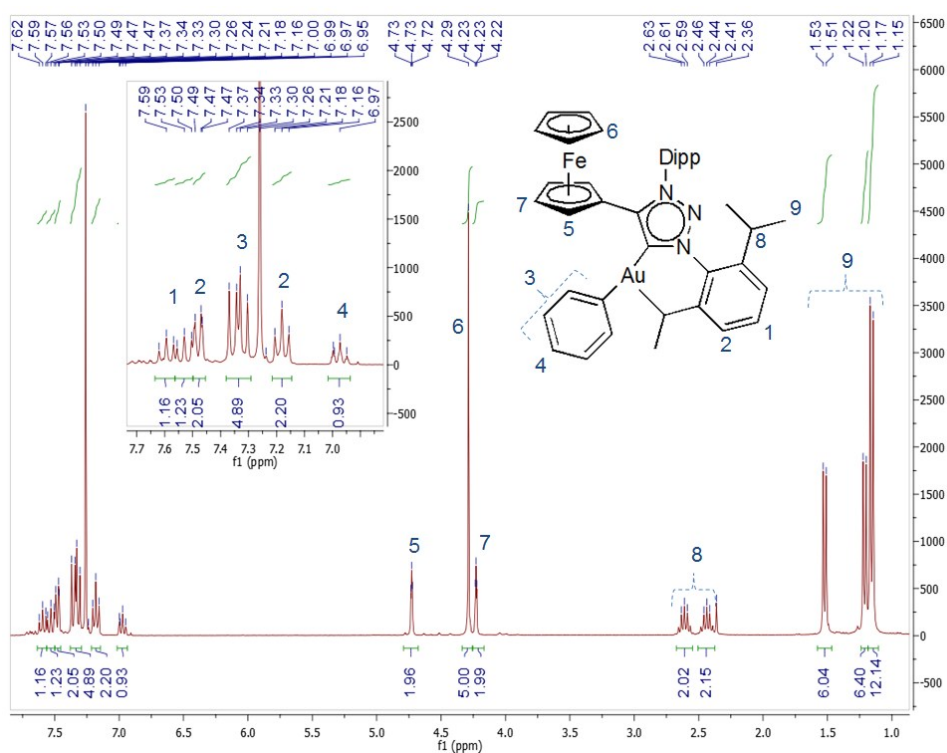


Figure S5. The  $^1\text{H}$  NMR spectrum of the triazolylidene phenyl gold(I) complex **3** in solvent  $\text{CDCl}_3$ .

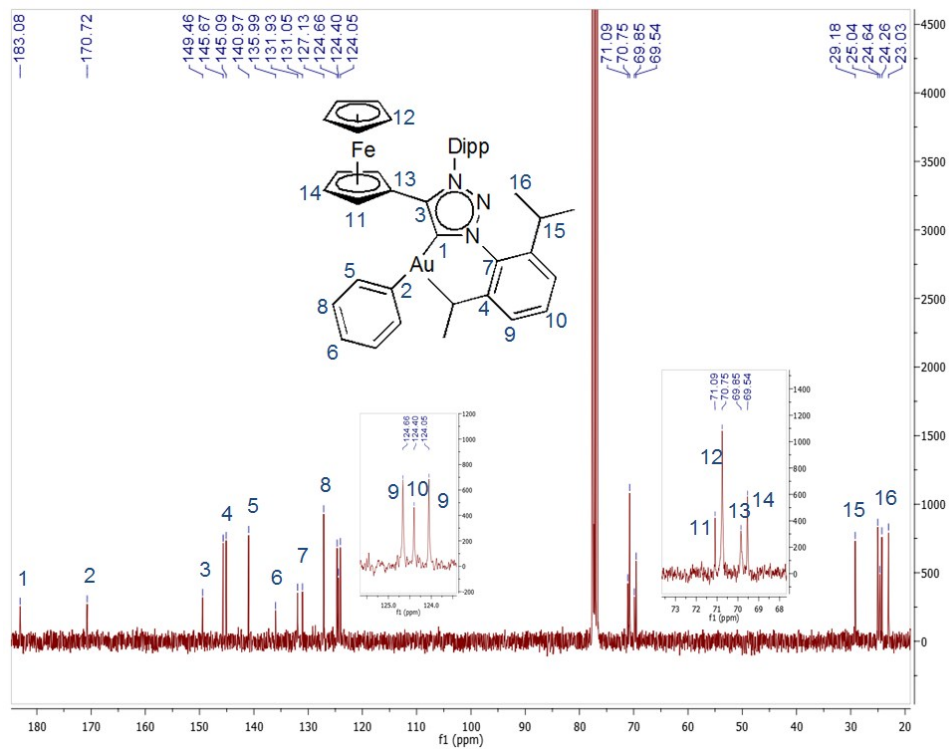


Figure S6. The  $^{13}\text{C}$  NMR spectrum of the triazolylidene phenyl gold(I) complex **3** in solvent  $\text{CDCl}_3$ .

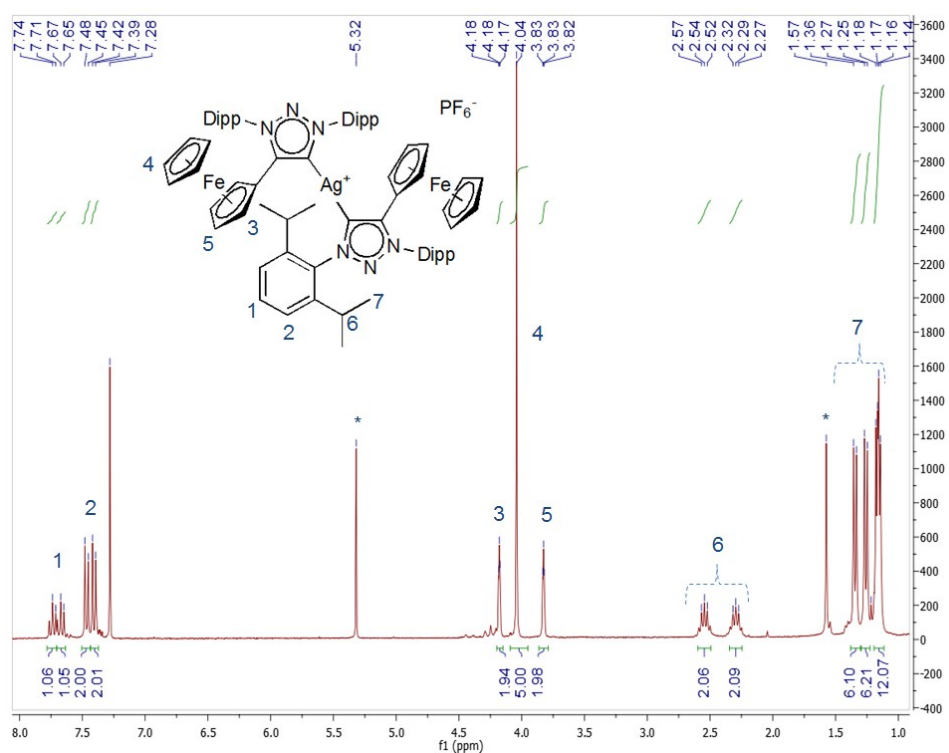


Figure S7. The  $^1\text{H}$  NMR spectrum of the cationic bis(triazolylidene) silver(I) complex **4** in solvent  $\text{CDCl}_3$ .



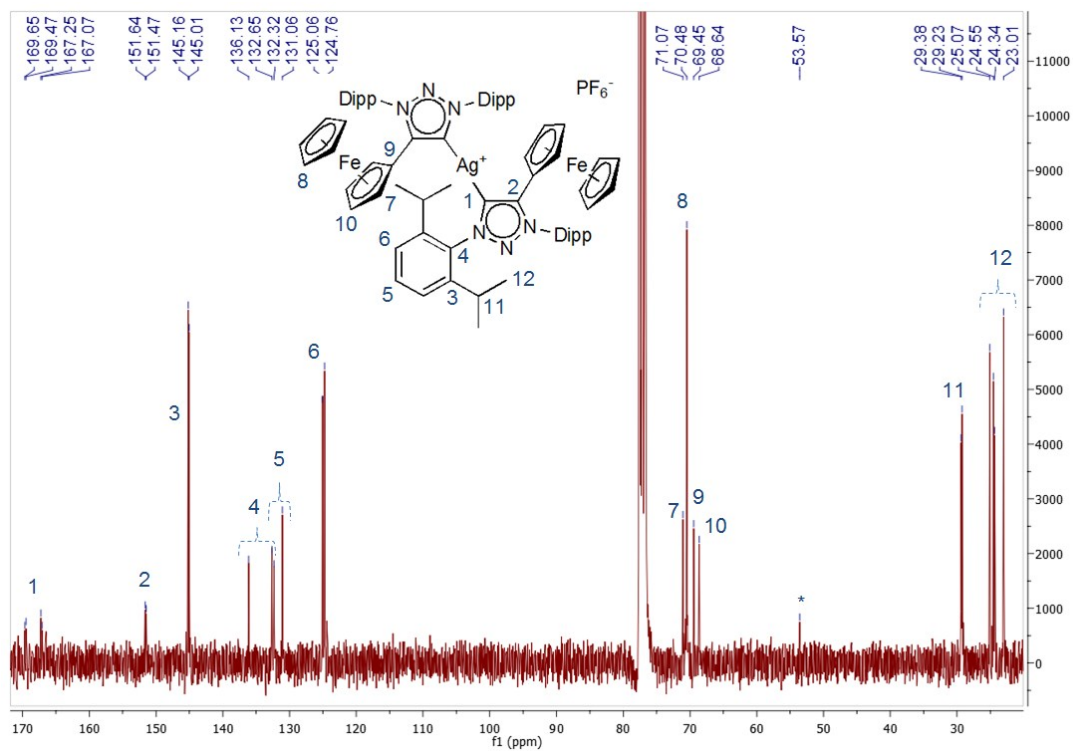


Figure S8. The  $^{13}\text{C}$  NMR spectrum of the cationic bis(triazolylidene) silver(I) complex **4** in solvent  $\text{CDCl}_3$ .

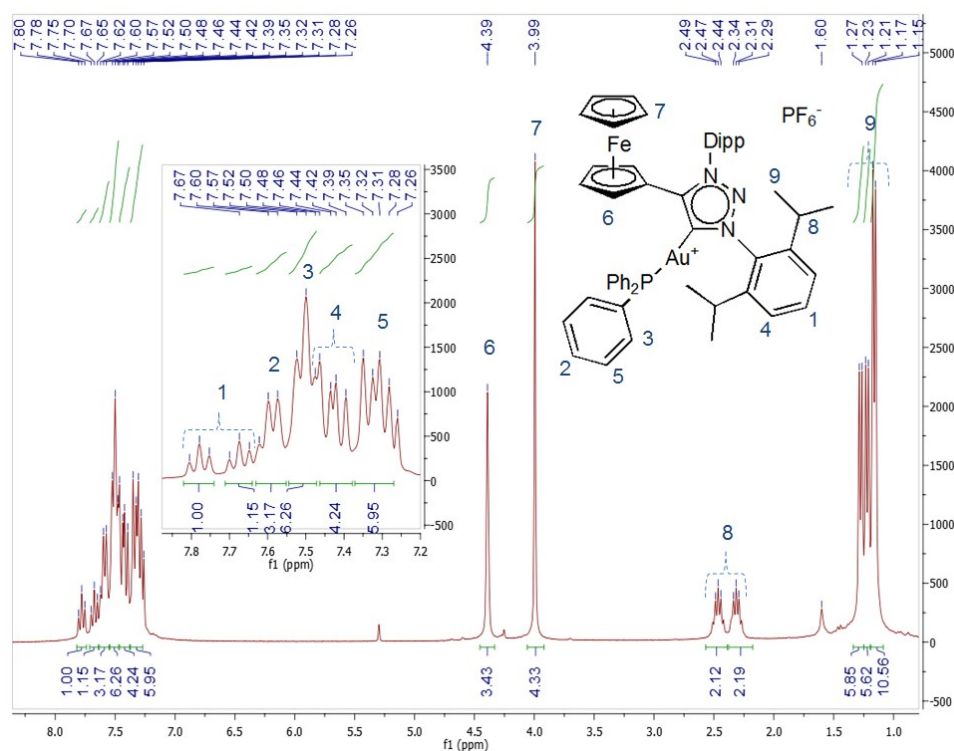


Figure S9. The  $^1\text{H}$  NMR spectrum of the cationic triazolylidene triphenylphosphine gold(I) complex **5** in solvent  $\text{CDCl}_3$ .

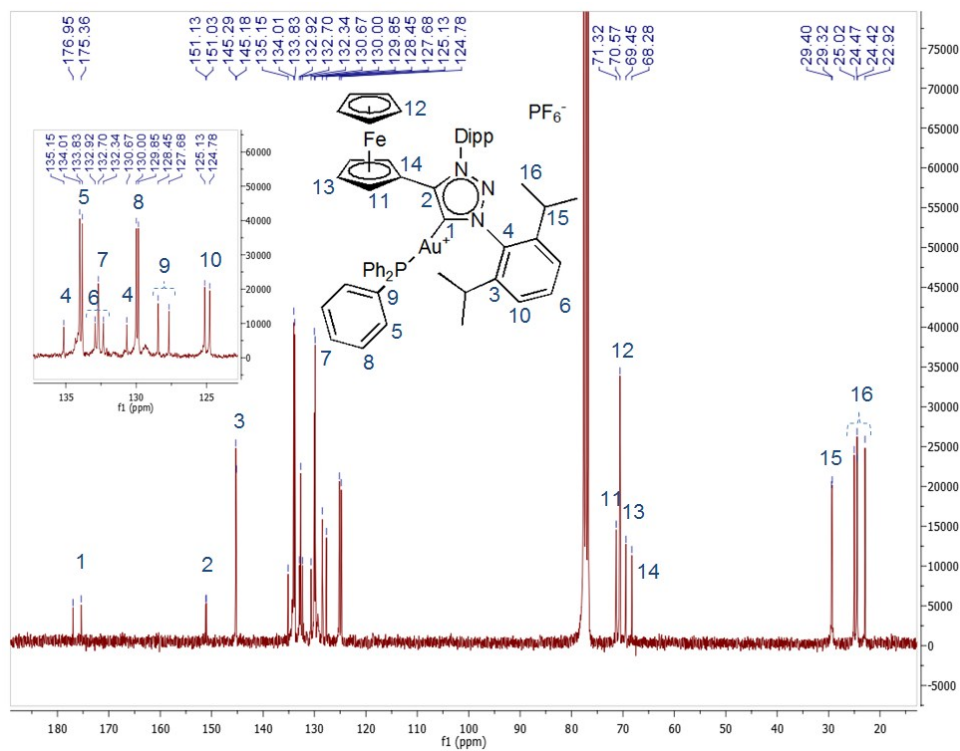


Figure S10. The <sup>13</sup>C NMR spectrum of the cationic triazolylidene triphenylphosphine gold(I) complex **5** in solvent CDCl<sub>3</sub>.

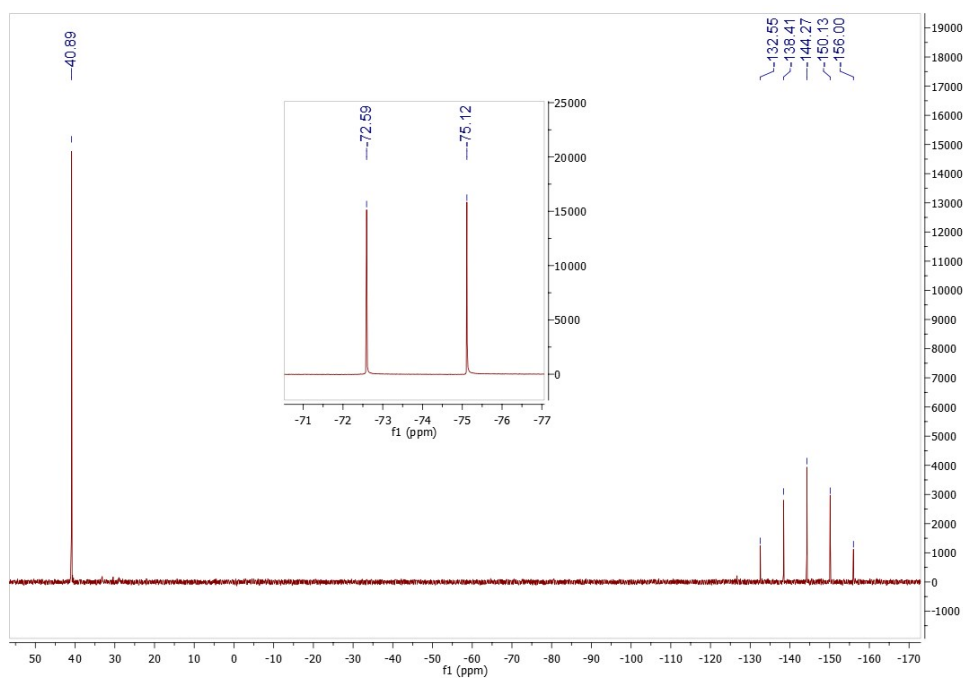


Figure S11. The <sup>31</sup>P NMR and the <sup>19</sup>F NMR (inside block) spectra of cationic triazolylidene triphenylphosphine gold(I) complex **5** in solvent CDCl<sub>3</sub>.



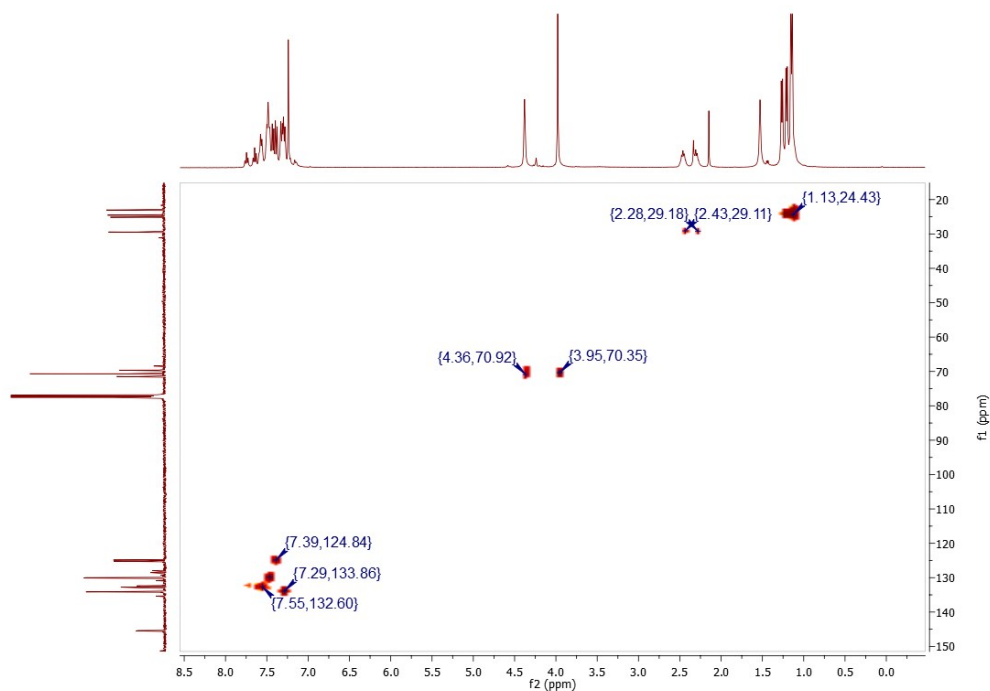


Figure S12. The HSQC (2D NMR) spectrum of the cationic triazolylidene triphenylphosphine gold(I) complex **5** in solvent  $\text{CDCl}_3$ .

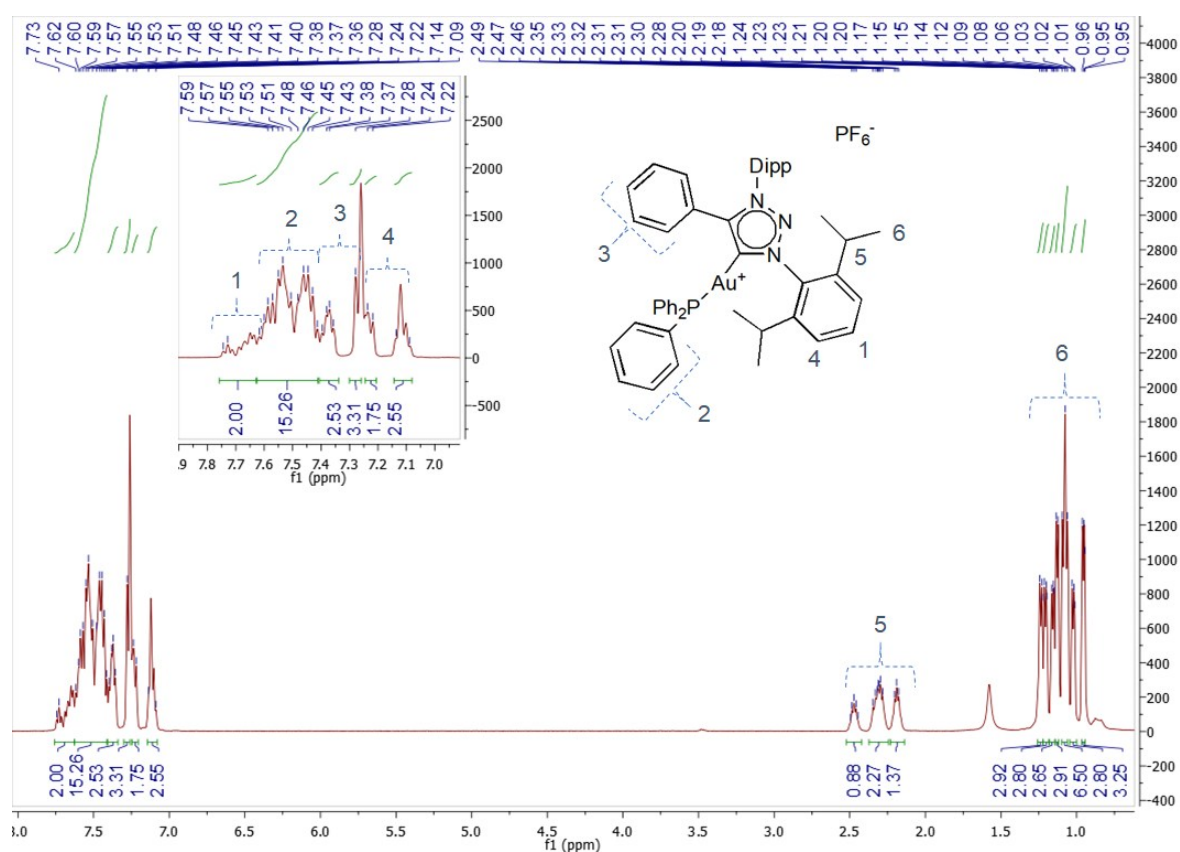


Figure S13. The  $^1\text{H}$  NMR spectrum of the cationic triazolylidene triphenylphosphine gold(I) complex **6** in solvent  $\text{CDCl}_3$ .

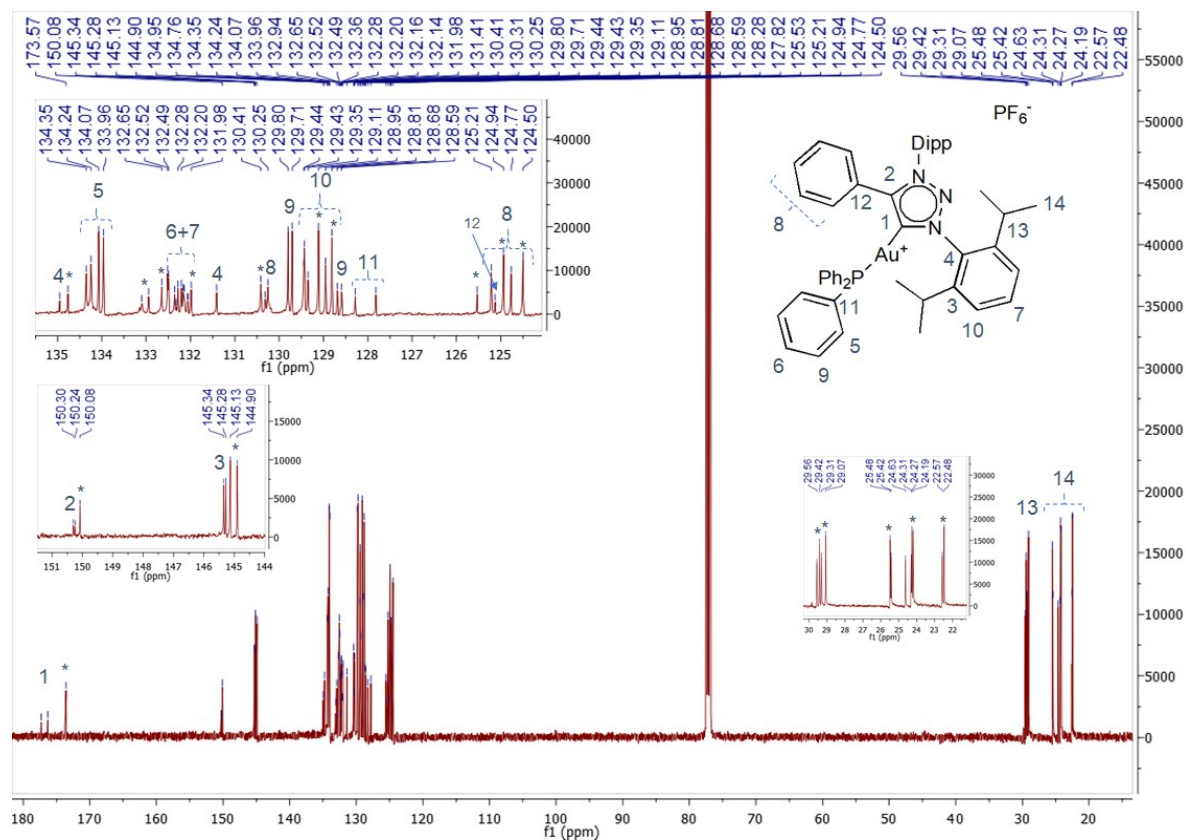
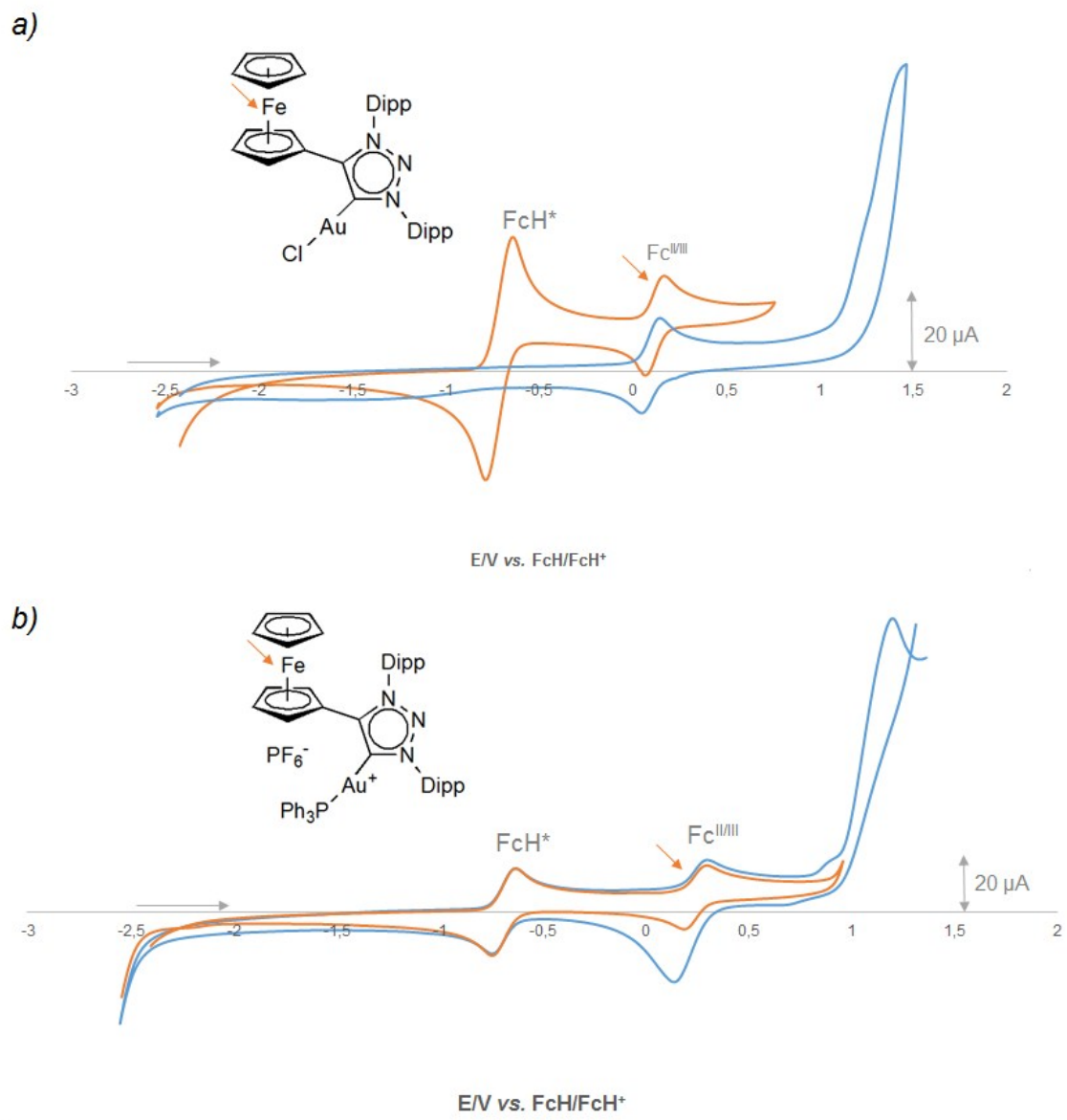


Figure S14. The  $^{13}\text{C}$  NMR spectrum of the cationic triazolylidene triphenylphosphine gold(I) complex **6** in solvent  $\text{CDCl}_3$ . The set of signals denoted (\*) belongs to **bis[(1,3-bis(2,6-diisopropylphenyl)-4-phenyl-1,2,3-triazol-5-ylidene) gold(I) hexafluorophosphate** complex (decomposition product).

### III. Cyclic voltammograms of complexes **2** and **5**



IV. Stability determination of **2** and **5** in  $d_6$ -DMSO by  $^1\text{H}$ -NMR spectroscopy and mass spectrometry

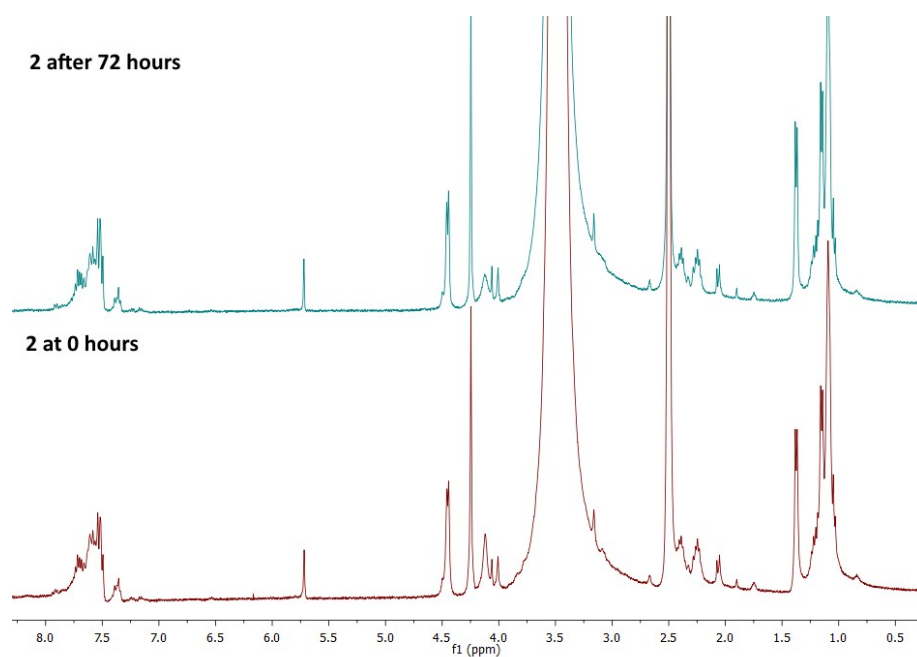


Figure S16.  $^1\text{H}$  NMR spectra of **2** in  $d_6$ -DMSO:  $\text{H}_2\text{O}$  (1:1) solution, recorded at 0 and 72 hours respectively.

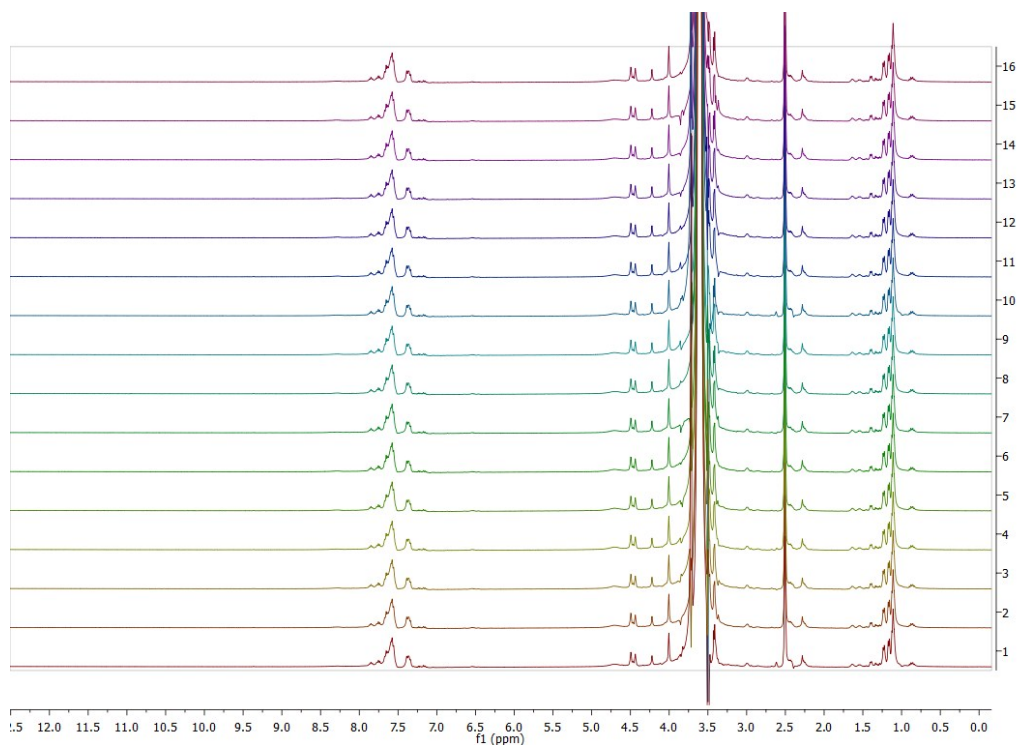


Figure S17.  $^1\text{H}$  NMR spectra of **5** in  $d_6$ -DMSO:  $\text{H}_2\text{O}$  (1:1) solution, recorded over 180 min time intervals, over a total period of 48 h.

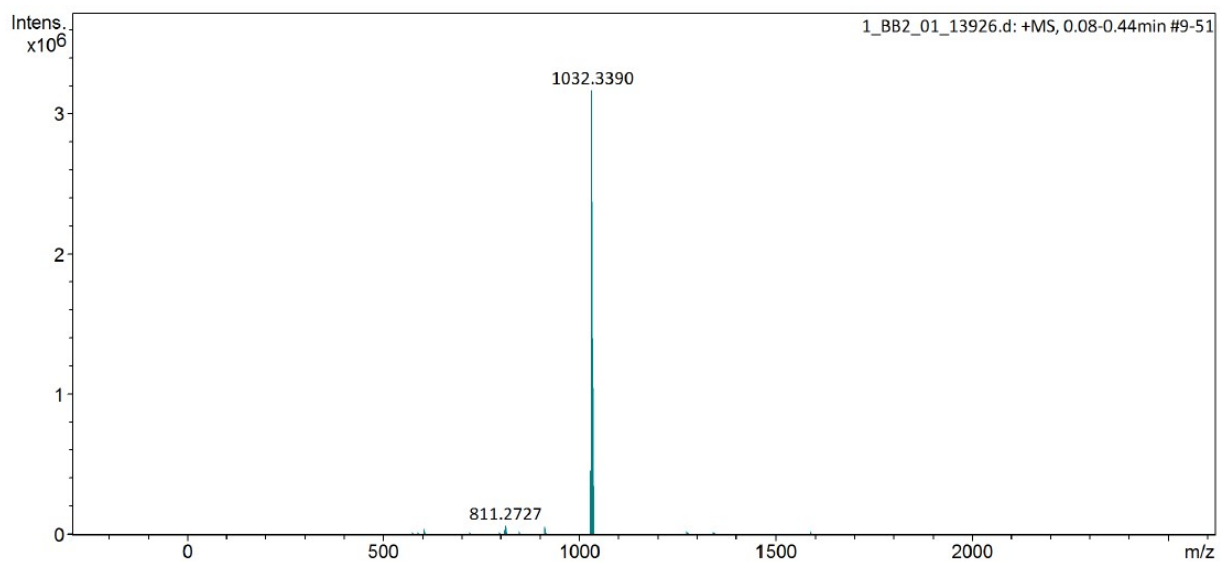


Figure S18. HRMS spectrum of **5** stored in DMSO at room temperature after 5 days.

V. Preliminary cytotoxicity screening of ligand salts **A**, **B** and complexes **2–6**

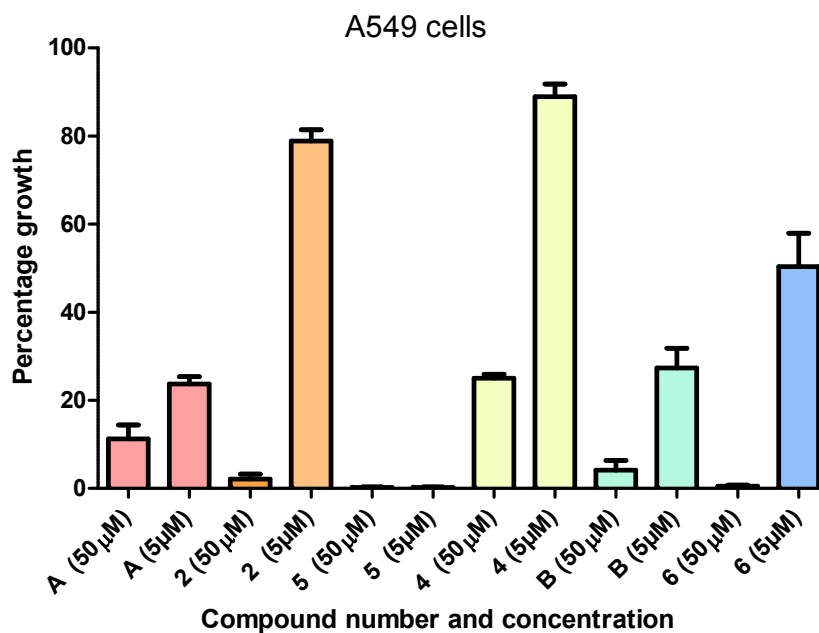


Figure S19. The percentage cell growth of A549 cells screened against ligand salts **A**, **B** and metal complexes **2–6** at 5 μM and 50 μM.

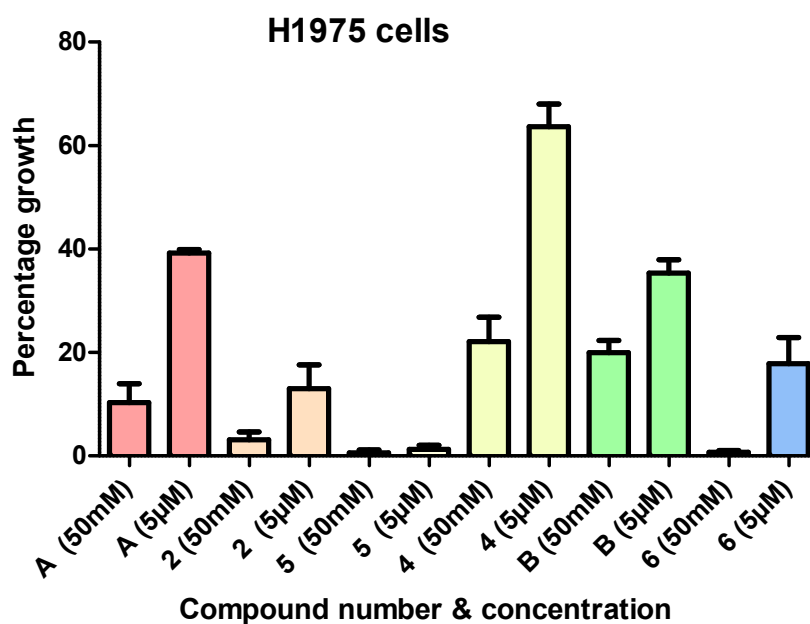


Figure S20. The percentage cell growth of H1975 cells screened against ligand salts **A**, **B** and metal complexes **2–6** at 5 μM and 50 μM.



VI. Fluorescence microscopy study on A549 cell line treated with 5

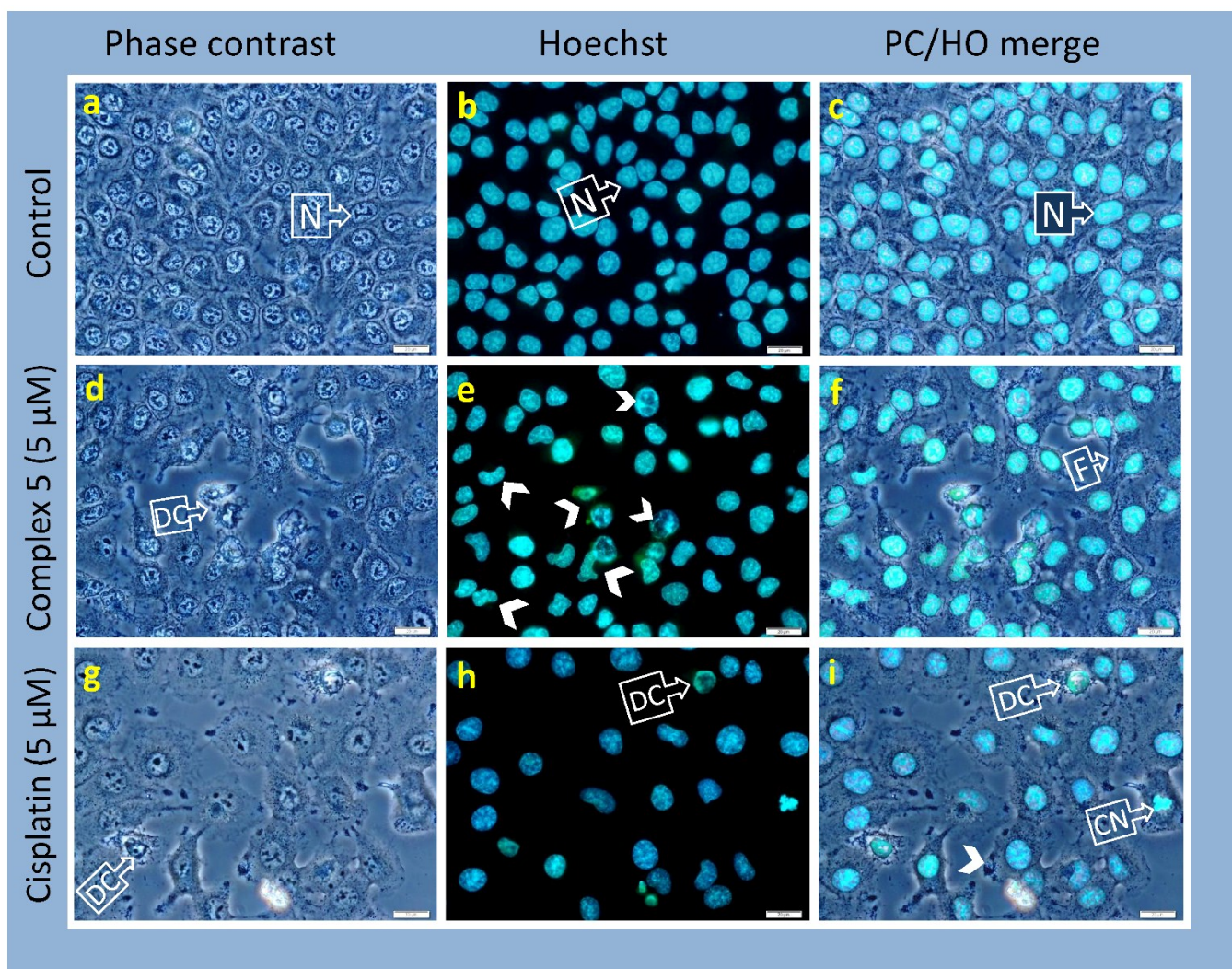


Figure. **S21**. A549 cells were treated with **5** at a concentration of 5  $\mu$ M for 18 hours. Cells show a change in nuclear morphology and the absence of necrosis. Staining the live unfixed cells with Hoechst and ethidium bromide shows the absence of necrosis and changes in nuclear morphology as indicated in e and f, rounding of cells and the formation of filopodia. Cells were viewed under 400 X magnification with an Olympus BX41 epifluorescence microscope and images were captured with an Olympus DP72 digital camera and processed using the Olympus CellSens software package.

Scalebar: 20 microns. Legend: N=nucleus, DC= detached cell, F=filopodia, CN=contracted nucleus

Untreated A549 cells typically have an oval nucleus; when treated with **5** there was a change in the shape of many nuclei as indicated by the arrows (Fig. S19, panel e). Although the cells appeared to be intact (Fig. S19, panel d) many nuclei lost their typical ovoid shape. Changes to the normal nuclear morphology is followed by cell death. Filopodia were also visible in the cells that were affected by **5**. Cisplatin treated cells were more granular (indicated by the arrow in Figure S19 panel i) than untreated cells and the cell density was decreased when compared to the control cells.

## VII. Crystal Structure Data for complexes **2–6**

### *Crystal data for 2*

$C_{36}H_{43}AuClFeN_3$  (M = 806.00 g/mol): triclinic, space group P-1,  $a = 12.4522(10)$  Å,  $b = 12.5760(9)$  Å,  $c = 13.8786(8)$  Å,  $\alpha = 116.729(2)^\circ$ ,  $\beta = 95.812(2)^\circ$ ,  $\gamma = 106.593(2)^\circ$ ,  $V = 1792.1(2)$  Å<sup>3</sup>,  $Z = 2$ ,  $T = 150$  K,  $D_{calc} = 1.494$  g/cm<sup>3</sup>,  $\mu(\text{MoK}\alpha) = 4.594$  mm<sup>-1</sup>, 75451 reflections measured ( $2.36^\circ \leq 2\theta \leq 28.28^\circ$ ), 8886 unique [ $R_{int} = 0.0848$ ,  $R_{sigma} = 0.0530$ ] which were used in all calculations. The final  $R_1$  was 0.0342 ( $I > 2\sigma(I)$ ) and  $wR_2$  was 0.0665 (all data).

### *Crystal data for 3*

$C_{42}H_{48}AuFeN_3$  (M = 847.65 g/mol): monoclinic, space group  $P2_1/n$ ,  $a = 10.7939(19)$  Å,  $b = 22.047(4)$  Å,  $c = 15.733(3)$  Å,  $\alpha = 90^\circ$ ,  $\beta = 90.785(7)^\circ$ ,  $\gamma = 90^\circ$ ,  $V = 3743.8(12)$  Å<sup>3</sup>,  $Z = 4$ ,  $T = 150$  K,  $D_{calc} = 1.504$  g/cm<sup>3</sup>,  $\mu(\text{MoK}\alpha) = 4.334$  mm<sup>-1</sup>, 136890 reflections measured ( $4.548^\circ \leq 2\theta \leq 54.496^\circ$ ), 8341 unique [ $R_{int} = 0.0490$ ,  $R_{sigma} = 0.0183$ ] which were used in all calculations. The final  $R_1$  was 0.0210 ( $I > 2\sigma(I)$ ) and  $wR_2$  was 0.0500 (all data).

### *Crystal data for 4*

$C_{73}H_{88}AgCl_2F_6Fe_2N_6P$  (M = 1484.93 g/mol): triclinic, space group P-1,  $a = 10.6222(5)$  Å,  $b = 15.5756(8)$  Å,  $c = 22.4569(10)$  Å,  $\alpha = 84.938(3)^\circ$ ,  $\beta = 82.430(2)^\circ$ ,  $\gamma = 73.391(2)^\circ$ ,  $V = 3524.2(3)$  Å<sup>3</sup>,  $Z = 2$ ,  $T = 150$  K,  $D_{calc} = 1.399$  g/cm<sup>3</sup>,  $\mu(\text{MoK}\alpha) = 0.841$  mm<sup>-1</sup>, 187604 reflections measured ( $4.192^\circ \leq 2\theta \leq 66.458^\circ$ ), 27000 unique [ $R_{int} = 0.0445$ ,  $R_{sigma} = 0.0315$ ] which were used in all calculations. The final  $R_1$  was 0.0374 ( $I > 2\sigma(I)$ ) and  $wR_2$  was 0.0921 (all data).

### Crystal data for 5

$C_{55}H_{59}AuCl_3F_6FeN_3P_2$  (M = 1298.16 g/mol): monoclinic, space group  $P2_1/c$ ,  $a = 17.8411(8)$  Å,  $b = 11.8539(6)$  Å,  $c = 26.3035(13)$  Å,  $\alpha = 90^\circ$ ,  $\beta = 97.359(2)^\circ$ ,  $\gamma = 90^\circ$ ,  $V = 5517.0(5)$  Å<sup>3</sup>,  $Z = 4$ ,  $T = 150$  K,  $D_{calc} = 1.563$  g/cm<sup>3</sup>,  $\mu(MoK\alpha) = 3.182$  mm<sup>-1</sup>, 221967 reflections measured ( $4.524^\circ \leq 2\theta \leq 57.852^\circ$ ), 14373 unique [ $R_{int} = 0.0522$ ,  $R_{sigma} = 0.0222$ ] which were used in all calculations. The final  $R_1$  was 0.0244 ( $I > 2\sigma(I)$ ) and  $wR_2$  was 0.0523 (all data).

### Crystal data for 6

$C_{50}H_{54}AuF_6N_3P_2$  (M = 1069.87 g/mol): monoclinic, space group  $P2_1/n$ ,  $a = 12.2845(10)$  Å,  $b = 34.873(3)$  Å,  $c = 22.3896(17)$  Å,  $\alpha = 90^\circ$ ,  $\beta = 101.251(2)^\circ$ ,  $\gamma = 90^\circ$ ,  $V = 9407.3(13)$  Å<sup>3</sup>,  $Z = 8$ ,  $T = 173$  K,  $D_{calc} = 1.511$  g/cm<sup>3</sup>,  $\mu(MoK\alpha) = 3.257$  mm<sup>-1</sup>, 190094 reflections measured ( $5.708^\circ \leq 2\theta \leq 56.794^\circ$ ), 23549 unique [ $R_{int} = 0.0469$ ,  $R_{sigma} = 0.0317$ ] which were used in all calculations. The final  $R_1$  was 0.0677 ( $I > 2\sigma(I)$ ) and  $wR_2$  was 0.1448 (all data).

## VIII. References

1. APEX3, (including SAINT and SADABS), BrukerAXS Inc., Madison, WI, 2017.; G. M. Sheldrick, Acta Crystallogr., Sect. A: Found. Crystallogr., 2015, 71, 3–8.; G. M. Sheldrick, Acta Crystallogr., Sect. C: Cryst. Struct. Commun., 2015, 71, 3–8.; O.V. Dolomanov, L.J. Bourhis, R.J. Gildea, J.A.K. Howard, H. Puschmann, J. Appl. Cryst., 2009, 42, 339-341.