

1 Supplementary Information

2 **Table S1.** PCR based primer specificity testing on 55 different fungal DNAs.

	Fungi	<i>Aspergillus</i>	<i>Penicillium</i>	<i>Cladosporium</i>	<i>Alternaria</i>	<i>Trichoderma</i>	<i>Aureobasidium</i>	<i>Stachybotrys</i>	<i>Tritirachium</i>	<i>Eurotiales</i>	<i>Saccharomycetale</i>	<i>Mucorales</i>	<i>Basidiomycetes</i>	<i>Ascomycetes</i>
<i>Acremonium antarcticum</i>	+	-	-	-	-	-	-	-	-	x	~	-	-	+
<i>Alternaria alternata</i>	+	-	-	-	+	-	~	-	-	x	~	-	~	+
<i>Alternaria botrytis/oudemansii</i>	+	-	-	-	~	-	-	-	-	-	-	-	~	+
<i>Ampelomyces quisqualis</i>	+	-	-	-	-	§	-	-	-	§	-	-	~	+
<i>Annulohypoxylon cohaerens</i>	+	x	-	-	-	§	-	-	-	-	~	-	~	+
<i>Anthracocystis flocculosa</i>	+	-	-	-	-	-	-	-	-	x	-	-	+	~
<i>Aspergillus niger</i>	+	+	-	-	-	-	-	-	-	+	-	-	-	+
<i>Aspergillus versicolor</i>	+	+	-	-	-	-	-	-	-	+	-	-	-	+
<i>Aureobasidium pullulans</i>	+	-	-	-	-	-	+	-	-	-	-	-	-	+
<i>Bjerkandera adusta</i>	+	-	-	-	-	-	-	-	-	x	-	-	+	+
<i>Botryotinia fuckeliana</i>	+	-	-	-	-	-	-	-	-	+	~	-	-	+
<i>Botrytis cinerea</i>	+	-	-	-	-	-	-	-	-	~	-	-	-	+
<i>Candida albicans</i>	+	-	-	-	-	-	-	-	-	-	+	-	-	+
<i>Cephalotrichum microsporum</i>	+	-	-	-	-	-	-	~	-	-	-	-	-	+
<i>Chaetomium globosum</i>	+	-	-	-	-	-	-	-	-	x	-	-	-	+
<i>Cladosporium herbarum-complex</i>	+	-	-	+	-	-	-	-	-	§	-	-	-	+
<i>Coprinopsis strossmayeri</i>	+	x	§	-	-	§	-	-	§	x	+	§	+	+
<i>Cyphellophora olivacea</i>	+	§	-	-	-	-	-	-	-	x	-	-	-	+
<i>Dacrymyces stillatus</i>	+	-	-	-	~	-	-	-	-	x	-	-	+	+
<i>Didymella</i>	+	-	-	-	-	-	-	-	-	§	-	-	~	+

<i>Sistotrema brinkmannii</i>	+	§	§	-	-	-	-	-	-	~	-	+	+
<i>Solicoccozyma terricola</i>	+	-	-	-	-	-	-	-	-	-	-	+	~
<i>Stachybotrys chartarum</i>	+	-	-	-	-	-	-	+	-	-	-	-	~
<i>Tilletiopsis washingtonensis</i>	+	-	-	-	-	-	-	-	-	~	-	+	+
<i>Trametes versicolor</i>	+	-	-	-	-	-	-	-	-	x	~	-	+
<i>Trichocladium asperum</i>	+	-	§	-	-	-	-	-	-	§	-	-	+
<i>Trichoderma harzianum</i>	+	-	-	-	-	+	-	-	-	§	-	-	§
<i>Tritirachium oryzae</i>	+	-	-	-	-	-	-	-	+	-	~	-	~
<i>Ustilago maydis</i>	+	-	-	-	-	-	-	-	-	-	-	+	+
<i>Wallemia sebi</i>	+	-	-	-	-	-	-	-	-	+	-	+	~

3 + positive; - negative; ~ weak band; § different fragment size; x smear;

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6 **Table S2.** qPCR primer efficiency, qPCR correlation coefficient, coverage of dynamic
7 range and detection limits of the indoor mold detection system's core assays
8 measured on serial dilutions of fungal DNA standard mixtures.

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Primer set	Efficiency [%]	r ² value	Dynamic range	Detection limit [pg DNA]
Fungi	90 – 92	≥ 0.9991	6-log	0.0300
Aspergillus	89 – 94	≥ 0.9975	4-log	0.2143
Penicillium	84 – 94	≥ 0.9970	6-log	0.0021
Cladosporium	92 – 101	≥ 0.9995	6-log	0.0021
Alternaria	91 – 99	≥ 0.9978	6-log	0.0021
Trichoderma	84 – 91	≥ 0.9981	6-log	0.0021
Aureobasidium	84 – 88	≥ 0.9964	6-log	0.0021
Stachybotrys	93 – 95	≥ 0.9989	4-log	0.2143
Tritirachium	90 – 96	≥ 0.9971	6-log	0.0021
Eurotiales	97 – 98	≥ 0.9991	6-log	0.0043
Saccharomycetales	82 – 89	≥ 0.9970	6-log	0.0021
Mucorales	82 – 101	≥ 0.9901	6-log	0.0021

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Basidiomycetes	88 – 92	≥ 0.9989	6-log	0.0043
Ascomycetes	83 – 88	≥ 0.9986	5 – 6-log	0.0236

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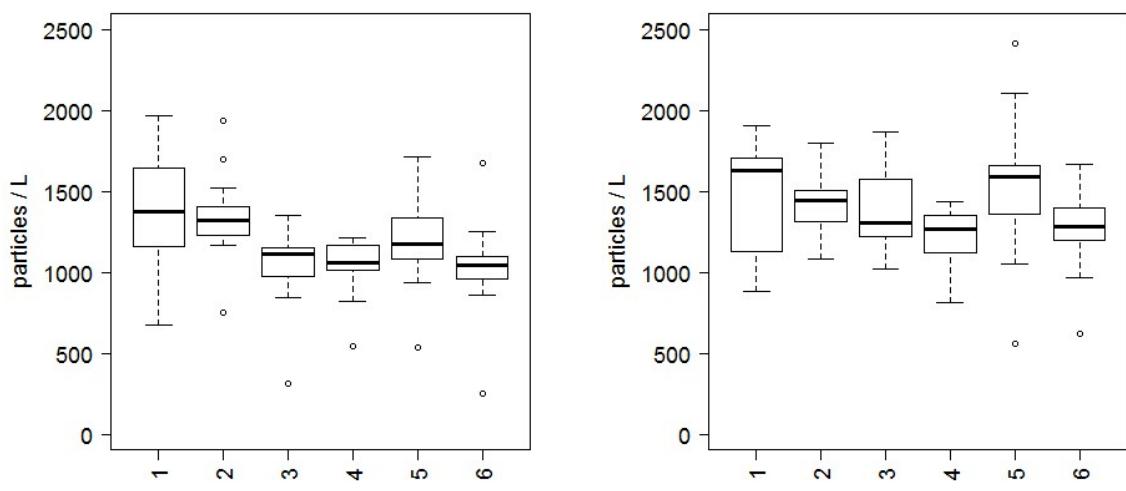
12

13 **Table S3.** Differences on qPCR detection of samples with single fungal DNAs
 14 compared to complex DNA mixtures consisting of 13 fungal species illustrated with
 15 qPCR measurements conducted with the specific assays.

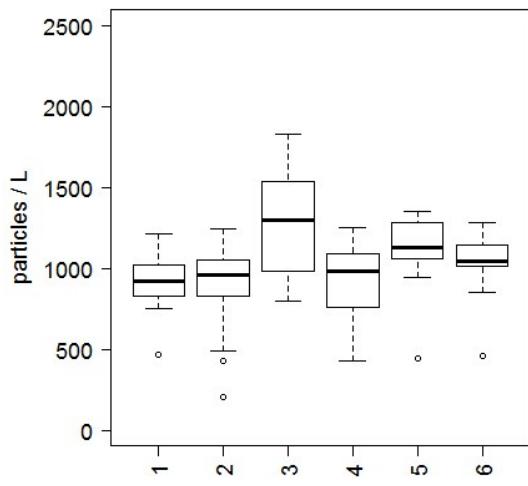
Assay	SINGLE		R²	Eff. [%]	MIX		R²	Eff. [%]	Diff. Eff. [%]
	Mean [Cq]	Std. Dev. [Cq]			Mean [Cq]	Std. Dev. [Cq]			
<i>Alternaria</i>	12.61 – 28.02	0.03 – 0.27	0.9996	82.0	12.62 – 26.96	0.02 – 0.22	0.985	92.4	10.4
	11.47 – 23.08	0.01 – 0.12			11.81 – 21.70	0.08 – 0.24			
<i>Aspergillus</i>	15.16 – 28.49	0.05 – 0.15	0.9987	102.0	14.42 – 28.49	0.04 – 0.22	0.985	92.8	-9.2
	17.65 – 31.66	0.00 – 0.12			18.25 – 32.28	0.07 – 0.94			
<i>Aureobasidium</i>	14.97 – 29.71	0.00 – 1.11	0.9914	89.0	14.91 – 28.76	0.01 – 0.35	0.991	96.0	7.0
	18.77 – 31.39	0.03 – 0.63			18.26 – 31.76	0.07 – 0.41			
<i>Basidiomycetes</i>	18.60 – 31.73	0.01 – 0.88	0.9992	105.6	18.59 – 32.11	0.03 – 0.54	0.998	96.3	-6.2
	16.88 – 31.12	0.01 – 0.23			16.84 – 30.69	0.01 – 0.35			
<i>Cladosporium</i>	17.02 – 32.76	0.09 – 0.23	0.9996	79.9	16.70 – 32.38	0.03 – 0.11	0.993	80.4	0.5
	14.68 – 27.72	0.02 – 0.26			16.30 – 30.93	0.02 – 0.17			
<i>Mucorales</i>	15.74 – 29.30	0.03 – 0.25	0.9998	98.1	15.63 – 29.34	0.04 – 0.19	0.992	97.2	-0.9
	15.99 – 29.63	0.02 – 0.20			15.78 – 30.47	0.00 – 0.37			
<i>Penicillium</i>	15.44 – 29.04	0.03 – 0.23	0.9999	96.2	15.10 – 29.44	0.02 – 0.15	0.992	90.2	-6.0
	16				16				

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20 **Fig. S1.** Particle count deviations within the bioaerosol chamber of 3 independent
 21 sampling campaigns (1 upper left; 2 upper right; 3 bottom left) during 6 bioaerosol
 22 collections with the Coriolis μ (1-6).

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24 **Table S4.** Relative abundances in % of the 50 most abundant OTUs in air samples.

25 Samples were taken in October 2015 and January 2016 from an outdoor reference, a
 26 vacant office and a wine cellar. At each campaign three samples were collected and
 27 subjected to high-throughput sequencing for fungal community analysis. Due to very
 28 low airborne fungal particles in the office in October 2015, data were obtained only

29 from one of the three samples. Data from separate samples were combined for the
 30 OTU abundance table. Total OTU numbers and Simpson's diversity index (D1=1-D)
 31 are shown for combined samples.

Sampling Site	OTU_ID	Oct 2015			Jan 2016		
		Outdoor	Office	Cellar	Outdoor	Office	Cellar
# samples		3	1	3	3	3	3
# OTUs		173	70	169	199	189	220
1-D		0,65	0,94	0,75	0,82	0,95	0,95
OTU_ID	Genus						
OTU_2	<i>Mycosphaerella</i>	567,0	38,6	217,7	381,7	82,5	122,0
OTU_1	<i>Apotrichum</i>	31,9	179,1	12,6	49,3	180,6	56,7
OTU_5	<i>Athelia</i>	0,1	4,1	402,3	0,0	0,0	1,8
OTU_6	<i>Bjerkandera</i>	0,6	0,0	9,5	141,8	31,7	90,0
OTU_7	<i>Ascochyta</i>	162,4	0,0	4,8	9,9	36,7	31,1
OTU_19	<i>Trametes</i>	0,3	10,7	0,7	95,4	36,6	86,5
OTU_11	<i>Homophron</i>	0,2	0,1	190,6	0,0	0,0	1,8
OTU_4	<i>Pseudeurotiium</i>	11,5	64,7	4,0	20,8	60,2	17,0
OTU_3	<i>Hypodiscus</i>	6,8	57,0	2,7	14,3	38,1	11,0
OTU_8	<i>Inocybe</i>	9,8	86,9	2,0	5,3	13,1	7,2
OTU_20	<i>Daedaleopsis</i>	0,0	0,0	0,0	2,1	38,1	64,1
OTU_14	<i>Alternaria</i>	41,2	0,0	8,5	28,1	1,7	0,9
OTU_15	<i>Ilyonectria</i>	5,3	18,6	1,9	8,6	30,4	6,6
OTU_9	<i>Hypodiscus</i>	2,5	40,4	1,9	6,4	12,6	5,9
OTU_13	<i>Oidiodendron</i>	4,8	15,4	1,5	6,2	24,3	8,3
OTU_10	<i>Hypodiscus</i>	2,3	25,6	1,2	4,3	17,4	4,4
OTU_16	<i>Exophiala</i>	3,1	19,8	0,8	6,4	17,9	4,4
OTU_30	<i>Penicillium</i>	0,4	0,0	0,3	0,7	4,1	45,7
OTU_17	<i>Clavulina</i>	1,9	25,1	0,5	3,1	14,1	4,0
OTU_27	<i>Penicillium</i>	0,2	6,4	6,5	0,3	1,1	22,9
OTU_37	<i>Penicillium</i>	0,0	1,0	1,3	2,0	3,8	28,5
OTU_12	<i>Saitozyma</i>	3,1	6,0	1,5	4,6	15,9	5,0
OTU_93	<i>Absidia</i>	0,0	0,0	5,7	0,0	0,0	30,4
OTU_32	<i>Mortierella</i>	1,0	25,1	0,3	2,6	5,2	1,3
OTU_24	<i>Malbranchea</i>	0,0	0,0	30,5	0,0	0,0	4,9
OTU_34	<i>Penidiella</i>	0,0	13,1	2,2	0,0	0,0	15,5
OTU_29	<i>Boletus</i>	5,0	6,3	0,9	2,5	11,9	3,1
OTU_115	<i>Peniophora</i>	0,1	0,0	0,0	11,7	1,7	15,6
OTU_67	<i>Letendraea</i>	9,4	14,5	4,5	0,4	0,2	0,0
OTU_36	<i>Zasmidium</i>	0,0	0,0	5,0	0,0	0,0	23,1
OTU_64	<i>Aspergillus</i>	0,0	12,6	6,5	0,0	1,1	5,7
OTU_86	<i>Aspergillus</i>	0,1	14,0	4,8	0,2	0,5	5,8
OTU_28	<i>Aureobasidium</i>	20,1	0,0	0,0	5,3	0,0	0,1
OTU_21	<i>Leptodontidium</i>	2,4	7,3	0,5	3,5	8,6	3,0
OTU_26	<i>Tetracladium</i>	1,2	6,1	0,6	2,6	11,0	1,9
OTU_38	<i>Trichoderma</i>	2,0	0,0	0,9	4,4	11,6	3,8
OTU_22	<i>Cryptendoxyla</i>	0,0	12,4	6,1	0,0	0,0	3,3
OTU_44	<i>Ischnoderma</i>	0,0	0,0	0,0	9,4	8,7	3,5
OTU_23	<i>Fusarium</i>	1,6	0,9	0,9	3,5	10,5	3,9
OTU_842	<i>Kendrickiella</i>	0,1	0,0	0,0	5,4	2,3	13,4
OTU_56	<i>Fellozyma</i>	2,1	5,1	0,3	1,7	9,0	2,1
OTU_75	<i>Tetracladium</i>	0,7	9,8	0,7	0,8	6,1	2,0
OTU_55	<i>Xylodon</i>	0,1	0,0	0,1	7,8	6,2	5,8
OTU_63	<i>Fusarium</i>	0,8	12,9	0,1	0,7	3,4	1,8
OTU_45	<i>Penicillium</i>	0,0	0,0	8,7	0,0	0,0	11,0
OTU_25	<i>Pezizales</i>	1,3	9,3	0,8	2,4	3,6	1,6
OTU_48	<i>Mycosphaerella</i>	0,0	0,0	0,4	0,0	0,0	17,5
OTU_1093	<i>Nectria</i>	1,4	6,3	0,3	0,8	7,8	1,3
OTU_245	<i>Fusarium</i>	0,4	14,2	0,3	0,6	1,8	0,1
OTU_320	<i>Phialophora</i>	0,1	14,9	0,0	0,1	0,8	0,7

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34 **Table S5.** Relative abundances (given in percent from detected abundances within
 35 each sample) represented in the heat map of figure 5. Bdl: Below detection limit.

Assay	<u>OUT1</u> [%]	<u>CEL1</u> [%]	<u>OFF1</u> [%]	<u>OUT2</u> [%]	<u>CEL2</u> [%]	<u>OFF2</u> [%]
<i>Aspergillus</i>	Bdl	Bdl	Bdl	Bdl	21.67	Bdl
<i>Penicillium</i>	0.94	6.06	3.57	Bdl	8.58	Bdl
<i>Cladosporium</i>	46.12	5.87	57.99	22.63	5.49	Bdl
<i>Alternaria</i>	2.50	0.30	Bdl	0.89	Bdl	Bdl
<i>Trichoderma</i>	Bdl	Bdl	15.29	Bdl	Bdl	Bdl
<i>Aureobasidium</i>	3.14	Bdl	Bdl	13.94	0.15	Bdl
<i>Stachybotrys</i>	Bdl	Bdl	Bdl	Bdl	Bdl	Bdl
Eurotiales	1.65	16.42	9.73	1.79	35.26	Bdl
Saccharomycetales	Bdl	Bdl	Bdl	Bdl	0.26	Bdl
Mucorales	Bdl	1.36	2.74	Bdl	3.57	Bdl
Basidiomycetes	45.65	69.99	10.68	60.74	25.03	100.00

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38 **Table S6.** Spearman's rank correlation of qPCR and sequencing data. Nd: Not
 39 detected.

Assay	<u>Correlation</u>	<u>p-value</u>
<i>Aspergillus</i>	0.78	0.23
<i>Penicillium</i>	0.80	0.33
<i>Cladosporium</i>	*	
<i>Alternaria</i>	1.00	0.08
<i>Trichoderma</i>	*	
<i>Aureobasidium</i>	0.80	0.33
<i>Stachybotrys</i>	Nd	
Eurotiales	1.00	0.08
Saccharomycetales	0.82	0.18
Mucorales	1.00	< 0.01
Basidiomycetes	0.80	0.33

40 * underrepresented in qPCR or sequencing data