

Reduction pathway-dependent cytotoxicity of reduced graphene oxide[†]

Qiurong Zhang,^a Xiaolei Liu,^a Hongyan Meng,^a Sijin Liu,^b and Chengdong Zhang^{*ca}

^a College of Environmental Science and Engineering, Nankai University, Tianjin 300350, China.

^b State Key Laboratory of Environmental Chemistry and Ecotoxicology, Research Center for Eco-Environmental Sciences, Chinese Academy of Sciences, Beijing 100085, China.

^c School of Environment, Beijing Normal University, Beijing 100857, China. E-mail: zhangchengdong@bnu.edu.cn

[†] Electronic supplementary information (ESI) available. See DOI:

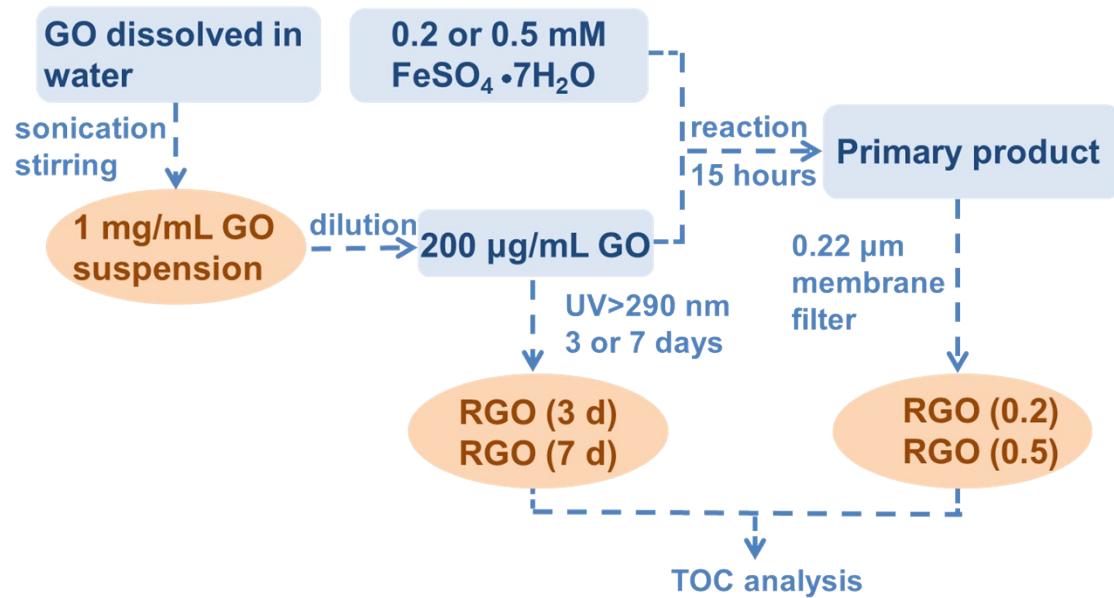


Fig. S1 Scheme of reduction procedures.

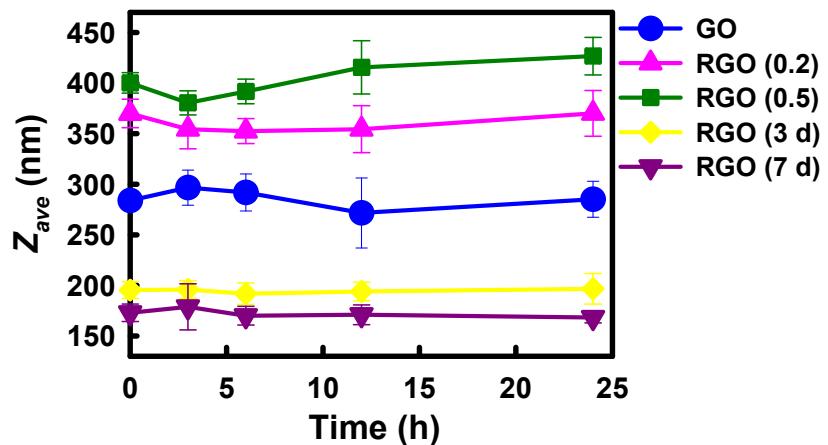


Fig. S2 The change of nanoparticle size with time. Twenty $\mu\text{g/mL}$ of GO/RGOs was incubated in culture medium and the size was analyzed by DLS ($n=3$).

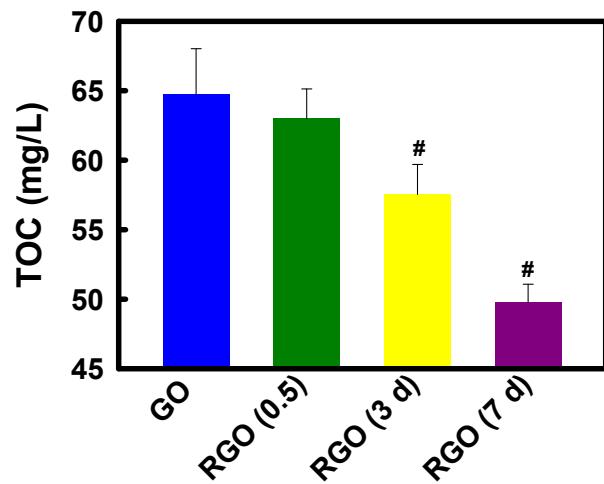


Fig. S3 Changes of TOC concentration in GO/RGOs colloidal solution after transformation (n=3). The suspension was sonicated for 30 min before analysis to ensure no precipitation occurring. #, $p < 0.05$ compared to initial GO concentration.

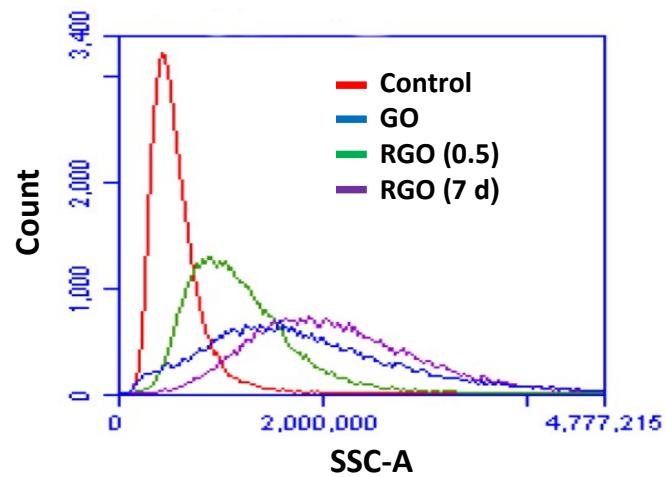


Fig. S4 Representative side-scatter histogram profile of cells after exposure to 20 $\mu\text{g/mL}$ GO/RGOs for 24 h

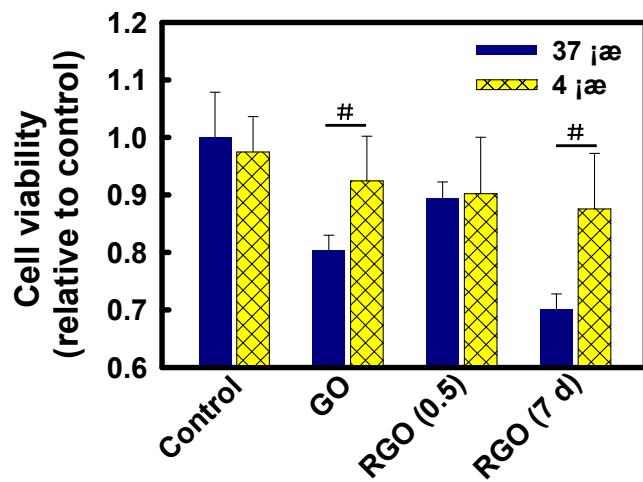


Fig. S5 The effects of temperature on cell viability in the presence of GO/RGOs. Cells were exposed to 20 $\mu\text{g/mL}$ GO/RGOs for 24 h at indicated temperature (n=5). #, $p < 0.05$ relative to the same treatment at 37 °C.

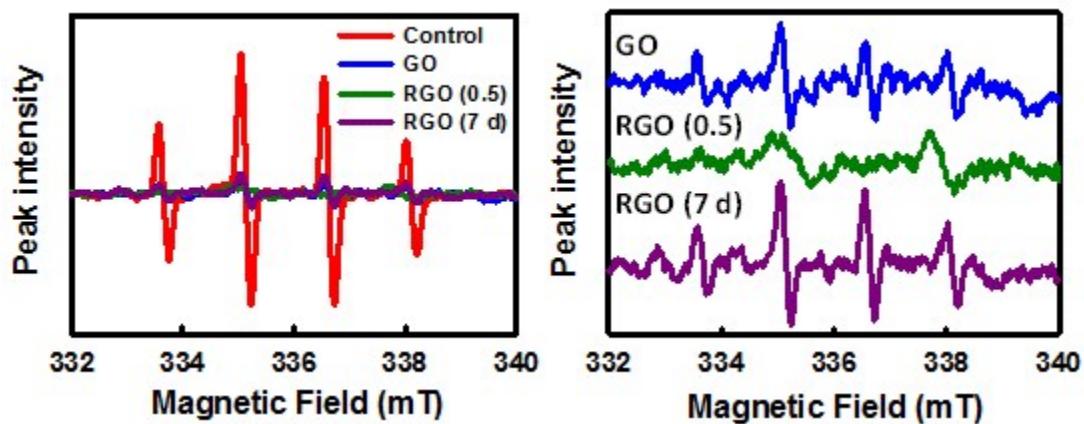


Fig. S6 EPR spectra showing the radical scavenging activity of GO/RGOs. The left panel shows the overall EPR spectra, and the right panel enlarges the differences among three nanoparticles.