Supporting information for

Unraveling the complexity of complex mixtures by combining highresolution pharmacological, analytical and spectroscopic techniques: Antidiabetic constituents in Chinese medicinal plants

Yong Zhao,^a Kenneth Thermann Kongstad,^a Yueqiu Liu,^a Chenhua He,^{a,b} and Dan Staerk*,^a

^a Department of Drug Design and Pharmacology, Faculty of Health and Medical Sciences,
 ^b College of Veterinary Medicine, Nanjing Agricultural University, Nanjing 210095, China

*Corresponding author. Address: Department of Drug Design and Pharmacology, Faculty of Health and Medical Sciences, University of Copenhagen, Universitetsparken 2, DK-2100 Copenhagen, Denmark. Tel.: +45 35336177; fax +45 35336001.

E-mail address: ds@sund.ku.dk (D. Staerk).

Table of Contents

Table S1. Plants tested for α -glucosidase, α -amylase and PTP1B inhibitory activity at a single concentration of 50 µg/mL.

Table S2. Identification of metabolites by HPLC-HRMS and NMR.

Figure S1. α-glucosidase (A) and PTP1B (B) inhibition curves of crude ethyl acetate extracts of

Persicaria bistorta (L.) Samp., Dioscorea bulbifera L., Boehmeria nivea (L.) Gaudich, and

Tinospora sagittata Gagnep.

Figure S2. UV and HRMS spectra of compound 8 obtained in the HPLC-HRMS mode.

Figure S3. ¹H-NMR spectrum of the mixture of compound 7 and 8.

Figure S4. ¹H-NMR spectrum of compound 7 and the mixture of compound 7 and 8.

Figure S5. HSQC spectrum of the mixture of compound 7 and 8.

Figure S6. HMBC spectrum of the mixture of compound 7 and 8.

Figure S7. COSY spectrum of the mixture of compound 7 and 8.

Figure S8. UV and HRMS spectra of compound 9 obtained in the HPLC-HRMS mode.

Figure S9. ¹H-NMR spectrum of compound 9.

Figure S10. HSQC spectrum of compound 9.

Figure S11. HMBC spectrum of compound 9.

Figure S12. COSY spectrum of compound 9.

Figure S13. UV and HRMS spectra of compound 10 obtained in the HPLC-HRMS mode.

Figure S14. ¹H-NMR spectrum of compound 10.

Figure S15. HSQC spectrum of compound 10.

Figure S16. HMBC spectrum of compound 10.

Figure S17. COSY spectrum of compound 10.

Figure S18. Triple high-resolution α -glucosidase/ α -amylase/PTP1B inhibition profiling of *P*. *bistorta*.

Figure S19. IC₅₀ curves of isolates and Acarbose (reference compound) in the α -glucosidase inhibition assays.

Figure S20. IC₅₀ curves of isolates and RK682 (reference compound) in the PTP1B inhibition assays.

Table S1. Plants tested for α -glucosidase, α -amylase and PTP1B inhibitory activity at a single concentration of 50 μ g/mL.

| No. | Plant | Botanical | Part used | Voucher | % i | nhibition at 50 | μ g/m L^a |
|-----------------|--|-----------------|-------------------|-----------------|-------------------|-------------------|-------------------|
| | | family | | specimen _ # | AGH | AM | PTP1B |
| 1 | Rhus chinensis Mill. | Anacardiaceae | Root | zyc003 | 2.3 | 6.4 | 95.5 |
| 2 | Oenanthe javanica (Bl.) DC. | Apiaceae | Seed | hch109 | -2.5 | 5.8 | 78.0 |
| 3 | Oenanthe javanica (Bl.) DC. | Apiaceae | Whole plant | zyc022 | -1.6 | 4.5 | 92.9 |
| 4 | Ligusticum striatum DC. | Apiaceae | Rhizome | hch114 | 33.2 | 8.4 | 42.9 |
| 5 | Rauvolfia verticillata (Lour.) Baill. | Apocynaceae | Root | zyc054 | 20.6 | 6.3 | 28.9 |
| 6 | Amorphophallus rivieri Durieu ex Riviere | Araceae | Tuber | zyc111 | 27.7 | 5.3 | 75.6 |
| 7 | Pinellia ternata (Thunb.) Breit. | Araceae | Rhizome | zyc092 | -2.1 | 3.6 | 88.1 |
| 8 | Ophiopogon japonicus (Thunb.) Ker Gawl. | Asparagaceae | Tuber | hch104 | -0.9 | 25.9 | 42.3 |
| 9 | Anemarrhena asphodeloides Bunge. | Asparagaceae | Rhizome | hch111 | 51.5 | 8.7 | 82.8 |
| 10 | Taraxacum mongolicum HandMazz. | Asteraceae | Whole plant | zyc203 | 4.2 | 6.8 | 52.7 |
| 11 | Senecio scandens BuchHam. | Asteraceae | Whole plant | zyc182 | 17.4 | 8.2 | 86.0 |
| 12 | Siegesbeckia orientalis L. | Asteraceae | Whole plant | zyc202 | 18.9 | 7.2 | 89.5 |
| 13 | Bidens bipinnata L. | Asteraceae | Whole plant | zyc184 | 23.3 | 3.2 | 105.8 |
| 14 | Bidens pilosa L. | Asteraceae | Whole plant | zyc184 | 28.0 | 17.1 | 102.5 |
| 15 | Commelina communis L. | Commelinaceae | Whole plant | zyc235 | 8.2 | 3.3 | 91.6 |
| 16 | Cornus officinalis Siebold & Zucc. | Cornaceae | Fruit | hch112 | 14.2 | 6.0 | 73.3 |
| 17 | Sedum erythrostictum Miq. | Crassulaceae | Whole plant | zyc263 | 61.9 | 10.3 | 87.8 |
| 18 | Trichosanthes kirilowii Maxim. | Cucurbitaceae | Root | hch107 | 2.7 | 2.6 | 24.9 |
| 19 | Dioscorea oppositifolia L. | Dioscoreaceae | Rhizome | hch103 | 5.7 | 6.2 | 3.8 |
| <mark>20</mark> | Dioscorea bulbifera L. | Dioscoreaceae | Rhizome | zyc040 | <mark>94.3</mark> | 16.8 | <mark>78.2</mark> |
| 21 | Eucommia ulmoides Oliv. | Eucommiaceae | Bark | hch113 | 5.8 | 0.8 | 73.1 |
| 22 | Phyllanthus urinaria L. | Euphorbiaceae | Whole plant | zyc268 | 37.3 | 5.2 | 105.1 |
| 23 | Ocimum basilicum L. | Lamiaceae | Whole plant | zyc118 | 53.2 | 7.5 | 78.9 |
| 24 | Prunella vulgaris L. | Lamiaceae | Whole plant | zyc125 | 53.0 | 2.4 | 102.1 |
| <mark>25</mark> | Tinospora sagittata Gagnep. | Menispermaceae | Rhizome | zyc145 | <mark>83.8</mark> | <mark>4.5</mark> | <mark>67.1</mark> |
| 26 | Melastoma dodecandrum Roxb. | Melastomataceae | Whole plant | zyc175 | 39.6 | 23.4 | 96.1 |
| 27 | Cistanche deserticola Y.C.Ma | Orobanchaceae | Rhizome | hch102 | -0.5 | 5.6 | 12.6 |
| 28 | Zea mays L. | Poaceae | Stigmas and style | hch108 | 23.8 | 4.1 | 82.7 |
| 29 | Lophatherum gracile Brongn. | Poaceae | Leaves | hch110 | 1.1 | 4.1 | 81.3 |
| 30 | Reynoutria multiflora (Thunb.) Moldenke | Polygonaceae | Tuber | hch101 | -2.9 | 5.5 | 23.5 |
| 31 | Persicaria bistorta (L.) Samp. | Polygonaceae | Rhizome | zyc085 | 88.9 | <mark>44.6</mark> | <mark>99.4</mark> |

| 32 | Portulaca oleracea L. | Portulacaceae | Whole plant | zyc225 | 10.8 | 5.7 | 95.7 |
|-----------------|--------------------------------------|------------------|-------------|--------|------|------------------|-------------------|
| 33 | Coptis chinensis Franch. | Ranunculaceae | Rhizome | hch106 | 20.4 | 1.7 | -3.7 |
| 34 | Semiaquilegia adoxoides (DC.) Makino | Ranunculaceae | Root | zyc126 | 39.1 | -0.3 | 77.9 |
| 35 | Potentilla kleiniana Wight & Arn. | Rosaceae | Whole plant | zyc243 | 6.2 | 11.2 | 49.9 |
| 36 | Gardenia jasminoides Ellis | Rubiaceae | Fruit | zyc315 | 8.1 | 6.1 | 56.2 |
| 37 | Paederia scandens (Lour.) Merr. | Rubiaceae | Whole plant | zyc309 | 60.1 | 4.2 | 95.0 |
| 38 | Dimocarpus longan Lour. | Sapindaceae | Seed | zyc295 | 41.3 | 9.8 | 27.4 |
| 39 | Scrophularia ningpoensis Hemsl. | Scrophulariaceae | Tuber | hch105 | 0.6 | 6.6 | 65.9 |
| <mark>40</mark> | Boehmeria nivea (L.) Gaudich | Urticaceae | Root | zyc105 | 84.8 | <mark>8.6</mark> | <mark>97.2</mark> |

^{*a*} Values expressed as mean of three independent experiments.

Table S2. Identification metabolites with HRMS and NMR data.

| Peak | Detected precursor ion $(m/z, positive)$ | Molecular formula | Compound | NMR Data ^a |
|------|---|-------------------|---|---|
| 1 | 155.0347 [M+H] ⁺ (calcd for C ₇ H ₇ O ₄ ⁺ : 155.0339) ΔM -5.0 ppm | $C_7H_6O_4$ | HO 2 ² 1 OH HO 3,4-Dihydroxybenzoic acid | ¹ H NMR (methanol- <i>d</i> ₄ , 600 MHz, δ in ppm, <i>J</i> in Hz) δ: 7.43 (1H, d, <i>J</i> = 1.6 Hz, H-2'), 7.41 (1H, dd, <i>J</i> = 8.1, 1.6 Hz, H-6'), 6.79 (1H, d, <i>J</i> = 8.1 Hz, H-5') |
| 2 | 291.0864 [M+H] ⁺ (calcd for C ₁₅ H ₁₅ O ₆ ⁺ : 291.0863) ΔM -0.3 ppm | C15H14O6 | HO B Catechin HO Catechin | ¹H NMR (methanol-<i>d</i>₄, 600 MHz, δ in ppm, <i>J</i> in Hz) δ: 6.84 (1H, d, <i>J</i> = 1.8 Hz, H-2'), 6.76 (1H, d, <i>J</i> = 8.2 Hz, H-5'), 6.72 (1H, dd, <i>J</i> = 8.2, 1.8 Hz, H-6'), 5.93 (1H, s, H-8), 5.86 (1H, s, H-6), 4.56 (1H, d, <i>J</i> = 7.4 Hz, H-2), 3.98 (1H, m, H-3), 2.85 (1H, dd, <i>J</i> = 16.0, 2.4 Hz, H-4b), 2.50 (1H, dd, <i>J</i> = 16.0, 8.6 Hz, H-4a). |
| 3 | 241.0706 [M+H] ⁺ (calcd for C ₁₁ H ₁₃ O ₆ ⁺ : 241.0707) ΔM 0.3 ppm | C11H12O6 | HO Eucomic acid | ¹ H NMR (methanol- <i>d</i> ₄ , 600 MHz, δ in ppm, <i>J</i> in Hz) δ: 7.07 (1H, d, <i>J</i> = 8.3 Hz, H-2' and H-6'), 6.66 (1H, d, <i>J</i> = 8.3 Hz, H-3' and H-5'), 2.96 (1H, d, <i>J</i> = 13.5 Hz, H-4a), 2.84 (1H, d, <i>J</i> = 16.2 Hz, H-2b), 2.82 (1H, d, <i>J</i> = 13.5 Hz, H-4b), 2.58 (1H, d, <i>J</i> = 16.2 Hz, H-2a) |
| 4 | 291.0872 [M+H] ⁺ (calcd for C ₁₅ H ₁₅ O ₆ ⁺ : 291.0863) ∆M -3.1 ppm | C15H14O6 | HO B C HO B C HO C HO C HO HO HO HO HO HO C HO HO HO HO HO HO HO HO HO HO | ¹ H NMR (methanol- d_4 , 600 MHz, δ in ppm, J in Hz) δ : 6.97 (1H, d, $J = 1.9$ Hz, H-2'), 6.80 (1H, dd, $J = 8.2, 1.9$ Hz, H-6'), 6.76 (1H, d, $J = 8.2$ Hz, H-5'), 5.94 (1H, d, $J = 2.2$ Hz, H-8), 5.91 (1H, d, $J = 2.2$ Hz, H-6), 4.69 (1H, m, H-3), 2.87 (1H, dd, $J = 16.8, 4.7$ Hz, H-4b), 2.73 (1H, dd, J = 16.8, 2.8 Hz, H-4a). |





| | | | | ¹ H NMR (acetone- d_6 , 600 MHz, δ in ppm, J in Hz) δ : 7.62 |
|----|---|---------------------|---------------|---|
| | | | | (1H, s, H-16), 7.47 (1H, d, J = 1.5 Hz, H-15), 6.94 (1H, d, |
| | | | | J = 1.5 Hz, H-14), 5.30 (1H, dd, J = 10.9, 5.5 Hz, H-12), |
| | | | | 4.84 (1H, m, H-2), 4.75 (1H, d, J = 5.9 Hz, H-6), 2.71 |
| | | | 0 | $(1H, m, H-4), 2.50 (1H, m, H-3\beta), 1.99 (1H, m, H-3\alpha),$ |
| | | | H 1 3 17 | 2.46 (1H, m, H-7 β), 2.26 (1H, m, H-7 α), 2.25 (1H, m, H- |
| | 345 1318 [M+H]+(calcd for C10H21O6+ | | | 5), 2.06 (1H, m, H-11β), 1.98 (1H, m, H-11α), 2.01 (1H, |
| 12 | 345.1333) AM 4.2 ppm | $C_{19}H_{20}O_{6}$ | 14 0 7 /H | m, H-1β), 1.77 (1H, m, H-1α), 1.90 (1H, m, H-10), 1.29 |
| | 5 1511555) zin 112 ppin | | | (3H, s, H-19). |
| | | | 0 | $^{13}\mathrm{C}$ NMR (acetone- $d_6,$ 150 MHz, δ in ppm) δ : 29.6 (C-1), |
| | | | Diosbulbin B | 77.2 (C-2), 39.4 (C-3), 42.7 (C-4), 42.7 (C-5), 77.9 (C-6), |
| | | | | 37.8 (C-7), 90.2 (C-8), 45.9 (C-9), 39.4 (C-10), 42.8 (C- |
| | | | | 11), 75.6 (C-12), 126.9 (C-13), 111.0 (C-14), 143.3 (C- |
| | | | | 15), 141.3 (C-16), 175.7 (C-17), 177.3 (C-18), 16.6 (C- |
| | | | | 19). |
| | | | | ¹ H NMR (acetone- d_6 , 600 MHz, δ in ppm, J in Hz) δ : 9.37 |
| | | | MeO | (1H, d, <i>J</i> = 9.2 Hz, H-5), 7.55 (1H, d, <i>J</i> = 8.8 Hz, H-9), |
| | 241 0860 [M+U]+ (colod for CHO.+ | | | 7.51 (1H, d, <i>J</i> = 8.8 Hz, H-10), 7.23 (1H, d, <i>J</i> = 2.8 Hz, H- |
| 13 | 241.0860 [M+H] ⁺ (caled for C ₁₅ H ₁₃ O ₃ ⁺ : 241.0859) ΔM -0.4 ppm | $C_{15}H_{12}O_3$ | | 6), 7.14 (1H, dd, <i>J</i> = 9.2, 2.8 Hz, H-6), 6.90 (1H, d, <i>J</i> = 2.4 |
| | | | HO | Hz, H-1), 6.82 (1H, d, <i>J</i> = 2.4 Hz, H-3), 4.09 (3H, s, 4- |
| | | | Flavanthrinin | OMe). |









 $C_{30}H_{48}O_4$

473.3625) ΔM -1.3 ppm

24



Hederagenin

¹H NMR (DMSO- d_6 , 600 MHz, δ in ppm, J in Hz) δ : 5.15 (1H, s, H-12), 3.43 (1H, overlapped, H-3), 3.32 (1H, overlapped, H-23a), 3.07 (1H, d, J = 10.6 Hz, H-23b), 2.74 (1H, dd, J = 3.2, 13.7 Hz, H-18), 1.89 (1H, t, J = 12.6Hz, H-16 β), 1.80 (2H, m, H-11 α , H-11 β), 1.66 (1H, m, H-15 β), 1.61 (1H, m, H-22 β), 1.60 (1H, m, H-19 β), 1.50 (1H, overlapped, H-9), 1.48 (1H, m, H-16 α), 1.47 (1H, m, H-1 β), 1.46 (2H, m, H-2), 1.43 (1H, m, H-7 β), 1.42 (1H, m, H-22 α), 1.40 (1H, m, H-6 α), 1.31 (1H, m, H-21 α), 1.23 (1H, m, H-6 β), 1.16 (1H, m, H-7 α), 1.12 (1H, m, H-21 α), 1.04 (1H, m, H-19 α), 0.97 (1H, m, H-15 α), 1.10 (1H, m, H-5 α), 1.09 (3H, s, H-27), 0.87 (9H, s, H-25, H-29, H-30), 0.84 (1H, m, H-1 α), 0.71 (3H, s, H-26), 0.53 (3H, s, H-24).

¹³C NMR (DMSO-*d*₆, 150 MHz, δ in ppm) δ: 37.7 (C-1),
26.3 (C-2), 70.0 (C-3), 41.6 (C-4), 46.1 (C-5), 17.3 (C-6),
31.8 (C-7), 38.6 (C-8), 46.9 (C-9), 35.9 (C-10), 22.8 (C-11), 121.3 (C-12), 143.6 (C-13), 40.8 (C-14), 27.0 (C-15),
22.5 (C-16), 45.1 (C-17), 40.6 (C-18), 45.6 (C-19), 30.2 (C-20), 33.2 (C-21), 31.9 (C-22), 64.4 (C-23), 12.4 (C-24),
15.2 (C-25), 16.7 (C-26), 25.5 (C-27), 32.6 (C-29), 23.2 (C-30).



C30H48O4

473.3625) ΔM 2.6 ppm

25



Pomolic acid

¹H NMR (DMSO- d_6 , 600 MHz, δ in ppm, J in Hz) δ : 5.15 (1H, s, H-12), 3.00 (1H, m, H-3), 2.48 (1H, m, H-16β), 2.39 (1H, m, H-18),1.89 (1H, m, H-11β), 1.83 (1H, m, H-11α), 1.69 (1H, m, H-15α), 1.68 (1H, m, H-2β), 1.61 (1H, m, H-21α), 1.59 (1H, m, H-22β), 1.58 (1H, m, H-9), 1.52 (1H, m, H-1β), 1.50 (1H, m, H-22α), 1.49 (1H, m, H-1α), 1.46 (1H, m, H-6β), 1.45 (1H, m, H-7β), 1.38 (1H, m, H-16α), 1.31 (1H, m, H-6α), 1.28 (3H, s, H-27), 1.26 (1H, m, H-20), 1.22 (1H, m, H-7α), 1.12 (1H, m, H-21β), 1.07 (3H, s, H-29), 0.89 (3H, s, H-23), 0.85 (3H, s, H-25), 0.87 (1H, m, H-15β), 0.84 (3H, s, H-30), 0.70 (3H, s, H-26),0.68 (3H, s, H-24), 0.67 (1H, m, H-5). ¹³C NMR (DMSO-*d*₆, 150 MHz, δ in ppm) δ: 37.9 (C-1), 27.9 (C-2), 76.7 (C-3), 38.2 (C-4), 54.6 (C-5), 17.9 (C-6), 32.6 (C-7), 39.1 (C-8), 46.5 (C-9), 38.1 (C-10), 23.0 (C-11), 126.8 (C-12), 138.6 (C-13), 40.9 (C-14), 27.9 (C-15), 25.0 (C-16), 46.8 (C-17), 53.1 (C-18), 73.4 (C-19), 41.3 (C-20), 25.7 (C-21), 37.1 (C-22), 28.1 (C-23), 15.8 (C-24), 15.4 (C-25), 16.4 (C-26), 23.8 (C-27), 178.8 (C-28), 26.1 (C-29), 16.1 (C-30).



| $\frac{28}{281.2475) \Delta M 3.9 \text{ ppm}} C_{18}H_{32}O_2 \qquad (4H, m, H-8 \text{ and } H-14), 1.62 (2H, p, J = 7.3 \text{ Hz}, H-3), 1.36-1.25 (14H, m, H-4, H-5, H-6, H-7, H-15, H-16 \text{ and} H-14) = 0.00 \text{ Hz}, H H_{32}O_2 = 0$ | 281.2464 [M+H] ⁺ (calcd for C ₁₈ H ₃₃ O ₂ ⁺ : 28 281.2475) ΔM 3.9 ppm | C ₁₈ H ₃₂ O ₂ | Linolenic acid | ¹ H NMR (chloroform- <i>d</i> , 600 MHz, δ in ppm, <i>J</i> in Hz) δ: 5.37-5.32 (4H, m, H-9, H-10, H-12, H-13), 2.76 (2H, t, <i>J</i> = 6.8 Hz, H-11), 2.33 (2H, t, <i>J</i> = 7.6 Hz, H-2), 2.03-2.06 (4H, m, H-8 and H-14), 1.62 (2H, p, <i>J</i> = 7.3 Hz, H-3), 1.36-1.25 (14H, m, H-4, H-5, H-6, H-7, H-15, H-16 and |
|--|--|--|----------------|--|
|--|--|--|----------------|--|

^{*a*}¹³C NMR data were obtained from HSQC and/or HMBC spectra. ^{*b*} Quaternary carbon was not determinated.

Figure S1. α-Glucosidase (A) and PTP1B (B) inhibition curves of crude ethyl acetate extracts of Persicaria bistorta (L.) Samp., Dioscorea bulbifera

L., Boehmeria nivea (L.) Gaudich, and Tinospora sagittata Gagnep.





Figure S2. UV and HRMS spectra of compound 8 obtained in the HPLC-HRMS mode.



Figure S3. ¹H-NMR spectrum of the mixture of compound 7 and 8 (600 MHz, Methanol- d_4).







Figure S5. HSQC spectrum of the mixture of compound 7 and 8 (600 MHz, Methanol- d_4).



Figure S6. HMBC spectrum of the mixture of compound 7 and 8 (600 MHz, Methanol- d_4).



Figure S7. COSY spectrum of the mixture of compound 7 and 8 (600 MHz, Methanol- d_4).



Figure S8. UV and HRMS spectra of compound 9 obtained in the HPLC-HRMS mode.



Figure S9. ¹H-NMR spectrum of compound **9** (600 MHz, DMSO-*d*₆).

Figure S10. HSQC spectrum of compound 9 (600 MHz, DMSO-*d*₆).



Figure S11. HMBC spectrum of compound 9 (600 MHz, DMSO-*d*₆).



Figure S12. COSY spectrum of compound 9 (600 MHz, DMSO-*d*₆).





Figure S13. UV and HRMS spectra of compound 10 obtained in the HPLC-HRMS mode.



Figure S14. ¹H-NMR spectrum of compound 10 (600 MHz, Methanol-*d*₄).



Figure S15. HSQC spectrum of compound 10 (600 MHz, Methanol-*d*₄).

Figure S16. HMBC spectrum of compound 10 (600 MHz, Methanol-*d*₄).







Figure S18. HPLC chromatogram of crude *P. bistorta* ethyl acetate extract monitored at 254 nm (black) as well as high-resolution α -glucosidase inhibition profile (red), high-resolution PTP1B inhibition profile (green) and high-resolution α -amylase inhibition profile (blue) [40 mg/mL, 10 μ L injection].





Figure S19. IC₅₀ curves of isolated compounds and Acarbose (reference compound) in the α -glucosidase inhibition assays.



