Supporting information

Facile, highly efficient and environmentally friendly transesterification mediated by platinum dioxide and nickel oxide under essentially neutral conditions

Binhao Teng, Jiangong Shi and Chunsuo Yao*

State Key Laboratory of Bioactive Substance and Function of Natural Medicines, Institute of Materia Medica, Chinese Academy of Medical Sciences & Peking Union Medical College, Beijing 100050, P. R. China

Table of Contents

1. General methods	S2
2. General procedures	.S2
3. Products characterization	
4. References	.S11
5. NMR spectra	S12
6. ESI MS spectra	.S40

1. General methods

All manipulations were carried out in autoclave vessel or glass reaction tube equipped with a magnetic stir bar. Unless otherwise mentioned, solvents and reagents were purchased from commercial sources. The transformation progress was detected by TLC or HPLC. NMR spectra were acquired at 500 MHz for ¹H and 125 MHz for ¹³C, respectively, on Varian INOVA 500 MHz (Bruker Corporation, Karlsruhe, Germany), in CDCl₃, with solvent peaks as references. ESI-MS and HR-ESI-MS data were measured using an AccuToFCS JMST100CS spectrometer (Agilent Technologies, Ltd, Santa Clara, CA, USA). Column chromatography (CC) was performed with silica gel (200-300 mesh, Qingdao Marine Chemical Inc. Qingdao, China). HPLC analysis was performed on an Agilent liquid chromatography 1260 (Agilent Technologies, Ltd, Santa Clara, CA, USA) with a YMC (250 × 5 mm i.d.) column packed with C18 (5 μ M). TLC was carried out with glass precoated silica gel GF254 plates (Qingdao Marine Chemical, Inc., Qingdao, China). Spots were visualized under UV light or by spraying with 7% H₂SO₄ in 95% EtOH followed by heating.

2. General Procedures

2.1 General Procedure for the PtO₂/H₂-catalyzed transesterification of esters.

Methyl 3,5-dimethoxybenzoate (30 mg, 0.000153 mol), ethanol (10.0 mL), platinic oxide (35 mg, 0.000153 mol) were placed in an autoclave vessel. Autoclave was pressurized with 1–2 bar of nitrogen followed by 1–2 bar of hydrogen gas. The reaction mixtures were then warmed to 60 °C, stirred for 6 h at 500 rpm, and concentrated *in vacuo* to provide a residue. Whereafter, the residue was solved in acetone, filtered through microporous filtering film. The filtrate was concentrated *in vacuo* to give the products (Table 2, entry 1).

2.2 General Procedure for the PtO₂/NiO and H₂-catalyzed transesterification of esters.

Methyl 3,5-dimethoxybenzoate (30 mg, 0.000153 mol), ethanol (10.0 mL), platinic oxide (35 mg, 0.000153 mol) and nickel protoxide (13.7 mg, 0.000184 mol) were placed in an autoclave vessel. The autoclave was pressurized with 1-2 bar of nitrogen followed by 1-2 bar of hydrogen gas. Then the pressure was increased to the desired 6 bar. Subsequently, the reaction mixtures was warmed

to 95 °C with the pressure increased to 7-8 bar and stirred for 15 h at 680 rpm. Then, the reaction mixture was concentrated *in vacuo* to provide a residue. The residue was solved in acetone, filtered through microporous filtering film, and the filtrate was concentrated *in vacuo* to give the products (Table 8, entry 1).

3. Products characterization

3.1 2-Furancaboxylic acid, ethyl ester.

¹H NMR (500 MHz, CDCl₃) δ 7.56 (d, J = 0.9 Hz, 1H), 7.16 (dd, J = 3.5, 0.5 Hz, 1H), 6.49 (dd, J = 3.5, 1.7 Hz, 1H), 4.35 (q, J = 7.1 Hz, 2H), 1.37 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 158.88, 146.26, 145.02, 117.83, 111.90, 61.09, 14.45. (+)-ESI *m/z*: 163.2 [M+Na]⁺. Spectral data were in agreement with those reported in the literature.¹

3.2 2-Naphthalene carboxylic acid, ethyl ester.



¹H NMR (500 MHz, CDCl₃) δ 8.62 (s, 1H), 8.12 - 8.03 (m, 1H), 7.96 (d, J = 8.1 Hz, 1H), 7.88 (d, J = 8.6 Hz, 2H), 7.63 - 7.48 (m, 2H), 4.45 (q, J = 7.1 Hz, 2H), 1.45 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 166.92, 135.60, 132.63, 131.06, 129.45, 128.27, 128.20, 127.87, 126.71, 125.38, 61.23, 14.53. (+)-ESI *m/z*: 223.2 [M+Na]⁺. Spectral data were in agreement with those reported in the literature.²

3.3 3,4-Dimethoxybenzoic acid, ethyl ester.



¹ H NMR (500 MHz, CDCl₃) δ 7.68 (dd, J = 8.4, 1.9 Hz, 1H), 7.54 (d, J = 1.9 Hz, 1H), 6.87 (d, J = 8.4 Hz, 1H), 4.35 (q, J = 7.1 Hz, 2H), 3.92 (s, 6H), 1.38 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 166.52, 152.98, 148.69, 123.59, 123.16, 112.07, 110.31, 60.92, 56.11, 56.11,

14.52. (+)-ESI m/z: 233.2 [M+Na]⁺. Spectral data were in agreement with those reported in the literature.³

3.4 3,4-Dimethoxybenzoic acid, butyl ester.



¹H NMR (500 MHz, CDCl₃) δ 7.67 (dd, J = 8.4, 1.9 Hz, 1H), 7.54 (d, J = 1.8 Hz, 1H), 6.87 (d, J = 8.4 Hz, 1H), 4.29 (t, J = 6.7 Hz, 2H), 3.92 (s, 3H), 1.79 - 1.68 (m, 2H), 1.46 (dt, J = 14.8, 7.4 Hz, 2H), 0.97 (t, J = 7.4 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 166.57, 152.98, 148.70, 123.57, 123.18, 112.09, 110.32, 64.81, 56.10, 56.10, 30.96, 19.41, 13.90. (+)-ESI *m/z*: 261.2 [M+Na]⁺. These data were in agreement with those reported in the literature.⁴

3.5 3,5-Dimethylbenzoic acid, ethyl ester.



¹H NMR (500 MHz, CDCl₃) δ 7.66 (s, 2H), 7.18 (s, 1H), 4.36 (q, *J* = 7.1 Hz, 2H), 2.36 (s, 6H), 1.39 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 167.11, 138.08, 134.58, 134.56, 130.51, 127.38, 127.35, 60.94, 21.30, 21.27, 14.49. (+)-ESI *m*/*z*: 201.2 [M+Na]⁺. These data were in agreement with those reported in the literature.⁵

3.6 3,5-Dimethoxybenzoic acid, 1-methylethyl ester.



¹H NMR (500 MHz, CDCl₃) δ 7.18 (d, J = 2.2 Hz, 2H), 6.63 (t, J = 2.2 Hz, 1H), 5.23 (hept, J = 6.29 Hz, 1H), 3.83 (s, 6H), 1.36 (d, J = 6.3 Hz, 6H). ¹³C NMR (125 MHz, CDCl₃) δ 165.98, 160.72 (2 × C), 132.98, 107.29 (2 × C), 105.49, 68.73, 55.72 (2 × C), 29.85, 22.06 (2 × C). (+)-ESI *m/z*: 247.2 [M+Na]⁺. These data were in agreement with those reported in the literature.⁶

3.7 3,5-Dimethoxybenzoic acid, ethyl ester.



¹H NMR (500 MHz, CDCl₃) δ 7.19 (d, J = 2.4 Hz, 2H), 6.64 (t, J = 2.4 Hz, 1H), 4.37 (q, J = 7.1 Hz, 2H), 3.83 (s, 6H), 1.39 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 166.50, 160.74 (2 × C), 132.53, 107.26 (2 × C), 105.69, 61.27, 55.69 (2 × C), 14.44. (+)-ESI *m/z*: 233.2 [M+Na]⁺. These data were in agreement with those reported in the literature.⁷

3.8 3,5-Dimethoxybenzoic acid, butyl ester.



¹H NMR (500 MHz, CDCl₃) δ 7.19 (d, J = 2.3 Hz, 2H), 6.64 (t, J = 2.3 Hz, 1H), 4.31 (t, J = 6.6 Hz, 2H), 3.83 (s, 6H), 1.79 - 1.68 (m, 2H), 1.52 - 1.42 (m, 2H), 0.98 (t, J = 7.4 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 166.56, 160.75 (2 × C), 132.57, 107.31 (2 × C), 105.58, 65.17, 55.71 (2 × C), 30.90, 19.42, 13.91. (+)-ESI m/z: 261.2 [M+Na]⁺. These data were in agreement with those reported in the literature.⁸

3.9 3,5-Dimethoxybenzoic acid, octyl ester.



¹H NMR (500 MHz, CDCl₃) δ 7.19 (d, J = 2.3 Hz, 2H), 6.64 (t, J = 2.3 Hz, 1H), 4.30 (t, J = 6.7 Hz, 2H), 3.83 (s, 6H), 1.80 - 1.71 (m, 2H), 1.48 - 1.39 (m, 2H), 1.38 - 1.24 (m, 10H), 0.88 (t, J = 6.8 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 166.56, 160.75 (2 × C), 132.58, 107.31 (2 × C), 105.59, 65.48, 55.70 (2 × C), 31.94, 29.39, 29.34, 28.84, 26.18, 22.79, 14.23. (+)-ESI *m/z*: 317.3 [M+Na]⁺.

3.10 3,5-Dimethoxybenzoic acid, phenylmethyl ester.



¹H NMR (500 MHz, CDCl₃) δ 7.44 (d, J = 6.7 Hz, 2H), 7.39 (t, J = 7.4 Hz, 2H), 7.35 (t, J = 7.1 Hz, 1H), 7.23 (d, J = 2.3 Hz, 2H), 6.65 (t, J = 2.3 Hz, 1H), 5.36 (s, 1H), 3.82 (s, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 166.33, 160.78 (2 × C), 136.13, 132.14, 128.73 (2 × C), 128.37, 128.28 (2 × C), 107.47 (2 × C), 105.83, 66.97, 55.72 (2 × C). (+)-ESI *m/z*: 295.2 [M+Na]⁺. These data were in agreement with those reported in the literature.⁹

3.11 Benzoic acid, ethyl ester.



¹H NMR (500 MHz, CDCl₃) δ 8.09 - 7.98 (m, 2H), 7.54 (tt, *J* = 10.6, 4.2 Hz, 1H), 7.43 (t, *J* = 7.7 Hz, 2H), 4.38 (q, *J* = 7.1 Hz, 2H), 1.39 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 166.74, 132.89, 130.61, 129.63 (2 × C), 128.40 (2 × C), 61.05, 14.43. (+)-ESI *m/z*: 173.2 [M+Na]⁺. These data were in agreement with those reported in the literature.¹⁰

3.12 Benzoic acid, butyl ester.



¹H NMR (500 MHz, CDCl₃) δ 8.10 - 8.00 (m, 2H), 7.59 - 7.51 (m, 1H), 7.43 (t, *J* = 7.7 Hz, 2H), 4.33 (t, *J* = 6.6 Hz, 2H), 1.76 (dt, *J* = 14.5, 6.6 Hz, 2H), 1.48 (dt, *J* = 14.8, 7.4 Hz, 2H), 0.98 (t, *J* = 7.4 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 166.80, 132.90, 130.67 (2 × C), 129.65 (2 × C), 128.43, 64.95, 30.92, 19.41, 13.89. (+)-ESI *m/z*: 201.2 [M+Na]⁺. These data were in agreement with those reported in the literature.¹¹

3.13 3-Fluorobenzoic acid, ethyl ester.



¹H NMR (500 MHz, CDCl₃) δ 7.83 (dt, J = 7.7, 1.3 Hz, 1H), 7.72 (ddd, J = 9.4, 2.7, 1.5 Hz, 1H), 7.41 (td, J = 8.0, 5.5 Hz, 1H), 7.28 - 7.20 (m, 1H), 4.38 (q, J = 7.1 Hz, 2H), 1.39 (t, J = 7.1 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 165.60, 162.67, 132.82, 130.06, 125.39, 119.98, 116.57, 61.47, 14.40. (+)-ESI m/z: 191.2 [M+Na]⁺. These data were in agreement with those reported in the literature.¹²

3.14 3-Methoxybenzoic acid, ethyl ester.



¹H NMR (500 MHz, CDCl₃) δ 7.64 (dt, J = 7.7, 1.3 Hz, 1H), 7.57 (dd, J = 2.7, 1.5 Hz, 1H), 7.34 (t, J = 7.9 Hz, 1H), 7.09 (ddd, J = 8.3, 2.7, 1.0 Hz, 1H), 4.38 (q, J = 7.1 Hz, 2H), 3.85 (s, 3H), 1.39 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 166.63, 159.67, 131.96, 129.46, 122.08, 119.45, 114.14, 61.19, 55.57, 14.47. (+)-ESI *m/z*: 203.2 [M+Na]⁺. These data were in agreement with those reported in the literature.¹²

3.15 2-Methoxybenzoic acid, ethyl ester.



¹H NMR (500 MHz, CDCl₃) δ 7.77 (dd, J = 7.9, 1.8 Hz, 1H), 7.42 - 7.48 (m, 1H), 6.98 - 6.93 (m, 2H), 4.34 (q, J = 7.1 Hz, 2H), 3.88 (s, 3H), 1.36 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 166.26, 159.17, 133.38, 131.52, 120.56, 120.16, 112.12, 60.84, 56.05, 14.38. (+)-ESI *m/z*: 203.2 [M+Na]⁺. These data were in agreement with those reported in the literature.¹²

3.16 4-Dimethylaminobenzoic acid, ethyl ester.



¹H NMR (500 MHz, CDCl₃) δ 7.93 (d, J = 8.9 Hz, 2H), 6.72 (d, J = 8.5 Hz, 2H), 4.32 (q, J = 7.1 Hz, 2H), 3.04 (s, 6H), 1.37 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 167.04, 152.97,

131.36 (2 × C), 131.36 (2 × C), 111.42, 60.35, 40.57 (2 × C), 14.62. (+)-ESI *m/z*: 216.2 [M+Na]⁺.

These data were in agreement with those reported in the literature.¹⁰

3.17 4-Ethylbenzoic acid, ethyl ester.



¹H NMR (500 MHz, CDCl₃) δ 7.96 (d, J = 7.8 Hz, 2H), 7.26 (d, J = 7.8 Hz, 2H), 4.36 (q, J = 7.2 Hz, 2H), 2.70 (q, J = 7.7 Hz, 2H), 1.38 (t, J = 7.2 Hz, 3H), 1.25 (t, J = 7.7 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 166.86, 149.75, 129.80 (2 × C), 128.13, 127.96 (2 × C), 60.88, 29.10, 15.40, 14.50. (+)-ESI *m/z*: 201.2 [M+Na]⁺. These data were in agreement with those reported in the literature.¹³

3.18 4-Trifluoromethylbenzoic acid, ethyl ester.



¹H NMR (500 MHz, CDCl₃) δ 8.16 (d, J = 8.1 Hz, 2H), 7.70 (d, J = 8.2 Hz, 2H), 4.42 (q, J = 7.1 Hz, 2H), 1.42 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 165.53, 134.48, 133.86, 130.09, 125.50, 123.8, 61.70, 14.41. (-)-ESI *m*/*z*: 217.8 [M-H]⁻. These data were in agreement with those reported in the literature.¹²

3.19 4-Ethenylbenzoic acid, ethyl ester.



¹H NMR (500 MHz, CDCl₃) δ 8.00 (d, J = 8.4 Hz, 2H), 7.46 (d, J = 8.3 Hz, 2H), 6.75 (dd, J = 17.6, 10.9 Hz, 1H), 5.86 (d, J = 17.6 Hz, 1H), 5.38 (d, J = 10.9 Hz, 1H), 4.37 (q, J = 7.1 Hz, 2H), 1.39 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 166.53, 141.95, 136.19, 129.98 (2 × C), 129.77, 126.19 (2 × C), 116.51, 61.06, 14.48. (+)-ESI *m/z*: 199.2 [M+Na]⁺. These data were in agreement with those reported in the literature.¹⁴

3.20 4-Methylbenzoic acid, ethyl ester.



¹H NMR (500 MHz, CDCl₃) δ 7.94 (d, J = 8.2 Hz, 2H), 7.23 (d, J = 8.0 Hz, 2H), 4.36 (q, J = 7.1 Hz, 2H), 2.40 (s, 3H), 1.39 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 166.81, 143.51, 129.67 (2 × C), 129.13 (2 × C), 127.91, 60.86, 21.75, 14.47. (+)-ESI *m/z*: 187.2 [M+Na]⁺. These data were in agreement with those reported in the literature.¹²

3.21 4-Methoxybenzene acetic acid, ethyl ester.



¹H NMR (500 MHz, CDCl₃) δ 7.20 (d, J = 8.7 Hz, 2H), 6.86 (d, J = 8.6 Hz, 2H), 4.14 (q, J = 7.1 Hz, 2H), 3.79 (s, 3H), 3.55 (s, 2H), 1.25 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 172.03, 158.77, 130.36 (2 × C), 126.35, 114.08 (2 × C), 60.88, 55.36, 40.64, 14.31. (+)-ESI *m/z*: 217.2 [M+Na]⁺. These data were in agreement with those reported in the literature.¹⁵

3.22 4-Methoxybenzoic acid, ethyl ester.



¹H NMR (500 MHz, CDCl₃) δ 8.00 (d, J = 8.9 Hz, 2H), 6.91 (d, J = 8.9 Hz, 2H), 4.35 (q, J = 7.1 Hz, 2H), 3.86 (s, 3H), 1.38 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 166.53, 163.38, 131.66 (2 × C), 123.10, 113.67 (2 × C), 60.77, 55.54, 14.52. (+)-ESI *m/z*: 203.2 [M+Na]⁺. These data were in agreement with those reported in the literature.¹²

3.23 4-Methoxybenzoic acid, butyl ester.



¹H NMR (500 MHz, CDCl₃) δ 7.99 (d, J = 8.8 Hz, 2H), 6.91 (d, J = 8.9 Hz, 2H), 4.29 (t, J = 6.6 Hz, 2H), 3.84 (s, 3H), 1.81 - 1.66 (m, 2H), 1.54 - 1.39 (m, 2H), 0.97 (t, J = 7.4 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 166.55, 163.35, 131.63 (2 × C), 123.10, 113.65 (2 × C), 64.63, 55.50, 30.96, 19.41, 13.89. (+)-ESI *m/z*: 231.2 [M+Na]⁺. These data were in agreement with those reported in the literature.¹⁶

3.24 4-Fluorobenzoic acid, ethyl ester.



¹H NMR (500 MHz, CDCl₃) δ 8.08 - 8.03 (m, 2H), 7.10 (t, *J* = 8.7 Hz, 2H), 4.36 (q, *J* = 7.1 Hz, 2H), 1.38 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (125 MHz, cdcl₃) δ 166.29, 164.80, 132.18, 126.86, 115.55, 61.21, 14.44. (+)-ESI *m/z*: 191.2 [M+Na]⁺. These data were in agreement with those reported in the literature.¹²

3.25 Ethyl 4-formylbenzoate.



¹H NMR (500 MHz, CDCl₃) δ 10.10 (s, 1H), 8.20 (d, J = 8.3 Hz, 2H), 7.95 (d, J = 8.4 Hz, 2H), 4.42 (q, J = 7.1 Hz, 2H), 1.42 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 191.80, 165.72, 139.23, 135.62, 130.29 (2 × C), 129.63 (2 × C), 61.76, 14.42. (+)-ESI *m/z*: 217.5 [M+K]⁺. These data were in agreement with those reported in the literature.¹²

3.26 Ethyl 3-nitrobenzoate.



¹H NMR (500 MHz, CDCl₃) δ 8.85 (t, J = 1.9 Hz, 1H), 8.43 – 8.38 (m, 1H), 8.36 (dd, J = 7.7, 1.4 Hz, 1H), 7.64 (t, J = 8.0 Hz, 1H), 4.44 (q, J = 7.1 Hz, 2H), 1.43 (t, J = 7.2 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 164.58, 148.39, 135.38, 132.37, 129.69, 127.40, 124.66, 62.07, 14.39. (+)-ESI m/z: 218.2 [M+Na]⁺. These data were in agreement with those reported in the literature.¹⁷

3.27 3-(3,5-Dimethoxyphenyl)-2-(4-methoxyphenyl)-5-Benzofuran carboxylic acid, ethyl ester



¹H NMR (500 MHz, CDCl₃) δ 8.20 (d, J = 1.7 Hz, 1H), 8.04 (dd, J = 8.6, 1.8 Hz, 1H), 7.64 (d, J = 8.7 Hz, 2H), 7.54 (d, J = 8.6 Hz, 1H), 6.87 (d, J = 8.5 Hz, 2H), 6.64 (d, J = 2.4 Hz, 2H), 6.54 (t, J = 2.3 Hz, 1H), 4.38 (q, J = 7.1 Hz, 2H), 3.83 (s, 3H), 3.79 (s, 7H), 1.39 (t, J = 7.1 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 167.00, 161.45 (2 × C), 160.16, 156.42, 152.16, 134.38, 130.60, 128.71 (2 × C), 126.23, 125.78, 122.77, 122.30, 114.12 (2 × C), 110.86, 107.82 (2 × C), 100.35, 77.42, 77.16, 76.91, 61.07, 55.61, 55.46, 29.85. (+)-HRESI *m/z*: 455.1581 [M+Na]⁺ (Calcd for C₂₆H₂₄O₆Na: 455.1473).

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S13







































230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 fl (ppm)











230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 fl (ppm)













6. ESI MS spectra

Single Mass Spectrum Deconvolution Report

Analysis Name: tengbh21.d	Instrument:	LC-MSD-Trap-SL	Print Date:	1/3/2018 11:09:24 AM
Method: TEST.M Sample Name: E56	Operator:	Operator	Acq. Date:	1/3/2018 11:08:05 AM
Analysis Info:				



Analysis Name: tengbh19.d	Instrument: LC-MSD-Trap	-SL Print Date: 1/3/2018 11:10:23 AM
Method: TEST.M	Operator: Operator	Acq. Date: 1/3/2018 10:57:18 AM
Sample Name: E54 Analysis Info:		



Analysis Name: tengbh08.d Method: TEST.M Sample Name: E28 Analysis Info:	Instrument: Operator:	LC-MSD-Trap-SL Operator	Print Date: 1/3/2018 10:39:51 AM Acq. Date: 1/3/2018 10:10:55 AM
Intens. x10 ⁷		Ņ	tengbh08.d: +MS, 0.0-0.2min #(3-13)



Analysis Name:	tengbh17.d	Instrument:	LC-MSD-Trap-SL	Print Date:	1/3/2018	10:55:35 AM
Method: TEST.M		Operator:	Operator	Acq. Date:	1/3/2018	10:49:49 AM
Sample Name: E48	1					
Analysis Info:			r l			
The second se	45 45					



Analysis Name: tengbh16.d Method: TEST.M Sample Name: E37 Analysis Info:

Operator:

Instrument: LC-MSD-Trap-SL Operator

Print Date: 1/3/2018 10:52:10 AM Acq. Date: 1/3/2018 10:46:09 AM



Analysis Name:	tengbh05.d	Instrument:	LC-MSD-Trap-SL	Print Date:	1/3/2018	10:03:53 AM
Method: TEST.M		Operator:	Operator	Acq. Date:	1/3/2018 9:	58:01 AM
Sample Name: E20	li -					
Analysis Info:						



Analysis Name:	tengbh00.d	Instrument:	LC-MSD-Trap-SL	Print Date:	1/3/2018 9:48:48 AM
Method: TEST.M		Operator:	Operator	Acq. Date:	1/3/2018 9:32:45 AM
Sample Name: E7 Analysis Info:					



Analysis Name: tengbh02.d	Instrument:	LC-MSD-Trap-SL	Print Date:	1/3/2018 9:52:55 AM
Method: TEST.M	Operator:	Operator	Acq. Date:	1/3/2018 9:45:52 AM
Sample Name: E17 Analysis Info:		1		



Analysis Name:	tengbh03.d	Instrument:	LC-MSD-Trap-SL	Print Date:	1/3/2018	10:00:20 AM
Method: TEST.M		Operator:	Operator	Acq. Date:	1/3/2018 9:	49:50 AM
Sample Name: E18 Analysis Info:						



Analysis Name:	tengbh06.d	Instrument:	LC-MSD-Trap-SL	Print Date:	1/3/2018	10:25:06 AM
Method: TEST.M		Operator:	Operator	Acq. Date:	1/3/2018	10:01:58 AM
Sample Name: E22						
Analysis Info:						
				100 C		



Analysis Name: tengbh18.d Method: TEST.M Sample Name: E49 Analysis Info: Instrument: LC-MSD-Trap-SL Operator: Operator Print Date: 1/3/2018 10:55:52 AM Acq. Date: 1/3/2018 10:53:22 AM



Analysis Name:	tengbh23.d	Instrument:	LC-MSD-Trap-SL	Print Date:	1/3/2018 11:41:57 AM
Method: TEST.M		Operator:	Operator	Acq. Date:	1/3/2018 11:16:27 AM
Sample Name: E58 Analysis Info:					



Analysis Name: tengbh25.d Method: TEST.M Sample Name: E66 Analysis Info:

d Instrument Operator:

Instrument:LC-MSD-Trap-SLOperator:Operator

 Print Date:
 1/3/2018
 11:27:40 AM

 Acq. Date:
 1/3/2018
 11:25:21 AM



Analysis Name:	tengbh04.d	Instrument:	LC-MSD-Trap-SL	Print Date:	1/3/2018	10:00:44 AM
Method: TEST.M		Operator:	Operator	Acq. Date:	1/3/2018 9:5	3:47 AM
Sample Name: E19						
Analysis Info:						



Analysis Name: tengbh	07.d Instrument: LC-MS	D-Trap-SL Print Date: 1/3/2018 10:34:29 AM
Method: TEST.M	Operator: Opera	tor Acq. Date: 1/3/2018 10:06:38 AM
Sample Name: E26 Analysis Info:		



Analysis Name:	tengbh09.d	Instrument:	LC-MSD-Trap-SL	Print Date:	1/3/2018	10:43:54 AM
Method: TEST.M		Operator:	Operator	Acq. Date:	1/3/2018 10	0:15:03 AM
Sample Name: E29 Analysis Info:						



Analysis Name:tengbh20.dInstrument:LC-MSD-Trap-SLPrint Date:1/3/201811:05:04 AMMethod:TEST.MOperator:OperatorAcq. Date:1/3/201811:03:12 AMSample Name:E55-2Analysis Info:



Analysis Name:	tengbh15.d	Instrument:	LC-MSD-Trap-SL	Print Date:	1/3/2018	10:51:45 AM
Method: TEST.M		Operator:	Operator	Acq. Date:	1/3/2018 1	l0:42:29 AM
Sample Name: E36	5					
Analysis Info:						
		200 0. XX				



Analysis Name: tengbh24.d	Instrument:	LC-MSD-Trap-SL	Print Date:	1/3/2018	11:42:15 AM
Method: TEST.M	Operator:	Operator	Acq. Date:	1/3/2018 1	L1:20:13 AM
Sample Name: E59					
Analysis Info:					



Analysis Name: tengbh10.d	Instrument:	LC-MSD-Trap-SL	Print Date:	1/3/2018 10:44:13 AM
Method: TEST.M	Operator:	Operator	Acq. Date:	1/3/2018 10:19:16 AM
Sample Name: E30				
Analysis Info:				



Analysis Name: tengbh11.d	Instrument:	LC-MSD-Trap-SL	Print Date:	1/3/2018 10:44:31 AM
Method: TEST.M	Operator:	Operator	Acq. Date:	1/3/2018 10:23:45 AM
Sample Name: E32				
Analysis Info:				



Analysis Name: tengbh01.d	Instrument: LC-MSD-Trap-SL	Print Date: 1/3/2018 9:51:49 AM
Method: TEST.M	Operator: Operator	Acq. Date: 1/3/2018 9:38:42 AM
Sample Name: E16		
Analysis Info:		
	80.05012	



Analysis Name:	tengbh22.d	Instrument:	LC-MSD-Trap-SL	Print Date:	1/3/2018	11:22:48 AM
Method: TEST.M		Operator:	Operator	Acq. Date:	1/3/2018 1	1:12:15 AM
Sample Name: E57 Analysis Info:						



Analysis Name: tengbh12.d	Instrument:	LC-MSD-Trap-SL	Print Date:	1/3/2018	10:47:35 AM
Method: TEST.M	Operator:	Operator	Acq. Date:	1/3/2018	10:29:49 AM
Sample Name: E34					
Analysis Info:					



Analysis Name: tengbh27.d Method: TEST.M Sample Name: E68 Analysis Info:

- d Instrument: Operator:
- Instrument:LC-MSD-Trap-SLOperator:Operator

 Print Date:
 1/3/2018
 11:46:59 AM

 Acq. Date:
 1/3/2018
 11:36:47 AM



Analysis Name:	tengbh26.d	Instrument:	LC-MSD-Trap-SL	Print Date:	1/3/2018	11:45:40 AM
Method: TEST.M		Operator:	Operator	Acq. Date:	1/3/2018 1	.1:31:07 AM
Sample Name: E67	1					
Analysis Info:						



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Page 1 of 2

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