Supporting information

Multiple hydrogen bond coordination in three-constituent deep eutectic solvent enhances lignin fractionation from biomass

Qinqin Xia, Yongzhuang Liu, Juan Meng, Wanke Cheng, Wenshuai Chen, Shouxin Liu, Yixing Liu, Jian Li, Haipeng Yu*

Key laboratory of Bio-based Material Science and Technology, Ministry of Education, Northeast Forestry University, Harbin, 150040, P. R. China

Contents

| Materials |
|---|
| Note S1. Measurement of the K–T solvatochromic parameters |
| Note S2. Calculation of CR, LF and LER |
| Note S3. Calculation of purity of LF |
| Table S1 Electron density and density Laplacian for the ChCl-glycerol and lignin-lignin at |
| the bond critical points |
| Table S2 Lewis acid strength and K-T parameters of the ChCl/glycerol DES with different |
| metal chlorides |
| Table S3 Assignment of ${}^{13}C-{}^{1}H$ cross-signals in the HSQC spectra of lignin fractions S8 |
| Table S4 $M_{\rm w}$, $M_{\rm n}$, and polydispersity ($M_{\rm w}/M_{\rm n}$) of MWL and 3c-DESL |
| Table S5 Analysis of carbohydrates that dissolved in the 3c-DES |
| Figure S1 Topological graph of ChCl-glycerol interactions |
| Figure S2 Typical LCC structure and lignin substructures |
| Figure S3 Photographs of the mixtures formed by adding metal chloride salts in the |
| ChCl/glycerol DES |
| Figure S4 3c-DES-4 at room temperature and low temperature (-20 °C) |
| Figure S5 UV spectra of the K–T dyes and the 3c-DESs with K–T dyes |
| Figure S6 The fractionation process of wood lignocellulose into CR, LF and dissolved |
| carbohydrates by oil bath heating with the 3c-DES |
| Figure S7 (a) LER and (b) purity of the LF pretreated with the 3c-DES-4 at different |
| temperatures and with increasing time |

| Figure S8 SEM image of the pseudolignin generated at 120 °C | .S15 |
|--|--------------|
| Figure S9 Comparison of the LERs between 3c-DES-4 and glycerol/AlCl ₃ · $6H_2O$ S | 15 |
| Figure S10 2D-HSQC NMR spectra of MWL and 3c-DESL | . S16 |
| Figure S11 ³¹ P NMR spectra of MWL and 3c-DESL | . S17 |
| References | S18 |

Materials:

Poplar wood powder was used as feedstock and dried at 103 °C for 24 h. ChCl, glycerol, and AlCl₃·6H₂O were purchased from Aladdin Reagent Co. Ltd. Acetone were purchased from Kermel Chemical Reagent Co., Ltd. 4-nitroaniline (NH₂), N,N-diethyl-4-nitroaniline (NEt₂), and Nile red (NR) were purchased from Sigma-Aldrich (St. Louis, MO). All the above chemicals and reagents were used without further purification.

Note S1. Measurement of the K-T solvatochromic parameters

The dyes of NR, NEt₂, NH₂, 4-nitroanisole (OMe) and 4-nitrophenol (OH) were considered for measuring the K–T solvatochromic parameters.^[1,2] NH₂ was used to determine β .^[3] NEt₂ was selected to determine π^* , because this dye will show distinct absorption band for high-polarity DES. NR was used as a solvatochromic indicator to determine α . Reichardt's dye 30 and NR are generally used to determine the α values. But when Reichardt's dye 30 was added in the 3c-DES, the dye faded to colorless immediately. There was no detectable absorption band in the UV spectra (400–700 nm). Therefore, NR was used as solvatochromic probe to calculate α values instead of Reichardt's dye 30.

The dyes were respectively dissolved in methanol and stored in amber glass vials. Each time 5 mL dye solution was added into a 5mL centrifuge tube. The methanol was evaporated by vacuum drying at 40 °C for 2 day. Then, 3 mL DES was put into the dried dye, and was transferred into a 1 cm² quartz cuvette. The absorption spectrum of the mixture was checked using UV-visible spectrophotometer (TU-1900, Beijing Purkinje General Instrument Co. Ltd, China) at room temperature in the wavelength range of 300–700 nm.

The calculation of α , β and π^* values use the following equations: ^[4]

$$\pi^* = 0.314 \times (27.52 - \nu_{NEt2})$$

$$\beta = 11.134 - \frac{3580}{\lambda(NH_2)_{max}} - 1.125\pi^*$$

$$\alpha = \frac{19.9657 - 1.0241\pi^* - \nu_{NR}}{1.6078}$$

Where π^* is normalized to give cyclohexane and DMSO,^[5] λ_{max} is the wavelength

corresponding to maximum absorption of dye and $v = 1/(\lambda_{max} \times 10^{-4})$.

Note S2. Calculation of CR, LF and LER

CR (wt%) is the weight percentage of non-dissolved solid residues to the wood feedstock. LF means the DES-extracted lignin which was precipitated with water after the removal of cellulose residues by filtration. LF (wt%) is calculated as the weight percentage of non-dissolved solid residues to the wood feedstock. LER (%) is calculated by the percentage of lignin fractions to the initial lignin content in wood feedstock. The initial lignin content in wood *feedstock* is measured as 19.12 \pm 0.32%.

$$CR (wt\%) = \frac{Non - dissolved \ solid \ residues}{Wood \ feedstock} \times 100\%$$
$$LF (wt\%) = \frac{DES - extracted \ lignin}{Wood \ feedstock} \times 100\%$$
$$LER (\%) = \frac{Lignin \ fractions}{Initial \ lignin \ content \ in \ wood \ feedstock} \times 100\%$$

Note S3. Calculation of purity of LF

The content of LF in the DES was determined by the acid hydrolysis method. First, the precipitated lignin by adding water was dried at 80 °C for 24 h. The oven-dried lignin (100 mg) was then reacted with 1.5 mL of 72% H_2SO_4 at room temperature with stirring for 2 h. The solution was diluted with DI water to a 3% H_2SO_4 concentration and refluxed for 2 h. Finally, it was filtered and the acid-insoluble lignin (Klason lignin) was determined by gravimetric analysis. The percentage of acid-insoluble lignin to the DES-extracted lignin was defined as the purity of LF.

| Substance | | Interestion | ρ | $^{\bigtriangledown 2} ho$ | |
|-----------------|-----|---|---------------|----------------------------|--|
| | | Interaction | (0.002–0.035) | (0.024–0.139) | |
| Ch+GlycerolCl- | | Cl_{44} ····H_{22}-C_{16} | 0.008 | 0.0237 | |
| | | Cl_{44} ····H_{25}– C_{18} | 0.005 | 0.0141 | |
| | | Cl_{44} ····H ₇ –O ₁ | 0.021 | 0.0560 | |
| | | Cl_{44} ···· H_{10} – C_3 | 0.012 | 0.0397 | |
| | | O ₁₅ …H ₃₅ -C ₂₉ | 0.014 | 0.0473 | |
| | | O_{15} ···H ₄₃ - C_{33} | 0.013 | 0.0421 | |
| | A 1 | O ₂₃ …H ₄₆ -O ₄₅ | 0.022 | 0.0680 | |
| | AI | O ₅₀ …H ₁₉ –O ₁₈ | 0.021 | 0.0671 | |
| Lignin…lignin | A2 | O ₃₈ …H ₁₉ –O ₁₈ | 0.021 | 0.0694 | |
| Liginii liginii | A3 | $O_{45} \cdots H_{19} - O_{18}$ | 0.027 | 0.0884 | |
| | | $O_{45} \cdots H_{14} - C_{12}$ | 0.009 | 0.0291 | |
| | A4 | O ₃₈ …H ₁₉ –O ₁₈ | 0.024 | 0.0737 | |

Table S1 The electron density (ρ) and density Laplacian $({}^{\bigtriangledown 2}\rho)$ for the ChCl-glycerol and lignin-lignin at the bond critical points calculated at the M06-2X/6-31+G^{**} level

| Metal | State | Lewis acid strength ^[6] | K–T parameters | | | |
|--------------------------------------|-----------------|---------------------------------------|----------------|------|------|--------|
| chloride | | | α | β | π* | LF (%) |
| NaCl | solid-liquid | -1.3697 | | | | |
| LiCl | liquid | -1.0262 | 0.99 | 0.63 | 1.13 | 2.02 |
| CaCl ₂ | solid-liquid | -1.3849 | | | | |
| ZnCl ₂ | crystallization | -0.6115 | | | | 3.38 |
| MgCl ₂ ·6H ₂ O | liquid | -0.5429 | 0.96 | 0.73 | 1.17 | 4.10 |
| FeCl ₃ | liquid | 1.4993 | | | | 11.44 |
| AlCl ₃ ·6H ₂ O | liquid | 2.4401 | 1.96 | 0.66 | 1.13 | 18.27 |

 Table S2 Lewis acid strength and K–T parameters of the ChCl/glycerol DES with different metal chlorides

| label | δ_C / δ_H | assignment | | | |
|----------------|-----------------------|---|--|--|--|
| OMe | 55.7/3.75 | C–H in methoxyls | | | |
| Αγ | 59.9/3.28-3.61 | C_{γ} -H _{γ} in β -O-4' substructures (A) | | | |
| Αα | 71.6/4.81 | C_{α} -H _{α} in β -O-4' substructures (A) | | | |
| $A_{\beta(G)}$ | 83.7/4.30 | C_{β} -H _{β} in β -O-4' substructures linked to G and H units (A) | | | |
| $A_{\beta(s)}$ | 85.9/4.11 | C_{β} -H _{β} in β -O-4' substructures linked to S units (A) | | | |
| Βγ | 70.9/3.77and4.15 | C_{γ} – H_{γ} in resinol substructures (B) | | | |
| Βα | 84.9/4.68 | C_{α} -H _{α} in resinol substructures (B) | | | |
| Ββ | 53.5/3.01 | C_{β} -H _{β} in resinol substructures (B) | | | |
| Сү | 62.8/3.65 | C_{γ} – H_{γ} in phenylcoumaran substructures (C) | | | |
| Ca | 87.0/5.45 | C_{α} -H _{α} in phenylcoumaran substructures (C) | | | |
| Сβ | 53.0/3.47 | C_{β} – H_{β} in phenylcoumaran substructures (C) | | | |
| S2,6 | 103.5/6.01 | $C_{2,6}$ -H _{2,6} in etherified syringyl units (S) | | | |
| G2 | 110.9/6.90 | C_2 – H_2 in G units (G) | | | |
| G5 | 115.0/6.73 | C_5 – H_5 in G units (G) | | | |
| G6 | 118.7/6.78 | C_6 – H_6 in G units (G) | | | |
| X2 | 70.55/3.06 | C ₂ –H ₂ in β –D–xylopyranoside | | | |
| X3 | 74.14/3.28 | C ₃ –H ₃ in β –D–xylopyranoside | | | |
| PB2,6 | 131.84/7.69 | C _{2,6} –H _{2,6} in p-hydroxybenzoate substructures (PB) | | | |

Table S3 Assignment of ¹³C–¹H cross-signals in the HSQC spectra of lignin fractions^[7]

| | MWL | 3c-DESL |
|---------------------|------|---------|
| $M_{ m w}$ | 6838 | 5466 |
| $M_{ m n}$ | 4550 | 3270 |
| $M_{ m w}/M_{ m n}$ | 1.50 | 1.67 |

Table S4 $M_{\rm w}$, $M_{\rm n}$, and polydispersity ($M_{\rm w}/M_{\rm n}$) of MWL and 3c-DESL

Table S5 Analysis of carbohydrates that dissolved in the 3c-DES

| | Content of carbohydrates (wt%) | | | | |
|-------------------------------|---------------------------------------|--------|----------|-------|----------|
| - | Glucose | Xylose | Fructose | 5-HMF | Furfural |
| Wood ^a | 22.7 | 20.0 | - | - | - |
| 3c-DES-extracted ^b | 1.95 | 5.12 | 1.86 | 8.31 | 3.44 |

Annotation: The glucose, xylose, and fructose content were analyzed by HPLC (LC-1260, refractive index detector RID-G1362A, Agilent, USA). A Bio-Rad Aminex HPX-87P column (300 mm×7.8 mm) was used with the following parameters: mobile phase: ultrapure water; flow rate: 0.6 mL/min; injection volume: 20 μ L; column temperature: 80 °C; refractive index detector, retention time: fructose (10.8 min), glucose (11.4 min), and xylose (12.8 min). Furfural and 5-HMF were analyzed by HPLC (Waters 600E, UV detector VMD-2996, USA) using an Acquity BEH C18 column (1.7 μ m, 2.1×50 mm), with the following parameters: mobile phase: acetonitrile:water (10:90, v/v); flow rate: 0.8 ml/min; injection volume: 20 μ L; column temperature: 30 °C; UV detector: detection wavelength of 280 nm; and retention time: 5-HMF (8.4 min), furfural (13.9 min). **a:** The value of glucose and xylose reported by our previous report^[8]. **b:** Liquid phase of 3c-DES after removal of lignin (120 °C, 4h), carbohydrates were calculated by relative mass percentage to wood.



Fig. S1 Topological graph of ChCl-glycerol interactions



Fig. S2 Typical LCC structure and lignin substructures



Fig. S3 Photographs of the mixtures formed by adding metal chloride salts in the ChCl/glycerol DES



Fig. S4 3c-DES-4 at room temperature and low temperature (-20 °C)



Fig. S5 (a) UV spectra of the K–T dyes: N,N-diethyl-4-nitroaniline (NEt₂), 4-nitroaniline (NH₂) and Nile red (NR) in ChCl/glycerol. UV spectra of the 3c-DESs with K–T dyes (b) NH₂, (c) NEt₂ and (d) NR.



Fig. S6 The fractionation process of wood lignocellulose into CR, LF and dissolved carbohydrates by oil bath heating with the 3c-DES







Fig. S8 SEM image of the pseudolignin generated at 120 °C for 4 h.



Fig. S9 Comparison of the lignin extraction rates (LERs) between 3c-DES-4 and

glycerol/AlCl₃·6H₂O (7:1)



Fig. S10 2D-HSQC NMR spectra of (a) MWL and (b) 3c-DESL in the side-chain region and aromatic region (insets)



Fig. S11 ³¹P NMR spectra of MWL and 3c-DESL

References

- C. Froschauer, M. Hummel, M. Iakovlev, A. Roselli, H. Schottenberger and H. Sixta, *Biomacromolecules*, 2013, 14, 1741–1750.
- [2] A. Duereh, Y. Sato, R. L. Smith Jr and H. Inomata, *ACS Sustainable. Chem. Eng.*, 2015, 3, 1881–1889.
- [3] P. G. Jessop, D. A. Jessop, D. Fu and L. Phan, Green Chem., 2012, 14, 1245–1259.
- [4] Z. Xue, X. Zhao, R. C. Sun and T. Mu, ACS Sustain. Chem. Eng., 2016. 4, 3864–3870.
- [5] M. J. Kamlet, J. L. Abboud and R. W. Taft, J. Am. Chem. Soc., 1977, 99, 6027-6038.
- [6] P. Panagiotopoulou, N. Martin and D. G. Vlachos, ChemSusChem, 2015, 8, 2046–2054.
- [7] M. Wu, J. -K. Liu and Z.-Y Yan, RSC Adv., 2016, 6, 6196–6204.
- [8] Y. Liu, W. Chen, Q. Xia, B. Guo, Q. Wang, S. Liu, et al., ChemSusChem, 2017, 10: 1692–1700.