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Evaluating polymer-supported isothiourea catalysis in industriallypreferable solvents for the acylative kinetic resolution of secondary and tertiary heterocyclic alcohols in batch and flow

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General Experimental

Reactions involving moisture sensitive reagents were carried out in flame-dried glassware under an argon or nitrogen atmosphere using standard vacuum line techniques, and using anhydrous solvents. Anhydrous solvents (THF and toluene) were obtained from an anhydrous solvent system (purified using an alumina column, Mbraun SPS-800). All other reactions were performed in standard glassware with no precautions to exclude air or moisture. Solvents and commercial reagents were used as supplied without further purification unless otherwise stated.

Room temperature (r.t.) refers to 20–25 °C. Temperatures of 0 °C and -78 °C were obtained using ice/water and CO₂(s)/acetone baths respectively. Reflux conditions were obtained using a DrySyn, oil bath or sand bath equipped with a contact thermometer.

'in vacuo' refers to the use of either a Büchi Rotavapor R-200 with a Büchi V-491 heating bath and Büchi V-800 vacuum controller, a Büchi Rotavapor R-210 with a Büchi V-491 heating bath and Büchi V-850 vacuum controller, a Heidolph Laborota 4001 with vacuum controller, an IKA RV10 rotary evaporator with a IKA HB10 heating bath and ILMVAC vacuum controller, or an IKA RV10 rotary evaporator with a IKA HB10 heating bath and Vacuubrand CVC3000 vacuum controller. Rotary evaporator condensers are fitted to Julabo FL601 Recirculating Coolers filled with ethylene glycol and set to -5 °C.

Analytical thin layer chromatography was performed on pre-coated aluminium plates (Kieselgel 60 F_{254} silica). TLC visualisation was carried out with ultraviolet light (254 nm), followed by staining with a 1% aqueous KMnO₄ solution. Manual column chromatography was performed in glass columns fitted with porosity 3 sintered discs over Kieselgel 60 silica using the solvent system stated. Automated chromatography was performed on a Biotage Isolera Four running Biotage OS578 with a UV/Vis detector using the method stated and cartridges filled with Kieselgel 60 silica.

Melting points were recorded on an Electrothermal 9100 melting point apparatus and are uncorrected.

Optical rotations were measured on a Perkin Elmer Precisly/Model-341 polarimeter operating at the sodium D line with a 100 mm path cell at 20 °C.

HPLC analyses were obtained using either a Shimadzu HPLC consisting of a DGU-20A5 degassing unit, LC-20AT liquid chromatography pump, SIL-20AHT autosampler, CMB-20A communications bus module, SPD-M20A diode array detector and a CTO-20A column oven; or a Shimadzu HPLC consisting of a DGU-20A5R degassing unit, LC-20AD liquid chromatography pump, SIL-20AHT autosampler, SPD-20A UV/Vis detector and a CTO-20A column oven. Separation was achieved using DAICEL CHIRALCEL OD-H and OJ-H columns or DAICEL CHIRALPAK AD-H and AS-H columns. All HPLC traces of enantiomerically-enriched compounds were compared with authentic racemic spectra.

¹H, and ¹³C nuclear magnetic resonance (NMR) spectra were acquired on either a Bruker Avance II 400 (¹H 400 MHz; ¹³C 101 MHz or a Bruker Avance II 500 (¹H 500 MHz; ¹³C 126 MHz) spectrometer at ambient temperature in the deuterated solvent stated. All chemical shifts are quoted in parts per million (ppm) and referenced to the residual solvent peak. All coupling constants, *J*, are quoted in Hz. Multiplicities are indicated by: s (singlet), d (doublet), t (triplet), q (quartet), sept (septet), dd (doublet of doublets), dt (doublet of triplets), dq (doublet of quartets), td (triplet of doublet), td (doublet of doublet of doublet), td (doublet of triplets), dt (doublet of triplets), dt (doublet of triplets), to denote benzyl, br to denote broad and app to denote apparent.

Infrared spectra were recorded on a Shimadzu IRAffinity-1 Fourier transform IR spectrophotometer fitted with a Specac Quest ATR accessory (diamond puck). Spectra were recorded of either thin films or solids, with characteristic absorption wave numbers (v_{max}) reported in cm⁻¹.

Continuous flow experiments. The catalyst resin was packed into an Omnifit column [borosilicate glass; length = 100 mm (70 mm adjustable bed height); internal diameter = 10 mm; maximum bed volume 5.6 mL]. A Gilson 305 HPLC pump was used to pump solvent for column equilibration and regeneration. A Legato 200 series syringe pump (World Precision Instruments) was used to deliver solutions of reagents. A Huber Ministat was used to circulate ethylene glycol at -5 °C.

Selectivity factors were calculated using the following equations, with all ees determined by chiral HPLC analysis. See reference 1a for the derivation, and alternative forms, of these equations.

$$s = \frac{\ln[(1-\operatorname{conv})(1-\operatorname{ee}_{\operatorname{alcohol}})]}{\ln[(1-\operatorname{conv})(1+\operatorname{ee}_{\operatorname{alcohol}})]} \quad \text{and,} \quad \operatorname{conv} = \frac{\operatorname{ee}_{\operatorname{alcohol}}}{\operatorname{ee}_{\operatorname{alcohol}} + \operatorname{ee}_{\operatorname{ester}}}$$

where both ee and conv are given as between 0 and 1

General Procedures

General procedure A: Preparation of α -substituted arylacetic acids

Following a modified literature method², *n*BuLi (2.2 equiv.) was added to a solution of HN^iPr_2 (2.2 equiv.) in anhydrous THF in a flame-dried round-bottomed flask under a N₂ atmosphere at 0 °C. The LDA solution was stirred for 30 minutes, the arylacetic acid (1.0 equiv.) was added and the reaction mixture stirred for 1 h at 0 °C. The dihaloalkane (2.2 equiv.) was added and the reaction stirred overnight at r.t.. HCl (20 mL) was added until pH 1 was reached. The aqueous layer was extracted with EtOAc (3 × 20 mL). The organic layers were combined, dried (Na₂SO₄), filtered and concentrated *in vacuo* to give the crude α -substituted aryl acetic acid.

General procedure B: Amidation of α -substituted arylacetic acids

Following a literature method³, the crude acid (1 equiv.) was dissolved in anhydrous THF and stirred at 0 °C under N₂. 1,1'-Carbonyldiimidazole (0.95 equiv.) was added and the mixture stirred for 1 h. The desired amine (1.2 equiv.) was added and the reaction mixture warmed to r.t. and stirred for 2 h. Et₂O (30 mL) and Na₂CO₃ (5 mL) were then added and the organic layer separated, washed with brine (30 mL), dried (MgSO₄), filtered and concentrated *in vacuo*.

General procedure C: Preparation of racemic pyrrolidine alcohol substrates

Following a literature method³, NaH (60% in mineral oil) (5 equiv.) was added to the amide (1 equiv.) in anhydrous THF under N₂, and the mixture stirred for 2 h. The reaction was then exposed to air and stirred for 16 h. On completion, NH₄Cl (30 mL) was added and the aqueous layer was extracted with EtOAc (3×30 mL). The organics were combined, dried (MgSO₄), filtered and concentrated *in vacuo*. On occasions when hydroperoxide formation was observed

by ¹H NMR spectroscopic analysis of the crude product, the mixture was dissolved in anhydrous MeOH (15 mL) and NaBH₄ (1.5 equiv.) was added and the mixture stirred for 3 h. 1 M HCl (10 mL) was added and the mixture stirred for a further 1 h. The product was extracted with CH_2Cl_2 (3 × 20 mL), washed with brine (20 mL), dried (MgSO₄), filtered and concentrated *in vacuo*.

General procedure D: Preparation of racemic esters

An anhydride (1.2 equiv.) and DMAP (10 mol%) were added to a solution of alcohol (1 equiv.) in CH₂Cl₂. *i*-Pr₂NEt (1.2 equiv.) was added and the reaction mixture was stirred at r.t. for 18 h. CH₂Cl₂ and added and the organic phase washed sequentially with 1 M HCl (2 × 10 mL), sat. aq. NaHCO₃ (2 × 10 mL) and brine (10 mL), dried (MgSO₄), filtered and concentrated *in vacuo*. The ester products were purified by Biotage® IsoleraTM 4 [SNAP Ultra 10 g, 40 mL min⁻¹, hexane:EtOAc (100:0 2 CV, 100:0 to 70:40 30 CV, (16 ml each))] to afford pure esters.

General Procedure E: Acylative kinetic resolution of secondary alcohols using solid supported isothiourea catalyst

A solid supported isothiourea catalyst (5 mol%), appropriate anhydride (0.55 equiv) and *i*-Pr₂NEt (0.55 equiv) were added to a solution of the appropriate alcohol (1 equiv.) in the required solvent at r.t.. The solution was allowed to stir for the time given. The reaction mixture was filtered under vacuum through a sintered funnel (porosity 4), and the catalyst resin washed with CH_2Cl_2 (30 mL). The filtrate was concentrated *in vacuo* and the residue purified by Biotage® IsoleraTM 4 [SNAP Ultra 10 g, 75 mL min⁻¹, hexane:EtOAc (100:0 2 CV, 100:0 to 70:30 30 CV, (16 ml each))] to afford the ester and alcohol.

For catalyst recycling studies, the catalyst resin was washed sequentially with $CH_2Cl_2/MeOH$ (1:1, 50 mL), MeOH (50 mL), THF (50 mL) and CH_2Cl_2 (50 mL) and then dried under high vacuum at 40 °C for 2 h.

General procedure F: Acylative kinetic resolution of tertiary alcohols using solid supported isothiourea catalyst

A solid supported isothiourea catalyst (5 mol%), appropriate anhydride (0.7 equiv.) and *i*-Pr₂NEt (0.6 equiv.) were added to a solution of the appropriate alcohol (1 equiv.) in the required solvent at r.t.. The solution was allowed to stir for the time given. The reaction mixture was filtered

under vacuum through a sintered funnel (porosity 4), and the catalyst resin washed with CH_2Cl_2 (30 mL). The filtrate was concentrated *in vacuo* and the residue purified by Biotage® IsoleraTM 4 [SNAP Ultra 10 g, 75 mL min⁻¹, hexane:EtOAc (100:0 2 CV, 100:0 to 70:30 30 CV, (16 ml each))] to afford the ester and alcohol.

For catalyst recycling studies, the catalyst resin was washed sequentially with $CH_2Cl_2/MeOH$ (1:1, 50 mL), MeOH (50 mL), THF (50 mL) and CH_2Cl_2 (50 mL) and then dried under high vacuum at 40 °C for 2 h.

General Procedure G: Hydrolysis of esters.

The appropriate ester (1.0 equiv.) was dissolved in MeOH (0.20 M) and 1 M aq. NaOH (3.0 equiv.) was added and the solution stirred at 50-55°C for 30 min for esters derived from secondary alcohols and heated at reflux for 24 h for esters derived from tertiary alcohols. The reaction was cooled to r.t. and concentrated *in vacuo*. The residue was acidified with 1 M HCl and extracted with EtOAc (\times 2). The combined organic fractions were washed sequentially with sat. aq. NaHCO₃ and brine, dried (MgSO₄), filtered and concentrated *in vacuo* to give the corresponding alcohol.

Synthesis of Polymer Supported Catalysts

Azide-functionalised Merrifield resin S1-M

 $\sqrt[N_3]{}$ = Merrifield resin-derived polystyrene support

Following a literature procedure,⁴ (chloromethyl)polystyrene resin (3.0 g, $f = 1.23 \text{ mmol g}^{-1}$) was added to NaN₃ (780 mg, 51 mmol) in DMSO (30 mL). The mixture was heated at 60 °C (without stirring) for 16 h and then cooled to r.t.. The suspension was filtered and washed sequentially with H₂O (500 mL), THF-MeOH 1:1 (250 mL), MeOH (250 mL) and THF (250 mL). The resulting solid was dried *in vacuo* for 24 h at 40 °C to afford (azidomethyl)polystyrene⁵ S1-M (12.6 g). IR v_{max} (solid, cm⁻¹) 2094 (N₃); Elemental analysis (%) C 85.61, H 6.75, N 5.48; $f = 1.20 \text{ mmol g}^{-1}$

Merrifield resin-supported (2R,3S)-HyperBTM derivative 2



(2*R*,3*S*)-3-Isopropyl-2-phenyl-8-(prop-2-yn-1-yloxy)-3,4-dihydro-2*H*-benzo[4,5]thiazolo[3,2*a*]pyrimidine **11** (1.86 g, 5.13 mmol, 1.1 equiv.), *i*Pr₂NEt (2.84 mL, 16.33 mmol, 3.5 equiv.) and CuI (44 mg, 0.233 mmol, 5 mol%) were added to a suspension of (azidomethyl)polystyrene **S1-M** (3.89 g, 4.665 mmol, f = 1.20 mmol/g, 1 equiv.) in THF:DMF 1:1 (52 mL) with slow stirring (100 rpm). The reaction mixture was stirred until disappearance of the azide band (~2094 cm⁻¹) was confirmed by IR (*ca.* 20 h). The suspension was filtered and washed sequentially with THF (1:1, 200 mL), H₂O (200 mL), H₂O-MeOH (1:1, 200 mL), MeOH (200 mL), MeOH-THF (1:1, 200 mL), THF (200 mL) and CH₂Cl₂ (200 mL) and the resin was dried *in vacuo* at 40 °C for 24 h to afford a pale brown resin (2*R*,3*S*)-8-((1-ethyl-1*H*-1,2,3-triazol-4-yl)methoxy)-3-isopropyl-2phenyl-3,4-dihydro-2*H*-benzo[4,5]thiazolo[3,2-*a*]pyrimidine⁵ on polymer support **2** (5.58 g, 100% yield). Elemental analysis (%) C 81.3, H 6.7, N 6.21 *f* = 0.89 mmol g⁻¹.

Azide-functionalised TentaGel resin S1-T



Following a literature procedure,⁴ TentaGel bromide resin (2.0 g, $f = 0.26 \text{ mmol g}^{-1}$) was added to NaN₃ (1.56 g, 51 mmol) in DMSO (60 mL). The mixture was heated at 60 °C (without stirring) for 16 h and then cooled to r.t.. The suspension was filtered and washed sequentially with H₂O (750 mL), THF-MeOH (1:1, 500 mL), MeOH (500 mL) and THF (500 mL). The resulting solid was dried *in vacuo* for 24 h at 40 °C to afford azide-functionalised Tentagel **S1-T** (2.0 g). Elemental analysis (%) C 65.14, H 8.9, N 1.425; $f = 0.34 \text{ mmol g}^{-1}$. Functionality (*f*) calculated according to literature method⁶

TentaGel resin-supported (2R,3S)-HyperBTM derivative 3



(2*R*,3*S*)-3-Isopropyl-2-phenyl-8-(prop-2-yn-1-yloxy)-3,4-dihydro-2*H*-benzo[4,5]thiazolo[3,2*a*]pyrimidine **1** (253 mg, 0.975 mmol, 1.1 equiv.), *i*Pr₂NEt (400 µL, 2.31 mmol, 3.5 equiv.) and CuI (6.3 mg, 0.033 mmol, 5 mol%) were added to a suspension of TentaGel azide **S1-T** (2.0 g, 0.66 mmol, f = 0.34 mmol/g, 1 equiv.) in THF:DMF (1:1, 35 mL) with slow stirring (100 rpm). The reaction mixture was stirred for 24 h. The suspension was filtered and washed sequentially with THF (1:1, 400 mL), H₂O (400 mL), H₂O-MeOH (1:1, 400 mL), MeOH (400 mL), MeOH-THF (1:1, 400 mL), THF (400 mL) and CH₂Cl₂ (400 mL) and the resin was dried *in vacuo* at 40 °C for 24 h to give TentaGel resin-supported (2*R*,3*S*)-HyperBTM derivative **3** as a brown resin (1.96 g, 88% yield). Elemental analysis (%) C 63.61, H 8.75, N 1.83. f = 0.26 mmol g⁻¹.

Azide-functionalised Wang resin S1-W



Following a literature procedure,⁴ brominated Wang resin (1.5 g, f = 0.5-1.5 mmol g⁻¹) was added to NaN₃ (1.17 g, 51 mmol) in DMSO (45 mL). The mixture was heated at 60 °C (without stirring) for 16 h and then cooled to r.t.. The suspension was filtered and washed sequentially with H₂O (500 mL), THF-MeOH (1:1, 250 mL), MeOH (250 mL) and THF (250 mL). The resulting solid was dried *in vacuo* for 24 h at 40 °C to afford azide-functionalised Wang resin **S1-W** (1.00 g). Elemental analysis (%) C 84.09, H 8.12, N 2.72; f = 0.65 mmol g⁻¹.

Functionality (*f*) calculated according to literature method.⁶

Wang resin-supported (2R,3S)-HyperBTM derivative 4



(2*R*,3*S*)-3-Isopropyl-2-phenyl-8-(prop-2-yn-1-yloxy)-3,4-dihydro-2*H*-benzo[4,5]thiazolo[3,2*a*]pyrimidine⁵ (374 mg, 1.07 mmol, 1.1 equiv.), *i*Pr₂NEt (594 µL, 3.41 mmol, 3.5 equiv.) and CuI (9.3 mg, 0.049 mmol, 5 mol%) were added to a suspension of Wang azide **S1-W** (1.5 g, 0.975 mmol, f = 0.65 mmol/g, 1 equiv.) in THF:DMF (1:1, 30 mL) with slow stirring (100 rpm). The reaction mixture was stirred for 24 h. The suspension was filtered and washed sequentially with THF (1:1, 300 mL), H₂O (300 mL), H₂O-MeOH (1:1, 300 mL), MeOH (300 mL), MeOH-THF (1:1, 300 mL), THF (300 mL) and CH₂Cl₂ (300 mL) and the resin was dried *in vacuo* at 40 °C for 24 h to give Wang resin-supported (2*R*,3*S*)-HyperBTM derivative **4** as a brown resin (1.47 g, 80% yield). Elemental analysis (%) C 82.16, H 7.07, N 2.755. f = 0.39 mmol g⁻¹.

Synthesis of Polymer Supported BTM 6



The multi-step sequence for the synthesis of polystyrene-supported variant of BTM **6**, started with an S_NAr reaction of the HCl salt of (*R*)-phenylglycinol **S2** with 2-chloro-6-methoxybenzo[*d*]thiazole **S3** followed by *in situ* mesylation and heating to afford cyclized product (*R*)-7-methoxy-2-phenyl-2,3-dihydrobenzo[*d*]imidazo[2,1-*b*]thiazole **S4**. This upon demethylation gave (*R*)-7-hydroxy-2-phenyl-2,3-dihydrobenzo[*d*]imidazo[2,1-*b*]thiazol-1-ium bromide **S5**. *O*-Propargylation of **S5** generated alkyne-substituted BTM derivative **5**. Finally, the attachment of Alkyne **5** to an azidomethyl polystyrene support **S1-M**, synthesized from commercially available (chloromethyl)polystyrene (Merrifield resin) by the procedure described by Pericàs, was achieved by a Cu-catalyzed azide-alkyne cycloaddition reaction⁷. The nitrogen content of polymer **S1-M**, determined by elemental analysis, was used to calculate the functionalization⁶ of **6** (0.89 mmol g⁻¹), with this value used to determine catalyst loading in all subsequent KRs. The extent of functionalization (*f*) (mmol/g) of **6** was analysed by nitrogen (%) elemental analysis using the formula outlined by Pericàs⁶ (*f* = %N × 1000 × (number of N atoms in functional unit)⁻¹ × (14.001)⁻¹ × 100⁻¹) and found to be 0.86 mmol g⁻¹.

(R)-7-Methoxy-2-phenyl-2,3-dihydrobenzo[d]imidazo[2,1-b]thiazole S4



Following a modification of a literature procedure,⁸ a yellow suspension of (R)-phenylglycinol S2 (3.61 g, 26.3 mmol, 1.05 equiv), iPr₂NEt (6.50 mL, 37.6 mmol, 1.5 equiv.), 2-chloro-6methoxybenzo[d]thiazole S3 (5.0 g, 25.0 mmol, 1 equiv.) and o-dichlorobenzene (15 mL) was heated at reflux (195 °C DrySyn[®]) until completion as judged by TLC (ca. 48 h). The orange mixture was allowed to cool to r.t., H₂O (40 mL) was added and the aqueous phase was extracted with CH_2Cl_2 (3 × 50 mL). The organic layers were combined, washed with brine, dried (MgSO₄), filtered and concentrated *in vacuo*. The resulting residue was triturated with hexane to afford the crude product as an off-white solid that was recrystallised from toluene to give (R)-2-((6-methoxybenzo[d]thiazol-2-yl)amino)-2-phenylethan-1-ol as fluffy colourless crystals (5.80 g, 82% yield). $[\alpha]_D^{20} = -36.5$ (c 1.0 CHCl₃); mp 130-133 °C; v_{max} (thin film, cm⁻¹) 1604 (C=C), 1548 (C=N); ¹H NMR (400 MHz, CDCl₃) $\delta_{\rm H}$: 3.80 (3H, s, OCH₃), 3.85 – 4.06 (2H, m, C(1)H₂), 4.80 (1H, dd, J 6.3, 4.0, C(2)H), 5.98 (1H, bs, NH), 6.88 (1H, dd, J 8.8, 2.6, C(5)ArH), 7.04 (1H, d, J 2.6, C(7)ArH), 7.22 – 7.40 (5H, m, PhC(2,3,4,5,6)H), 7.44 (1H, d, J 8.8, C(4)ArH); ¹³C{¹H} NMR (126 MHz, CDCl₃) δ_C: 56.0 (OCH₃), 62.4 (C(1)H₂), 66.8 (C(2)H), 105.4 (ArC(5)H), 113.8 (ArC), 119.1 (ArC), 127.0 (2 × PhCH), 128.2 (PhCH), 128.9 (2 × PhCH), 131.4 (PhC), 138.7 (ArC(7)), 145.5 (ArC(3a)), 155.3 (ArC(6)), 166.9 (ArC=N), HRMS (ESI⁺) C₁₆H₁₇N₂O₂S [M+H]⁺, found 301.1005, requires 301.1011 (-1.9 ppm).

(*R*)-2-((6-Methoxybenzo[*d*]thiazol-2-yl)amino)-2-phenylethan-1-ol (5.8g, 19.31 mmol, 1 equiv.) and Et₃N (10.80 mL, 77.24 mmol, 4 equiv.) in anhydrous CH₂Cl₂ was stirred at 0 °C. After 10 min methanesulfonyl chloride (1.74 mL, 25.1 mmol, 1.3 equiv.) was added with stirring. The ice/water bath was removed, and the reaction stirred for 15 mins. Once complete consumption of (*R*)-2-((6-methoxybenzo[*d*]thiazol-2-yl)amino)-2-phenylethan-1-ol was observed by TLC, *i*PrOH (0.3 mL) was added and the reaction was heated at reflux for 16 h. The reaction was quenched with 1 M NaOH (50 mL) and the biphasic mixture stirred vigorously for 30 mins. The aqueous layer was extracted with CH₂Cl₂ (3 × 50 mL) and the combined organic phases washed with brine (50 mL), dried (MgSO₄), filtered and concentrated *in vacuo* to afford the crude product

which was purified by Biotage® IsoleraTM 4 [SNAP Ultra 50 g, 100 mL min⁻¹, CH₂Cl₂ :EtOAc (95 : 5 5CV, 95 : 5 to 80 : 20 10 CV, 80 : 20 3 CV)] to give (*R*)-7-methoxy-2-phenyl-2,3-dihydrobenzo[*d*]imidazo[2,1-*b*]thiazole **S4** as a colourless crystalline solid (2.90 g, 53%); $[\alpha]_D^{20}$ = +58.2 (*c* 0.5 CHCl₃); mp 106-108 °C; ν_{max} (thin film) 1597 (C=C), 1573 (C=N); ¹H NMR (500 MHz, CDCl₃) δ_{H} : 3.68 (1H, app. t, *J* 8.6, C(3)*H*^AH^B), 3.78 (3H, s, OC*H*₃), 4.26 (1H, dd, *J* 10.1, 8.6, C(3)H^AH^B), 5.65 (1H, dd, *J* 10.1, 8.6, C(2)*H*), 6.60 (1H, d, *J* 8.5, C(5)*H*), 6.74 (1H, dd, *J* 8.6, 2.5, C(6)Ar*H*), 6.93 (1H, d, *J* 2.6, C(8)Ar*H*), 7.27-7.32 (1H, m, Ph*H*), 7.33-7.41 (4H, m, 4 × Ph*H*); ¹³C{¹H} NMR (126 MHz, CDCl₃) δ_C : 53.2 (*C*(3)H₂), 56.1 (OC*H*₃), 75.2 (*C*(2)H), 109.0 (Ar*C*(5)H), 109.7 (Ar*C*(6)H), 112.3 (Ar*C*(8)H), 126.6 (2 × Ph*C*H), 127.7 (Ph*C*H), 128.6 (Ph*C*(1)), 128.8 (2 × Ph*C*H), 131.4 (Ar*C*(8a)), 142.9 (Ar*C*(4a)), 155.2 (Ar*C*(7)), 167.4 (Ar*C*=N); HRMS (ESI⁺) C₁₆H₁₅N₂OS [M+H]⁺, found 283.0900, requires 283.0905 (-1.7 ppm).

(R)-7-Hydroxy-2-phenyl-2,3-dihydrobenzo[d]imidazo[2,1-b]thiazol-1-ium bromide S5



BBr₃ (33.6 mL, 33.60 mmol, 1 M in CH₂Cl₂, 10 equiv.) was added dropwise to a solution of (*R*)-7-methoxy-2-phenyl-2,3-dihydrobenzo[*d*]imidazo[2,1-*b*]thiazole **S3** (950 mg, 3.36 mmol, 1 equiv.) in CH₂Cl₂ (10 mL) at 0 °C. The solution was stirred at 0 °C for 2 h then warmed to r.t. and stirred for 16 h. The reaction was carefully quenched with MeOH (10 mL) at 0 °C and warmed to r.t. CH₂Cl₂ (10 mL) was added and the aqueous phase was extracted with CH₂Cl₂ (3 × 20 mL). The organic layers were combined, dried (MgSO₄), filtered and concentrated *in vacuo* to give (*R*)-7-hydroxy-2-phenyl-2,3-dihydrobenzo[*d*]imidazo[2,1-*b*]thiazol-1-ium bromide **S4** as a colorless solid (1.15 g, 97%); $[\alpha]_D^{20} = +93.6$ (*c* 1.0 MeOH), mp182-185 °C; v_{max} (thin film, cm⁻¹) 3026 (C-H), 1558 (C=N), 1550 (N-C); ¹H NMR (500 MHz, *d*₆-DMSO) δ_{H} : 4.34 (1H, dd, *J* 10.7, 8.4, C(3)*H*^AH^B), 4.92 (1H, app. t, *J* 10.7, C(3)H^AH^B), 5.92 (1H, dd, *J* 10.7, 8.4, C(2)*H*), 6.97 (1H, dd, *J* 8.7, 2.4, ArC(6)*H*), 7.36 (1H, d, *J* 8.7, ArC(5)*H*), 7.40-7.51 (4H, m, PhC(3,4,5)*H*, ArC(8)*H*), 7.53-7.58 (2H, m, PhC(2,6)*H*), 9.98 (1H, s, N*H*), 10.66 (1H, s, O*H*); ¹³C{¹H} NMR (126 MHz, *d*₆-DMSO) δ_{C} : 53.3 (*C*(3)H₂), 66.1 (*C*(2)H), 111.0 (ArC(8)H), 113.5 (ArC(5)H), 115.8 (ArC(6)H), 127.2 (PhC(2,6)H), 127.4 (ArC(8a)), 128.7 (ArC(4a)), 129.03 (PhC(4)H), 129.05 (Ph*C*(3,5)H), 138.8 (Ph*C*(1)), 154.9 (Ar*C*(7)), 168.7 (*C*=N); HRMS (ESI⁺) $C_{15}H_{13}N_2OS$ [M+H]⁺, found 269.0736, requires 269.0743 (-2.6 ppm).

(R)-2-Phenyl-7-(prop-2-yn-1-yloxy)-2, 3-dihydrobenzo[d]imidazo[2,1-b]thiazole 5



KOtBu (919 mg, 8.19 mmol, 2.6 equiv.) was added to a solution of (R)-7-hydroxy-2-phenyl-2,3dihydrobenzo[d]imidazo[2,1-b]thiazol-1-ium bromide S4 (1.1g, 3.15 mmol, 1 equiv.) in THF/DMSO (1 mL of each) and the reaction mixture was stirred at 0 °C for 2 h. Propargyl bromide (526 µL, 4.73 mmol, 80% in toluene, 1.5 equiv.) was added and the reaction mixture was allowed to warm to r.t. over 2 h. Brine (20 mL) was added and the aqueous phase was extracted with EtOAc (3×30 mL). The organic layers were combined, washed with brine, dried (MgSO₄), filtered and concentrated *in vacuo*. The residue was purified by Biotage® IsoleraTM 4 [SNAP Ultra 25 g, 75 mL min⁻¹, CH₂Cl₂:EtOAc (95 : 5 5CV, 95 : 5 to 60 : 40 10 CV, 60 : 40 5 CV)] to give (R)-2-phenyl-7-(prop-2-yn-1-yloxy)-2,3-dihydrobenzo[d]imidazo[2,1-b]thiazole 5 as a colorless semi-solid (0.89 g, 92%); $[a]_{D}^{20}$ +26.4 (c 1.0 CHCl₃); v_{max} (thin film, cm⁻¹) 1591 (C=C), 1575 (C=N); ¹H NMR (500 MHz, CDCl₃) δ_H: 2.53 (1H, t, J 2.4, C=H), 3.67 (1H, app. t, J 8.5, C(3)H^AH^B), 4.24 (1H, dd, J 10.2, 8.7, C(3)H^AH^B), 4.65 (2H, d, J 2.4, OCH₂), 5.64 (1H, dd, J 10.2, 8.2, C(2)H), 6.59 (1H, d, J 8.5, ArC(5)H), 6.83 (1H, dd, J 8.5, 2.5, ArC(6)H), 7.01 (1H, d, J 2.5, ArC(8)H), 7.26-7.32 (1H, m, PhH), 7.34-7.40 (4H, m, PhC(2,3,5,6)H); ¹³C{¹H} NMR (126 MHz, CDCl₃) δ_C: 53.0 (*C*(3)H₂), 57.0 (OCH₂), 75.5 (*C*(2)H), 75.9 (C=*C*H), 78.5 (*C*=CH), 108.8 (ArC(5)H), 111.3 (ArC(6)H), 113.7 (ArC(8)H), 126.6 ($2 \times PhCH$), 127.7 (PhC(4)H), 128.5 (PhC(1)), 128.8 (2 \times PhCH), 132.3 (ArC(8a)), 143.0 (ArC(4a)), 152.8 (ArC(7)), 167.1 (C=N); HRMS(ESI⁺) $C_{18}H_{15}N_2OS [M+H]^+$, found 307.0895, requires 307.0900 (-1.6 ppm).

Merrifield resin-supported (R)-BTM derivative 6



To a round bottomed flask containing (azidomethyl)polystyrene⁵ **S1-M** (1.6 g, 2.55 mmol, f = 1.20 mmol/g, 1 equiv.) suspended in THF:DMF (1:1, 60 mL) was added (*R*)-2-phenyl-7-(prop-2-yn-1-yloxy)-2,3-dihydrobenzo[*d*]imidazo[2,1-*b*]thiazole **5** (860 mg, 2.81 mmol, 1.1 equiv.), *i*Pr₂NEt (1.6 mL, 8.90 mmol, 3.5 equiv.) and CuI (24 mg, 0.126 mmol, 5 mol%) with slow stirring. The reaction mixture was stirred until disappearance of the azide band (~2094 cm⁻¹) was confirmed by IR (*ca.* 21 h). The suspension was filtered and washed sequentially with THF (130 mL), H₂O (130 mL), H₂O-MeOH (1:1, 130 mL), MeOH (130 mL), MeOH-THF (1:1, 200 mL), THF (100 mL) and CH₂Cl₂ (100 mL) and the resin was dried *in vacuo* at 35-40 °C for 24 h to give Merrifield resin-supported (*R*)-BTM derivative **6** as a dark brown resin (2.45 g, quantitative yield); Elemental analysis (%) C 79.84, H 6.71, N 6.04; f = 0.86 mmol/g

Synthesis of Racemic Alcohols

Racemic alcohols given below were prepared according to the literature reported^{5,9} from this group.



1-Allyl-3-hydroxy-3-phenylindolin-2-one 20



Phenylmagnesium bromide (3.0 M, 1.2 mL, 3.6 mmol) was added dropwise to a solution of 1allylindoline-2,3-dione (555 mg, 3.0 mmol) in anhydrous THF (30 mL) at -78 °C under a N₂ atmosphere. The solution stirred at -78 °C for 20 mins, then at 0 °C with the reaction monitored by TLC until completion (typically within 30 min). The reaction mixture reaction was poured into aqueous NH₄Cl solution (20 mL) and extracted with EtOAc (3 × 20 mL). The organic layers were combined, dried (MgSO₄), filtered and concentrated *in vacuo*. The residue was purified by flash column chromatography (CH₂Cl₂:EtOAc 9:1, R_F 0.40) to give 1-allyl-3-hydroxy-3phenylindolin-2-one **20** as a yellow powder (702 mg, 2.6 mmol, 88%). m.p. 132-134 °C {Lit. 134.5-135.5 °C}; ¹H NMR (400 MHz, CDCl₃) $\delta_{\rm H}$: 3.38 (1H, br s, OH), 4.28 (1H, ddt, *J* 16.4, 5.4, 1.7, CH_AH_BCH=CH₂), 4.46 (1H, ddt, *J* 16.4, 5.4, 1.7, CH_AH_BC=CH₂), 5.27 (2H, m, HRC=CH₂), 5.87 (1H, m, CH₂CH=CH₂), 6.90 (1H, app dt, *J* 7.8, 0.8, C(7)H), 7.08 (1H, app td, *J* 7.5, 1.0, C(5)H), 7.27-7.43 (7H, m, ArCH). Data were in accordance with those previously reported.¹⁰

3-Hydroxy-1-methyl-3-phenylindolin-2-one 21



Phenylmagnesium bromide (3.0 M, 1.5 mL, 4.4 mmol) was added dropwise to a solution of 1methylindoline-2,3-dione (600 mg, 3.7 mmol) in anhydrous THF (35 mL) at -78 °C under a N₂ atmosphere. The solution stirred at -78 °C for 20 mins, then at 0 °C with the reaction monitored by TLC until completion (typically within 30 min). The reaction mixture reaction was poured into aqueous NH₄Cl solution (20 mL) and extracted with EtOAc (3 × 20 mL). The organic layers were combined, dried (MgSO₄), filtered and concentrated *in vacuo*. The residue was purified by flash column chromatography (CH₂Cl₂:EtOAc 9:1, R_F 0.26) to give 3-hydroxy-1-methyl-3phenylindolin-2-one **21** as a yellow powder (730 mg, 3.0 mmol, 82%). m.p. 123-125 °C {Lit. 141-142 °C}; ¹H NMR (400 MHz, CDCl₃) δ_{H} : 3.25 (3H, s, NC*H*₃), 3.41 (1H, s, O*H*), 6.91 (1H, app dt, *J* 7.9, 0.7, C(7)*H*), 7.09 (1H, app td, *J* 7.5, 1.0, C(5)*H*), 7.26-7.41 (7H, m, ArC*H*); ¹³C NMR (100 MHz, CDCl₃) δ_{C} : 26.7 (NCH₃), 78.0 (*C*(3)), 108.7 (ArC(7)H), 123.6 (ArC(5)H), 124.9 (ArC(6)H), 125.3 (C(3)ArC(2,6)H), 128.3 (C(3)ArC(4)H), 128.6 (C(3)ArC(3,5)H), 129.9 (ArC(4)H), 131.5 (C(3)ArC(1)), 140.1 (ArC(3a)), 143.5 (ArC(7a)), 177.5 (*C*=O). Data were in accordance with those previously reported.¹¹

1-Allyl-3-hydroxy-3-(thiophen-2-yl)indolin-2-one 24



n-Butyl lithium (2.5 M in hexanes, 1.6 mL, 4.00 mmol) was added dropwise to a solution of thiophene (264 µL, 3.3 mmol) in anhydrous THF (13 mL) at 0 °C under N₂, and the reaction stirred for 1 hour. In a separate flask, 1-allylindoline-2,3-dione (561 mg, 3 mmol) was dissolved in anhydrous THF (15 mL), cooled to 0 °C under N₂, and the solution of 2-thienyllithium added dropwise over 15 mins. The reaction was stirred at 0 °C for 1 h. On completion, saturated aqueous NH₄Cl was added, and the aqueous phase extracted with EtOAc (3×50 mL). The organic layer was washed with brine $(3 \times 50 \text{ mL})$, dried (MgSO₄), filtered and concentrated in vacuo. The crude product was purified by Biotage® Isolera 4 chromatography (eluent: $0\% \rightarrow 30\%$ EtOAc in hexanes) to give 1-allyl-3-hydroxy-3-(thiophen-2-yl)indolin-2-one 24 as a vellow solid (594 mg, 2.2 mmol, 73%). m.p. 126-128 °C; ¹H NMR (500 MHz, CDCl₃) δ_H: 3.42 (1H, s, OH), 4.23-4.30 (1H, m, NCH_AH_B), 4.41-4.48 (1H, m, NCH_AH_B), 5.20-5.27 (2H, m, CH=CH₂), 5.80-5.89 (1H, m, CH=CH₂), 6.88 (1H, app d, J 7.8, ArC(7)H), 6.94 (1H, d, J 5.0, 3.7, C(3)ArC(4)H), 6.99 (1H, dd, J 3.6, 1.0, C(3)ArC(3)H), 7.14 (1H, app t, J 7.5, ArC(5)H), 7.32 (1H, dd, J 5.0, 1.0, C(3)ArC(5)H), 7.34 (1H, app td, J 7.7, 1.0, ArC(6)H), 7.54 (1H, app. d, J 7.5, ArC(4)H); ¹³C NMR (126 MHz, CDCl₃) δ_C: 42.7 (NCH₂), 75.5 (C(3)), 109.9 (ArC(7)H), 118.0 (CH=CH₂), 123.5 (ArC(5)H), 125.2 (ArC(4)H), 126.0 (C(3)ArC(4)H), 126.9 (C(3)ArC(5)H), 127.0 (C(3)ArC(3)H), 130.3 (ArC(6)H), 130.5 (ArC(3a)), 131.2 (CH=CH₂), 142.5 (C(3)ArC(2)), 143.6 (ArC(7a)), 175.9 (C=O); IR (neat) v_{max} cm⁻¹ 3337 (OH), 3103, 3057,

1692 (C=O), 1612, 1487, 1466, 1373, 1180, 1159; HRMS (NSI⁺) calculated for $C_{15}H_{13}NO_2SNa^+$ ([M+Na]⁺) requires 294.0559; found 294.0560 (+0.3 ppm).

1-Benzyl-6-chloro-3-hydroxy-3-phenylindolin-2-one 29



Phenylmagnesium bromide (3.0 M, 0.91 mL, 2.7 mmol) was added dropwise to a solution of 1benzyl-6-chloroindoline-2,3-dione (615 mg, 2.26 mmol) in anhydrous THF (22 mL) at -78 °C under a N₂ atmosphere. The solution stirred at -78 °C for 20 mins, then at 0 °C with the reaction monitored by TLC until completion (typically within 30 min). The reaction mixture reaction was poured into aqueous NH₄Cl solution (20 mL) and extracted with EtOAc (3 \times 20 mL). The organic layers were combined, dried (MgSO₄), filtered and concentrated in vacuo. The residue was dissolved in the minium amount of hot CH₂Cl₂ and then cooled in an ice bath to give 1benzyl-6-chloro-3-hydroxy-3-phenylindolin-2-one 29 as a colourless solid (541 mg, 1.55 mmol, 68%). m.p. 190-191 °C; ¹H NMR (500 MHz, CDCl₃) $\delta_{\rm H}$: 3.41 (1H, br s, OH), 4.81 (1H, d, J 15.7, CH_AH_BPh), 5.02 (1H, d, J 15.7, CH_AH_BPh), 6.78 (1H, d, J 1.8, ArC(7)H), 7.02 (1H, dd, J 7.9, 1.8, ArC(5)*H*), 7.19 (1H, d, *J* 7.9, ArC(4)*H*), 7.28-7.41 (10H, m, CH₂Ar*H* + C(3)Ar*H*); ¹³C NMR (126 MHz, CDCl₃) δ_C: 44.2 (CH₂Ph), 77.6 (C(3)), 110.4 (ArC(7)H), 123.5 (ArC(5)H), 125.2 (C(3)ArC(2,6)H), 126.0 (ArC(4)H), 127.2 (CH₂ArC(2,6)H), 128.1 (CH₂ArC(4)H), 128.6 (C(3)ArC(4)H), 128.8 (C(3)ArC(3,5)H), 129.1 (CH₂ArC(3,5)H), 129.9 (ArC(3a)), 134.8 (CH₂ArC(1)), 135.6 (ArC(6)), 139.6 (C(3)ArC(1)), 143.9 (ArC(7a)), 177.5 (C=O). Data were in accordance with those previously reported¹².



Following general procedure A, *n*BuLi (13.2 mL, 33 mmol, 2.5 M in hexanes), HNⁱPr₂ (4.6 mL, 33 mmol), 4-chlorophenylacetic acid (2.55 g, 15 mmol) and 1-bromo-2-chloroethane (2.75 mL, 33 mmol) in anhydrous THF (50 mL) gave 4-chloro-2-phenylbutanoic acid, which was used without further purification. Following general procedure B, 4-chloro-2-phenylbutanoic acid (3.60 g, 15.5 mmol), 1,1'-carbonyldiimidazole (2.39 g, 14.73 mmol) and allylamine (1.4 mL, 18.6 mmol) in anhydrous THF (50 mL) gave, after column chromatography (eluent CH₂Cl₂/EtOAc, 9:1; R_F 0.45) N-allyl-4-chloro-2-(4-chlorophenyl)butanamide as a cream solid (2.02 g, 7.45 mmol, 50%), mp 70-72 °C; v_{max} (ATR) 3296 (NH), 2970, 1639 (C=O), 1555, 1489, 1261; ¹H NMR (400 MHz, CDCl₃) $\delta_{\rm H}$: 2.06 (1H, app. dtd, J 14.6, 7.2, 4.8, C(3) $H^{\rm A}H^{\rm B}$), 2.45 (1H, app. dtd, J 14.9, 7.6, 4.9, C(3)H_AH_B), 3.28 (1H, ddd, J 11.1, 7.8, 4.8, C(4)H_AH_B), 3.46 (1H, ddd, J 11.6, 7.0, 4.9, C(4)H_AH_B), 3.65 (1H, app. t, J 7.5, C(2)H), 3.66-3.80 (2H, m, NCH₂CH=CH₂), 4.91-5.00 (2H, m, NCH₂CH=CH₂), 5.65 (1H, app. ddt, J 16.9, 10.7, 5.4, NCH₂CH=CH₂), 5.99-6.04 (1H, br s, NH), 7.16-7.24 (4H, m, ArC(2,3,5,6)H); $^{13}C{^1H}$ NMR (100 MHz, CDCl₃) δ_C : 35.8 (C(3)H₂), 42.0 (NCH₂), 43.0 (C(4)H₂), 48.8 (C(2)H), 116.2 (NCH₂CH=CH₂), 129.1 (ArC(3,5)H), 129.4 (ArC(2,6)H), 133.5 (ArC(4)), 133.9 (NCH₂CH=CH₂), 137.3 (ArC(1)), 172.0 (C=O); m/z (NSI) 272 ([M+H]⁺, 100%) C₁₃H₁₆NOCl₂⁺ ([M+H]⁺) requires 272.0603; found 272.0606 (+0.9 ppm).

Following general procedure C, *N*-allyl-4-chloro-2-(4-chlorophenyl)butanamide (2.02 g, 7.45 mmol) and NaH (60% in mineral oil) (1.49 mg, 37.25 mmol) in anhydrous THF (50 mL) gave, after column chromatography (eluent CH₂Cl₂/EtOAc, 4:1; $R_F 0.24$) 1-allyl-3-(4-chlorophenyl)-3-hydroxypyrrolidin-2-one **31** as a colourless solid (898 mg, 3.58 mmol, 48%), mp 76-78 °C; v_{max} (ATR) 3202 (OH), 2876, 1668 (C=O), 1489, 1271; ¹H NMR (400 MHz, CDCl₃) δ_{H} : 2.29 (1H, ddd, *J* 13.1, 7.3, 3.8, C(4)*H*_AH_B), 2.42 (1H, ddd, *J* 13.2, 8.4, 7.0, C(4)H_AH_B), 3.25 (1H, app. dt, *J* 10.0, 7.1, C(5)*H*_AH_B), 3.40 (1H, ddd, *J* 10.0, 8.5, 3.8, C(5)H_AH_B), 3.93 (1H, app. ddt, *J* 15.1, 6.1,

1.3, NC*H*_AH_BCH=CH₂), 4.01 (1H, app. ddt, *J* 15.1, 6.2, 1.3, NCH_A*H*_BCH=CH₂), 4.23 (1H, s, O*H*), 5.20-5.27 (2H, m, NCH₂CH=CH₂), 5.75 (1H, app. ddt, *J* 17.7, 9.7, 6.2, NCH₂CH=CH₂), 7.24-7.31 (4H, m, ArC(2,3,5,6)*H*); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ_{C} : 36.2 (*C*(4)H₂), 43.2 (*C*(5)H₂), 46.0 (N*C*H₂CH=CH₂), 78.5 (*C*(3)), 118.9 (NCH₂CH=CH₂), 126.8 (Ar*C*(3,5)H), 128.7 (Ar*C*(2,6)H), 131.7 (NCH₂CH=CH₂), 133.7 (Ar*C*(4)), 141.2 (Ar*C*(1)), 174.6 (*C*=O); *m*/*z* (NSI) 252 ([M+H]⁺, 100%) C₁₃H₁₅NO₂Cl⁺ ([M+H]⁺) requires 252.0786; found 252.0784 (-0.7 ppm).

1-Allyl-3-hydroxy-3-(thiophen-3-yl)pyrrolidin-2-one 32



Following general procedure A, nBuLi (8.8 mL, 22 mmol, 2.5 M in hexanes), HNⁱPr₂ (3.1 mL, 22 mmol), 3-thienylacetic acid (1.42 g, 10 mmol) and 1-bromo-2-chloroethane (1.82 mL, 22 mmol) in anhydrous THF (50 mL) gave 4-chloro-2-(thiophen-3-yl)butanoic acid, which was used without further purification. Following general procedure B, 4-chloro-2-(thiophen-3-yl)butanoic acid (1.89 g, 10 mmol), 1,1'-carbonyldiimidazole (1.54 g, 9.5 mmol) and allylamine (898 µL, 12 mmol) in anhydrous THF (50 mL) gave, after column chromatography (eluent Petrol/EtOAc, 7:3), impure 4-chloro-N-phenyl-2-(thiophen-3-yl)butanamide, which was used without further purification. Following general procedure C, 4-chloro-N-phenyl-2-(thiophen-3-yl)butanamide (1.23 g, 5.1 mmol) and NaH (60% in mineral oil) (1.02 g, 25.5 mmol) in anhydrous THF (50 mL) gave, after column chromatography (eluent $CH_2Cl_2/EtOAc$, 4:1; R_F 0.14), 1-allyl-3hydroxy-3-(thiophen-3-yl)pyrrolidin-2-one **32** as a yellow oil (459 mg, 2.06 mmol, 40%); v_{max} (ATR) 3345 (OH), 2964, 1674 (C=O), 1416, 1271; ¹H NMR (400 MHz, CDCl₃) δ_H: 2.38-2.54 (2H, m, C(4)H₂), 3.23-3.33 (1H, m, C(5)H_AH_B), 3.32 (1H, s, OH), 3.37 (1H, ddd, J 9.9, 8.4, 2.9, C(5)H_AH_B), 3.94 (1H, app. ddt, J 15.2, 6.1, 1.1, NCH_AH_BCH=CH₂), 4.01 (1H, app. ddt, J 15.1, 6.2, 1.1, NCH_AH_BCH=CH₂), 5.18-5.26 (2H, m, NCH₂CH=CH₂), 5.75 (1H, app. ddt, J 16.5, 10.4, 6.1, NCH₂CH=CH₂), 7.14 (1H, dd, J 5.0, 1.4, ArC(4)H), 7.28 (1H, dd, J 3.0, 1.4, ArC(2)H), 7.32 (1H, dd, J 5.0, 3.0, ArC(5)H); ${}^{13}C{}^{1}H$ NMR (100 MHz, CDCl₃) δ_{C} : 34.9 (C(4)H₂), 42.9 (C(5)H₂), 45.9 (NCH₂), 76.3 (C(3)), 118.7 (NCH₂CH=CH₂), 121.5 (ArC(2)H), 125.5 (ArC(4)H),

126.8 (Ar*C*(5)H), 131.7 (NCH₂*C*H=CH₂), 143.0 (Ar*C*(3)), 173.7 (*C*=O); m/z (NSI) 224 ([M+H]⁺, 100%) C₁₁H₁₄NO₂S⁺ ([M+H]⁺) requires 224.0740; found 224.0738 (-0.8 ppm).

1-Allyl-3-hydroxy-3-(thiophen-2-yl)pyrrolidin-2-one 34



Following general procedure A, nBuLi (8.8 mL, 22 mmol, 2.5 M in hexanes), HNⁱPr₂ (3.1 mL, 22 mmol), 2-thiopheneacetic acid (1.42 g, 10 mmol) and 1-bromo-2-chloroethane (1.82 mL, 22 mmol)in anhydrous THF (50 mL) gave 4-chloro-2-(thiophen-2-yl)butanoic acid, which was used without further purification. Following general procedure B, 4-chloro-2-(thiophen-2-yl)butanoic acid (1.89 g, 10 mmol), 1,1'-carbonyldiimidazole (1.54 g, 9.5 mmol) and allylamine (898 µL, 12 mmol) in anhydrous THF (25 mL) gave, after column chromatography (eluent Petrol/EtOAc, 7:3), impure N-allyl-4-chloro-2-(thiophen-2-yl)butanamide, which was used without further purification. Following general procedure C, N-allyl-4-chloro-2-(thiophen-2-yl)butanamide (559 mg, 2.3 mmol) and NaH (60% in mineral oil) (460 mg, 11.5 mmol) in anhydrous THF (25 mL) gave, after column chromatography (eluent CH₂Cl₂/EtOAc, 4:1; R_F 0.29), 1-allyl-3-hydroxy-3-(thiophen-2-yl)pyrrolidin-2-one **34** as a colourless oil (209 mg, 0.93 mmol, 9%); v_{max} (ATR) 3327 (OH), 2949, 1674 (C=O), 1271; ¹H NMR (400 MHz, CDCl₃) δ_H: 2.43-2.59 (2H, m, C(4)H₂), 3.26-3.40 (2H, m, C(5)H₂), 3.89 (1H, ddd, J 15.2, 6.1, 1.3, NCH_AH_BCH=CH₂), 3.98 (1H, app. ddt, J 15.2, 6.1, 1.3, NCH_AH_BCH=CH₂), 4.25 (1H, s, OH), 5.15-5.24 (2H, m, NCH₂CH=CH₂), 5.72 (1H, app. ddt, J 16.6, 10.6, 6.1, NCH₂CH=CH₂), 6.93 (1H, dd, J 5.1, 3.6, ArC(4)*H*), 7.01 (1H, dd, *J* 3.6, 1.2, ArC(3)*H*), 7.26 (1H, dd, *J* 5.1, 1.2, ArC(5)*H*); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ_{C} : 36.1 (C(4)H₂), 42.9 (C(5)H₂), 45.9 (NCH₂), 76.3 (C(3)), 118.7 (NCH₂CH=*C*H₂), 124.1 (Ar*C*(4)H), 125.5 (Ar*C*(5)H), 126.8 (Ar*C*(3)H), 131.6 (NCH₂CH=CH₂), 145.7 (ArC(2)), 173.7 (C=O); m/z (NSI) 224 ([M+H]⁺, 100%) C₁₁H₁₄NO₂S⁺ ([M+H]⁺) requires 224.0740; found 224.0739 (-0.3 ppm).

4-Chloro-N,2-diphenylpropanamide S6



Following general procedure B, 4-chloro-2-phenylbutanoic acid⁹ (990 mg, 5 mmol), 1,1'carbonyldiimidazole (770 mg, 4.75 mmol) and aniline (449 μ L, 6 mmol) in anhydrous THF (50 mL) gave, after column chromatography (eluent CH₂Cl₂/EtOAc, 95:5; R_F 0.26), 4-chloro-*N*,2diphenylpropanamide **S7** as a pale yellow solid (683 mg, 2.60 mmol, 53%), mp 102-104 °C; v_{max} (ATR) 3258 (NH), 2953, 1661 (C=O), 1597 (C=C), 1543, 1443, 1329; ¹H NMR (400 MHz, CDCl₃) δ_{H} : 2.28 (1H, dddd, *J* 14.5, 7.8, 6.8, 4.9, C(3)*H*_AH_B), 2.70 (1H, dddd, *J* 14.5, 7.8, 6.8, 4.9, C(3)H_A*H*_B), 3.44 (1H, ddd, *J* 11.1, 8.0, 4.5, C(4)*H*_AH_B), 3.65 (1H, ddd, *J* 11.5, 6.7, 5.0, C(4)H_A*H*_B), 3.92 (1H, app. t, *J* 7.4, C(2)*H*), 7.07-7.14 (1H, m, NHArC(4)*H*), 7.26-7.50 (9H, m, ArCH); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ_{C} : 35.7 (*C*(3)H₂), 43.2 (*C*(4)H₂), 50.5 (*C*(2)H), 120.0 (NHArC(3,5)H), 124.5 (NHArC(4)H), 127.9 (C(2)ArC(4)H), 128.1 (NHArC(2,6)H), 129.0 (C(2)ArC(3,5)H), 129.3 (C(2)ArC(2,6)H), 137.7 (NHArC(1)), 138.3 (C(2)ArC(1)), 172.4 (*C*=O); *m*/*z* (NSI) 274 ([M+H]⁺, 100%) C₁₆H₁₇ONCl⁺ ([M+H]⁺) requires 274.0993; found 274.0995 (+0.7 ppm).

3-Hydroxy-1,3-diphenylpyrrolidin-2-one 35



Following general procedure C, 4-chloro-*N*,2-diphenylpropanamide **S7** (683 mg, 2.6 mmol) and NaH (60% in mineral oil) (530 mg, 13.23 mmol) in anhydrous THF (25 mL) gave, after column chromatography (eluent CH₂Cl₂/EtOAc, 9:1; R_F 0.32), 3-hydroxy-1,3-diphenylpyrrolidin-2-one **35** as a yellow solid (208 mg, 0.83 mmol, 32%), mp 81-83 °C; v_{max} (ATR) 3358 (OH), 3059, 1676 (C=O), 1591, 1489, 1292; ¹H NMR (400 MHz, CDCl₃) δ_{H} : 2.50-2.63 (2H, m, C(4)*H*₂), 3.73 (1H, ddd, *J* 9.7, 8.2, 7.0, C(5)*H*_AH_B), 3.86 (1H, ddd, *J* 9.8, 7.9, 3.4, C(5)H_AH_B), 4.26 (1H, s, O*H*), 7.21-7.27 (1H, m, ArC*H*), 7.29-7.39 (3H, m, ArC*H*), 7.40-7.49 (4H, m, ArC*H*), 7.70-7.77 (2H, m, ArC*H*); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ_{C} : 35.4 (*C*(4)H₂), 44.5 (*C*(5)H₂), 79.4

(C(3)), 119.9 (NArC(2,6)H), 125.2 (NArC(4)H), 125.3 (C(3)ArC(2,6)H), 128.1 (C(3)ArC(4)H), 128.7 (C(3)ArC(3,5)H), 129.0 (NArC(3,5)H), 139.0 (NArC(1)), 142.0 (C(3)ArC(1)), 174.7 (C=O); m/z (NSI) 254 ([M+H]⁺, 100%) C₁₆H₁₆O₂N⁺ ([M+H]⁺) requires 254.1176; found 254.1175 (-0.2 ppm).

Synthesis of Racemic Esters

Racemic esters given below were prepared as previously reported^{5,9} from this group.



1-Allyl-2-oxo-3-phenylindolin-3-yl isobutyrate S16



Following general Procedure D, 1-allyl-3-hydroxy-3-phenylindolin-2-one 20 (42 mg, 0.16 mmol), isobutyric anhydride (30 µL, 0.18 mmol), DMAP (2.0 mg, 0.016 mmol, 10 mol%) and *i*Pr₂NEt (28 μ L, 0.16 mmol) were reacted in CH₂Cl₂ (2.0 mL) to give the crude product, which was purified by column chromatography (eluent EtOAc:hexane 3:7, $R_F 0.55$) to give 1-allyl-2oxo-3-phenylindolin-3-yl isobutyrate S16 as a yellow solid (42 mg, 0.12 mmol, 78%). m.p. 61-63 °C; ¹H NMR (400 MHz, CDCl₃) δ_H: 1.20 (3H, d, J 7.0, CH(CH₃)_A(CH₃)_B), 1.24 (3H, d, J 7.0, CH(CH₃)_A(CH₃)_B), 2.71 (1H, sept, J 7.0, CH(CH₃)₂), 4.35 (2H, m, NCH₂), 5.23 (1H, dq, J 10.4, 1.5, CH=CH_{cis}H_{trans}), 5.32 (1H, dq, J 17.2, 1.5, CH=CH_{cis}H_{trans}), 5.86 (1H, ddt, J 17.2, 10.4, 5.2, CH=CH₂), 6.91 (1H, app dt, J 7.9, 0.8, C(7)H), 7.08 (1H, app td, J 7.4, 1.0, C(5)H), 7.20 (1H, ddd, J 7.4, 1.5, 0.6, C(6)H), 7.31-7.39 (6H, m, ArH); ¹³C NMR (126 MHz, d₆-DMSO) δ_C: 18.98 (CH(CH₃)_A(CH₃)_B), 19.04 (CH(CH₃)_A(CH₃)_B), 33.4 (CH(CH₃)₂), 42.3 (CH₂CH=CH₂), 80.8 (C(3)), 110.2 (ArC(7)H), 117.5 (CH₂CH=CH₂), 123.5 (ArC(5)H), 123.9 (ArC(6)H), 126.1 (C(3)ArC(2,6)H), 128.6 (C(3)ArC(1)), 129.2 (C(3)ArC(3,5)H), 129.3 (C(3)ArC(4)H), 130.4 (ArC(4)H), 132.0 (CH₂CH=CH₂), 137.3 (ArC(3a)), 143.5 (ArC(7a)), 173.4 (C(2)=O), 174.8 (C(=O)CH(CH₃)₂); IR (neat) v_{max} cm⁻¹ 2972, 1724 (C=O), 1614 (C=O), 1466, 1348, 1146; HRMS (NSI⁺) calculated for $C_{21}H_{21}NO_3Na^+$ ([M+Na]⁺) requires 358.1414; found 314.1415 (+0.4 ppm).

1-Methyl-2-oxo-3-phenylindolin-3-yl isobutyrate S17



Following general Procedure D, 3-hydroxy-1-methyl-3-phenylindolin-2-one **21** (38 mg, 0.16 mmol), isobutyric anhydride (30 μ L, 0.18 mmol), DMAP (2.0 mg, 0.016 mmol, 10 mol%) and

*i*Pr₂NEt (28 µL, 0.16 mmol) were reacted in CH₂Cl₂ (2.0 mL) to give the crude product, which was purified by flash column chromatography (eluent CH₂Cl₂:EtOAc 9:1, R_F 0.69) to give 1-methyl-2-oxo-3-phenylindolin-3-yl isobutyrate **S17** as a colourless powder (39 mg, 0.12 mmol, 78%). m.p. 123-125 °C; ¹H NMR (400 MHz, CDCl₃) $\delta_{\rm H}$: 1.20 (3H, d, *J* 7.0, CH(CH₃)_A(CH₃)_B), 1.24 (3H, *J* 7.0, CH(CH₃)_A(CH₃)_B), 2.71 (1H, sept, *J* 7.0, CH(CH₃)₂), 3.24 (3H, s, NCH₃), 6.93 (1H, app dt, *J* 7.8, 0.7, C(7)*H*), 7.10 (1H, app td, *J* 7.5, 1.0, C(5)*H*), 7.10 (1H, ddd, *J* 7.3, 1.4, 0.5, C(4)*H*), 7.31-7.43 (6H, m, Ar*H*); ¹³C NMR (126 MHz, *d*₆-DMSO) $\delta_{\rm C}$: 18.96 (CH(CH₃)_A(CH₃)_B), 19.03 (CH(CH₃)_A(CH₃)_B), 26.9 (CH₃), 33.4 (CH(CH₃)₂), 76.4 (C(3)), 109.6 (ArC(7)H), 123.4 (ArC(5)H), 123.8 (ArC(6)H), 126.2 (C(3)ArC(2,6)H), 128.6 (C(3)ArC(1)), 129.1 (C(3)ArC(2,6)H), 129.2 (C(3)ArC(3,5)H), 129.3 (C(3)ArC(4)H), 130.7 (ArC(4)H), 137.2 (ArC(3a)), 144.6 (ArC(7a)), 173.7 (C(2)=O), 174.8 (C(=O)CH(CH₃)₂); IR (neat) v_{max} cm⁻¹ 2978, 1721 (C=O), 1613 (C=C). 1470, 1342, 1148; HRMS (NSI⁺) calculated for C₁₉H₁₉NO₃Na⁺ ([M+Na]⁺) requires 332.1257; found 332.1259 (+0.6 ppm).

1-Allyl-2-oxo-3-(thiophen-2-yl)indolin-3-yl isobutyrate S20



Following general Procedure D, 1-allyl-3-hydroxy-3-(thiophen-2-yl)indolin-2-one **24** (41 mg, 0.15 mmol), isobutyric anhydride (28 μ L, 0.17 mmol), DMAP (1.8 mg, 0.015 mmol, 10 mol%) and *i*Pr₂NEt (26 μ L, 0.15 mmol) were reacted in CH₂Cl₂ (2 mL) to give the crude product, which was purified by Biotage® Isolera 4 chromatography (eluent: $0\%\rightarrow20\%$ EtOAc in hexanes) to give 1-allyl-2-oxo-3-(thiophen-2-yl)indolin-3-yl isobutyrate **S20** as a yellow solid (46 mg, 0.13 mmol, 90%). m.p. 80-82 °C; ¹H NMR (500 MHz, CDCl₃) δ_{H} : 1.17 (3H, d, *J* 7.0 Hz, CH(CH₃)_A(CH₃)_B), 1.21 (3H, d, *J* 7.0 Hz, CH(CH₃)_A(CH₃)_B), 2.67 (1H, sept, *J* 7.0 Hz, CH(CH₃)₂), 4.30-4.40 (2H, m, NCH₂), 5.19-5.24 (1H, m, CH=CH_{trans}H_{cis}), 5.27-5.34 (1H, m, CH=CH_{trans}H_{cis}), 5.85 (1H, app ddt, *J* 17.2, 10.3, 5.1, CH=CH₂), 6.87-6.92 (2H, m, ArC(7)H + C(3)ArC(3)H), 6.94 (1H, d, *J* 5.0, 3.6, C(3)ArC(4)H), 7.11 (1H, app t, *J* 7.8, ArC(5)H), 7.34-7.39 (3H, m, C(3)ArC(5)H + ArC(4,6)H); ¹³C NMR (126 MHz, CDCl₃) δ_{C} : 18.7 (CH(CH₃)_A(CH₃)_B), 18.8 (CH(CH₃)_A(CH₃)_B), 33.7 (CH(CH₃)₂), 42.8 (NCH₂), 78.7 (C(3)), 109.8

(Ar*C*(7)H), 117.8 (CH=*C*H₂), 123.0 (Ar*C*(5)H), 123.8 (Ar*C*(4)H), 126.6 (C(3)Ar*C*(4)H), 127.1 (C(3)Ar*C*(5)H), 127.8 (C(3)Ar*C*(3)H), 127.9 (Ar*C*(3a)), 130.4 (Ar*C*(6)H), 131.2 (*C*H=CH₂), 139.3 (C(3)Ar*C*(2)), 143.4 (Ar*C*(7a)), 172.7 (*C*=O), 175.2 (*C*O₂R); IR (neat) v_{max} cm⁻¹ 3103, 3076, 2972, 2930, 1724 (C=O), 1695, 1614, 1487, 1464, 1371, 1350, 1184, 1143, 1098; HRMS (NSI⁺) calculated for C₁₉H₁₉NO₃SNa⁺ ([M+Na]⁺) requires 364.0983; found 364.0981 (-0.5 ppm).

1-Benzyl-7-chloro-2-oxo-3-phenylindolin-3-yl isobutyrate S25



Following general Procedure D, 1-benzyl-6-chloro-3-hydroxy-3-phenylindolin-2-one 29 (70 mg, 0.2 mmol), isobutyric anhydride (37 µL, 0.22 mmol), DMAP (2.4 mg, 0.02 mmol, 10 mol%) and *i*Pr₂NEt (35 μ L, 0.2 mmol) were reacted in CH₂Cl₂ (6 mL) to give the crude product, which was purified by Biotage® Isolera 4 chromatography (eluent: 0%→30% EtOAc in hexanes) to give 1benzyl-6-chloro-2-oxo-3-phenylindolin-3-yl isobutyrate S25 as a colourless solid (71 mg, 0.17 mmol, 85%). m.p. 120-122 °C; ¹H NMR (500 MHz, CDCl₃) δ_H: 1.23 (3H, d, J 7.0, CH(CH₃)_A(CH₃)_B), 1.27 (3H, d, J 7.0, CH(CH₃)_A(CH₃)_B), 2.74 (1H, sept, J 7.0, CH(CH₃)₂), 4.83 (1H, d, J 16.0, CH_AH_BPh), 4.98 (1H, d, J 16.0, CH_AH_BPh), 6.72 (1H, d, J 1.8, ArC(7)H), 7.04 (1H, dd, J 7.9, 1.8, ArC(5)H), 7.12 (1H, d, J 7.9, ArC(4)H), 7.24-7.39 (10H, m, CH₂ArH + C(3)ArH); ¹³C NMR (126 MHz, CDCl₃) δ_{C} : 18.8 (CH(CH₃)_A(CH₃)_B), 18.9 (CH(CH₃)_A(CH₃)_B), 33.9 (CH(CH₃)₂), 44.5 (CH₂Ph), 80.5 (C(3)), 110.5 (ArC(7)H), 123.2 (ArC(5)H), 124.8 (ArC(4)H), 126.3 (C(3)ArC(2,6)H), 127.0 (ArC(3a)), 127.3 (CH₂ArC(2,6)H), 127.9 (CH₂ArC(4)H), 128.9 (C(3)ArC(3,5)H), 129.0 (CH₂ArC(3,5)H), 129.2 (C(3)ArC(4)H), 135.2 (CH₂ArC(1)), 135.9 (ArC(6)), 136.4 (C(3)ArC(1)), 145.0 (ArC(7a)), 174.1 (C(2)=O), 175.3 (CO₂R). IR (neat) v_{max} cm⁻¹ 3030, 2974, 2934, 2876, 1736 (C=O), 1612, 1489, 1449, 1369, 1354, 1339, 1256, 1184, 1146, 1115, 1070; HRMS (NSI⁺) calculated for $C_{25}H_{26}N_2O_3{}^{35}Cl$ $([M+NH_4]^+)$ requires 437.1626; found 437.1626 (-0.1 ppm).

1-Allyl-3-(4-chlorophenyl)-2-oxopyrrolidin-3-yl acetate S27



Following general procedure D, 1-allyl-3-(4-chlorophenyl)-3-hydroxypyrrolidin-2-one **31** (40 mg, 0.16 mmol), acetic anhydride (20 µL, 0.21 mmol), DMAP (2 mg, 0.016 mmol, 10 mol %) and *i*Pr₂NEt (70 µL, 0.38 mmol) in CH₂Cl₂ (3 mL) gave, after column chromatography (eluent CH₂Cl₂/EtOAc, 9:1; R_F 0.29), 1-allyl-3-(4-chlorophenyl)-2-oxopyrrolidin-3-yl acetate **S27** as a colourless oil (44 mg, 0.15 mmol, 94%); v_{max} (ATR) 2974, 1742 (C=O), 1686 (C=O), 1371, 1223; ¹H NMR (400 MHz, CDCl₃) δ_{H} : 2.16 (3H, s, CH₃), 2.69-2.81 (2H, m, C(4)H₂), 3.38 (1H, app. dt, *J* 9.8, 7.6, C(5)H_AH_B), 3.53 (1H, ddd, *J* 9.8, 7.3, 5.1, C(5)H_AH_B), 3.91 (1H, app. ddt, *J* 15.2, 6.2, 1.3, NCH_AH_BCH=CH₂), 3.99 (1H, app. ddt, *J* 15.2, 5.9, 1.4, NCH_AH_BCH=CH₂), 5.20-5.29 (2H, m, NCH₂CH=CH₂), 5.74 (1H, app. ddt, *J* 17.1, 10.2, 6.1, NCH₂CH=CH₂), 7.32-7.37 (2H, m, ArC(2,6)H), 7.41-7.47 (2H, m, ArC(3,5)H); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ_{C} : 21.5 (CH₃), 31.5 (C(4)H₂), 43.1 (C(5)H₂), 45.9 (NCH₂), 82.8 (C(3)), 118.7 (NCH₂CH=CH₂), 126.7 (ArC(3,5)H), 128.8 (ArC(2,6)H), 131.6 (NCH₂CH=CH₂), 134.5 (ArC(4)), 137.5 (ArC(1)), 169.9 (C=O(CH₃)), 170.1 (C(2)=O); *m/z* (NSI) 294 ([M+H]⁺, 100%) C₁₅H₁₇NO₃Cl⁺ ([M+H]⁺) requires 294.0891; found 294.0896 (+1.5 ppm).

1-Allyl-2-oxo-3-(thiophen-3-yl)pyrrolidin-3-yl acetate S28



Following general procedure D, 1-allyl-3-hydroxy-3-(thiophen-3-yl)pyrrolidin-2-one **32** (36 mg, 0.16 mmol), acetic anhydride (21 μ L, 0.21 mmol), DMAP (2 mg, 0.016 mmol, 10 mol %) and *i*Pr₂NEt (70 μ L, 0.38 mmol) in CH₂Cl₂ (3 mL) gave, after column chromatography (eluent CH₂Cl₂/EtOAc, 9:1; R_F 0.30), 1-allyl-2-oxo-3-(thiophen-3-yl)pyrrolidin-3-yl acetate **S28** as a

colourless oil (40 mg, 0.15 mmol, 94%); v_{max} (ATR) 2978, 1730 (C=O), 1697 (C=O), 1423, 1240; ¹H NMR (400 MHz, CDCl₃) δ_{H} : 2.13 (3H, s, CH₃), 2.75-2.82 (2H, m, C(4)H₂), 3.34 (1H, app. q, *J* 8.6, C(5)*H*_AH_B), 3.49 (1H, app. dt, *J* 11.3, 6.1, C(5)H_AH_B), 3.90 (1H, dd, *J* 15.2, 6.0, NCH_AH_BCH=CH₂), 3.98 (1H, dd, *J* 15.4, 5.8, NCH_AH_BCH=CH₂), 5.17-5.26 (2H, m, NCH₂CH=CH₂), 5.73 (1H, app. ddt, *J* 16.2, 10.9, 5.9, NCH₂CH=CH₂), 7.23 (1H, d, *J* 5.1, ArC(4)*H*), 7.31-7.35 (1H, m, ArC(5)*H*), 7.42 (1H, br s, ArC(2)*H*); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ_{C} : 21.5 (CH₃), 31.4 (C(4)H₂), 42.9 (C(5)H₂), 45.9 (NCH₂), 81.3 (C(3)), 118.4 (NCH₂CH=CH₂), 122.5 (ArC(2)H), 125.7 (ArC(4)H), 126.6 (ArC(5)H), 131.7 (NCH₂CH=CH₂), 139.9 (ArC(3)), 170.0 (*C*=O(CH₃)), 170.1 (*C*(2)=O); *m*/*z* (NSI) 266 ([M+H]⁺, 100%) C₁₃H₁₆NO₃S⁺ ([M+H]⁺) requires 266.0844; found 266.0847 (+1.3 ppm).

1-Allyl-2-oxo-3-(thiophen-2-yl)pyrrolidin-3-yl acetate S30



Following general procedure D, 1-allyl-3-hydroxy-3-(thiophen-2-yl)pyrrolidin-2-one **34** (36 mg, 0.16 mmol), acetic anhydride (20 µL, 0.21 mmol), DMAP (2 mg, 0.016 mmol, 10 mol %) and iPr_2NEt (70 µL, 0.38 mmol) in CH₂Cl₂ (3 mL) gave, after column chromatography (eluent CH₂Cl₂/EtOAc, 9:1; R_F 0.46), 1-allyl-2-oxo-3-(thiophen-2-yl)pyrrolidin-3-yl acetate **S30** as a colourless oil (34 mg, 0.13 mmol, 79%); v_{max} (ATR) 2982, 1740 (C=O), 1699 (C=O), 1435, 1223; ¹H NMR (400 MHz, CDCl₃) δ_{H} : 2.13 (3H, s, CH₃), 2.79-2.93 (2H, m, C(4)H₂), 3.36 (1H, app. td, *J* 9.7, 7.7, C(5)H_AH_B), 3.49 (1H, ddd, *J* 9.6, 9.0, 2.9, C(5)H_AH_B), 3.88 (1H, app. ddt, *J* 15.3, 6.1, 1.3, NCH_AH_BCH=CH₂), 3.98 (1H, app. ddt, *J* 15.3, 5.8, 1.4, NCH_AH_BCH=CH₂), 5.16-5.25 (2H, m, NCH₂CH=CH₂), 5.66-77 (1H, m, NCH₂CH=CH₂), 6.99 (1H, dd, *J* 5.1, 3.7, ArC(4)H), 7.20 (1H, dd, *J* 3.7, 1.2, ArC(3)H), 7.34 (1H, dd, *J* 5.1, 1.2, ArC(5)H); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ_C : 21.5 (CH₃), 32.2 (C(4)H₂), 42.8 (C(5)H₂), 45.9 (NCH₂), 81.3 (C(3)), 118.4 (NCH₂CH=CH₂), 125.4 (ArC(3)H), 126.69 (ArC(5)H), 126.74 (ArC(4)H), 131.6 (NCH₂CH=CH₂), 141.0 (ArC(2)), 169.5 (C(2)=O), 169.9 (C=O(CH₃)); m/z (NSI) 266 ([M+H]⁺, 100%) C₁₃H₁₆NO₃S⁺ ([M+H]⁺) requires 266.0845; found 266.0847 (+0.6 ppm).

2-Oxo-1,3-diphenylpyrrolidin-3-yl acetate S31



Following general procedure D, 3-hydroxy-1,3-diphenylpyrrolidin-2-one **35** (41 mg, 0.16 mmol), acetic anhydride (20 µL, 0.21 mmol), DMAP (2.0 mg, 0.016 mmol, 10 mol %) and *i*Pr₂NEt (70 µL, 0.38 mmol) in CH₂Cl₂ (3 mL) gave, after column chromatography (eluent CH₂Cl₂/EtOAc, 9:1;R_F 0.62), 2-oxo-1,3-diphenylpyrrolidin-3-yl acetate **S31** as a clear oil (41 mg, 0.14 mmol, 86%); v_{max} (ATR) 3055, 1740 (C=O), 1701 (C=O), 1495, 1371, 1223; ¹H NMR (400 MHz, CDCl₃) δ_{H} : 2.20 (3H, s, *CH*₃), 2.87-3.01 (2H, m, C(4)*H*₂), 3.88 (1H, app. dt, *J* 9.6, 8.0, C(5)*H*_AH_B), 4.01 (1H, app. td, *J* 9.2, 3.0, C(5)H_AH_B), 7.15-7.21 (1H, m, ArC*H*), 7.31-7.42 (5H, m, ArC*H*), 7.53-7.59 (2H, m, ArC*H*), 7.65-7.71 (2H, m, ArC*H*); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ_{C} : 21.6 (*C*H₃), 30.9 (*C*(4)H₂), 44.7 (*C*(5)H₂), 83.7 (*C*(3)), 120.0 (NAr*C*(2,6)H), 125.2 (NAr*C*(4)H), 125.4 (C(3)Ar*C*(2,6)H), 128.7 (C(3)Ar*C*(4)H), 128.8 (C(3)Ar*C*(3,5)H), 128.9 (NAr*C*(3,5)H), 138.2 (NAr*C*(1)), 139.1 (C(3)Ar*C*(1)), 169.9 (*C*(=O)CH₃), 170.0 (*C*(2)=O); *m/z* (NSI) 296 ([M+H]⁺, 100%) C₁₈H₁₈NO₃⁺ ([M+H]⁺) requires 296.1281; found 296.1283 (+0.6 ppm).

Kinetic Resolution of Secondary Alcohols

Optimisation

Initial solvent screen using TentaGel and Wang-supported HyperBTM derivatives 3 and 4:



			TentaGel-HyperBTM 3		Wang-HyperBTM 4	
Solvent	Temp	(ⁱ PrCO) ₂ O (equiv.)	c	S	с	S
CHCl ₃	r.t.	0.55	35	46	37	50
CHCl ₃	0 °C	0.55	39	70	30	80
THF	r.t.	0.55	49	28	46	45
CH_2Cl_2	r.t.	0.55	49	38	42	33
CH ₃ CN	r.t.	0.55	50	21	46	19
EtOAc	r.t.	0.55	46	45	44	43
Et ₂ O	r.t.	0.55	47	80	47	50
Et ₂ O	0 °C	0.55	46	45	NP ^a	NP ^a
Toluene	r.t.	0.55	52	60	32	71
Toluene	0 °C	0.55	NP ^a	NP ^a	43	80

Reaction conditions: 7 (0.2 mmol), **3** or **4** (3 mol%), (*i*-PrCO)₂O (0.55 equiv.), *i*-Pr₂NEt (0.6 equiv.), solvent (0.2 M), 16 h. Conversion (c)and er determined by chiral HPLC analysis. Selectivity factors (*s*) calculated using er of **7** and reaction conversion (see ref. 1a), and rounded according to estimated associated errors (see ref. 1b). ^a NP = not performed

Industrially-preferable solvent screen using polymer supported-catalysts 2-4 and 6



Solvent	Catalyst	(ⁱ PrCO) ₂ O	ee (ester)				
(0.2 M)	(mol%)	(equiv.)		ee (alcohol)	c	S	
DMC	2		87.65	88.97	50	45	
DMC	3	0.55	92.11	77.19	46	60	
DMC	4	0.55	91.29	75.05	45	50	
DMC	6	0.55	92.95	34.37	27	38	
^t BuOAc	2	0.55	77.53	44.45	36	12	
^t BuOAc	3	0.55	89.11	81.27	48	46	
^t BuOAc	4	0.55	78.73	6.19	7	9	
^t PrOAc	2	0.55	86.41	94.08	52	49	
^t PrOAc	3	0.55	88.62	77.68	47	39	
^t PrOAc	4	0.55	79.42	62.7	44	16	
^t AmOH	2	0.55	86.47	17.51	17	16	
^t AmOH	3	0.55	88.33	84.29	49	43	
^t AmOH	4	0.55	74.26	2.66	3	7	
^t BuOMe	2	0.55	89.81	30.28	25	25	
^t BuOMe	3	0.55	88.21	77.35	47	37	
^t BuOMe	4	0.55	92.3	39.28	30	37	

Reaction conditions: 7 (0.2 mmol),**2**, **3**, **4** or **6** (5 mol%), (*i*-PrCO)₂O(0.55 equiv.), *i*-Pr₂NEt (0.6 equiv.), solvent (0.2 m), 0-r.t., 16 h. Conversion (c) and er determined by chiral HPLC analysis. Selectivity factors (*s*) calculated using er of **7** and reaction conversion (see ref. 1a), and rounded according to estimated associated errors (see ref. 1b). DMC = dimethyl carbonate, ^{*t*}BuOAc = *tert*-butyl acetate, ^{*i*}PrOAc = isopropyl acetate, ^{*t*}AmOH = *tert*-amyl alcohol, ^{*t*}BuOMe = *tert*-butyl methyl ether,



Figure 1 Screening of solvents using Merifield-HyperBTM 2



Figure 2 Screening of solvents using TentaGel-HyperBTM 3



Figure 3 Screening of solvents using Wang-HyperBTM 4



Figure 4 Screening of solvents using Merifield-BTM 6

Kinetic resolution of 1-(naphthalen-2-yl)ethan-1-ol 12 (Table 1, entries 13-16) Table 1: Reaction Optimisation



With Wang-HyperBTM:

Following General Procedure E, 1-(naphthalen-2-yl)ethan-1-ol **7** (34 mg, 0.2 mmol), Wang-HyperBTM **4** (15 mg, 5 mol%), *i*-Pr₂NEt (21 μ L, 0.12 mmol, 0.6 equiv.) and (*i*-PrCO)₂O (18 μ L, 0.11 mmol, 0.55 equiv.) in DMC (1 mL) for 16 h at room temperature gave the crude product, which was purified by Biotage® IsoleraTM 4 to give (*S*)-1-(naphthalen-2-yl)ethyl isobutyrate **8** (20 mg, 41%) and (*R*)-1-(naphthalen-2-yl)ethan-1-ol **7** (17 mg, 49%).

Chiral HPLC analysis:

(S)-1-(Naphthalen-2-yl)ethyl isobutyrate 8: $[\alpha]_D^{20} = -88$ (c 1.0, CHCl₃) Chiral HPLC analysis, Chiralcel OJ-H (98:2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (*R*): 9.8 min, t_R (*S*): 12.2 min, 4.364:95.636 er. *s* = 50.

(*R*)-1-(Naphthalen-2-yl)ethan-1-ol 7: mp 67-70 °C {Lit.¹³ 69-72 °C}. $[\alpha]_D^{20} = +35.5$ (*c* 1.0, CHCl₃) {Lit.⁸ $[\alpha]_D^{20} = +46.5$ (98% ee, *c* 1.2, CHCl₃); Chiral HPLC analysis, Chiralcel OJ-H

(90:10 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (S): 12.2 min, t_R (R): 19.5 min, 12.474:87.526 er.

With TentaGel-HyperBTM 3: Yield of (*R*)-1-(naphthalen-2-yl)ethan-1-ol 7 = 52% (18 mg) and (*S*)-1-(naphthalen-2-yl)ethyl isobutyrate 8 = 39% (19 mg)

Chiral HPLC analysis:

(S)-1-(Naphthalen-2-yl)ethyl isobutyrate 8: Chiralcel OJ-H (98:2 hexane:IPA, flow rate 1.0 mL min⁻¹, 254 nm, 30 °C) t_R (*R*): 9.4 min, t_R (S): 11.6 min, 3.946:96.054 er. s = 57.

(*R*)-1-(Naphthalen-2-yl)ethan-1-ol 7: Chiralcel OJ-H (90:10 hexane:IPA, flow rate 1.0 mL min⁻¹, 254 nm, 30 °C) t_R (*S*): 15.0 min, t_R (*R*): 19.1 min, 11.403:88.597 er.

With Merrifield-HyperBTM 2: Yield of (*R*)-1-(naphthalen-2-yl)ethan-1-ol 7 = 41% (14 mg) and (*S*)-1-(naphthalen-2-yl)ethyl isobutyrate 8 = 45% (22 mg)

Chiral HPLC analysis:

(*R*)-1-(Naphthalen-2-yl)ethan-1-ol 7: Chiralcel OJ-H (90:10 hexane:IPA, flow rate 1.0 mL min⁻¹, 254 nm, 30 °C) t_R (*S*): 15.0 min, t_R (*R*): 19.1 min, 5.5:94.5 er.

(*S*)-1-(Naphthalen-2-yl)ethyl isobutyrate 8: Chiralcel OJ-H (98:2 hexane:IPA, flow rate 1.0 mL min⁻¹, 254 nm, 30 °C) t_R (*R*): 9.4 min, t_R (*S*): 11.6 min, 6.2:93.8 er. s = 45.

With Merrifield-BTM 6: Yield of (*R*)-1-(naphthalen-2-yl)ethan-1-ol 7 = 70% (24 mg) and (*S*)-1-(naphthalen-2-yl)ethyl isobutyrate 8 = 25% (12 mg)

Chiral HPLC analysis:

(*S*)-1-(Naphthalen-2-yl)ethyl isobutyrate 8: Chiralcel OJ-H (98:2 hexane:IPA, flow rate 1.0 mL min⁻¹, 254 nm, 30 °C) t_R (*R*): 9.4 min, t_R (*S*): 11.6 min, 96.5:3.5 er. s = 38.

(*R*)-1-(Naphthalen-2-yl)ethan-1-ol 7: Chiralcel OJ-H (90:10 hexane:IPA, flow rate 1.0 mL min⁻¹, 254 nm, 30 °C) t_R (*S*): 15.0 min, t_R (*R*): 19.1 min, 67.2:32.8 er.

Chiral HPLC data for Table 2: Recycling experiments of supported catalysts 2-4 and 6 for the sequential KR of five secondary alcohols 9–13

Cycle 1: 2-Azido-1-(naphthalen-3-yl)ethan-1-ol 9



With Merrifield-HyperBTM 2:

Following General Procedure E, 2-azido-1-(naphthalen-3-yl)ethan-1-ol **9** (85 mg, 0.4 mmol), Merrifield-HyperBTM **2** (23 mg, 5 mol%), *i*-Pr₂NEt (42 μ L, 0.24 mmol, 0.6 equiv.) and (*i*-PrCO)₂O (36 μ L, 0.22 mmol, 0.55 equiv.) in DMC (2 mL) for 24 h at r.t. gave the crude product, which was purified by Biotage® IsoleraTM 4 to give (*S*)-2-azido-1-(naphthalen-3-yl)ethan-1-ol **9** (37 mg, 43%) and (*R*)-2-azido-1-(naphthalen-3-yl)ethyl isobutyrate **S7** (50 mg, 44%). The catalyst was recovered by simple filtration and washed with MeOH (250 ml), CH₂Cl₂ (250 ml), CHCl₃ (250 ml) and THF (250 ml) respectively. Finally, the washed catalyst was dried under vacuum for 3 hrs, weighed (23 mg) and used for next cycle.

(*R*)-2-Azido-1-(naphthalen-2-yl)ethyl isobutyrate S7: $[\alpha]_D^{20} = -103$ (*c* 1.0, CHCl₃) {Lit.⁵ $[\alpha]_D^{20}$ = -78 (75% ee, *c* 1.0, CHCl₃); Chiral HPLC analysis, Chiralcel AD-H (99.8:0.2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (*S*): 14.3 min, t_R (*R*): 25.5 min, 10.236:89.764 er.

(S)-2-Azido-1-(naphthalen-3-yl)ethan-1-ol 9: $[\alpha]_D^{20} = +94.7$ (c 1.0, CHCl₃); {Lit.¹⁴ $[\alpha]_D^{20} = +125.2$ (99 %ee, c 1.3, CHCl₃), Chiral HPLC analysis, Chiralcel OD-H (95:5 hexane:IPA, flow rate 1 mLmin⁻¹, 220 nm, 30 °C) t_R (*R*): 31.1 min, t_R (*S*): 35.4 min, 6.124:93.876 er. *s* = 25.

With TentaGel-HyperBTM 3 (used 47 mg): Yield of (S)-2-Azido-1-(naphthalen-3-yl)ethan-1-ol 9 = 52% (44 mg) and (R)-2-Azido-1-(naphthalen-2-yl)ethyl isobutyrate S7 = 41% (46 mg)

(*R*)-2-Azido-1-(naphthalen-2-yl)ethyl isobutyrate S7: $[\alpha]_D^{20} = -107$ (*c* 1.0, CHCl₃) {Lit.⁵ $[\alpha]_D^{20}$ = -78 (75% ee, *c* 1.0, CHCl₃); Chiralcel AD-H (99.8:0.2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (*S*): 13.4 min, t_R (*R*): 21.4 min, 8.747:91.253 er.

(S)-2-Azido-1-(naphthalen-3-yl)ethan-1-ol 9: $[\alpha]_D^{20} = +75$ (c 1.0, CHCl₃); {Lit.¹⁴ $[\alpha]_D^{20} = +125.2$ (99 %ee, c 1.3, CHCl₃), Chiral HPLC analysis, Chiralcel OD-H (95:5 hexane:IPA, flow rate 1 mLmin⁻¹, 220 nm, 30 °C) t_R (*R*): 30.2 min, t_R (*S*): 34.2 min, 14.640:85.360 er. *s* = 22.

The quantity of recovered catalyst after proper washing and vacuum drying was 47 mg.

With Wang-HyperBTM 4 (used 31 mg): Yield of (S)-2-Azido-1-(naphthalen-3-yl)ethan-1-ol 9 = 48% (41 mg) and (R)-2-Azido-1-(naphthalen-2-yl)ethyl isobutyrate S7 = 46% (52 mg).

(*R*)-2-Azido-1-(naphthalen-2-yl)ethyl isobutyrate S7: mp 35-37 °C; $[\alpha]_D^{20} = -107$ (*c* 1.0, CHCl₃) {Lit.⁵ $[\alpha]_D^{20} = -78$ (75% ee, *c* 1.0, CHCl₃); Chiral HPLC analysis, Chiralcel AD-H (99.8:0.2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (*S*): 13.4 min, t_R (*R*): 21.6 min, 8.220:91.780 er.

(*S*)-2-Azido-1-(naphthalen-3-yl)ethan-1-ol 9: mp 84-86 °C {Lit.¹⁵ 82-83 °C}. $[\alpha]_D^{20} = +85$ (*c* 1.0, CHCl₃) {Lit.¹⁴ $[\alpha]_D^{20} = +125.2$ (99 %ee, *c* 1.3, CHCl₃); Chiral HPLC analysis, Chiralcel OD-H (95:5 hexane:IPA, flow rate 1 mLmin⁻¹, 220 nm, 30 °C) t_R (*R*): 30.3 min, t_R (*S*): 34.0 min, 8.421:91.579 er. *s* = 29

The quantity of recovered catalyst after proper washing and vacuum drying was 31 mg.

With Merrifield-BTM 6 (used 23 mg): Yield of (*R*)-2-Azido-1-(naphthalen-3-yl)ethan-1-ol 9=82% (70 mg) and (*S*)-2-Azido-1-(naphthalen-2-yl)ethyl isobutyrate S7=13% (15 mg)
(S)-2-Azido-1-(naphthalen-2-yl)ethyl isobutyrate S7: $[\alpha]_D^{20} = +48$ (c 1.0, CHCl₃) {Lit.⁵ (ent, 75% ee) $[\alpha]_D^{20} = -78$ (c 1.0, CHCl₃); Chiral HPLC analysis, Chiralcel AD-H (99.8:0.2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (S): 15.1 min, t_R (R): 25.6 min, 69.814:30.186 er

(*R*)-2-Azido-1-(naphthalen-3-yl)ethan-1-ol 9: $[\alpha]_D^{20} = -7.25$ (*c* 1.0, CHCl₃) {Lit.¹⁴ (*ent*, 99% ee) $[\alpha]_D^{20} = +125.2$ (*c* 1.3, CHCl₃); Chiralcel OD-H (95:5 hexane:IPA, flow rate 1 mLmin⁻¹, 220 nm, 30 °C) t_R (*R*): 30.6 min, t_R (*S*): 35.1 min, 53.792:46.208 er. *s* = 2.

The quantity of recovered catalyst after proper washing and vacuum drying was 21 mg.

Cycle 2: Kinetic resolution of 4-phenylbut-3-yn-2-ol 10



Following General Procedure E, 4-phenylbut-3-yn-2-ol **10** (58 mg, 0.4 mmol), Merrifield-HyperBTM **2** (23 mg, 5 mol%), *i*-Pr₂NEt (42 μ L, 0.24 mmol, 0.6 equiv.) and (*i*-PrCO)₂O (36 μ L, 0.22 mmol, 0.55 equiv.) in DMC (2 mL) for 24 h at r.t. gave the crude product, which was purified by Biotage® IsoleraTM 4 to give (*R*)-4-phenylbut-3-yn-2-ol **10** (26 mg, 44%) and (*S*)-4-phenylbut-3-yn-2-yl isobutyrate **S8** (39 mg, 45%).

Following General Procedure G, (*S*)-4-phenylbut-3-yn-2-yl isobutyrate S8 was hydrolysed to corresponding (*S*)-4-phenylbut-3-yn-2-ol for HPLC analysis. $[\alpha]_D^{20} = -90$ (*c* 0.8, CHCl₃) {Lit.⁵ $[\alpha]_D^{20} = -23$ (77% ee, *c* 1.0, CHCl₃); Chiral HPLC analysis, Chiralcel OD-H (95:5 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (*R*): 11.2 min, t_R (*S*): 28.7 min, 19.002:80.998 er. *s* =10.

(*R*)-4-Phenylbut-3-yn-2-ol 10: $[\alpha]_D^{20} = +23$ (*c* 0.8, CHCl₃){Lit.¹⁶ $[\alpha]_D^{20} = +32.4$ (96% ee, *c* 0.966, CHCl₃); Chiral HPLC analysis, Chiralcel OD-H (95:5 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (*R*): 11.1 min, t_R (*S*): 27.7 min, 88.823:11.177 er.

The quantity of recovered catalyst after proper washing and vacuum drying was 21 mg.

With TentaGel-HyperBTM 3 (catalyst used is 47 mg): Yield of (*R*)-4-Phenylbut-3-yn-2-ol 10=46% (27 mg) and (*S*)-4-phenylbut-3-yn-2-yl isobutyrate S8 = 44% (38 mg).

Following General Procedure G, (*S*)-4-phenylbut-3-yn-2-yl isobutyrate S8 was hydrolysed to corresponding (*S*)-4-phenylbut-3-yn-2-ol for HPLC analysis. $[\alpha]_D^{20} = -61.5$ (*c* 0.8, CHCl₃) {Lit.⁵ $[\alpha]_D^{20} = -23$ (77% ee, *c* 1.0, CHCl₃); Chiral HPLC analysis, Chiralcel OD-H (95:5 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (*R*): 11.1 min, t_R (*S*): 27.6 min, 21.073:78.927 er.

(*R*)-4-Phenylbut-3-yn-2-ol 10: $[\alpha]_D^{20} = +21.4$ (*c* 0.8, CHCl₃)){Lit.¹⁶ $[\alpha]_D^{20} = +32.4$ (96% ee, *c* 0.966, CHCl₃);; Chiral HPLC analysis, Chiralcel OD-H (95:5 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (*R*): 11.1 min, t_R (*S*): 27.7 min, 81.202:18.798 er. *s* =7.

The quantity of recovered catalyst after proper washing and vacuum drying was 43 mg.

With Wang-HyperBTM 4 (catalyst used is 31 mg): Yield of (*R*)-4-Phenylbut-3-yn-2-ol 10 = 44% (26 mg) and (*S*)-4-phenylbut-3-yn-2-yl isobutyrate S8 = 42% (36 mg).

Following General Procedure G, (*S*)-4-phenylbut-3-yn-2-yl isobutyrate S8 was hydrolysed to corresponding (*S*)-4-phenylbut-3-yn-2-ol for HPLC analysis. $[\alpha]_D^{20} = -105$ (*c* 1.0, CHCl₃); {Lit.⁵ $[\alpha]_D^{20} = -23$ (77% ee, *c* 1.0, CHCl₃); Chiral HPLC analysis, Chiralcel OD-H (95:5 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (*R*): 11.2 min, t_R (*S*): 27.8 min, 13.368:86.632 er.

(*R*)-4-Phenylbut-3-yn-2-ol 10: $[\alpha]_D^{20} = +24$ (*c* 0.8, CHCl₃);){Lit.¹⁶ $[\alpha]_D^{20} = +32.4$ (96% ee, *c* 0.966, CHCl₃); Chiral HPLC analysis, Chiralcel OD-H (95:5 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (*R*): 11.1 min, t_R (*S*): 27.7 min, 88.132:11.868 er. *s* =15.

The quantity of recovered catalyst after proper washing and vacuum drying was 29 mg

With Merrifield-BTM 6: No reaction.

Cycle 3: trans-2-Phenylcyclopentan-1-ol 11



Following General Procedure E, *trans*-2-phenylcyclopentan-1-ol **11** (49 mg, 0.3 mmol), Merrifield-HyperBTM **2** (17 mg, 5 mol%), *i*-Pr₂NEt (31 µL, 0.18 mmol, 0.6 equiv.) and (*i*-PrCO)₂O (27 µL, 0.165 mmol, 0.55 equiv.) in DMC (1.5 mL) for 24 h at r.t gave the crude product, which was purified by Biotage® IsoleraTM 4 to give (1*R*,2*S*)-2-phenylcyclopentan-1-ol **11** (21 mg, 43%) and (1*S*,2*R*)-2-phenylcyclopentyl isobutyrate **S9** (26 mg, 37%).

(1*S*,2*R*)-2-Phenylcyclopentyl isobutyrate S9: $[\alpha]_D^{20} = +33.8 (c \ 1.0, \text{CHCl}_3) \{\text{Lit.}^5 [\alpha]_D^{20} = +67.2 (90\% \text{ ee, } c \ 1.0, \text{CHCl}_3); \text{ Following General Procedure G, } (1S,2R)-2-\text{phenylcyclopentyl isobutyrate S9 was hydrolysed to corresponding alcohol } (1S,2R)-2-\text{phenylcyclopentan-1-ol 11} for HPLC analysis. Chiral HPLC analysis, Chiralcel OD-H (98:2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (1S,2R): 14.4 min, t_R (1R,2S): 16.1 min, 90.046:9.954 er.$

(1R,2S)-2-Phenylcyclopentan-1-ol 11: $[\alpha]_D^{20} = -49.3$ (*c* 1.0, CHCl₃) {Lit.⁵ $[\alpha]_D^{20} = -60.7$ (95% ee, *c* 1.0, CHCl₃); Chiral HPLC analysis Chiralcel OD-H (98:2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (1*S*,2*R*): 14.4 min, t_R (1*R*,2*S*): 16.0 min, 4.997:95.003 er. *s* = 27.

The quantity of recovered catalyst after proper washing and vacuum drying was 17 mg.

With TentaGel-HyperBTM 3 (catalyst used is 35 mg): Yield of (1R,2S)-2-Phenylcyclopentan-1-ol 11 = 45% (22 mg) and (1S,2R)-2-Phenylcyclopentyl isobutyrate S9 = 46% (32 mg).

(1*S*,2*R*)-2-Phenylcyclopentyl isobutyrate S9: $[\alpha]_D^{20} = +35.6 (c \ 1.0, \text{CHCl}_3) \{\text{Lit.}^5 [\alpha]_D^{20} = +67.2 (90\% \text{ ee, } c \ 1.0, \text{CHCl}_3); \text{ Following General Procedure G, } (1S,2R)-2-\text{phenylcyclopentyl} \}$

isobutyrate **S9** was hydrolysed to corresponding alcohol (1*S*,2*R*)-2-phenylcyclopentan-1-ol **11** for HPLC analysis. Chiral HPLC analysis, Chiralcel OD-H (98:2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (1*S*,2*R*): 14.4 min, t_R (1*R*,2*S*): 16.1 min, 92.331:7.669 er. *s* = 29.

(1*R*,2*S*)-2-Phenylcyclopentan-1-ol 11: $[\alpha]_D^{20} = -45$ (*c* 1.0, CHCl₃) {Lit.⁵ $[\alpha]_D^{20} = -60.7$ (95% ee, *c* 1.0, CHCl₃); Chiral HPLC analysis Chiralcel OD-H (98:2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (1*S*,2*R*): 14.4 min, t_R (1*R*,2*S*): 16.1 min, 10.359:89.641 er.

The quantity of recovered catalyst after proper washing and vacuum drying was 30 mg.

With Wang-HyperBTM 4 (catalyst used is 23 mg): Yield of (1R,2S)-2-Phenylcyclopentan-1ol 11 = 45% (22 mg) and (1S,2R)-2-Phenylcyclopentyl isobutyrate S9 = 47% (33 mg).

(1*S*,2*R*)-2-Phenylcyclopentyl isobutyrate S9: $[\alpha]_D^{20} = +32.6 \ (c \ 1.0, \text{CHCl}_3) \ \{\text{Lit.}^5[\alpha]_D^{20} = +67.2 \ (90\% \text{ ee, } c \ 1.0, \text{CHCl}_3); \text{ Following General Procedure G, } (1S,2R)-2-\text{phenylcyclopentyl isobutyrate hydrolysed to corresponding alcohol } (1S,2R)-2-\text{phenylcyclopentan-1-ol 41 for HPLC} analysis. Chiral HPLC analysis, Chiralcel OD-H (98:2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (1S,2R): 14.4 min, t_R (1R,2S): 16.1 min, 92.068:7.932 er.$

(1R,2S)-2-Phenylcyclopentan-1-ol 11: $[\alpha]_D^{20} = -40.9$ (*c* 1.0, CHCl₃) {Lit.⁵ $[\alpha]_D^{20} = -60.7$ (95% ee, *c* 1.0, CHCl₃); Chiral HPLC analysis, Chiralcel OD-H (98:2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (1*S*,2*R*): 14.5 min, t_R (1*R*,2*S*): 16.1 min, 9.266:90.734 er. *s* = 29.

The quantity of recovered catalyst after proper washing and vacuum drying was 21 mg.

Cycle 4: Kinetic resolution of 1-(naphthalen-2-yl)prop-2-en-1-ol 12



Following General Procedure E, 1-(naphthalen-2-yl)prop-2-en-1-ol **12** (46 mg, 0.25 mmol), Merrifield-HyperBTM **2** (14 mg, 5 mol%), *i*-Pr₂NEt (26 μ L, 0.15 mmol, 0.6 equiv.) and (*i*-PrCO)₂O (23 μ L, 0.137 mmol, 0.55 equiv.) in DMC (1.25 mL) for 24 h at room temperature gave the crude product, which was purified by Biotage® IsoleraTM 4 to give (*R*)-1-(naphthalen-2yl)prop-2-en-1-ol **12** (22 mg, 48%) and (*S*)-1-(naphthalen-2-yl)allyl isobutyrate **S10** (30 mg, 47%).

(S)-1-(Naphthalen-2-yl)allyl isobutyrate S10: $[\alpha]_D^{20} = -54.7$ (*c* 1.0, CHCl₃) {Lit.¹⁷ $[\alpha]_D^{20} = +54$ (>99 %ee, *c* 1.0, CHCl₃); Chiral HPLC analysis Chiralcel OJ-H (95:5 hexane:IPA, flow rate 0.5 mLmin⁻¹, 211 nm, 30 °C) t_R (*R*): 15.6 min, t_R (*S*): 22.6 min, 12.460:87.540 er.

(*R*)-1-(Naphthalen-2-yl)prop-2-en-1-ol 12: $[\alpha]_D^{20} = -5.6 \ (c \ 1.0, \text{CHCl}_3) \ \{\text{Lit.}^{17} [\alpha]_D^{20} = +10.1 \ (94)^{10} \ (94)^{10} \ (95)^{10} \ (9$

The quantity of recovered catalyst after proper washing and vacuum drying was 17 mg.

With TentaGel-HyperBTM 3 (catalyst used is 29 mg): Yield of (*R*)-1-(Naphthalen-2-yl)prop-2-en-1-ol 12 = 56% (26 mg) and (*S*)-1-(Naphthalen-2-yl)allyl isobutyrate S10 = 39% (25 mg)

(S)-1-(Naphthalen-2-yl)allyl isobutyrate S10: $[\alpha]_D^{20} = -62.3$ (*c* 1.0, CHCl₃) {Lit.¹⁷ $[\alpha]_D^{20} = +54$ (>99 %ee, *c* 1.0, CHCl₃); Chiral HPLC analysis, Chiralcel OJ-H (95:5 hexane:IPA, flow rate 0.5 mLmin⁻¹, 211 nm, 30 °C) t_R (*R*): 15.6 min, t_R (*S*): 22.5 min, 10.258:89.742 er.

(*R*)-1-(Naphthalen-2-yl)prop-2-en-1-ol 12: $[\alpha]_D^{20} = -5$ (*c* 1.0, CHCl₃) {Lit.¹⁷ $[\alpha]_D^{20} = +10.1$ (94 %ee, *c* 0.8, CHCl₃); Chiral HPLC analysis, Chiralcel OJ-H (80:20 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (*S*): 11.4 min, t_R (*R*): 13.7 min, 15.227:84.773 er. *s* = 18.

The quantity of recovered catalyst after proper washing and vacuum drying was 25 mg.

With Wang-HyperBTM 4 (catalyst used is 19 mg): Yield of (*R*)-1-(Naphthalen-2-yl)prop-2en-1-ol 12 = 50% (23 mg) and (*S*)-1-(Naphthalen-2-yl)allyl isobutyrate S10 = 41% (26 mg).

(S)-1-(Naphthalen-2-yl)allyl isobutyrate S10: $[\alpha]_D^{20} = -53.1$ (*c* 1.0, CHCl₃) {Lit.¹⁷ $[\alpha]_D^{20} = +54$ (>99 %ee, *c* 1.0, CHCl₃); Chiral HPLC analysis, Chiralcel OJ-H (95:5 hexane:IPA, flow rate 0.5 mLmin⁻¹, 211 nm, 30 °C) t_R (*R*): 15.6 min, t_R (*S*): 22.6 min, 11.374:88.626 er.

(*R*)-1-(Naphthalen-2-yl)prop-2-en-1-ol 12: $[\alpha]_D^{20} = -4.8 \ (c \ 1.0, \text{CHCl}_3) \ \{\text{Lit.}^{17} [\alpha]_D^{20} = +10.1 \ (94)^{10} \ (94)^{10} \ (95)^{10} \ (9$

The quantity of recovered catalyst after proper washing and vacuum drying was 16 mg.

Cycle 5: Kinetic resolution of 2-methyl-1-(naphthalen-2-yl)propan-1-ol 13



Following General Procedure E, 2-methyl-1-(naphthalen-2-yl)propan-1-ol **13** (40 mg, 0.2 mmol), Merrifield-HyperBTM **2** (11 mg, 5 mol%), *i*-Pr₂NEt (21 μ L, 0.12 mmol, 0.6 equiv.) and (*i*-PrCO)₂O (18 μ L, 0.11 mmol, 0.55 equiv.) in DMC (1 mL) for 24 h at room temperature gave the crude product, which was purified by Biotage® IsoleraTM 4 to give (*R*)-2-methyl-1-(naphthalen-2-yl)propan-1-ol **13** (19 mg, 47%) and (*S*)-2-methyl-1-(naphthalen-2-yl)propyl isobutyrate **S11** (17 mg, 31%). (S)-2-Methyl-1-(naphthalen-2-yl)propyl isobutyrate S11: $[\alpha]_D^{20} = -67.5$ (*c* 1.0, CHCl₃) {Lit.⁵ $[\alpha]_D^{20} = -78$ (99% ee, *c* 1.0, CHCl₃); Chiral HPLC analysis, Chiralcel AD-H (99.8:0.2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (*R*): 6.9 min, t_R (*S*): 8.6 min, 3.935:96.065 er.

(*R*)-2-Methyl-1-(naphthalen-2-yl)propan-1-ol 13: $[\alpha]_D^{20} = +23.7$ (*c* 1.0, CHCl₃) {Lit.⁵ $[\alpha]_D^{20} = +25$ (82% ee, *c* 1.0, CHCl₃); Chiral HPLC analysis, Chiralcel OD-H (95:5 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (*S*): 14.1 min, t_R (*R*): 15.9 min, 0.301:99.699 er. *s* = 140.

The quantity of recovered catalyst after proper washing and vacuum drying was 9 mg.

With TentaGel-HyperBTM 3 (catalyst used is 23 mg): Yield of (*R*)-2-Methyl-1-(naphthalen-2-yl)propan-1-ol 13 = 55% (22 mg) and (*S*)-2-Methyl-1-(naphthalen-2-yl)propyl isobutyrate S11 = 26% (14 mg)

(S)-2-Methyl-1-(naphthalen-2-yl)propyl isobutyrate S11: $[\alpha]_D^{20} = -42$ (*c* 0.5, CHCl₃) {Lit.⁵ $[\alpha]_D^{20} = -78$ (99% ee, *c* 1.0, CHCl₃); Chiral HPLC analysis, Chiralcel AD-H (99.8:0.2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (*R*): 6.7 min, t_R (*S*): 8.2 min, 1.279:98.721 er.

(*R*)-2-Methyl-1-(naphthalen-2-yl)propan-1-ol 13: $[\alpha]_D^{20} = +20.6$ (*c* 1.0, CHCl₃) {Lit.⁵ $[\alpha]_D^{20} = +25$ (82% ee, *c* 1.0, CHCl₃); Chiral HPLC analysis, Chiralcel OD-H (95:5 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (*S*): 13.9 min, t_R (*R*): 15.6 min, 12.970:87.030 er. *s* = 172.

The quantity of recovered catalyst after proper washing and vacuum drying was 20 mg.

With Wang-HyperBTM 4 (catalyst used is 15 mg): Yield of (*R*)-2-Methyl-1-(naphthalen-2-yl)propan-1-ol 13 = 52% (21 mg) and (*S*)-2-Methyl-1-(naphthalen-2-yl)propyl isobutyrate S11 = 33% (18 mg).

(*S*)-2-Methyl-1-(naphthalen-2-yl)propyl isobutyrate S11: $[\alpha]_D^{20} = -46.4$ (*c* 1.0, CHCl₃) {Lit.⁵ $[\alpha]_D^{20} = -78$ (99% ee, *c* 1.0, CHCl₃); Chiralcel AD-H (99.8:0.2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (*R*): 7.2 min, t_R (*S*): 9.2 min, 1.037:98.963 er.

(*R*)-2-Methyl-1-(naphthalen-2-yl)propan-1-ol 13: $[\alpha]_D^{20} = +20.8$ (*c* 1.0, CHCl₃) {Lit.⁵ $[\alpha]_D^{20} = +25$ (82% ee, *c* 1.0, CHCl₃); Chiral HPLC analysis, Chiralcel OD-H (95:5 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (*S*): 14.1 min, t_R (*R*): 15.7 min, 12.201:87.799 er. *s* =194.

The quantity of recovered catalyst after proper washing and vacuum drying was 12 mg.

Kinetic Resolutions of Tertiary Alcohols

Reaction Optimisation



Solvent	Catalyst	(^{<i>i</i>} PrCO) ₂ O	(ⁱ PrCO) ₂ O (equiv.) ee (ester)	ee (alcohol)		<u>s</u>	
(0.17 M)	Catalyst	(equiv.)		ee (alconol)	С		
DMC	2	0.55	80.68	75.24	48	21	-
DMC	2	0.7	75.88	86.12	53	20	
DMC	3	0.55	88.83	54.73	38	29	
DMC	4	0.55	84.08	48.04	36	19	
DMC	6	0.55	32.72	0.43	1	2	
CHCl ₃	2	0.55	95.97	27.11	22	60	
Toluene	2	0.55	81.70	70.47	46	21	
^t BuOAc	2	0.55	76.68	7.7	9	8	
^t PrOAc	2	0.55	82.33	49.6	38	17	
EtOAc	2	0.55	81.61	73.42	47	22	
EtOAc	3	0.55	76.97	55.22	42	13	
EtOAc	4	0.55	80.83	71.40	47	20	
EtOAc	6	0.55	5.2	0.01	0.19	1	
^t AmOH	2	0.55					

Reaction conditions: 14 (0.1 mmol), **2**, **3**, **4** or **6** (5 mol%), (*i*-PrCO)₂O(0.55 to 0.7 equiv.), *i*-Pr₂NEt (0.6 equiv.), solvent (0.17 m), 0-r.t., 24 h. Conversion (c) and er determined by chiral HPLC analysis. Selectivity factors (*s*) calculated using er of **14** and reaction conversion (see ref. 1a), and rounded according to estimated associated errors (see ref. 1b). DMC = dimethyl carbonate, ^{*t*}BuOAc = *tert*-butyl acetate, ^{*i*}PrOAc = isopropyl acetate, ^{*t*}AmOH = *tert*-amyl alcohol.

Following General Procedure F, 3-allyl-1-benzyl-3-hydroxyindolin-2-one 14 (27.93 mg, 0.1 mmol), (*i*-PrCO)₂O (11.6 μ L, 0.07 mmol, 0.7 equiv.), Merrifield-HyperBTM 2 (5.7 mg, 0.005 mmol, 5 mol%) and *i*Pr₂NEt (10.5 μ L, 0.06 mmol) in DMC (0.6 mL) at r.t. for 24 h to give the crude products, which were purified by Biotage® IsoleraTM 4 to give *R*)-3-allyl-1-benzyl-2-oxoindolin-3-yl isobutyrate 15 (18 mg, 0.0515 mmol, 52%) and (*S*)-3-allyl-1-benzyl-3-hydroxyindolin-2-one 14 (13 mg, 0.047 mmol, 47%).

Chiral HPLC analysis: Chiralcel OD-H (98:2 hexane:IPA, flow rate 1 mLmin⁻¹, 254 nm, 30 °C); (*R*)-3-Allyl-1-benzyl-2-oxoindolin-3-yl isobutyrate 15: t_R (*R*): 8.3 min, t_R (*S*): 10.1 min, 87.773:12.227 er. (*S*)-3-Allyl-1-benzyl-3-hydroxyindolin-2-one 14: t_R (*R*): 32.6 min, t_R (*S*): 37.8 min, 6.938:93.062 er. (Table 3, entry 9)

Following **General Procedure F**, 3-allyl-1-benzyl-3-hydroxyindolin-2-one **14** (28 mg, 0.1 mmol), (*i*-PrCO)₂O (9.1 μ L, 0.055 mmol, 0.55 equiv.), Merrifield-HyperBTM **2** (5.7 mg, 0.005 mmol, 5 mol%) and *i*Pr₂NEt (10.5 μ L, 0.06 mmol) in DMC (0.6 mL) at r.t. for 24 h to give the crude products, which were purified by Biotage® IsoleraTM 4 to give (*R*)-3-allyl-1-benzyl-2-oxoindolin-3-yl isobutyrate **15** (16 mg, 0.0457 mmol, 46%) and (*S*)-3-allyl-1-benzyl-3-hydroxyindolin-2-one **14** (14 mg, 0.0501 mmol, 50%).

(*R*)-3-Allyl-1-benzyl-2-oxoindolin-3-yl isobutyrate 15: $[\alpha]_D^{20} = -19.5$ (*c* 1.0, CHCl₃) {Lit.⁹ (*ent*, 95% ee) $[\alpha]_D^{20} = +20$ (*c* 1.0, CHCl₃); Chiral HPLC analysis: Chiralcel OD-H (98:2 hexane:IPA, flow rate 1 mLmin⁻¹, 254 nm, 30 °C); ester: t_R (*R*): 8.3 min, t_R (*S*): 10.0 min, 90.188:9.812 er.

(*S*)-3-Allyl-1-benzyl-3-hydroxyindolin-2-one 14: $[\alpha]_D^{20} = -4.5$ (*c* 1.0, CHCl₃) {Lit.⁹ (*ent*, 88% ee)) $[\alpha]_D^{27} = +13$ (*c* 1.0, CHCl₃)}; Chiral HPLC analysis, Chiralcel OD-H (98:2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C); t_R (*R*): 32.5 min, t_R (*S*): 37.6 min, 13.247:86.753. *s* = 21. (Table 3, entry 6).

Tentagel-HyperBTM 3 (19 mg, 0.005 mmol, 5 mol%), r.t.

Chiral HPLC analysis: Chiralcel OD-H (98:2 hexane:IPA, flow rate 1 mLmin⁻¹, 254 nm, 30 °C); (*R*)-3-Allyl-1-benzyl-2-oxoindolin-3-yl isobutyrate 15: t_R (*R*): 8.2 min, t_R (*S*): 10.1 min, 94.414:5.586 er. (*S*)-3-Allyl-1-benzyl-3-hydroxyindolin-2-one 14: t_R (*R*): 32.8 min, t_R (*S*): 38.5 min, 22.637:77.363 er. (Table 3, entry 7)

Wang-HyperBTM 4 (13 mg, 0.005 mmol, 5 mol%), r.t.

Chiral HPLC analysis: Chiralcel OD-H (98:2 hexane:IPA, flow rate 1 mLmin⁻¹, 254 nm, 30 °C); (*R*)-3-Allyl-1-benzyl-2-oxoindolin-3-yl isobutyrate 15: t_R (*R*): 8.2 min, t_R (*S*): 10.0 min, 92.040:7.960 er. (*S*)-3-Allyl-1-benzyl-3-hydroxyindolin-2-one 14: t_R (*R*): 32.8 min, t_R (*S*): 38.6 min, 25.964:74.036 er. (Table 3, entry 8)

Merrifield-BTM 6 (5.81 mg, 0.01 mmol, 5 mol%), r.t.

Chiral HPLC analysis: Chiralcel OD-H (98:2 hexane:IPA, flow rate 1 mLmin⁻¹, 254 nm, 30 °C); (*R*)-3-Allyl-1-benzyl-2-oxoindolin-3-yl isobutyrate 15: t_R (*R*): 8.3 min, t_R (*S*): 10.1 min, 65.736:34.264 (*R*:*S*) er. (*S*)-3-Allyl-1-benzyl-3-hydroxyindolin-2-one 14: t_R (*R*): 32.9 min, t_R (*S*): 38.8 min, 49.376:50.624 (*R*:*S*) er. (Table 3, entry 10)

Table 4: Substrate Scope



Following General Procedure F, *tert*-butyl 3-allyl-3-hydroxy-2-oxoindoline-1-carboxylate 16 (28.9 mg, 0.1 mmol), isobutyric anhydride (11.6 μL, 0.07 mmol), Merrifield-HyperBTM 2 (5.7

mg, 0.005 mmol, 5 mol%) and *i*Pr₂NEt (10.5 μ L, 0.06 mmol) in DMC (1.2 mL) at r.t. for 24 h to give the crude products, which were purified by flash column chromatography (CH₂Cl₂:EtOAc 9:1) to give *tert*-butyl 3-allyl-3-(isobutyryloxy)-2-oxoindoline-1-carboxylate **S12** (19 mg, 0.052 mmol, 53%) and *tert*-butyl 3-allyl-3-hydroxy-2-oxoindoline-1-carboxylate **16** (9 mg, 0.031 mmol, 31%).

(*R*)-*tert*-Butyl 3-allyl-3-(isobutyryloxy)-2-oxoindoline-1-carboxylate S12: $[\alpha]_D^{20} = -10.6$ (*c* 0.5, CHCl₃) {Lit.⁹ (*ent*, 92% ee) $[\alpha]_D^{20} = +17$ (*c* 0.1, CHCl₃);}; Chiral HPLC analysis, Chiralcel OD-H (99:1 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (*R*): 4.7 min, t_R (*S*): 7.6 min, 77.241:22.759 (*R*:*S*) er..

(*S*)-*tert*-Butyl 3-allyl-3-hydroxy-2-oxoindoline-1-carboxylate 16: $[\alpha]_D^{20} = -34.6 (c \ 0.5, \text{CHCl}_3)$ {Lit.⁹(*ent*, 98% ee) $[\alpha]_D^{20} = +43 (c \ 0.1, \text{CHCl}_3)$ }; Chiral HPLC analysis, Chiralcel AD-H (95:5 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C)) t_R (*R*): 10.7 min, t_R (*S*): 13.4 min, 1.048:98.952 (*R*:*S*) er. *s* = 14.

1-Benzyl-3-ethyl-3-hydroxyindolin-2-one 17



Following General Procedure F, 1-benzyl-3-hydroxy-3-isopropylindolin-2-one **17** (28.1 mg, 0.1 mmol), isobutyric anhydride (11.6 μ L, 0.07 mmol), Merrifield-HyperBTM **2** (5.7 mg, 0.005 mmol, 5 mol%) and *i*Pr₂NEt (10.5 μ L, 0.06 mmol) in DMC (1.2 mL) at r.t. for 48 h to give the crude products, which were purified by Biotage® Isolera 4 chromatography (eluent: $0\% \rightarrow 30\%$ EtOAc in hexane) to give 1-benzyl-3-isopropyl-2-oxoindolin-3-yl isobutyrate **S13** (15 mg, 0.041 mmol, 42%) and 1-benzyl-3-hydroxy-3-isopropylindolin-2-one **17** (15 mg, 0.053 mmol, 53%).

(*R*)-1-Benzyl-3-ethyl-2-oxoindolin-3-yl isobutyrate S13: $[\alpha]_D^{20} = +26.3$ (*c* 1.0, CHCl₃) {Lit.⁹(*ent*, 93% ee) $[\alpha]_D^{20} = -30$ (*c* 1.0, CHCl₃)}; Chiral HPLC analysis, Chiralpak AD-H (98:2 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 8.2 min, t_R (*R*): 10.2 min, 12.511:87.489 er.

(*S*)-1-Benzyl-3-ethyl-3-hydroxyindolin-2-one 17: $[\alpha]_D^{20} = -49.5$ (*c* 1.0, CHCl₃) {Lit.⁹(*ent*, 70% ee) $[\alpha]_D^{25} = +59$ (*c* 1.0, CHCl₃)}; Chiral HPLC analysis, Chiralpak AD-H (98:2 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*R*): 38.4 min, t_R (*S*): 46.9 min, 19.578:80.422 er.

1-Benzyl-3-hydroxy-3-(trifluoromethyl)indolin-2-one 18



Following **General Procedure F**, 1-benzyl-3-hydroxy-3-(trifluoromethyl)indolin-2-one **18** (30.7 mg, 0.1 mmol), isobutyric anhydride (11.6 μ L, 0.07 mmol), Merrifield-HyperBTM **2** (5.7 mg, 0.005 mmol, 5 mol%) and *i*Pr₂NEt (10.5 μ L, 0.06 mmol) in DMC (1.2 mL) at r.t. for 20 h to give the crude products, which were purified by flash column chromatography (hexane:EtOAc 10:2) to give 1-benzyl-2-oxo-3-(trifluoromethyl)indolin-3-yl isobutyrate **S14** (16 mg, 0.042 mmol, 42%) and 1-benzyl-3-hydroxy-3-(trifluoromethyl)indolin-2-one **18** (11 mg, 0.035 mmol, 36%).

(*R*)-1-Benzyl-2-oxo-3-(trifluoromethyl)indolin-3-yl isobutyrate S14: $[\alpha]_D^{20} = +3.0$ (*c* 1.0, CHCl₃) {Lit.⁹ (*ent*, 70% ee) $[\alpha]_D^{20} = -8.0$ (*c* 0.1, CHCl₃)}; Chiral HPLC analysis, Chiralpak AD-H (99:1 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R (*S*): 12.7 min, t_R (*R*): 15.3 min, 19.991:80.009 er.

(S)-1-Benzyl-3-hydroxy-3-(trifluoromethyl)indolin-2-one 18: $[\alpha]_D^{20} = -52$ (c 0.5, CHCl₃) {Lit.⁹ (*ent*, >99% ee) $[\alpha]_D^{20} = +60$ (c 0.1, CHCl₃)}; Chiral HPLC analysis, Chiralpak AD-H (95:5) hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R (*R*): 15.8 min, t_R (*S*): 20.5 min, 0.305:99.695 er.

1-Benzyl-3-hydroxy-3-phenylindolin-2-one 19



Following General Procedure F, 1-benzyl-3-hydroxy-3-phenylindolin-2-one **19** (31.5 mg, 0.1 mmol), isobutyric anhydride (11.6 μ L, 0.07 mmol), M-HyperBTM (5.7 mg, 0.005 mmol, 5 mol%) and *i*Pr₂NEt (10.5 μ L, 0.06 mmol) in DMC (1.2 mL) at r.t. for 48 h to give the crude products, which were purified by Biotage® Isolera 4 chromatography (eluent: 0% \rightarrow 25% EtOAc in hexane) to give 1-benzyl-2-oxo-3-phenylindolin-3-yl isobutyrate **S15** (19 mg, 0.049 mmol, 49%) and 1-benzyl-3-hydroxy-3-phenylindolin-2-one **19** (15 mg, 0.047 mmol, 48%).

(*R*)-1-Benzyl-2-oxo-3-phenylindolin-3-yl isobutyrate S15: $[\alpha]_D^{20} = -93.5$ (*c* 1.0, CHCl₃) {Lit.⁹ (*ent*, 90% ee) $[\alpha]_D^{20} = +107$ (*c* 1.0, CHCl₃)}; Chiral HPLC analysis, Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.25 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 12.2 min, t_R (*R*): 25.6 min, 4.968:95.032 er.

(S)-1-Benzyl-3-hydroxy-3-phenylindolin-2-one 19: $[\alpha]_D^{20} = -51.5$ (*c* 1.0, CHCl₃) {Lit.⁹ (*ent*, 99.6% ee) 98% ee) $[\alpha]_D^{20} = +55$ (*c* 1.0, CHCl₃)}; Chiral HPLC analysis, Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.25 mL min⁻¹, 211 nm, 40 °C) t_R (*R*): 19.7 min, t_R (*S*): 25.5 min, 3.556:96.444 er.

1-Allyl-3-hydroxy-3-phenylindolin-2-one 20



Following **General Procedure F**, 1-allyl-3-hydroxy-3-phenylindolin-2-one **20** (26.5 mg, 0.1 mmol), isobutyric anhydride (11.6 μ L, 0.07 mmol), Merrifield-HyperBTM **2** (5.7 mg, 0.005 mmol, 5 mol%) and *i*Pr₂NEt (10.5 μ L, 0.06 mmol) in DMC (1.2 mL) at r.t. for 48 h to give the crude products, which were purified by flash column chromatography (Petrol:EtOAc 90:10) to give 1-allyl-2-oxo-3-phenylindolin-3-yl isobutyrate **S16** (17 mg, 0.050 mmol, 51%) and 1-allyl-3-hydroxy-3-phenylindolin-2-one **20** (11 mg, 0.041 mmol, 41%).

(*R*)-1-Allyl-2-oxo-3-phenylindolin-3-yl isobutyrate S16: $[\alpha]_D^{20} = -102.5$ (*c* 0.1, CHCl₃); Chiral HPLC analysis Chiralcel OD-H (99:1 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R (*R*): 11.1 min, t_R (*S*): 12.8 min, 92.201:7.799 er.

(S)-1-Allyl-3-hydroxy-3-phenylindolin-2-one 20: $[\alpha]_D^{20} = -82.5$ (*c* 0.1, CHCl₃); Chiral HPLC analysis Chiralcel OD-H (95:5 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R (S): 13.9 min, t_R (*R*): 16.0 min, 99.852:0.148 (er.

3-Hydroxy-1-methyl-3-phenylindolin-2-one 21



Following General Procedure F, 3-hydroxy-1-methyl-3-phenylindolin-2-one **21** (23.9 mg, 0.1 mmol), isobutyric anhydride (11.6 µL, 0.07 mmol), Merrifield-HyperBTM **2** (5.7 mg, 0.005

mmol, 5 mol%) and *i*Pr₂NEt (10.5 μ L, 0.06 mmol) in DMC (1.2 mL) at r.t. for 24 h to give the crude products, which were purified by flash column chromatography (Petrol:EtOAc 85:15) to give 1-methyl-2-oxo-3-phenylindolin-3-yl isobutyrate **S17** (15 mg, 0.048 mmol, 48%) and 3-hydroxy-1-methyl-3-phenylindolin-2-one **21** (11 mg, 0.045 mmol, 46%).

(*R*)-1-Methyl-2-oxo-3-phenylindolin-3-yl isobutyrate S17: $[\alpha]_D^{20} = -149$ (*c* 1.0, CHCl₃); Chiral HPLC analysis, Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R (*S*): 8.9 min, t_R (*R*): 12.2 min, 5.919:94.081 (*S*:*R*) er..

(*S*)-3-Hydroxy-1-methyl-3-phenylindolin-2-one 21: $[\alpha]_D^{20} = -77$ (*c* 1.0, CHCl₃); Chiral HPLC analysis Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R (*R*): 27.9 min, t_R (*S*): 31.0 min, 0.994:99.006 er.

1-Allyl-3-hydroxy-3-(6-methoxynaphthalen-2-yl)indolin-2-one 22



Following **General Procedure F**, 1-allyl-3-hydroxy-3-(6-methoxynaphthalen-2-yl)indolin-2-one **22** (34.5 mg, 0.1 mmol), isobutyric anhydride (11.6 μ L, 0.07 mmol), M-HyperBTM (5.7 mg, 0.005 mmol, 5 mol%) and *i*Pr₂NEt (10.5 μ L, 0.06 mmol) in DMC (1.2 mL) at r.t. for 24 h to give the crude products, which were purified by Biotage® Isolera 4 chromatography (eluent:0% \rightarrow 30% EtOAc in hexane) to give 1-allyl-3-(6-methoxynaphthalen-2-yl)-2-oxoindolin-3-yl isobutyrate **S18** (23 mg, 0.055 mmol, 55%) and 1-allyl-3-hydroxy-3-(6-methoxynaphthalen-2-yl)indolin-2-one **22** (14 mg, 0.040 mmol, 41%).

(*R*)-1-Allyl-3-(6-methoxynaphthalen-2-yl)-2-oxoindolin-3-yl isobutyrate S18: $[\alpha]_D^{20} = -98.7$ (*c* 1.0, CHCl₃); Chiral HPLC analysis, Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 18.7 min, t_R (*R*): 33.6 min, 12.592:87.408 er.

(*S*)-1-Allyl-3-hydroxy-3-(6-methoxynaphthalen-2-yl)indolin-2-one 22: $[\alpha]_D^{20} = -21.2$ (*c* 1.0, CHCl₃); Chiral HPLC analysis Chiralcel OJ-H (90:10 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*R*): 34.7 min, t_R (*S*): 47.7 min, 0.451:99.549 er.

1-Allyl-3-(3,5-bis(trifluoromethyl)phenyl)-3-hydroxyindolin-2-one 23



Following **General Procedure F**, 1-allyl-3-(3,5-bis(trifluoromethyl)phenyl)-3-hydroxyindolin-2-one **23** (40.1 mg, 0.1 mmol), isobutyric anhydride (11.6 μ L, 0.07 mmol), M-HyperBTM (5.7 mg, 0.005 mmol, 5 mol%) and *i*Pr₂NEt (10.5 μ L, 0.06 mmol) in DMC (1.2 mL) at r.t. for 48 h to give the crude products, which were purified by Biotage® Isolera 4 chromatography (Hexane:EtOAc 90:10) to give 1-allyl-3-(3,5-bis(trifluoromethyl)phenyl)-2-oxoindolin-3-yl isobutyrate **S19** (28 mg, 0.059 mmol, 59%) and 1-allyl-3-(3,5-bis(trifluoromethyl)phenyl)-3-hydroxyindolin-2-one **23** (15 mg, 0.037 mmol, 37%).

(*R*)-1-Allyl-3-(3,5-bis(trifluoromethyl)phenyl)-2-oxoindolin-3-yl isobutyrate S19: $[\alpha]_D^{20} = -$

60 (*c* 1.0, CHCl₃) {Lit.⁹ (*ent*, 90% ee) $[\alpha]_D^{20} = +102$ (*c* 2.0, CHCl₃)}; Chiral HPLC analysis – suitable conditions couldn't be found to separate the enantiomers of the ester using the chiral HPLC columns Chiralcel OD-H & OJ-H, and Chiralpak AD-H, AS-H, IA, IB, IC & ID. The ester was therefore hydrolysed following general procedure G and the resulting alcohol analysed by Chiral HPLC:

Chiralpak AS-H (99:1 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*R*): 6.9 min, t_R (*S*): 27.5 min, 81.747:18.253 er.

(S)-1-Allyl-3-(3,5-bis(trifluoromethyl)phenyl)-3-hydroxyindolin-2-one 23: $[\alpha]_D^{20} = -33.9$ (*c* 1.0, CHCl₃) {Lit.⁹ (*ent*, 92% ee) $[\alpha]_D^{20} = +37$ (*c* 2.0, CHCl₃)}; Chiral HPLC analysis Chiralpak AS-H (99:1 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*R*): 7.1 min, t_R (*S*): 28.7 min, 5.053:94.947 er.

1-Allyl-3-hydroxy-3-(thiophen-2-yl)indolin-2-one 24



Following **General Procedure F**, 1-allyl-3-hydroxy-3-(thiophen-2-yl)indolin-2-one **24** (27.1 mg, 0.1 mmol), isobutyric anhydride (11.6 μ L, 0.07 mmol), M-HyperBTM (5.7 mg, 0.005 mmol, 5 mol%) and *i*Pr₂NEt (10.5 μ L, 0.06 mmol) in DMC (1.2 mL) at r.t. for 48 h to give the crude products, which were purified by Biotage® Isolera 4 chromatography (Hexane:EtOAc 80:20) to give 1-allyl-2-oxo-3-(thiophen-2-yl)indolin-3-yl isobutyrate **S20** (18 mg, 0.052 mmol, 53%) and 1-allyl-3-hydroxy-3-(thiophen-2-yl)indolin-2-one **24** (17 mg, 0.040 mmol, 41%).

(*R*)-1-Allyl-2-oxo-3-(thiophen-2-yl)indolin-3-yl isobutyrate S20: $[\alpha]_D^{20} = -127.5$ (*c* 0.2, CHCl₃); Chiral HPLC analysis Chiralcel OD-H (99:1 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R (*R*): 13.1 min, t_R (*S*): 17.0 min, 87.916:12.084 er.

(*S*)-1-Allyl-3-hydroxy-3-(thiophen-2-yl)indolin-2-one 24: $[\alpha]_D^{20} = +4.5$ (*c* 0.5, CHCl₃); Chiral HPLC analysis Chiralcel OD-H (95:5 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R (*S*): 17.8 min, t_R (*R*): 22.5 min, 99.254:0.746 er.

1-Allyl-3-hydroxy-3-(pyridin-3-yl)indolin-2-one 25



Following **General Procedure F**, 1-allyl-3-hydroxy-3-(pyridin-3-yl)indolin-2-one **25** (26.6 mg, 0.1 mmol), isobutyric anhydride (11.6 μ L, 0.07 mmol), M-HyperBTM (5.7 mg, 0.005 mmol, 5 mol%) and *i*Pr₂NEt (10.5 μ L, 0.06 mmol) in DMC (1.2 mL) at r.t. for 48 h to give the crude products, which were purified by Biotage® Isolera 4 chromatography (Hexane:EtOAc 75:25) to give 1-allyl-2-oxo-3-(pyridin-3-yl)indolin-3-yl isobutyrate **S21** (19 mg, 0.056 mmol, 56%) and 1-allyl-3-hydroxy-3-(pyridin-3-yl)indolin-2-one **25** (9 mg, 0.033 mmol, 34%).

(S)-1-Allyl-2-oxo-3-(pyridin-3-yl)indolin-3-yl isobutyrate S21: $[\alpha]_D^{20} = -83.4$ (c 0.5, CHCl₃) {Lit.⁹ (*ent*, 88% ee) $[\alpha]_D^{20} = +123$ (c 0.5, CHCl₃)}; Chiral HPLC analysis Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*R*): 14.8 min, t_R (*S*): 21.8 min, 15.803:84.197 er.

(*R*)-1-Allyl-3-hydroxy-3-(pyridin-3-yl)indolin-2-one 25: $[\alpha]_D^{20} = -16.6$ (*c* 1.0, CHCl₃) {Lit.⁹ (*ent*, 96.6% ee) $[\alpha]_D^{20} = +38$ (*c* 0.5, CHCl₃)}; Chiral HPLC analysis Chiralpak AD-H (92:8 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 20.1 min, t_R (*R*): 22.7 min, 0.586:99.414 er.

1-Benzyl-3-hydroxy-3-(2-methylprop-1-en-1-yl)indolin-2-one 26



Following General Procedure F, 1-benzyl-3-hydroxy-3-(2-methylprop-1-en-1-yl)indolin-2-one **26** (29.3 mg, 0.1 mmol), isobutyric anhydride (11.6 μ L, 0.07 mmol), Merrifield-HyperBTM **2** (5.7 mg, 0.005 mmol, 5 mol%) and *i*Pr₂NEt (10.5 μ L, 0.06 mmol) in DMC (1.2 mL) at r.t. for 48 h to give the crude products, which were purified by flash column chromatography (hexane:EtOAc 4:1) to give 1-benzyl-3-(2-methylprop-1-en-1-yl)-2-oxoindolin-3-yl isobutyrate **S22** (15 mg, 0.041 mmol, 41%) and 1-benzyl-3-hydroxy-3-(2-methylprop-1-en-1-yl)indolin-2-one **26** (16 mg, 0.054 mmol, 55%).

(*R*)-1-Benzyl-3-(2-methylprop-1-en-1-yl)-2-oxoindolin-3-yl isobutyrate S22: $[\alpha]_D^{20} = -43$ (*c* 1.0, CHCl₃) {Lit.⁹ (*ent*, 94% ee) $[\alpha]_D^{20} = +65$ (*c* 0.1, CHCl₃)}; Chiral HPLC analysis Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R (*S*): 8.9 min, t_R (*R*): 17.2 min, 7.530:92.470 er.

(S)-1-Benzyl-3-hydroxy-3-(2-methylprop-1-en-1-yl)indolin-2-one 26: $[\alpha]_D^{20} = -46.8$ (c 1.0, CHCl₃) {Lit.⁹ (*ent*, 74% ee) $[\alpha]_D^{20} = +97$ (c 0.1, CHCl₃)}; Chiral HPLC analysis Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R (*R*): 32.7 min, t_R (*S*): 34.7 min, 16.801:83.199 er

1-Benzyl-4-chloro-3-hydroxy-3-methylindolin-2-one 27



Following **General Procedure F**, 1-benzyl-4-chloro-3-hydroxy-3-methylindolin-2-one **27** (28.8 mg, 0.1 mmol), isobutyric anhydride (11.6 μ L, 0.07 mmol), Merrifield-HyperBTM **2** (5.7 mg, 0.005 mmol, 5 mol%) and *i*Pr₂NEt (10.5 μ L, 0.06 mmol) in DMC (1.2 mL) at r.t. for 48 h to give the crude products, which were purified by Biotage® Isolera 4 chromatography (hexane:EtOAc 80:20) to give 1-benzyl-4-chloro-3-methyl-2-oxoindolin-3-yl isobutyrate **S23** (14 mg, 0.039 mmol, 39%) and 1-benzyl-4-chloro-3-hydroxy-3-methylindolin-2-one **27** (16 mg, 0.055 mmol, 56%).

(*R*)-1-Benzyl-4-chloro-3-methyl-2-oxoindolin-3-yl isobutyrate S23: $[\alpha]_D^{20} = +19$ (*c* 1.0, CHCl₃) {Lit.⁹ (*ent*, 93% ee) $[\alpha]_D^{20} = -21$ (*c* 1.0, CHCl₃)}; Chiral HPLC analysis, Chiralpak AD-H (98:2 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 8.6 min, t_R (*R*): 10.3 min, 10.001:89.999 (*S*:*R*) er.

(S)-1-Benzyl-4-chloro-3-hydroxy-3-methylindolin-2-one 27: $[\alpha]_D^{20} = -23.9$ (c 1.0, CHCl₃) {Lit.⁹ (*ent*, 86% ee) $[\alpha]_D^{20} = +32$ (c 1.0, CHCl₃)}; Chiral HPLC analysis Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*R*): 17.6 min, t_R (*S*): 20.3 min, 21.819:78.181 (*R*:*S*) er.

1-Benzyl-3-hydroxy-5-methyl-3-phenylindolin-2-one 28



Following **General Procedure F**, 1-benzyl-3-hydroxy-5-methyl-3-phenylindolin-2-one **28** (32.9 mg, 0.1 mmol), isobutyric anhydride (11.6 μ L, 0.07 mmol), Merrifield-HyperBTM **2** (5.7 mg, 0.005 mmol, 5 mol%) and *i*Pr₂NEt (10.5 μ L, 0.06 mmol) in DMC (1.2 mL) at r.t. for 48 h to give the crude products, which were purified by Biotage® Isolera 4 chromatography (hexane:EtOAc 80:20) to give 1-benzyl-5-methyl-2-oxo-3-phenylindolin-3-yl isobutyrate **S24** (20 mg, 0.050 mmol, 50%) and 1-benzyl-3-hydroxy-5-methyl-3-phenylindolin-2-one **28** (15 mg, 0.045 mmol, 46%).

(*R*)-1-Benzyl-5-methyl-2-oxo-3-phenylindolin-3-yl isobutyrate S24: $[\alpha]_D^{20} = -98.4$ (*c* 1.0, CHCl₃) {Lit.⁹ (*ent*, 94% ee) $[\alpha]_D^{20} = +107$ (*c* 1.0, CHCl₃)}; Chiral HPLC analysis Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 11.3 min, t_R (*R*): 25.2 min, 10.298:89.702 er.

(S)-1-Benzyl-3-hydroxy-5-methyl-3-phenylindolin-2-one 28: $[\alpha]_D^{20} = -28.7$ (*c* 1.0, CHCl₃) {Lit.⁹ (*ent*, 83% ee) $[\alpha]_D^{20} = +23.7$ (*c* 1.0, CHCl₃)}; Chiral HPLC analysis Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*R*): 12.4 min, t_R (*S*): 15.9 min, 4.610:95.390 er.

1-Benzyl-6-chloro-3-hydroxy-3-phenylindolin-2-one 29



Following **General Procedure F**, 1-benzyl-6-chloro-3-hydroxy-3-phenylindolin-2-one **29** (35 mg, 0.1 mmol), isobutyric anhydride (11.6 μ L, 0.07 mmol), Merrifield-HyperBTM **2** (5.7 mg, 0.005 mmol, 5 mol%) and *i*Pr₂NEt (10.5 μ L, 0.06 mmol) in DMC (1.2 mL) at r.t. for 48 h to give the crude products, which were purified by Biotage® Isolera 4 chromatography (hexane:EtOAc 80:20) to give 1-benzyl-6-chloro-2-oxo-3-phenylindolin-3-yl isobutyrate **S25** (20 mg, 0.047 mmol, 48%) and 1-benzyl-6-chloro-3-hydroxy-3-phenylindolin-2-one **29** (13 mg, 0.037 mmol, 37%).

(*R*)-1-Benzyl-6-chloro-2-oxo-3-phenylindolin-3-yl isobutyrate S25: $[\alpha]_D^{20} = -120$ (*c* 0.5, CHCl₃); Chiral HPLC analysis Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 7.9 min, t_R (*R*): 15.5 min, 20.665:79.335 er.

(*S*)-1-Benzyl-6-chloro-3-hydroxy-3-phenylindolin-2-one 29: $[\alpha]_D^{20} = -48.8$ (*c* 0.5, CHCl₃); Chiral HPLC analysis Chiralpak AD-H (92:8 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*R*): 13.8 min, t_R (*S*): 15.9 min, 18.545:81.455 er.

1-Benzyl-7-chloro-3-hydroxy-3-methylindolin-2-one 30



Following **General Procedure F**, 1-benzyl-7-chloro-3-hydroxy-3-methylindolin-2-one **30** (28.8 mg, 0.1 mmol), isobutyric anhydride (11.6 μ L, 0.07 mmol), Merrifield-HyperBTM **2** (5.7 mg, 0.005 mmol, 5 mol%) and *i*Pr₂NEt (10.5 μ L, 0.06 mmol) in DMC (1.2 mL) at r.t. for 24 h to give

the crude products, which were purified by Biotage® Isolera 4 chromatography (hexane:EtOAc 87:13) to give 1-benzyl-7-chloro-3-methyl-2-oxoindolin-3-yl isobutyrate **S26** (18 mg, 0.050 mmol, 50%) and 1-benzyl-7-chloro-3-hydroxy-3-methylindolin-2-one **30** (11 mg, 0.038 mmol, 38%).

(*R*)-1-Benzyl-7-chloro-3-methyl-2-oxoindolin-3-yl isobutyrate S26: $[\alpha]_D^{20} = -13.5$ (*c* 1.0, CHCl₃) {Lit.⁹ (*ent*, 77% ee) $[\alpha]_D^{20} = +20$ (*c* 3.0, CHCl₃)}; Chiral HPLC analysis Chiralpak AD-H (99:1 hexane:IPA, flow rate 1.5 mL min⁻¹, 254 nm, 40 °C) t_R (*S*): 11.2 min, t_R (*R*): 12.9 min, 24.370:75.630 er.

(S)-1-Benzyl-7-chloro-3-hydroxy-3-methylindolin-2-one 30: $[\alpha]_D^{20} = -37.6$ (*c* 0.5, CHCl₃) {Lit.⁹ (*ent*, 99% ee) $[\alpha]_D^{20} = +39$ (*c* 2.0, CHCl₃)}; Chiral HPLC analysis Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*R*): 11.6 min, t_R (*S*): 13.8 min, 9.044:90.956 er.

Recyclability of Merrifield-HyperBTM 2

Cycle 1: 1-Benzyl-3-hydroxy-3-phenylindolin-2-one 19



Following **General Procedure F**, 1-benzyl-3-hydroxy-3-phenylindolin-2-one **19** (150 mg, 0.47 mmol), isobutyric anhydride (55.2 μ L, 0.33 mmol), Merrifield-HyperBTM **2** (27 mg, 0.023 mmol, 5 mol%) and *i*Pr₂NEt (49.7 μ L, 0.28 mmol) in DMC (2.8 mL) at r.t. for 48 h to give the crude products, which were purified by Biotage® Isolera 4 chromatography (hexane:EtOAc 80:20) to give 1-benzyl-2-oxo-3-phenylindolin-3-yl isobutyrate **S15** (93 mg, 0.24 mmol, 51%) and 1-benzyl-3-hydroxy-3-phenylindolin-2-one **19** (65 mg, 0.21 mmol, 43%).

(*R*)-1-Benzyl-2-oxo-3-phenylindolin-3-yl isobutyrate S15: Chiral HPLC analysis Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.25 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 12.1 min, t_R (*R*): 25.5 min, 10.228:89.772 er

(S)-1-Benzyl-3-hydroxy-3-phenylindolin-2-one 19: Chiral HPLC analysis Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.25 mL min⁻¹, 211 nm, 40 °C) $t_R(R)$: 19.5 min, $t_R(S)$: 25.1 min, 0.088:99.912 er.

Cycle 2: 1-Allyl-3-hydroxy-3-phenylindolin-2-one 20



Following **General Procedure F**, 1-allyl-3-hydroxy-3-phenylindolin-2-one **20** (121.4 mg, 0.45 mmol), isobutyric anhydride (53.1 μ L, 0.32 mmol), Merrifield-HyperBTM **2** (26 mg, 0.022 mmol, 5 mol%) and *i*Pr₂NEt (47.8 μ L, 0.27 mmol) in DMC (2.7 mL) at r.t. for 48 h to give the crude products, which were purified by flash column chromatography (Petrol:EtOAc 90:10) to give 1-allyl-2-oxo-3-phenylindolin-3-yl isobutyrate **S16** (75 mg, 0.233 mmol, 49%) and 1-allyl-3-hydroxy-3-phenylindolin-2-one **20** (561 mg, 0.211 mmol, 46%).

(*R*)-1-Allyl-2-oxo-3-phenylindolin-3-yl isobutyrate S16: Chiral HPLC analysis Chiralcel OD-H (99:1 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R (*R*): 11.6 min, t_R (*S*): 13.3 min, 92.344:7.656 er.

(S)-1-Allyl-3-hydroxy-3-phenylindolin-2-one 20: Chiral HPLC analysis Chiralcel OD-H (95:5 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R (S): 13.9 min, t_R (R): 15.9 min, 99.852:0.148 er.

Cycle 3: 1-Benzyl-3-hydroxy-3-phenylindolin-2-one 19



Following **General Procedure F**, 1-benzyl-3-hydroxy-3-phenylindolin-2-one (127.7 mg, 0.40 mmol), isobutyric anhydride (47.0 μ L, 0.28 mmol), Merrifield-HyperBTM **2** (23 mg, 0.020 mmol, 5 mol%) and *i*Pr₂NEt (42.3 μ L, 0.24 mmol) in DMC (2.4 mL) at r.t. for 48 h to give the crude products, which were purified by Biotage® Isolera 4 chromatography (hexane:EtOAc 80:20) to give 1-benzyl-2-oxo-3-phenylindolin-3-yl isobutyrate **S15** (80 mg, 0.21 mmol, 51%) and 1-benzyl-3-hydroxy-3-phenylindolin-2-one **19** (55 mg, 0.17 mmol, 43%).

(*R*)-1-Benzyl-2-oxo-3-phenylindolin-3-yl isobutyrate S15: Chiral HPLC analysis Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.25 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 12.1 min, t_R (*R*): 25.4 min, 7.797:92.203 er.

(S)-1-Benzyl-3-hydroxy-3-phenylindolin-2-one 19: Chiral HPLC analysis Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.25 mL min⁻¹, 211 nm, 40 °C) $t_R(R)$: 19.4 min, $t_R(S)$: 25.1 min, 0.276:99.724 er.

Cycle 4: 3-Hydroxy-1-methyl-3-phenylindolin-2-one 21



Following **General Procedure F**, 3-hydroxy-1-methyl-3-phenylindolin-2-one **21** (88.43 mg, 0.37 mmol), isobutyric anhydride (42.9 μ L, 0.25 mmol), Merrifield-HyperBTM **2** (21 mg, 0.018 mmol, 5 mol%) and *i*Pr₂NEt (38.6 μ L, 0.22 mmol) in DMC (2.2 mL) at r.t. for 24 h to give the crude products, which were purified by flash column chromatography (Petrol:EtOAc 85:15) to give 1-methyl-2-oxo-3-phenylindolin-3-yl isobutyrate **S17** (57 mg, 0.184 mmol, 50%) and 3-hydroxy-1-methyl-3-phenylindolin-2-one **21** (42 mg, 0.175 mmol, 47%).

(*R*)-1-Methyl-2-oxo-3-phenylindolin-3-yl isobutyrate S17: Chiral HPLC analysis Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R (*S*): 8.9 min, t_R (*R*): 12.2 min, 5.178:94.822 er.

(S)-3-Hydroxy-1-methyl-3-phenylindolin-2-one 21: Chiral HPLC analysis Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R (*R*): 27.8 min, t_R (*S*): 30.9 min, 0.718:99.282 er.

Cycle 5: 1-Benzyl-3-hydroxy-3-phenylindolin-2-one 19



Following **General Procedure F**, 1-benzyl-3-hydroxy-3-phenylindolin-2-one **19** (111 mg, 0.35 mmol), isobutyric anhydride (40.9 μ L, 0.24 mmol), Merrifield-HyperBTM **2** (20 mg, 0.018 mmol, 5 mol%) and *i*Pr₂NEt (36.8 μ L, 0.211 mmol) in DMC (2.1 mL) at r.t. for 48 h to give the crude products, which were purified by Biotage® Isolera 4 chromatography (hexane:EtOAc 80:20) to give 1-benzyl-2-oxo-3-phenylindolin-3-yl isobutyrate **S15** (65 mg, 0.169 mmol, 48%) and 1-benzyl-3-hydroxy-3-phenylindolin-2-one **19** (50 mg, 0.159 mmol, 45%).

(*R*)-1-Benzyl-2-oxo-3-phenylindolin-3-yl isobutyrate S15: Chiral HPLC analysis Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.25 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 12.1 min, t_R (*R*): 25.3 min, 6.376:93.624 er.

(S)-1-Benzyl-3-hydroxy-3-phenylindolin-2-one 19: Chiral HPLC analysis Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.25 mL min⁻¹, 211 nm, 40 °C) $t_R(R)$: 19.5 min, $t_R(S)$: 25.2 min, 0.983:99.017 er.

Cycle 6: 1-Allyl-3-hydroxy-3-(pyridin-3-yl)indolin-2-one 25



Following **General Procedure F**, 1-allyl-3-hydroxy-3-(pyridin-3-yl)indolin-2-one **25** (75.0 mg, 0.28 mmol), isobutyric anhydride (32.7 μ L, 0.197 mmol), Merrifield-HyperBTM **2** (16 mg, 0.014 mmol, 5 mol%) and *i*Pr₂NEt (29.4 μ L, 0.169 mmol) in DMC (1.7 mL) at r.t. for 48 h to give the crude products, which were purified by Biotage® Isolera 4 chromatography (Hexane:EtOAc 75:25) to give 1-allyl-2-oxo-3-(pyridin-3-yl)indolin-3-yl isobutyrate **S21** (54 mg, 0.161 mmol, 57%) and 1-allyl-3-hydroxy-3-(pyridin-3-yl)indolin-2-one **25** (29 mg, 0.109 mmol, 39%).

(S)-1-Allyl-2-oxo-3-(pyridin-3-yl)indolin-3-yl isobutyrate S21: Chiral HPLC analysis Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) $t_R(R)$: 15.2 min, t_R (S): 22.5 min, 16.134:83.866 er.

(*R*)-1-Allyl-3-hydroxy-3-(pyridin-3-yl)indolin-2-one 25: Chiral HPLC analysis Chiralpak AD-H (92:8 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 20.1 min, t_R (*R*): 22.7 min, 0.010:99.990 er.

Cycle 7: 1-Benzyl-3-hydroxy-3-phenylindolin-2-one 19



Following **General Procedure F**, 1-benzyl-3-hydroxy-3-phenylindolin-2-one **19** (88.9 mg, 0.282 mmol), isobutyric anhydride (32.7 μ L, 0.197 mmol), Merrifield-HyperBTM **2** (16 mg, 0.014 mmol, 5 mol%) and *i*Pr₂NEt (29.4 μ L, 0.169 mmol) in DMC (1.7 mL) at r.t. for 48 h to give the crude products, which were purified by Biotage® Isolera 4 chromatography

(hexane:EtOAc 80:20) to give 1-benzyl-2-oxo-3-phenylindolin-3-yl isobutyrate **S15** (54 mg, 0.140 mmol, 50%) and 1-benzyl-3-hydroxy-3-phenylindolin-2-one **19** (40 mg, 0.127 mmol, 45%).

(*R*)-1-Benzyl-2-oxo-3-phenylindolin-3-yl isobutyrate S15: Chiral HPLC analysis Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.25 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 12.1 min, t_R (*R*): 25.4 min, 7.315:92.685 er.

(S)-1-Benzyl-3-hydroxy-3-phenylindolin-2-one 19: Chiral HPLC analysis Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.25 mL min⁻¹, 211 nm, 40 °C) $t_R(R)$: 19.5 min, $t_R(S)$: 25.1 min, 0.151:99.849 er.

Cycle 8: 1-Benzyl-3-hydroxy-5-methyl-3-phenylindolin-2-one 28



Following **General Procedure F**, 1-benzyl-3-hydroxy-5-methyl-3-phenylindolin-2-one **28** (87.0 mg, 0.264 mmol), isobutyric anhydride (30.6 μ L, 0.185 mmol), Merrifield-HyperBTM **2** (15 mg, 0.013 mmol, 5 mol%) and *i*Pr₂NEt (27.6 μ L, 0.158 mmol) in DMC (1.6 mL) at r.t. for 48 h to give the crude products, which were purified by Biotage® Isolera 4 chromatography (hexane:EtOAc 80:20) to give 1-benzyl-5-methyl-2-oxo-3-phenylindolin-3-yl isobutyrate **S24** (55 mg, 0.138 mmol, 52%) and 1-benzyl-3-hydroxy-5-methyl-3-phenylindolin-2-one **28** (38 mg, 0.115 mmol, 44%).

(*R*)-1-Benzyl-5-methyl-2-oxo-3-phenylindolin-3-yl isobutyrate S24: Chiral HPLC analysis Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 11.3 min, t_R (*R*): 24.8 min, 10.583:89.417 er.

(S)-1-Benzyl-3-hydroxy-5-methyl-3-phenylindolin-2-one 28: Chiral HPLC analysis Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) $t_R(R)$: 12.4 min, t_R (S): 15.9 min, 1.127:98.873 er.

Cycle 9: 1-Benzyl-3-hydroxy-3-phenylindolin-2-one 19



Following General Procedure F, 1-benzyl-3-hydroxy-3-phenylindolin-2-one **19** (77.7 mg, 0.246 mmol), isobutyric anhydride (28.6 μ L, 0.173 mmol), Merrifield-HyperBTM **2** (14 mg, 0.012 mmol, 5 mol%) and *i*Pr₂NEt (25.8 μ L, 0.148 mmol) in DMC (1.5 mL) at r.t. for 48 h to give the crude products, which were purified by Biotage® Isolera 4 chromatography (hexane:EtOAc 80:20) to give 1-benzyl-2-oxo-3-phenylindolin-3-yl isobutyrate **S15** (50 mg, 0.13 mmol, 53%) and 1-benzyl-3-hydroxy-3-phenylindolin-2-one **19** (35 mg, 0.111 mmol, 45%).

(*R*)-1-Benzyl-2-oxo-3-phenylindolin-3-yl isobutyrate S15: Chiral HPLC analysis Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.25 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 12.1 min, t_R (*R*): 25.4 min, 7.964:92.036 er.

(S)-1-Benzyl-3-hydroxy-3-phenylindolin-2-one 19: Chiral HPLC analysis Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.25 mL min⁻¹, 211 nm, 40 °C) $t_R(R)$: 19.4 min, $t_R(S)$: 25.1 min, 0.112:99.888 er.

Cycle 10: 1-Benzyl-3-hydroxy-3-(2-methylprop-1-en-1-yl)indolin-2-one 26



Following **General Procedure F**, 1-benzyl-3-hydroxy-3-(2-methylprop-1-en-1-yl)indolin-2-one **26** (72.3 mg, 0.246 mmol), isobutyric anhydride (28.6 μ L, 0.173 mmol), Merrifield-HyperBTM **2** (14 mg, 0.012 mmol, 5 mol%) and *i*Pr₂NEt (25.8 μ L, 0.148 mmol) in DMC (1.5 mL) at r.t. for 48 h to give the crude products, which were purified by flash column chromatography (hexane:EtOAc 4:1) to give 1-benzyl-3-(2-methylprop-1-en-1-yl)-2-oxoindolin-3-yl isobutyrate

S22 (43 mg, 0.118 mmol, 48%) and 1-benzyl-3-hydroxy-3-(2-methylprop-1-en-1-yl)indolin-2-one **26** (35 mg, 0.119 mmol, 55%).

(*R*)-1-Benzyl-3-(2-methylprop-1-en-1-yl)-2-oxoindolin-3-yl isobutyrate S22: Chiral HPLC analysis Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R (*S*): 8.9 min, t_R (*R*): 17.4 min, 8.365:91.635 er

(S)-1-Benzyl-3-hydroxy-3-(2-methylprop-1-en-1-yl)indolin-2-one 26: Chiral HPLC analysis Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R (*R*): 32.9 min, t_R (S): 34.6 min, 10.526:89.474 er.

Kinetic Resolution of Tertiary Alcohols in Flow

Optimisation



Racemic

Entry ^a	Substrate	Solvent	Temp.	Anhydride (equiv.), <i>i</i> Pr ₂ NEt (equiv.)	C	S
1		DMC	10 °C	(<i>i</i> -PrCO) ₂ O (0.8 equiv.)	43	11
				<i>i</i> Pr ₂ NEt (0.8 equiv.)		
2		DMC	10 °C	(<i>i</i> -PrCO) ₂ O (1.5 equiv.)	56	9
				$i Pr_2 NEt (1.5 equiv.)$		
3	HO N Bn 14	DMC	10 °C	(<i>i</i> -PrCO) ₂ O (2.0 equiv.)	58	9
				<i>i</i> Pr ₂ NEt (2.0 equiv.)		
		EtOAc	20 °C	(<i>i</i> -PrCO) ₂ O (0.8 equiv.)	44	10
				<i>i</i> Pr ₂ NEt (0.8 equiv.)		
5		EtOAc	0 °C	(<i>i</i> -PrCO) ₂ O (0.8 equiv.)	40	17
5				<i>i</i> Pr ₂ NEt (0.8 equiv.)		
6		EtOAc	0 °C	(<i>i</i> -PrCO) ₂ O (2.0 equiv.)	63	13
				<i>i</i> Pr ₂ NEt (2.0 equiv.)		
7	HO	Toluene	20 °C	(CH ₃ CO) ₂ O (0.8 equiv.)	37	36
				<i>i</i> Pr ₂ NEt (0.8 equiv.)		
8	O N Allyl		0 °C	(CH ₃ CO) ₂ O (1.0 equiv.)	40	42
		Toluene		$iPr_2NEt (1.0 equiv)$		
	31					

^a All reactions were carried out using the same catalyst bed of 600 mg Merrifield-HyperBTM with 0.1 ml/min flow rate.

General procedure: Description of Kinetic Resolution in Continuous Flow



A packed bed reactor consisting of a vertically-mounted Omnifit glass chromatography column [borosilicate glass; length = 100 mm (70 mm adjustable bed height); internal diameter = 10 mm; maximum bed volume 5.6 mL], with a glass cooling jacket was loaded with PS-HyperBTM resin (600 mg; f = 0.89 mmol g⁻¹). The resin was allowed to swell to its maximum volume by pumping CHCl₃ at 1 µL min⁻¹ for 30 min at r.t. using a Gilson 305 HPLC pump. The column was then cooled by circulating ethylene glycol (-5 °C) using a Huber Ministat over 10 min, during which time CHCl₃ was pumped through the packed bed reactor at 100 μ L min⁻¹. Two syringes were used to inject reagents using a Legato 200 series syringe pump by World Precision Instruments. The first syringe was filled with a solution of the appropriate alcohol (0.5 mmol, 1.0 equiv.) in solvent (5 mL) and the second syringe with a mixture of the appropriate anhydride (1.0 equiv. for oxindole-substrates and 0.8 equiv. for pyrrolidinone substrates) and *i*-Pr₂NEt (1.0 equiv. for oxindole-substrates and 0.8 equiv. for pyrrolidinone substrates) in solvent (5 mL total volume). Both solutions were injected at 50 μ L min⁻¹, mixed in a T-type mixing chamber, and passed through the reactor at a combined flow rate of 100 μ L min⁻¹. After complete addition of the reagents from the syringes, a Gilson 305 HPLC pump was connected, and CHCl₃ was pumped at 100 µL min⁻¹ for 30 min to ensure elution of the products. A solution of 10% MeOH in CHCl₃ was then pumped at 200 μ L min⁻¹ for 30 min to wash the column and avoid cross contamination. The column was then prepared for the next KR by pumping the desired solvent (CHCl₃, EtOAc or PhMe) at 200 μ L min⁻¹ for 30 min. The ester product and remaining alcohol were separated by column chromatography using a Biotage® Isolera[™] 4 [SNAP Ultra 10 g, 40 mL min⁻¹, CH₂Cl₂:EtOAc (100:0 2 CV, 100:0 to 90:10 30 CV, (16 ml each))].

Packed Column Set-up and Effect of Swelling:



Continuous Flow Set-up:



3-Allyl-1-benzyl-3-hydroxyindolin-2-one 14



In CHCl₃:

(*R*)-3-Allyl-1-benzyl-2-oxoindolin-3-yl isobutyrate 15: Yield: 43%; $[\alpha]_D^{20} = -18.0$ (*c* 0.1, CHCl₃); Chiral HPLC analysis - Chiralpak OD-H (98:2 hexane:IPA, flow rate 1.0 mL min⁻¹, 254 nm, 30 °C) t_R (*R*): 8.4 min, t_R (*S*): 9.8 min, 97.369:2.631 er.

(*S*)-3-Allyl-1-benzyl-3-hydroxyindolin-2-one 14: Yield: 44%; $[\alpha]_D^{20} = -13.6$ (*c* 0.1, CHCl₃); Chiral analysis HPLC analysis - Chiralpak OD-H (98:2 hexane:IPA, flow rate 1.0 mL min⁻¹, 254 nm, 30 °C) t_R (*R*): 32.7 min, t_R (*S*): 37.9 min, 6.296:93.704 er.

In EtOAc:

(*R*)-3-Allyl-1-benzyl-2-oxoindolin-3-yl isobutyrate 15: Yield: 60%; Chiral HPLC analysis - Chiralpak OD-H (98:2 hexane:IPA, flow rate 1.0 mL min⁻¹, 254 nm, 30 °C) t_R (*R*): 32.2 min, t_R (*S*): 37.6 min, 77.959:22.041 er.

(*S*)-3-Allyl-1-benzyl-3-hydroxyindolin-2-one 14: Yield: 33%; Chiral analysis HPLC analysis - Chiralpak OD-H (98:2 hexane:IPA, flow rate 1.0 mL min⁻¹, 254 nm, 30 °C) $t_R(R)$: 32.6 min, $t_R(S)$: 37.3 min, 1.796:98.204 er.

1-Benzyl-3-hydroxy-3-phenylindolin-2-one 19



In CHCl₃:

(*R*)-1-Benzyl-2-oxo-3-phenylindolin-3-yl isobutyrate S15: Yield: 34%; $[\alpha]_D^{20} = -115.1$ (*c* 0.1, CHCl₃); Chiral HPLC analysis - Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.25 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 12.2 min, t_R (*R*): 25.6 min, 1.094:98.906 er.

(*S*)-1-Benzyl-3-hydroxy-3-phenylindolin-2-one 19: Yield: 48%; $[\alpha]_D^{20} = -35.4$ (*c* 0.1, CHCl₃); Chiral HPLC analysis - Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.25 mL min⁻¹, 211 nm, 40 °C) t_R (*R*): 19.6 min, t_R (*S*): 25.3 min, 18.301:81.699 er.

In EtOAc:

(*R*)-1-Benzyl-2-oxo-3-phenylindolin-3-yl isobutyrate S15: Yield: 48%; Chiral HPLC analysis - Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.25 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 12.2 min, t_R (*R*): 25.6 min, 4.626:95.374 er.

(S)-1-Benzyl-3-hydroxy-3-phenylindolin-2-one 19: Yield: 43%; Chiral HPLC analysis - Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.25 mL min⁻¹, 211 nm, 40 °C) t_R (*R*): 19.5 min, t_R (*S*): 25.2 min, 4.192:95.808 er.
1-Allyl-3-hydroxy-3-(pyridin-3-yl)indolin-2-one 25



In CHCl₃:

(*S*)-1-Allyl-2-oxo-3-(pyridin-3-yl)indolin-3-yl isobutyrate S21: Yield: 41%; $[\alpha]_D^{20} = -127.2$ (*c* 0.1, CHCl₃); Chiral HPLC analysis - Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*R*): 14.8 min, t_R (*S*): 21.6 min, 3.549:96.451 er.

(*R*)-1-Allyl-3-hydroxy-3-(pyridin-3-yl)indolin-2-one 25: Yield: 45%; $[\alpha]_D^{20} = -32.8$ (*c* 0.1, CHCl₃); Chiral HPLC analysis - Chiralpak AD-H (92:8 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 20.1 min, t_R (*R*): 22.7 min, 5.289:94.715 er.

In EtOAc:

(S)-1-Allyl-2-oxo-3-(pyridin-3-yl)indolin-3-yl isobutyrate S21: Yield: 54%; Chiral HPLC analysis - Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) $t_R(R)$: 14.9 min, $t_R(S)$: 21.8 min, 10.729:89.271 er.

(*R*)-1-Allyl-3-hydroxy-3-(pyridin-3-yl)indolin-2-one 25: Yield: 43%; Chiral HPLC analysis - Chiralpak AD-H (92:8 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 19.7 min, t_R (*R*): 22.2 min, 1.906:98.094 er.

1-Benzyl-7-chloro-3-hydroxy-3-methylindolin-2-one 30



In CHCl₃:

(*R*)-1-Benzyl-7-chloro-3-methyl-2-oxoindolin-3-yl isobutyrate S26: Yield: 49%; $[\alpha]_D^{20} = -21.4$ (*c* 0.1, CHCl₃); Chiral HPLC analysis - Chiralpak AD-H (99:1 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 11.3 min, t_R (*R*): 12.9 min, 8.003:91.997 er.

(*S*)-1-Benzyl-7-chloro-3-hydroxy-3-methylindolin-2-one 30: Yield: 40%; $[\alpha]_D^{20} = -35.8$ (*c* 0.1, CHCl₃); Chiral HPLC analysis - Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*R*): 11.6 min, t_R (*S*): 13.9 min, 0.784:99.216 er.

In EtOAc:

(*R*)-1-Benzyl-7-chloro-3-methyl-2-oxoindolin-3-yl isobutyrate S26: Yield: 73%; Chiral HPLC analysis - Chiralpak AD-H (99:1 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 10.9 min, t_R (*R*): 12.7 min, 34.005:65.995 er.

(S)-1-Benzyl-7-chloro-3-hydroxy-3-methylindolin-2-one 30: Yield: 24%; Chiral HPLC analysis - Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) $t_R(R)$: 11.5 min, $t_R(S)$: 13.8 min, 0.544:99.456 er.

1-Allyl-3-hydroxy-3-phenylindolin-2-one 20



In CHCl₃:

(*R*)-1-Allyl-2-oxo-3-phenylindolin-3-yl isobutyrate S16: Yield: 34%; $[\alpha]_D^{20} = -125.3$ (*c* 0.1, CHCl₃); Chiral HPLC analysis - Chiralcel OD-H (99:1 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R (*R*): 11.0 min, t_R (*S*): 12.8 min, 98.093:1.907 er.

(*S*)-1-Allyl-3-hydroxy-3-phenylindolin-2-one 20: Yield: 57%; $[\alpha]_D^{20} = -38.3$ (*c* 0.1, CHCl₃); Chiral HPLC analysis - Chiralcel OD-H (95:5 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R (*S*): 13.8 min, t_R (*R*): 15.8 min, 82.999:17.001 (er.

1-Benzyl-3-hydroxy-5-methyl-3-phenylindolin-2-one 28



In CHCl₃:

(*R*)-1-Benzyl-5-methyl-2-oxo-3-phenylindolin-3-yl isobutyrate S24: Yield: 36%; $[\alpha]_D^{20} = -107.5 \ (c \ 0.1, \text{CHCl}_3)$; Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 11.3 min, t_R (*R*): 25.1 min, 3.832:96.168 er.

(*S*)-1-Benzyl-3-hydroxy-5-methyl-3-phenylindolin-2-one 28: Yield: 56%; $[\alpha]_D^{20} = -18.4$ (*c* 0.1, CHCl₃); Chiral HPLC analysis - Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*R*): 12.4 min, t_R (*S*): 15.9 min, 17.311:82.686 er.

In EtOAc:

(*R*)-1-Benzyl-5-methyl-2-oxo-3-phenylindolin-3-yl isobutyrate S24: Yield: 41%; Chiral HPLC analysis - Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 11.2 min, t_R (*R*): 24.9 min, 5.851:94.149 er.

(*S*)-1-Benzyl-3-hydroxy-5-methyl-3-phenylindolin-2-one 28: Yield: 54%; Chiral HPLC analysis - Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) $t_R(R)$: 12.4 min, $t_R(S)$: 15.8 min, 17.547:82.453 er.

1-Allyl-3-hydroxy-3-(6-methoxynaphthalen-2-yl)indolin-2-one 22



In EtOAc:

(*R*)-1-Allyl-3-(6-methoxynaphthalen-2-yl)-2-oxoindolin-3-yl isobutyrate S18: Yield: 51%; Chiral HPLC analysis - Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 18.7 min, t_R (*R*): 33.6 min, 9.315:90.685 er.

(*S*)-1-Allyl-3-hydroxy-3-(6-methoxynaphthalen-2-yl)indolin-2-one 22: Yield: 47%; Chiral HPLC analysis - Chiralcel OJ-H (90:10 hexane:IPA, flow rate 1.5 mL min⁻¹, 254 nm, 40 °C) t_R (*R*): 35.0 min, t_R (*S*): 47.8 min, 5.178:94.822 er.

1-Allyl-3-(4-chlorophenyl)-3-hydroxypyrrolidin-2-one 31



In PhMe:

(*S*)-1-Allyl-3-(4-chlorophenyl)-2-oxopyrrolidin-3-yl acetate S27: Yield: 33%; $[\alpha]_D^{20} = -22.8$ (*c* 0.1, CHCl₃); Chiral HPLC analysis - Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R (*R*): 20.4 min, t_R (*S*): 30.2 min, 4.205:95.795 er.

(*R*)-1-Allyl-3-(4-chlorophenyl)-3-hydroxypyrrolidin-2-one 31: Yield: 57%; $[\alpha]_D^{20} = -29.2$ (*c* 0.1, CHCl₃); Chiral HPLC analysis - Chiralpak AD-H (95:5 hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) t_R (*S*): 18.8 min, t_R (*R*): 21.4 min, 19.811:80.189 er.

1-Allyl-3-hydroxy-3-(thiophen-3-yl)pyrrolidin-2-one 32



In PhMe:

(*S*)-1-Allyl-2-oxo-3-(thiophen-3-yl)pyrrolidin-3-yl acetate S28: Yield: 41%; $[\alpha]_D^{20} = +17.7$ (*c* 0.1, CHCl₃); Chiral HPLC analysis - Chiralpak AD-H (95:5 hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) t_R (*R*): 20.8 min, t_R (*S*): 23.5 min, 6.793:93.207 er.

(*R*)-1-Allyl-3-hydroxy-3-(thiophen-3-yl)pyrrolidin-2-one 32: Yield: 50%; $[\alpha]_D^{20} = -33.1$ (*c* 0.1, CHCl₃); Chiral HPLC analysis - Chiralpak AD-H (95:5 hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) t_R (*S*): 25.8 min, t_R (*R*): 32.5 min, 14.503:85.497 er.

1-Benzyl-3-hydroxy-3-phenylpyrrolidin-2-one 33



In PhMe:

(*S*)-1-Benzyl-2-oxo-3-phenylpyrrolidin-3-yl acetate S29: Yield: 46%; $[\alpha]_D^{20} = -22.5$ (*c* 0.1, CHCl₃); Chiral HPLC analysis - Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*R*): 11.8 min, t_R (*S*): 18.5 min, 9.629:90.371 er.

(*R*)-1-Benzyl-3-hydroxy-3-phenylpyrrolidin-2-one 33: Yield: 43%; $[\alpha]_D^{20} = -27.0$ (*c* 0.1, CHCl₃); Chiral HPLC analysis - Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 11.7 min, t_R (*R*): 12.7 min, 7.025:92.975 er.

Scale up in PhMe:

(S)-1-Benzyl-2-oxo-3-phenylpyrrolidin-3-yl acetate S29: Yield: 37%; $[\alpha]_D^{20} = -24.7$ (*c* 0.1, CHCl₃); Chiral HPLC analysis - Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*R*): 11.8 min, t_R (*S*): 18.5 min, 5.267:94.733 er.

(*R*)-1-Benzyl-3-hydroxy-3-phenylpyrrolidin-2-one 33: Yield: 57%; $[\alpha]_D^{20} = -15.9$ (*c* 0.1, CHCl₃); Chiral HPLC analysis - Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 11.6 min, t_R (*R*): 12.7 min, 21.021:78.975 er.

1-Allyl-3-hydroxy-3-(thiophen-2-yl)pyrrolidin-2-one 34



In PhMe:

(*R*)-1-Allyl-2-oxo-3-(thiophen-2-yl)pyrrolidin-3-yl acetate S30: Yield: 48%; $[\alpha]_D^{20} = +11.6$ (*c* 0.1, CHCl₃); Chiral HPLC analysis - Chiralpak OD-H (95:5 hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) t_R (*S*): 20.9 min, t_R (*R*): 28.9 min, 10.279:89.721 er.

(*S*)-1-Allyl-3-hydroxy-3-(thiophen-2-yl)pyrrolidin-2-one 34: Yield: 41%; $[\alpha]_D^{20} = -50.1$ (*c* 0.1, CHCl₃); Chiral HPLC analysis - Chiralpak AD-H (98:2 hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) t_R (*R*): 58.9 min, t_R (*S*): 65.7 min, 6.365:93.635 er.

3-Hydroxy-1,3-diphenylpyrrolidin-2-one 35



In PhMe:

(*S*)-2-Oxo-1,3-diphenylpyrrolidin-3-yl acetate S31: Yield: 45%; $[\alpha]_D^{20} = +26.6 (c \ 0.1, \text{CHCl}_3)$; Chiral HPLC analysis - Chiralpak AD-H (90:10 hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) t_R (*R*): 32.3 min, t_R (*S*): 38.2 min, 9.153:90.847 er.

(*R*)-3-Hydroxy-1,3-diphenylpyrrolidin-2-one 35: Yield: 42%; $[\alpha]_D^{20} = -129.1(c \ 0.1, \text{ CHCl}_3)$; Chiral HPLC analysis - Chiralpak AD-H (95:5 hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) t_R (*S*): 19.2 min, t_R (*R*): 26.8 min, 5.717:94.283 er.

1-Allyl-3-hydroxy-3-(4-methoxyphenyl)pyrrolidin-2-one 36



In PhMe:

(*S*)-1-Allyl-3-(4-methoxyphenyl)-2-oxopyrrolidin-3-yl acetate S32: Yield: 44%; $[\alpha]_D^{20} = -13.6 (c \ 0.1, \text{CHCl}_3)$; Chiral HPLC analysis - Chiralpak AD-H (95:5 hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) t_R (*R*): 31.8 min, t_R (*S*): 45.5 min, 12.686:87.314 er.

(*R*)-1-Allyl-3-hydroxy-3-(4-methoxyphenyl)pyrrolidin-2-one 36: Yield: 40%; $[\alpha]_D^{20} = -39.5$ (*c* 0.1, CHCl₃); Chiral HPLC analysis - Chiralpak AD-H (95:5 hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) t_R (*S*): 38.5 min, t_R (*R*): 42.3 min, 8.165:91.835 er.

NMR Spectra





















220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 f1 (ppm)







S93











S97





220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 f1(ppm)



S100



220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 f1 (ppm)

HPLC Spectra

Reaction Optimisation for Secondary alcohol (Table 1, entry 15):

1-(Naphthalen-2-yl)ethyl isobutyrate 8



HPLC data for 8:Chiralcel OJ-H (98:2 hexane:IPA, flow rate 1 mLmin⁻¹, 254 nm, 30 °C)) t_R (*R*): 9.8 min, t_R (*S*): 12.2 min, 4.364:95.636 er.



(R)-1-(Naphthalen-2-yl)ethan-1-ol 7



HPLC data for 7.Chiralcel OJ-H (90:10 hexane:IPA, flow rate 1 mLmin⁻¹, 254 nm, 30 °C)) t_R (*S*): 12.2 min, t_R (*R*): 19.5 min, 12.474:87.526 er.





PDA Ch3 254nm

Peak#	Ret. Time	Area%
1	15.241	12.474
2	19.485	87.526
Total		100.000

Recyclability of Merrifield-HBTM (2), TentaGel-HBTM (3), Wang-HBTM (4) and Merrifield-BTM (6) for the KR of 5 selected secondary alcohols:

With Merrifield-HBTM (2); cycle 1 to cycle 5:

2-Azido-1-(naphthalen-2-yl)ethyl isobutyrate S7



HPLC data for S7: Chiralcel AD-H (99.8:0.2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (*S*): 14.3 min, t_R (*R*): 25.5 min, 10.236:89.764 er.



2-Azido-1-(naphthalen-3-yl)ethan-1-ol 9



HPLC data for 9: Chiralcel OD-H (95:5 hexane:IPA, flow rate 1 mLmin⁻¹, 220 nm, 30 °C) t_R (*R*): 31.1 min, t_R (*S*): 35.4 min, 6.124:93.876 er.







4-Phenylbut-3-yn-2-yl isobutyrate S8



HPLC data for S8: Following hydrolysis to alcohol. Chiralcel OD-H (95:5 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) $t_R(R)$: 11.2 min, $t_R(S)$: 28.7 min, 19.002:80.998 er.





HPLC data for 10: Chiralcel OD-H (95:5 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (*R*): 11.1 min, t_R (*S*): 27.7 min, 88.823:11.177 er.




(1*S*,2*R*)-2-Phenylcyclopentyl isobutyrate **S9**



HPLC data for S9: Following hydrolysis to alcohol. Chiralcel OD-H (98:2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (1*S*,2*R*): 14.4 min, t_R (1*R*,2*S*): 16.1 min, 90.046:9.954 er.





HPLC data for 11: Chiralcel OD-H (98:2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (1*S*,2*R*): 14.4 min, t_R (1*R*,2*S*): 16.0 min, 4.997:95.003 er.





1-(Naphthalen-2-yl)allyl isobutyrate S10



HPLC data for S10: Chiralcel OJ-H (95:5 hexane:IPA, flow rate 0.5 mLmin⁻¹, 211 nm, 30 °C) $t_R(R)$: 15.6 min, $t_R(S)$: 22.6 min, 12.460:87.540 er.





1-(Naphthalen-2-yl)prop-2-en-1-ol 12



HPLC data for 12: Chiralcel OJ-H (80:20 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (*S*): 11.4 min, t_R (*R*): 13.7 min, 15.133:84.867 er.



2-Methyl-1-(naphthalen-2-yl)propyl isobutyrate S11



HPLC data for S11: Chiralcel AD-H (99.8:0.2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) $t_R(R)$: 6.9 min, $t_R(S)$: 8.6 min, 3.935:96.065 er.





rean#	iver. mine	Alea /0
1	6.935	3.935
2	8.610	96.065
Total		100.000

2-Methyl-1-(naphthalen-2-yl)propan-1-ol 13



HPLC data for 13: Chiralcel OD-H (95:5 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (*S*): 14.1 min, t_R (*R*): 15.9 min, 0.301:99.699 er.





i cuit#	rtet. Thine	7400470
1	14.094	0.301
2	15.927	99.699
Total		100.000

With TentaGel-HBTM (3)

2-Azido-1-(naphthalen-2-yl)ethyl isobutyrate S7



HPLC data for S7: Chiralcel AD-H (99.8:0.2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (*S*): 13.4 min, t_R (*R*): 21.4 min, 8.747:91.253 er.



2-Azido-1-(naphthalen-3-yl)ethan-1-ol 9



HPLC data for 9: Chiralcel OD-H (95:5 hexane:IPA, flow rate 1 mLmin⁻¹, 220 nm, 30 °C)) t_R (*R*): 30.2 min, t_R (*S*): 34.2 min, 14.640:85.360 er.





4-Phenylbut-3-yn-2-yl isobutyrate S8



HPLC data for S8: Following hydrolysis to ester. Chiralcel OD-H (95:5 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) $t_R(R)$: 11.1 min, $t_R(S)$: 27.6 min, 21.073:78.9 27 er.





HPLC data for 10: Chiralcel OD-H (95:5 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (*S*): 11.1 min, t_R (*R*): 27.7 min, 81.202:18.798 er.





(1*S*,2*R*)-2-Phenylcyclopentyl isobutyrate **S9**



HPLC data for S9: Following hydrolysis to ester. Chiralcel OD-H (98:2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (1*S*,2*R*): 14.4 min, t_R (1*R*,2*S*): 16.1 min, 92.331:7.7669 er.





HPLC data for 11: Chiralcel OD-H (98:2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (1*S*,2*R*): 14.4 min, t_R (1*R*,2*S*): 16.1 min, 10.359:89.641 er.





1-(Naphthalen-2-yl)allyl isobutyrate S10



HPLC data for S10: Chiralcel OJ-H (95:5 hexane:IPA, flow rate 0.5 mLmin⁻¹, 211 nm, 30 °C) $t_R(R)$: 15.6 min, $t_R(S)$: 22.5 min, 10.258:89.742 er.





1-(Naphthalen-2-yl)prop-2-en-1-ol 12



HPLC data for 12: Chiralcel OJ-H (80:20 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (*S*): 11.4 min, t_R (*R*): 13.7 min, 15.227:84.773 er.





2-Methyl-1-(naphthalen-2-yl)propyl isobutyrate S11



HPLC data for S11: Chiralcel AD-H (99.8:0.2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) $t_R(R)$: 6.7 min, $t_R(S)$: 8.2 min, 1.279:98.721 er.





PDA Ch1 211nm

Peak#	Ret. Time	Area%
1	6.733	1.279
2	8.245	98.721
Total		100.000

2-Methyl-1-(naphthalen-2-yl)propan-1-ol 13



HPLC data for 13: Chiralcel OD-H (95:5 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (*S*): 13.9 min, t_R (*R*): 15.6 min, 12.970:87.030 er.





Peak#	Ret. Time	Area%
1	13.943	12.970
2	15.626	87.030
Total		100.000

With Wang-HBTM (4)

2-Azido-1-(naphthalen-2-yl)ethyl isobutyrate S7



HPLC data for S7: Chiralcel AD-H (99.8:0.2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) $t_R(S)$: 13.4 min, $t_R(R)$: 21.6 min, 8.220:91.780 er.



2-Azido-1-(naphthalen-3-yl)ethan-1-ol 9



HPLC data for 9: Chiralcel OD-H (95:5 hexane:IPA, flow rate 1 mLmin⁻¹, 220 nm, 30 °C) t_R (*R*): 30.3 min, t_R (*S*): 34.0 min, 8.421:91.579 er.



	30.277	0.421
2	33.956	91.579
Total		100.000

4-Phenylbut-3-yn-2-yl isobutyrate S8



HPLC data for S8: Following hydrolysis to alcohol. Chiralcel OD-H (95:5 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) $t_R(R)$: 11.2 min, $t_R(S)$: 27.8 min, 13.368:86.6 32 er.





HPLC data for 10: Chiralcel OD-H (95:5 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (*S*): 11.1 min, t_R (*R*): 27.7 min, 88.132:11.868 er.





(1*S*,2*R*)-2-Phenylcyclopentyl isobutyrate **S9**



HPLC data for S9: Following hydrolysis to alcohol. Chiralcel OD-H (98:2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (1*S*,2*R*): 14.4 min, t_R (1*R*,2*S*): 16.1 min, 92.068:7.932 er.





	/III <u>~ I IIIII</u>	
Peak#	Ret. Time	Area%
1	14.389	92.068
2	16.053	7.932
Total		100.000



HPLC data for 11: Chiralcel OD-H (98:2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (1*S*,2*R*): 14.5 min, t_R (1*R*,2*S*): 16.1 min, 9.266:90.734 er.





1-(Naphthalen-2-yl)allyl isobutyrate S10



HPLC data for S10: Chiralcel OJ-H (95:5 hexane:IPA, flow rate 0.5 mLmin⁻¹, 211 nm, 30 °C) $t_R(R)$: 15.6 min, $t_R(S)$: 22.6 min, 11.374:88.626 er.





100.000

Tota

30.0 min 1-(Naphthalen-2-yl)prop-2-en-1-ol 12



HPLC data for 12: Chiralcel OJ-H (80:20 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (*S*): 11.5 min, t_R (*R*): 13.7 min, 15.382:84.618 er.





Peak#	Ret. Time	Area%
1	11.451	15.382
2	13.732	84.618
Total		100.000

2-Methyl-1-(naphthalen-2-yl)propyl isobutyrate S11



HPLC data for S11: Chiralcel AD-H (99.8:0.2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) $t_R(R)$: 7.2 min, $t_R(S)$: 9.2 min, 1.037:98.963 er.





PDA Ch1 211nm

Peak#	Ret. Time	Area%
1	7.186	1.037
2	9.188	98.963
Total		100.000

2-Methyl-1-(naphthalen-2-yl)propan-1-ol 13



HPLC data for 13: Chiralcel OD-H (95:5 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (*S*): 14.1 min, t_R (*R*): 15.7 min, 12.201:87.799 er.





PDA Ch1 211nm		
Peak#	Ret. Time	Area%
1	14.051	12.201
2	15.736	87.799
Total		100.000

With Merrifield-BTM (6)

2-Azido-1-(naphthalen-2-yl)ethyl isobutyrate S7



HPLC data for S7: Chiralcel AD-H (99.8:0.2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (*S*): 15.1 min, t_R (*R*): 25.6 min, 69.814:30.186 er.



100.000

Total

2-Azido-1-(naphthalen-3-yl)ethan-1-ol 9



HPLC data for 9: Chiralcel OD-H (95:5 hexane:IPA, flow rate 1 mLmin⁻¹, 220 nm, 30 °C) t_R (*R*): 30.6 min, t_R (*S*): 35.1 min, 53.792:46.208 er.





1	30.586	53.792
2	35.076	46.208
Total		100.000

Reaction Optimisation for Tertiary heterocyclic alcohol:



Chiralcel OD-H (98:2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) - **Racemic ester:** t_R (*R*): 8.2 min, t_R (*S*): 10.0 min



Chiralcel OD-H (98:2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) - **Racemic alcohol:** t_R (*R*): 32.7 min, t_R (*S*): 38.4 min



Реак#	Ret. Time	Area%
1	32.656	49.922
2	38.440	50.078
Total		100.000

Table 3 entry 6:

HPLC data for 15: Chiralcel OD-H (98:2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C). **ester:** $t_R(R)$: 8.3 min, $t_R(S)$: 10.0 min, 90.188:9.812 (*R*:*S*) er.



HPLC data for 14: Chiralcel OD-H (98:2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C); **alcohol**: $t_R(R)$: 32.5 min, $t_R(S)$: 37.6 min, 13.247:86.753 (*R*:*S*) er



Table 3 entry 9:

HPLC data for 15: Chiralcel OD-H (98:2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C); ester: $t_R(R)$: 8.3 min, $t_R(S)$: 10.1 min, 87.773:12.227 (*R*:S) er.



HPLC data for 14: Chiralcel OD-H (98:2 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C); **alcohol**: $t_R(R)$: 32.6 min, $t_R(S)$: 37.8 min, 6.938:93.062 (*R*:*S*) er



Table 4

tert-butyl 3-allyl-3-(isobutyryloxy)-2-oxoindoline-1-carboxylate S12



Chiralcel OD-H (99:1 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C)

Racemic ester: $t_R(R)$: 4.6 min, $t_R(S)$: 7.3 min mAU



HPLC data for S12: Chiralcel OD-H (99:1 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C) t_R (*R*): 4.7 min, t_R (*S*): 7.6 min, 77.241:22.759 (*R*:*S*) er.



<Peak Table>

PDA C	h1 211nm	
Peak#	Ret. Time	Area%
1	4.688	77.241
2	7.586	22.759
Total		100.000





HPLC data for 16: Chiralcel AD-H (95:5 hexane:IPA, flow rate 1 mLmin⁻¹, 211 nm, 30 °C)) t_R (*R*): 10.7 min, t_R (*S*): 13.4 min, 1.048:98.952 (*R*:*S*) er.



<Peak Table>

PDA Ch1 211nm			
Peak#	Ret. Time	Area%	
1	10.706	1.048	
2	13.402	98.952	
Total		100.000	

1-Benzyl-3-isopropyl-2-oxoindolin-3-yl isobutyrate S13



HPLC data for S13: Chiralpak AD-H (98:2 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 8.2 min, t_R (*R*): 10.2 min, 12.511:87.489 er.



1-Benzyl-3-hydroxy-3-isopropyl-indolin-2-one 17



HPLC data for 17: Chiralpak AD-H (98:2 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) $t_R(R)$: 38.4 min, $t_R(S)$: 46.9 min, 19.578:80.422 (*S:R*) er.



PDA Ch1 211nm			
Peak#	Ret. Time	Area%	
1	38.428	19.578	
2	46.929	80.422	
Total		100.000	

1-Benzyl-2-oxo-3-(trifluoromethyl)indolin-3-yl isobutyrate S14



HPLC data for S14: Chiralpak AD-H (99:1 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) $t_R(S)$: 12.7 min, $t_R(R)$: 15.3 min, 19.991:80.009 er.


1-Benzyl-3-hydroxy-3-(trifluoromethyl)indolin-2-one 18



HPLC data for 18: Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) $t_R(R)$: 15.8 min, $t_R(S)$: 20.5 min, 0.305:99.695 er.



1-Benzyl-2-oxo-3-phenylindolin-3-yl isobutyrate S15



HPLC data for S15: Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.25 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 12.2 min, t_R (*R*): 25.6 min, 4.968:95.032 (*S*:*R*) er.



1-Benzyl-3-hydroxy-3-phenylindolin-2-one 19



HPLC data for 19: Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.25 mL min⁻¹, 211 nm, 40 °C) $t_R(R)$: 19.7 min, $t_R(S)$: 25.5 min, 3.556:96.444 (*R*:S) er.



1-Allyl-2-oxo-3-phenylindolin-3-yl isobutyrate S16



HPLC data for S16: Chiralcel OD-H (99:1 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) $t_R(R)$: 11.1 min, $t_R(S)$: 12.8 min, 92.201:7.799 (*R*:*S*) er.



Peak#	Ret. Time	Area%
1	11.149	92.201
2	12.810	7.799
Total		100.000

1-Allyl-3-hydroxy-3-phenylindolin-2-one 20



HPLC data for 20: Chiralcel OD-H (95:5 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R (*S*): 13.9 min, t_R (*R*): 16.0 min, 99.852:0.148 (*S*:*R*) er.



1-Methyl-2-oxo-3-phenylindolin-3-yl isobutyrate S17



HPLC data for S17: Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) $t_R(S)$: 8.9 min, $t_R(R)$: 12.2 min, 5.919:94.081 er.



3-Hydroxy-1-methyl-3-phenylindolin-2-one 21



HPLC data for 21: Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) $t_R(R)$: 27.9 min, $t_R(S)$: 31.0 min, 0.994:99.006 er.



1-Allyl-3-(6-methoxynaphthalen-2-yl)-2-oxoindolin-3-yl isobutyrate S18



HPLC data for S18: Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 18.7 min, t_R (*R*): 33.6 min, 12.592:87.408 er.



100.000

Total

1-Allyl-3-hydroxy-3-(6-methoxynaphthalen-2-yl)indolin-2-one 22



HPLC data for 22: Chiralcel OJ-H (90:10 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) $t_R(R)$: 34.7 min, $t_R(S)$: 47.7 min, 0.451:99.549 er.





Peak#	Ret. Time	Area%
1	34.678	0.451
2	47.703	99.549
Total		100.000

1-Allyl-3-(3,5-bis(trifluoromethyl)phenyl)-3-hydroxyindolin-2-one **23**(following hydrolysis of the ester, **1-allyl-3-(3,5-bis(trifluoromethyl)phenyl)-2-oxoindolin-3-yl isobutyrate S19**)



HPLC data for S19: Chiralpak AS-H (99:1 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) $t_R(R)$: 6.9 min, $t_R(S)$: 27.5 min, 81.747:18.253 er.



FUAC	4112111111	
Peak#	Ret. Time	Area%
1	6.857	81.747
2	27.531	18.253
Total		100.000

1-Allyl-3-(3,5-bis(trifluoromethyl)phenyl)-3-hydroxyindolin-2-one 23



HPLC data for 23: Chiralpak AS-H (99:1 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) $t_R(R)$: 7.1 min, $t_R(S)$: 28.7 min, 5.053:94.947 (*R*:*S*) er.



1-Allyl-2-oxo-3-(thiophen-2-yl)indolin-3-yl isobutyrate S20



HPLC data for S20: Chiralcel OD-H (99:1 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) $t_R(R)$: 13.1 min, $t_R(S)$: 17.0 min, 87.916:12.084 (*R*:*S*) er.



1-Allyl-3-hydroxy-3-(thiophen-2-yl)indolin-2-one 24



HPLC data for 24: Chiralcel OD-H (95:5 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R (*S*): 17.8 min, t_R (*R*): 22.5 min, 99.254:0.746 (*S*:*R*) er.





PDA Ch1 211nm			
Peak#	Ret. Time	Area%	
1	17.838	99.254	
2	22.539	0.746	
Total		100.000	

1-Allyl-2-oxo-3-(pyridin-3-yl)indolin-3-yl isobutyrate S21



HPLC data for S21: Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) $t_R(R)$: 14.8 min, $t_R(S)$: 21.8 min, 15.803:84.197 er.



Peak#	Ret. Time	Area%
1	14.830	15.803
2	21.791	84.197
Total		100.000

1-Allyl-3-hydroxy-3-(pyridin-3-yl)indolin-2-one 25



HPLC data for 25: Chiralpak AD-H (92:8 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 20.1 min, t_R (*R*): 22.7 min, 0.586:99.414 er.





PD/	Ą	Ch1	21	1	۱m
_				_	

Peak#	Ret. Lime	Area%
1	20.060	0.586
2	22.670	99.414
Total		100.000

1-Benzyl-3-(2-methylprop-1-en-1-yl)-2-oxoindolin-3-yl isobutyrate S22



HPLC data for S22: Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R (*S*): 8.9 min, t_R (*R*): 17.2 min, 7.530:92.470 er.



-eak#	Ret. Time	Area%	
1	8.937	7.530	
2	17.214	92.470	
Total		100.000	

1-Benzyl-3-hydroxy-3-(2-methylprop-1-en-1-yl)indolin-2-one 26



HPLC data for 26: Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) $t_R(R)$: 32.7 min, $t_R(S)$: 34.7 min, 16.801:83.199 er.





Peak#	Ret. Time	Area%
1	32.698	16.801
2	34.653	83.199
Total		100.000

1-Benzyl-4-chloro-3-methyl-2-oxoindolin-3-yl isobutyrate S23



HPLC data for S23: Chiralpak AD-H (98:2 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) $t_R(S)$: 8.6 min, $t_R(R)$: 10.3 min, 10.001:89.999 (*S:R*) er.



Peak#	Ret. Time	Area%
1	8.643	10.001
2	10.332	89.999
Total		100.000

1-Benzyl-4-chloro-3-hydroxy-3-methylindolin-2-one 27



HPLC data for 27: Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) $t_R(R)$: 17.6 min, $t_R(S)$: 20.3 min, 21.819:78.181 (*R*:*S*) er.





PDA Ch1 211nm			
Peak#	Ret. Time	Area%	
1	17.635	21.819	
2	20.321	78.181	
Total		100.000	

23 min 1-Benzyl-5-methyl-2-oxo-3-phenylindolin-3-yl isobutyrate S24



HPLC data for S24: Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 11.3 min, t_R (*R*): 25.2 min, 10.298:89.702 er.



PDA Ch1 211nm			
Peak#	Ret. Time	Area%	
1	11.331	10.298	
2	25.196	89.702	
Total		100.000	

1-Benzyl-3-hydroxy-5-methyl-3-phenylindolin-2-one 28



HPLC data for 28: Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) $t_R(R)$: 12.4 min, $t_R(S)$: 15.9 min, 4.610:95.390 (*R*:*S*) er.



PDA Ch1 211nm			
Peak#	Ret. Time	Area%	
1	12.446	4.610	
2	15.891	95.390	
Total		100.000	

1-Benzyl-6-chloro-2-oxo-3-phenylindolin-3-yl isobutyrate **S25**



HPLC data for S25: Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) $t_R(S)$: 7.9 min, $t_R(R)$: 15.5 min, 20.665:79.335 er.



1-Benzyl-6-chloro-3-hydroxy-3-phenylindolin-2-one 29



HPLC data for 29: Chiralpak AD-H (92:8 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) $t_R(R)$: 13.8 min, $t_R(S)$: 15.9 min, 18.545:81.455 er.



Detector A Channel 1 211nm

Peak#	Ret. Time	Area%
1	14.138	50.005
2	16.324	49.995
Total		100.000



 PDA Ch1 211nm

 Peak# Ret. Time
 Area%

 1
 13.817
 18.545

 2
 15.963
 81.455

 Total
 100.000

1-Benzyl-7-chloro-3-methyl-2-oxoindolin-3-yl isobutyrate S26



HPLC data for S26: Chiralpak AD-H (99:1 hexane:IPA, flow rate 1.5 mL min⁻¹, 254 nm, 40 °C) t_R (*S*): 11.2 min, t_R (*R*): 12.9 min, 24.370:75.630 (*S*:*R*) er.



1-Benzyl-7-chloro-3-hydroxy-3-methylindolin-2-one 30



HPLC data for 30: Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) $t_R(R)$: 11.6 min, $t_R(S)$: 13.8 min, 9.044:90.956 (*R*:S) er.



Recyclability of catalyst for the KR of tertiary heterocyclic alcohols

Cycle 1: 1-Benzyl-2-oxo-3-phenylindolin-3-yl isobutyrate S15



HPLC data for S15: Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.25 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 12.1 min, t_R (*R*): 25.5 min, 10.228:89.772 er.



1-Benzyl-3-hydroxy-3-phenylindolin-2-one 19



HPLC data for 19: Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.25 mL min⁻¹, 211 nm, 40 °C) $t_R(R)$: 19.5 min, $t_R(S)$: 25.1 min, 0.088:99.912 er.



PDA Ch1 211nm				
Peak#	Ret. Time	Area%		
1	19.517	0.088		
2	25.109	99.912		
Total		100.000		

Cycle 2:

1-Allyl-2-oxo-3-phenylindolin-3-yl isobutyrate S16



HPLC data for S16: Chiralcel OD-H (99:1 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) $t_R(R)$: 11.6 min, $t_R(S)$: 13.6 min, 92.344:7.656 er.



1-Allyl-3-hydroxy-3-phenylindolin-2-one 20



HPLC data for 20: Chiralcel OD-H (95:5 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R (*S*): 13.9 min, t_R (*R*): 15.9 min, 99.852:0.148 er.





PDA Ch1 211nm		
Peak#	Ret. Time	Area%
1	13.926	99.852
2	15.936	0.148
Total		100.000

Cycle 3:

1-Benzyl-2-oxo-3-phenylindolin-3-yl isobutyrate S15



HPLC data for S15: Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.25 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 12.1 min, t_R (*R*): 25.4 min, 7.797:92.203 (*S*:*R*) er.



1-Benzyl-3-hydroxy-3-phenylindolin-2-one 19



HPLC data for 19: Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.25 mL min⁻¹, 211 nm, 40 °C) $t_R(R)$: 19.4 min, $t_R(S)$: 25.1 min, 0.276:99.724 er.



Peak#	Ret. Time	Area%
1	19.427	0.276
2	25.100	99.724
Total		100.000

Cycle 4:

1-Methyl-2-oxo-3-phenylindolin-3-yl isobutyrate S17



HPLC data for S17: Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) $t_R(S)$: 8.9 min, $t_R(R)$: 12.2 min, 5.178:94.822 er.



3-Hydroxy-1-methyl-3-phenylindolin-2-one 21



HPLC data for 21: Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) $t_R(R)$: 27.8 min, $t_R(S)$: 30.9 min, 0.718:99.282 er.



Cycle 5:

1-Benzyl-2-oxo-3-phenylindolin-3-yl isobutyrate **S15**



HPLC data for S15: Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.25 mL min⁻¹, 211 nm, 40 °C) $t_R(S)$: 12.1 min, $t_R(R)$: 25.3 min, 6.376:93.624 er.





1-Benzyl-3-hydroxy-3-phenylindolin-2-one 19



HPLC data for 19: Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.25 mL min⁻¹, 211 nm, 40 °C) $t_R(R)$: 19.5 min, $t_R(S)$: 25.2 min, 0.983:99.017 (*R*:S) er.







1-Allyl-2-oxo-3-(pyridin-3-yl)indolin-3-yl isobutyrate S21



HPLC data for S21: Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) $t_R(R)$: 15.2 min, $t_R(S)$: 22.5 min, 16.134:83.866 er.




1-Allyl-3-hydroxy-3-(pyridin-3-yl)indolin-2-one 25



HPLC data for 25: Chiralpak AD-H (92:8 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) $t_R(S)$: 20.1 min, $t_R(R)$: 22.7 min, 0.010:99.990 er.





1-Benzyl-2-oxo-3-phenylindolin-3-yl isobutyrate **S15**



HPLC data for S15: Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.25 mL min⁻¹, 211 nm, 40 °C) $t_R(S)$: 12.1 min, $t_R(R)$: 25.4 min, 7.315:92.685 er.



1-Benzyl-3-hydroxy-3-phenylindolin-2-one 19



HPLC data for 19: Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.25 mL min⁻¹, 211 nm, 40 °C) $t_R(R)$: 19.5 min, $t_R(S)$: 25.1 min, 0.151:99.849 er.







1-Benzyl-5-methyl-2-oxo-3-phenylindolin-3-yl isobutyrate S24



HPLC data for S24: Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 11.3 min, t_R (*R*): 24.8 min, 10.583:89.417 er.



1-Benzyl-3-hydroxy-5-methyl-3-phenylindolin-2-one 28



HPLC data for 28: Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) $t_R(R)$: 12.4 min, $t_R(S)$: 15.9 min, 1.127:98.873 er.







1-Benzyl-2-oxo-3-phenylindolin-3-yl isobutyrate S15



HPLC data for S15: Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.25 mL min⁻¹, 211 nm, 40 °C) $t_R(S)$: 12.1 min, $t_R(R)$: 25.4 min, 7.964:92.036 er.



1-Benzyl-3-hydroxy-3-phenylindolin-2-one 19



HPLC data for 19: Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.25 mL min⁻¹, 211 nm, 40 °C) $t_R(R)$: 19.4 min, $t_R(S)$: 25.1 min, 0.112:99.888 er.



PDA Ch1 211nm			
Peak#	Ret. Time	Area%	
1	19.485	50.059	
2	25.210	49.941	
Total		100.000	



Cycle 10:

 $1\mbox{-}Benzyl-3\mbox{-}(2\mbox{-}methylprop-1\mbox{-}en-1\mbox{-}yl)\mbox{-}2\mbox{-}oxoindolin-3\mbox{-}yl isobutyrate {\bf S22}$



HPLC data for S22: Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R (*S*): 8.9 min, t_R (*R*): 17.4 min, 8.365:91.635 er.



1-Benzyl-3-hydroxy-3-(2-methylprop-1-en-1-yl)indolin-2-one 26



HPLC data for 26: Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) $t_R(R)$: 32.9 min, $t_R(S)$: 34.6 min, 10.526:89.474 er.





Flow HPLCs - KRs in CHCl₃

3-Allyl-1-benzyl-2-oxoindolin-3-yl isobutyrate 15







3-Allyl-1-benzyl-3-hydroxyindolin-2-one 14



Chiralpak OD-H (98:2 hexane:IPA, flow rate 1.0 mL min⁻¹, 254 nm, 30 °C) $t_R(R)$: 32.7 min, $t_R(S)$: 37.9 min, 6.296:93.704 er.



1-Benzyl-2-oxo-3-phenylindolin-3-yl isobutyrate S15





Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.25 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 12.2 min, t_R (*R*): 25.6 min, 1.094:98.906 er.

1-Benzyl-3-hydroxy-3-phenylindolin-2-one 19



Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.25 mL min⁻¹, 211 nm, 40 °C) t_R (*R*): 19.6 min, t_R (*S*): 25.319 min, 18.301:81.699 er.



Total	100.000	
3000		
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_		



1-Allyl-2-oxo-3-(pyridin-3-yl)indolin-3-yl isobutyrate S21



PDA Multi 1 211nm,4nm



Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*R*): 14.8 min, t_R (*S*): 21.6 min, 3.342:96.658 er.

1-Allyl-3-hydroxy-3-(pyridin-3-yl)indolin-2-one 25



Chiralpak AD-H (92:8 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 20.1 min, t_R (*R*): 22.7 min, 5.429:94.571 er.



1-Benzyl-7-chloro-3-methyl-2-oxoindolin-3-yl isobutyrate S26





Chiralpak AD-H (99:1 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 11.3 min, t_R (*R*): 12.9 min, 8.003:91.997 er.

1-Benzyl-7-chloro-3-hydroxy-3-methylindolin-2-one **30**







1-Allyl-2-oxo-3-phenylindolin-3-yl isobutyrate S16



Chiralcel OD-H (99:1 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) $t_R(R)$: 11.0 min, $t_R(S)$: 12.8 min, 98.093:1.907 er.



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PDA Ch1 211nm			
Peak#	Ret. Time	Area%	
1	11.176	98.516	
2	12.941	1.484	
Total		100.000	

1-Allyl-3-hydroxy-3-phenylindolin-2-one 20



Chiralcel OD-H (95:5 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) t_R (*S*): 13.8 min, t_R (*R*): 15.8 min, 82.999:17.001 er.



1-Benzyl-5-methyl-2-oxo-3-phenylindolin-3-yl isobutyrate S24



Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 11.3 min, t_R (*R*): 25.1 min, 3.832:96.168 er.



1-Benzyl-3-hydroxy-5-methyl-3-phenylindolin-2-one 28



Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) $t_R(R)$: 12.4 min, $t_R(S)$: 15.9 min, 17.3:82.686 er.



Flow HPLCs - KRs in EtOAc

3-Allyl-1-benzyl-2-oxoindolin-3-yl isobutyrate **S15** (Ester hydrolyzed to alcohol for HPLC analysis)



Chiralpak OD-H (98:2 hexane:IPA, flow rate 1.0 mL min⁻¹, 254 nm, 30 °C) $t_R(R)$: 32.2 min, $t_R(S)$: 37.6 min, 77.959:22.041 er.



100.000

Total

3-Allyl-1-benzyl-3-hydroxyindolin-2-one 16



Chiralpak OD-H (98:2 hexane:IPA, flow rate 1.0 mL min⁻¹, 254 nm, 30 °C) $t_R(R)$: 32.6 min, $t_R(S)$: 37.3 min, 1.796:98.204 er.





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Peak#	Ret. Time	Area%
1	32.556	1.796
2	37.344	98.204
Total		100.000

1-Benzyl-2-oxo-3-phenylindolin-3-yl isobutyrate S15



Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.25 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 12.2 min, t_R (*R*): 25.6 min, 4.626:95.374 er.



1-Benzyl-3-hydroxy-3-phenylindolin-2-one 19



Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.25 mL min⁻¹, 211 nm, 40 °C) t_R (*R*): 19.5 min, t_R (*S*): 25.2 min, 4.192:95.808 er.



1-Allyl-2-oxo-3-(pyridin-3-yl)indolin-3-yl isobutyrate S21



Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) $t_R(R)$: 14.9 min, $t_R(S)$: 21.8 min, 10.729:89.271 er.





PDA Ch1 211nm		
Peak#	Ret. Time	Area%
1	14.887	10.729
2	21.757	89.271
Total		100.000

1-Allyl-3-hydroxy-3-(pyridin-3-yl)indolin-2-one 25



Chiralpak AD-H (92:8 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 19.7 min, t_R (*R*): 22.2 min, 1.906:98.094 er.



100.000

Total

1-Benzyl-7-chloro-3-methyl-2-oxoindolin-3-yl isobutyrate S26



Chiralpak AD-H (99:1 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 10.9 min, t_R (*R*): 12.7 min, 34.005:65.995 er.



1-Benzyl-7-chloro-3-hydroxy-3-methylindolin-2-one 30



Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) $t_R(R)$: 11.5 min, $t_R(S)$: 13.8 min, 0.544:99.456 er.



2	13.786	99.456	
Total		100.000	

1-Benzyl-5-methyl-2-oxo-3-phenylindolin-3-yl isobutyrate S24



Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 11.2 min, t_R (*R*): 24.9 min, 5.851:94.149 er.



1-Benzyl-3-hydroxy-5-methyl-3-phenylindolin-2-one 28



Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) $t_R(R)$: 12.4 min, $t_R(S)$: 15.8 min, 17.547:82.453 er.





 PDA Ch1 211nm

 Peak#
 Ret. Time
 Area%

 1
 12.362
 17.547

 2
 15.799
 82.453

 Total
 100.000

1-Allyl-3-(6-methoxynaphthalen-2-yl)-2-oxoindolin-3-yl isobutyrate S18



Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 18.7 min, t_R (*R*): 33.6 min, 9.315:90.685 er.



1-Allyl-3-hydroxy-3-(6-methoxynaphthalen-2-yl)indolin-2-one 22



Chiralcel OJ-H (90:10 hexane:IPA, flow rate 1.5 mL min⁻¹, 254 nm, 40 °C) $t_R(R)$: 35.0 min, $t_R(S)$: 47.8 min, 5.178:94.822 er.



Peak#	Ret. Time	Area%
1	35.005	5.178
2	47.793	94.822
Total		100.000

Flow HPLCs - KRs in PhMe

1-Allyl-3-(4-chlorophenyl)-2-oxopyrrolidin-3-yl acetate S27



Chiralpak AD-H (95:5 hexane:IPA, flow rate 1.0 mL min⁻¹, 211 nm, 30 °C) $t_R(R)$: 20.4 min, $t_R(S)$: 30.2 min, 4.205:95.795 er.



1-Allyl-3-(4-chlorophenyl)-3-hydroxypyrrolidin-2-one 31



Chiralpak AD-H (95:5 hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) t_R (S): 18.8 min, t_R (R): 21.4 min, 19.811:80.189 er.





100.000

Total
1-Allyl-2-oxo-3-(thiophen-3-yl)pyrrolidin-3-yl acetate S28



Chiralpak AD-H (95:5 hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) t_R (*R*): 20.8 min, t_R (*S*): 23.5 min, 6.793:93.207 er.



Total

100.000

1-Allyl-3-hydroxy-3-(thiophen-3-yl)pyrrolidin-2-one 32



Chiralpak AD-H (95:5 hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) t_R (*S*): 25.8 min, t_R (*R*): 32.5 min, 14.503:85.497 er.





PDA Ch1 211nm				
Peak#	Ret. Time	Area%		
1	25.840	14.503		
2	32.509	85.497		
Total		100.000		

1-Benzyl-2-oxo-3-phenylpyrrolidin-3-yl acetate S29



Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) $t_R(R)$: 11.8 min, $t_R(S)$: 18.5 min, 9.629:90.371 er.



1-Benzyl-3-hydroxy-3-phenylpyrrolidin-2-one 33



Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 11.7 min, t_R (*R*): 12.7 min, 7.025:92.975 er.



Total

100.000

Scale-up

1-Benzyl-2-oxo-3-phenylpyrrolidin-3-yl acetate S29



Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) $t_R(R)$: 11.8 min, $t_R(S)$: 18.5 min, 5.267:94.733 (*R*:S) er.



Peak#	Ret. Time	Area%
1	11.842	5.267
2	18.936	94.733
Total		100.000



Chiralpak AD-H (90:10 hexane:IPA, flow rate 1.5 mL min⁻¹, 211 nm, 40 °C) t_R (*S*): 11.6 min, t_R (*R*): 12.7 min, 21.021:78.975 (*S*:*R*) er.



1-Allyl-2-oxo-3-(thiophen-2-yl)pyrrolidin-3-yl acetate S30



Chiralpak OD-H (95:5 hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) t_R (S): 20.9 min, t_R (R): 28.9 min, 10.279:89.721 er.





Peak#	Ret. Time	Area%
1	20.944	10.279
2	28.927	89.721
Total		100.000

1-Allyl-3-hydroxy-3-(thiophen-2-yl)pyrrolidin-2-one 34



Chiralpak AD-H (98:2 hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) t_R (*R*): 58.9 min, t_R (*S*): 65.7 min, 6.365:93.635 er.



2-Oxo-1,3-diphenylpyrrolidin-3-yl acetate S31



Chiralpak AD-H (90:10 hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) $t_R(R)$: 32.3 min, $t_R(S)$: 38.2 min, 9.153:90.847 er.





100.000

Total



Chiralpak AD-H (95:5 hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) t_R (*S*): 19.2 min, t_R (*R*): 26.8 min, 5.717:94.283 er.





 Peak#
 Ret. Time
 Area%

 1
 19.188
 5.717

 2
 26.780
 94.283

 Total
 100.000

1-Allyl-3-(4-methoxyphenyl)-2-oxopyrrolidin-3-yl acetate S32



Chiralpak AD-H (95:5 hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) t_R (*R*): 31.8 min, t_R (*S*): 45.5 min, 12.686:87.314 er.



1-Allyl-3-hydroxy-3-(4-methoxyphenyl)pyrrolidin-2-one 36



Chiralpak AD-H (95:5 hexane:IPA, flow rate 1 mL min⁻¹, 211 nm, 30 °C) t_R (S): 38.5 min, t_R (R): 42.3 min, 8.165:91.835 er.





PDA Ch1 211nm				
Pe	ak#	Ret. Time	Area%	
	1	38.523	8.165	
	2	42.291	91.835	
٦	Fotal		100.000	

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