

Supporting Information

The identification and use of robust transaminases from a domestic drain metagenome

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1. Taxonomic assignment of drain Class III TAmS

Table S1. Protein length, predicted molecular weight (MW) (calculated using ExPASy ProtParam), protein annotation and organism and percentage identity to closest homologue in NCBI database. Data have been deposited in GenBank and the accession number of each enzyme coding sequence is listed below beside the pQR number.

pQR	Accession number	Length (aa)	Predicted MW (kDa)	Function	Taxonomic Assignment	% Ident. to NCBI
2188	MK121624	483	52.3	Aspartate aminotransferase family protein	<i>Pseudoxanthomonas sp.</i>	97
2189	MK121625	484	52.3	Aminotransferase	<i>Sphingopyxis sp.</i>	96
2190	MK121626	490	54.6	Adenosylmethionine-8-amino-7-oxononanoate transaminase	<i>Perlucidibaca sp.</i>	71
2191	MK121627	450	49.2	Glutamate-1-semialdehyde 2,1-aminomutase	<i>Novosphingobium aromaticivorans</i>	63
2192	MK121628	701	77.8	Glutamate-1-semialdehyde aminotransferase	<i>Herbaspirillum lusitanum</i>	77
2193	MK121629	417	44.4	Acetylornithine aminotransferase	<i>Azonexus hydrophilus</i>	85
2194	MK121630	456	48.1	Glutamate-1-semialdehyde aminotransferase	<i>Candidatus Propionivibrio aalborgensis</i>	83
2195	MK121631	478	51.5	Omega amino acid-pyruvate aminotransferase	<i>Acidovorax sp.</i>	99
2196	MK121632	469	51.1	Aspartate aminotransferase family protein	<i>Mesorhizobium</i>	90
2197	MK121633	414	44.5	Acetylornithine aminotransferase	<i>Alkanindiges illinoisensis</i>	78
2198	MK121634	454	47.7	Aspartate aminotransferase family protein	<i>Azospira oryzae</i>	99
2199	MK121635	468	49.8	Aspartate aminotransferase family protein	<i>Alkanindiges illinoisensis</i>	76
2200	MK121636	460	50.9	Aminotransferase	<i>Pseudoxanthomonas mexicana</i>	97
2201	MK121637	1494, 504	54.5	Lysine 6-aminotransferase	<i>Pseudoxanthomonas mexicana</i>	97
2202	MK121638	394	42.4	Acetylornithine aminotransferase	<i>Candidatus Accumolibacter phosphatis</i>	83
2203	MK121639	433	45.7	Aspartate aminotransferase family protein	<i>Azonexus hydrophilus</i>	96

2204	MK121640	401	42.8	Acetylornithine transaminase	<i>Sphingopyxis sp.</i>	98
2205	MK121641	447	48.4	Adenosylmethionine-8-amino-7-oxononanoate transaminase	<i>Roseateles depolymerans</i>	76
2206	MK121642	436	45.9	Glutamate-1-semialdehyde aminotransferase	<i>Pseudoxanthomonas sp.</i>	98
2207	MK121643	402	42.9	Acetylornithine transaminase	<i>Azospira oryzae</i>	98
2208	MK121644	1380, 466	50.1	Aspartate aminotransferase family protein	<i>Sphingomonas sp.</i>	83
2209	MK121645	404	42.4	Acetylornithine aminotransferase	<i>Curvibacter delicatus</i>	69
2210	MK121646	416	44.3	Acetylornithine aminotransferase	<i>Pseudoxanthomonas sp.</i>	94
2211	MK121647	412	44.2	Acetylornithine aminotransferase	<i>Pseudomonas lavalieres</i>	100
2212	MK121648	423	46.2	Denosylmethionine-8-amino-7-oxononanoate transaminase	<i>Sphingopyxis</i>	94
2213	MK121649	453	47.9	Aminotransferase	<i>Sphingopyxis</i>	87
2214	MK121650	459	49.5	Omega amino acid-pyruvate aminotransferase	<i>Aquabacterium parvum</i>	70
2215	MK121651	414	44.1	Acetylornithine aminotransferase	<i>Pseudoxanthomonas sp</i>	96
2216	MK121652	396	42.0	Acetylornithine aminotransferase	<i>Dechloromonas aromatica</i>	87

2. Percentage Identity Matrix

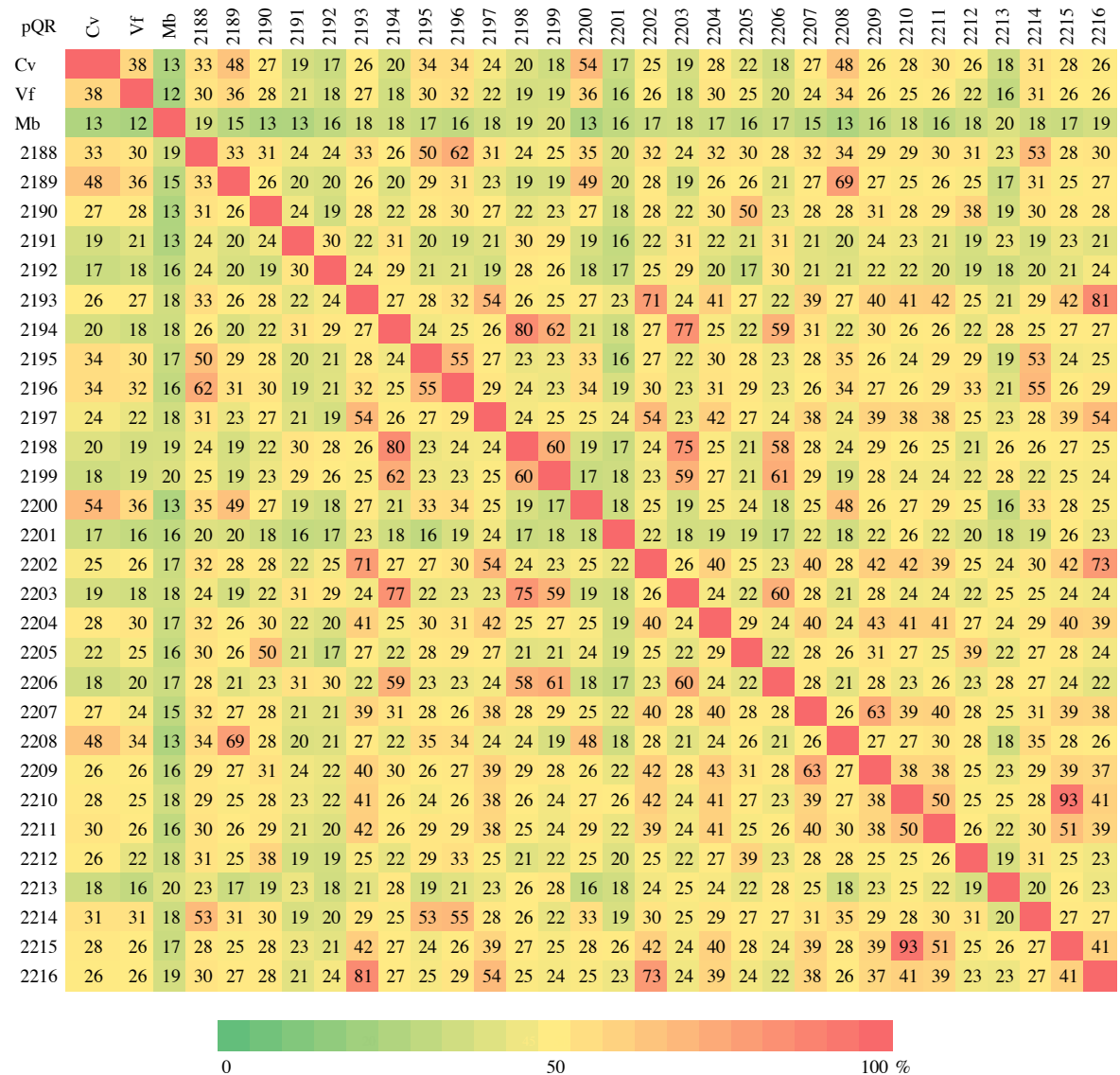


Figure S1. Heat map of the percentage identity of 29 cloned drain TAmS. Percentage identity matrix generated using Clustal Omega Multiple Sequence Alignment tool and visualised using Excel. Cv: *Chromobacterium violaceum* CV2025 accession number WP_011135573.1, Vf: *Vibrio fluvialis* JS17 accession number AEA39183.1, Mb: *Mycobacterium vanbaalenii* ABM15291.1.

3. Expression of 29 TAmS

In the following SDS-Page gels, lane 1 is always ladder - NEB Broad Range protein ladder (10-250 kDa) (Figure S2-S4) or NEB color protein standard (Figure S5-S9), each protein is in two wells – the first is the cell free extract (CFE) and the second is the total protein (TP) fraction.

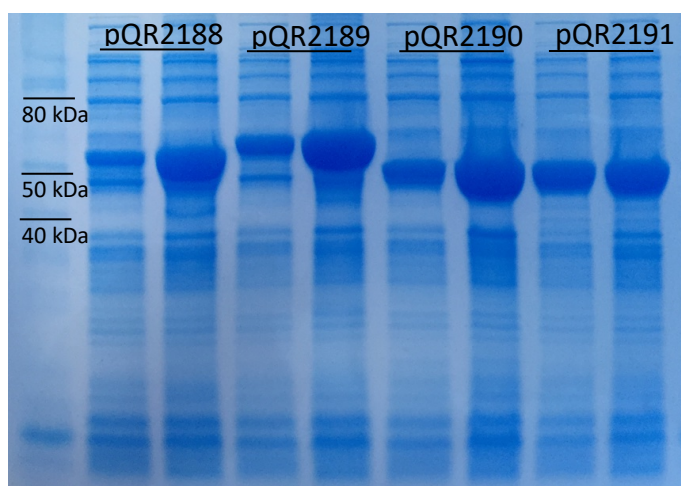


Figure S2. SDS-Page gel of pQR2188-2191.

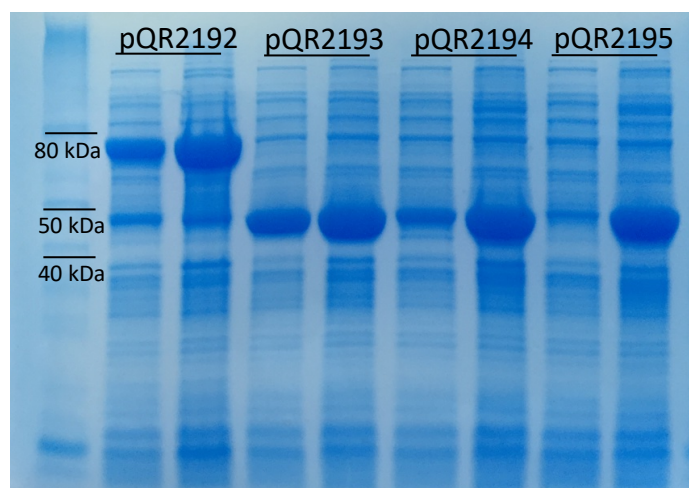


Figure S3. SDS-Page gel of pQR2192-2195.

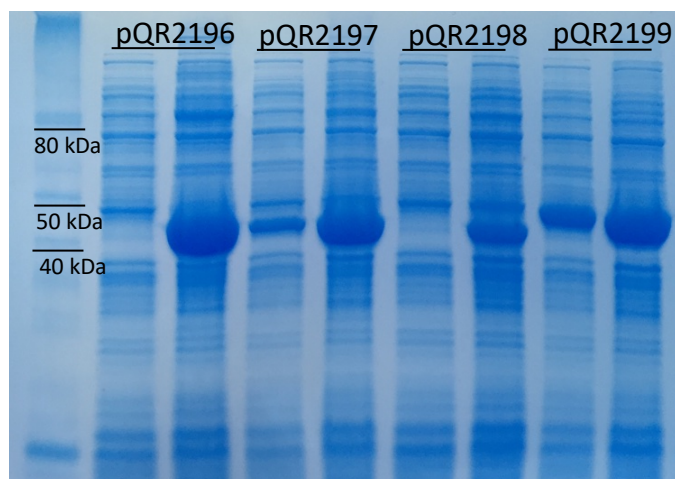


Figure S4. SDS-Page gel of pQR2196-2199.

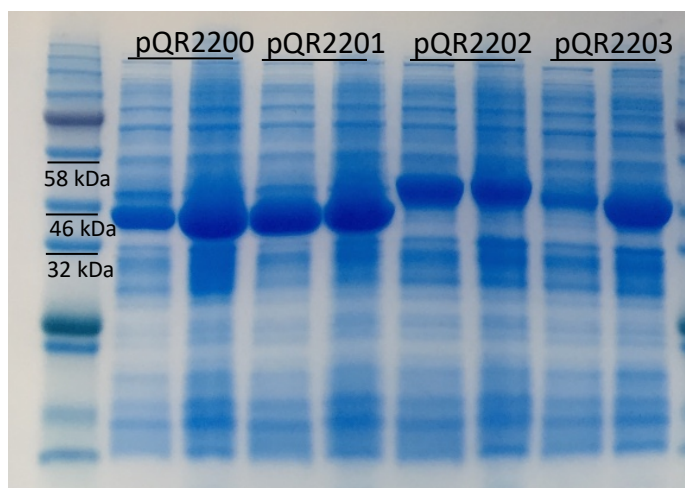


Figure S5. SDS-Page gel of pQR2200-2203.

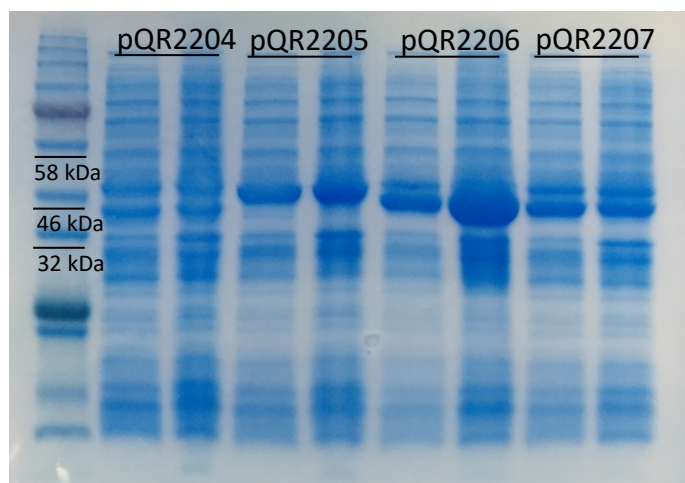


Figure S6. SDS-Page gel of pQR2204-2207.

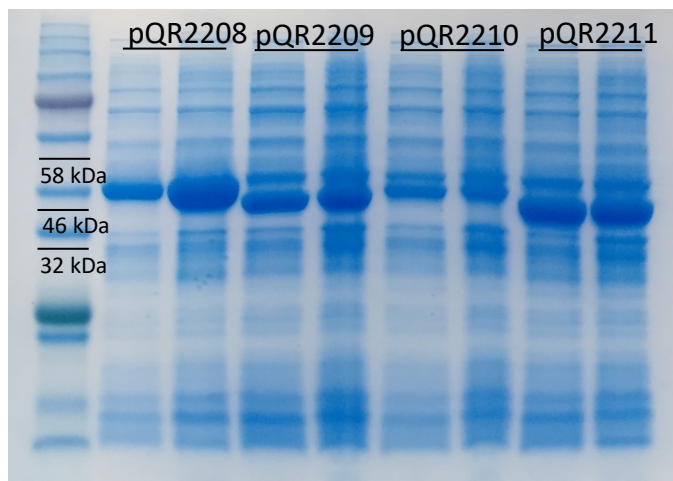


Figure S7. SDS-Page gel of pQR2208-2211.

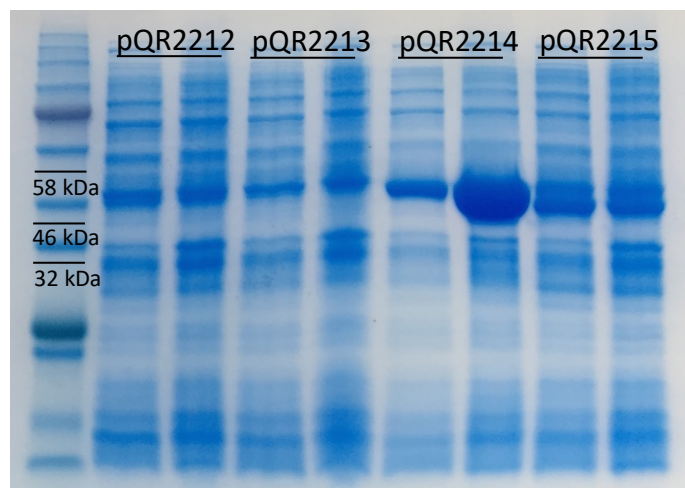


Figure S8. SDS-Page gel of pQR2212-2215.

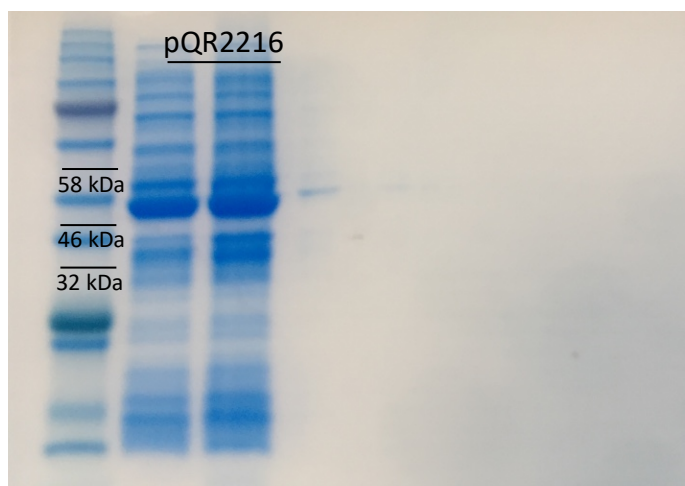


Figure S9. SDS-Page gel of pQR2216.

4. DNA Sequences

>pQR2188

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> pQR2209

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CATGA

> pQR2210

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> pQR2211

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CGCCAAGCTCGCGCAGGGCTGA

> pQR2212

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> pQR2213

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> pQR2214

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> pQR2215

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GGTGCTGGATCTCGCCGCGGCACAGGGCCTGCTGATGCTGCAGGCCGGCCCCGATGT
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> pQR2216

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GGTCCCGCCTGTTTCGATGTGCGAGGGCAAGGAGTATCTCGACGCCTTGTCCGGCATTGC
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CGTCCGGGTTCCCTACGGCGATCTCGACGCCATCAAGGCGGTGGCCGAGCACAACAA
GAACATCGTCGCGGTGATGTTTGAATCATCCAGGGCGAAGGCGGCATCCATCTCGTC
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TGTGCGACGAAGTCCAGTGCGGCATGGGACGAACCGGCAAATGGTTCGGCTTCCAGA
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TGCCGCCGCTGAACTTCACCGAAAACGATGCCCGTGAGCTGGTCGACCGTGTGCGAC
CGCTGATCAAGGCATTCCTCGCAGGGTGA

5. Amino acid sequence

Protein sequences correspond to cloned genes; amino acids in **bold** correspond to the vector sequence.

> pQR2188

MGSSHHHHHHSSGLVPRGSHMSADDTPSALAEHYARQNLDAPGSLDHFWMPTANKQF
KAKPRLLASASGMYYKDVDGNEVLDATEAGLWCCNAGHARPRIVEAVRQQIGTLDFAFNFS
MSSPLPFKLAERLAALAPGDLNRVFFSNSGSEAVDSALKIALAYHRVRGEGQRTRFIGREK
GYHGVGFGGMSVGGPLNNRKWFGPGLPAVSHIRHTLDVARNAFSKGLPPHGIELAEDLER
QIALYDASTIAAVIVEPVSGSAGVVIPPEGYLQRLREICDKHGILLIFDEVITGFGRVGHAFGA
QRFVGTDPMITAAKGITNGCVPMGATFVSERLDAFMNGPDNAIDMFHGYTYSGHPLACA
AALATLDTYEEEHLFDKALSLGDYWQEALHSLKGLPNIIDIRNIGLVGAIELAPRAGAPGTRA
YDVFARAFHEGHLLTRVTGDVIALSPPLIVEKDHIDRIVNVLADTIRATA**EEEEHHH**

> pQR2189

MGSSHHHHHHSSGLVPRGSHMPRNHDIAELRRLDVAHHLPAQADWAEIEKLGGSRITHAE
GCYIHDGDGHRILDGMAGLWCVNVGYGREELVEAAAAQMRELPFYNTFFKTATPPTVTLA
AKIASLTGNRLPHIFFNASGSEANDTVFRMVRHYWKLKGEPKRTVFISRWNAYHGSTVAGV
SLGGMKAMHAQGDLPPIGIEHVRQPYSFGEGQGMTEEEFCDACVHAIEDKILEVGPENCAA
FIGEPVQGAGGVVIPPCKGYWPKVEAVARKYGLLVVSDEVICGFGRTGKMWGHETMGFTPD
LMSMAKGLSSGYLPISATAVATHVVDVLKTGGDFVHGFTYSGHPVAAAVALKNIEIIEREGLV
ERTGSVTGPHLAKALATLNDHPLVGETRSIGLLGAVEIVGEKVTRARFGGAEGTAGPMARD
ACIANGLMVRGIRDLSVMCPPLIISTEQIDEMVAIRKSLDEVMPKLRAL**EEEEHHH**

> pQR2190

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VWLED FEGRRYLDVSSWWVNLFGHANPRINNAVKEQLDTLEHVILAGFTHEPIVELSERL
VQLAPKGLTRCFYADNGSAATEIALKMSLHFWRNVGKAEKTRFICLENGYHGETLGSLSVT
DIPFSAFYAPLLKDHLRAPSPDCSRRDEGESWESFSRRQFAAMEALLEKHHAIEVSAVILEP
LVQGAAGMKMYHPVYLTLLREACDRYGVHLIADEIAVGFGRTGTLFACEQAGITPDFLCLSK
GLTAGYLPMSVVMTTDTVYNFYDSYESLKGFLHSHSYTGNALAARAALASLDIFASDNVLE
KNKLLAATMTDALRGLGDHGHVLEVRQTGMIAAVELVQDRRTRQPFDRERRGLQIFQHA
LDKGVLLRPIGSVVYFIPPYVITPEEIRLMVDVAAAIDVATAGTASRPGPGNIAL**EEEEHHH**

> pQR2191

MGSSHHHHHHSSGLVPRGSHMSGQRDQELRARAANKVMPSSAFGHVGTALLPANYPQFF
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AEKFVEIVSHADWAFFCKNGTDATTIARTIARAQTGRRKILIAEGSYHGAAPWCNPFPAQTV
PEDRAHMLTFTFNDIASLEAAVAEAGDDLAGIATPDKHEAFANQEFPTQDYARRCREICDA
SGAVLVVDDVRAGFRLAVDCSWATVGVPDLSCWGKCFANGYSISAVMGSNRVKQGADS
IFATGSFWQSAISMAAALATLDIIRDGKVIEKTVRLGQRLRDGLDEVSRRHGFTLNQGTGPVQ
MPQILFEGDPDFRVGFVAWTSAMIDRGFYLHPWHNMFLCDAMTEEDIDQTIEAADSFAFATVR
AALPTLQPHERVLAFLSARA**EEEEHHH**

> pQR2192

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PLIDAALVDLVIDRFLQGDVDYVSNAVPTYPDGLDTEVFSMAALRQAWQQATSTFDHEHV
TPYLRDSGKFR LAVVSGEHDYSGERWTVDEPADFDVITQIFAHFAPRLDFSWTEVLALCHT
QPQLFAANQHLIRNEGAHMG TGQKLWKRAKNVIAGGNMLLSKRPEMFLPEQWPAYFSRA
QGCTVWDLDNQAYTDMIMIGTNTLGYGHPEVDDAVRRTIDAGNMSTFNCPEEVYLAekli
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HLLPGLEPNGVPESLRGTIYPFNYNNAEALVNSQDIGVIKMEVSRNHGPEDEGFLHKVRE
LATARGIVLIFDECTSGFRQTFGGLHKLYGVEPDMAMFGKALGNGYAITATIGRREVMEAAQ
TTFISSTFWTERIGPTAALKTLEVMERERSWDITQTGLAITERWKTLAARHGLSINTNGLPA
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GRDVMSELLKGPICHAGFKRLEHHHHHH

> pQR2193

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AAIKLARFYGHKKGIELPTVIVMEKSFHGRMTATLSATGNYKVQVGFELVAGFVRVPYGD
DAIRAVAEQNPNIIVAVMLEVIQGEGLHREPAYYQGVRLCDADHDLWLMICDEVQCGMGR
TGKWFQYQVGVQPDIALAKGLGSGVPIGACMAGGRAAGLFGPGNHGSTFGGNPLVCA
AALTLDLCEEEGLLANAENIGKLIRQLAAGLADARGVVDIRGHGLMIGIELDRPCGVLVTQ
GLAAGLLINVTGDTVVRLLPPLNFSERDASELVDRMIPLIKAFLAGEHHHHHH

> pQR2194

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LLCAMLPSLDMVRLVSSGTEATMSAIRLARGHTGRDLLIKFEGCYHGHSDGLLVKAGSGLL
TFGNPSSGGVPADVAQHTMVLNDYVQGLEAAFTHEGDRIAIVIVEPVAGNMNLIAPLPAF
LKTMRALCTQHGAFLIFDEVMTGFRVGPQCAQGFYGITPDLTTLGKVIGGGMPVGAFFGK
REIMEKIAPLGPVYQAGTLSGNPVAVAAGLATRLIQAPGFYDALAASTRALCAGLTEAAKR
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> pQR2195

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SNIAIVIVEPFGSGSAGVIPPVGYLQRLREICTQNNILLIFDEVISGFGRSGAFTGAEAFGVTP
DILNFAKQVTNGAQPLGGVIASKEIYDTFMAAGGPEYMLEFPHGYTYSAPHPVACAAGIAALD
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> pQR2196

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> pQR2197

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> pQR2198

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> pQR2199

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>pQR2200

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> pQR2201

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> pQR2202

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> pQR2203

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> pQR2204

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> pQR2205

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> pQR2206

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> pQR2207

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> pQR2208

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> pQR2209

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> pQR2210

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> pQR2211

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> pQR2212

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> pQR2213

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> pQR2214

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> pQR2215

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> pQR2216

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KGLGSGVPIGACLAGGKAAGLFGPGNHGSTFGGNPLVATAALT TIAVIEEEGLLDNAAKIGV
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6. Other Substrates

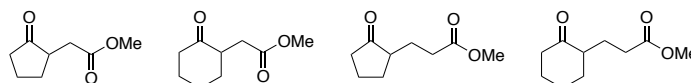


Figure S10. Substrates that showed no activity with any of the enzymes.

7. Analytical Methods

7.1 Achiral methods

Quantitative analysis of enzymatic reactions was analytical reverse phase HPLC analysis with respect of chemical standards using an Agilent 1260 Infinity or Dionex Ultimate 3000 with an Ace 5 C18 column 150×4.6 mm. Elution was carried out at 1 mL/min with a linear gradient of acetonitrile/0.1% TFA in water, injection volume 10 μ L and column temperature 30 $^{\circ}$ C. Chromatograms showing retention times included below.

Method A:

Products were detected at 250 nm using a linear gradient 15 - 72% acetonitrile over 15 min.

This method was used to detect acetophenone.

Method B:

Products were detected at 204 nm using a linear gradient 5 - 72% acetonitrile over 15 min.

This method was used to detect all amine products **18b-25b**.

7.2 Chiral methods

Enantiomeric excess of **18b** and **20b** was determined by GC analysis using an Agilent 7820A GC System with a Supelco Beta Dex 225 capillary GC column $30 \text{ m} \times 250 \mu\text{m} \times 0.25 \mu\text{m}$ with flame ionization detector at 300 $^{\circ}$ C, a temperature gradient and 1 μ L injection volume. Chromatograms showing retention times included below.

Method A:

Initial temperature 140 $^{\circ}$ C, 1 min hold, ramp 5 $^{\circ}$ C/min to 210 $^{\circ}$ C, 2 min hold. This method was used to detect **18b**.

Method B:

Initial temperature 150 $^{\circ}$ C, 1 min hold, ramp 10 $^{\circ}$ C/min to 210 $^{\circ}$ C, 3 min hold. This method was used to detect **TFA-20b**

8. HPLC and GC Traces

8.1. HPLC traces

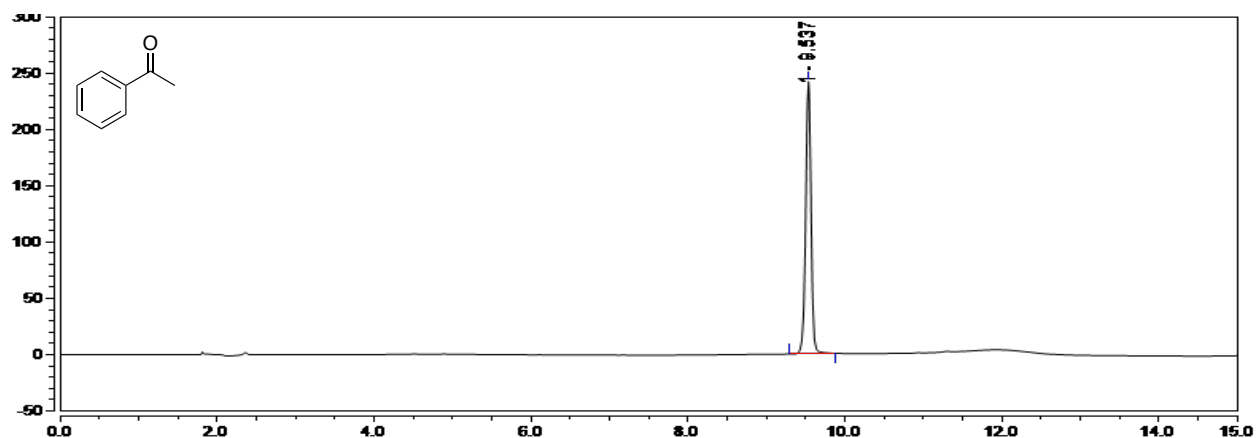


Figure S11. Chromatogram of acetophenone using HPLC method A. Retention time 9.5 min

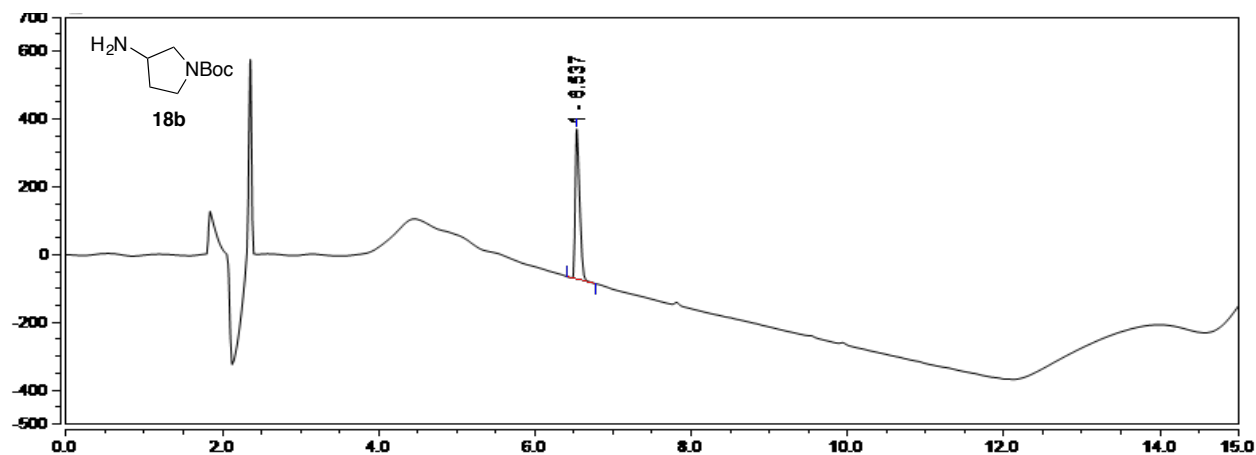


Figure S12. Chromatogram of amine **18b** using HPLC method B. Retention time 6.5 min.

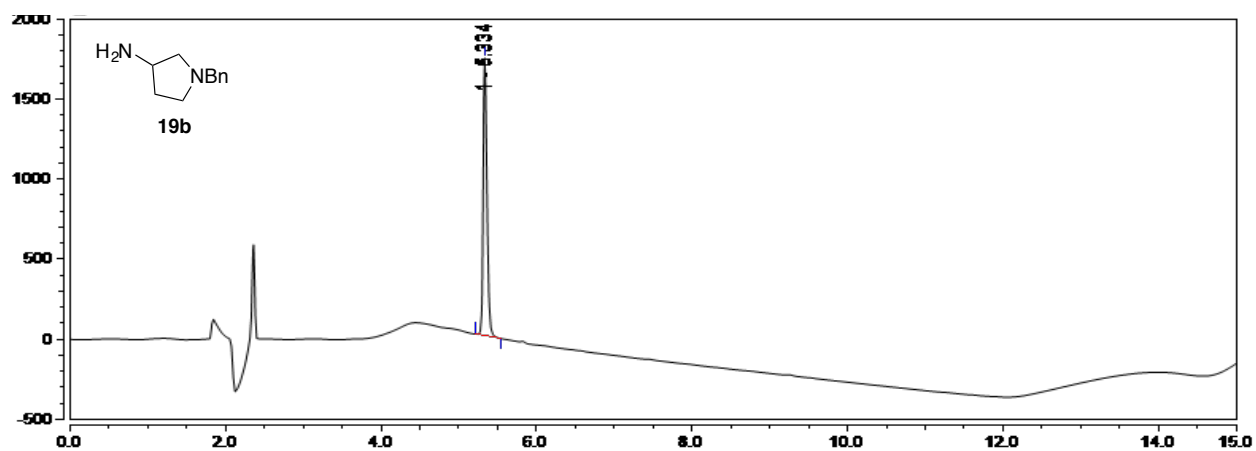


Figure S13. Chromatogram of amine **19b** using HPLC method B. Retention time 5.3 min.

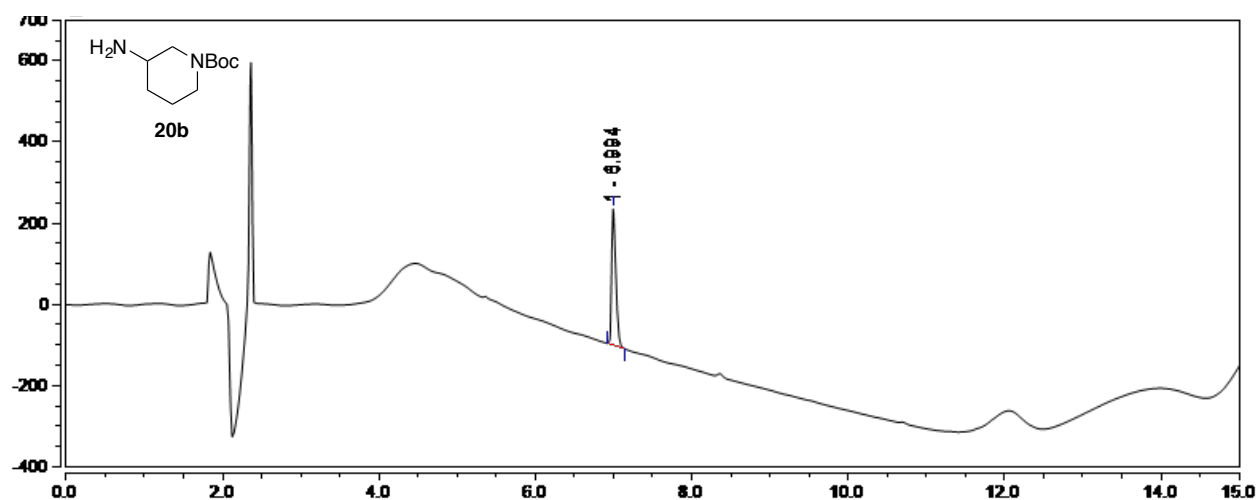


Figure S14. Chromatogram of amine **20b** using HPLC method B. Retention time 7.0 min.

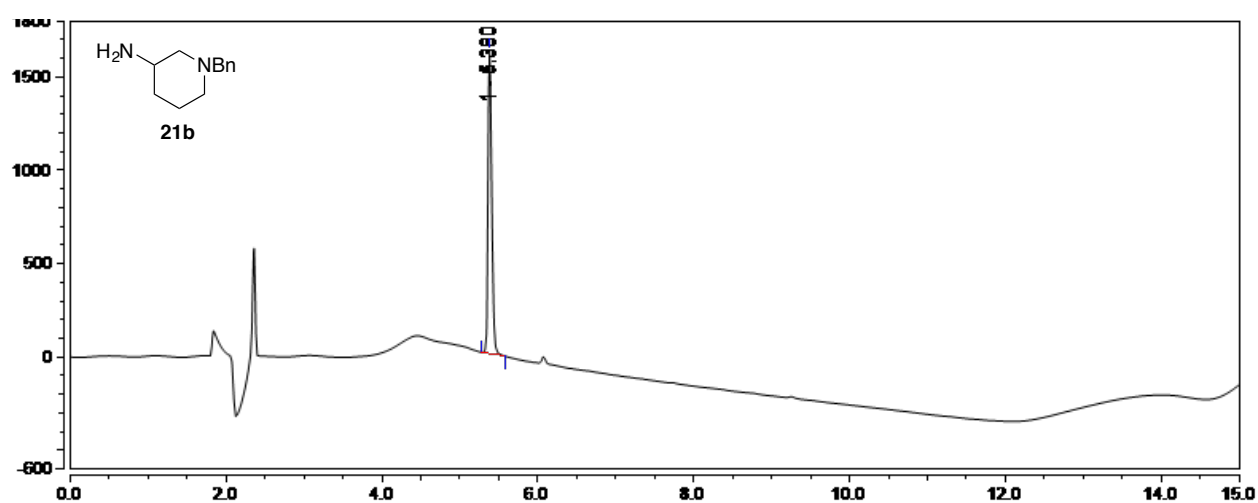


Figure S15. Chromatogram of amine **21b** using HPLC method B. Retention time 5.4 min.

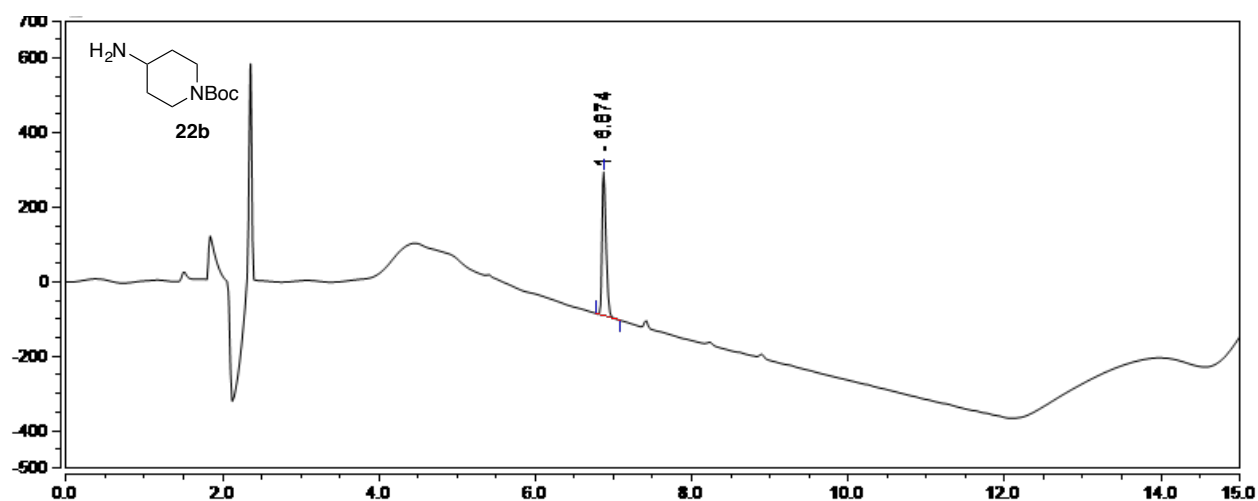


Figure S16. Chromatogram of amine **22b** using HPLC method B. Retention time 6.7 min.

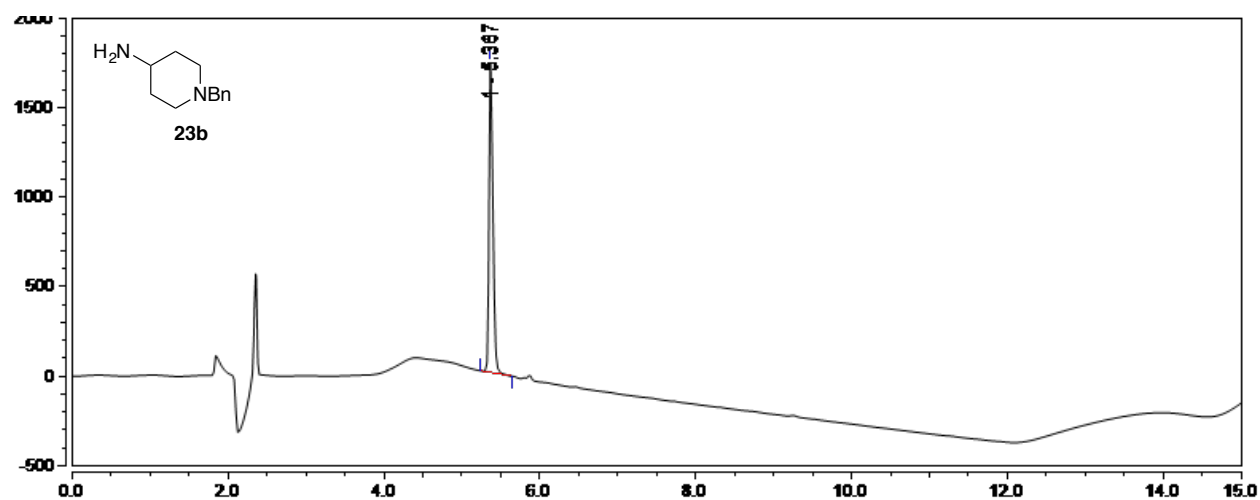


Figure S17. Chromatogram of amine **23b** using HPLC method B. Retention time 5.4 min.

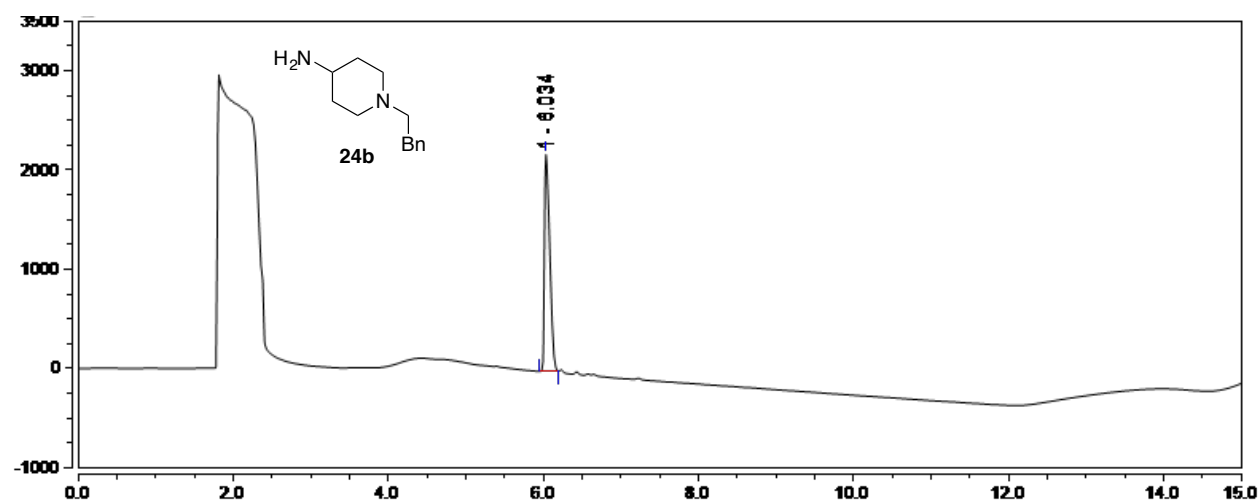


Figure S18. Chromatogram of amine **24b** using HPLC method B. Retention time 6.0 min.

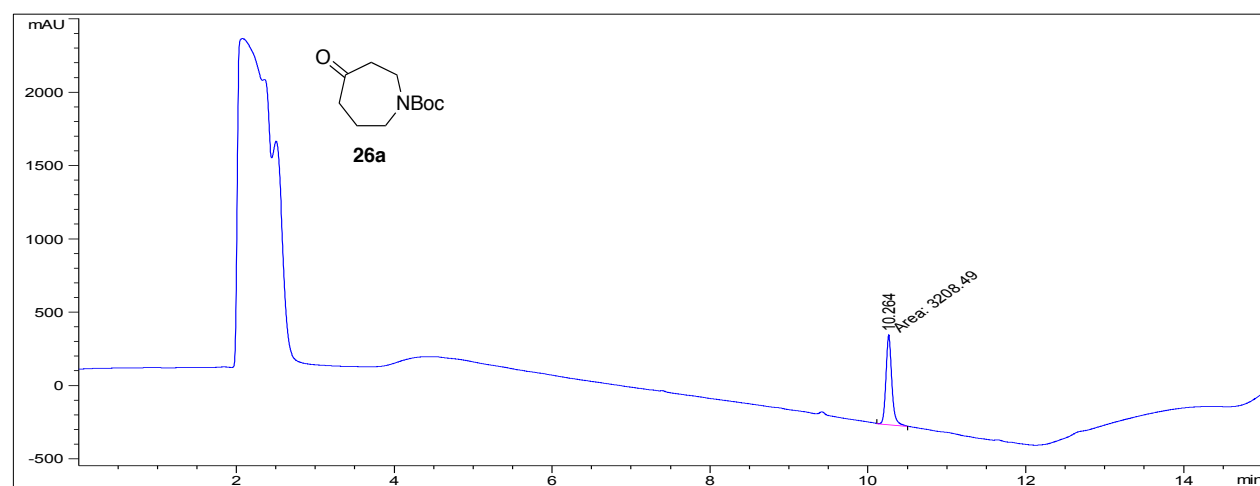


Figure S19. Chromatogram of ketone **26a** using HPLC method B. Retention time 10.3 min.

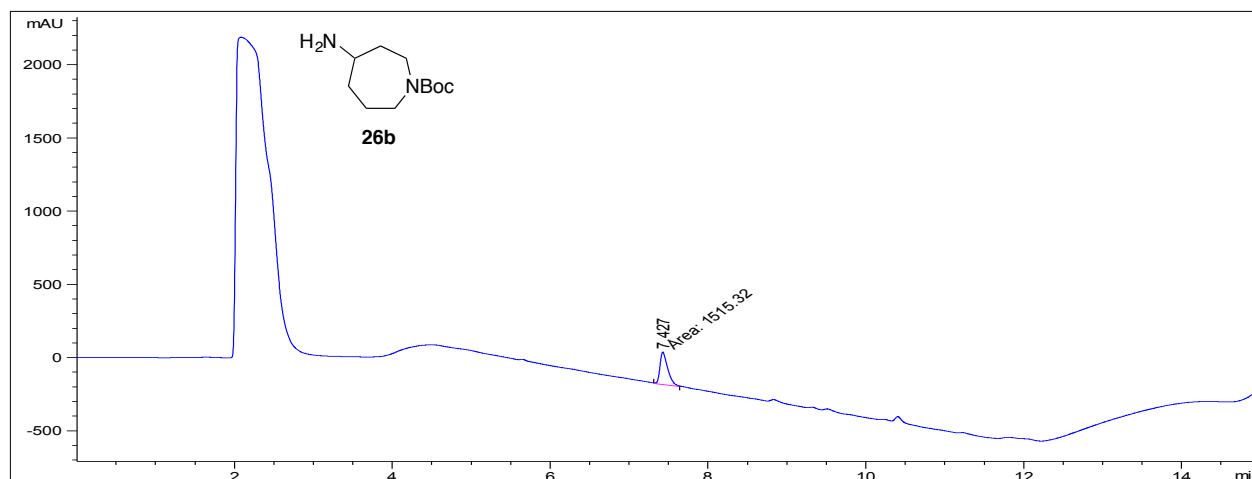


Figure 20. Chromatogram of amine **26b** using HPLC method B. Retention time 7.4 min.

8.2. GC Traces

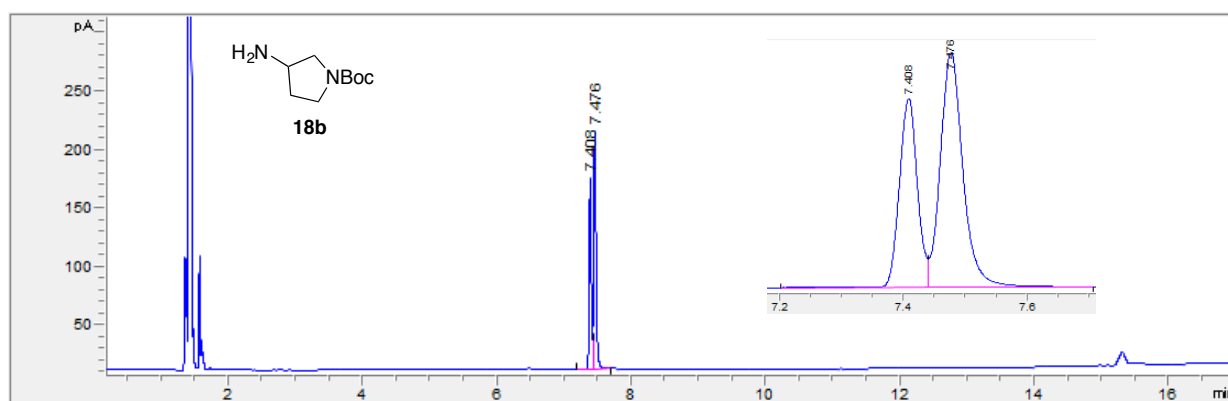


Figure S21. Chromatogram of racemic **18b** using GC method A. Retention times 7.4 & 7.5 min.

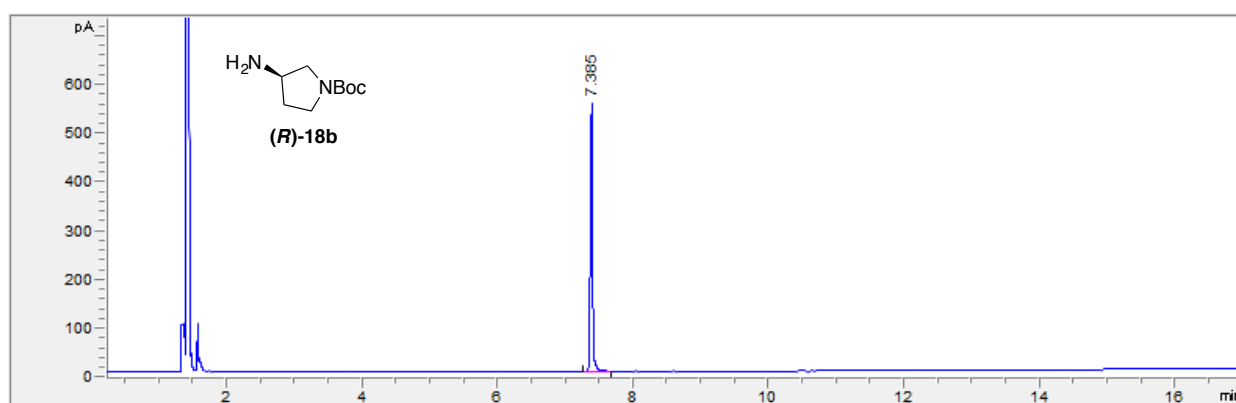


Figure S22. Chromatogram of (*R*)-**18b** using GC method A. Retention times 7.3 min.

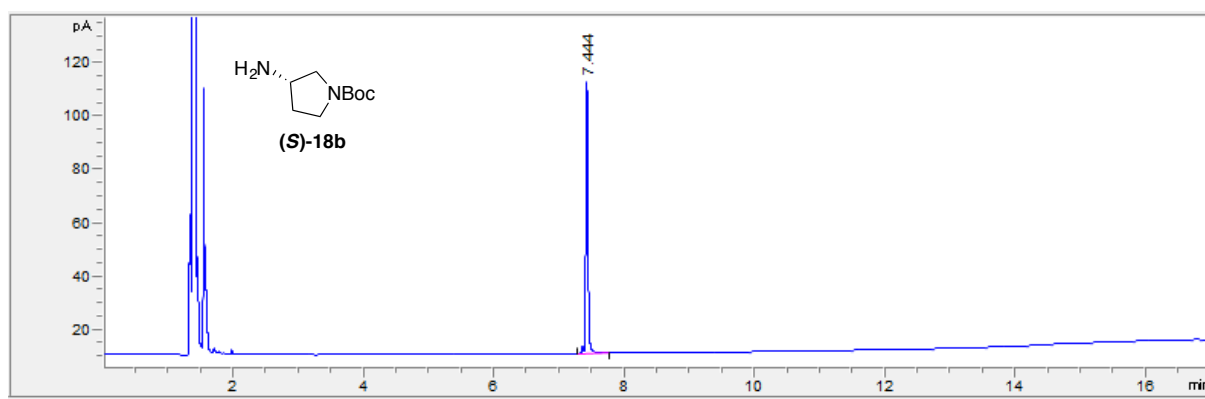


Figure S23. Chromatogram of **(S)-18b** using GC method A. Retention times 7.4 min.

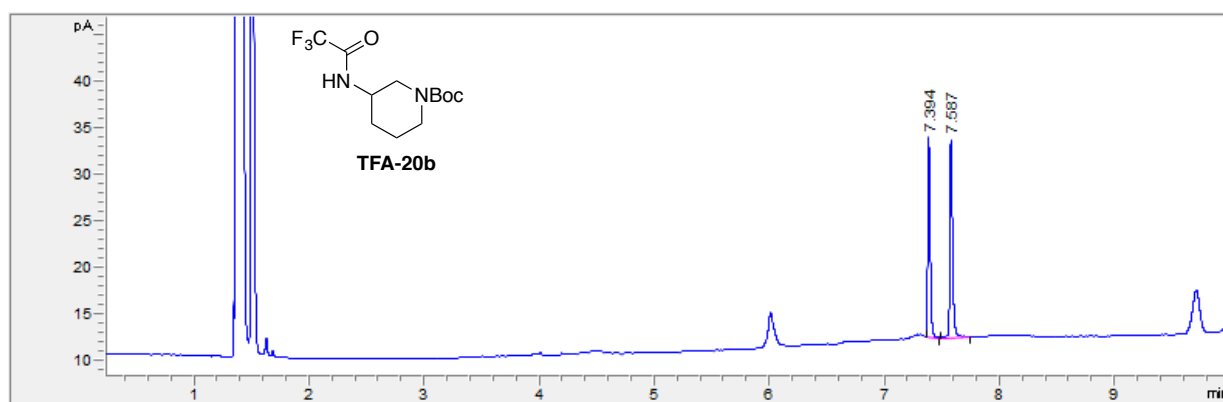


Figure S24. Chromatogram of racemic **TFA-20b** using GC method A. Retention times 7.4 and 7.6 min. Enantioselectivity inferred from **18b**.

9. Calibration Curves

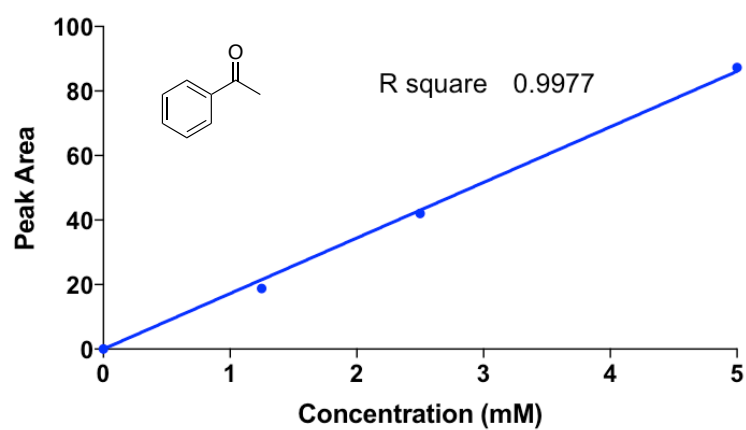


Figure S25. Calibration curve for acetophenone using HPLC method A.

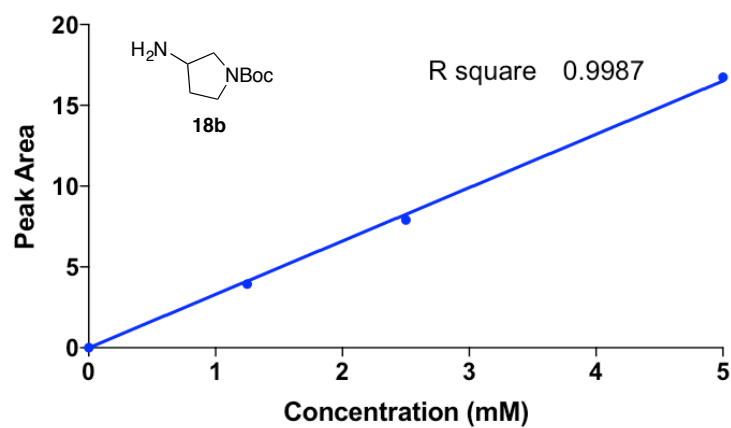


Figure S26. Calibration curve for amine **18b** using HPLC method B.

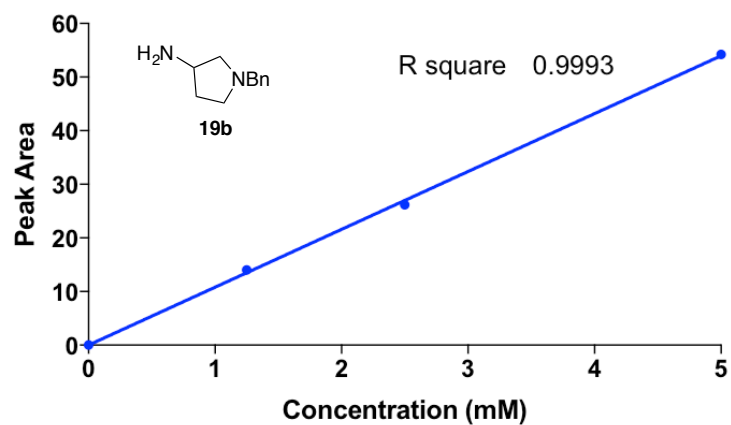


Figure S27. Calibration curve for amine **19b** using HPLC method B.

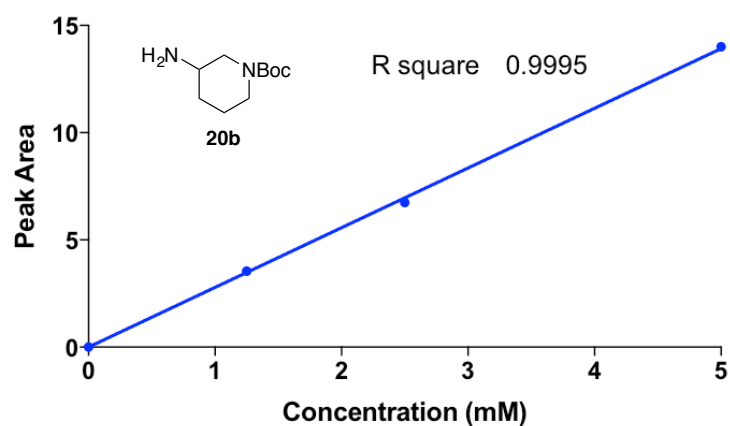


Figure S28. Calibration curve for amine **20b** using HPLC method B.

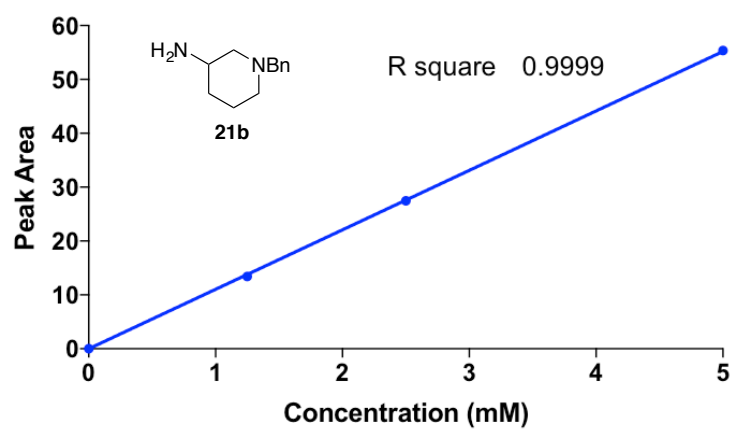


Figure S29. Calibration curve for amine **21b** using HPLC method B.

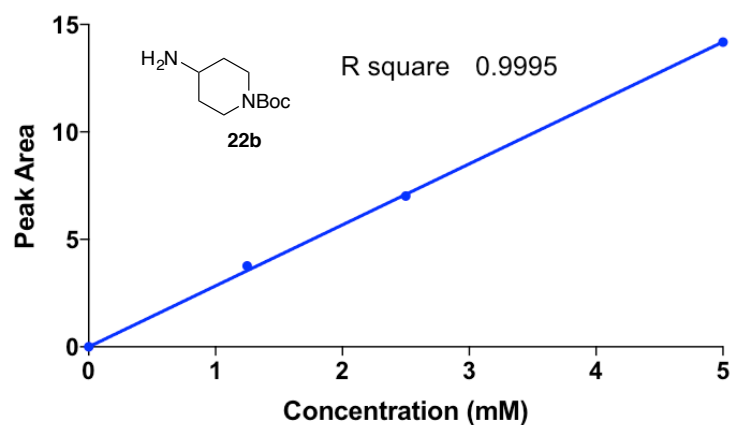


Figure S30. Calibration curve for amine **22b** using HPLC method B.

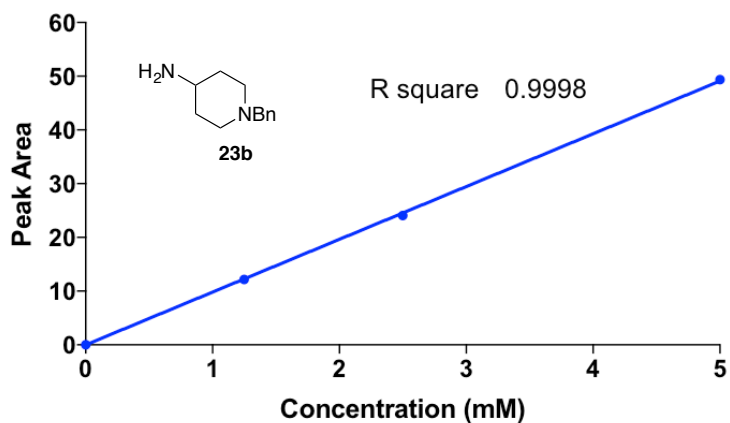


Figure S31. Calibration curve for amine **23b** using HPLC method B.

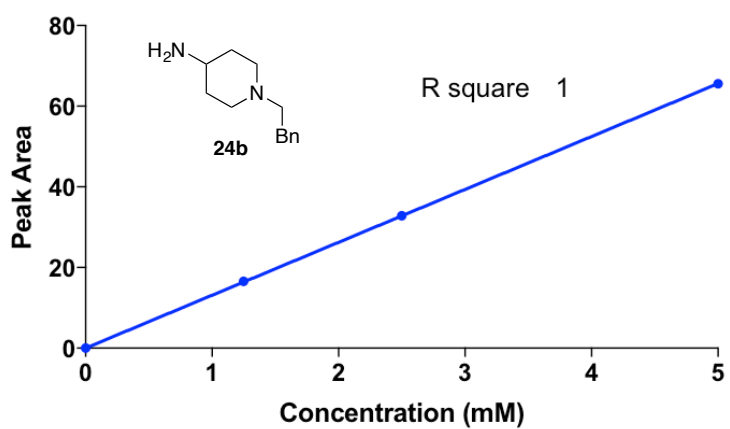


Figure S32. Calibration curve for amine **24b** using HPLC method B.

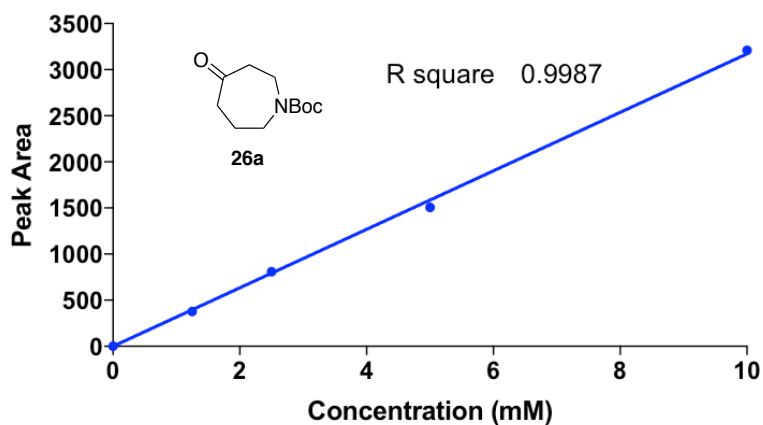


Figure S33. Calibration curve for ketone **26a** using HPLC method B.

10. Purification of pQR2189

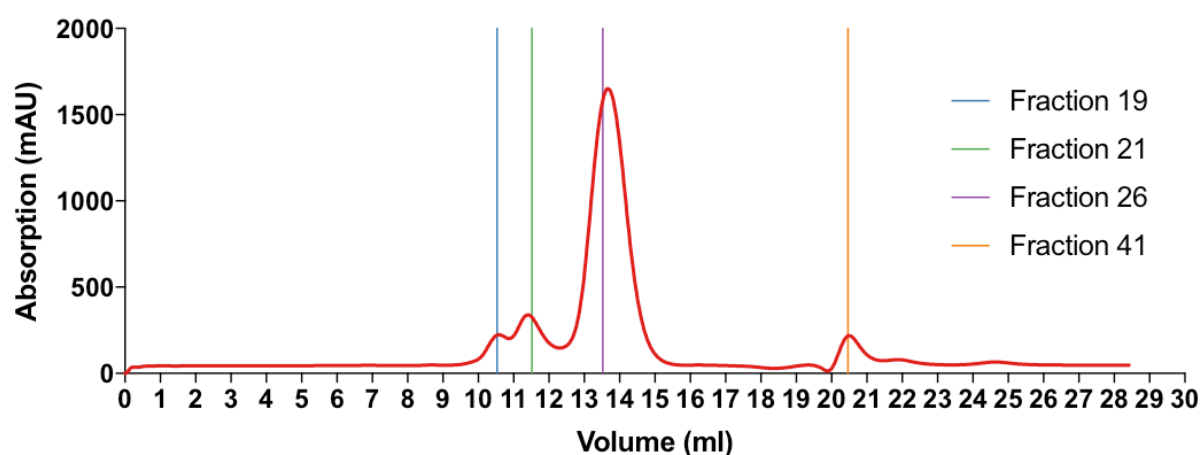


Figure S34. Trace of gel filtration of pQR2189 and His-tag purification. Vertical lines show the fractions that were run on a gel shown in Figure S35.

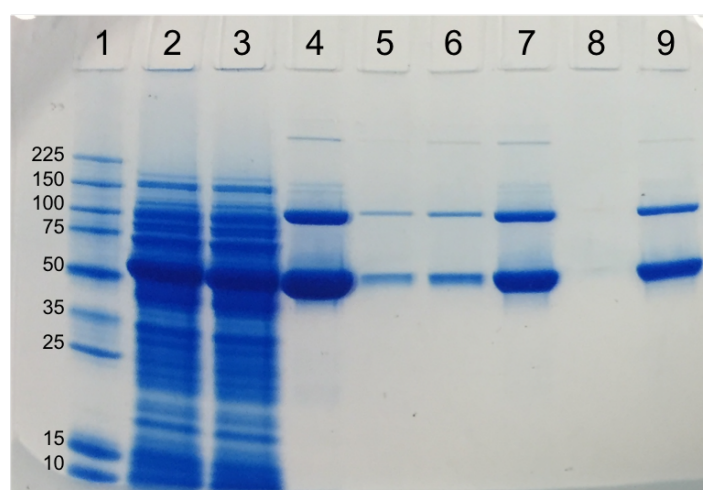


Figure S35. SDS-Page gel of pQR2189 purification. **Lane 1:** Promega broad range protein molecular marker. **Lane 2:** TP fraction. **Lane 3:** CFE fraction. **Lane 4:** pQR2189 after His-tag purification. **Lane 5:** Gel filtration fraction 19. **Lane 6:** Gel filtration fraction 21. **Lane 7:** Gel filtration fraction 26. **Lane 8:** Gel filtration fraction 41. **Lane 9:** Pure protein fractions used in kinetics studies.

11. Kinetics Graphs

11.1. Pyruvate and (S)-MBA

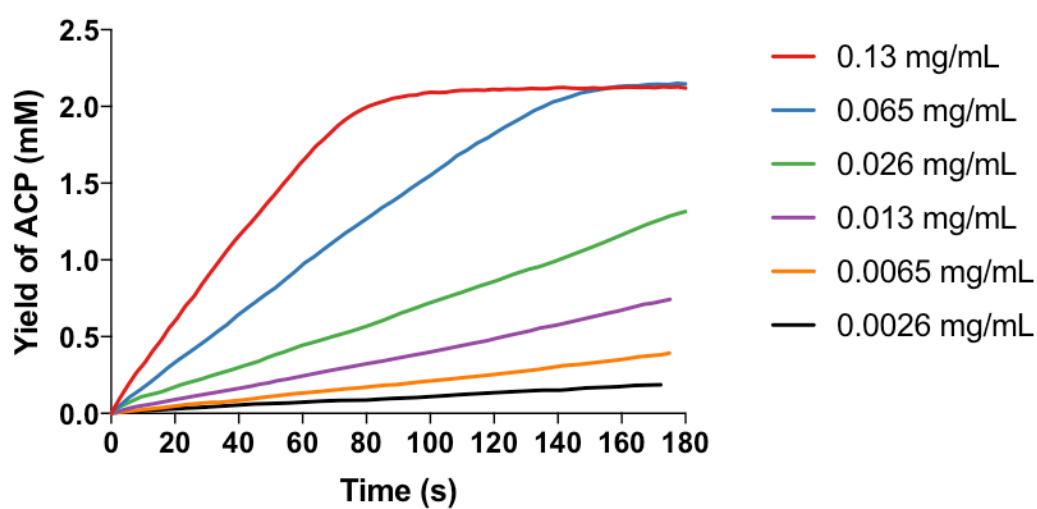


Figure S36. Effect of enzyme concentration on acetophenone formation.^{1,2}

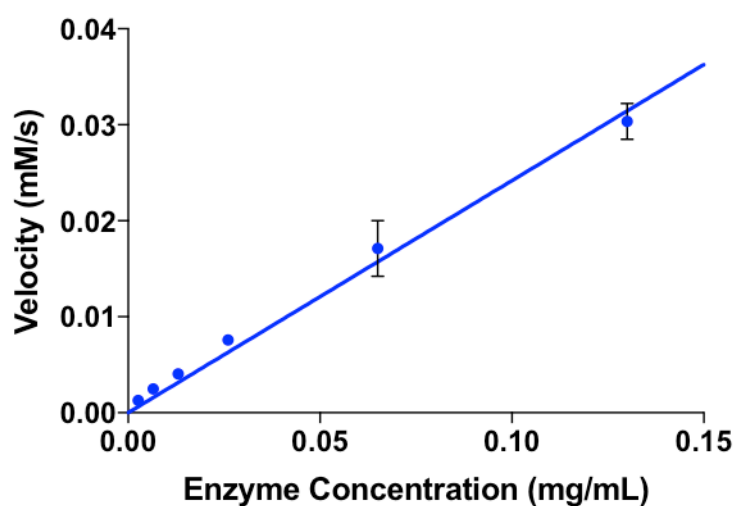


Figure S37. Enzyme concentration vs. velocity.

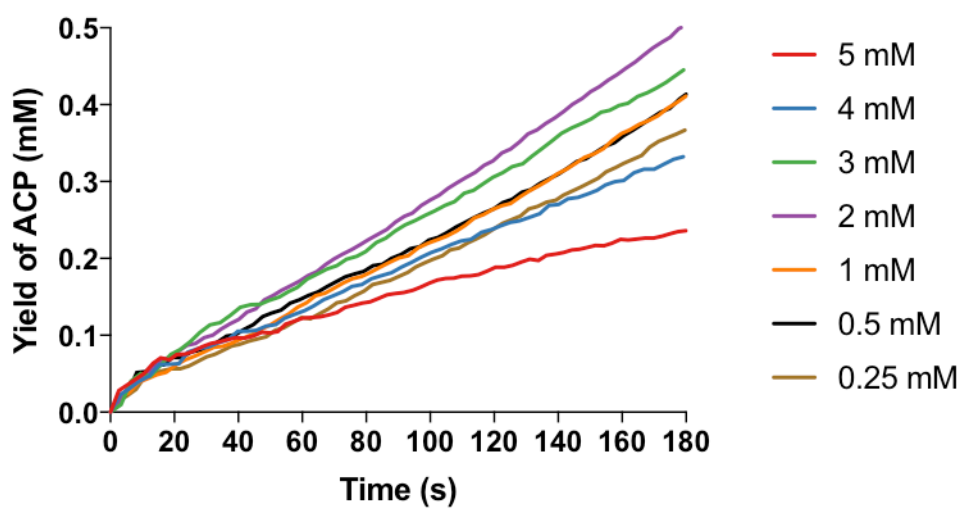


Figure S38. Effect of varying pyruvate concentration on acetophenone formation.^{1,2}

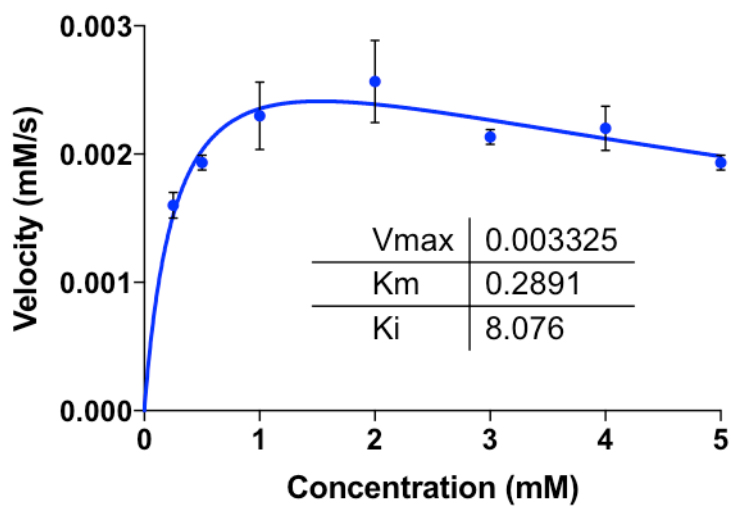


Figure S39. Michaelis-Menten plot of pyruvate concentration, $K_m = 0.29$ mM.

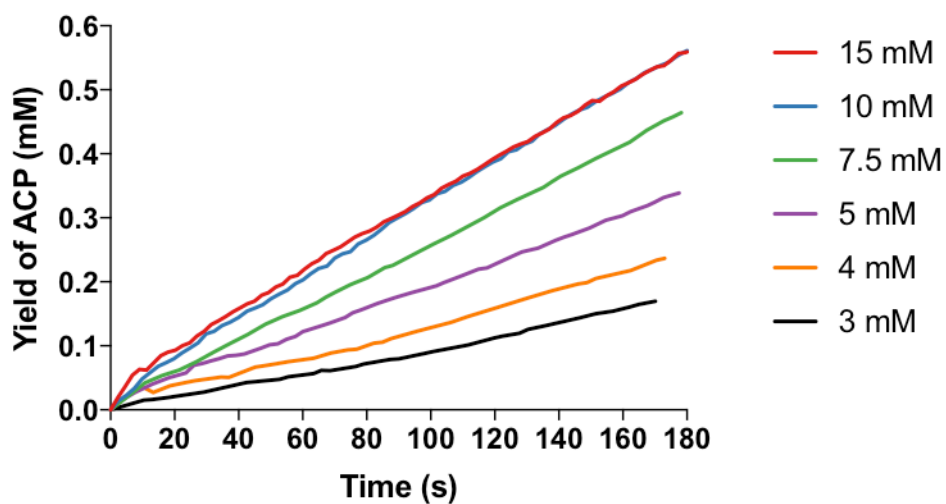


Figure S40. Effect of varying (S)-MBA (**S**)-**7** concentration on acetophenone formation.^{1,2}

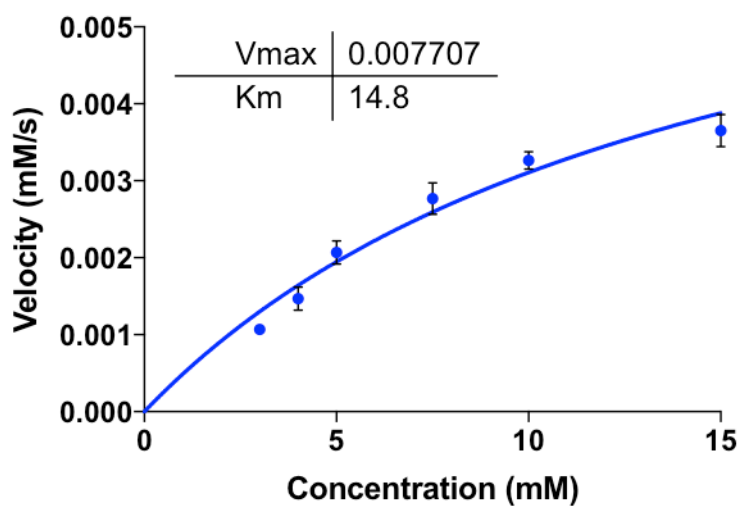


Figure S41. Michaelis-Menten plot of (S)-MBA concentration, $K_m = 14.8$ mM.

11.2. 1-Boc-3-pyrrolidinone and IPA

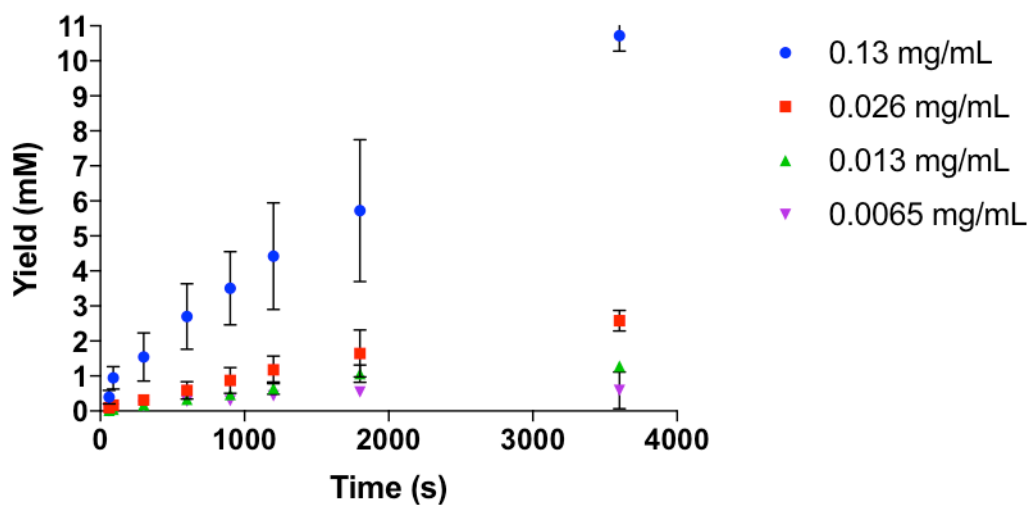


Figure S42. Effect of enzyme concentration on formation of **18b**.²

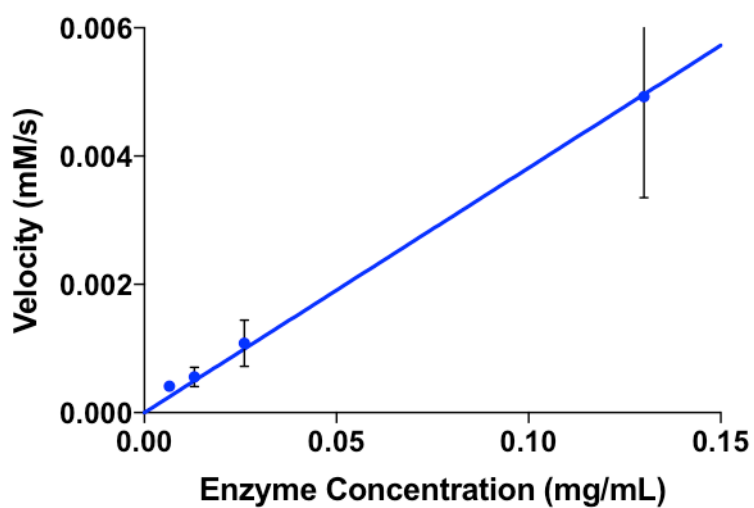


Figure S43. Enzyme concentration vs. velocity.

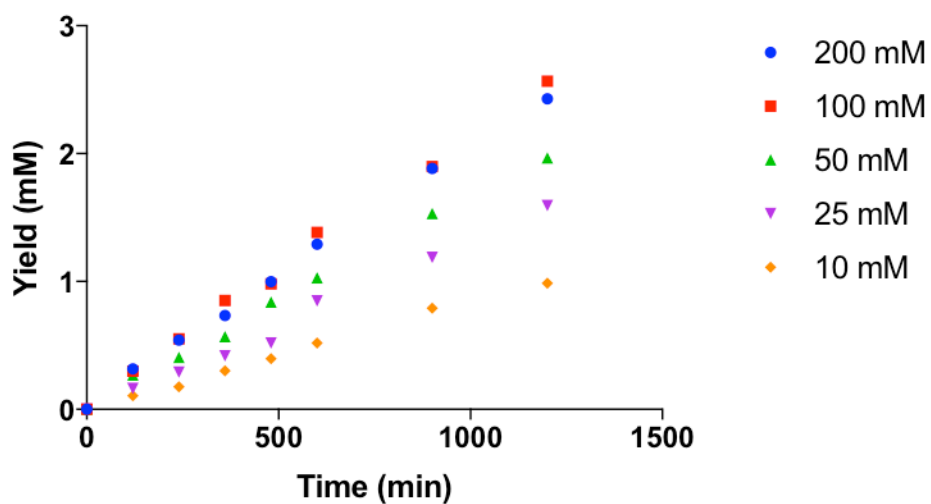


Figure S44. Effect of varying ketone **18a** concentration on formation of **18b**.²

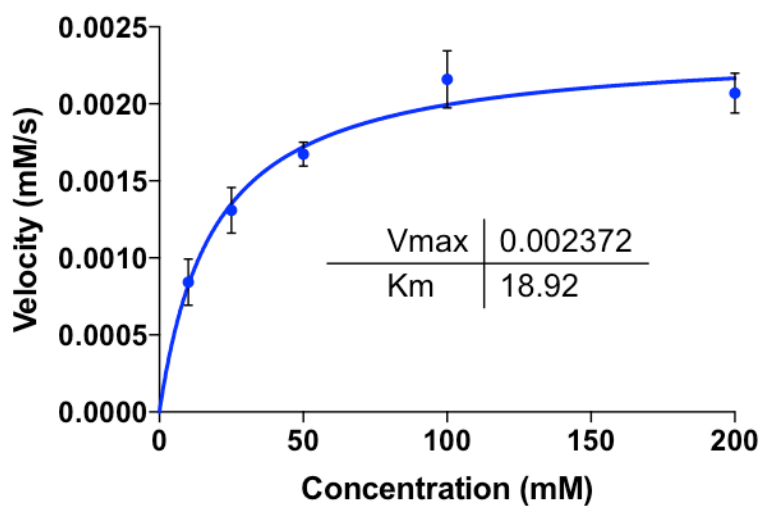


Figure S45. Michaelis-Menten plot of 1-boc-3-pyrrolidinone **18a**, $K_m = 18.9$ mM.

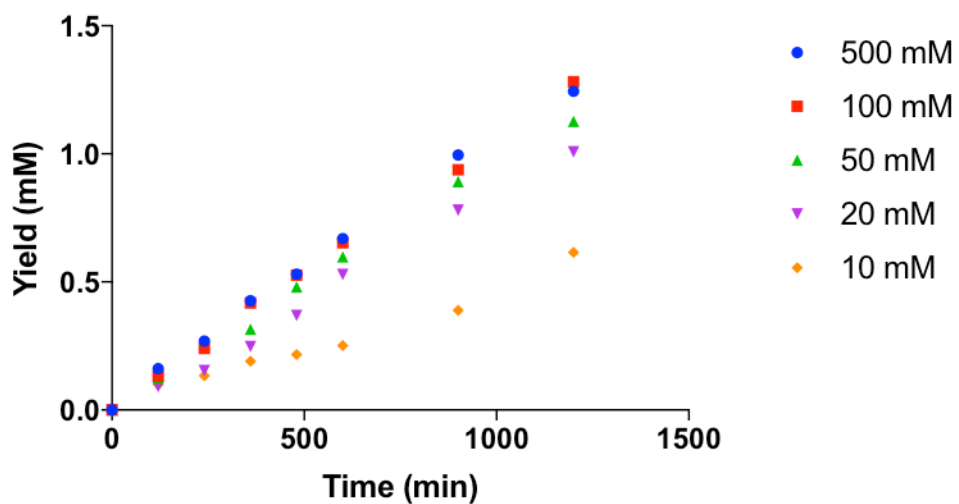


Figure S46. Effect of varying IPA 27 concentration on formation of 18b.²

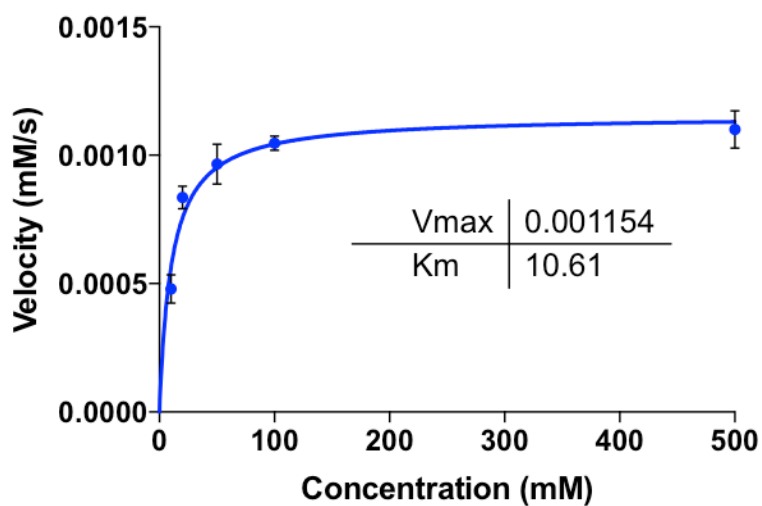


Figure S47. Michaelis-Menten plot of IPA 27, $K_m = 10.6$ mM

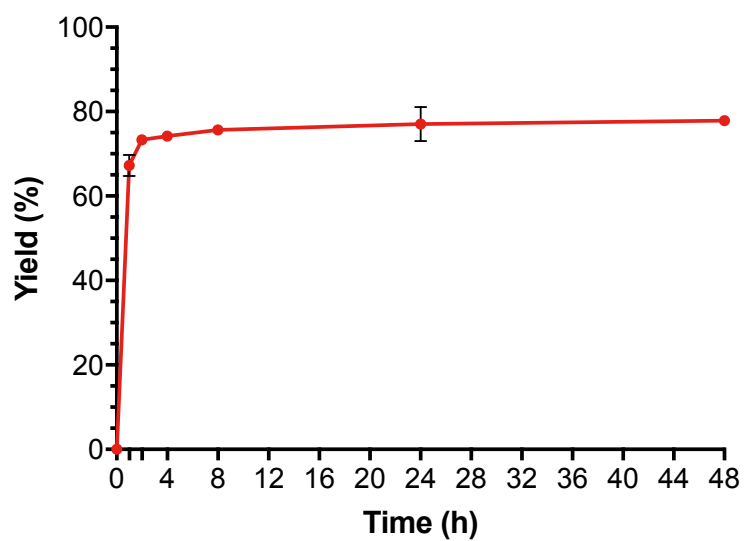


Figure S48. Percentage yield of **18b** using IPA **27** as an amine donor over time for pQR2189 using crude cell lysate.

12. NMR Spectra

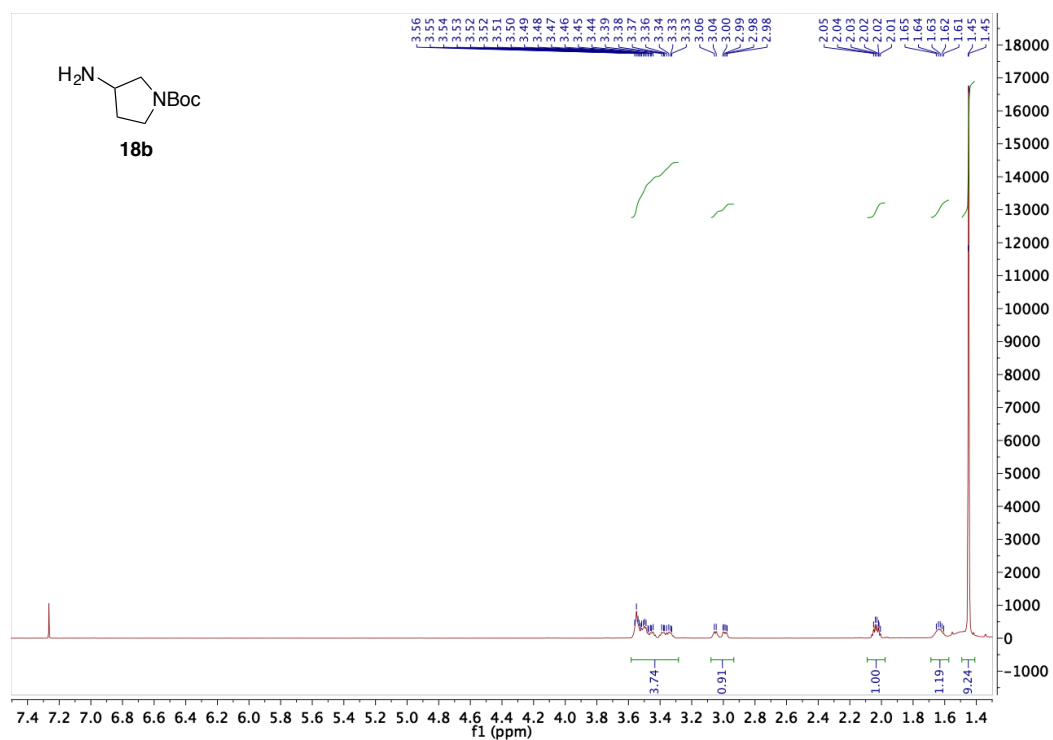


Figure S49. ¹H NMR spectrum of amine **18b** in CDCl₃.

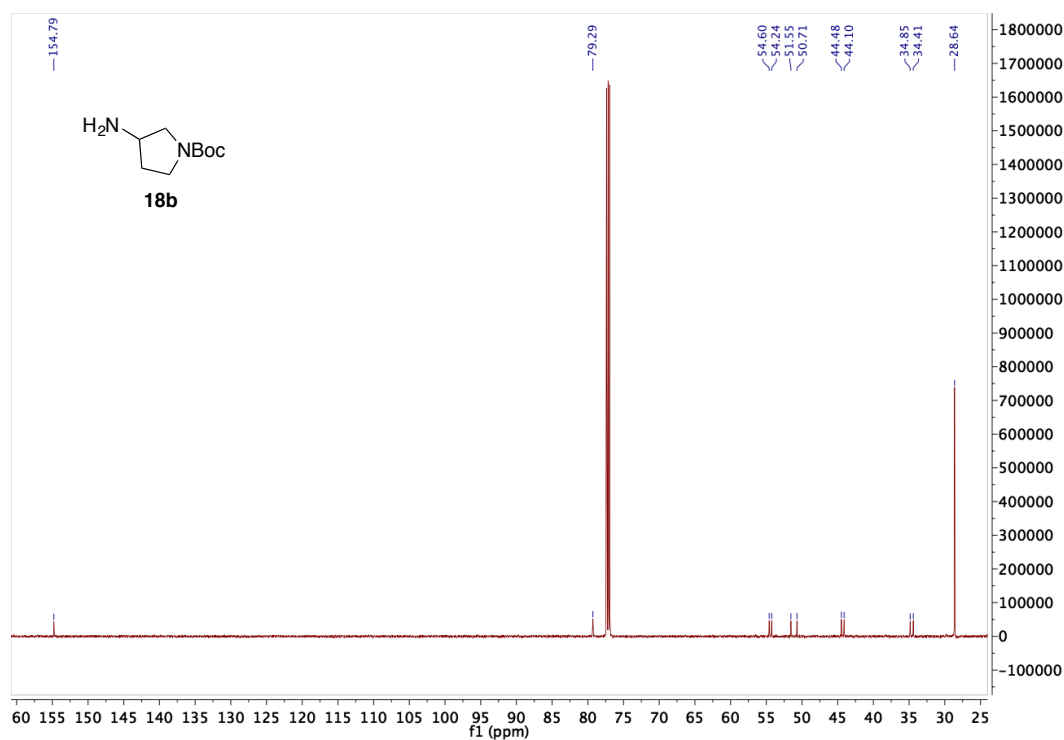


Figure S50. ¹³C NMR spectrum of amine **18b** in CDCl₃.

13. References

- 1 S. Schätzle, M. Höhne, E. Redestad, K. Robins and U. T. Bornscheuer, *Anal. Chem.*, 2009, **81**, 8244–8248.
- 2 N. Al-Haque, P. A. Santacoloma, W. Neto, P. Tufvesson, R. Gani and J. M. Woodley, *Biotechnol. Prog.*, 2012, **28**, 1186–1196.