

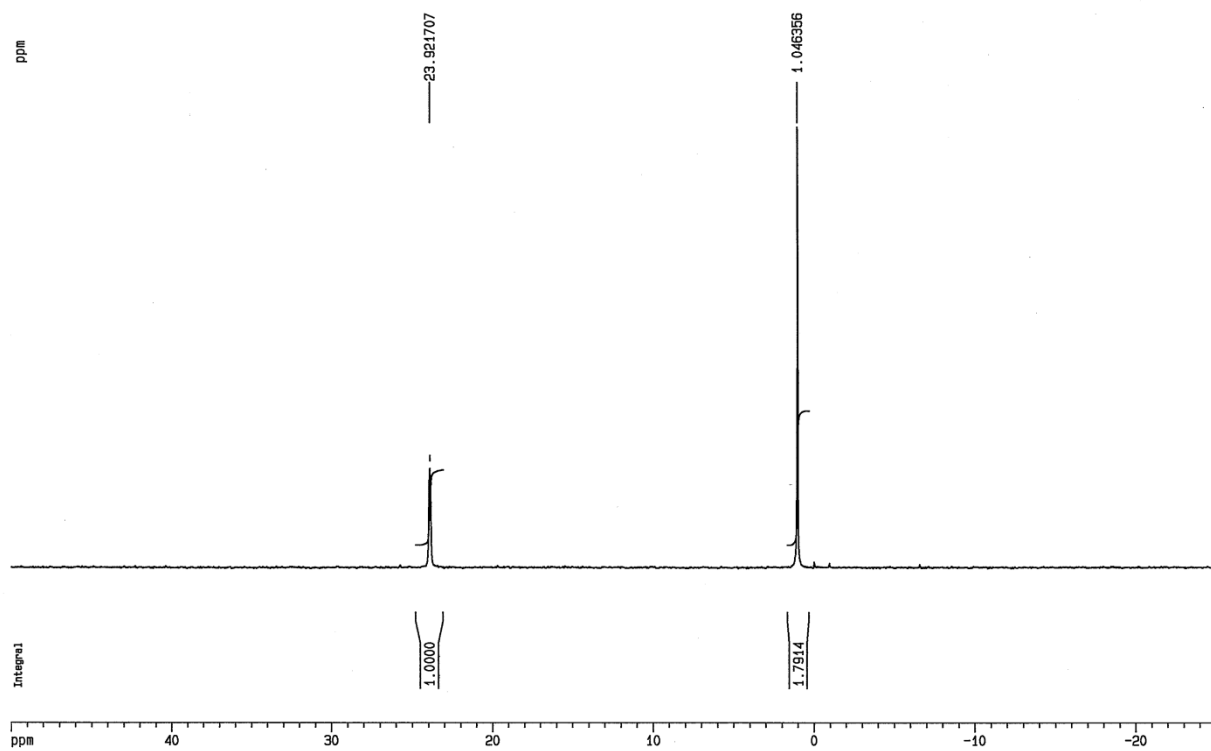
## ELECTRONIC SUPPORTING INFORMATION

### **Au@Zirconium-Phosphonate Nanoparticles as Effective Catalytic System for the Chemoselective and Switchable Reduction of Nitroarenes**

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**Figure S1a.**  $^{31}\text{P}$ NMR spectra of  $\text{ZP(AEP)}_{0.72}$

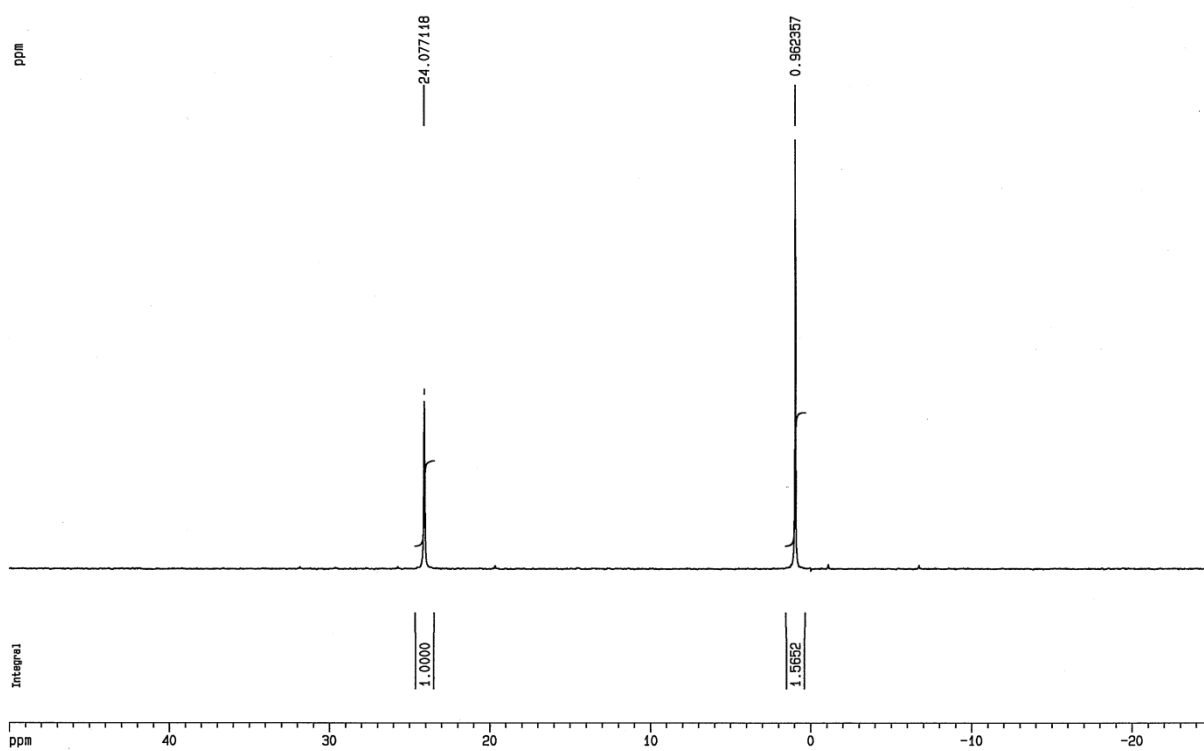


Figure S1b.  $^{31}\text{P}$ NMR spectra of ZP(AEP)<sub>0.78</sub>

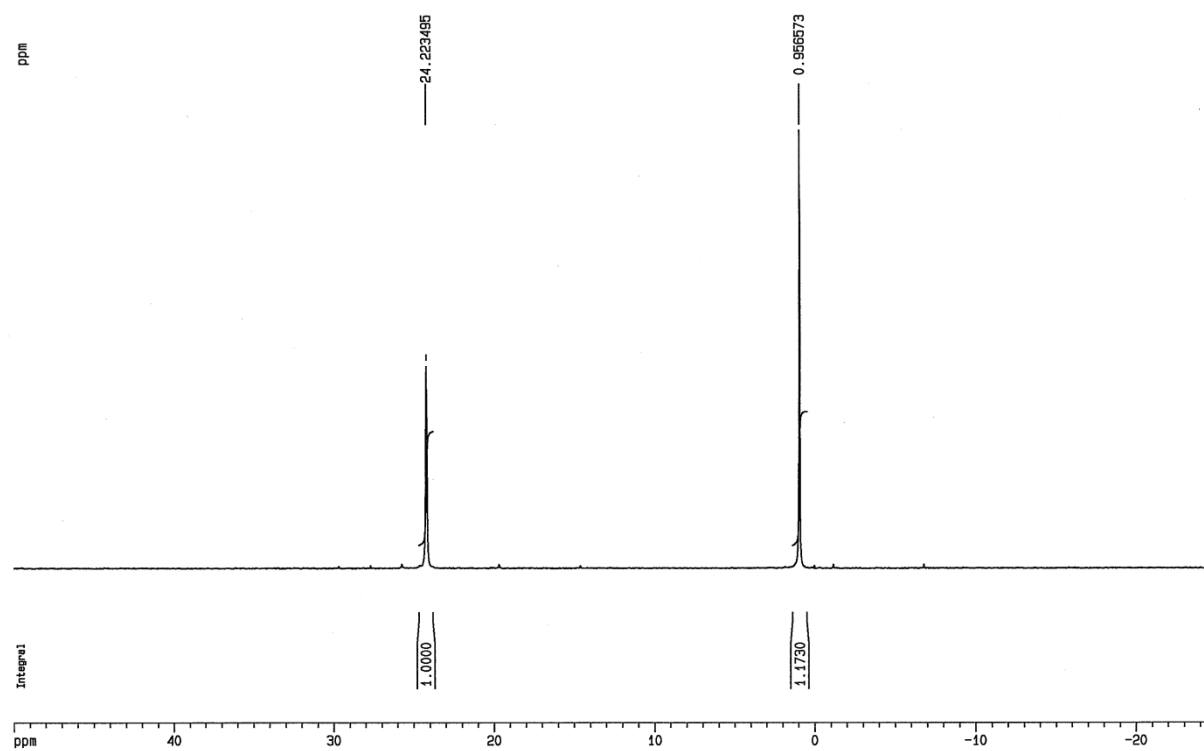
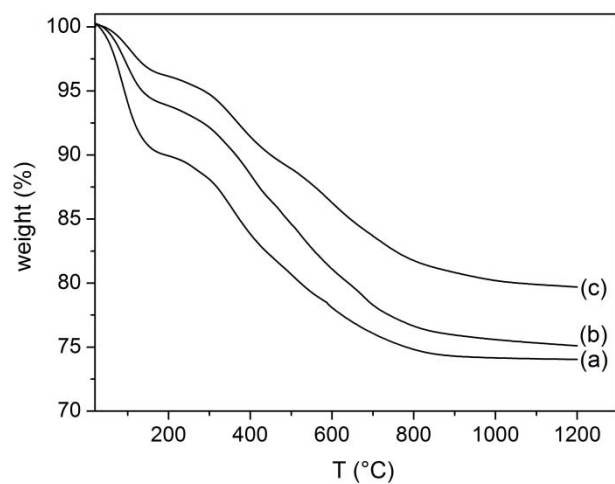
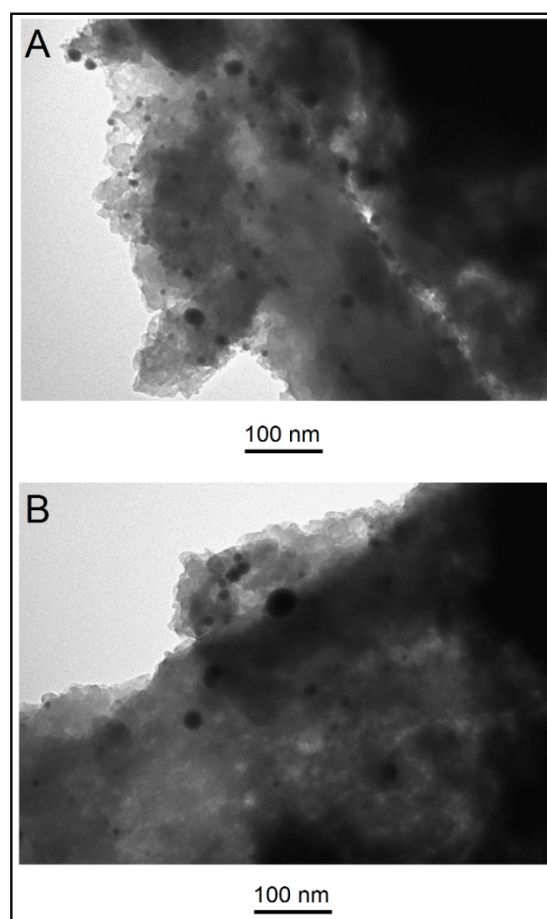


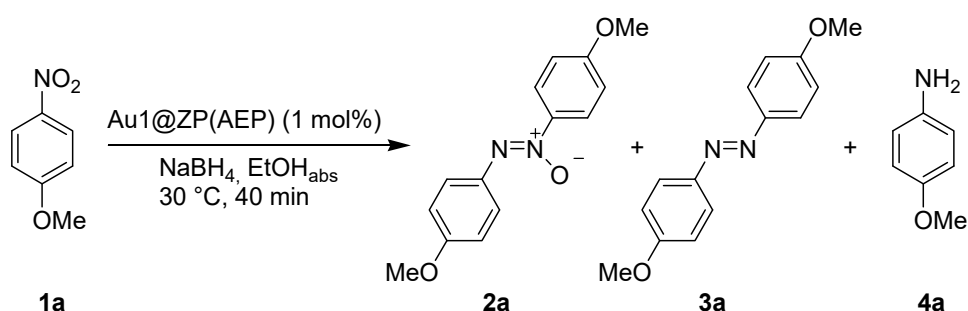
Figure S1c.  $^{31}\text{P}$ NMR spectra of ZP(AEP)<sub>0.92</sub>



**Figure S2.** Thermogravimetric curves of ZP(AEP)<sub>0.72</sub> (a); ZP(AEP)<sub>0.78</sub> (b); ZP(AEP)<sub>0.93</sub> (c)



**Figure S3.** Representative TEM image of Au1@ZP(AEP) after five runs in EtOH 96 % (A) and EtOH abs (B) under batch conditions.

**Table S1.** Recovery and reuse of catalyst in EtOH<sub>abs</sub> at 30 °C for a reaction time of 40 min.

Entry <sup>a</sup>	Run	Medium	C (%) <sup>b</sup>	2:3:4 ratio <sup>b</sup>
1	Run 1	EtOH <sub>abs</sub>	95	3:1:96
2	Run 2	EtOH <sub>abs</sub>	94	7:1:92
3	Run 3	EtOH <sub>abs</sub>	95	11:1:88
4	Run 4	EtOH <sub>abs</sub>	95	14:1:85
5	Run 5	EtOH <sub>abs</sub>	94	19:1:80

<sup>a</sup>Reaction condition: **1** (0.1 mmol), Au1@ZP(AEP) (1 mol%), NaBH<sub>4</sub> (6 eq), reaction medium: EtOH<sub>abs</sub> (1.8 mL), at 30 °C for 40 min. <sup>b</sup> Conversion and product ratios determined by GLC analyses.

#### Typical procedure for a sustainable protocol for the preparation of **4a** under flow conditions

A premixed mixture of **1a** (1.53 g, 10 mmol), NaBH<sub>4</sub> (1.13 g, 3 eq) and EtOH abs. (40 mL) was charged into a glass column functioning as a reservoir. The equipment was connected, by using the appropriate valves, to a pump and installed into a box thermostated at 30 °C. The reaction mixture was continuously pumped (flow rate: 0.1 mL · min<sup>-1</sup>; residence time: 50 min) through the catalyst column (15 cm, stainless steel HPLC column) equipped with a 120 psi back pressure regulator, and the reaction was monitored by GC. Product was collected in fractions and gold content in the EtOH solution was periodically measured by ICP-OES analysis. After completion 20 mL of EtOH was used to wash the catalyst inside the reactor. After removal and recovery of the solvent via distillation (90% of solvent recovered) the product was separated simple by washing the residue with cyclopenthyl methyl ether (3 x 15 mL). The solvent was evaporated by distillation (95% of solvent recovered) obtaining the pure product (yield of **4a** 97%). The catalyst inside the column was dried under N<sub>2</sub> flow at 180 °C and reused in the next cycle.

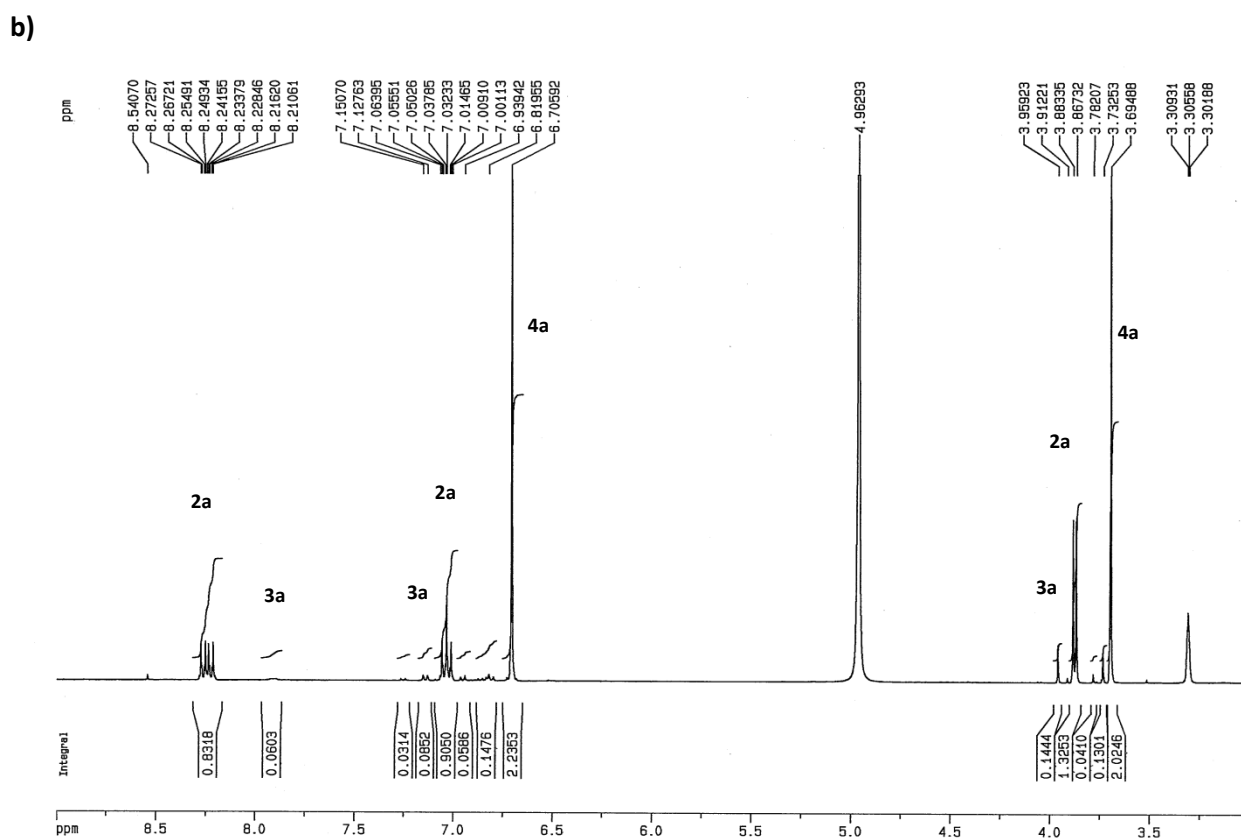
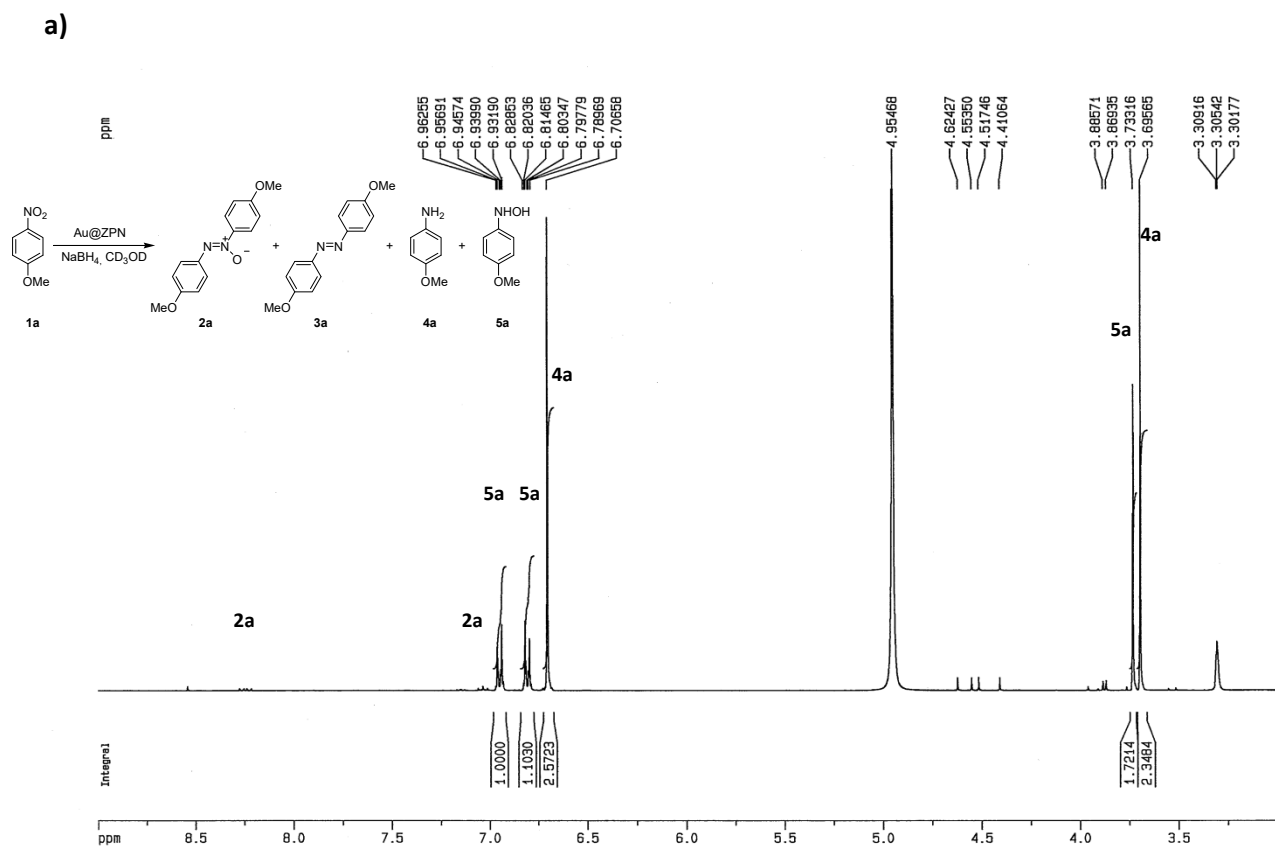
E-factor calculation for product **4a** (yield 97%): E-factor = [1.5 g (4-nitroanisole, **1a**) + 1.13 g (NaBH<sub>4</sub>) + 32 g (EtOH) + 39 g (CPME) – 1.2 g (4-methoxyaniline, **4a**) – 29 g (recovered EtOH)] – 37 g (recovered CPME) / 1.2 g (4-methoxyaniline, **4a**) = 5.4

#### Typical procedure for a sustainable protocol for the preparation of **2a** under flow conditions

A premixed mixture of **1a** (2.29 g, 15 mmol), NaBH<sub>4</sub> (1.7 g, 3 eq) and EtOH 96 %. (60 mL) was charged into a glass column functioning as a reservoir. The equipment was connected, by using the appropriate valves, to a pump and installed into a box thermostated at 30 °C. The reaction mixture was continuously pumped

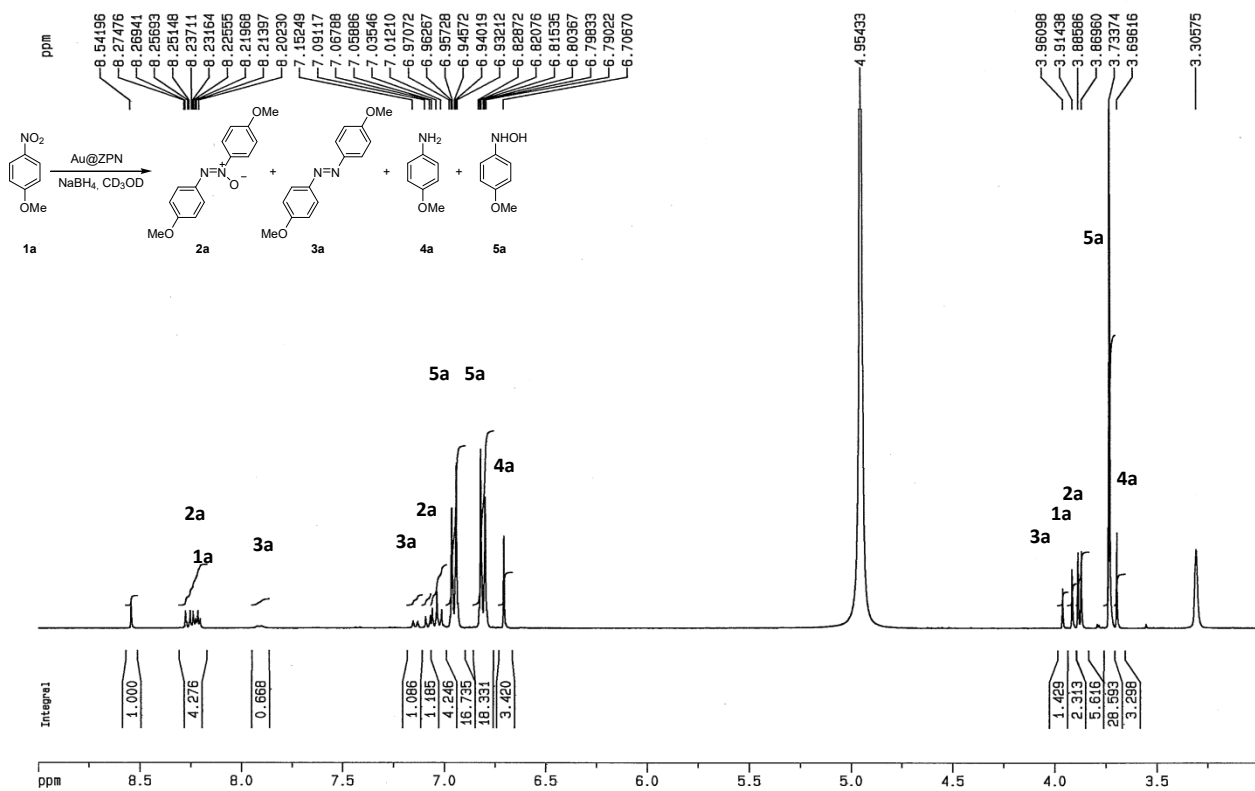
(flow rate: 0.1 mL · min<sup>-1</sup>; residence time: 50 min) through the catalyst column (15 cm, stainless steel HPLC column) equipped with a 120 psi back pressure regulator, and the reaction was monitored by GC. Product was collected in fractions and gold content in the EtOH solution was periodically measured by ICP-OES analysis. After completion 20 mL of EtOH was used to wash the catalyst inside the reactor. After removal and recovery of the solvent via distillation (90% of solvent recovered) the product was separated simple by washing the residue with cyclopenthyl methyl ether (3 x 18 mL). The solvent was evaporated by distillation (95% of solvent recovered) obtaining the pure product (yield of **2a** 90%). The catalyst inside the column was dried under N<sub>2</sub> flow at 180 °C and reused in the next cycle.

E-factor calculation for product **2a** (yield 90%): E-factor = [2.3 g (4-nitroanisole, **1a**) + 1.7 g (NaBH<sub>4</sub>) + 47 g (EtOH) + 46 g (CPME) – 3.5 g (azoxyderivative, **2a**) – 42 g (recovered EtOH)] – 44 g (recovered CPME)] / 3.5 g (4-methoxyaniline, **2a**) = 2.1

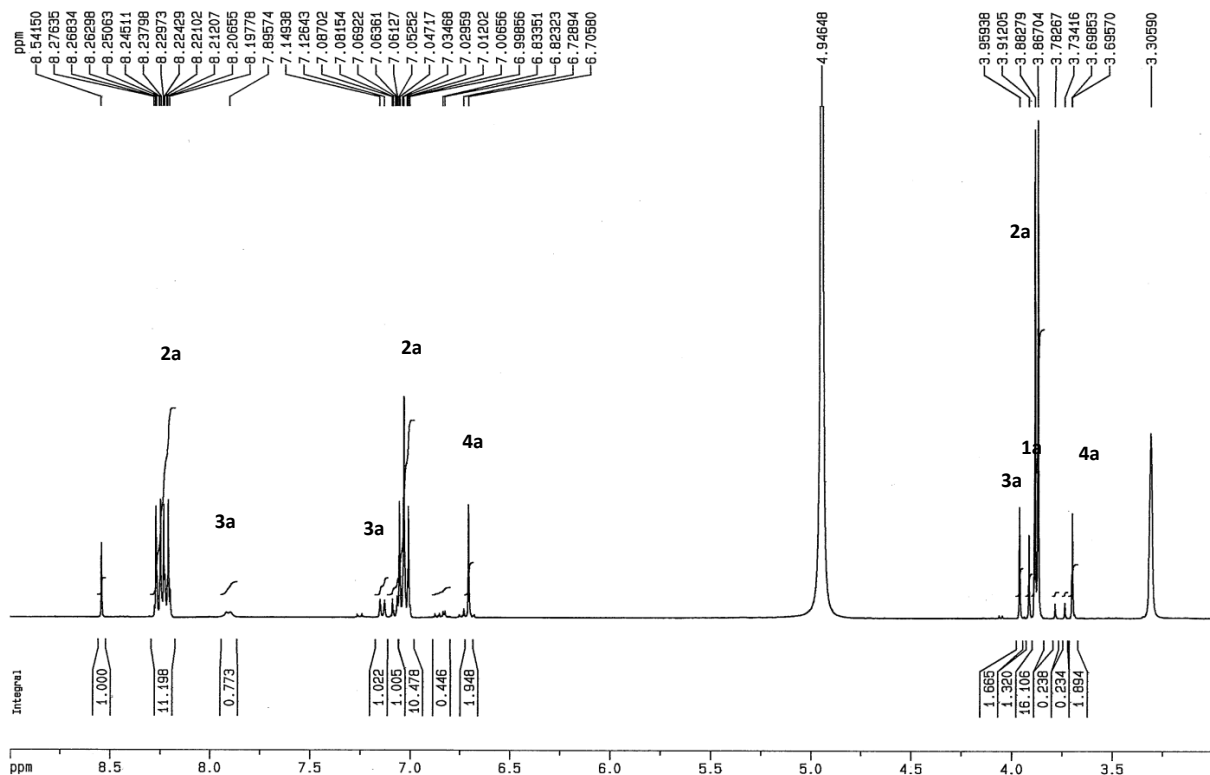


**Figure S4:**  $^1\text{H-NMR}$  spectrum of crude mixture of reduction of **1a** in  $\text{CD}_3\text{OD}$  with  $\text{NaBH}_4$ , reaction time 20 min, immediately after filtration on celite (**a**) and after 1h from filtration (**b**)

a)



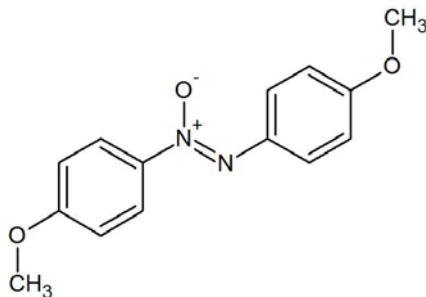
b)





**FigureS5:**  $^1\text{H-NMR}$  spectrum of crude mixture of reduction of **1a** in  $\text{CD}_3\text{OD}/\text{D}_2\text{O}$  (96:4) with  $\text{NaBH}_4$ , reaction time 20 min, immediately after filtration on celite (**a**) and after 1h from filtration (**b**).

**4,4'-dimethoxy-azoxybenzene (2a)** <sup>[1-2]</sup>



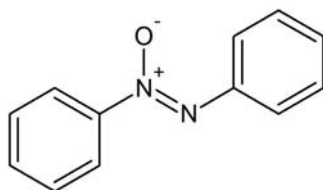
Yellow pale solid, mp 117-119 °C.

$^1\text{H-NMR}$  ( $\text{CDCl}_3$ , 400 MHz)  $\delta$ : 3.869 (s, 3H), 3.872 (s, 3H), 6.95 (d, 2H,  $J = 8.9$  Hz), 6.97 (d, 2H,  $J = 8.7$  Hz), 8.24 (d, 2H,  $J = 9.1$  Hz), 8.28 (d, 2H,  $J = 9.1$  Hz).

$^{13}\text{C-NMR}$  ( $\text{CDCl}_3$ , 100.6 MHz)  $\delta$ : 55.4, 55.6, 113.5 (2C), 113.7 (2C), 123.7 (2C), 127.8 (2C), 137.9, 141.6, 160.1, 161.8.

GC-MS ( $m/z$ ): 258 (100), 215 (7), 137 (10), 121 (70), 107 (57), 92 (29), 77 (36), 64 (21), 52 (9%).

**Azoxybenzene (2b)** <sup>[1-4]</sup>



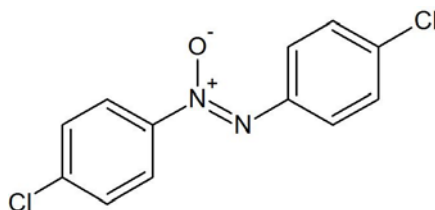
Yellow pale solid, mp 36-37 °C

$^1\text{H-NMR}$  ( $\text{CDCl}_3$ , 400 MHz)  $\delta$ : 7.38-7.42 (m, 1H), 7.47-7.59 (m, 5H), 8.17 (d, 2H,  $J = 7.6$  Hz), 8.31 (d, 2H,  $J = 8.2$  Hz).

GC-MS ( $m/z$ ): 198 (100), 170 (38), 169 (97), 141 (56), 115 (25), 105 (65), 91 (100), 77 (100), 65 (92), 64 (72), 51 (100).

$^{13}\text{C-NMR}$  ( $\text{CDCl}_3$ , 100.6 MHz)  $\delta$ : 122.3 (2C), 125.5 (2C), 128.7 (2C), 128.8 (2C), 129.6, 131.6, 143.9, 148.3.

**4,4'-dichloro-azoxybenzene (2c)** <sup>[1-4]</sup>



Yellow pale solid, mp 152-155 °C

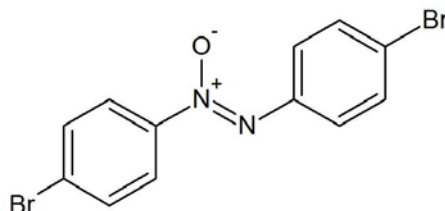
$^1\text{H-NMR}$  ( $\text{CDCl}_3$ , 400 MHz)  $\delta$ : 7.44 (d, 2H,  $J = 8.8$  Hz), 7.48 (d, 2H,  $J = 8.8$  Hz), 8.16 (d, 2H,  $J = 8.8$  Hz), 8.25 (d, 2H,  $J$

= 8.8 Hz).

$^{13}\text{C}$ -NMR ( $\text{CDCl}_3$ , 100.6 MHz)  $\delta$ : 123.7 (2C), 127.1 (2C), 128.9 (2C), 129.0 (2C), 135.2, 138.0, 142.2, 146.5.

GC-MS ( $m/z$ ): 268 (29), 266 (44), 203 (11), 168 (9), 139 (18), 125 (45), 111 (100), 90 (32), 75 (44), 63 (18), 50 (13).

#### 4,4'-dibromo-azoxybenzene (2d) <sup>[1-4]</sup>



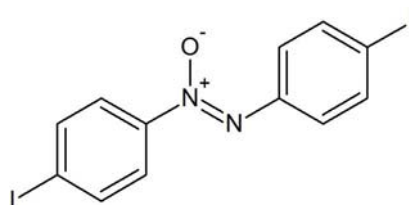
Yellow pale solid, mp 169-171 °C

$^1\text{H}$ -NMR ( $\text{CDCl}_3$ , 400 MHz)  $\delta$ : 7.61 (d, 2H, J = 8.8 Hz), 7.66 (d, 2H, J = 9.0 Hz), 8.08 (d, 2H, J = 8.8 Hz), 8.18 (d, 2H, J = 9.0 Hz).

$^{13}\text{C}$ -NMR ( $\text{CDCl}_3$ , 100.6 MHz)  $\delta$ : 123.6, 123.90 (2C), 126.5, 127.2 (2C), 131.98 (2C), 132.04 (2C), 142.6, 147.1.

GC-MS ( $m/z$ ): 358 (31), 356 (62), 354 (32), 249 (12), 247 (12), 185 (26), 183 (19), 171 (29), 169 (31), 168 (46), 157 (96), 155 (100), 145 (10), 143 (10), 140 (11), 90 (66), 76 (51), 75 (47), 63 (38), 50 (31).

#### 4,4'-diiodo-azoxybenzene (2e) <sup>[1,21]</sup>



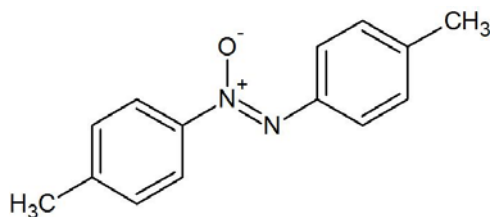
Yellow pale solid, mp 199-200 °C

$^1\text{H}$ -NMR ( $\text{CDCl}_3$ , 400 MHz)  $\delta$ : 7.81 (d, 2H, J = 8.8 Hz), 7.5 (d, 2H, J = 8.9 Hz), 7.92 (d, 2H, J = 8.8 Hz), 8.02 (d, 2H, J = 8.9 Hz).

$^{13}\text{C}$ -NMR ( $\text{CDCl}_3$ , 100.6 MHz)  $\delta$ : 95.7, 98.7, 123.9 (2C), 127.2 (2C), 138.0 (2C), 138.1 (2C), 143.1, 147.8.

GC-MS ( $m/z$ ): 450 (94), 295 (9), 233 (13), 231 (23), 217 (17), 203 (100), 168 (34), 140 (18), 90 (52), 76 (75), 63 (30), 50 (27).

#### 4,4'-dimethyl-azoxybenzene <sup>[1-4]</sup>



Yellow pale solid, mp 65-68 °C

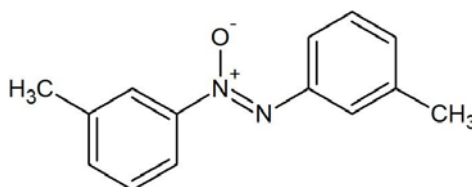
<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz) δ: 2.42 (s, 3H), 2.44 (s, 3H), 7.29 (m, 4H), 8.12 (d, 2H, J = 8.4 Hz), 8.19 (d, 2H, J = 8.6 Hz).

<sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz) δ: 21.3, 21.5, 122.1 (2C), 125.6 (2C), 129.3 (4C), 140.0, 141.8, 141.9, 146.2.

GC-MS (m/z): 226 (48), 198 (47), 183 (21), 155 (8), 119 (9), 105 (24), 104 (38), 91 (100), 79 (34), 77 (35), 65 (50), 51 (11).

### 3,3'-dimethyl-azoxybenzene <sup>[1-4]</sup>

C<sub>14</sub>H<sub>14</sub>N<sub>2</sub>O - 226.274 g/mol - P.f. = liq



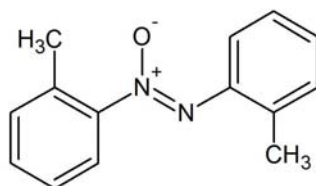
Yellow pale solid, mp 34-36 °C

<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz) δ: 2.44 (s, 3H), 2.47 (s, 3H), 7.21 (d, 1H, J = 7.4 Hz), 7.35-7.43 (m, 3H), 7.97 (s, 1H), 7.98 (d, 1H, J = 7.4), 8.08-8.11 (m, 2H).

<sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz) δ: 21.4, 21.5, 119.5, 122.5, 122.8, 126.0, 128.5, 128.6, 130.3, 132.3, 138.4, 139.0, 144.0, 148.4.

GC-MS (m/z): 226 (48), 198 (47), 183 (21), 155 (8), 119 (9), 105 (24), 104 (38), 91 (100), 79 (34), 77 (35), 65 (50), 51 (11).

### 2,2'-dimethyl-azoxybenzene <sup>[1-3]</sup>



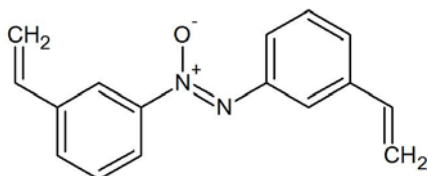
Yellow pale solid, mp 53-56 °C

<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz) δ: 2.38 (s, 3H), 2.53 (s, 3H), 7.24-7.42 (m, 6H, J = 8.4 Hz), 7.68 (d, 1H, J = 8.7 Hz), 8.04 (d, 1H, J = 7.8 Hz).

<sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz) δ: 18.39, 18.42, 121.5, 123.6, 126.0, 126.6, 128.6, 130.0, 130.8, 131.2, 131.8, 134.1, 142.8, 149.5 .

GC-MS (m/z): 226 (32), 225 (45), 211 (92), 209 (33), 194 (7), 183 (9), 168 (10), 119 (7), 104 (32), 91 (100), 79 (27), 77 (26), 65 (52), 51 (13).

### 3,3'-divinyl azoxybenzene <sup>[5]</sup>



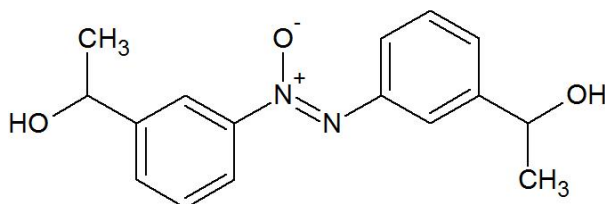
Yellow solid, mp 34-36 °C

<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz) δ: 5.33 (d, 1H, J = 10.9 Hz), 5.40 (d, 1H, J = 10.9 Hz), 5.84 (d, 1H, J = 17.6 Hz), 5.90 (d, 1H, J = 17.6 Hz), 6.78 (dd, 1H, J<sub>1</sub> = 17.6 Hz, J<sub>2</sub> = 10.4 Hz), 6.80 (dd, 1H, J<sub>1</sub> = 17.6 Hz, J<sub>2</sub> = 10.4 Hz), 7.40-7.50 (m, 3H), 7.60 (d, 1H, J = 7.6 Hz), 8.09 (m, 1H), 8.18 (m, 1H), 8.20 (s, 1H), 8.34 (s, 1H).

<sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz) δ: 114.8, 115.9, 119.9, 121.5, 123.4, 124.7, 127.3, 128.8, 128.9, 129.3, 135.5, 136.2, 138.2, 138.6, 144.2, 148.6.

GC-MS (m/z): 250 (75), 222 (33), 221 (100), 208 (2%), 207 (9), 195 (9), 178 (12), 165 (7), 131 (7), 116 (20), 103 (100), 91 (44), 90 (45), 89 (36), 77 (83), 65 (20), 51 (25).

### 3,3'-di(1-idroxyethyl)azoxybenzene



Yellow pale oil

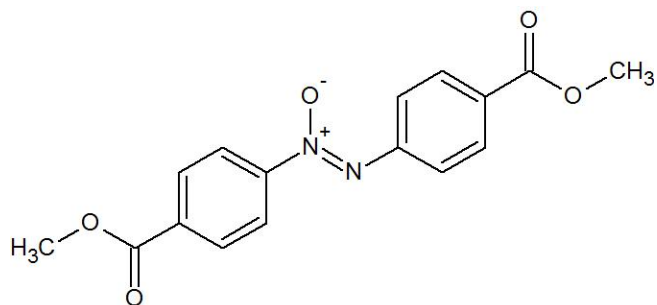
<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz) δ: 1.50 (s, 3H), 1.52 (s, 3H), 2.52 (s broad, 2H), 4.94 (m, 2H), 7.37-7.48 (m, 3H), 7.53 (d, 1H, J = 7.6 Hz), 8.04 (d, 1H, J = 7.8 Hz), 8.08 (s, 1H), 8.13 (d, 1H, J = 8.0 Hz), 8.25 (s, 1H).

<sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz) δ: 25.1, 25.3, 69.7, 70.0, 119.3, 121.2, 122.7, 124.3, 126.7, 128.6, 128.8, 128.9, 143.9, 146.5, 147.1, 148.2.

GC-MS (m/z): 286 (41), 270 (4), 242 (5), 195 (5), 181 (15), 169 (8), 149 (8), 121 (88), 103 (100), 93 (30), 91 (24), 77 (56), 65 (20), 51 (11).

### 4,4'-(metoxycarbonyl)azoxybenzene <sup>[3,4]</sup>

C<sub>16</sub>H<sub>14</sub>N<sub>2</sub>O<sub>5</sub> - 314.293 g/mol - P.f. = 197-200 °C



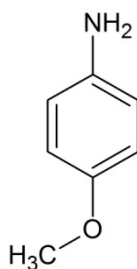
Yellow pale solid, mp 197-200 °C

$^1\text{H-NMR}$  ( $\text{CDCl}_3$ , 400 MHz)  $\delta$ : 3.95 (s, 3H), 3.97 (s, 3H), 8.14-8.20 (m, 6H), 8.38 (d, 2H,  $J = 8.6$  Hz).

$^{13}\text{C-NMR}$  ( $\text{CDCl}_3$ , 100.6 MHz)  $\delta$ : 52.3, 52.6, 122.5 (2C), 125.3 (2C), 130.2 (2C), 130.4 (2C), 130.7, 133.3, 146.9, 150.9, 165.7, 166.2.

GC-MS ( $m/z$ ): 314 (27), 298 (13), 283 (11), 227 (7), 163 (20), 151 (10), 135 (100), 120 (22), 119 (18), 104 (28), 103 (30), 90 (10), 76 (34), 63 (15), 59 (14), 50 (13).

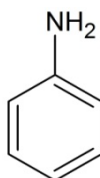
#### 4-methoxyaniline (4a) <sup>[6-8]</sup>



$^1\text{H-NMR}$  ( $\text{CDCl}_3$ , 400 MHz)  $\delta$ : 3.21 (s broad, 2H), 3.74 (s, 3H), 6.64 (d, 2H,  $J = 8.8$  Hz), 6.75 (d, 2H,  $J = 8.8$  Hz).

GC-MS ( $m/z$ ): 123 (70%), 109 (7), 108 (100%), 80 (53), 65 (5), 53 (15%).

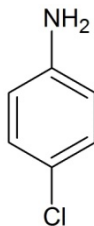
#### Aniline (4b) <sup>[4,6-8]</sup>



$^1\text{H-NMR}$  ( $\text{CDCl}_3$ , 400 MHz)  $\delta$ : 3.47 (s broad, 2H), 6.70 (m, 2H), 6.77 (m, 1H), 7.16 (m 1H).

GC-MS ( $m/z$ ): 93 (100), 77 (7), 66 (35), 65 (21), 61 (10), 51 (10).

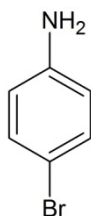
**4-chloroaniline (4c)** <sup>[4, 6-8]</sup>



<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz) δ: 3.99 (s broad, 2H), 6.61 (d, 2H, J = 8.8 Hz), 7.10 (d, 2H, J = 8.8 Hz).

GC-MS (m/z): 129 (33), 127 (100), 102 (2%), 100 (9), 92 (13), 65 (12), 63 (5%).

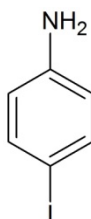
**4-bromoaniline (4d)** <sup>[4,6,8]</sup>



<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 200 MHz) δ: 3.52 (s broad, 2H), 6.56 (d, 2H, J = 8.8 Hz), 7.24 (d, 2H, J = 8.8 Hz).

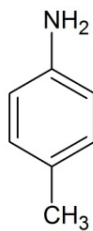
GC-MS (m/z): 173 (100), 171 (100), 145 (11), 143 (11), 119 (8), 117 (8), 92 (100), 91 (42), 74 (14), 66 (31), 65 (100), 64 (44%), 63 (70), 62 (36), 61 (23), 52 (29), 51 (13), 50 (23).

**4-iodoaniline (4e)** <sup>[8]</sup>



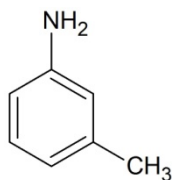
GC-MS (m/z): 219 (100), 127 (12), 92 (42), 65 (43), 63 (11), 62 (6), 52 (5).

**4-methylaniline (4f)** <sup>[4,7,8]</sup>



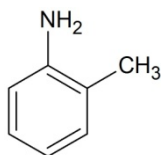
<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz) δ: 2.24 (s, 3H), 3.30 (s broad, 2H), 6.62 (d, 2H, J = 7.9 Hz), 6.97 (d, 2H, J = 7.9 Hz).  
GC-MS (m/z): 107 (84), 106 (100), 89 (8), 79 (15), 77 (19), 63 (6), 52 (9), 51 (10).

### 3-methylaniline (4g) <sup>[4,8]</sup>



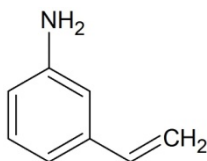
<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz) δ: 2.26 (s, 3H), 3.49 (s broad, 2H), 6.53-6.62 (m, 3H), 7.06 (m, 1H)  
GC-MS (m/z): 107 (100), 106 (100), 91 (16), 89 (20), 80 (30), 79 (65), 77 (69), 65 (19), 63 (17), 53 (24), 52 (24), 51 (25).

### 2-methylaniline (4e) <sup>[8]</sup>



<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz) δ: 2.19 (s, 3H), 3.83 (s broad, 2H), 6.70-6.75 (m, 2H), 7.03-7.07 (m, 2H).  
GC-MS (m/z): 107 (100), 106 (100), 91 (4), 80 (10), 79 (16), 77 (24), 63 (6), 53 (10), 52 (12), 51 (12)..

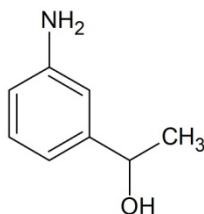
### 3-vinylaniline (4i) <sup>[6,7]</sup>



<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz) δ: 3.58 (s broad, 2H), 5.20 (dd, 1H, J 10.8, 0.8 Hz), 5.70 (dd, 1H, J = 17.6, 0.9 Hz), 6.58-6.67 (m, 2H), 6.74 (s, broad, 1H), 6.83 (d, 1H, J = 7.6), 7.11 (m, 1H).

GC-MS (m/z): 119 (100), 118 (58), 117 (17), 104 (7), 93 (12), 91 (34), 77 (4), 65 (14), 63 (8), 51 (6%).

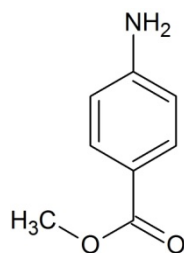
**3-(1-idroxyethyl)aniline (4j)** <sup>[6,7,9]</sup>



<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 200 MHz) δ: 1.47 (d, 3H, J = 6.4 Hz), 2.73 (s broad, 2H), 4.81 (q, 1H, J = 6.4 Hz), 6.57-6.63 (m, 1H), 6.72-6.77 (m, 2H), 7.14 (m, 1H).

GC-MS (m/z): 137 (44), 122 (23), 94 (100), 93 (17), 91 (6), 77 (26), 65 (15), 51 (5).

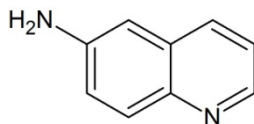
**4-(metoxycarbonyl)aniline (4k)** <sup>[4,6,7]</sup>



<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 200 MHz) δ: 3.85 (s, 3H), 4.06 (s broad, 2H), 6.63 (d, 2H, J = 8.2 Hz), 7.85 (d, 2H, J = 8.2 Hz).

GC-MS (m/z): 151 (100), 121 (50), 120 (100), 108 (9), 93 (2), 92 (100), 66 (210), 65 (100%), 63 (37), 52 (18), 51 (6).

**6-aminoquinoline (4l)** <sup>[7]</sup>



<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 200 MHz) δ: 3.92 (s broad, 2H), 6.89 (d, 1H, J = 2.6 Hz), 7.15 (dd, 1H, J = 9.0, 2.6 Hz), 7.26 (dd, 1H, J = 8.4, 4.2 Hz), 7.89 (dd, 1H, J = 8.4, 1.6 Hz), 7.90 (d, 1H, J = 9.0 Hz), 8.65 (dd, 1H, J = 4.2, 1.6 Hz).

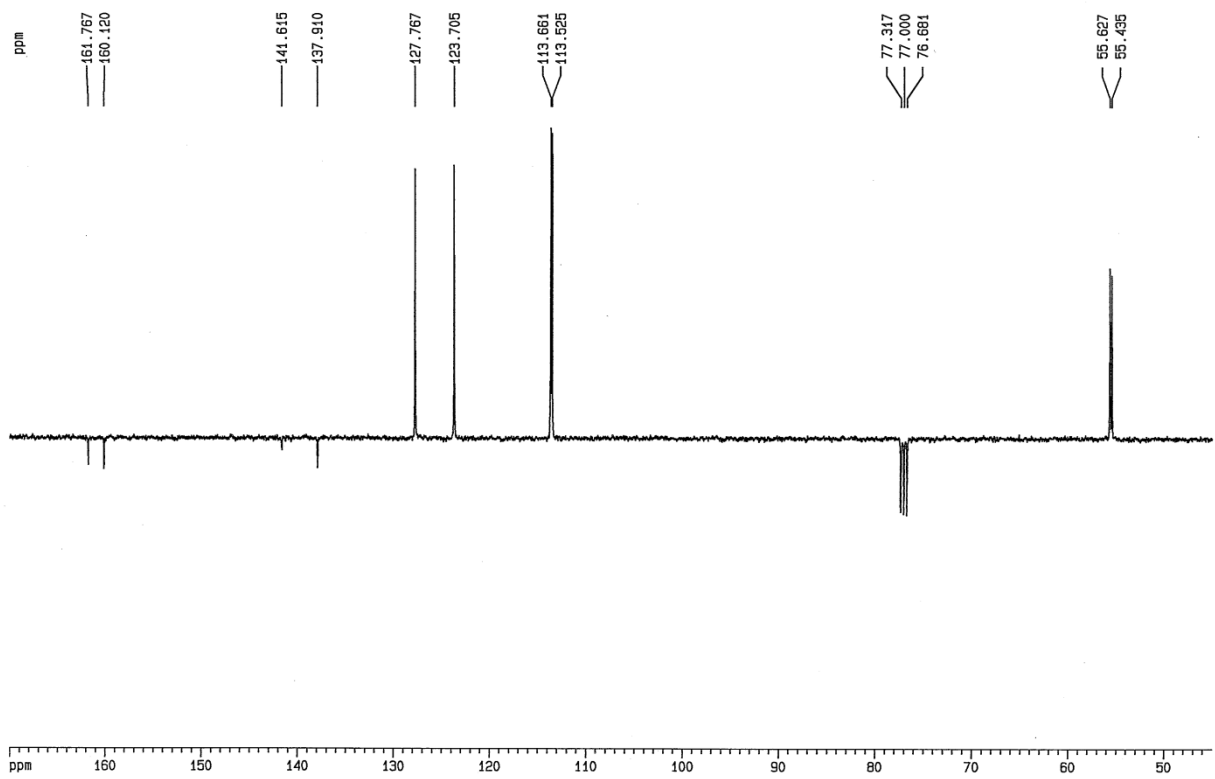
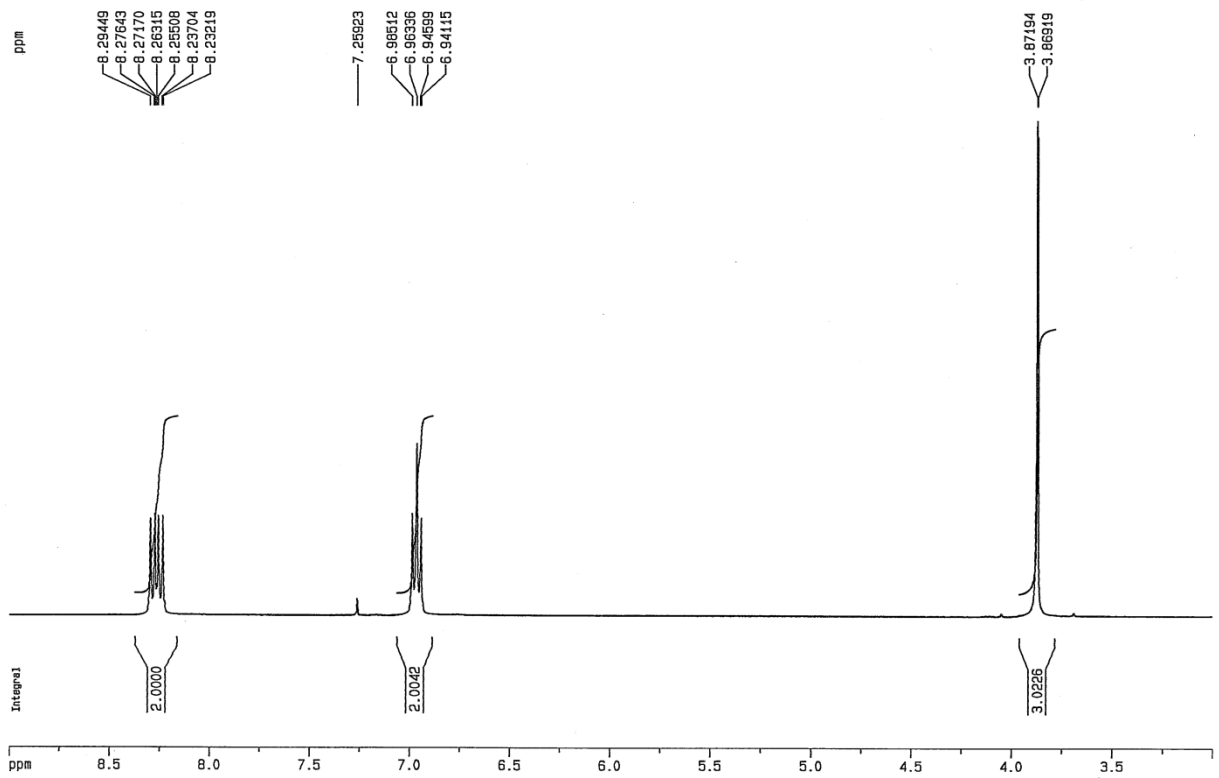
GC-MS (m/z): 145 (100), 144 (100), 127 (8), 118 (53), 117 (100), 116 (100), 91 (27), 90 (100), 89 (100), 72 (46), 63 (75), 52 (32), 51 (21).



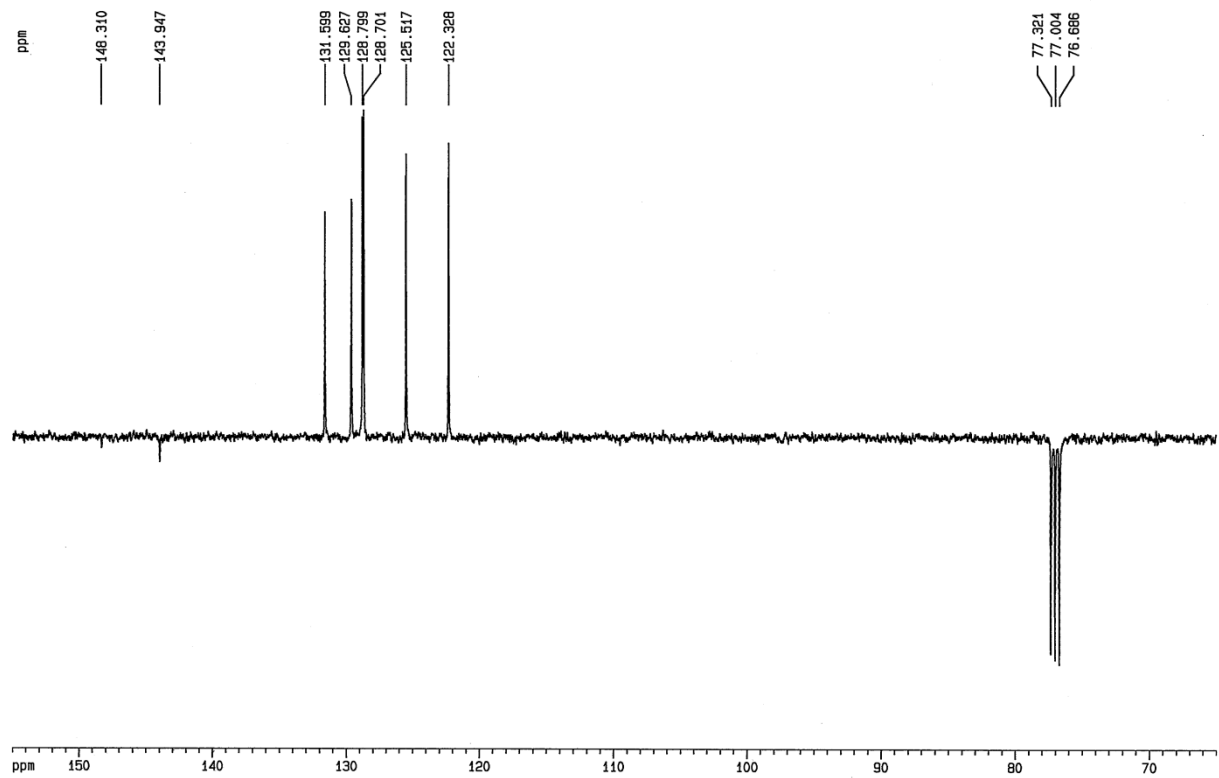
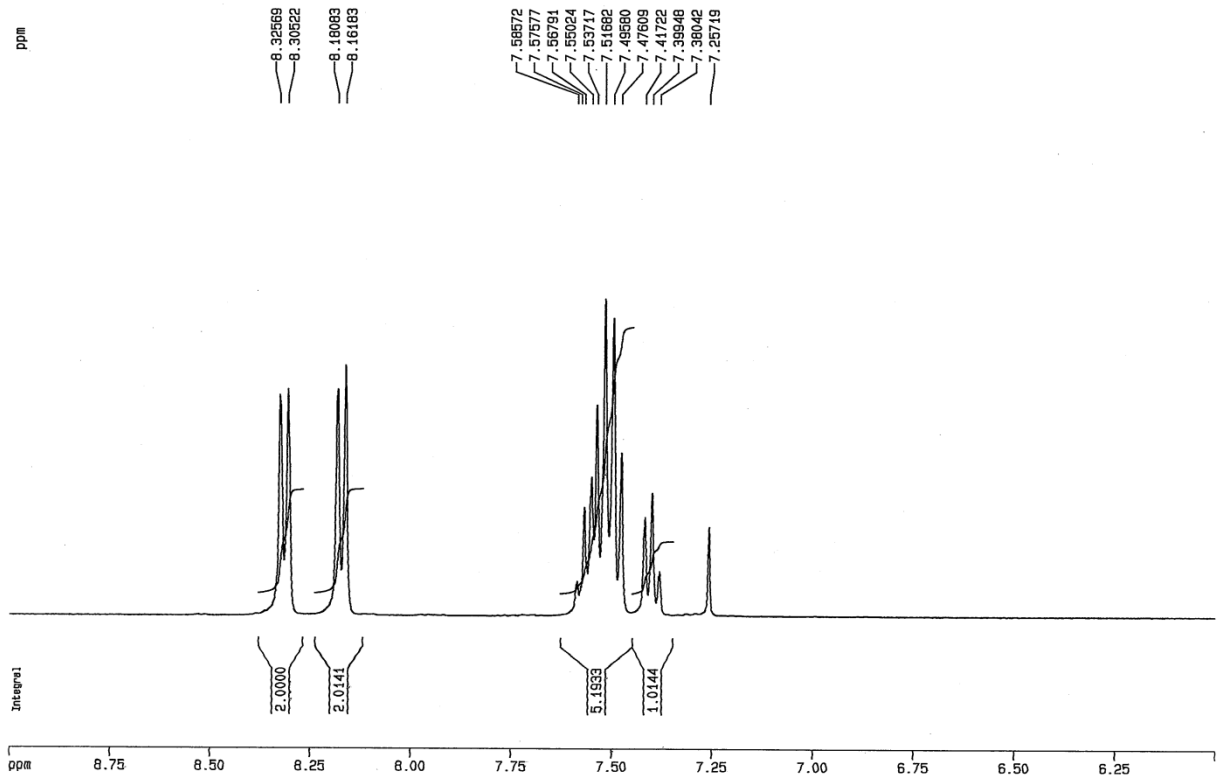
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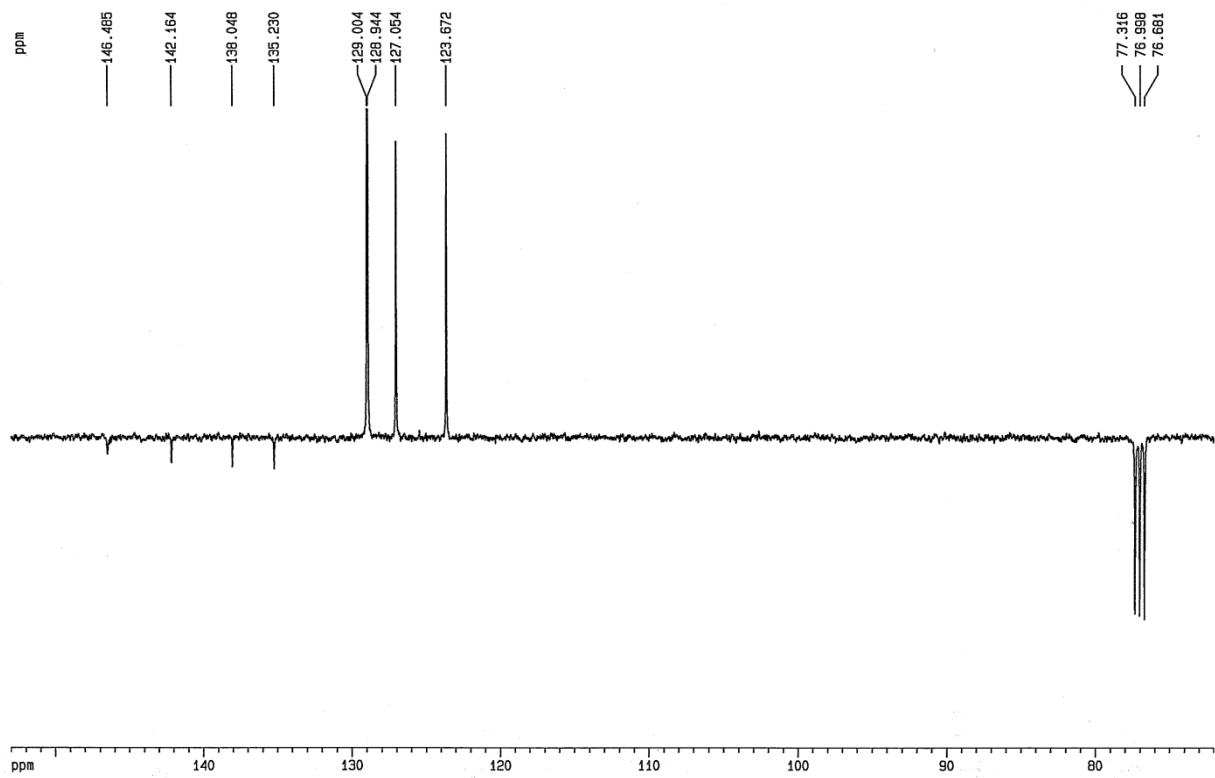
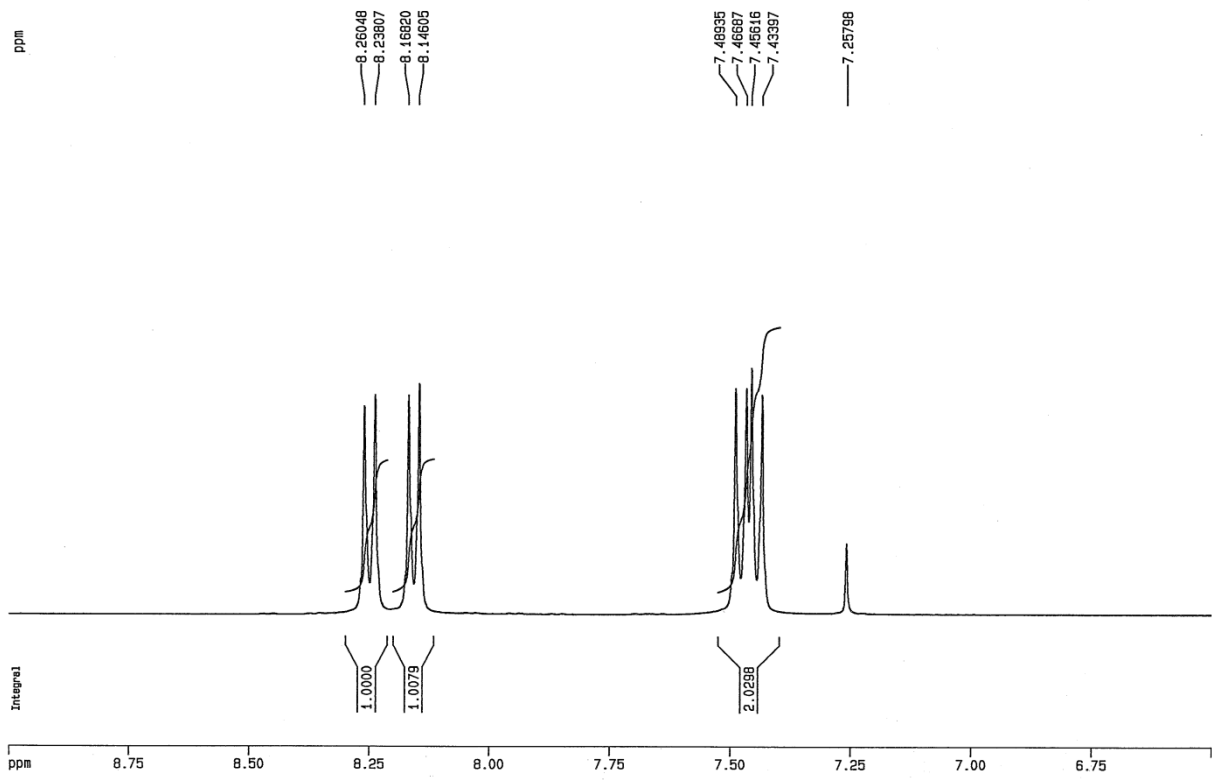
2a: R = 4-OMe



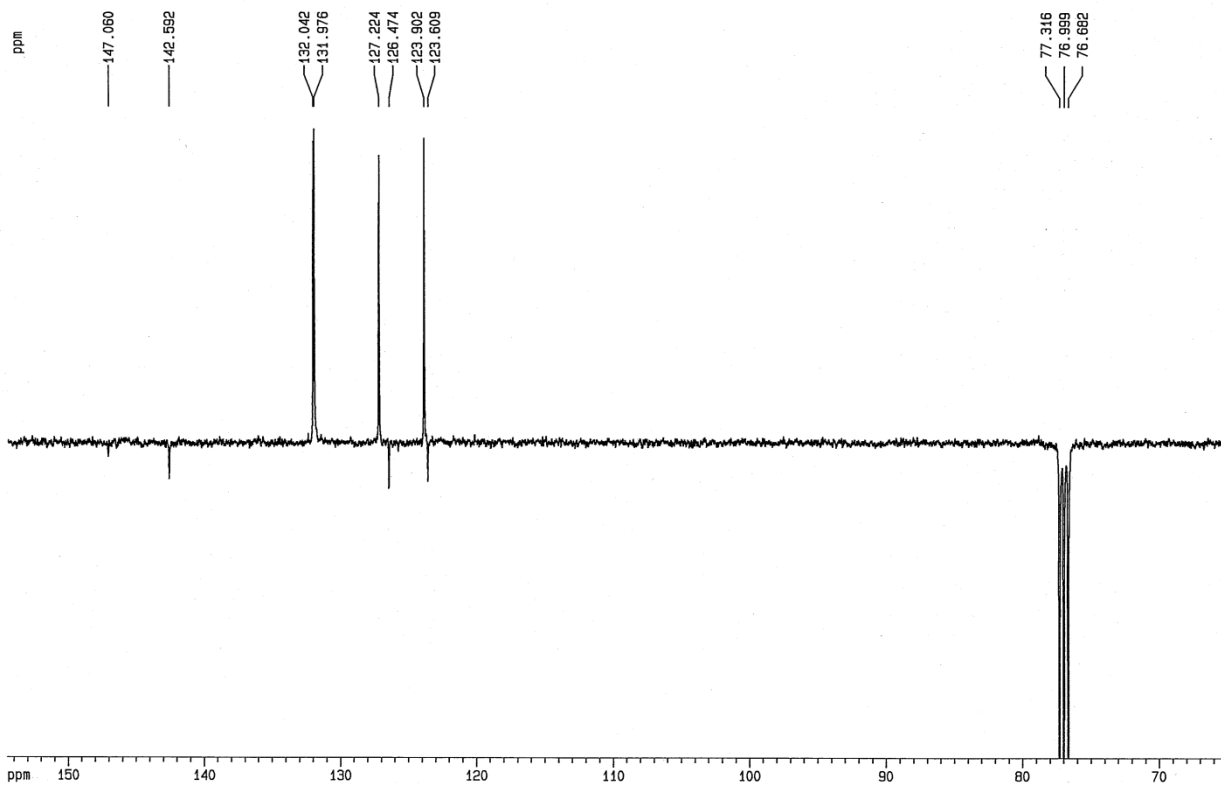
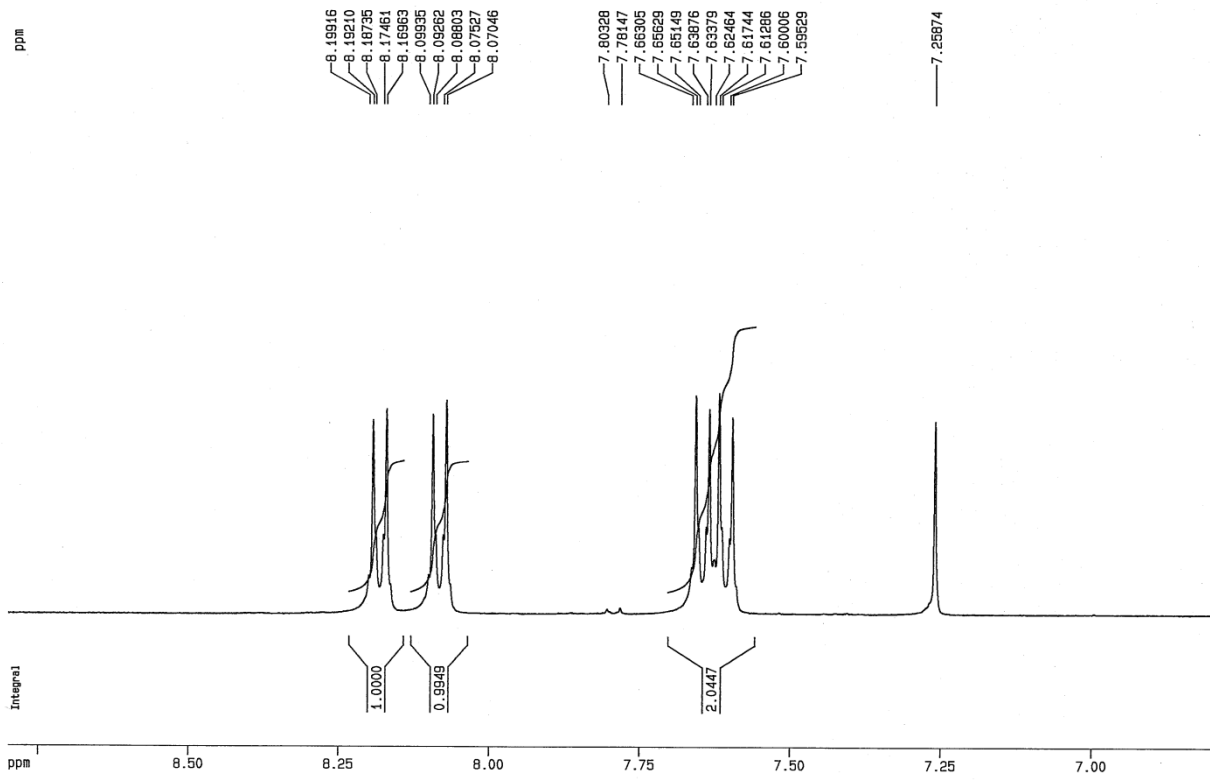
2b: R = H



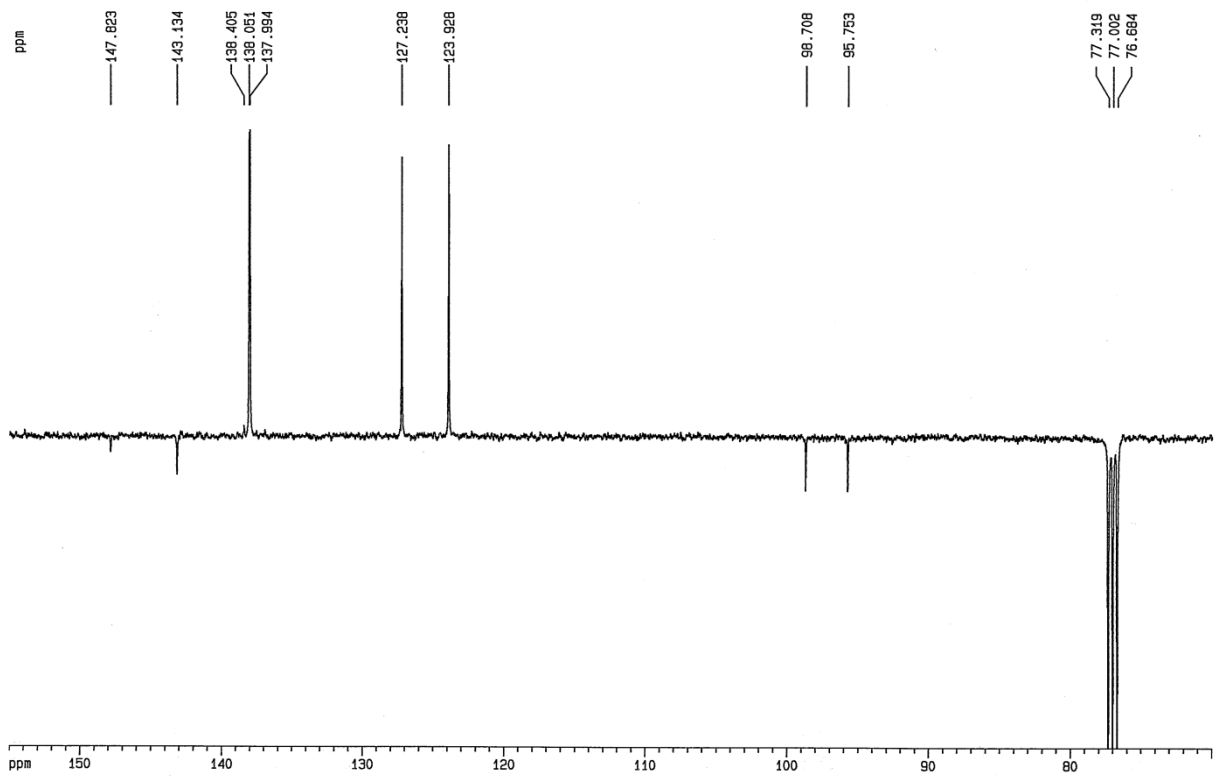
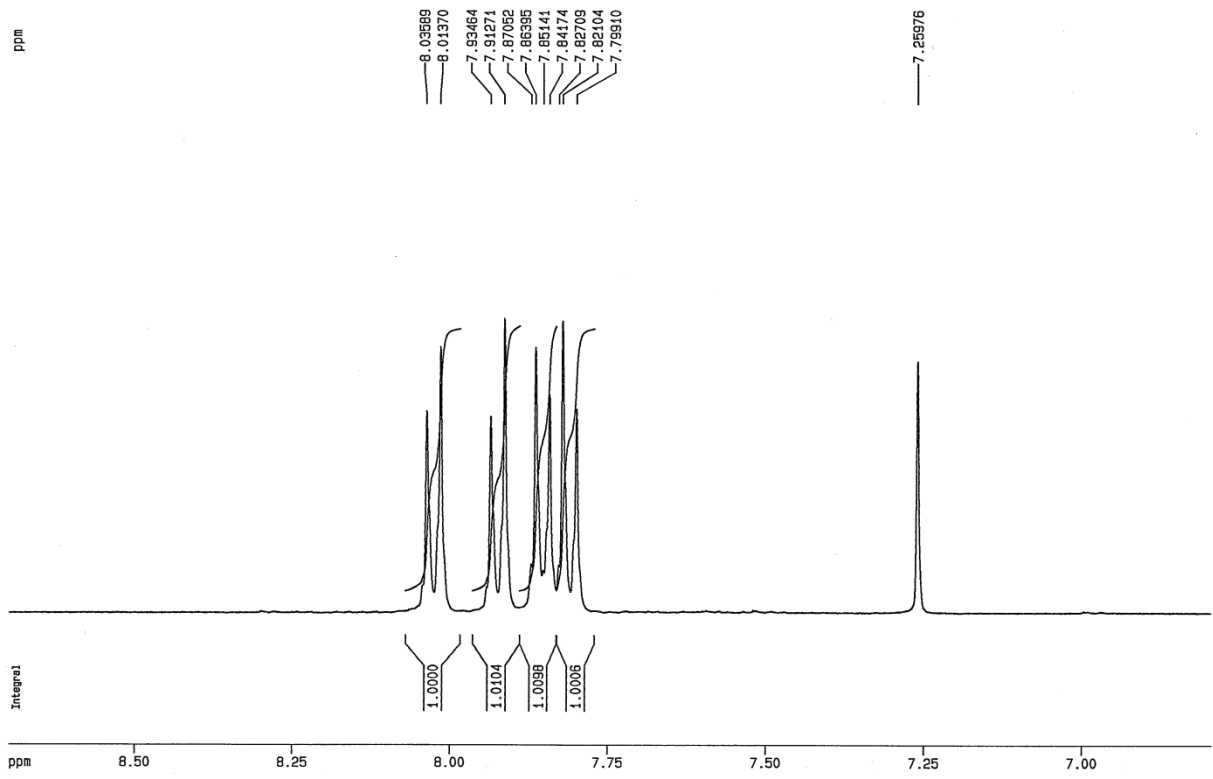
2c: R = 4-Cl



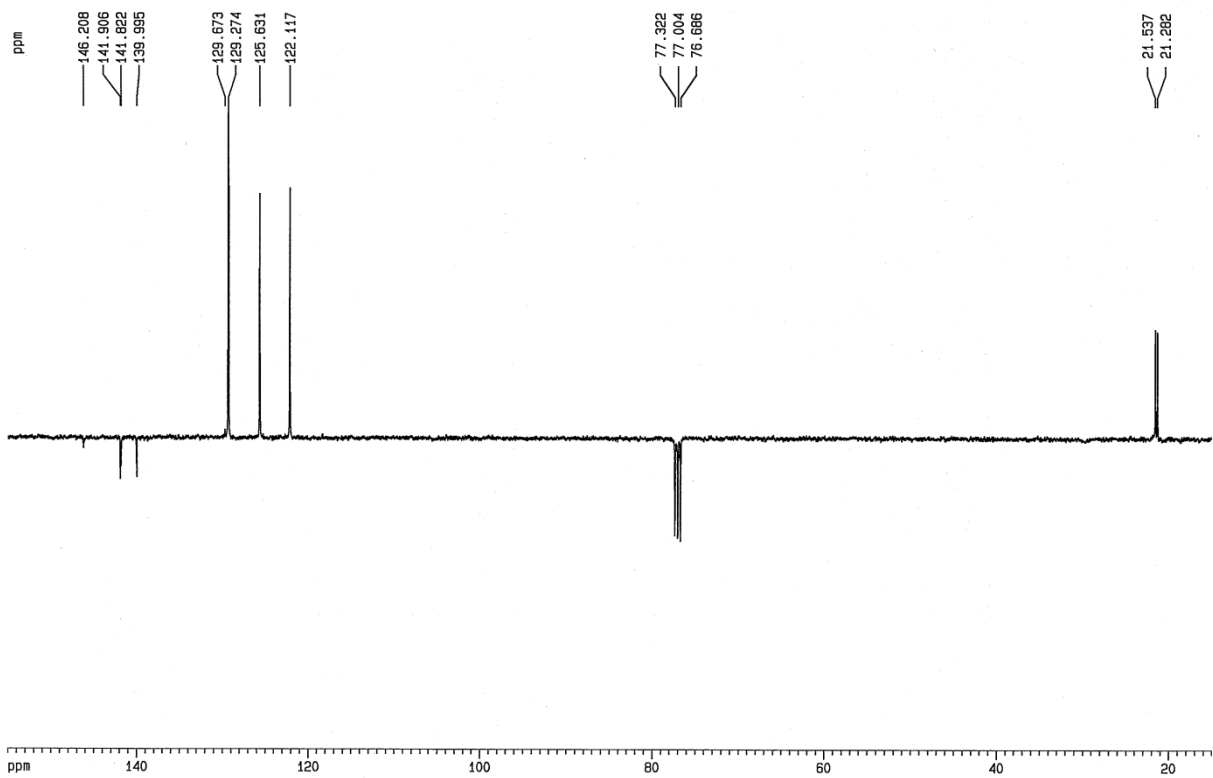
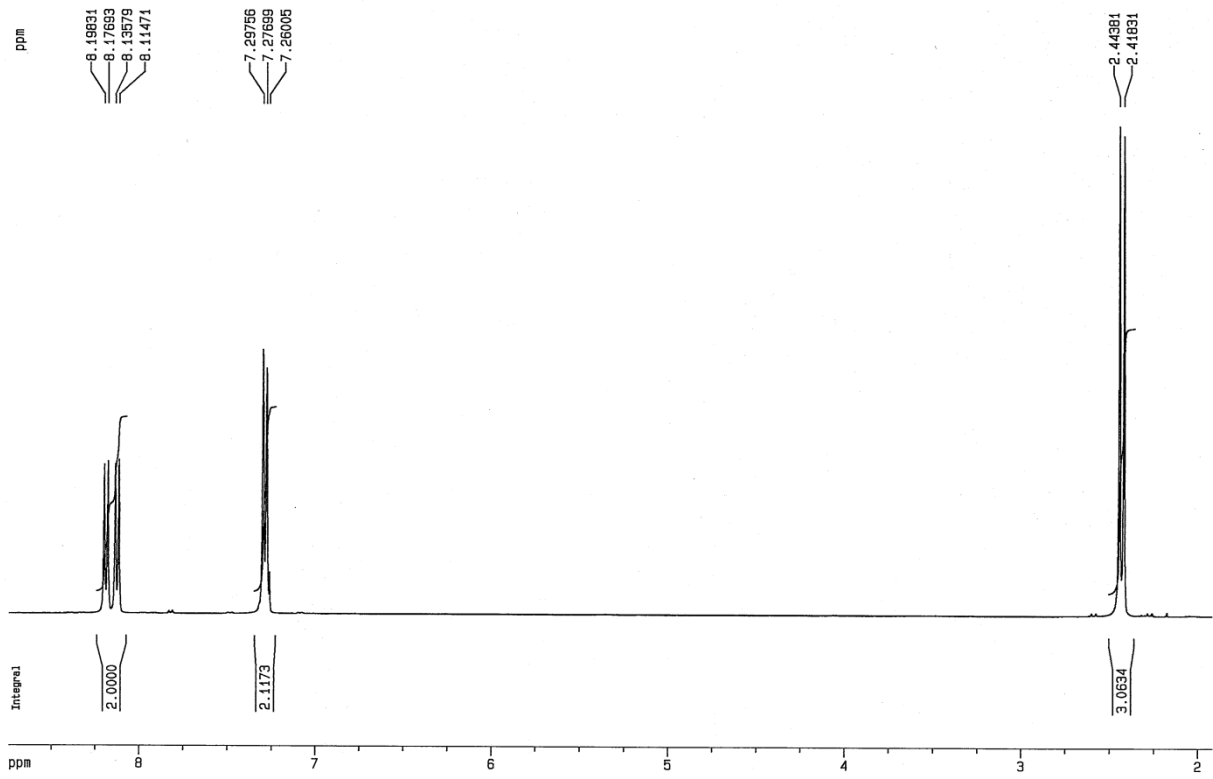
2d: R = 4-Br



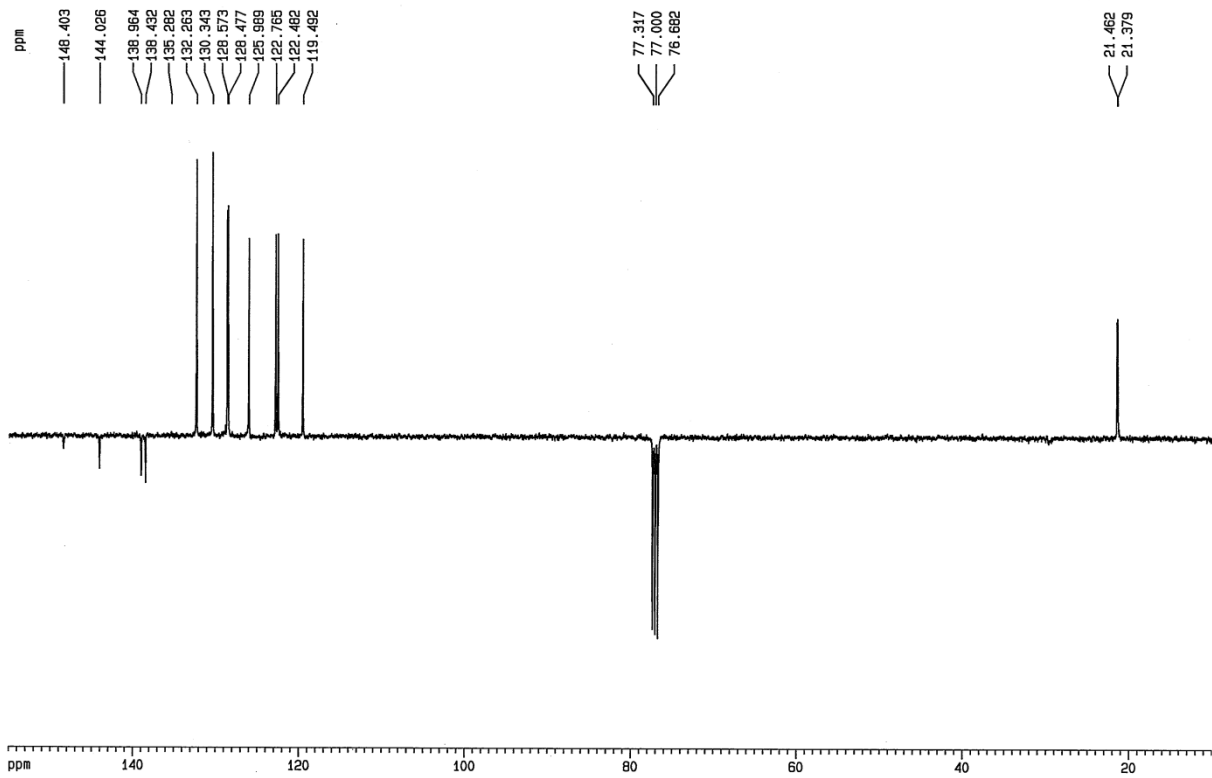
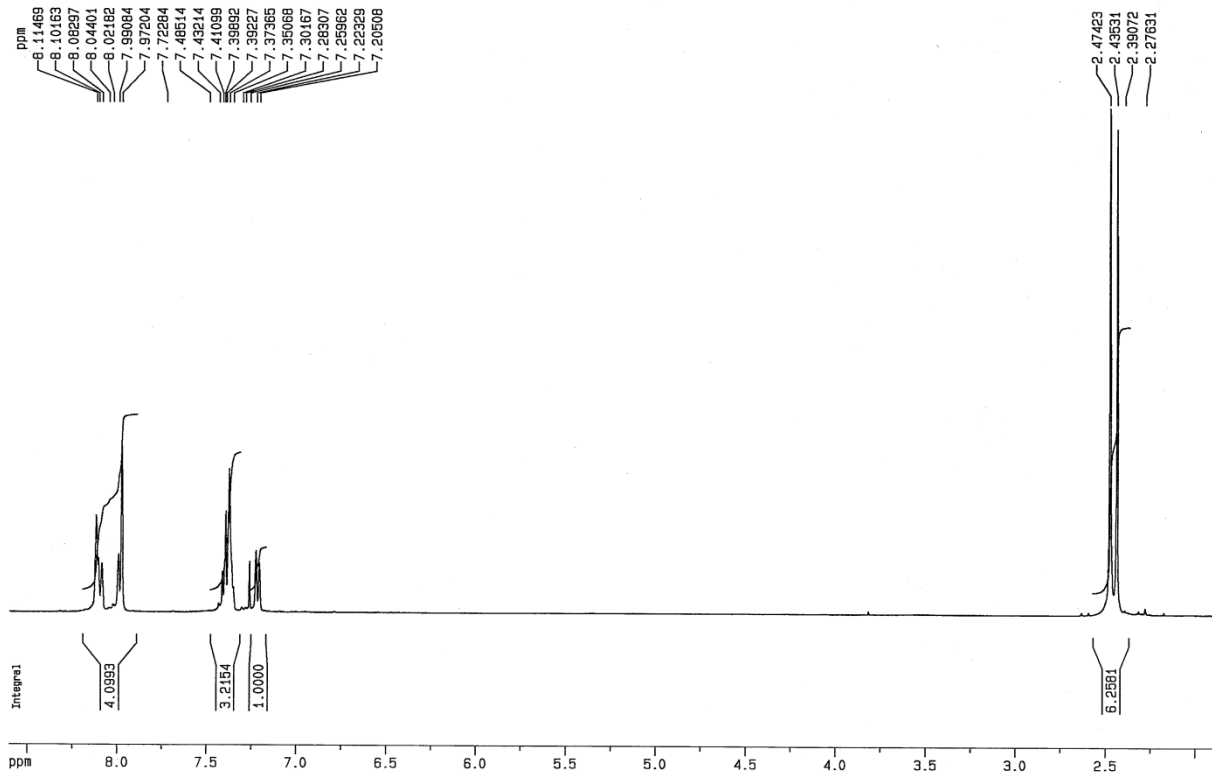
2e: R = 4-I



2f: R = 4-Me

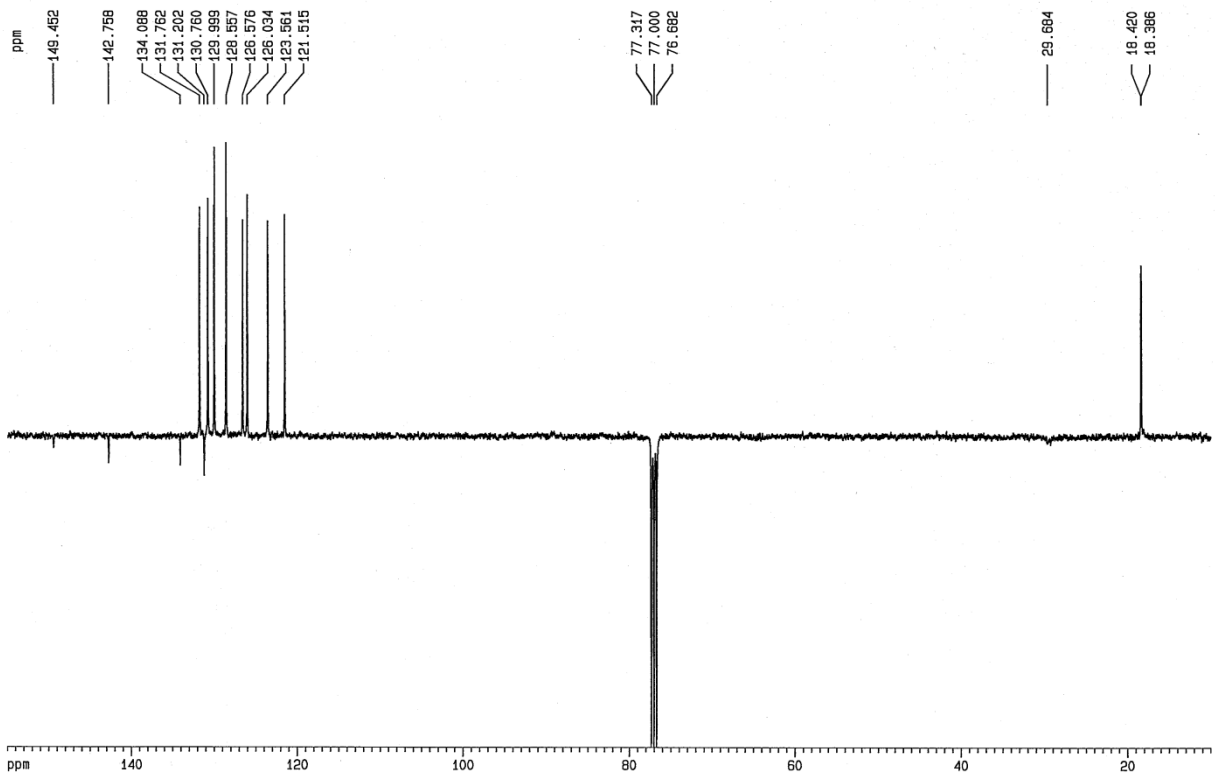
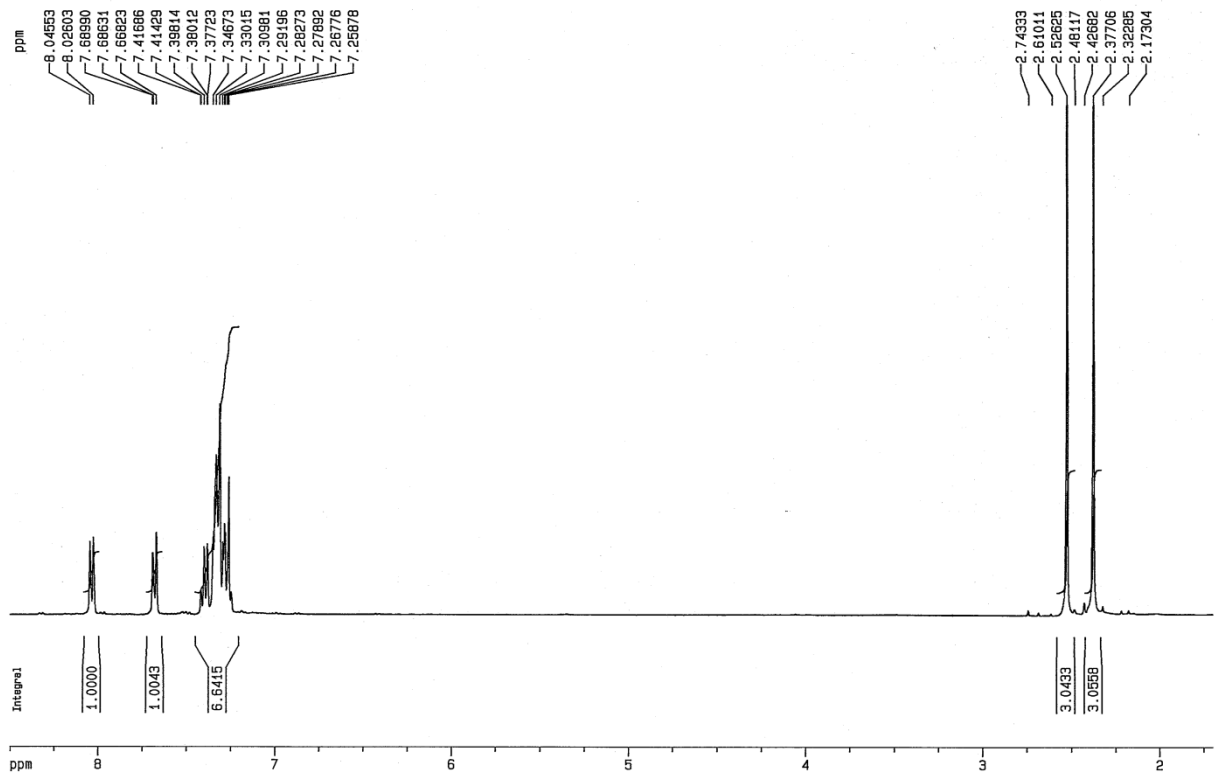


2g: R = 3-Me

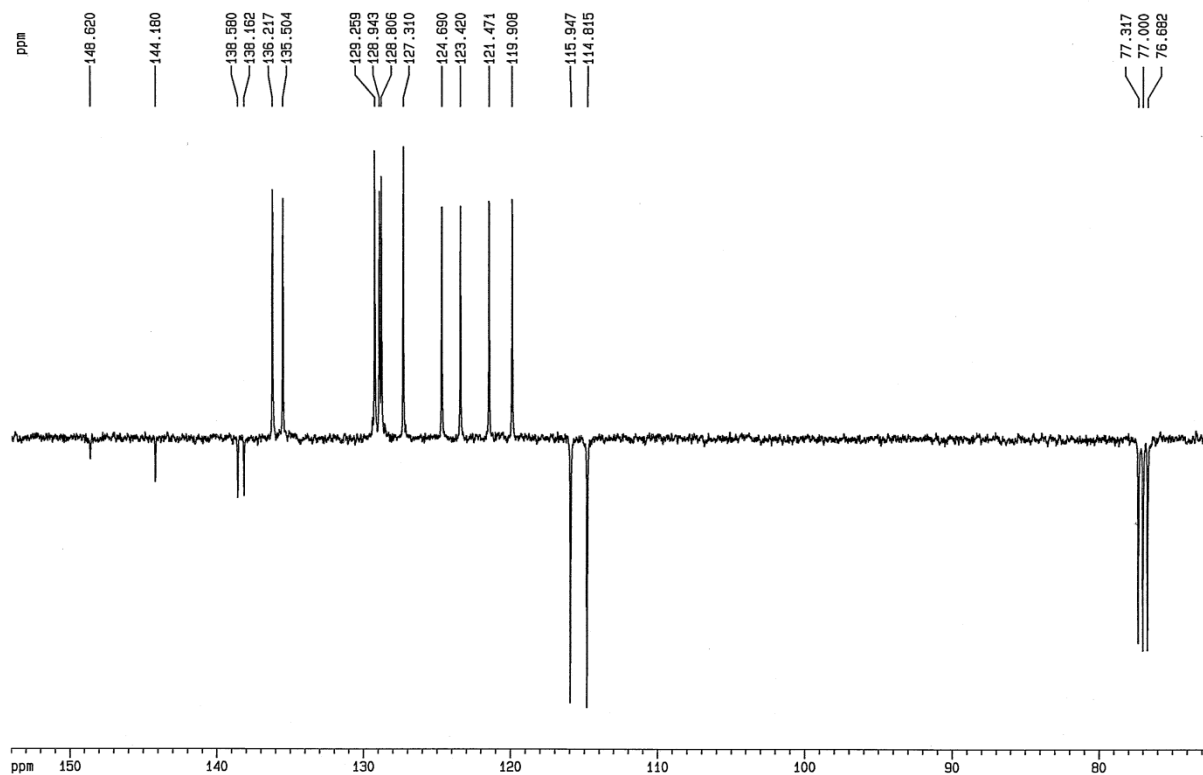
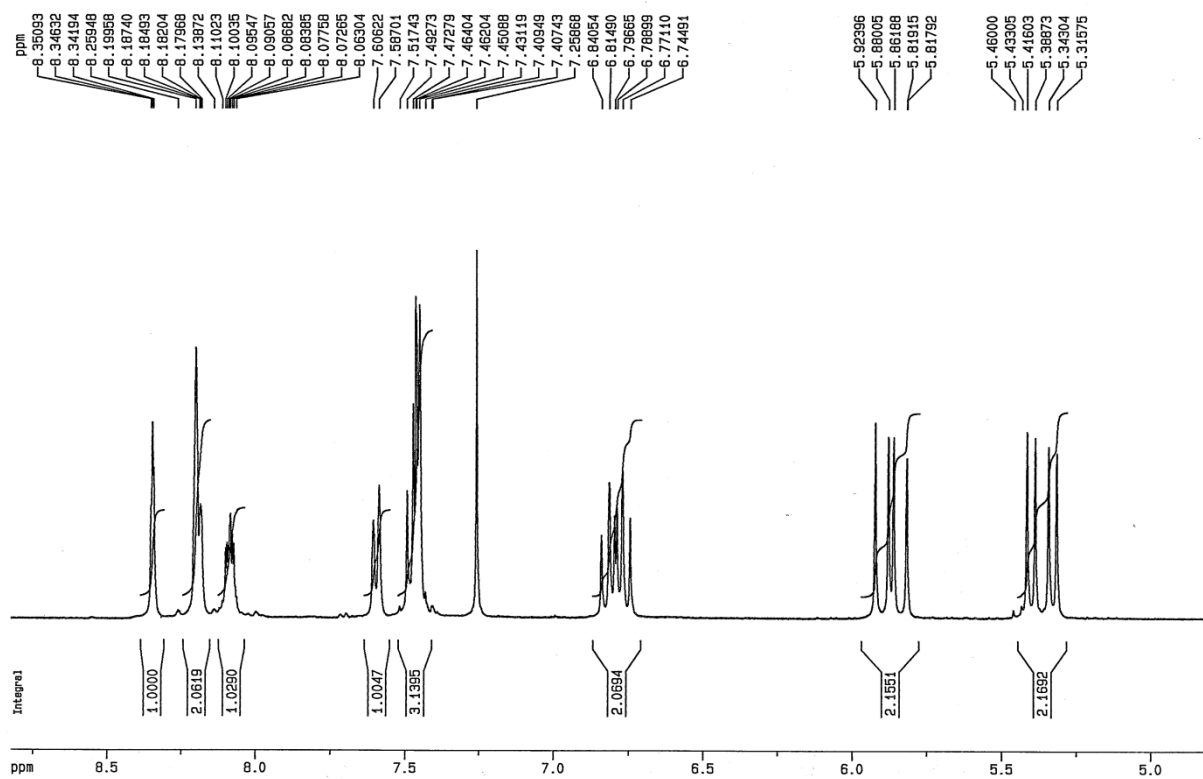




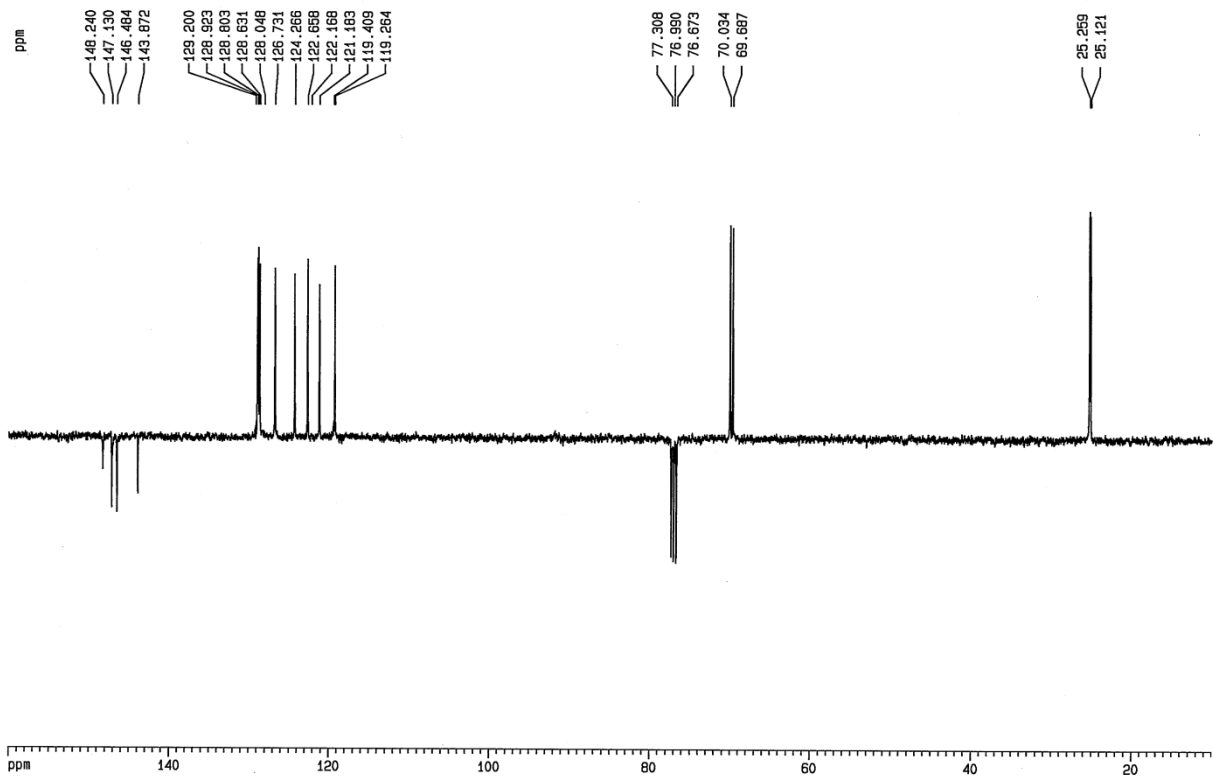
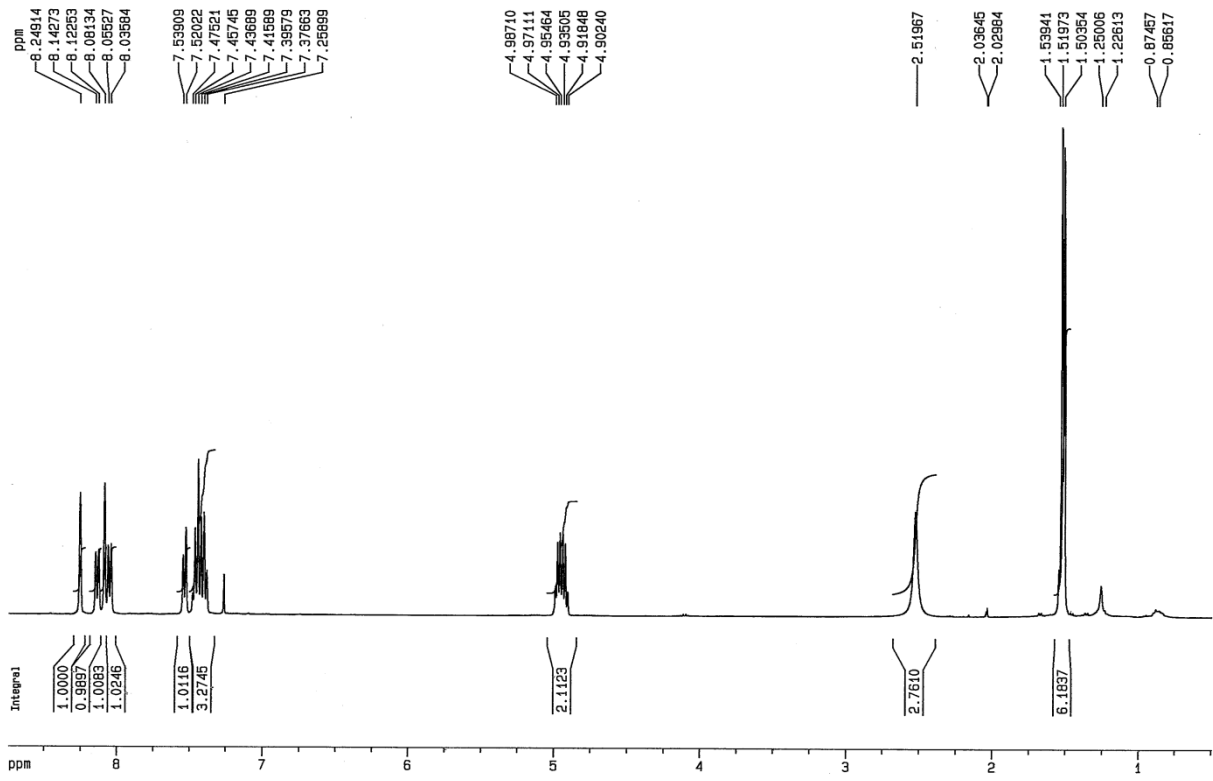
2h: R = 2-Me



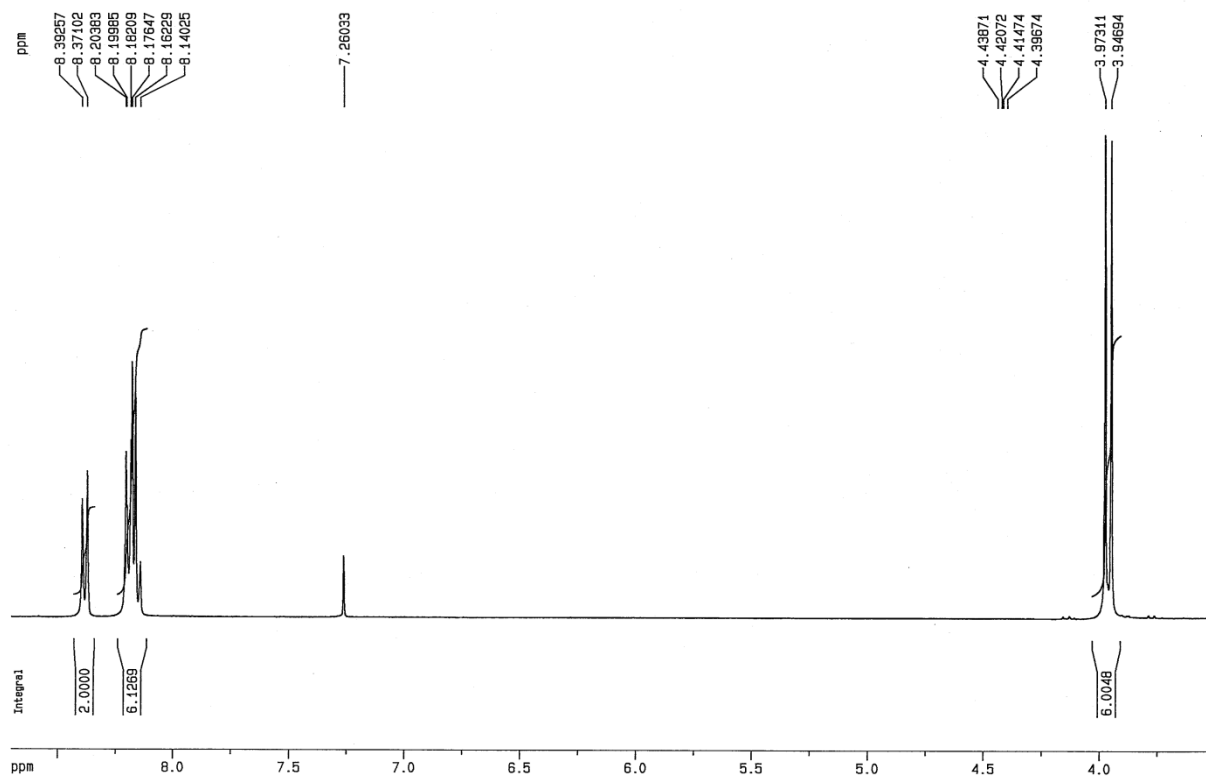
2i: R = 3-CH=CH<sub>2</sub>

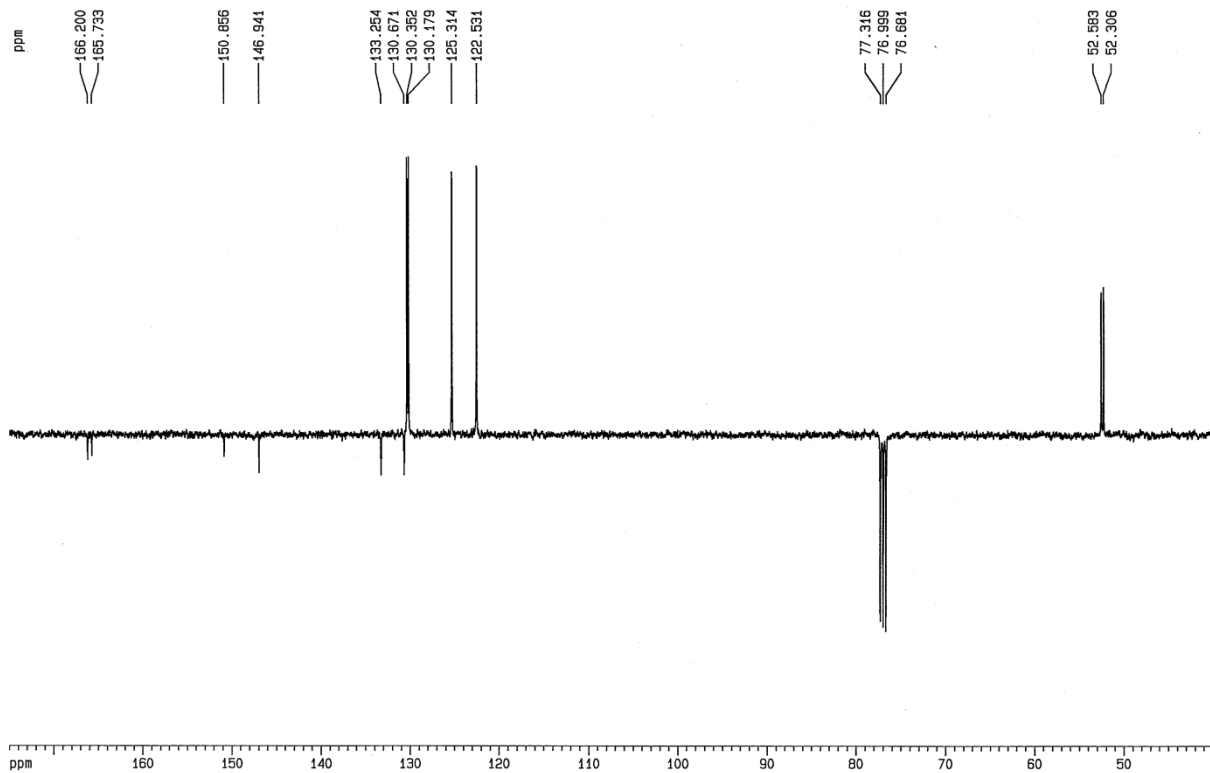


2j: R = 3-CH(OH)Me

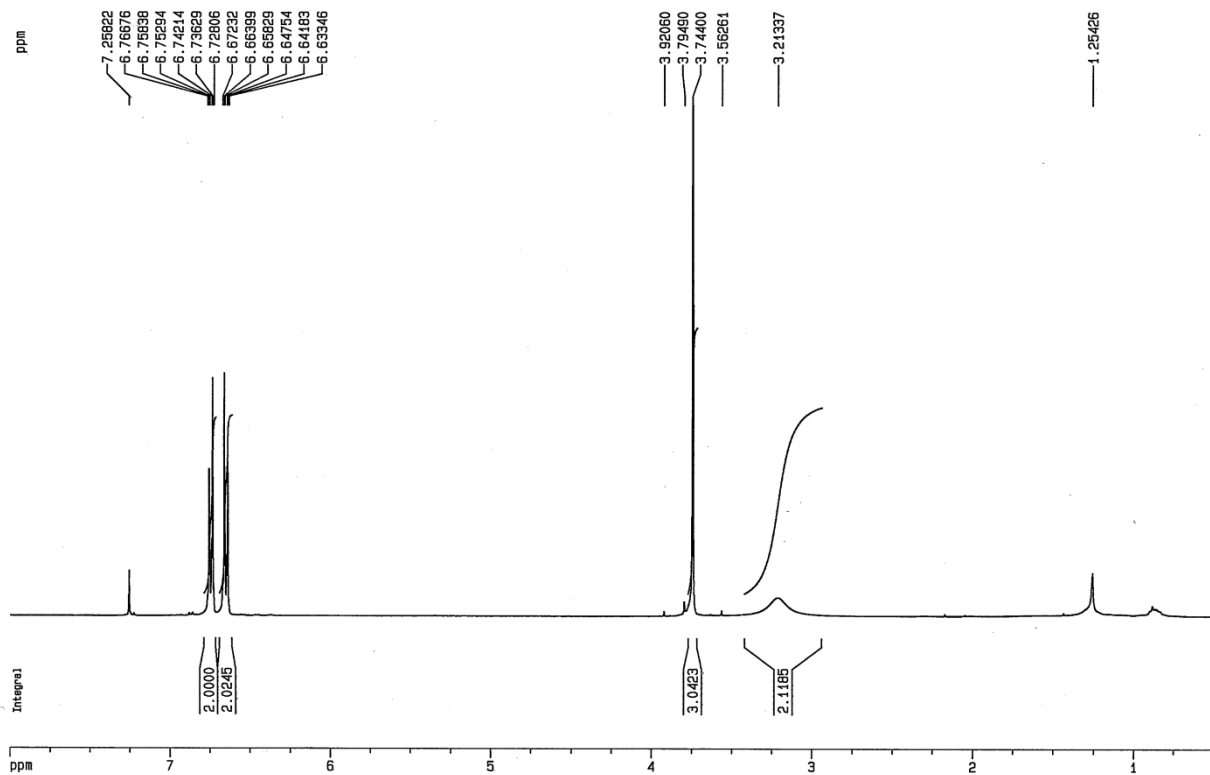


2k: R = 4-COOMe

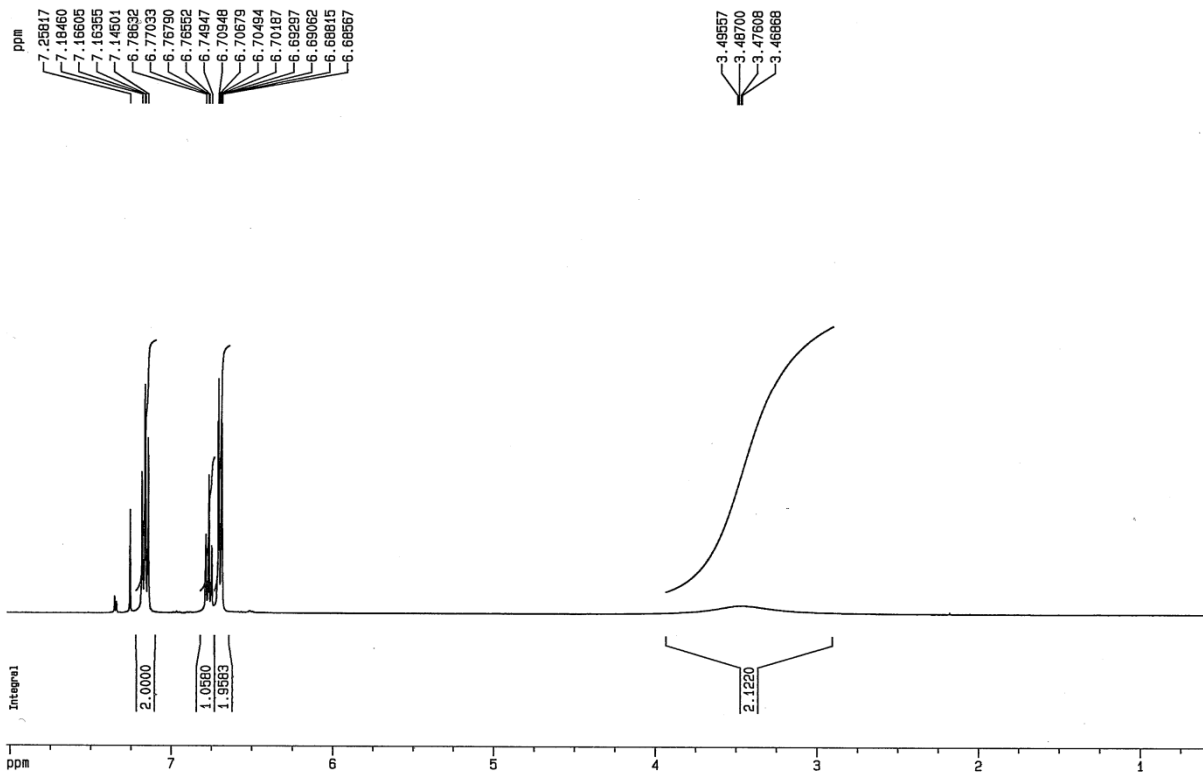




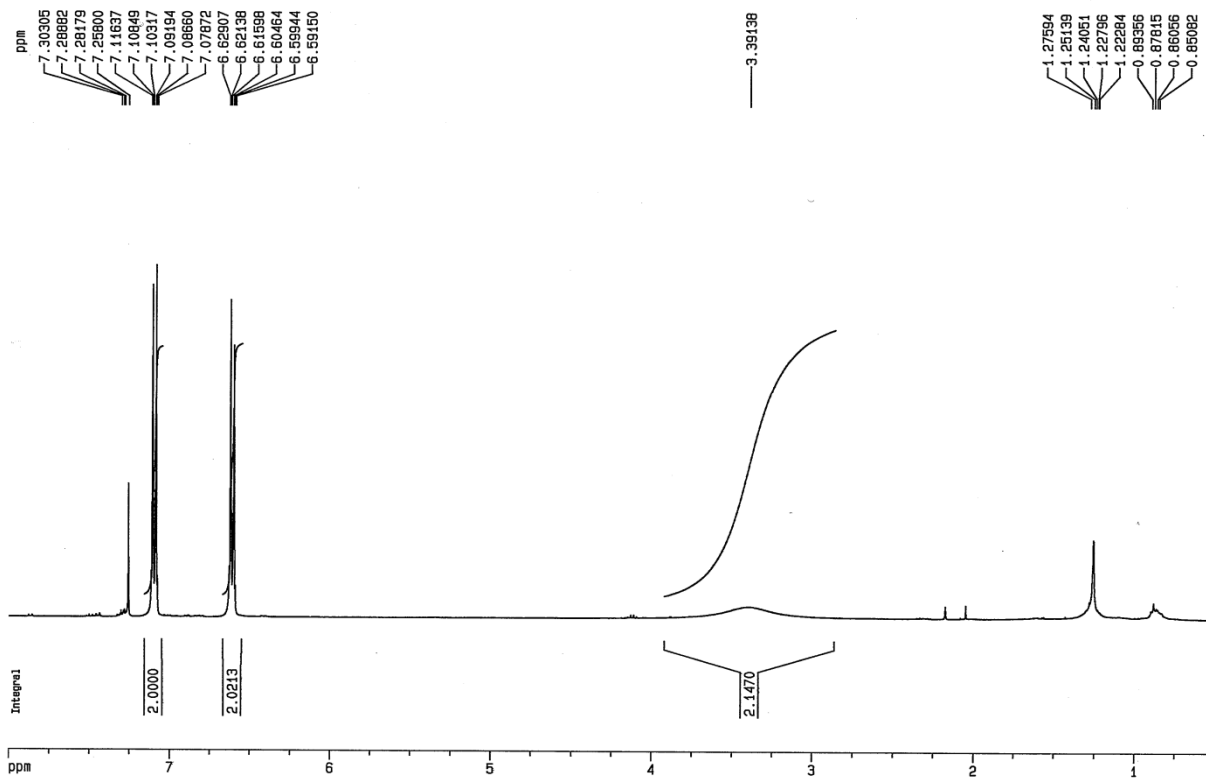
4a: R = 4-OMe



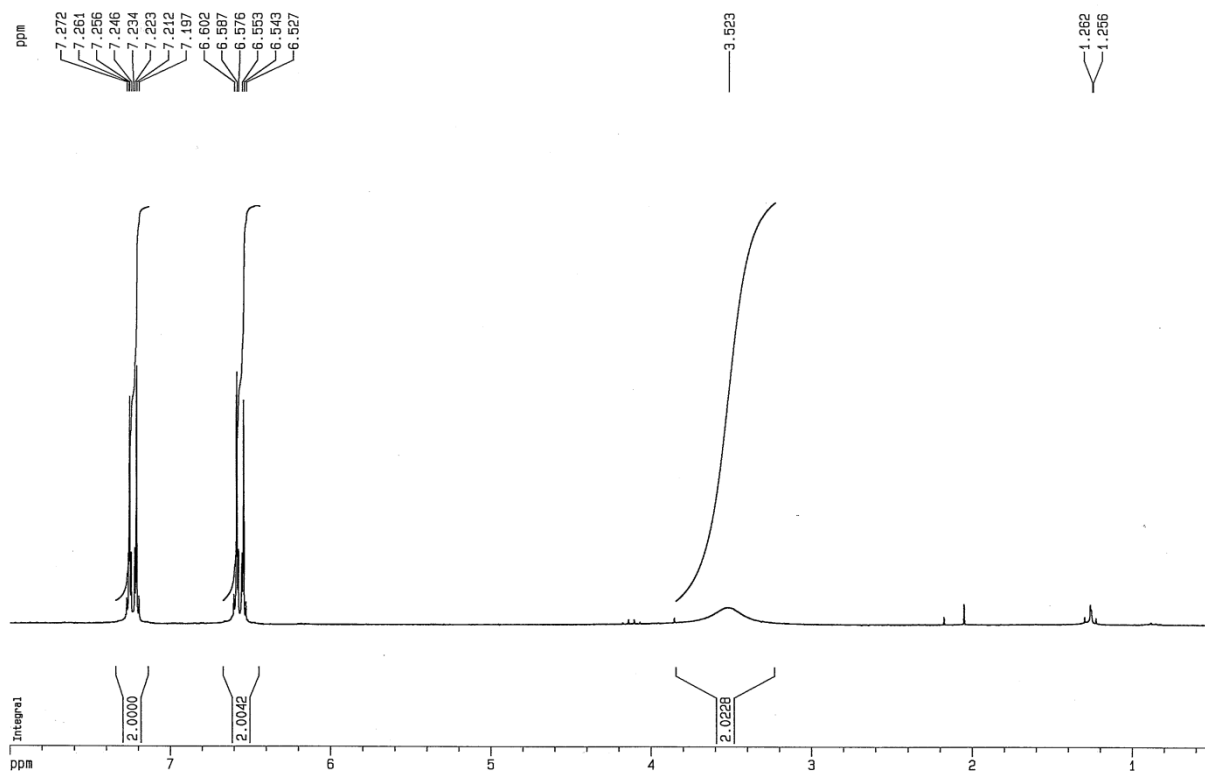
4b: R = H



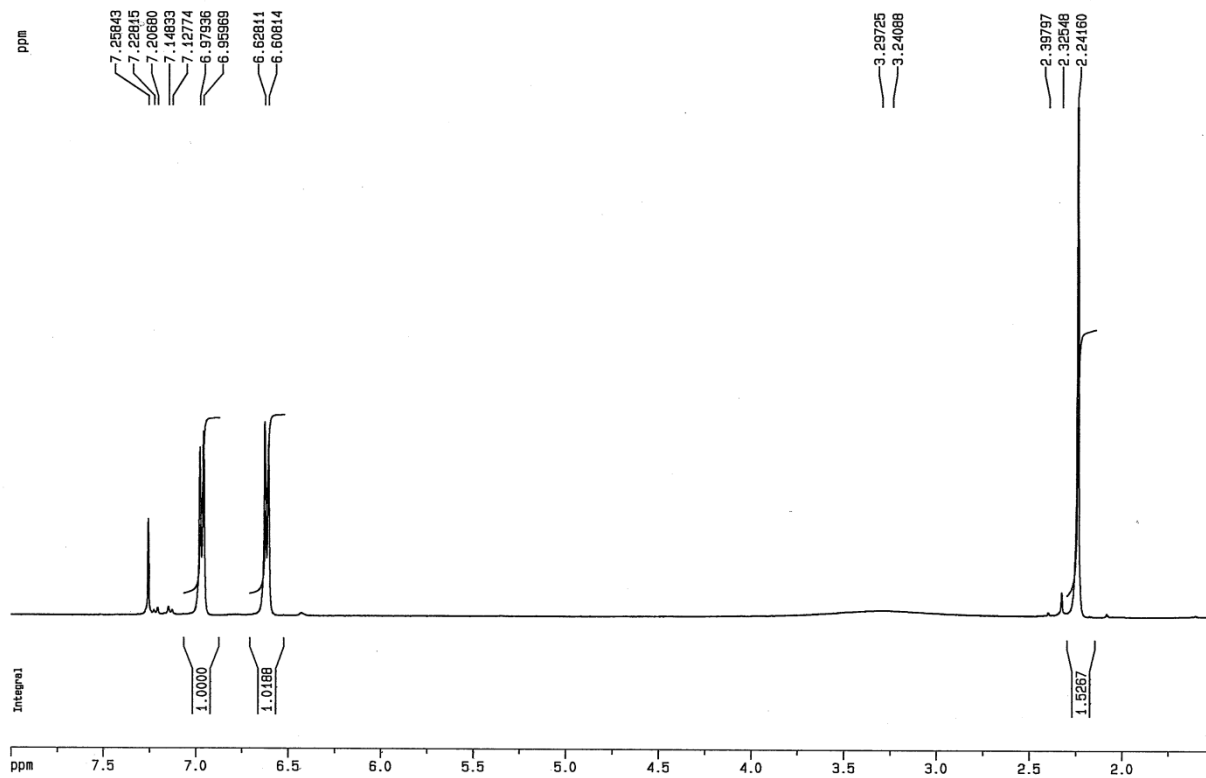
4c: R = 4-Cl



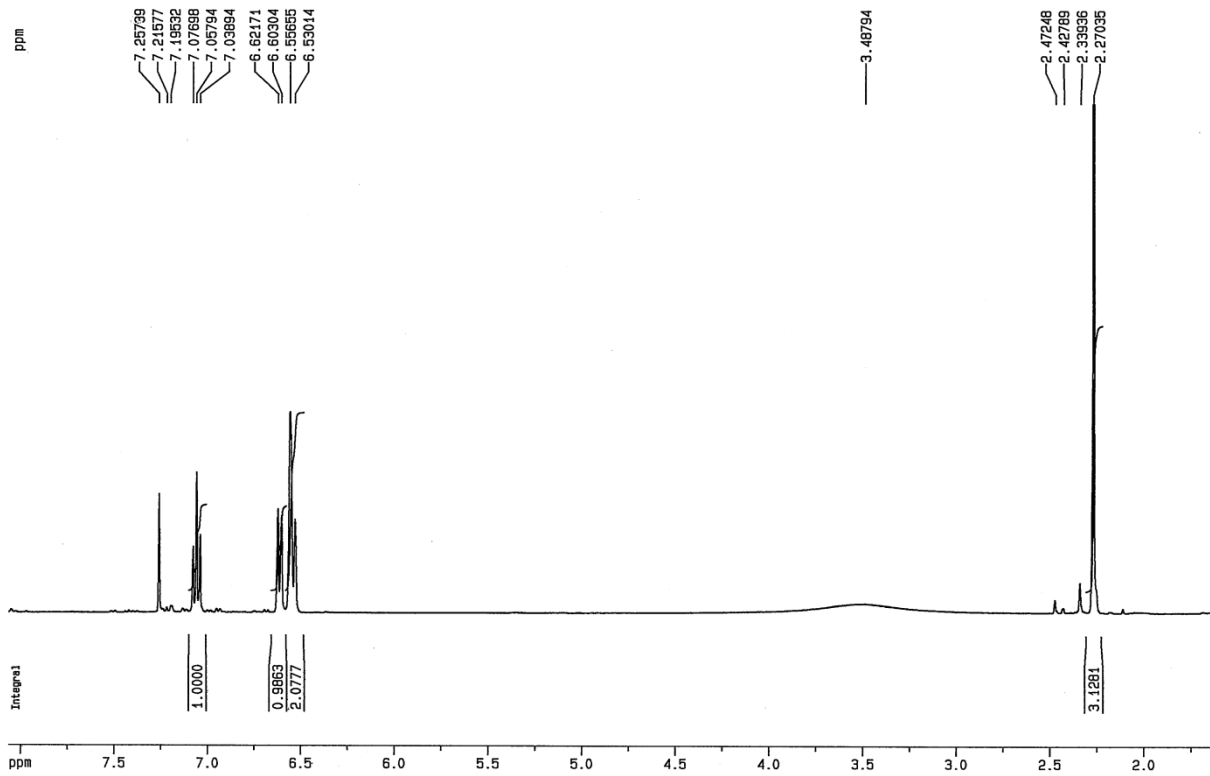
4d: R = 4-Br



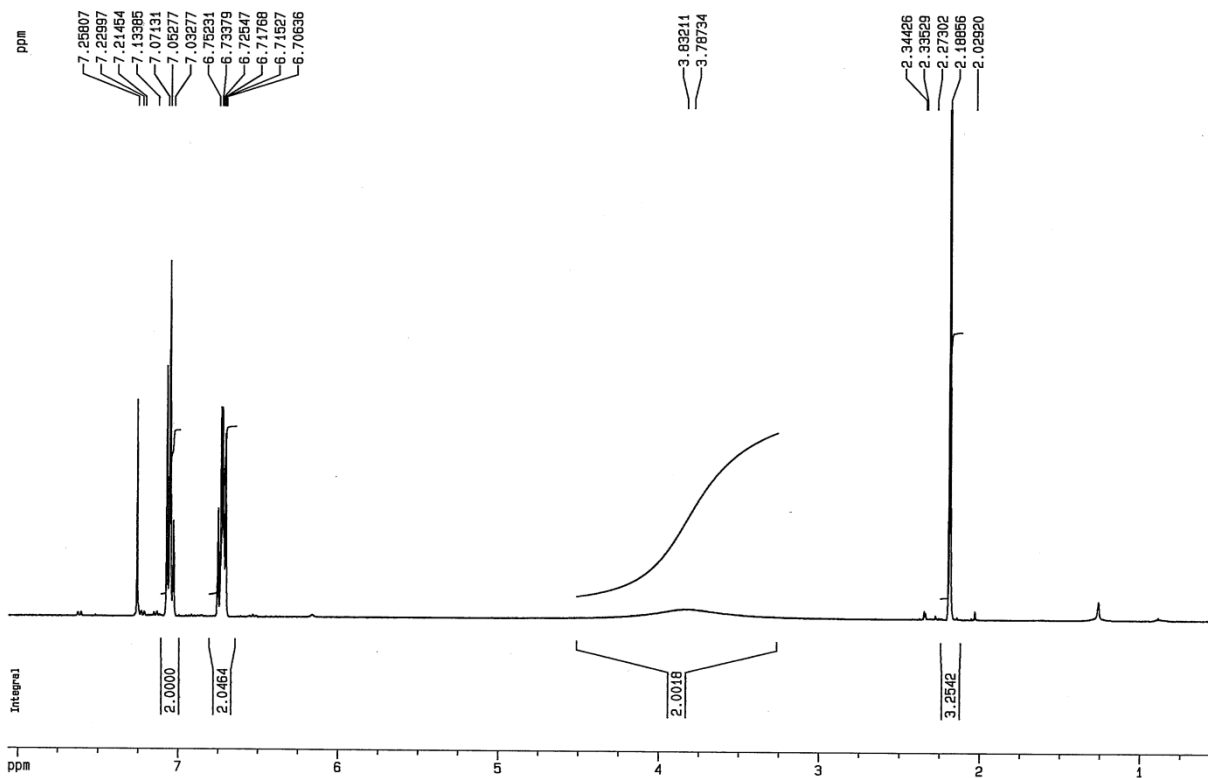
4f: R = 4-Me



4g: R = 3-Me

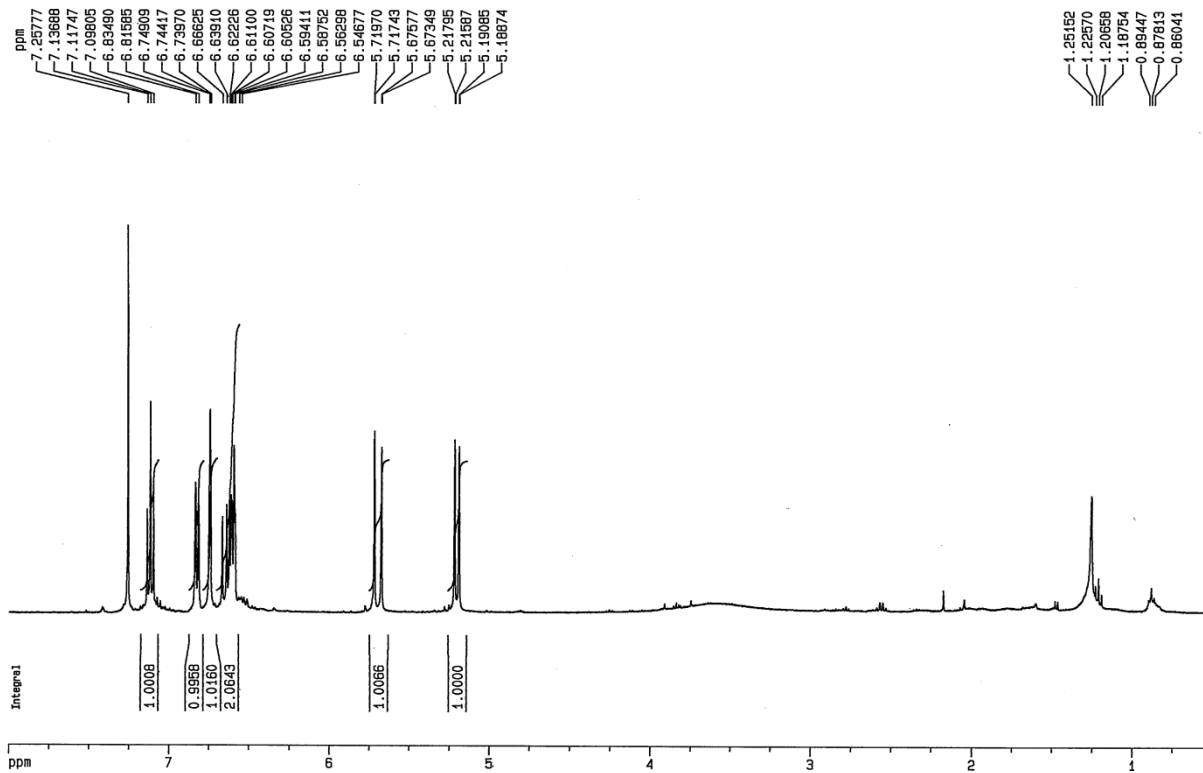


4h: R = 2-Me

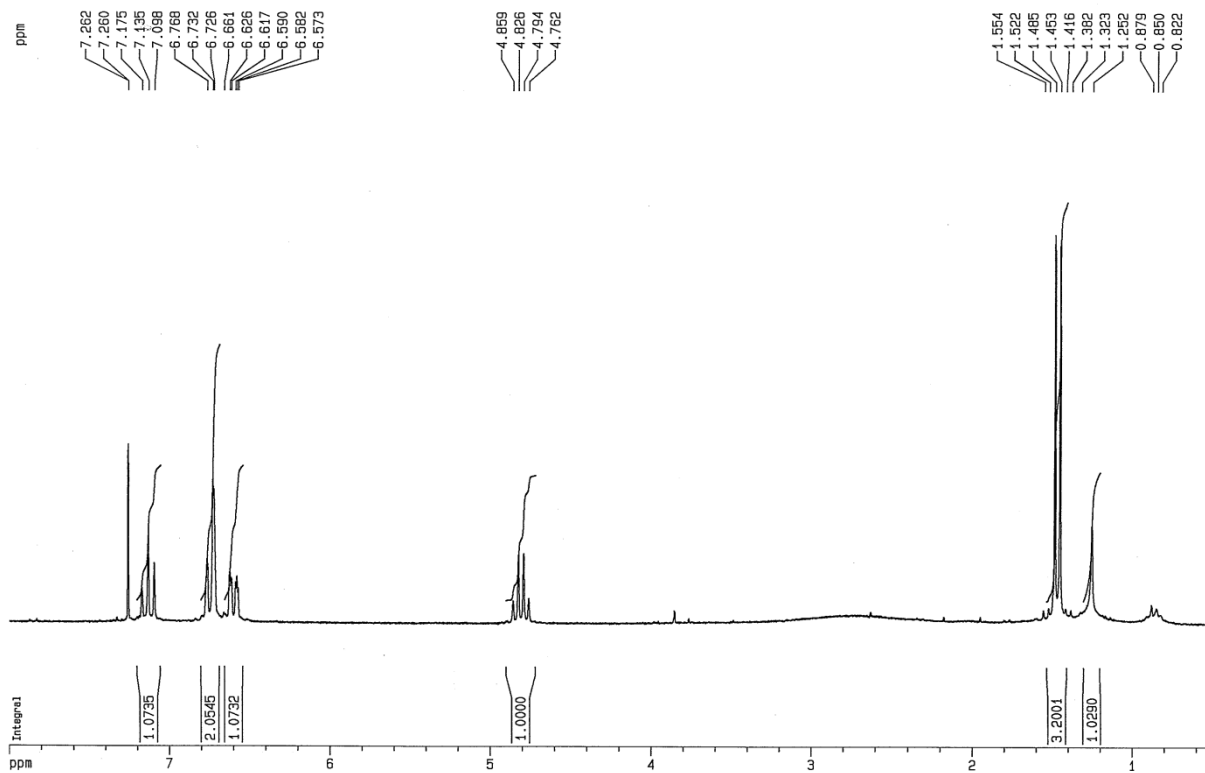


4i: R = 3-CH=CH<sub>2</sub>

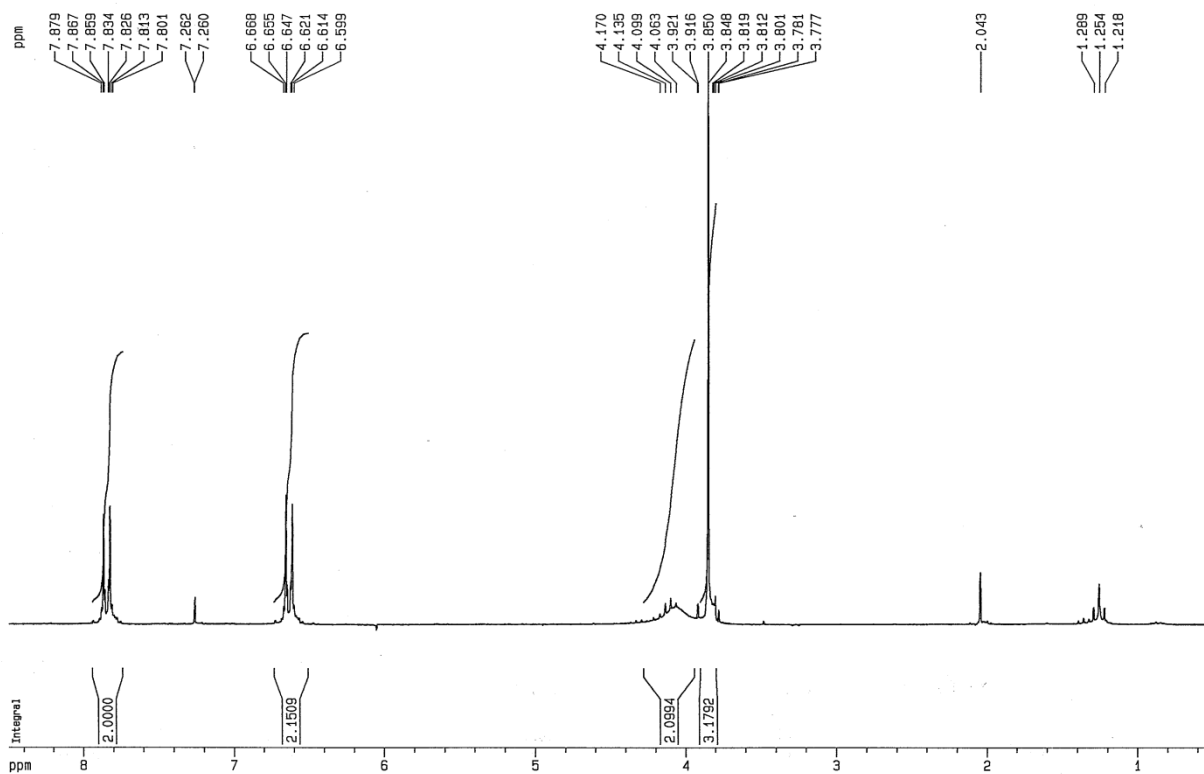




4j: R = 3-CH(OH)Me



4k: R = 4-COOMe



4l: 6-nitroquinoline

