

Supporting information

Solventless and Metal-free Regioselective Hydrofluorination of Functionalized Alkynes and Allenes: An Efficient Protocol for the Synthesis of *gem*-Difluorides

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Section 1. General experimental details

¹H NMR and ¹³C NMR spectra were recorded at 400 MHz and 100 MHz respectively, using CDCl₃ as a solvent. The chemical shifts are reported in δ (ppm) values relative to CHCl₃ (δ 7.26 ppm for ¹H NMR), multiplicities are indicated by s (singlet), d (doublet), t (triplet), q (quartet), p (pentet), h (heptet), m (multiplet) and br (broad). Coupling constants, *J*, are reported in Hertz.

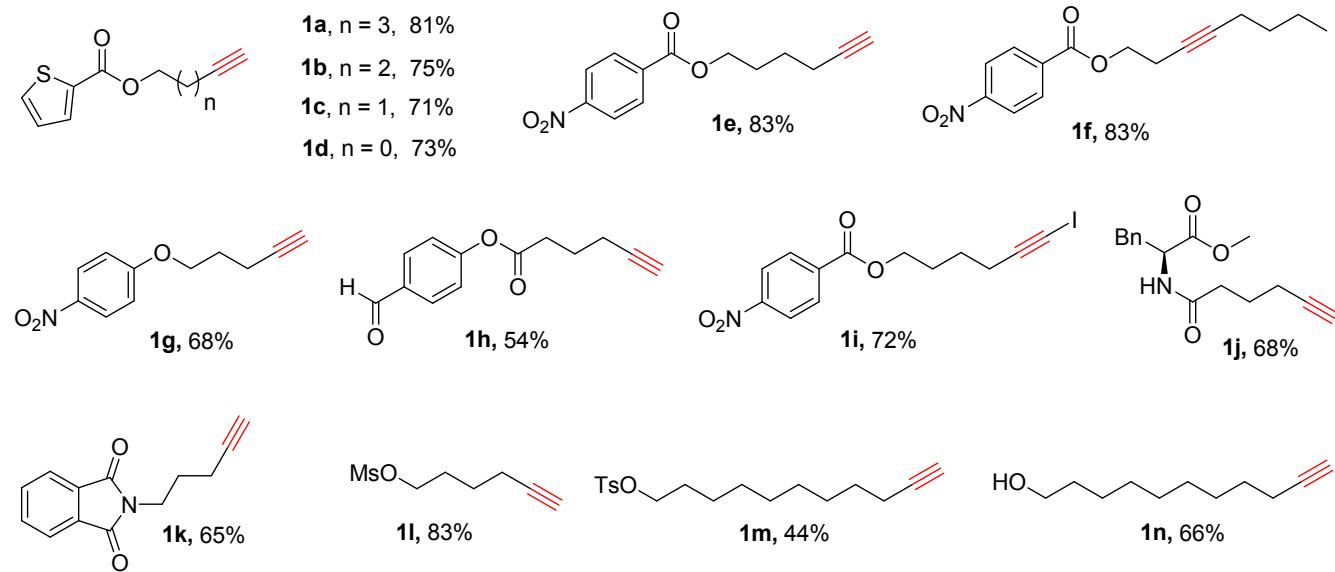
Solvents like DCM, Et₂O, Toluene, DMF were chemically dried using a commercial solvent purification system. Other solvents like DCE, dioxane, EtOAc and DMSO were dried with activated 4 Å molecular sieves overnight. Anhydrous hydrogen fluoride (HF) gas cylinder was purchased from Synquest Laboratories Inc.

All other reagents and solvents were employed without further purification. The products were purified using a CombiFlash system. TLC was developed on Merck silica gel 60 F254 aluminum sheets and KMnO₄ stain was used for TLC developing. KMnO₄ stain was prepared by mixing KMnO₄ (1.5 g), K₂CO₃ (10 g), and NaOH (10 wt%, 1.25 mL) in 200 mL water. All NMR solvents were purchased from Cambridge Isotope Laboratories, Inc.

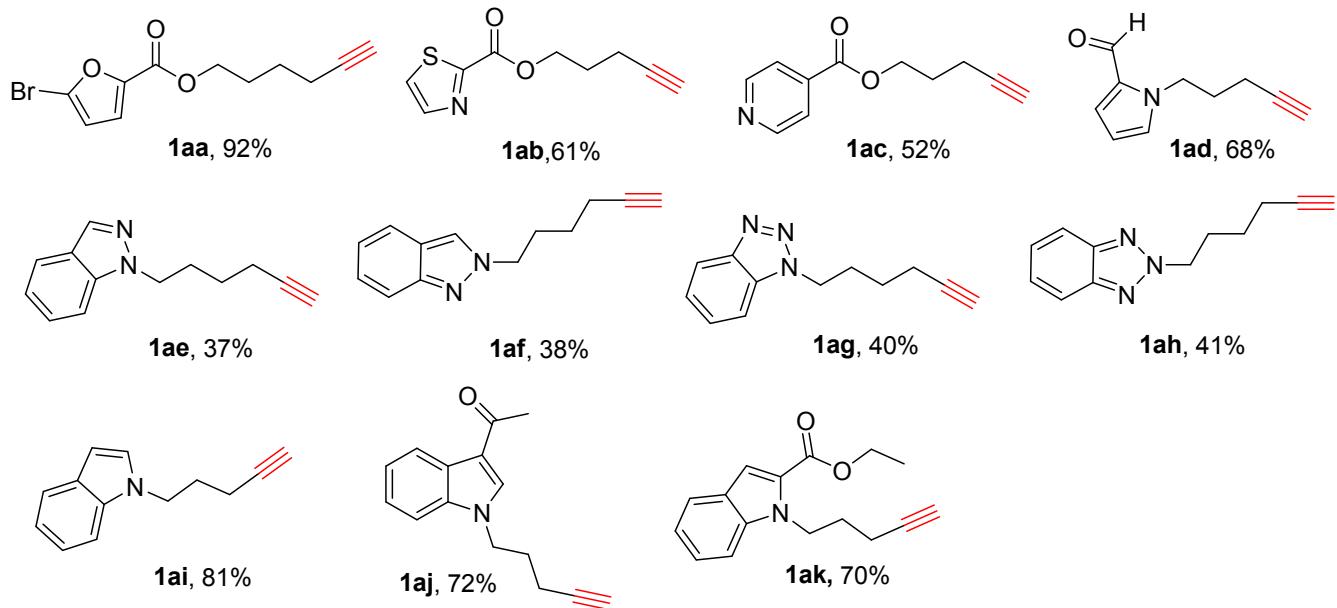
KHSO₄-13HF¹ and DMPU-12HF² were freshly prepared in our lab with the same method in the literature.

Section 2. Preparation of substrates

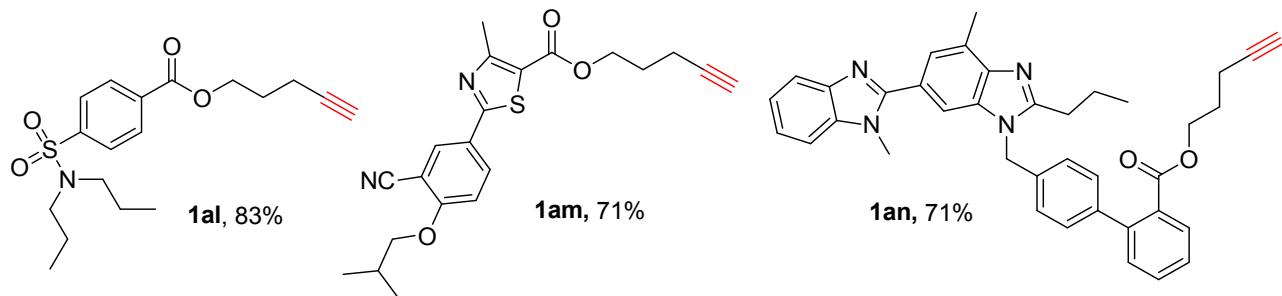
Alkyne substrates 1 (the identity of these substrates was confirmed by ¹H and ¹³C NMR spectra. Substrates **1j**, ³ **1k**, ⁴ **1l**, ⁵ **1m**, ⁶ **1ai**, ⁷ **1aj**⁸ were prepared using literature methods and the NMR spectra obtained were in accord with the literature. Substrate **1n** was purchased from Sigma-Aldrich).



Heterocyclic substrates:



Natural product and drug derivatives substrates:



Allene substrates ⁹:

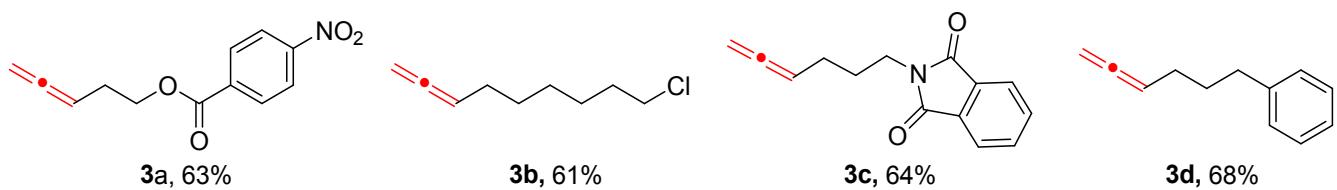


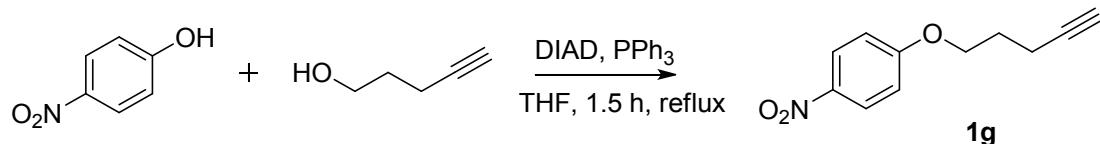
Figure S 1. Prepared alkyne and allene substrates.

General synthetic procedure for the synthesis of esters **1a**, **1b**, **1c**, **1d**, **1e**, **1f**.



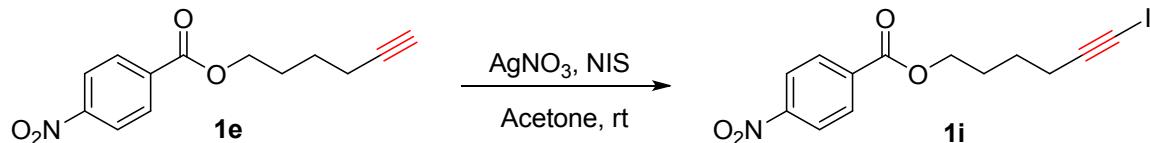
A 20-mL vial fitted with a stirring bar was charged with alcohol (2 mmol), Et₃N (2 equiv) and dry DCM (10 mL). The mixture was cooled down to 0 °C. Acyl chloride (1.2 equiv) and 5 mg DMAP were then added sequentially. The mixture was stirred overnight and then it was diluted with 50 mL DCM, washed with 1M aqueous HCl (2 x 20 mL), saturated NaHCO₃ (20 mL) and brine (20 mL), sequentially. The organic layer was then dried with Na₂SO₄ and concentrated. The residue was purified with column chromatography to afford the desired esters **1a** (yield: 93%), **1b** (yield: 75%), **1c** (yield: 71%), **1d** (yield: 73%), **1e** (yield: 83%), **1f** (yield: 83%).

General synthetic procedure for the synthesis of ether **1g.**



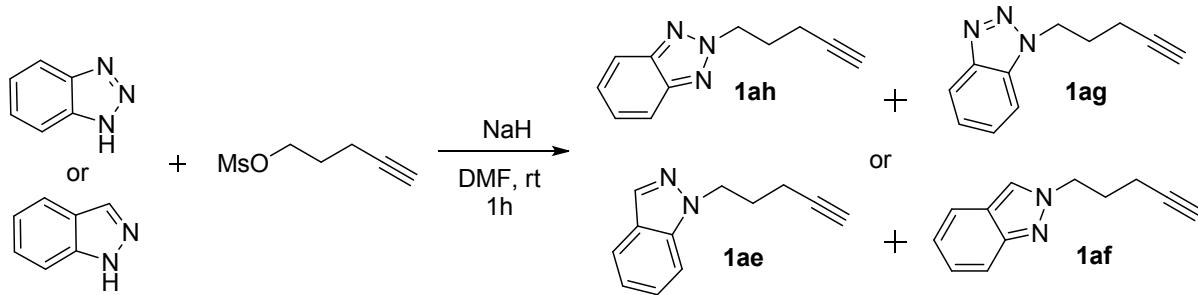
A solution of phenol (3 mmol), 3-methyl-3-buten-1-ol (3.6 mmol), triphenylphosphine (3.6 mmol), diisopropyl azodicarboxylate (3.6 mmol) in THF (27 mL) was heated for 1.5 hours at reflux. After concentrated in vacuo, the residue was purified by flash column chromatography to give the desired ethers **1g** (yield: 68%).

Synthetic procedure for the synthesis of ester **1i.**



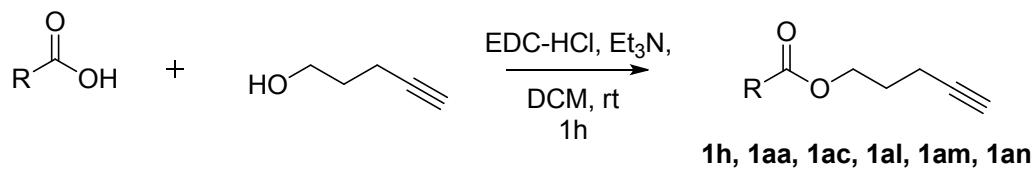
Alkyne **1e** (1.5 mmol) and silver nitrate (0.3 mmol) were stirred in acetone (10 mL) for 5 minutes and then N-iodosuccinimide (3.6 mmol) was added. The reaction was stirred at rt for 5 hours until complete consumption of the starting material according to TLC. It was then poured onto iced water. The aqueous layer was extracted with hexane (3 times), and the combined organic extracts were dried over MgSO₄, filtered, and the solvent removed by evaporation under reduced pressure. The iodo-alkyne was isolated by flash column chromatography (yield: 72%).

General synthetic procedure for the synthesis of **1ae, **1af**, **1ag**, **1ah**.**



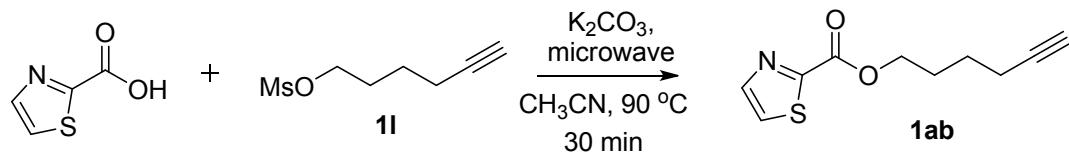
A 50-mL flask fitted with a stirring bar was charged with a solution of starting material (2 mmol) in DMF (10 mL). The mixture was cooled down to 0 °C and NaH (1.5 equiv) was then added. The mesylate was also added and the mixture was stirred at rt for 1h. The reaction was quenched by NH₄Cl (1 M) and diluted with CH₂Cl₂ (50 mL). After being washed with 5% LiCl aqueous solution (2 x 20 mL), the aqueous layer was extracted with CH₂Cl₂ (20 mL). The combined organic extracts were washed with water (20 mL), dried and concentrated in vacuo. The resulting residue was purified by silica gel flash chromatography to afford **1ah** (yield: 41%), **1ag** (yield: 40%), **1ae** (yield: 37%), **1af** (yield: 38%).

General synthetic procedure for the preparation of esters **1h**, **1aa**, **1ac**, **1al**, **1am**, **1an**.



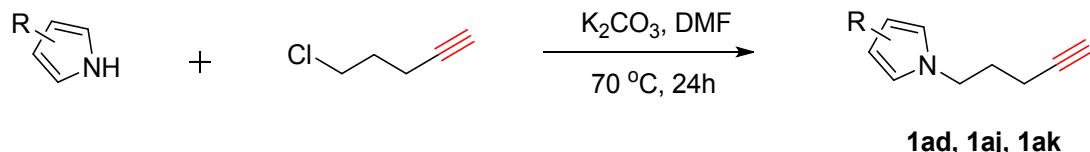
A 50-mL flask fitted with a stirring bar was charged with a solution of alcohol (2 mmol), EDCI (1.2 equiv), triethylamine (1.5 equiv), and DMAP (0.1 equiv) in dichloromethane (10 mL). Nicotinic acid (1 equiv) was then added at 0 °C, and the reaction mixture was stirred overnight at room temperature. After the reaction was complete, the resulting mixture was diluted with DCM (50 mL), washed by 1 N HCl (2 x 20 mL), 1 N aqueous NaHCO₃ (2 x 20 mL), and brine (1 x 20 mL). The organic layer was dried (Na₂SO₄) and evaporated in vacuo. The resulting residue was purified by column chromatography to afford the desired ester **1h** (yield: 54%), **1aa** (yield: 92%), **1ac** (yield: 52%), **1al** (yield: 83%), **1am** (yield: 71%), **1an** (yield: 71%).

General synthetic procedure for the preparation of ester **1ab**.



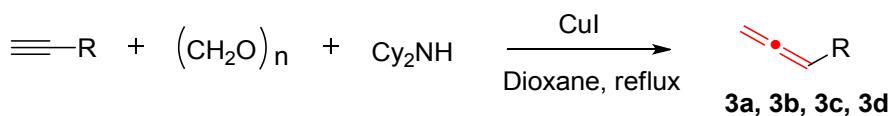
A 10-mL microwave tube fitted with a stirring bar was charged with a solution of acid starting material (1.5 mmol), K_2CO_3 (1.5 equiv), mesylate (1 equiv) in CH_3CN (2 mL). The mixture was stirred under microwave conditions at 90 °C for 30 mins. The reaction was concentrated and purified directly by silica gel flash chromatography to afford **1ab** (yield: 61%).

General synthetic procedure for the preparation of **1ad**, **1aj**, **1ak**.



To a 10-mL flask were added 5-chloro-1-pentyne (2.4 mmol), indole or pyrrole derivatives (2.0 mmol), anhydrous potassium carbonate (2.0 mmol), and DMF (2 mL). The mixture was stirred for 24 h at 70 °C in an oil bath, cooled, diluted with water (40 mL), and extracted with CH_2Cl_2 (3*10 mL). The combined organic phase was washed with water (4*20 mL), dried over sodium sulfate, and concentrated in vacuo. The crude was purified with flash chromatography to afford **1ad** (yield: 68%), **1aj** (yield: 72%), **1ak** (yield: 70%).

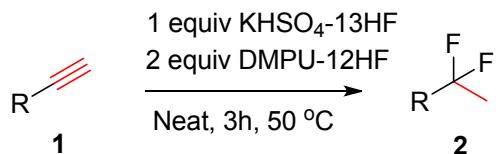
General synthetic procedure for the preparation of allene **3a**, **3b**, **3c**, **3d**.⁹



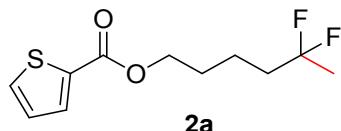
A 50-mL flask fitted with a stirring bar was charged with $(CH_2O)_n$ (10 mmol), CuI (2 mmol), dioxane (20 mL), alkyne (4 mmol), and amine (7.2 mmol) sequentially. The flask was equipped with a reflux condenser under an argon atmosphere and the resulting mixture was stirred under reflux. When the reaction was complete as determined by TLC, it was cooled to rt. Water (25 mL) and ether (50 mL) were added and then the aqueous solution was separated and extracted with ether (3 × 25 mL). The organic layer was then washed with brine and dried over anhydrous Na_2SO_4 . Evaporation and flash column chromatography on silica gel afforded the terminal allene **3a** (yield: 63%), **3b** (yield: 61%), **3c** (yield: 64%), **3d** (yield: 68%).

Section 3. General procedure for hydrofluorination of alkynes

3.1 Hydrofluorination of alkynes.



An 8-mL polytetrafluoroethylene (PTFE) vial fitted with a stirring bar was charged with alkyne starting material (0.2 mmol). DPMU-12HF (134 μ L, 24 equiv based on HF) and KHSO₄-13HF (52 μ L, 13 equiv based on HF) were then added in one portion at room temperature. The reaction was heated to 50 °C and stirred for 3 hours. The progress of the reaction was monitored by TLC (visualized by KMnO₄ stain). The product usually shows a slightly more polar spot than the starting material on TLC (R_f difference < 0.1 in most cases). The reaction was cooled down to 0 °C with an ice bath and calcium chloride powder was added to quench the reaction until no bubbling was observed. The crude was mixed with silica gel and concentrated. The resulting residue was purified with flash chromatography.



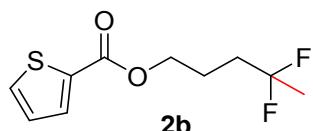
Colorless oil, isolated yield: 81%.

¹H NMR (400 MHz, CDCl₃) δ = 7.80 (dd, J = 3.7, 1.1 Hz, 1H), 7.55 (dd, J = 5.0, 1.1 Hz, 1H), 7.10 (dd, J = 5.0, 3.7 Hz, 1H), 4.31 (t, J = 6.4 Hz, 2H), 1.98 – 1.85 (m, 2H), 1.85 – 1.76 (m, 2H), 1.67 – 1.61 (m, 2H), 1.64 – 1.55 (t, J = 20 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ = 162.20, 133.80, 133.36, 132.32, 127.74, 124.07 (t, J = 237.7 Hz), 64.63, 37.48 (t, J = 25.6 Hz), 28.35, 23.28 (t, J = 28.0 Hz), 19.35 (t, J = 4.6 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ = -90.75 – -90.90 (m, 2F).

HRMS: (APCI⁺) [M+H] cal. for C₁₁H₁₅F₂O₂S: 249.0761; found: 249.0756.



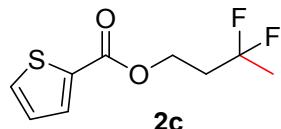
Colorless oil, isolated yield: 75%.

¹H NMR (500 MHz, CDCl₃) δ = 7.85 – 7.79 (dd, *J* = 3.6, 1.2 Hz, 1H), 7.59 (dd, *J* = 4.8, 1.2 Hz, 1H), 7.14 (dd, *J* = 4.8, 3.6 Hz, 1H), 4.37 (t, *J* = 5.8 Hz, 2H), 2.10 – 1.93 (m, 4H), 1.66 (t, *J* = 18.4 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ = 162.13, 133.64, 133.48, 132.45, 127.79, 123.84 (t, *J* = 238.0 Hz), 64.27, 34.61 (t, *J* = 26.0 Hz), 23.46 (t, *J* = 27.9 Hz), 22.25 (t, *J* = 4.5 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ = -89.72 – -93.27 (m, 2F).

HRMS: (APCI⁺) [M+H] cal. for C₁₀H₁₃F₂O₂S: 235.0604; found: 235.0599.



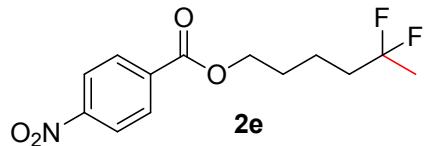
Colorless oil, isolated yield: 71%.

¹H NMR (500 MHz, CDCl₃) δ = 7.82 (dd, *J* = 3.8, 1.2 Hz, 1H), 7.58 (dd, *J* = 5.0, 1.2 Hz, 1H), 7.12 (dd, *J* = 5.0, 3.8 Hz, 1H), 4.50 (t, *J* = 6.5 Hz, 2H), 2.36 (tt, *J* = 15.2, 6.5 Hz, 2H), 1.71 (t, *J* = 18.7 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ = 161.87, 133.68, 133.32, 132.65, 127.83, 122.90 (t, *J* = 238.1 Hz), 59.30 (t, *J* = 6.2 Hz), 37.11 (t, *J* = 26.2 Hz), 23.79 (t, *J* = 27.4 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ = -89.43 (qt, *J* = 18.7, 15.2 Hz, 2F).

HRMS: (APCI⁺) [M+H] cal. for C₉H₁₁F₂O₂S: 221.0448; found: 221.0442.



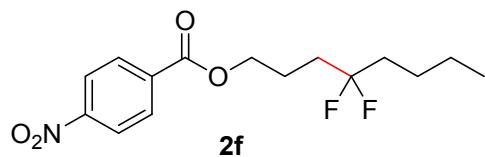
White solid, isolated yield: 83%.

¹H NMR (500 MHz, CDCl₃) δ = 8.30 (d, *J* = 8.9 Hz, 2H), 8.21 (d, *J* = 8.9 Hz, 2H), 4.40 (t, *J* = 6.5 Hz, 1H), 1.98 – 1.90 (m, 2H), 1.89 – 1.84 (m, 2H), 1.70 – 1.62 (m, 2H), 1.61 (t, *J* = 18.7 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ = 164.79, 150.65, 135.75, 130.79, 124.11 (t, *J* = 238.0 Hz), 123.68, 65.59, 37.57 (t, *J* = 25.6 Hz), 28.41, 23.51 (t, *J* = 28.0 Hz), 19.41 (t, *J* = 4.5 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ = -90.73 – -91.36 (m, 2F).

HRMS: (EI⁺) [M] cal. for C₁₃H₁₅F₂NO₄: 287.0969; found: 287.0966.



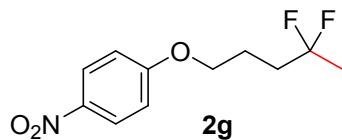
White solid, isolated yield: 81%.

¹H NMR (400 MHz, CDCl₃) δ = 8.22 – 8.13 (m, 2H), 8.12 – 8.03 (m, 2H), 4.30 (t, *J* = 6.1 Hz, 1H), 1.93 – 1.81 (m, 4H), 1.80 – 1.67 (m, 2H), 1.38 – 1.31 (m, 2H), 1.29 – 1.20 (m, 2H), 0.80 (t, *J* = 7.2 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ = 164.63, 150.61, 135.54, 130.72, 124.80 (t, *J* = 240.6 Hz), 123.62, 65.27, 36.38 (t, *J* = 25.1 Hz), 32.99 (t, *J* = 26.2 Hz), 29.74, 24.44 (t, *J* = 4.3 Hz), 22.50, 21.82 (t, *J* = 4.2 Hz), 13.88.

¹⁹F NMR (376 MHz, CDCl₃) δ = -97.54 – -97.72 (m, 0.07F), -98.62 – -98.79 (m, 2F).

HRMS: (EI⁺) [M] cal. for C₁₅H₁₉F₂NO₄: 315.1282; found: 315.1281.



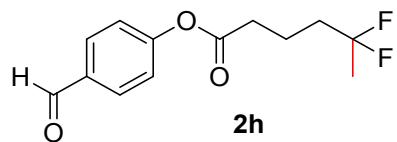
White solid, isolated yield: 46%.

¹H NMR (400 MHz, CDCl₃) δ = 7.99 – 7.92 (m, 2H), 6.74 – 6.65 (m, 2H), 3.87 (t, *J* = 5.7 Hz, 2H), 1.93 – 1.74 (m, 4H), 1.42 (t, *J* = 18.4 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ = 163.79, 141.54, 125.91, 123.84 (t, *J* = 238.2 Hz), 114.35, 67.84, 34.43 (t, *J* = 25.9 Hz), 23.56 (t, *J* = 27.9 Hz), 22.42 (t, *J* = 4.3 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ = -91.67 – -92.00 (m, 2F).

HRMS: (EI⁺) [M] cal. for C₁₁H₁₃F₂NO₃: 245.0863; found: 245.0866.



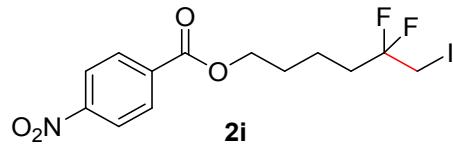
Colorless oil, isolated yield: 54%.

¹H NMR (500 MHz, CDCl₃) δ = 9.87 (s, 1H), 7.84 (d, *J* = 8.0 Hz, 2H), 7.00 (d, *J* = 8.0 Hz, 2H), 2.47 (t, *J* = 7.0 Hz, 2H), 2.02 – 1.76 (m, 4H), 1.63 (t, *J* = 18.4 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ = 191.47, 178.41, 161.87, 132.57, 129.59, 123.86 (t, *J* = 237.8 Hz), 116.01, 36.95 (t, *J* = 25.7 Hz), 33.16, 23.29 (t, *J* = 27.8 Hz), 17.85 (t, *J* = 4.7 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ = -90.94 – -91.25 (m, 2F).

HRMS: (EI⁺) [M] cal. for C₁₃H₁₄F₂O₃: 256.0911; found: 256.0906.



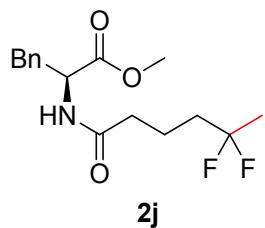
White solid, isolated yield: 72%.

¹H NMR (400 MHz, CDCl₃) δ = 8.41 (d, *J* = 8.7 Hz, 2H), 8.32 (d, *J* = 8.7 Hz, 2H), 4.52 (t, *J* = 6.4 Hz, 2H), 3.54 (t, *J* = 14.3 Hz, 2H), 2.42 – 2.18 (m, 2H), 2.07 – 1.92 (m, 2H), 1.89 – 1.73 (m, 2H).

¹³C NMR (100 MHz, CDCl₃) δ = 164.50, 150.41, 135.41, 130.54, 123.43, 120.75 (t, *J* = 242.5 Hz), 65.13, 34.73 (t, *J* = 25.0 Hz), 27.96, 18.82 (t, *J* = 4.0 Hz), 3.55 (t, *J* = 31.8 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ = -94.76 – -94.96 (m, 2F).

HRMS: (EI⁺) [M] cal. for C₁₃H₁₄F₂INO₄: 412.9936; found: 412.9919.



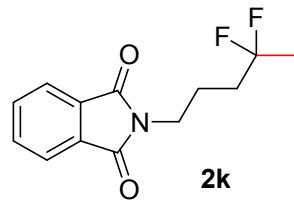
White solid, isolated yield: 68%.

¹H NMR (400 MHz, CDCl₃) δ = 7.30 – 7.18 (m, 3H), 7.09 – 7.03 (m, 2H), 5.92 (d, *J* = 7.7 Hz, 1H), 4.87 (dt, *J* = 7.8, 6.0 Hz, 1H), 3.71 (s, 3H), 3.13 (dd, *J* = 13.9, 5.7 Hz, 1H), 3.04 (dd, *J* = 13.9, 6.2 Hz, 1H), 2.19 (t, *J* = 6.7 Hz, 2H), 1.86 – 1.72 (m, 4H), 1.54 (t, *J* = 18.5 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ = 172.08, 171.65, 135.78, 129.17, 128.56, 127.13, 124.04 (t, *J* = 237.9 Hz), 52.92, 52.32, 37.86, 36.93 (t, *J* = 25.6 Hz), 35.45, 23.23 (t, *J* = 27.9 Hz), 18.67 (t, *J* = 4.7 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ = -90.49 – -90.81 (m, 2F).

HRMS: (APCI⁺) [M+H] cal. for C₁₆H₂₂F₂NO₃: 314.1568; found: 314.1566.



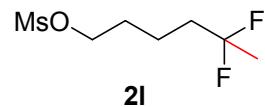
White solid, isolated yield: 65%.

¹H NMR (400 MHz, CDCl₃) δ = 7.78-7.76 (m, 2H), 7.66-7.64 (m, 2H), 3.66 (t, *J* = 6.6 Hz, 2H), 1.88-1.78 (m, 4H), 1.51 (t, *J* = 18.4 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ = 168.20, 133.91, 131.92, 123.61 (t, *J* = 238.1 Hz), 123.18, 37.31, 35.30 (t, *J* = 26.1 Hz), 23.25 (t, *J* = 27.9 Hz), 21.99 (t, *J* = 4.3 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ = -90.89 – -91.12 (m, 2F).

HRMS: (EI⁺) [M] cal. for C₁₃H₁₃F₂NO₂: 253.0914; found: 253.0907.

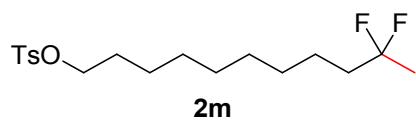


Colorless oil, isolated yield: 83%.

¹H NMR (400 MHz, CDCl₃) δ = 4.26 (t, *J* = 6.3 Hz, 2H), 3.03 (s, 3H), 2.03 – 1.78 (m, 4H), 1.69 – 1.60 (m, 2H), 1.61 (t, *J* = 18.4 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ = 123.91 (t, *J* = 237.8 Hz), 69.39, 37.33, 37.14 (t, *J* = 25.7 Hz), 28.71, 23.30 (t, *J* = 27.9 Hz), 18.81 (t, *J* = 4.6 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ = -91.27 – -91.58 (m, 2F).

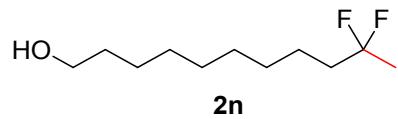


White solid, isolated yield: 44%.

¹H NMR (400 MHz, CDCl₃) δ = 7.73 (d, *J* = 8.3 Hz, 2H), 7.28 (d, *J* = 8.3 Hz, 2H), 3.96 (t, *J* = 6.5 Hz, 2H), 2.39 (s, 3H), 1.81 – 1.69 (m, 2H), 1.61 – 1.52 (m, 2H), 1.51 (t, *J* = 18.4 Hz, 3H), 1.38 (dt, *J* = 12.0, 7.6 Hz, 2H), 1.26 – 1.10 (m, 10H).

¹³C NMR (100 MHz, CDCl₃) δ = 144.58, 133.20, 129.75, 127.83, 124.38 (t, *J* = 237.4 Hz), 70.61, 37.88 (t, *J* = 25.3 Hz), 29.19, 29.16, 29.14, 28.81, 28.76, 25.26, 23.18 (t, *J* = 28.2 Hz), 22.64 (t, *J* = 4.5 Hz), 21.59.

¹⁹F NMR (376 MHz, CDCl₃) δ = -90.36 – -90.59 (m, 2F).

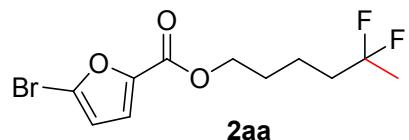


White needle crystal, isolated yield: 66%.

¹H NMR (400 MHz, CDCl₃) δ = 3.80 (t, *J* = 6.6 Hz, 2H), 2.07 – 1.89 (m, 2H), 1.77 – 1.70 (m, 2H), 1.73 (t, *J* = 18.4 Hz, 3H), 1.66 – 1.55 (m, 2H), 1.56 – 1.40 (m, 10H).

¹³C NMR (100 MHz, CDCl₃) δ = 124.43 (t, *J* = 237.4 Hz), 63.04, 37.93 (t, *J* = 25.3 Hz), 32.75, 29.38, 29.34, 29.29, 25.68, 23.18 (t, *J* = 28.1 Hz), 22.70 (t, *J* = 4.4 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ = -90.22 – -90.55 (m, 2F).



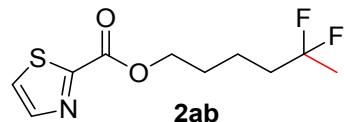
Colorless oil, isolated yield: 92%.

¹H NMR (400 MHz, CDCl₃) δ = 7.01 (d, *J* = 3.5 Hz, 1H), 6.35 (d, *J* = 3.5 Hz, 1H), 4.21 (t, *J* = 6.5 Hz, 2H), 1.87 – 1.73 (m, 2H), 1.73 – 1.65 (m, 2H), 1.55 – 1.46 (m, 2H), 1.49 (t, *J* = 18.4 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ = 157.65, 146.30, 127.54, 124.07 (t, *J* = 237.7 Hz), 120.06, 113.94, 77.39, 77.07, 76.75, 64.74, 37.49 (t, *J* = 25.6 Hz), 28.34, 23.34 (t, *J* = 28.0 Hz), 19.29 (t, *J* = 4.6 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ = -90.67 – -90.90 (m, 2F).

HRMS: (APCI⁺) [M+H] cal. for C₁₁H₁₄BrF₂O₃: 311.0094; found: 311.0090.



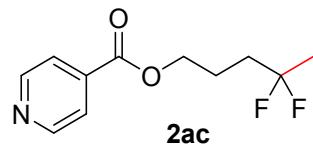
Colorless oil, isolated yield: 78%.

¹H NMR (400 MHz, CDCl₃) δ = 8.03 (d, *J* = 3.0 Hz, 1H), 7.64 (d, *J* = 3.1 Hz, 1H), 4.43 (t, *J* = 6.6 Hz, 2H), 1.97 – 1.83 (m, 4H), 1.68 – 1.60 (m, 2H), 1.59 (t, *J* = 18.4 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ = 159.99, 158.27, 145.00, 125.27, 124.00 (t, *J* = 238.0 Hz), 66.03, 37.44 (t, *J* = 25.6 Hz), 28.23, 23.30 (t, *J* = 28.0 Hz), 19.26 (t, *J* = 4.6 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ = -90.78 – -91.07 (m, 2F).

HRMS: (APCI⁺) [M+H] cal. for C₁₀H₁₄F₂NO₂S: 250.0713; found: 250.0710.



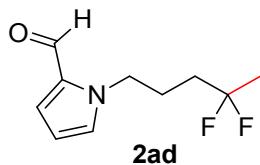
Colorless oil, isolated yield: 71%.

¹H NMR (400 MHz, CDCl₃) δ = 8.63 (d, *J* = 4.8 Hz, 2H), 7.68 (d, *J* = 4.8 Hz, 2H), 4.24 (t, *J* = 5.7 Hz, 2H), 1.91 – 1.78 (m, 4H), 1.48 (t, *J* = 18.4 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ = 164.89, 150.53, 137.17, 123.58 (d, *J* = 237.9 Hz), 122.69, 64.85, 34.46 (t, *J* = 26.1 Hz), 23.39 (t, *J* = 27.9 Hz), 21.96 (t, *J* = 4.4 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ = -91.52 – -91.82 (m, 2F).

HRMS: (APCI⁺) [M+H] cal. for C₁₁H₁₄F₂NO₂: 239.0993; found: 230.0989.



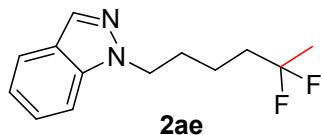
Dark red oil, isolated yield: 53%.

¹H NMR (400 MHz, CDCl₃) δ = 9.38 (s, 1H), 6.87 – 6.73 (m, 2H), 6.09 (dd, *J* = 3.8, 2.6 Hz, 1H), 4.22 (t, *J* = 7.0 Hz, 2H), 1.90 – 1.75 (m, 2H), 1.75 – 1.60 (m, 2H), 1.43 (t, *J* = 18.4 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ = 179.43, 131.39, 131.34, 125.17, 124.02 (d, *J* = 238.1 Hz), 109.85, 48.36, 34.72 (t, *J* = 25.9 Hz), 24.77 (t, *J* = 4.2 Hz), 23.44 (t, *J* = 27.9 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ = -90.67 – -90.90 (m, 2F).

HRMS: (APCI⁺) [M+H] cal. for C₁₀H₁₄F₂NO: 202.1043; found: 202.1039.



Colorless oil, isolated yield: 75%.

¹H NMR (400 MHz, CDCl₃) δ = 7.92 (d, *J* = 0.6 Hz, 1H), 7.66 (dt, *J* = 8.1, 0.9 Hz, 1H), 7.39 – 7.24 (m, 2H), 7.07 (ddd, *J* = 7.9, 6.1, 1.7 Hz, 1H), 4.37 – 4.28 (m, 2H), 1.96 – 1.86 (m, 2H), 1.85 – 1.71 (m, 2H), 1.49 – 1.39 (m, 2H), 1.46 (t, *J* = 18.4 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ = 139.38, 132.91, 126.24, 124.05 (t, *J* = 6.4 Hz), 124.02, 121.21, 120.50, 108.87, 48.47, 37.43 (t, *J* = 25.6 Hz), 29.74, 29.45, 23.24 (t, *J* = 28.0 Hz), 20.17 (t, *J* = 4.6 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ = -90.53 – -90.88 (m, 2F).

HRMS: (APCI⁺) [M+H] cal. for C₁₃H₁₇F₂N₂: 239.1360; found: 239.1356.



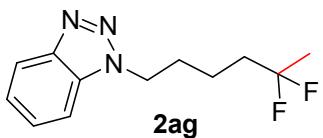
Off-white solid, isolated yield: 76%.

¹H NMR (400 MHz, CDCl₃) δ = 7.93 (s, 1H), 7.74 (d, *J* = 8.7 Hz, 1H), 7.68 (d, *J* = 8.4 Hz, 1H), 7.42 – 7.21 (m, 1H), 7.11 (dd, *J* = 7.8, 7.2 Hz, 1H), 4.45 (t, *J* = 7.0 Hz, 2H), 2.17 – 2.02 (m, 2H), 1.96 – 1.83 (m, 2H), 1.58 – 1.51 (m, 2H), 1.58 (t, *J* = 18.4 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ = 148.80, 125.76, 123.85 (t, *J* = 237.0 Hz), 122.55, 121.63, 121.54, 119.96, 117.27, 53.27, 37.22 (t, *J* = 25.7 Hz), 30.12, 23.22 (t, *J* = 27.9 Hz), 19.82 (t, *J* = 4.5 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ = -90.96 – -91.10 (m, 2F).

HRMS: (APCI⁺) [M+H] cal. for C₁₃H₁₇F₂N₂: 239.1360; found: 239.1357.



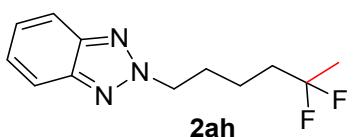
White solid, isolated yield: 80%.

¹H NMR (400 MHz, CDCl₃) δ = 8.10 (d, *J* = 8.4 Hz, 1H), 7.61 – 7.47 (m, 2H), 7.41 (t, *J* = 7.4 Hz, 1H), 4.69 (t, *J* = 7.0 Hz, 2H), 2.16 – 2.04 (m, 3H), 1.99 – 1.84 (m, 3H), 1.65 – 1.50 (m, 2H), 1.58 (t, *J* = 18.4 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ = 146.00, 132.89, 127.34, 123.95 (t, *J* = 237.0 Hz), 123.93, 120.09, 109.21, 47.80, 37.18 (t, *J* = 25.7 Hz), 29.23, 23.32 (t, *J* = 27.9 Hz), 20.01 (t, *J* = 4.5 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ = -90.58 – -90.93 (m, 2F).

HRMS: (APCI⁺) [M+H] cal. for C₁₃H₁₆F₂N₃: 240.1312; found: 240.1309.



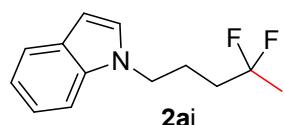
Light yellow oil, isolated yield: 82%.

¹H NMR (400 MHz, CDCl₃) δ = 7.96 (dd, J = 6.5, 3.0 Hz, 2H), 7.48 (dd, J = 6.5, 3.0 Hz, 2H), 4.84 (t, J = 7.0 Hz, 2H), 2.33 – 2.18 (m, 2H), 2.10 – 1.91 (m, 2H), 1.73 – 1.56 (m, 2H), 1.65 (t, J = 18.4 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ = 144.39, 126.38, 124.03 (t, J = 237.9 Hz), 118.04, 56.21, 37.31 (t, J = 25.7 Hz), 29.67, 23.37 (t, J = 27.9 Hz), 19.95 (t, J = 4.4 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ = -90.60 – -90.83 (m, 2F).

HRMS: (APCI⁺) [M+H] cal. for C₁₃H₁₆F₂N₃: 240.1312; found: 240.1309.



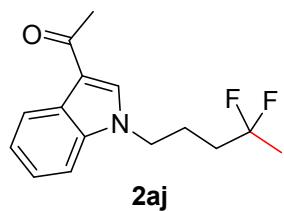
Colorless oil, isolated yield: 27%.

¹H NMR (400 MHz, CDCl₃) δ = 7.73 (d, J = 7.9 Hz, 1H), 7.43 (d, J = 8.2 Hz, 1H), 7.31 (t, J = 7.6 Hz, 1H), 7.20 – 7.18 (m, 2H), 6.60 (d, J = 3.1 Hz, 1H), 4.29 (t, J = 7.0 Hz, 2H), 2.22 – 2.08 (m, 2H), 1.98 – 1.82 (m, 2H), 1.65 (t, J = 18.4 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ = 135.84, 127.53, 126.20 (t, J = 238.5 Hz), 121.52, 121.00, 119.33, 109.14, 101.35, 45.62, 35.06 (t, J = 26.1 Hz), 23.51 (t, J = 27.5 Hz), 23.48.

¹⁹F NMR (376 MHz, CDCl₃) δ = -91.11 – -91.37 (m, 2F).

HRMS: (APCI⁺) [M+H] cal. for C₁₃H₁₆F₂N: 224.1251; found: 224.1247.



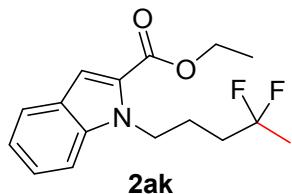
White solid, isolated yield: 60%.

¹H NMR (400 MHz, CDCl₃) δ = 8.50 – 8.44 (m, 1H), 7.81 (s, 1H), 7.48 – 7.35 (m, 3H), 4.30 (t, J = 7.2 Hz, 1H), 2.60 (s, 3H), 2.24 – 2.13 (m, 2H), 2.02 – 1.87 (m, 2H), 1.67 (t, J = 18.4 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ = 192.96, 136.63, 134.48, 126.35, 123.66 (t, *J* = 238.4 Hz), 123.38, 122.70, 122.59, 117.24, 109.63, 46.30, 34.82 (t, *J* = 25.9 Hz), 27.64, 23.64 (t, *J* = 27.8 Hz), 23.11 (t, *J* = 3.7 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ = -91.18 – -91.45 (m, 2F).

HRMS: (APCI⁺) [M+H] cal. for C₁₅H₁₈F₂NO: 266.1356; found: 266.1354.



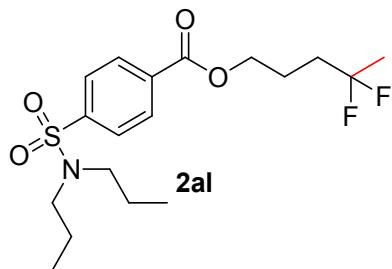
Colorless oil, isolated yield: 44%.

¹H NMR (400 MHz, CDCl₃) δ = 7.67 (d, *J* = 8.0 Hz, 1H), 7.39 (d, *J* = 8.4 Hz, 1H), 7.35 (d, *J* = 7.0 Hz, 1H), 7.32 (s, 1H), 7.15 (t, *J* = 7.2 Hz, 1H), 4.61 (t, *J* = 7.2 Hz, 2H), 4.37 (q, *J* = 7.1 Hz, 2H), 2.05 – 1.98 (m, 2H), 1.96 – 1.83 (m, 2H), 1.57 (t, *J* = 20.4, 18.4 Hz, 3H), 1.41 (t, *J* = 7.1 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ = 161.98, 138.95, 127.30, 125.96, 125.08, 123.99 (t, *J* = 237.0 Hz), 122.70, 120.62, 110.72, 110.23, 60.57, 43.96, 35.12 (t, *J* = 25.9 Hz), 23.88 (t, *J* = 3.8 Hz), 23.36 (t, *J* = 27.9 Hz), 14.34.

¹⁹F NMR (376 MHz, CDCl₃) δ = -90.53 – -90.76 (m, 2F).

HRMS: (APCI⁺) [M+H] cal. for C₁₆H₂₀F₂NO₂: 296.1462; found: 296.1459.



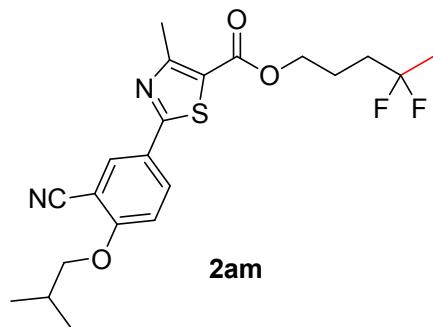
Colorless oil, isolated yield: 83%.

¹H NMR (400 MHz, CDCl₃) δ = 8.25 – 8.10 (m, 2H), 7.99 – 7.83 (m, 2H), 4.45 (t, *J* = 6.0 Hz, 2H), 3.17 – 3.12 (m, 4H), 2.18 – 1.93 (m, 4H), 1.69 (t, *J* = 18.4 Hz, 2H), 1.65 – 1.55 (m, 4H), 0.91 (t, *J* = 7.4 Hz, 6H).

¹³C NMR (100 MHz, CDCl₃) δ = 165.15, 144.37, 133.40, 130.21, 127.05, 123.76 (t, *J* = 238.1 Hz), 64.80, 49.94, 34.64 (t, *J* = 26.0 Hz), 23.52 (t, *J* = 27.9 Hz), 22.17 (t, *J* = 4.4 Hz), 21.95, 11.17.

¹⁹F NMR (376 MHz, CDCl₃) δ = -91.26 – -91.53 (m, 2F).

HRMS: (ESI⁺) [M+Na] cal. for C₁₈H₂₇F₂NO₄SnA: 414.1527; found: 414.1522.



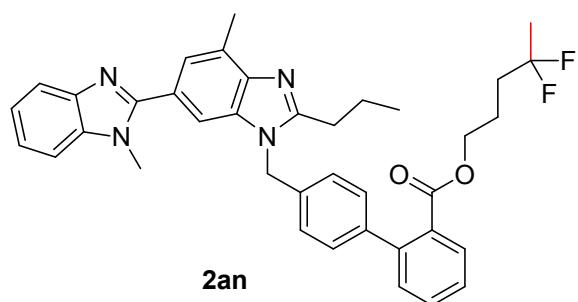
White solid, isolated yield: 71%.

¹H NMR (400 MHz, CDCl₃) δ = 8.16 (d, *J* = 2.3 Hz, 1H), 8.08 (dd, *J* = 8.9, 2.3 Hz, 1H), 7.00 (d, *J* = 8.9 Hz, 1H), 4.33 (t, *J* = 6.0 Hz, 2H), 3.89 (d, *J* = 6.5 Hz, 2H), 2.75 (s, 3H), 2.20 (dt, *J* = 13.3, 6.7 Hz, 1H), 2.08 – 1.89 (m, 4H), 1.64 (t, *J* = 18.4 Hz, 3H), 1.08 (d, *J* = 6.7 Hz, 6H).

¹³C NMR (100 MHz, CDCl₃) δ = 167.29, 162.49, 161.85, 161.37, 132.52, 132.06, 125.89, 123.72 (t, *J* = 237.0 Hz), 121.44, 115.34, 112.58, 102.94, 75.67, 64.45, 34.58 (t, *J* = 26.1 Hz), 28.12, 23.47 (t, *J* = 27.9 Hz), 22.15 (t, *J* = 4.4 Hz), 19.02, 17.48.

¹⁹F NMR (376 MHz, CDCl₃) δ = -91.26 – -91.53 (m, 2F).

HRMS: (ESI⁺) [M+Na] cal. for C₂₁H₂₄F₂N₂NaO₃S: 445.1373; found: 445.1370.



Colorless syrup, isolated yield: 71%.

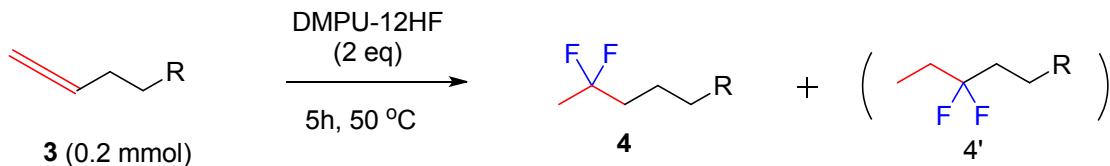
¹H NMR (500 MHz, CDCl₃) δ = 7.78 (dd, *J* = 10.7, 5.8 Hz, 2H), 7.51 – 7.32 (m, 5H), 7.29 – 7.21 (m, 5H), 7.08 (d, *J* = 8.1 Hz, 2H), 5.42 (s, 2H), 4.01 (t, *J* = 5.5 Hz, 2H), 3.78 (s, 3H), 2.94 – 2.88 (m, 2H), 2.75 (s, 3H), 1.86 (dt, *J* = 13.2, 6.6 Hz, 2H), 1.56 – 1.51 (m, 4H), 1.45 (t, *J* = 18.4 Hz, 3H), 1.04 (t, *J* = 7.3 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ = 168.32, 156.47, 154.66, 143.16, 142.79, 141.61, 141.27, 136.59, 134.97, 131.37, 130.68, 130.62, 129.93, 129.45, 128.94, 127.39, 125.95, 123.84, 123.80, 123.78 (t, *J* = 237.5 Hz), 122.46, 122.28, 119.49, 109.45, 108.85, 64.12, 46.96, 34.30 (t, *J* = 25.8 Hz), 31.74, 29.75, 23.15 (t, *J* = 27.9 Hz), 21.88, 21.76 (t, *J* = 4.5 Hz), 16.85, 14.02.

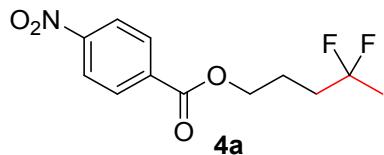
¹⁹F NMR (376 MHz, CDCl₃) δ = -90.53 – -90.85 (m, 2F).

HRMS: (ESI⁺) [M+H] cal. for C₃₈H₃₉F₂N₄O₂: 621.3041; found: 621.3037.

3.2 Hydrofluorination of alkenes.



An 8-mL polytetrafluoroethylene (PTFE) vial fitted with a stirring bar was charged alkyne starting material (0.2 mmol). DPMU-12HF (134 μL, 24 equiv based on HF) was then added in one portion at room temperature. The reaction was heated to 50 °C and stirred for 5 hours. The progress of reaction was monitored by TLC (visualized by KMnO₄ stain). The product usually shows a slightly more polar spot than the starting material on TLC (*R*_f difference < 0.1 in most cases). The reaction was cooled down to 0 °C with ice bath and calcium chloride powder was added to quench the reaction until no bubbling was observed. The crude was mixed with silica gel and concentrated. The result residue was purified with flash chromatography.



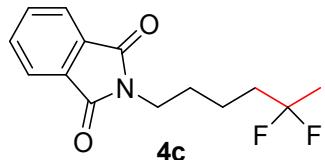
Yellow solid, isolated yield: 66% (**4a** : **4a'** = 66 : 1)

¹H NMR (400 MHz, CDCl₃) δ = 8.28 (d, J = 8.9 Hz, 2H), 8.19 (d, J = 8.8 Hz, 2H), 4.41 (t, J = 5.7 Hz, 2H), 2.05 – 1.92 (m, 4H), 1.63 (t, J = 18.4 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ = 164.14, 150.14, 135.07, 130.26, 123.34 (t, J = 238.0 Hz), 123.13, 64.68, 34.13 (t, J = 26.1 Hz), 23.04 (t, J = 27.8 Hz), 21.68 (t, J = 4.4 Hz).

¹⁹F NMR (376 MHz, CDCl₃) δ = -91.26 – -91.73 (m, 2F), -99.80 – -99.97 (m, 0.03F).

HRMS: (EI⁺) [M] cal. for C₁₂H₁₃F₂NO₄: 273.0813; found: 273.0808.



White solid, isolated yield: 61% (**4c** : **4c'** = 5 : 1)

¹H NMR (400 MHz, CDCl₃) δ = 7.86 – 7.80 (m, 2H), 7.73 – 7.67 (m, 2H), 3.75 – 3.65 (m, 2H), 1.97 – 1.79 (m, 2H), 1.78 – 1.66 (m, 2H), 1.62 – 1.47 (m, 4H), 0.98 (t, J = 7.5 Hz, 0.45H).

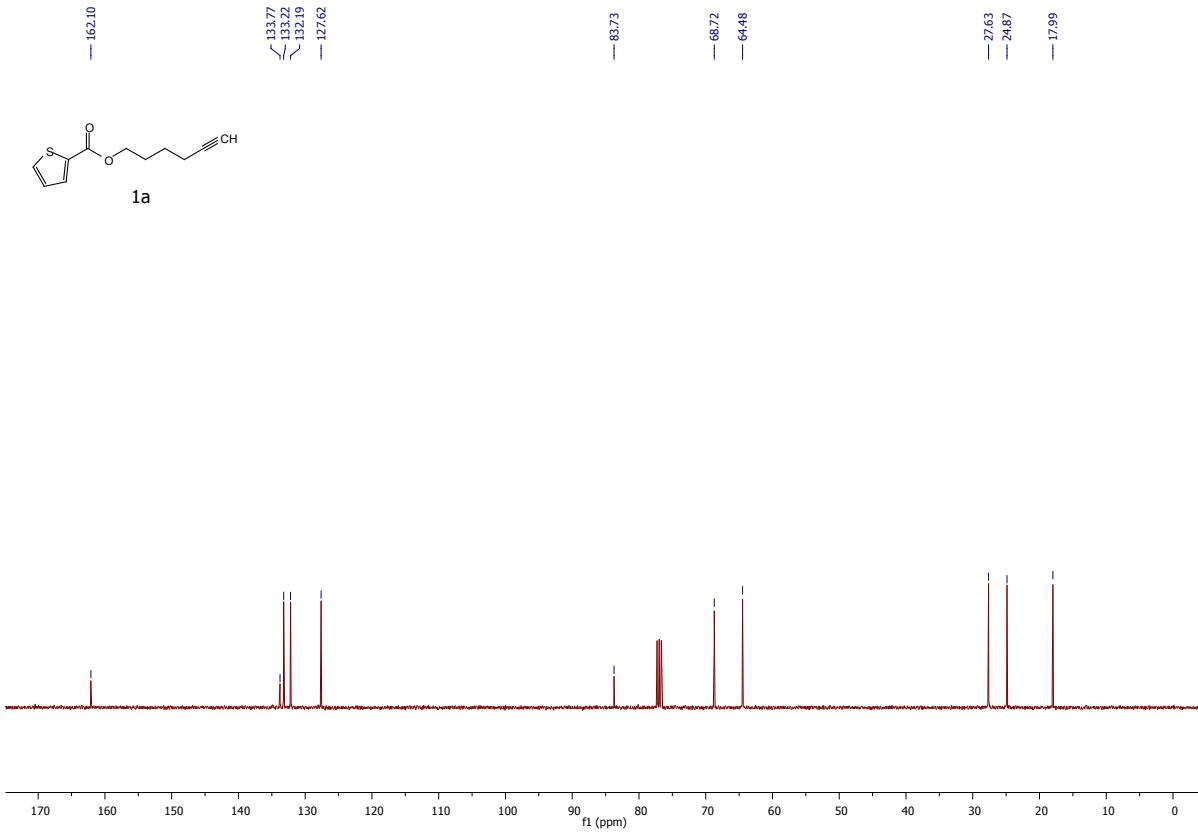
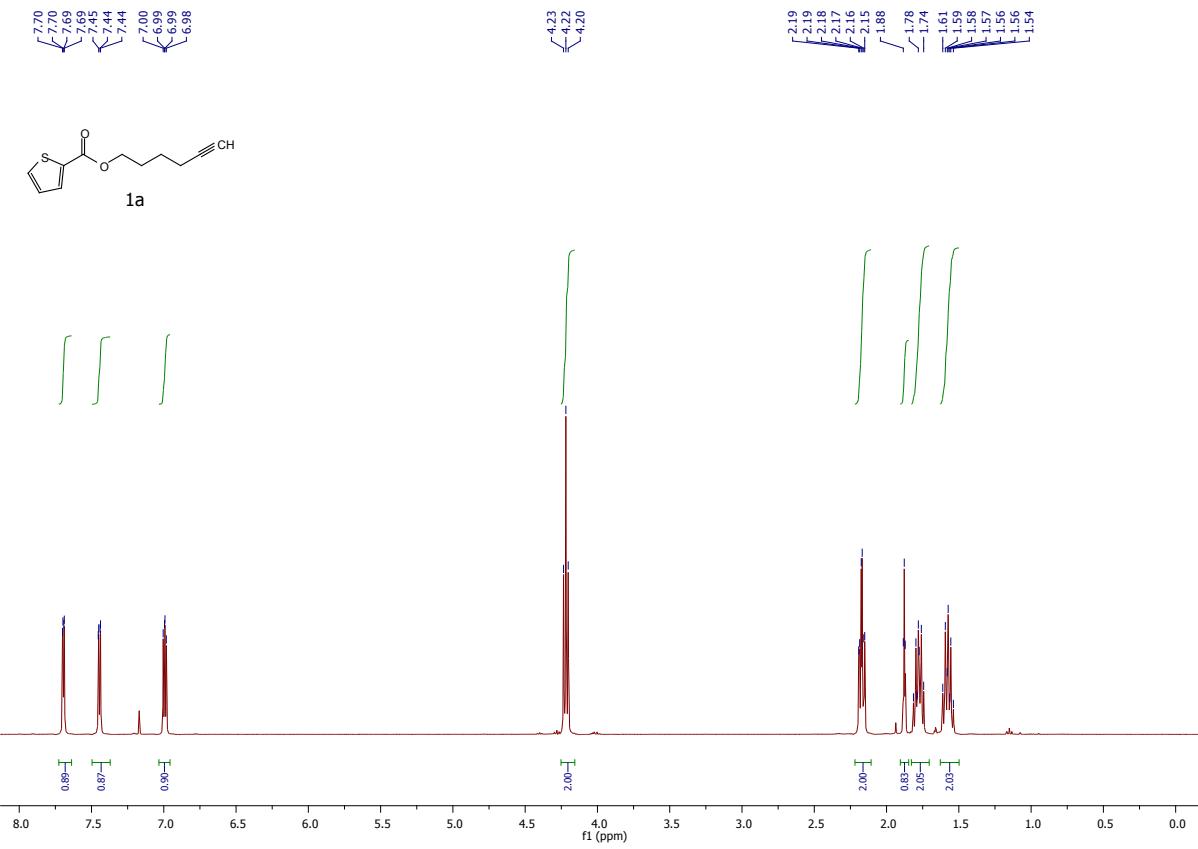
¹³C NMR (100 MHz, CDCl₃) δ = 168.34, 133.91, 132.05, 124.02 (t, J = 237.0 Hz), 123.20, 37.52, 37.36 (t, J = 26.2 Hz), 34.84, 33.39 (t, J = 26.2 Hz), 32.45, 29.60 (t, J = 26.2 Hz), 28.21, 23.51, 23.23, 22.96, 23.23 (t, J = 27.9 Hz), 21.69 (t, J = 4.0 Hz), 20.00 (t, J = 4.6 Hz, 1H).

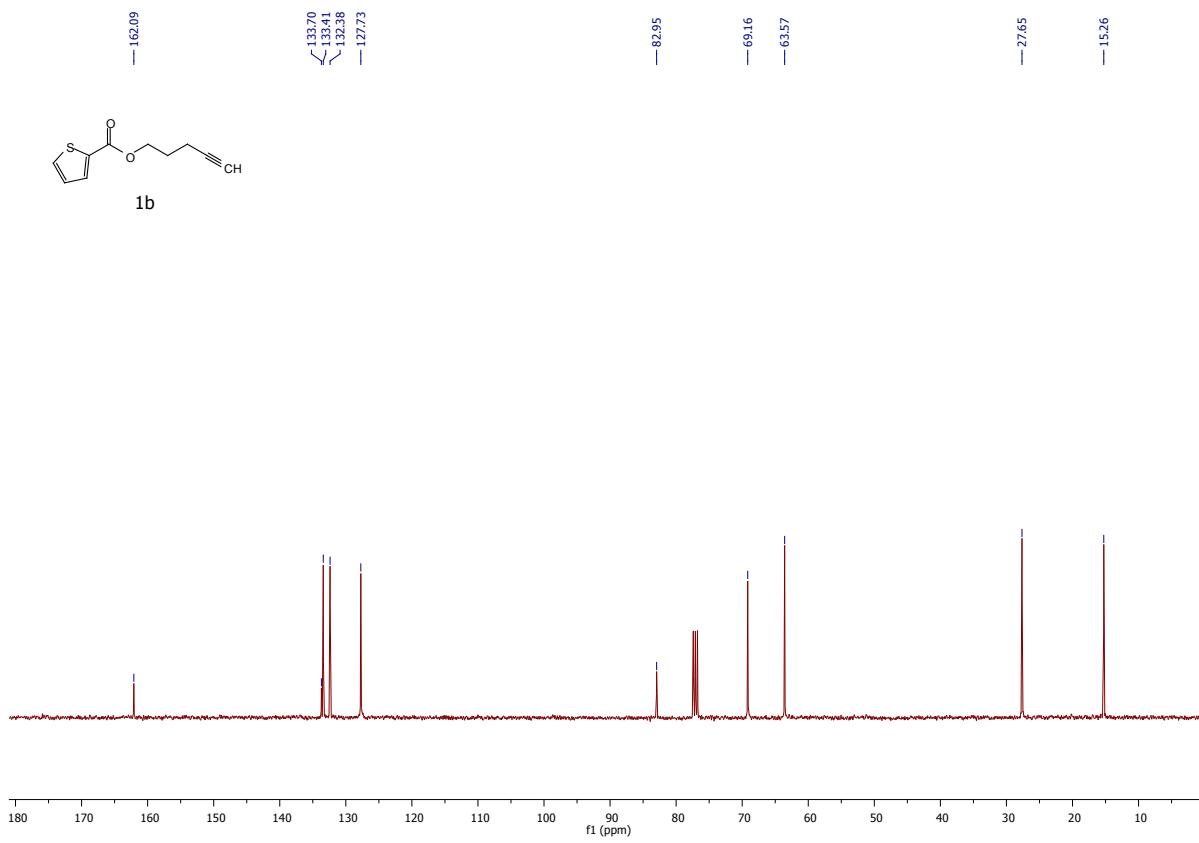
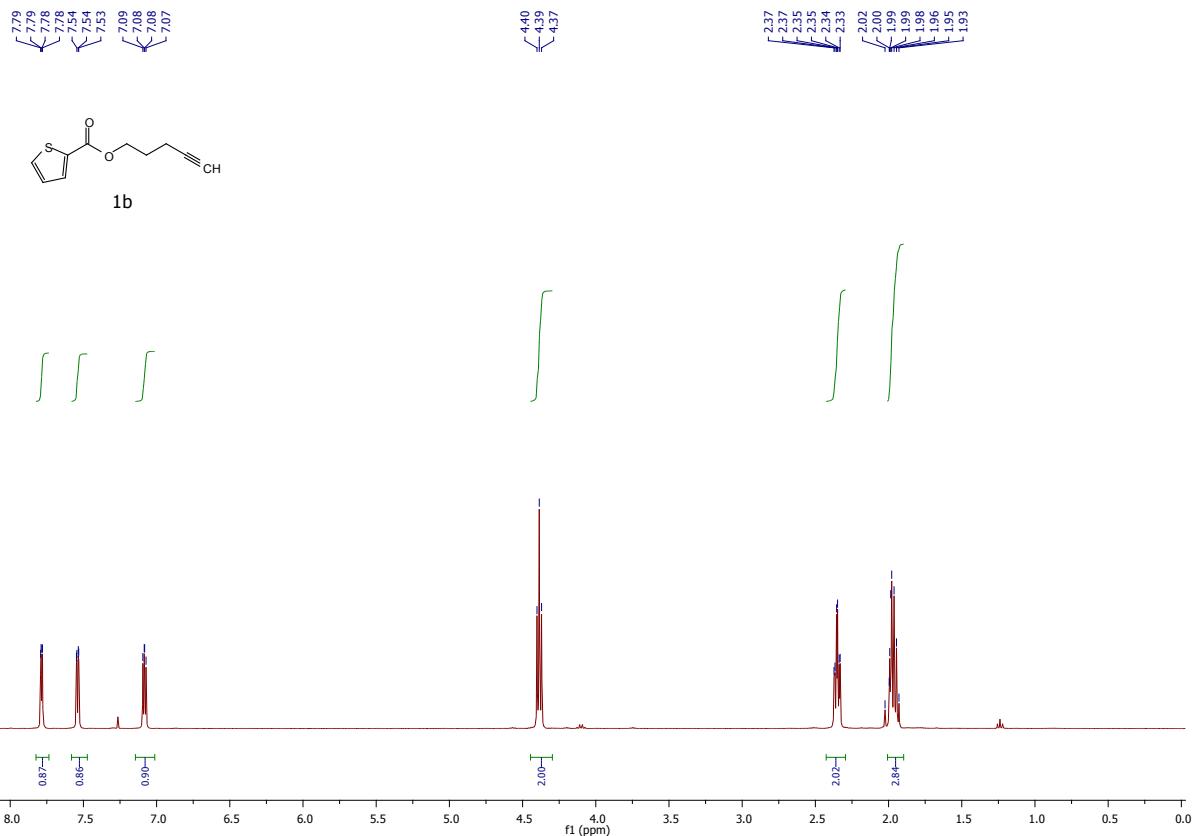
¹⁹F NMR (376 MHz, CDCl₃) δ = -90.58 – -90.89 (m, 2F), -100.39 – -100.60 (m, 0.36F).

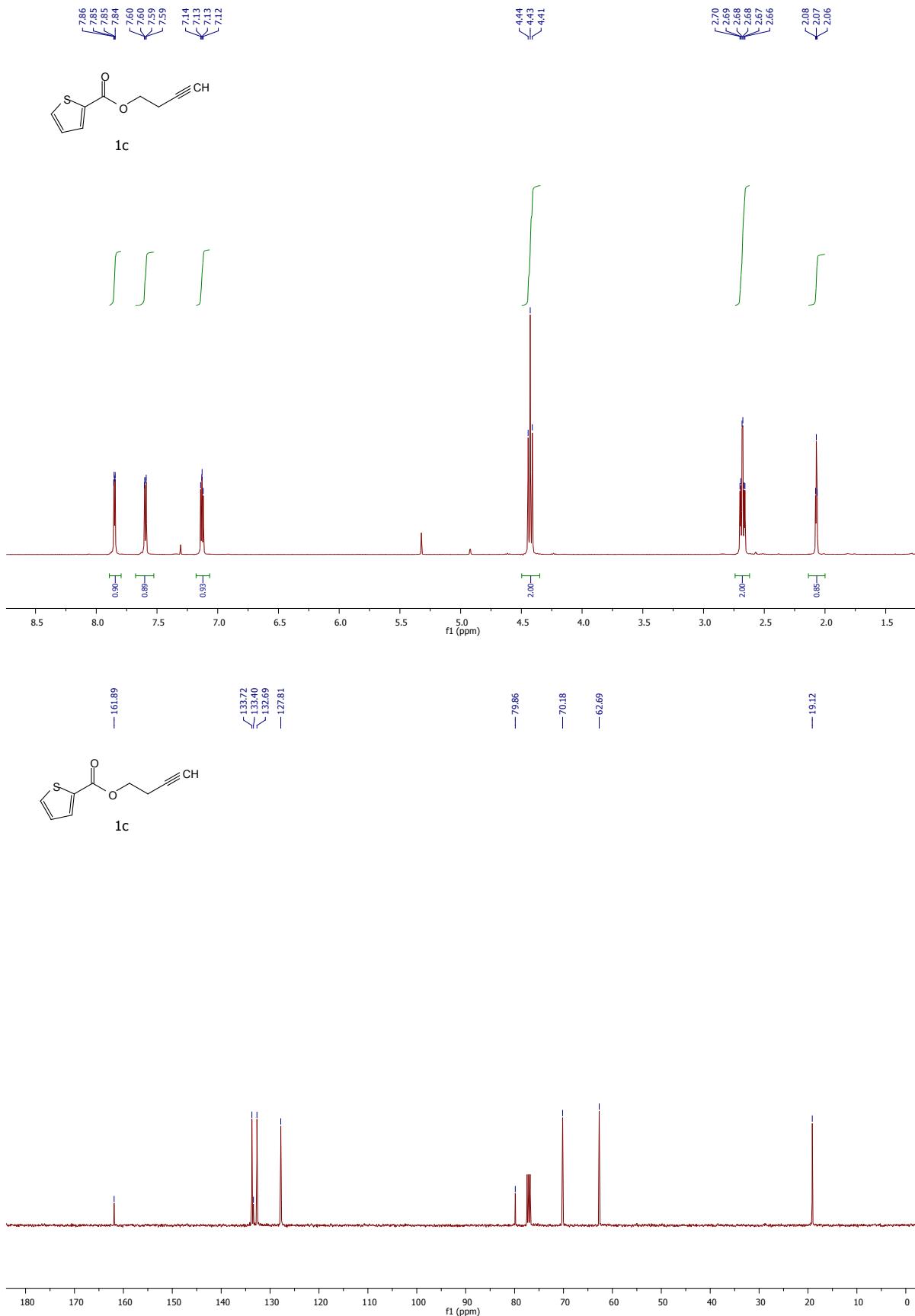
HRMS: (EI⁺) [M] cal. for C₁₄H₁₅F₂NO₂: 267.1071; found: 267.1067.

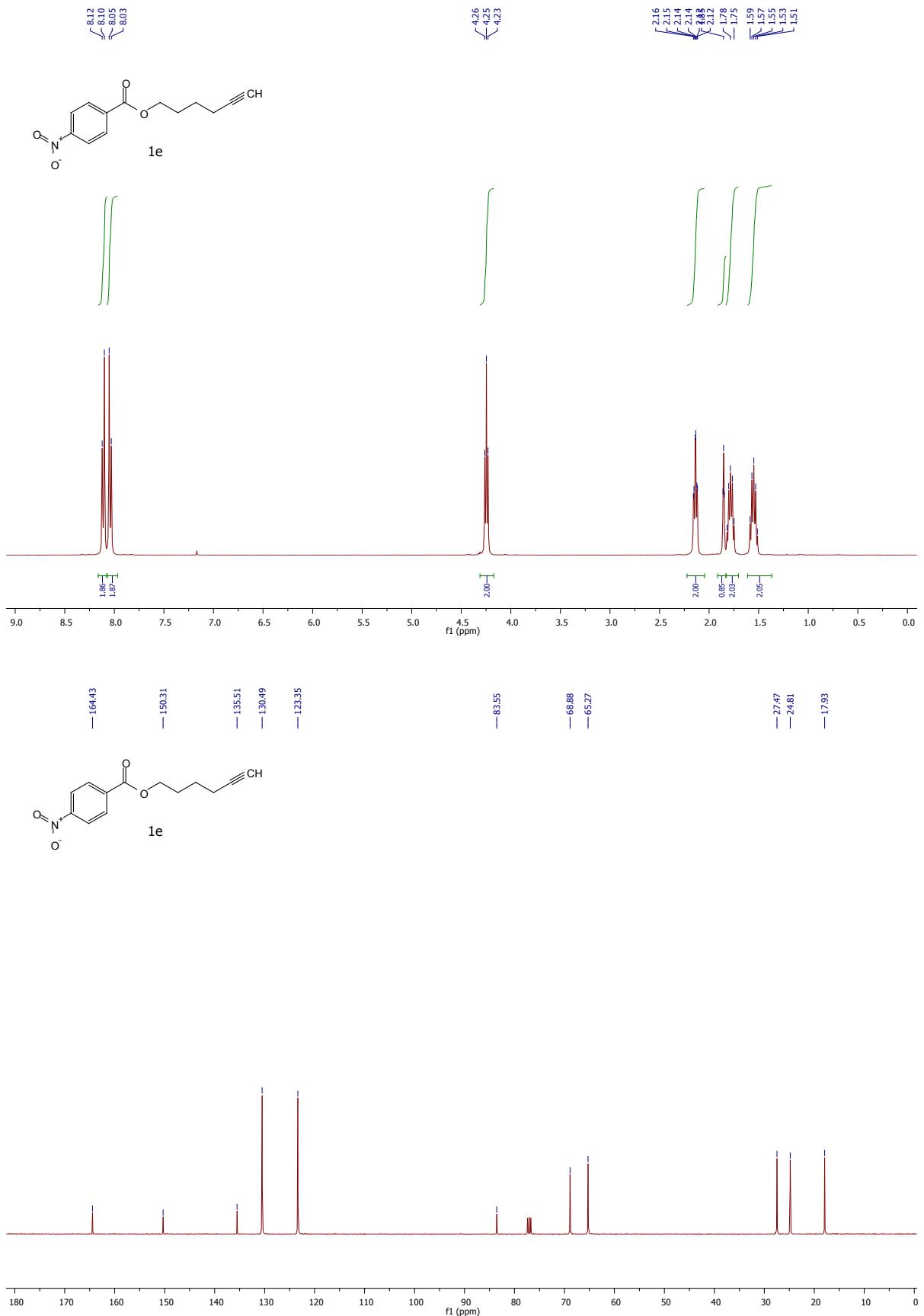
Section 4. Spectral data of starting materials and products

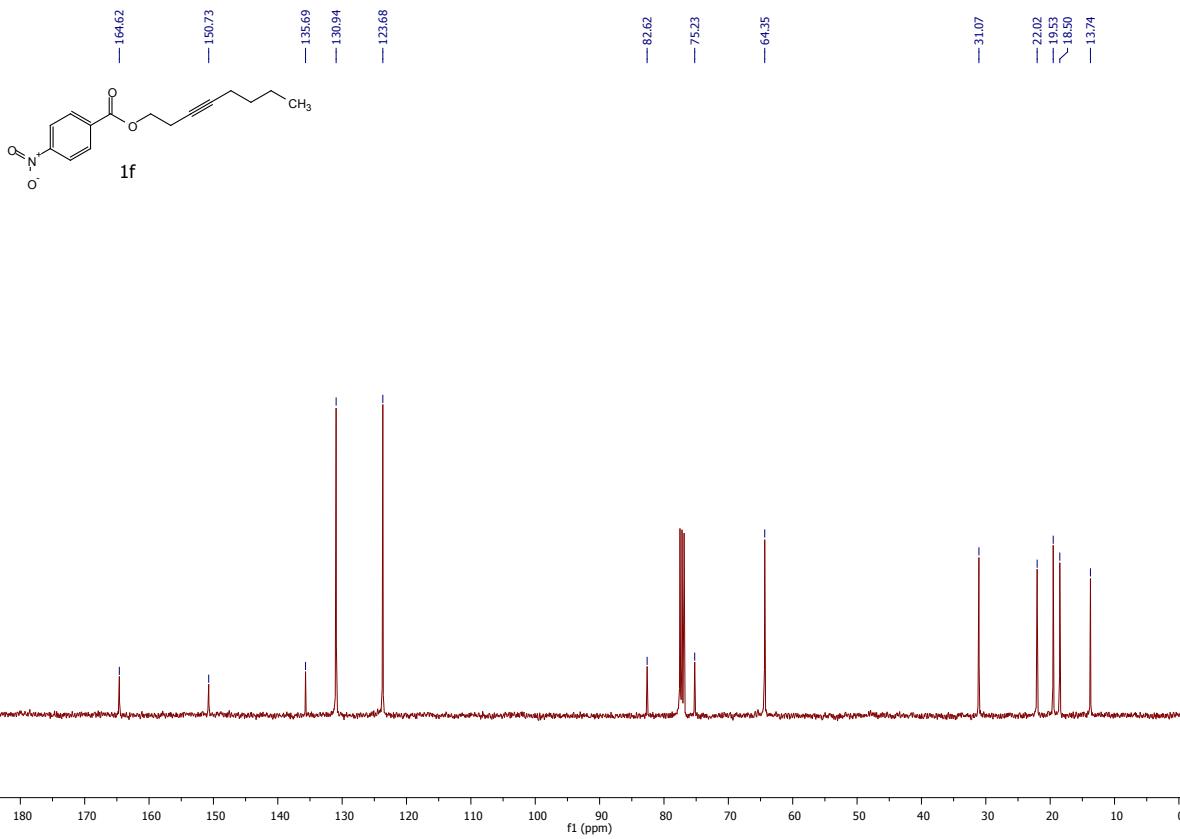
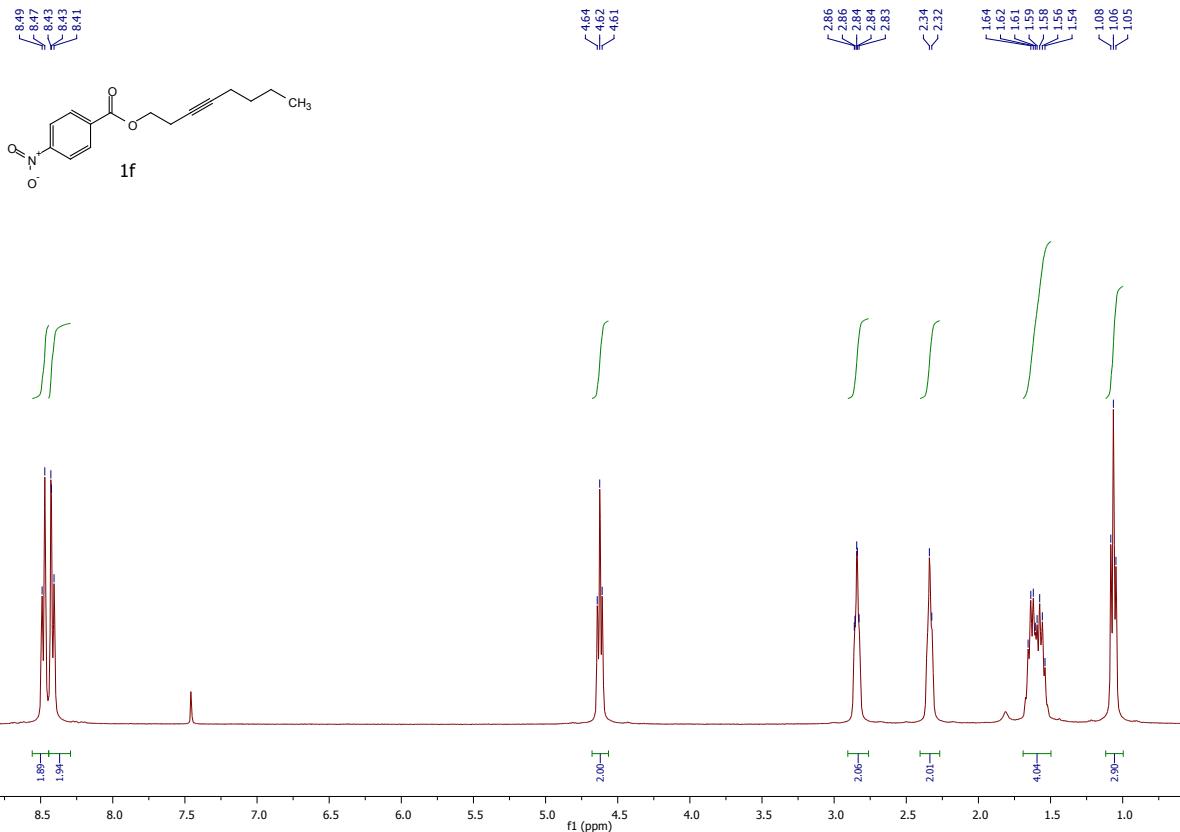
4.1 Starting materials NMR spectra.

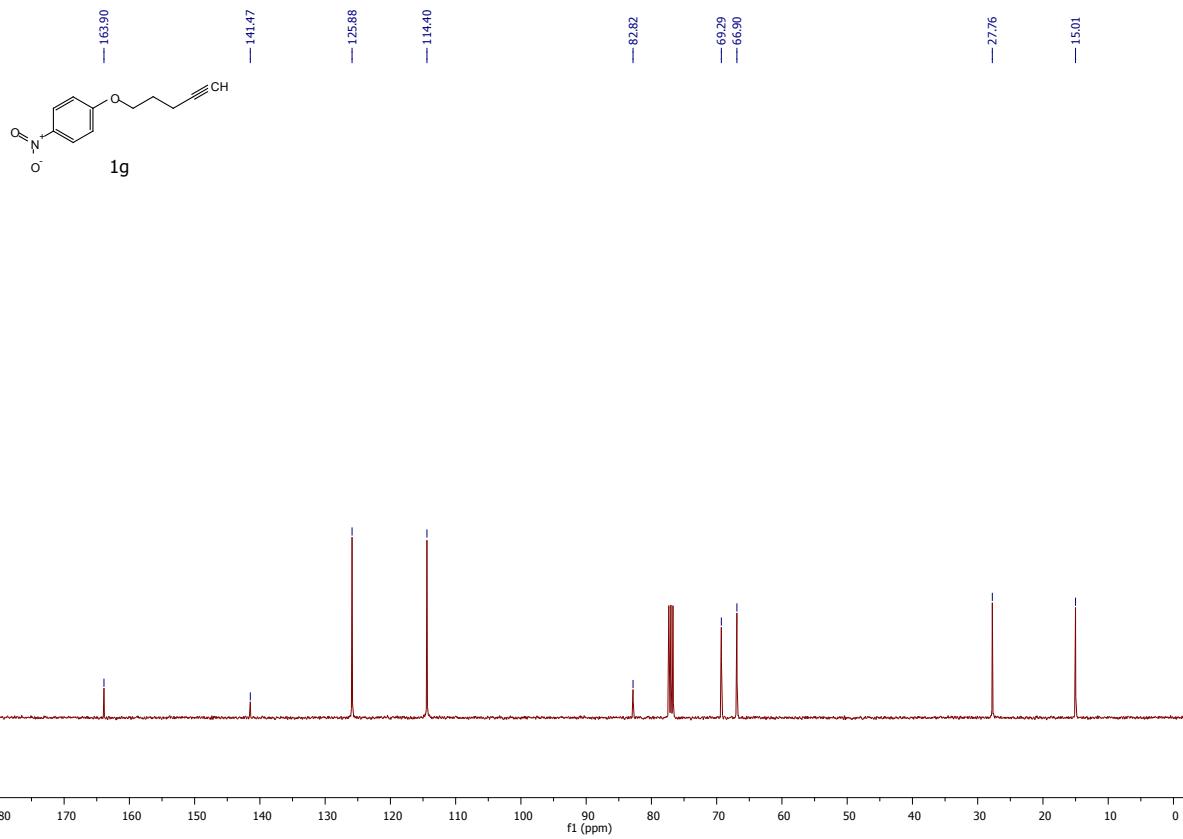
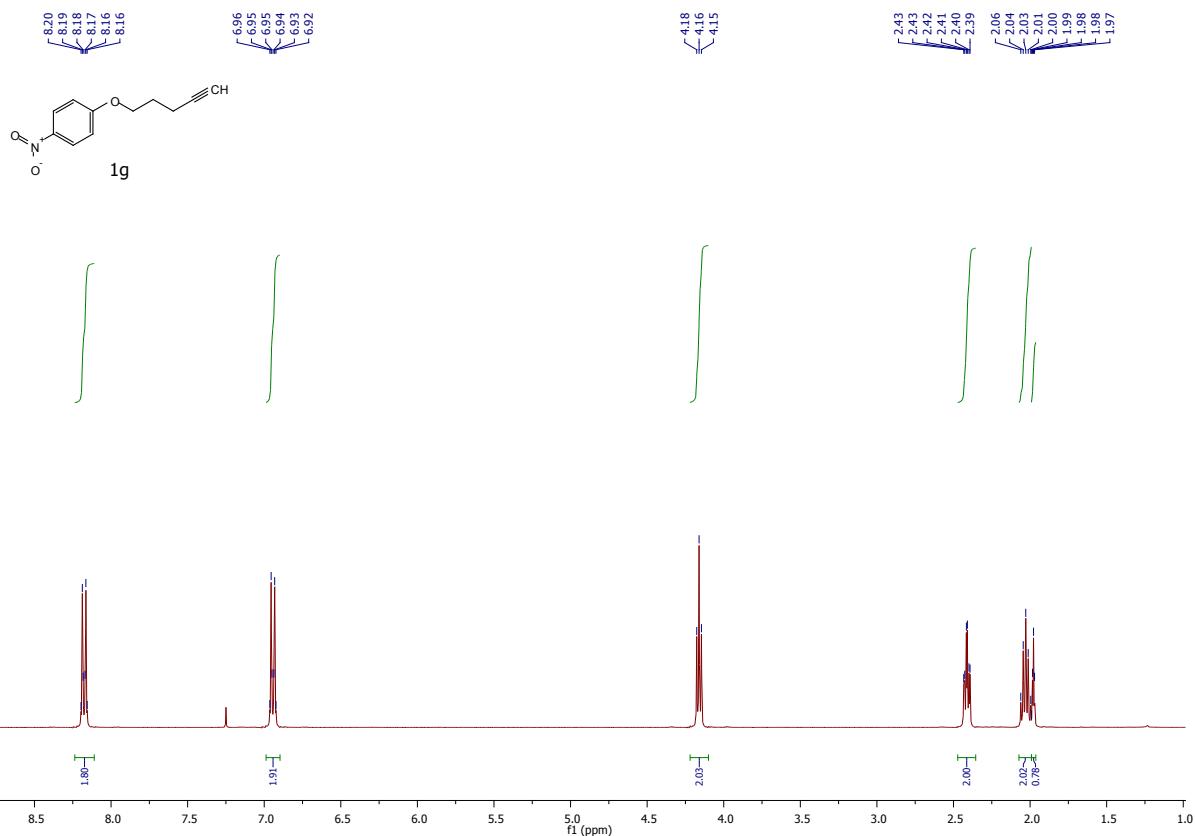


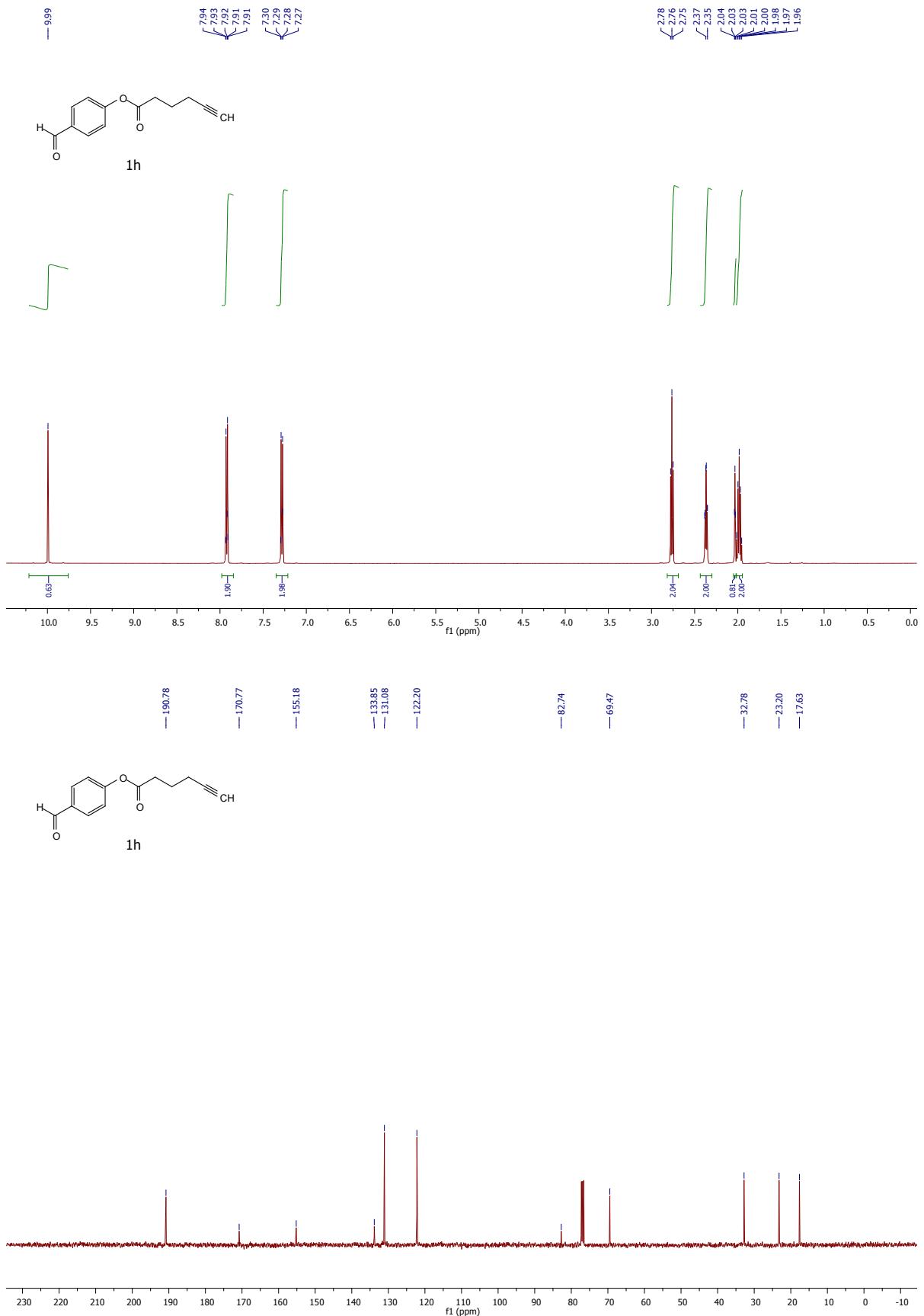


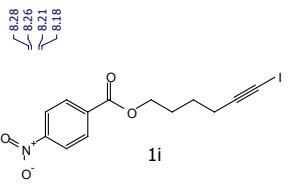








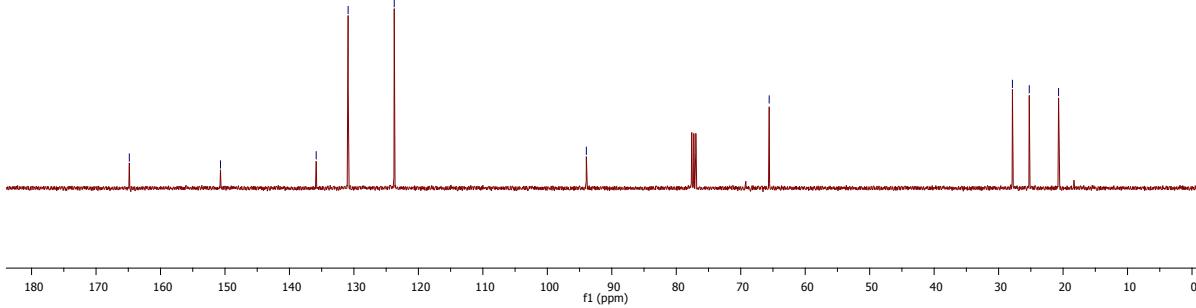
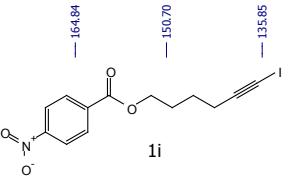
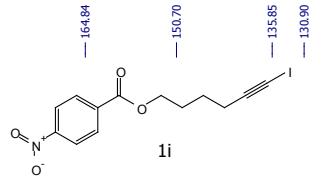
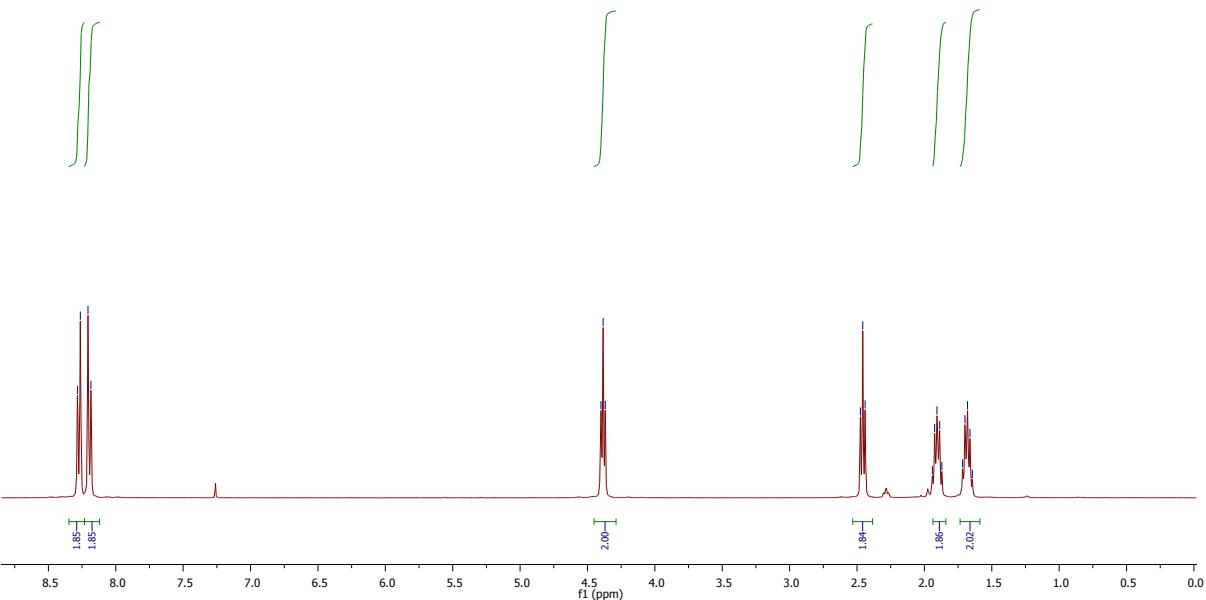


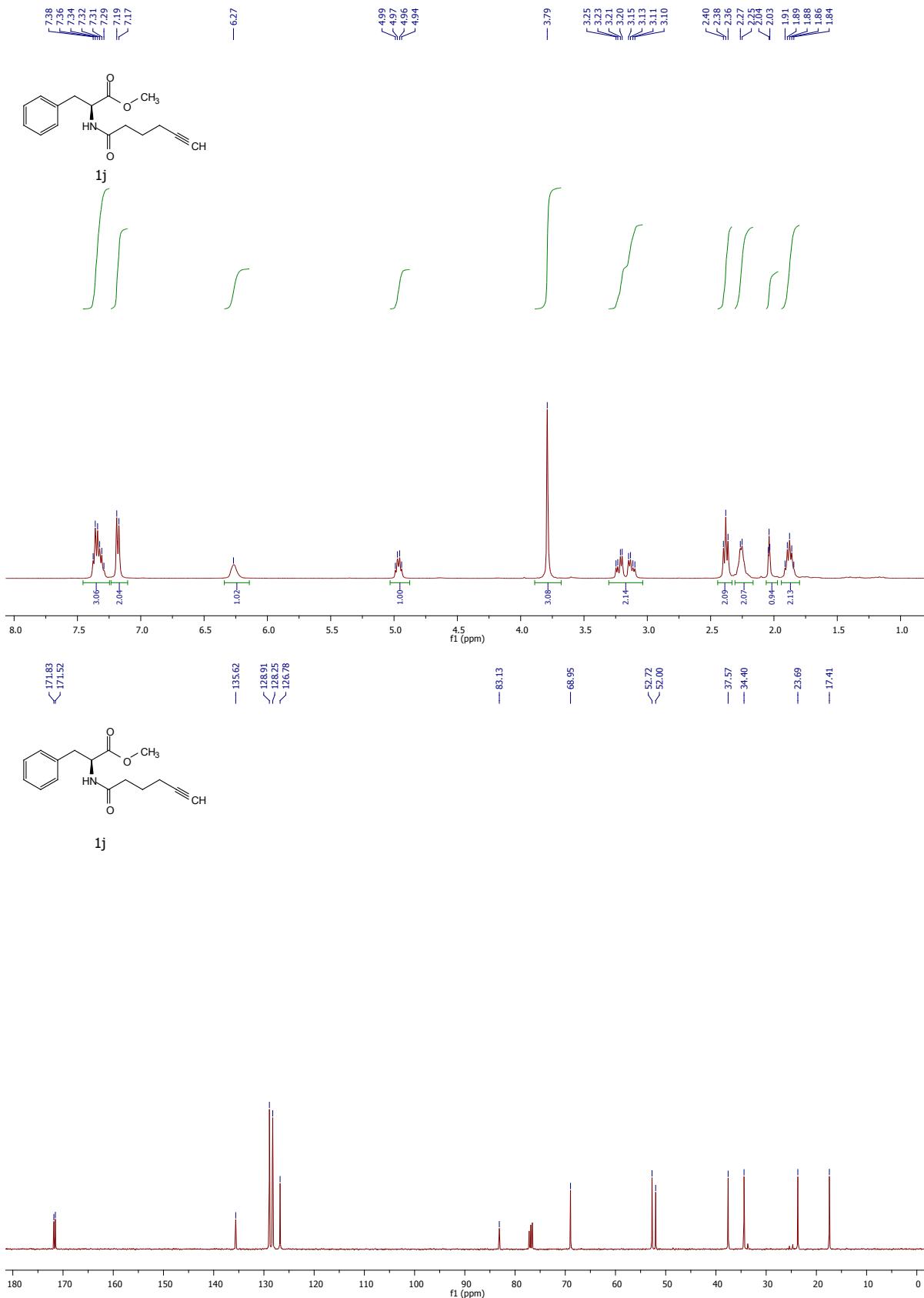


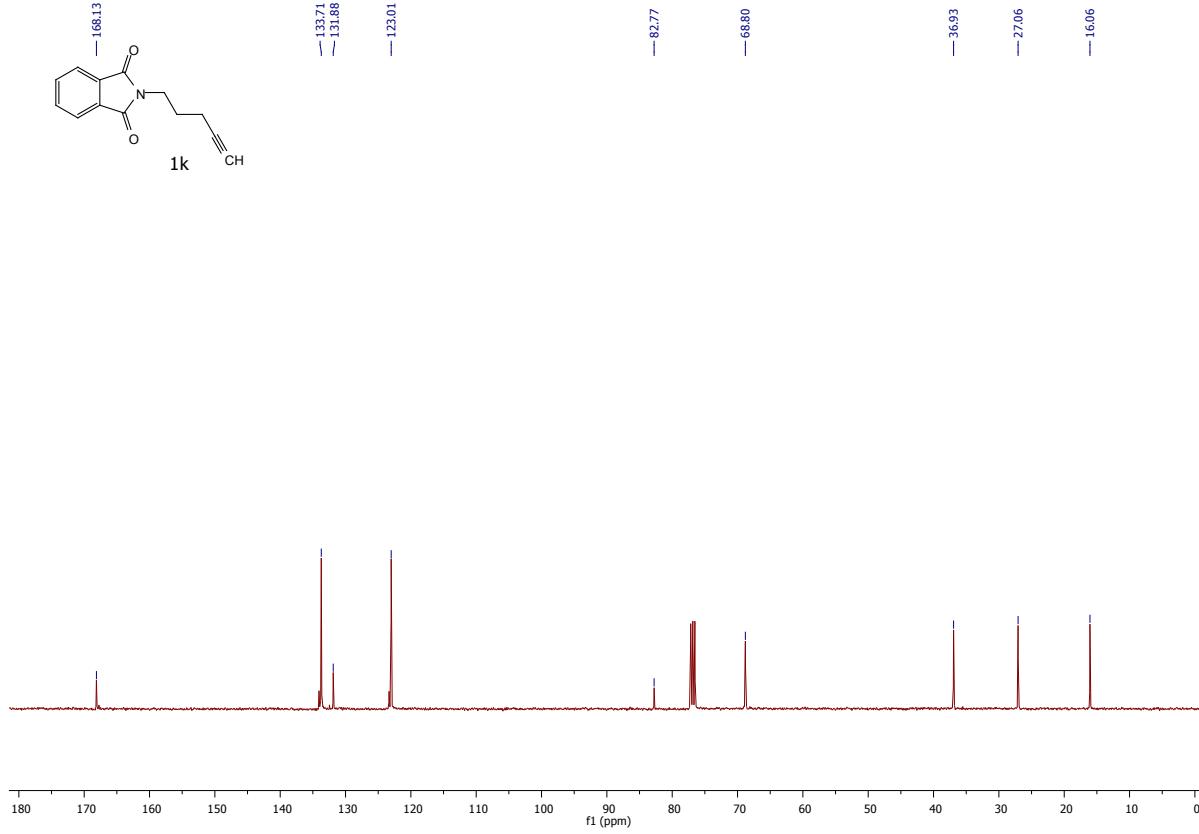
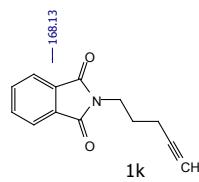
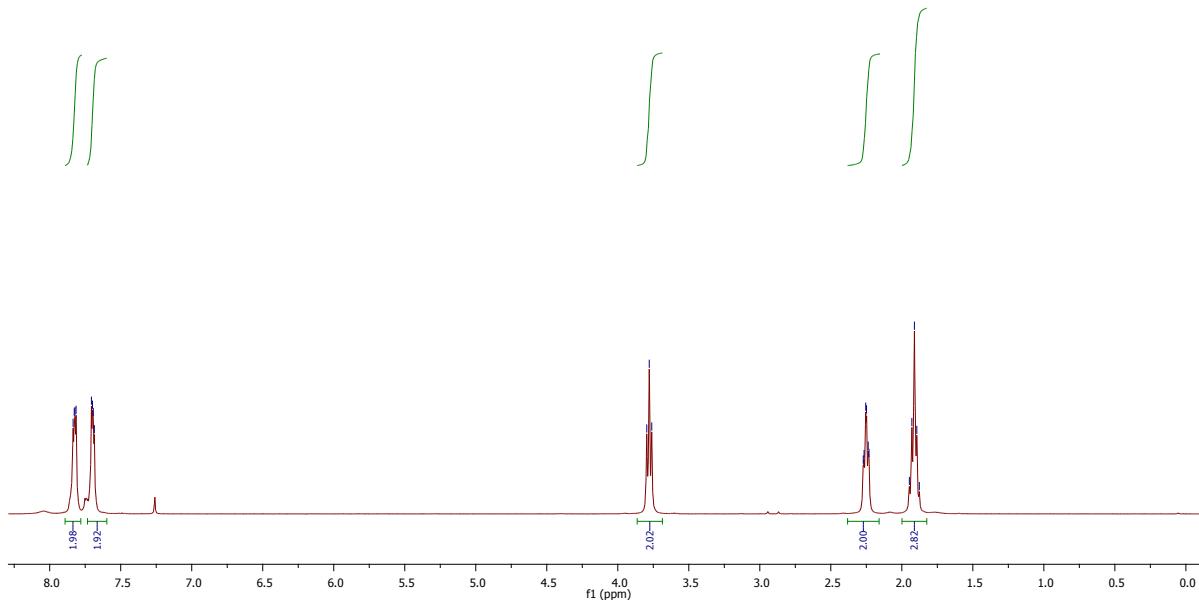
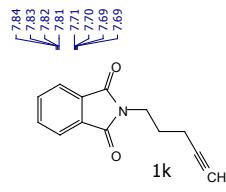
8.26
8.21
8.18

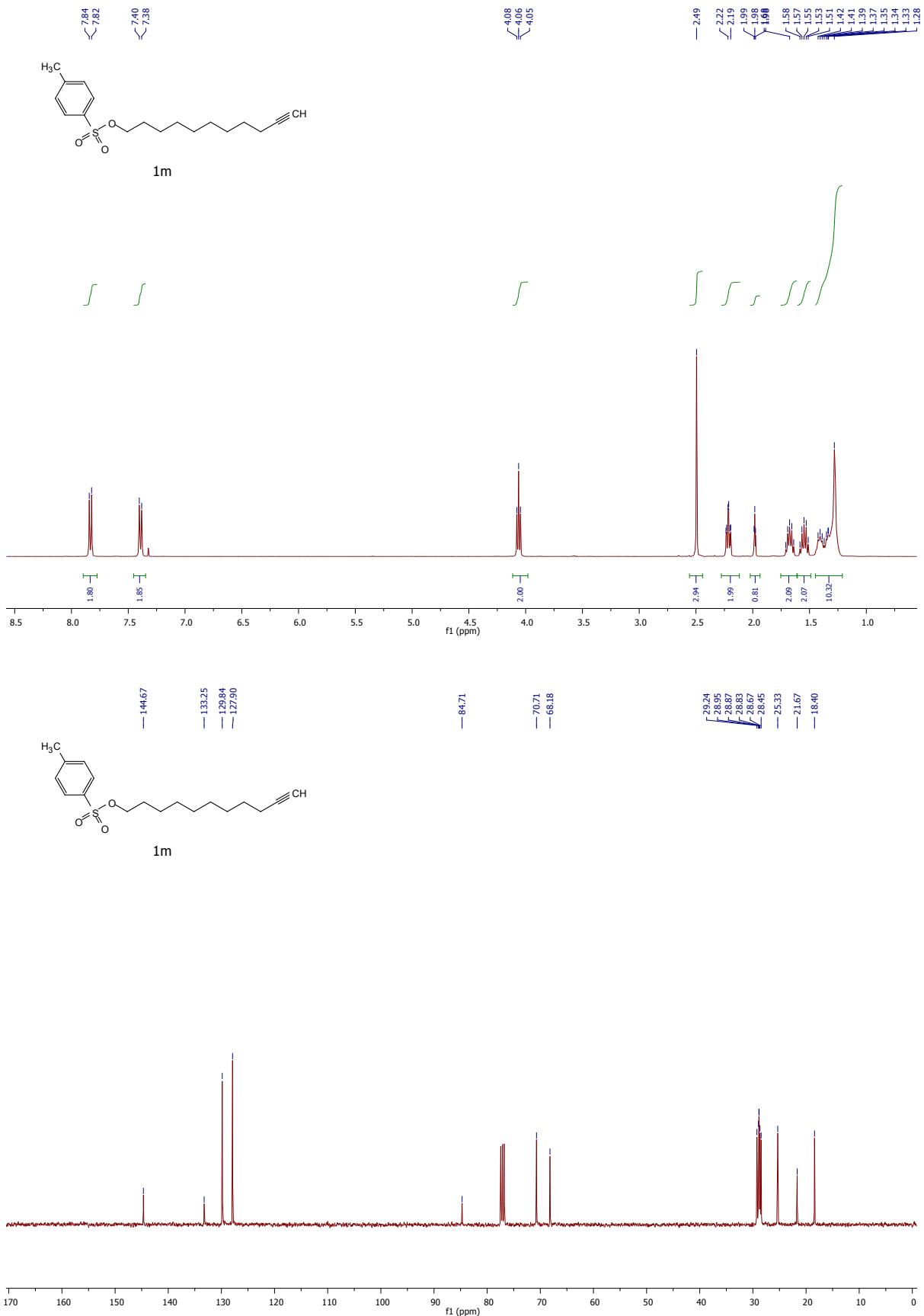
4.40
4.38
4.37

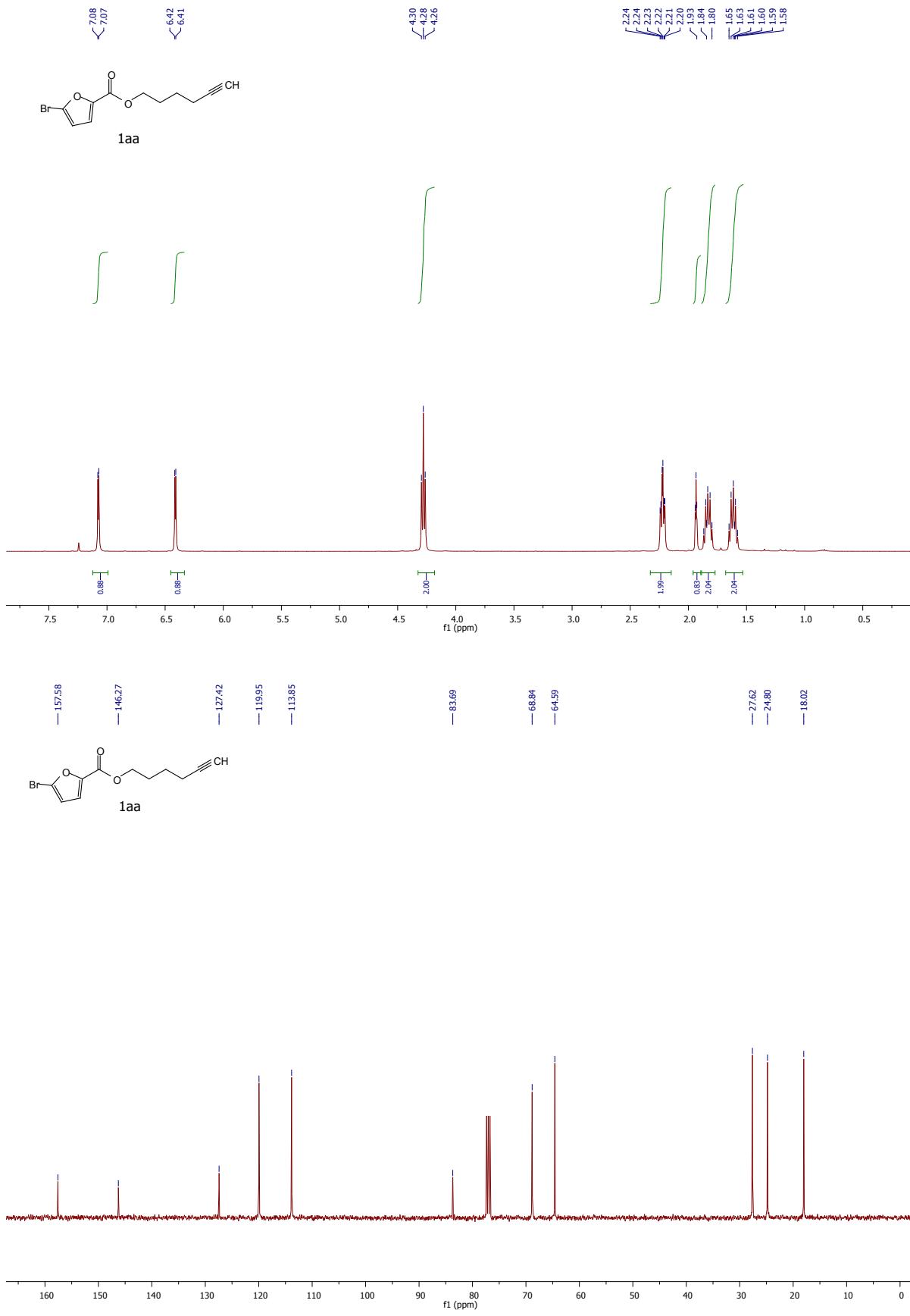
2.47
2.46
2.44
1.92
1.91
1.89
1.87
1.72
1.70
1.68
1.66
1.64

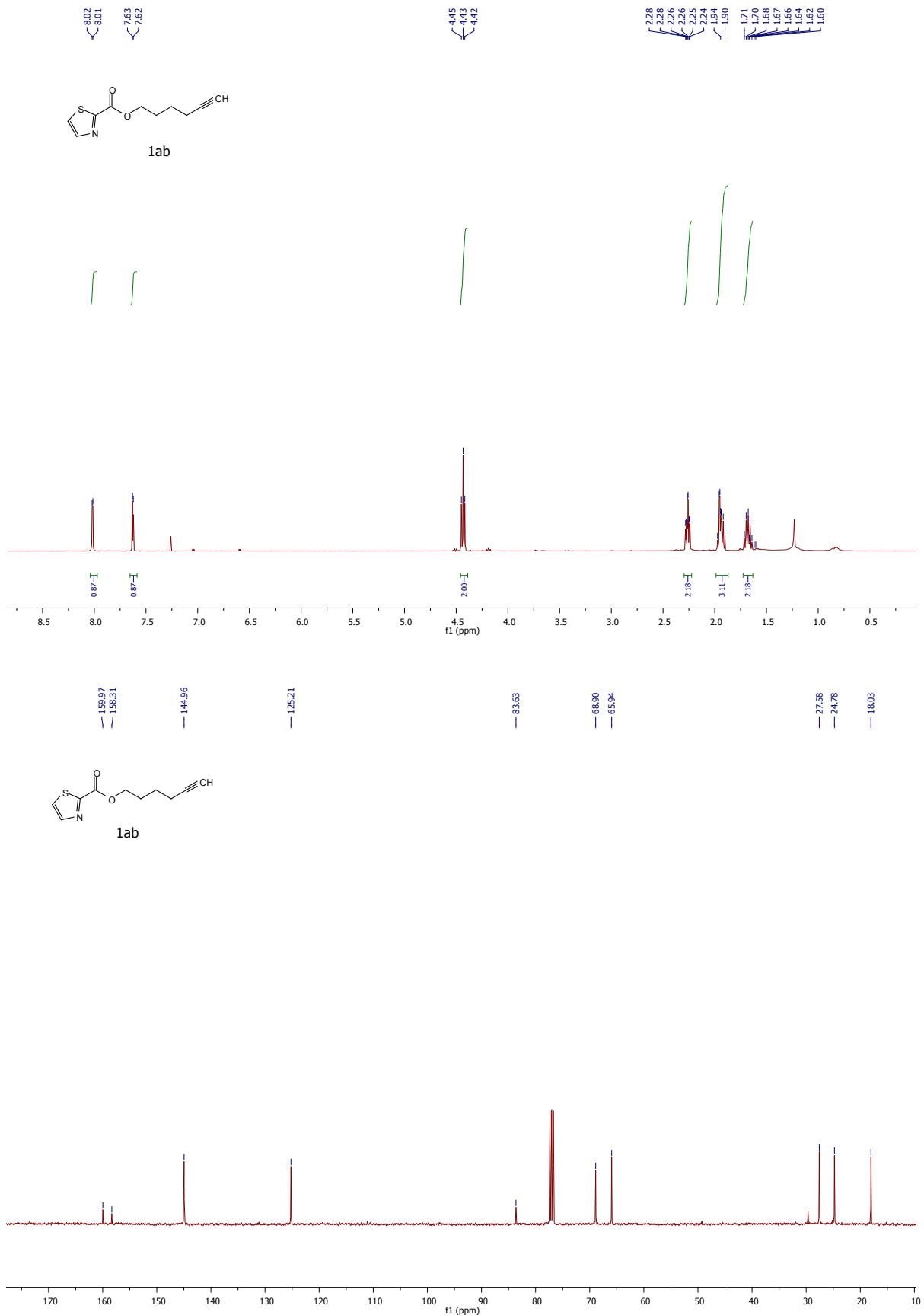


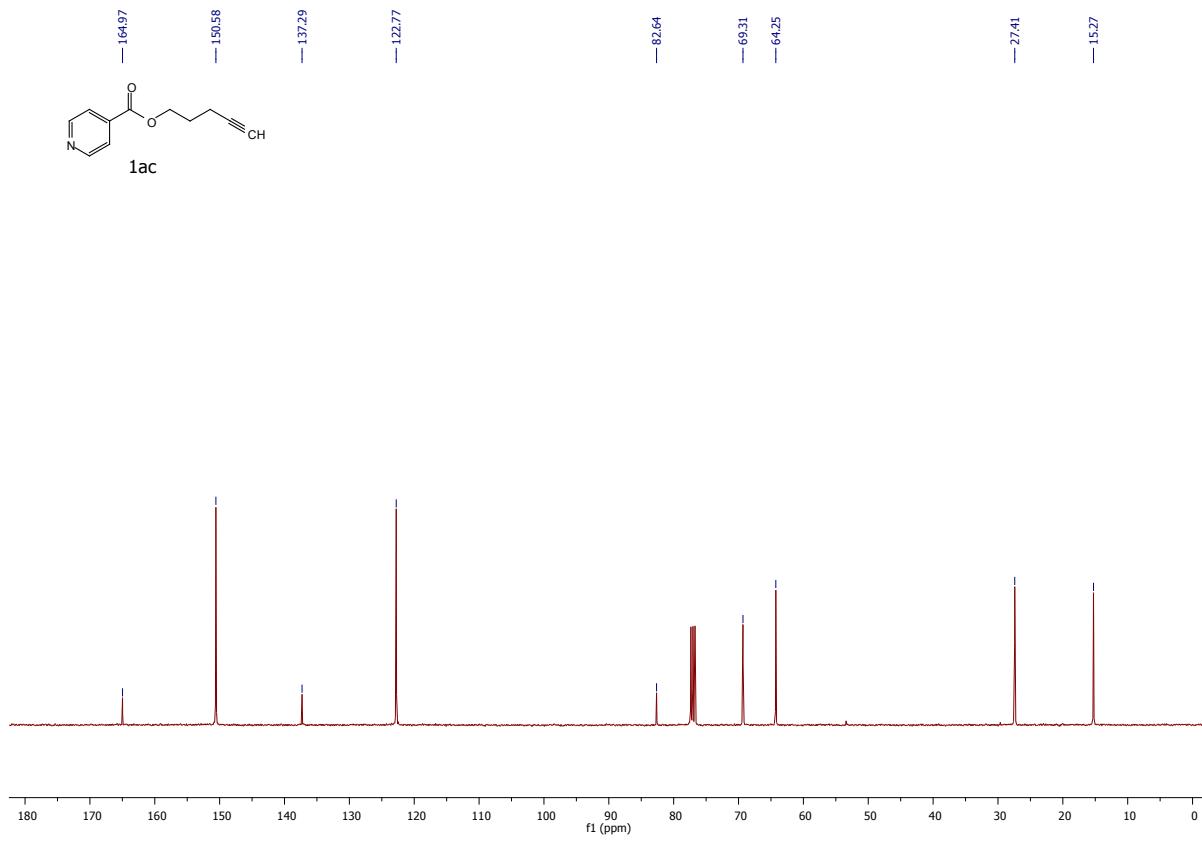
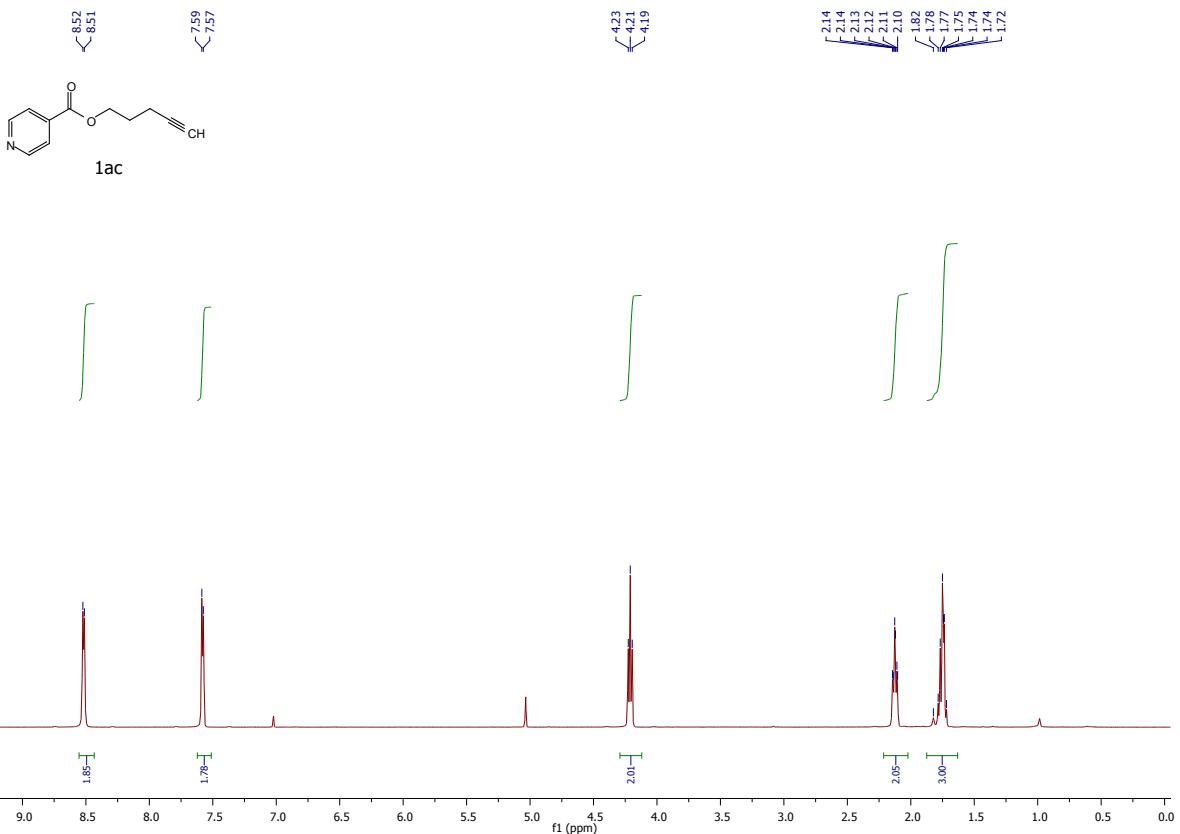


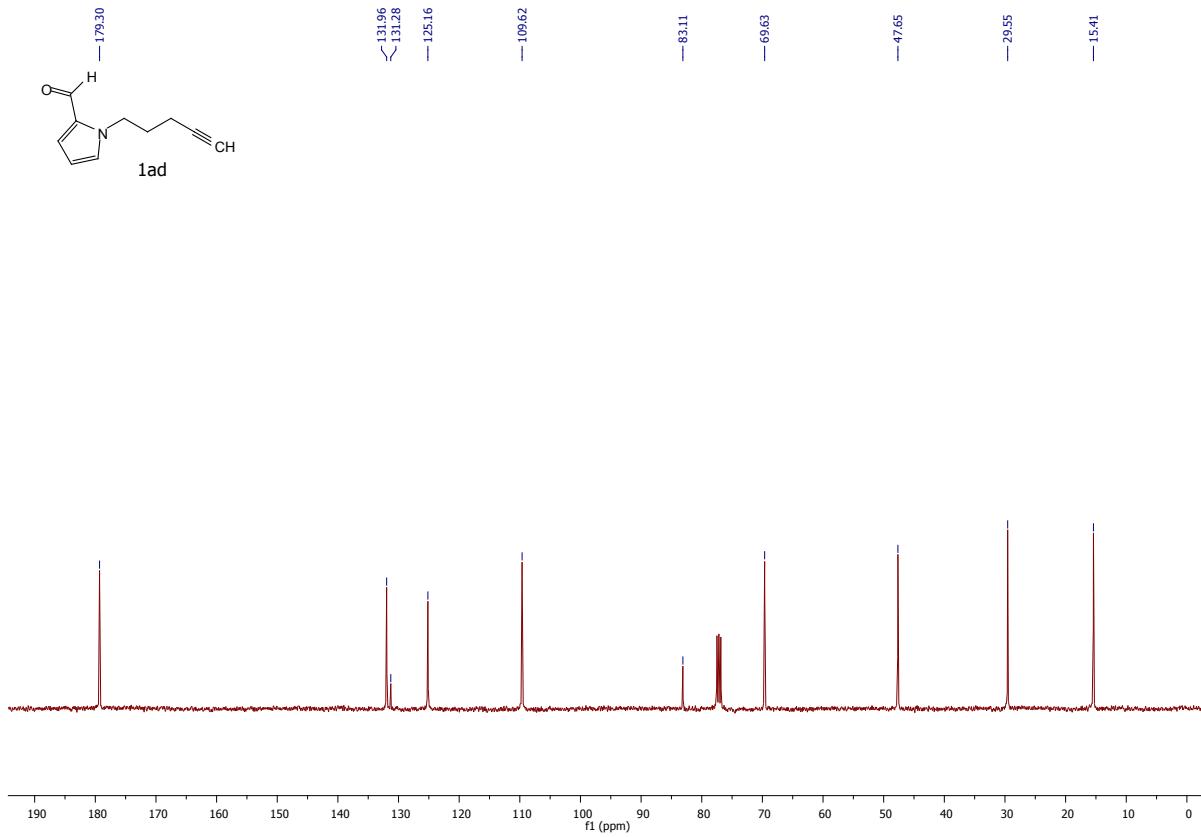
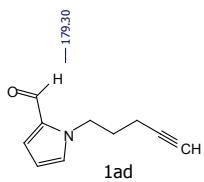
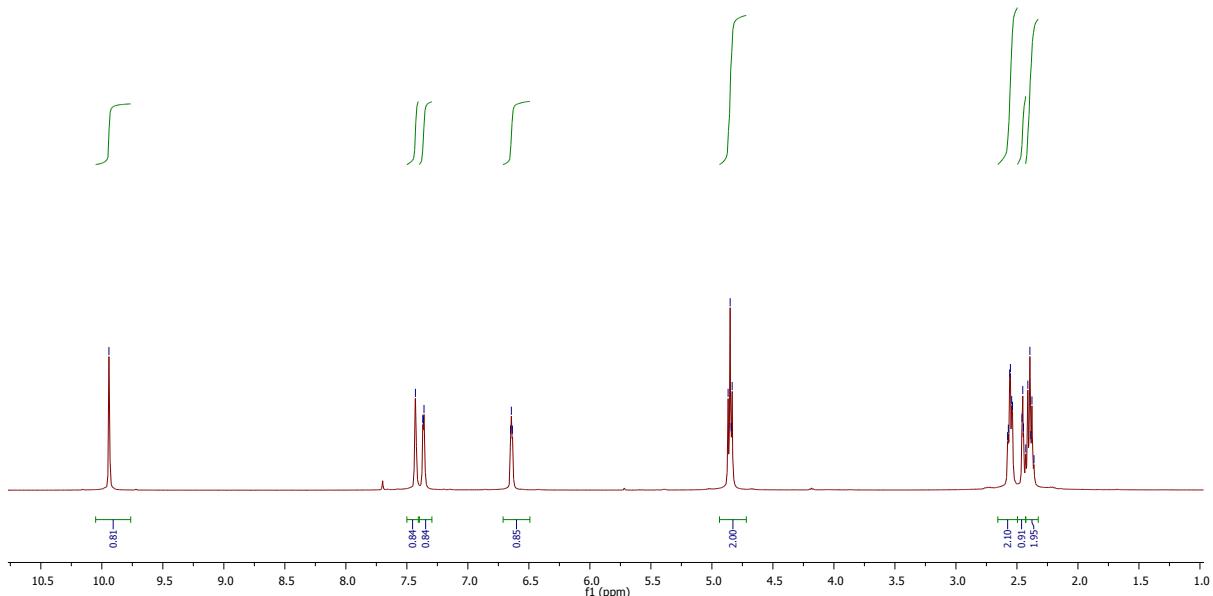
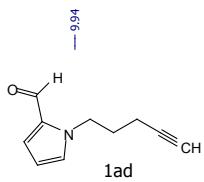


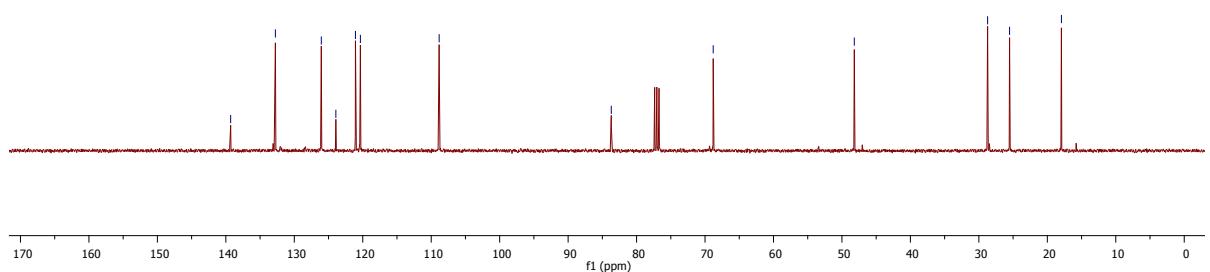
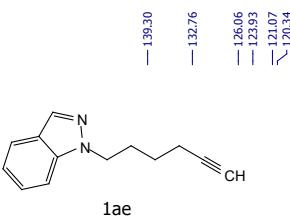
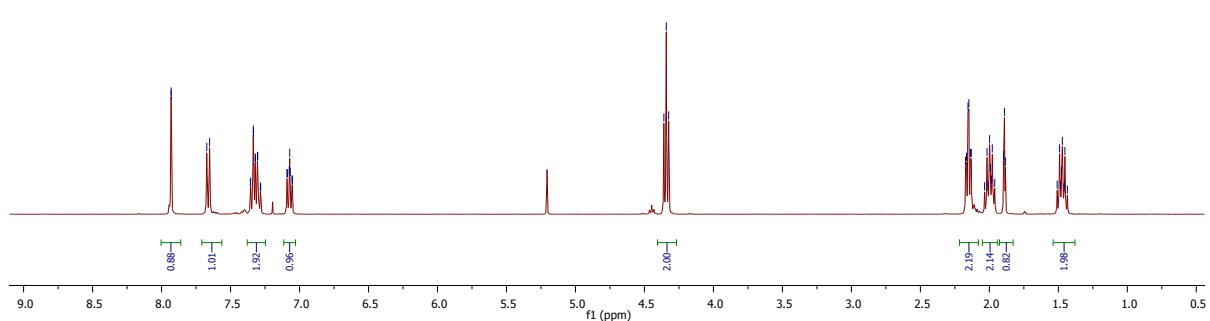
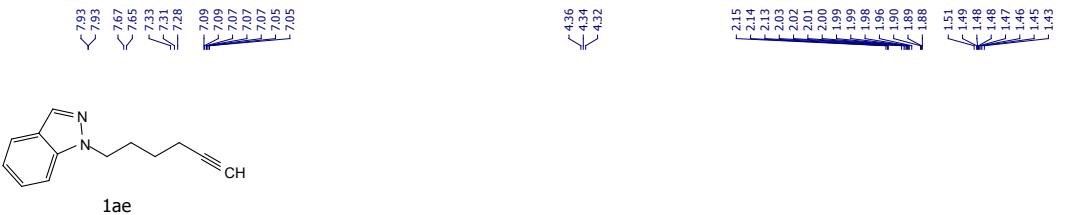


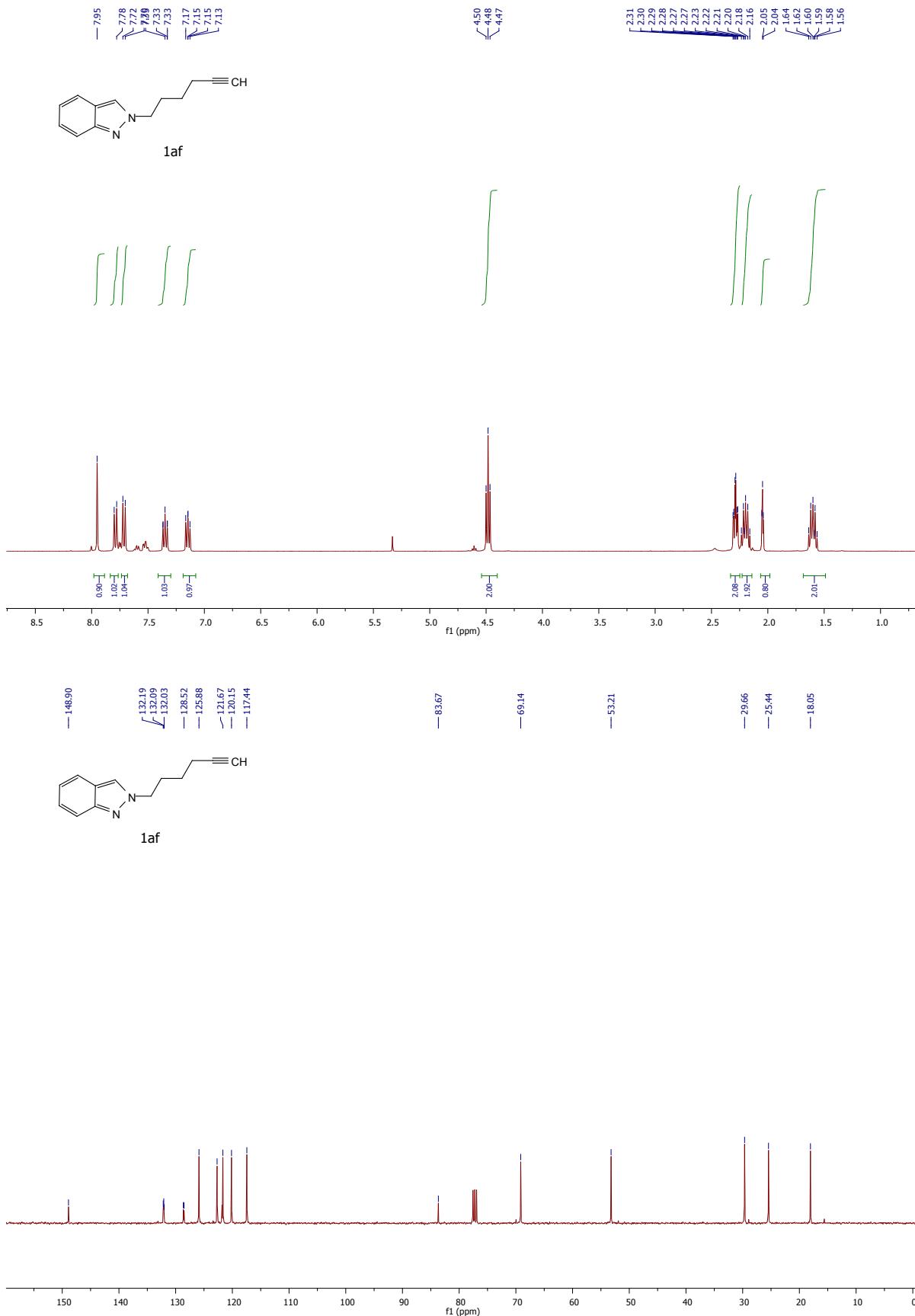


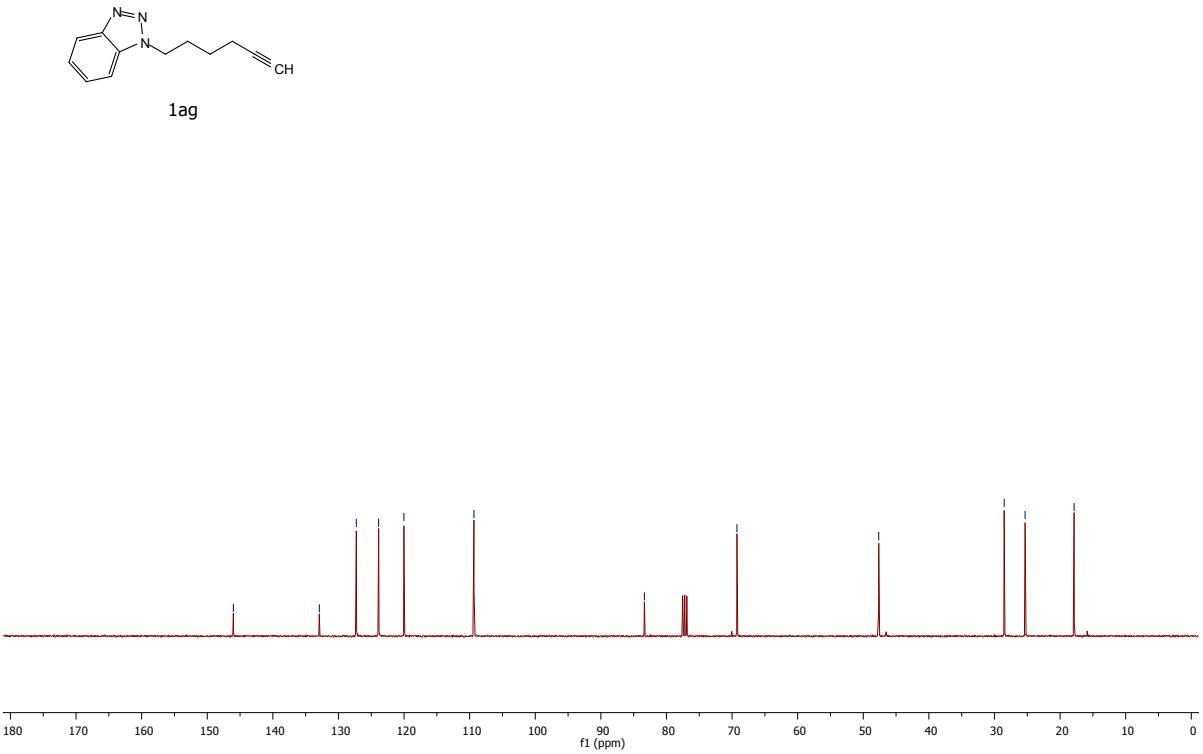
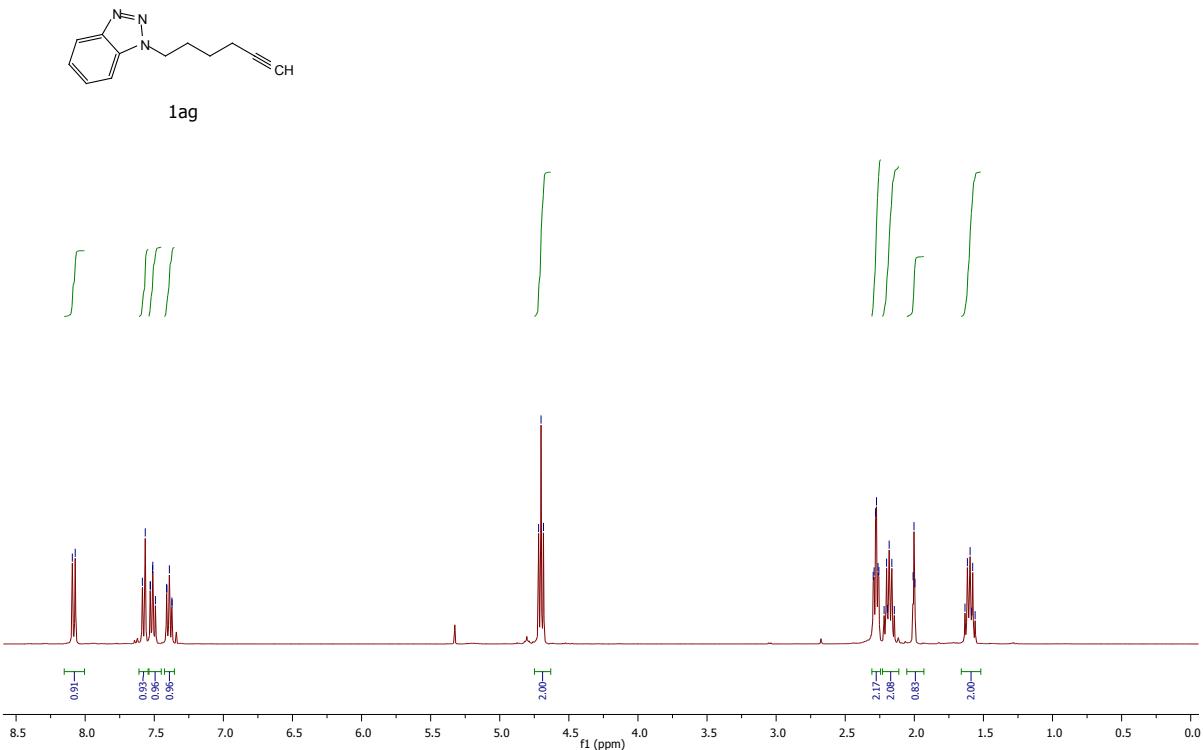


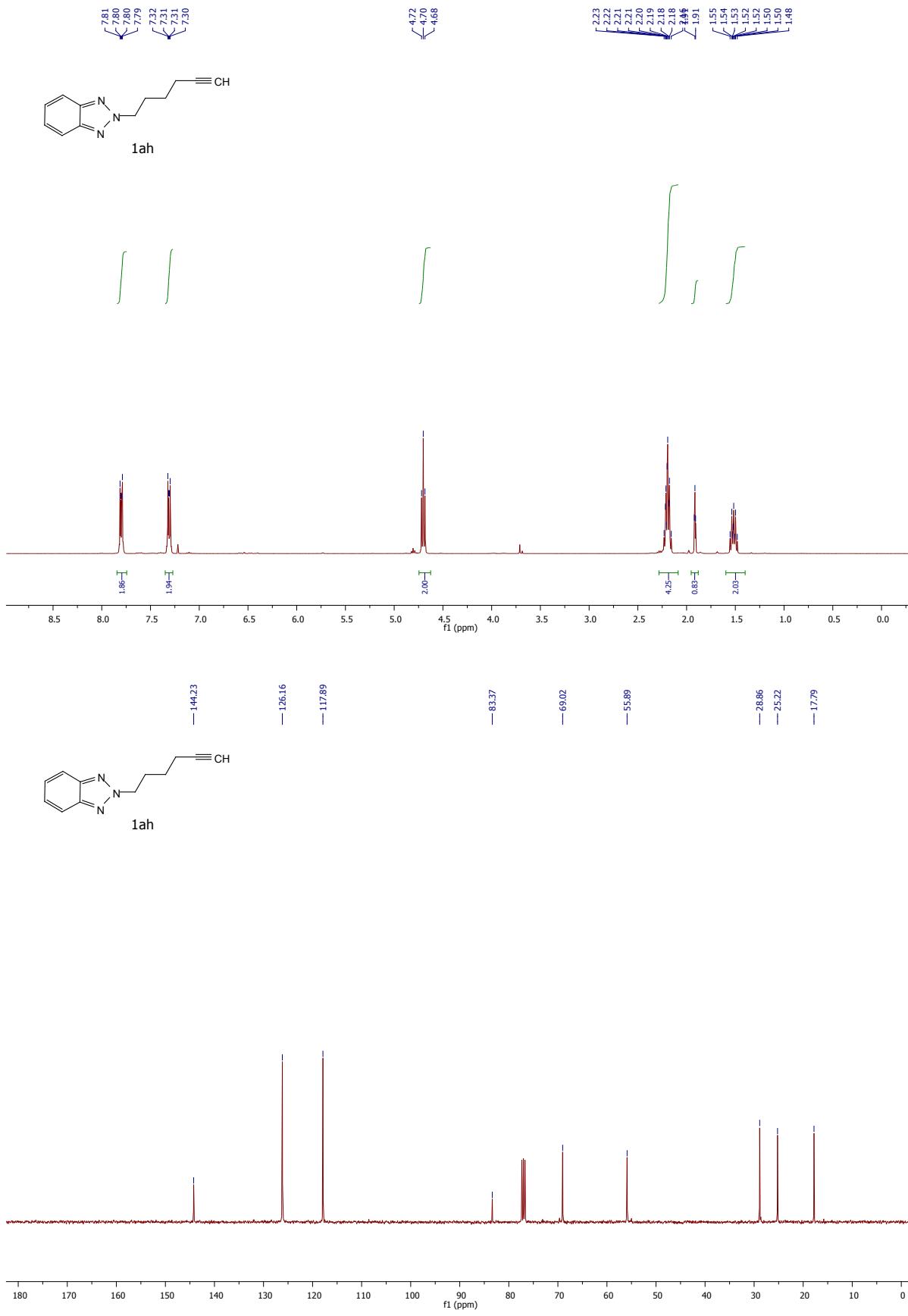


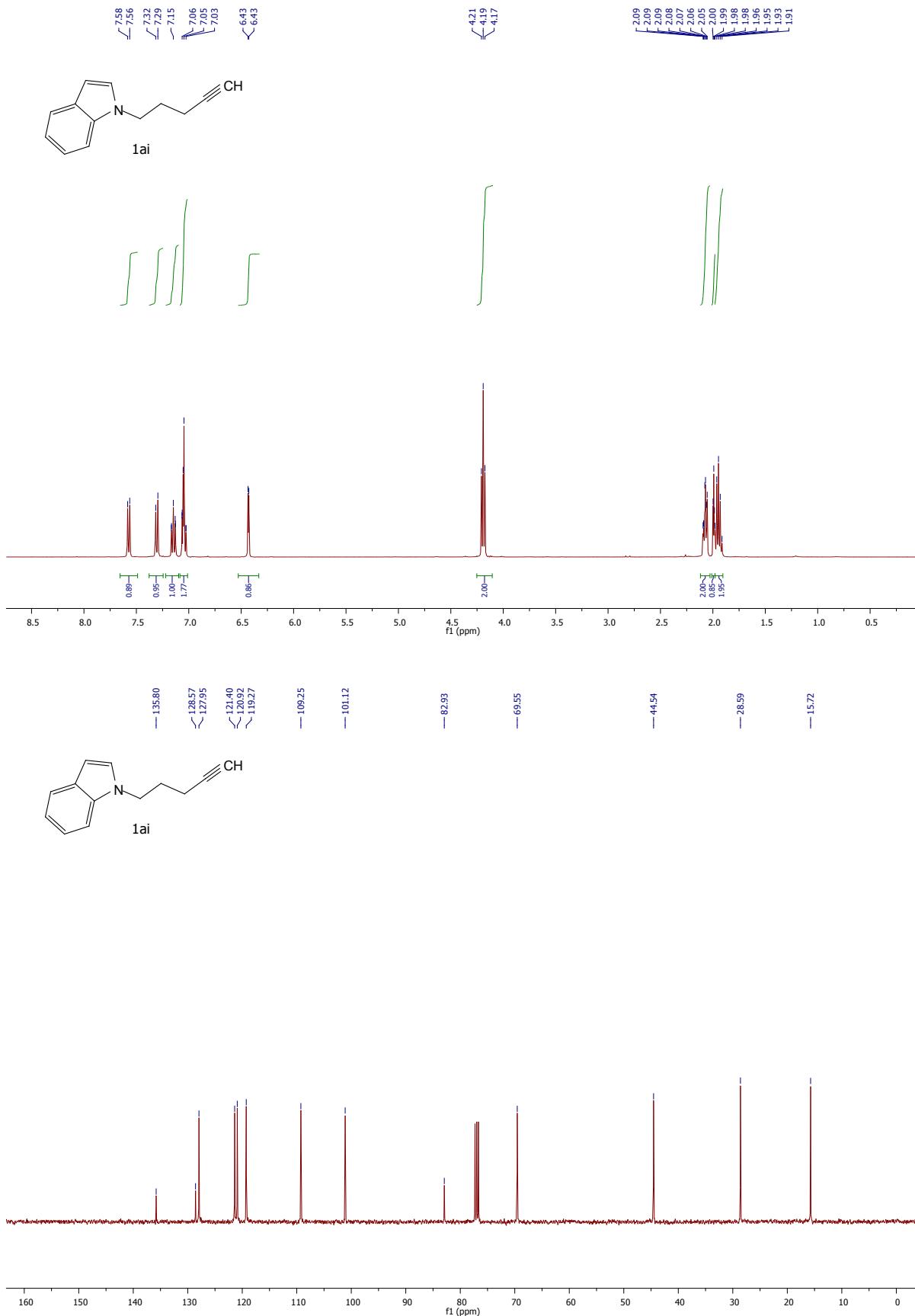


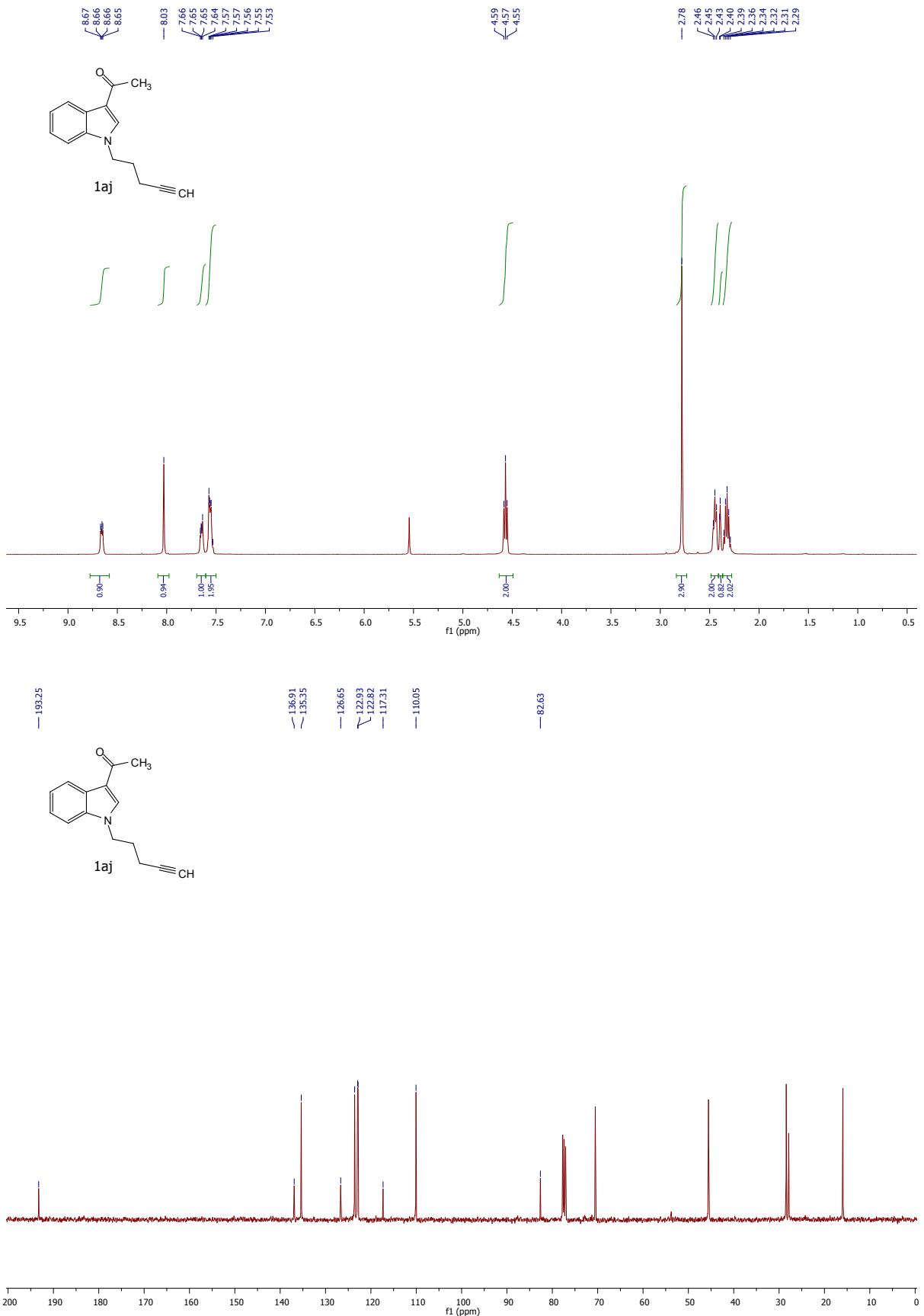




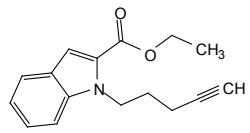








7.66
— 7.48
— 7.32
7.16
7.15
7.13

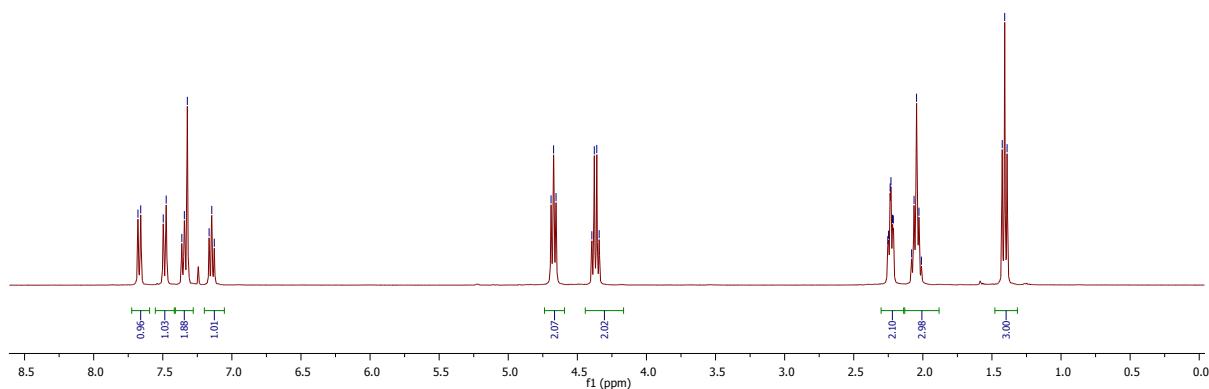


1ak

4.67
— 4.65
— 4.40
4.38
4.36
4.34

2.25
2.25
2.24
2.23
2.22
2.21
2.06
2.05
2.03
2.01
2.01

1.43
— 1.41
1.39



161.95
— 139.17

127.36
— 125.90
— 125.01
— 122.59
— 120.60

110.72
< 110.39

83.52

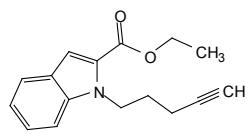
69.06

60.53

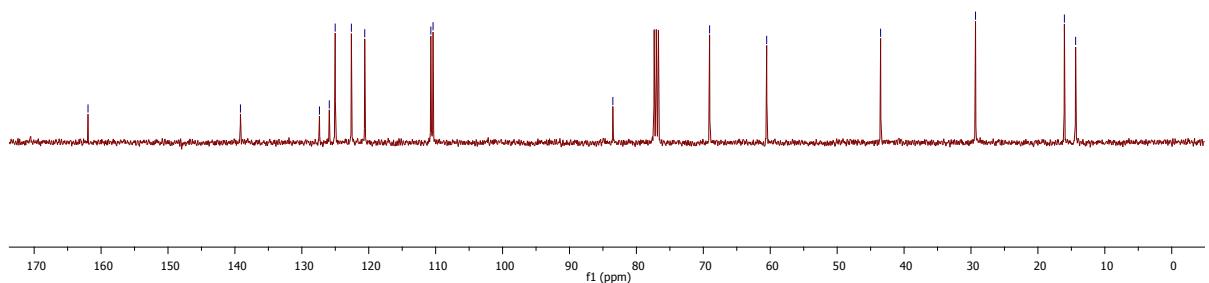
43.51

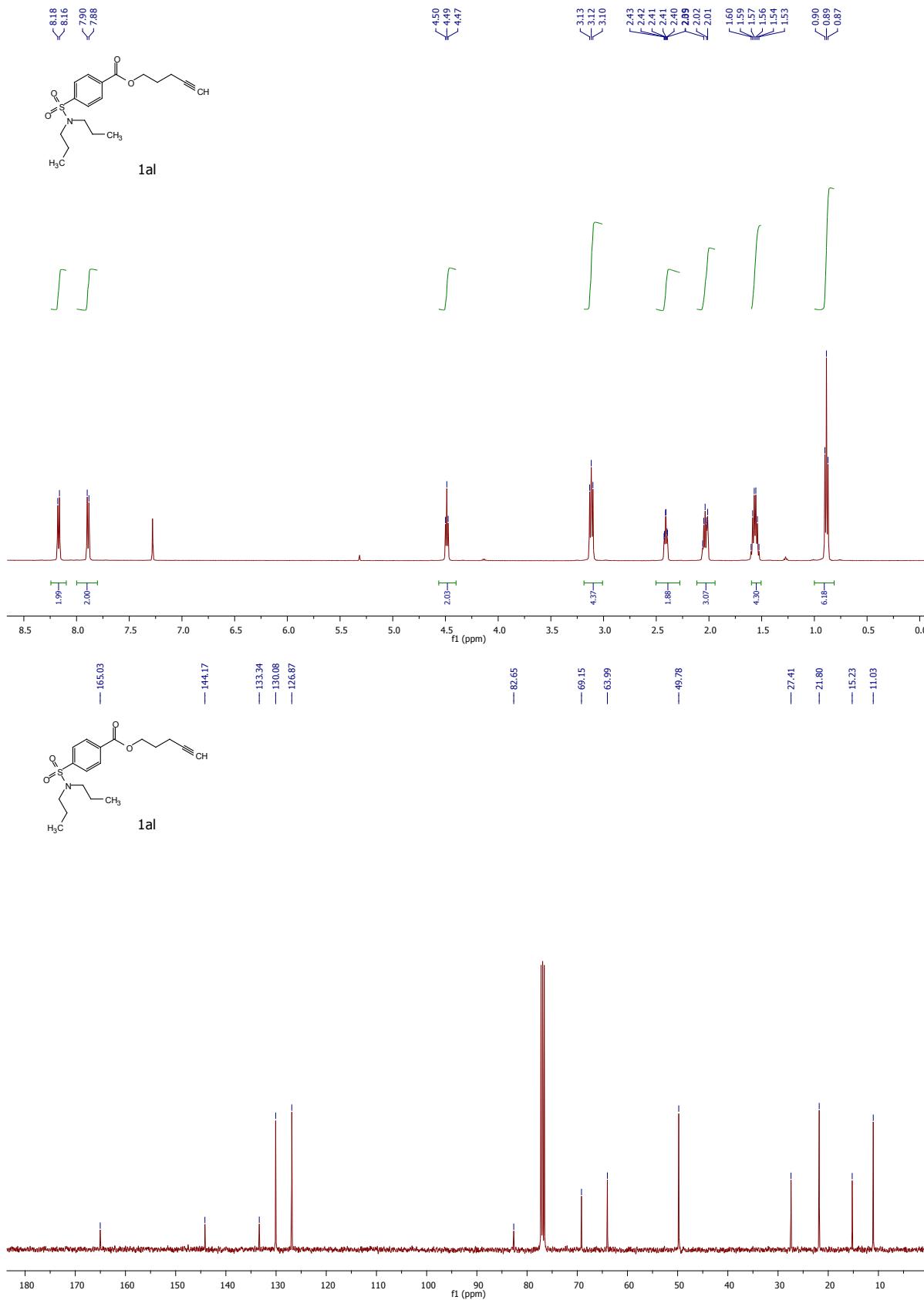
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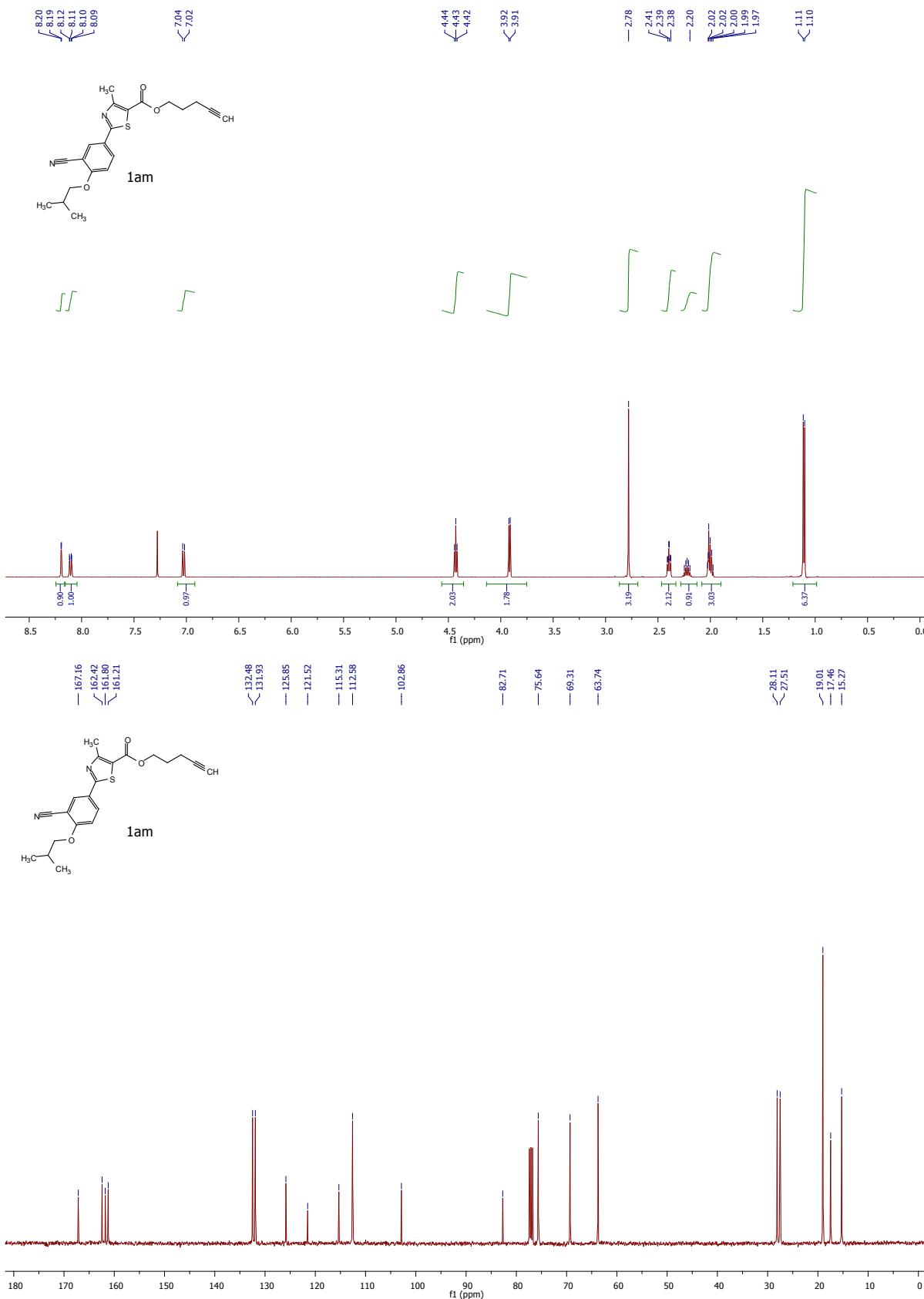
16.04
— 14.36

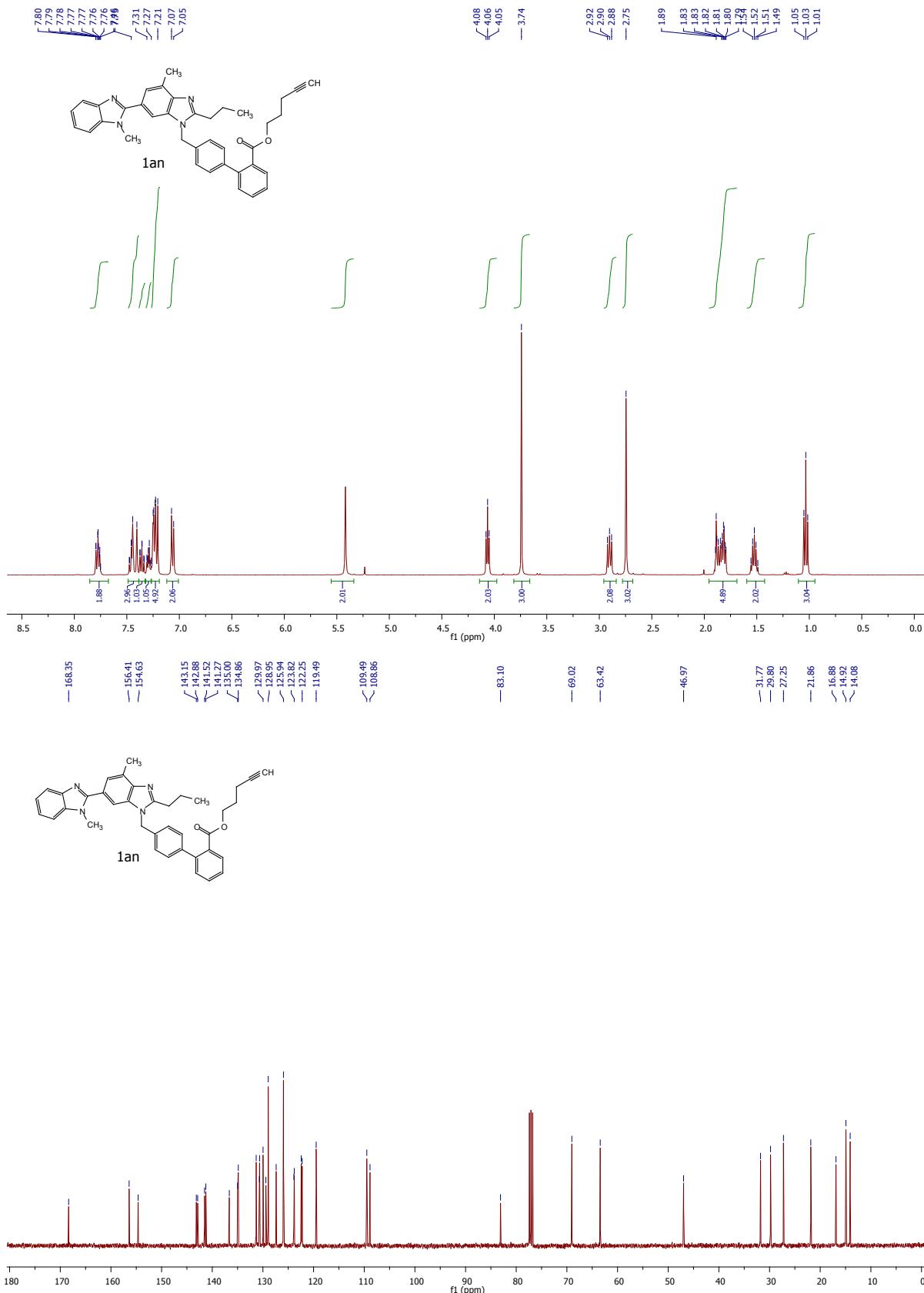


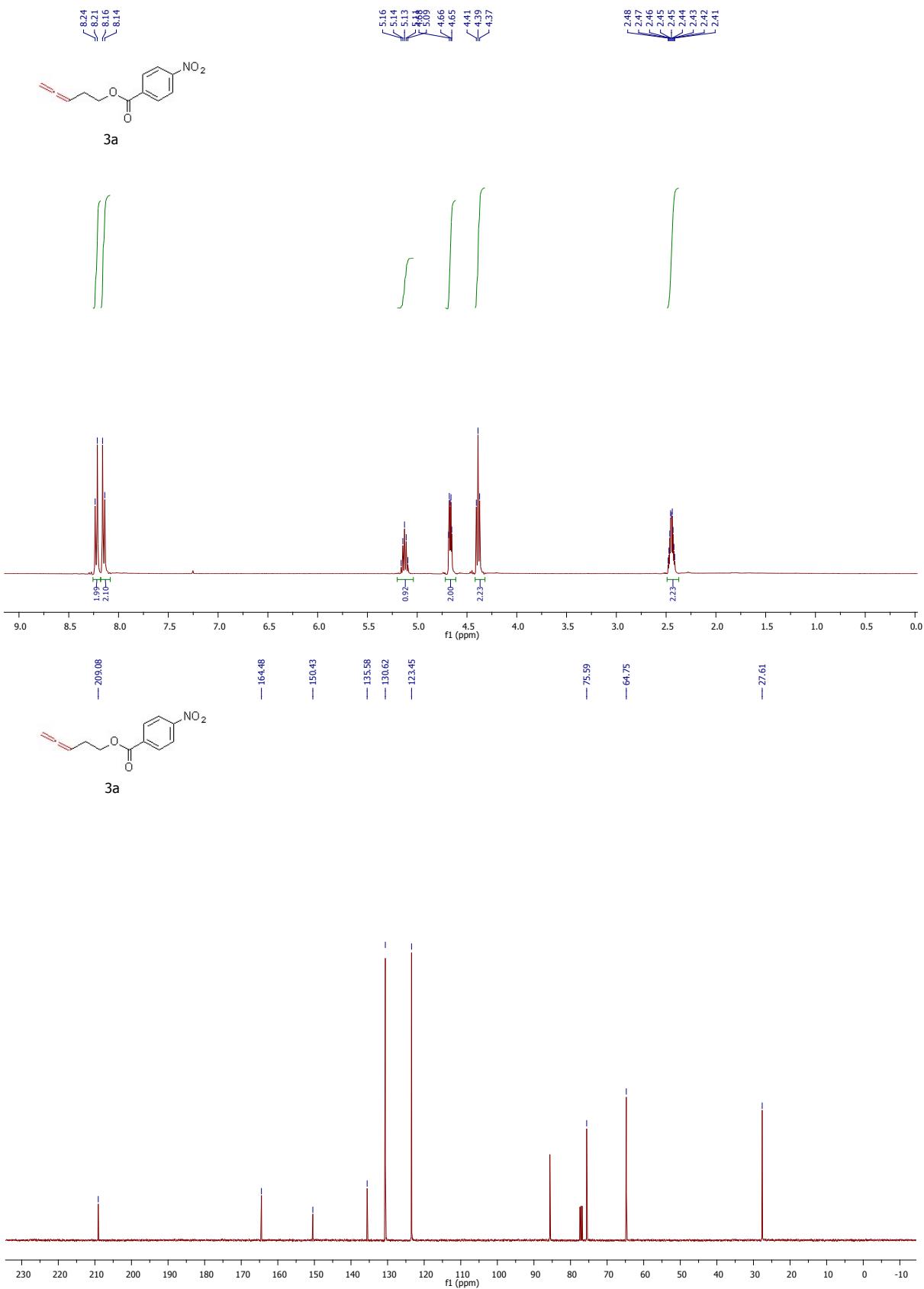
1ak

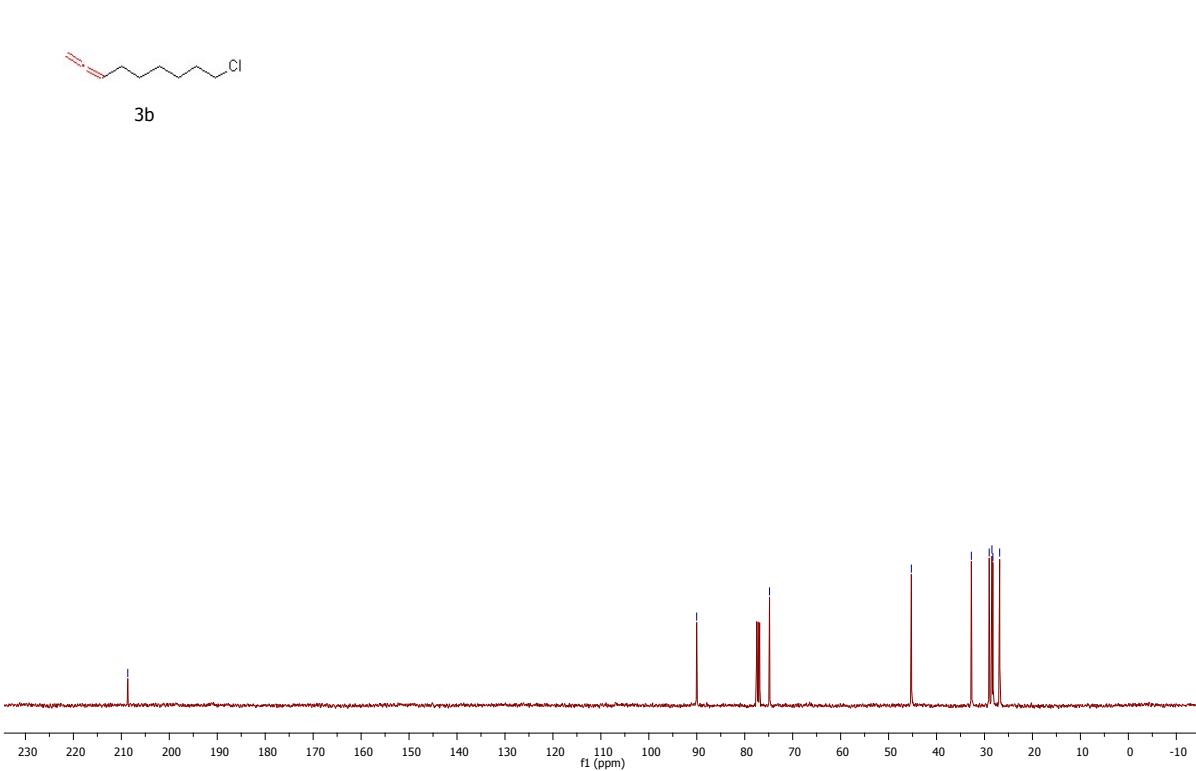
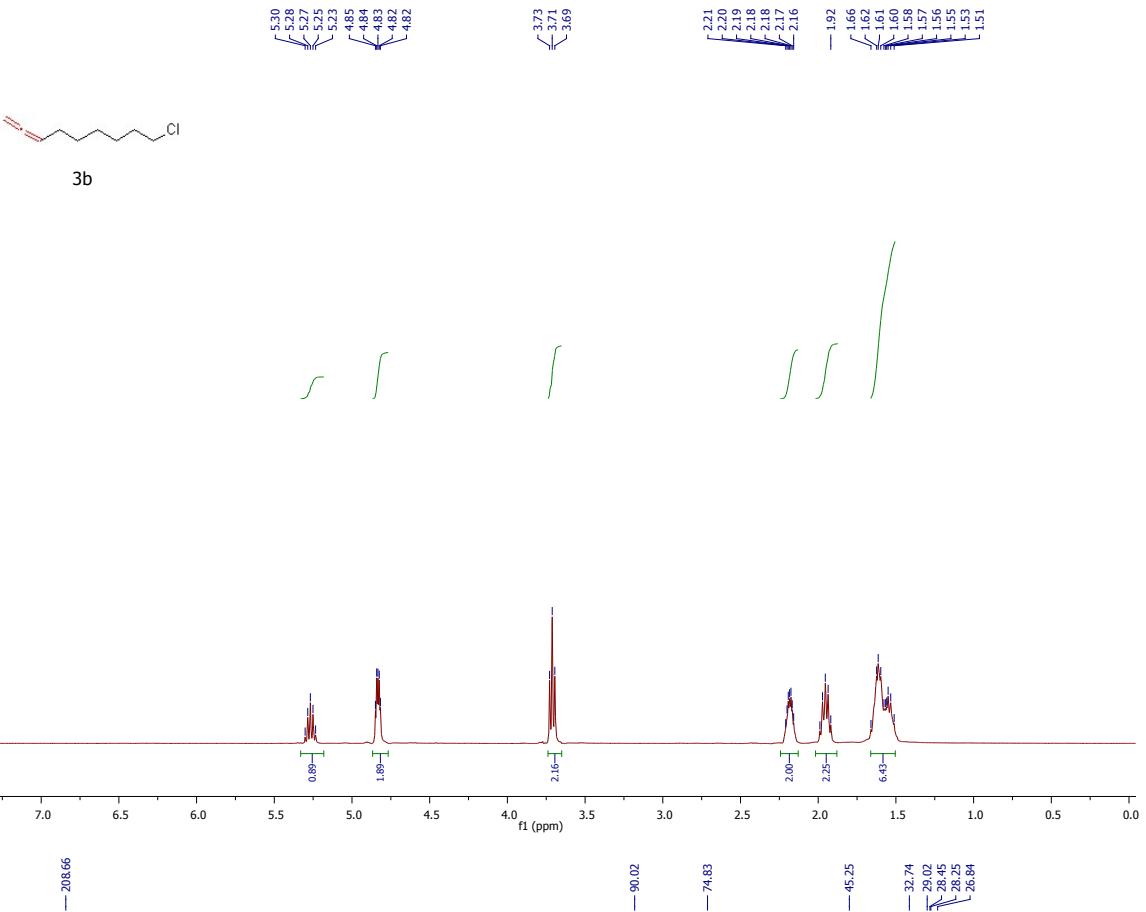










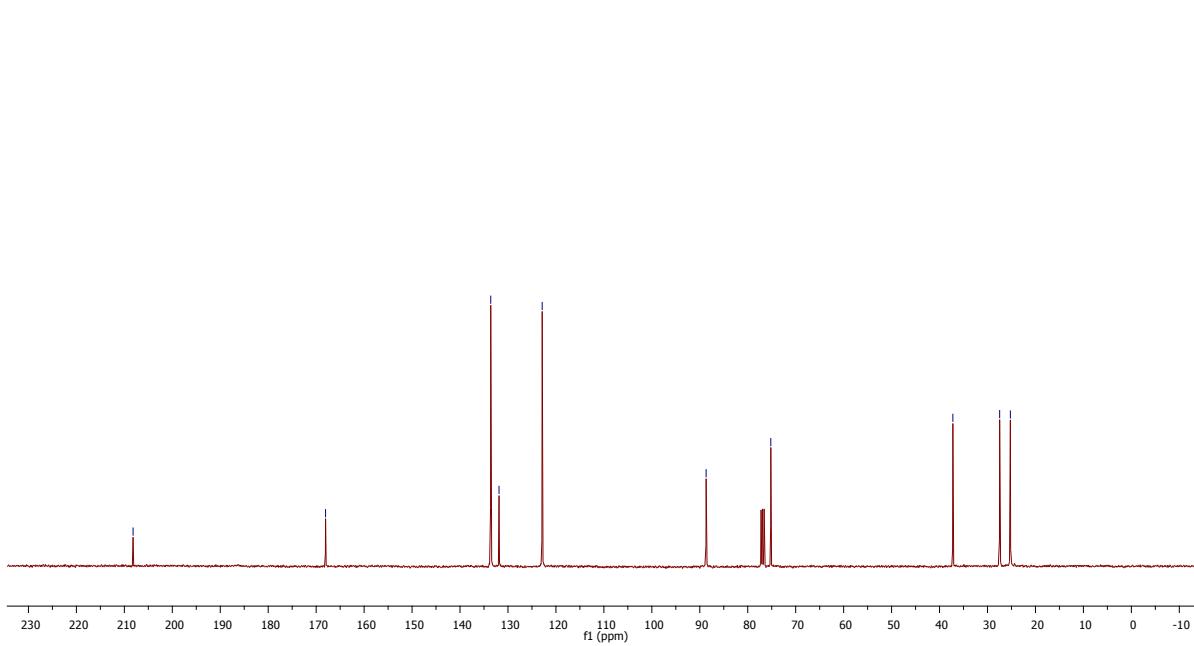
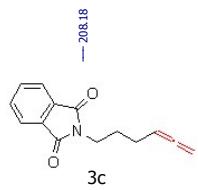
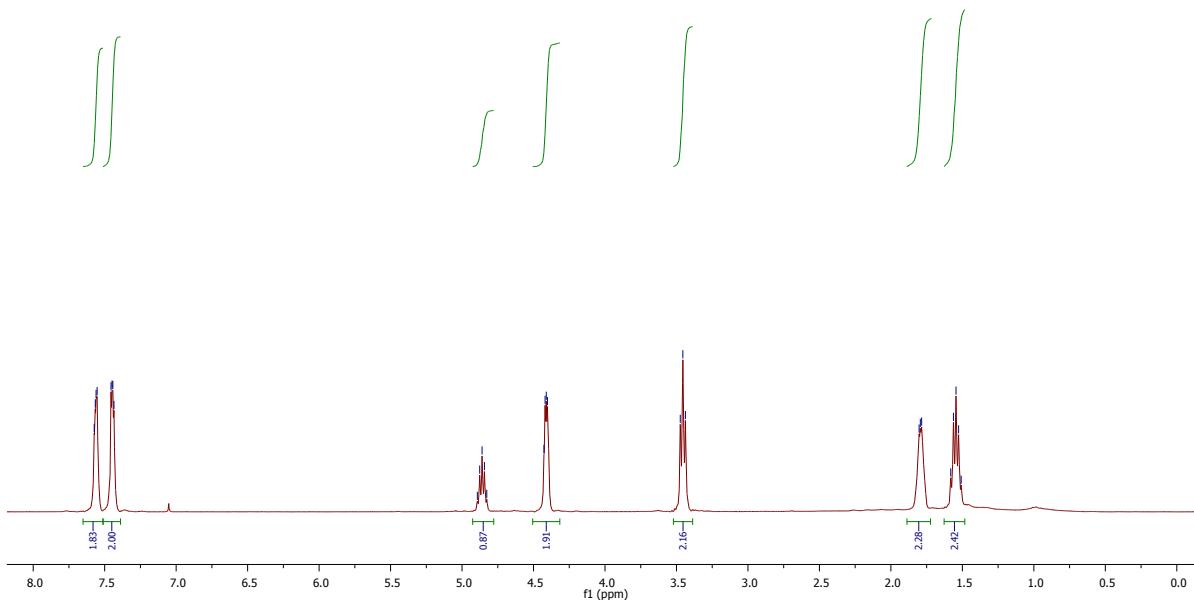
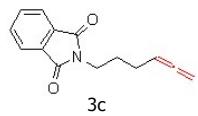


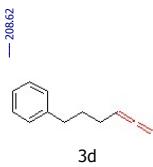
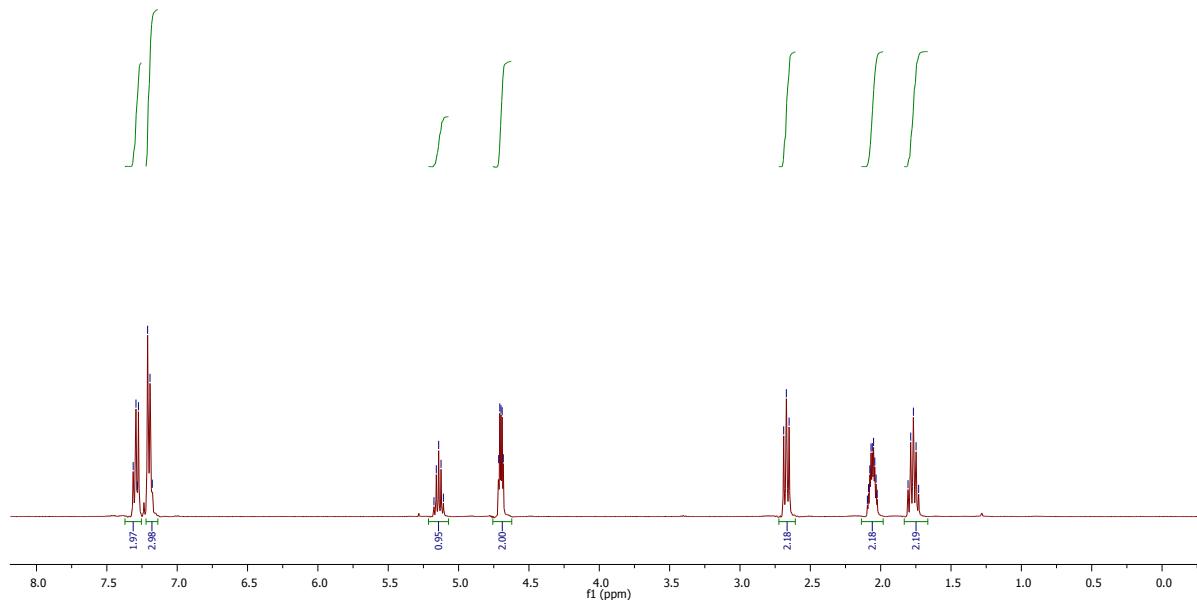
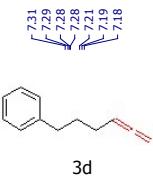
7.57
7.56
7.55
7.46
7.45
7.44
7.43

4.89
4.88
4.86
4.84
4.83
4.43
4.42
4.41
4.40

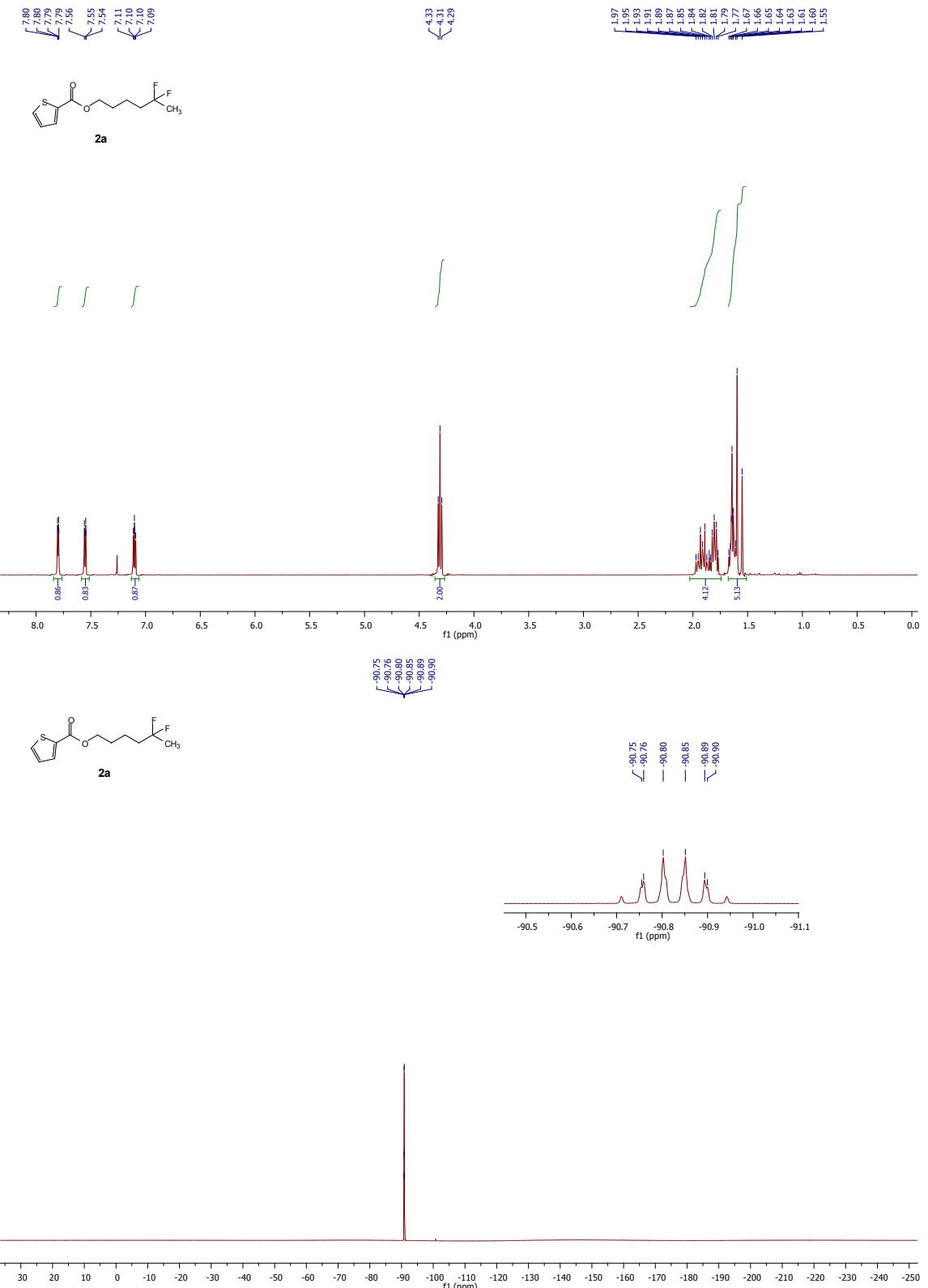
3.47
3.46
3.44

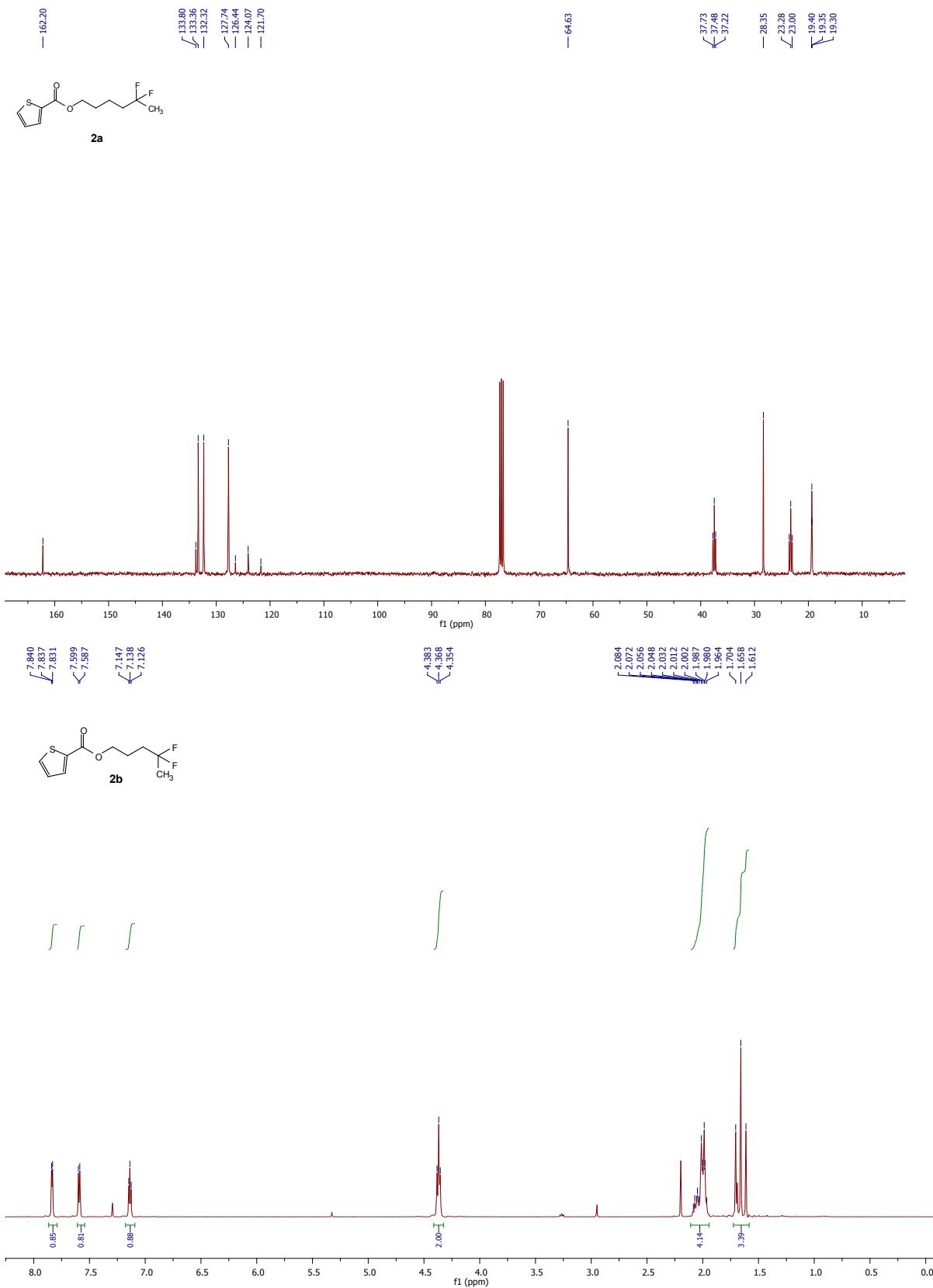
1.80
1.79
1.78
1.58
1.56
1.54
1.53
1.51

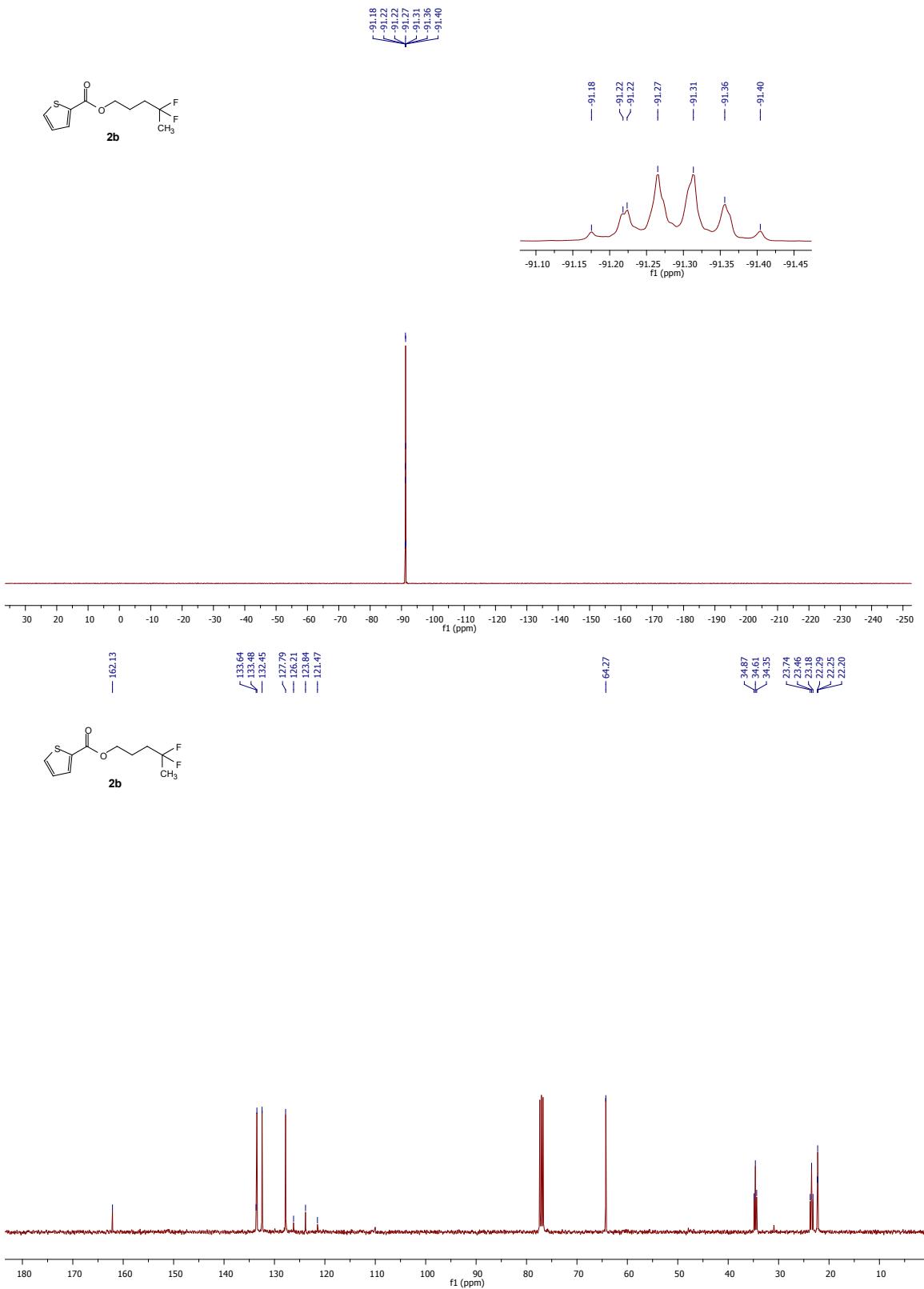


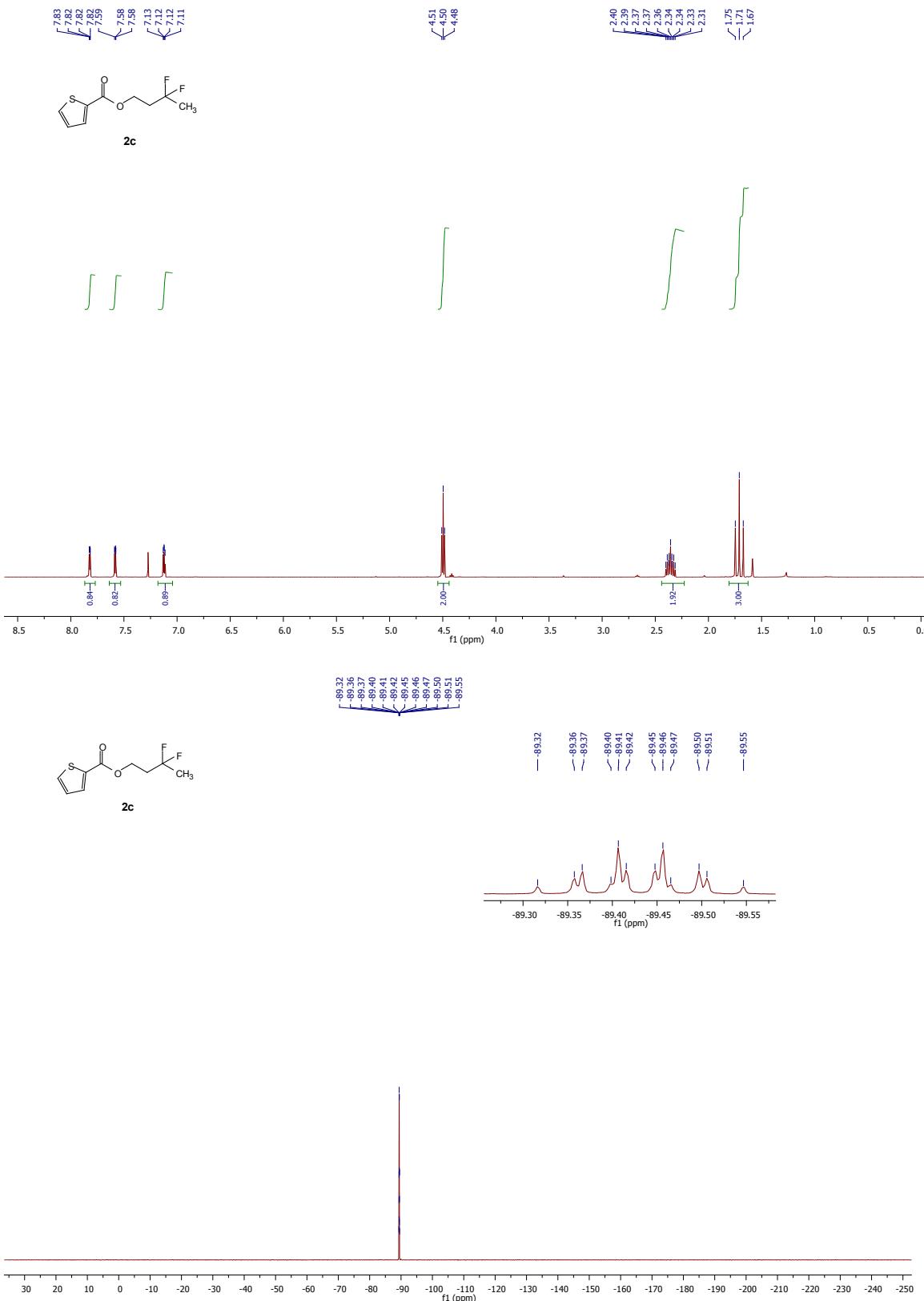


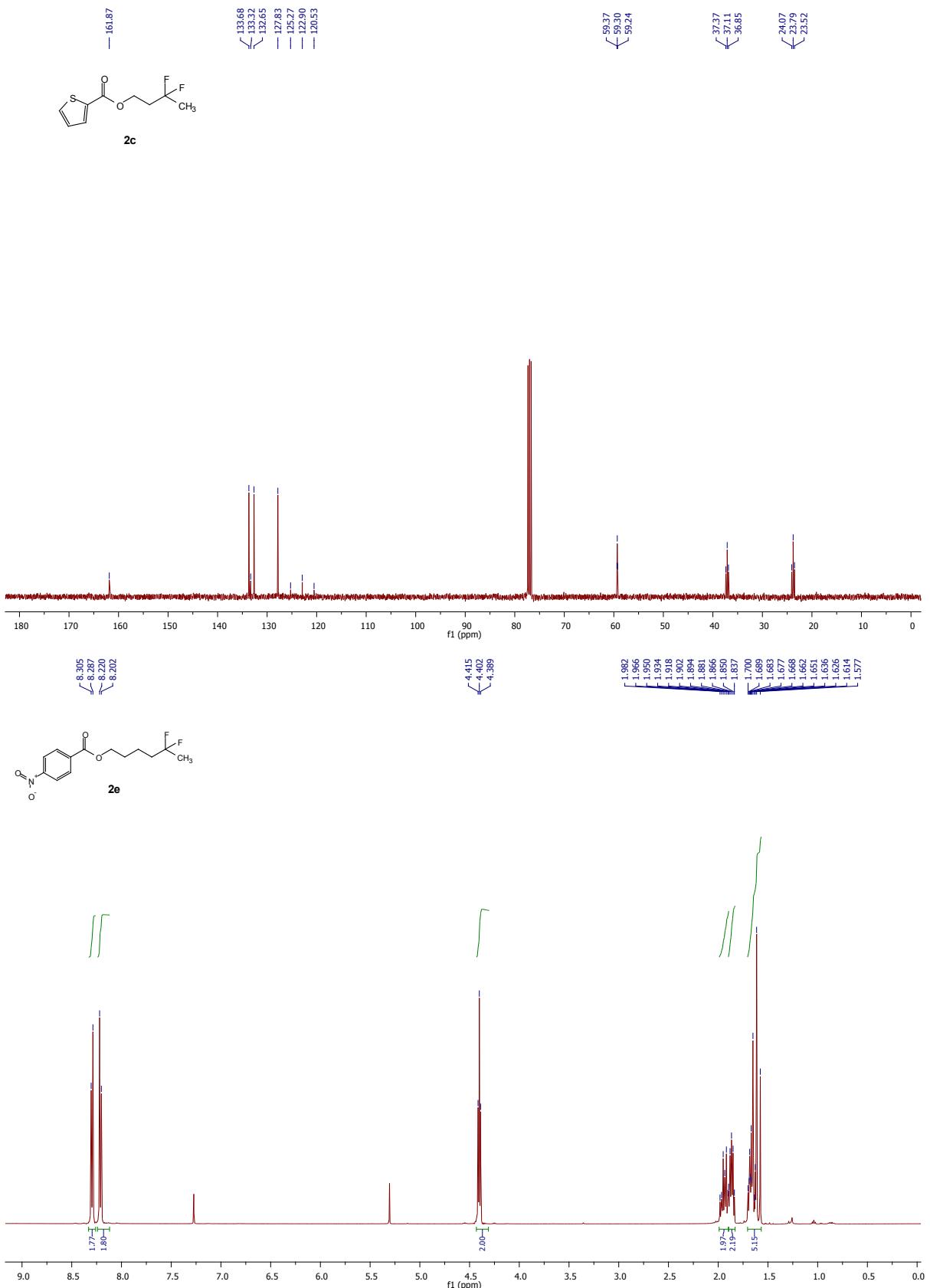
4.2 Products NMR spectra.

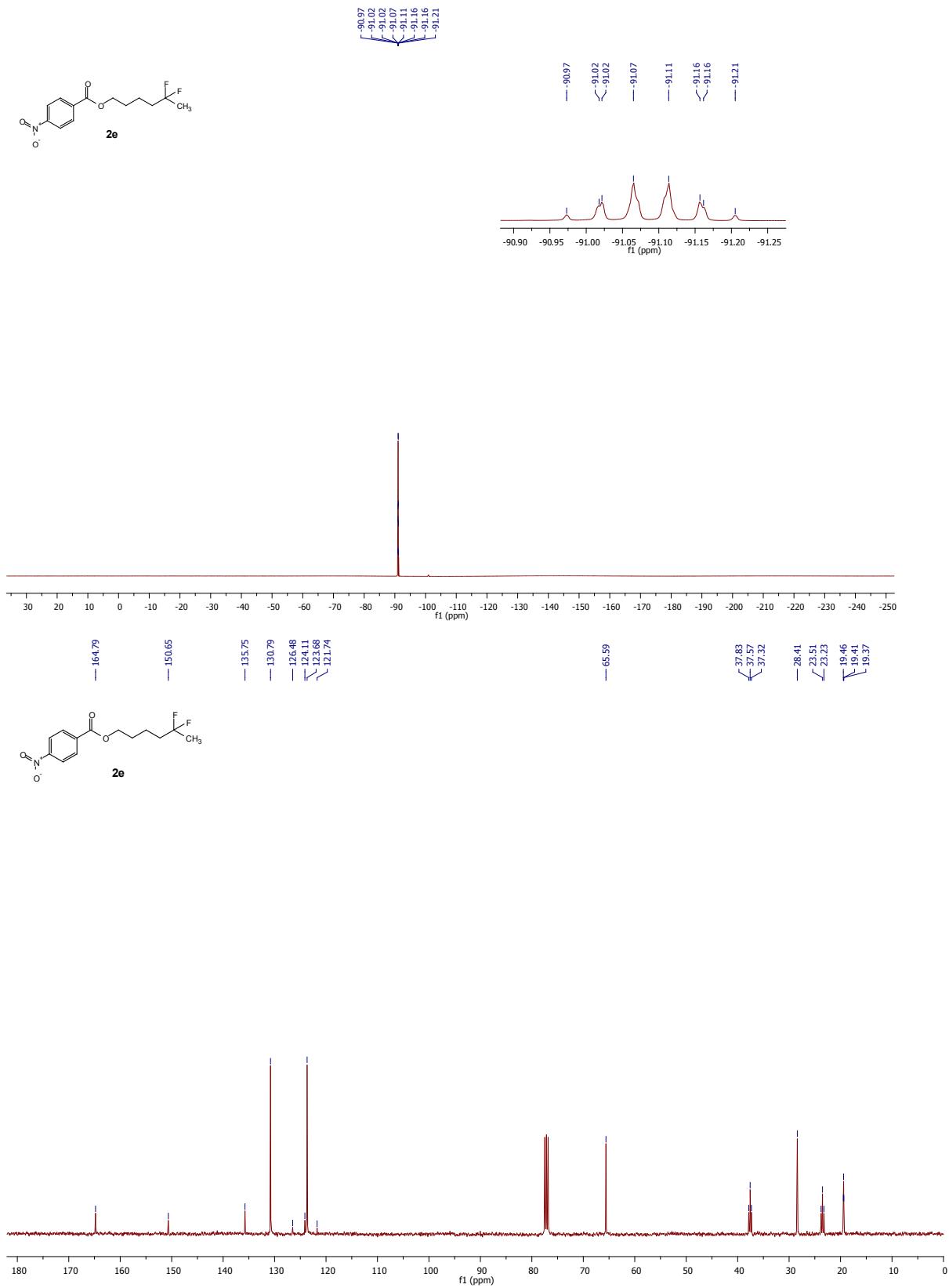


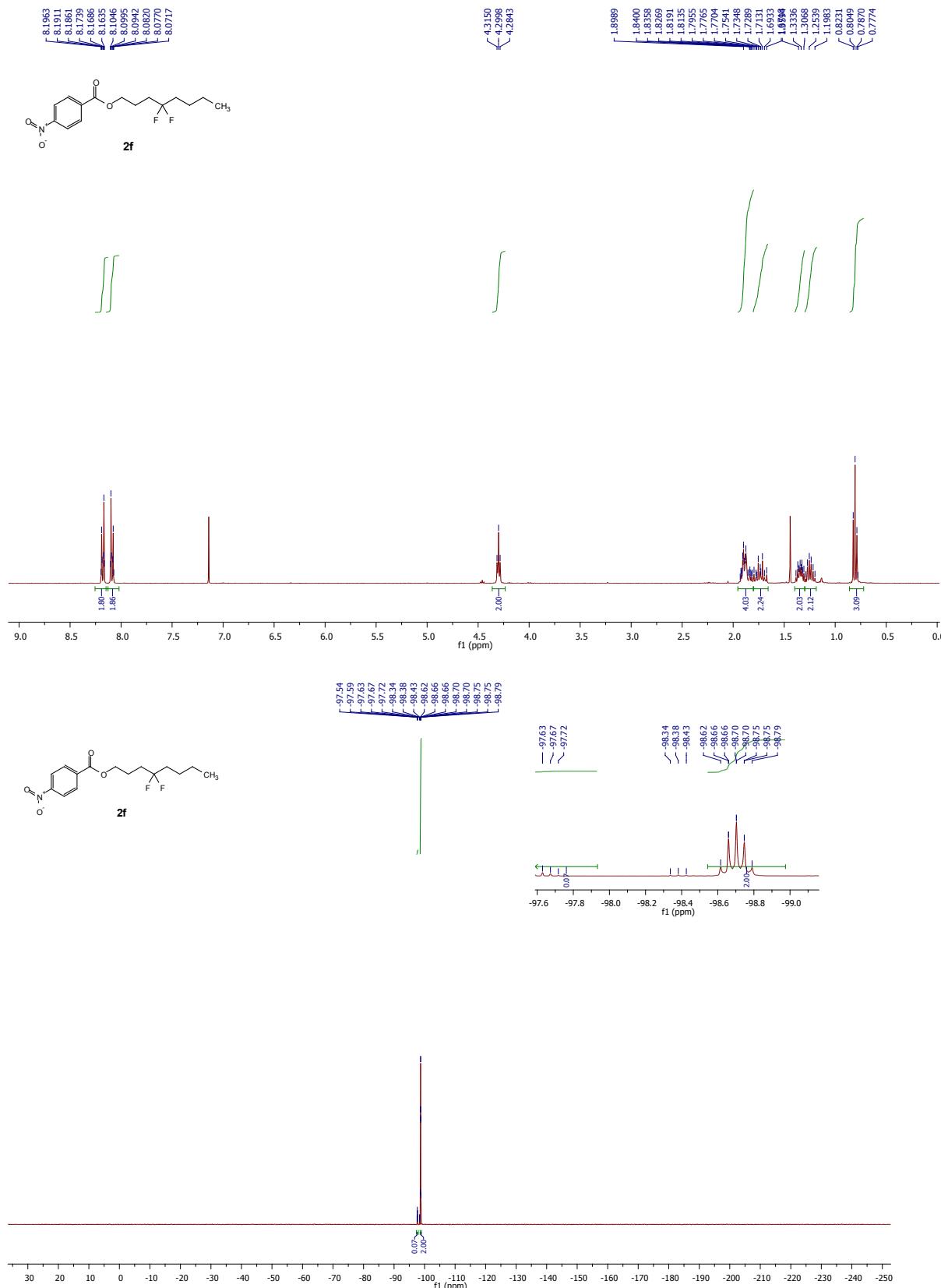


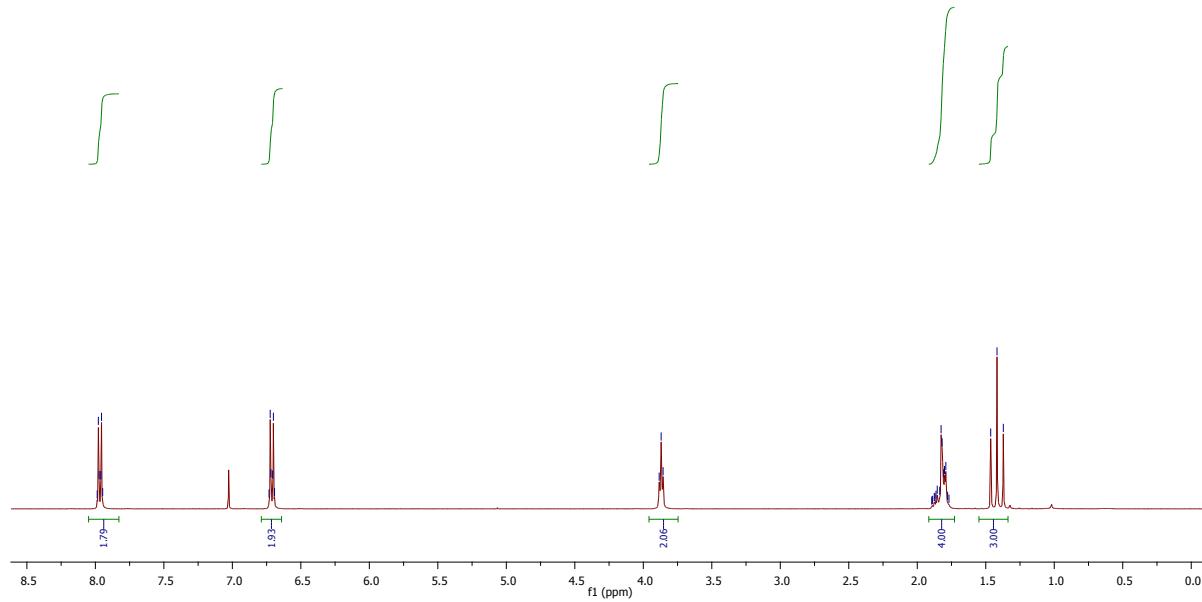
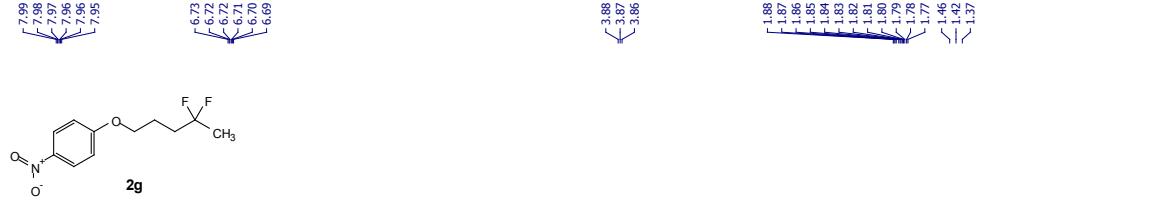
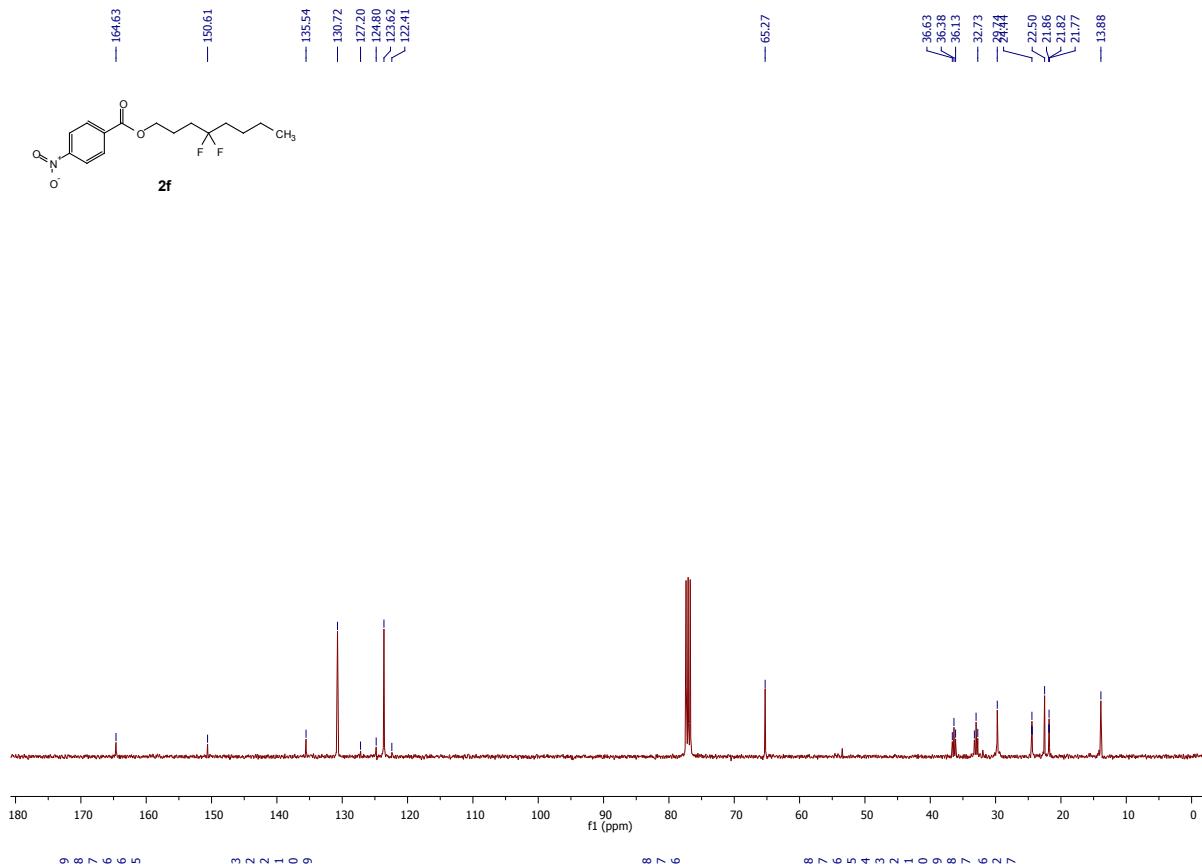


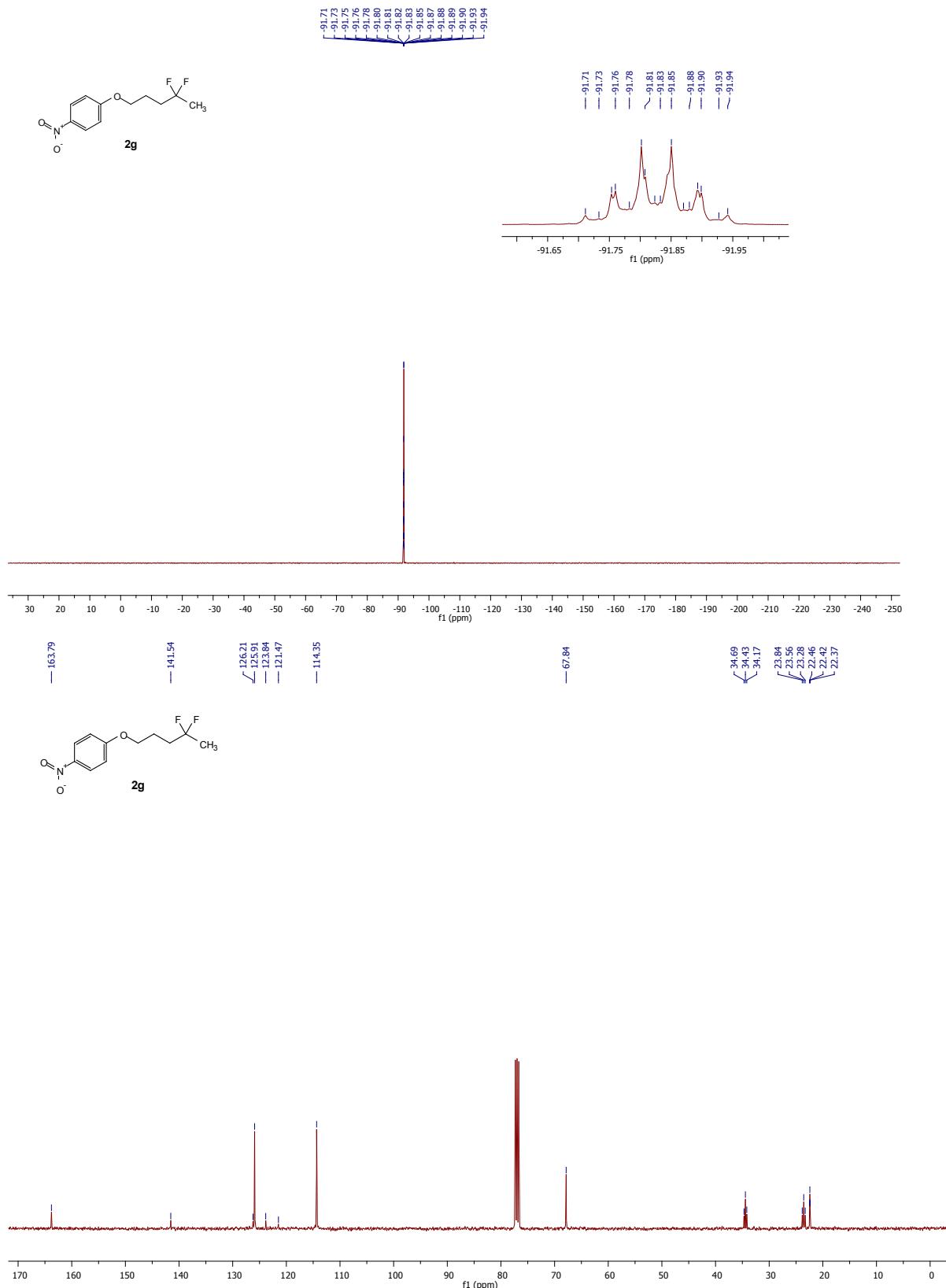


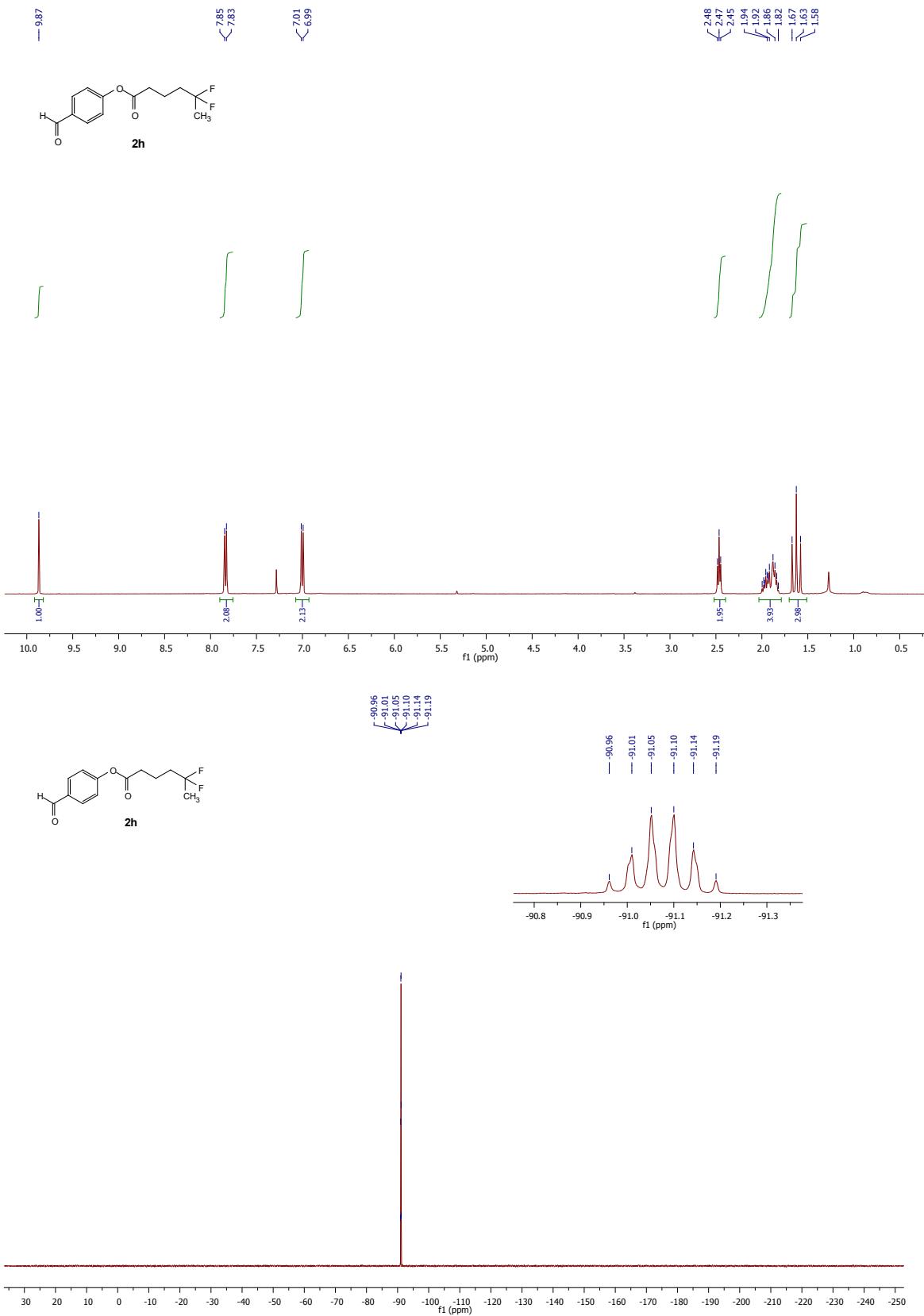


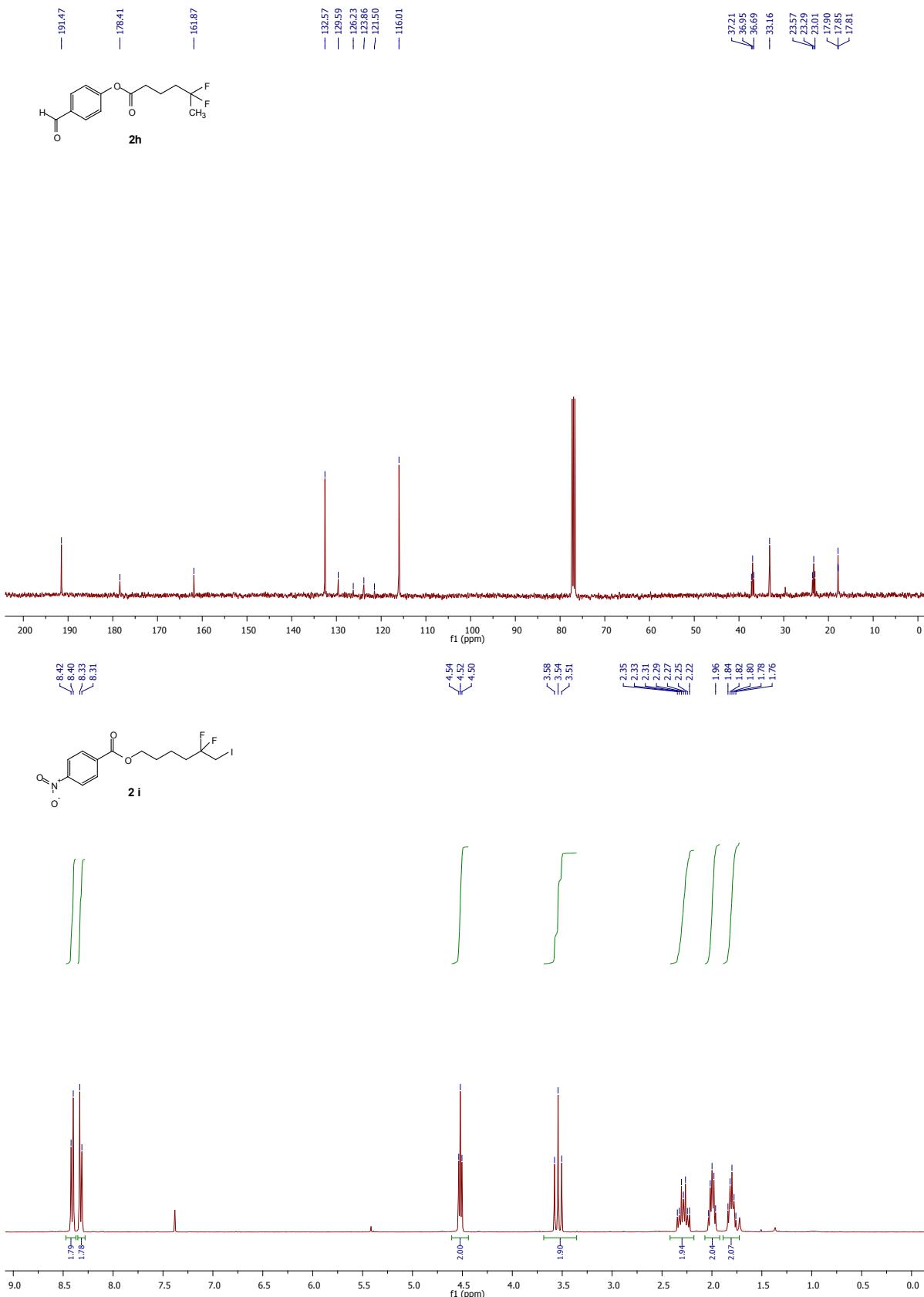


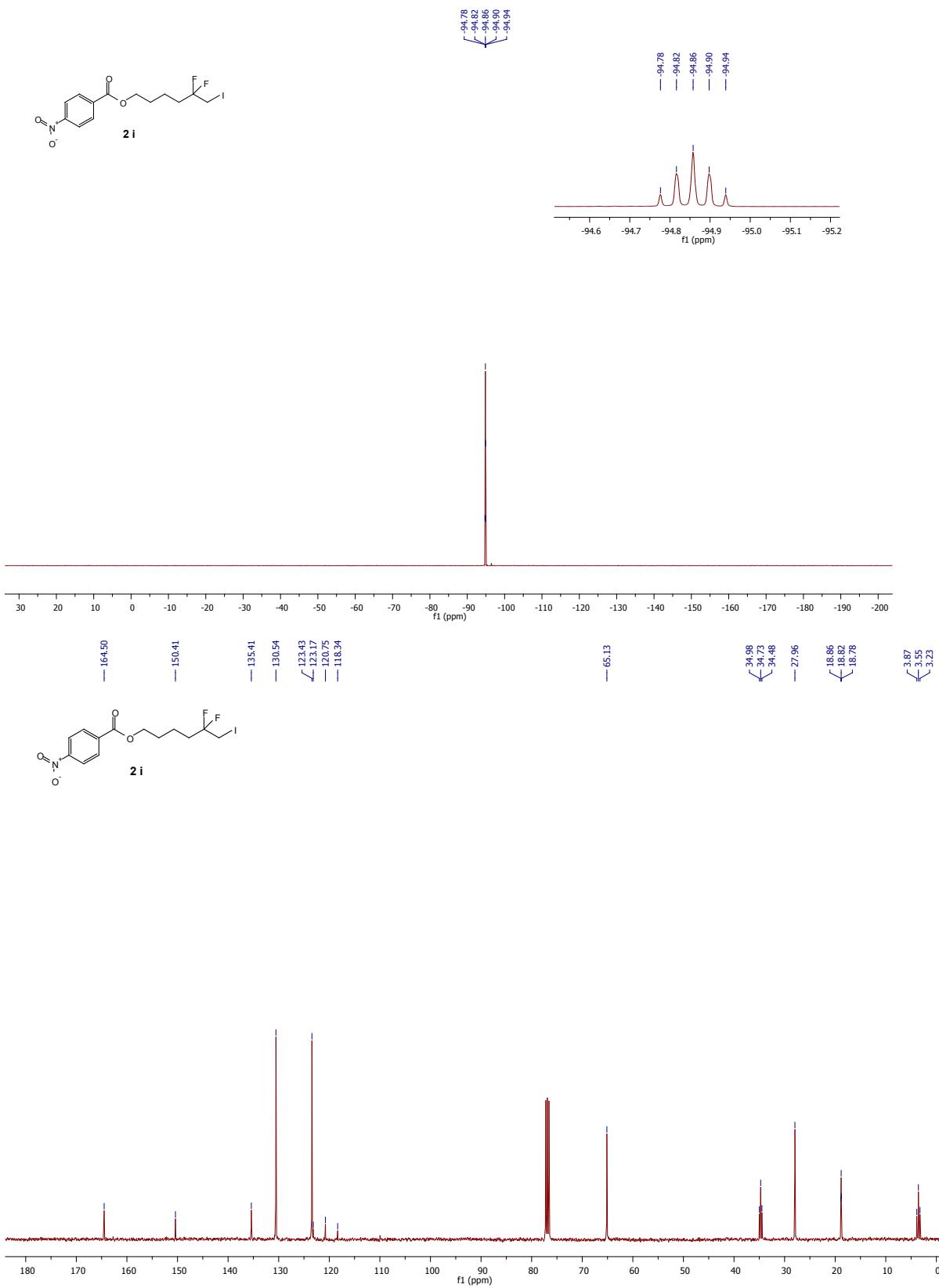


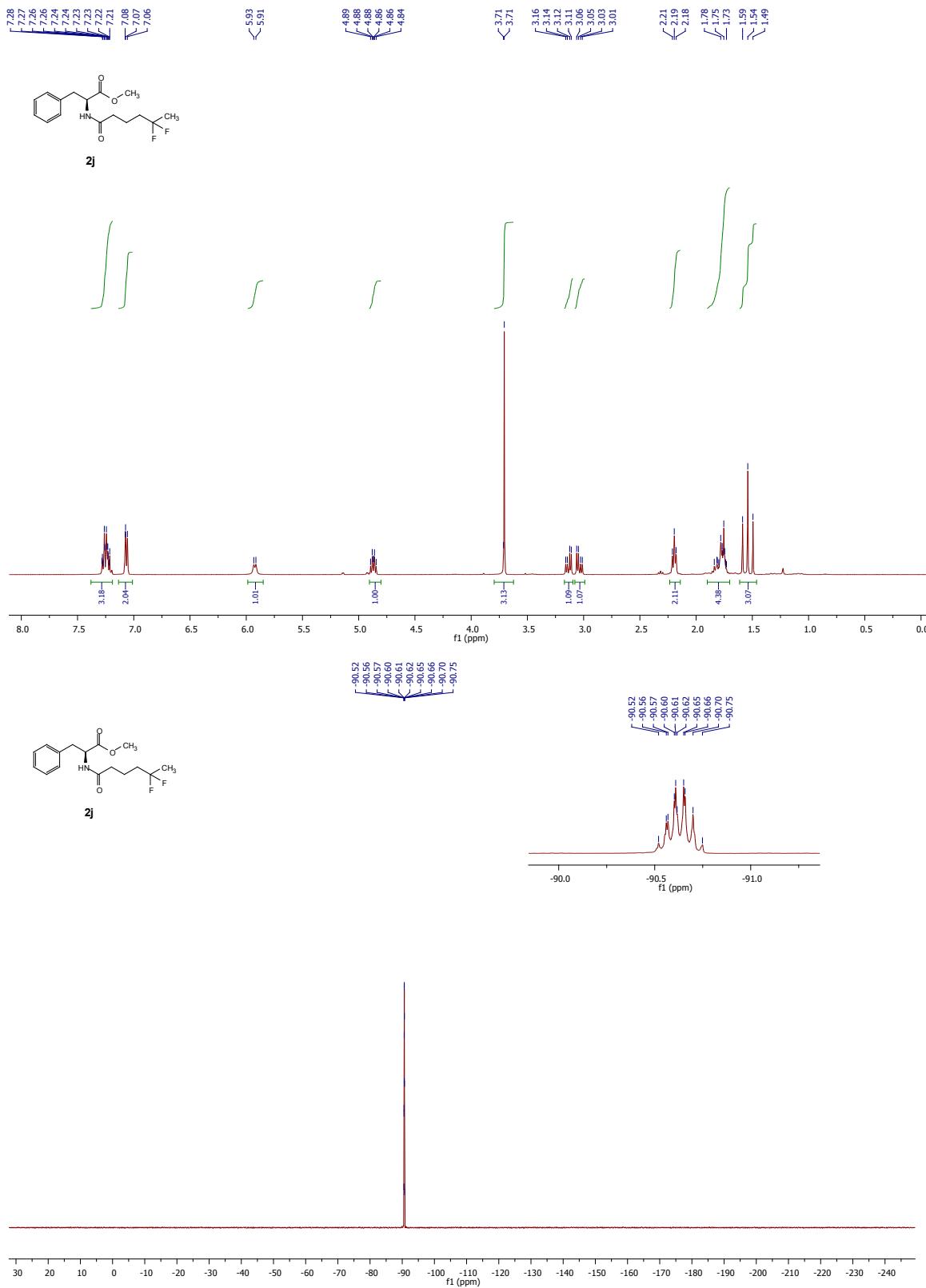


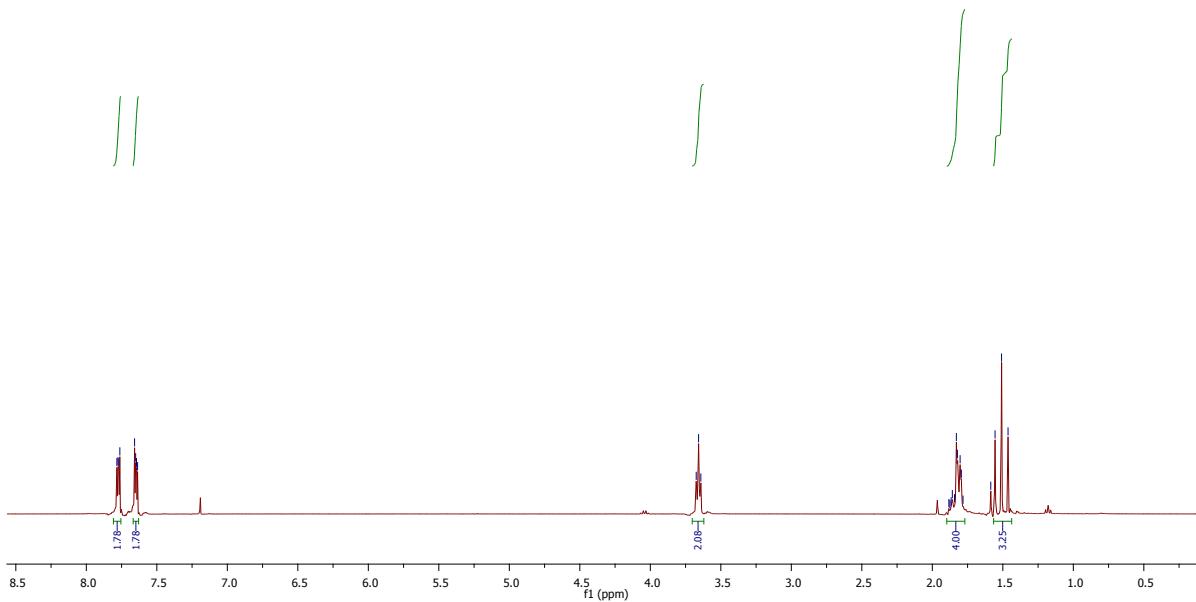
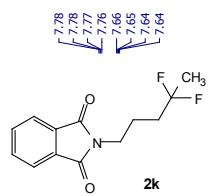
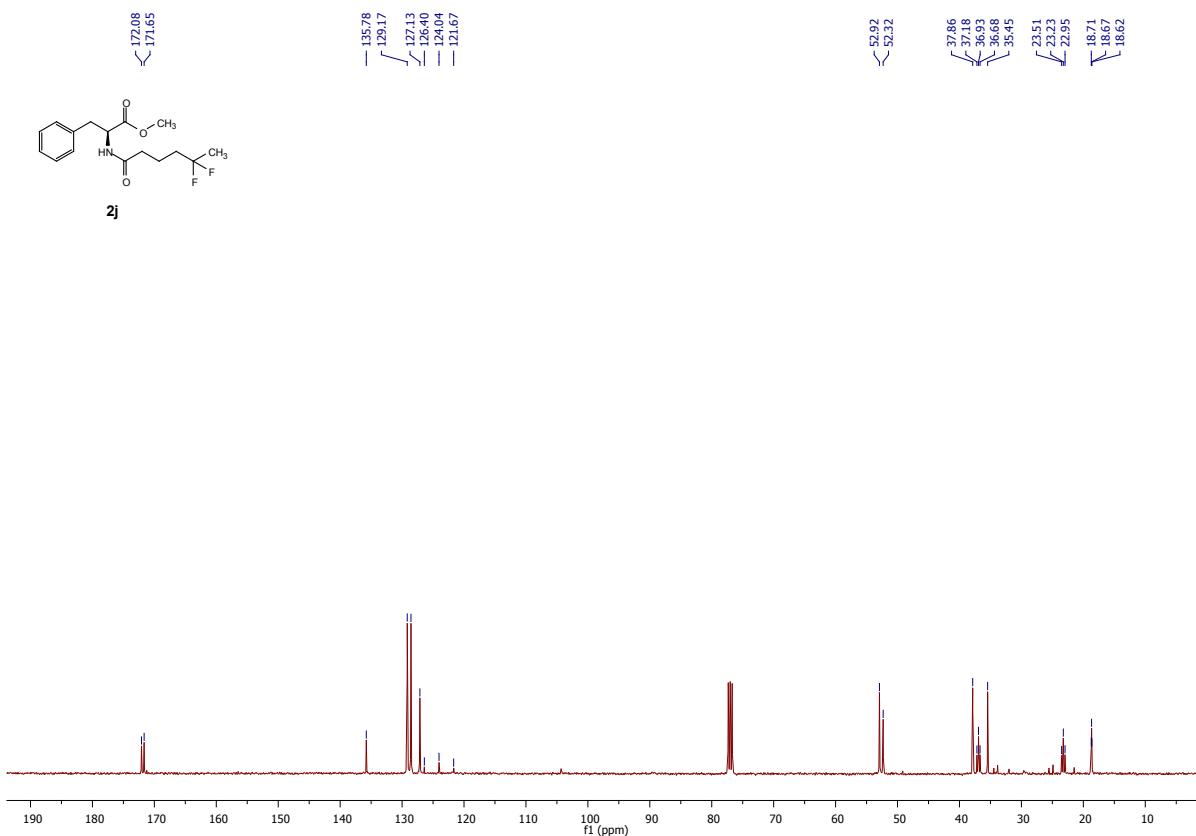
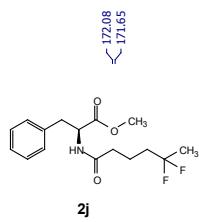




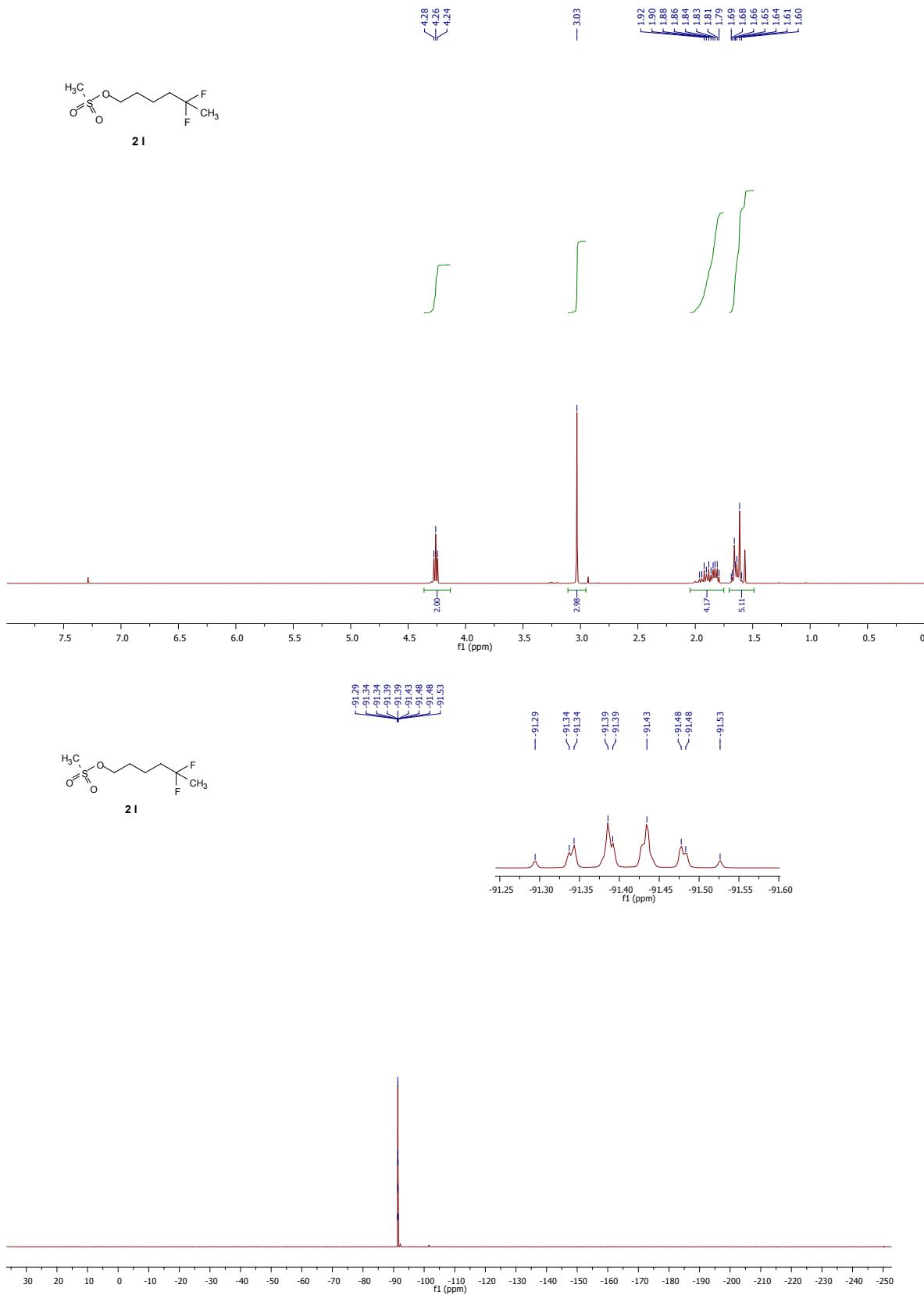


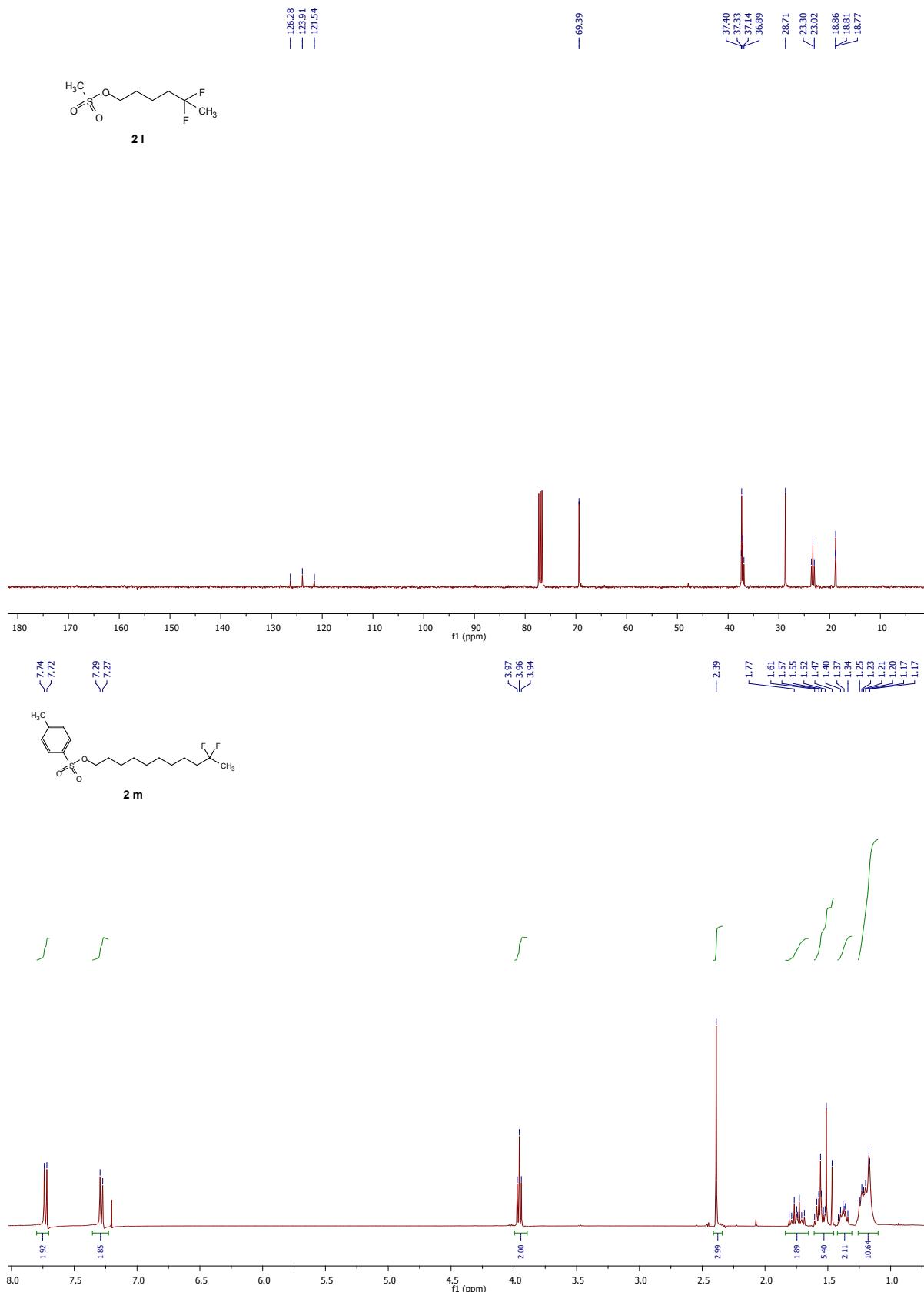


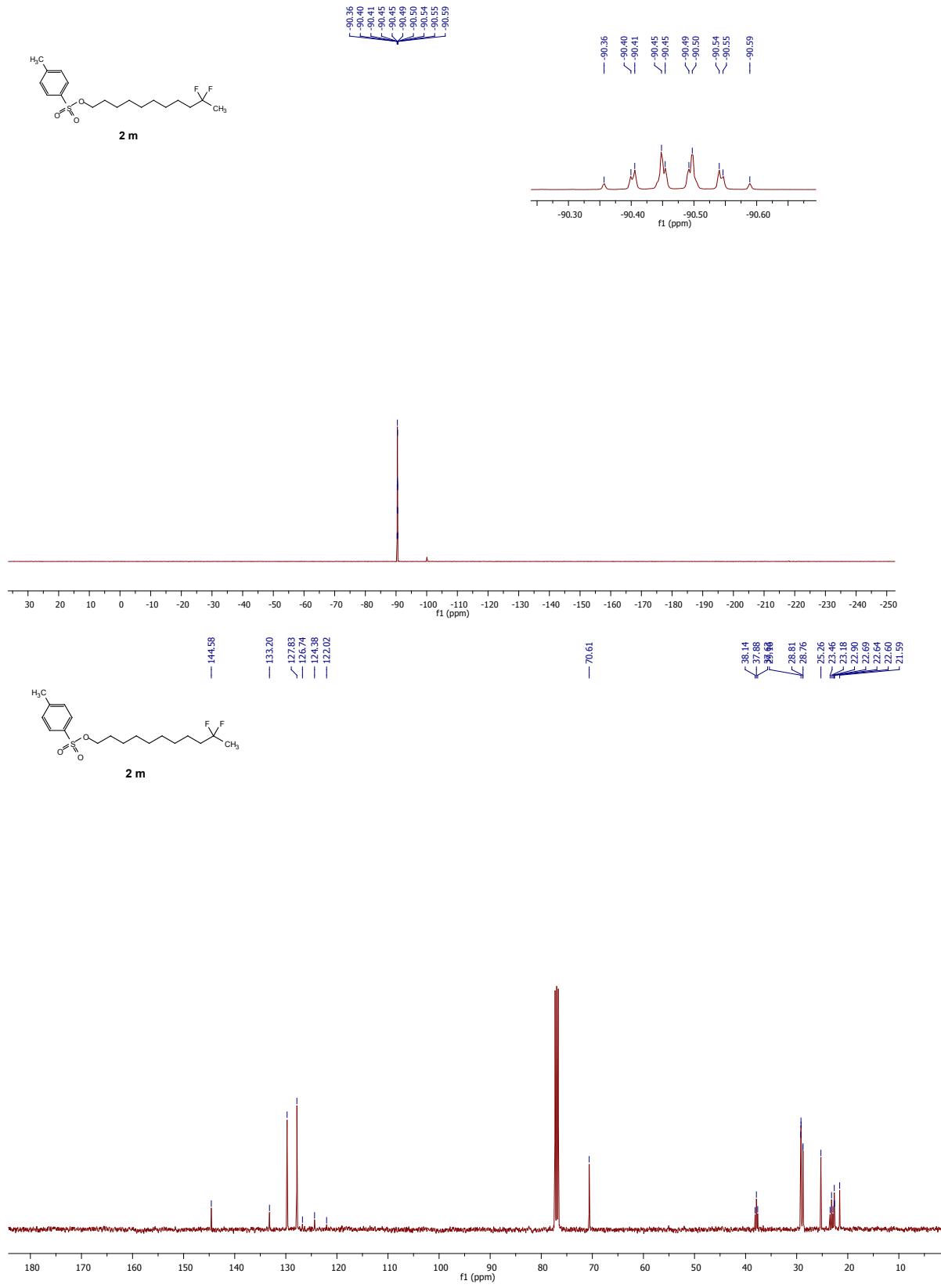


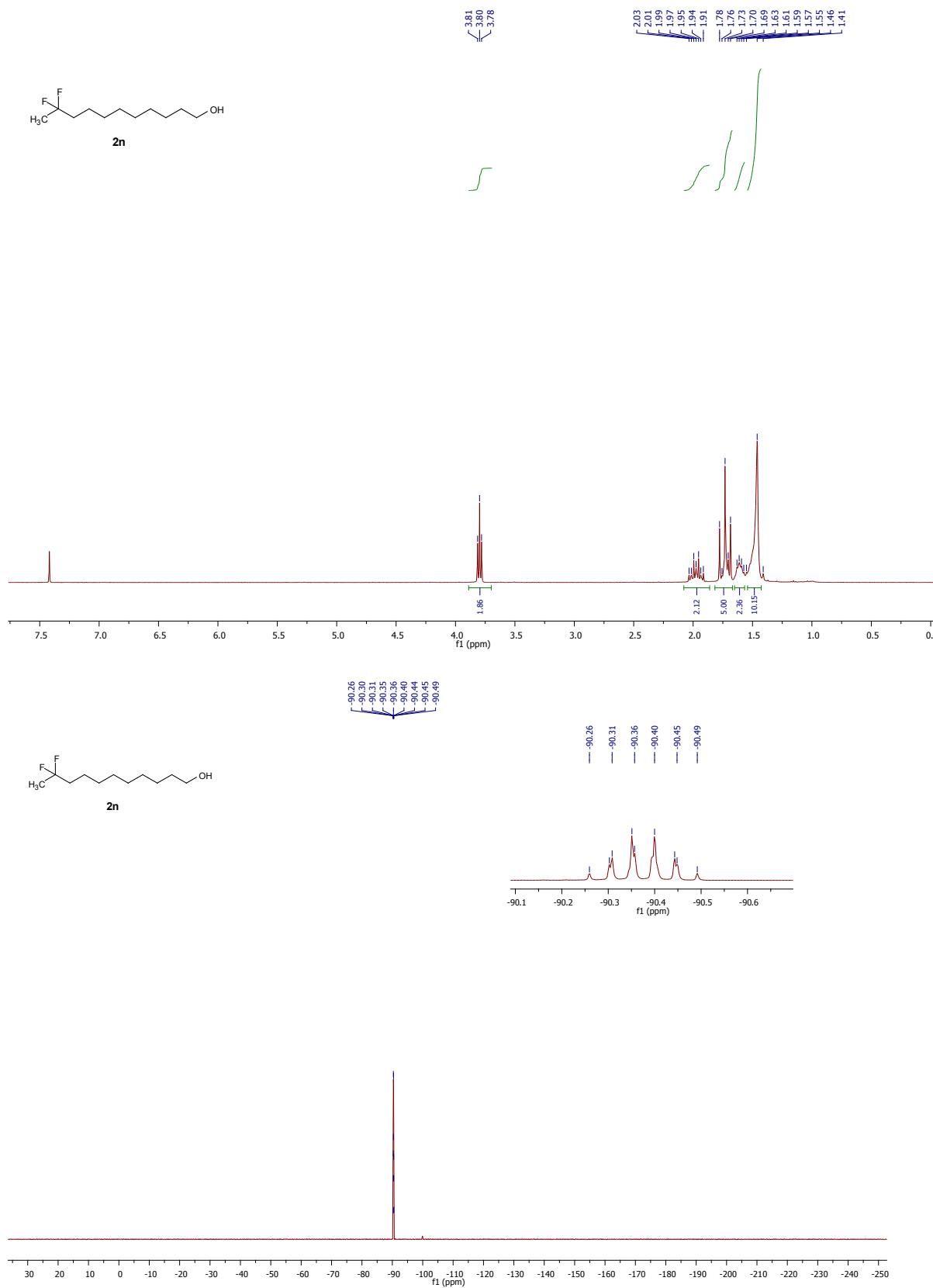


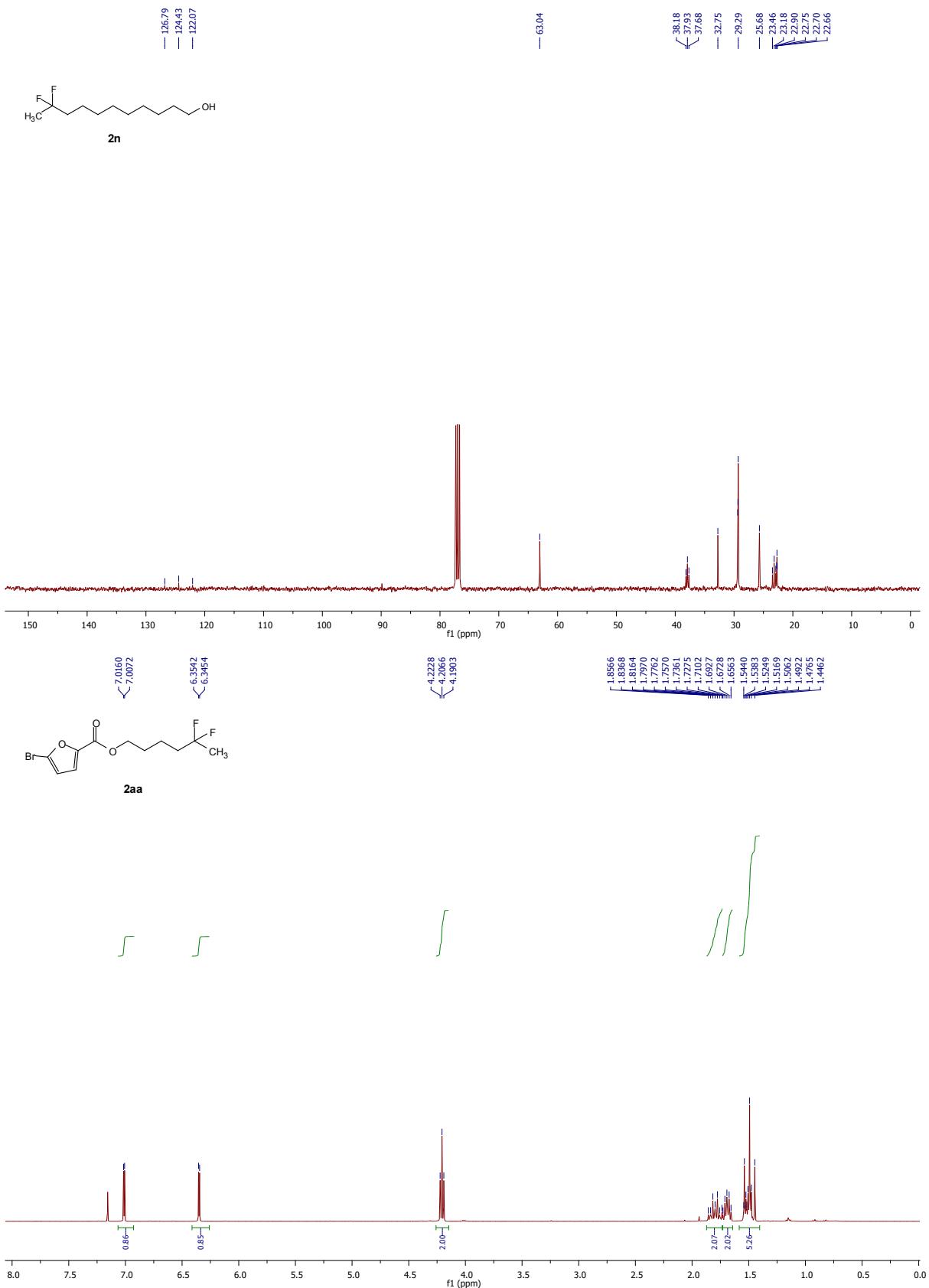


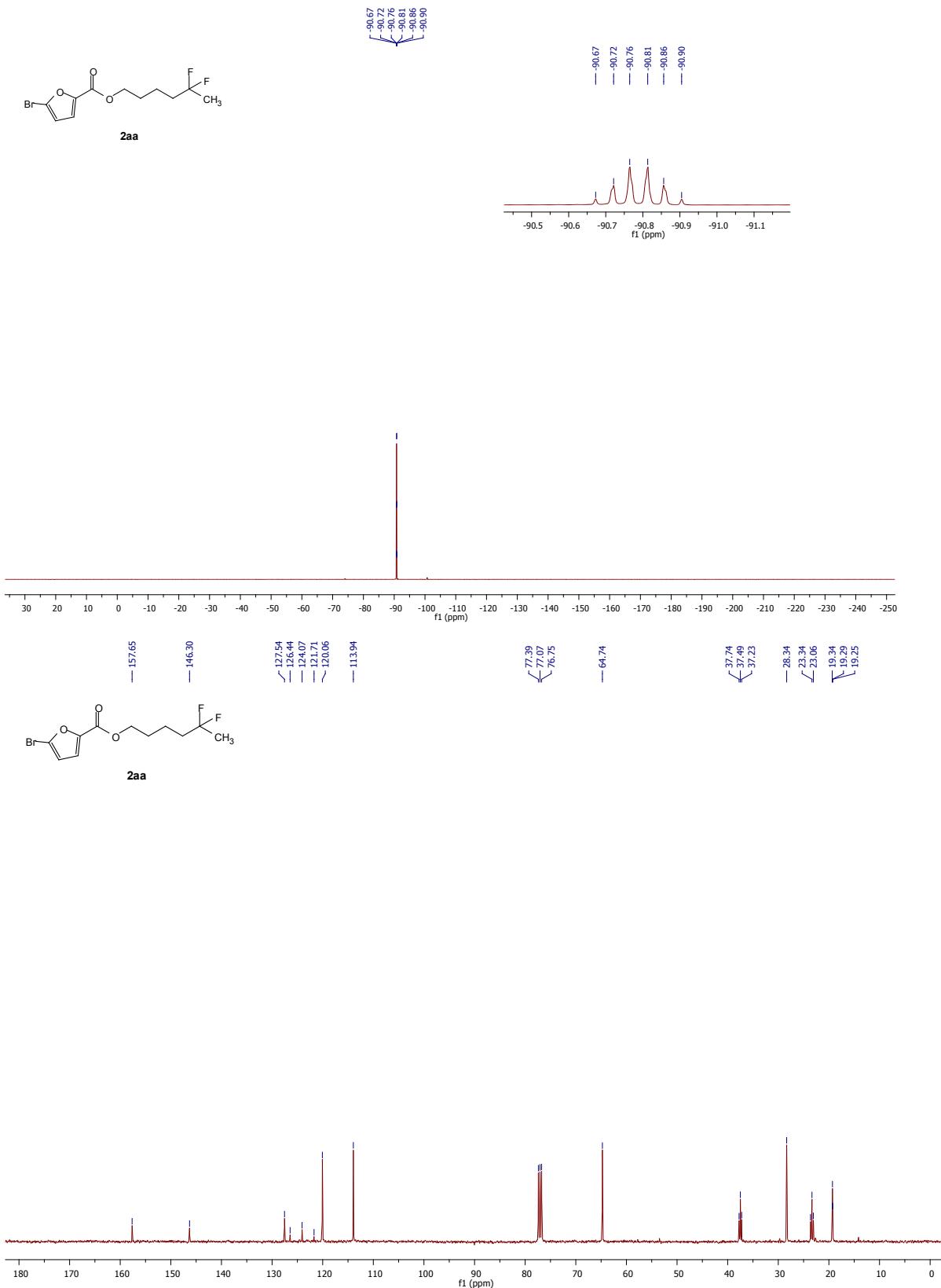


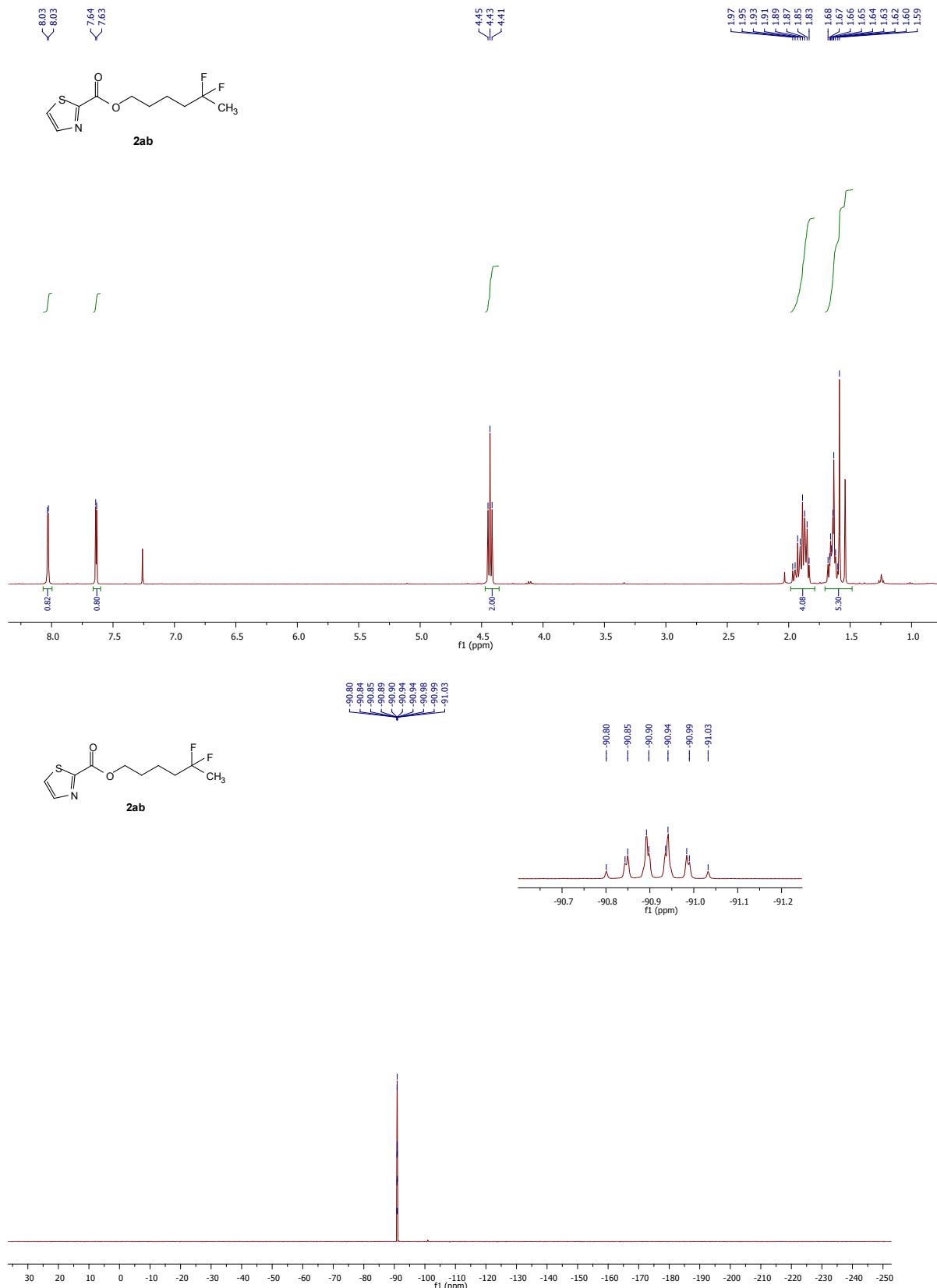


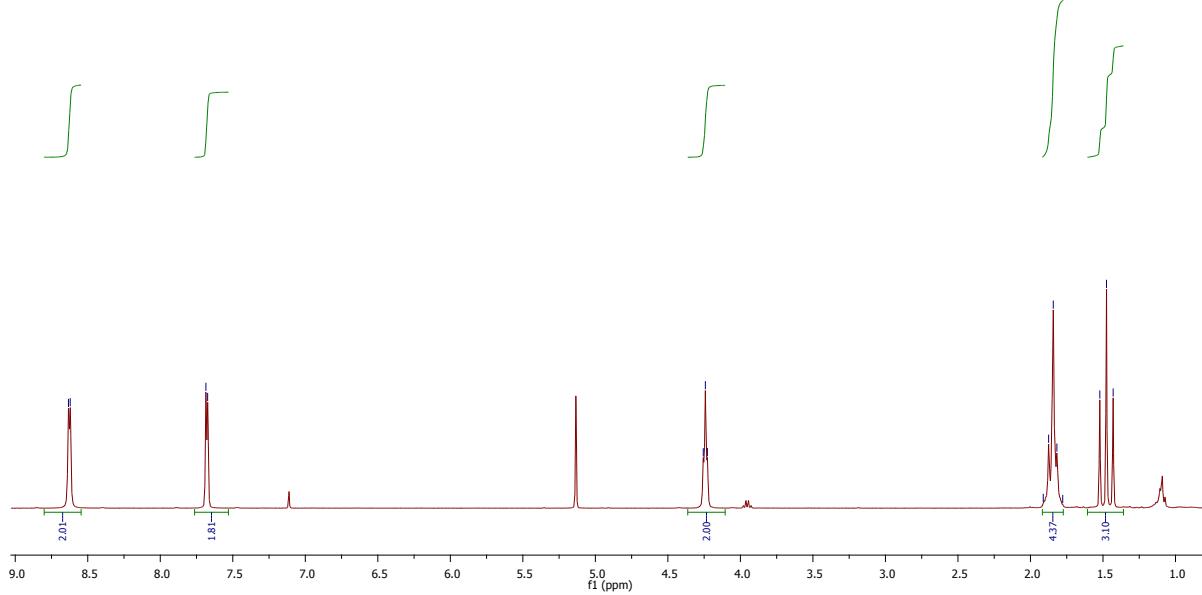
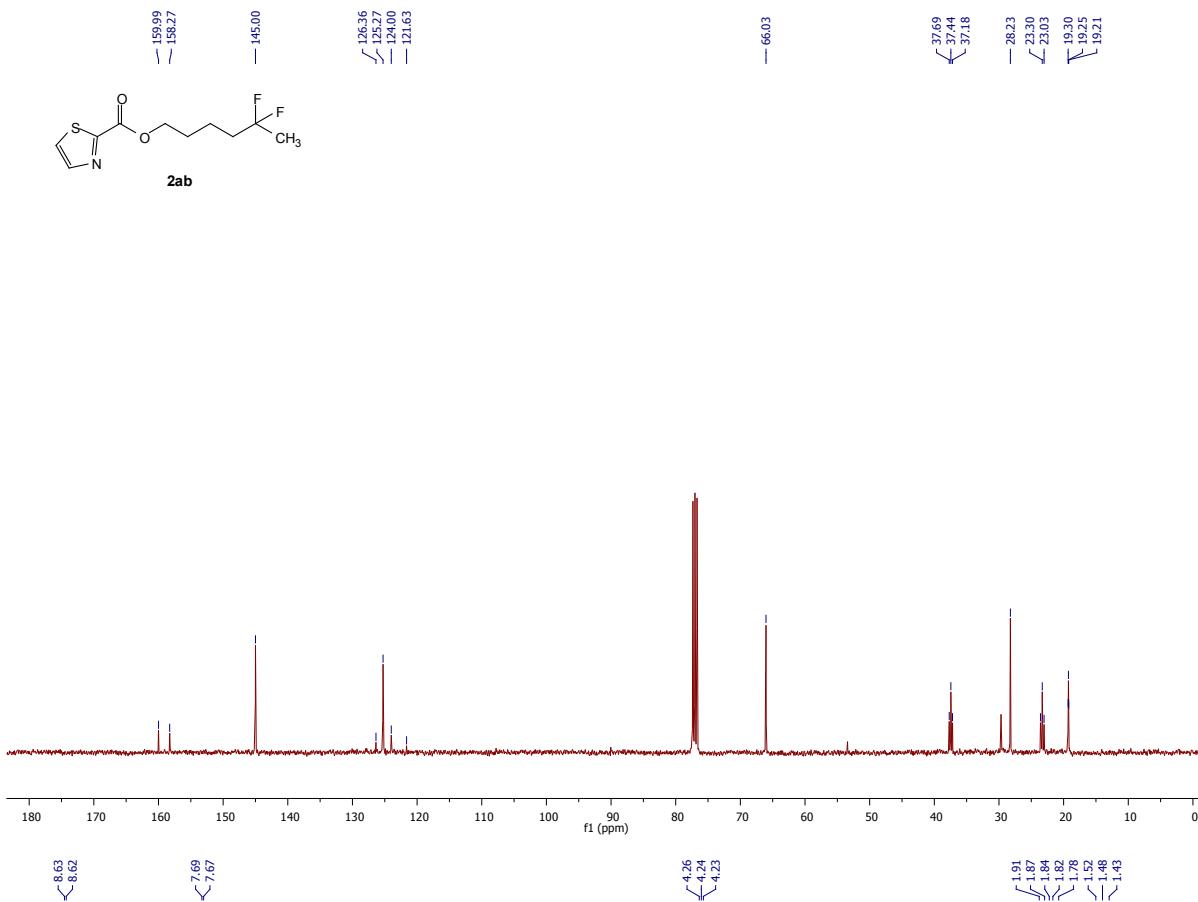


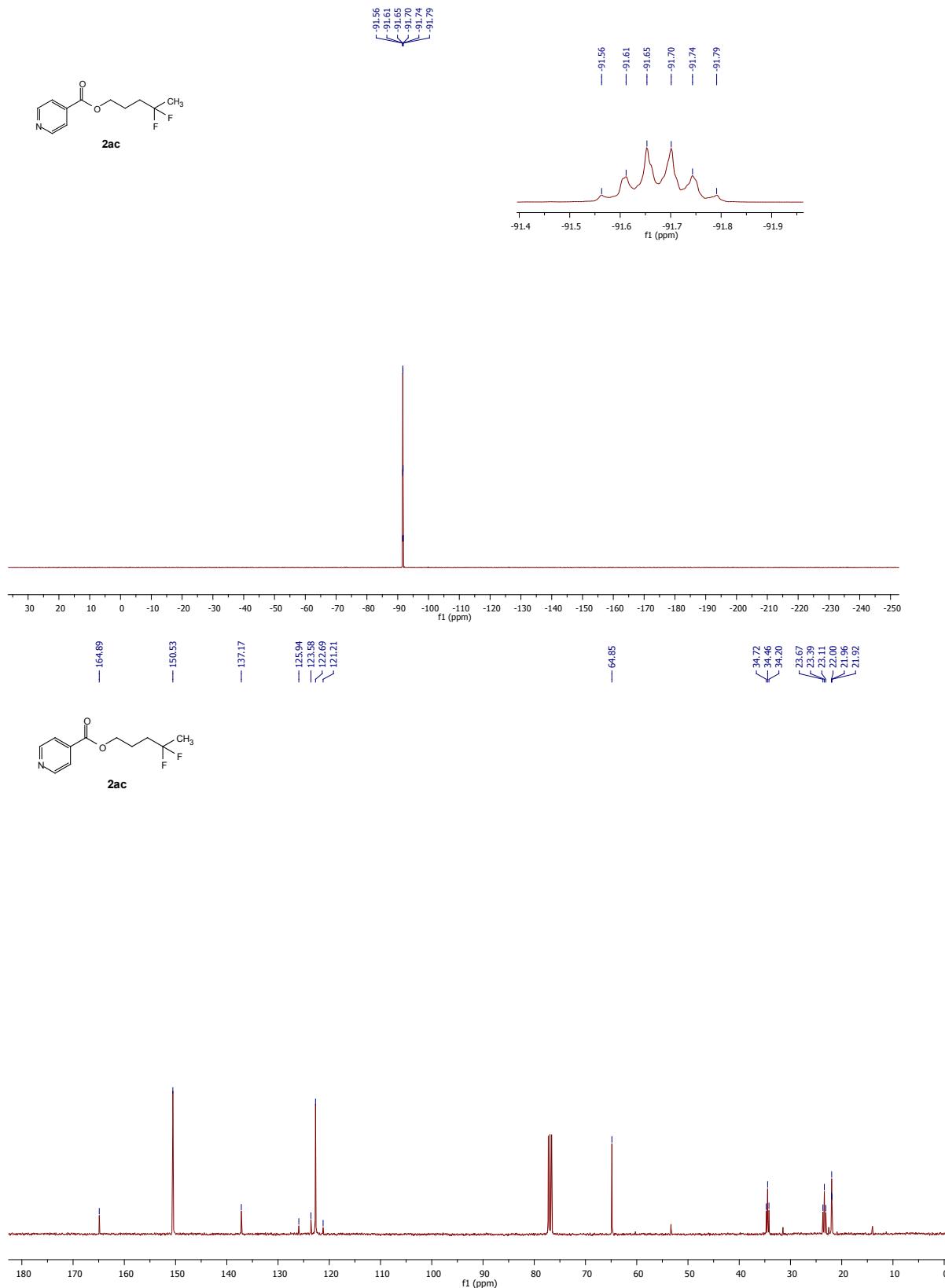


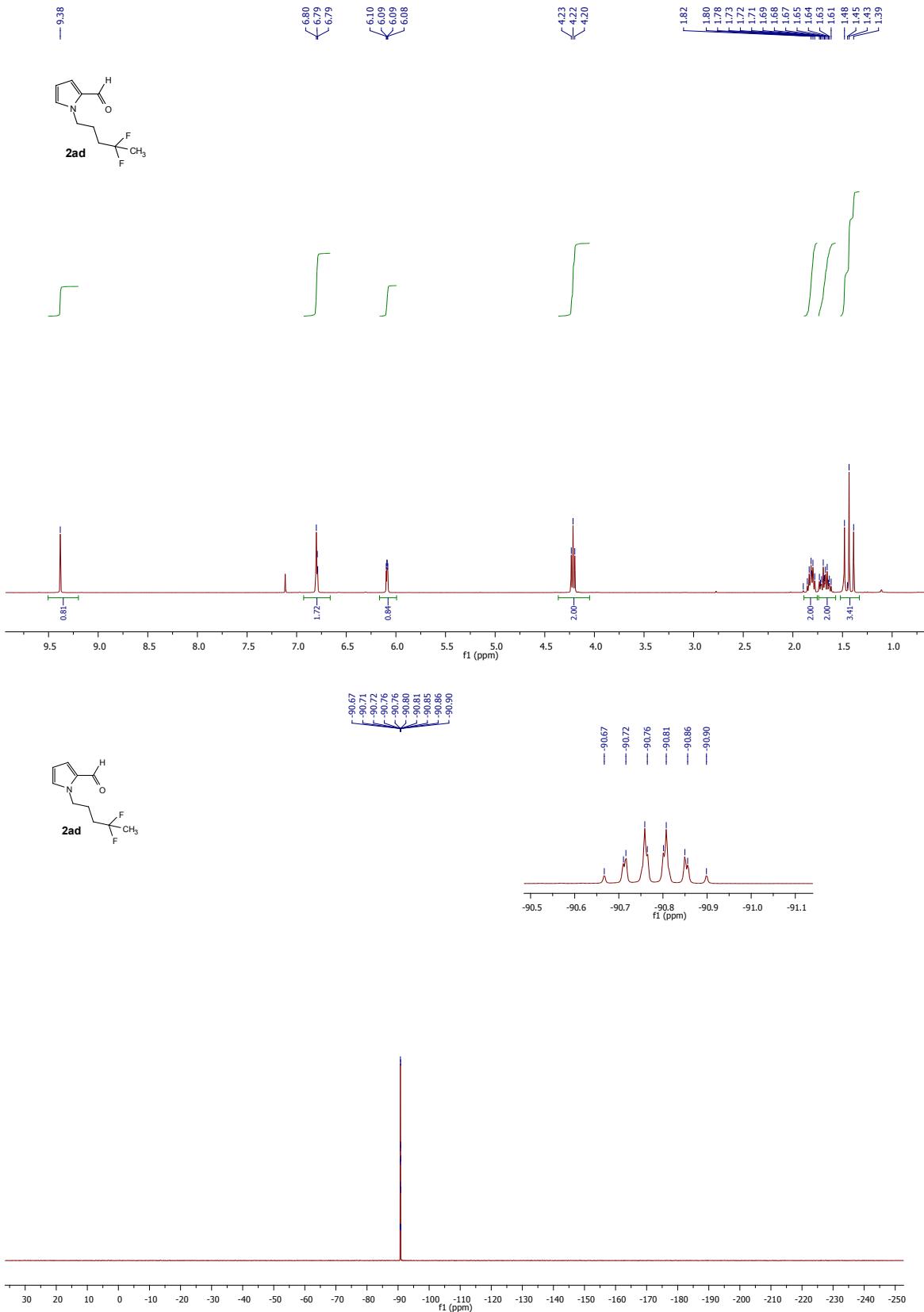


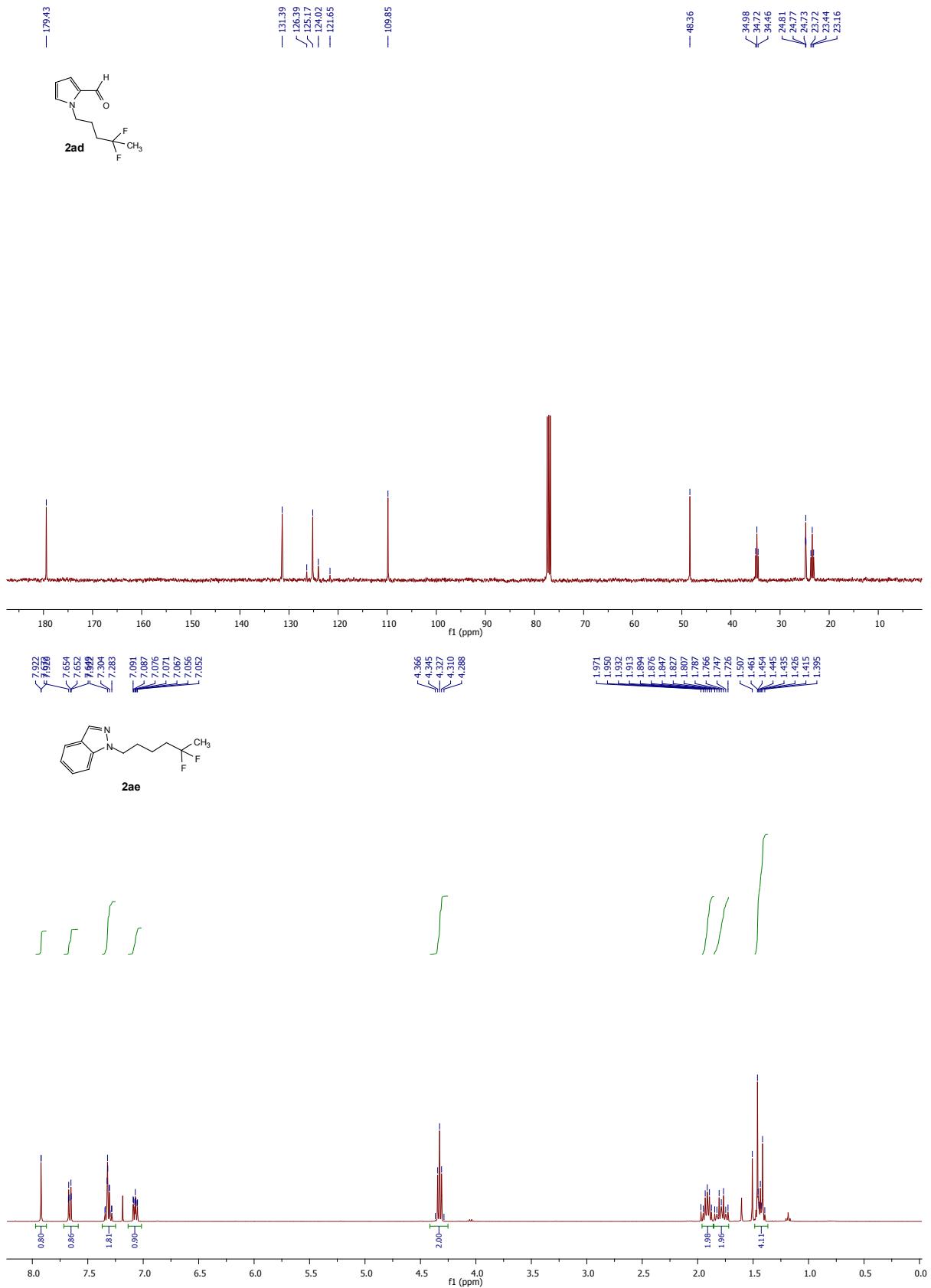


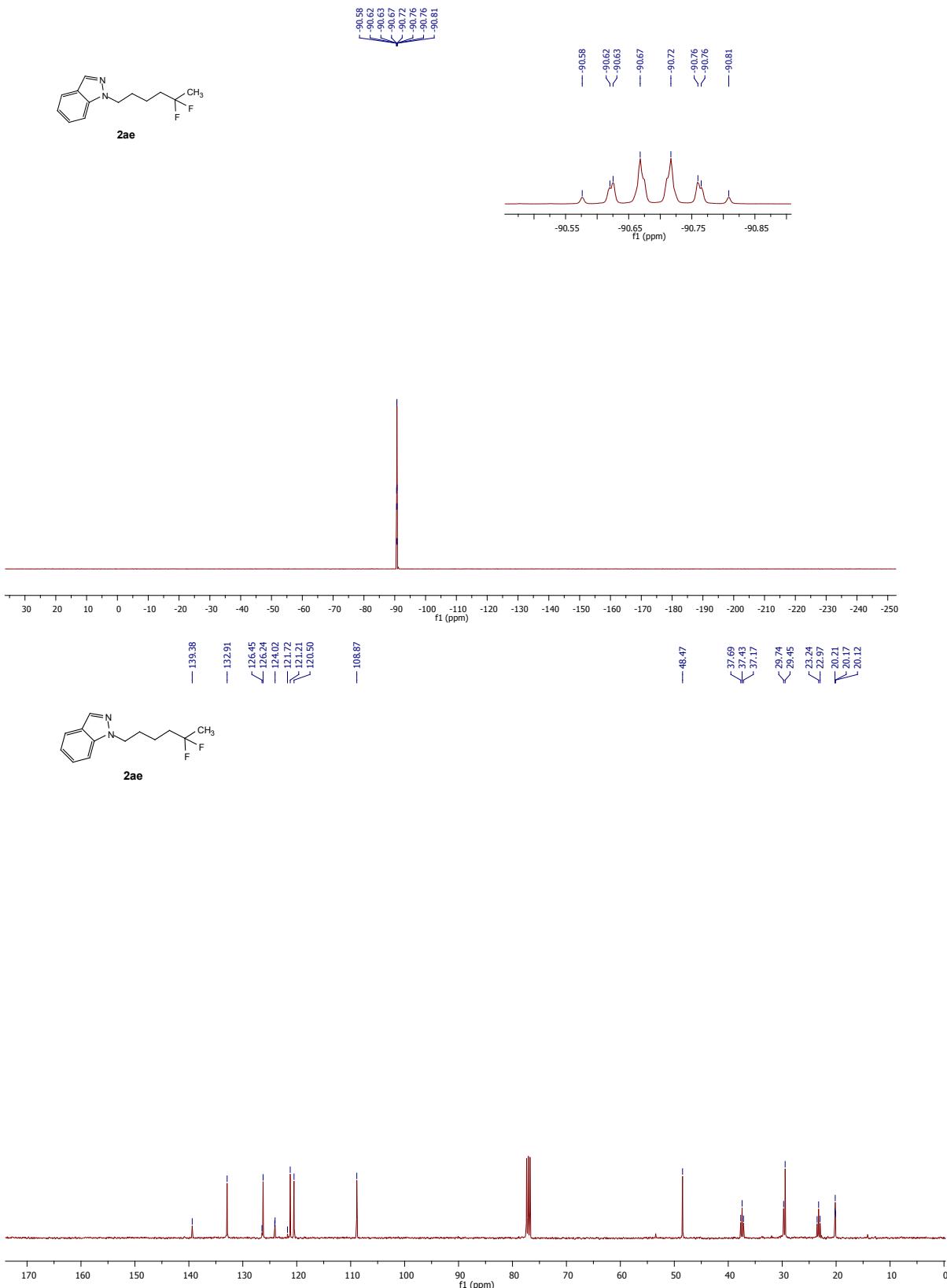


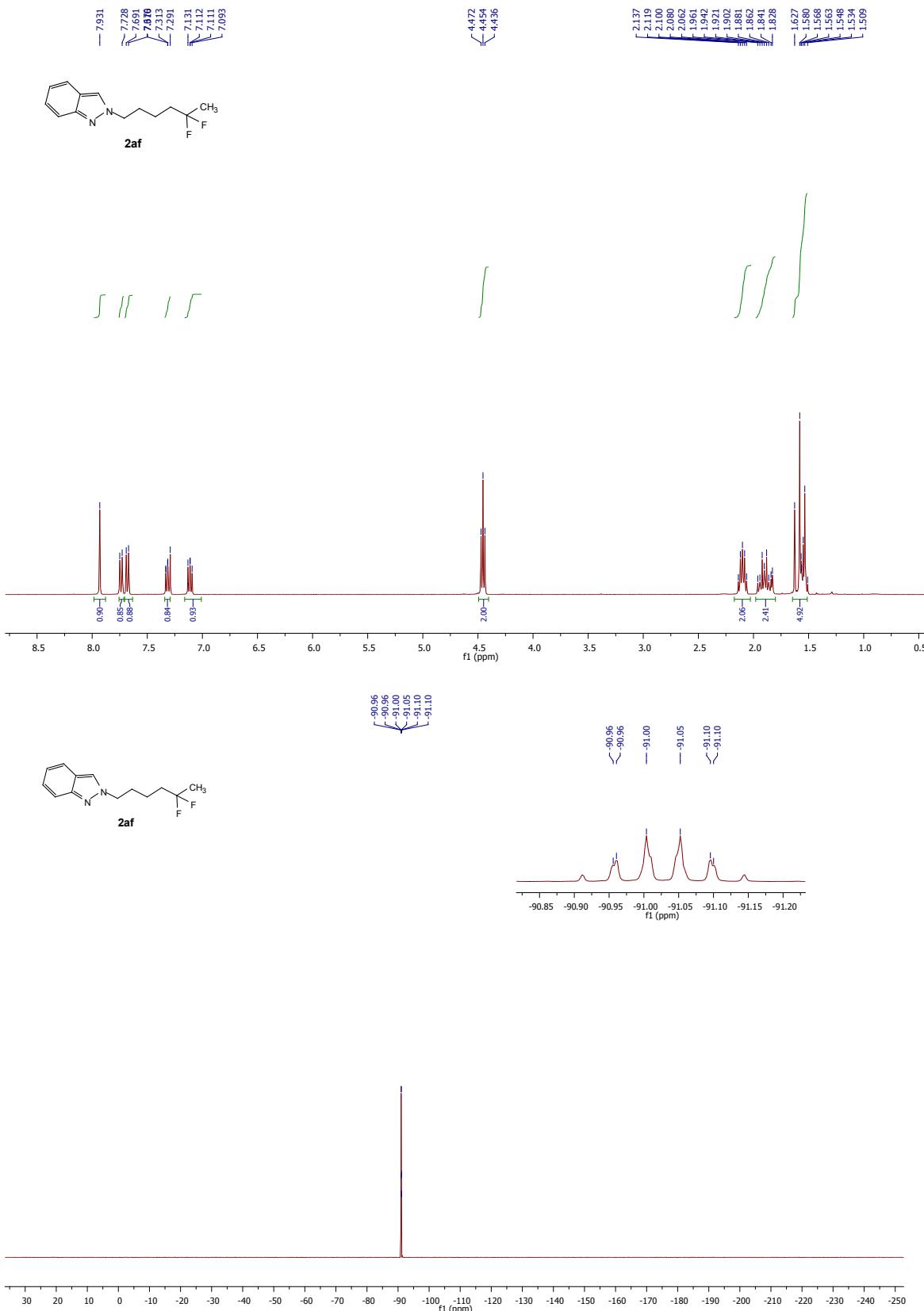


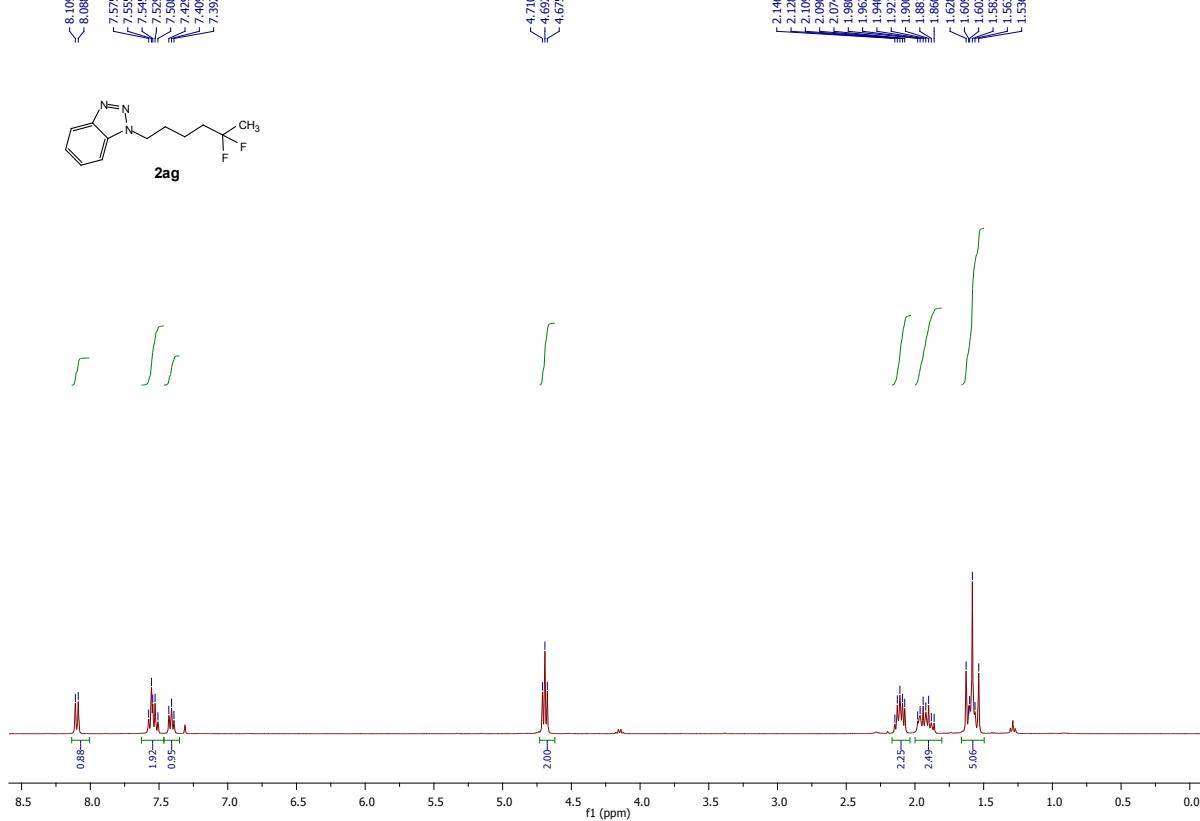
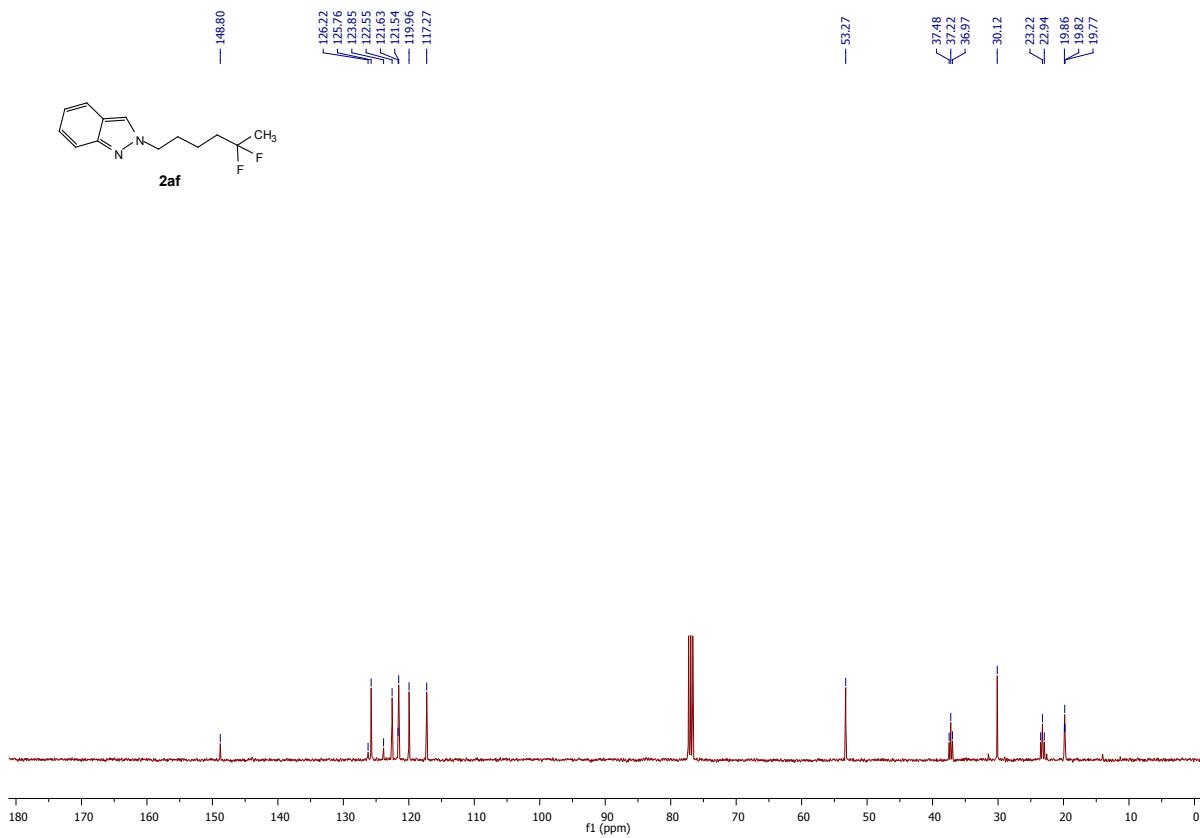


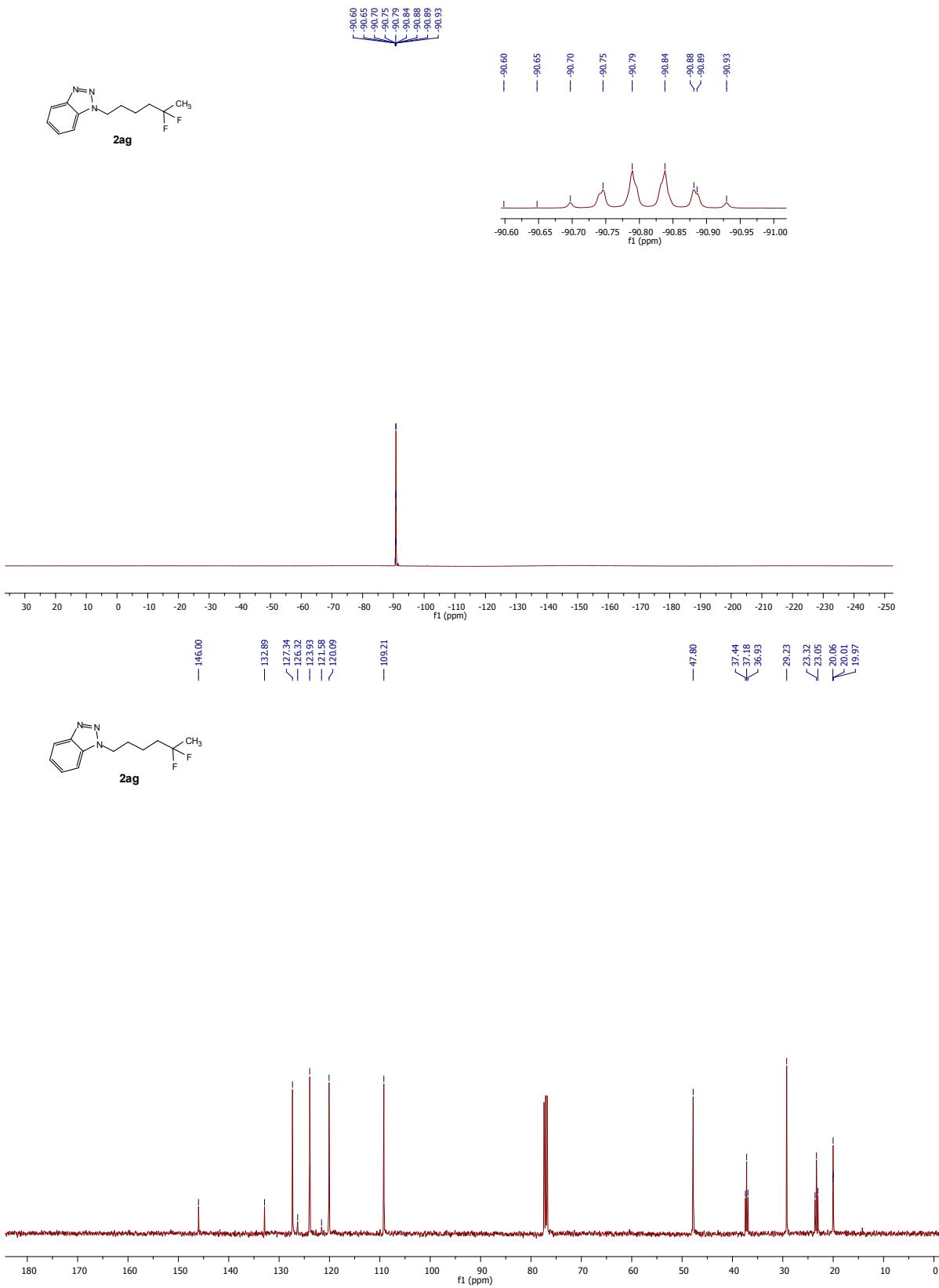


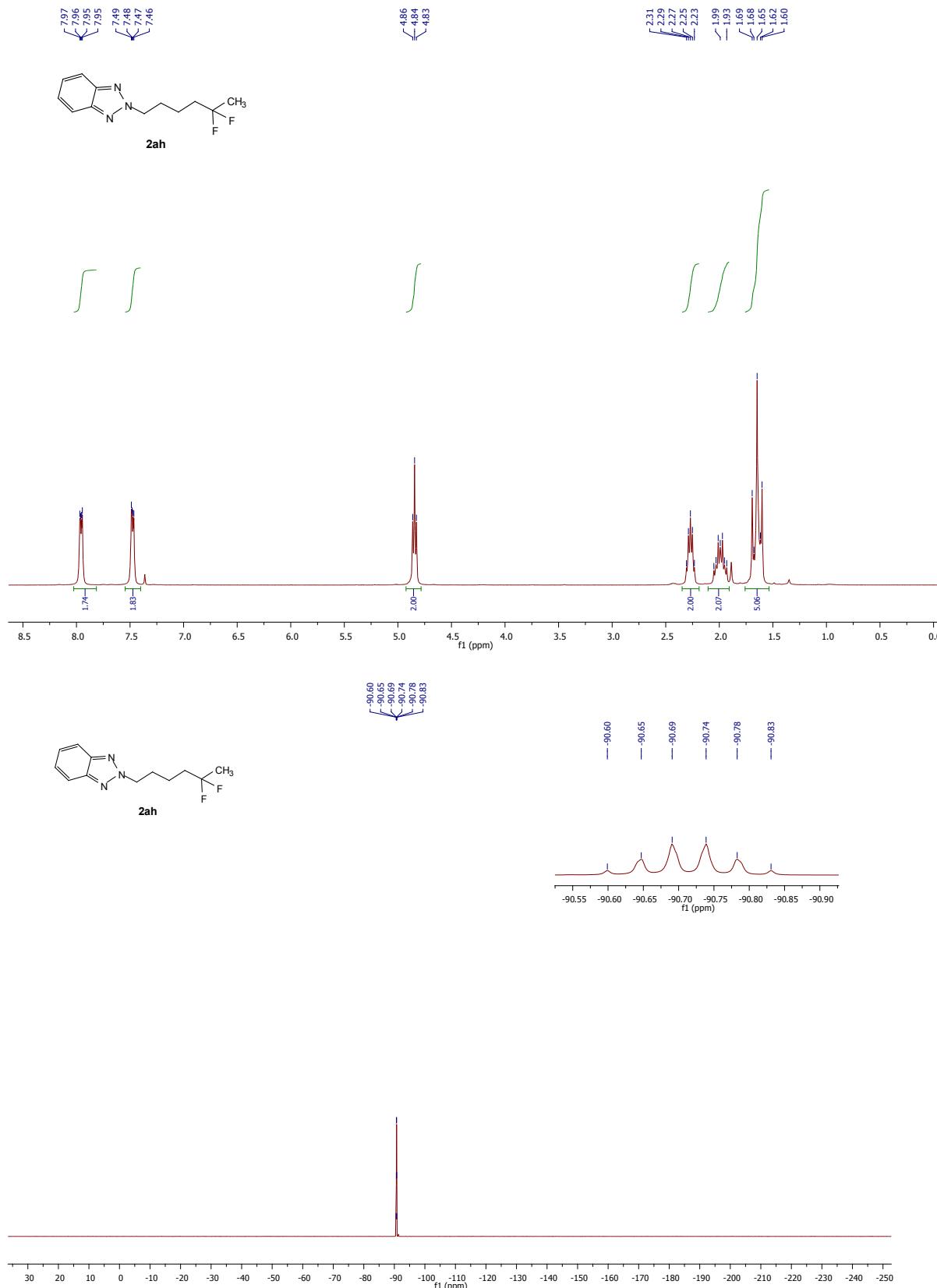


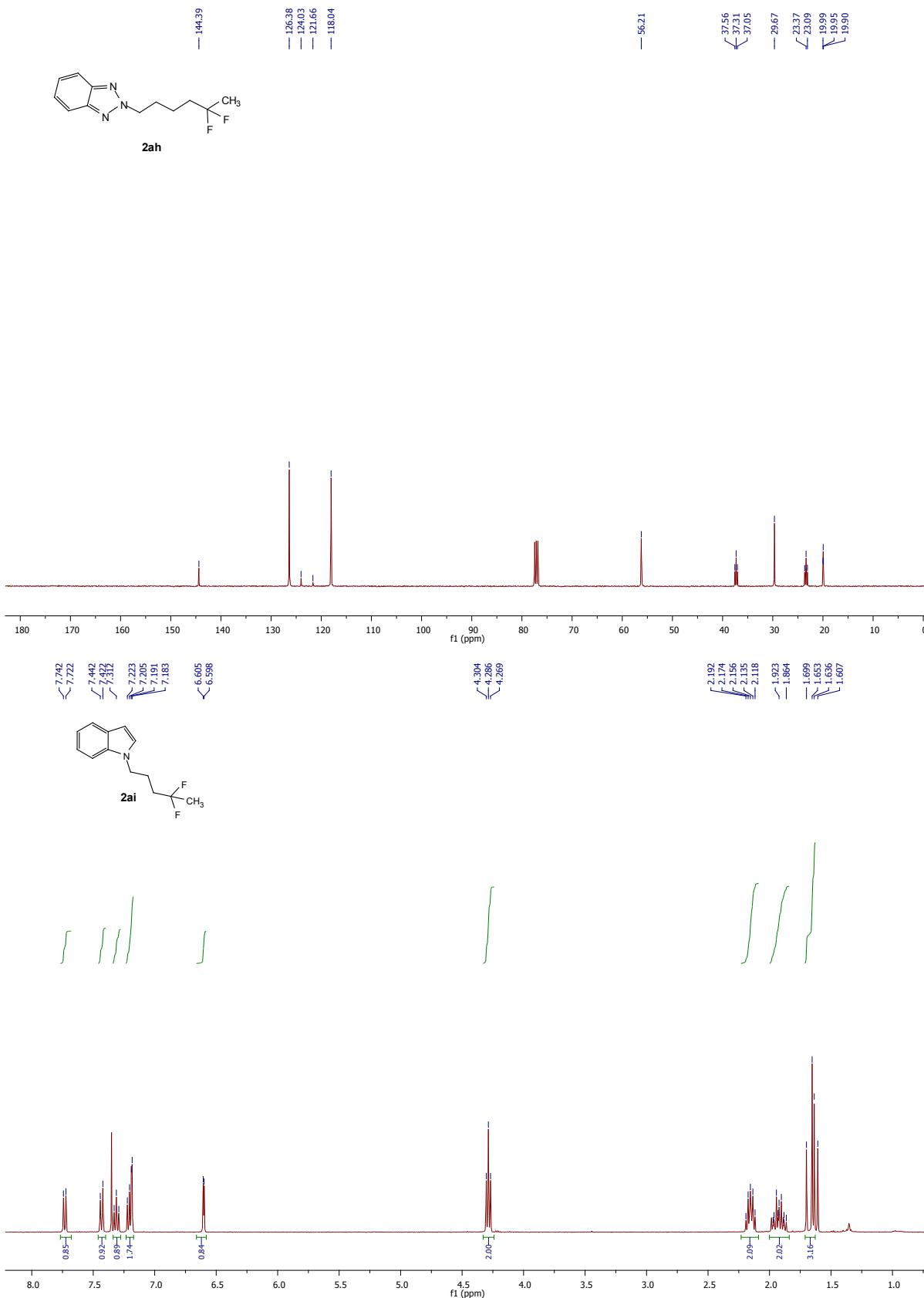


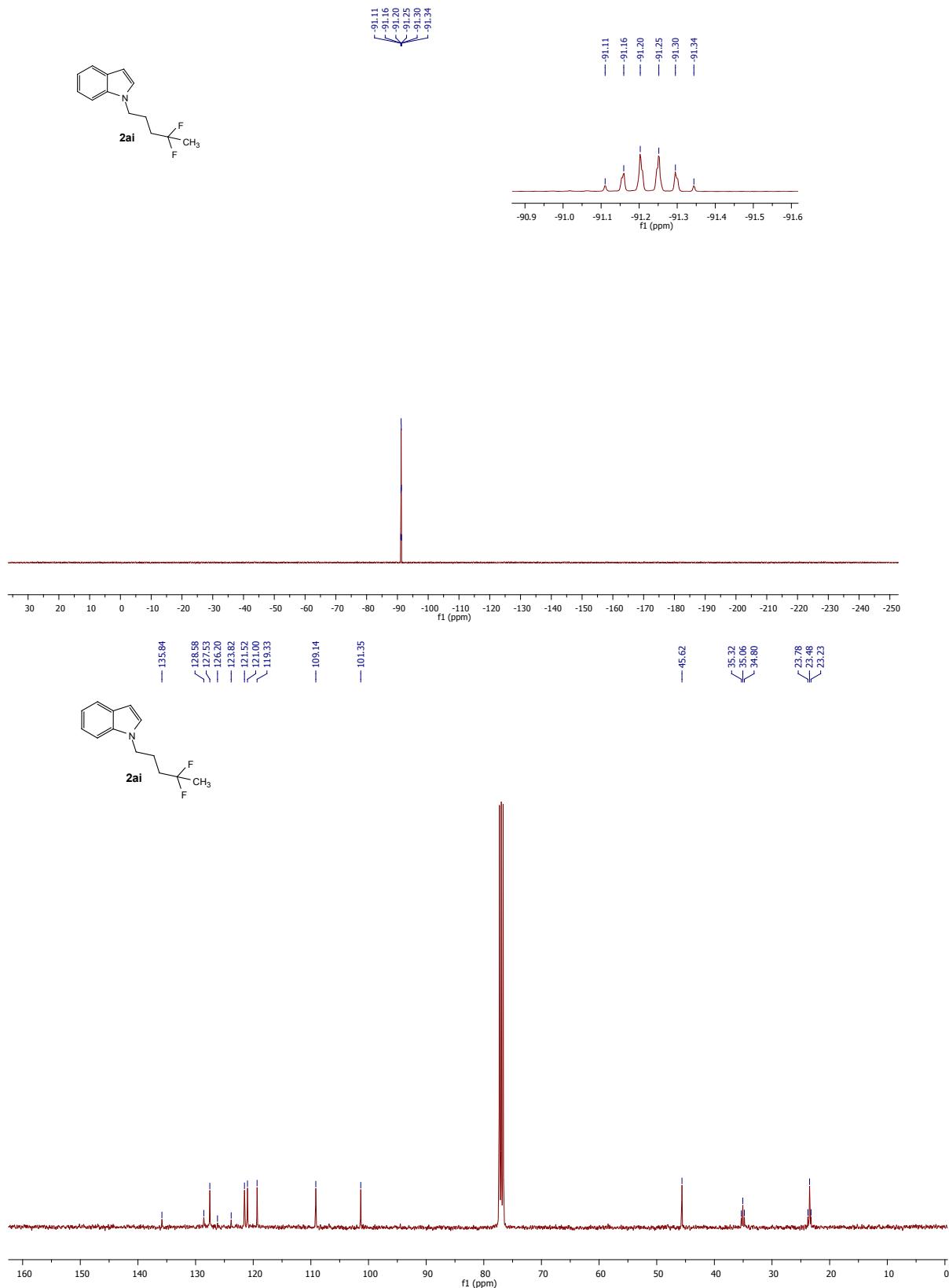


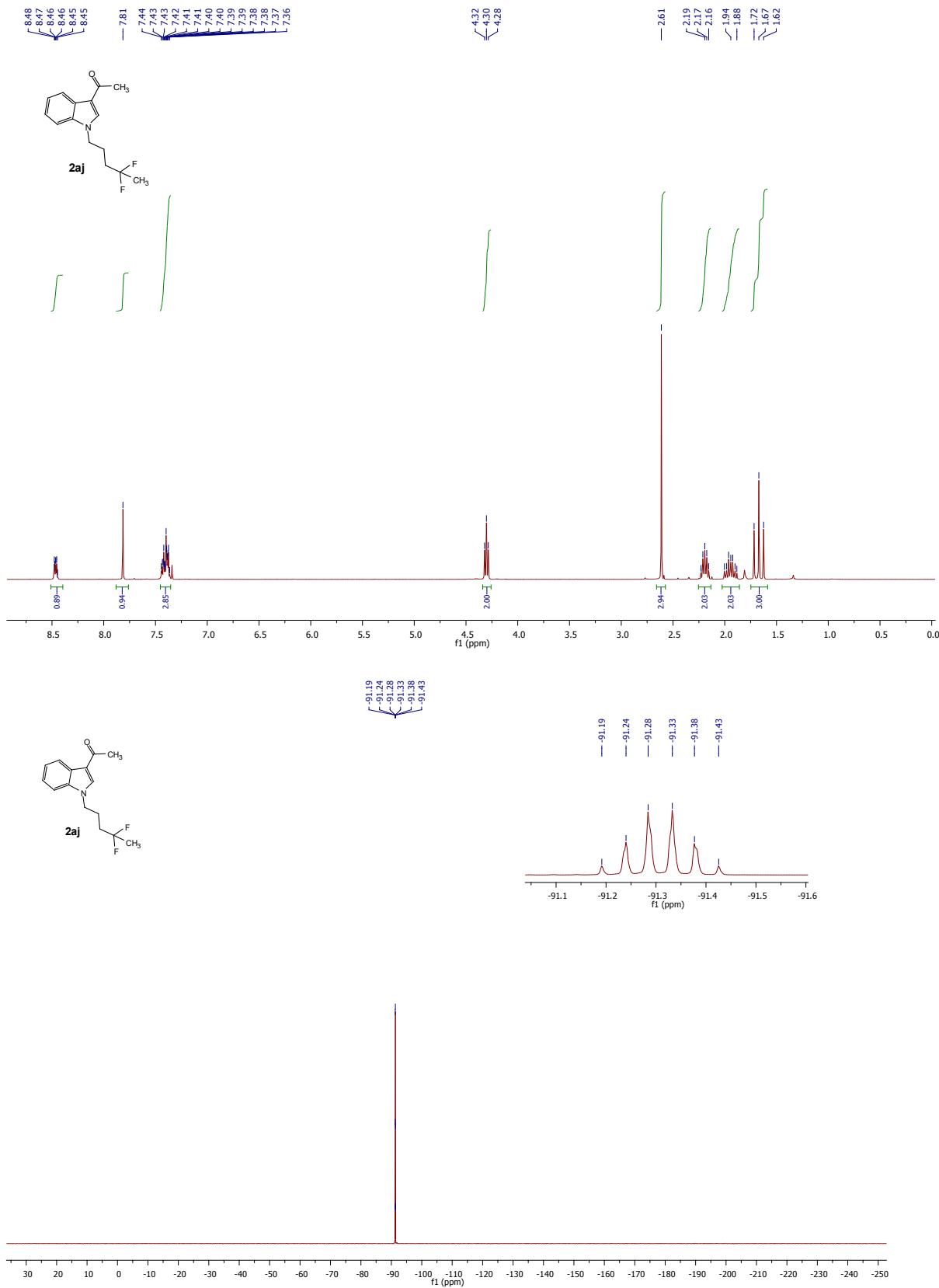


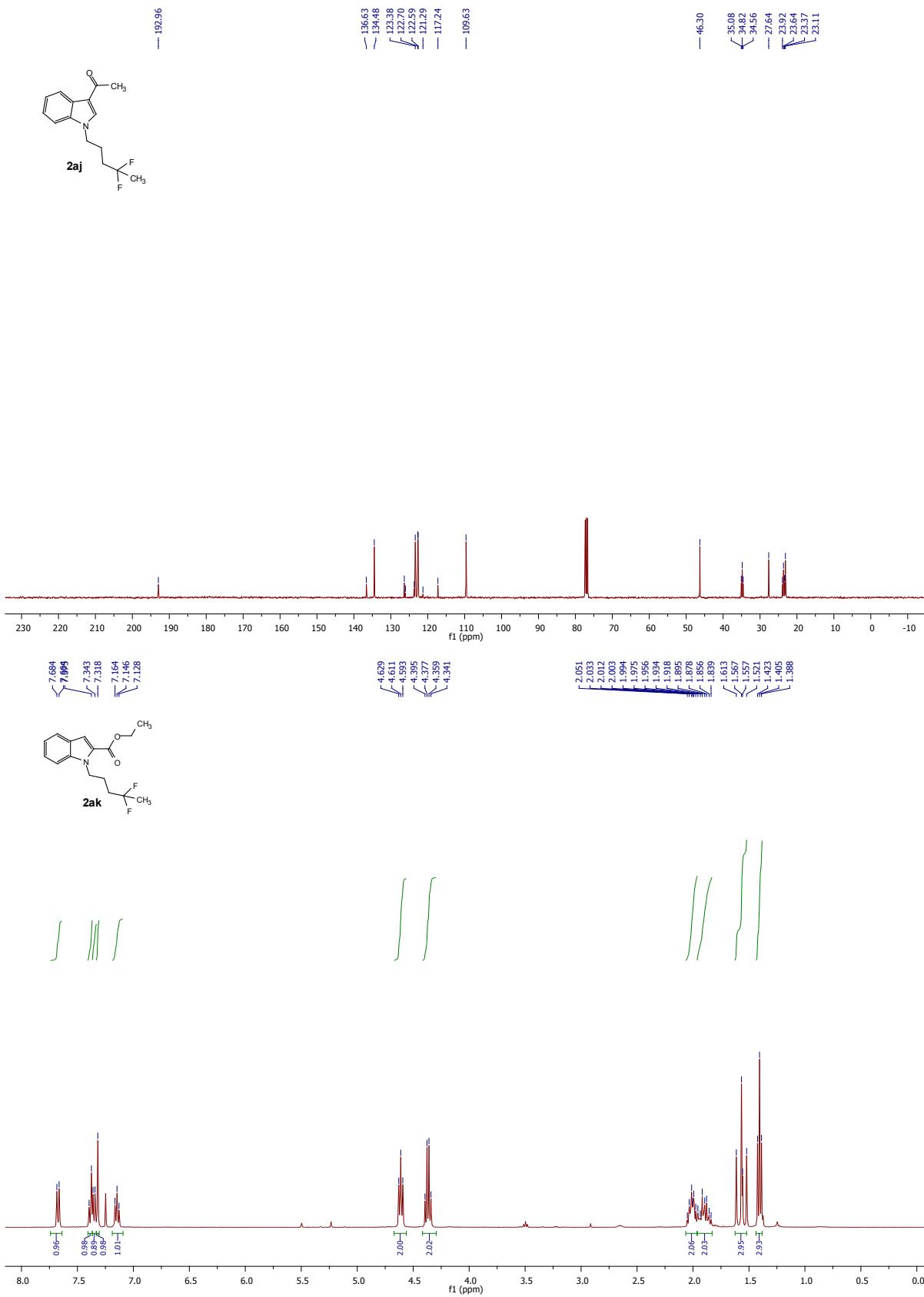


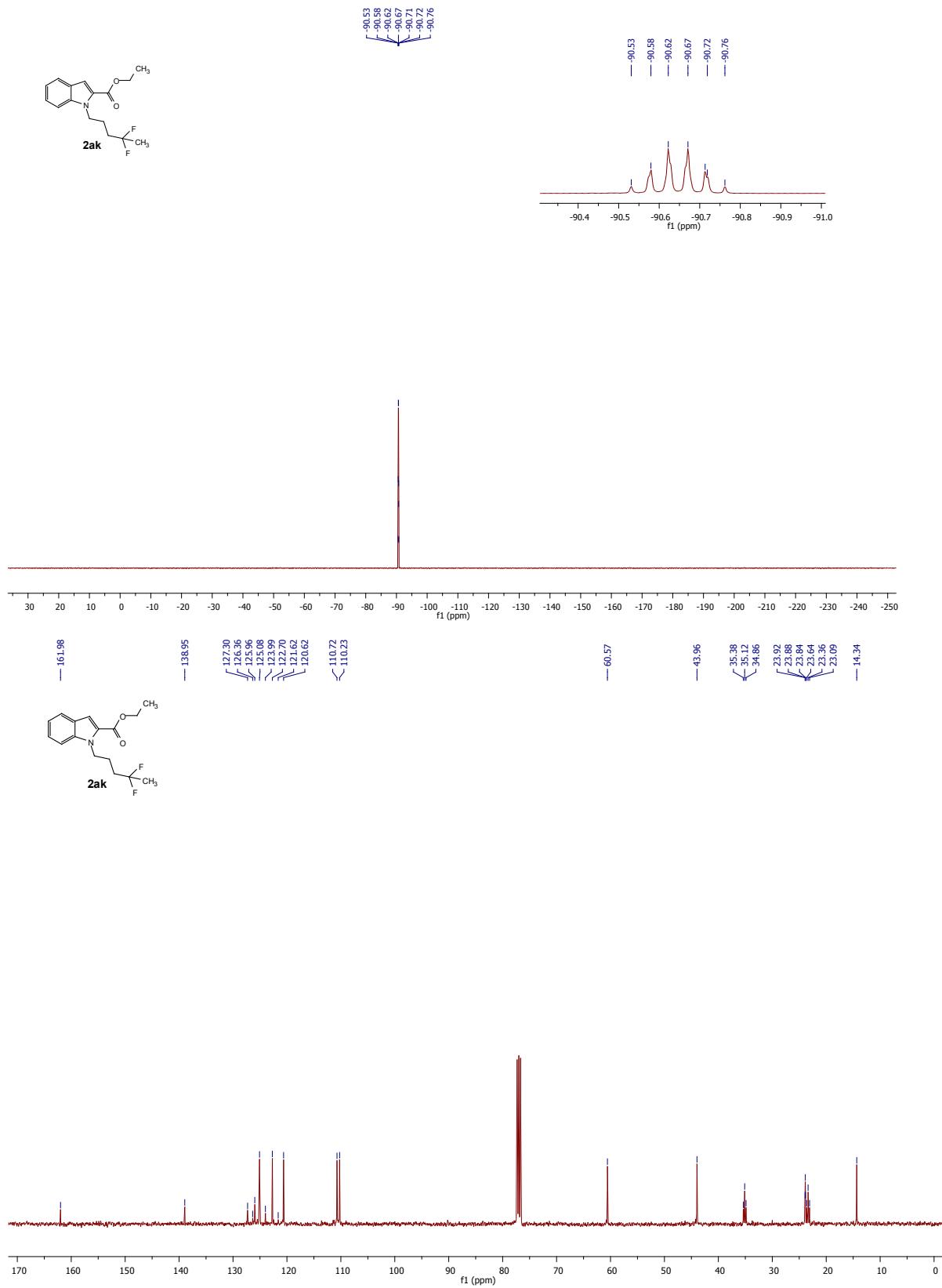


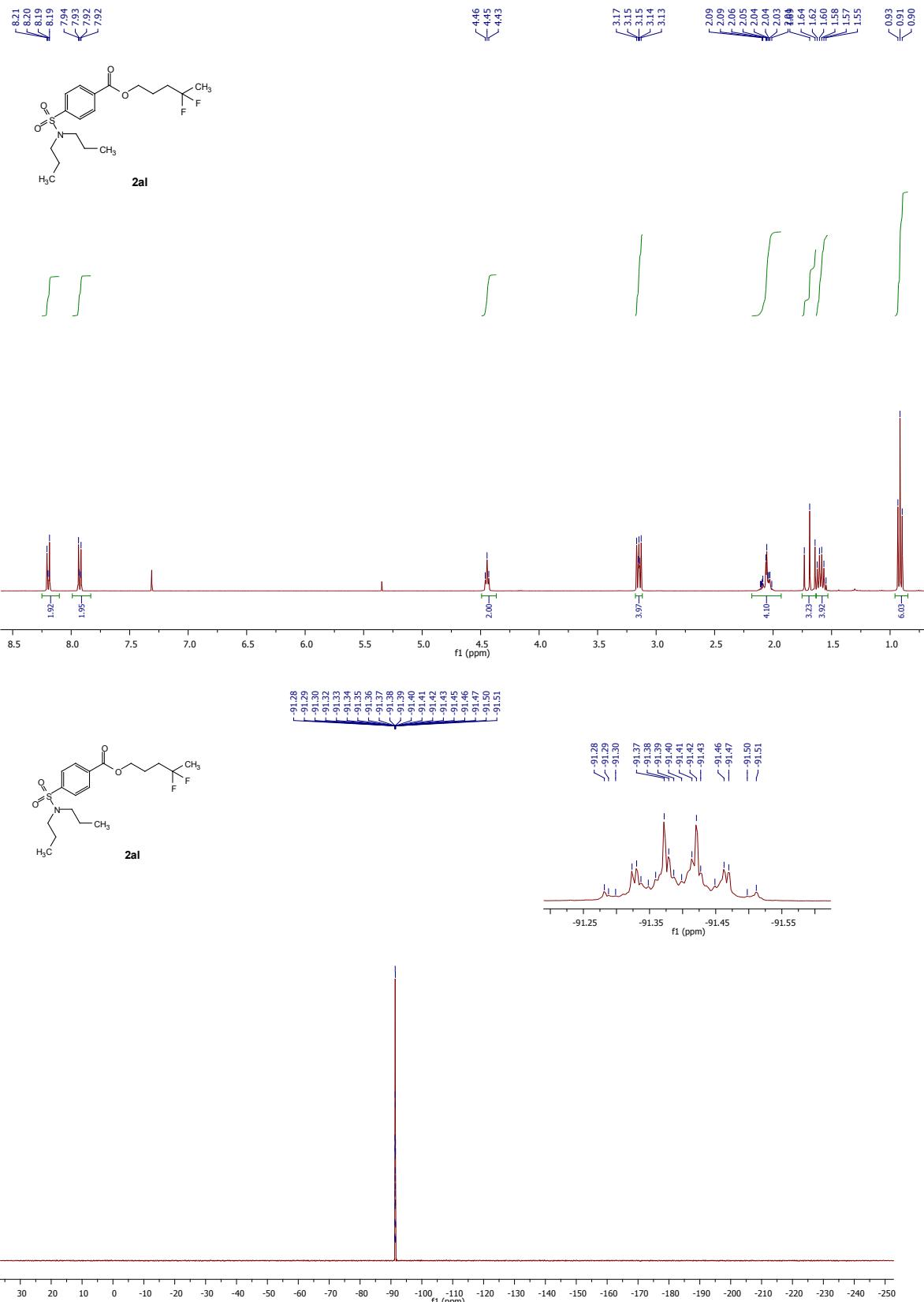


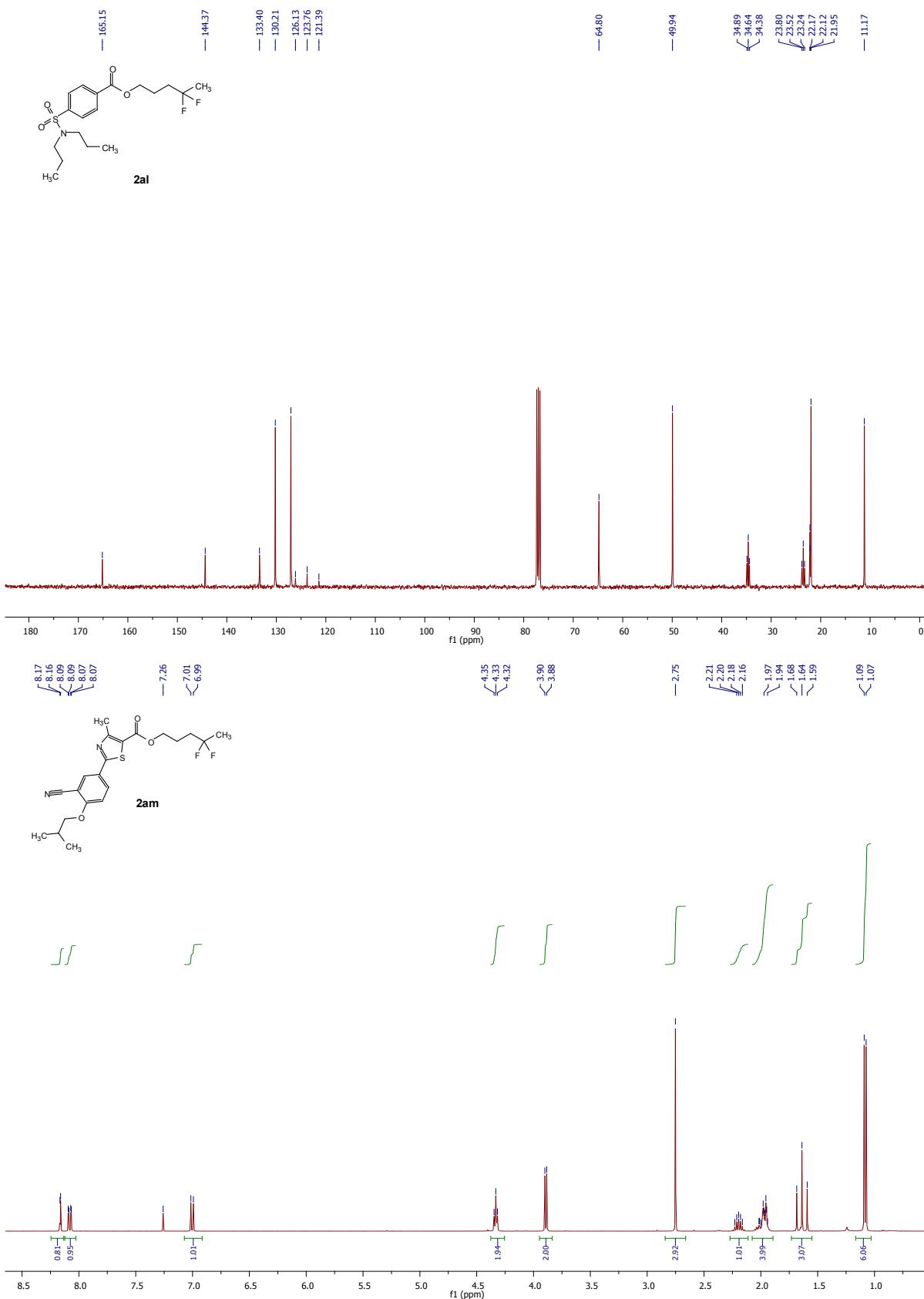


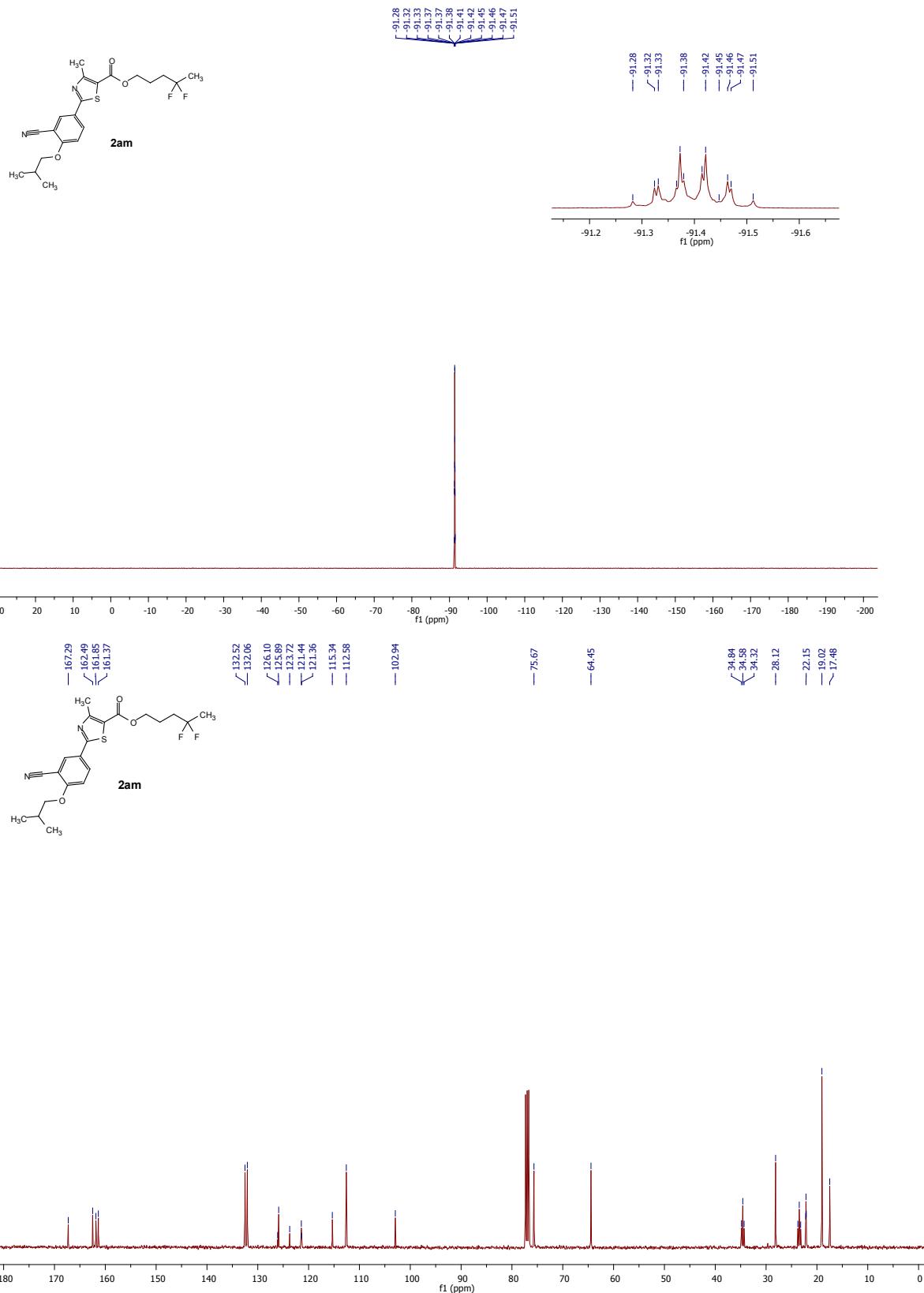


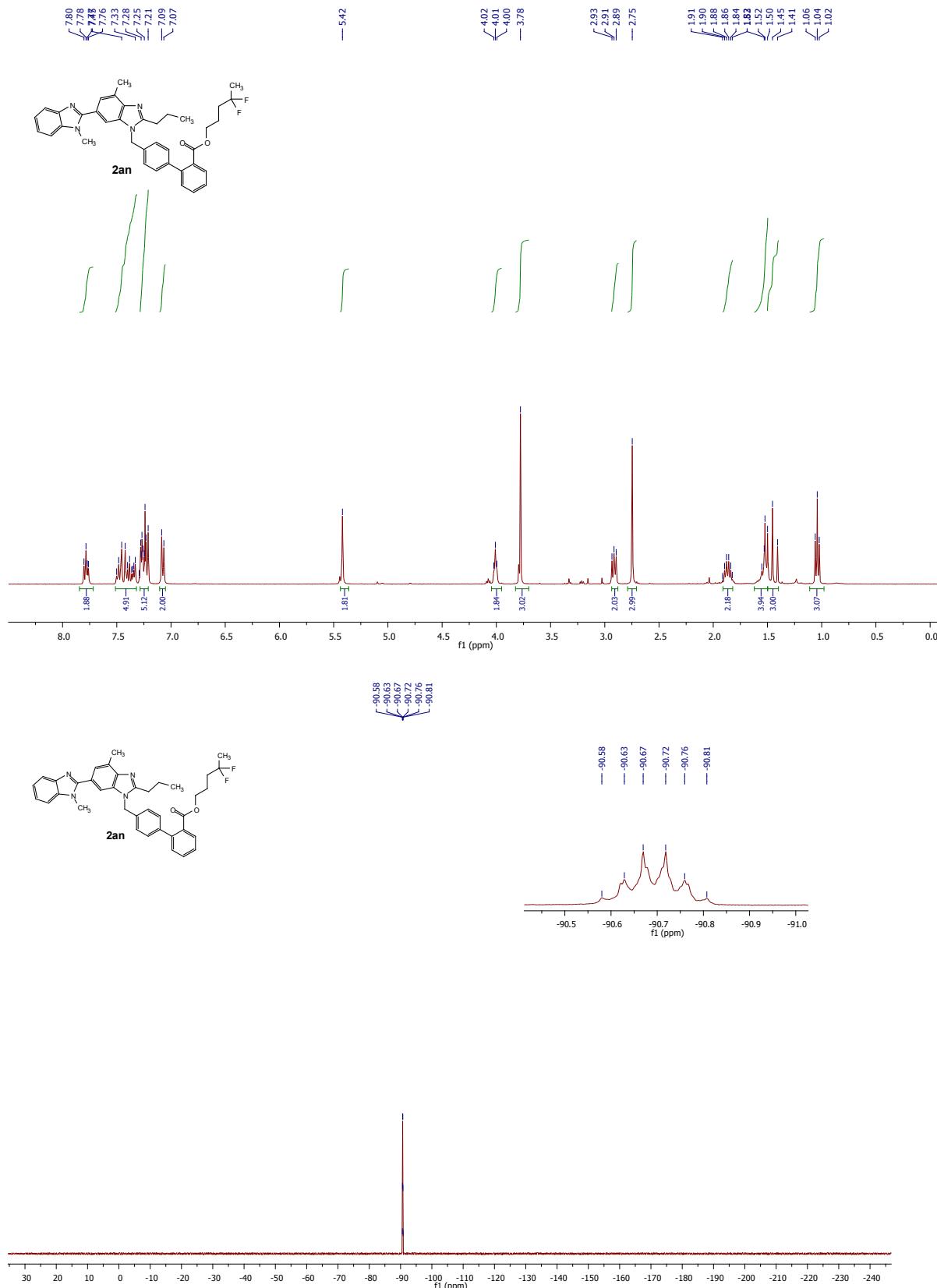


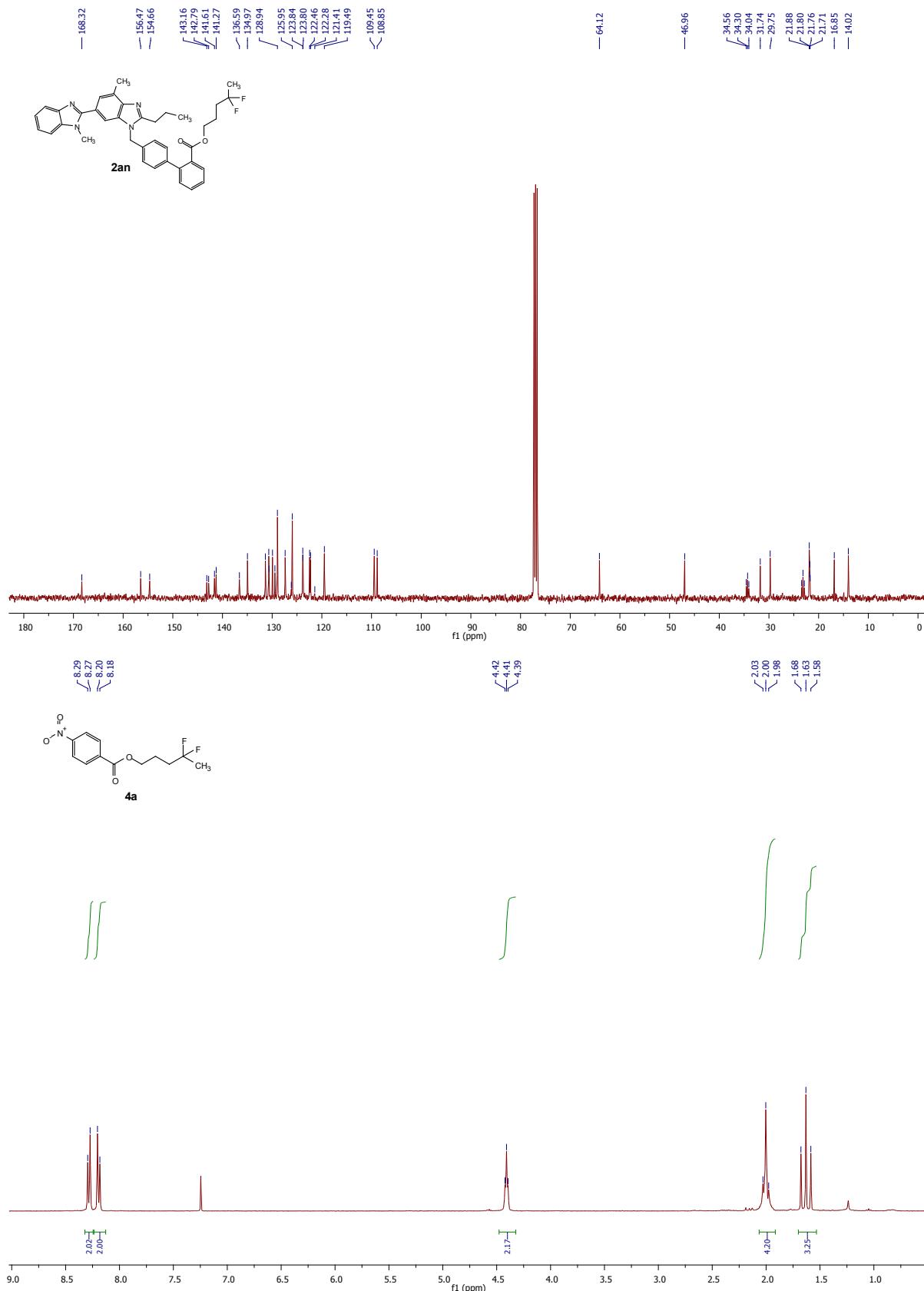




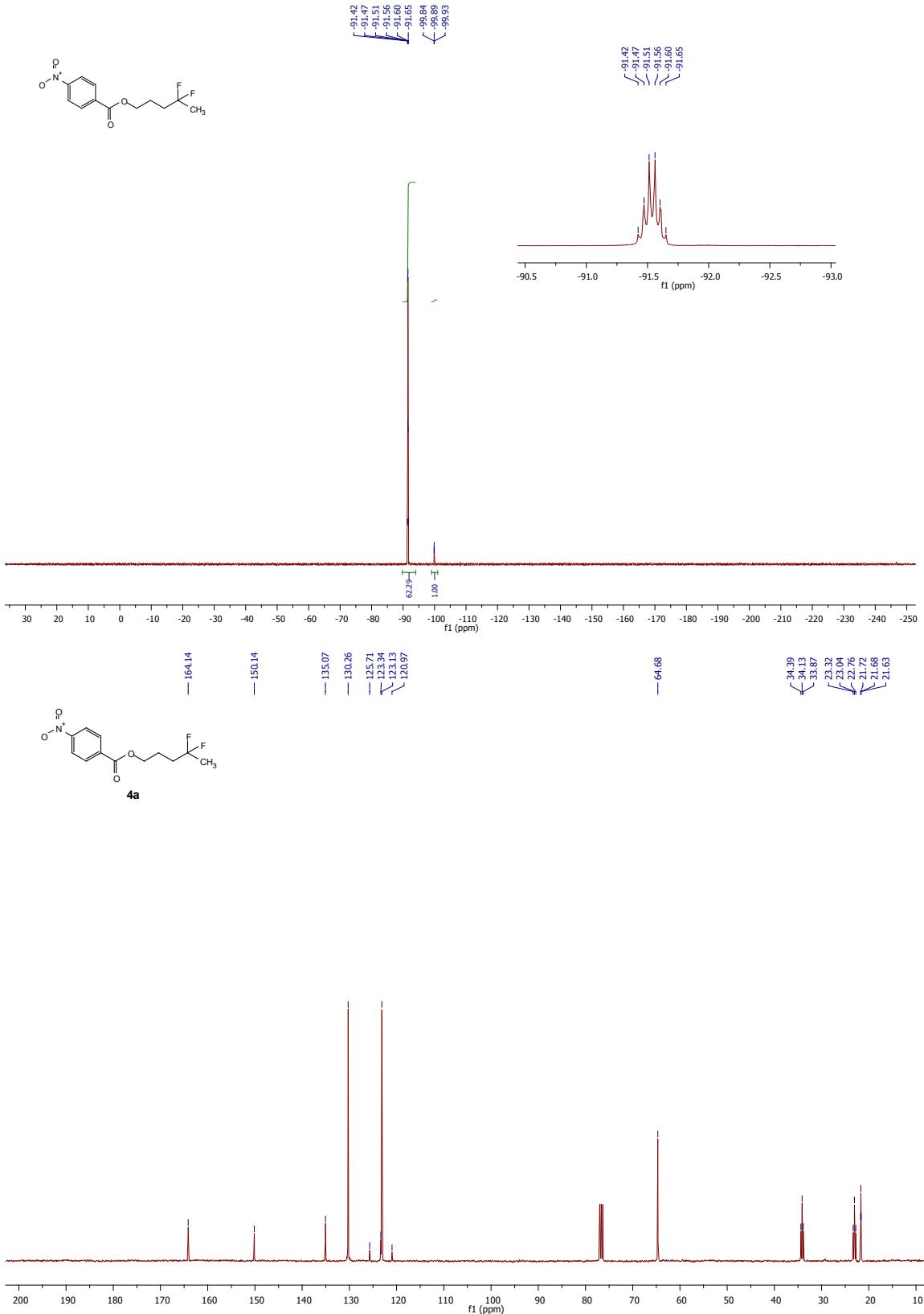


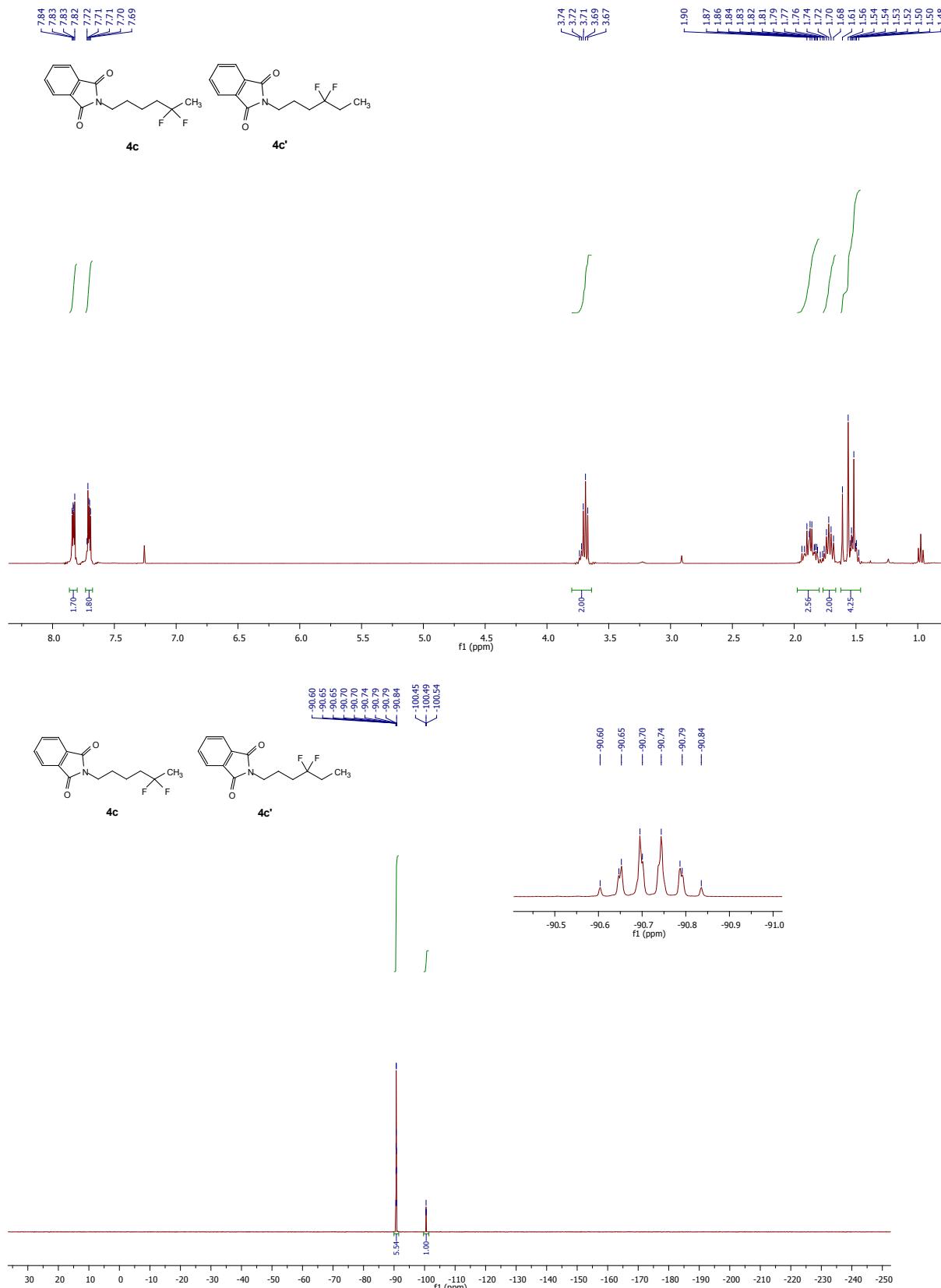


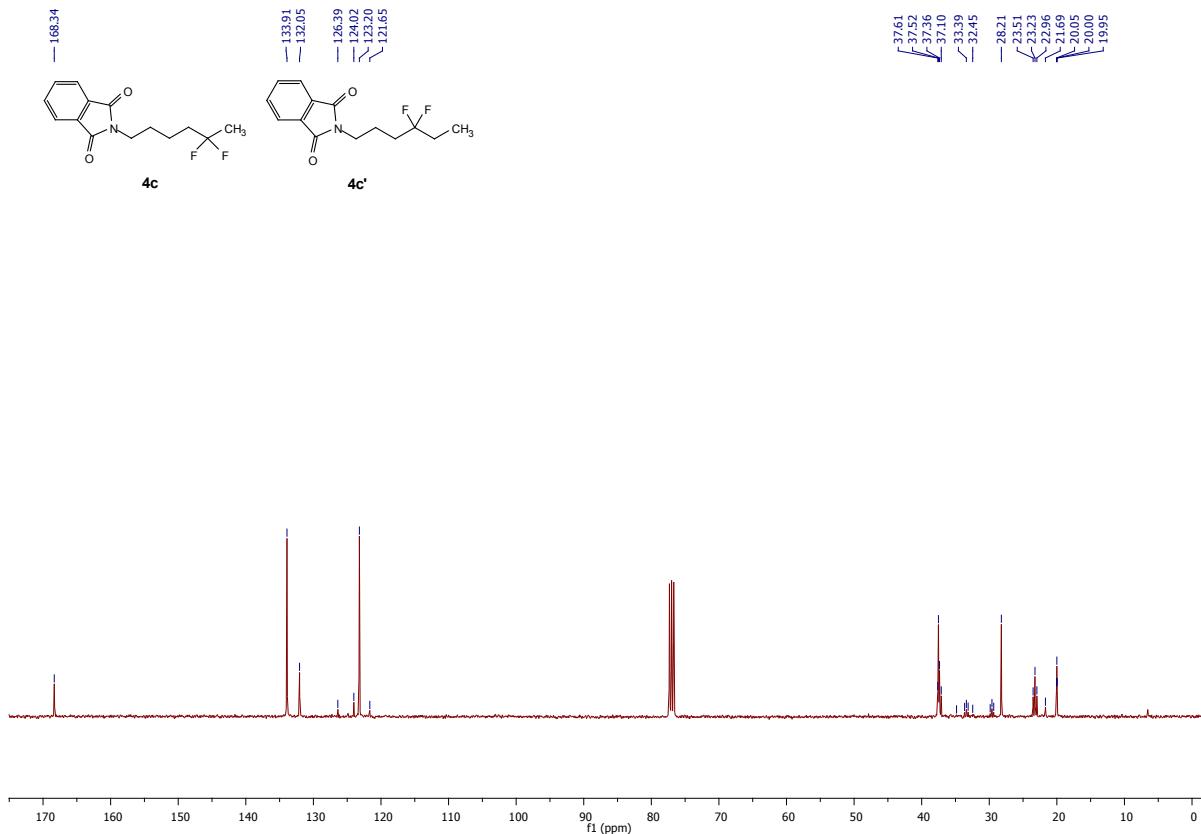




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Section 5. References

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