## **Supplementary Information**

# Streamlining routine organic chemistry reactions by the employment of high shear mixers.

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#### **General Information**

#### NMR

Nuclear Magnetic Resonance spectra of <sup>1</sup>H and <sup>13</sup>C were recorded in a Bruker Avance spectrometer (500 MHz). NMR spectra were obtained in deuterated chloroform (CDCl<sub>3</sub>. The chemical shifts are expressed in ppm ( $\delta$ ), referenced to internal CHCl<sub>3</sub> ( $\delta$  7.26), <sup>13</sup>CDCl<sub>3</sub> ( $\delta$  77.16). and the coupling constants (J) are expressed in Hz.

#### HSM

High Shear Mixing experiments were carried out using an IKA ULTRA-TURRAX<sup>®</sup> T 18 digital apparatus at 6000, 8000 and 10000 rpm.

#### SYNTHESIS OF AZIDOSTEROIDS

For the synthesis of azidosteroids, sodium azide (1.5 eq) was added to a solution of 26substituted-22-oxocholestanes **1a** or **1b** (500 mg, 0.8 mmol) in DMF (6 mL). The reaction mixture was stirred using a magnetic stirrer (300 rpm) or the HSM (6000, and 8000 rpm) at 25 °C and 65 °C. DMF was removed from aliquots of the reaction mixtures, and dried samples were analyzed by <sup>1</sup>H-NMR

#### SYNTHESIS OF PENTA AND TETRAACETYLATED GLUCOSE

Glucose (2 g, 11.1 mmol) was suspended in 5 ml of  $CH_2Cl_2$  and 5.8 ml of  $Ac_2O$  (5.5 eq); then 5% mol of DMAP was added and stirred at 300, 8000, and 10000 rpm. Reactions were followed by <sup>1</sup>H-NMR. After complete conversion, the organic phase was extracted with  $CH_2Cl_2$  and the extract was washed with a saturated solution of NaHCO<sub>3</sub> (5 x 100 mL), water (3 x 100 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, and concentrated under vacuum. The pentaacetylated glucose (4) was obtained quantitatively, as a syrup. For a selective deprotection reaction, the peracetylated glucose 4 (2 g, 5.12 mmol) was dissolved in 10 ml DMF, and 1.5 eq of hydrazine acetate were added. The reaction mixture was stirred at 300, 8000, and 10000 rpm. The organic phase was extracted with AcOEt (100 mL) and washed with saturated NaHCO<sub>3</sub> solution (3 x 100 mL), water (3 x 100 mL), dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and concentrated under reduced pressure to obtain a syrup of the tetraacetylated glucose (5).



<sup>1</sup>H NMR Spectrum of (25*R*)-26-bromo-22-oxocholest-5-ene-3β,16β-diyl diacetate (**1a**, 500 MHz, CDCl<sub>3</sub>).



<sup>13</sup>C NMR Spectrum of (25*R*)-26-bromo-22-oxocholest-5-ene-3β,16β-diyl diacetate (**1a**, 125 MHz, CDCl<sub>3</sub>).



<sup>1</sup>H Spectrum of (25*R*)-26-iodo-22-oxocholest-5-ene-3β,16β-diyl diacetate (**1b**, 500 MHz, CDCl<sub>3</sub>).



<sup>13</sup>C NMR Spectrum of (25*R*)-26-iodo-22-oxocholest-5-ene-3β,16β-diyl diacetate (**1b**, 125 MHz, CDCl<sub>3</sub>).



<sup>1</sup>H NMR Spectrum of (25*R*)-26-azido-22-oxocholest-5-ene-3β,16β-diyl diacetate (**2**, 500 MHz, CDCl<sub>3</sub>).



<sup>13</sup>C NMR Spectrum of (25*R*)-26-azido-22-oxocholest-5-ene-3β,16β-diyl diacetate (**2**, 125 MHz, CDCl<sub>3</sub>).



COSY 2D NMR experiment of (25*R*)-26-azido-22-oxocholest-5-ene-3β,16β-diyl diacetate (**2**, CDCl<sub>3</sub>).



HSQC 2D NMR experiment of (25*R*)-26-azido-22-oxocholest-5-ene-3β,16β-diyl diacetate (2, CDCl<sub>3</sub>).



HMBC 2D NMR experiment of (25*R*)-26-azido-22-oxocholest-5-ene-3β,16β-diyl diacetate (**2**, CDCl<sub>3</sub>).



<sup>1</sup>H NMR Spectrum of  $\xi$ -D-glucopyranose pentaacetate (4, 500 MHz, CDCl<sub>3</sub>).



Spectrum of ξ-D-glucopyranose pentaacetate (4, 125 MHz, CDCl<sub>3</sub>).



COSY 2D NMR experiment of ξ-D-glucopyranose pentaacetate (4, CDCl<sub>3</sub>).



HSQC 2D NMR experiment of  $\xi$ -D-glucopyranose pentaacetate (4, CDCl<sub>3</sub>).



<sup>1</sup>H NMR Spectrum of ξ-D-glucopyranose tetraacetate (5, 500 MHz, CDCl<sub>3</sub>).



<sup>13</sup>C NMR Spectrum of ξ-D-glucopyranose tretraacetate (4, 125 MHz, CDCl<sub>3</sub>).



COSY 2D NMR experiment of ξ-D-glucopyranose tetraaacetate (4, CDCl<sub>3</sub>).



HSQC 2D NMR experiment of ξ-D-glucopyranose tetraaacetate (4, CDCl<sub>3</sub>).