Supplementary Information

Enhancement of radionuclide bio-decontamination by screening highly efficient microalgae for Sr biomineralization

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Figure S1. Fabrication process of the proposed microalgae screening microplatform: (I) Electrode array,

(II) PDMS microfluidic chip and (III) assembly of microalgae screening microplatform.



Figure S2. Experimental setup for microalgae screening, including function generator, syringe pump, microalgae screening microplatform, optical microscope, CCD camera and image acquisition system.



Figure S3. Microscopic images of microalgae: (a) *C. vulgaris KMMCC9* and (b) *C. vulgaris KCTC AG10002* in 3 mM NaHCO₃. The scale bar in the images is 10 μ m. (c) A phylogenetic tree based on 18s rRNA sequences of several *C. vulgaris* strain, *C. sp.* strain, *C. sorokiniana, C. pituita, C. variabilis*, and *C. parva* using multiple sequence alignment ClustalW program. Note that the two strains (marked by *) are the outgroups of each other, however, close enough to perform further tests.



Figure S4. Histochemical method for detection of non-radioactive Sr absorbed in *C. vulgaris KMMCC9* after several hours' treatment of media containing non-radioactive Sr.



Figure S5. The viability of the microalgae in the assay buffer (i.e., NaHCO₃ solution) during DEP-based screening.

(a) p-DEP plates

(b) n-DEP plates



Figure S6. Colonies on the plates with (a) p-DEP group and (b) n-DEP group. The colonies numbers of *C. vulgaris KMMCC9* and *C. vulgaris KCTC AG10002* in p-DEP or n-DEP group were confirmed by colony counting. Note that the smaller and greener are *C. vulgaris KMMCC9* and the larger and light greens are *C. vulgaris KCTC AG10002*, which were confirmed by 18s rRNA sequencing.