Supporting Information

An integrated multi-molecular sensor for simultaneous BRAFV600E protein and DNA single point mutation detection in circulating tumour cells

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Table S1. Oligonucleotide sequences used in experiments. *<u>Underlined and bold</u> representRPA primer sequences on ligation probes.

| Oligos | 5'-Sequence-3' |
|------------------------------------|---|
| BRAF ^{V600E} Fwd Ligation | -PO- |
| Probe | AGACATCGATCTGGACGAGGGAAAGAGTTGTACCTAAAT |
| BRAF ^{V600E} Rev Ligation | TGTATAGGAATCCCACTGAATTTTTCCCATCGAGATTTC |
| Probe | |
| BRAF ^{V600E} RPA Fwd | SH-C3-ATTTAGGTACAACTCTTTCCCTCGTC |
| Primer | |
| BRAF ^{V600E} RPA Rev | TGTATAGGAATCCCACTGAATTTTTC |
| Primer | |
| BRAF ^{V600E} PCR Fwd | ATAGGTGATTTTGGTCTAGCTACAGA |
| Primer | |
| BRAF ^{V600E} PCR Rev | AGTAACTCAGCAGCATCTCAGG |
| Primer | |



Fig. S1. Electrode surface functionalization for $BRAF^{V600E}$ protein detection. Each overlaid differential pulse voltammetric signal represented a stepwise molecular functionalization to prepare the eventual protein sensing surface.



Fig. S2. (a) Capture and (b) release efficiencies of different SK-MEL-28 cell counts in 1 mM PBS. (insets) Corresponding actual number of captured and released cells.



Fig. S3. (a) Capture efficiencies of 100 SK-MEL-28 cells spiked in 10x diluted blood in 1 mM PBS under ac-EHD (left bar, blue) and hydrodynamic fluid flow conditions (right bar, grey), n=3. (b) DC induced cell release performance for cuaptured cells under ac-EHD (right, blue bar) and under hydrodynamic fluid flow (right, grey bar) conditons. Error bars represent the standard error of three repeats (n=3).