Electronic Supplementary Material (ESI) for MedChemComm. This journal is © The Royal Society of Chemistry 2018

Supporting Information

Copper(II) complexes based on quinoline derived Schiff-base ligands: Synthesis,

characterization, HSA/DNA binding ability, and anticancer activity

Kun Hu*a, Chensi Liua, Jingui Lia, Fupei Liang*a,b

- ^aState Key Laboratory Cultivation Base for Chemistry and Molecular Engineering of Medicinal Resources, School of Chemistry and Pharmacy, Guangxi Normal University, 15 Yucai Road, Guilin 541004, P. R. China., Fax, 086-773-2120958; E-mail: fliangoffice@yahoo.com.
- ^bGuangxi Key Laboratory of Electrochemical and Magnetochemical Functional Materials, College of Chemistry and Bioengineering, Guilin University of Technology, Guilin 541004, China.
- *Corresponding authors: <u>hukun216@163.com</u> (Kun Hu); <u>fliangoffice@yahoo.com</u> (Fupei Liang).

Contents:

Complex	C1	C2	C3
Empirical formula	$C_{18}H_{14}CuN_4O_8$	$C_{19}H_{16}Cl_2CuN_2O_2$	$C_{19}H_{16}CuN_2O_7S$
Formula weight	477.87	438.78	479.94
Crystal system	Triclinic	Monoclinic	Triclinic
Space group	<i>P</i> -1	$P2_{1}/n$	<i>P</i> -1
<i>a</i> (Å)	10.093(3)	9.784 (8)	8.818 (5)
<i>b</i> (Å)	10.384(3)	12.520 (11)	9.995 (6)
<i>c</i> (Å)	10.818(3)	15.403 (13)	13.151 (7)
α (°)	78.634(4)	90.00	99.756 (9)
eta (°)	65.151(4)	101.907 (13)	104.779 (8)
γ (°)	71.085(4)	90.00	103.630 (8)
Volume (Å ³)	970.8(5)	1846 (3)	1056.2 (10)
Ζ	2	4	2
Calculated density(Mg/m ³)	1.635	1.579	1.459
<i>F</i> (000)	486	892	475
θ range for data collection (°)	2.1-26.5	2.1-26.5	1.7-25.4
Reflections collected/unique	11785/4023	21771/3793	8547/3853
Goodness-of-fit on F^2	1.045	1.047	1.057
R _{int}	0.025	0.036	0.047
Final <i>R</i> indices $[I > 2\sigma(I)]$	0.0332	0.0254	0.0680
R indices (all data)	0.0414	0.0321	0.0910

 Table S1 Crystal data and structure refinement for complexes C1-C3.

Table S2 Selected bond lengths (Å) and bond angles (°) for complexes C1-C3.

C1					
Cu1–O5	2.0128(18)	Cu1-N1	1.984(2)		
Cu1-O6	1.9820(19)	Cu1-N2	1.9448(19)		
O6-Cu1-O8	57.05(7)	N2-Cu1-O5	159.13(8)		
O5-Cu1-O6	85.01(9)	N2-Cu1-O8	110.91(10)		
O5-Cu1-O8	85.44(10)	N2-Cu1-N1	94.96(8)		
O5-Cu1-N1	94.89(8)	N1-Cu1-O6	156.73(8)		
N2-Cu1-O6	93.05(8)	N1-Cu1-O8	99.71(8)		
C2					
Cu1-Cl1	2.2272 (19)	Cu1-N1	1.987 (2)		
Cu1-Cl2	2.2255 (15)	Cu1-N2	1.9855 (19)		
Cl2-Cu1-Cl1	101.64 (3)	N2-Cu1-Cl1	133.52 (5)		
N1-Cu1-Cl1	100.30 (6)	N2-Cu1-Cl2	96.67 (7)		
N1-Cu1-Cl2	138.00 (6)	N2-Cu1-N1	93.51 (7)		
C3 (Symmetry code: (i) $-x + 1, -y + 2, -z + 2$.)					
Cu1-O1	2.009 (4)	Cu1–O4i	2.223 (4)		
Cu1-O2	1.989 (4)	Cu1-N1	1.958 (5)		
Cu1-N2	1.979 (5)	N1-Cu1-O2	96.09 (17)		
O1–Cu1–O4i	99.69 (16)	N1-Cu1-O4i	97.82 (17)		
O2-Cu1-O1	70.86 (16)	N1-Cu1-N2	94.9 (2)		
O2-Cu1-O4i	104.74 (17)	N2-Cu1-O1	93.77 (18)		
N1-Cu1-O1	160.35 (18)	N2-Cu1-O2	159.58 (19)		
N2-Cu1-O4i	90.75 (19)	N1-Cu1-O2	96.09 (17)		

Fig. S1 FT-IR (KBr) spectra of ligand L1.



Fig. S2 The HRMS (ESI) spectra of ligand L1 in the methanol solution.







Fig. S4 FT-IR (KBr) spectra of ligand L2.



Fig. S5 The HRMS (ESI) spectra of ligand L2 in the methanol solution.







Fig. S7 FT-IR (KBr) spectra of complex C1.



Fig. S8 The HRMS (ESI) spectra of complex C1 in the methanol solution.



Fig. S9 FT-IR (KBr) spectra of complex C2.



Fig. S10 The HRMS (ESI) spectra of complex C2 in the methanol solution.



Fig.S11 FT-IR (KBr) spectra of complex C3.



Fig. S12 The HRMS (ESI) spectra of complex C3 in the methanol solution.









Fig. S14 Emission spectra of HSA (1 μ M; λ ex=280 nm) in the presence increasing concentrations of the ligand L1 (A) and L2 (B) (0, 1.0, 2.0, 3.0, 4.0 and 5.0 μ M).



Fig.S15 Stern-Volmer (A) and (B) Hill plots for HSA interacting with the test compounds at 295 K. Conditions: HSA= 1.0μ M; complexes: 0, 1.0, 2.0, 3.0, 4.0 and 5.0 μ M.



Fig. S16 UV-Vis absorption spectra of HSA (2.5 μ M) in the presence of ligand L2 and complex C3 (2.5 μ M).



Fig. S17 Synchronous fluorescence spectra of HSA (1.0 μM, black line) in presence of increasing amounts of complex C1 (A, B), C2 (C, D) and C3 (E, F) (0-5.0 μM) with a wavelength difference of Δλ=15 nm (A, C1; C, C2, and E, C3) and Δλ=60 nm (B, C1; D, C2, and F, C3).







Fig. S18 The high-performance liquid chromatographic (HPLC) spectra of HSA and HSA-C3 mixture. Condition: HSA 2.5 μ M; C3, L2 2.5 μ M; Separation was carried out on a Sinochrom ODS-AP column (150 mm × 4.6 mm id, 300 Å pore size, 5 μ m particle sizes; Elite, Dalian, China). The absorbance of the effluent was detected at 280 nm. The flow rate was set to 0.5 mL/min with isocratic elution. Mobile phase A: ACN containing 0.1% formic acid; mobile phase B: water containing 0.1% formic acid.



Fig. S19 Fluorescence quenching spectra of DNA-EB (10 μM) in the presence of ligand L1 (A) and L2 (B) (0-30 μM).







Fig. S21 The fluorescence excitation spectra of complex C1-C3 (30 μ M).



Fig. S22 Overall structure of the HSA.



Fig. S23 Molecular docked structure of the complexes interacting with HSA (PDB ID: 1E7H). A) **C1** and B) **C2**.



Fig. S24 Molecular docked model of the complex with DNA (PDB ID: 1BNA). A) C1 and B) C2.



