

Supplementary Information

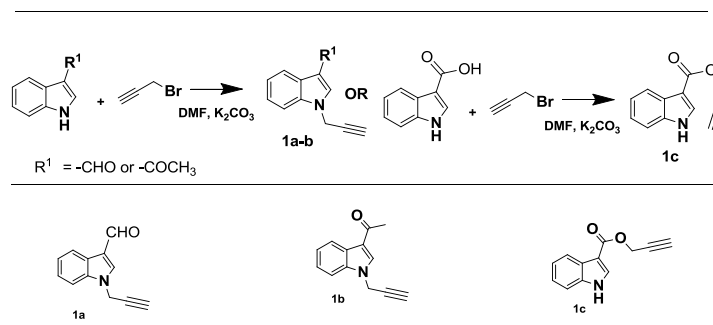
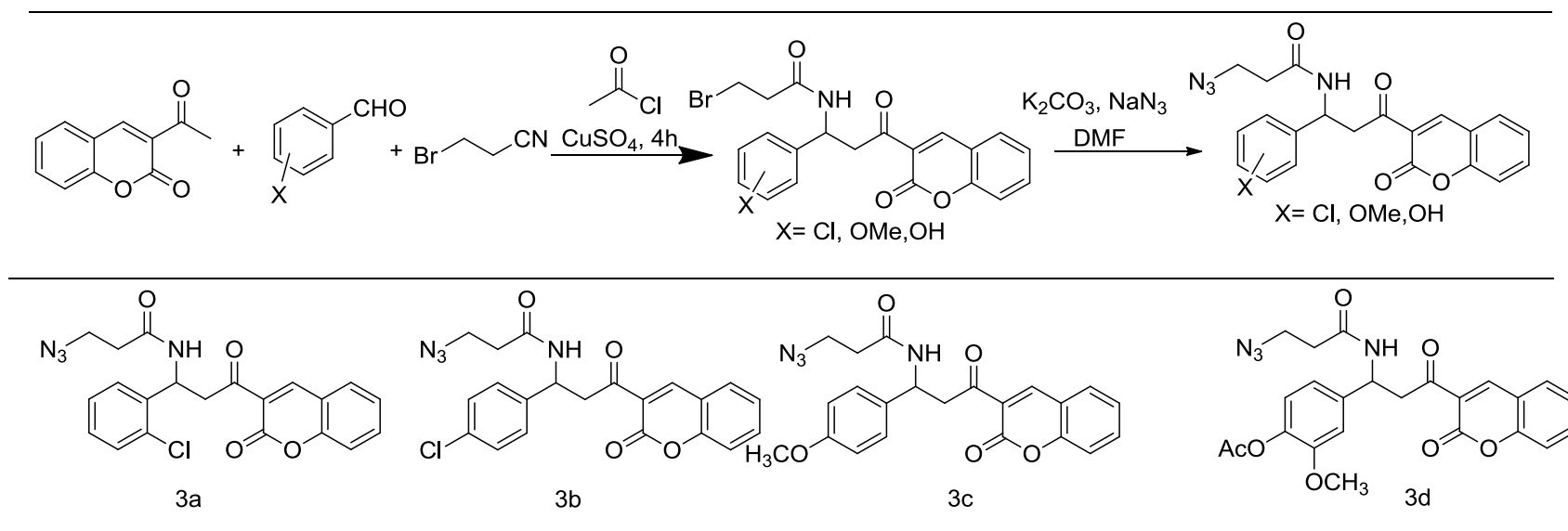
MCR-Click Synthesis, Molecular docking and Cytotoxicity evaluation of a new series of Indole-Triazole-Coumarin hybrid Peptidomimetics

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Table 1.1. List of alkyne functionalized indole scaffolds synthesized.**Table 1.2.** List of Coumarin azides 3a-3d synthesized via alternate Mannich reaction.

Experimental section

2.1. General

All reactions were carried out with oven-dried glassware. Starting materials were purchased from Aldrich and Merck. Alkynes and azides were synthesized based on literature procedure. High resolution mass spectra were measured with DMSO as solvent. IR spectra were recorded on a JASCO-FTIR-4100 spectrometer. ¹H and ¹³C NMR spectra were recorded on a Bruker FT-400 & 100MHz, using TMS as an internal standard.

2.2. General procedure for the synthesis of indole functionalized alkynes **1a-1c**.

Indole derivative (1mmol) and potassium carbonate (414.63mg, 3mmol) were dissolved in minimum amount of DMF, and stirred at 50°C for 10 min. After cooling the reaction mixture to room temperature, propargyl bromide (118.96mg, 1mmol) was added to this and stirred for 4 h and then poured into ice cold water. The solid product predicated was filtered and dried under vacuum to obtain indole functionalized alkynes **1a-1c**.

1-(prop-2-yn-1-yl)-1H indole-3-carbaldehyde (1a)

White solid ; 159 mg (yield 87%); Mp: 90-93°C ; ¹H NMR (DMSO-d₆ , 400 MHz) δH (ppm): 9.94 (s, 1H); 8.37-8.35(d, J=8Hz, 1H); 8.15-8.12 (m, 1H); 7.66-7.64 (m, 1H); 7.38 - 7.28 (m, 2H); 5.24 (s, 2H); 3.43 (s, 1H); ¹³C NMR (DMSO-d₆, 100 MHz) δC(ppm): 185.4, 140.56, 137.05, 125.17, 124.23, 123.43, 121.78, 118.34, 111.85, 78.43, 77.20, 40.45; ; IR (KBr) ν max : 3198, 2121, 1656, 1644, 1615, 1579, 1528, 1471, 1441, 1308, 1244, 1163, 1041, 770, 741, 562 cm⁻¹; MS (ESI): m/z = 184 (M + 1).

1-(1-(prop-2-yn-1-yl)-1H-indol-3-yl) ethanone (1b)

White solid ; 167mg (yield 85%); Mp: 103-105°C; ¹H NMR (DMSO-d₆, 500 MHz) δH(ppm): 8.40(s, 1H), 7.91(s, 1H), 7.35-7.32(m, 1H), 7.26 (s, 2H), 4.92 (s, 2H), 2.17 (s, 1H), 1.56(s, 3H); ¹³CNMR(DMSOd₆, 125MHz); δC(ppm): 195.40, 140.56, 137.05, 125.17, 124.23,

123.28,121.59,118.05,111.64,78.43,77.20,40.45,25.1; IR (KBr) ν max : 3218, 2125, 1676, 1628, 1578,1523, 1388, 1211, 932, 762, 747 cm⁻¹; MS(HRMS): m/z = 221.0739 (M+Na).

Prop-2-yn-1-yl 1H-indole-3- carboxylate (1c)

White solid ; 163mg (yield 82%);Mp: 80-82°C; ¹H NMR (DMSO-d₆ , 400 MHz) δ H (ppm): 12.00 (s, 1H), 8.05-8.00 (m, 2H), 7.66-7.64 (d, J=8Hz, 1H), 7.35-7.27 (m, 2H), 5.23 (s, 2H), 3.56 (s,1H);¹³CNMR (DMSO-d₆,100MHz) δ C(ppm): 163.83,136.87,126.60,122.98,121.91,121.12, 120.78,112.91,111.71,79.69,76.96,51.48 ; IR (KBr) ν max :3298, 3282, 2120,1698, 1698, 1619, 1539, 1271, 1245, 1183, 1149, 1095, 916, 774, 749, 626cm⁻¹;MS(ESI): m/z = 222 (M +Na).

2.3. General procedure for the synthesis of coumarin functionalized azides3a-3b**.**

A mixture of corresponding benzaldehyde (1 mmol), 3-acetylcoumarin (188 mg, 1mmol), and 3-bromopropionitrile (133 mg, 1 mmol) in acetonitrile (8 ml) were stirred in the presence of catalytic amount of CuSO₄ at room temperature for 4 h. Subsequently the reaction mixture was poured into ice cold water and extracted with CH₂Cl₂ (15 mLx2). Evaporation of the solvent followed by purification on a silica gel column (100–200 mesh), by eluting with ethyl acetate/hexane (3:1) afforded the corresponding β -amidoketonebromide derivative **2**. **2** (1 mmol), K₂CO₃ (414 mg, 3 mmol), NaN₃ (65 mg, 1 mmol) were dissolved in dimethylacetamide and stirred for 6–8 h and then poured into ice cold water. The precipitate was filtered and dried under vacuum to afford the azides**3a-3c**.

3-bromo-N-(1-(4-chlorophenyl)-3-oxo-3-(2-oxo-2H-Chromen-3-yl)propyl) propanamide (2b)

¹H NMR (500 MHz, DMSO-d₆): δ H (ppm): 8.588 (s, 1H), 8.171 (s, 1H), 7.944-7.929 (d, J = 7.5Hz, 1H), 7.776-7.742 (t, 1H), 7.526-7.482 (m, 4H), 7.302-7.284 (t, 1H), 6.795-6.780 (d, 2H), 5.179-5.138 (t, 1H), 3.891-3.844 (dd, J = 8Hz, J = 16Hz, 1H), 3.452-3.404 (dd, J = 8Hz and J=20Hz, 1H), 3.190-3.176 (t, 2H), 2.364-2.308 (t, 2H);¹³C NMR (100 MHz, DMSO-d₆) δ C(ppm): 199.79, 175.35, 159.92, 153.63, 143.68, 139.07, 134.91, 129.76, 128.92, 125.46, 123.82, 121.88, 116.13, 113.20, 50.16, 47.53, 38.23, 29.45.;FT-IR

(KBr) $\gamma_{\max}$: 3423.03, 3292.86, 2923.56, 2852.20, 1725.98, 1676.80, 1608.42, 1560.13, 1527.35, 1455.03, 1349.93, 1225.54, 1174.44, 1108.87, 1024.02, 759.82 cm^{-1}

3-bromo-N-(1-(4-methoxyphenyl)-3-oxo-3-(2-oxo-2H-Chromen-3-yl)propyl) propanamide(2c)

^1H NMR (500 MHz, $\text{DMSO}-d_6$): δH (ppm): 8.31 (s, 1H), 8.04 (s, 1H), 7.78-7.76 (d, $J = 8\text{Hz}$, 1H), 7.50-7.49 (t, 1H), 7.44-7.41 (t, 1H), 7.01-7.08 (d, $J=8\text{Hz}$, 2H), 6.79-6.78 (d, $J=7.5\text{Hz}$, 2H), 5.23-5.21 (t, 1H), 3.98 (s, 3H), 3.78-3.74 (dd, $J = 8\text{Hz}$, $J = 16\text{Hz}$, 1H) 3.49-3.44 (dd, $J = 8\text{Hz}$ and $J=20\text{Hz}$, 1H), 3.41-3.40 (t, 2H), 2.40-2.39 (t, 2H) ; ^{13}C NMR (100 MHz, $\text{DMSO}-d_6$) δC (ppm): 200.59, 174.05, 160.86, 156.06, 153.63, 139.77, 134.28, 129.76, 128.98, 125.75, 123.72, 121.88, 118.23, 114.23, 109.20, 56.28, 51.06, 47.69, 39.57, 29.08; FT-IR (KBr) $\gamma_{\max}$: 3421.10, 3068.19, 2923.56, 2850.27, 1725.98, 1681.62, 1606.41, 1580.38, 1561.09, 1510.95, 1455.03, 1422.24, 1364.39, 1303.64, 1251.58, 1172.51, 1119.48, 1028.84, 922.77, 831.16 cm^{-1} ; MS(ESI): $m/z = 459.3$ (M +1).

3-bromo-N-(1-(4-hydroxy-3-methoxyphenyl)-3-oxo-3-(2-oxo-2H-Chromen-3-yl)propyl) propanamide (2d)

Yellowish solid ; 396.46mg (yield 86%); ^1H NMR (500 MHz, $\text{DMSO}-d_6$): δH (ppm): 8.29 (s, 1H), 8.075 (s, 1H), 7.819-7.803 (d, $J = 8\text{Hz}$, 1H), 7.592-7.578 (t, 1H), 7.503-7.488 (t, 1H), 7.156-7.140 (d, $J=8\text{Hz}$, 2H), 6.894-6.879 (d, $J=7.5\text{Hz}$, 2H), 5.125-5.112 (t, 1H), 3.969 (s, 3H), 3.799-3.755 (dd, $J = 8\text{Hz}$, $J = 16\text{Hz}$, 1H) 3.497-3.449 (dd, $J = 8\text{Hz}$ and $J=20\text{Hz}$, 1H), 3.273-3.255 (t, 2H), 2.214-2.204 (t, 2H), 2.176 (s, 3H) ; ^{13}C NMR (100 MHz, $\text{DMSO}-d_6$) δC (ppm): 199.59, 172.77, 168.06, 159.02, 155.75, 150.82, 142.77, 139.04, 134.91, 129.96, 128.98, 125.05, 122.72, 120.88, 119.23, 114.20, 110.29, 106.22, 56.16, 52.67, 49.61, 47.62, 37.66, 29.96; FT-IR (KBr) $\gamma_{\max}$: 3429.78, 2923.56, 2853.17, 1760.69, 1724.05, 1677.77, 1607.38, 1560.13, 1509.99, 1489.74, 1455.99, 1422.24, 1370.18, 1269.9, 1199.51, 1123.33, 1083.8, 1031.73, 914.093, 862.989 cm^{-1}

3-azido-N-(1-(4-chlorophenyl)-3-oxo-3-(2-oxo-2H-Chromen-3-yl)propyl) propanamide(3b)

Brown solid ; 348.37mg (yield 82%); ^1H NMR (500 MHz, $\text{DMSO}-d_6$): δH (ppm): 8.581 (s, 1H), 8.400 (s, 1H), 7.942-7.926 (d, $J = 8\text{Hz}$, 2H), 7.773-7.744 (t, 2H), 7.521-7.482 (m, 2H), 7.304-7.284 (t, 1H), 6.796-6.780 (d, $J=8\text{Hz}$, 2H), 5.149-5.138 (t, 1H), 3.601-3.554

(dd, J = 8Hz, J = 16Hz, 1H) 3.184-3.136 (dd, J = 8Hz and J=20Hz, 1H), 2.127-2.115 (t, 2H), 1.314-1.300 (t, 2H) ; ^{13}C NMR (100 MHz, DMSO- d_6) $\delta\text{C}(\text{ppm})$: 199.75, 175.35, 159.82, 153.66, 143.74, 139.77, 134.91, 129.76, 128.98, 125.75, 123.72, 121.88, 116.23, 113.20, 51.16, 47.69, 45.92, 34.26; FT-IR (KBr) γ_{max} : 3423, 2107, 1720, 1655, 1606, 1489, 1456, 1407, 1267, 1213, 1121, 1037, 857, 755, 712 cm^{-1} .

3-azido-N-(1-(4-methoxyphenyl)-3-oxo-3-(2-oxo-2H-Chromen-3-yl)propyl) propanamide (3c)

^1H NMR (500 MHz, DMSO- d_6): δH (ppm): 8.52 (s, 1H), 8.19 (s, 1H), 7.93-7.92 (d, J = 7Hz, 1H), 7.74-7.71 (t, 1H), 7.52-7.42 (m, 2H), 7.07-7.05 (d, J=8Hz, 2H), 6.49-6.48 (d, J = 8.5Hz, 1H), 5.08-4.94 (t, 1H), 3.91 (s, 3H), 3.62-3.57 (dd, J = 8Hz, J = 16Hz, 1H) 3.26-3.21 (dd, J = 8Hz and J=20Hz, 1H), 2.72-2.70 (t, 2H), 1.98-1.96 (t, 2H) ; ^{13}C NMR (100 MHz, DMSO- d_6) $\delta\text{C}(\text{ppm})$: 201.21, 172.06, 160.52, 156.86, 153.63, 140.74, 134.11, 131.51, 130.66, 128.25, 125.15, 121.85, 120.91, 116.27, 112.92, 56.33, 51.15, 47.23, 46.11, 33.52; FT-IR (KBr) γ_{max} : 3423.03, 3069.16, 2932.23, 2836.77, 2102.99, 1725.98, 1658.48, 1606.41, 1563.99, 1510.95, 1489.74, 1303.64, 1250.61, 1173.47, 1113.69, 1030.77, 985.447, 923.736, 854.311, 831.169 cm^{-1} ; MS(ESI): m/z = 420.3 (M).

3-azido-N-(1-(4-hydroxy-3-methoxyphenyl)-3-oxo-3-(2-oxo-2H-Chromen-3-yl)propyl)propanamide (3d)

Brownish solid ; 382.76mg (yield 80%); ^1H NMR (500 MHz, DMSO- d_6): δH (ppm): 8.57 (s, 1H), 8.39 (s, 1H), 7.94-7.92 (d, J = 8Hz, 1H), 7.76-7.73 (t, 1H), 7.59-7.49 (m, 2H), 6.97-6.96 (d, J = 7Hz, 1H), 5.14-4.95 (t, 1H), 3.73 (s, 3H), 3.63-3.58 (dd, J = 8Hz, J = 16Hz, 1H), 3.38-3.34 (dd, J = 8Hz and J=20Hz, 1H), 2.46-2.44 (t, 2H), 2.04(s, 3H), 1.68-1.66 (t, 2H) ; ^{13}C NMR (100 MHz, DMSO- d_6) $\delta\text{C}(\text{ppm})$: 200.59, 173.05, 169.06, 160.82, 156.66, 149.67, 139.77, 137.04, 134.91, 129.76, 128.98, 125.75, 123.72, 121.88, 119.23, 114.20, 110.09, 106.22, 56.16, 52.57, 48.69, 47.69, 35.66, 29.26; FT-IR(KBr) γ_{max} : 2936, 2834, 2108, 1729, 1607, 1510, 1455, 1415, 1384, 1270, 1202, 1122, 1030, 872, 757, 524 cm^{-1}

2.4. General procedure for the Cu (I) 1, 3-dipolar cycloaddition reaction for the formation of indole-triazole-coumarin.

An equimolar amount 1-(prop-2-yn-1-yl)-1H indole-3-carbaldehyde **1a** (91.5342 mg, 0.5 mmol) and the coumarinazide **3b** (239.0744 mg, 0.5 mmol) were dissolved in minimum amount of DMSO. To this, 2 ml of *t*-BuOH, 1 ml of water, CuSO₄·5H₂O (200 mg) and sodium ascorbate (150 mg) were added and stirred in room temperature for 12 h. and then poured in to cold water. The precipitated click product was filtered, washed with water and dried under vacuum to afford **4d** in pure form (555.79 mg, 84%).

N-(1-(4-chlorophenyl)-3-oxo-3-(2-oxo-2H-chromen-3-yl)propyl)-3-(4-((3-formyl-1H-indol-1-yl)methyl)-1H-1,2,3-triazol-1-yl)propanamide (**4b**)

Brownish solid ; 522.92 mg (yield 86%); ¹H NMR (400 MHz, DMSO-d₆): δH (ppm): 9.880(s, 1H), 8.412-8.361(d, J=20 Hz, 1H), 8.361-8.341(d, J=8 Hz, 1H), 8.300-8.279(d, J=8 Hz, 1H), 8.216(s, 1H), 8.120(s, 1H), 7.736-7.719(d, J= 7 Hz, 1H), 7.669(s, 1H), 7.638-7.176(m, 8H), 6.365-6.341(m, 1H), 6.223(s, 2H), 5.933-5.871(dd, J= 8Hz and J=16Hz, 1H), 5.483-5.433(dd, J= 8Hz and J= 20Hz, 1H), 5.061-5.018(m, 2H), 1.356-1.236(t, 2H); ¹³C NMR (100 MHz, DMSO-d₆) δC (ppm): 199.93, 185.42, 177.79, 166.54, 159.39, 141.11, 139.87, 137.08, 131.96, 129.58, 125.30, 124.30, 124.16, 123.33, 123.26, 123.11, 121.66, 121.58, 121.55, 117.87, 112.58, 111.59, 112.58, 111.66, 111.24, 90.92, 77.21, 48.13, 46.13, 36.50; IR(KBr) ν_{max} : 3309, 2935, 2745, 2676, 2491, 1641, 1580, 1528, 1452, 1393, 1315, 1241, 1196, 1030, 750, 428 cm⁻¹; HRMS m/z ; 608.1699 (calc. 608.1701)

N-(1-(4-hydroxy-3-methoxyphenyl)-3-oxo-3-(2-oxo-2H chromen-3-yl) propyl)-3-(4-((3-formyl-1H-indol-1-yl) methyl)-1H-1,2,3-triazol-1-yl) propanamide (**4d**)

Brownish solid ; 555.79 mg (yield 84 %); ¹H NMR (400 MHz, DMSO-d₆): δ 9.937 (s, 1H), 8.908-8.876 (d, J= 12 Hz, 1H), 8.408 (s, 1H), 8.408 (t, 1H), 8.358(s, 1H), 8.213(s, 1H), 8.119-7.118 (9H, m), 7.643(s, 1H), 6.378(s, 2H), 6.341-6.223(t, 1H), 5.927-5.905 (d, J= 8 Hz, 1H), 5.753-5.714(d, J= 16Hz, 1H), 5.061-5.014(t, 2H), 5.061-5.014(t, 2H), 3.980(s, 3H), 1.234(s, 3H); ¹³C NMR (100 MHz, DMSO-d₆) δC (ppm): 199.93, 185.42, 177.79, 166.54, 159.39, 156.23, 141.11, 139.87, 137.08, 131.96, 129.58, 125.30, 124.30, 123.33, 123.11, 121.66, 117.87, 112.58, 111.24, 90.92, 77.21, 51.75, 48.13, 46.13, 29.46; IR(KBr) ν_{max} : 2922, 1719, 1655, 1608, 1530, 1488, 1465, 1401, 1384, 1321, 1269, 1201, 1170, 1123, 1032, 752 cm⁻¹; HRMS m/z ; 662.2232 (calc. 662.2251).

3-(4-((3-acetyl-1H-indol-1-yl)methyl)-1H-1,2,3-triazol-1-yl)-N-(1-(4-chlorophenyl)-3-oxo-3-(2-oxo-2H-chromen-3-yl)propyl)propanamide (5b)

Brownish solid ; 516.32mg (yield 83%); ¹H NMR (400 MHz, DMSO-d₆): δH (ppm): 8.582(s, 1H), 8.381(s, 1H), 8.309(s, 1H), 8.215-8.199(d, J= 6Hz ,1H), 8.215(s, 1H), 7.979(s, 1H), 7.628-7.612(d, J= 6Hz,1H),7.481(s, 1H), 7.435(s, 1H), 7.319-7.262(m, 6H), 5.245(s, 2H), 5.199-5.018(m,1H), 4.304-4.199(m, 2H), 3.319-3.262(dd, J=8Hz,16 Hz,1H), 3.199-3.018(dd, J= 10Hz, 1H), 2.460-2.352(m,2H),1.236(s,3H);¹³CNMR(100MHz,DMSO-d₆)δC(ppm): 195.13,185.22,168.60,167.48,160.72,141.07,140.56,139.86,138.65,137.31,137.05,129.48,128.98,125.13,124.22,124.15,124.06,123.32, 123.27,123.09,121.65,121.57,121.54,118.12,118.04,112.57,112.01,111.61,11.58,111.23,60.72,51,69,50.91,48.10,28.54,28.32;IR(KBr) ν_{max}:3218,2923,1716,1680,1627,1609,1525,1488,1455,1388,1340,1275,1207,1090,1013,944, 931,827,753,681,657 cm⁻¹; HRMS m/z ; 622.1854 (calc. 622.1857).

3-(4-((3-acetyl-1H-indol-1-yl)methyl)-1H-1,2,3-triazol-1-yl)-N-(1-(4-hydroxy-3-methoxyphenyl)-3-oxo-3-(2-oxo-2H-chromen-3-yl)propyl)propanamide (5d)

Brownish solid ; 540.55 mg (yield 80%);¹H NMR (400 MHz, DMSO-d₆): δH (ppm): 9.9321 (s, 1H), 8.380(s, 1H),8.304(s,1H),8.215 (s,1H),7.652-7.633 (d, J=8Hz, 1H), 7.498-7.482 (d, J=6Hz, 1H), 7.416(s, 1H),7.321-7.260(m,7H),6.378(s, 2H), 6.341-6.223(t, 1H),5.199-5.042(t, 2H), 4.304(s, 3H), 3.938-3.876(dd, J=10 Hz and J=12 Hz,1H),3.860-3.839(dd, J= 8Hz,1H),2.432-2.414(t, 2H), 1.911(s,3H),1.239(s,3H);¹³CNMR(100MHz,DMSO-d₆)δC(ppm): 195.23,185.40,168.05,162.39,160.72,148.15,147.81,141.07,140.56,139.86,138.65,137.31,129.48,128.98,125.13,124.22,123.32,121.65, 118.12,112.57,111.65,111.23,78.43,77.20,51.73,44.96,31.78,28.32,20.04;IR(KBr)ν_{max}:2927,2851,1718,1681,1608,1513,1488,1455,1428,1387,1364,1319,1271,1215,1123, 1084, 1031, 933, 859, 822, 756, 611, 559cm⁻¹; HRMS m/z; 676.2400 (calc. 676.2407).

(1-(3-((1-(4-chlorophenyl)-3-oxo-3-(2-oxo-2H-chromen-3-yl) propyl)amino)-3-oxopropyl)- 1H-1,2,3-triazol-4-yl)methyl 1H –indole-3-carboxylate (6b)

Brownish solid ; 505.47 mg (yield 81 %); ^1H NMR (400 MHz, DMSO- d_6): δH (ppm): 9.911(s, 1H), 8.296(s, 1H), 8.240(s, 1H), 8.134(s, 1H), 7.995(s, 2H), 7.769(s, 2H), 7.769-7.636(m, 2H), 7.491-7.045(m, 6H), 5.404(s, 2H), 5.328(t, 1H), 5.068-4.921(t, 2H), 4.83-4.718(dd, J = 20 Hz ,1H), 3.938-3.876(dd, J = 10 Hz and 12 Hz, 1H), 1.296-1.239(m, 2H); ^{13}C NMR (100MHz, DMSO- d_6) δC (ppm): 194.03, 185.17, 168.21, 167.69, 147.81, 141.05, 140.05, 140.65, 140.53, 139.87, 137.32, 137.04, 125.13, 124.28, 124.23, 124.14, 124.05, 123.30, 123.24, 123.08, 121.65, 121.57, 121.54, 112.57, 112.03, 111.65, 111.62, 111.59, 85.32, 60.92, 51.68, 48.12, 28.54; IR (KBr) ν_{max} : 2974, 2932, 2738, 2676, 2490, 1748, 1655, 1591, 1528, 1456, 397, 1360, 1337, 1314, 1243, 1170, 1243, 1170, 1122, 1089, 1036, 989, 939, 821, 754 cm^{-1} ; HRMS m/z ; 624.1650 (calc. 624.1650).

(1-(3-((1-(4-hydroxy-3-methoxyphenyl)-3-oxo-3-(2-oxo-2H-chromen-3-yl)propyl)amino)-3-oxopropyl)-1H-1,2,3-triazol-4-yl)methyl 1H-indole-3-carboxylate (6d)

Brownish solid ; 542.13 mg (yield 80%); ^1H NMR (400 MHz, DMSO- d_6): δH (ppm): 9.876(s, 1H), 8.301-8.244(d, J =22 Hz , 1H), 8.135(s, 1H), 8.041(s, 1H), 8.021(s, 1H), 7.716-7.660(d, J =22 Hz , 1H), 7.497(s, 1H), 7.458(s, 1H), 7.425(s, 1H), 7.335-7.229(m, 6H), 5.224(s, 2H), 5.115-4.876(t, 1H), 4.660-4.643(t, 3H), 4.237(s, 3H), 3.753-3.658(dd, J =16Hz, 1H), 3.458-3.367(dd, J =10 Hz, 1H), 1.474-1.449(t, 2H), 1.239 (s, 3H); ^{13}C NMR (100MHz, DMSO- d_6) δC (ppm): 194.37, 185.43, 167.69, 163.00, 160.72, 148.15, 147.81, 140.58, 139.94, 137.04, 135.38, 133.09, 130.09, 129.20, 128.57, 128.27, 127.49, 125.23, 125.15, 124.24, 123.29, 121.58, 118.03, 117.00, 111.64, 78.45, 77.19, 63.24, 44.52, 36.50, 31.89, 29.41; IR (KBr) ν_{max} : 2923, 1704, 1608, 1532, 1509, 1488, 1456, 1394, 1268, 1199, 1122, 1034, 1031, 861, 754, 606, 518 cm^{-1} ; HRMS m/z ; 678.2194 (calc. 678.2200).

2.5. Cell culture and treatment

The MCF-7 cells were maintained in RPMI medium 1640 supplemented with 10% fetal bovine serum as well as 100 $\mu\text{g}/\text{mL}$ streptomycin, 100 U/mL penicillin, 2 mM L-glutamine and Earle's BSS adjusted to contain 1.5 g/L Na bicarbonate, 0.1 mM nonessential amino acids, and 1.0 mM of Na pyruvate in a humidified atmosphere containing 5% CO_2 at 37°C.

2.6. Cytotoxicity assay

Cytotoxicity was assessed using a 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay. The MCF-7 cells in the log phase were seeded in 96-well plates at a concentration of 1.0×10^4 cells/well and incubated overnight at 37°C in $5\% \text{CO}_2$ humidified environment. The cells were then treated with different concentrations of the sample **4d**, **5d** and **6d** such as 10, 20, 30, 40, 50, 60, 70, 80, 90, and 100 $\mu\text{M/mL}$ (dissolved with RPMI medium 1640), respectively. Controls were also cultivated under same conditions without the addition of compound solution. The treated cells were incubated for 48 h and then subjected for MTT assay. The stock concentration (5 mg/mL) of MTT-(3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) was prepared and 100 μL of MTT was added in each compound solution treated wells and incubated for 4 h. Purple colour formazone crystals were observed and these crystals were dissolved in 100 μL of dimethyl sulphoxide (DMSO), and read at 620 nm in a multi well ELISA plate reader (Thermo, Multiskan).

2.7. Cell morphology

To study the morphological changes, MCF-7 cells were grown (1×10^5 cells/cover slip) and incubated with at its IC_{50} concentration and then they were fixed in a mixture of methanol and acetic acid (3:1, v/v). The cover slips were gently mounted on glass slides for the morphometric analysis. Morphological changes of MCF-7 cells were analyzed under a Nikon (Japan) bright field inverted light microscope at 40 magnification. Followed by this, DAPI (4,6-diamidino-2-phenylindole, dihydrochloride) staining was carried out. For this, the MCF-7 cells incubated with compound at its IC_{50} concentration for 48 h and then they were fixed in a mixture of methanol and acetic acid (3:1, v/v) prior to washing with PBS. The washed cells were then stained with 1 mg/mL DAPI (4,6-diamidino-2-phenylindole, dihydrochloride) for 20 min. in a dark atmosphere. The stained images were recorded using a fluorescent microscope with appropriate excitation filter.

2.8. Molecular docking study

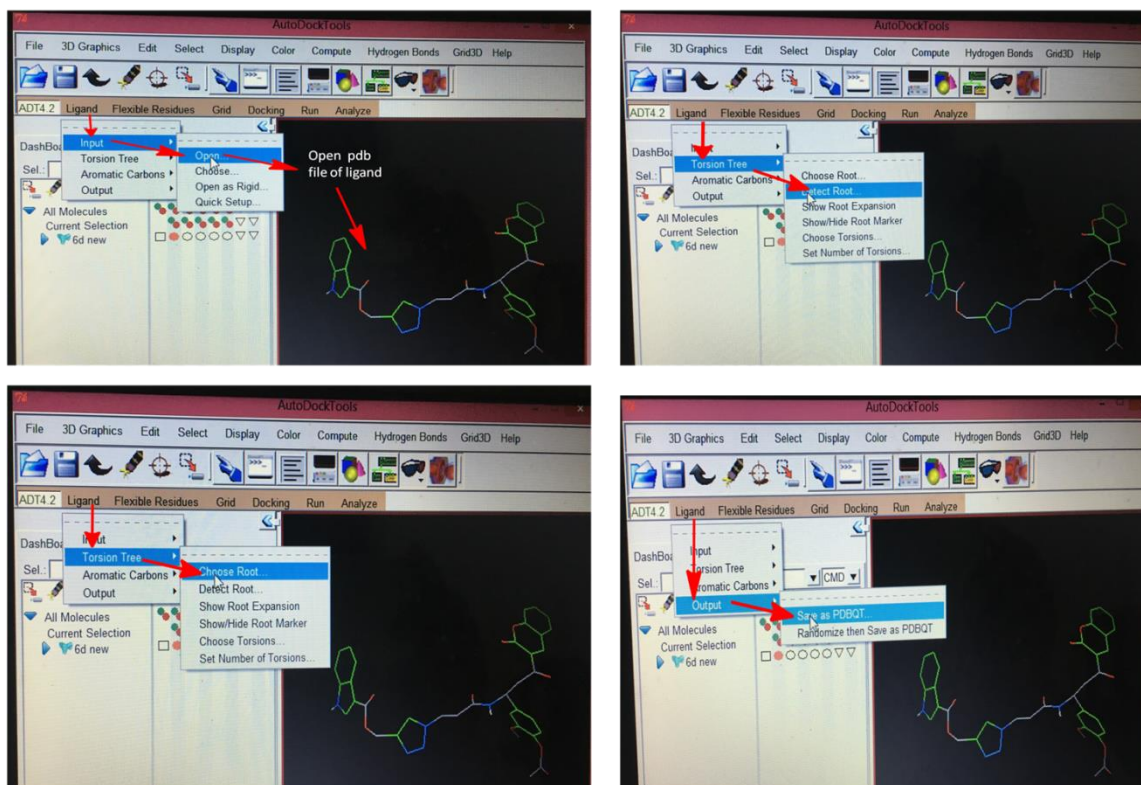
Docking studies were carried out using the Auto Dock Vina 1.1.2 program for the synthesized compounds. AutoDock vina 1.1.2 predicts the bound conformations of a small, flexible ligand to a nonflexible macromolecular target of known structure. The technique combines simulated annealing for conformation searching with a rapid grid-based method of energy

evaluation based on the AMBER force field. For this, the pdb structure of 1GII was retrieved from the Brookhaven protein database (<http://www.rcsb.org>).

STEP 1: Conversion of pdb file of ligand into pdbqt file using Autodock tool

The 3D structure of the synthesized compounds were provided using Marvin Sketch 5.8.3,2012, ChemAxon(<http://www.chemaxon.com>) and was converted to pdbqt coordinate by Autodock Tools (ADT; version 1.5.4). While converting the ligand as pdbqt, assign torsion free and detect root and then assign torsion free and choose root then save as pdbqt.

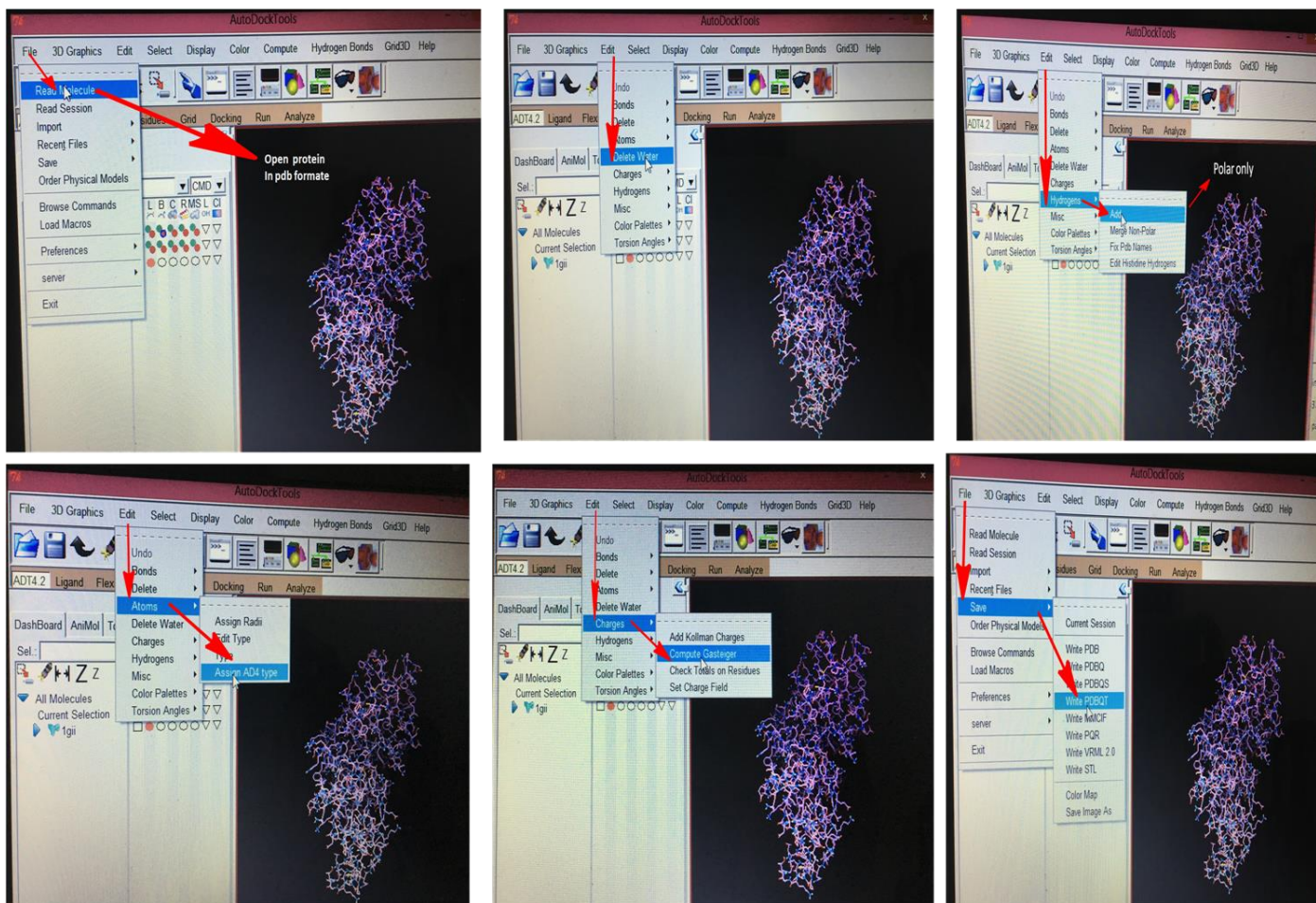
STEP 1



STEP 2: Conversion of pdb file of protien into pdbqt file using Autodock tool

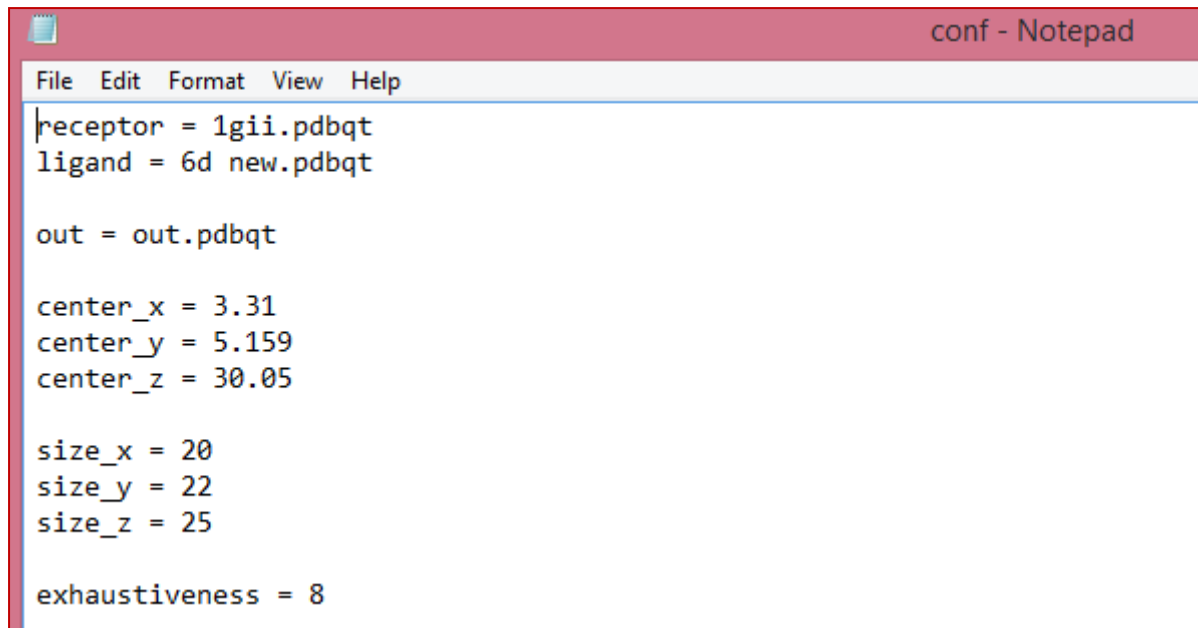
The pdb structure of 1GII (protein) was retrieved from the Brookhaven protein database (<http://www.rcsb.org>). The Protein was treated in a slightly different way. After removing all water, polar hydrogens were added, and charges assigned using the Kollman united atom library using Autodock Tools (ADT; version 1.5.4).

STEP 2



STEP 3: Note the parameters into conf file

Now to assign the docking site, all maps were calculated with 1 Å spacing between grid points by Auto grid. The center of the grid box was placed at the center of donepezil with coordinates x =3.31, y =5.159, z =30.05 with exhaustiveness 8. The dimensions of the active site box were set at 20 X22 X 25 Å. Note these parameters into conf file.



```
receptor = 1gii.pdbqt
ligand = 6d new.pdbqt

out = out.pdbqt

center_x = 3.31
center_y = 5.159
center_z = 30.05

size_x = 20
size_y = 22
size_z = 25

exhaustiveness = 8
```

STEP 4: Now Open the command prompt and run the command.

In the fourth step open the command prompt and run the command. The commands given in the box.

```
C:/vina>vina --config conf.txt --log log.txt
```

After running this command, put next command to split out file using vinasplit

```
Vina_split --input out.pdbqt
```

```

Command Prompt - vina --config conf.txt --log log.txt
Microsoft Windows [Version 6.3.9600]
(c) 2013 Microsoft Corporation. All rights reserved.

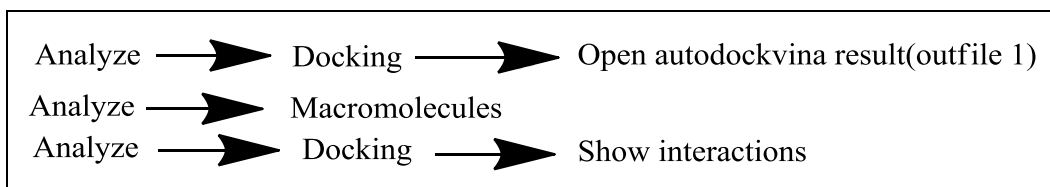
C:\Users\User>cd c:/vina

c:\vina>vina --config conf.txt --log log.txt
#####
# If you used AutoDock Vina in your work, please cite:      #
#                                                            #
# O. Trott, A. J. Olson,                                     #
# AutoDock Vina: improving the speed and accuracy of docking #
# with a new scoring function, efficient optimization and    #
# multithreading, Journal of Computational Chemistry 31 (2010) #
# 455-461                                                    #
# DOI 10.1002/jcc.21334                                     #
# Please see http://vina.scripps.edu for more information.   #
#####
Detected 4 CPUs
Reading input ... done.
Setting up the scoring function ... done.
Analyzing the binding site ... done.
Using random seed: -697182416
Performing search ...
0% 10 20 30 40 50 60 70 80 90 100%
|-----|-----|-----|-----|-----|-----|-----|-----|
*_

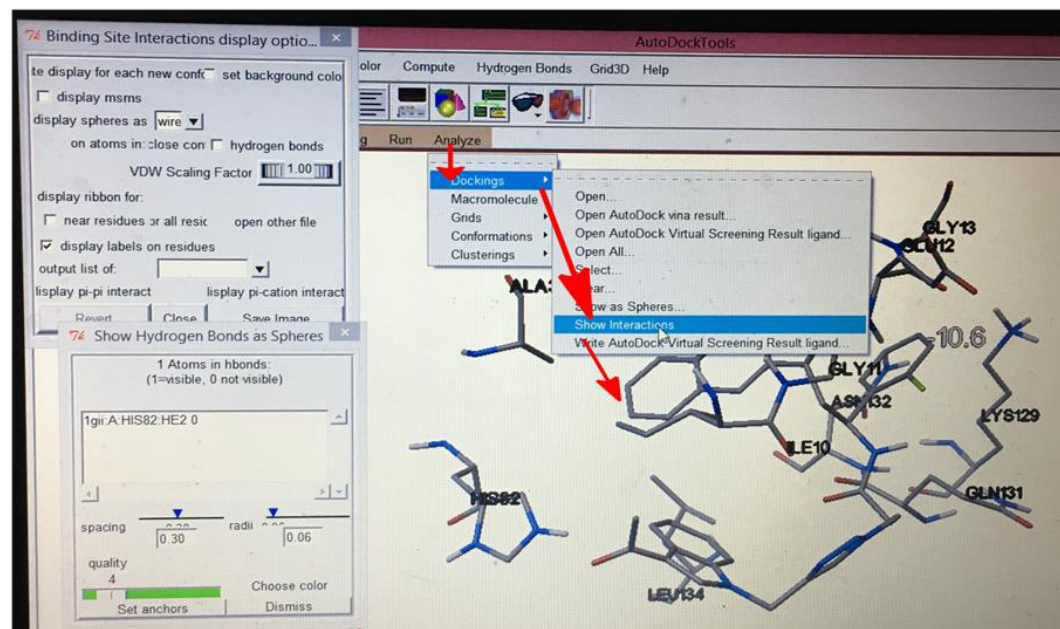
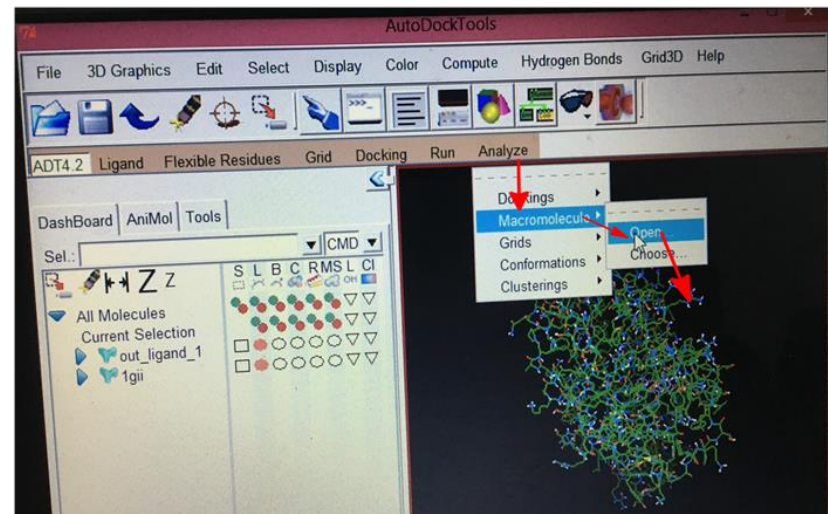
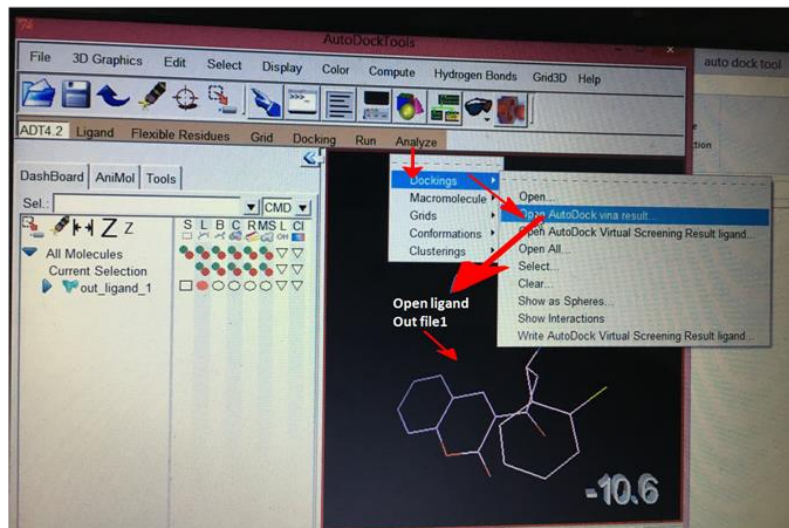
```

STEP 5: Analyse the docking interaction using autodock tool.

After finishing the vinasplit, analyse the docking interaction using autodock tool.

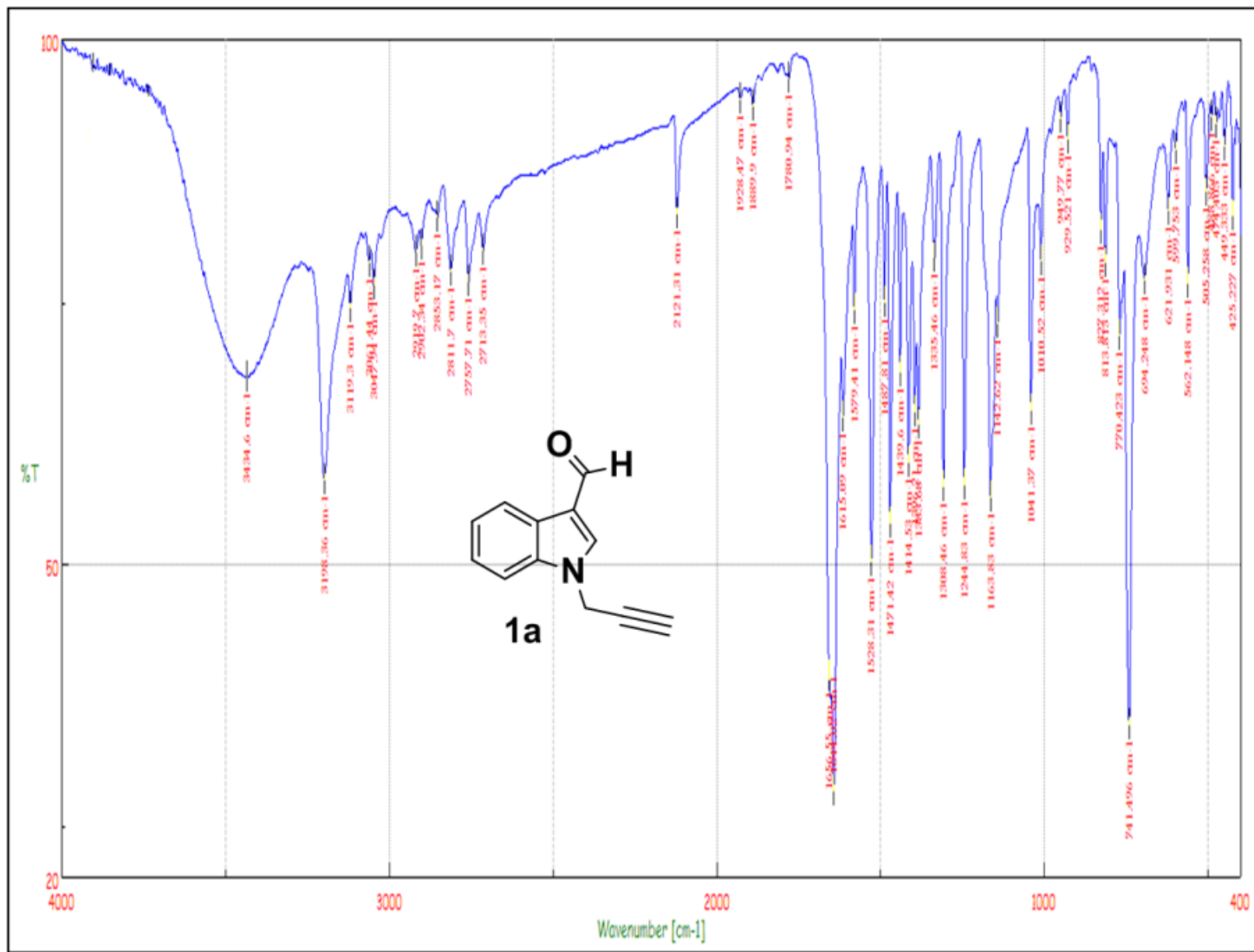


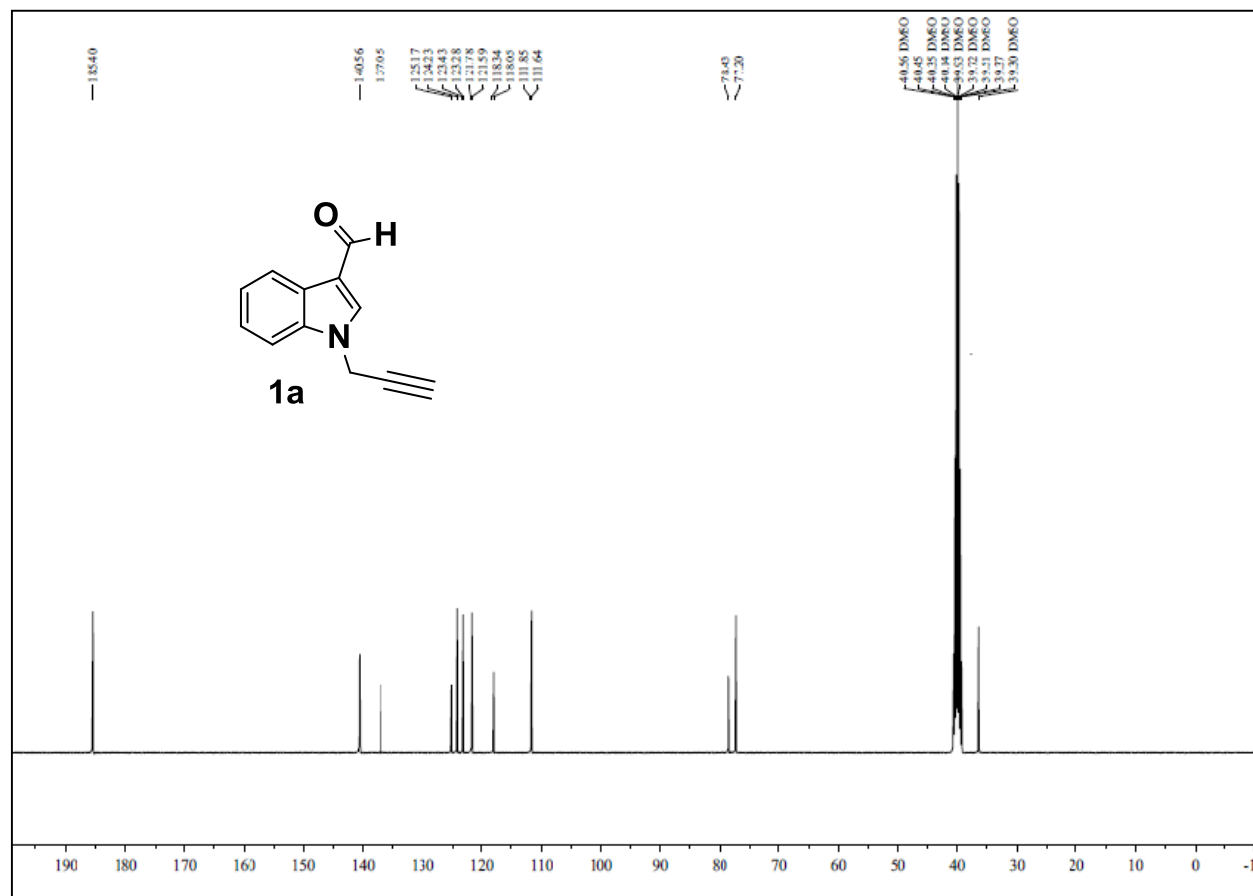
STEP 5

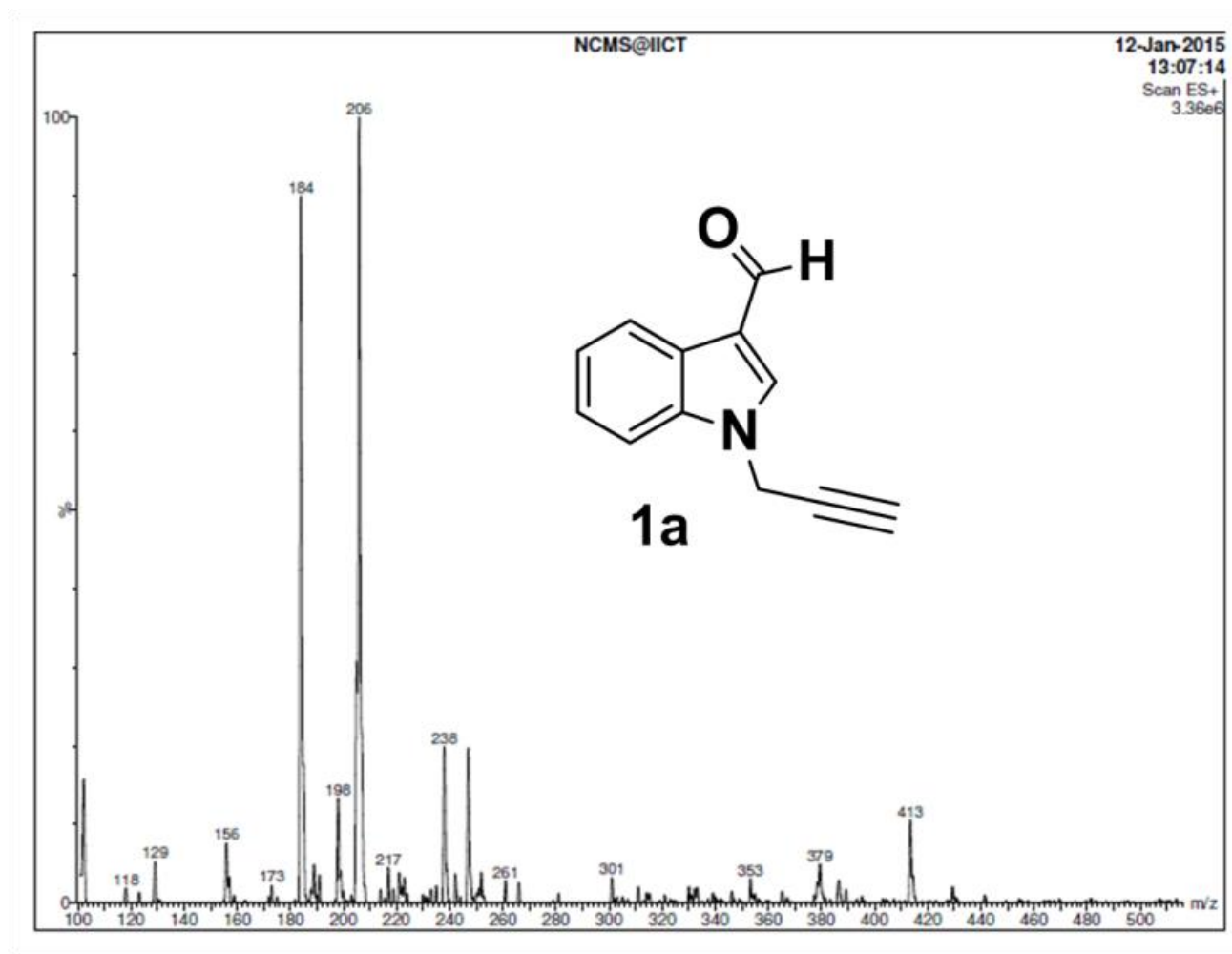


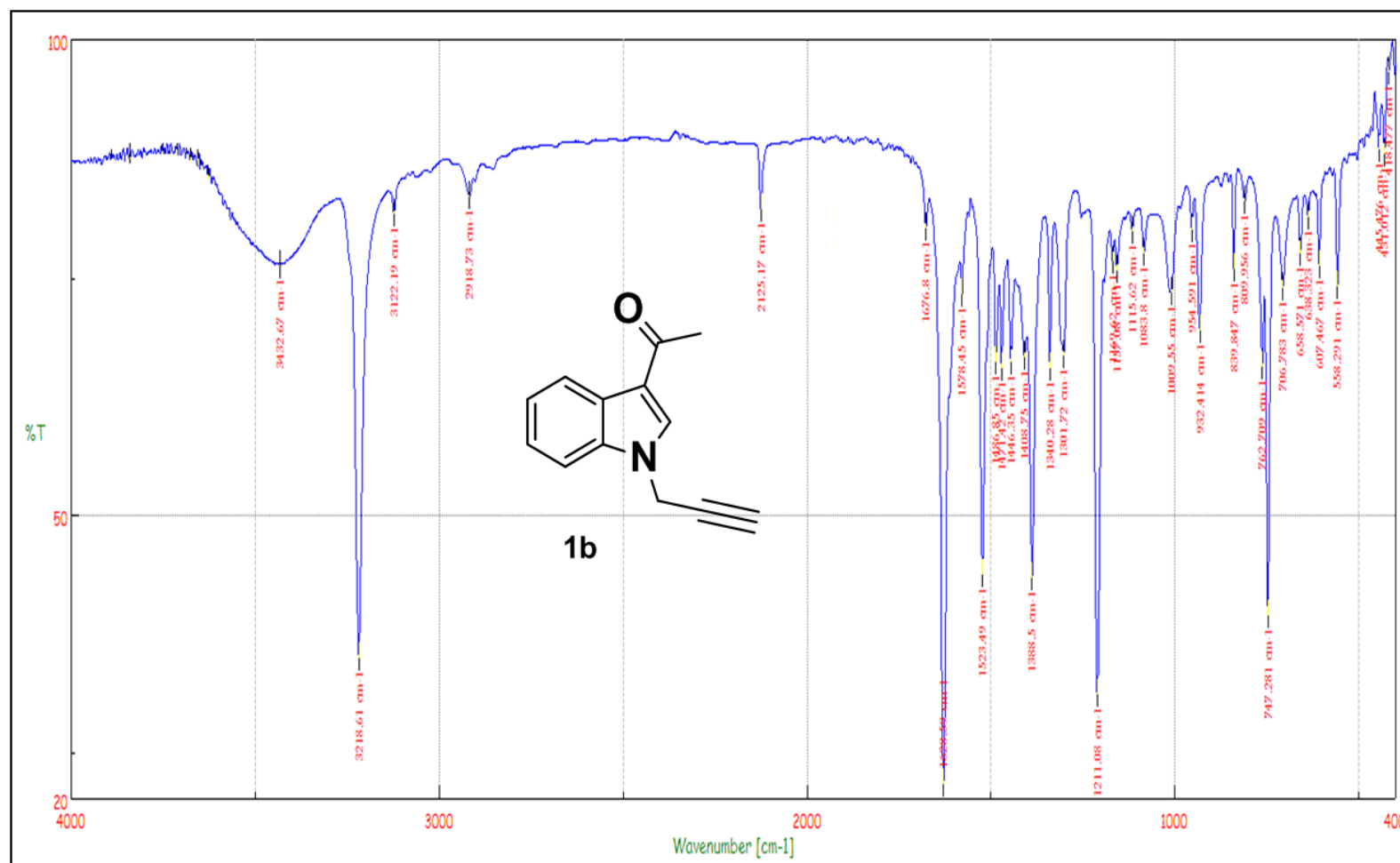
2.9. Western blot Analysis

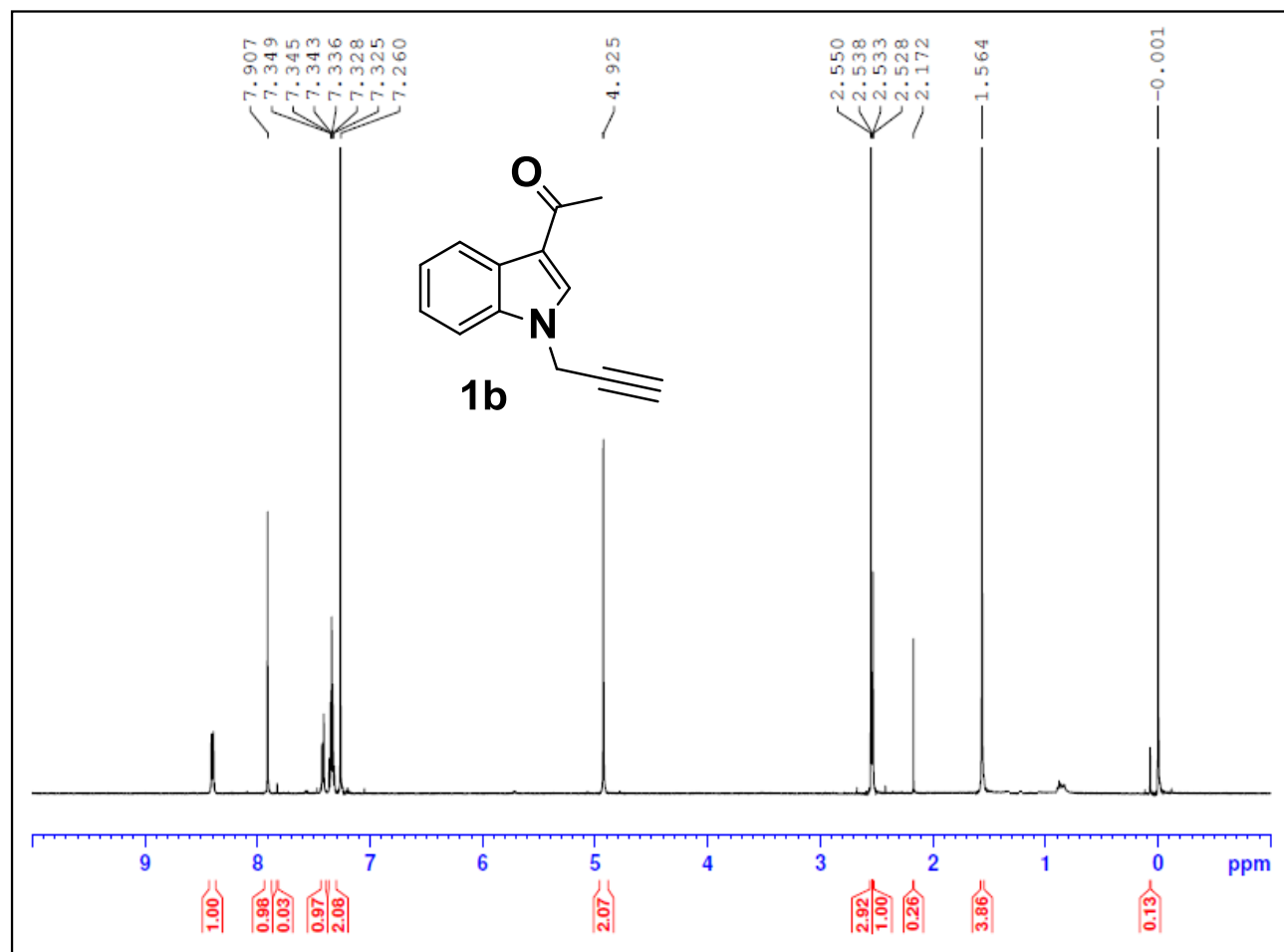
Western blotting was performed to detect the proteins of CDK2. The MCF-7 cells (1×10^6) were seeded onto 100-mm culture dishes in the presence or absence of **6a** were treated for 48 hrs. Cells were then washed twice with ice-cold PBS and incubated in lysis buffer. The lysates were centrifuged at $10,000 \times g$ for 5 min at 4 °C, and were used as the cell protein extracts. Each of the extracts was applied to 12% SDS polyacrylamide gel electrophoresis after which the proteins were transferred onto a nitrocellulose membrane, and then blocked for 1 h using 10% skim milk in water. After washing in a PBS containing 0.1% Tween 20 for 3 times, the primary antibodies were added at a v/v ratio of 1:1000. After overnight incubation at 4 °C, the primary antibodies were washed away and the secondary antibodies were added for 1 h incubation at room temperature. Finally, the enhanced chemiluminescence detection reagents were used to develop the signal of the membrane.

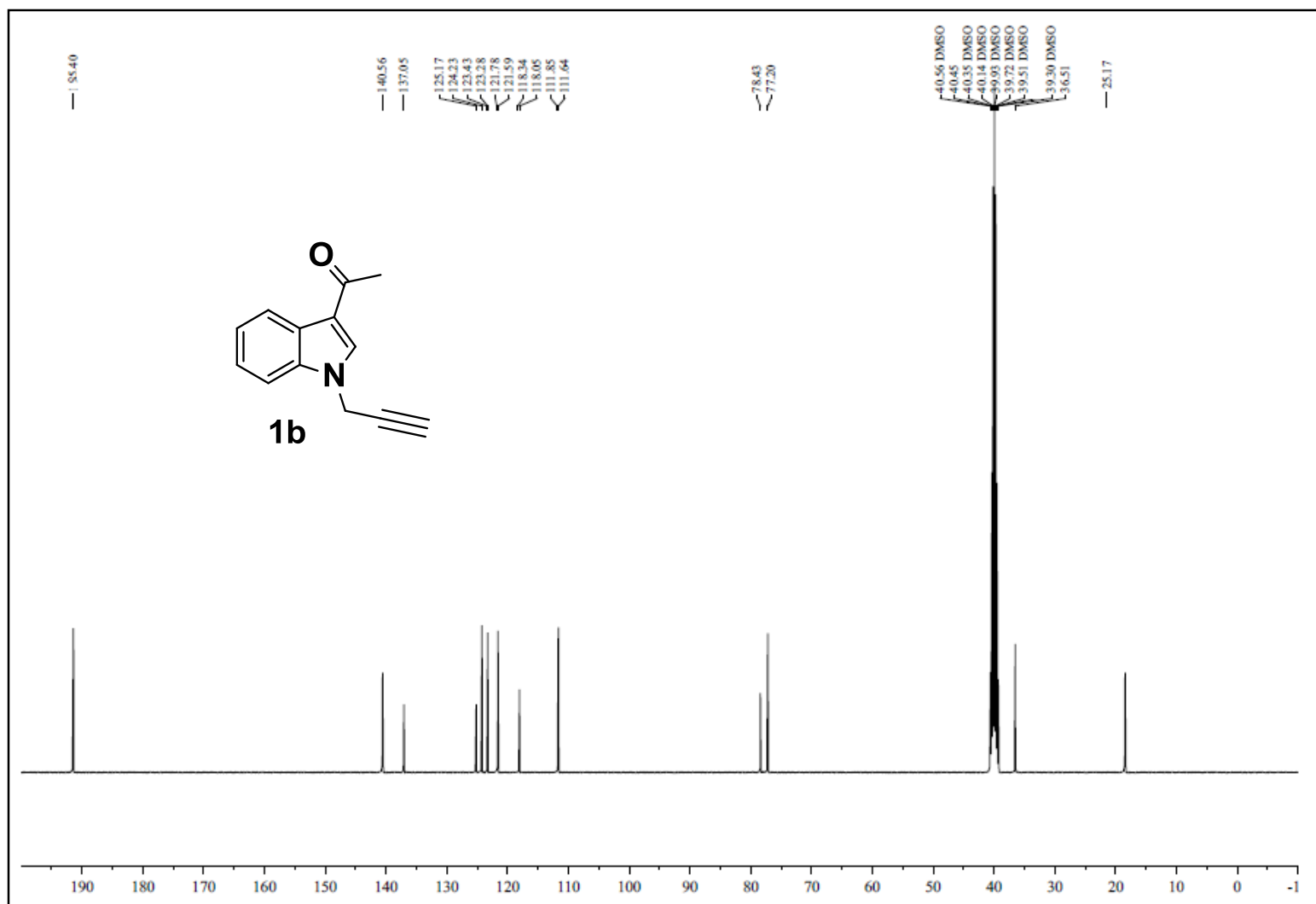
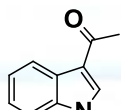


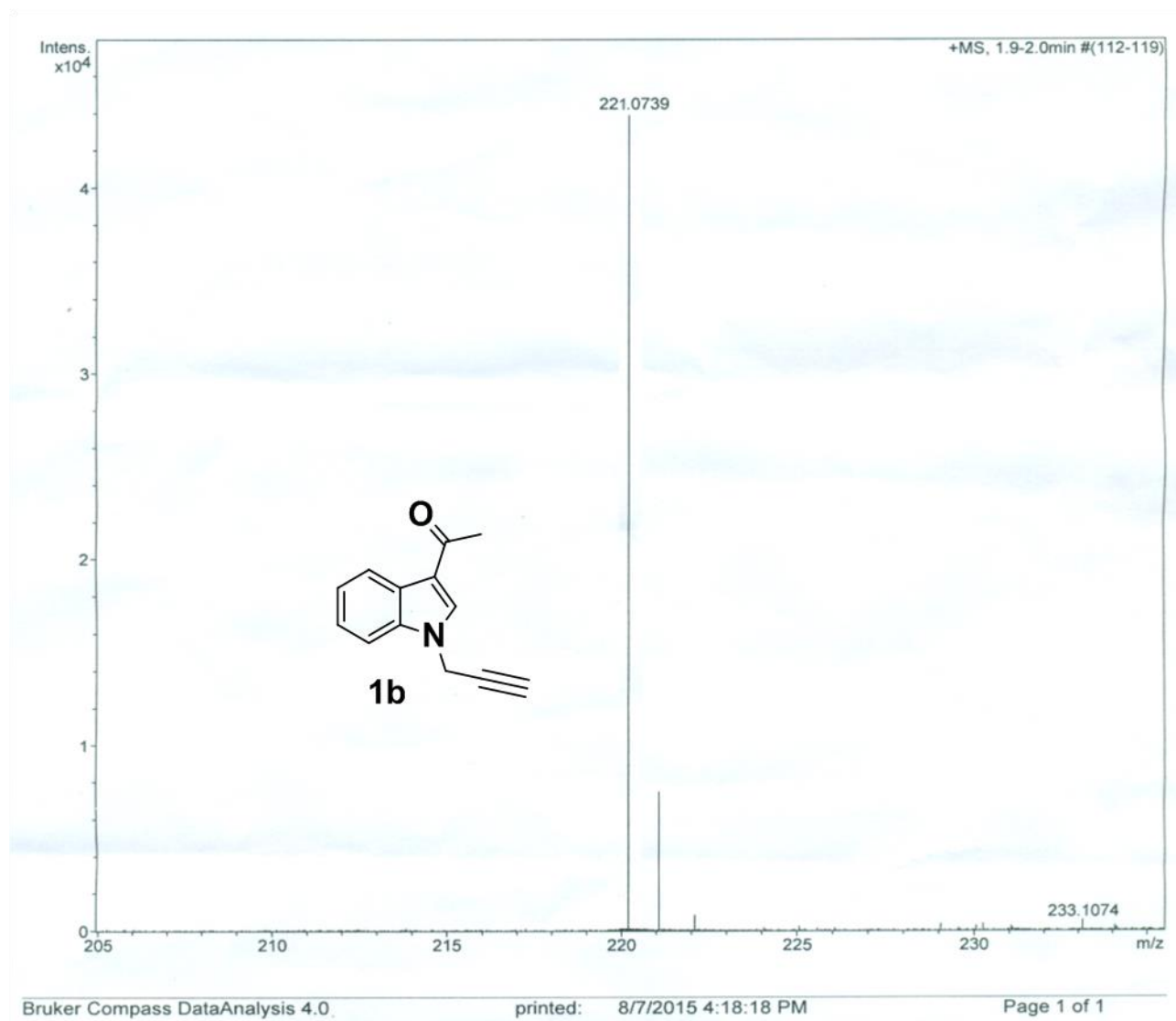


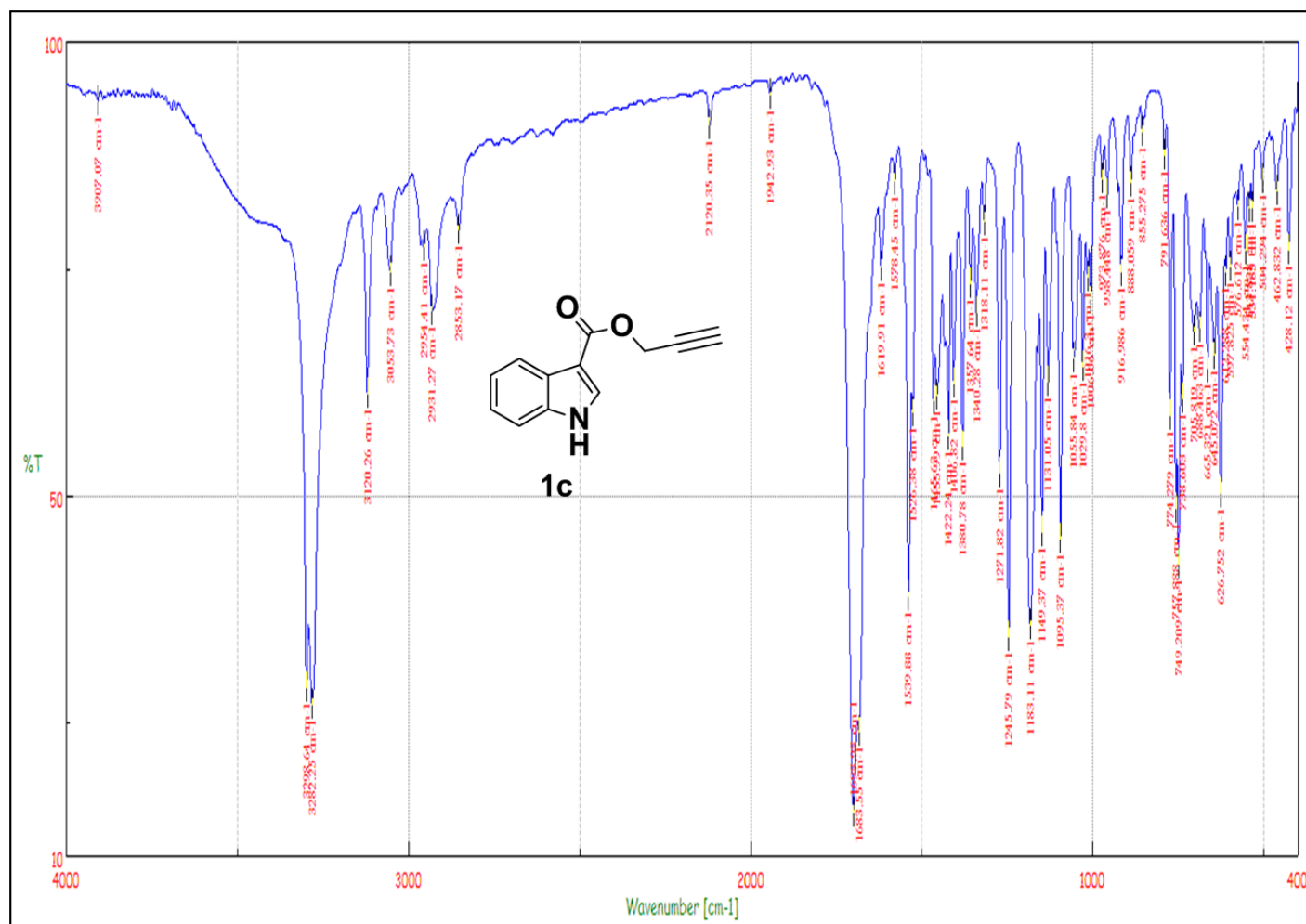


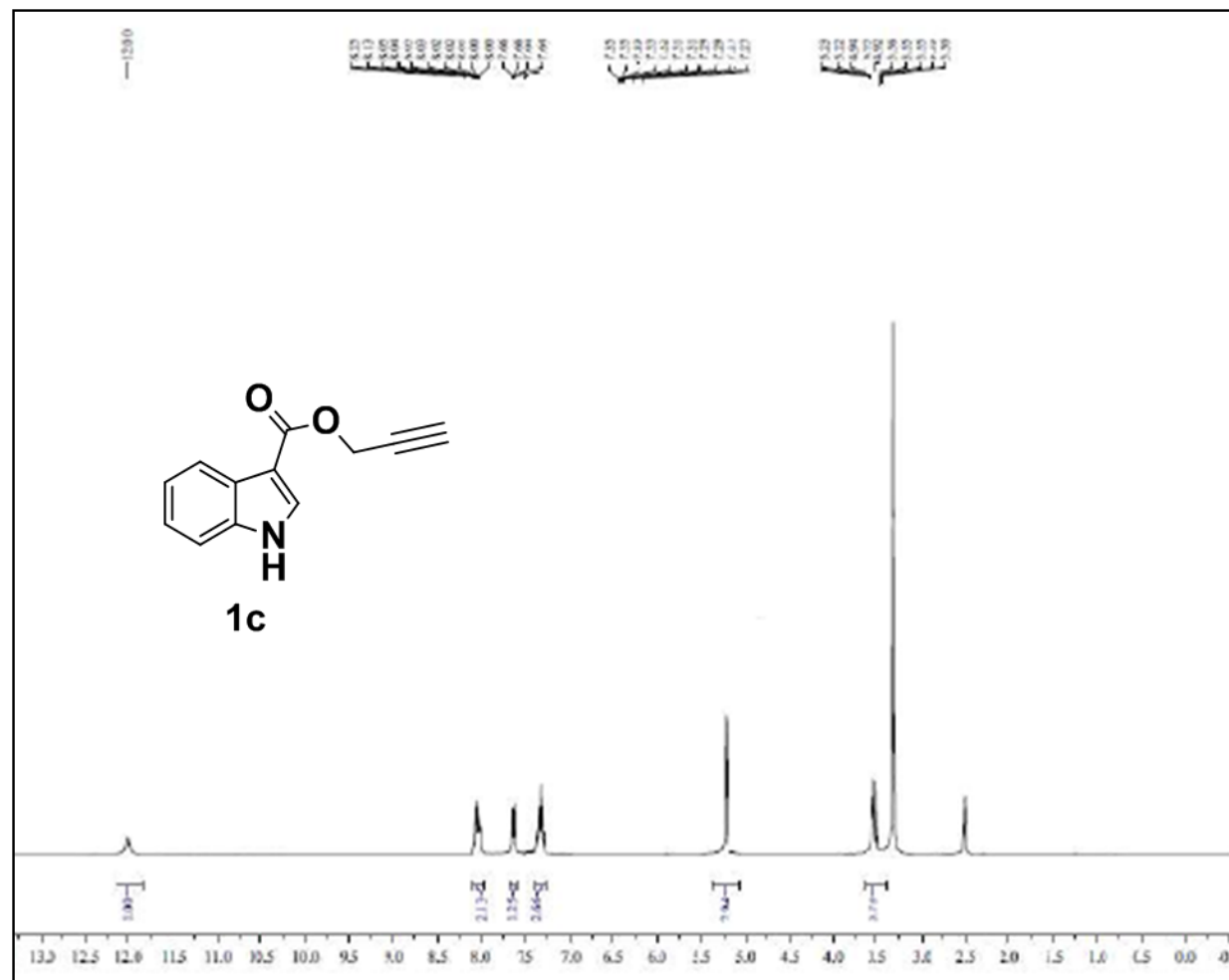


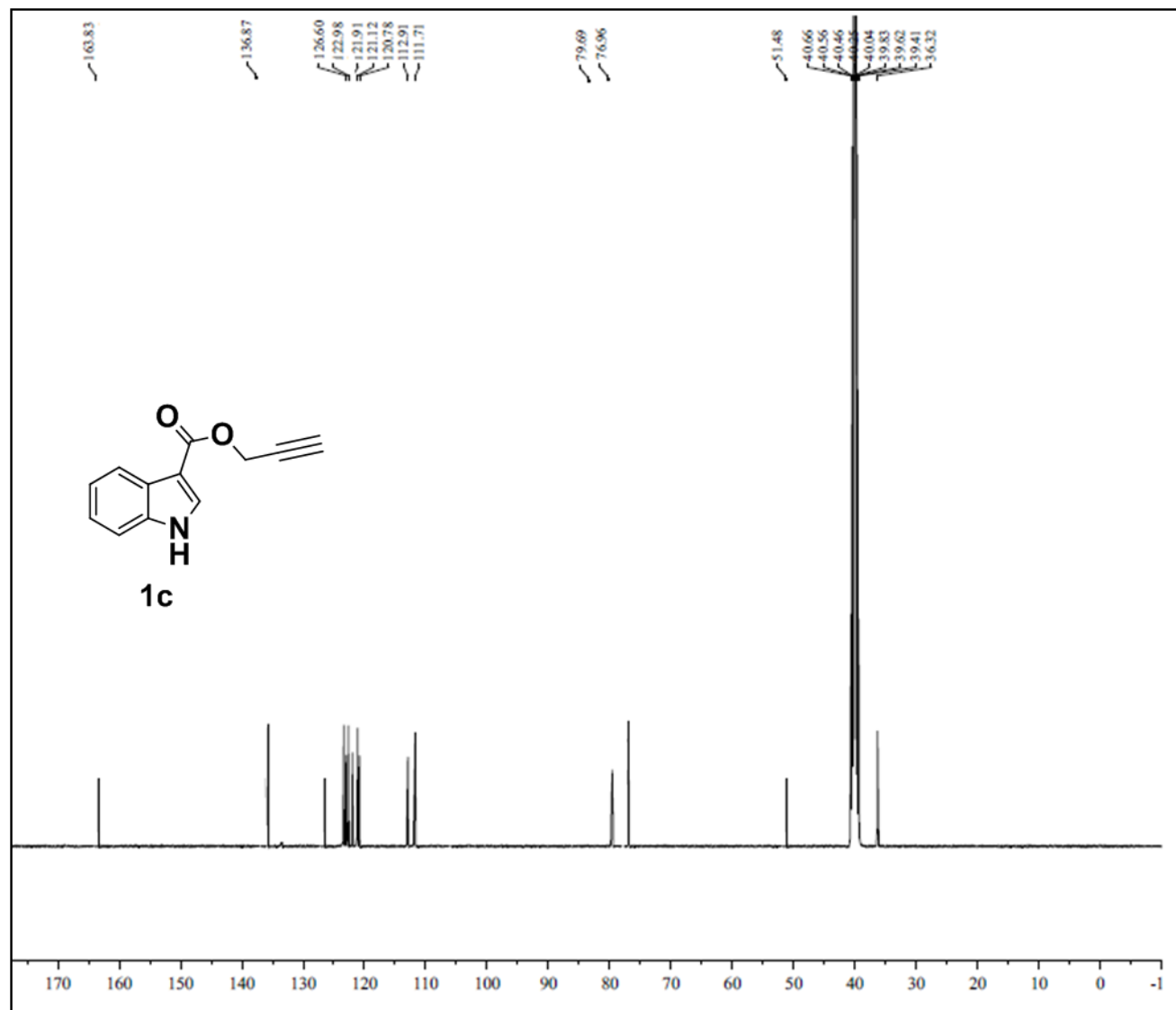


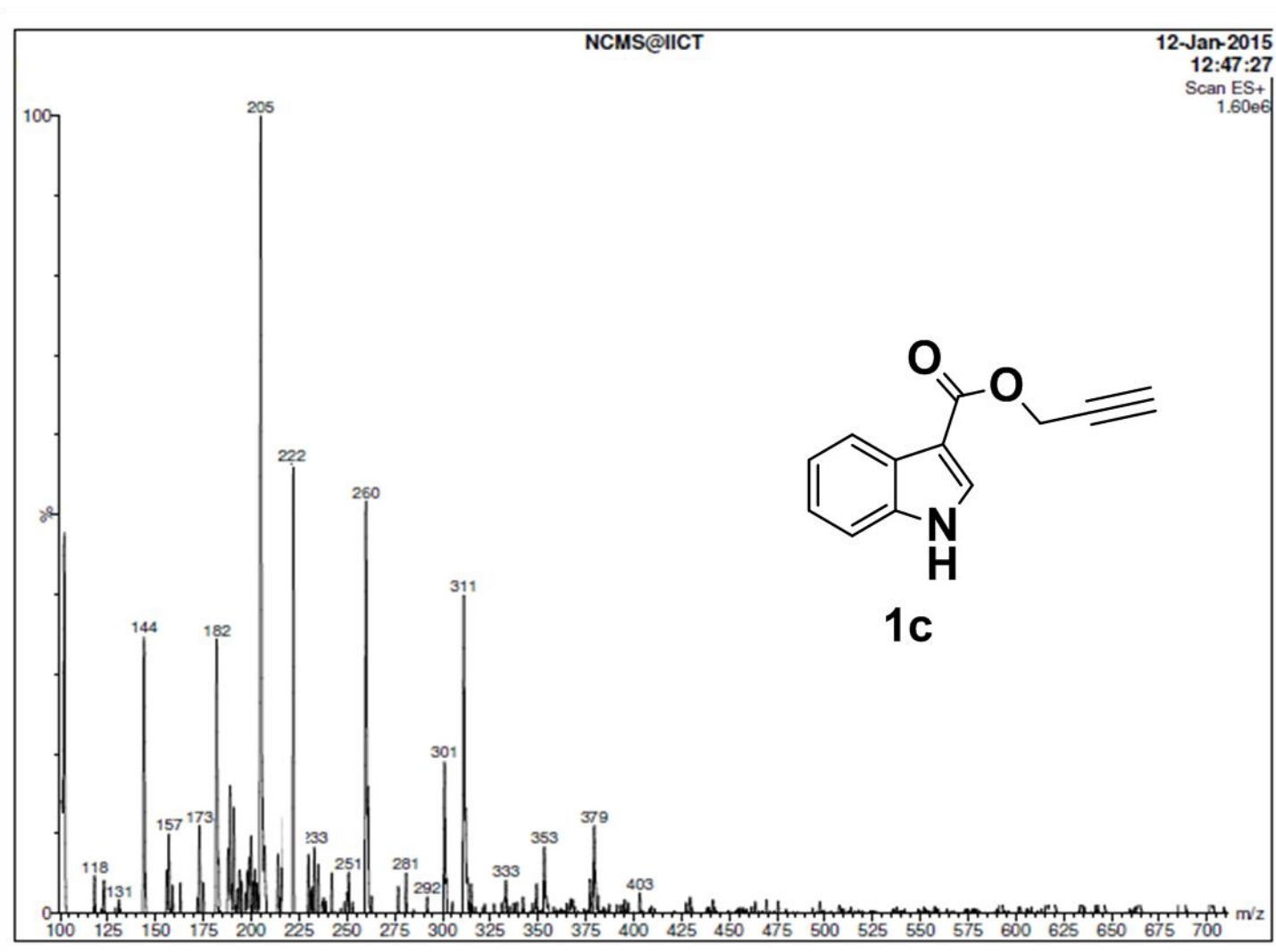


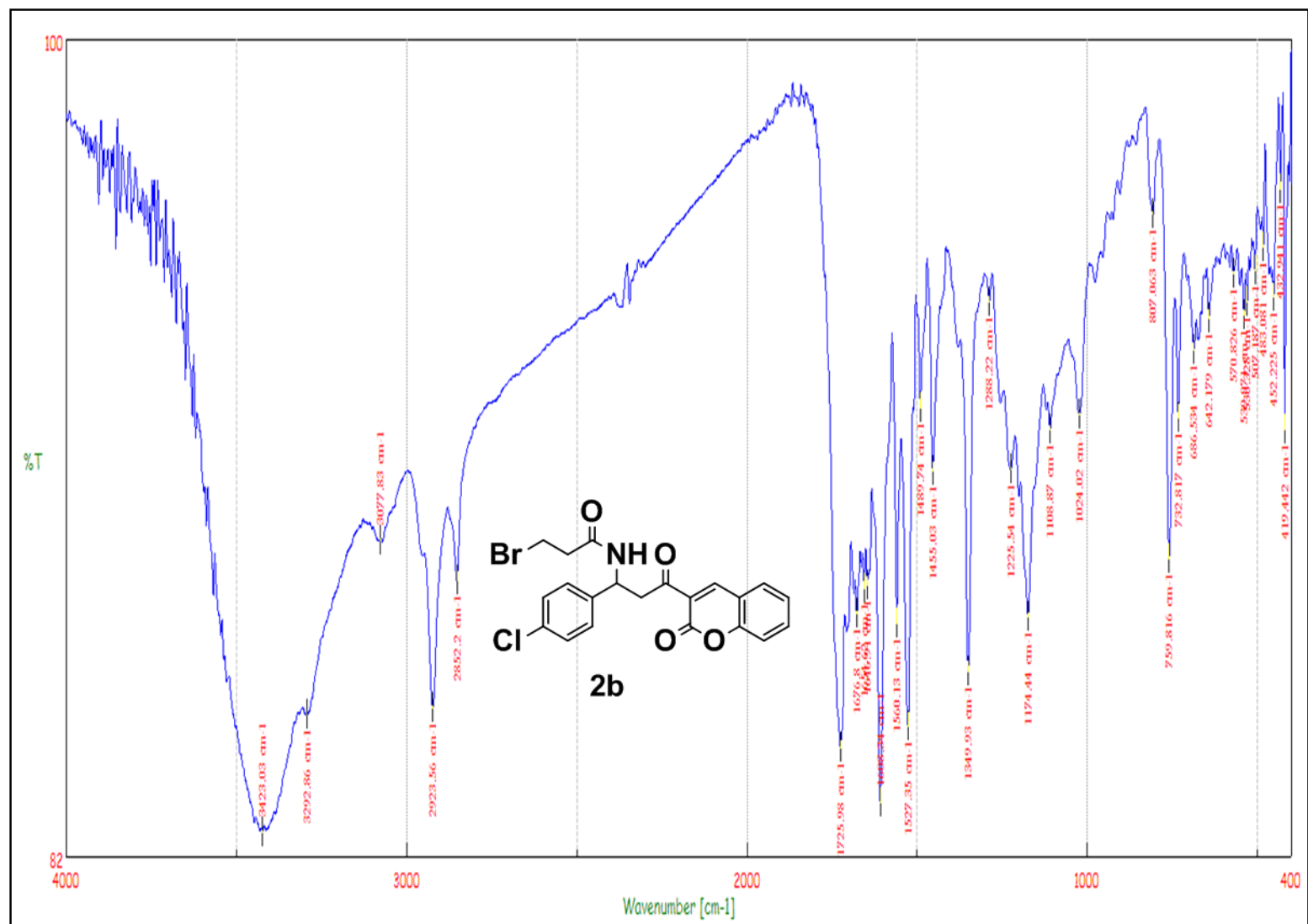


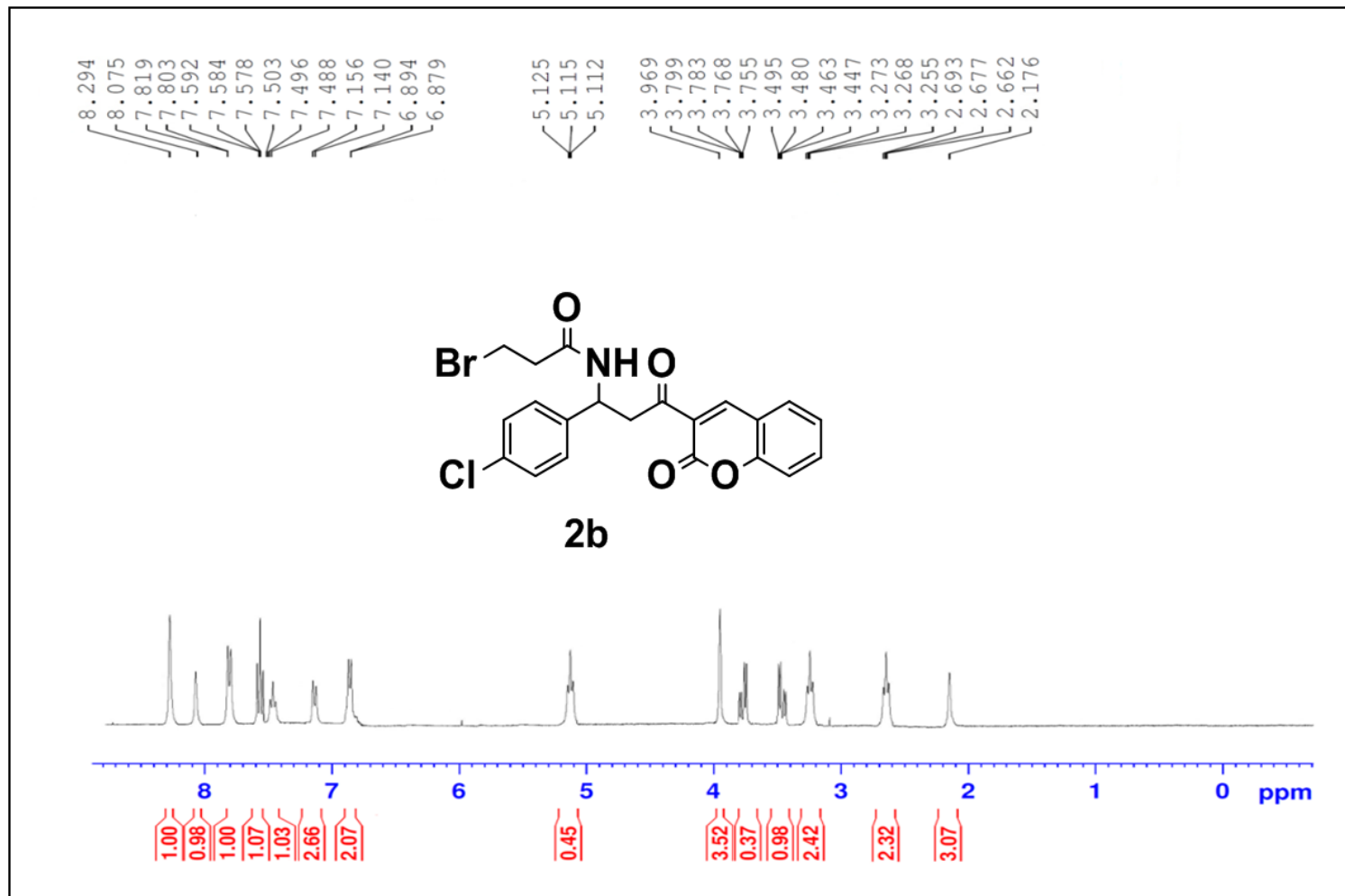






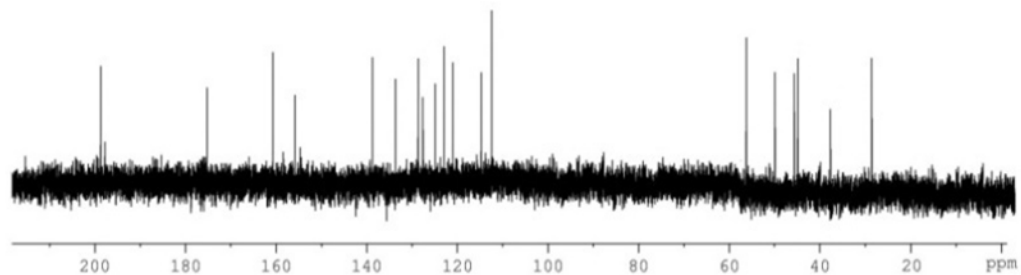
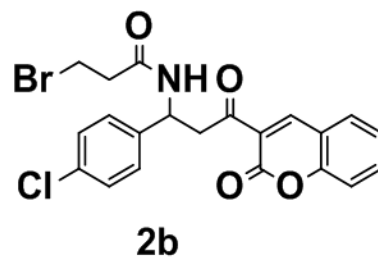






S-3
C13CPD CDC13 {D:\Yoosuf} NIIST 1

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175.35
159.92
153.63
143.68
139.07
134.91
129.76
128.92
125.46
123.82
121.88
116.13
113.20
50.16
47.53
38.23
29.45



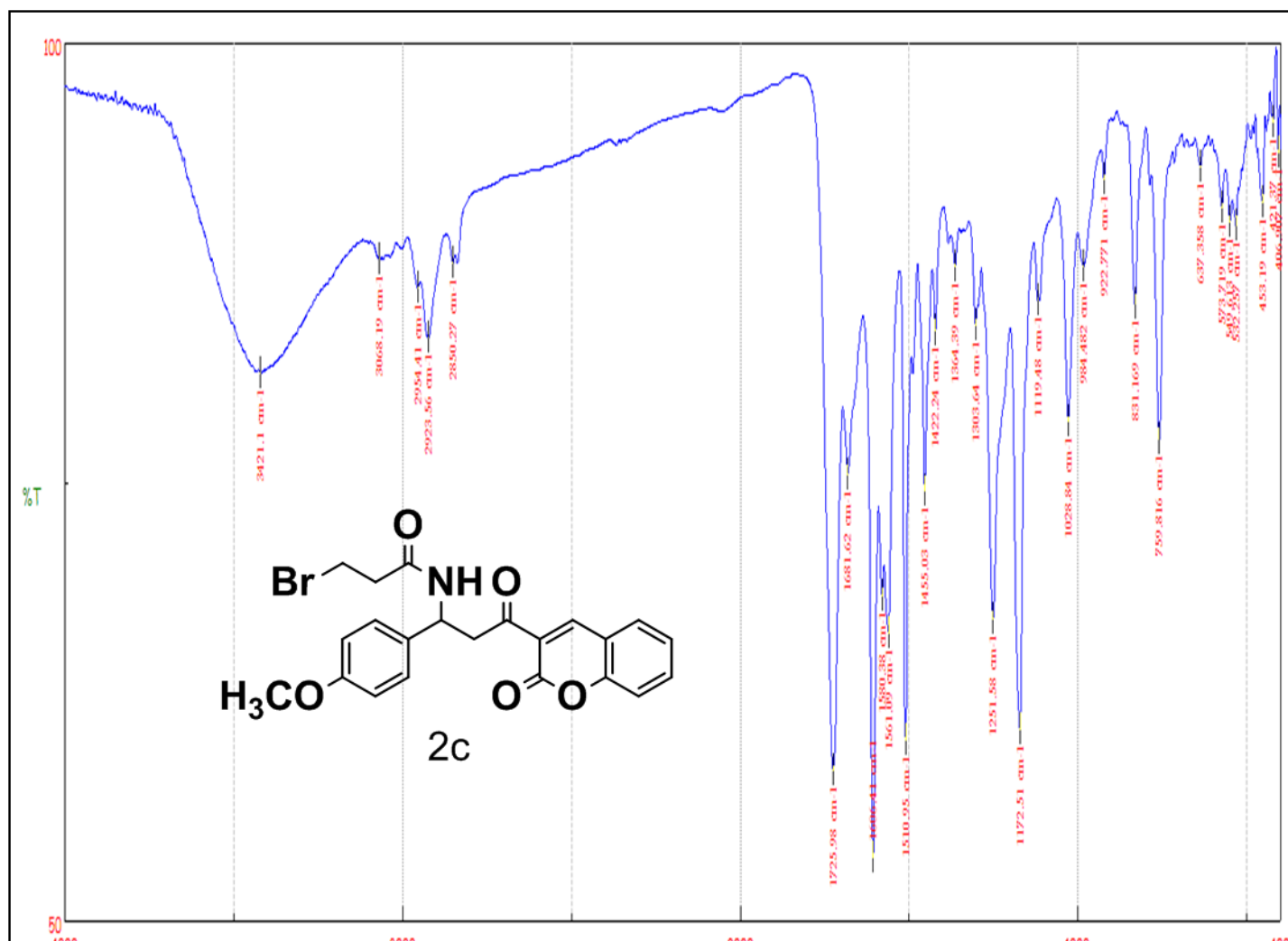
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EXPNO 2
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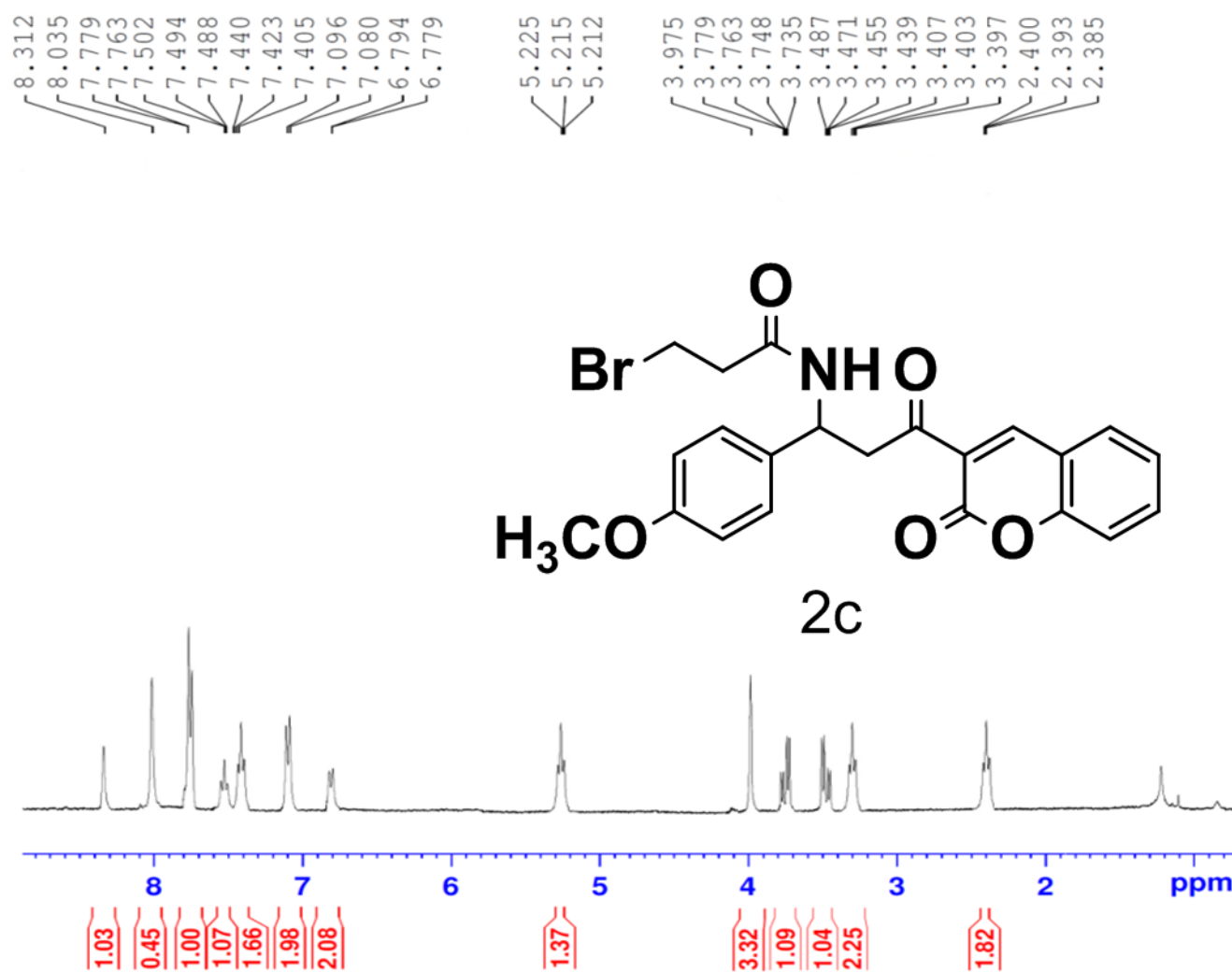
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FIDRES 0.454131 Hz
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RG 32800
RW 16.800 usec
DE 6.00 usec
TE 297.1 K
D1 2.00000000 sec
d11 0.03000000 sec
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SFO1 125.7703643 MHz

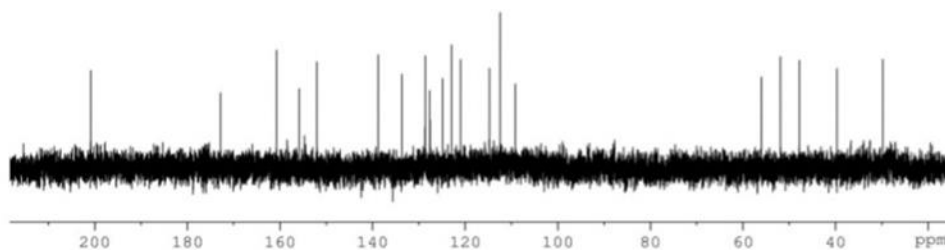
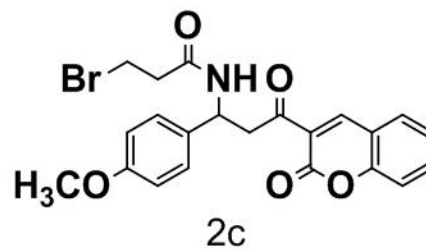
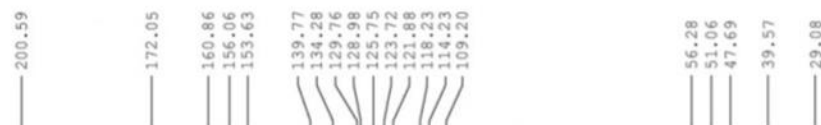
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SFO2 500.1320005 MHz

F2 - Processing parameters
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SSB 0
LB 1.00 Hz
GB 0
PC 1.40





S-1
C13CPD CDC13 {D:\Yoosuf} NIIST 7



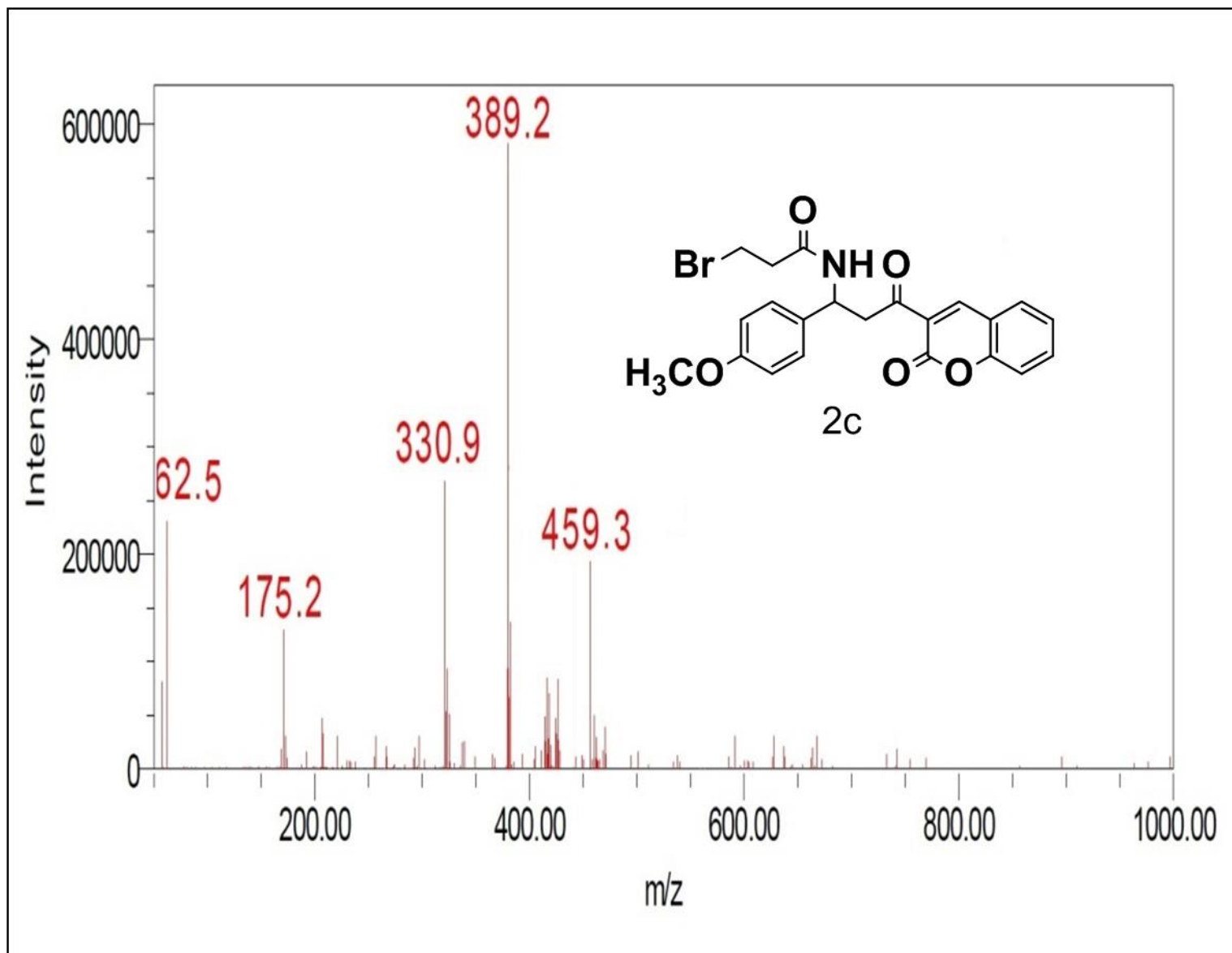
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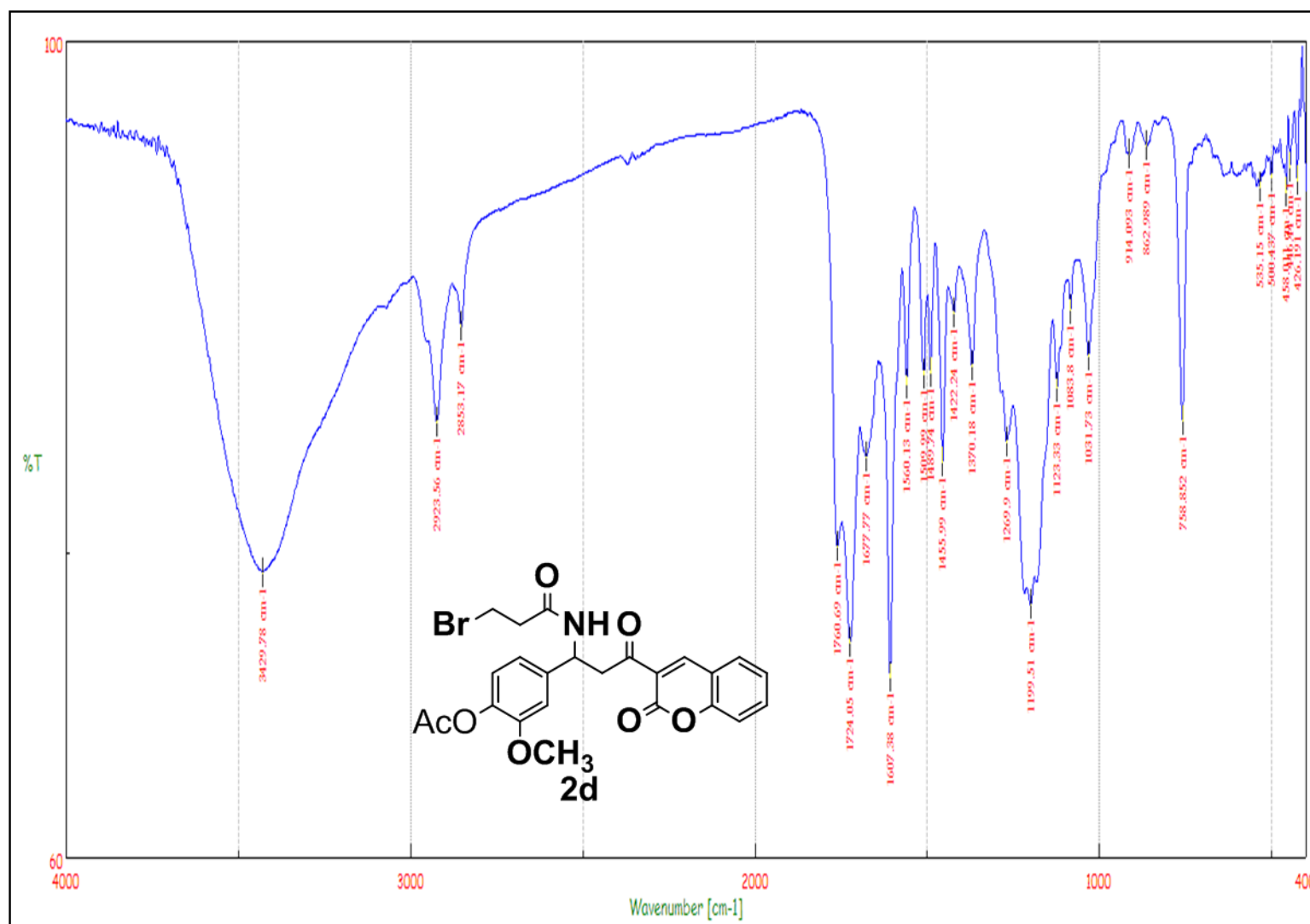
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DELTA 1.8999999 sec
TD0 1

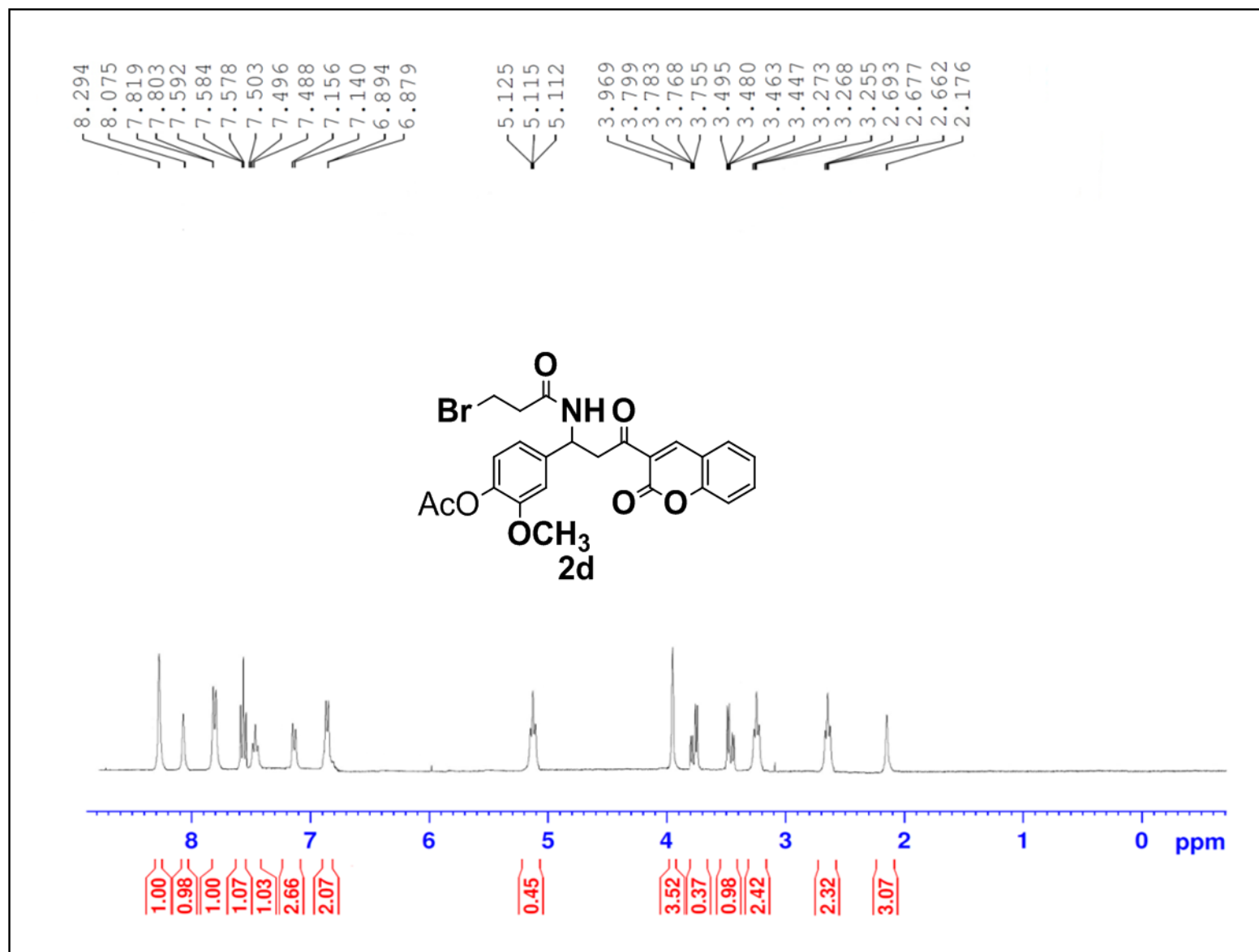
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SFO1 125.7703643 MHz

===== CHANNEL f2 =====
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PCPD2 80.00 usec
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PL13 120.00 dB
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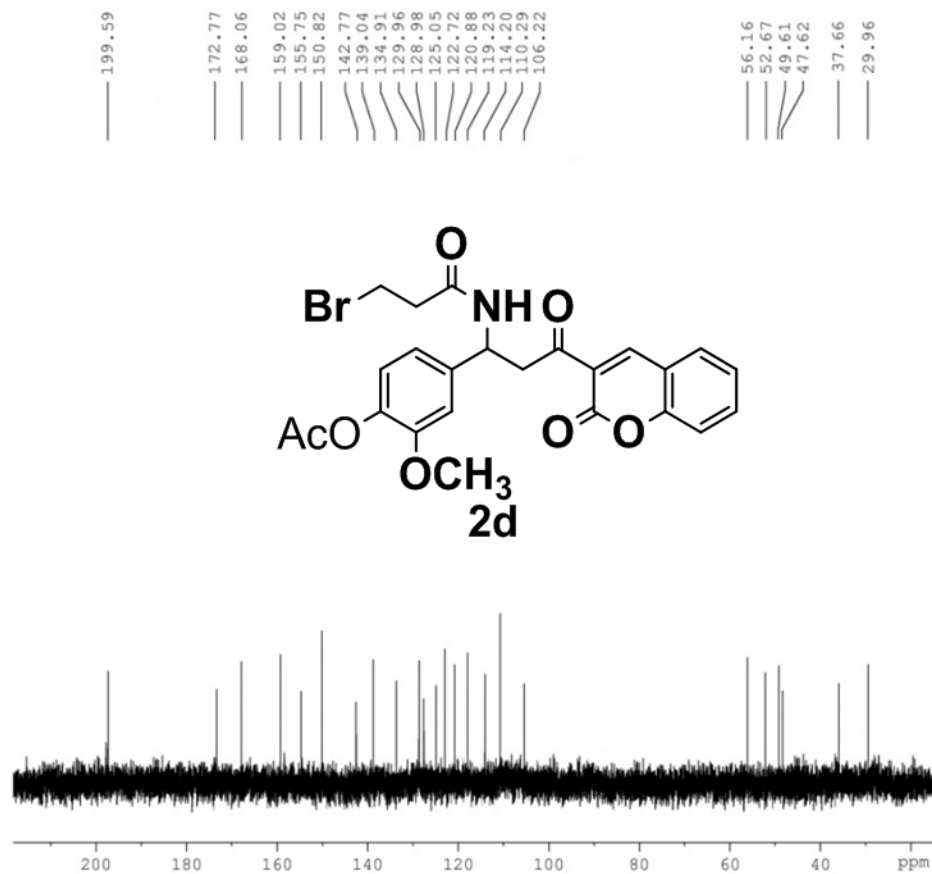
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S-16
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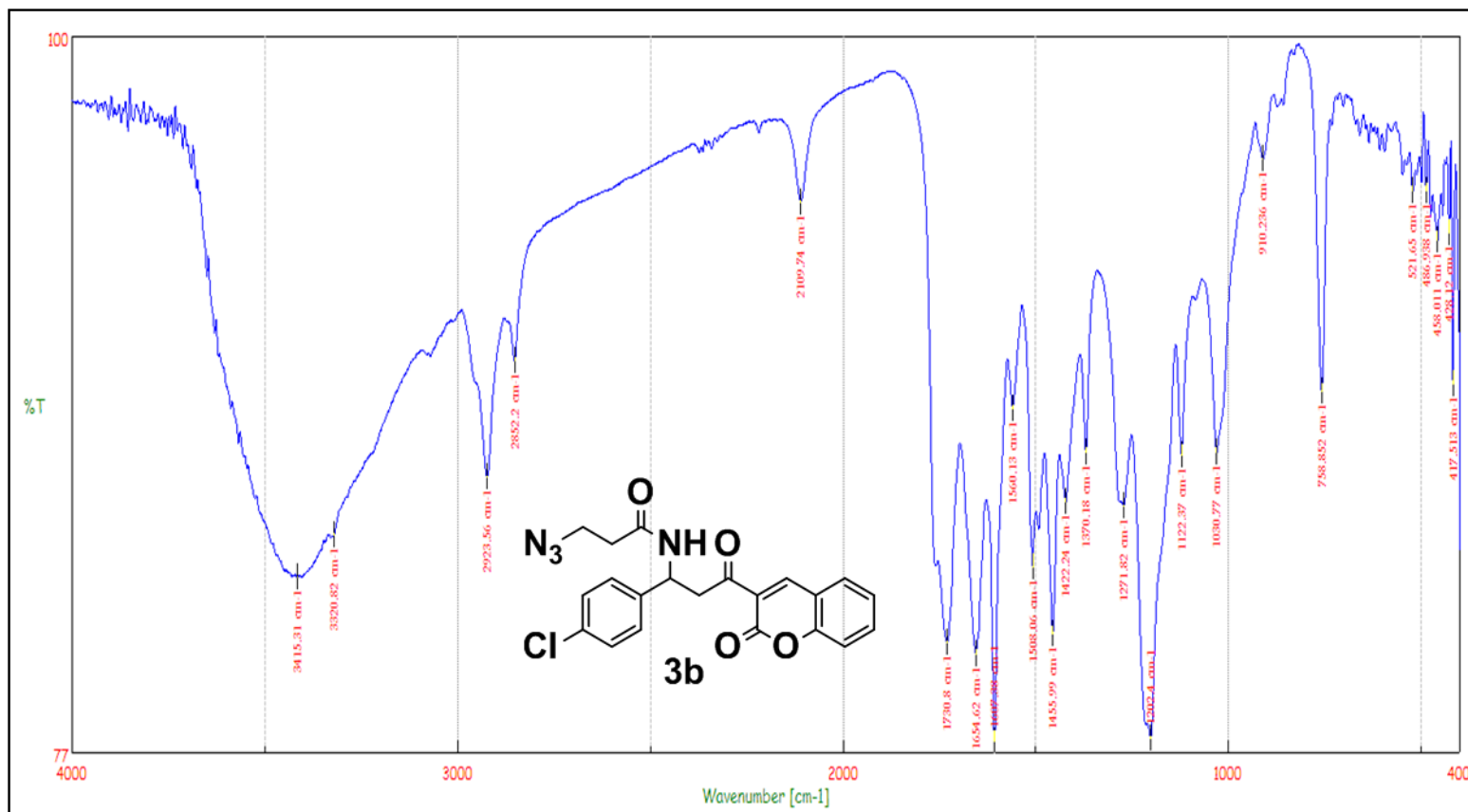
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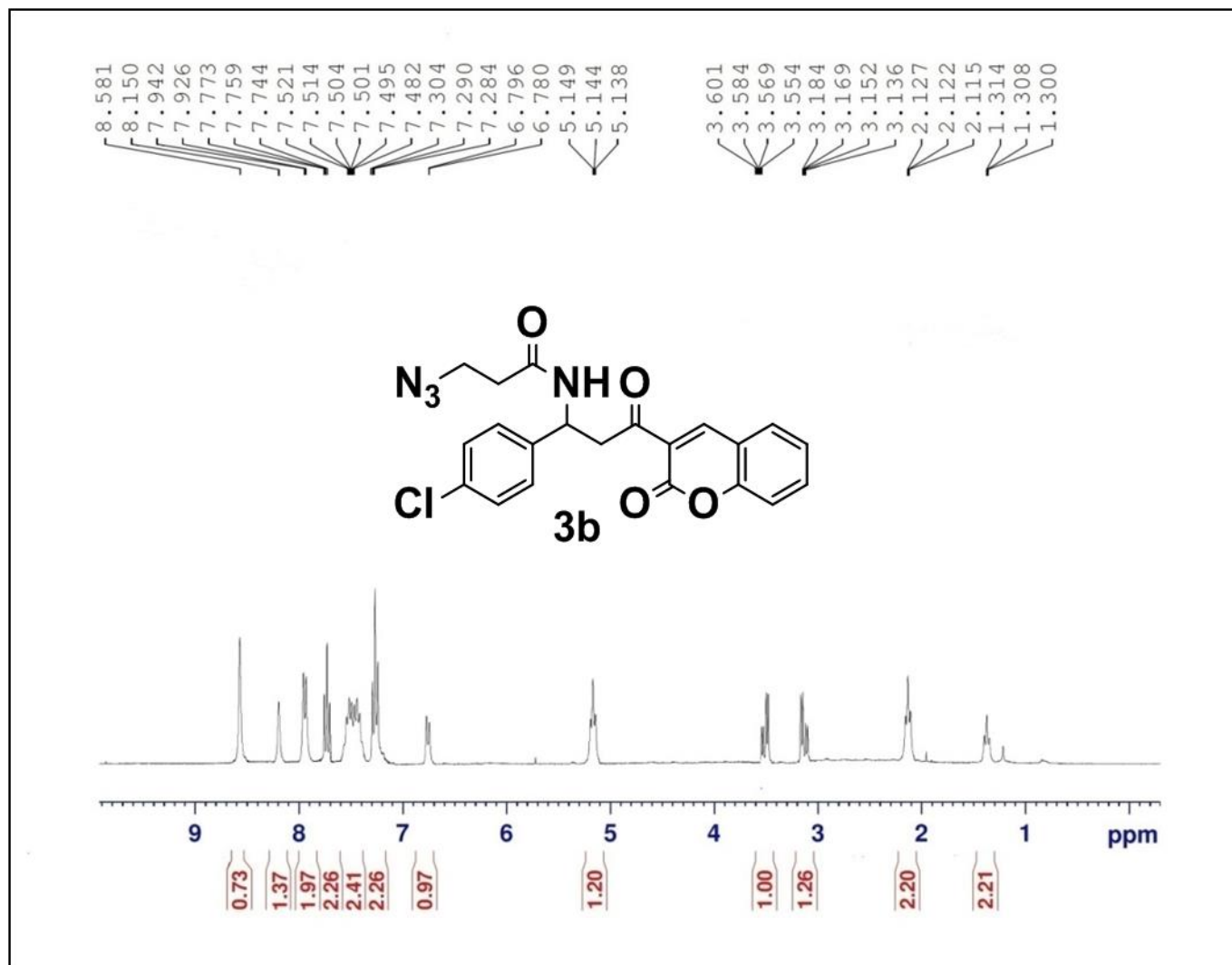
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TD 65536
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NS 200
DS 4
SWH 29761.904 Hz
FIDRES 0.454131 Hz
AQ 1.1010548 sec
RG 32800
DW 16.800 usec
DE 6.00 usec
TE 297.1 K
D1 2.00000000 sec
d11 0.03000000 sec
DELTA 1.89999998 sec
TD0 1

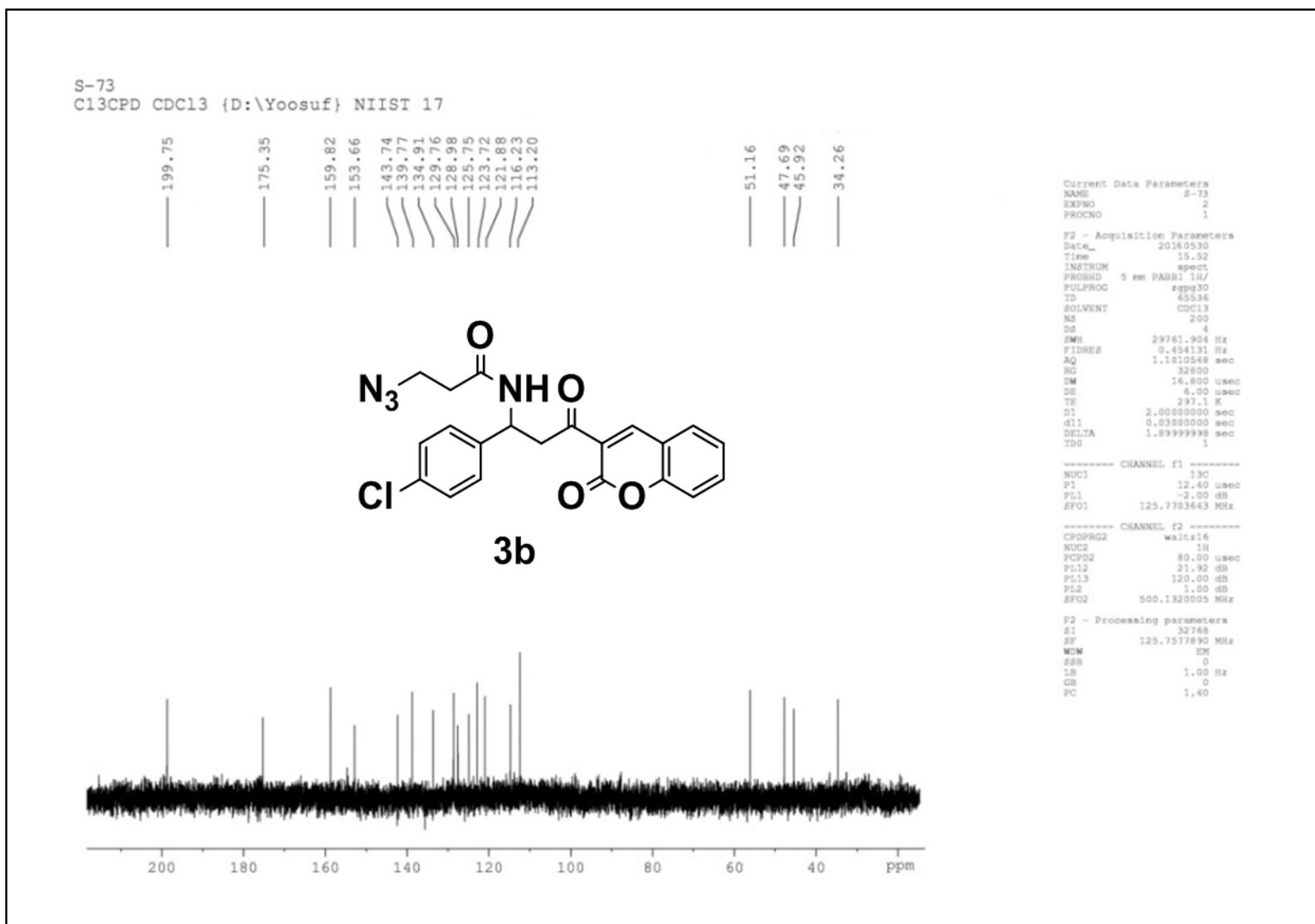
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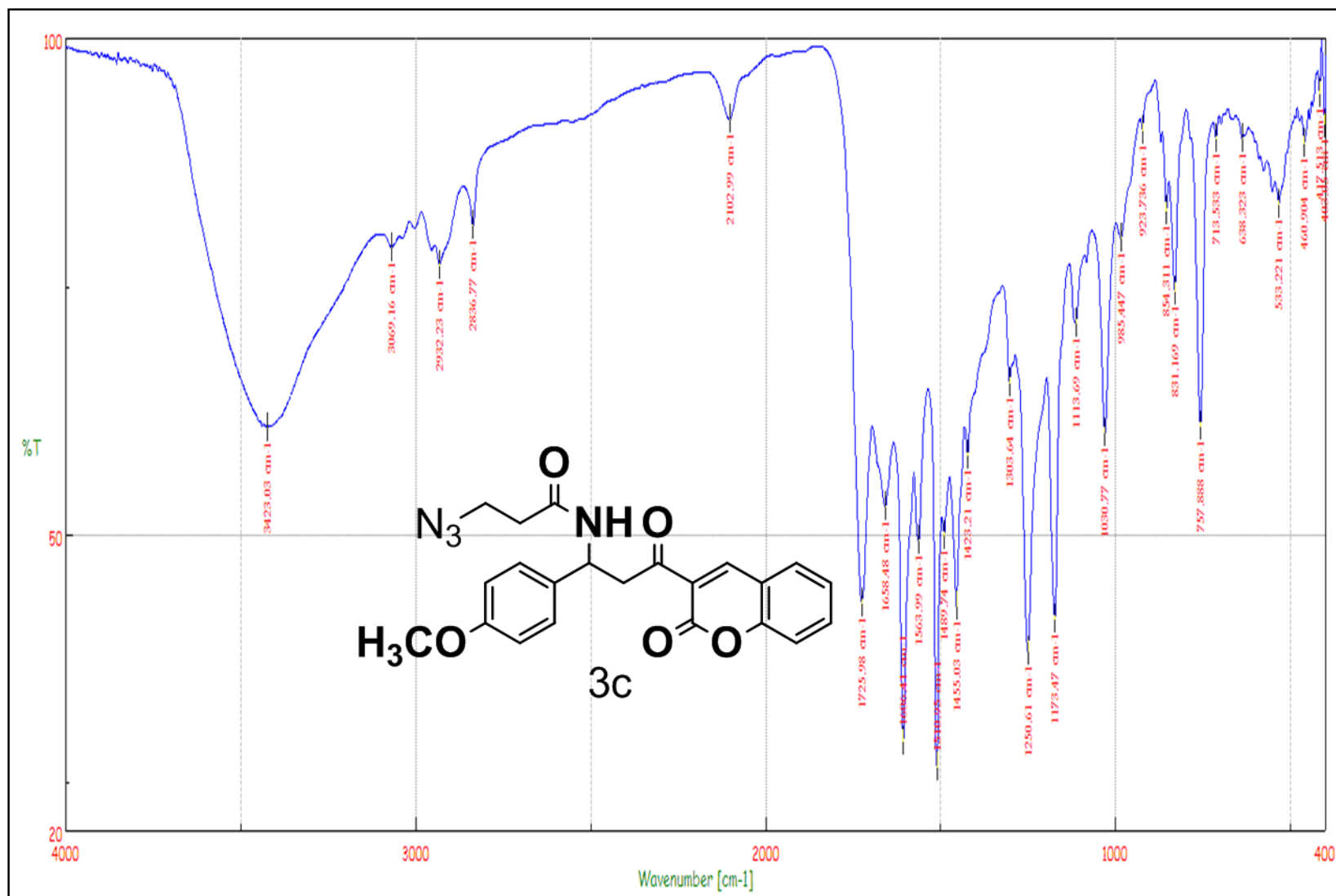
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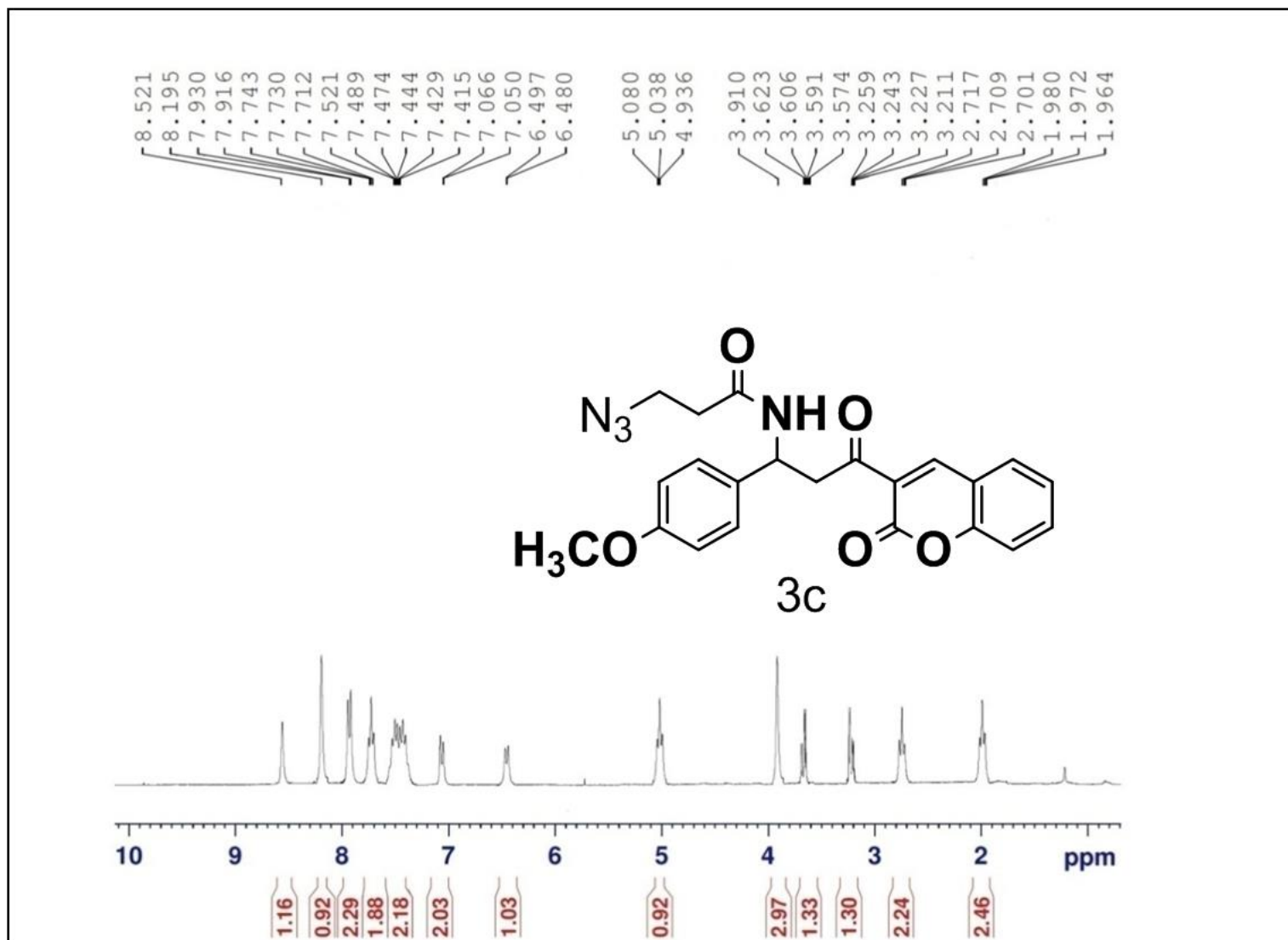
F2 - Processing parameters
SI 32768
SF 125.7577890 MHz
WOW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



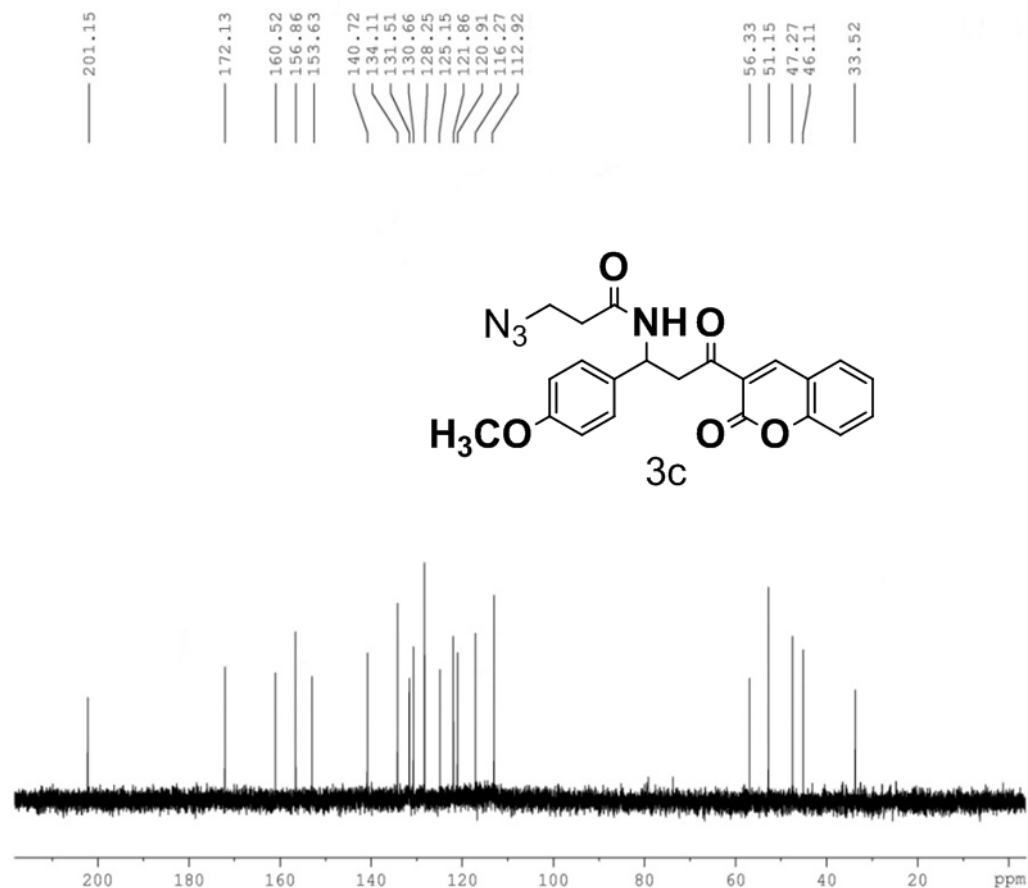








S-5
C13CPD CDC13 {D:\Yoosuf} NIIST 16



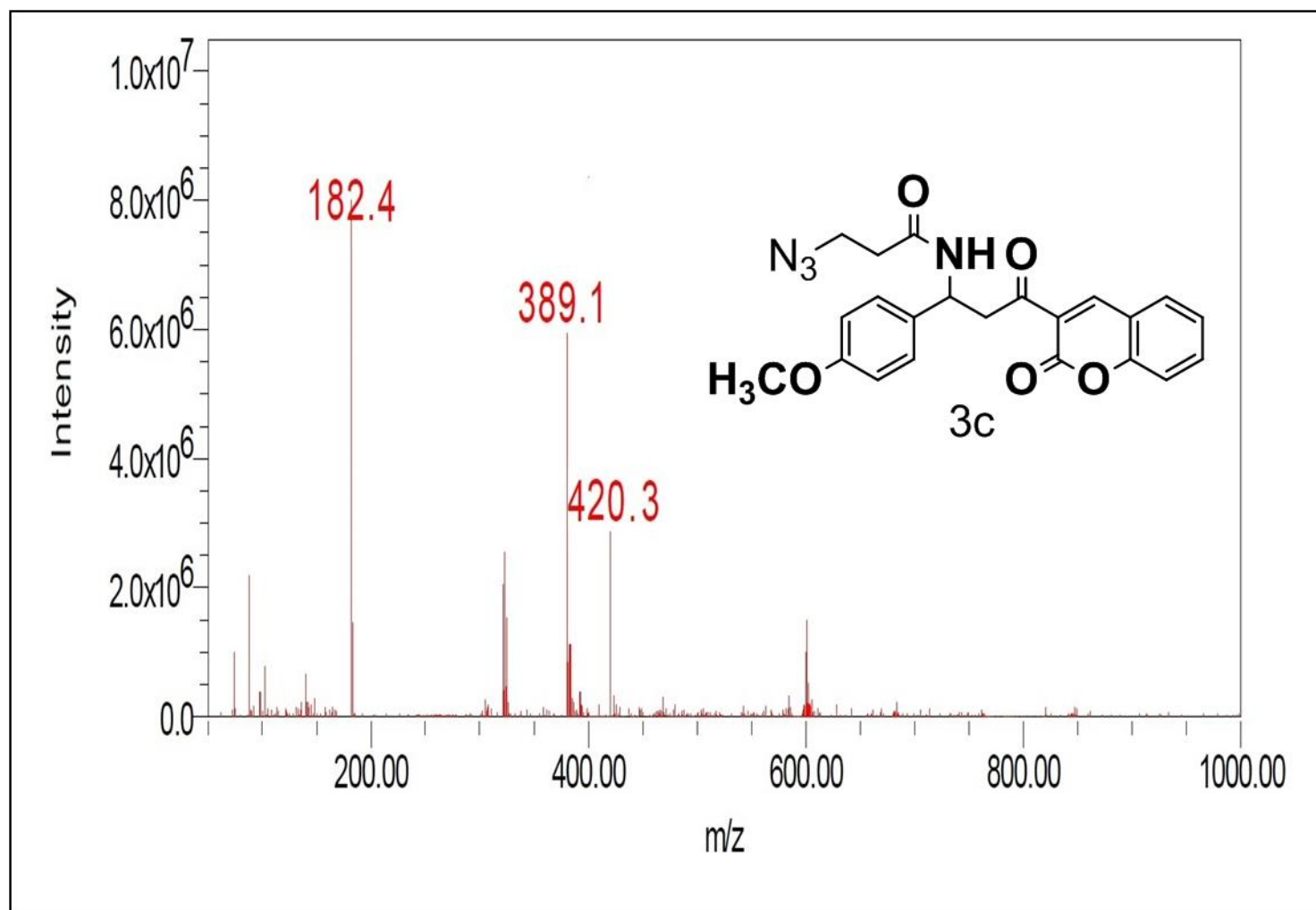
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EXPNO 2
PROCNO 1

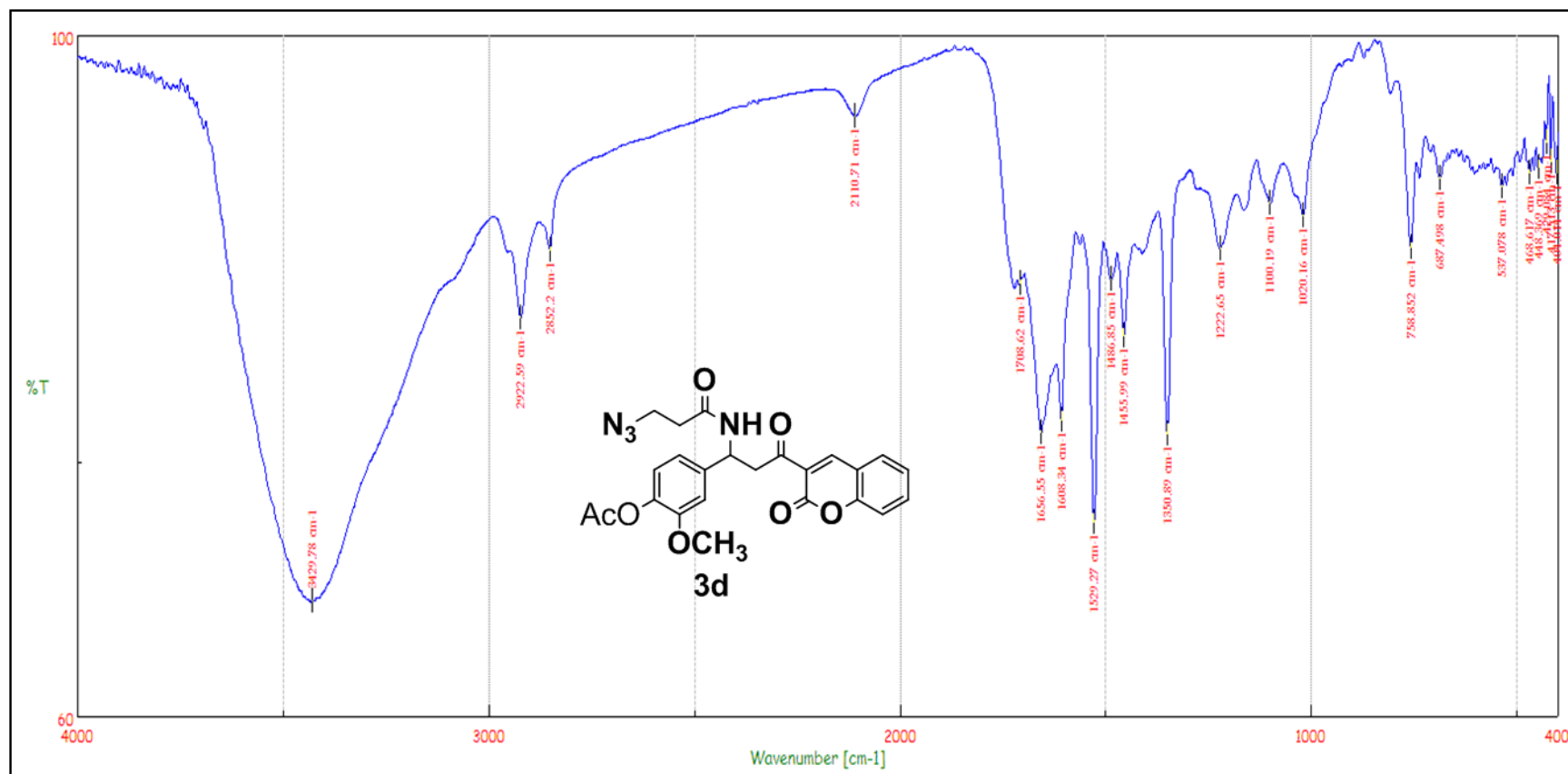
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FIDRES 0.454131 Hz
AQ 1.1010548 sec
RG 32800
DW 16.800 usec
DE 6.00 usec
TE 297.0 K
D1 2.0000000 sec
d11 0.0300000 sec
DELTA 1.8999999 sec
TDO 1

----- CHANNEL f1 -----
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P1 12.40 usec
PL1 -2.00 dB
SFO1 125.7703643 MHz

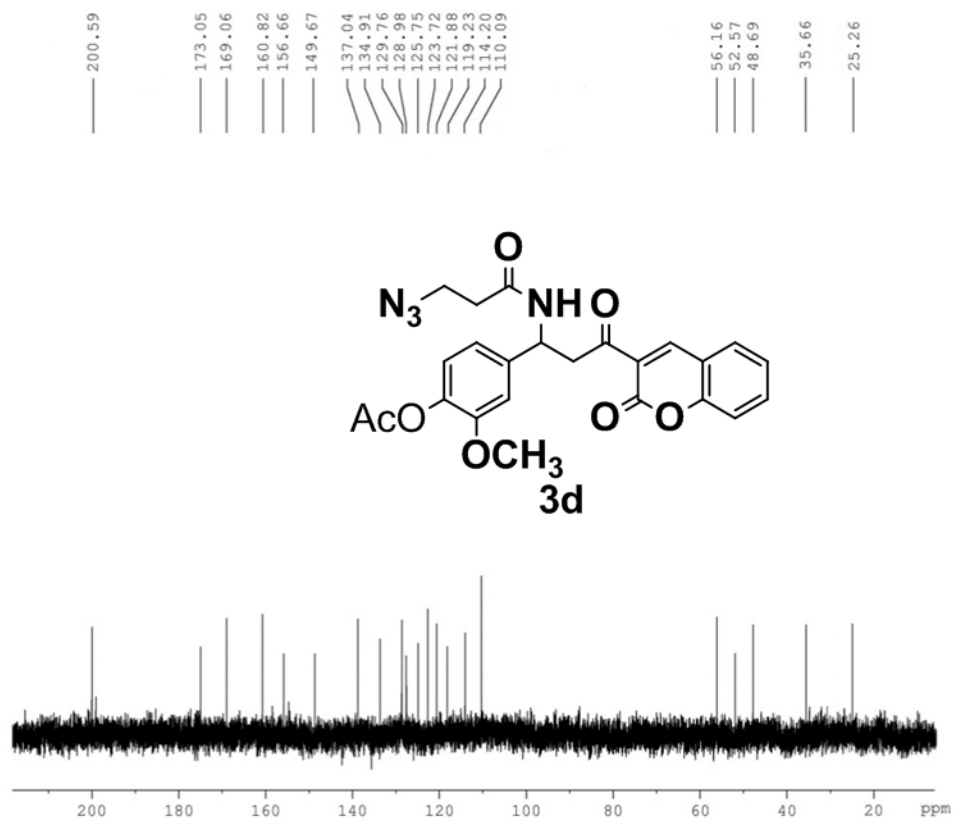
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NUC2 1H
PCPD2 80.00 usec
PL12 21.92 dB
PL13 120.00 dB
PL2 1.00 dB
SFO2 500.1320005 MHz

F2 - Processing parameters
SI 32768
SF 125.7577890 MHz
WCH EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40





S-7
C13CPD CDC13 {D:\Yoosuf} NIIST 16



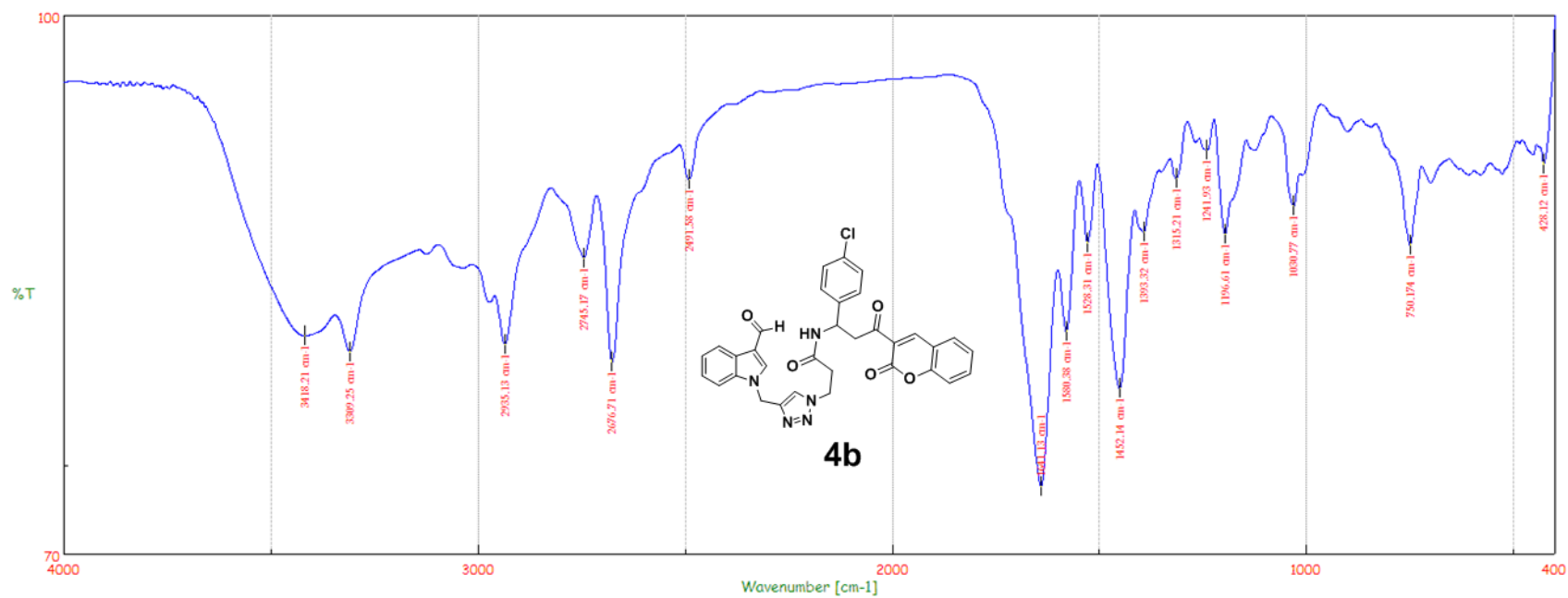
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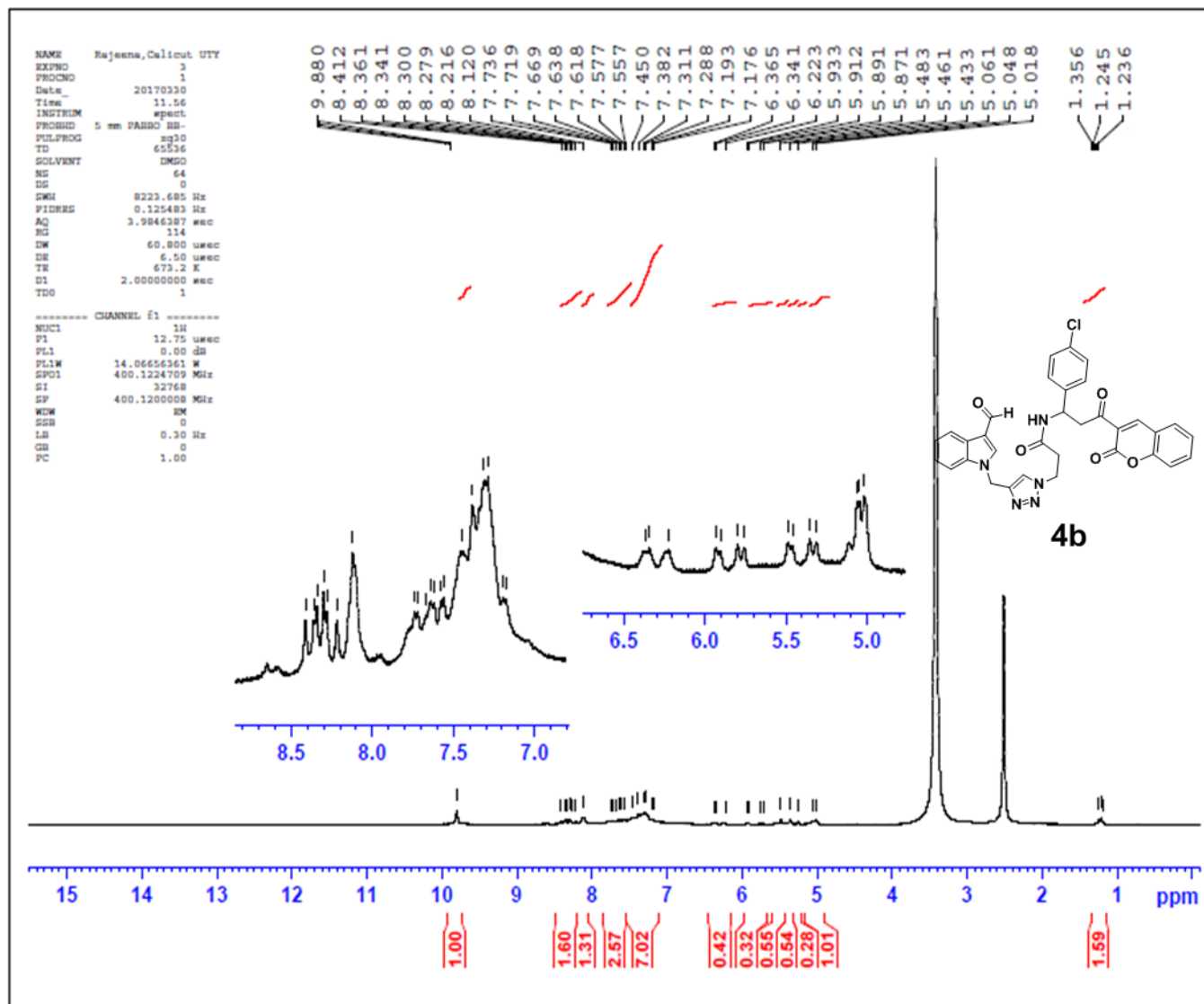
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Date_: 20160530
Time: 15.52
INSTRUM: spect
PROBHD: 5 mm PABBI 1H/
PULPROG: zgpg30
TD: 65536
SOLVENT: CDC13
NS: 200
DS: 4
SWH: 29761.904 Hz
FIDRES: 0.454131 Hz
AQ: 1.1010548 sec
RG: 32800
DM: 16.800 usec
DE: 6.00 usec
TE: 297.1 K
D1: 2.00000000 sec
d11: 0.03000000 sec
DELTA: 1.89999998 sec
TD0: 1

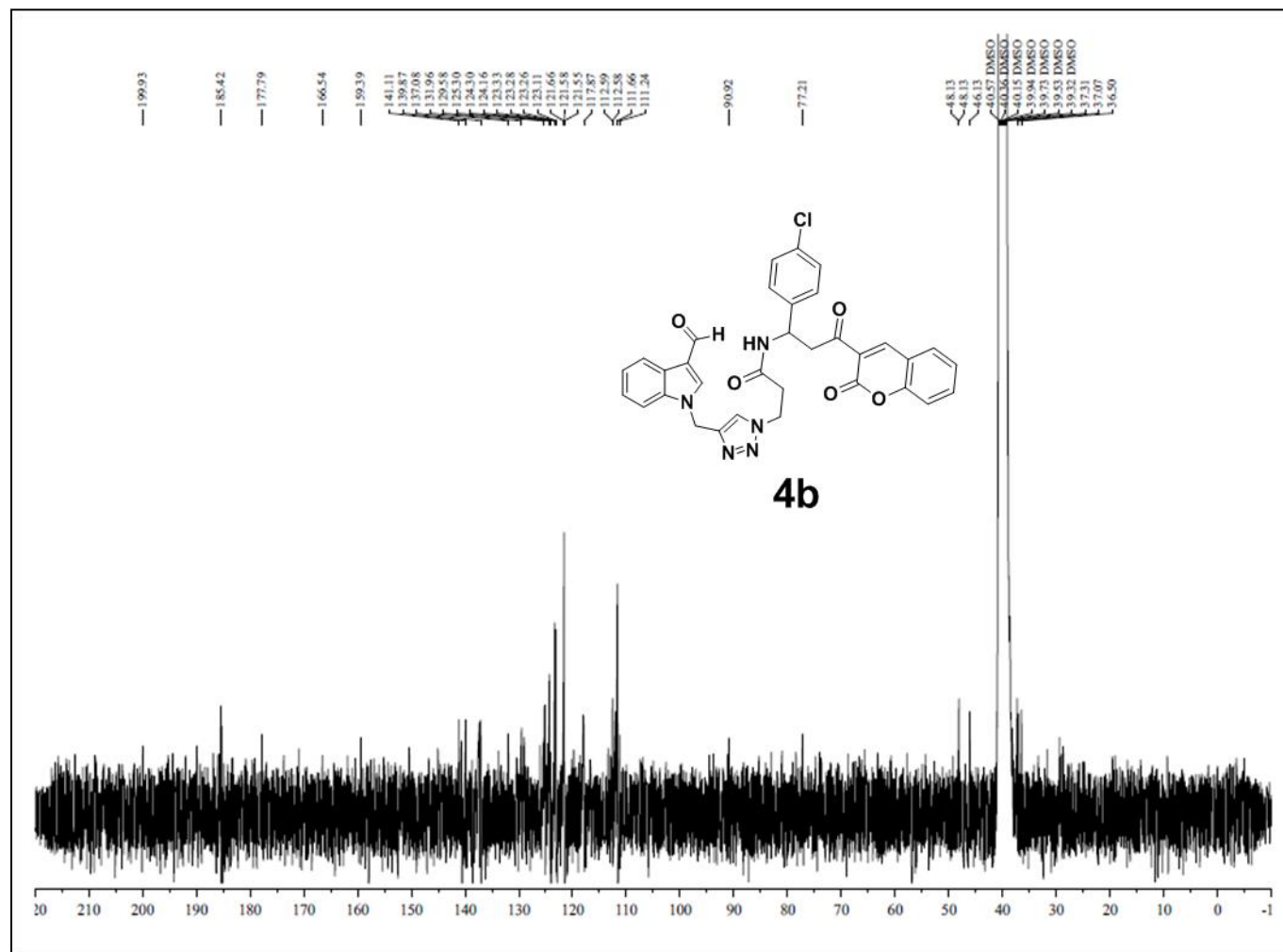
----- CHANNEL f1 -----
NUC1: 13C
P1: 12.40 usec
PL1: -2.00 dB
SFO1: 125.7703643 MHz

----- CHANNEL f2 -----
CPDPRG2: waltz16
NUC2: 1H
PCPD2: 80.00 usec
PL12: 21.92 dB
PL13: 120.00 dB
PL2: 1.00 dB
SFO2: 500.1320005 MHz

F2 - Processing parameters
S1: 32768
SF: 125.7577890 MHz
MCM: EM
SSB: 0
LB: 1.00 Hz
GB: 0
PC: 1.40







Elemental Composition Report

Page 1

Single Mass Analysis

Tolerance = 20.0 PPM / DBE: min = -1.5, max = 50.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions

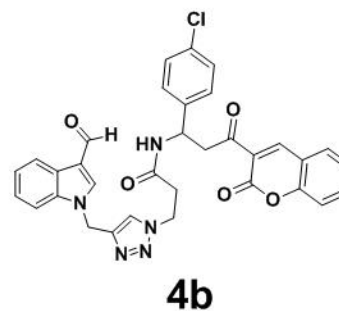
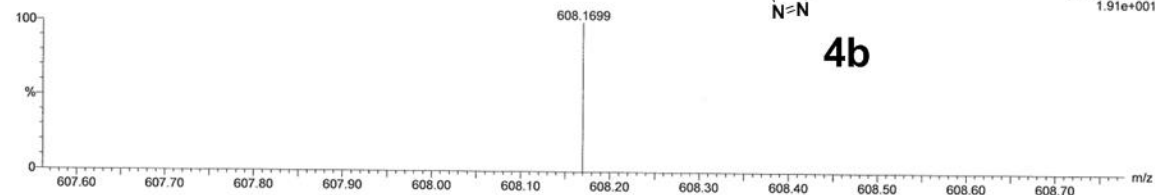
21 formula(e) evaluated with 1 results within limits (up to 50 closest results for each mass)

Elements Used:

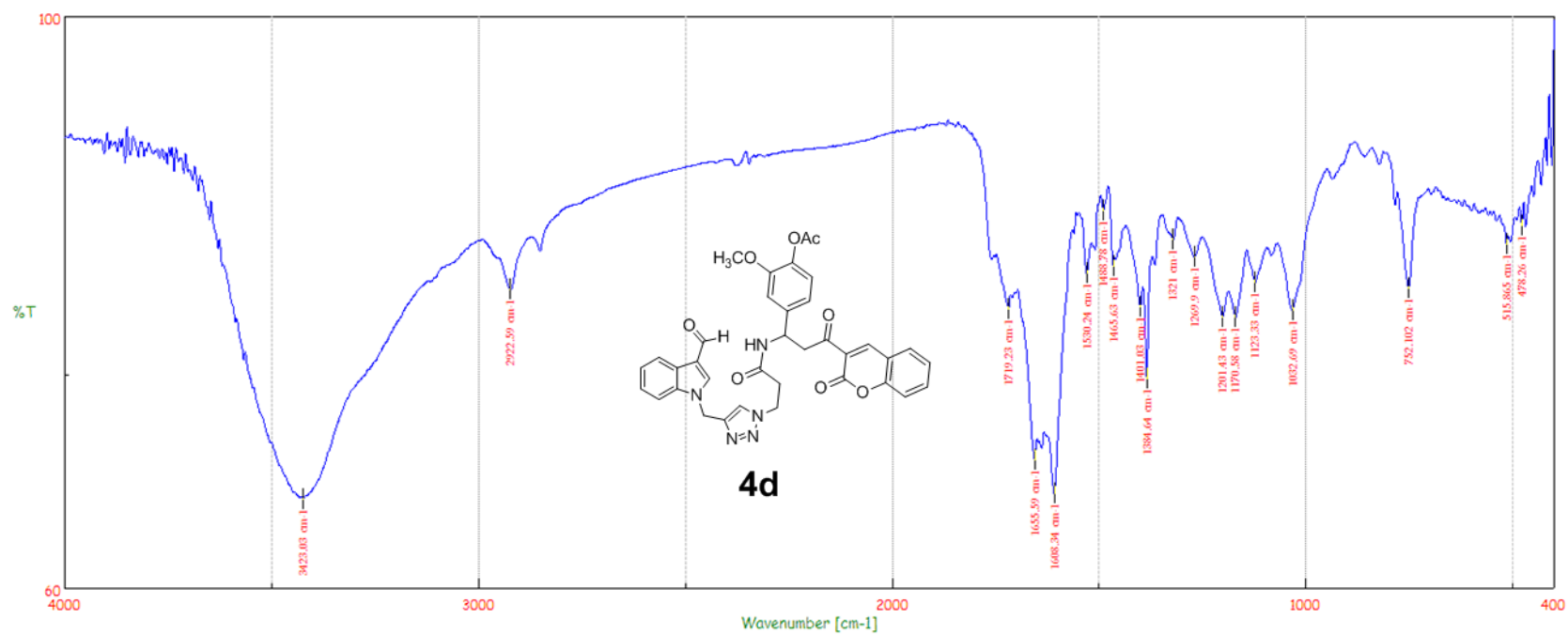
C: 33-33 H: 15-28 N: 2-7 O: 2-6 Cl: 1-2

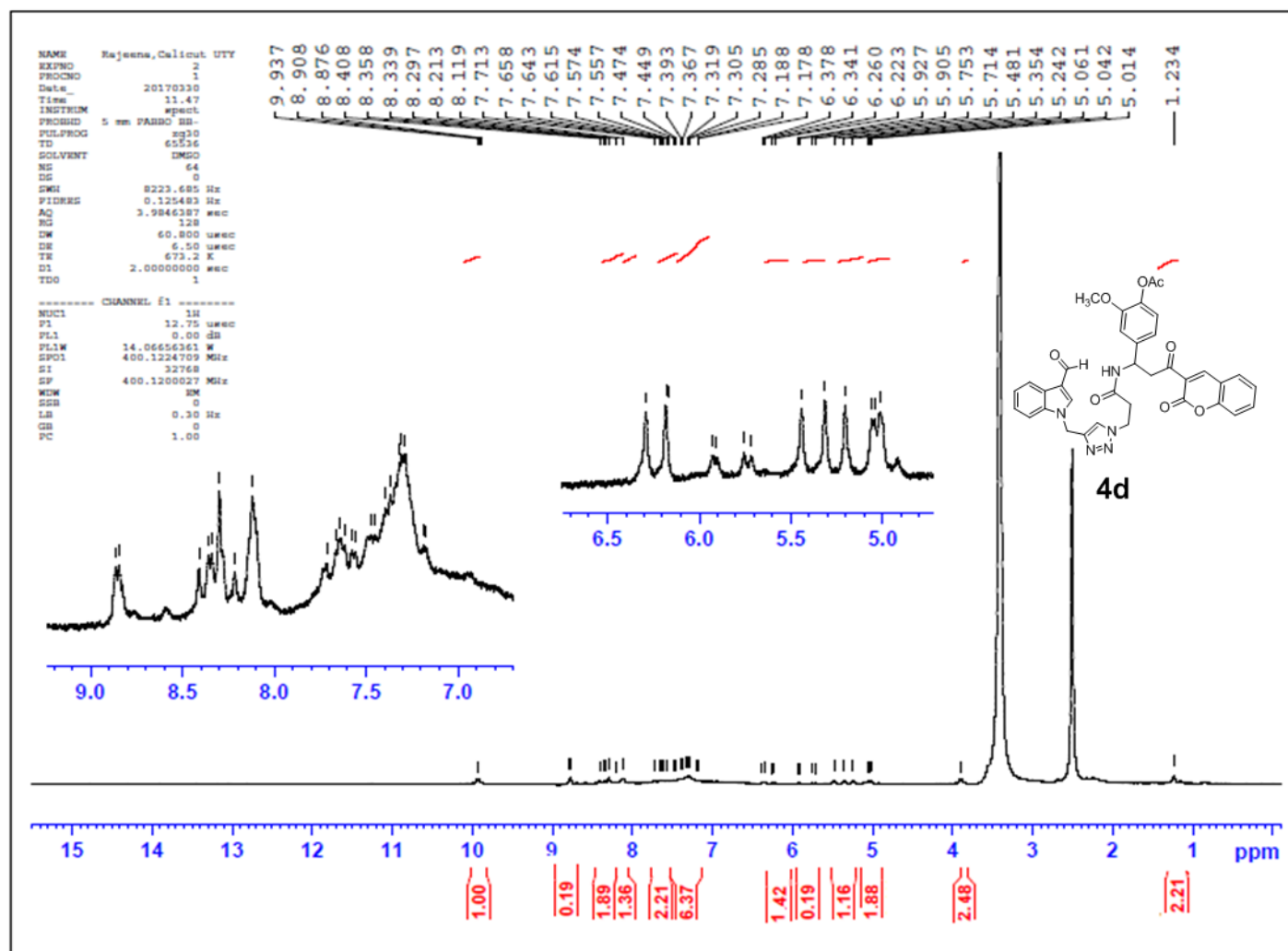
C43H38ClN7O6

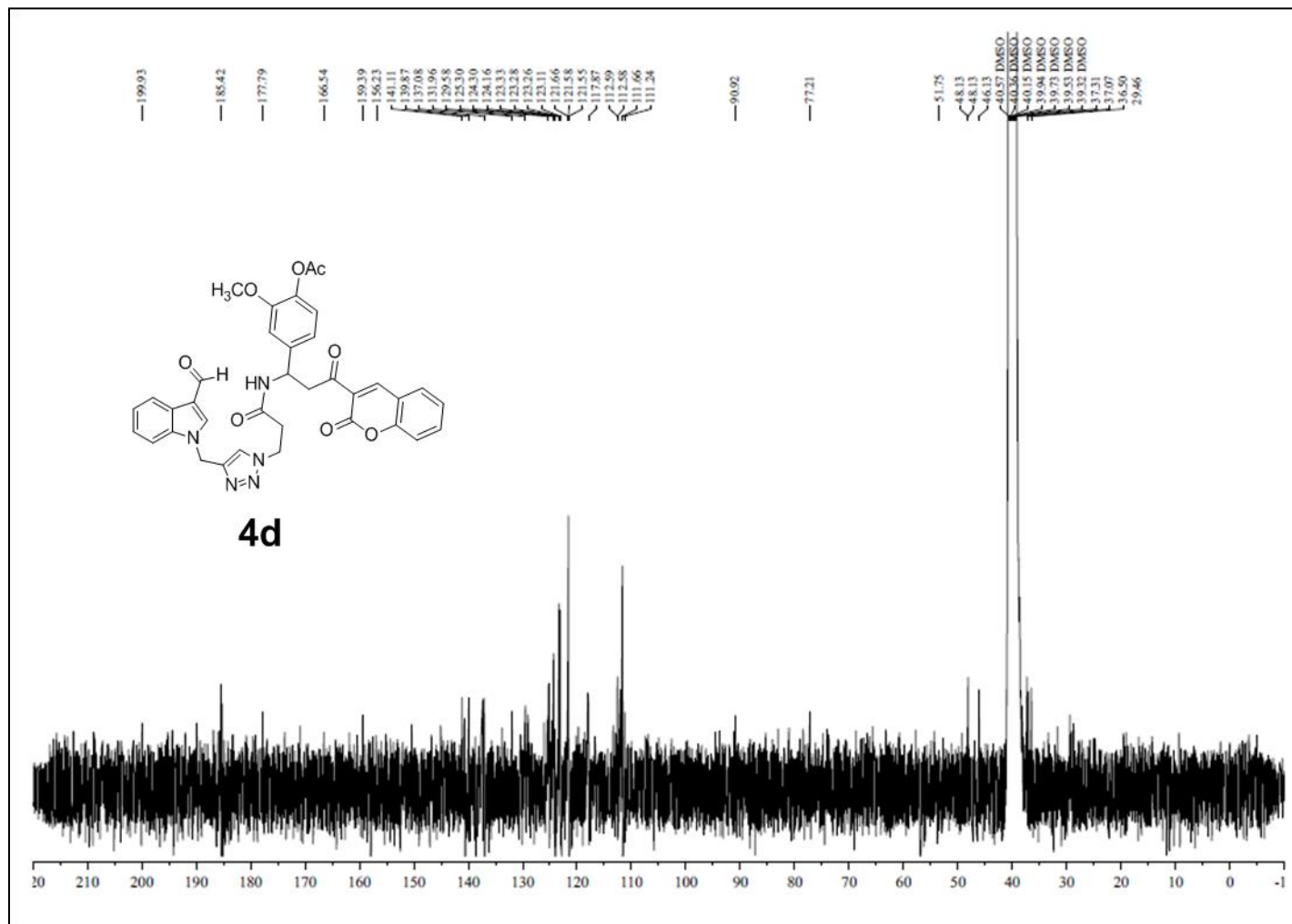
CC1 5 (0.119)

1: TOF MS ES+
1.91e+001

Minimum:									
Maximum:		5.0	20.0	-1.5					
				50.0					
Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (Norm)	Formula		
608.1699	608.1701	-0.2	-0.3	22.5	14.5	0.0	C33	H27	N5 O5 Cl







Elemental Composition Report

Page 1

Single Mass Analysis

Tolerance = 20.0 PPM / DBE: min = -1.5, max = 50.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions

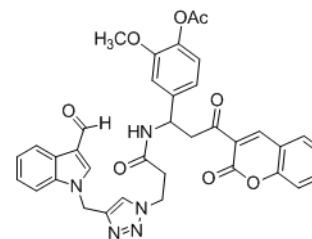
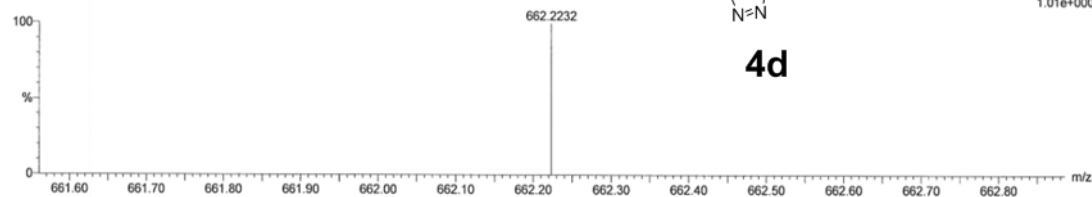
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Elements Used:

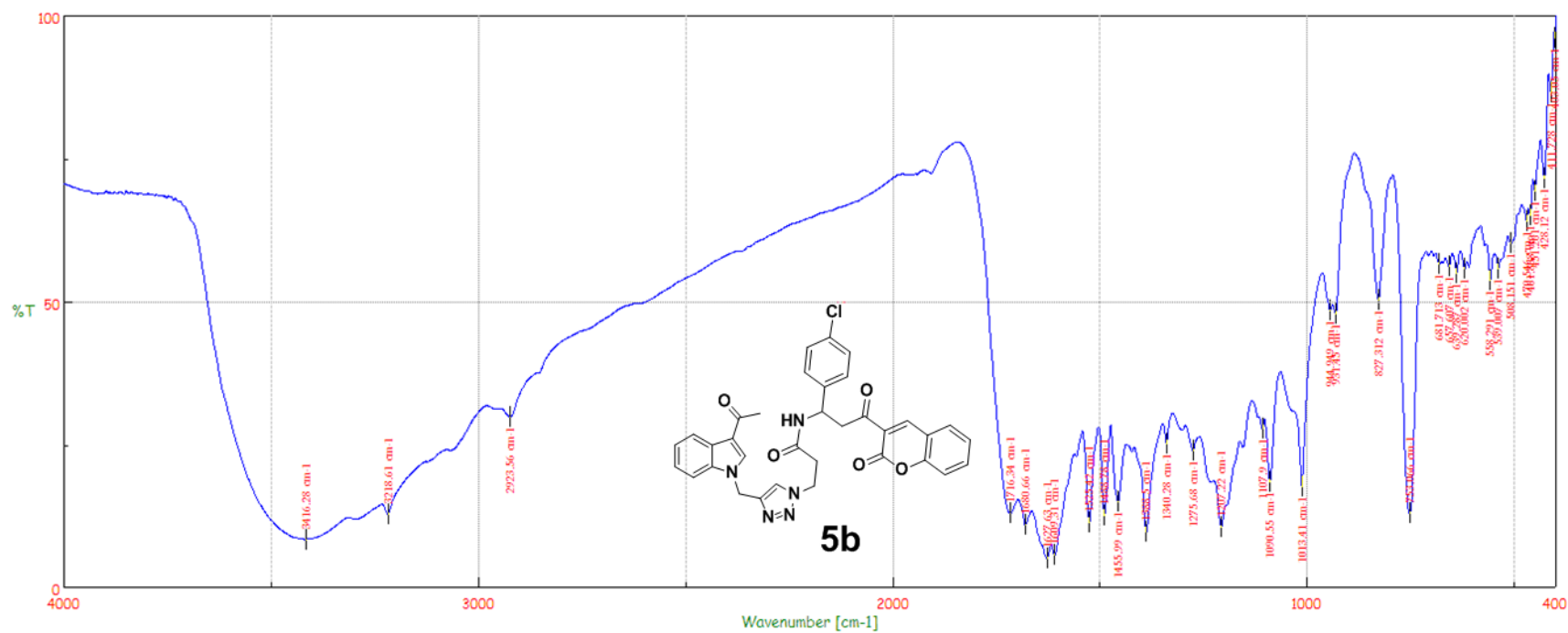
C: 36-36 H: 15-34 N: 2-7 O: 2-9

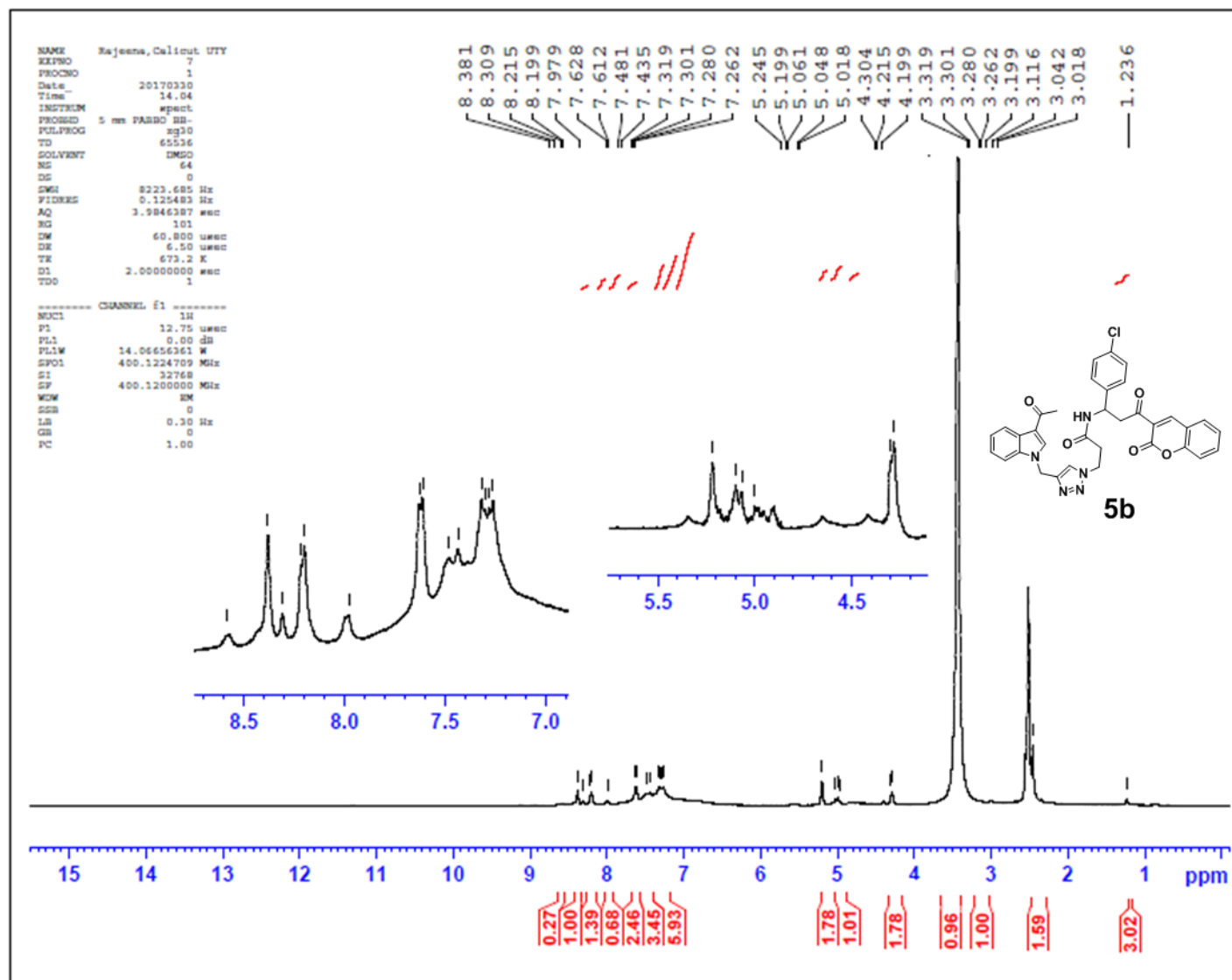
C₄₃H₃₈N₇O₆

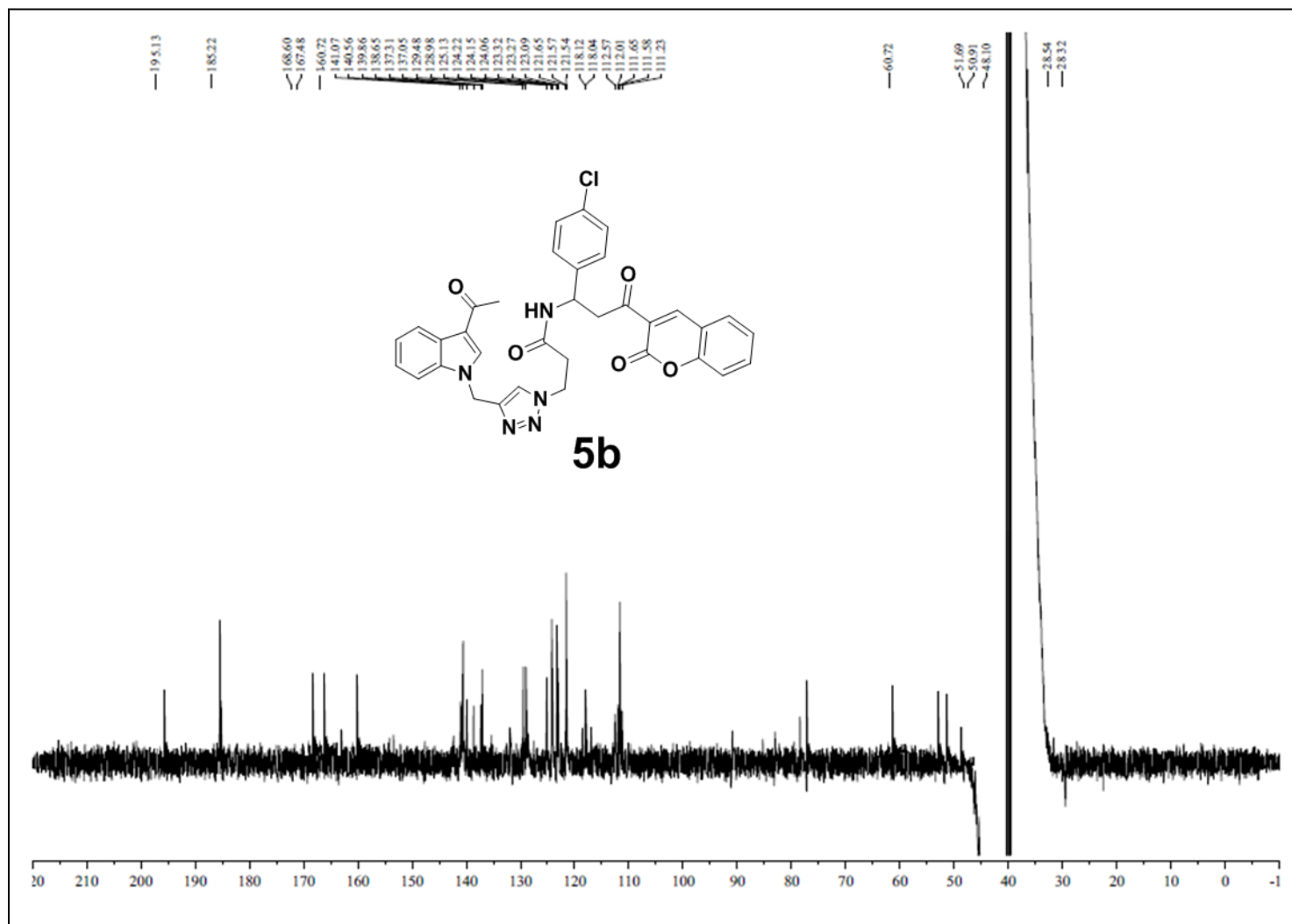
CC4 112 (2.447)

1: TOF MS ES+
1.01e+000**4d**Minimum: -1.5
Maximum: 50.0

Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (Norm)	Formula
662.2232	662.2251	-1.9	-2.9	23.5	12.5	0.0	C ₃₆ H ₃₂ N ₅ O ₈







Elemental Composition Report

Page 1

Single Mass Analysis

Tolerance = 20.0 PPM / DBE: min = -1.5, max = 50.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions

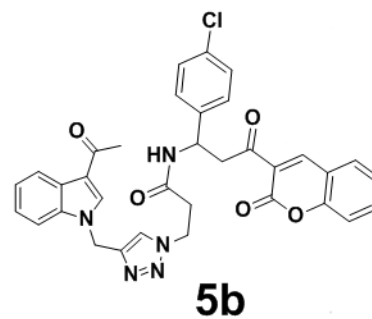
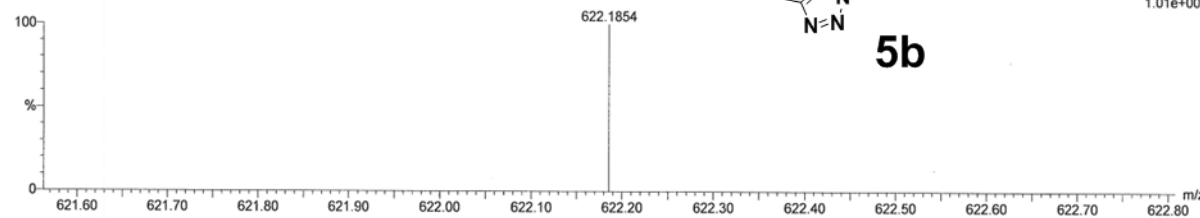
21 formula(e) evaluated with 1 results within limits (up to 50 closest results for each mass)

Elements Used:

C: 34-34 H: 15-30 N: 2-7 O: 2-6 Cl: 1-2

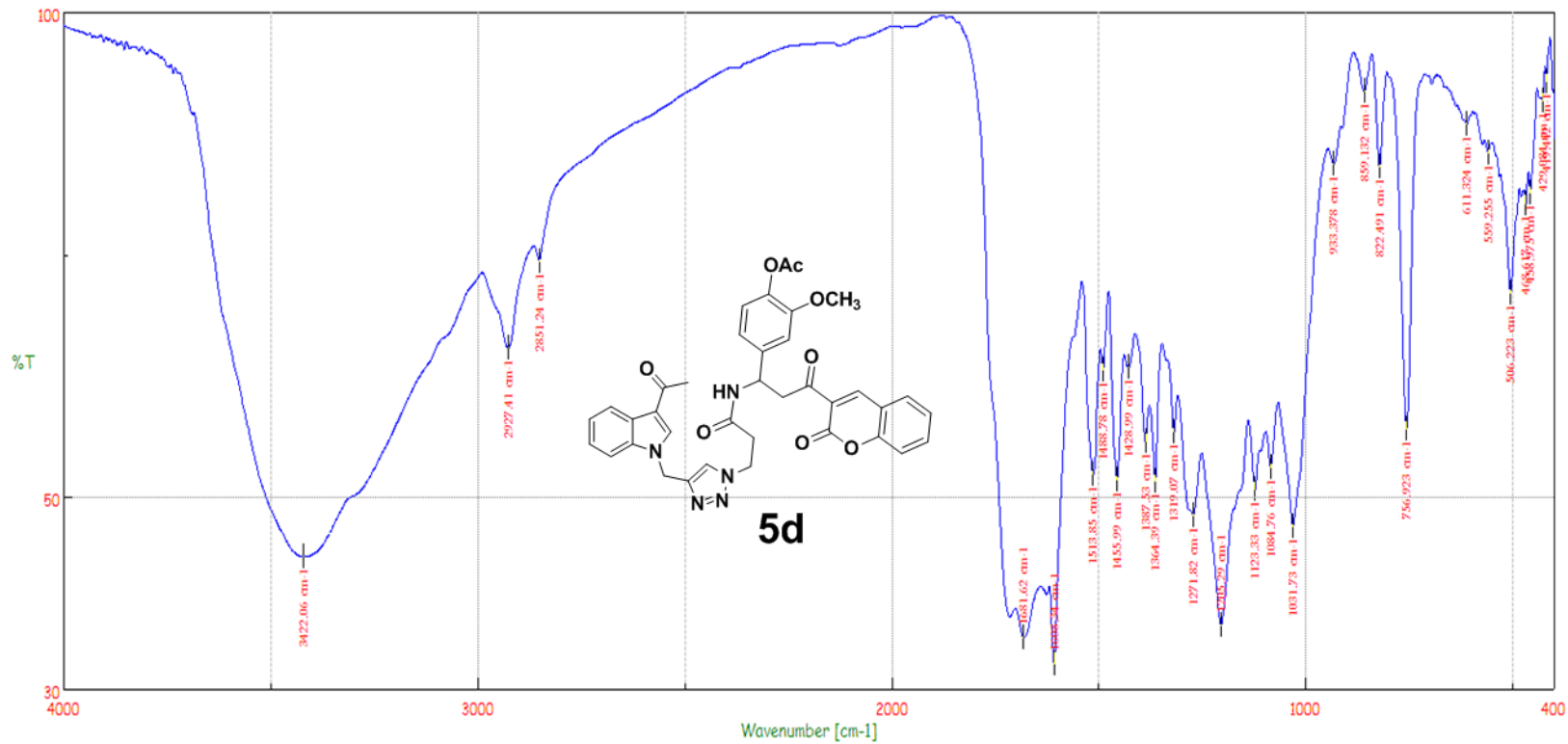
C₄₃H₃₈ClN₇O₆

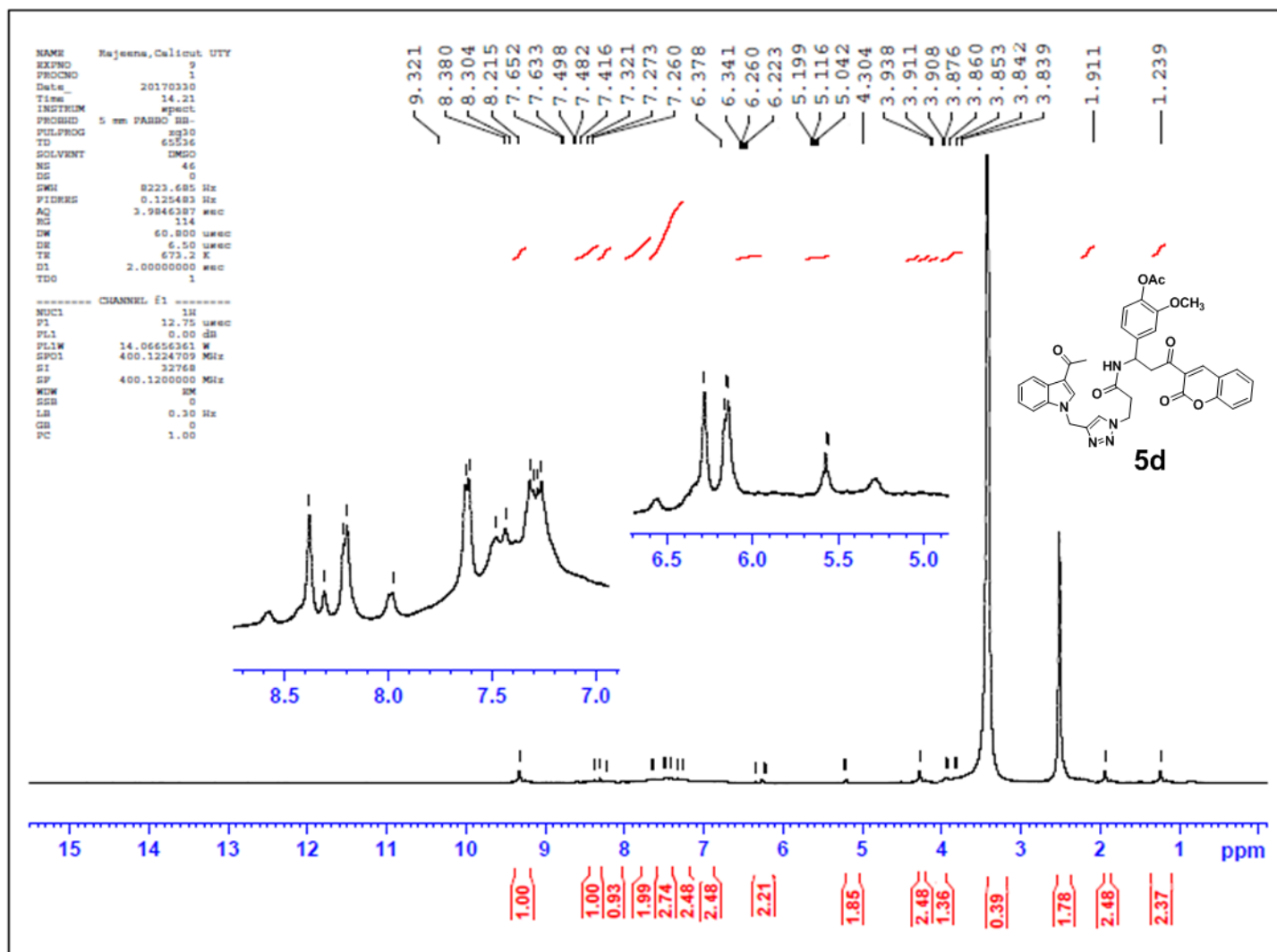
CC14 8 (0.175)

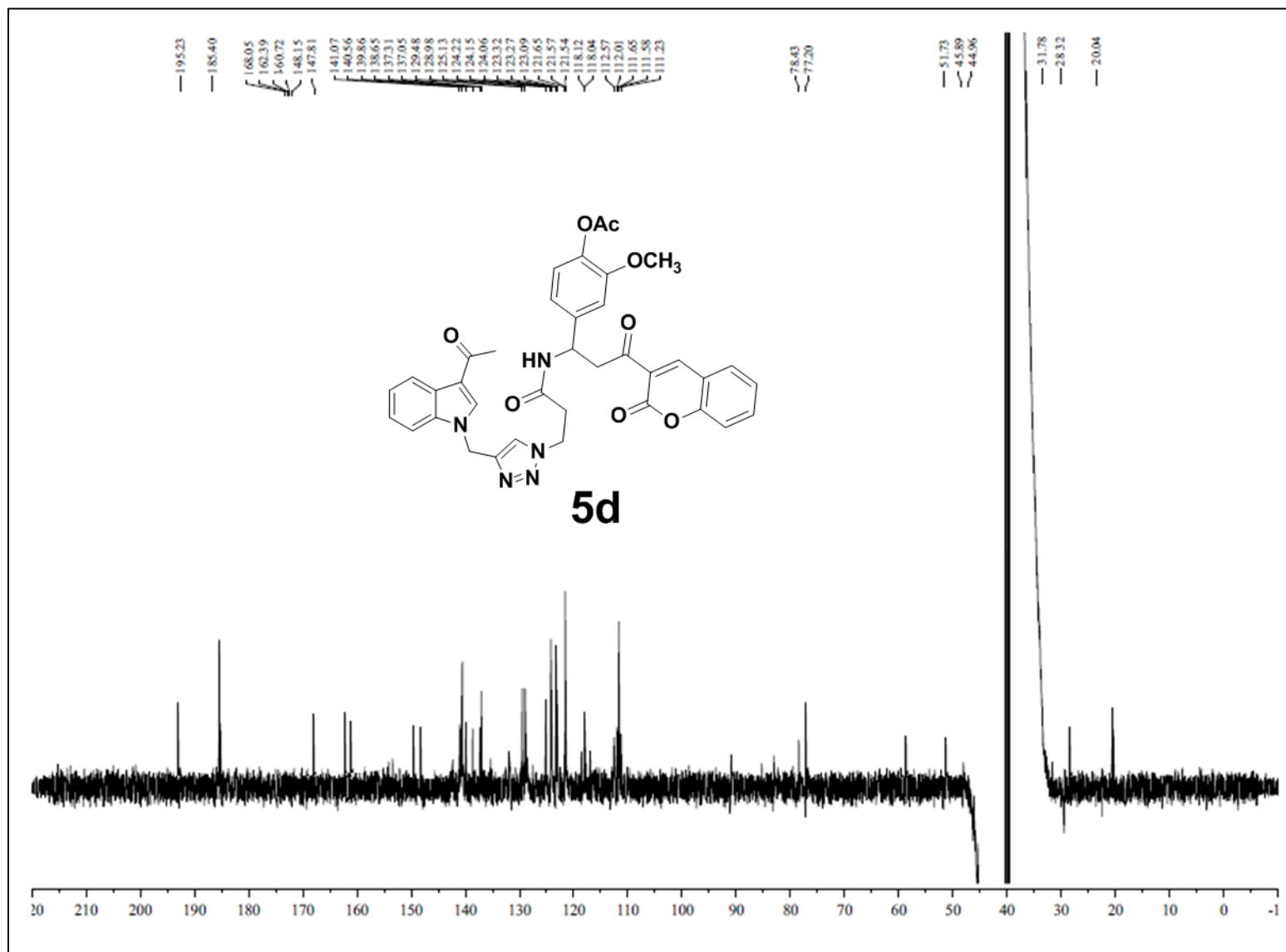
1: TOF MS ES+
1.01e+000

Minimum: -1.5
Maximum: 50.0

Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (Norm)	Formula
622.1854	622.1857	-0.3	-0.5	22.5	12.1	0.0	C ₃₄ H ₂₉ N ₅ O ₅ Cl







Elemental Composition Report

Page 1

Single Mass Analysis

Tolerance = 20.0 PPM / DBE: min = -1.5, max = 50.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions

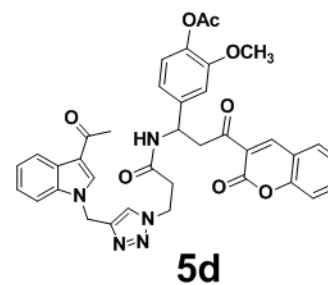
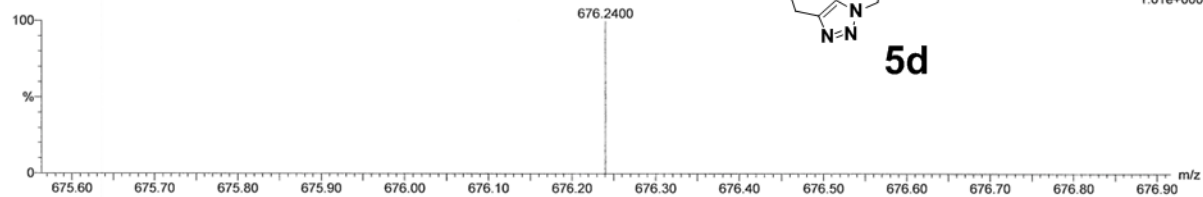
24 formula(e) evaluated with 1 results within limits (up to 50 closest results for each mass)

Elements Used:

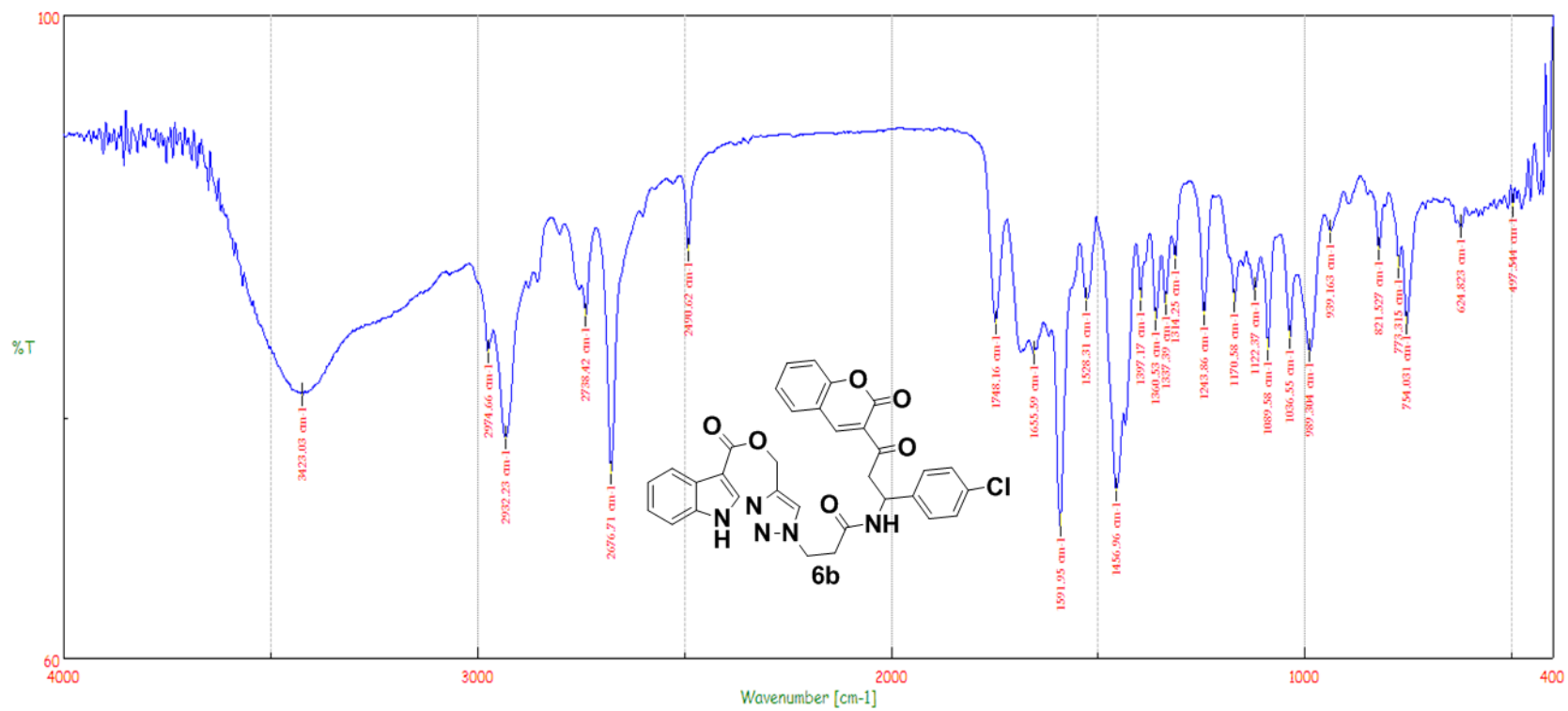
C: 37-37 H: 15-34 N: 2-7 O: 2-9

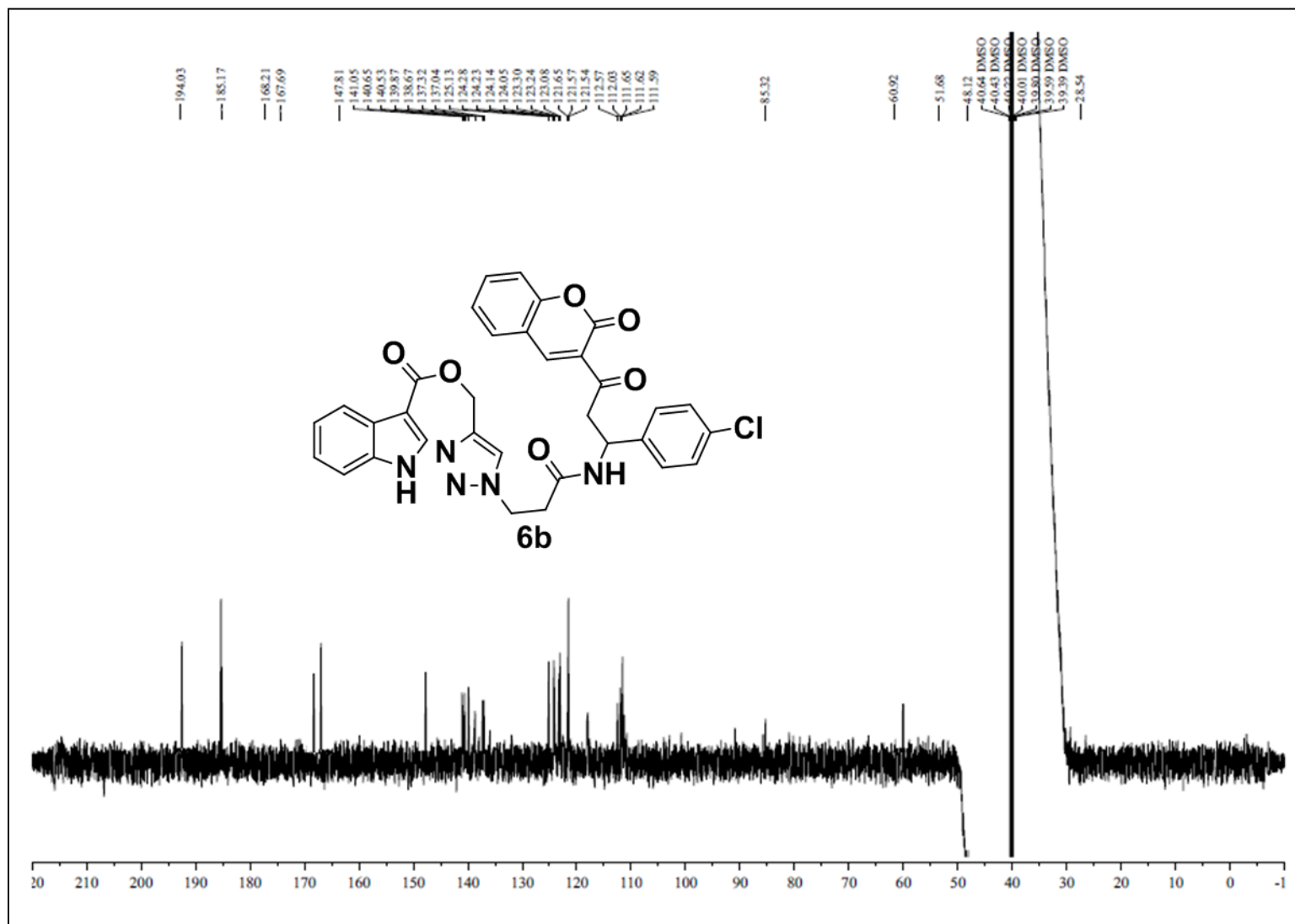
C₄₃H₃₈CIN₇O₆

CC16 38 (0.837)

1: TOF MS ES+
1.01e+000

Minimum:									
Maximum:		5.0	20.0	-1.5					
				50.0					
Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (Norm)	Formula		
676.2400	676.2407	-0.7	-1.0	23.5	12.5	0.0	C37	H34	N5 O8





Elemental Composition Report

Page 1

Single Mass Analysis

Tolerance = 20.0 PPM / DBE: min = -1.5, max = 50.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions

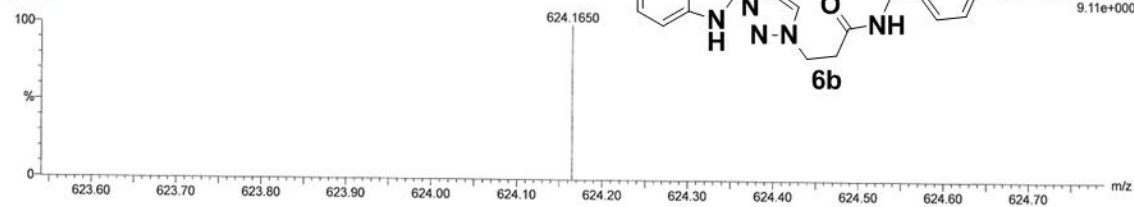
19 formula(e) evaluated with 1 results within limits (up to 50 closest results for each mass)

Elements Used:

C: 33-33 H: 15-27 N: 2-7 O: 2-6 Cl: 1-2

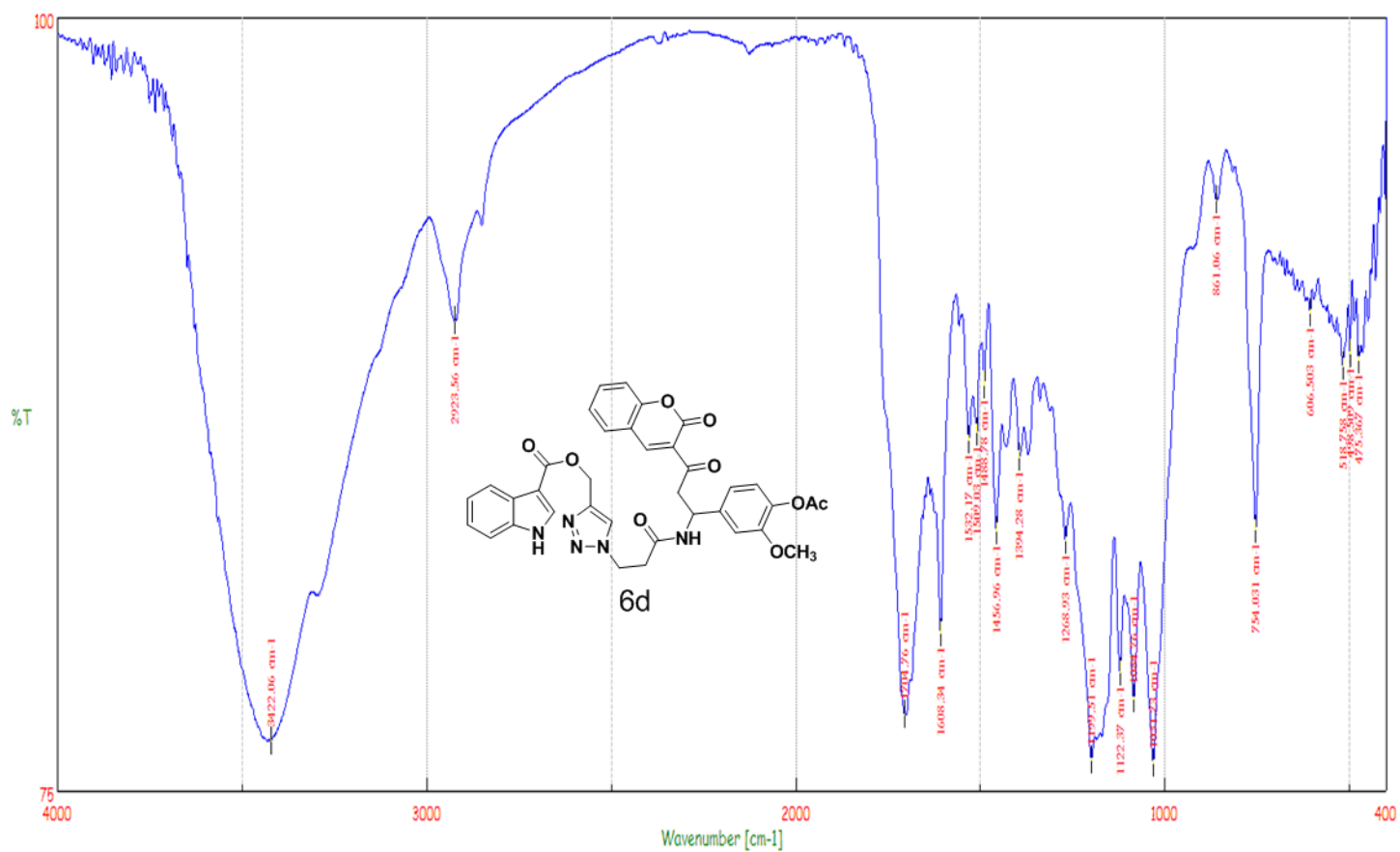
C₄₃H₃₈ClN₇O₆

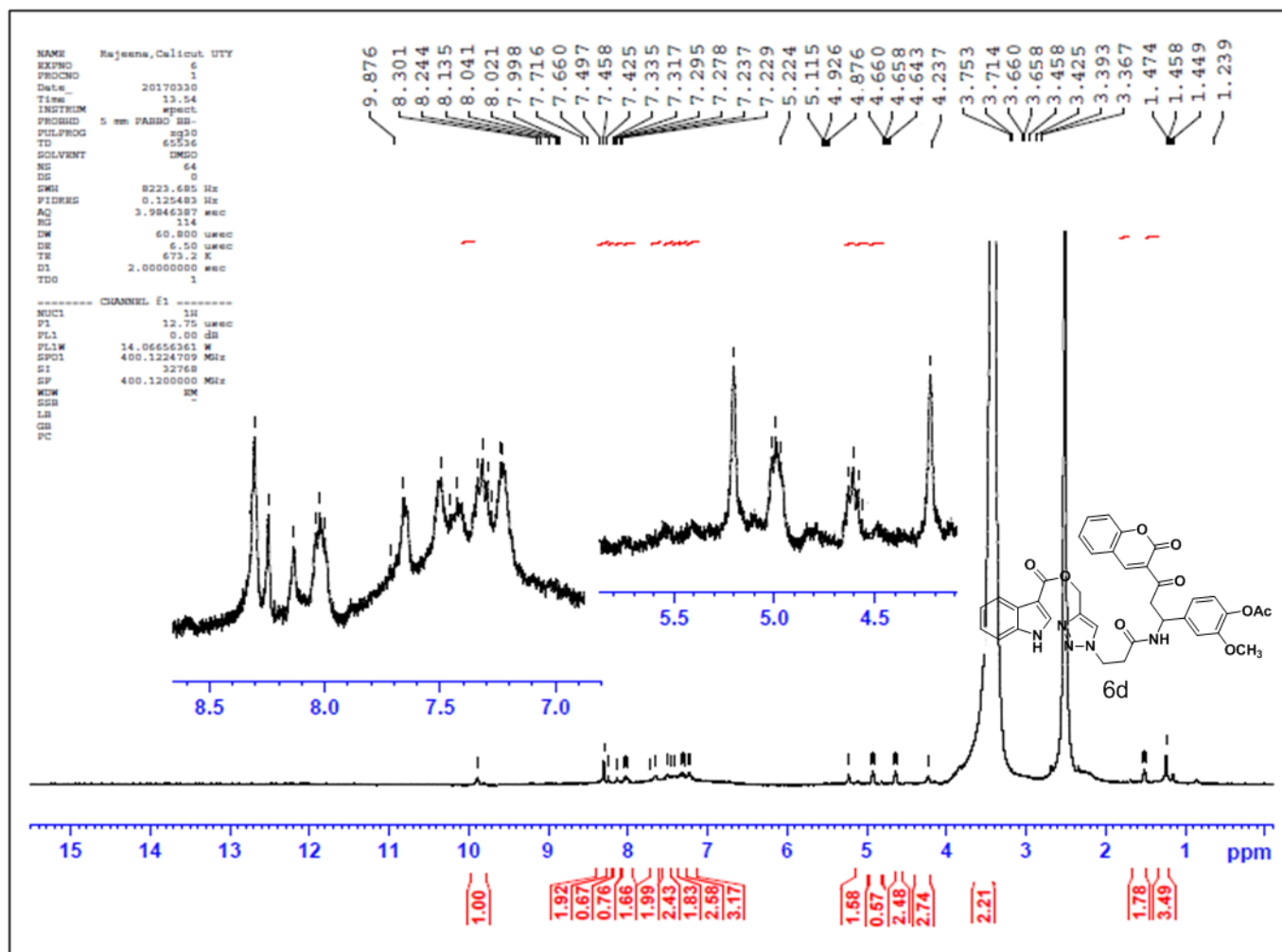
CC6 5 (0.119)

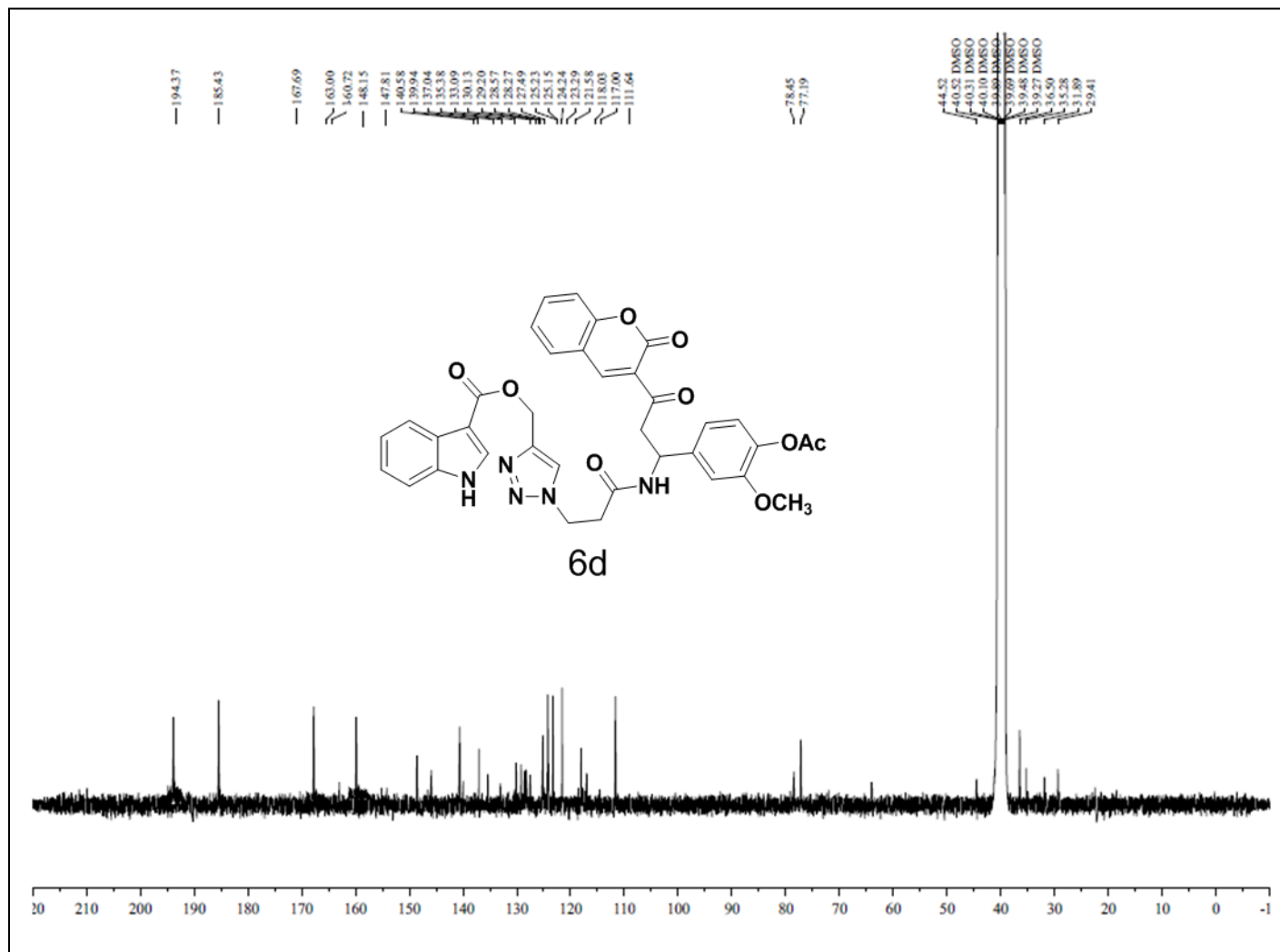


Minimum: -1.5
Maximum: 5.0 20.0 50.0

Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (Norm)	Formula
624.1650	624.1650	0.0	0.0	22.5	12.6	0.0	C ₃₃ H ₂₇ N ₅ O ₆ Cl







Elemental Composition Report

Page 1

Single Mass Analysis

Tolerance = 20.0 PPM / DBE: min = -1.5, max = 50.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 3

Monoisotopic Mass, Even Electron Ions

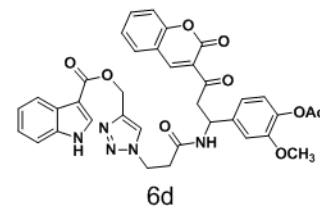
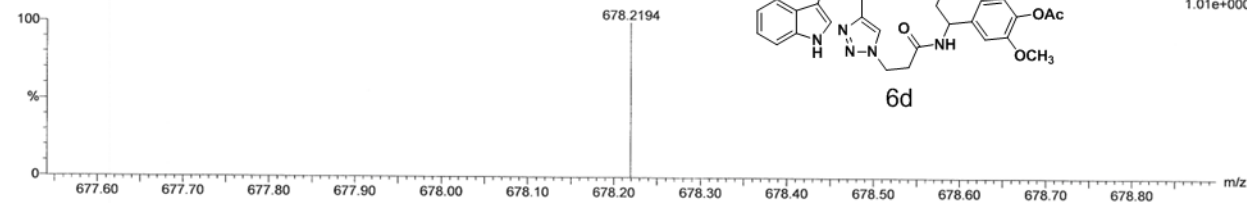
27 formula(e) evaluated with 1 results within limits (up to 50 closest results for each mass)

Elements Used:

C: 36-36 H: 15-32 N: 2-7 O: 2-9

C₄₃H₃₈N₇O₆

678.2194 (1.922)

1: TOF MS ES+
1.01e+000

Minimum:				-1.5				
Maximum:		5.0	20.0	50.0				
Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	i-FIT (Norm)	Formula	
678.2194	678.2200	-0.6	-0.9	23.5	12.5	0.0	C ₃₆ H ₃₂ N ₅ O ₉	

Figure S1. The super imposition of the compound 4d, 5d, 6a and 6d with parent ligand in the active site of 1G11.

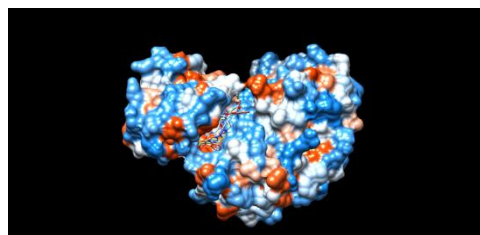


Figure S2. Chemical structure of some Triazole containing potent anticancer compounds

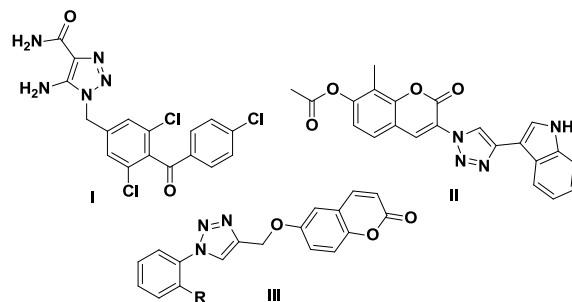


Table S3. Drug property descriptors of the compounds from molinspiration property calculation service

Compound	Molecular weight	miLog p	TPSA	NO. of atoms	n O N	nOHNH
4a	608.05	3.80	129.10	44	10	1
4b	608.05	3.85	129.10	44	10	1
4c	603.63	3.23	138.34	45	11	1
4d	619.63	2.51	158.56	46	12	2
5a	622.08	3.91	129.10	45	10	1
5b	622.08	3.96	129.10	45	10	1
5c	617.66	3.34	138.34	46	11	1
5d	633.66	2.62	158.56	47	12	2
6a	608.05	3.68	139.96	44	10	2
6b	608.05	3.73	139.96	44	10	2
6c	603.63	3.10	149.19	45	11	2
6d	619.63	2.39	169.42	46	12	3

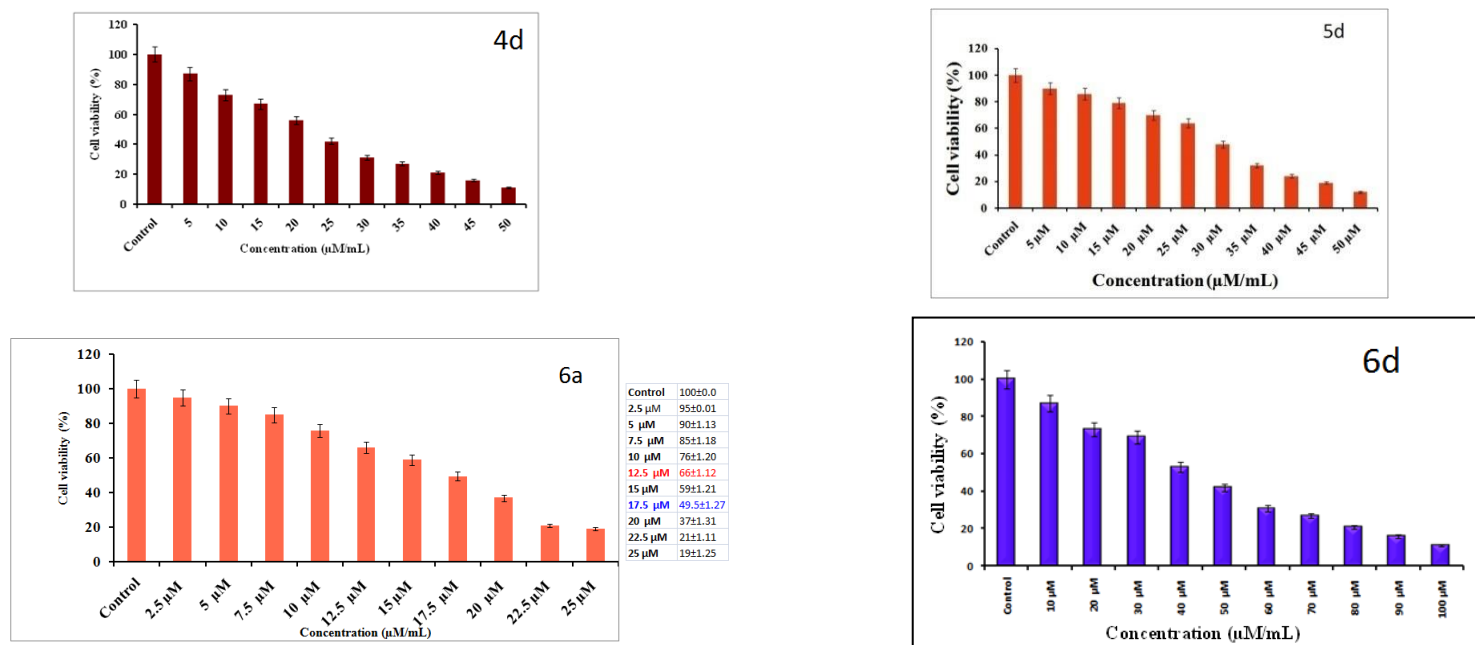
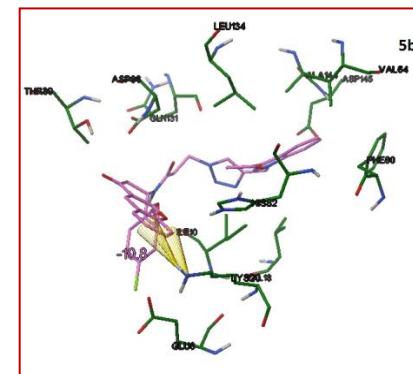
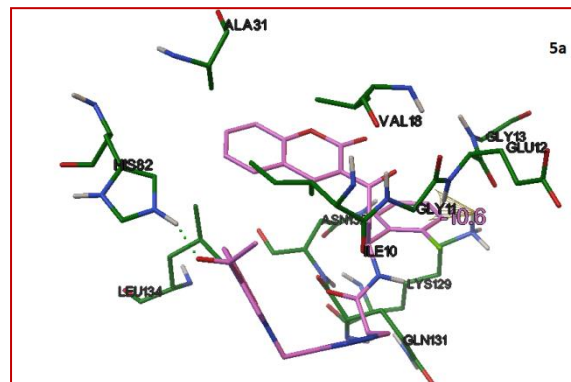
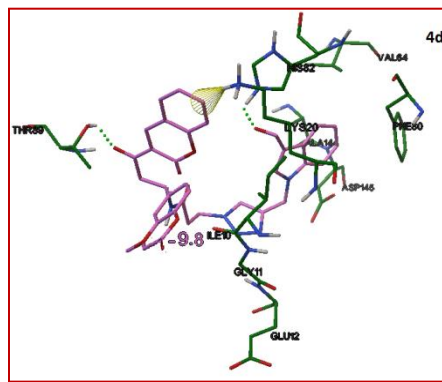
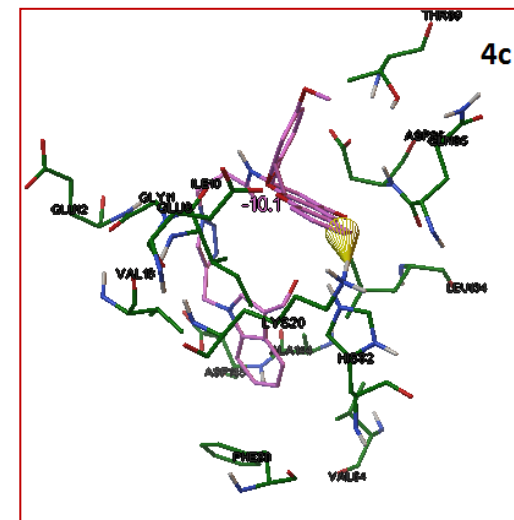
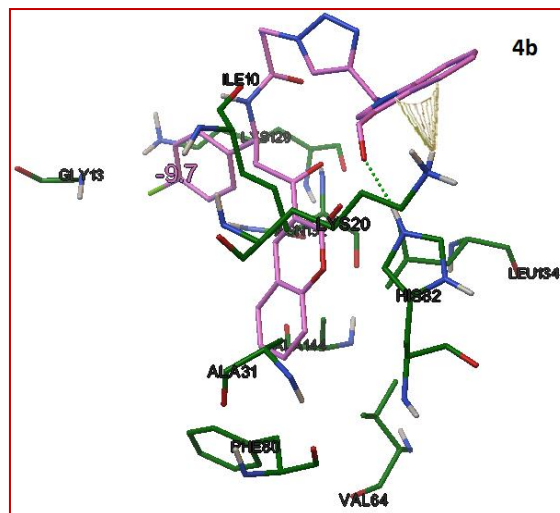
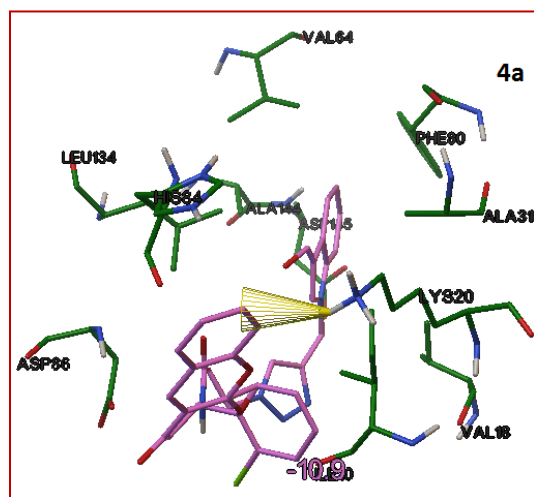
Figure S3. MTT assay results confirming the in vitro cytotoxicity effect of 4d, 5d, 6a and 6d against the MCF-7 cells. The detected IC₅₀ concentration was 20,30 and 17.5, 40 μ M respectively

Figure S4. The binding mode of the synthesised compounds in the active site of CDK2-1GII.



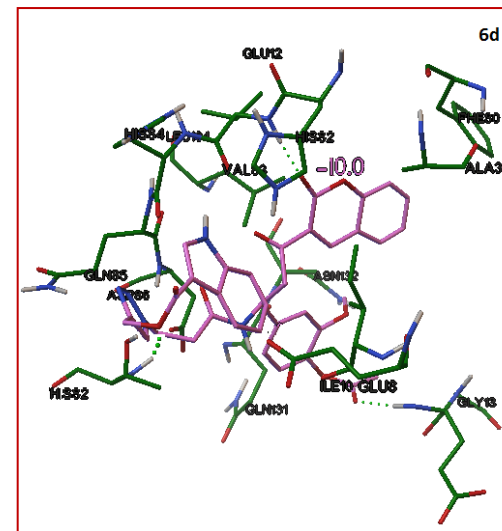
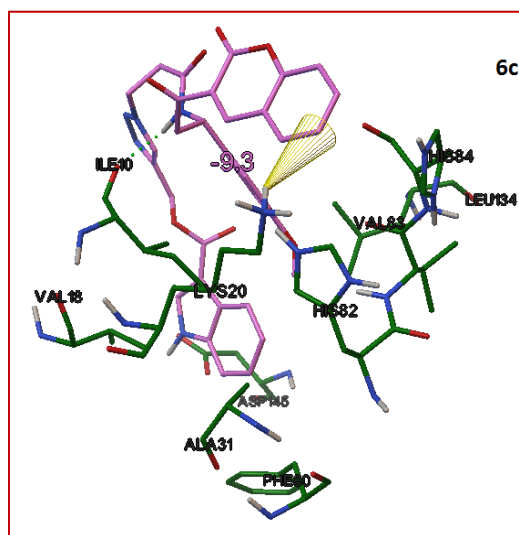
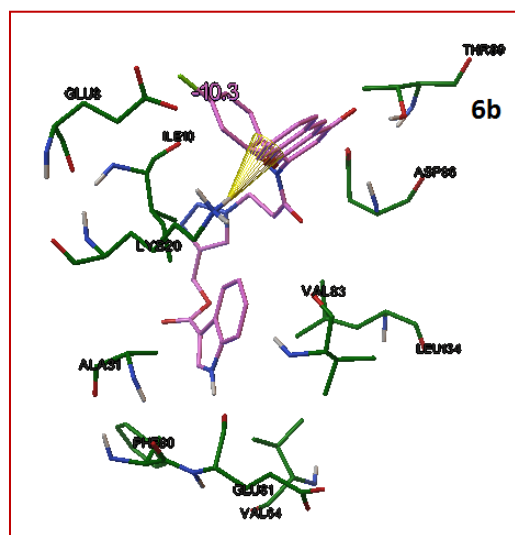
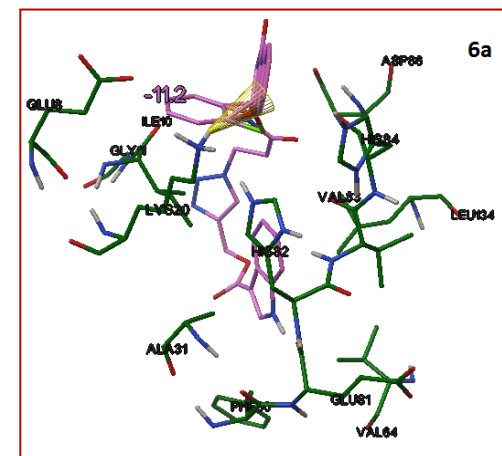
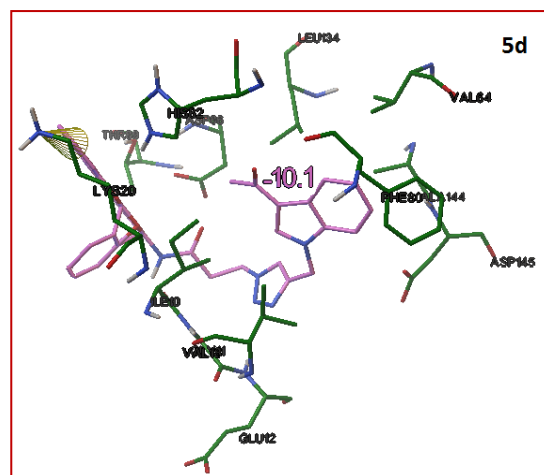
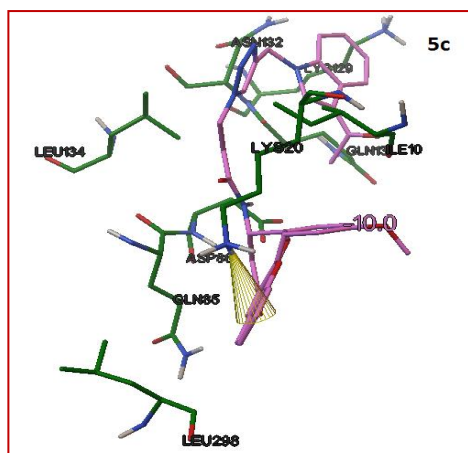


Figure S5. The super imposition of the compounds 6d (Red),5d (Green) , 4d(Pink) and (B) 6a (red) with parent ligand(blue) in the active site of CDK2-1GII.

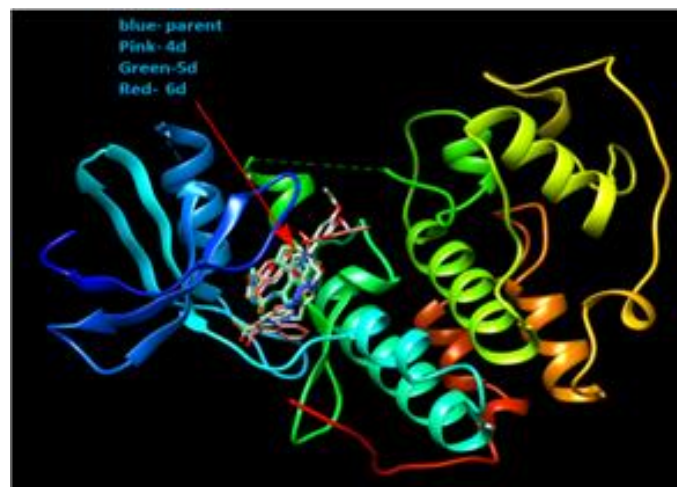


Figure S6. Bright field inverted light microscopy images of MCF-7 treated with 4d, 5d and 6d. (a) control cells, (b) cells treated with 4d, 5d and 6d at IC 25, (c) cells treated with 4d, 5d and 6d at IC 50 and Fluorescence microscopy images of: (d)control cells, (e)cells treated with 4d, 5d and 6d at IC 25, and (f) cells treated with 4d, 5d and 6dat IC 50

