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Electronic Supplementary Information

For

Pyridil-benzimidazole stator as platform for luminescent turnstiles

by

Bérangère Godde, Dialia, Ritaine, Abdelaziz Jouaiti,*, Matteo Mauro, and Mir Wais Hosseini*

Molecular Tectonics Laboratory, UMR UDS-CNRS 7140, icFRC, University of Strasbourg, Institut Le Bel, 4, rue Blaise Pascal, F-67000 Strasbourg, France.

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1. Experimental section

1.1 General considerations

Reagents and solvents were used as received unless differently stated. ¹H- and ¹³C-NMR spectra were recorded at 298 K on either Bruker AV300, Bruker AV400 or Bruker AV500 spectrometers in deuterated solvents and the residual solvent peak was used as the internal reference. All the chemical shifts (δ) are reported in ppm. Electron spray ionization Mass Spectrometry (ESI-MS) was performed by the "Service Spectrometrie de Masse" at the Faculty of Chemistry of the University of Strasbourg. Compound **2**¹, **9** and **10**² were synthetized as described in literature.

1.2 Synthetic procedures

The synthesis of compounds **3** and **4** were carried out by following the procedure reported elsewhere³ with some minor modifications: THF was used instead of EtOAc in each reaction.

Synthesis of 5.



Compound 4 (2.5 g, 15 mmol, 1eq) and Na₂S₂O₅ (4.3g, 23 mmol, 1.5 eq) were dissolved in 300 mL of ethanol under argon atmosphere. 4-Pyridinecarboxaldehyde (1.4 mL, 15 mmol, 1 eq) was slowly added under argon. The mixture was refluxed for 15 hours. Ice was then added. A pale-yellow solid was obtained and filtrated over a glass frit and washed with water. Compound 5 was purified by column chromatography (SiO₂, CH₂Cl₂ to CH₂Cl₂/MeOH: 100 to 95/5). (2.7 g, 11 mmol, 70%).

¹**H** NMR (500 MHz, DMSO): $\delta = 13.37$ (s, 1H, NH), 8.69 (d, J = 6.2 Hz, 2H, Hh), 8.15 (d, J = 6.2 Hz, 2H, Hg), 6.64 (s, 2H, Hc), 3.90 (s, 6H, Ha). ¹³C NMR (125 MHz, DMSO) $\delta = 150.40$ (Ch), 150.36;150.33 (Ce,f), 147.41(Cd), 137.19 (Cb), 120.43(Cg), 55.67 (Ca). **ESI-MS (ESI⁺):** m/z calculated for $[C_{14}H_{14}N_3O_2]^+ [M+H^+] = 256.11$, found = 256.10.





Compound **5** (1.0 g, 3.9 mmol, 1 eq.) was dissolved in degassed DMF (100 mL). Copper iodide (0.12 g, 0.59 mmol, 0.15 eq.), Cs_2CO_3 (3.2 g, 9.8 mmol, 2.5 eq.), 1,10-Phenanthroline(0.18 g, 0.98 mmol, 0.25 eq) and iodomethane (0.4 mL, 6.27 mmol, 1.6 eq) were then added under argon. The mixture was heated at 100 °C during 20 hours. The solvent was then removed, and the resulting crude was poured in CH_2Cl_2 (100 mL). The organic phase was washed twice with water before being dried with MgSO₄ and evaporated to dryness. Column chromatography (SiO₂, $CH_2Cl_2/MeOH$ 100 to 97/3) afforded **6** as an orange/red solid (0.61 g, 2.3 mmol, 58%).

¹**H NMR** (500 MHz, CDCl₃): δ = 8.77 (d, J = 6.1 Hz, 2H, Hh), 7.71 (d, J = 6.1 Hz, 2H, Hg), 6.63 (d, J = 8.5 Hz, 1H, Hc'), 6.57 (d, J = 8.5 Hz, 1H, Hc), 4.13 (s, 3H, Hj), 3.99 (s, 3H, Ha'), 3.93 (s, 3H, Ha).

¹³C NMR (126 MHz, CDCl₃): $\delta = 150.25$ (Ch), 150.13 (Ce), 146.34 (Ci), 141.99 (Cf), 137.91;135.19 (Cb,b'), 127.62 (Cd), 123.92(Cg), 104.15;101.95 (Cc,c'), 56.12 (Ca), 56.01 (Ca'), 34.52 (Cj).

MS (ESI⁺): m/z calculated for $[C_{15}H_{16}N_3O_2]^+ [M+H^+] = 270.12$, found = 270.12.

Synthesis of 7.



Compound 6 (190 mg, 0.705 mmol, 1eq) was dissolved in 35 mL of dry CH_2Cl_2 under argon. Then 8, 0 mL of BBr₃ (1M in CH_2Cl_2 , 10 eq) were dropwise added at -78 °C. The reaction was then allowed to reach room temperature. After 20 hours ice was added, the precipitate was filtrated over a glass frit, washed with cold water and dried under vacuum affording compound 7 as a yellow solid. (0.170g, 82%).

NMR ¹**H** (DMSO, 500 MHz): δ = 10.13 (s, 2H), 8.98 (d, *J* = 6.3 Hz, 2H), 8.02 (d, *J* = 5.3 Hz, 2H), 6.78 – 6.69 (m, 2H), 4.13 (s, 3H).

RMN ¹³C (DMSO, 125 MHz): δ = 151.94 (Ch), 148.83;148.79 (Ce,f), 146.66 (Cd,i), 138.03 (Cb), 124.80 (Cg), 116.83 (Cb') 111.01;109,87 (Ce,c'), 34.71 (Cj).

MS (ESI⁺): m/z calculated for $[C_{13}H_{12}N_3O_2]^+[M+H^+] = 242.09$, found = 242.09.

Synthesis of T1.



Compound **10** (0.36 g, 0.58 mmol, 1 eq.) was dissolved in DMF (400 mL) and the resulting solution degassed by vacuum argon cycles 5 times before addition of Cs_2CO_3 (0.76 g, 2.3 mmol, 4 eq.) under argon. The mixture was degassed again before adding compound **7** (0.14 g, 0.58 mmol, 1 eq.) under argon. The mixture was heated to 90 °C for 48 hours. The solvent was evaporated and the residue was dissolved in a mixture of CH_2Cl_2 (50 mL) and water (50 mL). The two phases were separated and the aqueous phase was extracted three times with CH_2Cl_2 . The combined organic phases were washed with water (50 mL) and with a solution of NH_4Cl (10%) (50 mL). The organic layer was dried over MgSO4 and evaporated to dryness. The crude was purified by column chromatography (SiO₂, $CH_2Cl_2/MeOH$ 100 to 95/5) to give **T1** as a yellow oil (0.13 g, 0.31 mmol, 33%).

¹**H NMR** (500 MHz, CD₃CN): δ = 8.71 (m, 2H, Hj), 7.73 (d, J = 5.2 Hz, 2H, Hi), 7.45 (t, J = 7.9 Hz, 1H, Ha), 6.67 (d, J = 8.5, 1H, Hc), 6.57 (d, J = 8.5, 1H, Hc²), 6.20 (dd, J = 7.9, 3.7 Hz, 2H, Hb), 4.29 – 4.23 (m, 2H), 4.22 – 4.18 (m, 2H), 4.16 – 4.01 (m, 4H), 4.10 (s, 3H, Hk), 3.92 – 3.76 (m, 4H), 3.68 – 3.61 (m, 6H), 3.61 – 3.49 (m, 14H).

¹³C NMR (125 MHz, CD₃CN): δ = 163.29;163.27 (Cl,l'), 150.72 (Cg), 150.63 (Cj), 146.08 (Ce), 142.26 (Ca), 142.13 (Ch), 138.92;135.80 (Cd,d'), 129.01 (Cf), 124.65(Ci), 107.32;105.04

(Cc,c'), 102.05;101.95 (Cb,b'), 71.35, 71.28, 71.26, 71.24, 71.22, 71.19, 71.11, 70.31, 70.22, 70.14, 70.02, 69.96, 69.27, 66.12, 66.07, 35.04(Ck).

HRMS (ESI⁺): m/z calculated for $[C_{34}H_{45}N_4O_{10}]^+[M+H^+] = 669.31$, found = 669.31.

Synthesis of T1-Ag⁺.



T1 (27.1 mg, 40.5 μ mol, 1 eq.) was dissolved in degassed CH₃CN (2 mL). To this solution, AgCF₃SO₃ (10.4 mg, 40.5 μ mol, 1 eq.) dissolved in CH₃CN (696 μ L) was added with an eppendorf and the reaction mixture was stirred at room temperature for 2 hours in the dark under argon. After removal of the solvent, T1-Ag⁺ was obtained quantitatively as a yellow solid.

¹**H** NMR (500 MHz, CD₃CN) : $\delta = 8.73$ (d, J = 6.1 Hz, 2H, Hj), 7.70 (d, J = 6.1 Hz, 2H, Hi), 7.41 (t, J = 7.9 Hz, 1H, Ha), 6.76 (d, J = 8.6 Hz, 1H, Hc'), 6.71 (d, J = 8.6 Hz, 1H, hc), 6.18 (dd, J = 7.9, 3.5 Hz, 2H, Hb,b'), 4.27 – 4.19 (m, 4H), 4.16 – 4.10 (m, 2H), 4.10 – 4.05 (m, 2H), 4.08 (s, 3H, Hk), 3.87 – 3.82 (m, 2H), 3.77 – 3.72 (m, 2H), 3.67 – 3.60 (m, 4H), 3.60 – 3.54 (m, 5H), 3.60 – 3.47 (m, 11H).

¹³C NMR (125 MHz, CD₃CN): $\delta = 163.28;163.21(Cl,l'), 151.33$ (Cg), 151.21 (Cj), 144.44 (Ce), 142.56 (Ca), 142.34 (Ch), 141.57, (Cd,d')128,05 (Cf), 124.83 (Ci), 108.18,106.91(Cc,c'), 102.00;101.84 (Cb,b'), 71.41, 71.25, 71.22, 71.14, 71.10, 71.00, 70.40, 70.19, 70.11, 70.02, 69.99, 69.94, 66.34, 66.17, 35.48 (Ck).

MS (ESI⁺): m/z calculated for $[C_{34}H_{44}N_4O_{10}Ag]^+[M]^+ = 775.21$, found = 775.21.

2. Measurement of binding constants

The NMR dosage was conducting as following: In an NMR tube, the turnstile was dissolved in CD₃CN (500 μ L, concentration of 1.36×10⁻³ M). Successive additions of a solution of AgCF₃SO₃ in CD₃CN (concentration of 4.9.10⁻² M) were done using a 50 μ L microsyringe. The data were then compilated in a file and analysed with a titration software, Hypspec (http://www.hyperquad.co.uk/HypSpec.htm) which uses multivariate factor analysis to obtain globally optimized parameters. The mathematical analysis of the titration data was done with a titration software, Hypspec (http://www.hyperquad.co.uk/HypSpec.htm) which uses multivariate factor analysis to obtain globally optimized parameters. The mathematical analysis of the titration data was done with a titration software, Hypspec (http://www.hyperquad.co.uk/HypSpec.htm) which uses multivariate factor analysis to obtain globally optimized parameters. The fit of the complete titration data was good with $\sigma_{1:1}$ =0.02 and $\sigma_{2:1}$ =0.03 (Fig. S3). The simulation for the speciation was done with the software Hyperquad.



Figure S1. ¹H NMR (CD₃CN, 500 MHz, 298 K) spectra of T1 after addition of 0 to 25 equivalents of CF₃SO₃Ag.



Figure S2. ¹H NMR ¹H (CD₃CN, 500 MHz, 298 K) spectra of **T1** after addition of 0 to 25 equivalents of CF₃SO₃Ag.

a)



c)



Figure S3. Plot of the variation of the experimental (red trace) and calculated (black trace) chemical shift as a function of the number of the CF_3SO_3Ag equivalents added a) hydrogens Hn b) hydrogens of the polyethylenglycol chain Hi c) hydrogens of the polyethylenglycol chain Hi.



Figure S4. Simulated speciation profiles for titrations of T1 with Ag (I) at different concentrations representing the proportion of T1 (black trace), the 1:1 Ag⁺/T1 complex (red trace) and the 2:1 Ag⁺/T1 (blue trace), function of the number of equivalents of CF₃SO₃Ag.

- a) $[T1] = 1.0 \times 10^{-3} M$.
- b) $[T1] = 1.0 \times 10^{-4} M$.
- c) [T1] = 1.0×10^{-5} M.

3. Photophysical measurements

Electronic absorption spectra of samples in solution at room temperature were recorded on a double-beam Perkin-Elmer Lambda 650 S spectrophotometer and baseline corrected. Steadystate emission spectra were recorded on a HORIBA Jobin-Yvon IBH FL-322 Fluorolog 3 spectrometer equipped with a 450 W xenon arc lamp, double grating excitation and emission monochromators (2.1 nm mm⁻¹; 1200 grooves mm⁻¹) and a PPD850 photomultiplier tube. Emission and excitation spectra were corrected for source intensity (lamp and grating) and emission spectral response (detector and grating) by standard correction curves. Time-resolved measurements were performed using the time-correlated single-photon-counting (TCSPC) option on the Fluorolog 3. NanoLEDs (λ_{exc} = 340 or 370 nm; FWHM <1 ns) with repetition rates between 10 kHz and 1 MHz used to excite the sample. The excitation source were mounted on the sample chamber at 90° to a double grating emission monochromator (2.1 nm mm⁻¹ dispersion; 1200 grooves mm⁻¹) and collected by a PPD850 single-photon-counting detector. The photons collected at the detector were correlated by a time-to-amplitude converter (TAC) to the excitation pulse. Signals were collected using an IBH DataStation Hub photon counting module and data analysis was performed using the commercially available DAS6 software (HORIBA Jobin Yvon IBH). The goodness of fit was assessed by minimizing the reduced chisquare function (χ) and visual inspection of the weighted residuals. Photoluminescence quantum yields (PLQY) were measured in optically dilute solution (optical density <0.1 at the excitation wavelength) and compared to reference emitters by the method of Demas and Crosby⁴. Quinine sulfate in 0.05 M H₂SO₄ was used as reference at room temperature (PLQY = 0.53)⁵. All the solvents were spectrophotometric grade.

4. Supplementary Figures



Figure S5. Solvent effect for T1. Absorption spectra of samples of T1 in toluene (red wine trace), in CH_2Cl_2 (dark trace), CH_3CN (red trace) and DMF (olive trace).





b)



c)



Figure S6. a) ESI MS spectra of the sample **T1** obtained after addition of 25 equivalents of CF_3SO_3Ag to **T1** (1.10⁻³ M) b) Zoom and enlarged image of a) corresponding to the 1:1 (ie [T+Ag]⁺) c) Zoom and enlarged image of a) corresponding to the 2:1 Ag⁺/**T1(**ie [T1+2Ag+CF_3SO_3]⁺) (c).



5. Chemical characterization of the compounds: ¹H-, ¹³C-NMR and MS spectra



Figure S7. ¹H (500 MHz, top) and ¹³C NMR (125 MHz, bottom) spectra of compound 5 (DMSO, 298K).





Figure S8. ESI-MS spectrum of compound 5 and enlarged image (bottom).



Figure S9. 1 H (500 MHz, top) and 13 C NMR (125 MHz, bottom) spectra of compound 6 (CDCl₃, 298K).









Figure S11. 1 H (500 MHz, top) and 13 C NMR (125 MHz, bottom) spectra of compound 7 (DMSO, 298K).



Figure S12. ESI-MS spectrum of 7 (top) and enlarged image (bottom).



Figure S13. ¹H (500 MHz, top) and ¹³C NMR (125 MHz, bottom) spectra of compound T1 (CD₃CN, 298K).



Figure S14. HRMS spectrum of T1 (top) and enlarged image (bottom).



Figure S15. ¹H (500 MHz, top) and ¹³C NMR (125 MHz, bottom) spectra of compound T1-Ag⁺ (CD₃CN, 298K).



Figure S16. ESI-MS spectrum obtained after addition of one equivalent of CF_3SO_3Ag to **T1** (top), enlarged image and simulation of the 1:1 (ie $[T1+Ag]^+$) complex.

6. References

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