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Supporting information

MoS₂-Au@Pt nanohybrid as a sensing platform for

electrochemical nonenzymatic glucose detection

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Synthesis of Au nanoseeds, MoS₂-Au and MoS₂-Au@Pt nanocomposite

Au nanoseeds were prepared by using $Na_3C_6H_5O_7 \cdot 2H_2O$ and $HAuCl_4$. First, 700 µL HAuCl_4 (10 mM) was added into 49.3 mL ultrapure water. Then, the mixture was heated to approximately 150 °C on a hotplate. After that, 5 mL $Na_3C_6H_5O_7 \cdot 2H_2O$ (10 mM) was inject into the solution for reacting 25 minutes to get the Au nanoparticles.

For MoS₂-Au nanocomposite synthesis, 4 mL (0.025 mg mL⁻¹) of MoS₂ nanosheet dispersion was mixed with 400 μ L of PVP (5%) under vigorous stirring for 2 min to obtain a stable mixture. Then, 200 μ L HAuCl₄·3H₂O (10 mM) was dropped into the mixture and mixed for 10 minutes. After this, the reaction mixture was heated to 60 °C for 5 minutes in the microwave reactor. Finally, the product of MoS₂-Au nanocomposite was purified at least twice by centrifugation.

For MoS₂-Au@Pt nanocomposite synthesis. First, 0.3 mL aqueous MoS₂-Au seed solution was added to 2 mL ultrapure water. Then 0.5 mL 10 mM CTAB solution was mixed with the seed solution for 5 min under vigorous stirring. After that, 1 mL AA (100 mM) was injected quickly into the mixed solution. At intervals of 5 s, 100 μ L 5 mM H₂PtCl₆ was added to the solution under stirring. Then, the mixture was heated on a hotplate at approximately 100 °C for 6 min to grow Pt nanoshells. Finally, the product of MoS₂-Au@Pt nanocomposite was purified by washing and centrifugation (5000 rpm for 10 min) to remove excess reagent three times with acetone and water.



Fig. S1. TEM images of the (A) MoS₂-Au nanocomposite, (B) MoS₂-Pt nanocomposite and (C) Au@Pt nanocomposite. Histograms of nanoparticles' size corresponding to (D) MoS₂-Au nanocomposite, (E) MoS₂-Pt nanocomposite and (F) Au@Pt nanocomposite.



Fig. S2. AFM images of MoS₂ nanosheet.



Fig. S3. Cyclic voltammograms of six modified electrodes in 0.1 M NaOH without and with 3 mM glucose at a scan rate of 100 mV s⁻¹.



Fig. S4. (A) Cyclic voltammograms of MoS₂-Au@Pt/GCE in 0.1 M NaOH containing 3 mM glucose at various scan rates ranging from 10, 50, 100, 150, 200 to 300 mV s⁻¹. (B) The plot of glucose oxidation peak currents versus the square root of scan rate.



Fig. S5. (A) Amperometric responses of MoS₂-Au@Pt/GCE in 0.1 M NaOH containing 1 mM glucose by using different applied potentials. (B) The peak current increment versus the applied potential.



Fig. S6. Successive 200 Cyclic voltammograms of MoS_2 -Au@Pt/GCE in 0.1 M NaOH containing 3 mM glucose at a scan rate of 100 mV s⁻¹.

Electrode material	Detection limit (µM)	Linear range (mM)	References
Pt–Au alloy	6	0.006-11	1
PtNi-ERGO	10	0.5-35	2
Pt-Pd nanoflakes	20.6	0-16	3
Pt-Au/MWCNT	10	0-24.44	4
Macroporous Au-Pt	25	1-20	5
NiO/Pt/ERGO	0.2	0.05-5.66	6
PtNCs/GR	30	1–25	7
Ni-MoS ₂	0.31	0-4	8
Au-Pd-MoS ₂	400	0.5-20	9
MoS ₂ -Au@Pt	1.08	0.01-3	our work

Table S1 Comparison of different electrode materials for nonenzymatic glucose determination.

ERGO: electrochemically reduced graphene oxide. MWCNT: multi-wall carbon nanotubes. PtNCs: Pt nanoclusters. GR: graphene.

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