

Chiral copper-salen complex grafted over functionalized mesoporous silica as an efficient catalyst for the asymmetric Henry reactions and synthesis of potent drug (R)-isoproterenol

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Section-I

1. GENERAL INFORMATION:

1.1 Chemicals:

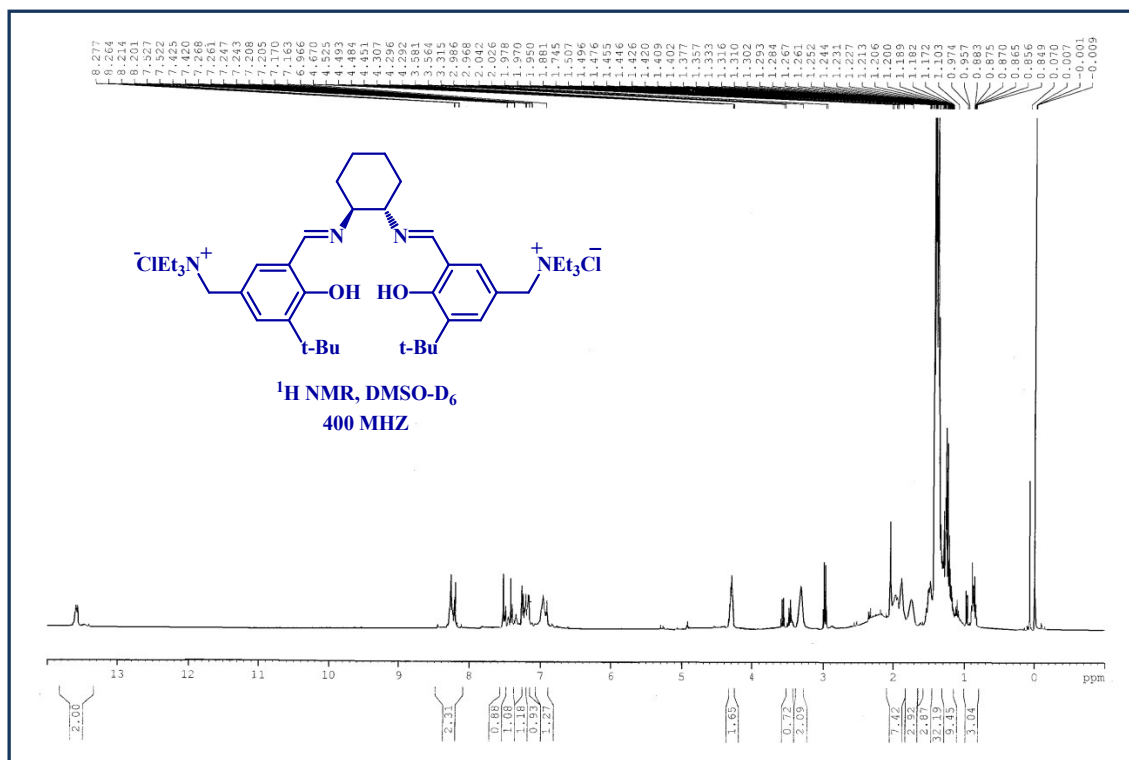
3-*tert*-butyl-2-hydroxybenzaldehyde (**1**), (1*S*, 2*S*)-(+)-1,2-diaminocyclohexane (**4**) Pluronic P123 (EO₂₀PO₇₀EO₂₀, EO = ethylene oxide, PO = propyleneoxide, M_{av} = 5800), tetraethoxyorthosilicate (TEOS), 3-aminopropyl triethoxysilane (3-APTES, **8**), 4-formylbenzoicacid, (**10**), Cu(OAc)₂. H₂O, all aldehydes, nitromethane and nitroethane were acquired as reagent grade and were used devoid of additional purification. All the solvents were dried according to standard procedures. TLC-analysis were performed using TLC Silica gel 60 F₂₅₄.

1.2 Characterization techniques:

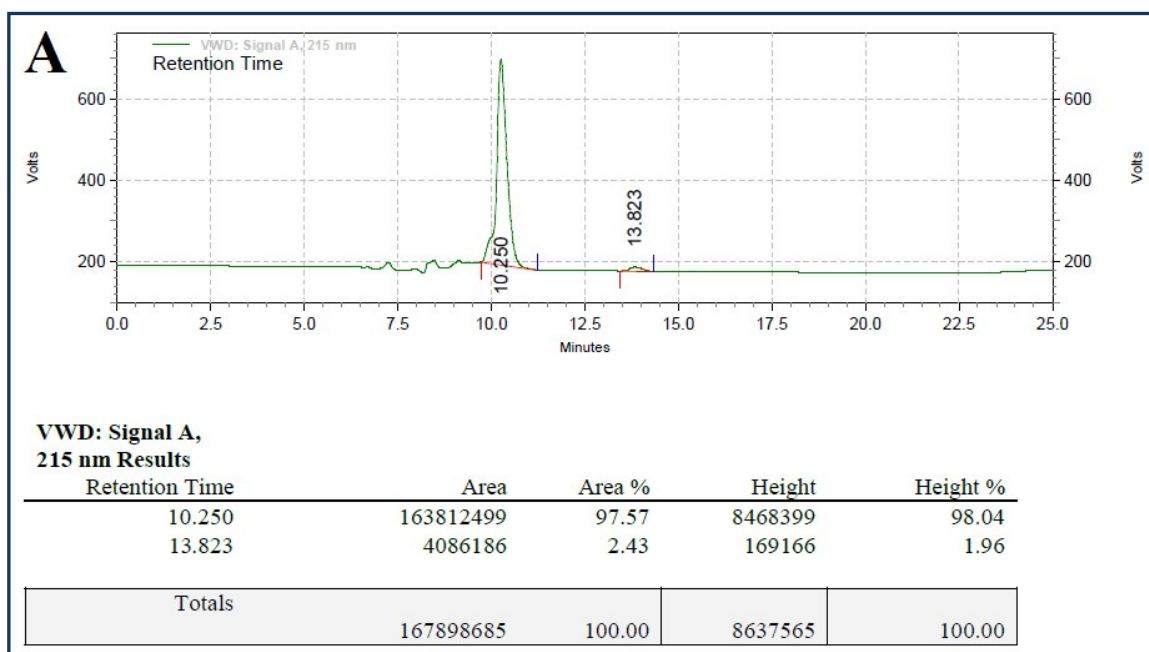
A Shimadzu UV 2401PC coupled with an integrating sphere attachment was used for the recording of UV-Visible spectra. BaSO₄ was applied as background standard. The Perkin-Elmer FT-IR 783 spectrophotometer was used to record FT-IR spectra of the samples in the range from 400 to 4000 cm⁻¹ using KBr pellet as support. Powder X-ray diffraction (PXRD) patterns of the sample was tested with a Bruker D8 Advance X-ray diffractometer operated at a voltage of 40 kV and a current of 40 mA using Ni-filtered Cu K α (λ =0.15406 nm) radiation. TEM images of the mesoporous silica supported Cu-salen catalyst were obtained using a JEOL JEM 2010 transmission electron microscope. A Mettler Toledo TGA/DTA 851e device was used for TGA. A Quantachrome Autosorb 1C surface area analyzer was employed for N₂-sorption desorption analysis at 77 K. For the bulk elemental analysis, the Cu(II)@AFS-1 was digested with acid to dissolved them into clear liquid and then Cu content were analyzed by a Shimadzu AA-6300 atomic absorption spectrophotometer (AAS) fitted with a double beam monochromator. Carbon, hydrogen and nitrogen contents of Cu(II)@AFS-1 were examined utilizing a Perkin Elmer 2400 Series II CHN analyzer. ¹H spectrums of the products were recorded on 400 MHz NMR instruments using CDCl₃ as solvent. Enantiomeric excesses were examined by HPLC (Agilent, Model 1220) using Ultron using a Chiralcel® OD-H column (wavelengths 215 nm). 2-propanol/hexane system was used as eluent. Optical rotations were (described as: $[\alpha]_D^{25}$ (c = in g per 100 ml, solvent)) tested using Digipol 781 Automatic Polari meter Rudolph equipment.

2. CHARACTERISATION DATA OF THE Cu(II)@AFS-1 CATALYST:

Fig. S1. ¹H NMR spectra of chiral Schiff base ligand (1)



3.COMPARISON OF CATALYTIC ACTIVITY BETWEEN PhCHO AND CH₃NO₂ CATALYZED BY HOMOGENEOUS Cu(II)-SALEN COMPLEX (2) AND HETEROGENEOUS CHIRAL Cu(II)@AFS-1 CATALYST FOR THE HENRYREACTION^a



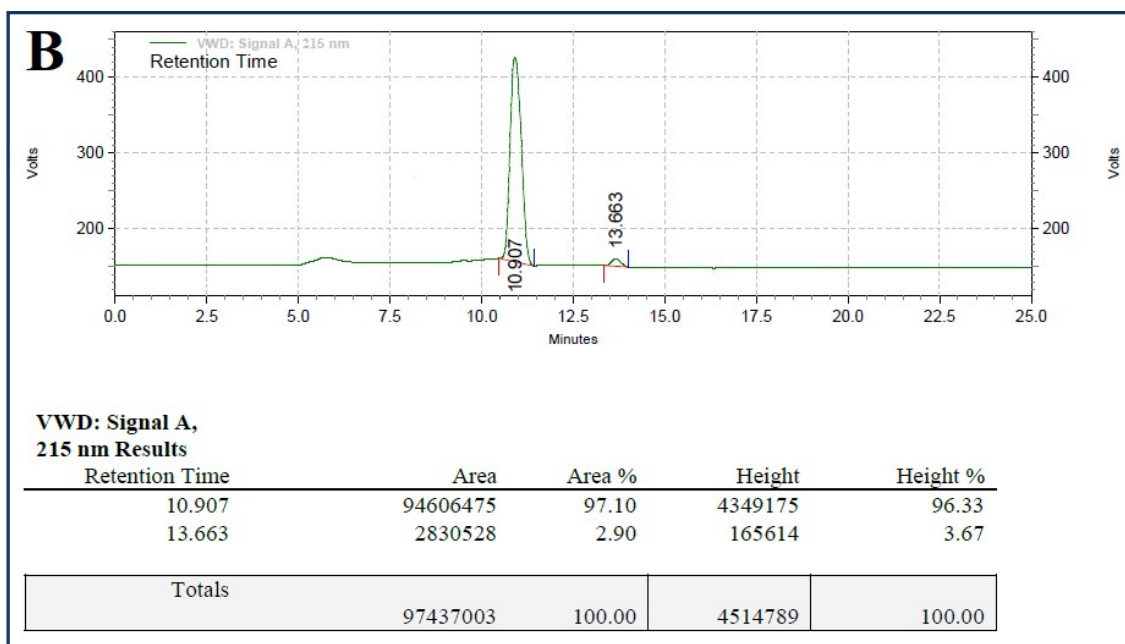


Figure S2. HPLC chromatograms of (R)-2-nitro-1-phenylethanol (**2d**) over homogeneous chiral Cu(II) salen complex (**A**) and heterogeneous chiral Cu(II)@AFS-1 catalyst (**B**).

4. SYNTHESIS AND CHARACTERIZATION DATA OF CHIRAL DRUG (R)-ISOPROTERENOL

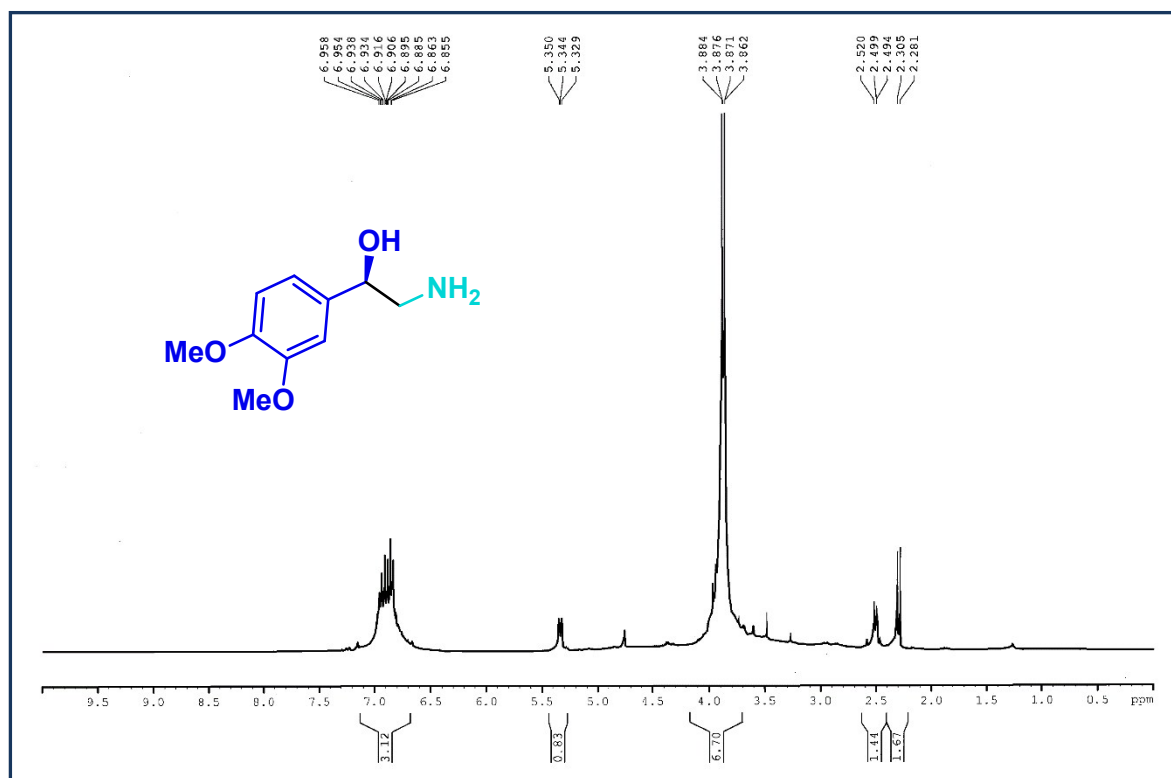
4.1 (R)-(-)-1-(3,4-dimethoxyphenyl)-2-nitroethanol (**2f**)^[1]:

(R)-1-(3,4-dimethoxyphenyl)-2-nitroethanol (**2f**) was synthesized from 3,4-dimethoxy benzaldehyde and nitromethane according to our optimized reaction conditions as stated in Table 2. The characterization data of the compound **2f** are stated in Section S2.

4.2(R)-(-)-2-Amino-1-(3,4-dimethoxyphenyl)ethanol (**3**)^[1]:

10% Pd/C (230 mg) was slowly and cautiously added to the stirred ethanolic solution of **2f** (2.95 mmol) under H₂ atmosphere for 24 h. After completion of the reaction catalyst was filtered out through the small pad of Celite. Then the solvent was removed under reduced pressure to furnish compound **3** (94% yield). ¹H NMR (400 MHz, CDCl₃) δ 6.96-6.85 (m, 3H), 5.35-5.33 (m, 1H), 3.88 (bs, 3H), 3.86 (bs, 3H), 2.51 (d, *J* = 10.4 Hz, 1H), 2.29 (d, *J* = 9.6 Hz, 1H); [α]_D²⁵ = -25.3 (*c* 1.0, EtOH)^[1] {[α]_D²⁵ = -24.0 (*c* 1.08, EtOH)}.

Fig. S3. ^1H NMR spectrum of (R)-(-)-2-Amino-1-(3,4-dimethoxyphenyl)ethanol (**3**):



4.3 (R)-(-)-1-(3,4-Dimethoxyphenyl)-2-(isopropylamino) ethanol (**4**)^[1]

A mixture of compound **3** (2.2 mmol) and acetone (4.7 mmol) in ethanol (4 mL) was stirred at RT for 1 h. Then after cooling the mixture to 0 °C (ice bath), NaBH_4 (3.3 mmol) was added to it. Thereafter the reaction mixture was further stirred for 1 h and finally the product compound (**4**) was separated through column chromatography (EtOAc/MeOH) as yellow coloured solid (95% yield). ^1H NMR (400 MHz, CDCl_3) δ 6.87-6.77 (m, 3H), 4.89 (bs, 1H), 3.79 (bs, 6H), 2.78-2.75 (m, 1H), 2.69-2.65 (m, 2H), 1.20-1.17 (m, 6H). $[\alpha]_{\text{D}}^{25} = -27.2$ (c 2.51, Acetone)^[1] $\{[\alpha]_{\text{D}}^{25} = -32.7$ (c 3.00, Acetone) $\}$, mp 126 °C^[2] mp: 126-128 °C, 91% ee; HPLC analysis was performed using Chiralcel OD-H column having 85/15 n-hexane/*i*-PrOH as mobile phase, flow rate 1.0 ml/min, retention time (*R*): 10.34 min (major), (*S*): 15.70 min (minor).

Fig. S4. ^1H NMR spectrum of (R)-(-)-1-(3,4-Dimethoxyphenyl)-2-(isopropylamino)ethanol (**4**):

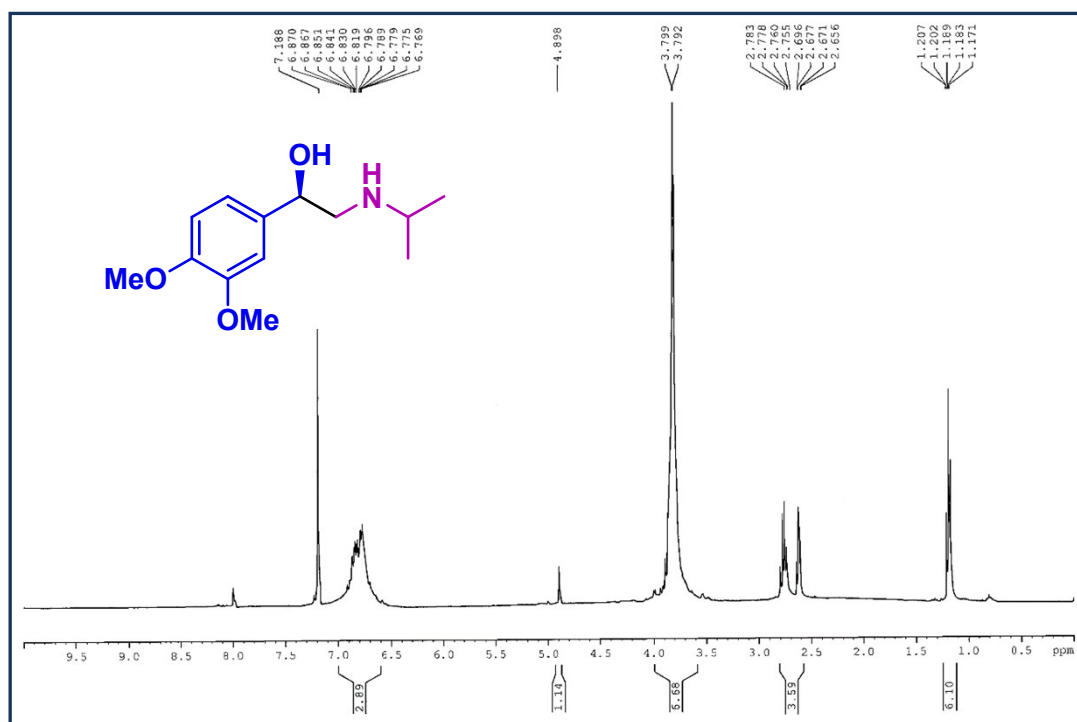
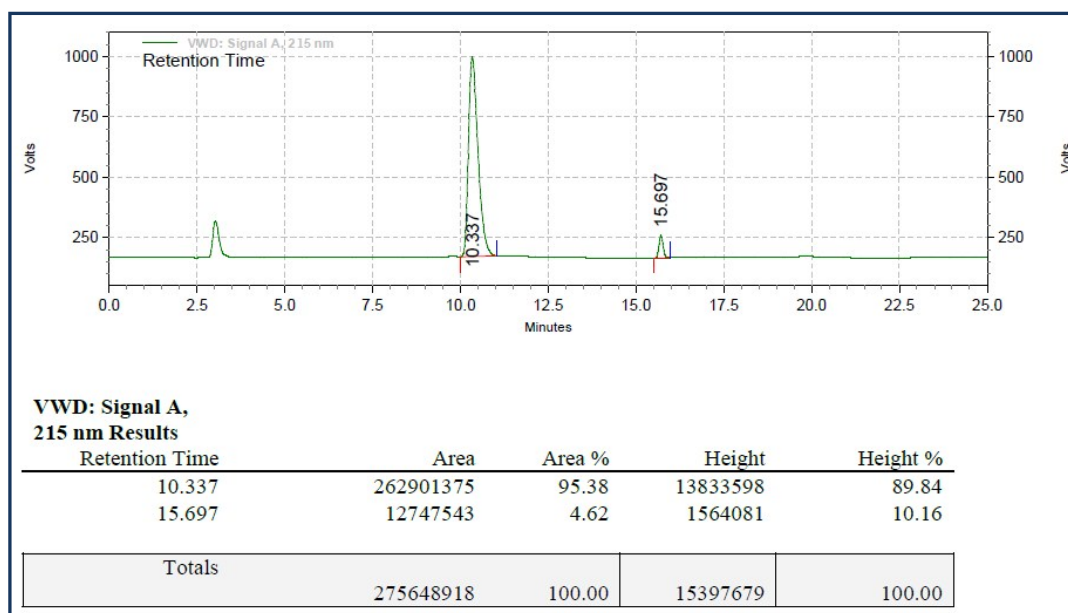


Fig. S5. HPLC chromatogram of **4**:

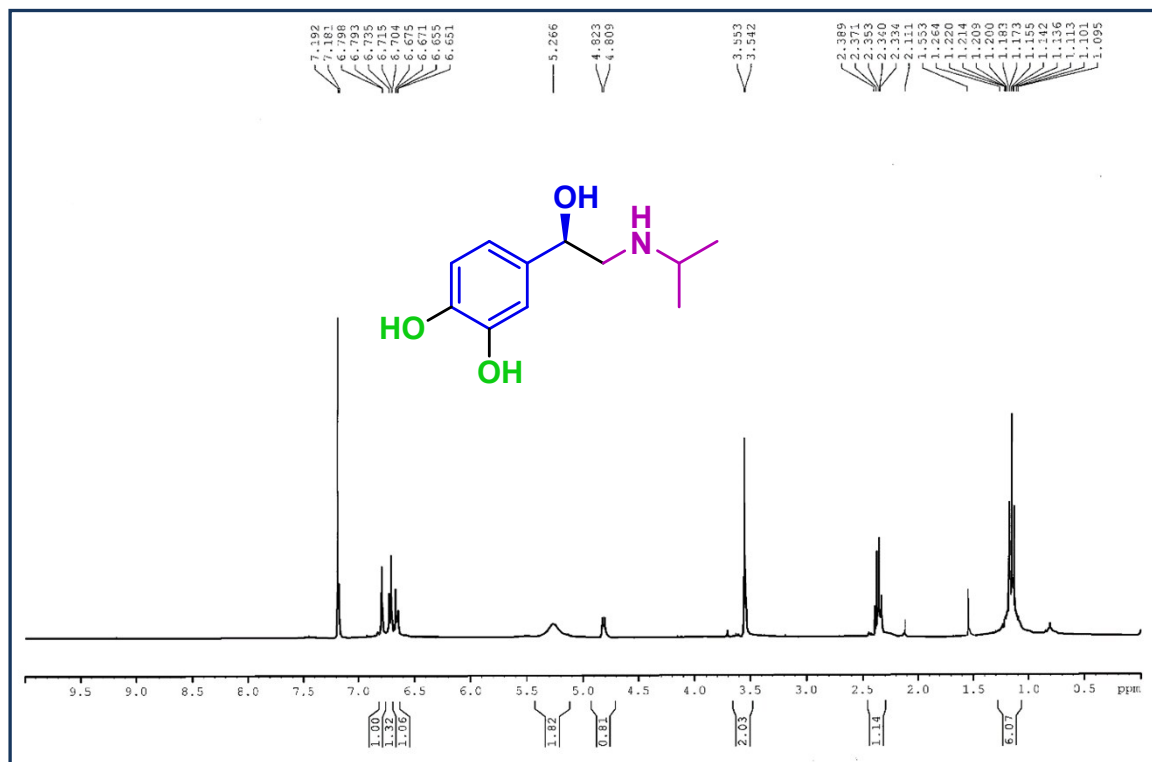


4.4 (R)-(-)-Isoproterenol^[3-5]

The solution of EtSH (2 mL) in DCM was cooled to 0 °C (ice bath) and AlCl₃ (12 mmol) was added to it. Then the solution was warmed to RT and the compound **4** (0.578 mmol) was added. Thereafter stirring for 20 h the reaction mixture was poured into triple distilled water followed by acidified with dil. HCl and extracted with DCM. Then the solvent was evaporated and the product was collected by column chromatography (CHCl₃/MeOH) as a colourless solid (96% yield); ¹H NMR (400 MHz, CDCl₃) δ 6.79-6.65 (m, 3H), 5.27 (br s, 2H), 4.81 (d, *J* = 5.6 Hz, 1H), 3.55-3.54 (m, 2H), 2.38-2.33 (m, 1H), 1.26-1.09 (m, 6H);

IR(neat): 3310, 2992, 1609, 1485, 1263, 1082, 889, 779 cm^{-1} ; Anal. calcd. for $\text{C}_{11}\text{H}_{17}\text{NO}_3$: C 62.54, H 8.11, N 6.63; found: C 62.72, H 8.09, N 6.54%. $[\alpha]_{\text{D}}^{25} = -41.2$ (c 1.06, 2M HCl)^[3-5] $[\alpha]_{\text{D}}^{25} = -42.3$ (c 1.00, 2M HCl), mp 162 $^{\circ}\text{C}$ ^[3-5] mp:163-164 $^{\circ}\text{C}$. The overall yield of the drug molecule is 82%.

Fig. S6. ^1H NMR spectrum of (R)-(-)-Isoproterenol:



5. COMPARATIVE STUDY:

Table S1. Comparison of the catalytic activity of Cu(II)@AFS-1 with related catalysts:

Catalyst	Reaction Conditions	Yield ^a (%)	ee ^b (%)	TON	Ref
Chiral macrocyclic [H ₄] salen ligand, Cu(OAc) ₂ ·H ₂ O	Benzaldehyde, CH ₃ NO ₂ , THF, RT, 20 h	84	93	11.2	6
Fam-1 ligand, Et ₂ Zn	Benzaldehyde, CH ₃ NO ₂ , THF, -50 $^{\circ}\text{C}$, 16 h	83	87	1.53	7
Macrocyclic salen ligand, CuCl ₂ ·2H ₂ O	Benzaldehyde, CH ₃ NO ₂ , EtOH + DCM, 2,6-lutidine, RT, 30 h.	86	91	8.6	8
C ₁ -symmetric primary-secondary diamines ligand, CuBr	Benzaldehyde, CH ₃ NO ₂ , THF, NMM, RT, 24 h, N ₂ -atm.	81	82	8.1	9
Chiral tetrahydrosalen ligand, (CuOTf) ₂ ·C ₆ H ₅ CH ₃	Benzaldehyde, CH ₃ NO ₂ , MeOH, 4 Å MS, 40 $^{\circ}\text{C}$, 20 h	90	92	90	10
Heterogeneous chiral Cu(II)@AFS-1	Benzaldehyde, CH₃NO₂, DCM, RT, 15 h	96	94	449	This work

^aYields are referred to those of isolated pure products. ^bee was determined from the HPLC analysis.

Section-II

1. CHARACTERIZATION DATA AND HPLC CHROMATOGRAMS OF THE PURE PRODUCTS:

(R)-2-nitro-1-(4-nitrophenyl)ethanol^[11] (2a, Table 1)

The desired product was isolated by column chromatography over silica gel (ethyl acetate/hexane 15/85) as yellow oil. $[\alpha]_D^{25} = -10.8$ (c 1.01, CH_2Cl_2), lit.^[11] $[\alpha]_D^{25} = -10.0$ (c 1.06, CH_2Cl_2); 91% ee; HPLC analysis was performed using Chiralcel OD-H column having 85/15 n-hexane/*i*PrOH as mobile phase, flow rate 1.0 ml/min, retention time (*R*): 17.79 min (major), (*S*): 24.53 min (minor).

Fig.S7. ^1H NMR spectrum (CDCl_3 , 400 MHz):

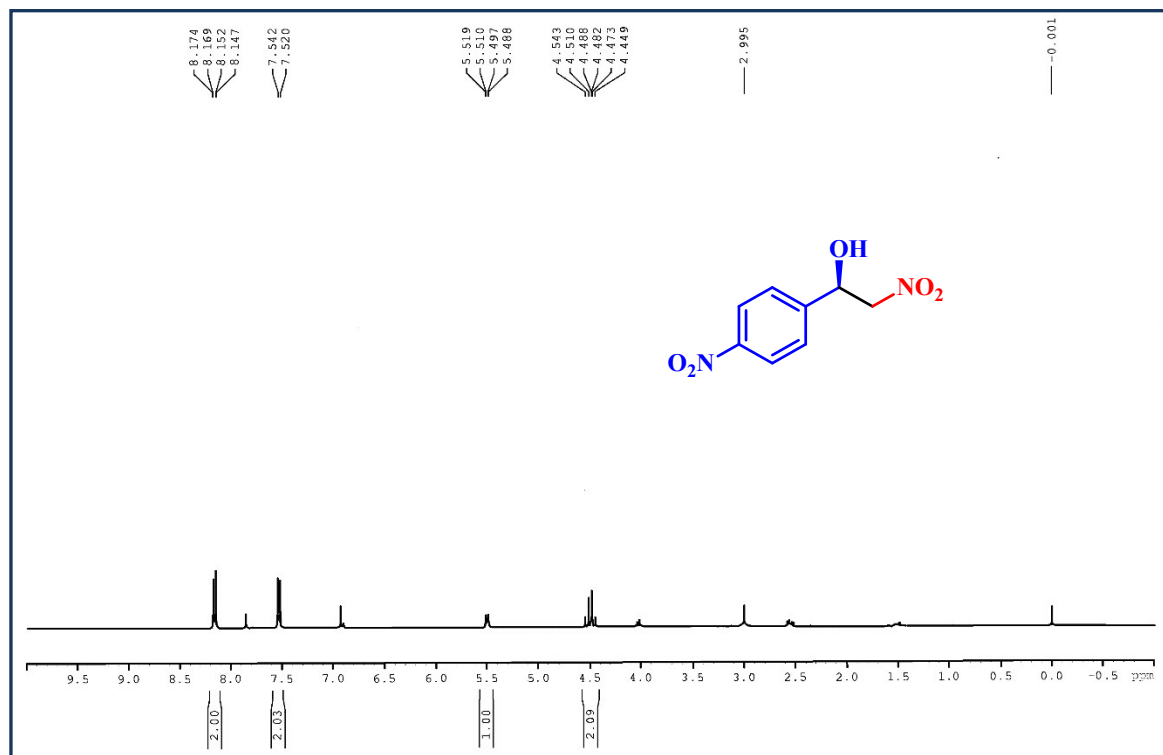
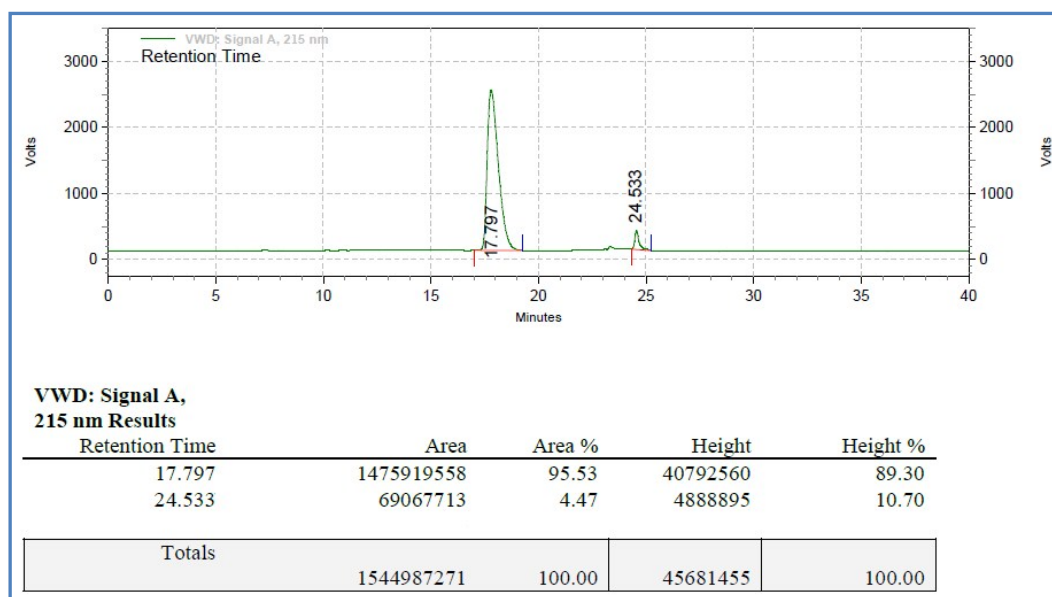


Fig. S8. HPLC chromatogram:



(R)-1-(4-chlorophenyl)-2-nitroethanol^[11] (2b, Table 1)

The desired product was isolated by column chromatography over silica gel (ethyl acetate/hexane 15/85) as yellowish oil. $[\alpha]_D^{25} = -25.1$ (c 1.09, CH_2Cl_2), lit.^[11] $[\alpha]_D^{25} = +24.7$ (c 1.13, CH_2Cl_2); 90% ee; HPLC analysis was performed using Chiralcel OD-H column having 85/15 n-hexane/*i*-PrOH as mobile phase, flow rate 1.0 ml/min, retention time (*R*): 10.39 min (major), (*S*): 12.46 min (minor).

Fig.S9. ^1H NMR spectrum(CDCl_3 , 400 MHz):

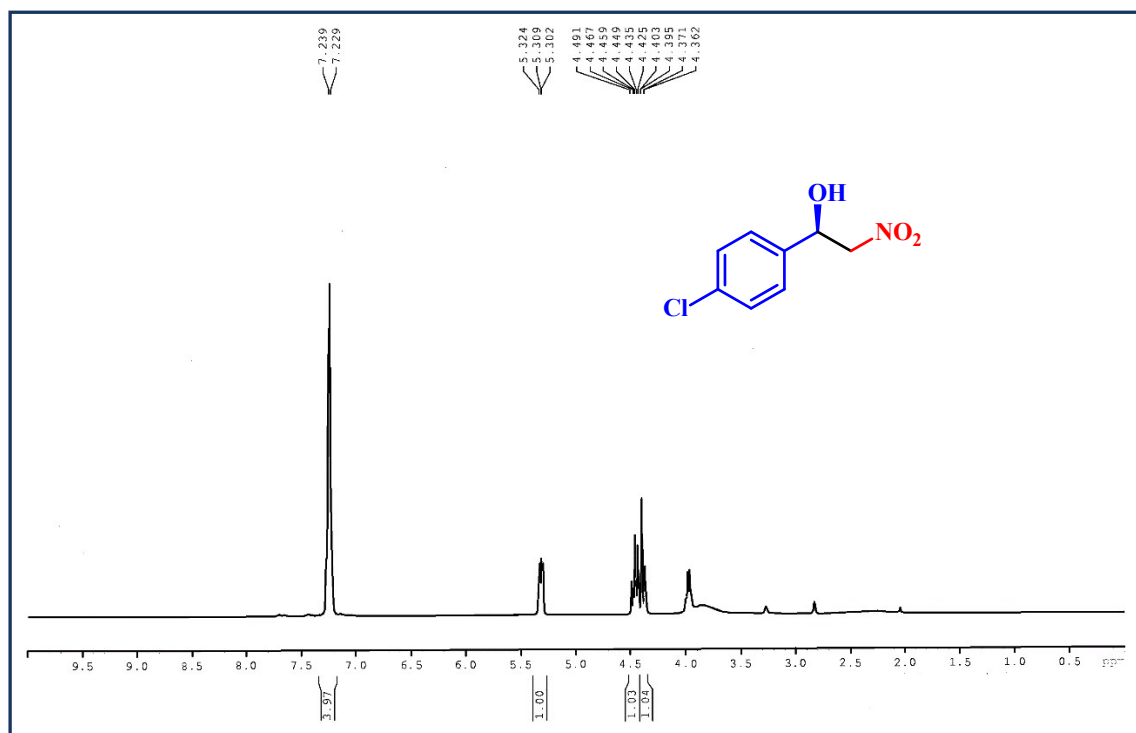
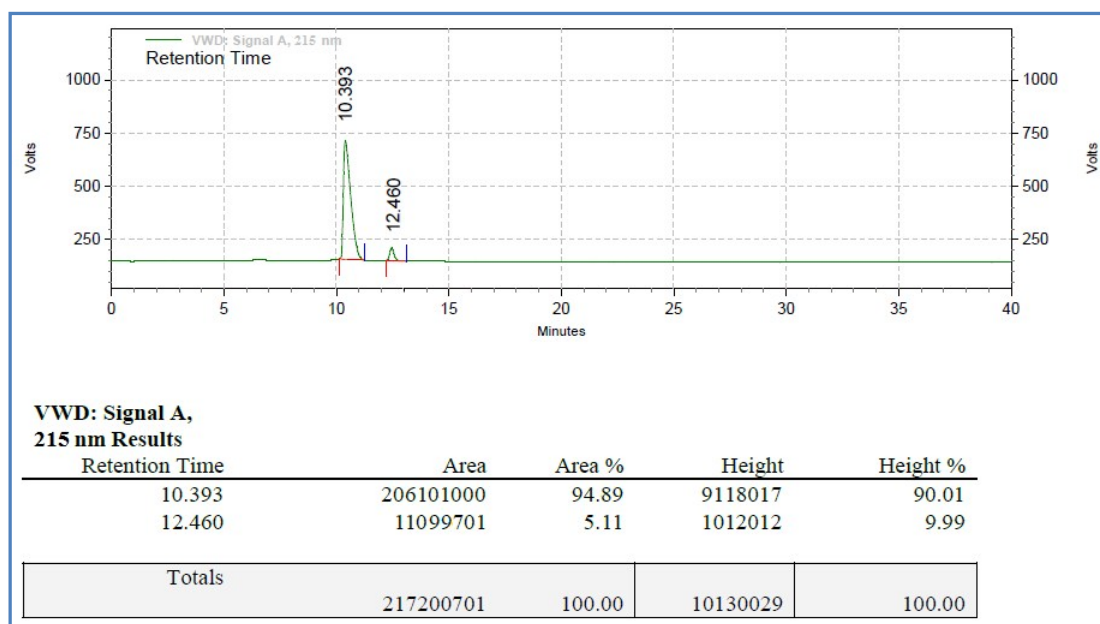


Fig. S10. HPLC chromatogram:



(R)-2-nitro-1-(p-tolyl)ethanol^[11] (2c, Table 1)

The desired product was isolated by column chromatography over silica gel (ethyl acetate/hexane 15/85) as yellowish oil. $[\alpha]_D^{25} = -37.9$ (c 1.1, CH_2Cl_2), lit.^[11] $[\alpha]_D^{25} = -37.2$ (c 1.14, CH_2Cl_2); 87% ee; HPLC analysis was performed using Chiralcel OD-H column having 85/15 n-hexane/*i*-PrOH as mobile phase, flow rate 1.0 ml/min, retention time (*R*): 10.35 min (major), (*S*): 13.55 min (minor).

Fig.S11. ^1H NMR spectrum(CDCl_3 , 400 MHz):

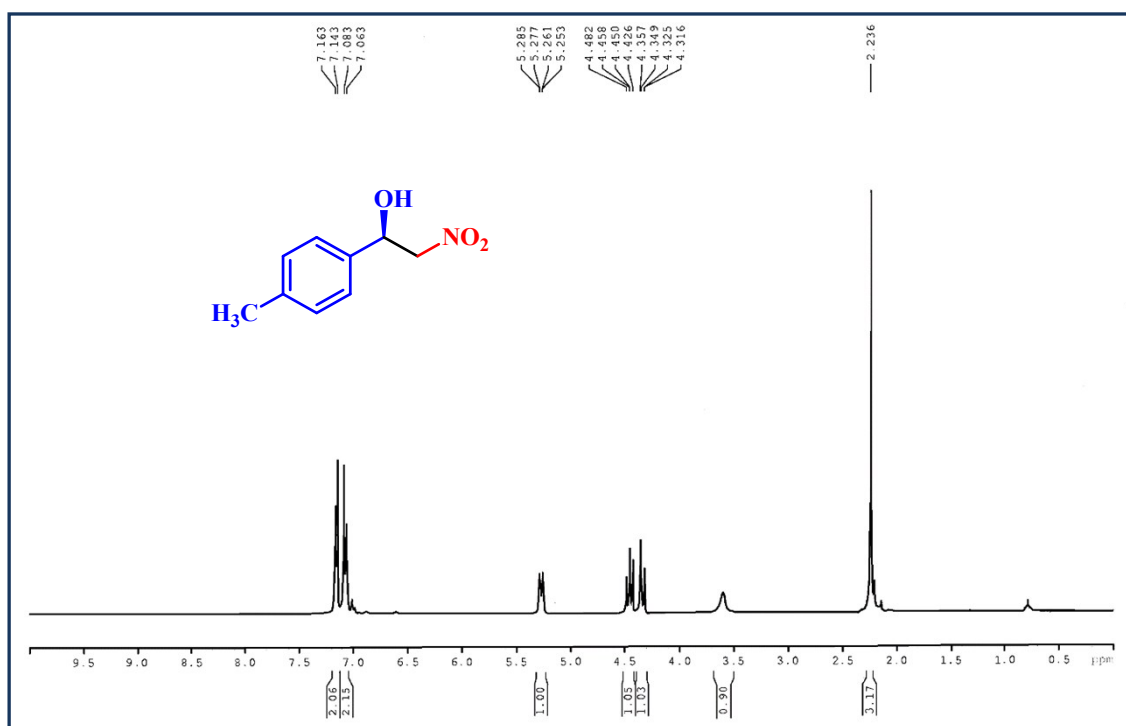
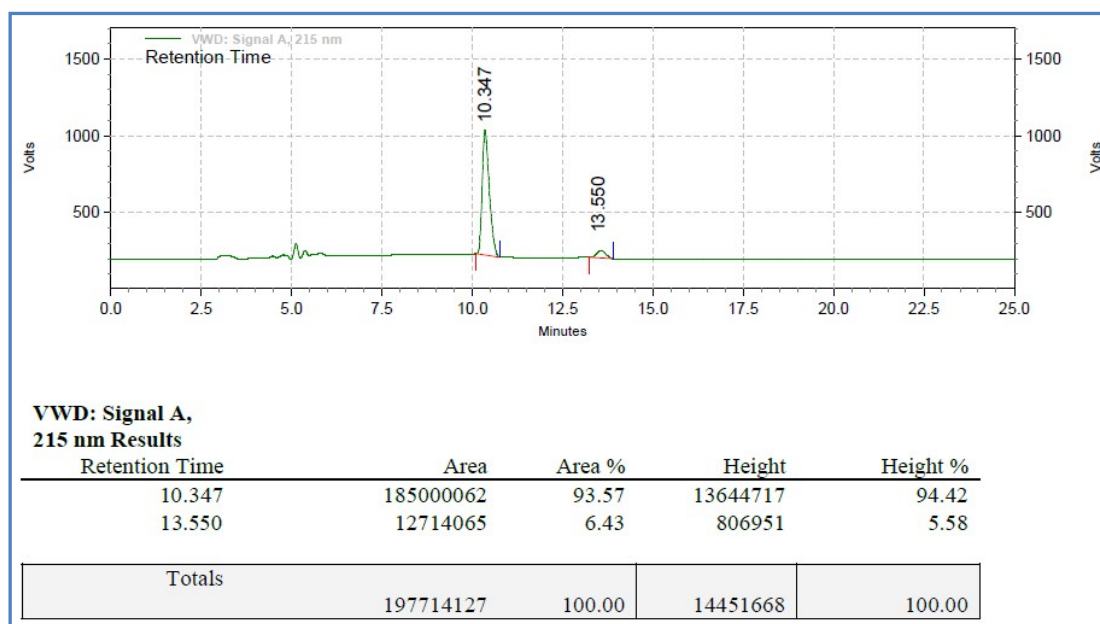


Fig. S12. HPLC chromatogram:



(R)-2-nitro-1-phenylethanol^[10](2d, Table 1)

The desired product was isolated by column chromatography over silica gel (ethyl acetate/hexane 15/85) as pale-yellow oil. $[\alpha]_D^{25} = -34.8$ (c 0.23, CH_2Cl_2), lit.^[10] $[\alpha]_D^{25} = -39.5$ (c 0.2, CH_2Cl_2); 94% ee; HPLC analysis was performed using Chiralcel OD-H column having 85/15 n-hexane/*i*-PrOH as mobile phase, flow rate 1.0 ml/min, retention time (*R*): 10.91 min (major), (*S*): 13.66 min (minor).

Fig.S13. ^1H NMR spectrum(CDCl_3 , 400 MHz):

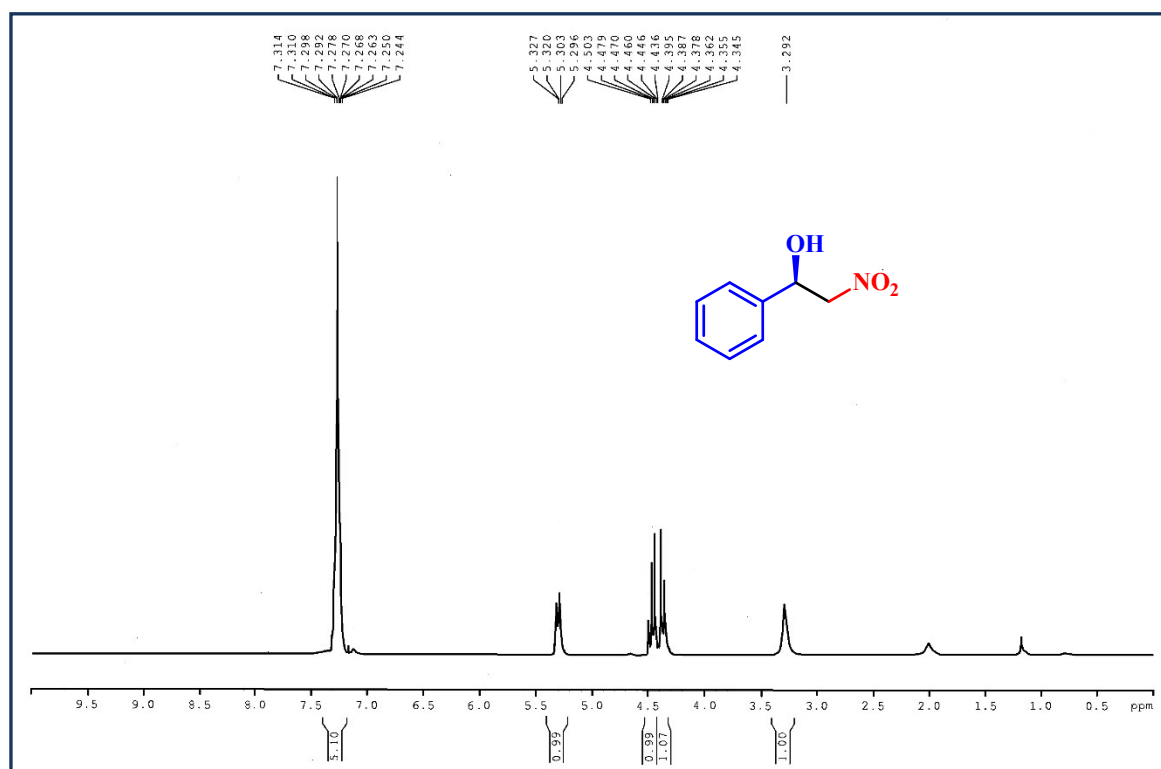
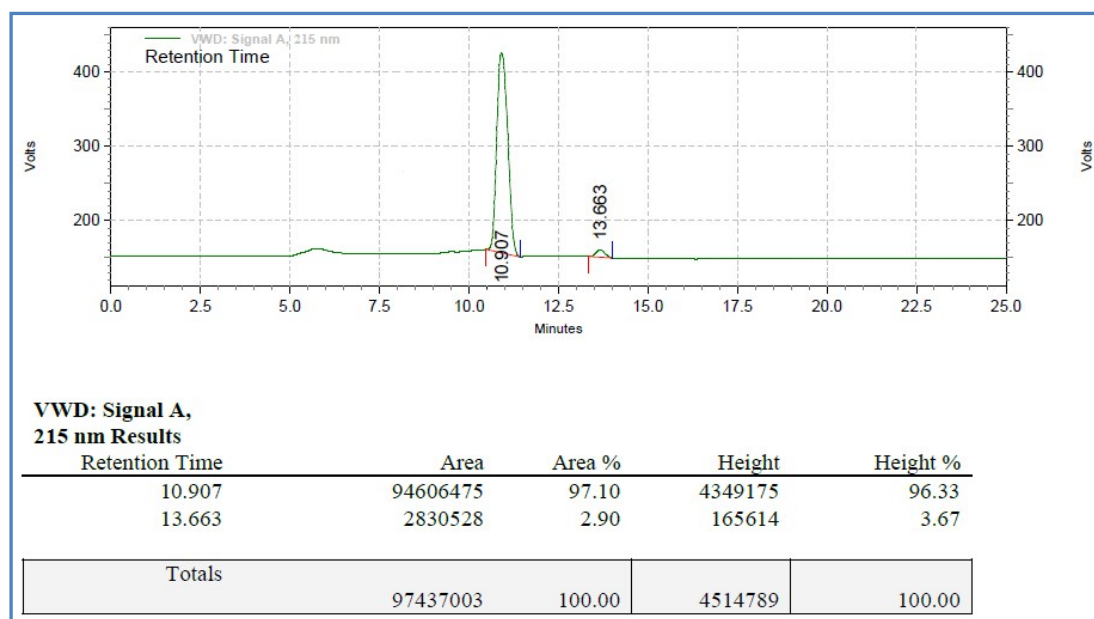


Fig. S14. HPLC chromatogram:



(R)-1-(4-methoxyphenyl)-2-nitroethanol^[11](2e, Table 1)

The desired product was isolated by column chromatography over silica gel (ethyl acetate/hexane 15/85) as yellow oil. $[\alpha]_D^{25} = -27.3$ (c 1.1, CH_2Cl_2), lit.^[12] $[\alpha]_D^{25} = -29.8$ (c 1, CH_2Cl_2); 85% ee; HPLC analysis was performed using Chiralcel OD-H column having 85/15 n-hexane/*i*-PrOH as mobile phase, flow rate 1.0 ml/min, retention time (*R*): 10.39 min (major), (*S*): 14.10 min (minor).

Fig.S15. ^1H NMR spectrum(CDCl_3 , 400 MHz):

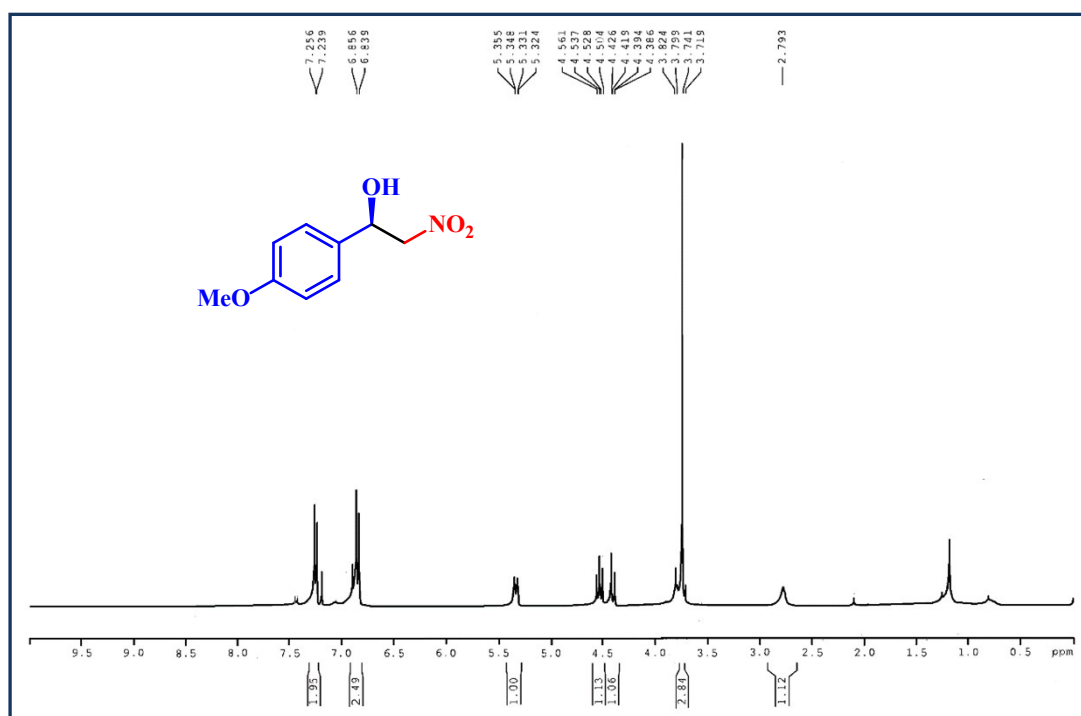
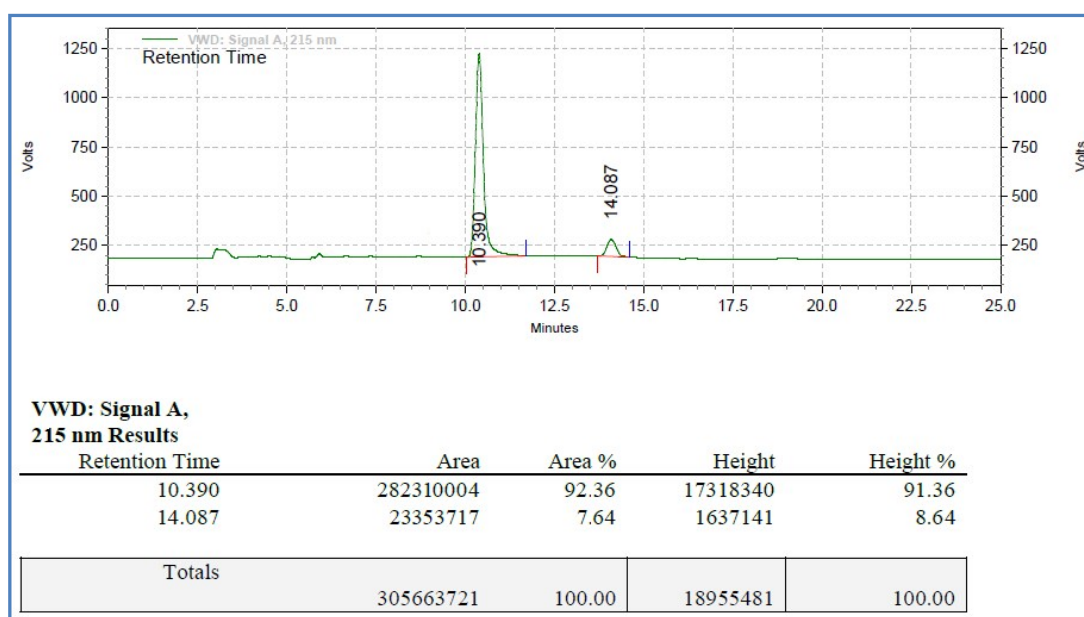


Fig. S16. HPLC chromatogram:



(R)-1-(3,4-dimethoxyphenyl)-2-nitroethanol^[1] (2f, Table 1)

The desired product was isolated by column chromatography over silica gel (ethyl acetate/hexane 15/85) as yellowish oil. $[\alpha]_D^{25} = -28.0$ (c 1.98, CH_2Cl_2), lit.^[1] $[\alpha]_D^{25} = -27.1$ (c 2.01, CH_2Cl_2); 93% ee; HPLC analysis was performed using Chiralcel OD-H column having 85/15 n-hexane/*i*-PrOH as mobile phase, flow rate 1.0 ml/min, retention time (*R*): 16.52 min (major), (*S*): 22.16 min (minor).

Fig.S17. ^1H NMR spectrum(CDCl_3 , 400 MHz):

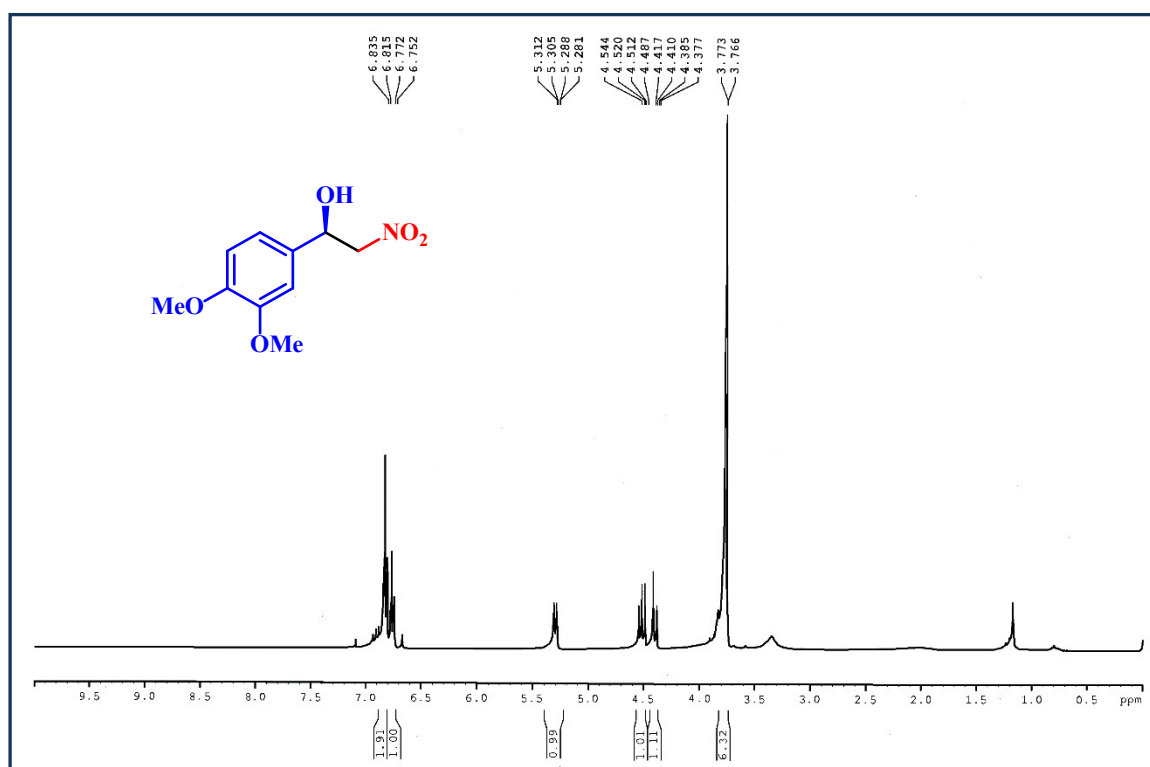
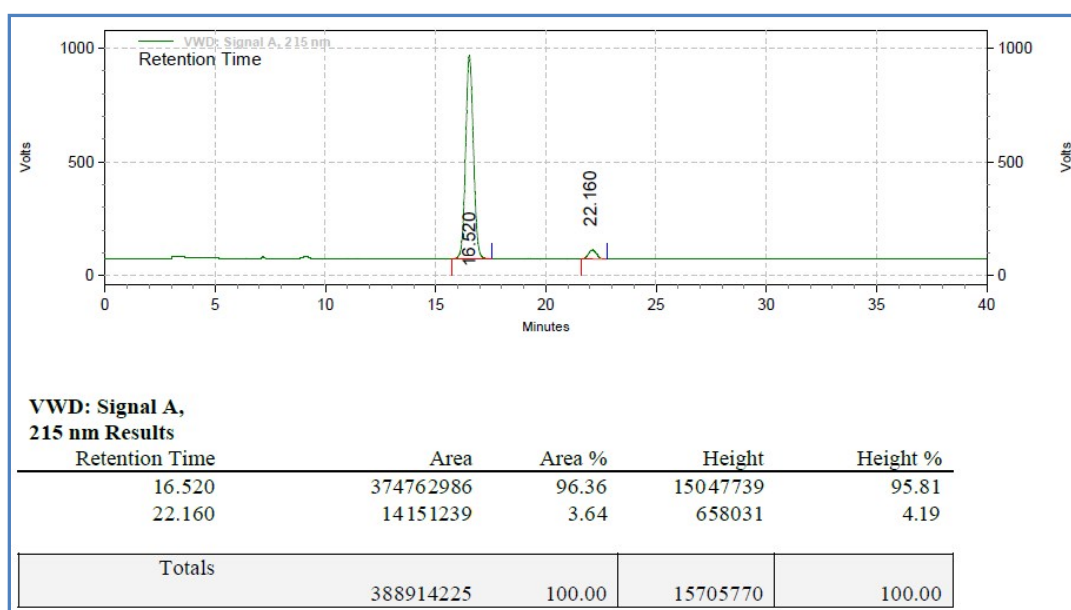


Fig. S18. HPLC chromatogram:



(R)-1-(3-chlorophenyl)-2-nitroethanol^[11] (2g, Table 1)

The desired product was isolated by column chromatography over silica gel (ethyl acetate/hexane 15/85) as yellowish oil. $[\alpha]_D^{25} = -28.6$ (c 1, CH_2Cl_2), lit.^[11] $[\alpha]_D^{25} = -27.2$ (c 1.05, CH_2Cl_2); 93% ee; HPLC analysis was performed using Chiralcel OD-H column having 85/15 n-hexane/*i*PrOH as mobile phase, flow rate 1.0 ml/min, retention time (*R*): 9.77 min (major), (*S*): 12.97 min (minor).

Fig.S19. ^1H NMR spectrum (CDCl_3 , 400 MHz):

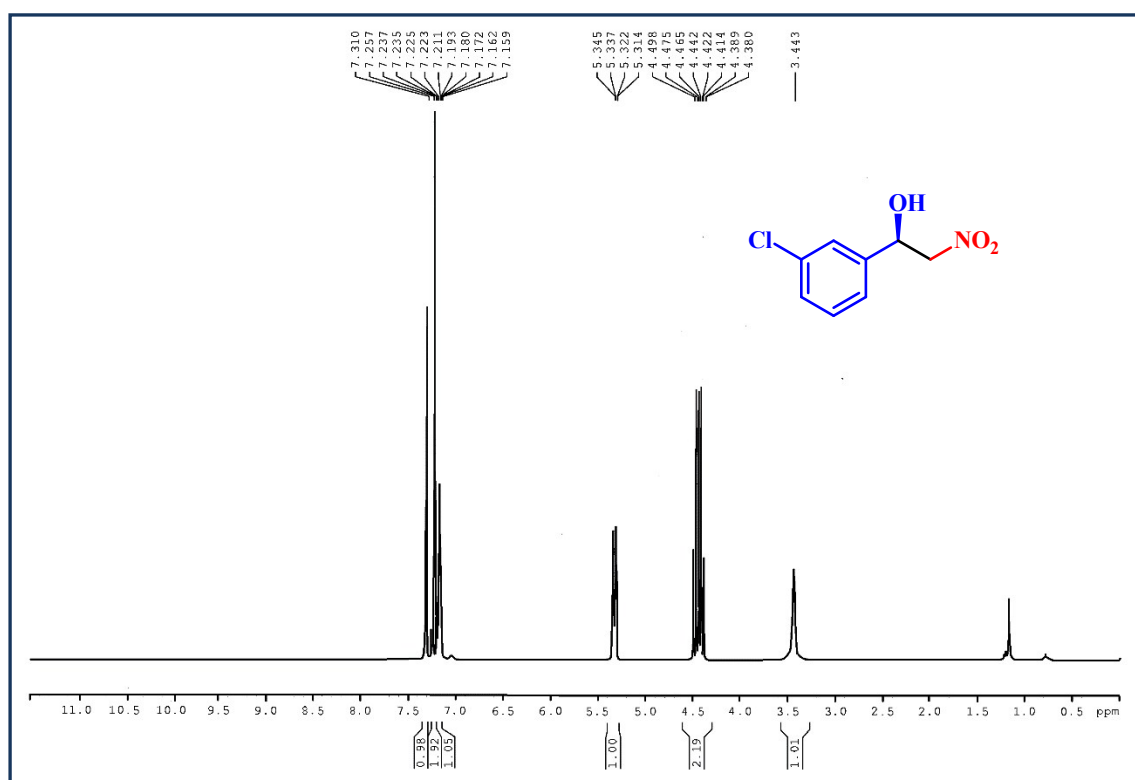
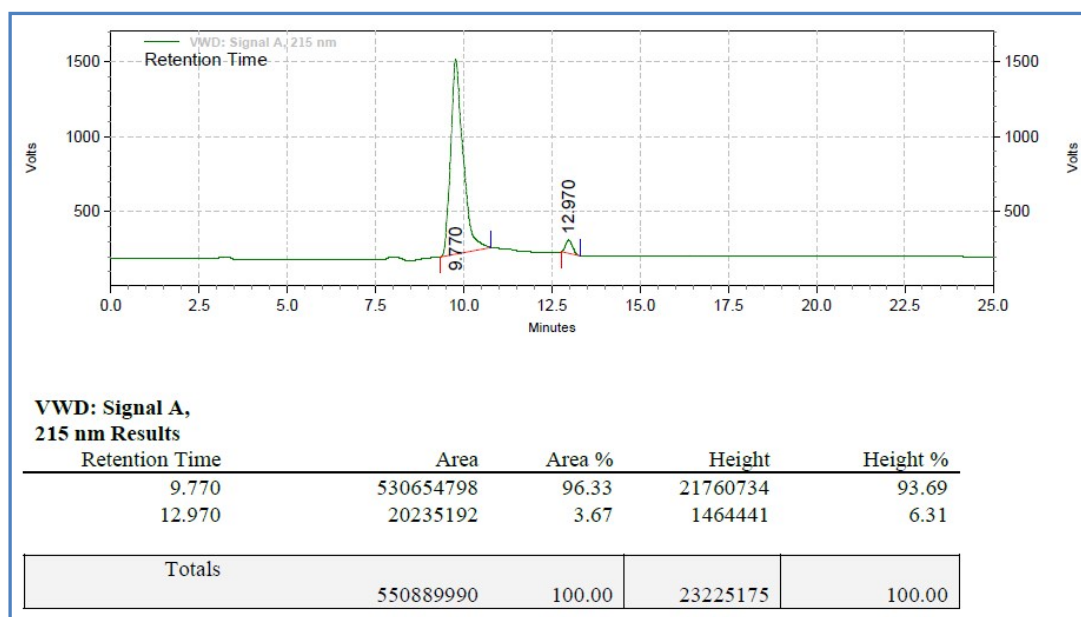


Fig. S20. HPLC chromatogram:



(S)-1-(furan-2-yl)-2-nitroethanol^[13] (2h, Table 1)

The desired product was isolated by column chromatography over silica gel (ethyl acetate/hexane 15/85) as pale-yellow oil. $[\alpha]_D^{25} = -34.9$ (c 0.26, CH_2Cl_2), lit.^[14] $[\alpha]_D^{25} = -37.1$ (c 0.24, CH_2Cl_2); 88% ee; HPLC analysis was performed using Chiralcel OD-H column having 85/15 n-hexane/*i*PrOH as mobile phase, flow rate 1.0 ml/min, retention time (S): 18.59 min (major), (R): 22.46 min (minor).

Fig.S21. ^1H NMR spectrum(CDCl_3 , 400 MHz):

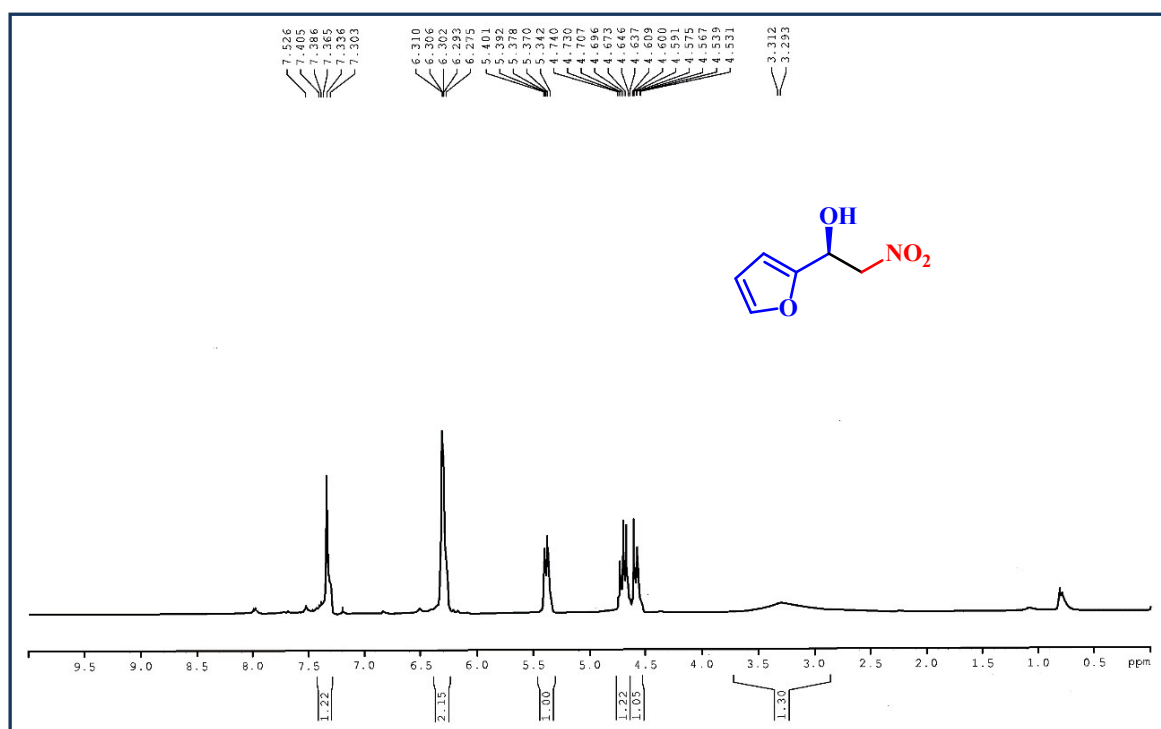
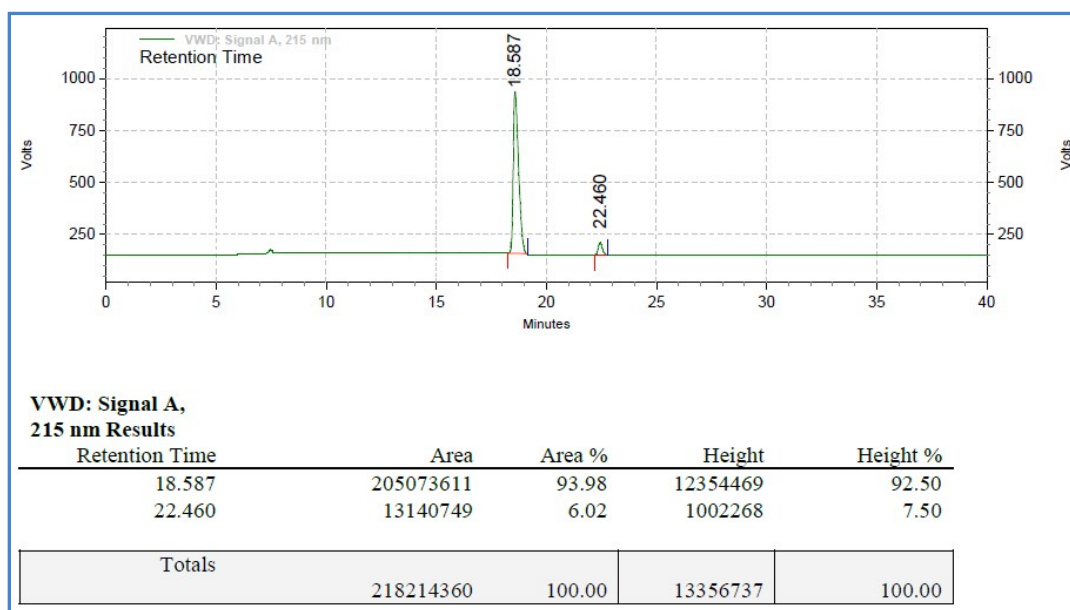


Fig. S22. HPLC chromatograms:



(S)-2-nitro-1-(thiophen-2-yl)ethanol^[13] (2i, Table 1)

The desired product was isolated by column chromatography over silica gel (ethyl acetate/hexane 15/85) as yellowish oil. $[\alpha]_D^{25} = -35.9$ (c 0.33, CH_2Cl_2), lit.^[14] $[\alpha]_D^{25} = -27.5$ (c 0.28, CH_2Cl_2); 90% ee; HPLC analysis was performed using Chiralcel OD-H column having 85/15 n-hexane/*i*PrOH as mobile phase, flow rate 1.0 ml/min, retention time (*S*): 10.91 min (major), (*R*): 13.38 min (minor).

Fig.S23. ^1H NMR spectrum(CDCl_3 , 400 MHz):

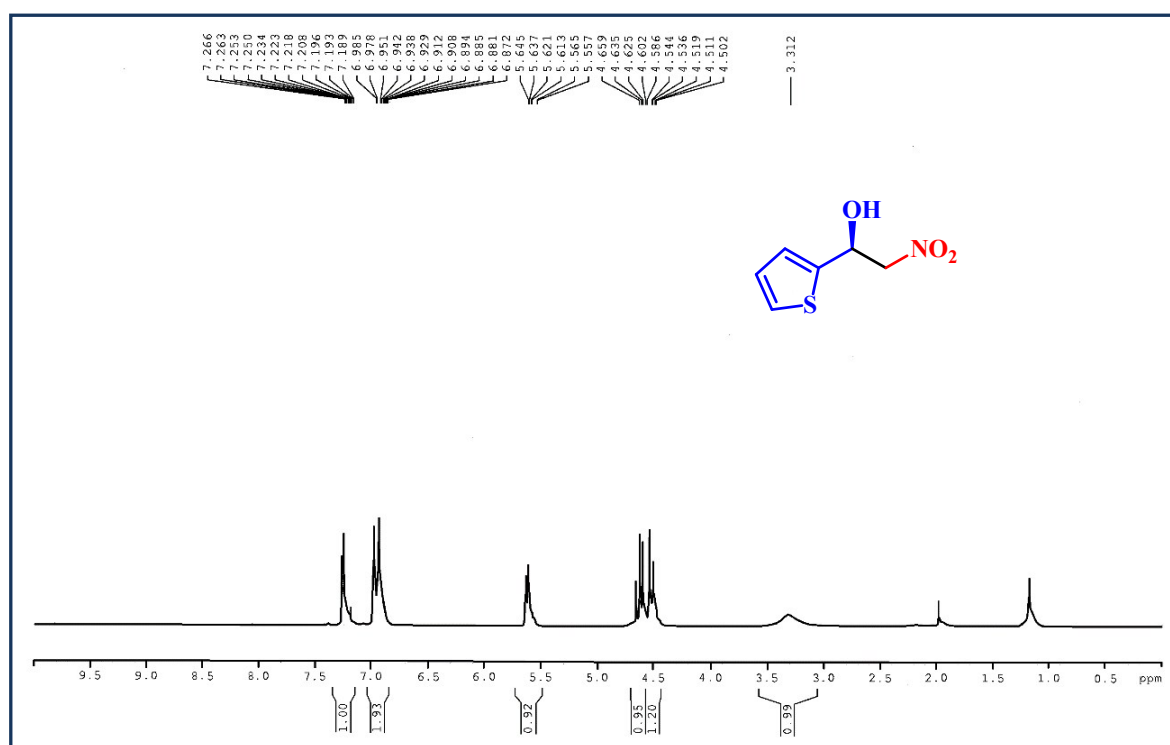
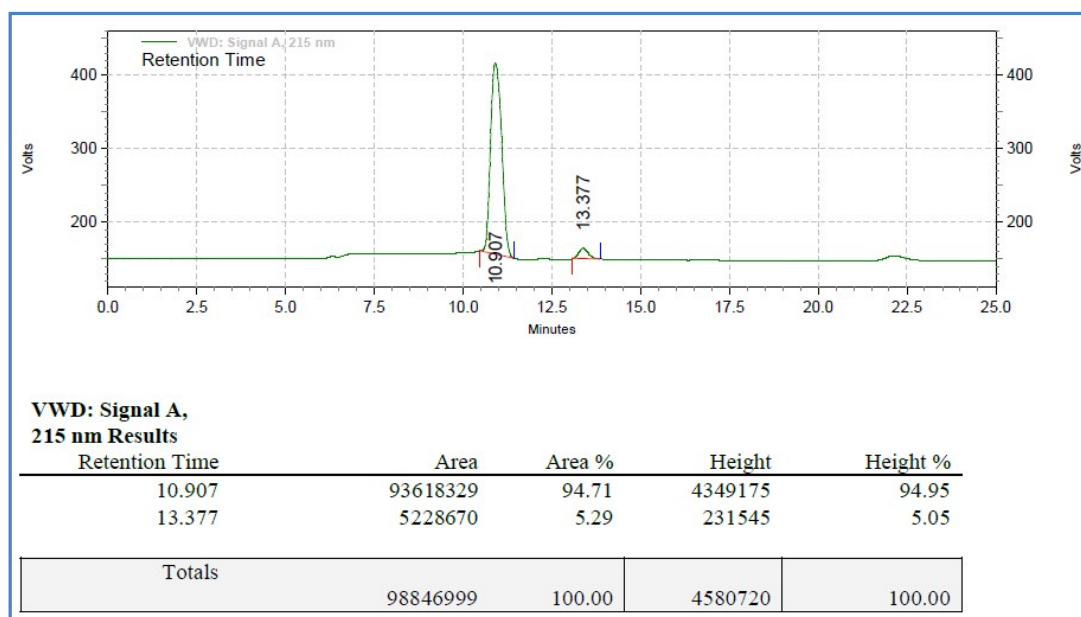


Fig. S24. HPLC chromatograms:



(R)-1-nitrooctan-2-ol^[7] (2j, Table 2)

The desired product was isolated by column chromatography (ethyl acetate/hexane 10/90) as light yellowish oil. $[\alpha]_D^{25} = -11.8$ (c 1.09, CH_2Cl_2), lit.^[15] $[\alpha]_D^{25} = -12.3$ (c 1.2, CH_2Cl_2); 81% ee; HPLC analysis was performed using Chiralcel OD-H column having 90/10 n-hexane/*i*-PrOH as mobile phase, flow rate 0.8 ml/min, retention time (*R*): 9.31 min (major), (*S*): 14.53 min (minor).

Fig. S25. ^1H NMR spectrum(CDCl_3 , 400 MHz):

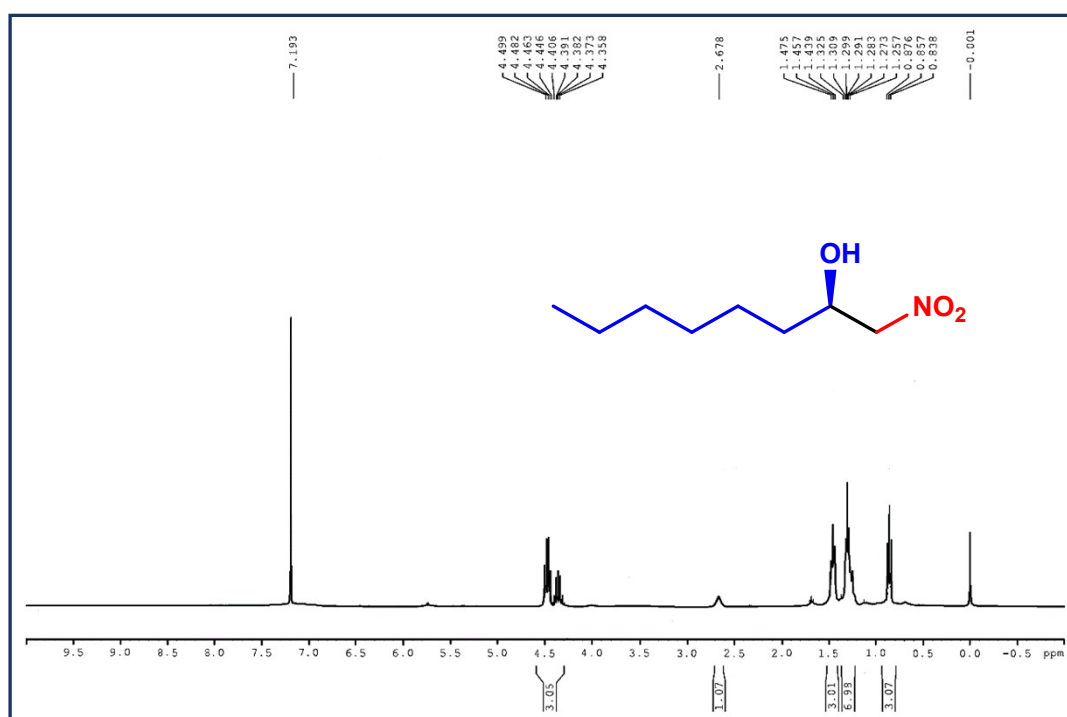
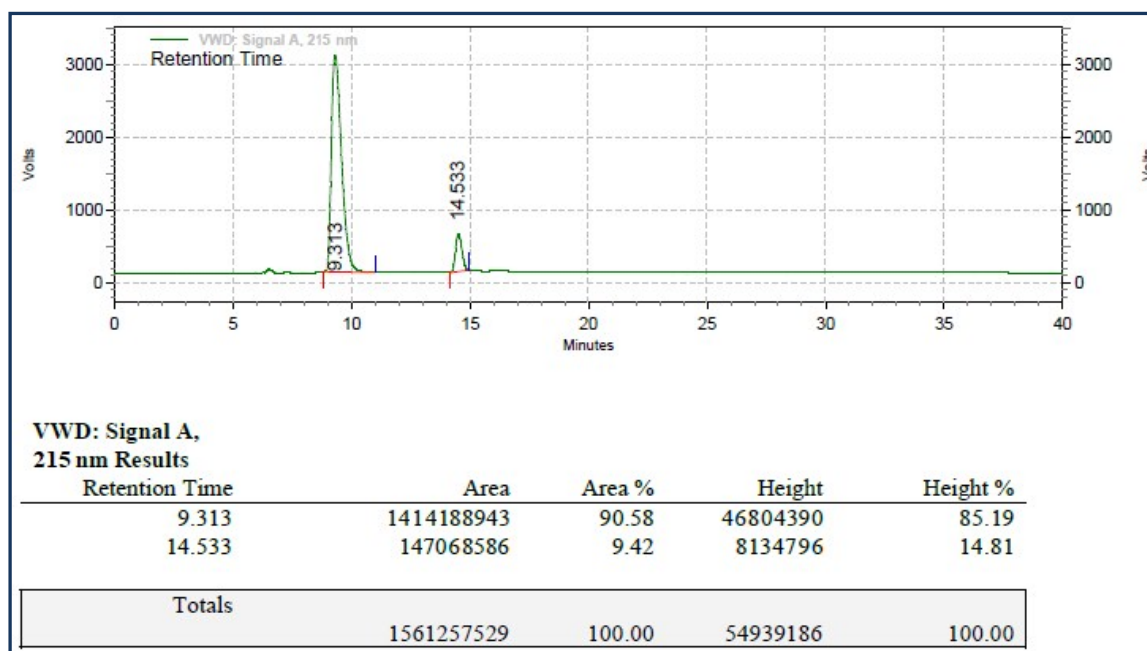


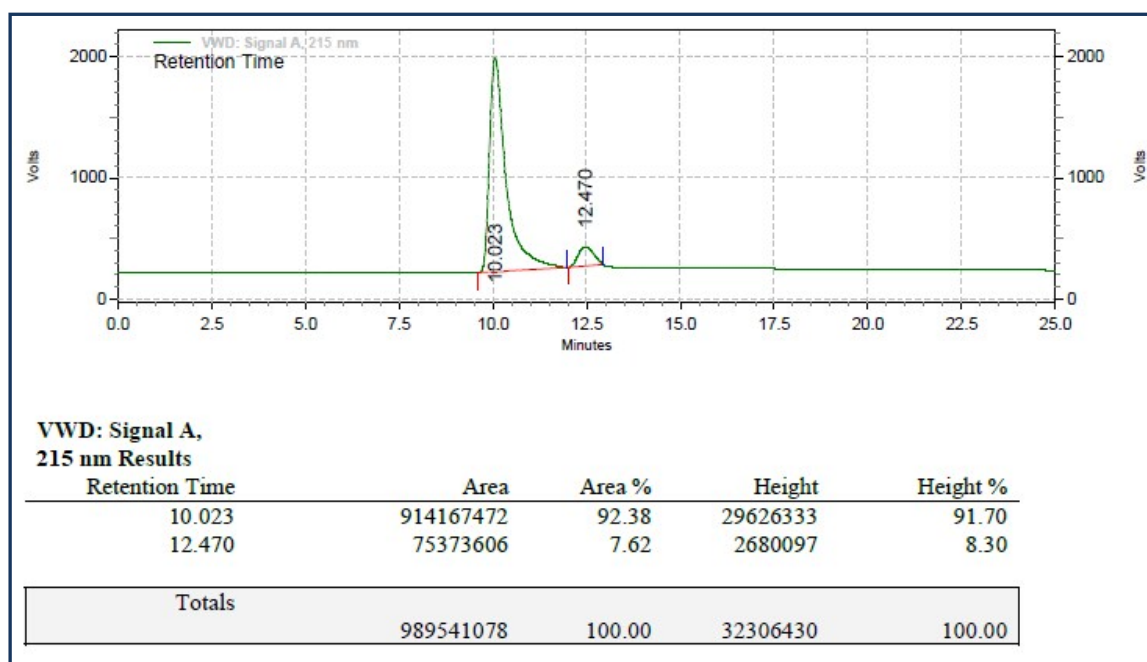
Fig. S26. HPLC chromatograms:



(R)-1-cyclohexyl-2-nitroethanol^[11] (2k, Table 2)

The desired product was isolated by column chromatography (ethyl acetate/hexane 10/90) as pale-yellow oil. $[\alpha]_D^{25} = -15.2$ (c 0.98, CHCl_3), lit.^[11] $[\alpha]_D^{25} = -14.7$ (c 1.03, CHCl_3); 85% ee; HPLC analysis was performed using Chiralcel OD-H column having 85/15 n-hexane/*i*-PrOH as mobile phase, flow rate 1.0 ml/min, retention time (*R*): 10.02 min (major), (*S*): 12.47 min (minor).

Fig. S27. HPLC chromatograms:



2-Nitro-1-(4-nitrophenyl)propan-1-ol^[5] (3a, Table 3)

The desired product was isolated by column chromatography over silica gel (ethyl acetate/hexane 15/85) as pale-yellow solid in 96% yield. Enantiomeric excess (ee_{syn} : 95%, ee_{anti} : 84%) of diastereomers were determined by HPLC analysis (Chiralcel OD-H, 85/15 n-hexane/*i*PrOH as mobile phase, flow rate 1.0 ml/min). Retention time, t_R (*anti* minor): 7.52 min, t_R (*anti* major): 9.56 min, t_R (*syn* major): 12.95 min, t_R (*syn* minor): 14.96 min. Diastereomeric ratio (*syn*/*anti* = 3.2:1) was determined by ^1H NMR. ^1H NMR (400 MHz, CDCl_3): δ = 8.29-8.24 (m, 2H), 7.61-7.58 (m, 2H), 5.56 (d, J = 2.8 Hz, 0.29H, *anti*), 5.19 (d, J = 8.4 Hz, 0.91H, *syn*), 4.79-4.70 (m, 1H, *syn* + *anti*), 3.07-3.00 (br, 1H, *syn* + *anti*), 1.50-1.47 (m, 0.86H, *anti*), 1.43-1.37 (m, 2.76H, *syn*).

Fig. S28. ^1H NMR spectrum (CDCl_3 , 400 MHz):

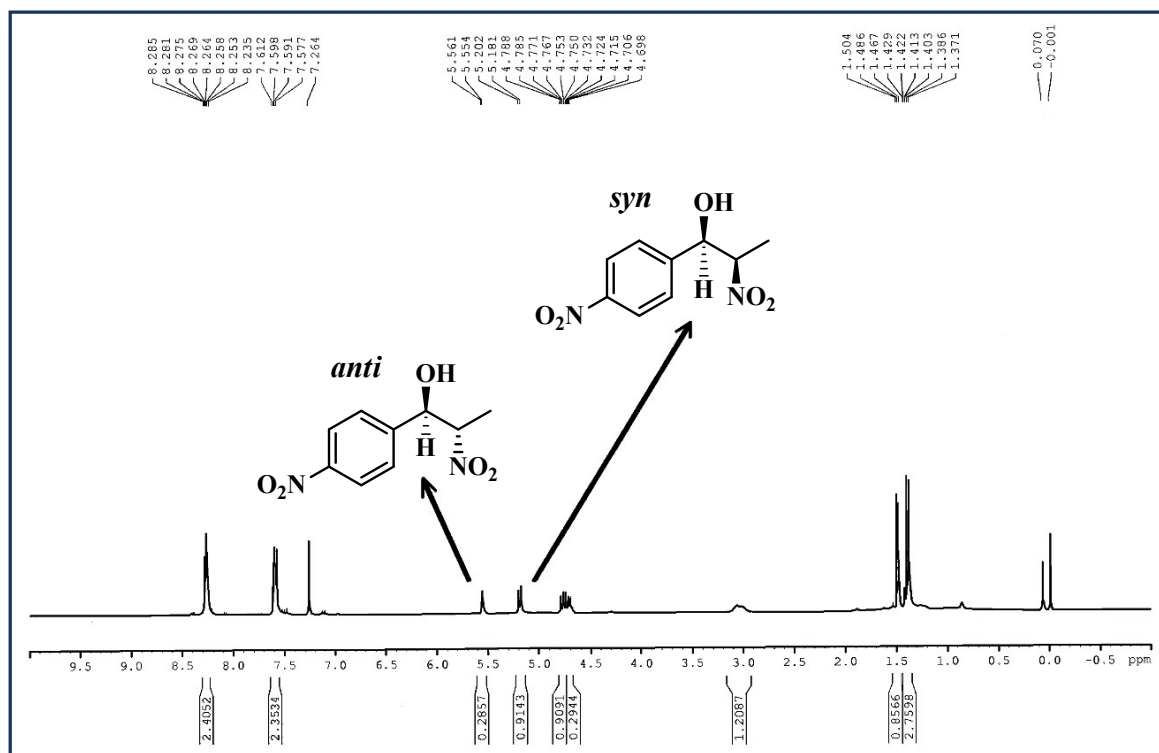
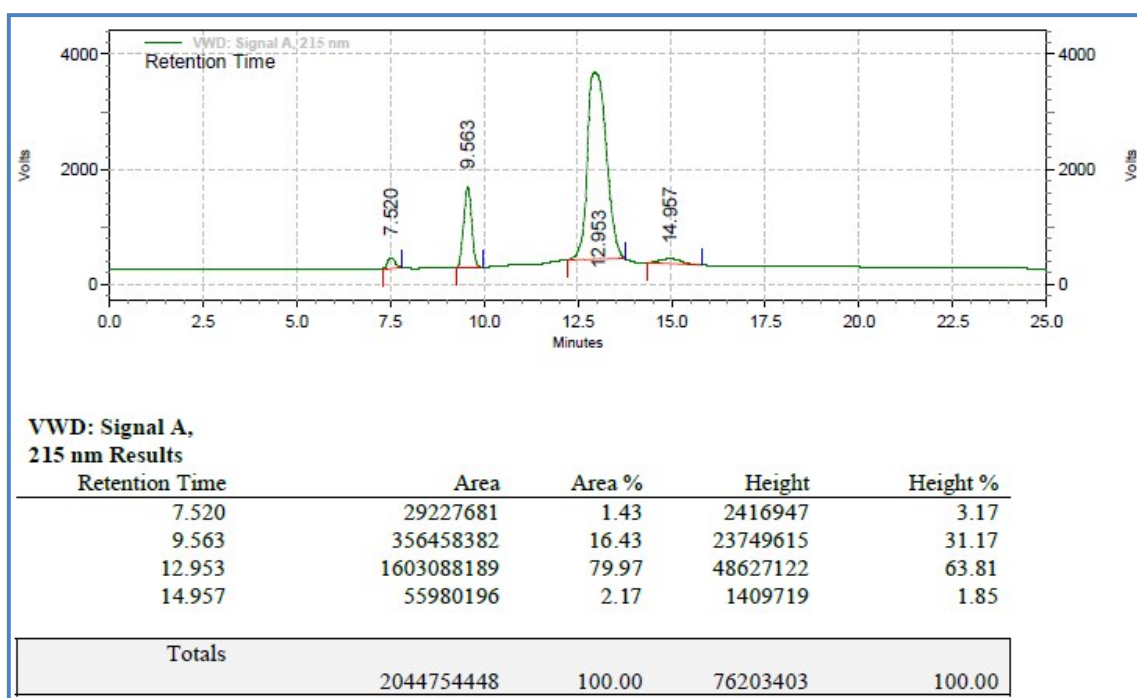


Fig. S29. HPLC chromatograms:



1-(4-chlorophenyl)-2-nitropropan-1-ol^[16] (3b, Table 3)

The desired product was isolated by column chromatography over silica gel (ethyl acetate/hexane 15/85) as pale-yellow solid in 95% yield. Enantiomeric excess (ee_{syn} : 95%, ee_{anti} : 66%) of diastereomers were determined by HPLC analysis (Chiralcel OD-H, 85/15 n-hexane/*i*PrOH as mobile phase, flow rate 1.0 ml/min). Retention time, t_R (*anti* minor): 4.54min, t_R (*anti* major): 5.54 min, t_R (*syn* major): 8.03 min, t_R (*syn* minor): 9.26 min. Diastereomeric ratio (*syn/anti*, 3:1) was determined by ^1H NMR. ^1H NMR (400 MHz, CDCl_3): δ = 7.30-7.20 (m, 4H), 5.28 (d, J = 3.2 Hz, 0.29H, *anti*), 4.92 (d, J = 8.8 Hz, 0.88H, *syn*), 4.66-4.62 (m, 0.88H, *syn*), 4.59-4.55 (m, 0.29H, *anti*), 3.10 (br, 1H, *syn* + *anti*), 1.51-1.47 (m, 0.88H, *anti*), 1.40-1.39 (m, 2.63H, *syn*).

Fig. S30. ^1H NMR spectrum (CDCl_3 , 400 MHz):

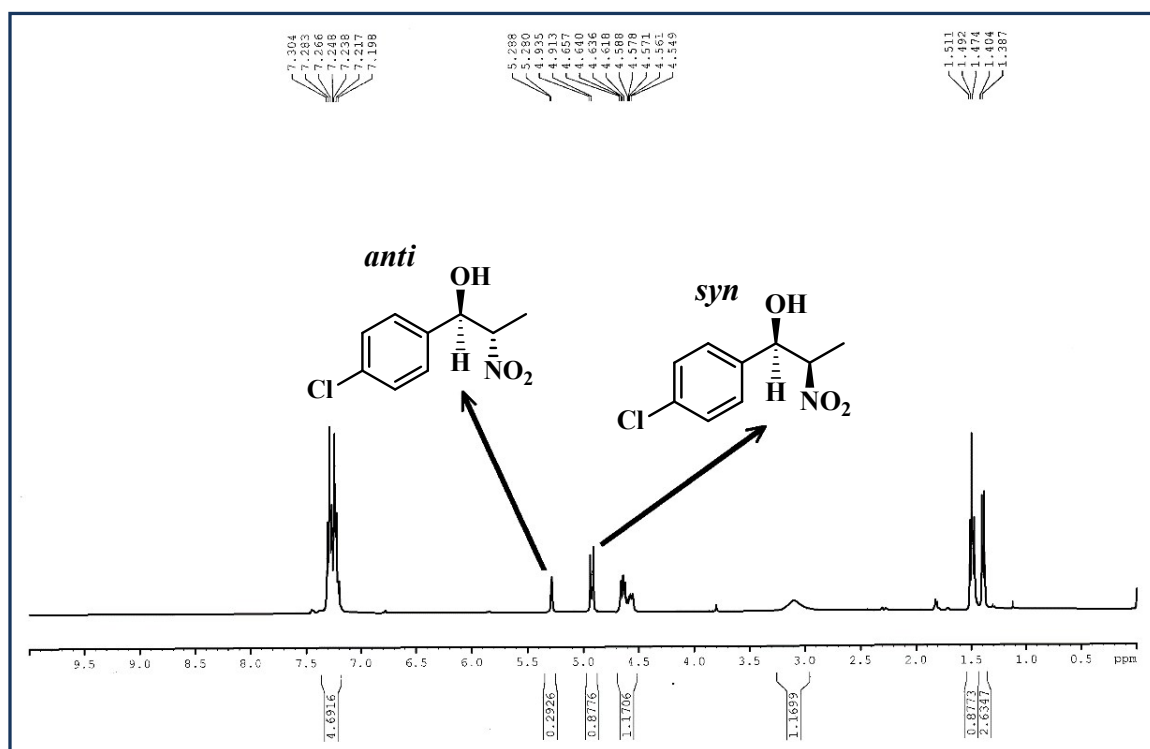
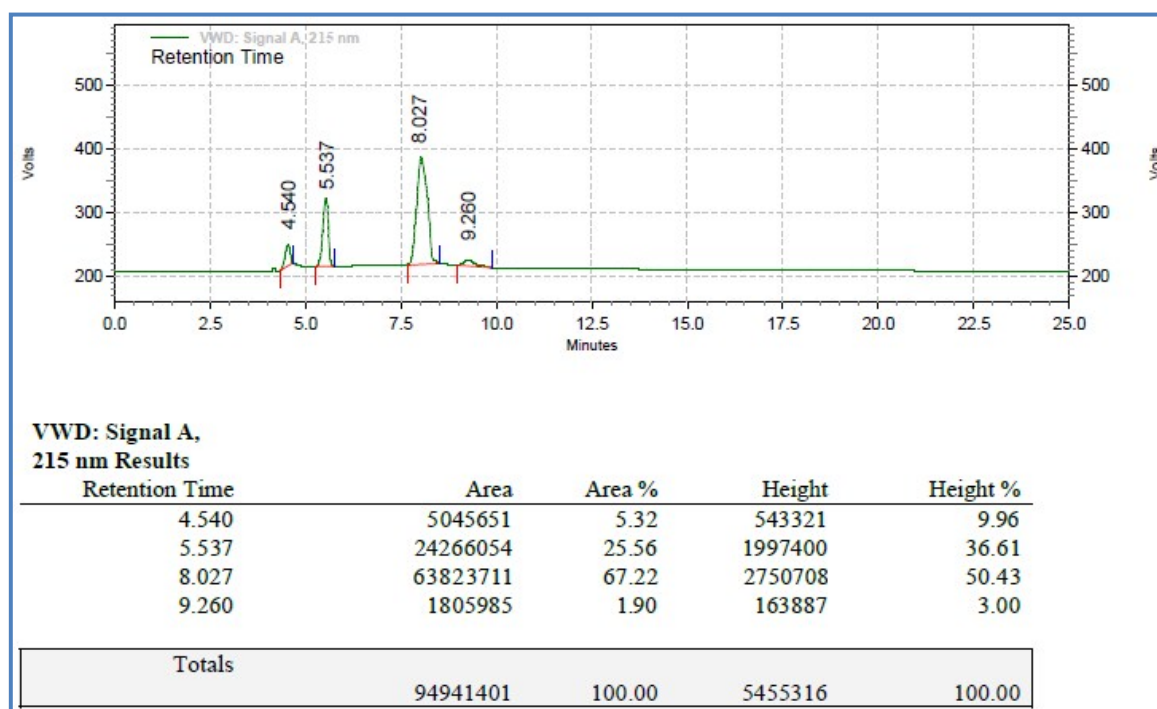


Fig. S31. HPLC chromatograms:



2. FT-IR SPECTRUM OF THE REUSED Cu(II)@AFS-1 CATALYST:

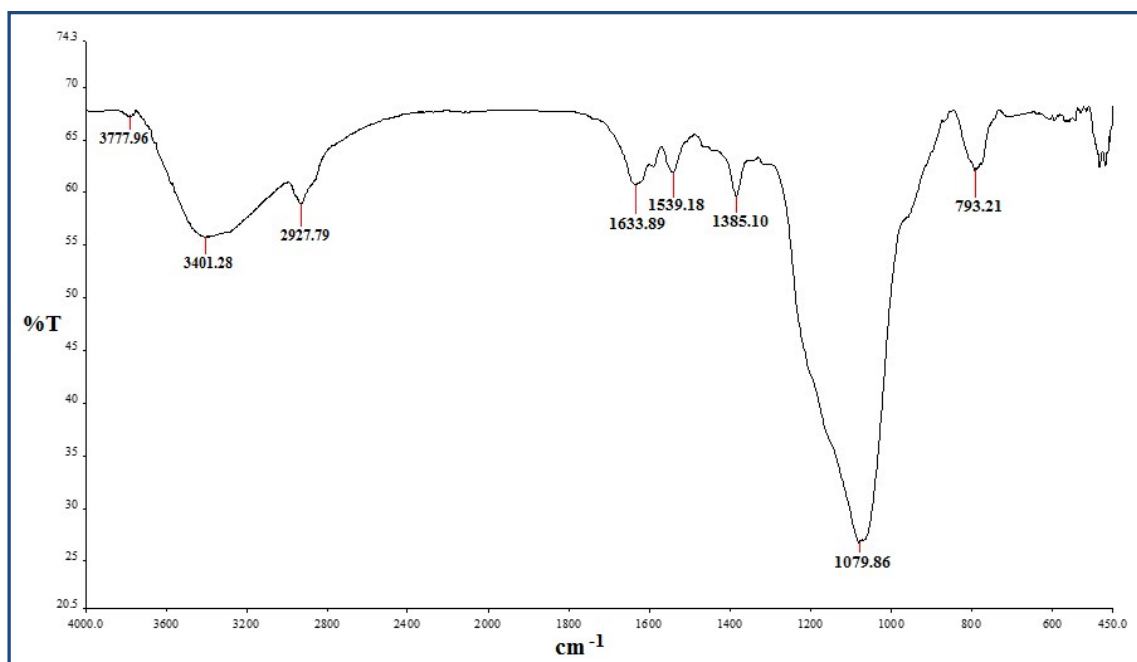


Fig. S32. FT-IR spectrum of the reused Cu(II)@AFS-1 catalyst.

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